

Supporting Information

Ir/XuPhos-Catalyzed Direct Asymmetric Reductive Amination of Ketones with Secondary Amines

Zhou Luo,^{[a]#} Tianxiang Fan,^{[a]#} Jingyan Luo,^[a] Yuanyuan Liu,^{*[a]} and Junliang Zhang^{*[b]}

^[a]Shanghai Key Laboratory of Green Chemistry and Chemical Processes, School of Chemistry and Molecular Engineering, East China Normal University, Shanghai 200062, China.

^[b]Department of Chemistry, Fudan University, 2005 Songhu Road, Shanghai, 200438, China.

E-mail : yyluo@chem.ecnu.edu.cn

junliangzhang@fudan.edu.cn

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1. General Information:

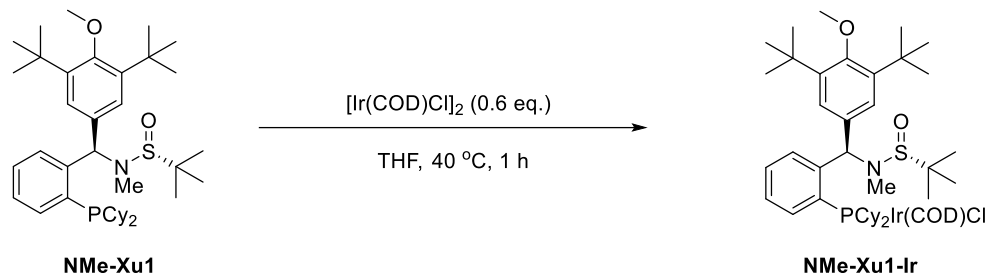
Unless otherwise noted, Materials obtained from commercial suppliers were used directly without further purification. ^1H NMR spectra were recorded on a BRUKER 500 (or 600) MHz spectrometer in CDCl_3 . Chemical shifts are reported in ppm with tetramethylsilane (TMS: 0 ppm) with the solvent resonance as the internal standard. Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = doublet of doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz), and integration. ^{13}C NMR spectra were recorded on a BRUKER 500 (125 MHz) or 600 (150 MHz) spectrometer in CDCl_3 with complete proton decoupling. Chemical shifts are reported in δ units, parts per million (ppm), and were referenced to CDCl_3 (δ 7.26 or 77.0 ppm) as the internal standard. All products were further characterized by HRMS (high resolution mass spectra). The enantiomeric excesses of the products were determined by chiral stationary phase HPLC using a Chiralpak OJ-H, OD-H, AD-H, AS-H, or determined by ^1H NMR using D-(-)-Mandelic acid as chemical shift reagent..

Anhydrous tetrahydrofuran (THF), toluene, 1,4-Dioxane and diethyl ether (Et_2O) were distilled from sodium and benzophenone to use; Anhydrous dichloromethane (DCM) and 1,2-dichloroethane (DCE) were distilled from CaH_2 . Unless otherwise noted, analytical grade solvents and commercially available reagents were used directly.

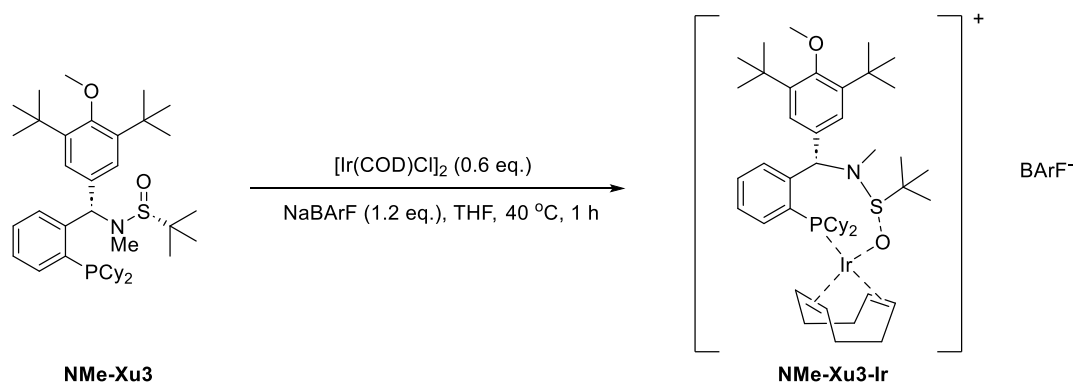
Reactions were monitored by thin layer chromatography (TLC) using silicycle pre-coated silica gel plates. Flash column chromatography was performed on silica gel 60 (particle size 200-400 mesh ASTM, purchased from Yantai, China) and eluted with petroleum ether/ethyl acetate.

2. Experimental Procedure and Characterization Data

2.1 Typical procedure for the synthesis of Ir catalyst.

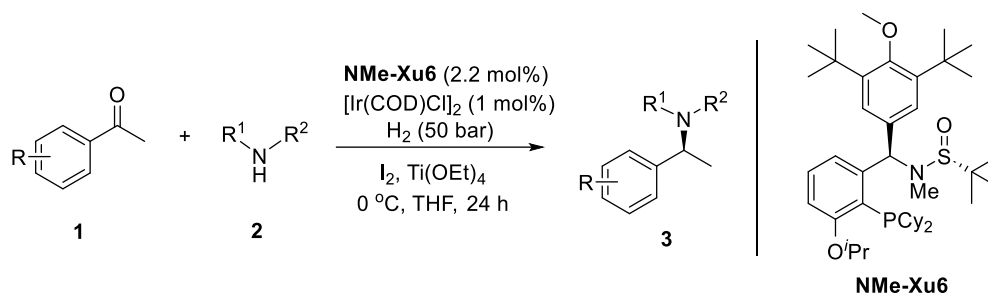


To an oven-dried Schlenk tube was added **NMe-XuPhos**^[1] (1 mmol) and $[\text{Ir}(\text{COD})\text{Cl}]_2$ (0.6 mmol). After vacuuming and refilled with nitrogen for three times, anhydrous THF (5 mL) was added under nitrogen atmosphere. The Schlenk tube was transferred to a heating pot and heated at 40 °C for 1 hour. Upon completion of the reaction, the solution was concentrated under reduced pressure. The crude mixture was then purified by column chromatography on silica gel with PE : EA = 5:1 as eluent to afford the **NMe-Xu1-Ir** as a yellow solid.



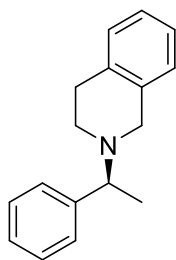
To an oven-dried Schlenk tube was added **NMe-XuPhos**^[1] (1 mmol), NaBARF (1.2 mmol) and $[\text{Ir}(\text{COD})\text{Cl}]_2$ (0.6 mmol). After vacuuming and refilled with nitrogen for three times, anhydrous THF (5 mL) was added under nitrogen atmosphere. The Schlenk tube was transferred to a heating pot and heated at 40 °C for 1 hour. Upon completion of the reaction, the solution was concentrated under reduced pressure. The crude mixture was then purified by column chromatography on silica gel with PE: DCM = 1:1 as eluent to afford the **NMe-Xu3-Ir** as a light yellow solid.

2.2 General procedure for the preparation of the chiral products.



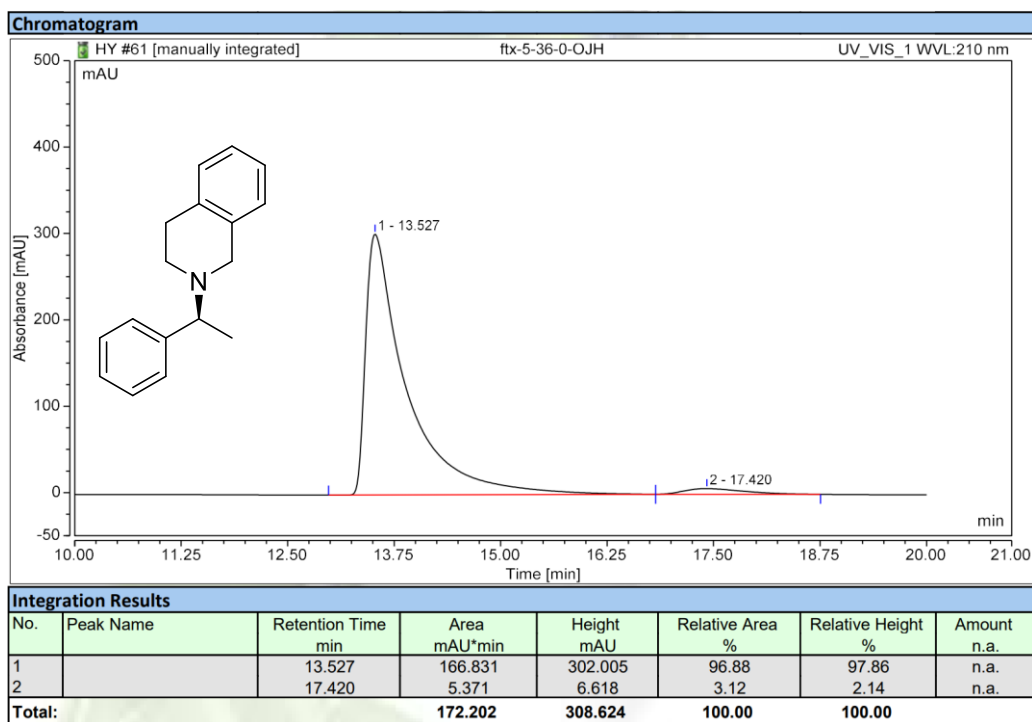
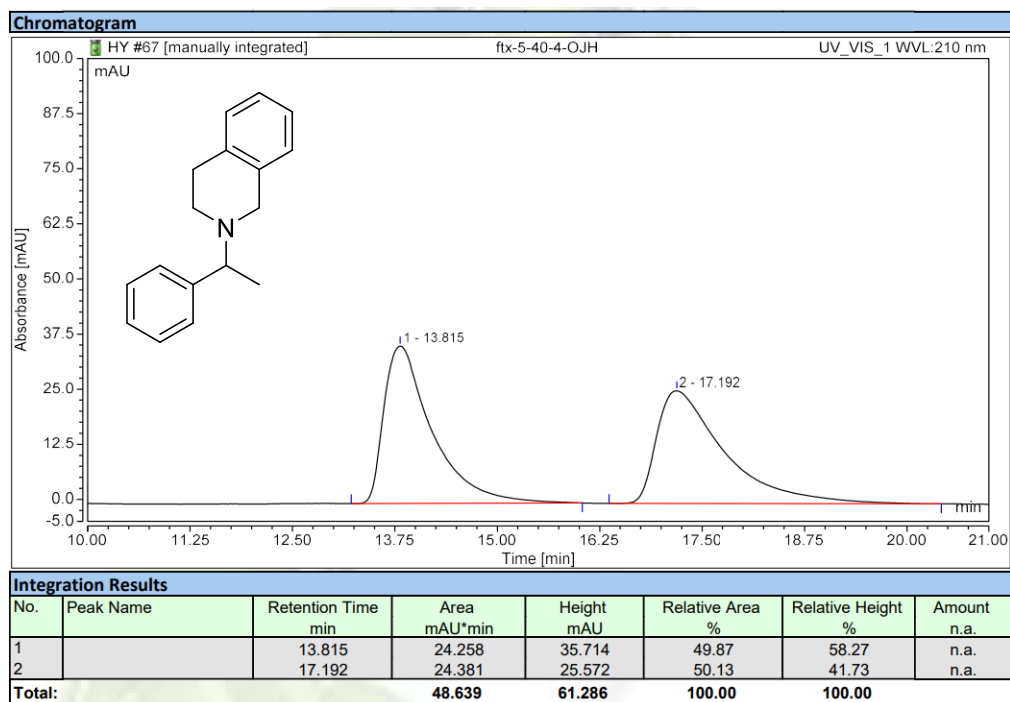
To a vial containing a magnetic stirring bar was added **1** (0.2 mmol), **2** (0.22 mmol), I_2 (2 mol%), $\text{Ti}(\text{OEt})_4$ (0.4 mmol), $[\text{Ir}(\text{COD})(\text{Cl})]_2$ (1 mol %), and **N-Me-Xu6** (2.2 mol %) under nitrogen atmosphere, anhydrous THF (2 mL) was added and stirred for 30 minutes. Then the vial was transferred in Parr steel autoclave, which was purged three times with hydrogen and finally pressurized to 50 atm. The reaction mixture was stirred at $0\text{ }^\circ\text{C}$ for 24 h. The hydrogen gas was released slowly and the solution was quenched with $\text{Na}_2\text{SO}_4 \cdot 10\text{H}_2\text{O}$ and filtered. The organic phase was concentrated and purified by column chromatography and then analyzed by chiral HPLC or using ^1H NMR with chemical shift reagent to determine the enantiomeric excesses.

(*S*)-2-(1-phenylethyl)-1,2,3,4-tetrahydroisoquinoline (**3a**).^[2]

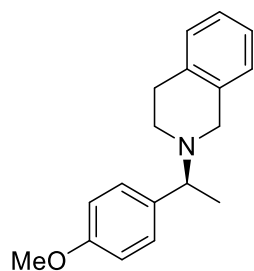


PE:EA = 20:1; Yellow oil, 43.6 mg (92% yield), 94% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.41 – 7.32 (m, 2H), 7.10 – 7.05 (m, 3H), 7.01 – 6.96 (m, 1H), 6.88 (d, J = 8.5 Hz, 2H), 3.57 – 3.52 (m, 2H), 2.92 – 2.72 (m, 3H), 2.62 – 2.55 (m, 1H), 1.41 (d, J = 6.6 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 158.58, 135.15, 134.26, 130.68, 128.65, 128.32, 126.11, 125.32, 125.11, 113.68, 73.60, 58.20, 53.38, 28.21, 20.09. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, t_{R} = 9.8 min (major), t_{R} = 13.6 min (minor). $[\alpha]_{\text{D}}^{20}$ = +3.3 (c 0.7, CHCl_3). The absolute

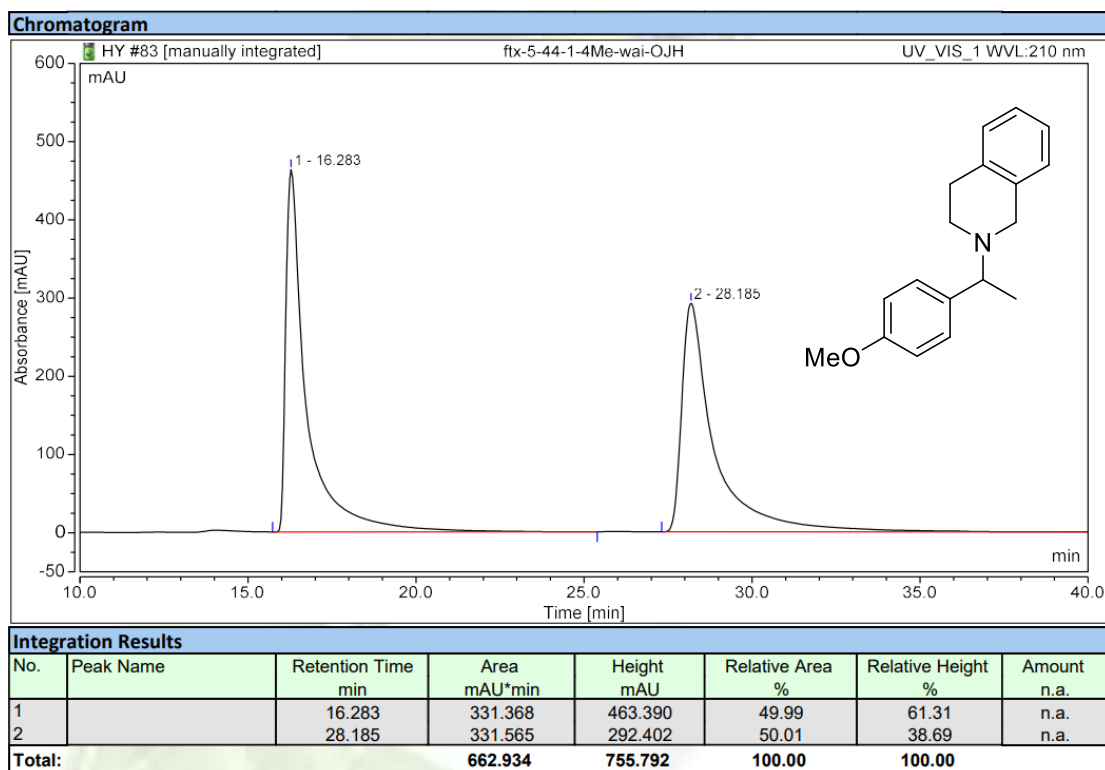
configuration was determined by comparison of the optical rotation value with the data in reference [3] ($[\alpha]_D^{20} = +9.2$ (c 0.5, CHCl_3)). The absolute configuration of other hydrogenation products was assigned by analogy to that of **3a**.

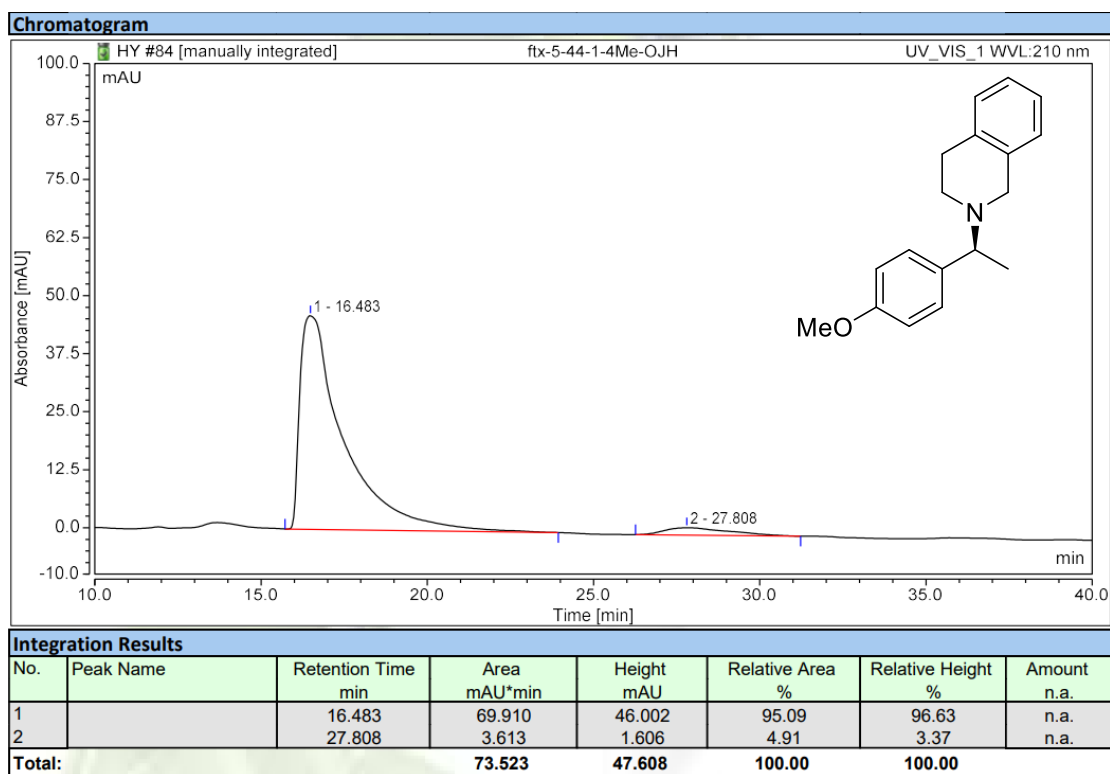


(S)-2-(1-(4-methoxyphenyl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3b). [4]

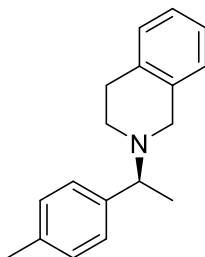


PE:EA = 5:1; Yellow oil, 50.8 mg (95% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.30 (m, 2H), 7.16 – 7.07 (m, 3H), 7.04 – 7.00 (m, 1H), 6.90 (d, J = 8.6 Hz, 2H), 3.84 (s, 3H), 3.62 – 3.52 (m, 2H), 2.94 – 2.77 (m, 3H), 2.66 – 2.60 (m, 1H), 1.48 (d, J = 6.6 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 158.50, 135.18, 134.59, 130.58, 128.59, 128.57, 126.77, 125.94, 125.47, 113.57, 63.62, 55.22, 53.49, 47.85, 29.29, 20.03. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, t_R = 7.5 min (major), t_R = 9.8 min (minor).

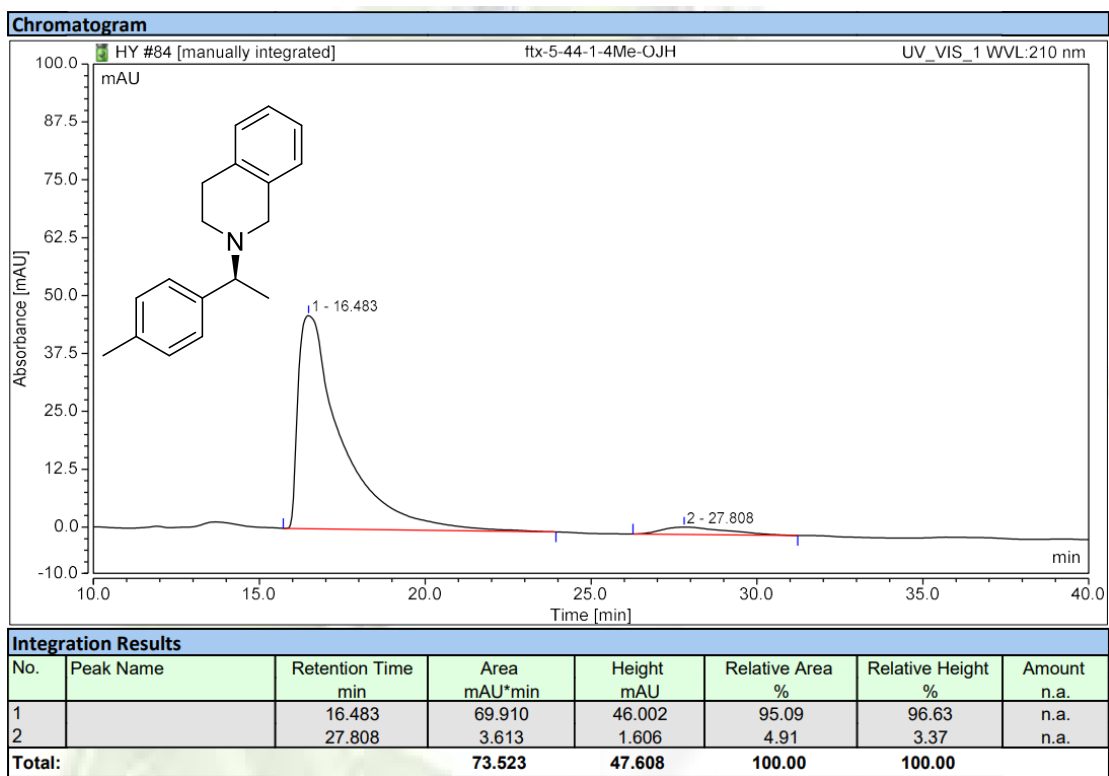
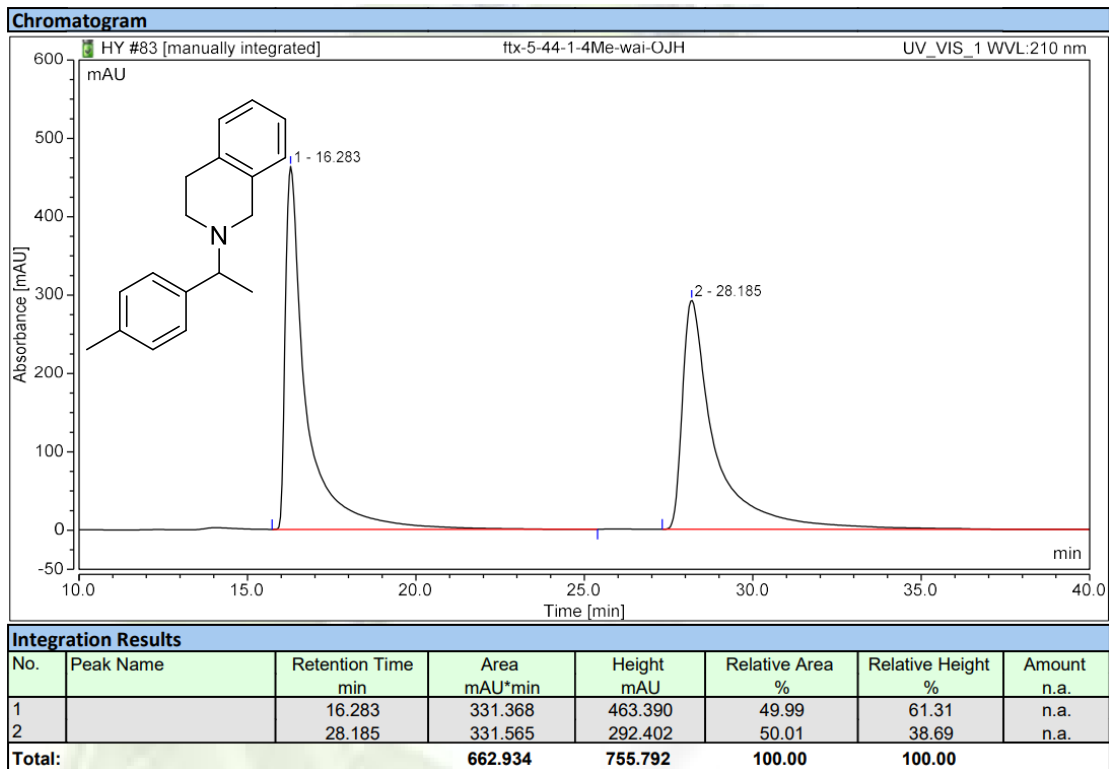




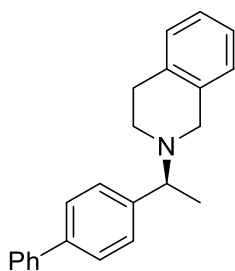
(S)-2-(1-(*p*-tolyl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3c).



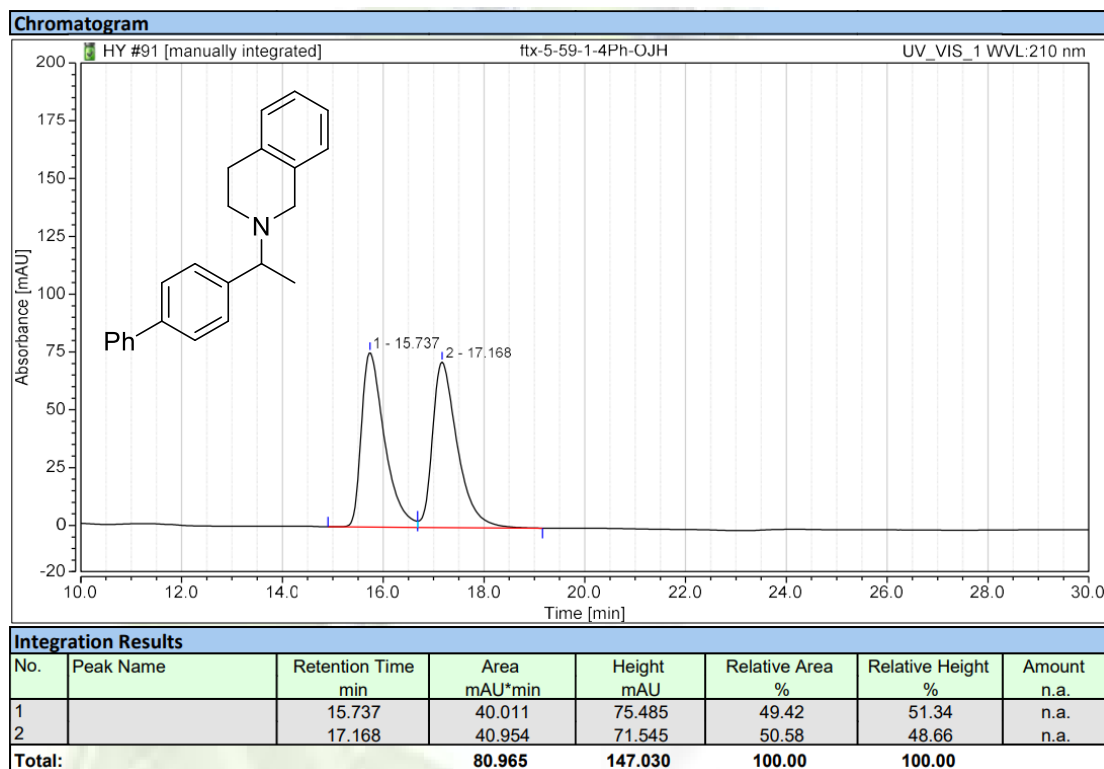
PE:EA = 30:1; Yellow oil, 46.7 mg (93% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.34 – 7.31 (m, 2H), 7.16 – 7.08 (m, 3H), 7.02 – 6.98 (m, 1H), 6.90 (d, J = 8.5 Hz, 2H), 3.79 (s, 3H), 3.59 – 3.51 (m, 2H), 2.90 – 2.69 (m, 3H), 2.65 – 2.58 (m, 1H), 1.40 (d, J = 6.6 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 158.51, 135.20, 134.49, 130.50, 128.57, 128.51, 126.75, 125.89, 125.40, 113.57, 63.60, 55.20, 53.38, 47.71, 29.21, 20.09. HPLC: Chiralpak OJ-H, hexane : isopropanol = 99:1, 0.5 mL/min, 210 nm, t_{R} = 16.5 min (major), t_{R} = 27.8 min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{18}\text{H}_{21}\text{NNa}$: 274.1572, found 274.1568.

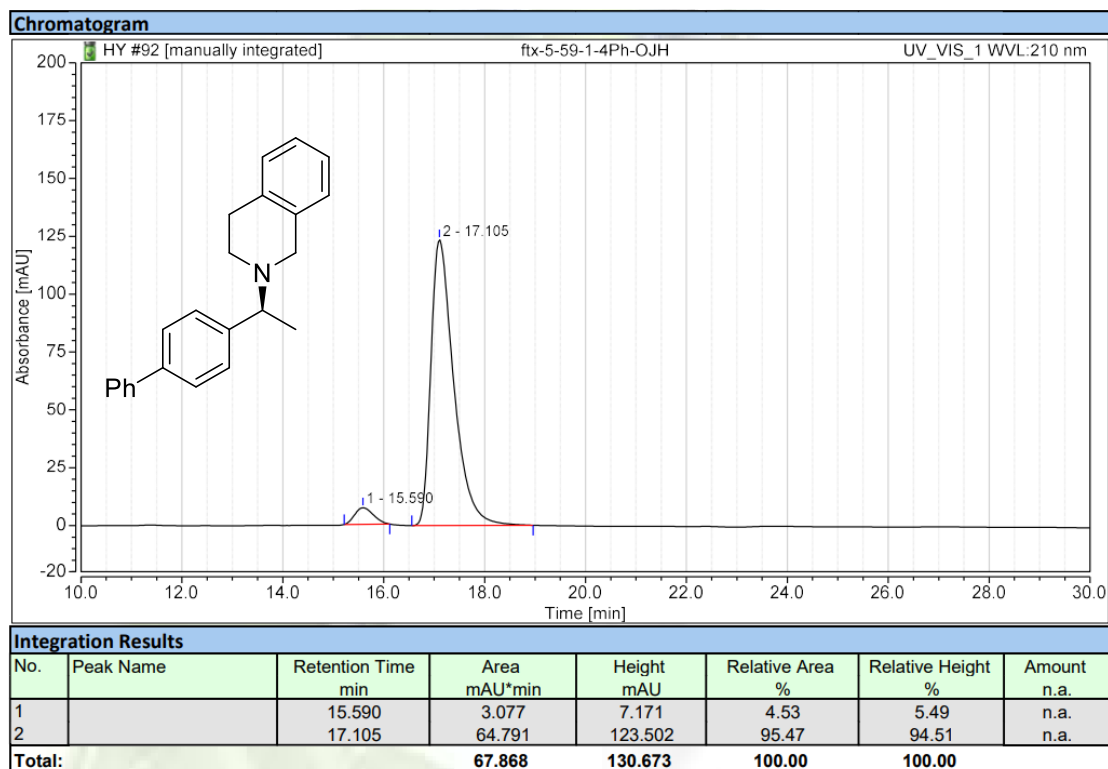


(S)-2-(1-([1,1'-biphenyl]-4-yl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3d).

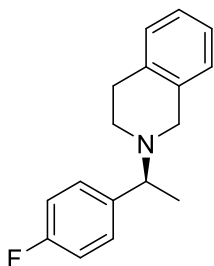


PE:EA = 20:1; Yellow oil, 50.8 mg (90% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.75 – 7.60 (m, 4H), 7.59 – 7.45 (m, 4H), 7.45 – 7.37 (m, 1H), 7.23 – 7.13 (m, 3H), 7.13 – 7.05 (m, 1H), 3.94 (d, $J = 14.8$ Hz, 1H), 3.76 – 3.64 (m, 2H), 3.09 – 2.86 (m, 3H), 2.79 – 2.68 (m, 1H), 1.59 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 143.33, 140.87, 139.73, 135.11, 134.54, 128.67, 128.56, 127.91, 127.06, 126.96, 126.73, 125.94, 125.47, 64.00, 53.52, 47.97, 29.30, 20.01. HPLC: Chiralpak OJ-H, hexane: isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 10.3$ min (major), $t_{\text{R}} = 15.7$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{23}\text{H}_{23}\text{NNa}$: 336.1728, found 336.1725.



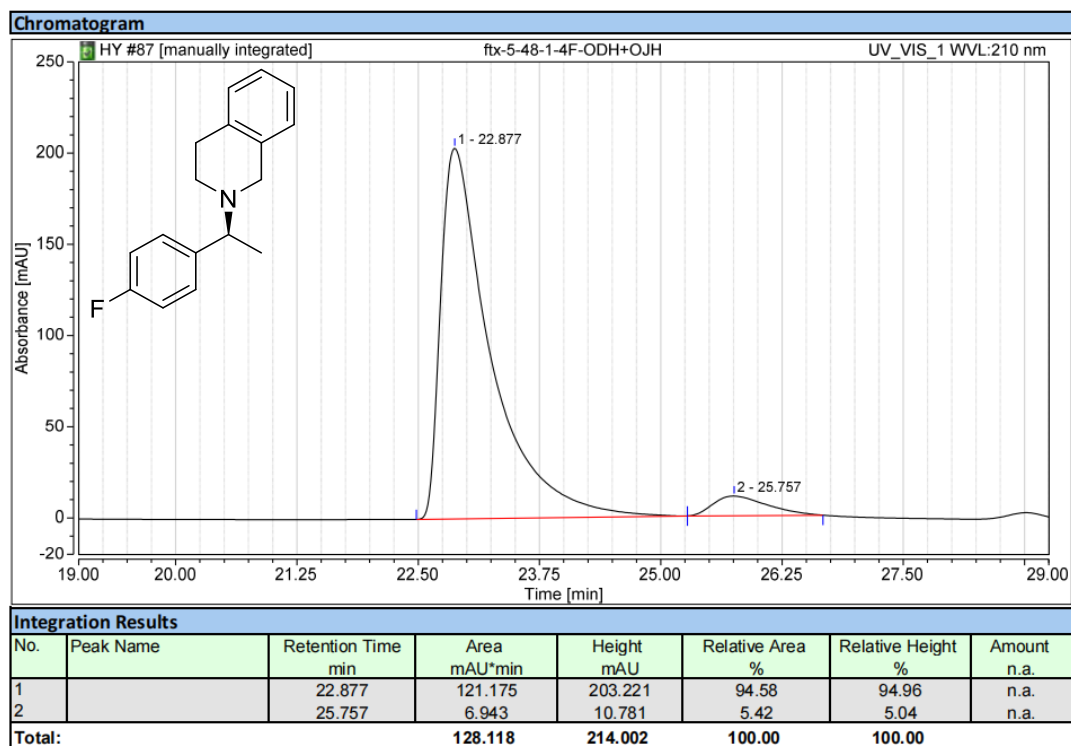
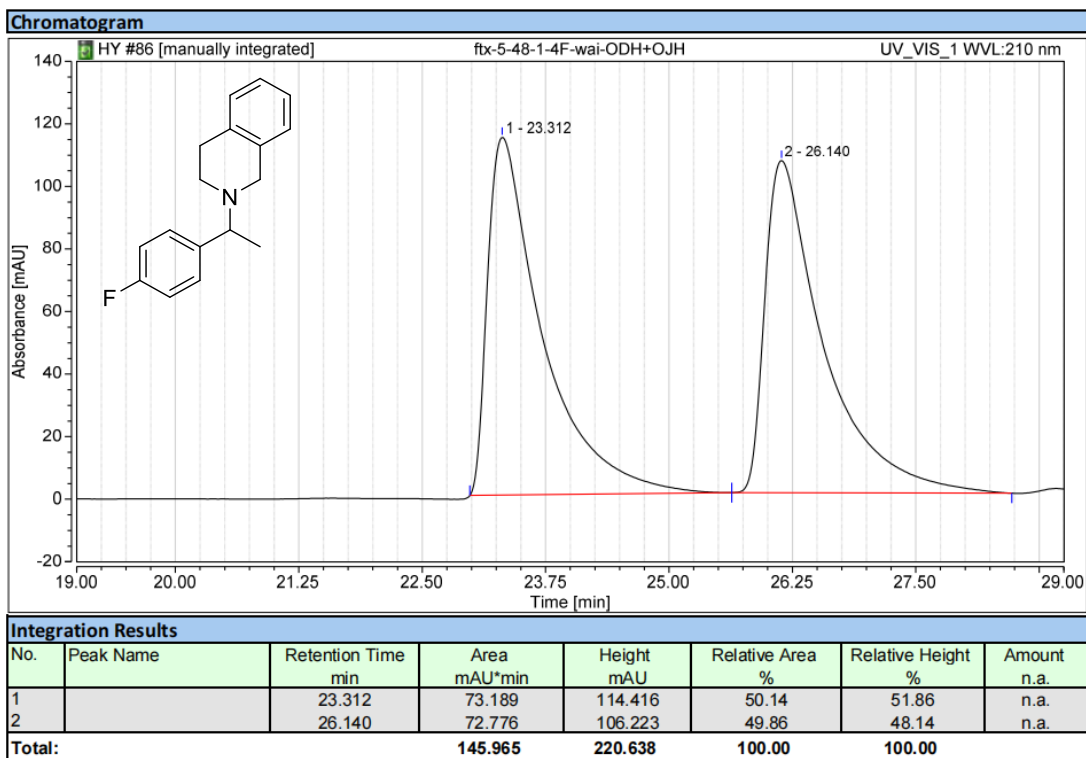


(S)-2-(1-(4-fluorophenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3e).

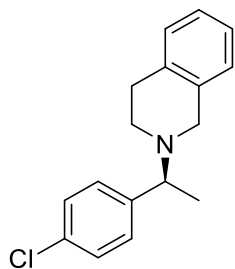


PE:EA = 20:1; Yellow oil, 48.5 mg (95% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.30 (m, 2H), 7.16 – 7.07 (m, 3H), 7.04 – 7.00 (m, 1H), 6.90 (d, J = 8.6 Hz, 2H), 3.84 (s, 3H), 3.62 – 3.52 (m, 2H), 2.94 – 2.77 (m, 3H), 2.66 – 2.60 (m, 1H), 1.48 (d, J = 6.6 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 158.50, 135.18, 134.59, 130.58, 128.59, 128.57, 126.77, 125.94, 125.47, 113.57, 63.62, 55.22, 53.49, 47.85, 29.29, 20.03. HPLC: Chiralpak ODH+OJ-H column, hexane: isopropanol = 98:2,

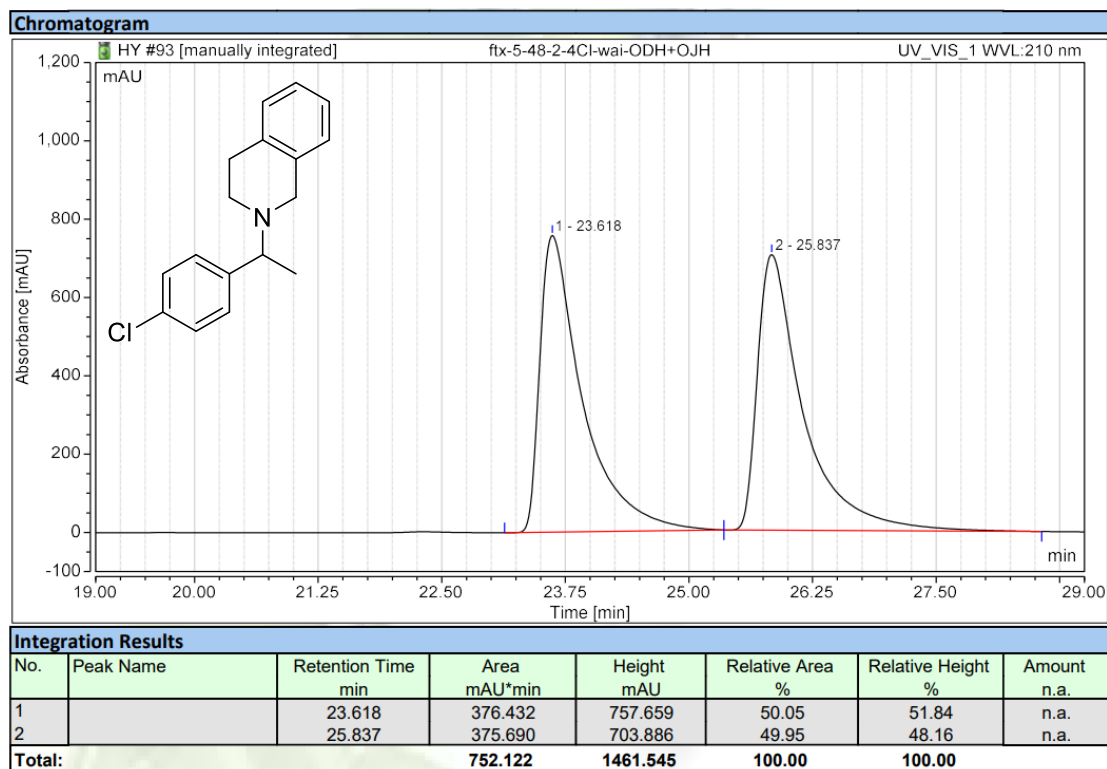
flow rate = 0.5 mL/min, UV detection at $\lambda = 210$ nm, $t_R = 22.9$ min (major), $t_R = 25.7$ min (minor). HRMS (ESI) m/z: $[M+H]^+$ calculated for $C_{17}H_{19}FN$: 256.1502, found 336.156.1494.

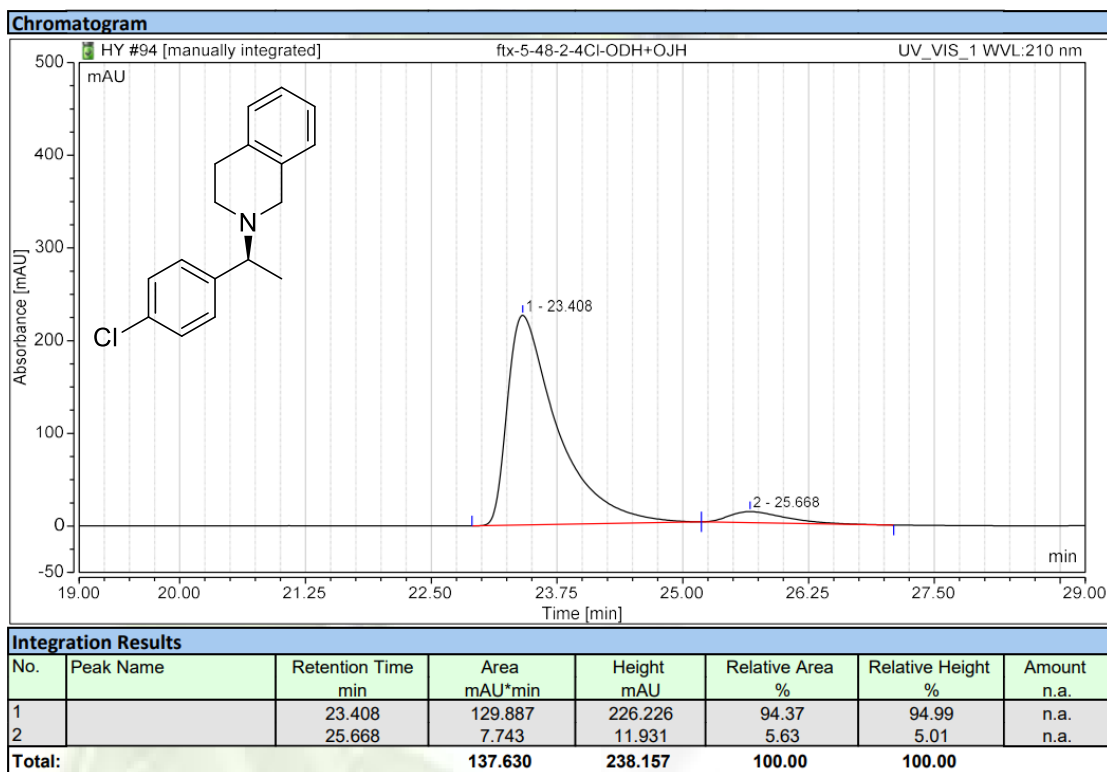


(S)-2-(1-(4-chlorophenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3f).

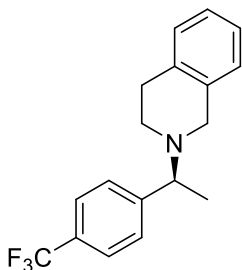


PE:EA = 20:1; Yellow oil, 50.5 mg (95% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.28 (m, 4H), 7.15 – 7.06 (m, 3H), 7.02 – 6.96 (m, 1H), 3.80 (d, J = 14.7 Hz, 1H), 3.61 – 3.49 (m, 2H), 2.94 – 2.71 (m, 3H), 2.67 – 2.58 (m, 1H), 1.44 (d, J = 6.7 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 143.00, 134.92, 134.47, 132.45, 128.82, 128.61, 128.45, 126.73, 126.05, 125.56, 63.68, 53.48, 47.95, 29.22, 20.01. HPLC: Chiralpak ODH + OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, t_{R} = 23.4 min (major), t_{R} = 25.7 min (minor). HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{17}\text{H}_{19}\text{NCl}$: 272.1206, found 272.1199.

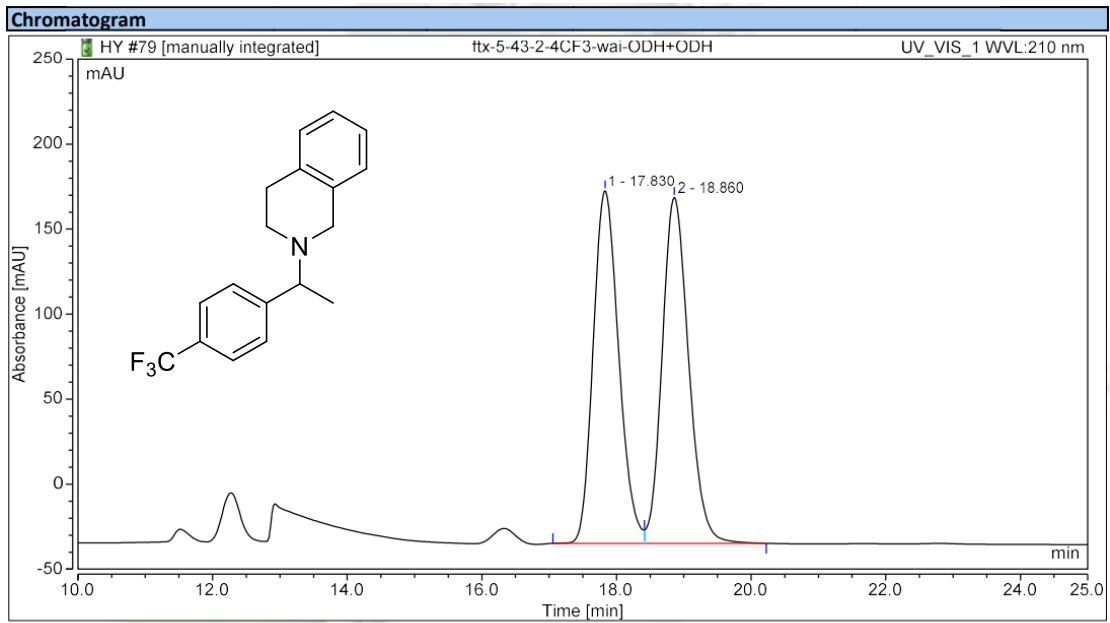




(S)-2-(1-(4-(trifluoromethyl)phenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3g).

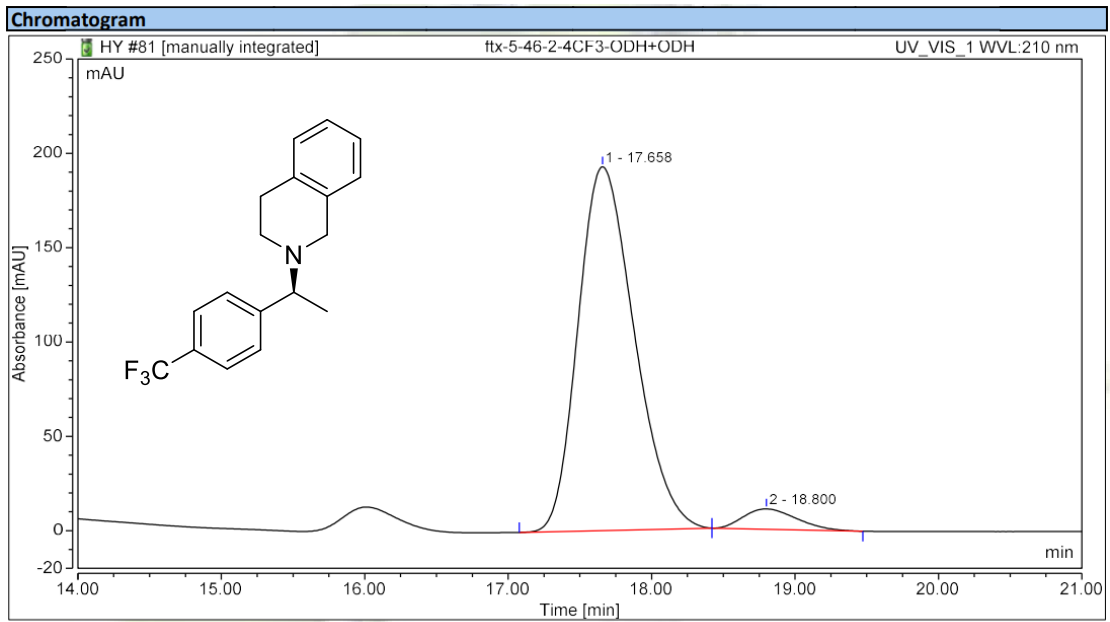


PE:EA = 20:1; Yellow oil, 55.6 mg (91% yield), 90% *ee*; ¹H NMR (500 MHz, CDCl₃) δ 7.62 (d, *J* = 8.1 Hz, 2H), 7.55 (d, *J* = 8.1 Hz, 2H), 7.20 – 7.11 (m, 3H), 7.06 – 6.99 (m, 1H), 3.86 (d, *J* = 14.7 Hz, 1H), 3.70 – 3.56 (m, 2H), 3.00 – 2.74 (m, 3H), 1.49 (d, *J* = 6.7 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 128.63, 127.74, 126.74, 126.14, 125.63, 125.35, 125.32, 64.03, 53.50, 48.08, 29.17, 20.06. HPLC: Chiralpak ODH+ODH, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, *t*_R = 17.6 min (major), *t*_R = 18.8 min (minor). HRMS (ESI) *m/z*: [M+Na]⁺ calculated for C₁₈H₁₈F₃NNa: 328.3340, found 328.3334.



Integration Results

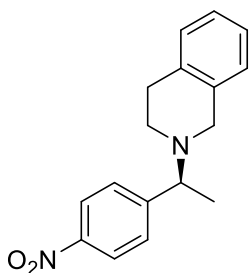
No.	Peak Name	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount n.a.
1		17.830	91.047	207.487	49.58	50.47	n.a.
2		18.860	92.607	203.613	50.42	49.53	n.a.
Total:			183.654	411.101	100.00	100.00	



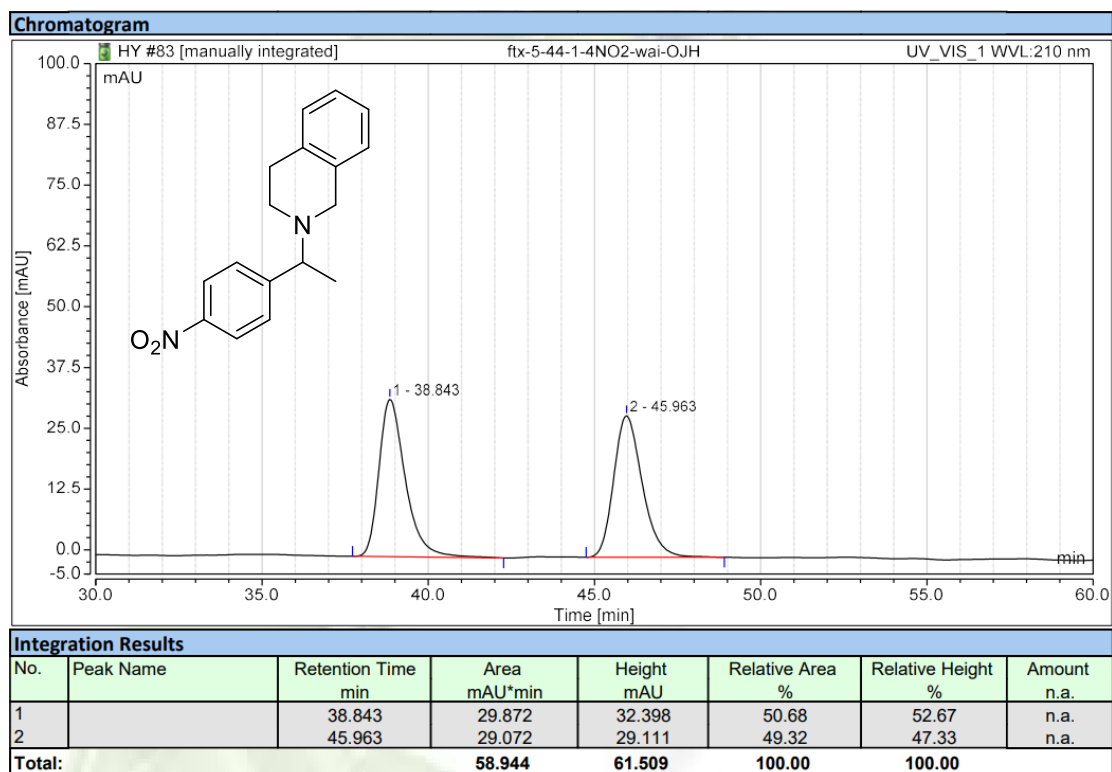
Integration Results

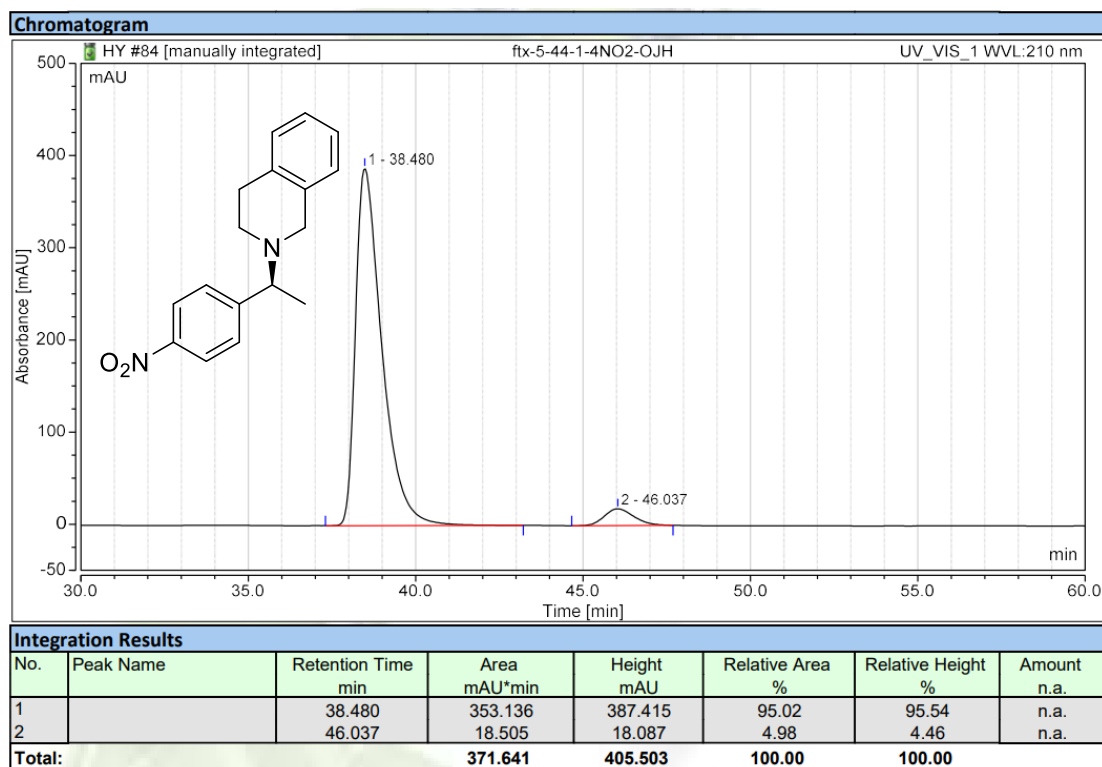
No.	Peak Name	Retention Time min	Area mAU*min	Height mAU	Relative Area %	Relative Height %	Amount n.a.
1		17.658	87.504	192.838	95.00	94.72	n.a.
2		18.800	4.608	10.751	5.00	5.28	n.a.
Total:			92.112	203.589	100.00	100.00	

(S)-2-(1-(4-nitrophenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3h).

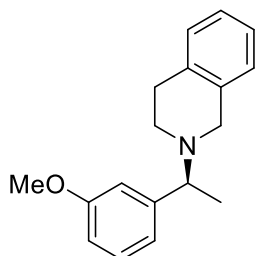


PE:EA = 20:1; Yellow oil, 50.8 mg (90% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 8.31 – 8.17 (m, 2H), 7.62 (d, $J = 8.3$ Hz, 2H), 7.22 – 7.11 (m, 3H), 7.06 – 6.98 (m, 1H), 3.86 (d, $J = 14.6$ Hz, 1H), 3.77 – 3.55 (m, 2H), 2.99 – 2.60 (m, 4H), 1.50 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 134.41, 128.63, 127.74, 126.74, 126.14, 125.63, 125.35, 125.32, 123.17, 64.03, 53.50, 48.08, 29.17, 20.06. HPLC: Chiralpak OJ-H, hexane : isopropanol = 97:3, 0.5 mL/min, 210 nm, $t_{\text{R}} = 38.5$ min (major), $t_{\text{R}} = 46.0$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_2\text{Na}$: 305.1266, found 305.1269.

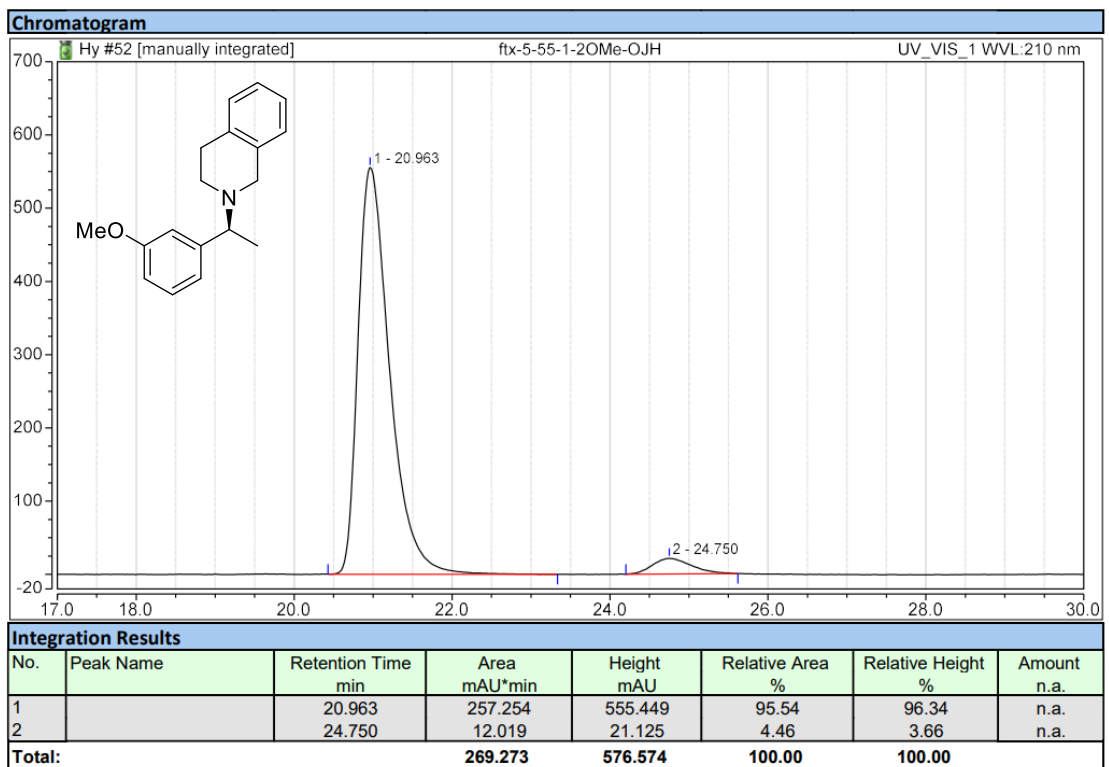
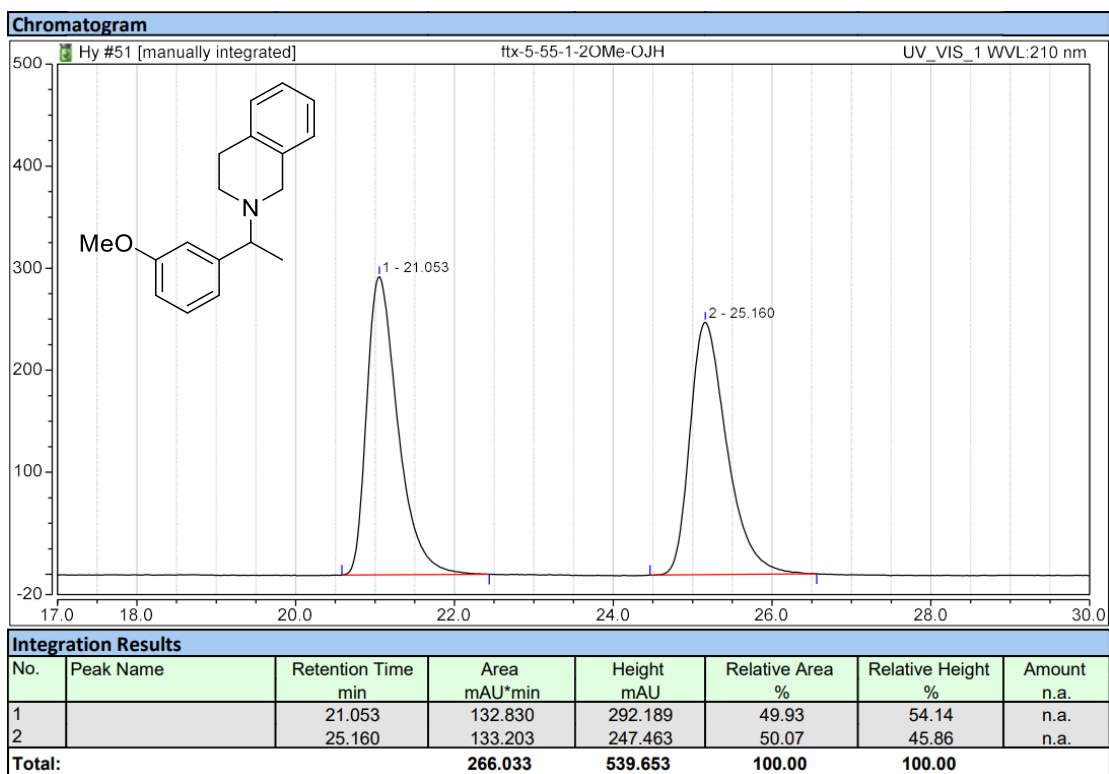




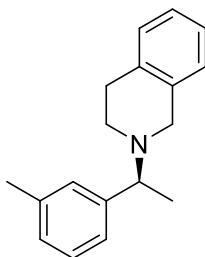
(S)-2-(1-(3-methoxyphenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3i).



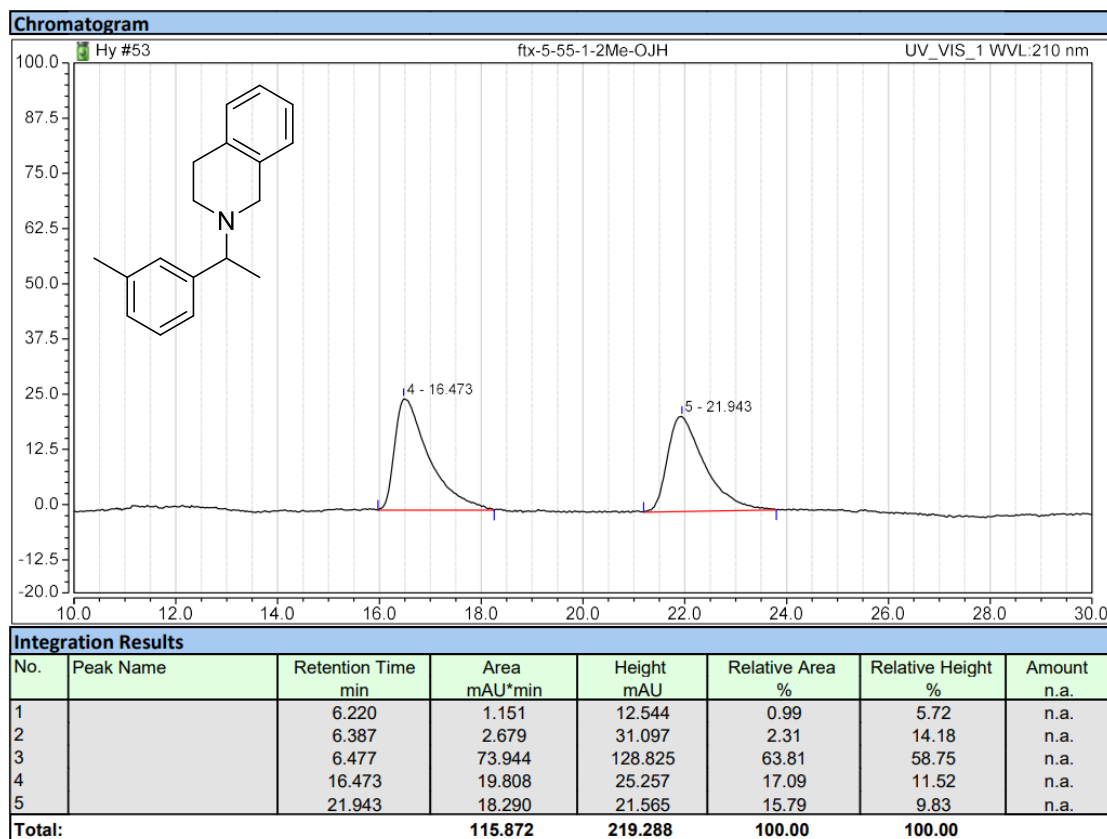
PE:EA = 20:1; Yellow oil, 50.3 mg (94% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.27 (d, $J = 8.1$ Hz, 1H), 7.17 – 7.09 (m, 3H), 7.05 – 6.96 (m, 3H), 6.89 – 6.76 (m, 1H), 3.87 (d, $J = 14.9$ Hz, 1H), 3.84 (s, 3H), 3.65 – 3.48 (m, 2H), 2.97 – 2.76 (m, 3H), 2.69 – 2.56 (m, 1H), 1.49 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 159.67, 146.19, 135.17, 134.62, 129.23, 128.59, 126.78, 125.97, 125.50, 119.97, 112.95, 112.28, 64.49, 55.21, 53.61, 48.09, 29.32, 20.26. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 9.9$ min (major), $t_{\text{R}} = 14.1$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{18}\text{H}_{21}\text{NONa}$: 290.1521, found 290.1510.

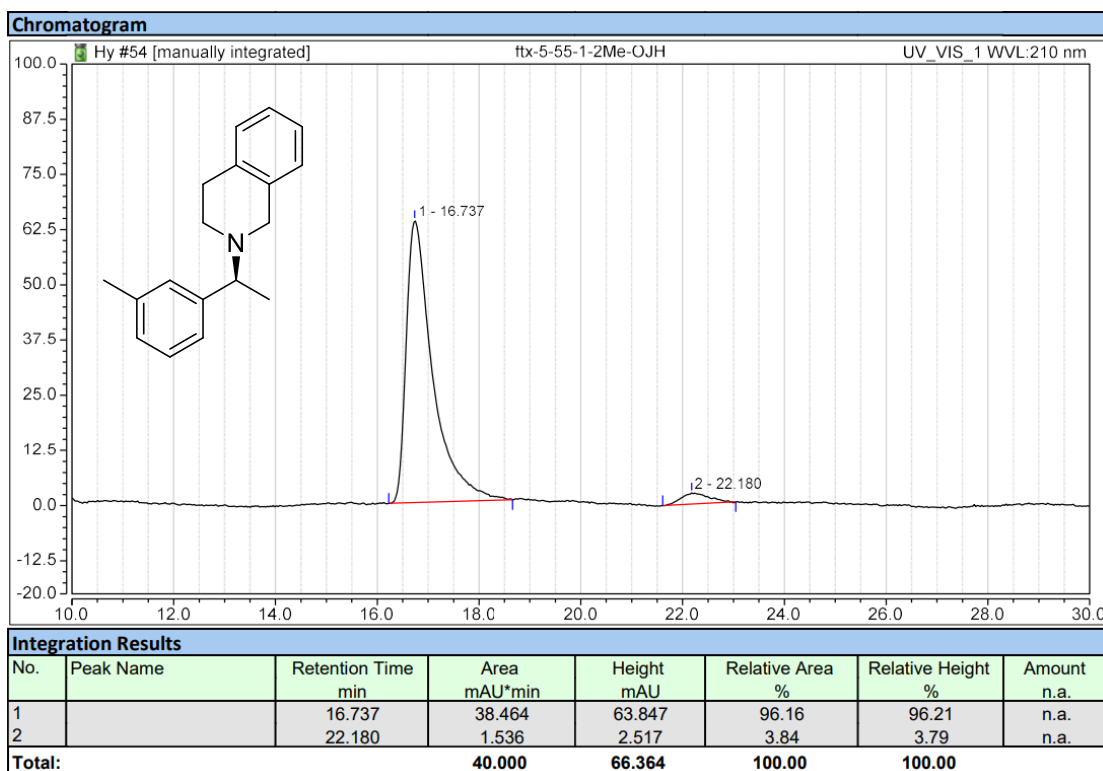


(S)-2-(1-(*m*-tolyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3j).

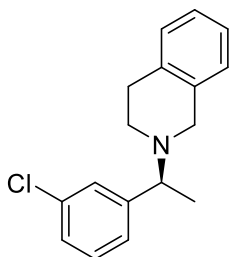


PE:EA = 20:1; Yellow oil, 47.7 mg (95% yield), 92% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.27 – 7.17 (m, 3H), 7.15 – 7.06 (m, 4H), 7.06 – 6.98 (m, 1H), 3.85 (d, J = 14.8 Hz, 1H), 3.66 – 3.47 (m, 2H), 2.98 – 2.76 (m, 3H), 2.71 – 2.57 (m, 1H), 2.38 (s, 3H), 1.49 (d, J = 6.7 Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 144.29, 137.92, 134.63, 128.63, 128.22, 128.19, 127.71, 126.83, 126.00, 125.53, 124.69, 77.29, 64.51, 53.69, 48.12, 29.30, 21.52, 20.31. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, t_{R} = 14.2 min (major), t_{R} = 17.5 min (minor). HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{18}\text{H}_{22}\text{N}$: 252.1752, found 252.1759.

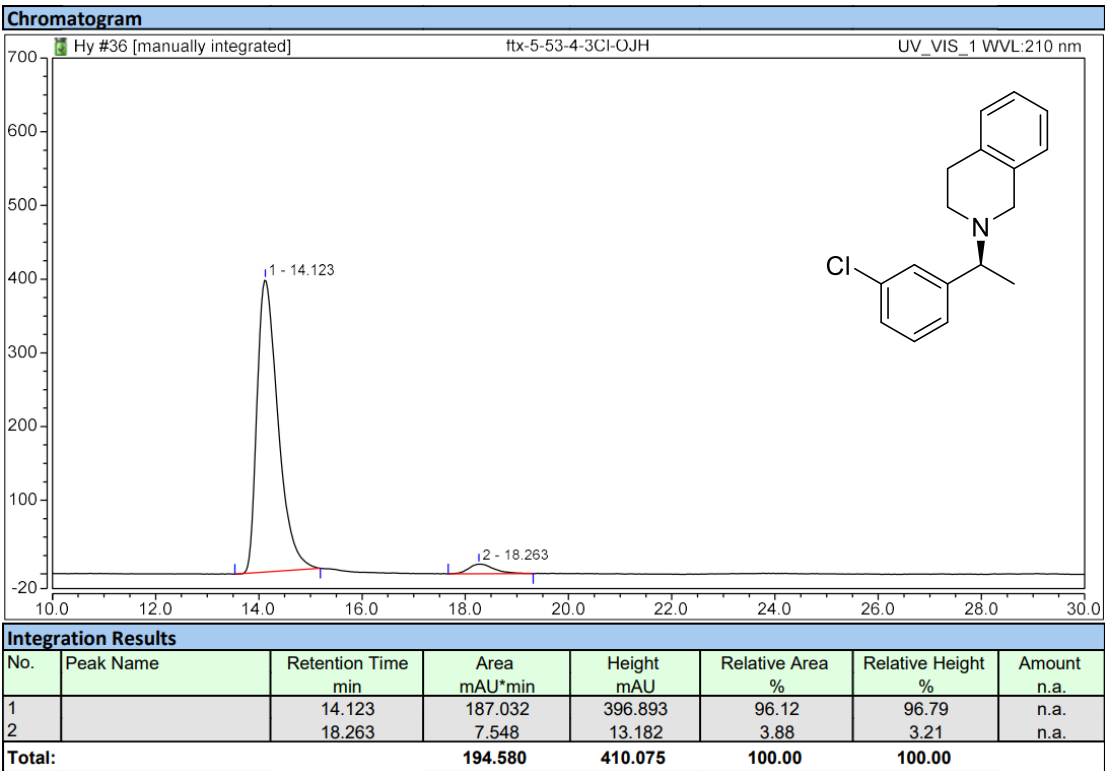
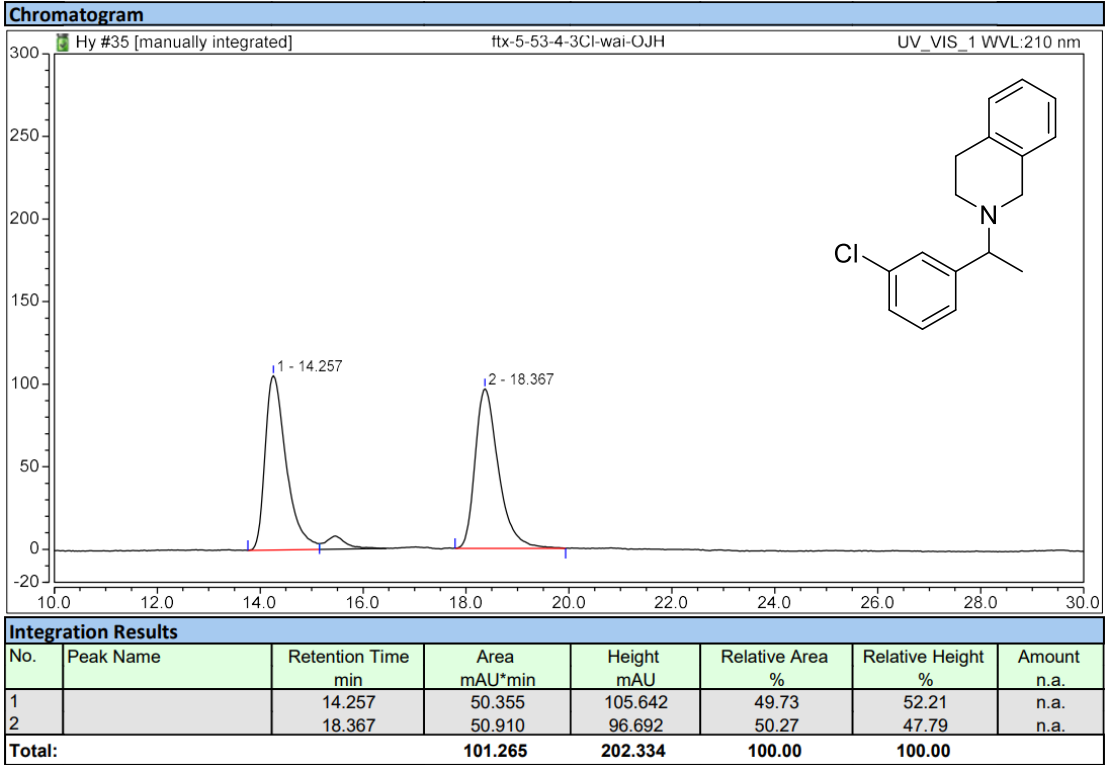




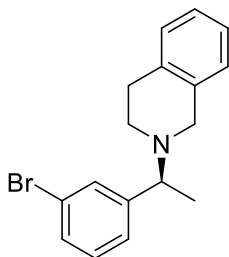
(S)-2-(1-(3-chlorophenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3k).



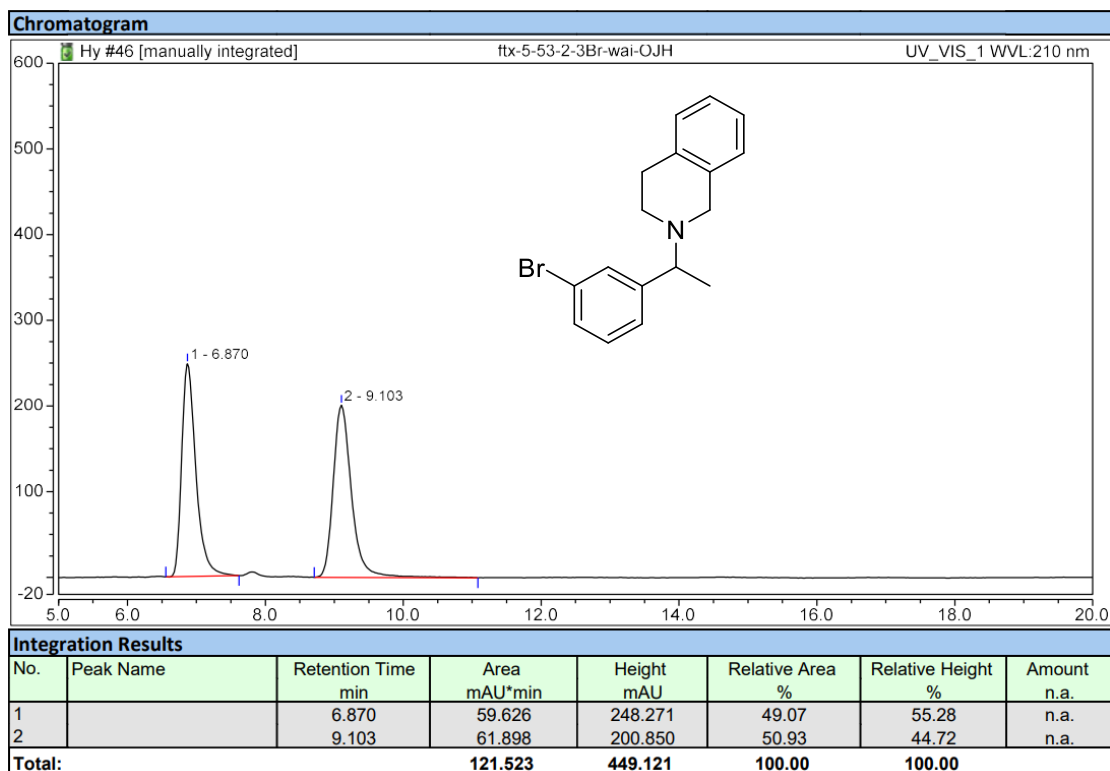
PE:EA = 20:1; Yellow oil, 49.4 mg (91% yield), 92% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.44 – 7.38 (m, 1H), 7.32 – 7.26 (m, 2H), 7.25 – 7.21 (m, 1H), 7.16 – 7.06 (m, 3H), 7.04 – 6.97 (m, 1H), 3.81 (d, $J = 14.7$ Hz, 1H), 3.63 – 3.47 (m, 2H), 2.95 – 2.72 (m, 3H), 2.69 – 2.60 (m, 1H), 1.45 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 146.74, 134.92, 134.49, 134.21, 129.60, 128.61, 127.54, 127.09, 126.74, 126.05, 125.68, 125.56, 63.96, 53.47, 47.97, 29.25, 19.96. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 14.1$ min (major), $t_{\text{R}} = 18.3$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{ClNNa}$: 294.1025, found 294.1012.

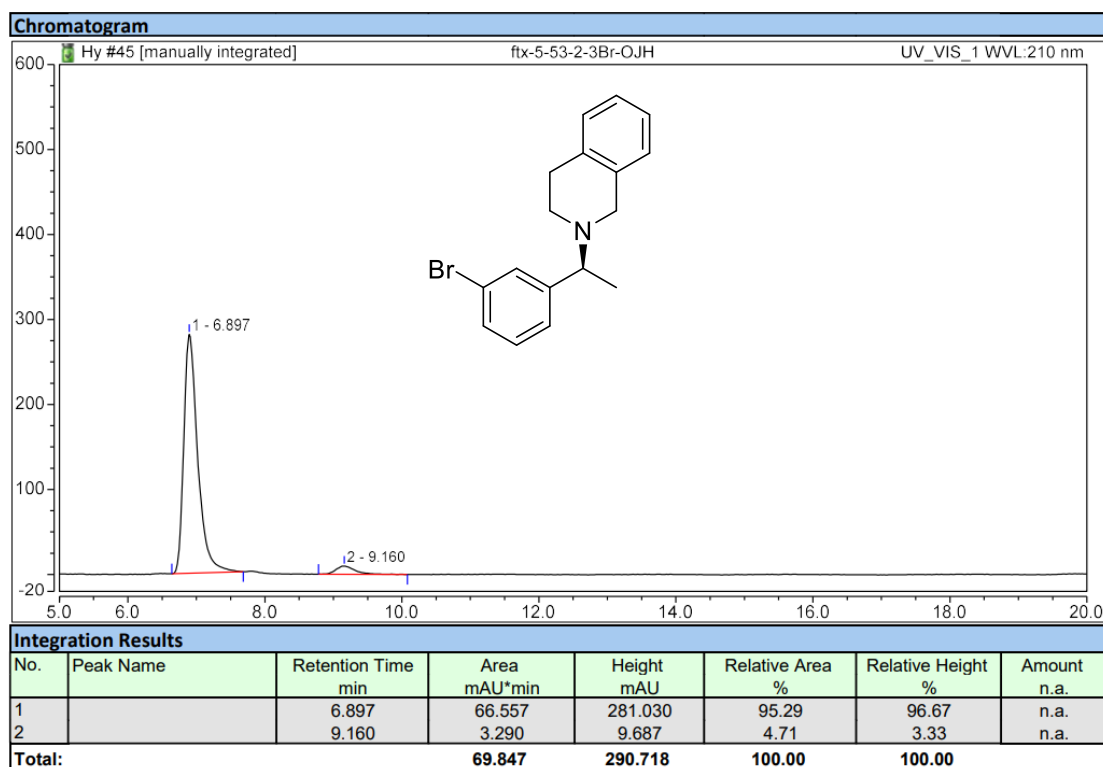


(S)-2-(1-(3-bromophenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3l).

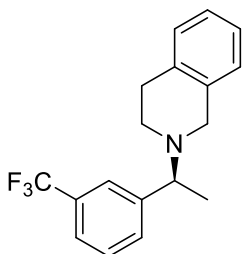


PE:EA = 20:1; Yellow oil, 60.0 mg (95% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.61 (t, $J = 1.8$ Hz, 1H), 7.47 – 7.40 (m, 1H), 7.39 – 7.34 (m, 1H), 7.24 (t, $J = 7.8$ Hz, 1H), 7.21 – 7.11 (m, 3H), 7.09 – 7.01 (m, 1H), 3.85 (d, $J = 14.7$ Hz, 1H), 3.70 – 3.48 (m, 2H), 3.01 – 2.76 (m, 3H), 2.73 – 2.63 (m, 1H), 1.49 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 147.02, 134.88, 134.45, 130.43, 130.01, 129.92, 128.59, 126.72, 126.12, 126.04, 125.55, 122.52, 63.91, 53.44, 47.95, 29.23, 19.97. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_R = 6.9$ min (major), $t_R = 9.2$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{BrNNa}$: 338.0520, found 338.0504.

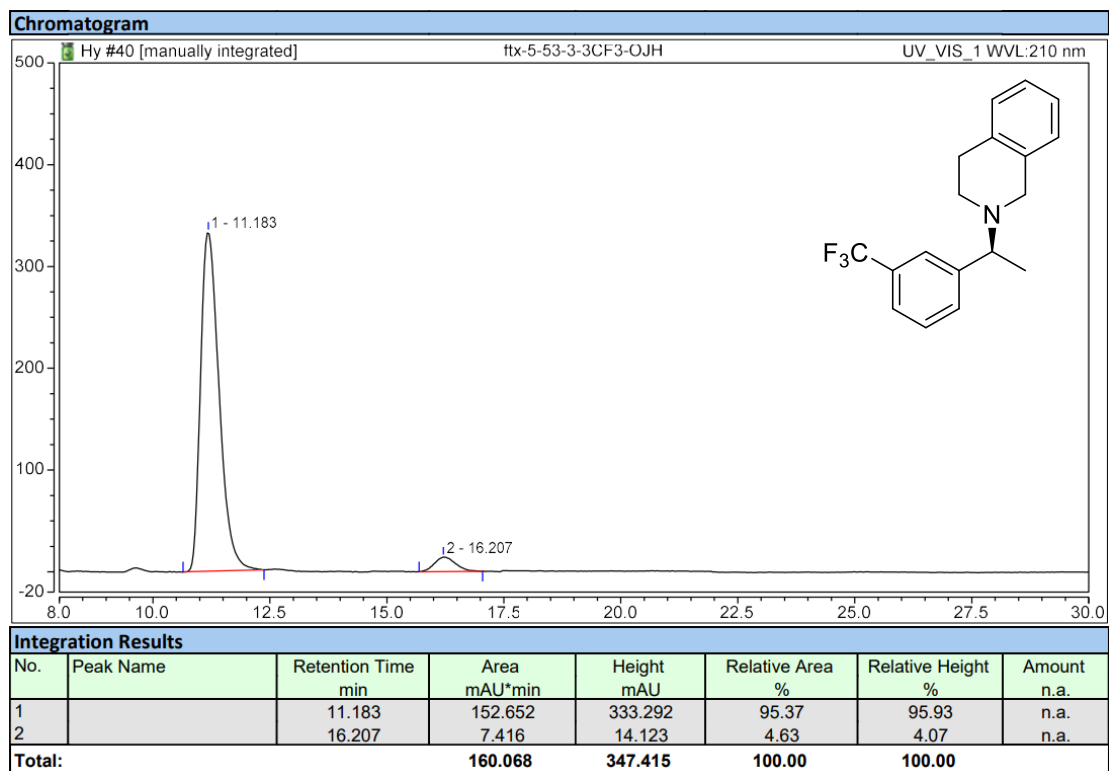
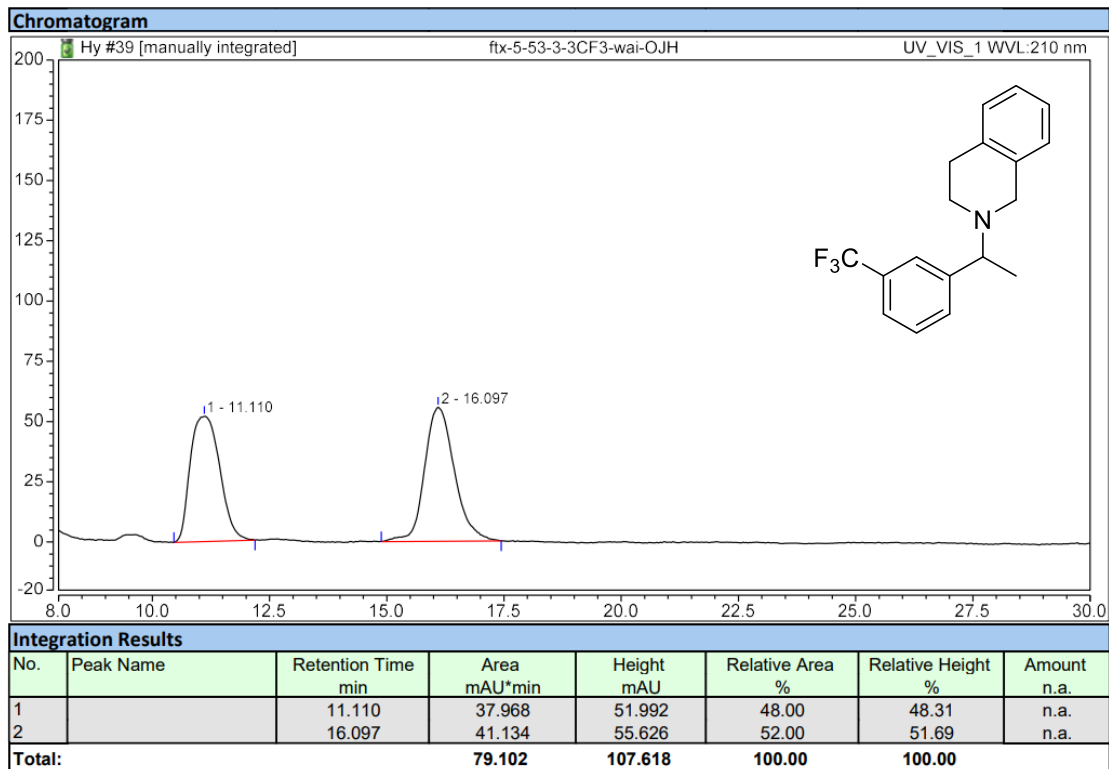




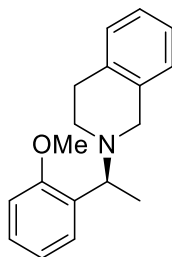
(S)-2-(1-(3-(trifluoromethyl)phenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3m).



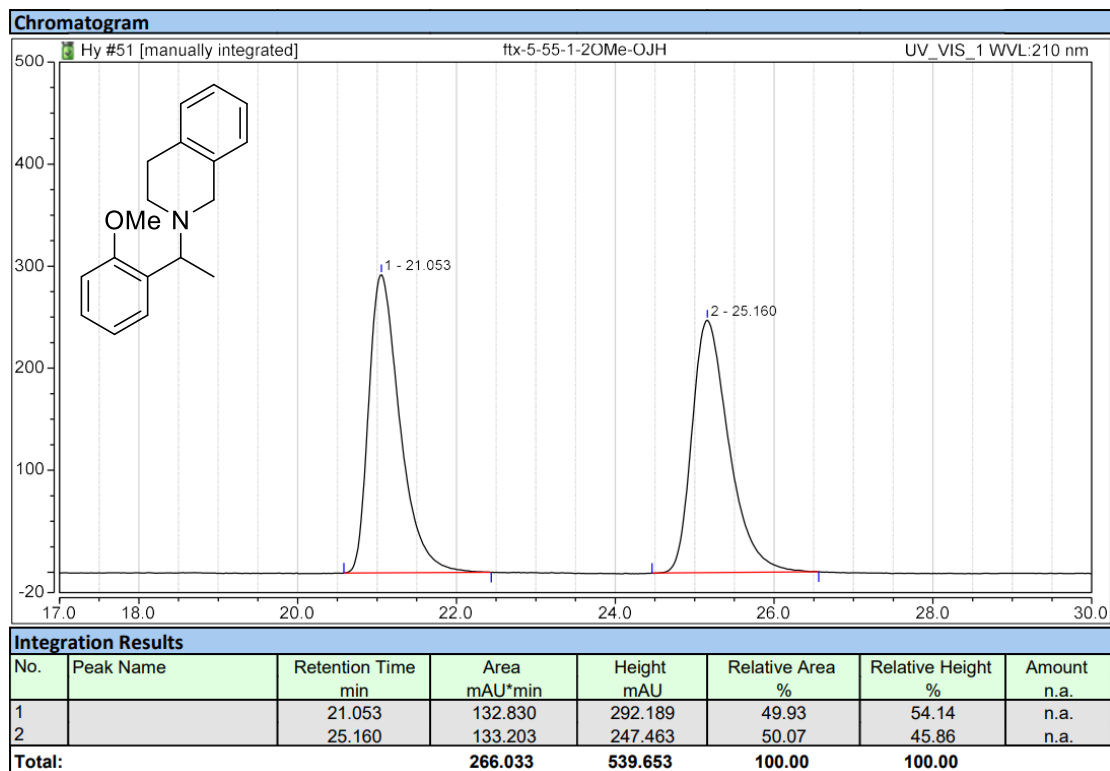
PE:EA = 20:1; Yellow oil, 58.0 mg (95% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.68 (s, 1H), 7.64 – 7.60 (m, 1H), 7.58 – 7.53 (m, 1H), 7.50 – 7.42 (m, 1H), 7.23 – 7.08 (m, 3H), 7.07 – 6.99 (m, 1H), 3.87 (d, $J = 14.7$ Hz, 1H), 3.72 – 3.55 (m, 2H), 2.98 – 2.74 (m, 3H), 2.73 – 2.57 (m, 1H), 1.50 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 145.72, 134.84, 134.47, 130.85, 128.80, 128.64, 126.75, 126.11, 125.61, 124.17, 124.14, 123.84, 63.98, 53.45, 47.99, 29.21, 19.97. ^{19}F NMR (471 MHz, CDCl_3) δ -62.38. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 11.2$ min (major), $t_{\text{R}} = 16.2$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{18}\text{H}_{18}\text{NF}_3\text{Na}$: 328.3340, found 328.3331.

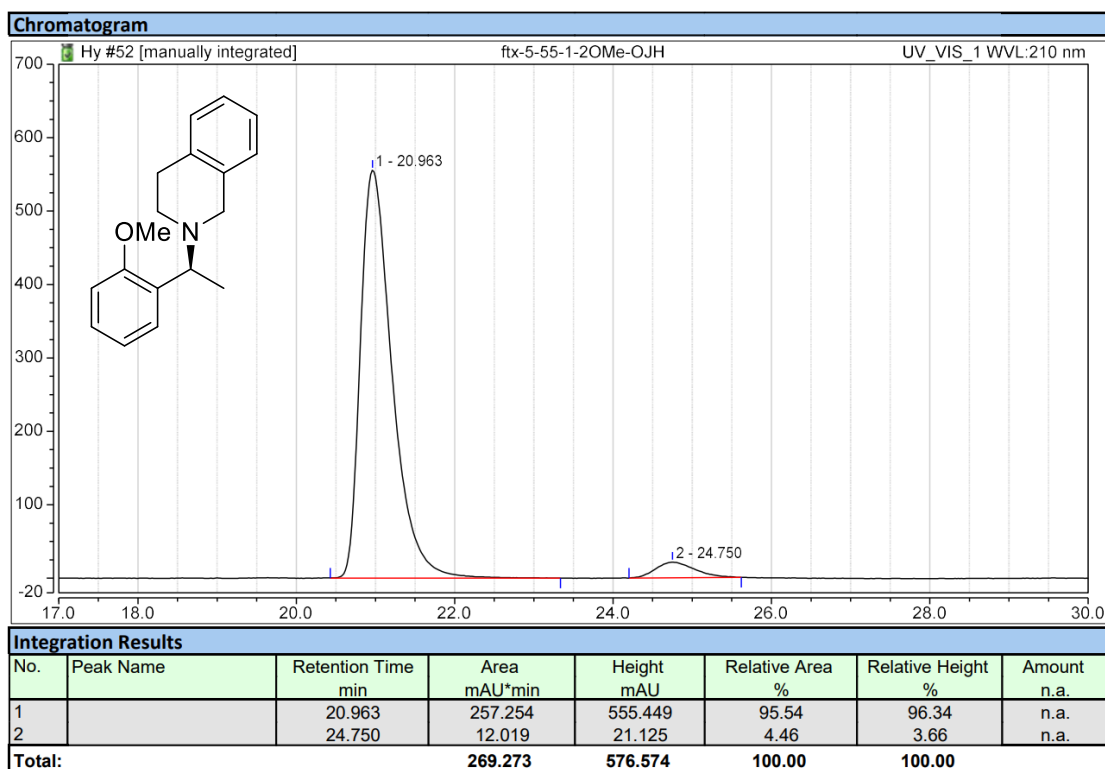


(S)-2-(1-(2-methoxyphenyl)ethyl)-1,2,3,4-tetrahydroisoquinoline (3n).

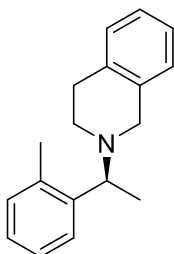


PE:EA = 20:1; Yellow oil, 51.3 mg (96% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.60 – 7.53 (m, 1H), 7.28 – 7.22 (m, 1H), 7.17 – 7.08 (m, 3H), 7.07 – 7.02 (m, 1H), 6.99 (t, $J = 7.5$ Hz, 1H), 6.92 (d, $J = 8.2$ Hz, 1H), 4.13 (q, $J = 6.7$ Hz, 1H), 3.91 (d, $J = 14.8$ Hz, 1H), 3.87 (s, 3H), 3.60 (d, $J = 14.8$ Hz, 1H), 2.97 – 2.77 (m, 3H), 2.67 – 2.56 (m, 1H), 1.44 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 156.95, 135.42, 134.74, 132.61, 128.56, 127.62, 127.43, 126.79, 125.89, 125.41, 120.68, 110.50, 55.88, 55.43, 53.74, 48.25, 20.15. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 20.9$ min (major), $t_{\text{R}} = 24.8$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{18}\text{H}_{21}\text{NONa}$: 290.1521, found 290.1516.

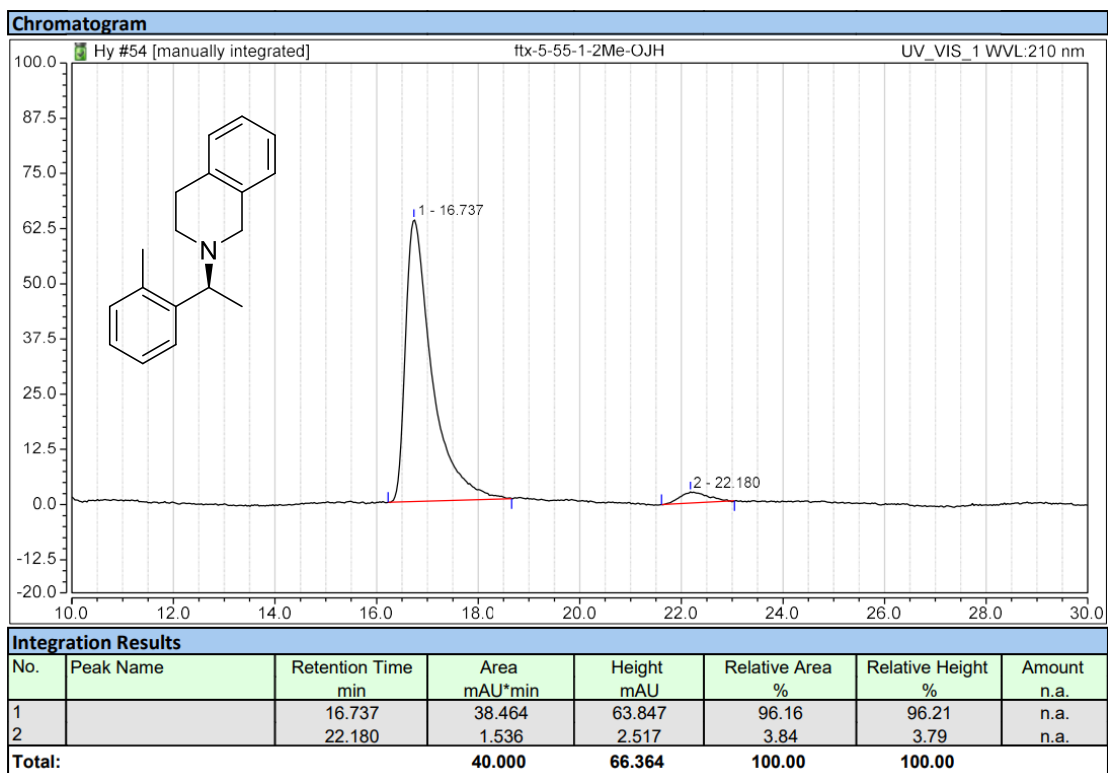
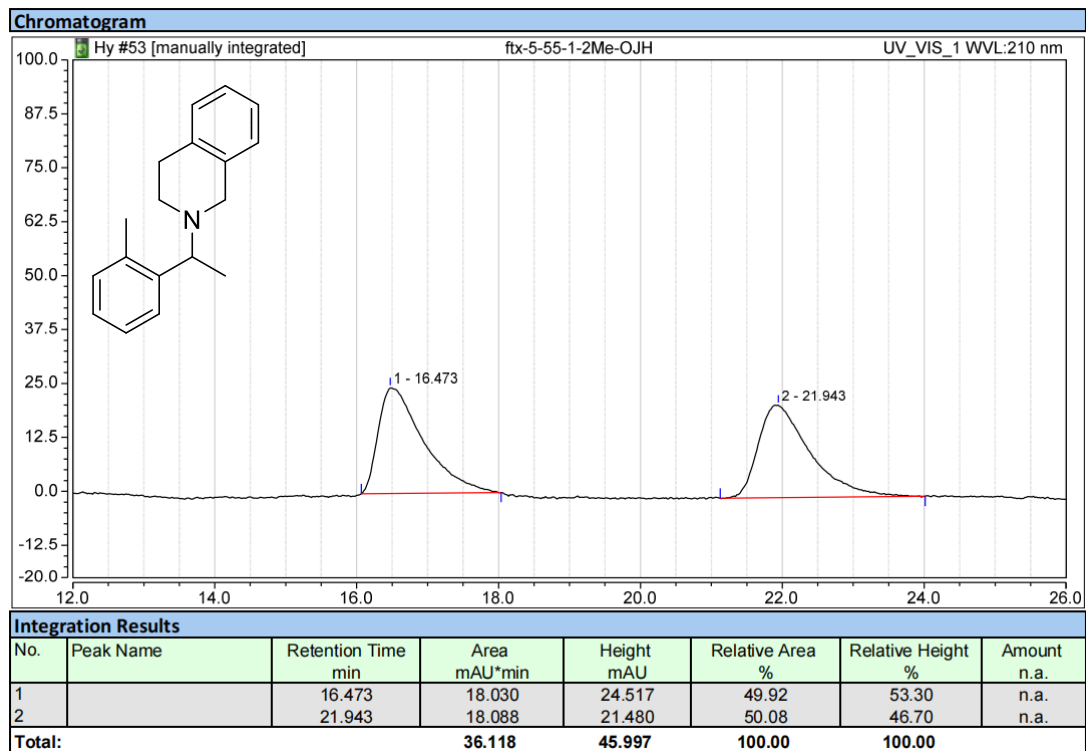




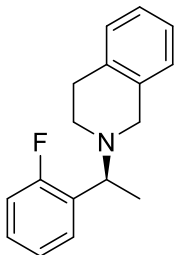
(S)-2-(1-(*o*-tolyl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3o).



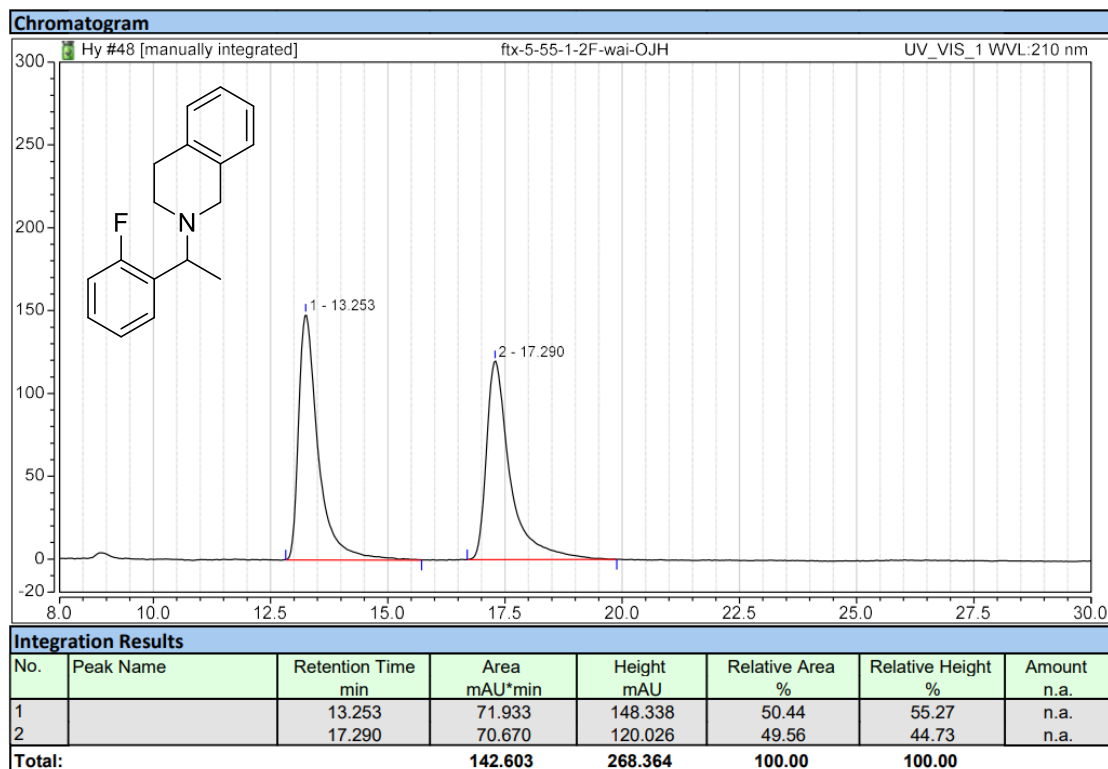
PE:EA = 20:1; Yellow oil, 47.2 mg (94% yield), 92% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.58 (d, $J = 7.6$ Hz, 1H), 7.27 – 7.10 (m, 6H), 7.08 – 7.02 (m, 1H), 3.91 (d, $J = 14.8$ Hz, 1H), 3.78 (q, $J = 6.6$ Hz, 1H), 3.61 (d, $J = 14.7$ Hz, 1H), 3.00 – 2.89 (m, 1H), 2.88 – 2.74 (m, 2H), 2.71 – 2.61 (m, 1H), 2.42 (s, 3H), 1.44 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 143.24, 135.66, 135.29, 134.76, 130.33, 128.57, 126.78, 126.60, 126.32, 126.13, 125.93, 125.46, 60.09, 53.67, 48.22, 29.38, 19.54, 19.16. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, t_R = 16.7 min (major), t_R = 22.2 min (minor). HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{18}\text{H}_{22}\text{N}$: 252.1752, found 252.1749.

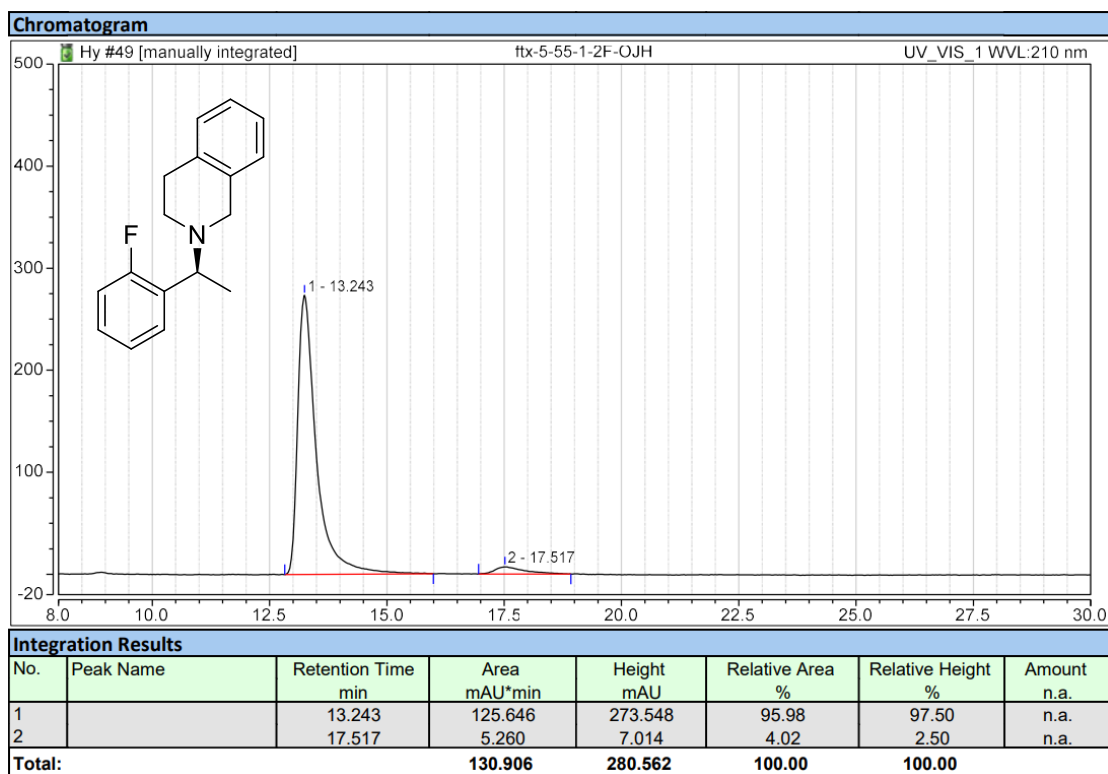


(S)-2-(1-(2-fluorophenyl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3p).

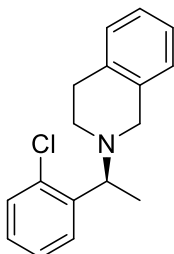


PE: EA = 20:1; Yellow oil, 48.0 mg (94% yield), 92% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.59 – 7.52 (m, 1H), 7.32 – 7.22 (m, 1H), 7.22 – 6.99 (m, 6H), 4.08 (q, $J = 6.8$ Hz, 1H), 3.88 (d, $J = 14.7$ Hz, 1H), 3.63 (d, $J = 14.7$ Hz, 1H), 3.01 – 2.77 (m, 3H), 2.72 – 2.60 (m, 1H), 1.53 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 161.72, 159.77, 135.09, 134.55, 130.49, 130.39, 128.74, 128.71, 128.64, 128.27, 128.20, 126.81, 126.06, 125.57, 124.13, 124.10, 115.45, 115.26, 56.17, 53.49, 47.98, 29.43, 19.74. ^{19}F NMR (471 MHz, CDCl_3) δ -118.60. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_{\text{R}} = 13.2$ min (major), $t_{\text{R}} = 17.5$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{FNNa}$: 278.1321, found 278.1312.

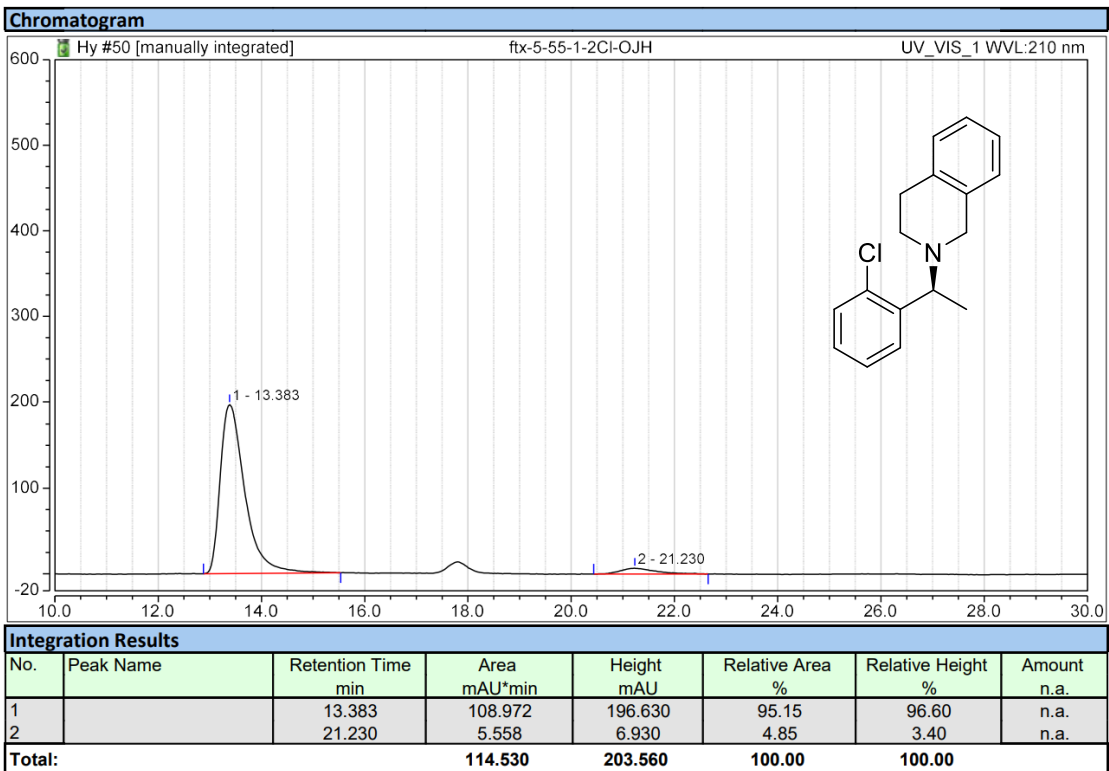
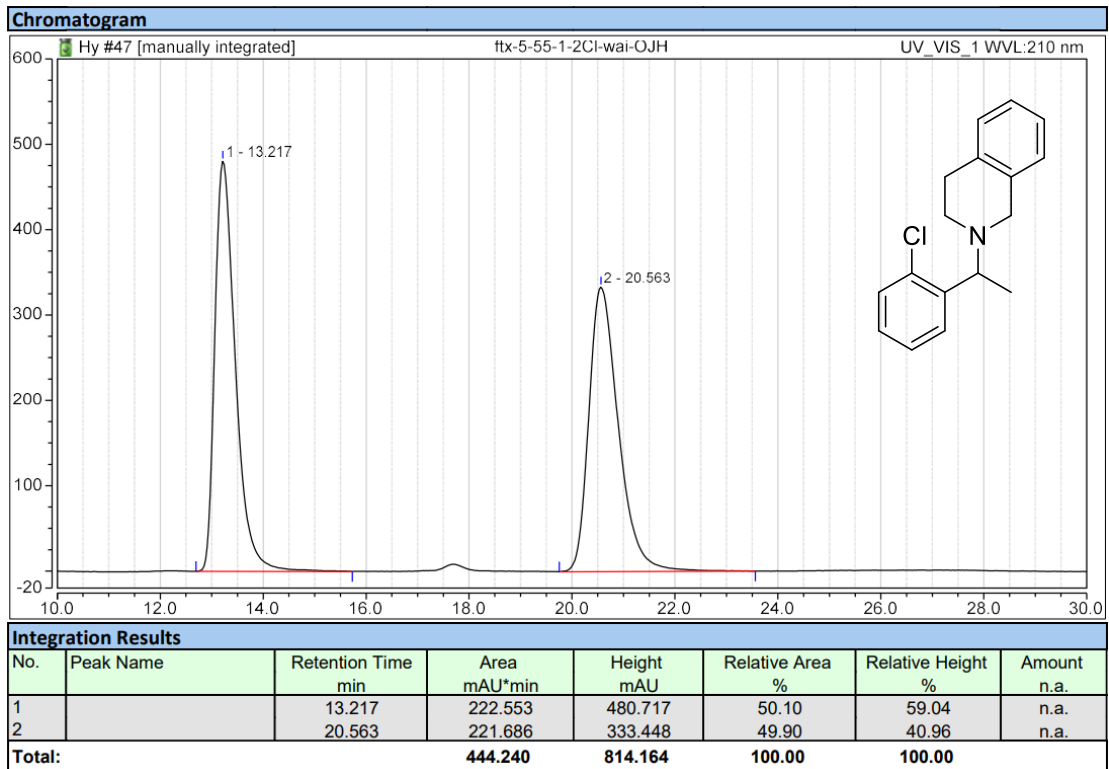




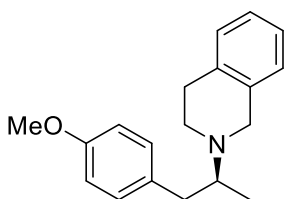
(S)-2-(1-(2-chlorophenyl) ethyl)-1,2,3,4-tetrahydroisoquinoline (3q).



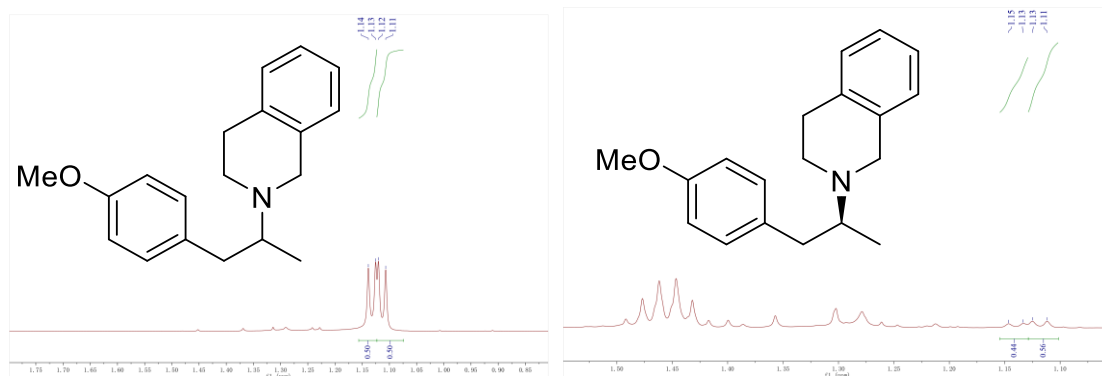
PE:EA = 20:1; Yellow oil, 51.1 mg (94% yield), 90% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.77 – 7.60 (m, 1H), 7.45 – 7.38 (m, 1H), 7.34 – 7.27 (m, 1H), 7.26 – 7.12 (m, 4H), 7.11 – 7.04 (m, 1H), 4.12 (q, $J = 6.6$ Hz, 1H), 3.97 (d, $J = 14.7$ Hz, 1H), 3.64 (d, $J = 14.7$ Hz, 1H), 3.02 – 2.90 (m, 1H), 2.89 – 2.77 (m, 2H), 2.76 – 2.65 (m, 1H), 1.47 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 142.50, 135.02, 134.58, 133.38, 129.40, 128.56, 128.32, 127.69, 127.05, 126.76, 126.01, 125.52, 59.96, 53.75, 48.39, 29.33, 20.12. HPLC: Chiralpak OJ-H, hexane : isopropanol = 98:2, 0.5 mL/min, 210 nm, $t_R = 13.4$ min (major), $t_R = 21.2$ min (minor). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{17}\text{H}_{18}\text{ClNNa}$: 294.1025, found 294.1031.



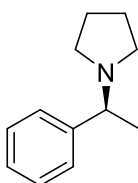
(S)-2-(1-(4-methoxyphenyl)propan-2-yl)-1,2,3,4-tetrahydroisoquinoline (3r).^[21]



PE:EA = 20:1; Yellow oil, 52.9 mg (94% yield), 14% *ee*; ¹H NMR (500 MHz, CDCl₃) δ 7.24 – 7.12 (m, 5H), 7.12 – 7.05 (m, 1H), 6.91 – 6.84 (m, 2H), 3.87 (s, 2H), 3.83 (s, 3H), 3.11 – 3.00 (m, 2H), 2.99 – 2.94 (m, 2H), 2.93 – 2.87 (m, 2H), 2.58 – 2.46 (m, 1H), 1.08 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 157.75, 135.44, 134.64, 132.54, 130.10, 128.73, 126.67, 125.87, 125.43, 113.63, 61.26, 55.17, 51.50, 46.09, 38.49, 29.89, 13.99. Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as chemical shift reagent. HRMS (ESI) *m/z*: [M+H]⁺ calculated for C₂₀H₂₆NO: 296.2014, found 296.2026.

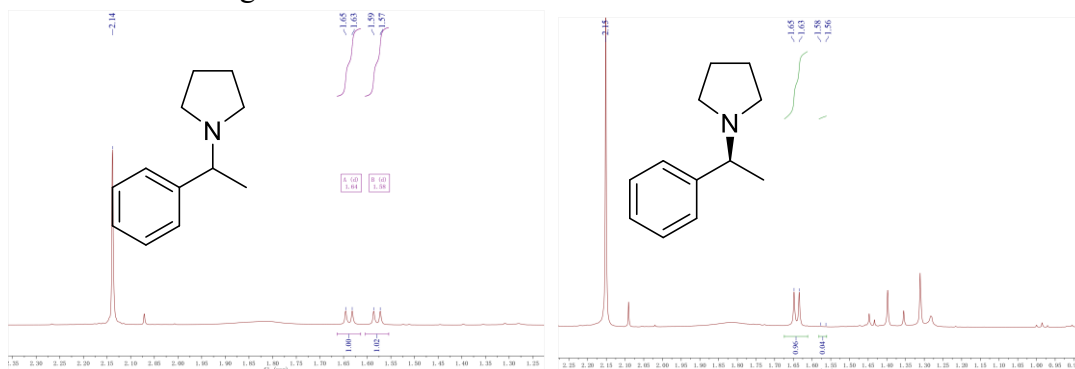


(S)-1-(1-phenylethyl)pyrrolidine (3s).^[22]

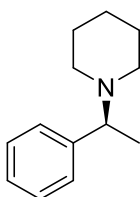


PE:EA = 2:1; Yellow oil, 32.2 mg (92% yield), 94% *ee*; ¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.30 (m, 4H), 7.27 – 7.22 (m, 1H), 3.20 (q, *J* = 6.6 Hz, 1H), 2.63 – 2.49 (m, 2H), 2.45 – 2.33 (m, 2H), 1.82 – 1.75 (m, 4H), 1.43 (dd, *J* = 6.6, 1.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 145.73, 128.22, 127.20, 126.78, 66.00, 52.98, 23.39, 23.18. Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as

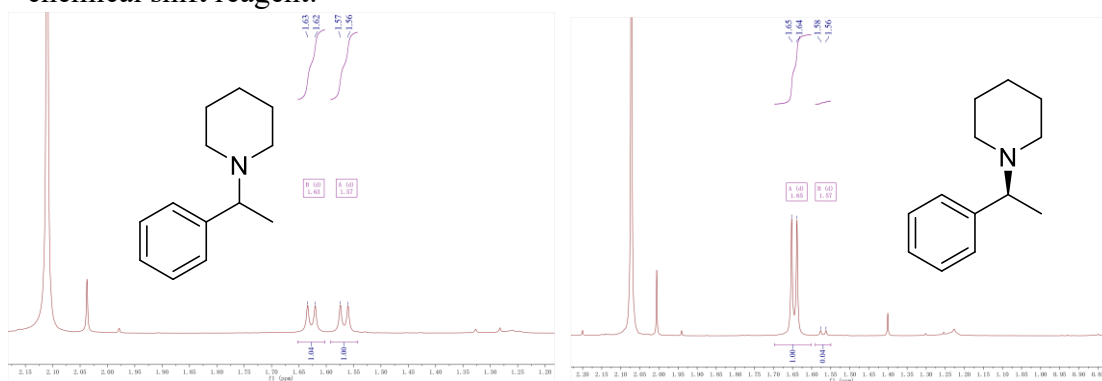
chemical shift reagent.



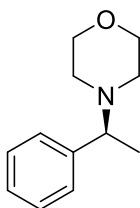
(S)-1-(1-phenylethyl)piperidine (3t).^[2]



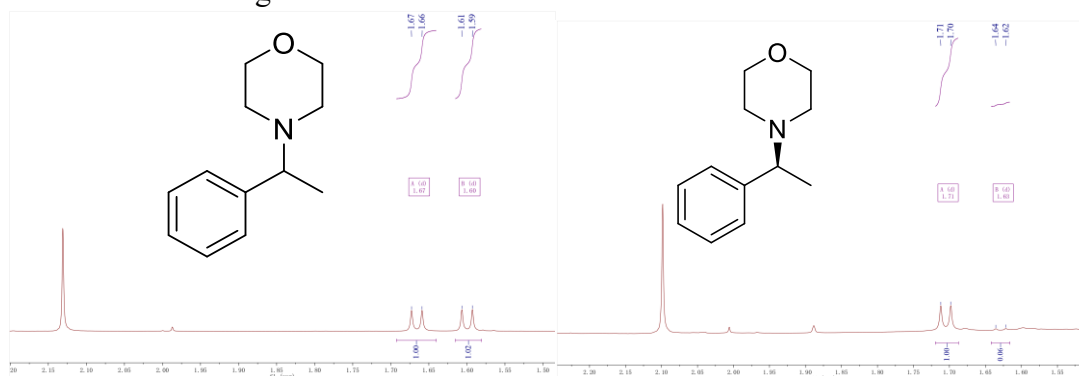
PE:EA = 5:1; Yellow oil, 34.1 mg (90% yield), 92 % *ee*, ¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.29 (m, 3H), 7.28 – 7.19 (m, 2H), 3.38 (q, *J* = 6.8 Hz, 1H), 2.38 – 2.29 (m, 4H), 1.65 – 1.43 (m, 6H), 1.36 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 143.91, 127.98, 126.63, 125.37, 65.18, 51.49, 26.26, 24.60, 19.40. Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as chemical shift reagent.



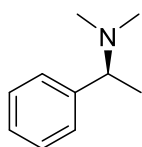
(S)-4-(1-phenylethyl)morpholine (3u).^[2]



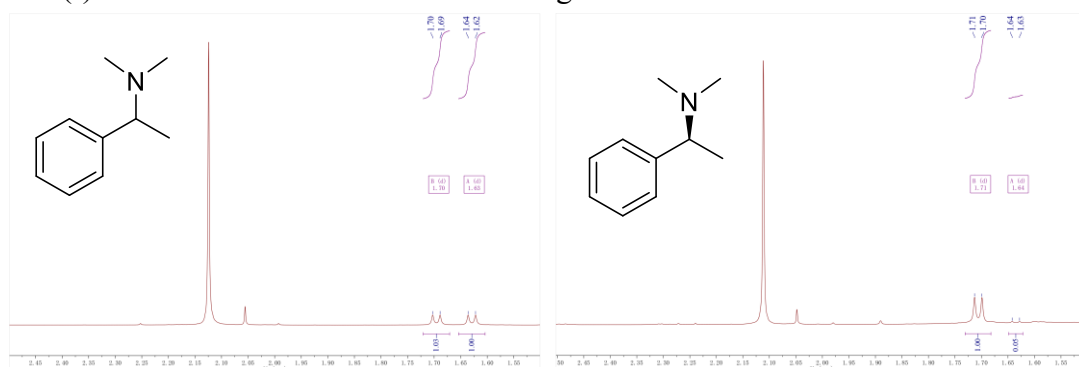
PE:EA = 5:1; Yellow oil, 34.4 mg (90% yield), 89% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.34 (d, $J = 4.4$ Hz, 4H), 7.29 – 7.24 (m, 1H), 3.75 – 3.68 (m, 4H), 3.32 (q, $J = 6.6$ Hz, 1H), 2.57 – 2.46 (m, 2H), 2.44 – 2.34 (m, 2H), 1.38 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.92, 128.26, 127.60, 126.94, 67.21, 65.37, 51.29, 19.81. Enantiomeric excess was determined by ^1H NMR using D-(-)-Mandelic acid as chemical shift reagent.



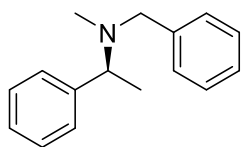
(S)-N,N-dimethyl-1-phenylethan-1-amine(3v)^[2]



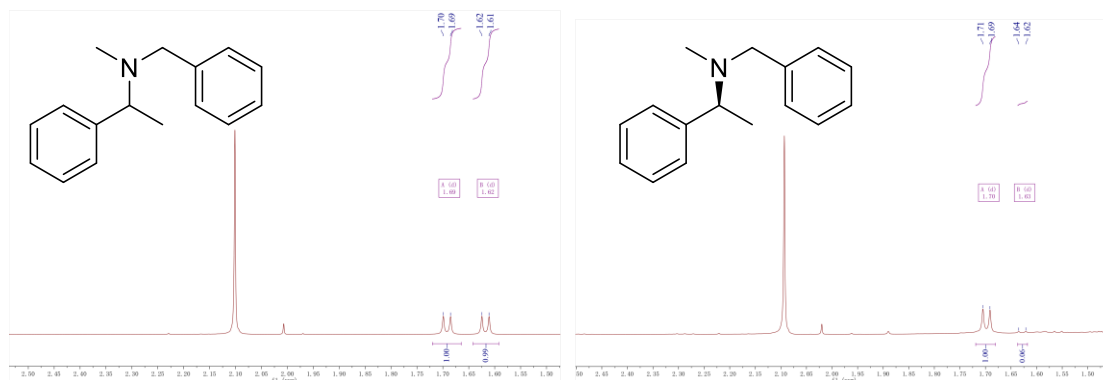
DCM:MeOH = 5:1; Yellow oil, 27.1 mg (91% yield), 91% *ee*; ^1H NMR (500 MHz, CDCl_3) δ 7.36 – 7.32 (m, 3H), 7.29 – 7.24 (m, 2H), 3.27 (d, $J = 6.7$ Hz, 1H), 2.22 (s, 6H), 1.40 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 144.08, 128.18, 127.51, 126.86, 65.98, 43.25, 20.24. Enantiomeric excess was determined by ^1H NMR using D-(-)-Mandelic acid as chemical shift reagent.



(*S*)-*N*-benzyl-*N*-methyl-1-phenylethan-1-amine(**3w**)^[2]

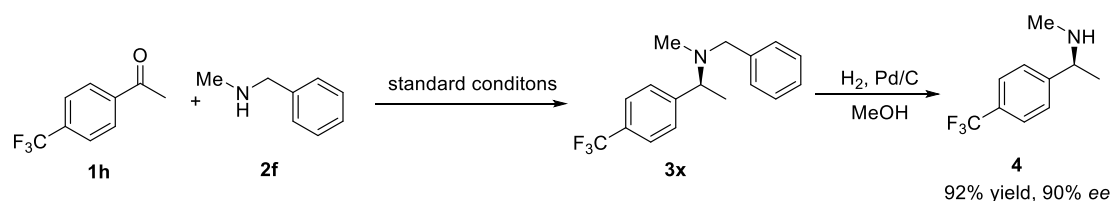


PE:EA = 5:1; Yellow oil, 42.3 mg (94% yield), 89% *ee*; ¹H NMR (500 MHz, CDCl₃) δ 7.47 – 7.42 (m, 2H), 7.40 – 7.29 (m, 6H), 7.30 – 7.23 (m, 2H), 3.73 – 3.55 (m, 2H), 3.35 (d, *J* = 13.3 Hz, 1H), 2.18 (s, 3H), 1.47 (d, *J* = 6.7 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 144.17, 140.10, 128.72, 128.15, 128.13, 127.67, 126.76, 126.69, 63.21, 58.86, 38.34, 18.37. Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as chemical shift reagent.



2.3 General procedure for the preparation of **4** and **5**.

2.3.1 General procedure for the preparation of **4**^[2]



Step1:

To a vial containing a magnetic stirring bar was added **1h** (0.2 mmol), **2f** (0.22 mmol), I₂ (2 mol%), Ti(OEt)₄ (0.4 mmol), [Ir(COD)(Cl)]₂ (1 mol %), and **N-Me-Xu6** (2.2 mol %) under nitrogen atmosphere, anhydrous THF (2 mL) was added and stirred

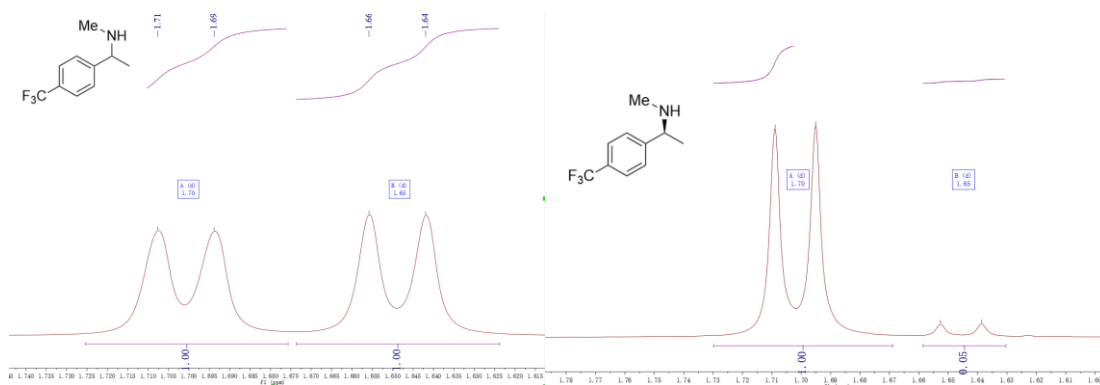
for 30 minutes. Then the vial was transferred in Parr steel autoclave, which was purged three times with hydrogen and finally pressurized to 50 atm. The reaction mixture was stirred at 0 °C for 24 h. The hydrogen gas was released slowly and the solution was quenched with Na₂SO₄·10H₂O and filtered. The organic phase was concentrated to give the crude products **3x**, which were used in next step without purification.

Step2:

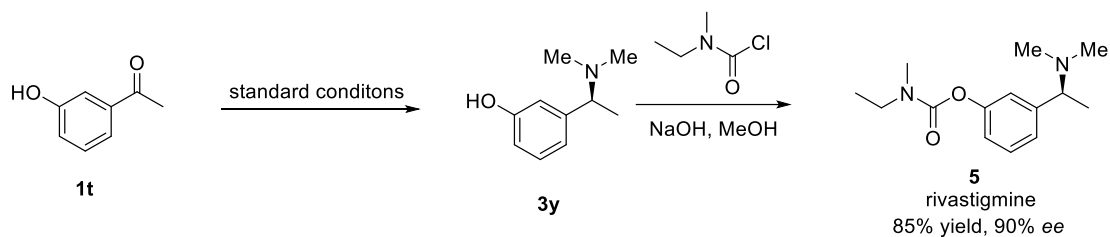
Pd(OH)₂ on carbon (10%, 4.5 mg) was added to the solution of **3x** (58.6 mg, 0.2 mmol) and acetic acid (1 drop) in MeOH (2 mL). The resulting mixture was transferred to an autoclave, which was charged with 20 atm of H₂, and stirred at r.t. for 10 h. The hydrogen gas was released slowly and the solution was filter and quenched with aqueous sodium bicarbonate solution. The organic phase was concentrated and purified with column chromatography to afford **4**

(S)-N-methyl-1-(4-(trifluoromethyl)phenyl)ethan-1-amine(**4**)^[5]

PE:EA=5:1, Yellow oil, 7.3 mg, 92% yield, 90% *ee*. ¹H NMR (500 MHz, CDCl₃) δ 7.64 (d, J = 8.1 Hz, 2H), 7.51 (d, J = 8.0 Hz, 2H), 3.89 (q, J = 6.7 Hz, 1H), 2.35 (s, 3H), 1.49 (d, J = 6.7 Hz, 3H). Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as chemical shift reagent.



2.3.2 General procedure for the preparation of rivastigmine. ^[2]



Step1:

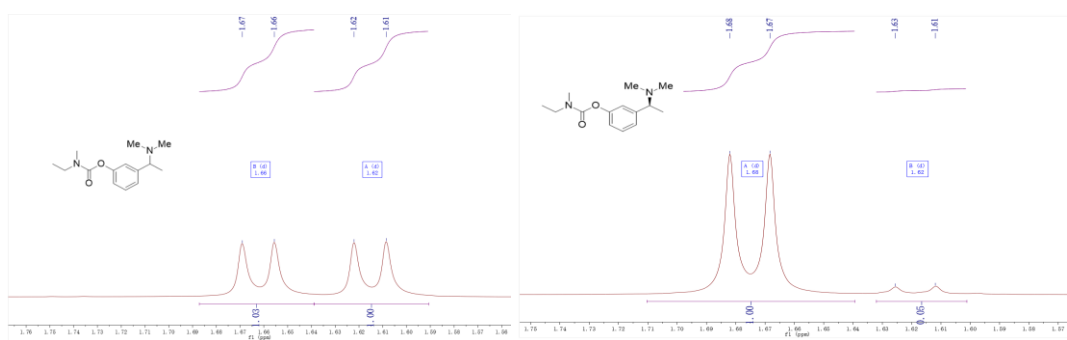
To a vial containing a magnetic stirring bar was added **1t** (0.2 mmol), dimethylamine (0.22 mmol), I₂ (2 mol%), Ti(OEt)₄ (0.4 mmol), [Ir(COD)(Cl)]₂ (1 mol %), and **N-Me-Xu6** (2.2 mol %) under nitrogen atmosphere, anhydrous THF (2 mL) was added and stirred for 30 minutes. Then the vial was transferred in Parr steel autoclave, which was purged three times with hydrogen and finally pressurized to 50 atm. The reaction mixture was stirred at 0 °C for 24 h. The hydrogen gas was released slowly and the solution was quenched with Na₂SO₄·10H₂O and filtered. The organic phase was concentrated to give the crude products **3y**, which were used in next step without purification.

Step2:

NaOH (0.25 mmol) was added to the solution of **3y** in CH₃CN (1.5 mL). The the above solution was stirred for 1 h at room temperature, then ethyl(methyl)carbamoyl chloride (0.22 mmol) was added. The reaction mixture was stirred for 12 h, quenched by aq. NH₄Cl solution. CH₃CN was removed under vacuum and the solution was extracted by EtOAc (5 mL*3). The organic phase was dried over anhydrous Na₂SO₄, concentrated and purified with column chromatography (EtOAc/PE) to give rivastigmine.

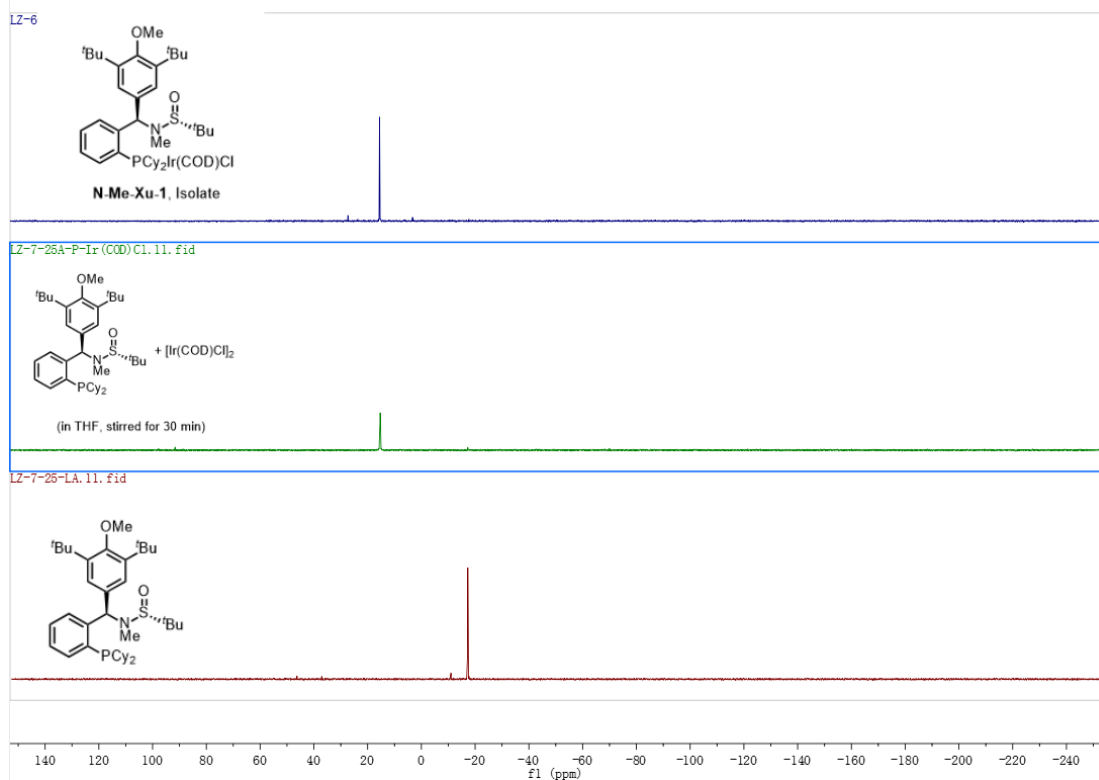
(S)-3-[1-(Dimethylamino)ethyl]phenyl Ethyl-(methyl)carbamate(rivastigmine):

PE:EA=20:1, 42.5 mg (85% yield for 2 steps), 90% *ee*, ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.27 (m, 1H), 7.15 – 7.01 (m, 3H), 3.52 – 3.39 (m, 2H), 3.29 (q, J = 6.7 Hz, 1H), 3.04 (d, J = 37.5 Hz, 3H), 2.23 (s, 6H), 1.39 (d, J = 6.7 Hz, 3H), 1.28 – 1.19 (m, 3H). Enantiomeric excess was determined by ¹H NMR using D-(-)-Mandelic acid as chemical shift reagent.

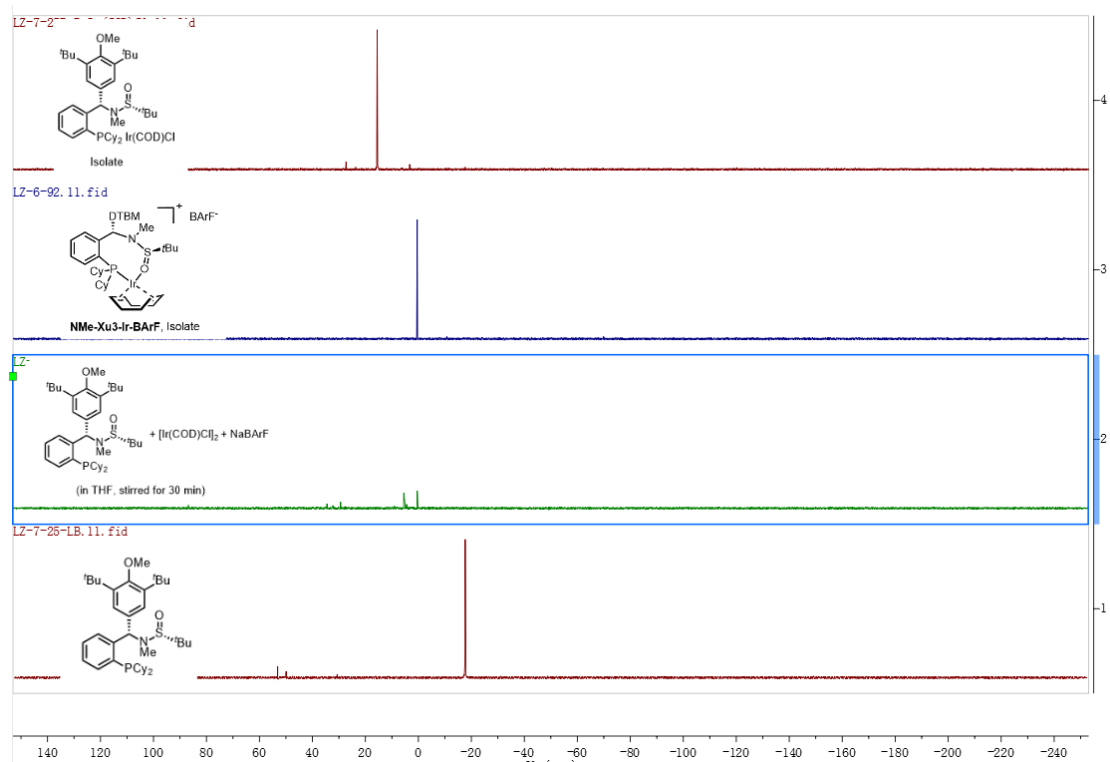


2.3.3 ^{31}P NMR spectra of ligands and Ir catalysts.

^{31}P NMR spectra for N-Me-Xu1 and Ir complexes



³¹P NMR spectra for N-Me-Xu3 and Ir complexes



3. X-Ray Crystal Data

Crystal Structure Information of NMe-Xu1-Ir

0.1 mL of DCM was added to a 10 mL oven-dried glass sample bottle with 10 mg pure NMe-Xu1-Ir to dissolve the sample, then 8 mL n-hexane was slowly added to the solution, sealed with perforated paper, and then the solvent was slowly dried at room temperature to obtain crystals. Single crystal X-ray diffraction data were collected on Bruker Smart Apex II CCD diffractometer. The crystal structure has been deposited at the Cambridge Crystallographic Data Centre (CCDC): 2371593

Formula: C₄₇H₇₄ IrNO₂PS

Flack parameter 0.066(7)

Bond precision:	C-C = 0.0115 Å	Wavelength=1.54178	
Cell:	a=44.8783(13)	b=11.8093(3)	c=21.0380(6)
	alpha=90	beta=115.469(1)	gamma=90
Temperature:	187 K		
	Calculated	Reported	
Volume	10066.2(5)	10066.2(5)	
Space group	C 2	C 2	
Hall group	C 2y	C 2y	
Moiety formula	C ₄₇ H ₇₄ Cl Ir N O ₂ P S [+ solvent]	C ₄₇ H ₇₄ Cl Ir N O ₂ P S [+ solvent]	
Sum formula	C ₄₇ H ₇₄ Cl Ir N O ₂ P S [+ solvent]	C ₄₇ H ₇₄ Cl Ir N O ₂ P S	
Mr	975.77	975.75	
Dx, g cm ⁻³	1.288	1.288	
Z	8	8	
Mu (mm ⁻¹)	6.560	6.560	
F000	4032.0	4032.0	
F000'	4009.46		
h, k, lmax	54, 14, 25	54, 14, 25	
Nref	18554 [9765]	18311	
Tmin, Tmax	0.403, 0.455	0.525, 0.753	
Tmin'	0.304		

Correction method= # Reported T Limits: Tmin=0.525 Tmax=0.753
AbsCorr = MULTI-SCAN

Data completeness= 1.88/0.99 Theta (max) = 68.562

Crystal Structure Information of NMe-Xu3-Ir-BArF

0.1 mL of DCM was added to a 10 mL oven-dried glass sample bottle with 10 mg pure NMe-Xu3-Ir-BArF to dissolve the sample, then 8 mL n-hexane was slowly added to the solution, sealed with perforated paper, and then the solvent was slowly dried at room temperature to obtain crystals. Single crystal X-ray diffraction data were collected on Bruker Smart Apex II CCD diffractometer. The crystal structure has been deposited at the Cambridge Crystallographic Data Centre (CCDC): 2328618

Formula: $C_{79}H_{86}BF_{24}IrNO_2PS$

Flack parameter -0.012(3)

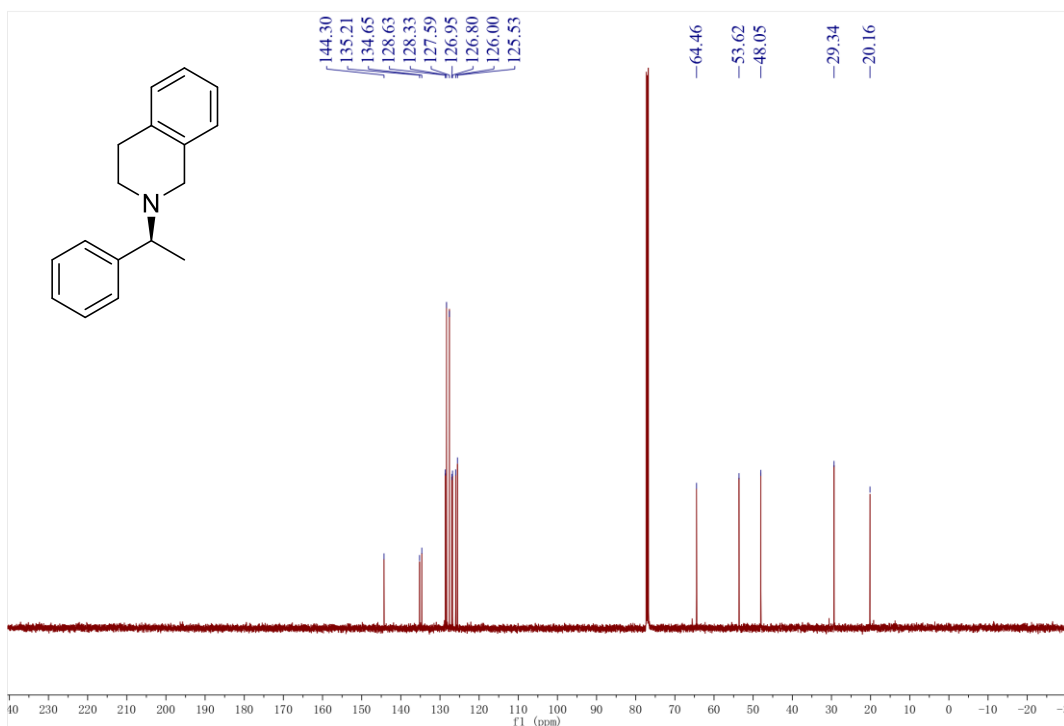
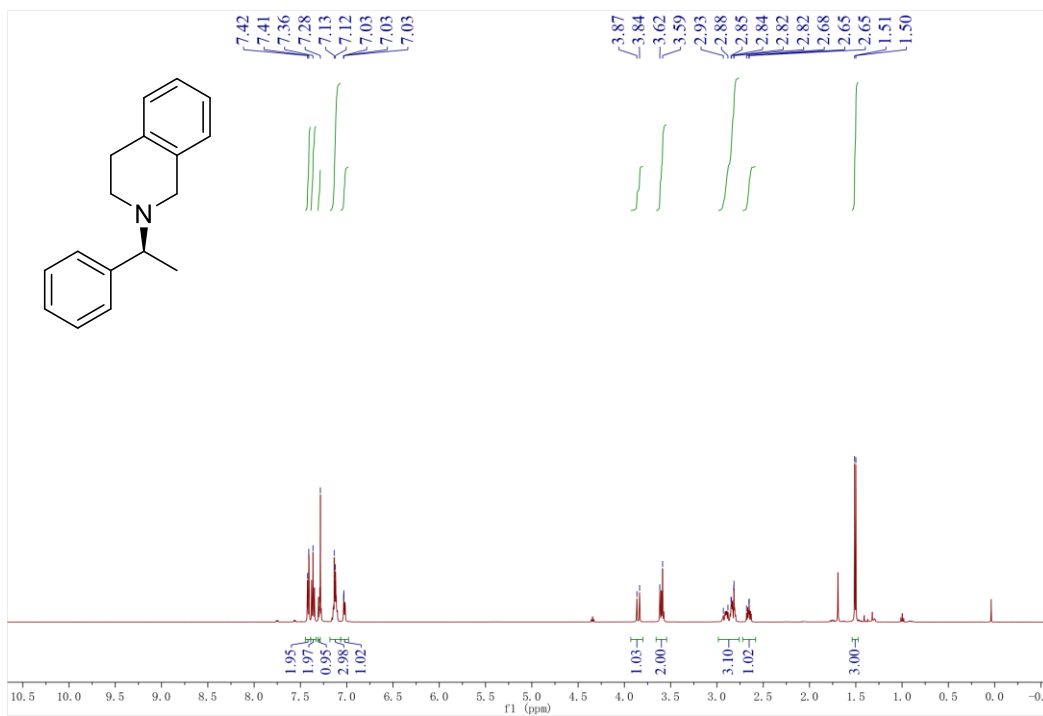
Bond precision:	C-C = 0.0105 Å	Wavelength=1.54184
Cell:	a=14.1597(1) b=14.8103(1) c=41.8365(4)	
	alpha=90 beta=90 gamma=90	
Temperature: 170 K		
	Calculated	Reported
Volume	8773.51(12)	8773.51(12)
Space group	P 21 21 21	P 21 21 21
Hall group	P 2ac 2ab	P 2ac 2ab
Moiety formula	C32 H12 B F24, C47 H74 Ir N O2 P S [+ solvent]	C47 H74 Ir N O2 P S, C32 H12 B F24
Sum formula	C79 H86 B F24 Ir N O2 P S [+ solvent]	C79 H86 B F24 Ir N O2 P S
Mr	1803.55	1803.52
Dx, g cm ⁻³	1.365	1.365
Z	4	4
Mu (mm ⁻¹)	4.157	4.157
F000	3648.0	3648.0
F000'	3644.32	
h, k, lmax	16, 17, 49	16, 17, 49
Nref	15652[8599]	15626
Tmin, Tmax	0.218, 0.633	0.198, 1.000
Tmin'	0.140	
Correction method= # Reported T Limits: Tmin=0.198 Tmax=1.000 AbsCorr = MULTI-SCAN		
Data completeness= 1.82/1.00	Theta(max)= 67.078	
R(reflections)= 0.0373(15445)	wR2(reflections)= 0.0982(15626)	
S = 1.022	Npar= 1002	

4. References

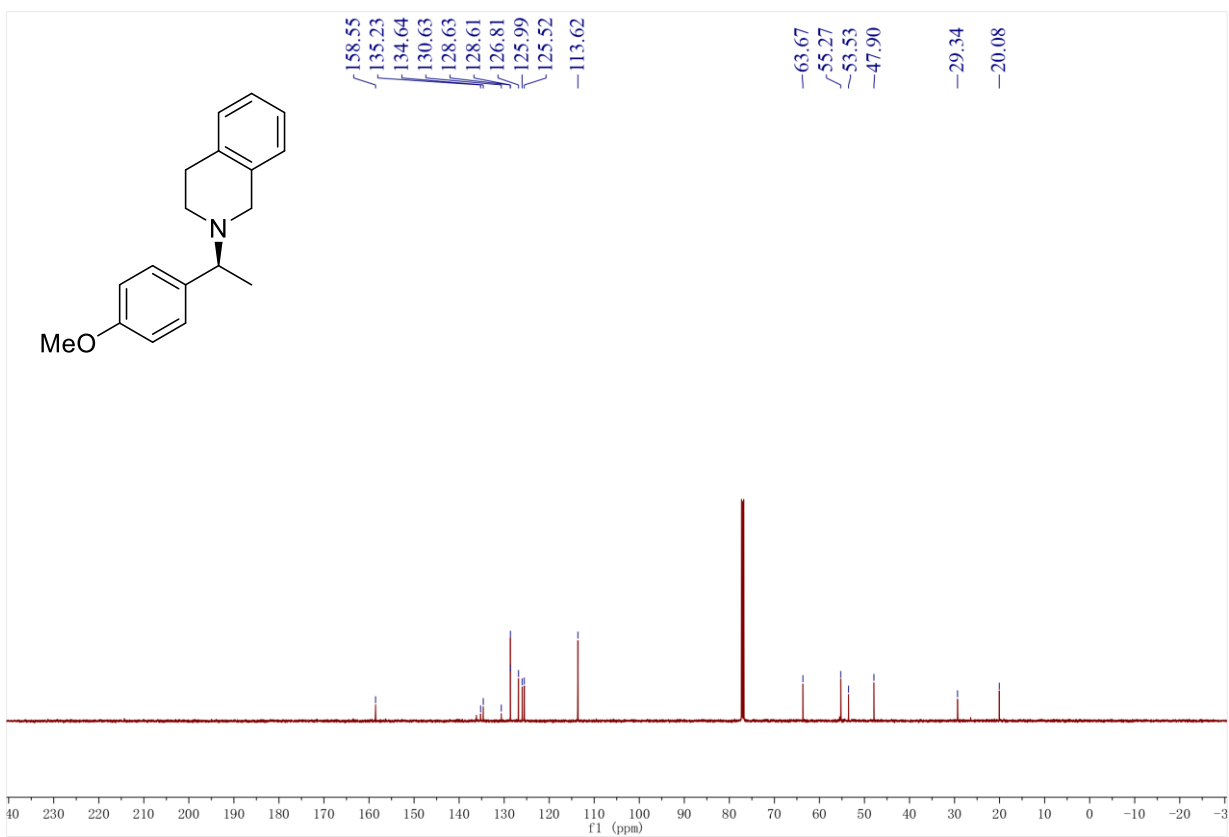
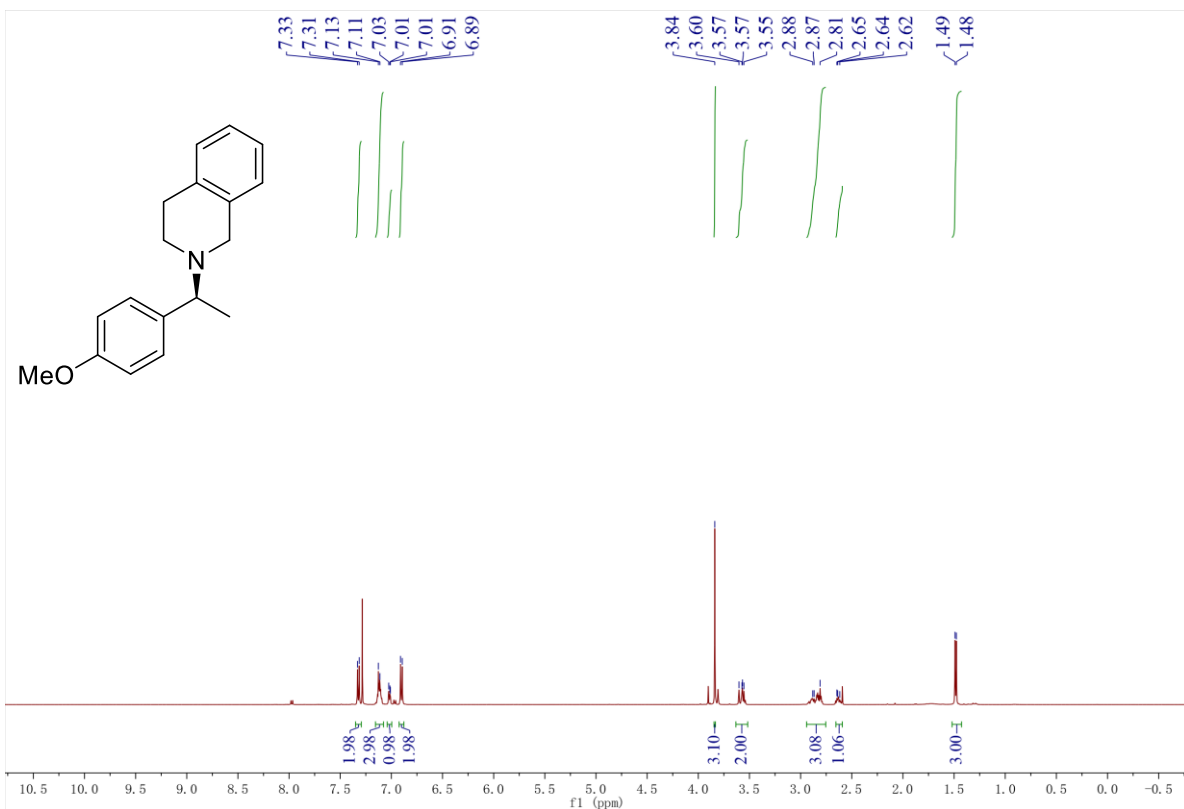
- [1] Wang, Y.; Wang, L. Chen, M.; Tu, Y.; Liu, Y.; Zhang, J. Palladium/Xu-Phos-catalyzed asymmetric carboamination towards isoxazolidines and pyrrolidines. *Chem. Sci.*, **2021**, *12*, 8241 - 8245.
- [2] Wu, Z.; Du, S.; Gao, G.; Yang, W.; Yang, X.; Huang, H.; Chang, M. Secondary Amines as Coupling Partners in Direct Catalytic Asymmetric Reductive Amination, *Chem. Sci.*, **2019**, *10*, 4509 - 4514.
- [3] Benmekhbi, L.; Louafi, F.; Roisnel, T.; Hurvois, J.-P. Synthesis of Tetrahydroisoquinoline Alkaloids and Related Compounds through the Alkylation of Anodically Prepared α -Amino Nitriles. *J. Org. Chem.*, **2016**, *81*, 6721 - 6739.
- [4] Kumpaty, H. J.; Bhattacharyya, S. Efficient synthesis of N-alkyl tetrahydroisoquinolines by reductive amination, *Synthesis*, **2005**, *13*, 2205 - 2209.
- [5] Wen, Y.; Fernández-Sabaté, M.; Lledós, A.; Sciortino, G.; Eills, J.; Marco-Rius, I.; Riera, A.; Verdaguer, X. Cyclometallated Imides as Templates for the H-Bond Directed Iridium-Catalyzed Asymmetric Hydrogenation of N-Methyl, N-Alkyl and N-Aryl Imines. *Angew. Chem. Int. Ed.*, **2024**, *63*, e202404955.

5. NMR Spectra of Hydrogenation Products

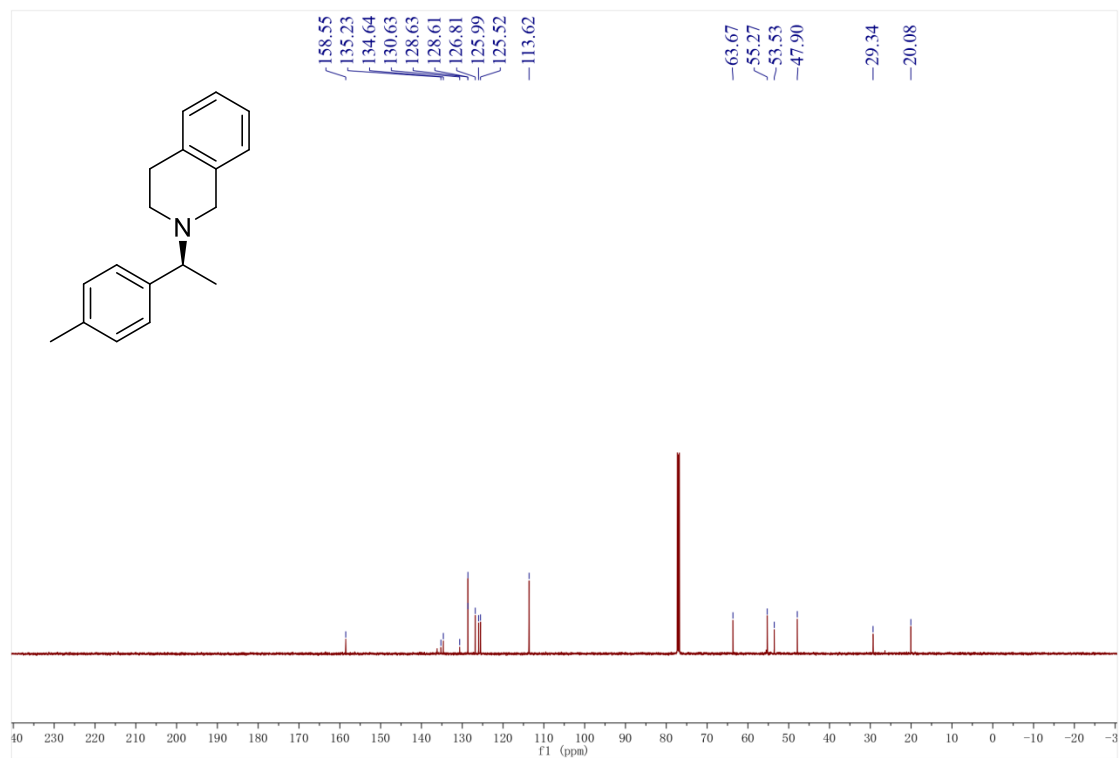
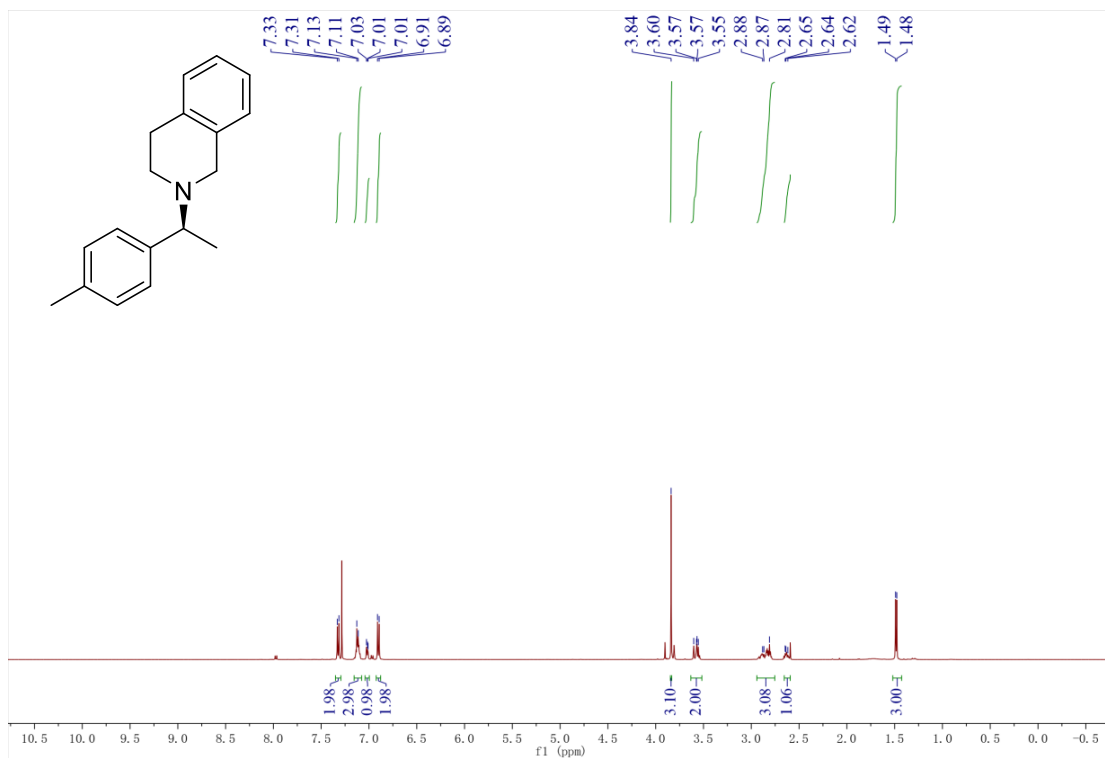
(S)- 3a



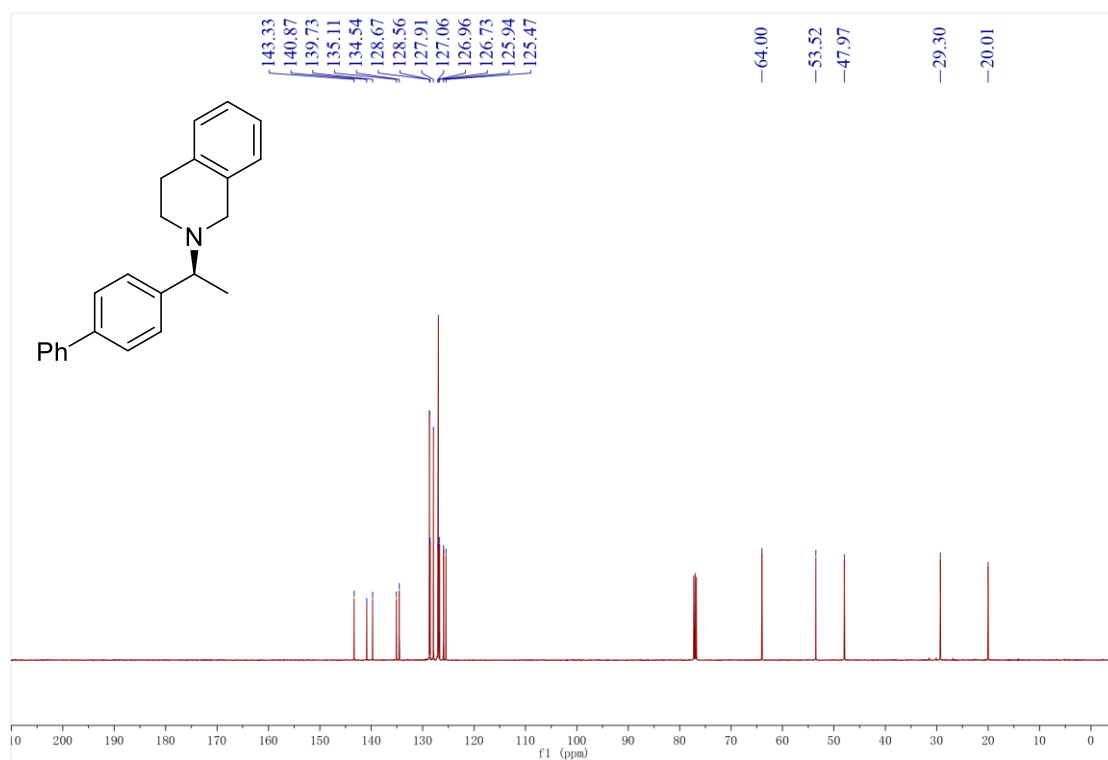
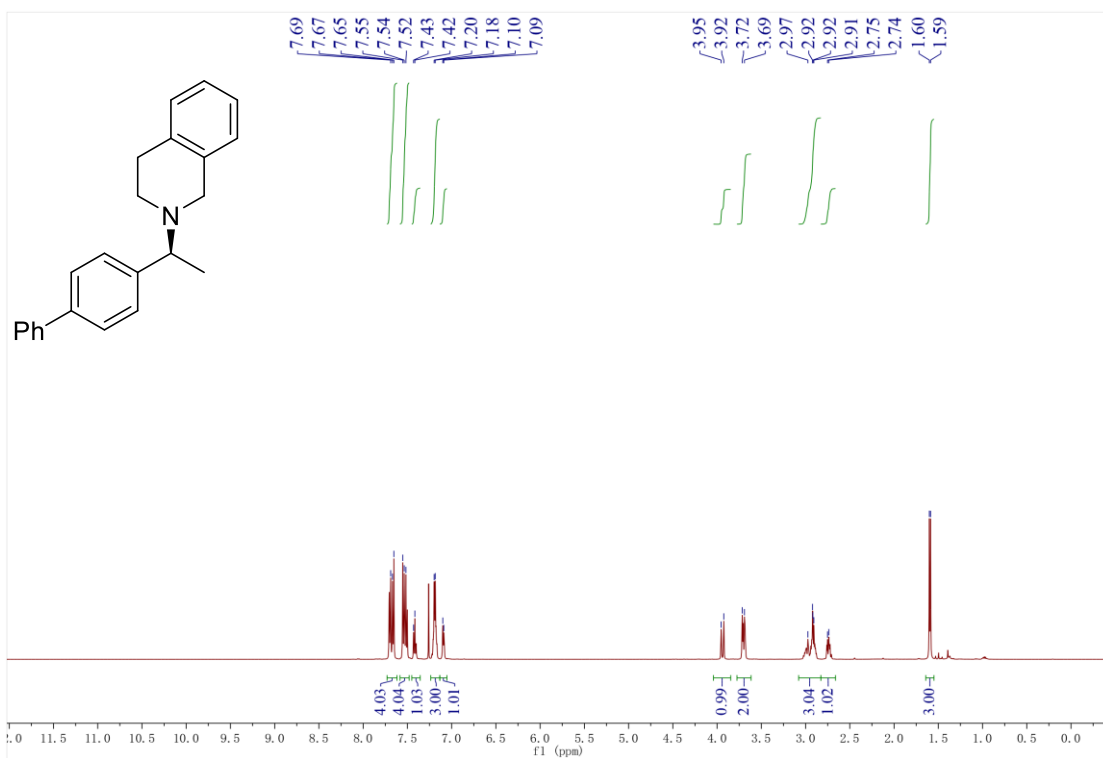
(S)-3b



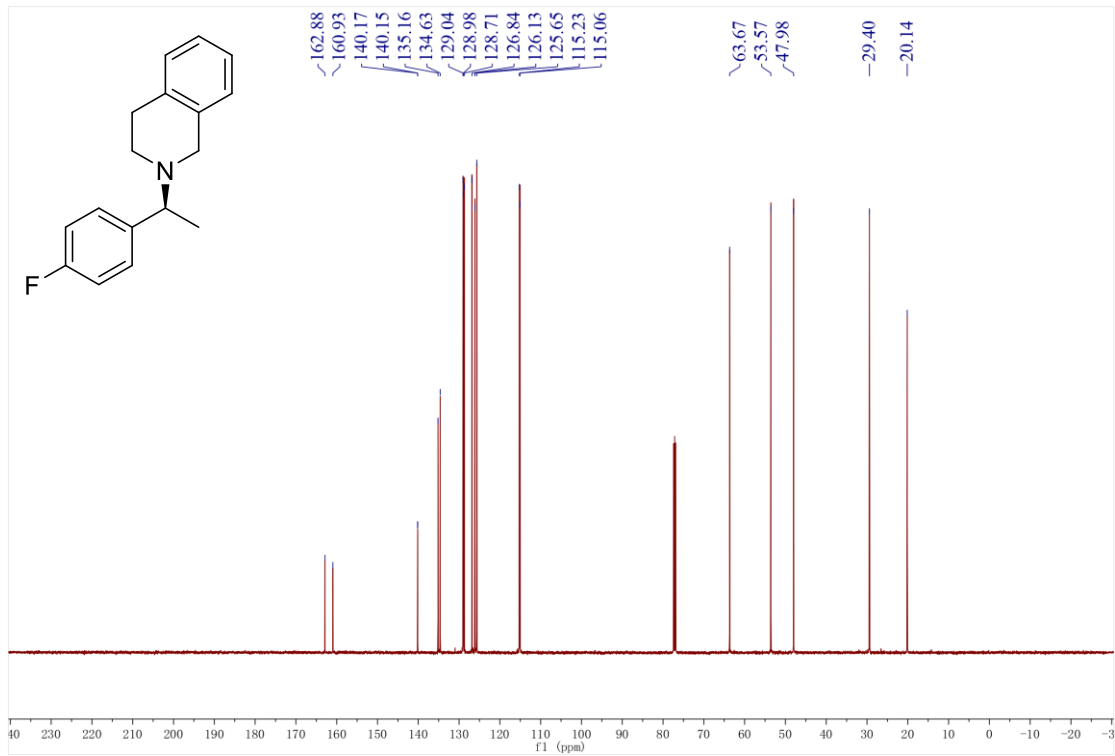
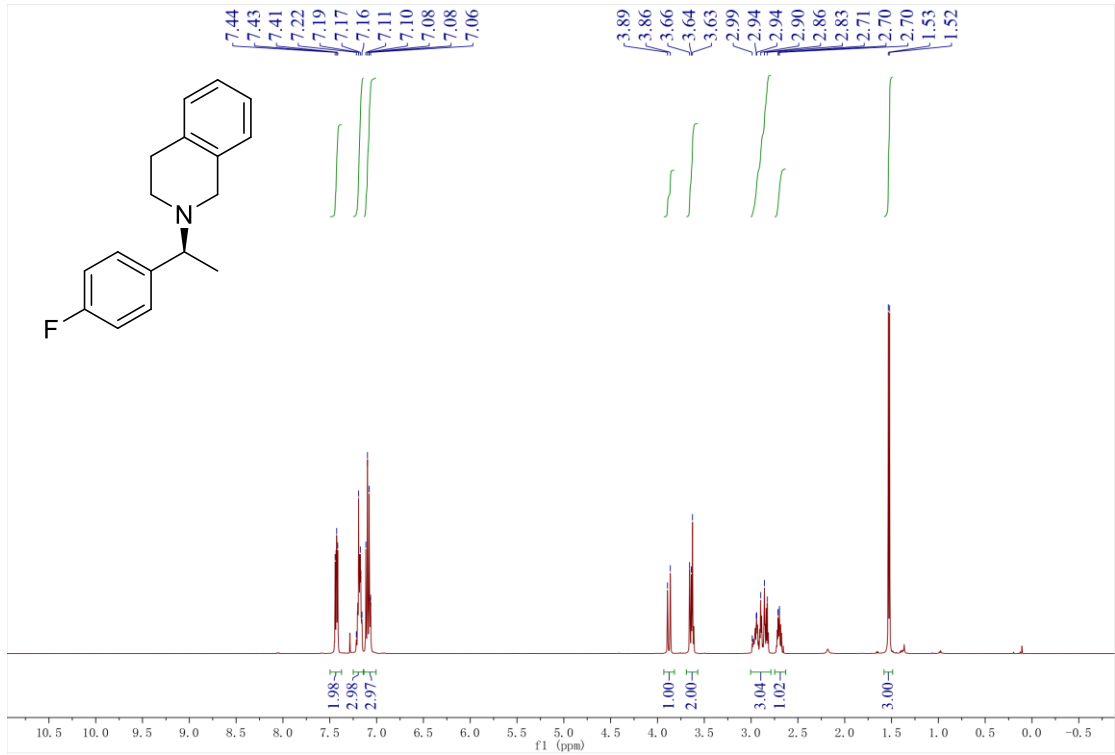
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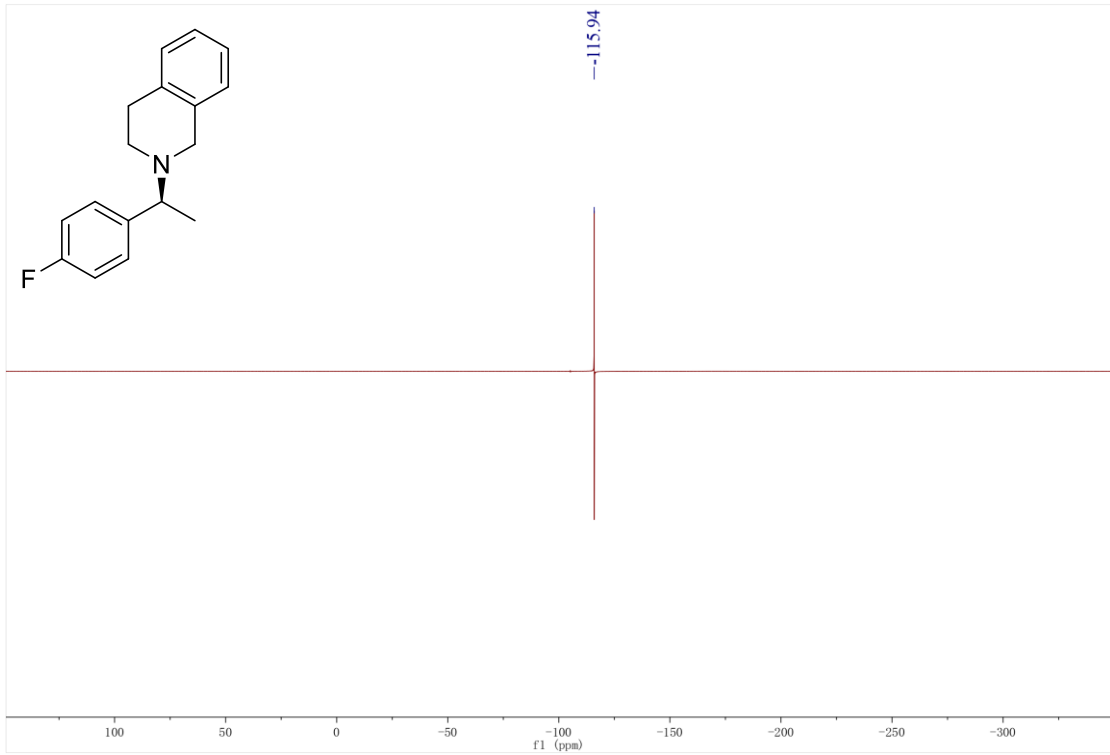


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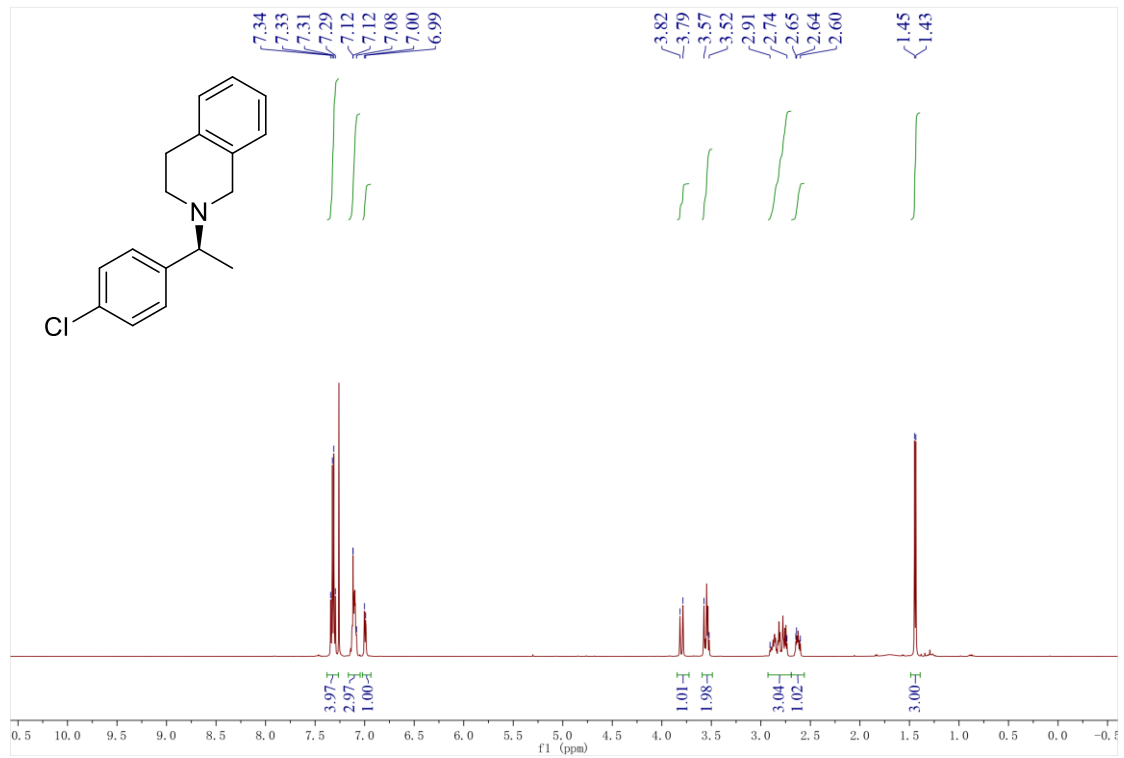


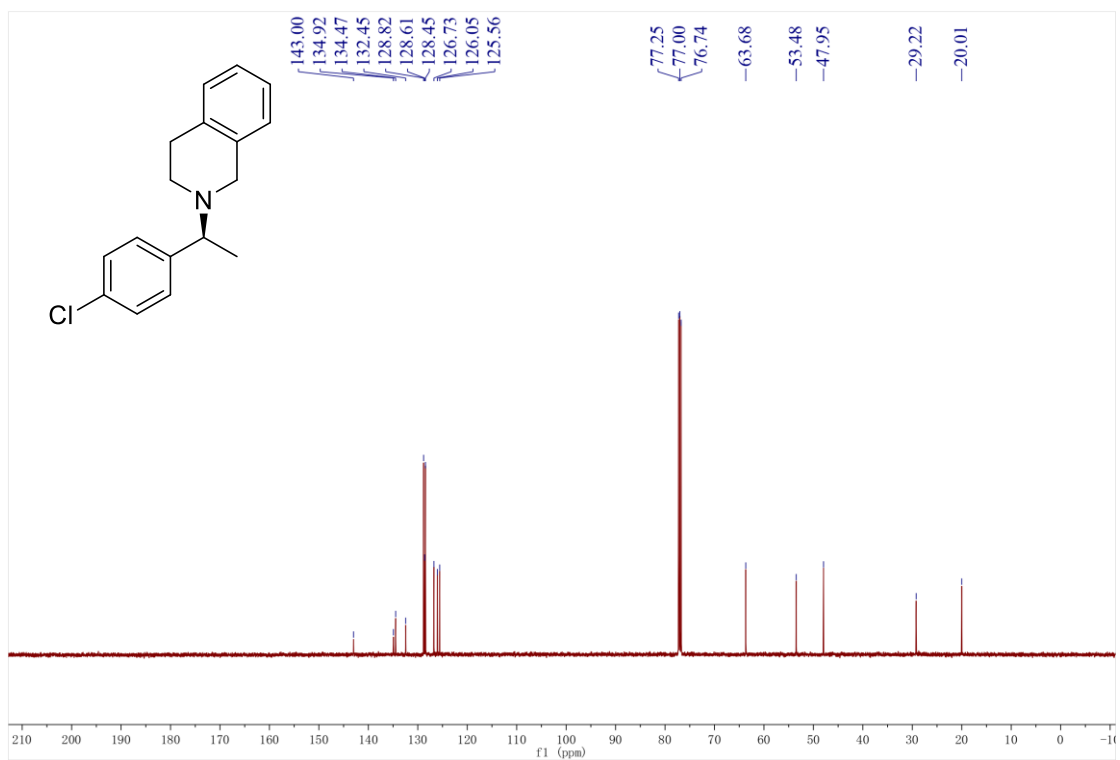
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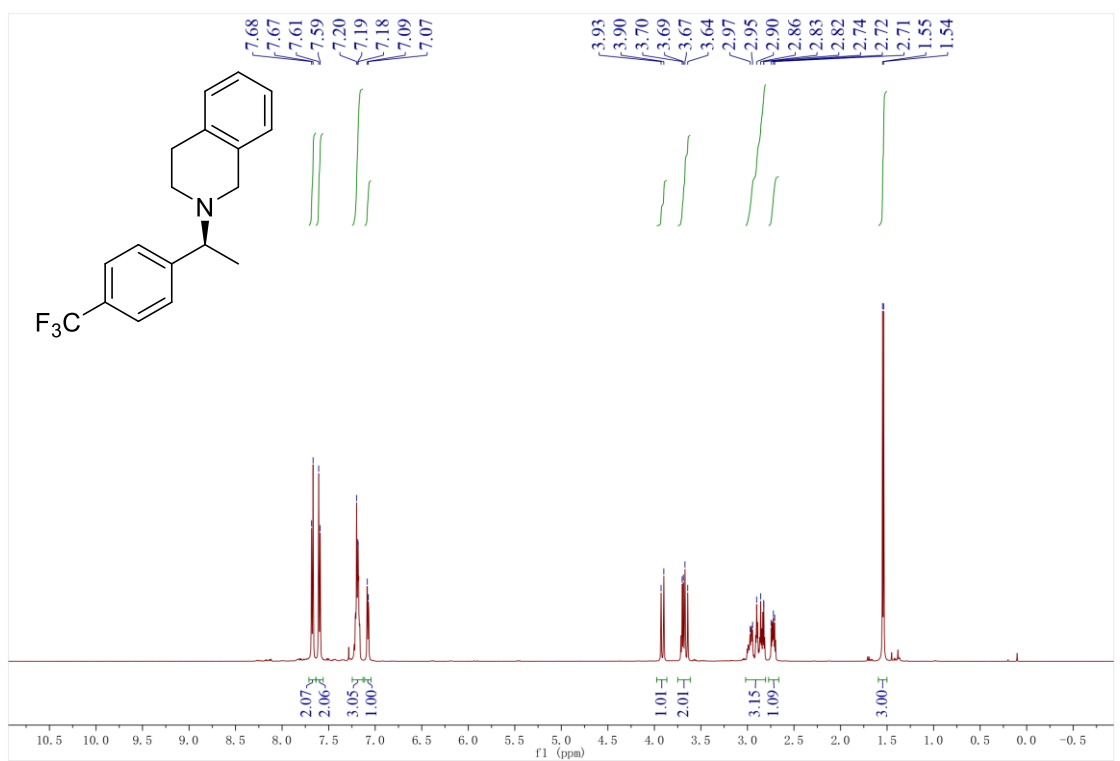


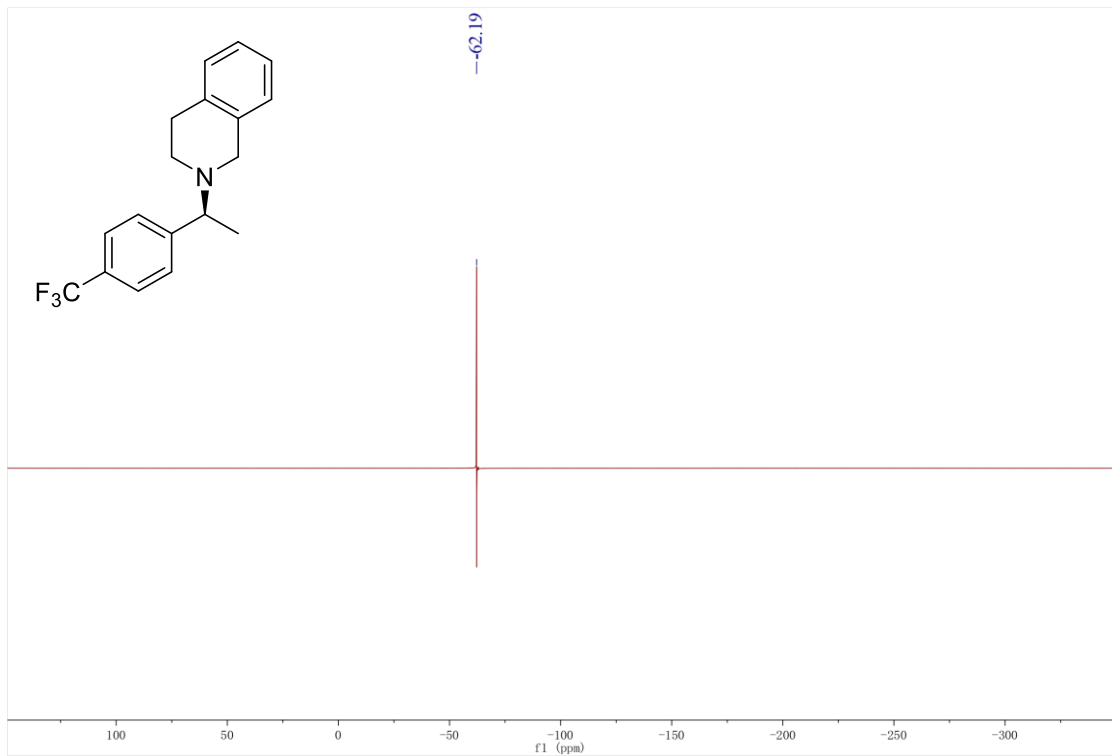
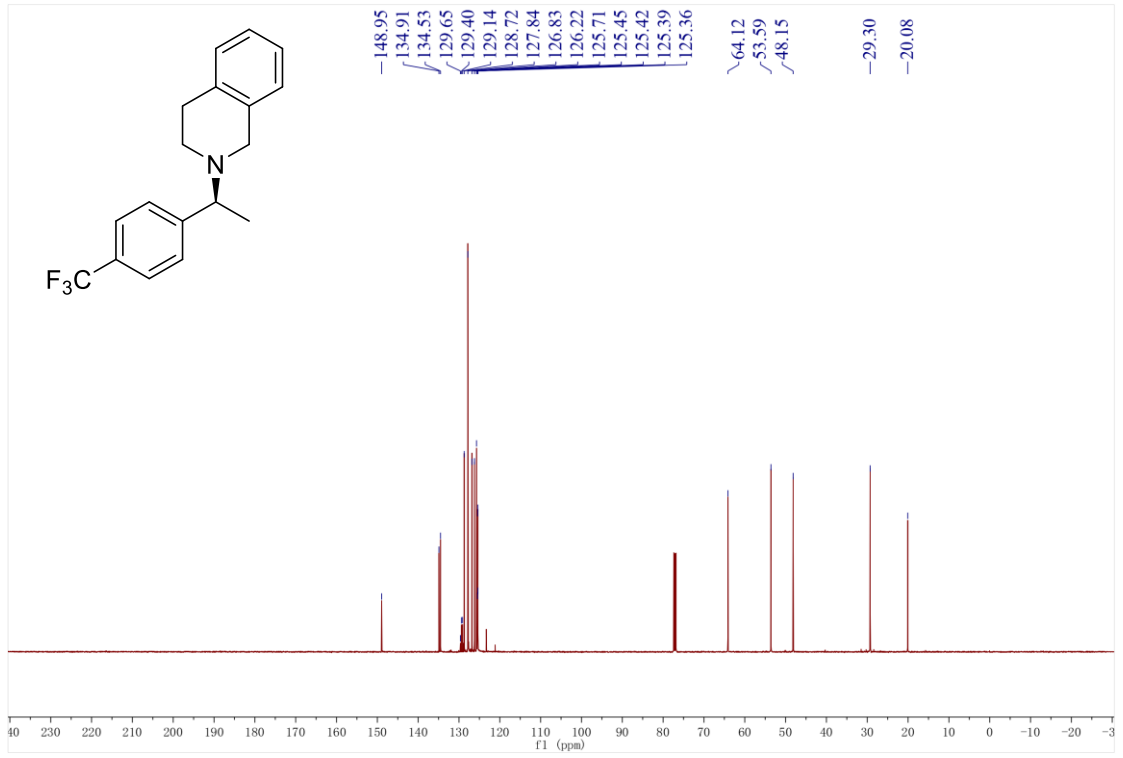
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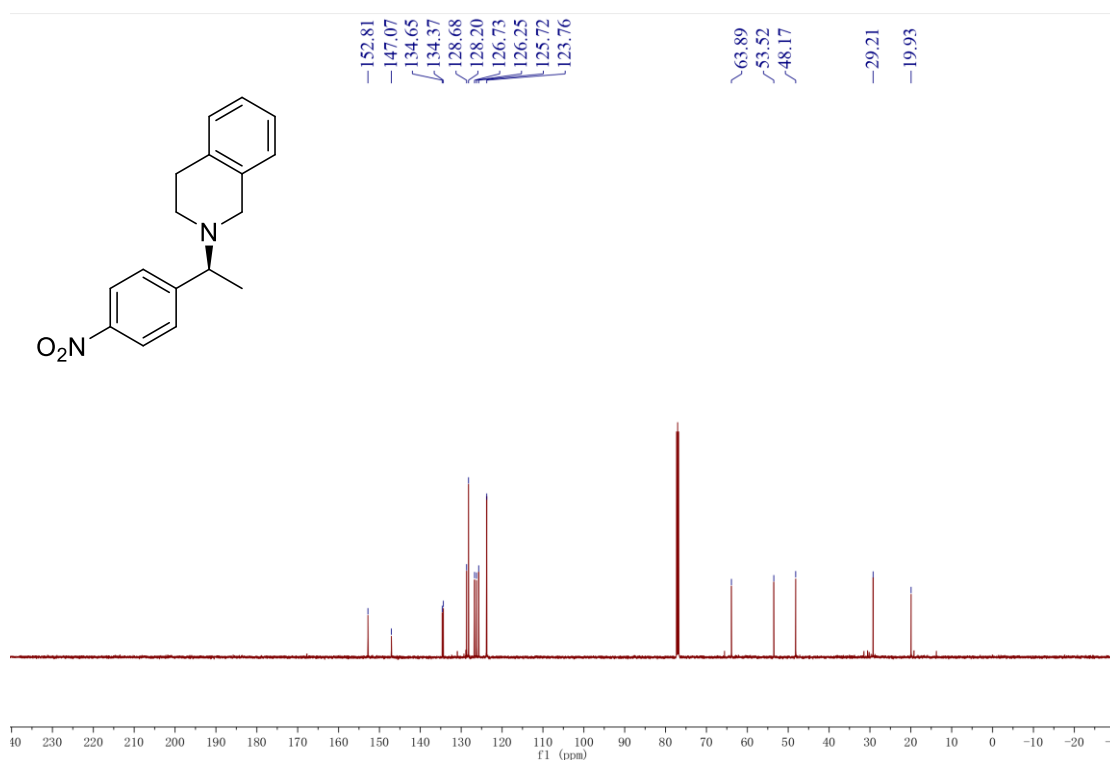
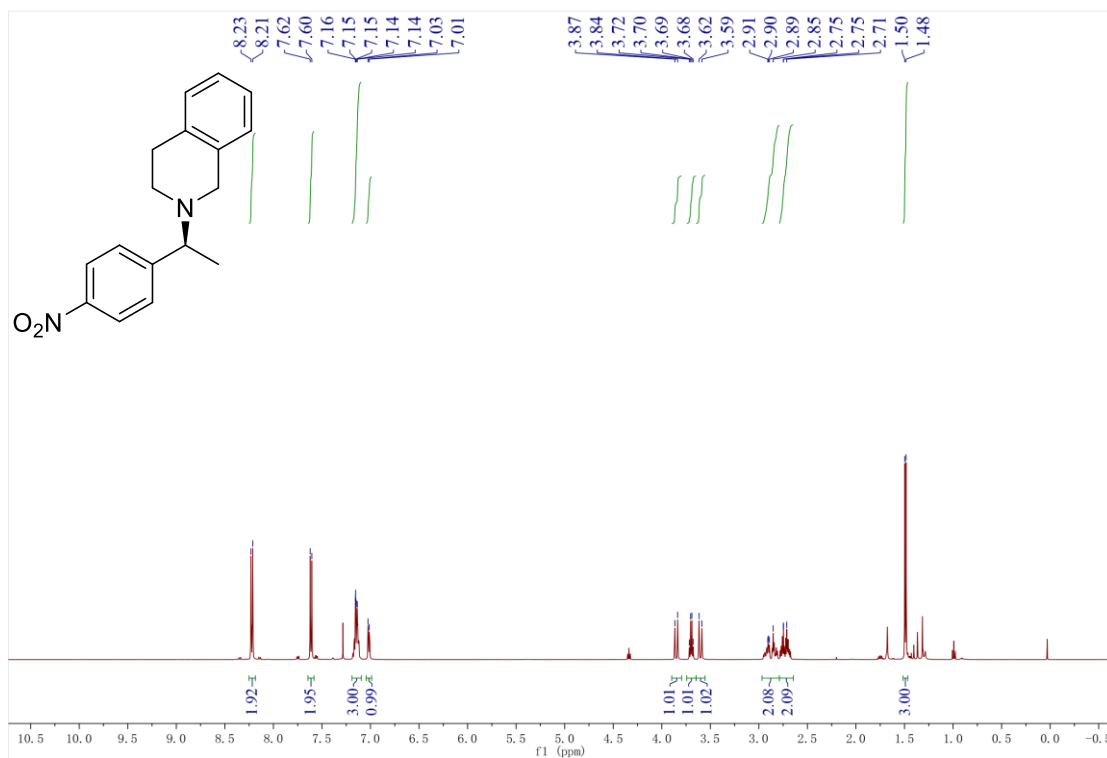


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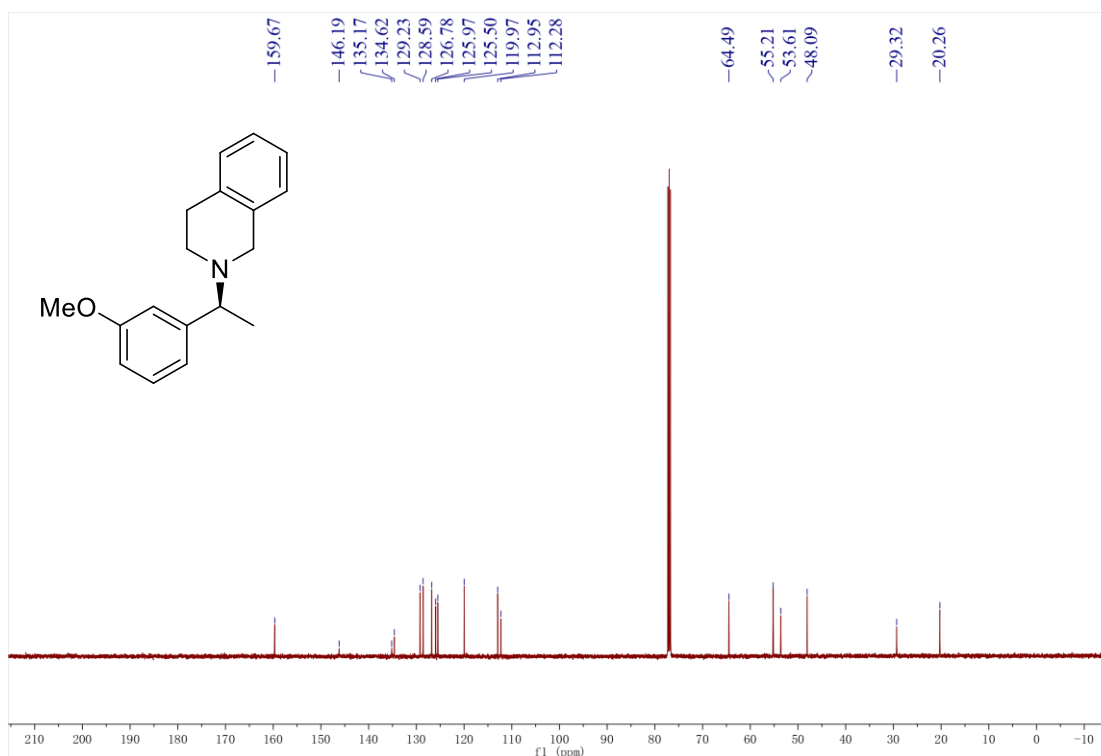
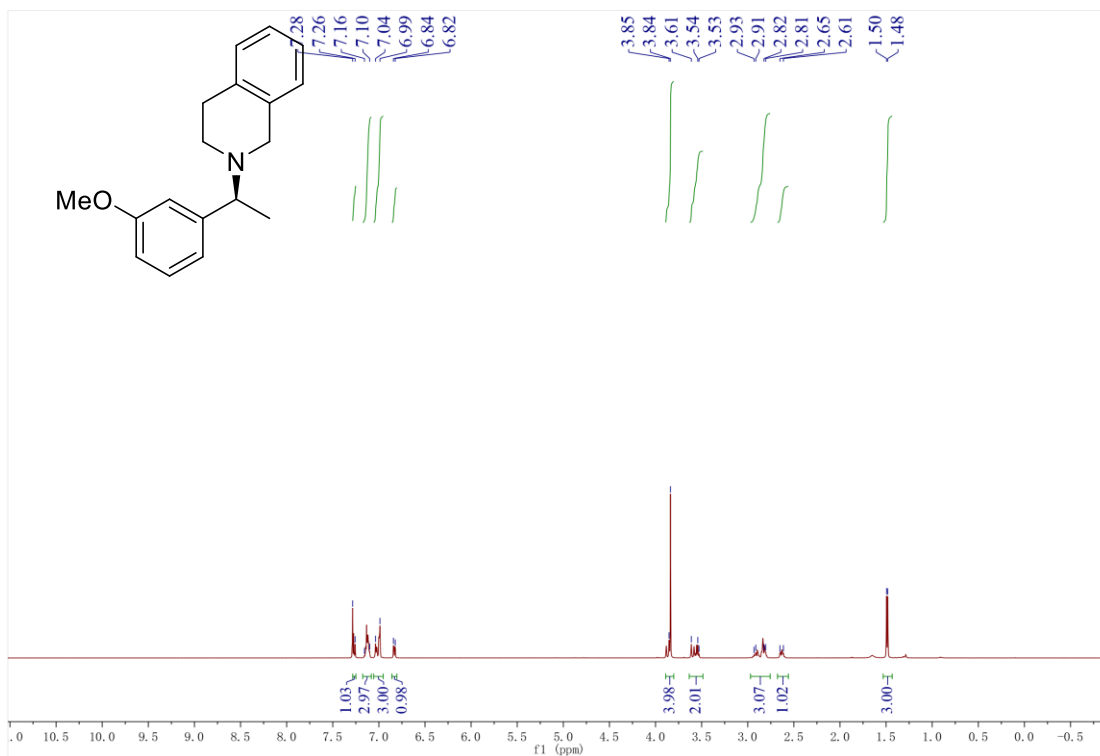




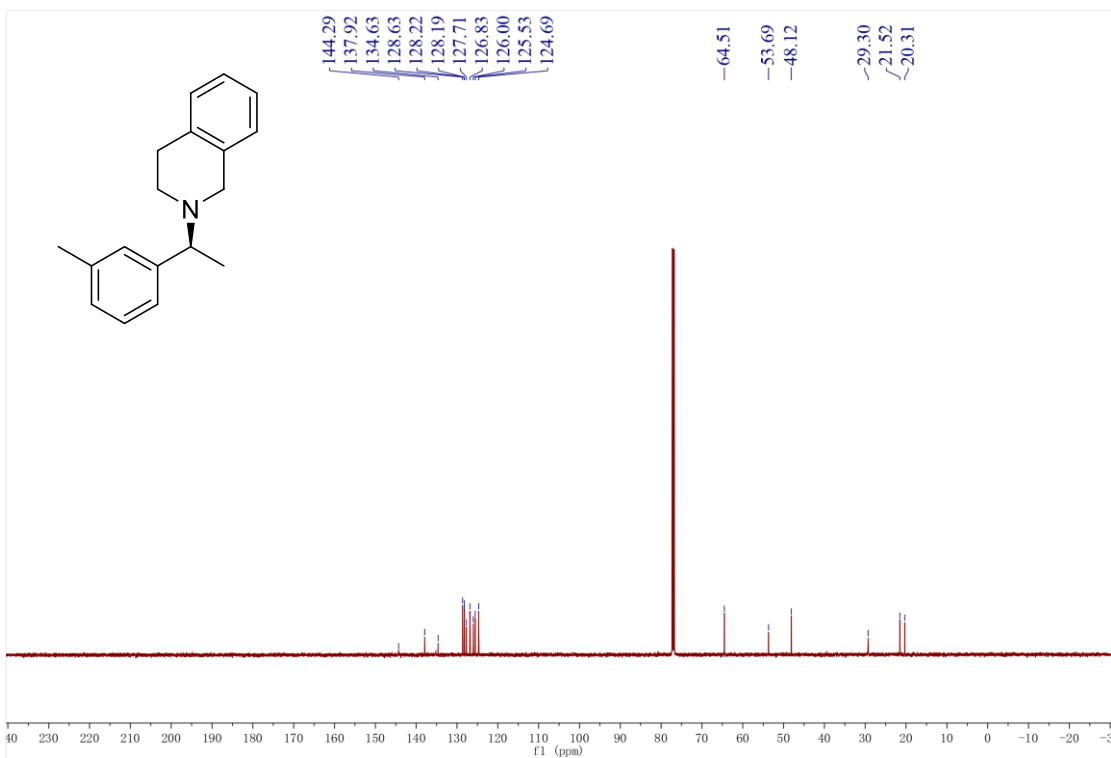
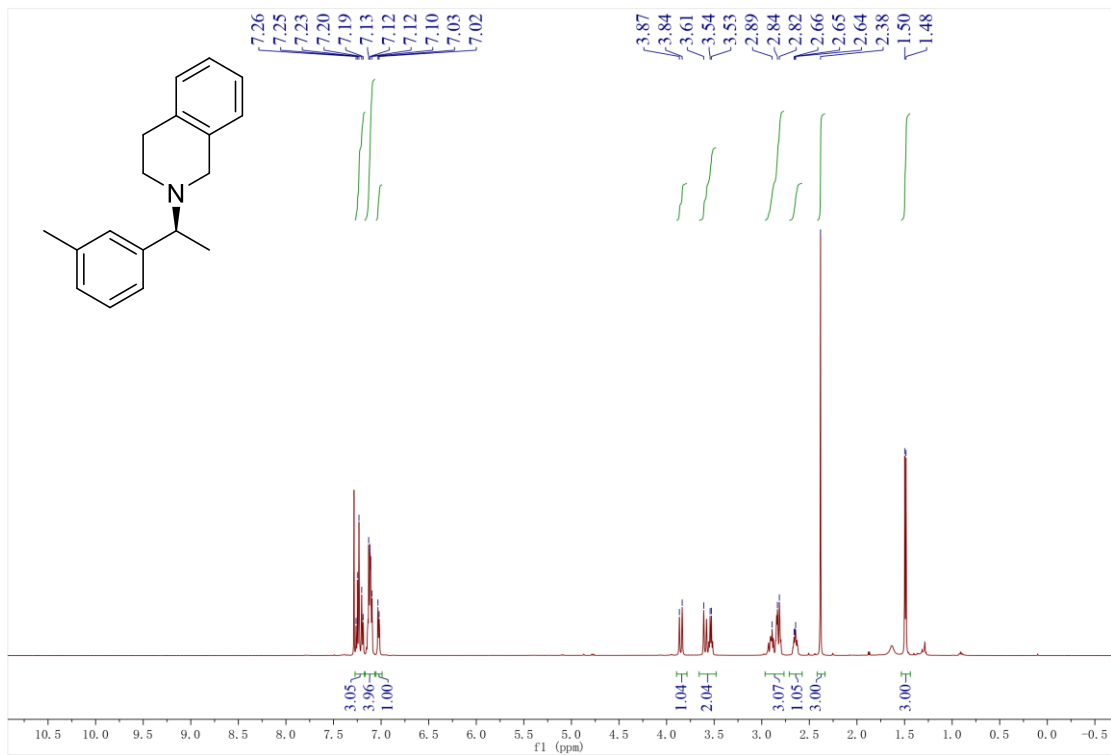
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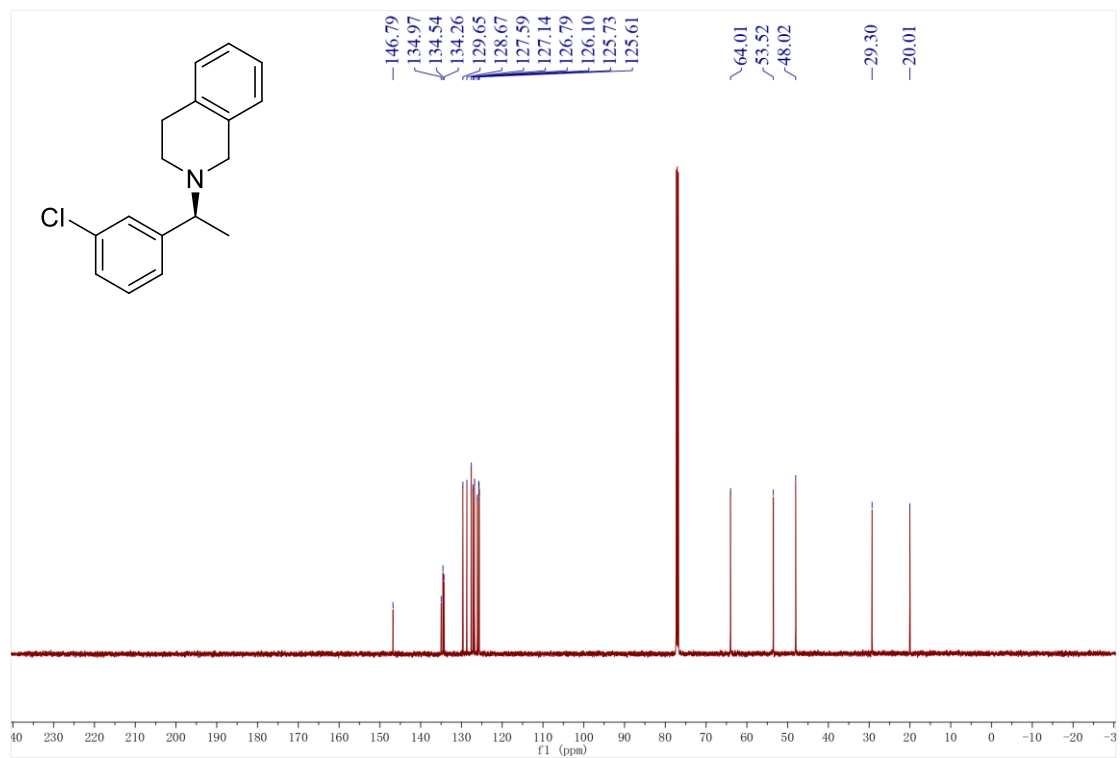
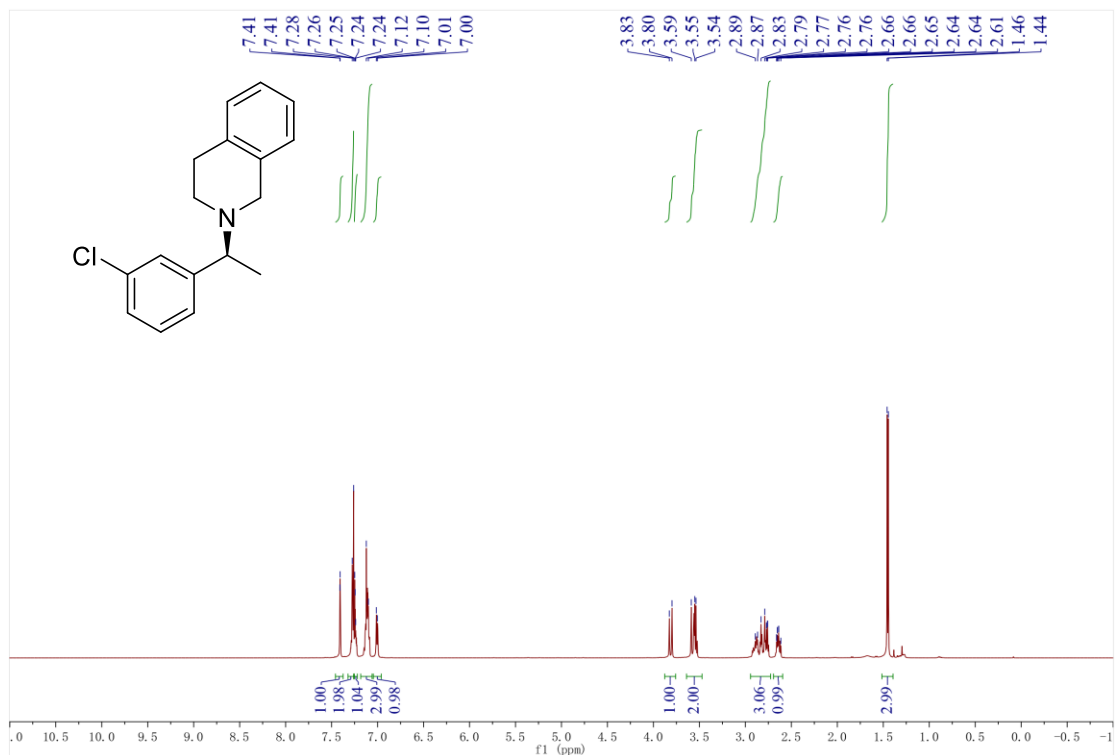
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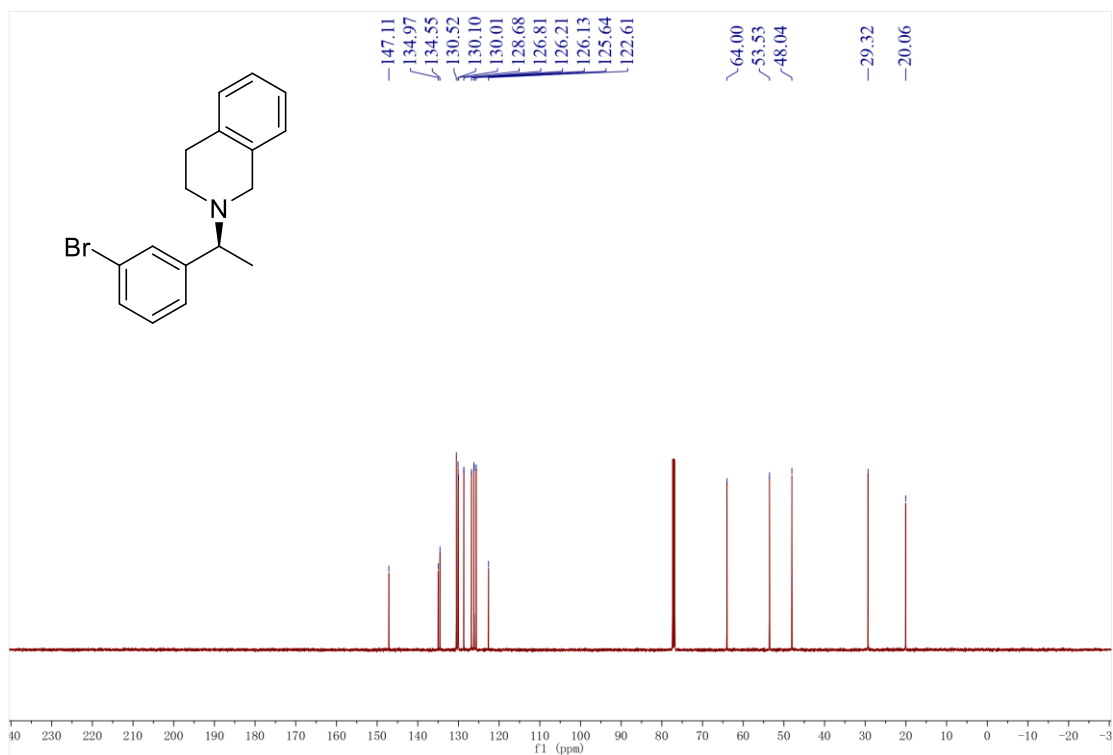
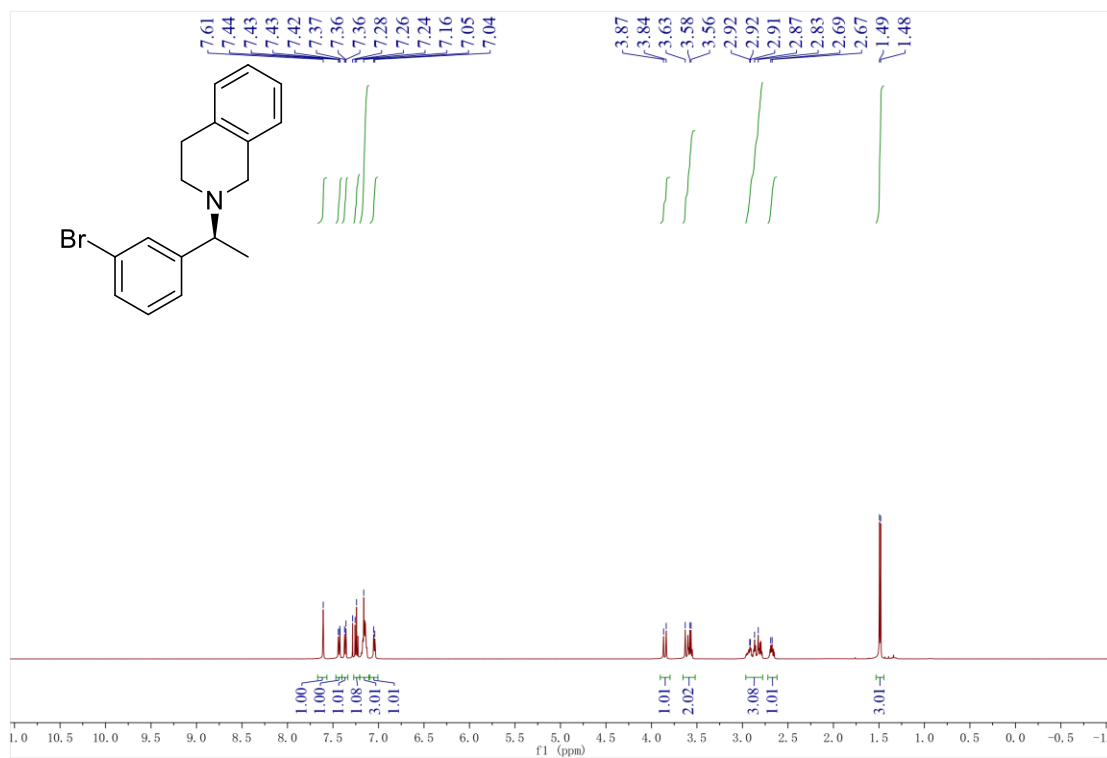
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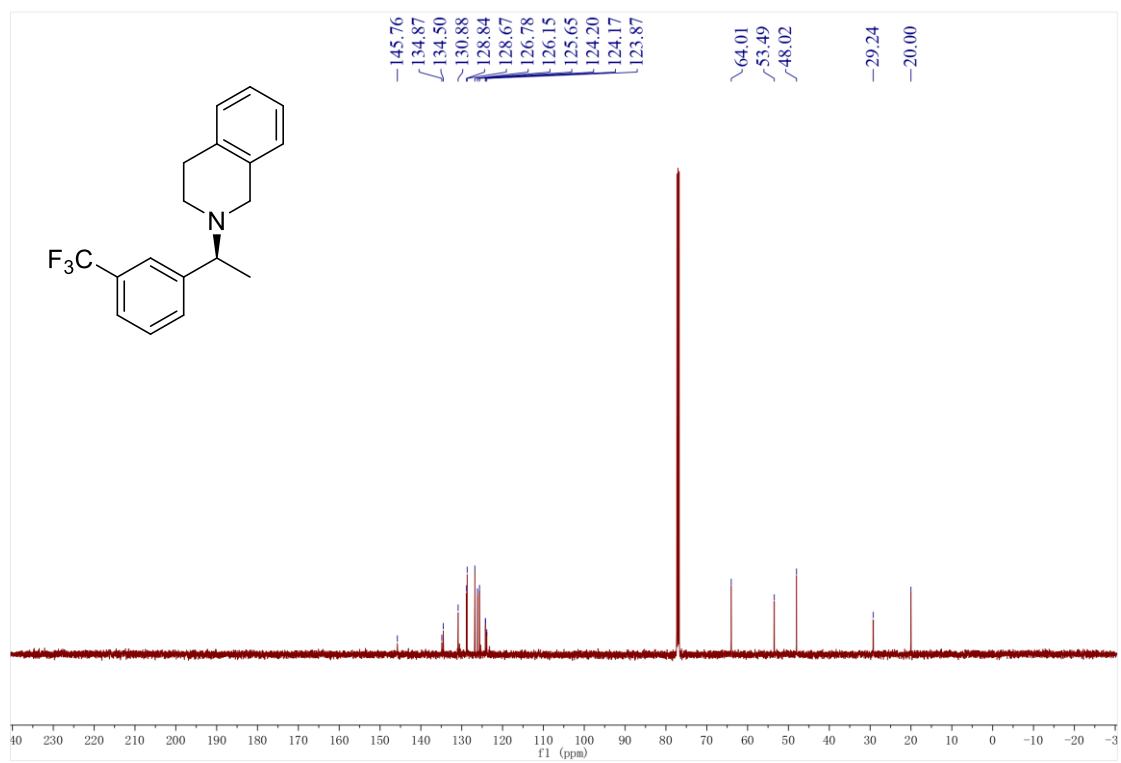
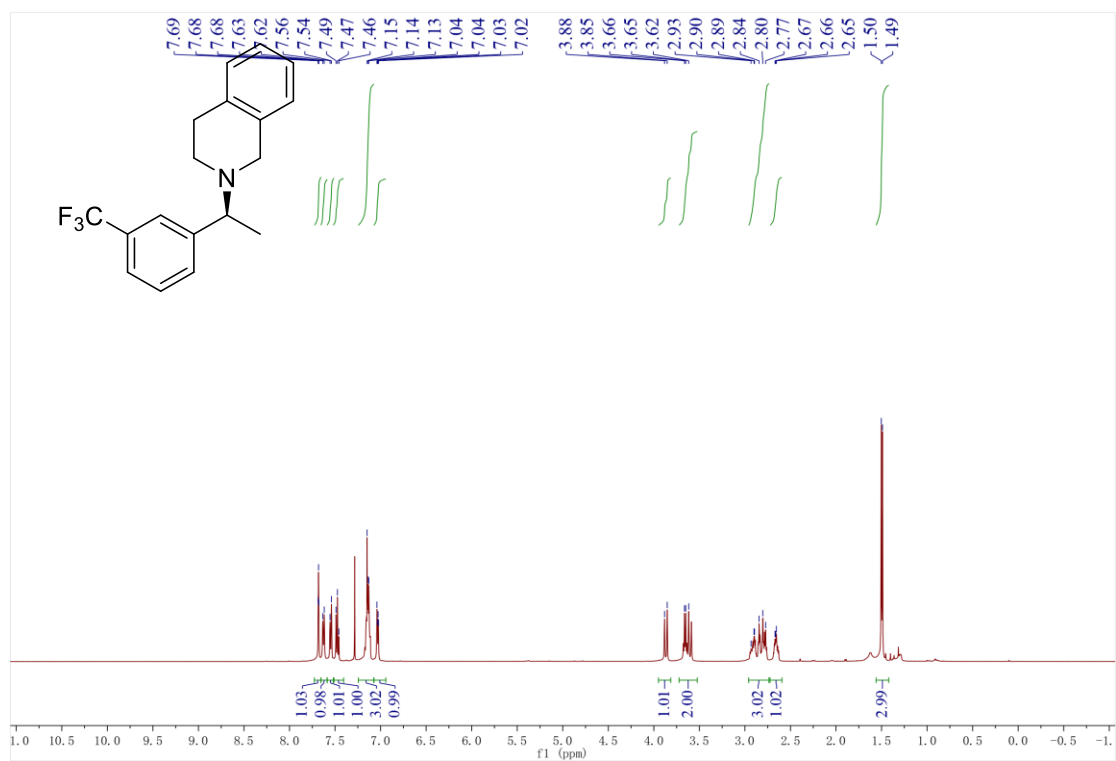
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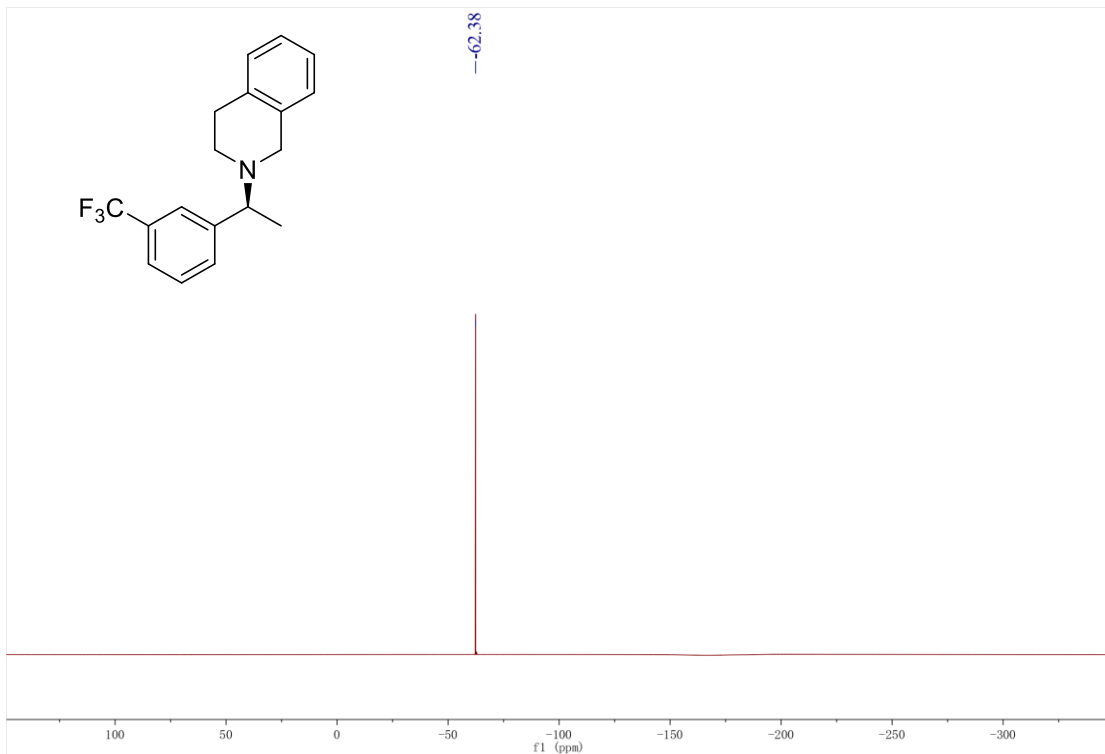


(S)-31

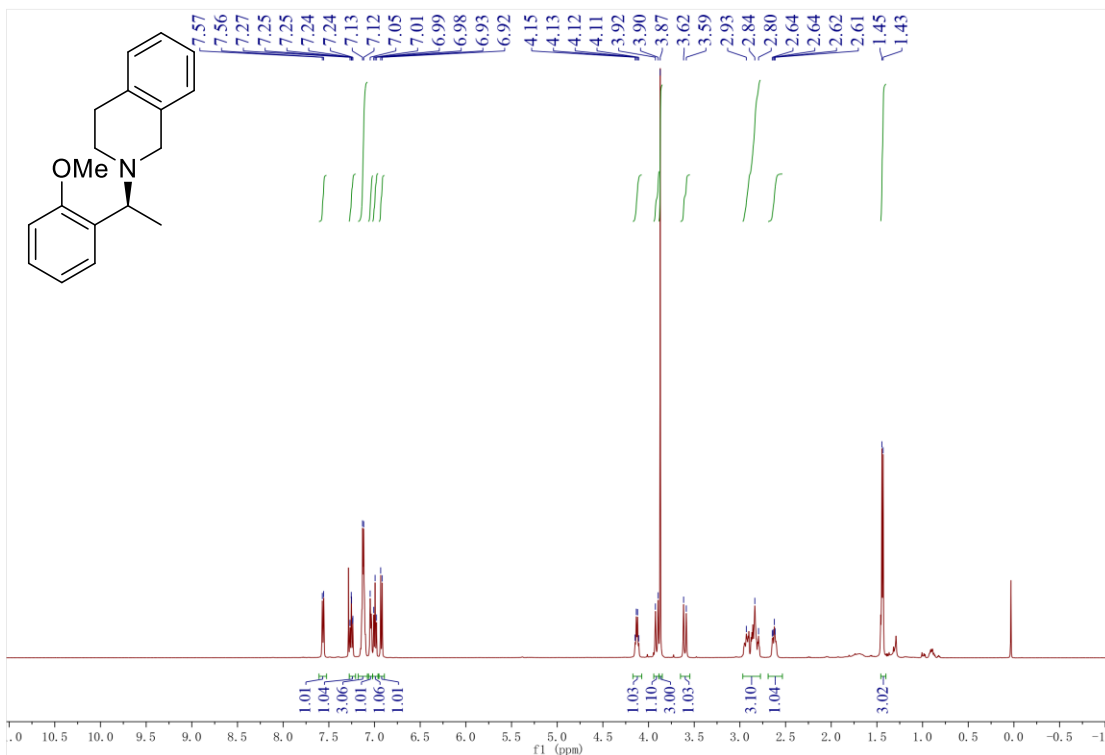


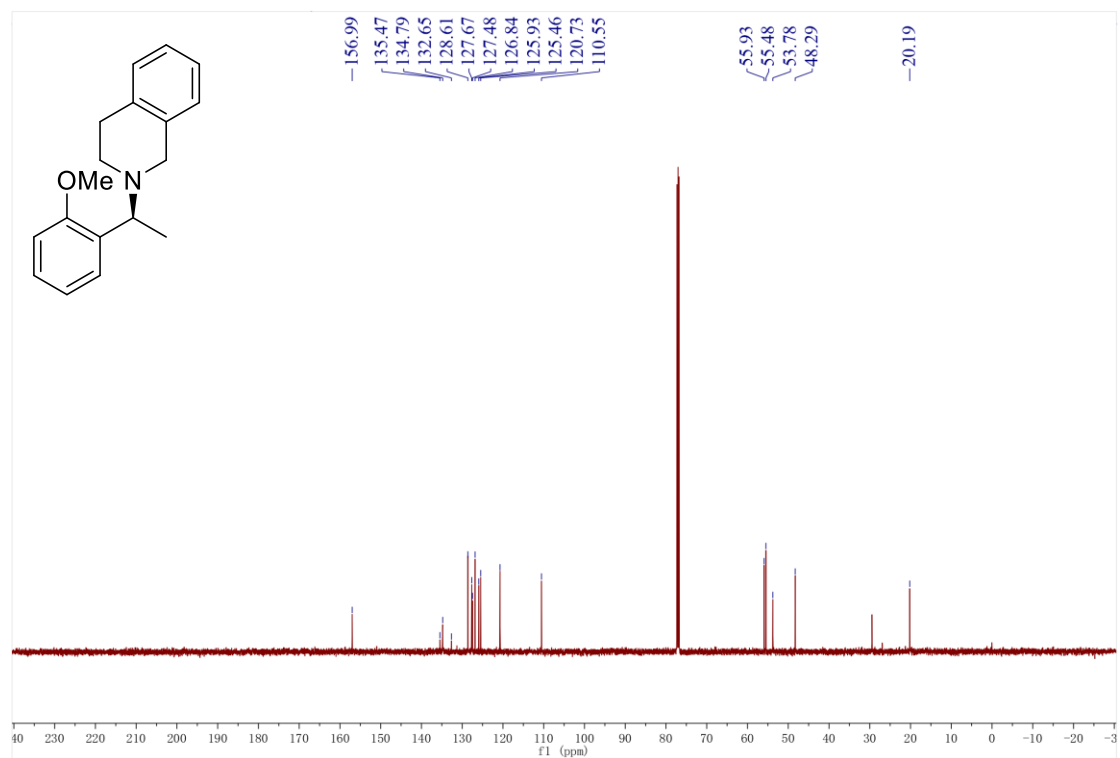
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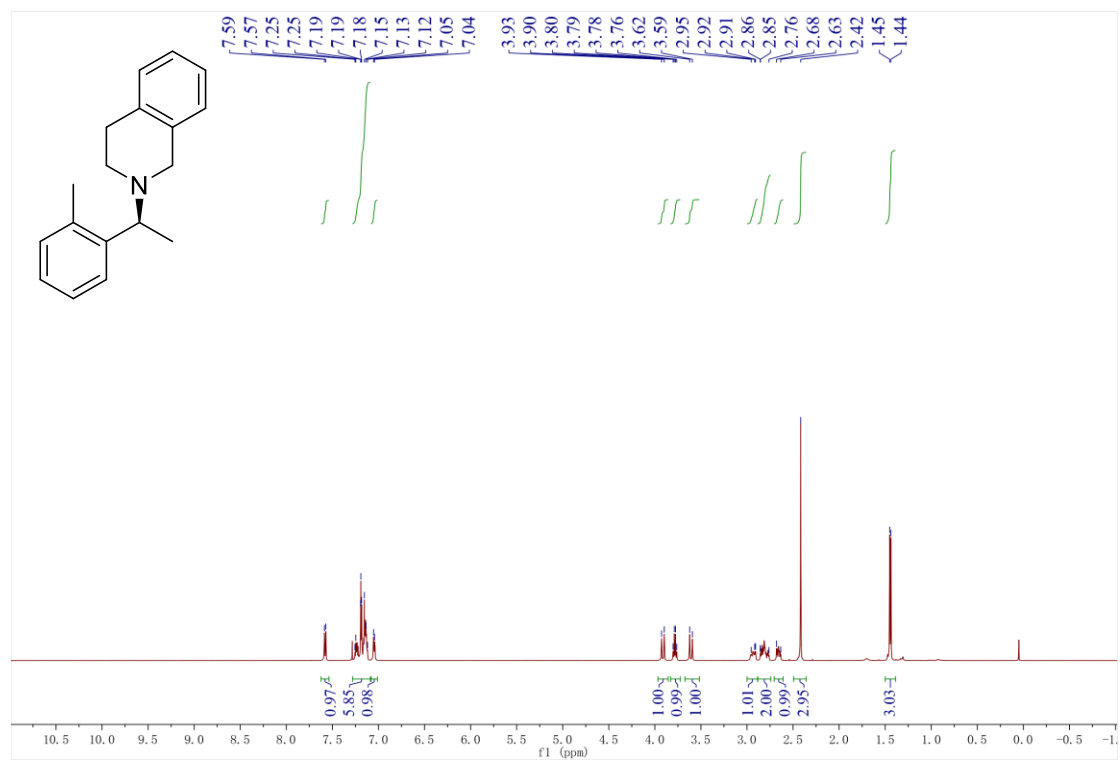


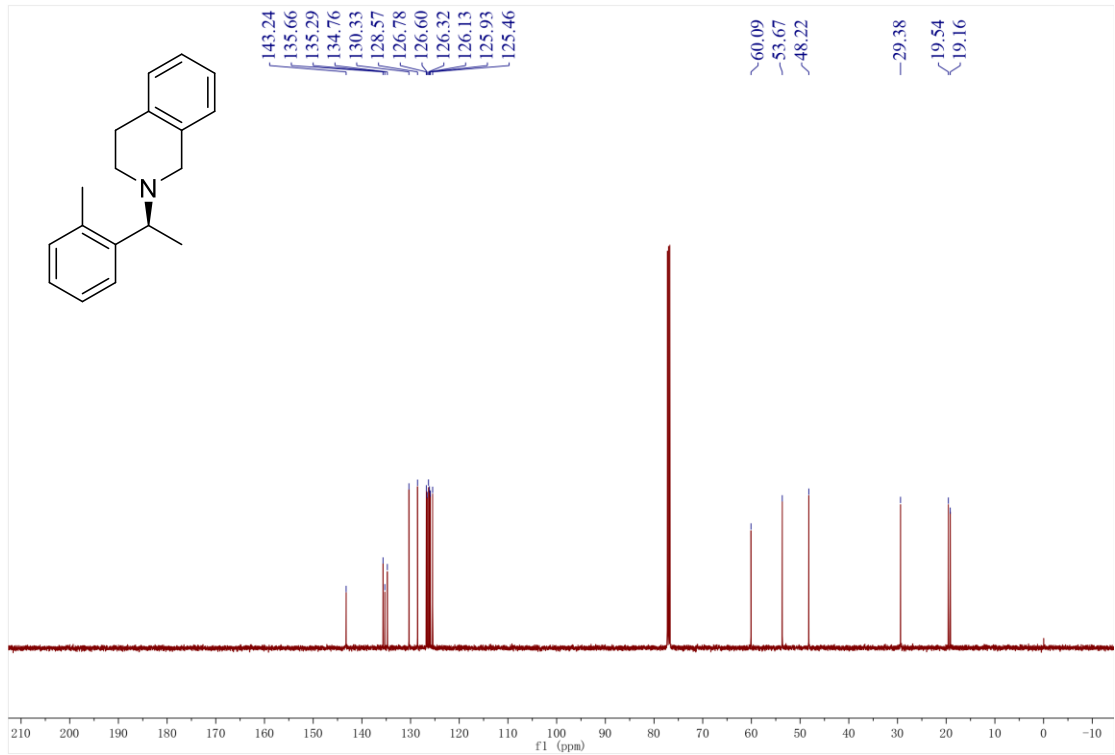
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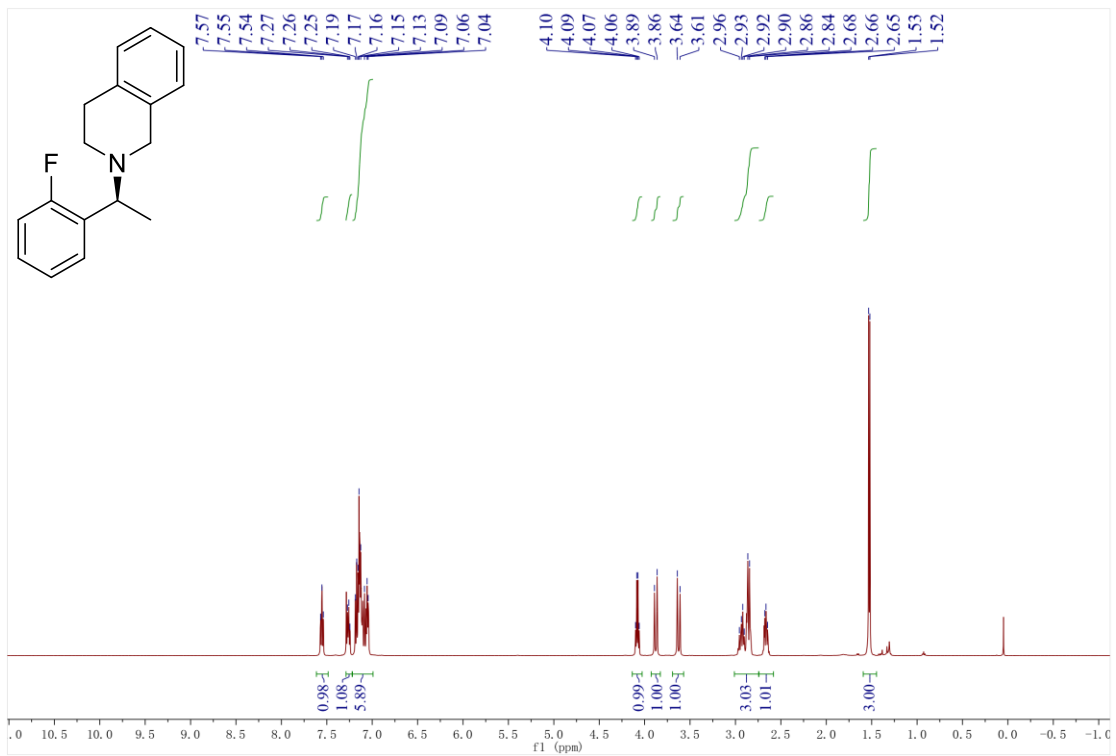


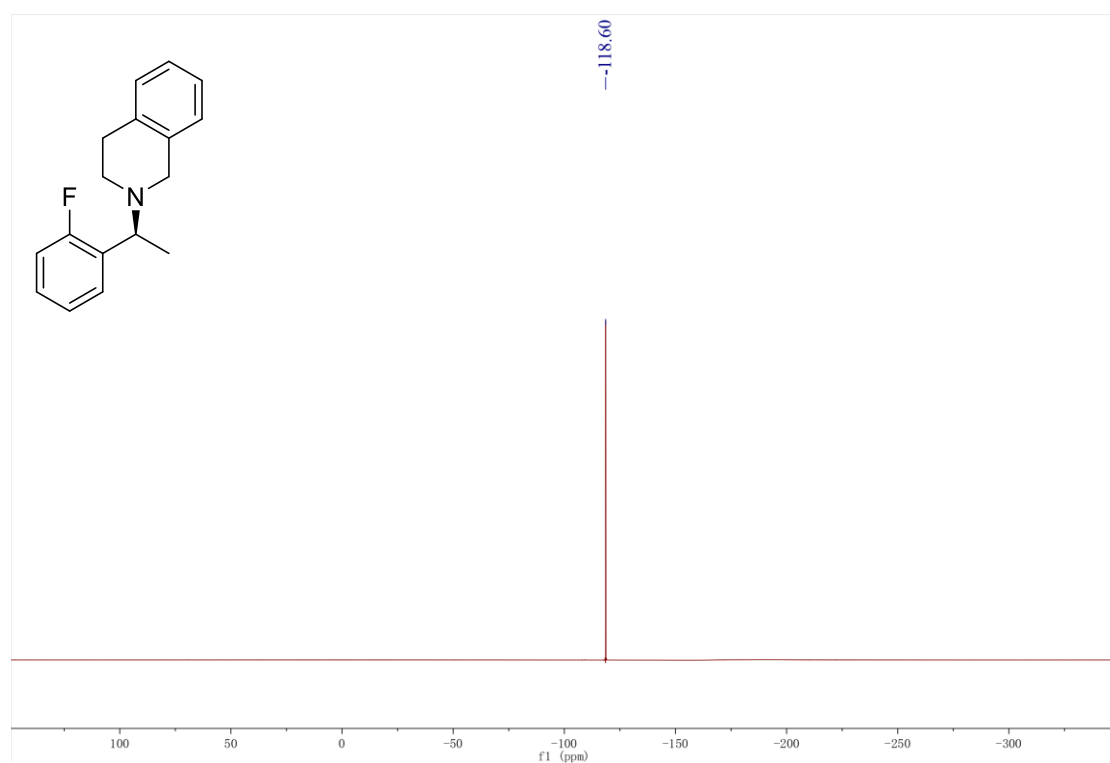
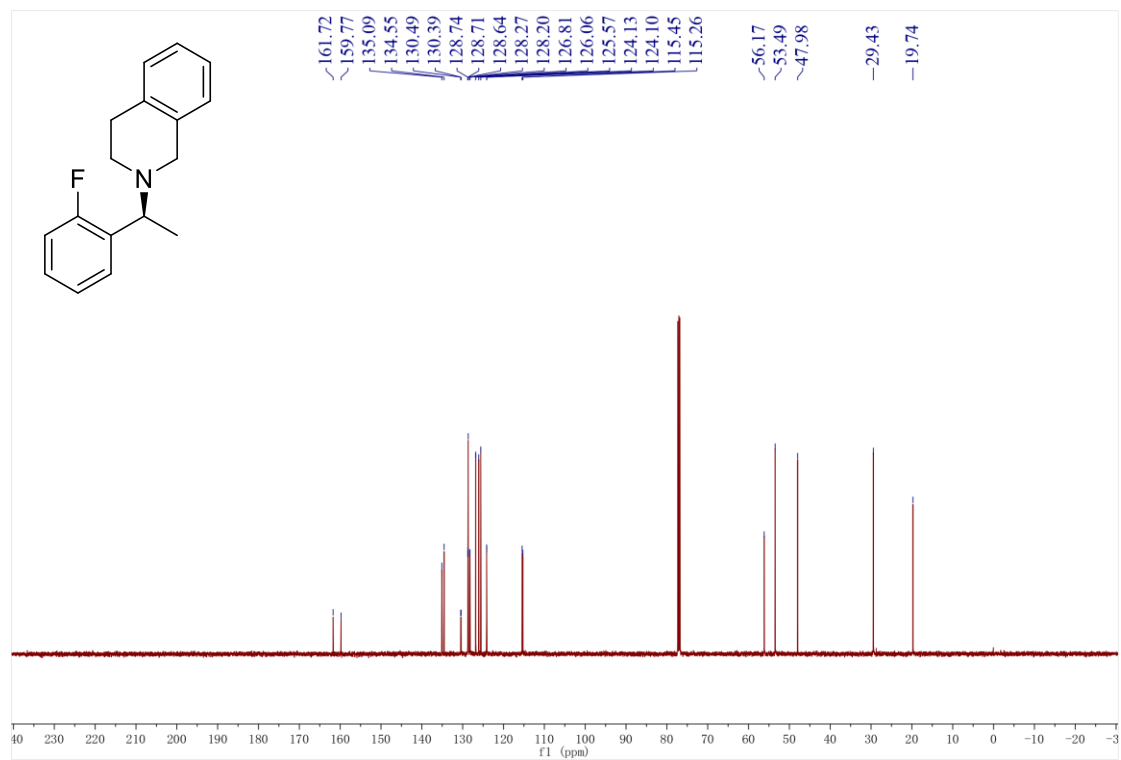
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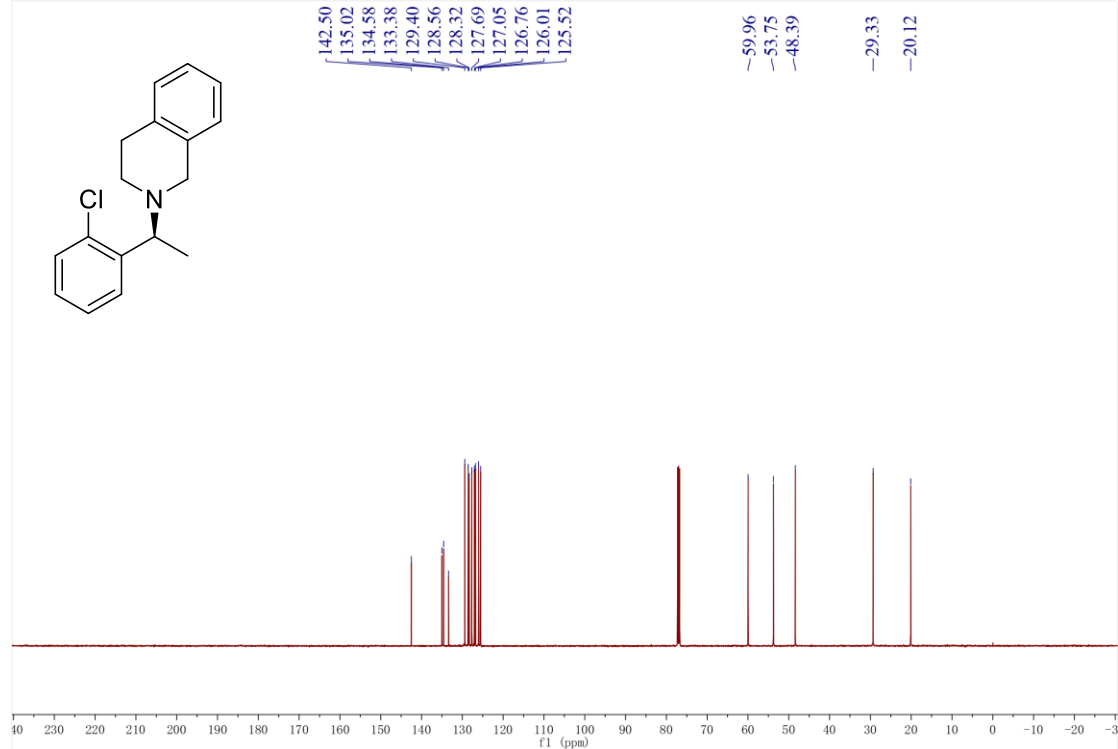
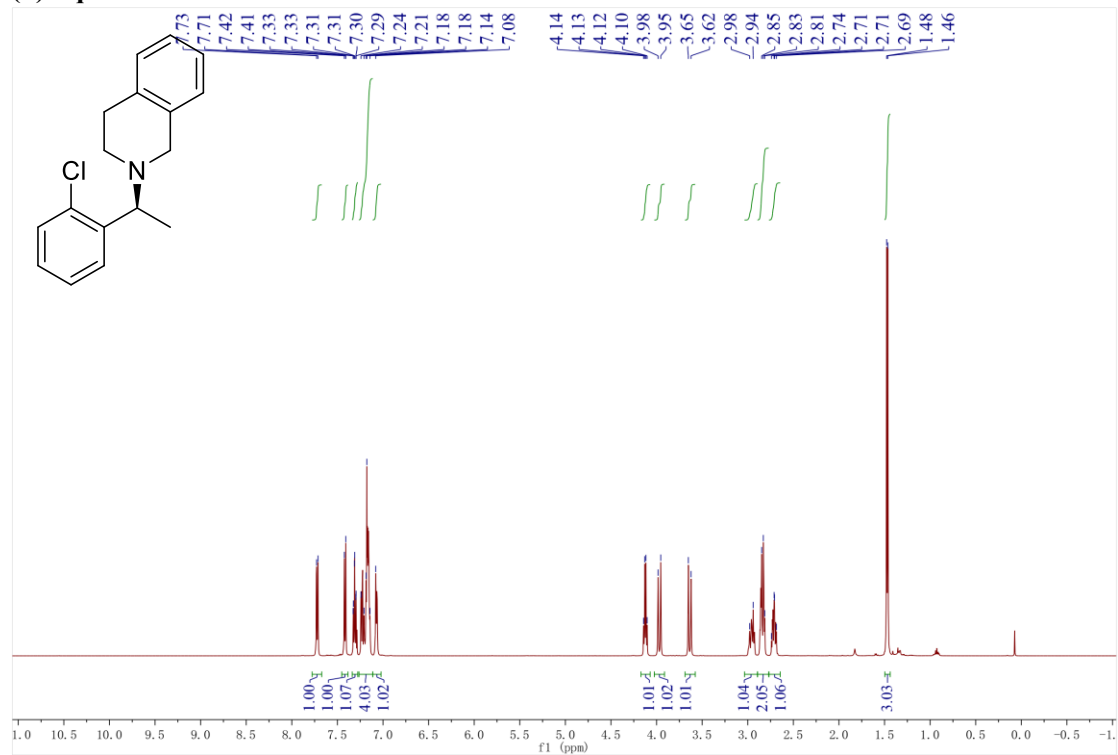


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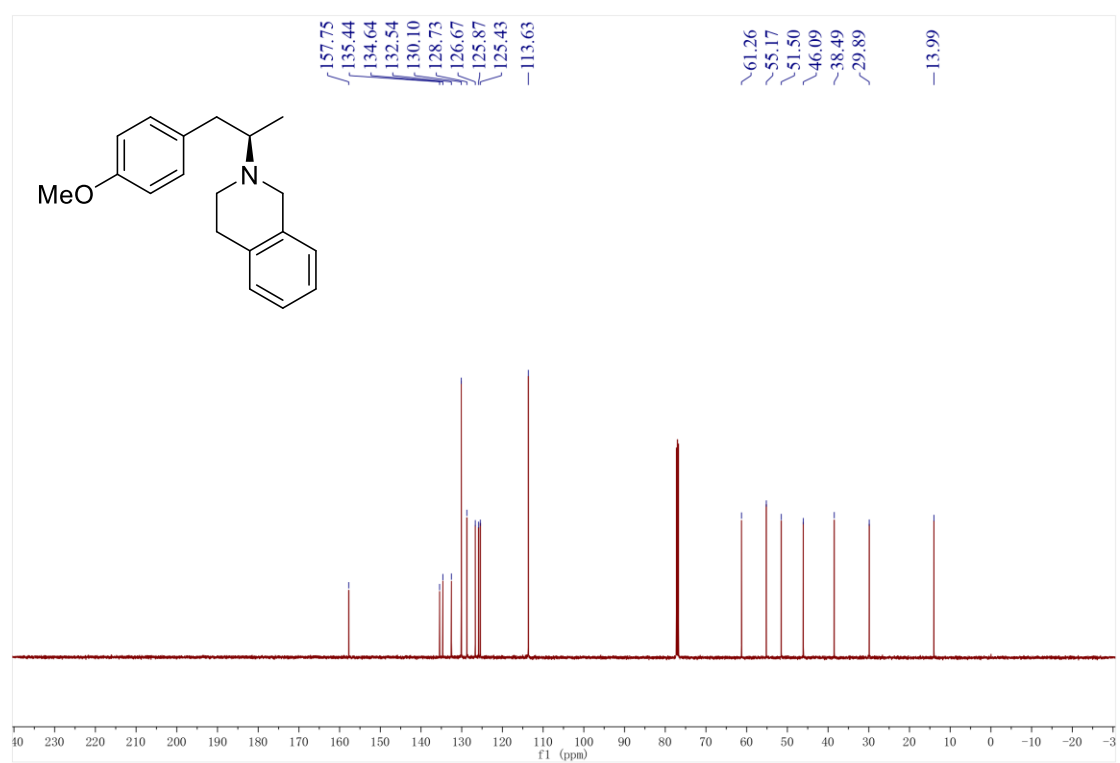
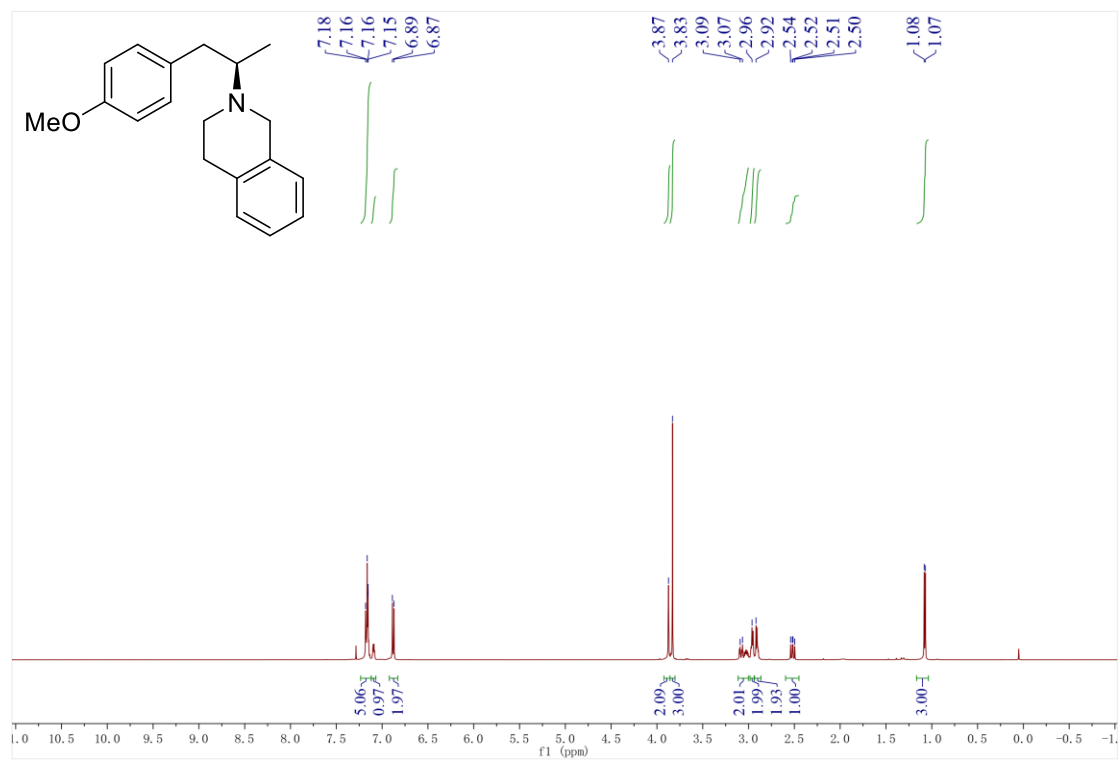




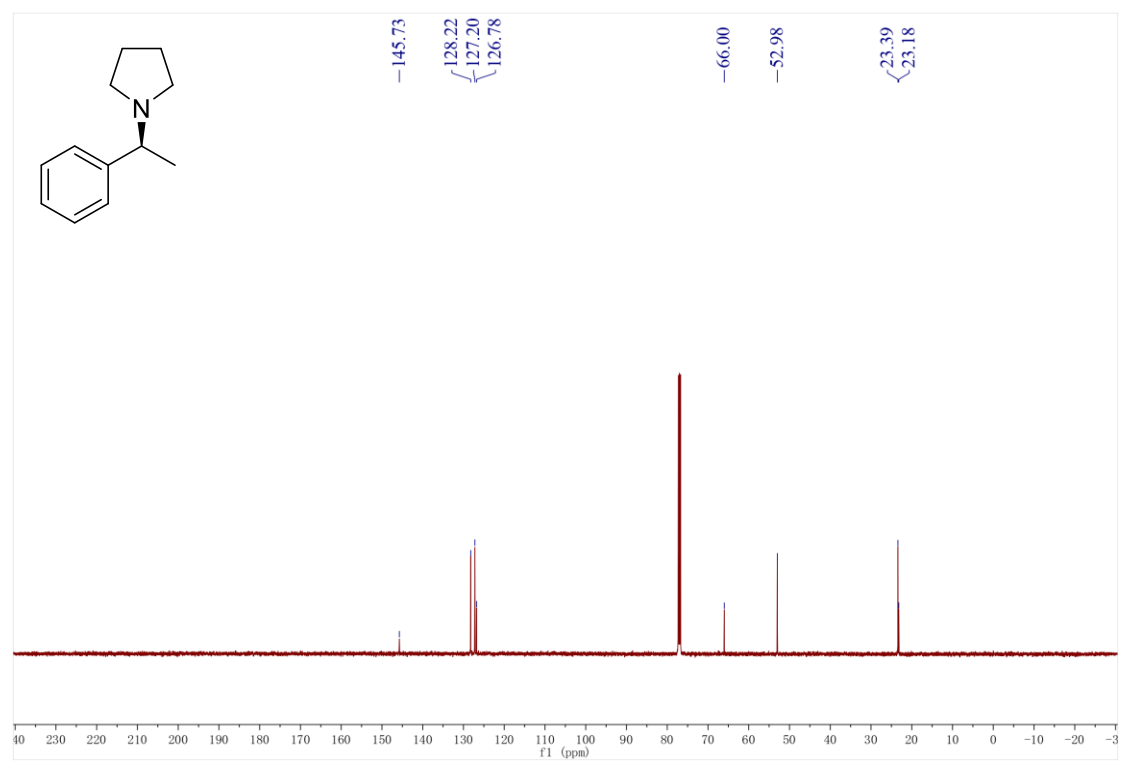
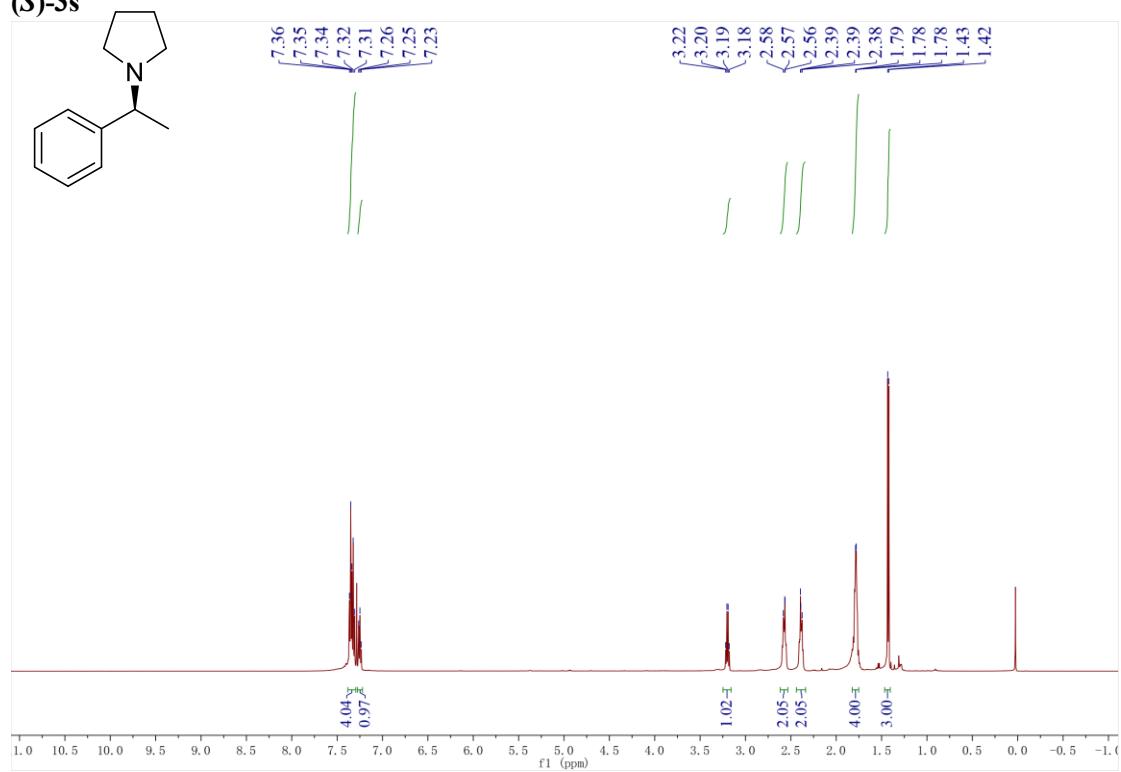
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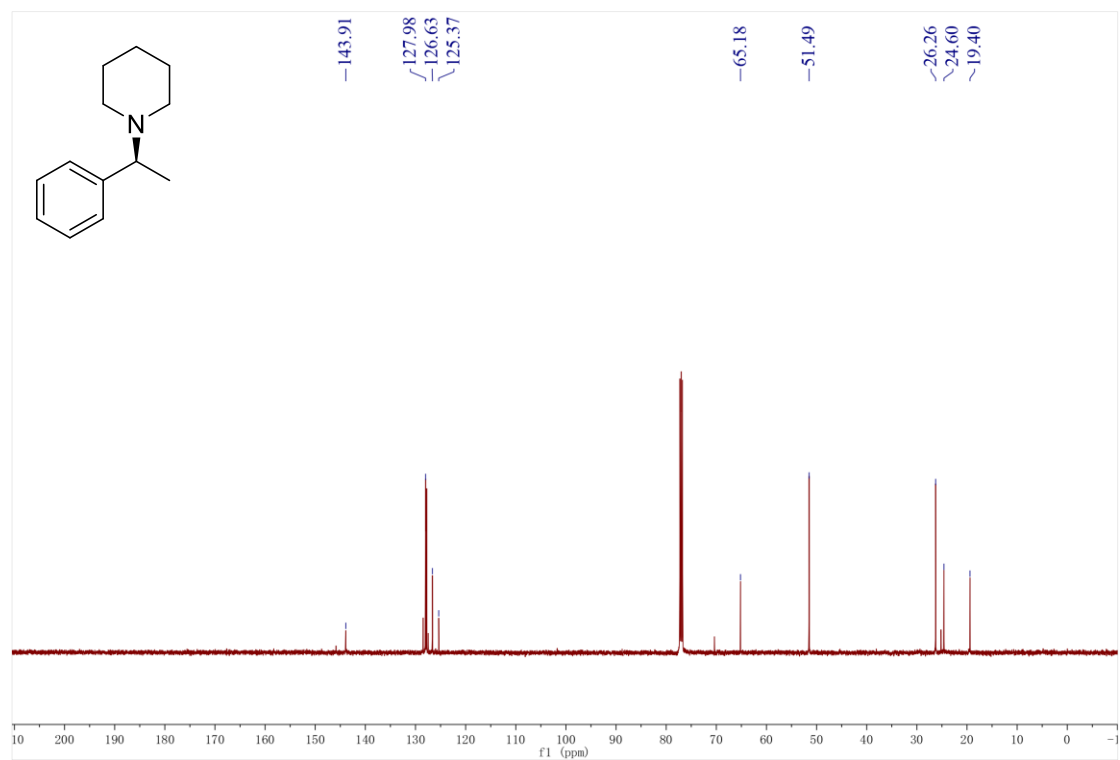
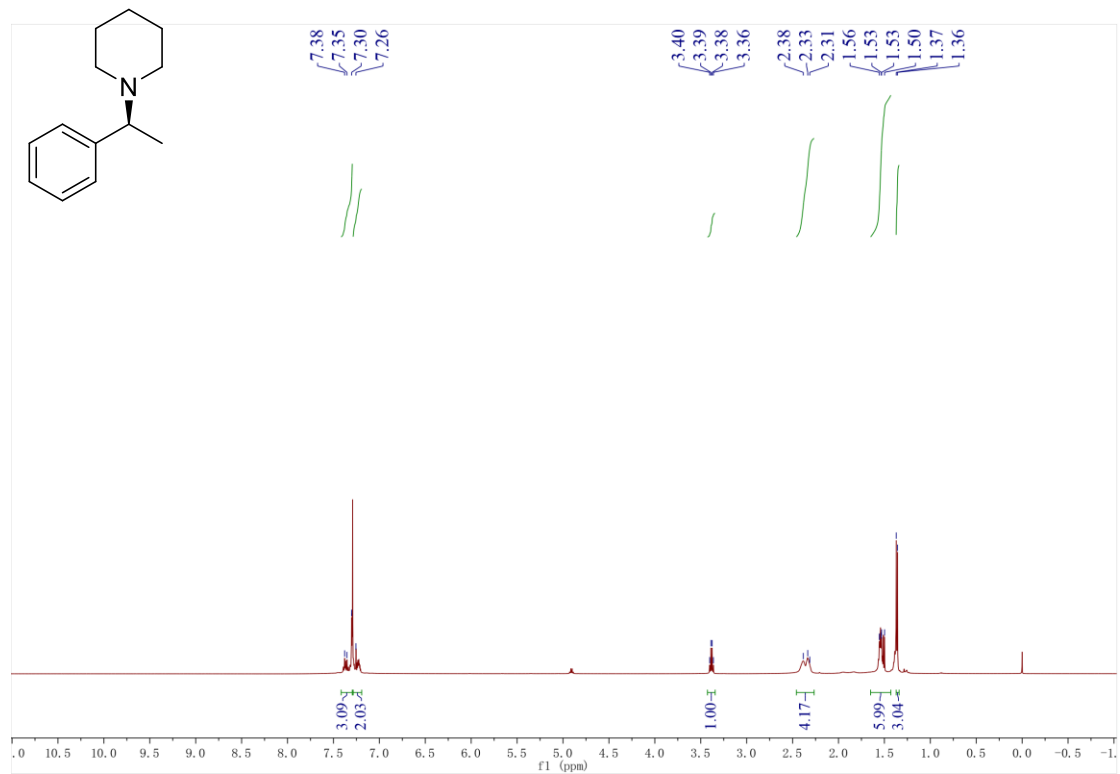
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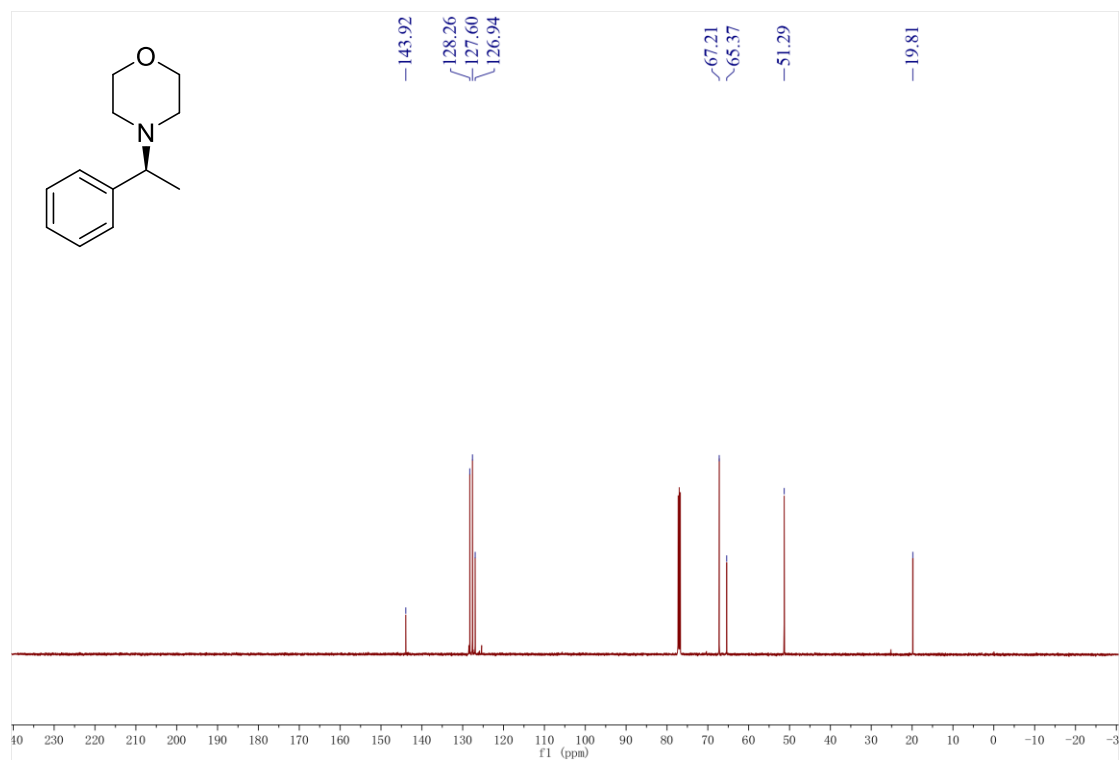
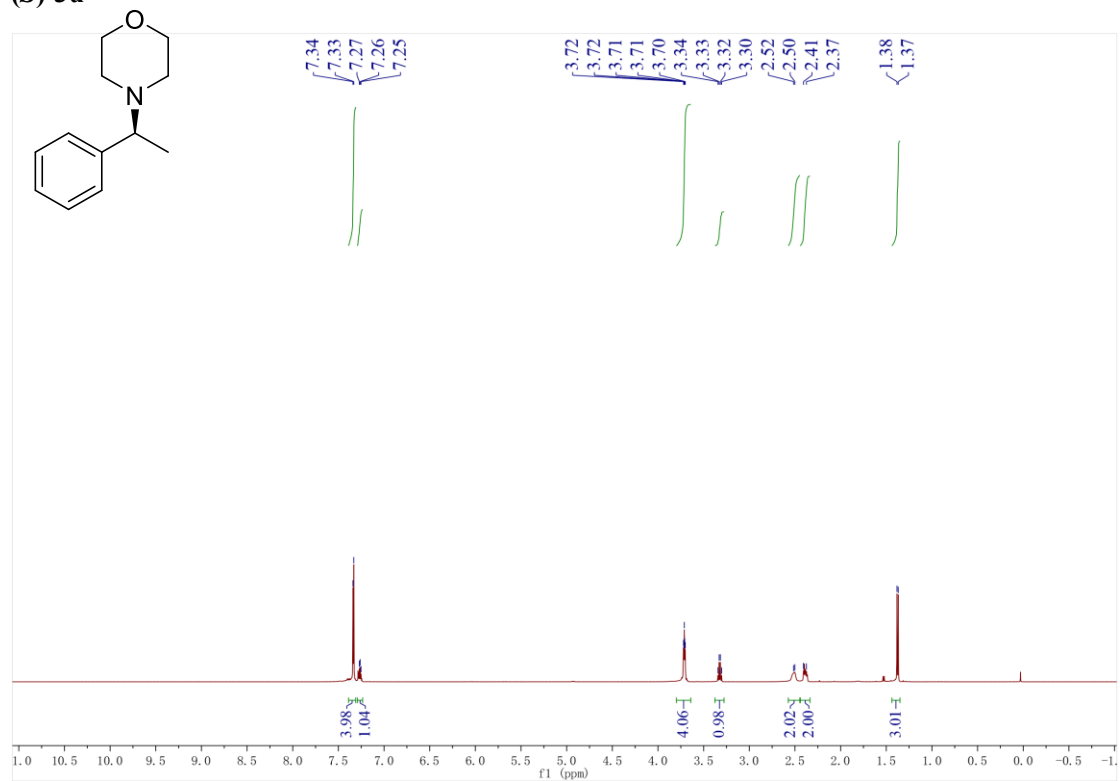
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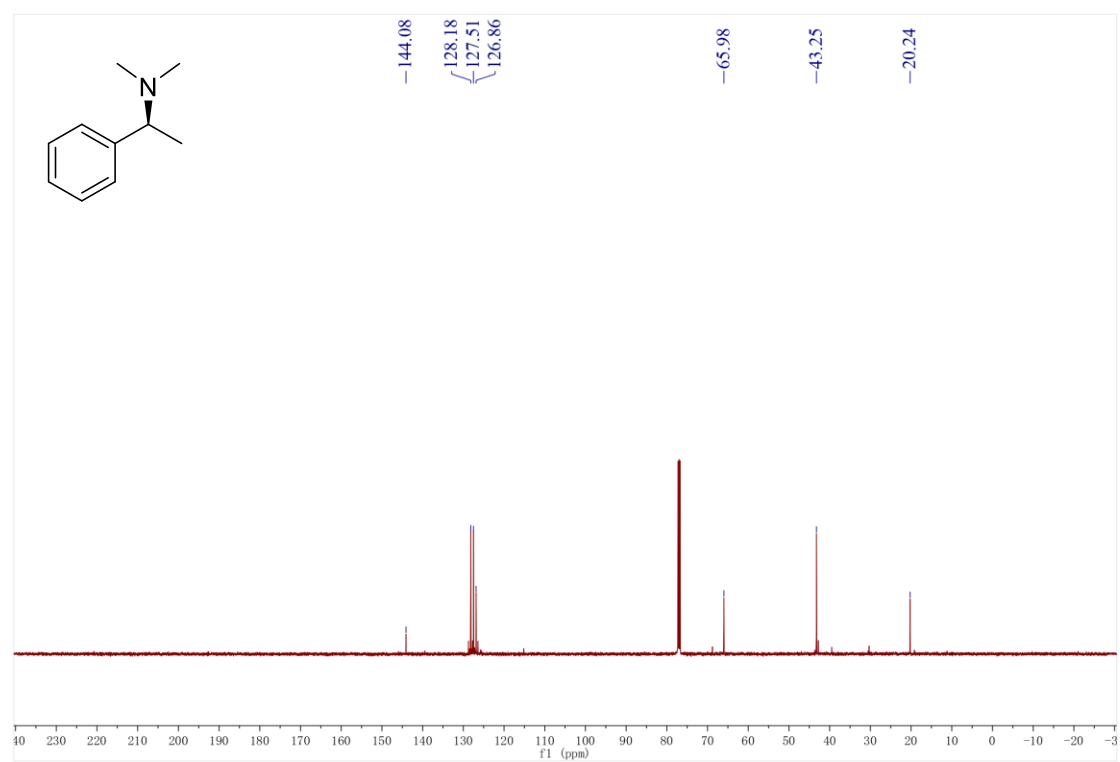
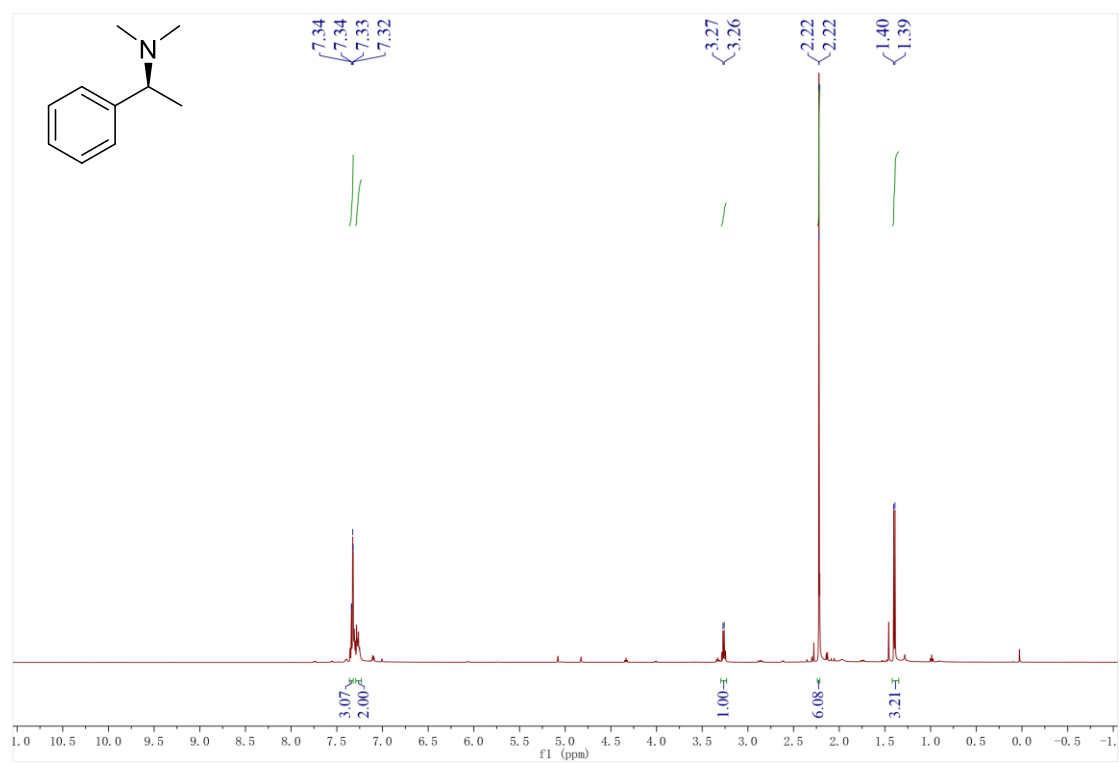
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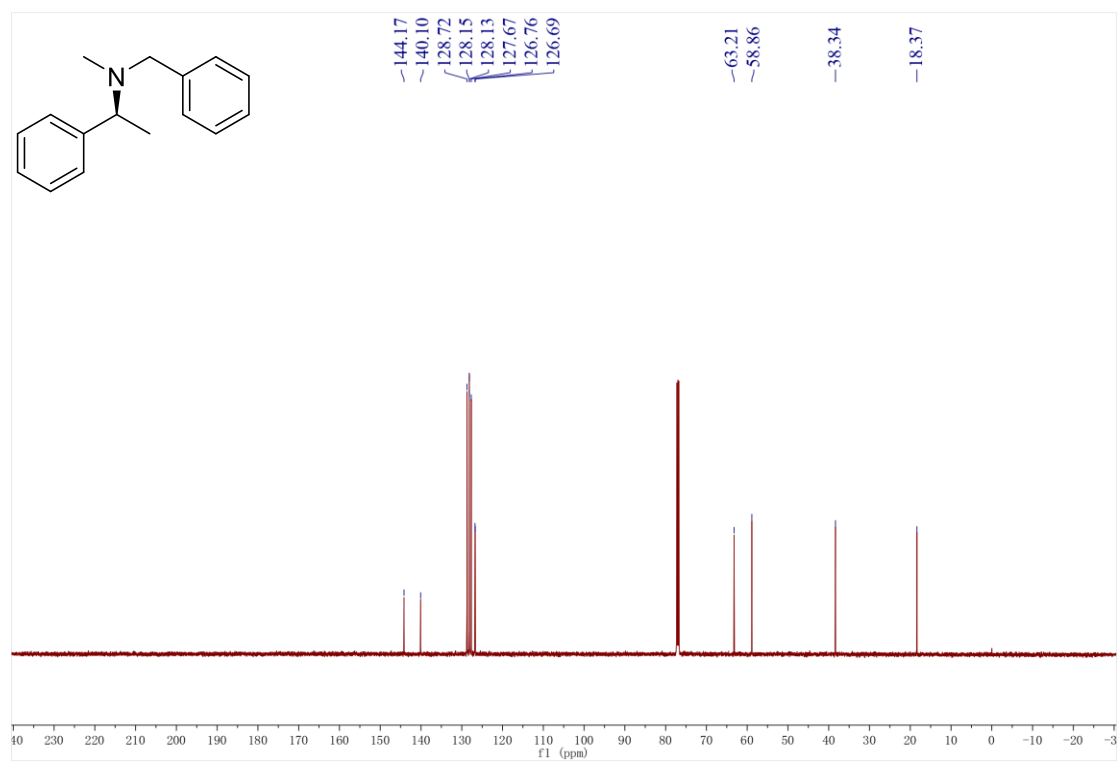
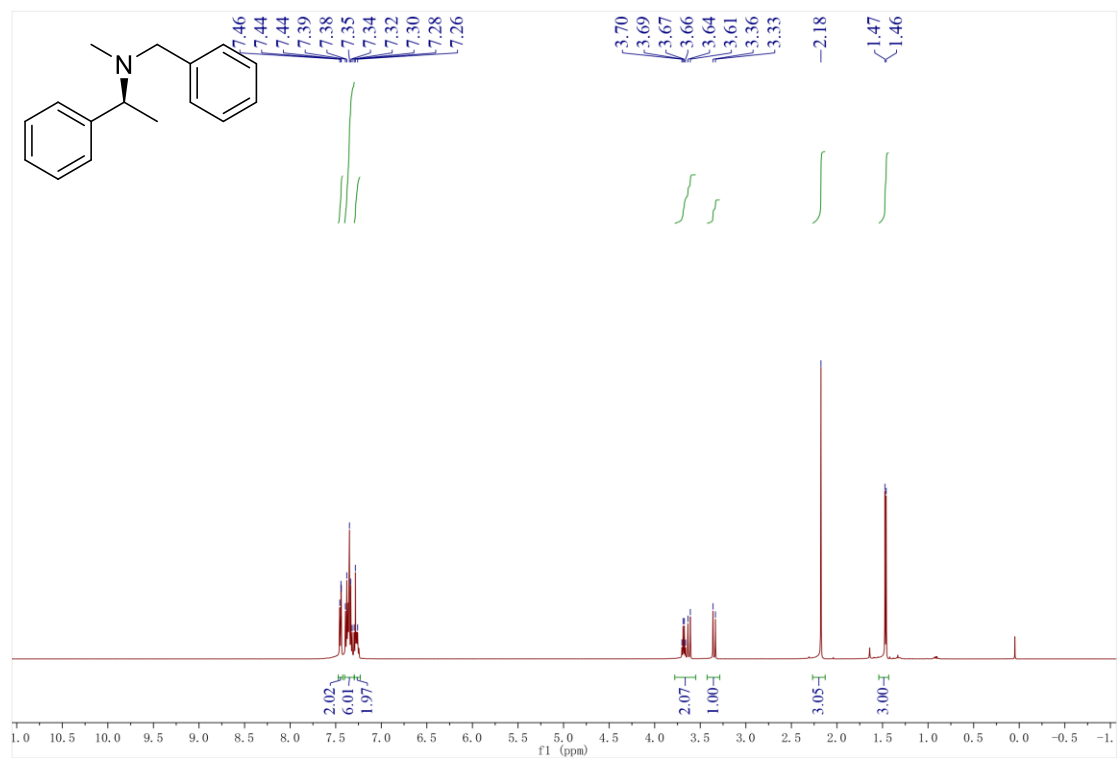
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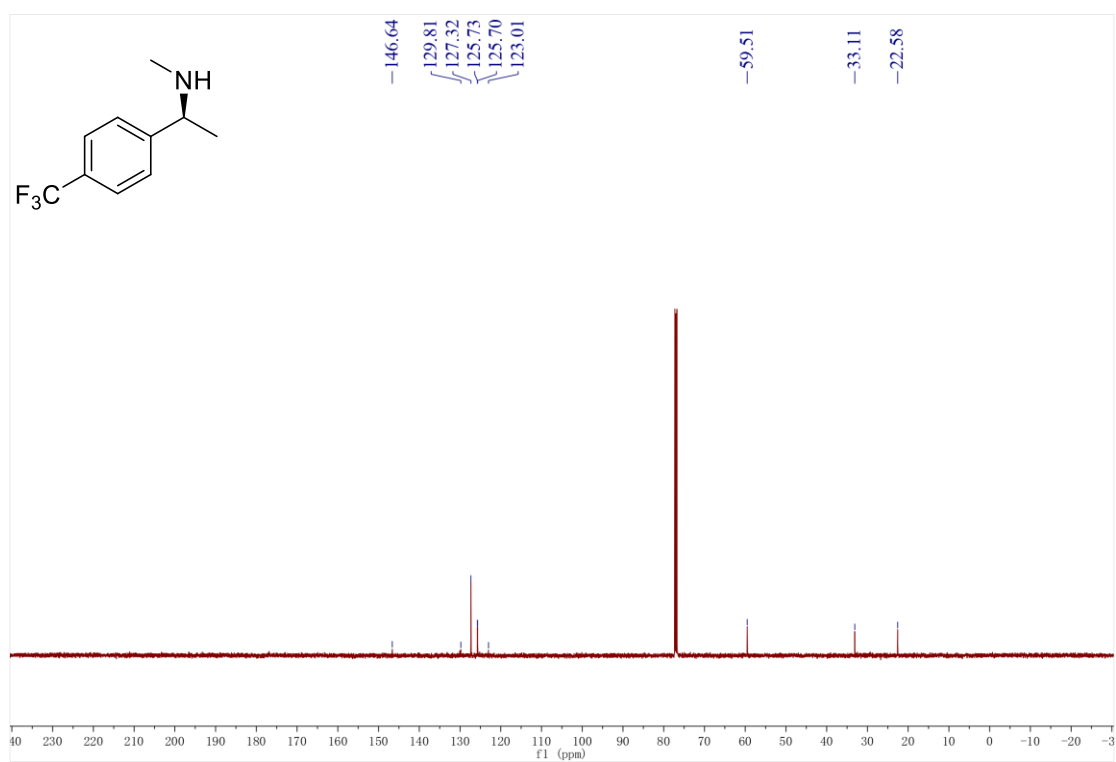
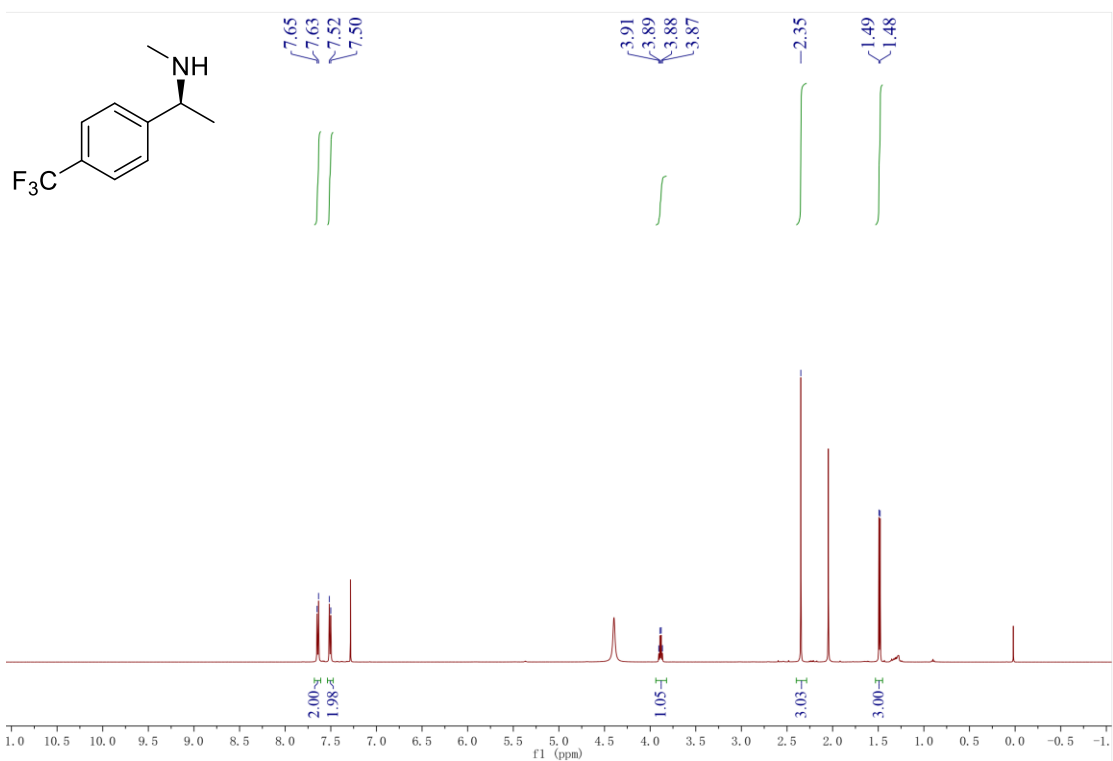
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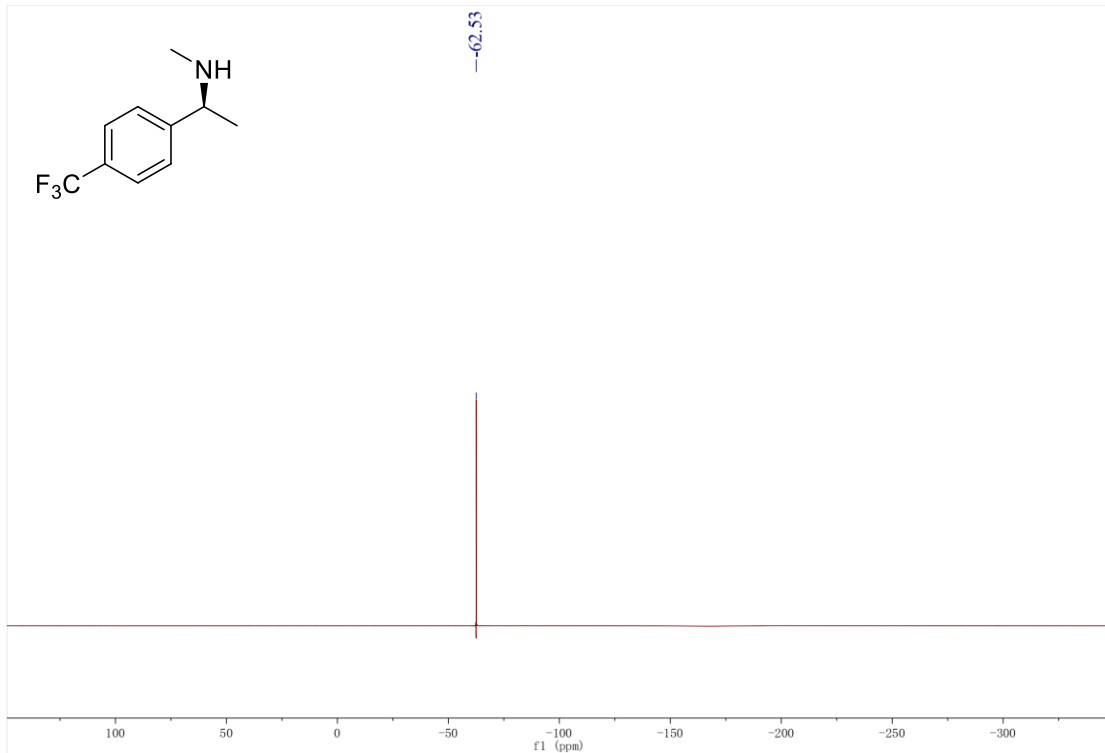


(S)-3w



(S)-4





(S)-5

