

Supporting Information

HSAB Theory Guiding Electrophilic Substitution Reactions of *o*-Carborane

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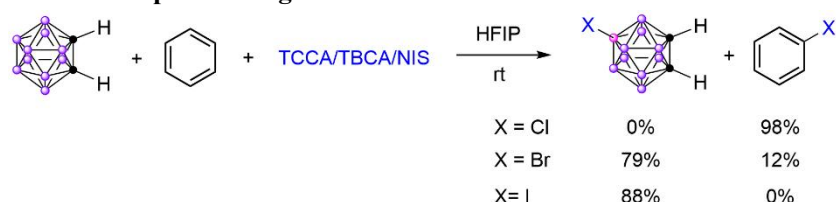
1. General Information

^1H , ^{13}C , ^{11}B and ^{19}F NMR spectra were recorded on Bruker Advance III600 (400) spectrometer at 600 (400), 151 (101), 193 (128) and 565 (377) MHz, respectively. All chemical shifts were reported in δ units with references to the residual solvent resonances of the deuterated solvents for proton and carbon chemical shifts, and to external $\text{BF}_3\cdot\text{OEt}_2$ (0.00 ppm) for boron chemical shifts. High Resolution Mass Spectra (HRMS (ESI-TOF)) were recorded on an Bruker Mass spectrometer using ESI-TOF (electrospray ionization-time of flight). GC-MS analyses were performed on SHIMADZU GCMS-QP 2020. Starting *o*-carborane and *m*-carborane were purchased from Zhengzhou Yuanli technology. All other chemicals were purchased from Aldrich, Acros Organics, J&K Chemicals, Energy Chemical, Aladdin, Macklin or TCI and used without further purification. TLC samples for carborane-containing compounds were stained with 1 wt.% PdCl_2 in 6 M HCl and were developed with high heat using a heat gun.

2. Experimental Section

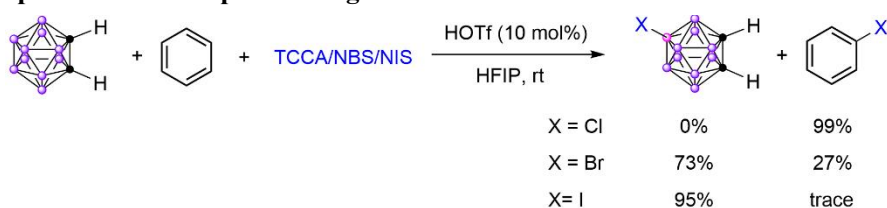
2.1 Comparison of electrophilic substitution reactions between *o*-carborane and benzene

(a) Catalyst-free electrophilic halogenation.



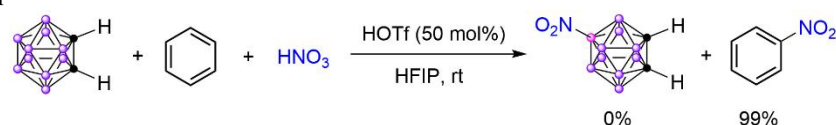
General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol) and electrophile TCCA (0.067 mmol), TBCA (0.067 mmol) or NIS (0.02 mmol) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.4 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

(b) HOTf-promoted electrophilic halogenation.



General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol) and electrophile TCCA (0.067 mmol), TBCA (0.067 mmol) or NIS (0.02 mmol) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOTf (10 mol%) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

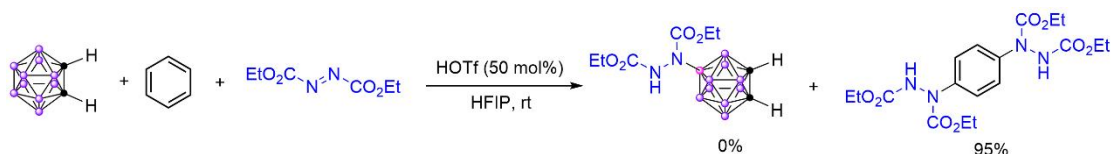
(c) Electrophilic nitration.



General procedure: Benzene (2.0 equiv, 0.40 mmol), 68% HNO_3 (1.0 equiv, 0.20 mmol) and

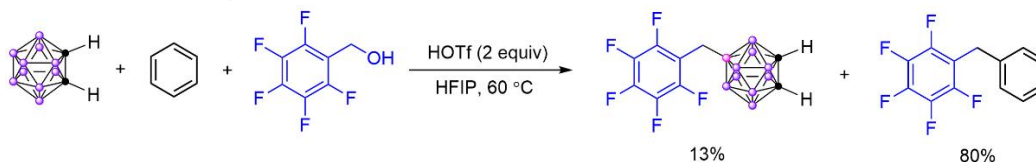
HOTf (50 mol%) were successively added to the solution of *o*-carborane (2.0 equiv, 0.40 mmol) in HFIP (4.0 mL). The resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

(d) Electrophilic amination.



General procedure: Benzene (2.0 equiv, 0.40 mmol), DEAD (1.0 equiv, 0.20 mmol), HOTf (50 mol%) were successively added to the solution of *o*-carborane (2.0 equiv, 0.40 mmol) in HFIP (4.0 mL). The resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

(e) Friedel-Crafts alkylation.



General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol) and 2,3,4,5,6-pentafluorobenzyl alcohol (1.0 equiv, 0.20 mmol) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOTf (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at 60 °C for 12 h under air atmosphere. Then, the reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

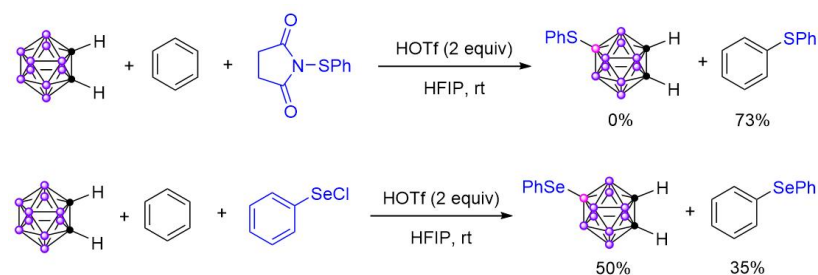
2.2 Electrophilic chalcogenation/Friedel-Crafts acylation/sulfonylation

(a) Electrophilic chalcogenation ^[1]



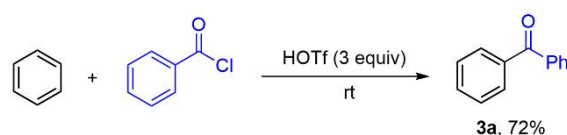
General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) and 1-(phenylthio)pyrrolidine-2,5-dione or phenylselenenyl chloride (2.0 equiv, 0.40 mmol) were mixed in HFIP (2.0 mL). Then to this solution was added HOTf (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction was completed, the solution was portioned between EtOAc and saturated aqueous NaHCO_3 . The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product.

(b) Competing experiments towards electrophilic chalcogenation



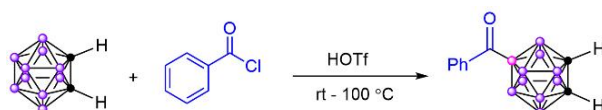
General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol) and electrophiles (1.0 equiv, 0.20 mmol) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOFt (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. The reaction solution was monitored by ^{11}B NMR and ^1H NMR spectroscopy to gain the product yield.

(c) Friedel-Crafts acylation of benzene. ^[2]



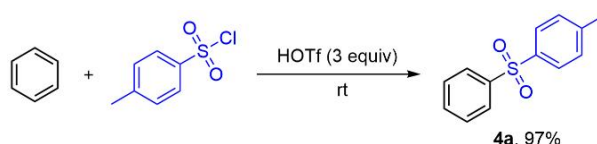
General procedure: Benzene (1.0 equiv, 0.20 mmol) and benzoyl chloride (2.0 equiv, 0.4 mmol) were mixed in a glass tube, then HOFt (3.0 equiv, 0.60 mmol) was carefully and slowly added, the reaction was stirred at ambient temperature for 12 h under air atmosphere. 1 mL of EtOAc was added to the reaction tube for dilution, then saturated aqueous NaHCO_3 was added to neutralize to $\text{pH} = 6 - 7$, and then extracted with EtOAc (20 mL \times 3). The organic phase was collected and dried with Na_2SO_4 . Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **3a**.

(d) Friedel-Crafts acylation of *o*-carborane.



General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) was mixed in HOFt (1.0 mL). Then to this solution was added benzoyl chloride (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO_3 . The organic portions were combined. No product was detected by ^{11}B NMR and GC-MS.

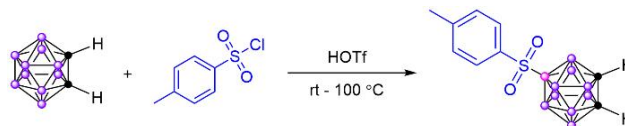
(e) Electrophilic sulfonylation of benzene. ^[3]



General procedure: Benzene (1.0 equiv, 0.20 mmol) and tosyl chloride (2.0 equiv, 0.4 mmol) were weighed into a glass reaction tube, then HOFt (3.0 equiv, 0.60 mmol) was carefully and slowly added, the reaction was stirred at ambient temperature for 12 h under air atmosphere. 1 mL of EtOAc was added to the reaction tube for dilution, then saturated aqueous NaHCO_3 was added to neutralize to $\text{pH} = 6 - 7$, and then extracted with EtOAc (20 mL \times 3). The organic phase was

collected and dried with Na₂SO₄. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **4a**.

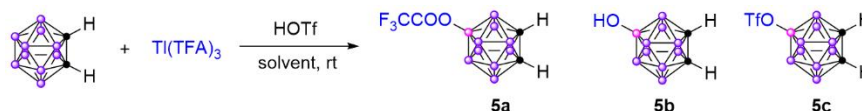
(f) Electrophilic sulfonylation of *o*-carborane.



General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) was mixed in HOTf (1.0 mL). Then to this solution was added tosyl chloride (2.0 equiv, 0.4 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. No product was detected by ¹¹B NMR and GC-MS.

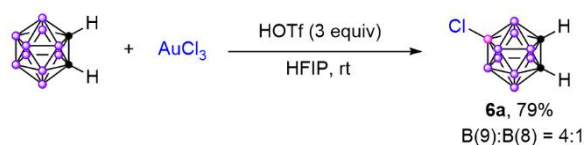
2.3 Electrophilic metallization

(a) Electrophilic thallation of *o*-carborane ^[4]



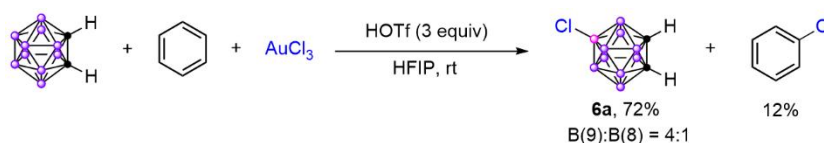
General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) and thallium(III) trifluoroacetate (2.0 equiv, 0.40 mmol) were mixed in HFIP or TFA (2.0 mL). Then to this solution was added HOTf (6.0 equiv, 1.20 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **5a**, **5b** and **5c**.

(b) Electrophilic auration of *o*-carborane ^[5]



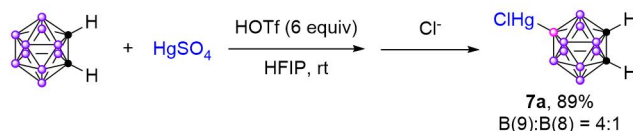
General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) and AuCl₃ (2.0 equiv, 0.40 mmol) were mixed in HFIP (2.0 mL). Then to this solution was added HOTf (3.0 equiv, 0.60 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **6a**.

(c) Competing experiment towards electrophilic auration.



General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol) and AuCl₃ (1.0 equiv, 0.20 mmol) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOTf (3.0 equiv, 0.60 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ¹¹B NMR and ¹H NMR spectroscopy to gain the product yield.

(d) Electrophilic mercuration of *o*-carborane.

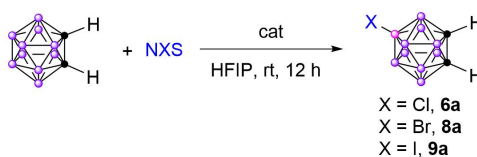


General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol) and HgSO₄ (2.0 equiv, 0.40 mmol) were mixed in HFIP (2.0 mL). Then to this solution was added HOTf (6.0 equiv, 1.20 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **7a**.

2.4 Pd(II)-catalyzed B(9)-functionalization of *o*-carborane

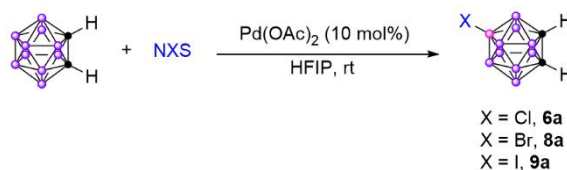
(a) Pd(II)-catalyzed B(9)-halogenation of *o*-carborane ^[5]

Table S1. B(9)-halogenation of *o*-carborane.^a



entry	NXS (1.0 equiv)	Cat (10 mol%)	Yield (%)
1	NCS	w/o	n.r.
2	NCS	Pd(OAc) ₂	36 (trace) ^b
3	NBS	w/o	26
4	NBS	Pd(OAc) ₂	69 (6) ^b
5	NIS	w/o	74
6	NIS	Pd(OAc) ₂	71 (13) ^b

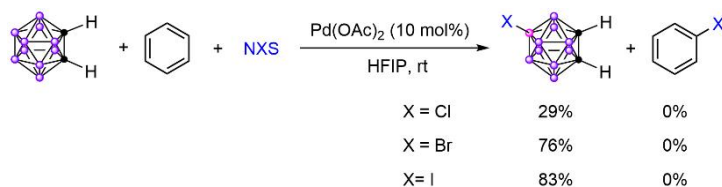
[a] reaction conditions: *o*-carborane (0.2 mmol), NXS (0.2 mmol) in 2 mL HFIP was stirred at room temperature for 12 hours. [b] yields of the dihalogenated products.



General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol), NXS (2.0 equiv, 0.40 mmol) and Pd(OAc)₂ (10 mol%) were mixed in HFIP (2.0 mL). Then the resulting mixture was stirred in a

closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **6a/8a/9a**.

(b) Comparison of Pd(II)-catalyzed B-H/C-H halogenation of *o*-carborane and benzene



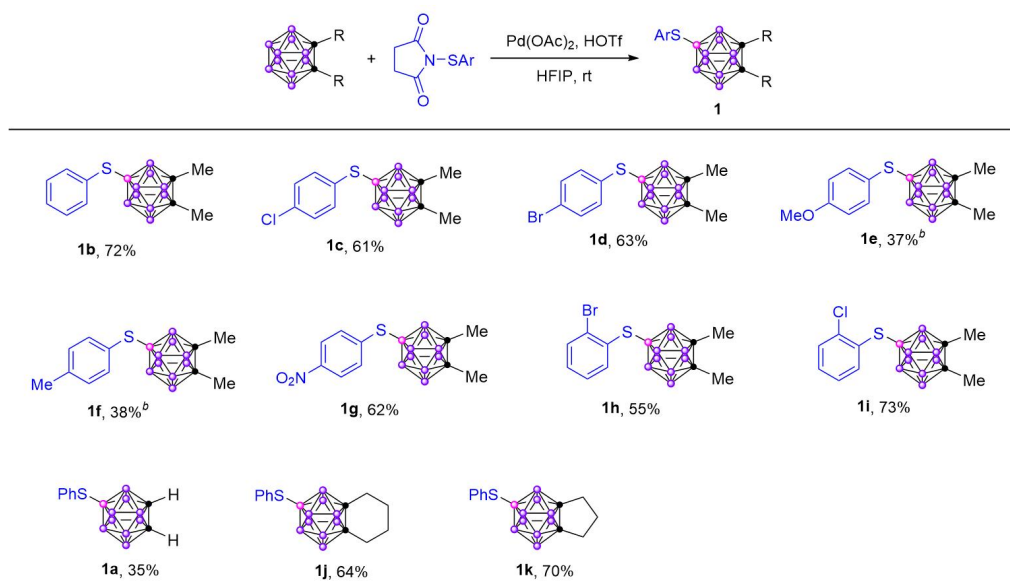
General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol), NXS (1.0 equiv, 0.20 mmol) and Pd(OAc)₂ (10 mol%) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ¹¹B NMR and ¹H NMR spectroscopy to gain the product yield.

(c) Pd(II)-catalyzed B(9)-thiolation of *o*-carborane



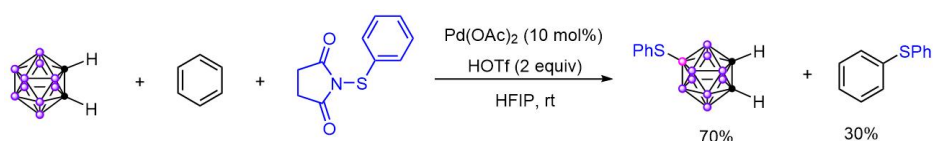
General procedure: *o*-Carborane (1.0 equiv, 0.20 mmol), the sulfur reagent (1.5 equiv, 0.30 mmol) and Pd(OAc)₂ (10 mol%) were mixed in HFIP (2.0 mL). Then to this solution was added HOTf (1.0 equiv, 0.20 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature or 60°C for 12 h – 24 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product **1a-1k**.

Table S2. Scope of Pd(II)-catalyzed B(9)-thiolation of *o*-carborane.^a



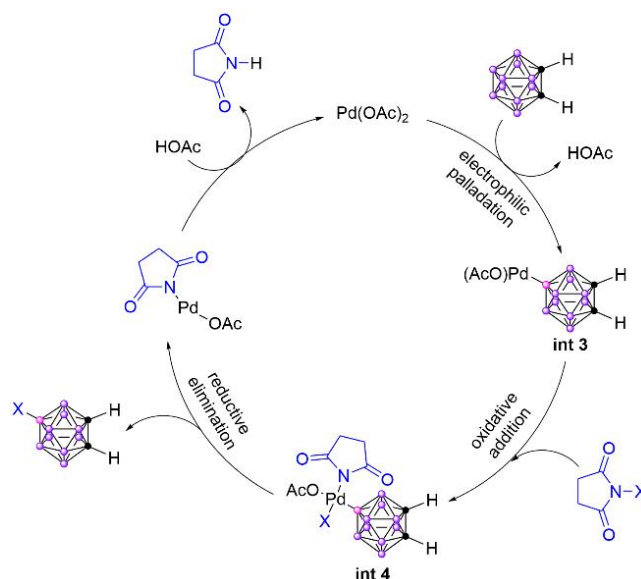
[a] Reaction conditions: carboranes (0.2 mmol, 1.0 equiv), arylthiolation reagent (0.3 mmol, 1.5 equiv), Pd(OAc)₂ (10 mol%) and HOTf (0.2 mmol, 1.0 equiv) in HFIP (2 mL) at room temperature under an air atmosphere. [b] At 60 °C for 24 hours.

(d) Comparison of Pd(II)-catalyzed B-H/C-H thiolation of *o*-carborane and benzene



General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol), 1-(phenylthio)pyrrolidine-2,5-dione (1.0 equiv, 0.20 mmol) and Pd(OAc)₂ (10 mol%) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOTf (2.0 equiv, 0.40 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ¹¹B NMR and ¹H NMR spectroscopy to gain the product yield.

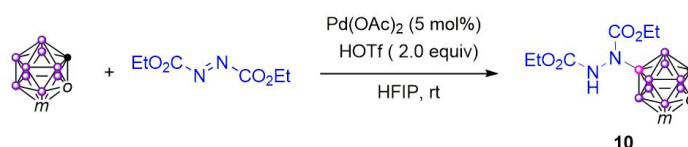
(e) Proposed mechanism for Pd(II)-catalyzed B(9)-H halogenation/thiolation of *o*-carborane



Scheme S1. The plausible mechanism of Pd(II)-catalyzed B(9)-halogenation of *o*-carborane

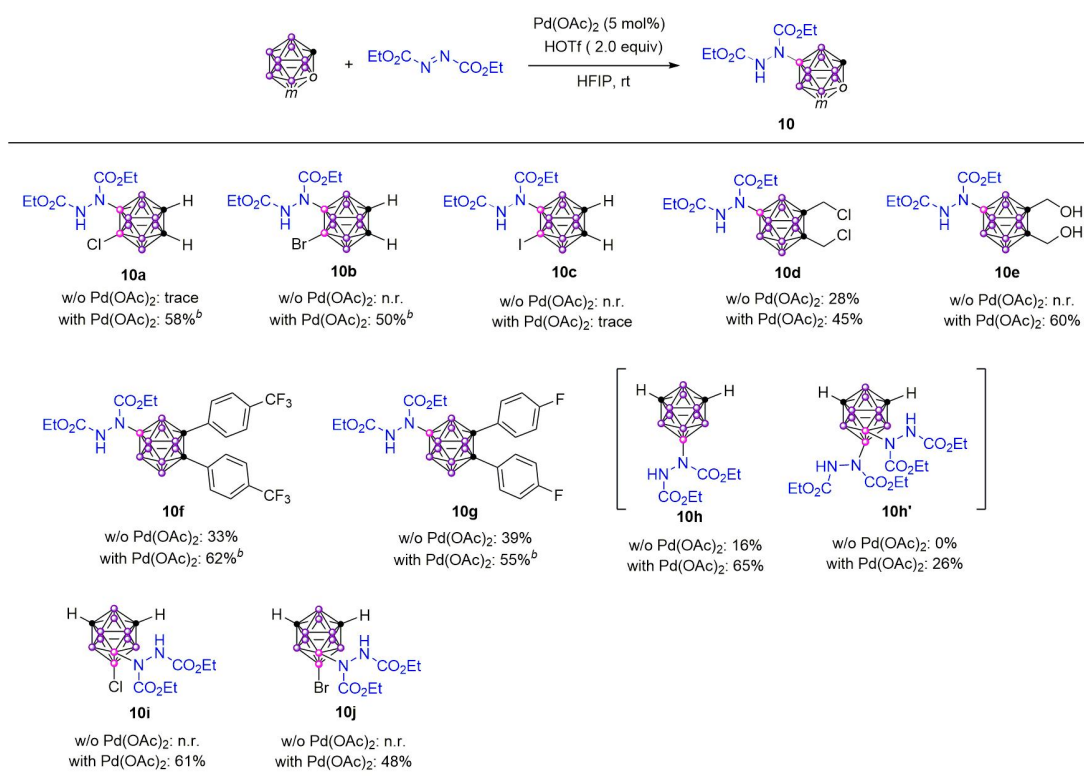
Based on the experiment results and our previous mechanism studies, a plausible mechanism of Pd(II)-catalyzed B(9)-halogenation and thiolation of *o*-carborane is proposed, as shown in **Scheme S1**. We consider that the first step in the catalytic cycle is the formation of B–Pd intermediate **int 3** through electrophilic palladation process. Then the halogenating reagent oxidative addition occurs to form the Pd(IV) intermediate **int 4**. B–X reductive elimination from **int 4** produces the target product, meanwhile the catalyst palladium is reduced from the Pd(IV) to the Pd(II), entering the catalytic cycle.

(f) Pd(II)-catalyzed B(9)-amination of *o*-carborane [6]



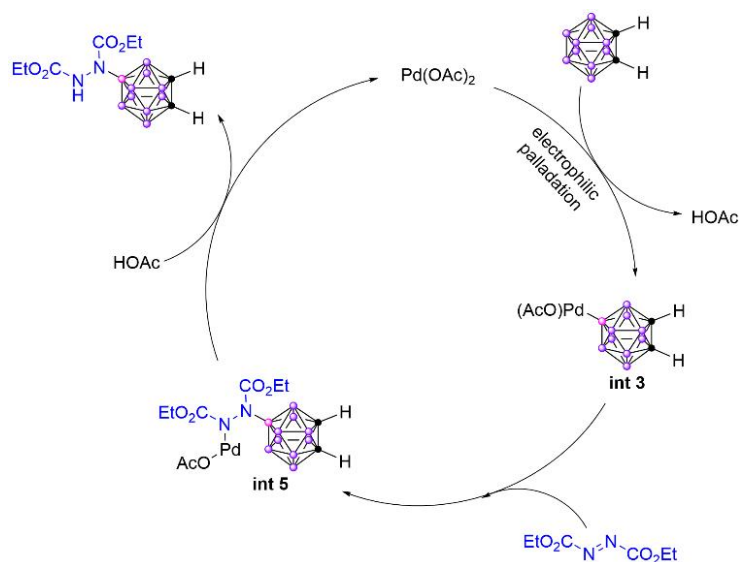
General procedure: *o/m*-Carborane (1.0 equiv, 0.20 mmol), DEAD (1.1 equiv, 0.22 mmol) and Pd(OAc)₂ (5 mol%) were mixed in HFIP (2.0 mL). Then to this solution was added HOTf (1.0 equiv, 0.20 mmol) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. After the reaction solution was portioned between EtOAc and saturated aqueous NaHCO₃. The organic portions were combined. Then removal of organic solvents under reduced pressure, the residue was subjected to flash column chromatography on silica gel (200-300 mesh) to give the product (**10a-10j**).

Table S3. Selective B(9)-amination of *o*-carborane.^a



[a] Reaction conditions: carboranes (0.2 mmol, 1.0 equiv), DEAD (0.22 mmol, 1.1 equiv), Pd(OAc)₂ (5 mol%) and HOTf (0.2 mmol, 1.0 equiv) in HFIP (2 mL) at room temperature under an air atmosphere. [b] At 60 °C for 12 hours.

(g) Proposed mechanism for Pd(II)-catalyzed B(9)-amination of *o*-carborane

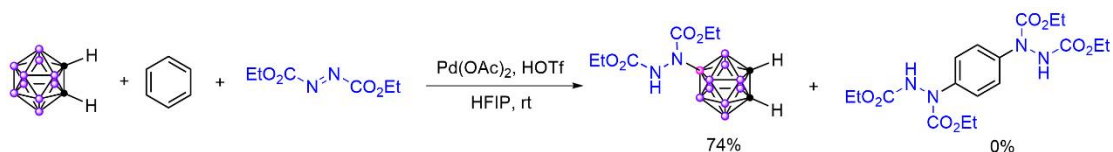


Scheme S2. The plausible mechanism of Pd(II)-catalyzed B(9)-amination of *o*-carborane

Based on the experiment results and our previous mechanism studies, a plausible mechanism of Pd(II)-catalyzed B(9)-amination of *o*-carborane is proposed, as shown in **Scheme S2**. We consider that the first step in the catalytic cycle is the formation of B–Pd intermediate **int 3** through electrophilic palladation process. Then the aminating reagent (DEAD) electrophilic addition occurs to form the B–N intermediate **int 5**, following protonation to form the target product and

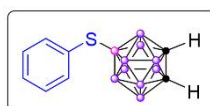
the catalyst palladium enters the catalytic cycle.

(h) Comparison of Pd(II)-catalyzed B-H/C-H amination of *o*-carborane and benzene



General procedure: *o*-Carborane (2.0 equiv, 0.40 mmol), DEAD (1.0 equiv, 0.20 mmol) and Pd(OAc)₂ (10 mol%) were mixed in HFIP (4.0 mL). Then to this solution was added benzene (2.0 equiv, 0.40 mmol), HOTf (50 mol%) and the resulting mixture was stirred in a closed flask at ambient temperature for 12 h under air atmosphere. Then, the reaction solution was monitored by ¹¹B NMR and ¹H NMR spectroscopy to gain the product yield. Then, the reaction solution was monitored by ¹¹B NMR and ¹H NMR spectroscopy to gain the product yield.

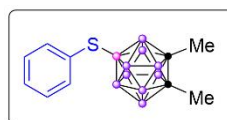
3. Characterization data



1a: Yield 35%; White solid; **Eluent:** PE/EA = 50:1 (*R_f* = 0.28), Mp:124-126 °C.

¹H NMR (600 MHz, CDCl₃) δ: 7.43 (d, *J* = 5.2 Hz, 2H), 7.24 (s, 1H), 7.23 (d, *J* = 1.4 Hz, 2H), 3.50 (s, 1H), 3.41 (s, 1H). **¹³C{¹H} NMR (151 MHz, CDCl₃)** δ: 135.4, 134.9, 128.5, 126.9, 52.7, 47.5. **¹¹B{¹H} NMR (193 MHz, CDCl₃)** δ: 7.5 (1B), -2.3 (1B), -8.6 (2B), -13.9 (2B), -14.6 (2B), -15.5 (2B). **IR:** 621, 718, 1069, 2603, 3258 cm⁻¹.

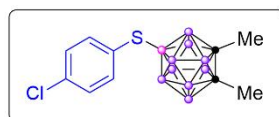
HRMS (ESI-TOF): *m/z* calcd for C₈B₁₀H₁₇S [M+H]⁺: 253.2053. Found: 253.2051.



1b: Yield 72%; White solid; **Eluent:** PE/EA = 25:1 (*R_f* = 0.32).

¹H NMR (600 MHz, CDCl₃) δ: 7.45 (d, *J* = 5.5 Hz, 2H), 7.25 (s, 1H), 7.23 (d, *J* = 0.8 Hz, 2H), 1.99 (s, 3H), 1.98 (s, 3H). **¹³C{¹H} NMR (151 MHz, CDCl₃)** δ: 135.8, 134.8, 128.4, 126.7, 71.9, 66.5, 23.2, 21.9. **¹¹B{¹H} NMR (193 MHz, CDCl₃)** δ: 5.3 (1B), -4.8 (1B), -8.9 (2B), -9.8 (4B), -10.3 (2B).

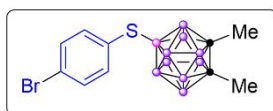
HRMS (ESI-TOF): *m/z* calcd for C₁₀B₁₀H₂₁S [M+H]⁺: 281.2367. Found: 281.2366.



1c: Yield 61%; White solid; **Eluent:** PE/EA = 25:1 (*R_f* = 0.28).

¹H NMR (600 MHz, CDCl₃) δ: 7.35 (d, *J* = 8.3 Hz, 2H), 7.19 (d, *J* = 8.5 Hz, 2H), 1.99 (s, 3H), 1.98 (s, 3H). **¹³C{¹H} NMR (151 MHz, CDCl₃)** δ: 136.1, 134.4, 132.9, 128.6, 71.9, 66.7, 23.2, 22.0. **¹¹B{¹H} NMR (193 MHz, CDCl₃)** δ: 5.1 (1B), -4.8 (1B), -8.9 (2B), -9.8 (4B), -10.3 (2B).

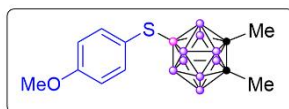
HRMS (ESI-TOF): *m/z* calcd for C₁₀B₁₀H₂₀SCl [M+H]⁺: 316.1951. Found: 316.1953.



1d: Yield 63%; White solid; **Eluent:** PE/EA = 100:1 (R_f = 0.28).

^1H NMR (600 MHz, CDCl_3) δ : 7.34 (d, J = 8.5 Hz, 2H), 7.28 (d, J = 8.3 Hz, 2H), 1.99 (s, 3H), 1.98 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3)** δ : 136.5, 135.0, 131.5, 121.1, 71.9, 66.7, 23.3, 22.0. **$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)** δ : 5.0 (1B), -4.8 (1B), -8.9 (2B), -9.8 (4B), -10.2 (2B).

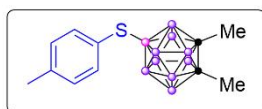
HRMS (ESI-TOF): m/z calcd for $\text{C}_{10}\text{B}_{10}\text{H}_{20}\text{SBr}$ $[\text{M}+\text{H}]^+$: 360.1458. Found: 360.1456.



1e: Yield 37%; White solid; **Eluent:** PE/EA = 25:1 (R_f = 0.26).

^1H NMR (600 MHz, CDCl_3) δ : 7.33 (d, J = 8.4 Hz, 2H), 6.77 (d, J = 8.6 Hz, 2H), 3.78 (s, 3H), 1.98 (s, 3H), 1.98 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3)** δ : 158.8, 136.2, 126.7, 114.0, 71.8, 66.2, 55.2, 23.3, 22.0. **$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)** δ : 5.7 (1B), -4.7 (1B), -8.9 (2B), -9.8 (4B), -10.3 (2B).

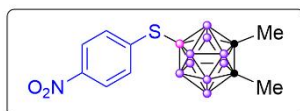
HRMS (ESI-TOF): m/z calcd for $\text{C}_{11}\text{B}_{10}\text{H}_{23}\text{SO}$ $[\text{M}+\text{H}]^+$: 311.2473. Found: 311.2468.



1f: Yield 38%; White solid; **Eluent:** PE (R_f = 0.26).

^1H NMR (600 MHz, CDCl_3) δ : 7.31 (d, J = 7.9 Hz, 2H), 7.04 (d, J = 7.9 Hz, 2H), 2.31 (s, 3H), 1.98 (s, 3H), 1.98 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3)** δ : 136.5, 134.8, 132.2, 129.2, 71.8, 66.3, 23.2, 22.0, 21.1. **$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)** δ : 5.5 (1B), -4.7 (1B), -8.9 (2B), -9.7 (4B), -10.3 (2B).

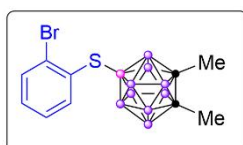
HRMS (ESI-TOF): m/z calcd for $\text{C}_{11}\text{B}_{10}\text{H}_{22}\text{SNH}_4$ $[\text{M}+\text{NH}_4]^+$: 312.2790. Found: 312.2785.



1g: Yield 62%; White solid; **Eluent:** PE/EA = 25:1 (R_f = 0.32).

^1H NMR (600 MHz, CDCl_3) δ : 8.06 (d, J = 8.8 Hz, 2H), 7.56 (d, J = 8.6 Hz, 2H), 2.02 (s, 3H), 2.01 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3)** δ : 146.4, 145.8, 134.3, 123.3, 72.2, 67.7, 23.3, 22.0. **$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)** δ : 4.2 (1B), -4.8 (1B), -8.8 (2B), -9.8 (6B).

HRMS (ESI-TOF): m/z calcd for $\text{C}_{10}\text{B}_{10}\text{H}_{20}\text{SNO}_2$ $[\text{M}+\text{H}]^+$: 326.2218. Found: 326.2216.

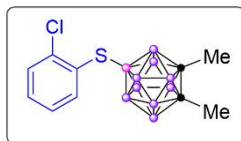


1h: Yield 55%; White solid; **Eluent:** PE/EA = 25:1 (R_f = 0.26).

^1H NMR (600 MHz, CDCl_3) δ : 7.57 (d, J = 8.0 Hz, 2H), 7.18 (t, J = 7.5 Hz, 1H), 7.06 (t, J = 7.6 Hz, 1H), 1.99 (s, 3H), 1.98 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3)** δ : 137.6, 137.2, 132.9, 130.3, 128.4, 127.2, 71.9, 66.9, 23.3, 22.0. **$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)** δ : 4.9 (1B), -4.7

(1B), -9.0 (2B), -9.8 (4B), -10.3 (2B).

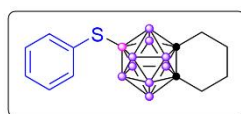
HRMS (ESI-TOF): m/z calcd for $C_{10}B_{10}H_{20}SBr$ $[M+H]^+$: 360.1458. Found: 360.1457.



1i: Yield 73%; White solid; **Eluent:** PE/EA = 25:1 (R_f = 0.28).

1H NMR (600 MHz, $CDCl_3$) δ : 7.55 (d, J = 7.4 Hz, 1H), 7.38 (d, J = 7.8 Hz, 1H), 7.18 – 7.11 (m, 2H), 1.99 (s, 3H), 1.98 (s, 3H). **$^{13}C\{^1H\}$ NMR (151 MHz, $CDCl_3$)** δ : 139.0, 137.8, 135.0, 129.5, 128.4, 126.6, 71.9, 66.8, 23.3, 22.0. **$^{11}B\{^1H\}$ NMR (193 MHz, $CDCl_3$)** δ : 4.8 (1B), -4.8 (1B), -9.0 (2B), -9.8 (4B), -10.3 (2B).

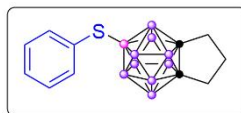
HRMS (ESI-TOF): m/z calcd for $C_{10}B_{10}H_{20}SCl$ $[M+H]^+$: 316.1951. Found: 316.1950.



1j: Yield 64%; White solid; **Eluent:** PE/EA = 50:1 (R_f = 0.32).

1H NMR (600 MHz, $CDCl_3$) δ : 7.44 (d, J = 6.4 Hz, 2H), 7.23 (s, 1H), 7.22 (s, 2H), 2.39 (s, 4H), 1.56 – 1.51 (m, 4H). **$^{13}C\{^1H\}$ NMR (151 MHz, $CDCl_3$)** δ : 135.9, 134.9, 128.4, 126.7, 71.6, 66.2, 32.7, 31.6, 19.7, 19.4. **$^{11}B\{^1H\}$ NMR (193 MHz, $CDCl_3$)** δ : 4.7 (1B), -5.2 (1B), -8.7 (2B), -9.8 (2B), -10.4 (2B), -12.3 (2B).

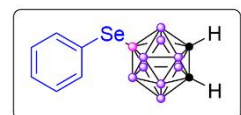
HRMS (ESI-TOF): m/z calcd for $C_{12}B_{10}H_{23}S$ $[M+H]^+$: 307.2525. Found: 307.2524.



1k: Yield 70%; White solid; **Eluent:** PE/EA = 50:1 (R_f = 0.32).

1H NMR (600 MHz, CD_2Cl_2) δ : 7.44 (d, J = 6.2 Hz, 2H), 7.23 (s, 1H), 7.22 (s, 2H), 2.48 – 2.38 (m, 6H). **$^{13}C\{^1H\}$ NMR (151 MHz, $CDCl_3$)** δ : 135.9, 134.9, 128.4, 126.7, 82.9, 77.8, 34.4, 33.3, 32.3. **$^{11}B\{^1H\}$ NMR (193 MHz, $CDCl_3$)** δ : 4.2 (1B), -6.0 (1B), -7.5 (2B), -9.7 (2B), -11.8 (2B), -12.5 (2B).

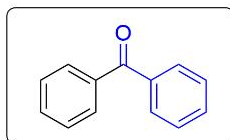
HRMS (ESI-TOF) m/z calcd for $C_{11}B_{10}H_{21}S$ $[M+H]^+$: 293.2368. Found: 293.2369.



2a: White solid; **Eluent:** PE/EA = 50:1 (R_f = 0.28), Mp: 94-97 °C.

1H NMR (400 MHz, $CDCl_3$) δ : 7.58 (d, J = 7.2 Hz, 2H), 7.28 (d, J = 3.2 Hz, 1H), 7.26 – 7.20 (m, 2H), 3.55 (s, 2H). **$^{13}C\{^1H\}$ NMR (101 MHz, $CDCl_3$)** δ : 136.2, 129.1, 128.6, 127.0, 53.7, 49.4. **$^{11}B\{^1H\}$ NMR (128 MHz, $CDCl_3$)** δ : 3.0 (1B), -2.0 (1B), -8.4 (2B), -14.0 (4B), -15.1 (2B). **IR:** 720, 846, 1023, 1134, 2556, 3052 cm^{-1} .

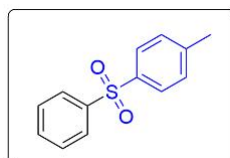
HRMS (ESI-TOF): m/z calcd for $C_8B_{10}H_{16}SeK$ $[M+K]^+$: 339.1047. Found: 339.1070. ^[1]



3a: Yield 72%; White solid; **Eluent:** PE/EA = 50:1 (R_f = 0.32), Mp: 47-49 °C.

^1H NMR (400 MHz, CD_3CN) δ : 7.76 (d, J = 7.2 Hz, 4H), 7.64 (t, J = 7.4 Hz, 2H), 7.52 (t, J = 7.6 Hz, 4H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CD_3CN)** δ : 197.2, 138.5, 133.4, 130.7, 129.3. **IR:** 634, 688, 943, 1269, 1448, 1646 cm^{-1} .

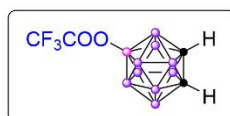
HRMS (ESI-TOF): m/z calcd for $\text{C}_{13}\text{H}_{10}\text{ONH}_4$ $[\text{M}+\text{NH}_4]^+$: 200.1070. Found: 200.1076. ^[2]



4a: Yield 97%; White solid; **Eluent:** DCM/EA = 20:1 (R_f = 0.28), Mp: 127-129 °C.

^1H NMR (400 MHz, CD_3CN) δ : 8.59 (d, J = 7.2 Hz, 2H), 8.49 (d, J = 8.1 Hz, 2H), 8.29 (t, J = 7.3 Hz, 1H), 8.23 (t, J = 7.4 Hz, 2H), 8.04 (d, J = 8.0 Hz, 2H), 3.05 (s, 3H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CD_3CN)** δ : 145.8, 142.9, 139.6, 134.3, 131.1, 130.5, 128.4, 128.2, 21.5. **IR:** 548, 651, 688, 723, 813, 1101, 1155, 1293, 1448 cm^{-1} .

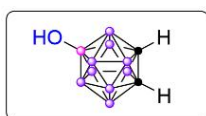
HRMS (ESI-TOF): m/z calcd for $\text{C}_{13}\text{H}_{12}\text{O}_2\text{SNa}$ $[\text{M}+\text{Na}]^+$: 225.0450. Found: 255.0456. ^[3]



5a: White solid; **Eluent:** PE/DCM = 3:1 (R_f = 0.28), Mp: 100-102 °C.

^1H NMR (400 MHz, CDCl_3) δ : 3.54 (s, 2H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)** δ : 155.8 (q, J = 41.41), 114.5 (q, J = 286.84), 50.5, 44.3. **$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)** δ : 10.0 (1B), -3.5 (1B), -10.4 (2B), -115.0 (2B), -16.1 (2B), -17.2 (2B). **^{19}F NMR (377 MHz, CDCl_3)** δ : -75.88. **IR:** 506, 725, 873, 1028, 1162, 1388, 1771, 2613, 3091 cm^{-1} .

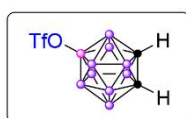
HRMS (ESI-TOF): m/z calcd for $\text{C}_4\text{B}_{10}\text{H}_{11}\text{O}_2\text{F}_3$ $[\text{M}+\text{H}]^+$: 257.1789. Found: 257.1791. ^[4]



5b: White solid; **Eluent:** PE/DCM = 1:1 (R_f = 0.28), Mp: 312-314 °C.

^1H NMR (400 MHz, CDCl_3) δ : 3.38 (s, 1H), 3.26 (s, 1H), 1.84 (s, 1H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)** δ : 49.1, 37.1. **$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)** δ : 14.7 (1B), -3.8 (1B), -10.2 (2B), -15.2 (2B), -16.8 (2B), -17.7 (2B). **IR:** 465, 721, 1028, 1122, 2548, 3070, 3272 cm^{-1} .

HRMS (ESI-TOF): m/z calcd for $\text{C}_2\text{B}_{10}\text{H}_{12}\text{O}_1$ $[\text{M}+\text{H}]^+$: 161.1965. Found: 161.1961. ^[4]

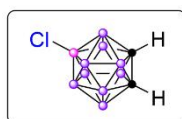


5c: Colorless oil; **Eluent:** PE/DCM = 4:1 (R_f = 0.26).

^1H NMR (400 MHz, CDCl_3) δ : 3.55 (s, 2H). **$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)** δ : 118.3 (q, J = 319.16 Hz), 50.6, 44.1. **$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)** δ : 9.8 (1B), -3.6 (1B), -10.4 (2B), -15.2

(2B), -16.0 (2B), -17.1 (2B). ^{19}F NMR (377 MHz, CDCl_3) δ : -75.91. IR: 509, 612, 886, 968, 1028, 1096, 1209, 1388, 2601, 3076 cm^{-1} .

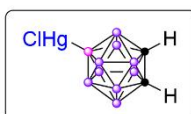
HRMS (ESI-TOF): m/z calcd for $\text{C}_3\text{B}_{10}\text{H}_{11}\text{SF}_3\text{O}_3$ $[\text{M}+\text{H}]^+$: 293.1459. Found: 293.1463. [4]



6a: White solid; Eluent: DCM/EA = 50:1 (R_f = 0.32), Mp: 221-223 $^{\circ}\text{C}$.

^1H NMR (400 MHz, CDCl_3) δ : 3.57 (s, 1H), 3.45 (s, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 52.1, 44.4. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) δ : 7.4 (1B), -2.1 (1B), -8.8 (2B), -13.8 (2B), -15.1 (2B), -16.3 (2B). IR: 531, 723, 873, 972, 1007, 1140, 1713, 2570, 3063 cm^{-1} .

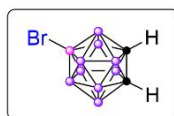
HRMS (ESI-TOF): m/z calcd for $\text{C}_2\text{B}_{10}\text{H}_{12}\text{Cl}$ $[\text{M}+\text{H}]^+$: 180.1599. Found: 180.1598. [5]



7a: White solid; Eluent: PE/EA = 3:1 (R_f = 0.26), Mp: 266-268 $^{\circ}\text{C}$.

^1H NMR (400 MHz, CDCl_3) δ : 3.74 (s, 1H), 3.54 (s, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 58.9, 57.0. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) δ : 3.5 (1B), -1.0 (1B), -8.1 (2B), -12.5 (4B), -13.7 (2B). IR: 716, 852, 966, 1141, 1211, 2564, 3031 cm^{-1} .

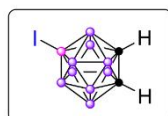
HRMS (ESI-TOF): m/z calcd for $\text{C}_2\text{B}_{10}\text{H}_{11}\text{HgClNH}_4$ $[\text{M}+\text{NH}_4]^+$: 397.1574. Found: 397.1581.



8a: White solid; Eluent: DCM/EA = 50:1 (R_f = 0.32), Mp: 182-184 $^{\circ}\text{C}$.

^1H NMR (400 MHz, CDCl_3) δ : 3.62 (s, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 53.2, 46.7. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) δ : 0.1 (1B), -1.6 (1B), -8.3 (2B), -13.5 (2B), -14.5 (2B), -15.7 (2B). IR: 725, 846, 966, 1137, 1207, 2595, 3058 cm^{-1} .

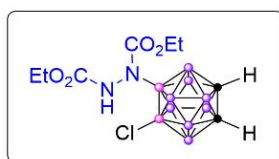
HRMS (ESI-TOF): m/z calcd for $\text{C}_2\text{B}_{10}\text{H}_{11}\text{BrNH}_4$ $[\text{M}+\text{NH}_4]^+$: 241.1372. Found: 241.1390. [5]



9a: White solid; Eluent: DCM/EA = 50:1 (R_f = 0.32), Mp: 119-120 $^{\circ}\text{C}$.

^1H NMR (600 MHz, CDCl_3) δ : 3.87 (s, 1H), 3.66 (s, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3) δ : 54.8, 50.7. $^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3) δ : -0.9 (1B), -7.5 (2B), -12.8 (2B), -13.4 (2B), -14.8 (2B), -16.6 (1B). IR: 725, 832, 966, 1137, 1204, 2579, 3048 cm^{-1} .

HRMS (ESI-TOF): m/z calcd for $\text{C}_2\text{B}_{10}\text{H}_{11}\text{IK}$ $[\text{M}+\text{K}]^+$: 309.0542. Found: 309.0525. [5]

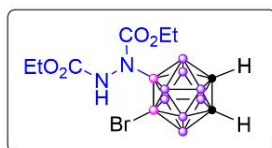


10a: Yield 58%; White solid; Eluent: PE/EA = 2:1 (R_f = 0.26).

^1H NMR (600 MHz, CDCl_3) δ : 6.70 – 6.30 (m, 1H), 4.13 (s, 4H), 3.66 (s, 1H), 3.56 (s, 1H), 1.24 (t, J = 7.1 Hz, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3) δ : 157.7, 156.3, 62.5, 61.6, 44.4, 43.1, 14.4,

14.2. $^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3) δ : 7.3 (1B), 6.4 (1B), -9.7 (1B), -10.8 (1B), -16.2 (4B), -18.8 (2B).

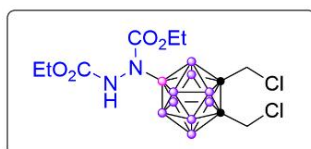
HRMS (ESI-TOF): m/z calcd for $\text{C}_8\text{B}_{10}\text{H}_{22}\text{N}_2\text{O}_4\text{Cl}$ $[\text{M}+\text{H}]^+$: 354.2243. Found: 354.2245.



10b: Yield 50%; White solid; **Eluent**: PE/EA = 2:1 (R_f = 0.32).

^1H NMR (600 MHz, CDCl_3) δ : 6.75 – 6.35 (m, 1H), 4.22 – 4.08 (m, 4H), 3.59 (s, 2H), 1.27 (t, J = 7.1 Hz, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3) δ : 157.6, 156.3, 62.6, 61.6, 45.3, 45.2, 14.5, 14.2. $^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3) δ : 7.6 (1B), -0.4 (1B), -9.1 (1B), -10.4 (1B), -16.0 (4B), -18.3 (2B).

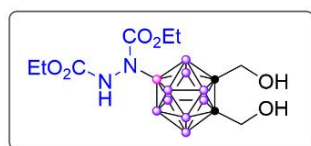
HRMS (ESI-TOF): m/z calcd for $\text{C}_8\text{B}_{10}\text{H}_{22}\text{N}_2\text{O}_4\text{Br}$ $[\text{M}+\text{H}]^+$: 398.1750. Found: 398.1749.



10d: Yield 45%; White solid; **Eluent**: PE/EA = 2:1 (R_f = 0.26).

^1H NMR (400 MHz, CDCl_3) δ : 6.52 – 6.05 (m, 1H), 4.20 (s, 2H), 4.15 (s, 4H), 1.25 (t, J = 7.1 Hz, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 157.9, 156.4, 62.4, 61.7, 43.9, 42.9, 14.4, 14.3. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) 8.5 (1B), -4.3 (1B), -10.9 (4B), -12.5 (4B).

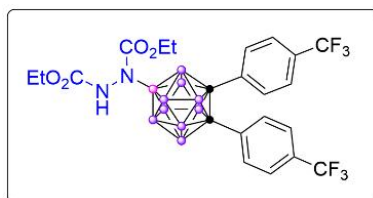
HRMS (ESI-TOF): m/z calcd for $\text{C}_{10}\text{B}_{10}\text{H}_{25}\text{N}_2\text{O}_4\text{Cl}_2$ $[\text{M}+\text{H}]^+$: 416.2173. Found: 416.2174. ^[6]



10e: Yield 60%; White solid; **Eluent**: PE/EA = 4:1 (R_f = 0.32)

^1H NMR (400 MHz, CDCl_3) δ : 6.50 – 6.10 (m, 1H), 4.17 – 4.01 (m, 4H), 2.01 (s, 3H), 2.00 (s, 3H), 1.21 (t, J = 7.1 Hz, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 158.0, 156.3, 69.5, 64.4, 62.0, 61.3, 23.5, 21.4, 14.3, 14.2. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) 8.7 (1B), -6.2 (1B), -10.9 (8B).

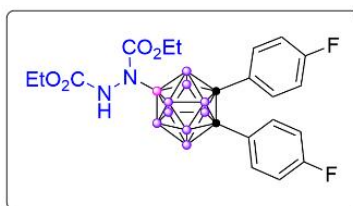
HRMS (ESI-TOF): m/z calcd for $\text{C}_{10}\text{B}_{10}\text{H}_{26}\text{N}_2\text{O}_4\text{Na}$ $[\text{M}+\text{Na}]^+$: 369.2793. Found: 369.2798.



10f: Yield 62%; White solid; **Eluent**: PE/EA = 4:1 (R_f = 0.28).

^1H NMR (400 MHz, CDCl_3) δ : 7.56 (t, J = 8.0 Hz, 4H), 7.44 (d, J = 8.2 Hz, 4H), 6.55 – 6.13 (m, 1H), 4.26 – 4.05 (m, 4H), 1.29 – 1.21 (m, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ : 157.9, 156.6, 132.7 (dd, J_1 = 71.71 Hz, J_2 = 31.31 Hz), 131.2, 131.0, 125.6 (q, J = 3.03 Hz), 123.1 (q, J = 273.71 Hz), 79.4, 74.4, 62.4, 61.7, 14.5, 14.3. $^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3) δ : 9.8 (1B), -3.0 (1B), -10.7 (8B). ^{19}F NMR (377 MHz, CDCl_3) δ : -63.23 (d, J = 2.6 Hz).

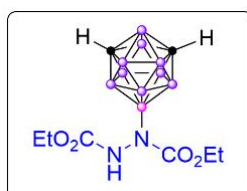
HRMS (ESI-TOF): m/z calcd for $C_{22}B_{10}H_{29}N_2O_4F_6$ $[M+H]^+$: 607.3040. Found: 607.3041. ^[6]



10g: Yield 55%; White solid; **Eluent:** PE/EA = 4:1 (R_f = 0.28).

¹H NMR (400 MHz, CDCl₃) δ : 7.49 – 7.36 (m, 4H), 6.84 (t, J = 8.4 Hz, 4H), 6.54 – 6.15 (m, 1H), 4.27 – 4.06 (m, 4H), 1.32 – 1.20 (m, 6H). **¹³C{¹H} NMR (101 MHz, CDCl₃)** δ : 163.7 (d, J = 253.51 Hz), 158.0, 156.6, 132.9 (d, J = 9.09 Hz), 126.3, 125.3, 115.6 (d, J = 22.22 Hz), 80.6, 75.2, 62.3, 61.6, 14.4, 14.3. **¹¹B{¹H} NMR (128 MHz, CDCl₃)** δ : 9.5 (1B), -3.0 (1B), -10.7 (8B). **¹⁹F NMR (377 MHz, CDCl₃)** δ : -109.24.

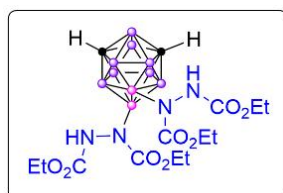
HRMS (ESI-TOF): m/z calcd for $C_{20}B_{10}H_{29}N_2O_4F_2$ $[M+H]^+$: 507.3103. Found: 507.3101. ^[6]



10h: Yield 65%; White solid; **Eluent:** PE/EA = 4:1 (R_f = 0.26), Mp:115-117 °C.

¹H NMR (400 MHz, CDCl₃) δ : 6.61 – 6.26 (m, 1H), 4.27 – 4.05 (m, 4H), 2.84 (s, 2H), 1.25 (t, J = 7.1 Hz, 6H). **¹³C{¹H} NMR (101 MHz, CDCl₃)** δ : 158.1, 156.5, 62.4, 61.7, 51.4, 14.4, 14.3. **¹¹B{¹H} NMR (128 MHz, CDCl₃)** δ : 2.2 (1B), -7.4 (2B), -11.0 (1B), -14.0 (2B), -15.5 (2B), -19.0 (1B), -22.0 (1B). **IR:** 743, 1065, 1225, 1277, 1674, 2613, 3056, 3262 cm^{-1} .

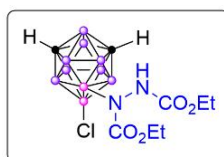
HRMS (ESI-TOF): m/z calcd for $C_8B_{10}H_{23}N_2O_4$ $[M+H]^+$: 319.2660. Found: 319.2661. ^[6]



10h': Yield 26%; White solid; **Eluent:** PE/EA = 2:1 (R_f = 0.28).

¹H NMR (400 MHz, CDCl₃) δ : 7.65 – 6.69 (m, 2H), 4.17 (s, 8H), 2.73 (s, 2H), 1.31 – 1.17 (m, 12H). **¹³C{¹H} NMR (101 MHz, CDCl₃)** δ : 157.6, 156.4, 62.2, 61.6, 46.0, 14.5, 14.3. **¹¹B{¹H} NMR (128 MHz, CDCl₃)** δ : 0.8 (1B), 0.5 (1B), -9.5 (2B), -16.0 (4B), -24.0 (2B).

HRMS (ESI-TOF): m/z calcd for $C_{14}B_{10}H_{32}N_4O_8K$ $[M+K]^+$: 503.2801. Found: 503.2839.

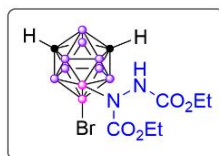


10i: Yield 61%; White solid; **Eluent:** PE/EA = 4:1 (R_f = 0.32).

¹H NMR (600 MHz, CDCl₃) δ : 6.75 – 6.30 (m, 1H), 4.21 (d, J = 7.0 Hz, 4H), 2.83 (s, 1H), 2.79 (s, 1H), 1.28 – 1.24 (m, 6H). **¹³C{¹H} NMR (151 MHz, CDCl₃)** δ : 156.5, 62.7, 61.8, 47.6, 14.5,

14.3. $^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3) δ : 1.3 (1B), -0.3 (1B), -7.5 (1B), -8.4 (1B), -15.3 (4B), -23.9 (2B).

HRMS (ESI-TOF): m/z calcd for $\text{C}_8\text{B}_{10}\text{H}_{22}\text{N}_2\text{O}_4\text{Cl}$ $[\text{M}+\text{H}]^+$: 354.2243. Found: 354.2247.



10j: Yield 48%; White solid; **Eluent**: PE/EA = 4:1 (R_f = 0.32).

^1H NMR (600 MHz, CDCl_3) δ : 6.80 – 6.45 (m, 1H), 4.20 (s, 4H), 2.90 (s, 1H), 2.86 (s, 1H), 1.32 – 1.20 (m, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CDCl_3) δ : 157.8, 156.6, 62.7, 61.8, 49.2, 48.9, 14.3,

14.2. $^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3) δ : 1.6 (1B), -7.2 (3B), -14.8 (4B), -22.7 (2B).

HRMS (ESI-TOF): m/z calcd for $\text{C}_8\text{B}_{10}\text{H}_{22}\text{N}_2\text{O}_4\text{Br}$ $[\text{M}+\text{H}]^+$: 398.1750. Found: 398.1751.

4. References

1. Bai, F.; Yan, H. **2021**. DOI:10.27235/d.cnki.gnjju.2021.002442.
2. Chena, J.; Chena, S.; Tanga, Y.; Moub, C.; Tsaia, F. *J. Mol. Catal. A. Chem.* **2009**, *307*, 88–92.
3. Falconnet, A.; Arndt, J.; Hashmi, A. S. K.; Schaub, T. *Eur. J. Org. Chem.* **2022**, e202200477, 1-5.
4. Ma, Y.-N.; Ren, H.; Wu, Y.; Li, N.; Chen, F.; Chen, X.-N. *J. Am. Chem. Soc.* **2023**, *145*, 7331–7342.
5. Lu, W.; Wu, Y.; Ma, Y.-N.; Chen, F.; Chen, X.-N. *Inorg. Chem.* **2023**, *62*, 885–892.
6. Wang, Y.; Gao, Y.; Guo, W.; Zhao, Q.-Y.; Ma, Y.-N.; Chen, X.-N. *Org. Chem. Front.*, **2022**, *9*, 4975–4980.

5. X-ray Structure Determination

Single crystal XRD analysis of the **7a** compound was recorded on a SuperNova Dual diffractometer using graphite-monochromated $\text{Cu K}\alpha$ radiation, $\lambda = 1.54184 \text{ \AA}$. The linear absorption coefficients, scattering factors for the atoms, and anomalous dispersion corrections were taken from the International Tables for X-Ray Crystallography. Empirical absorption corrections were applied. Structures were solved using direct methods (SHELXT) and refined by full-matrix least-squares (SHELXL) interfaced with the programme OLEX2. Crystal data and details of data collection and structure refinements are given in **Table S2**.

Further details on the crystal structure investigations can be obtained free of charge from The Cambridge Crystallographic Data Centre via <https://www.ccdc.cam.ac.uk/structures/> by quoting the depository numbers CCDC- 2358875 (**7a**).

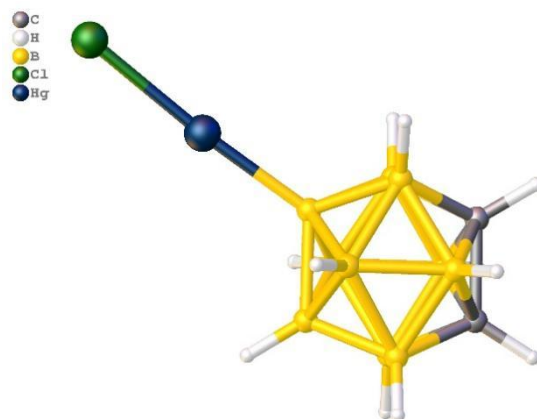


Figure S2. Molecular Structure of **7a**

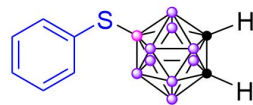
Table S1. Crystal Data and Summary of Data Collection and Refinement.

compound	7a
formula	C ₂ H ₁₁ B ₁₀ ClHg
Formula weight	379.25
Temperature/K	293(2)
Crystal system	monoclinic
Space group	P2 ₁ /n
a/Å	6.73953(18)
b/Å	21.6921(6)
c/Å	7.3452(2)
α/°	90
β/°	91.737(2)
γ/°	90
Volume/Å ³	1073.33(5)
Z	4
ρ _{calc} /cm ³	2.347
μ/mm ⁻¹	27.444
F(000)	680.0
Crystal size/mm ³	0.138 × 0.123 × 0.059
Radiation	Cu Kα (λ = 1.54184)
Reflections collected	3825
Independent reflections	[R _{int} = 0.0512, R _{sigma} = 0.0634]
Goodness-of-fit on F ²	1.044
Final R indexes [I ≥ 2σ (I)]	R1 = 0.0434, wR2 = 0.1166
Final R indexes [all data]	R1 = 0.0508, wR2 = 0.1238

6. NMR spectra

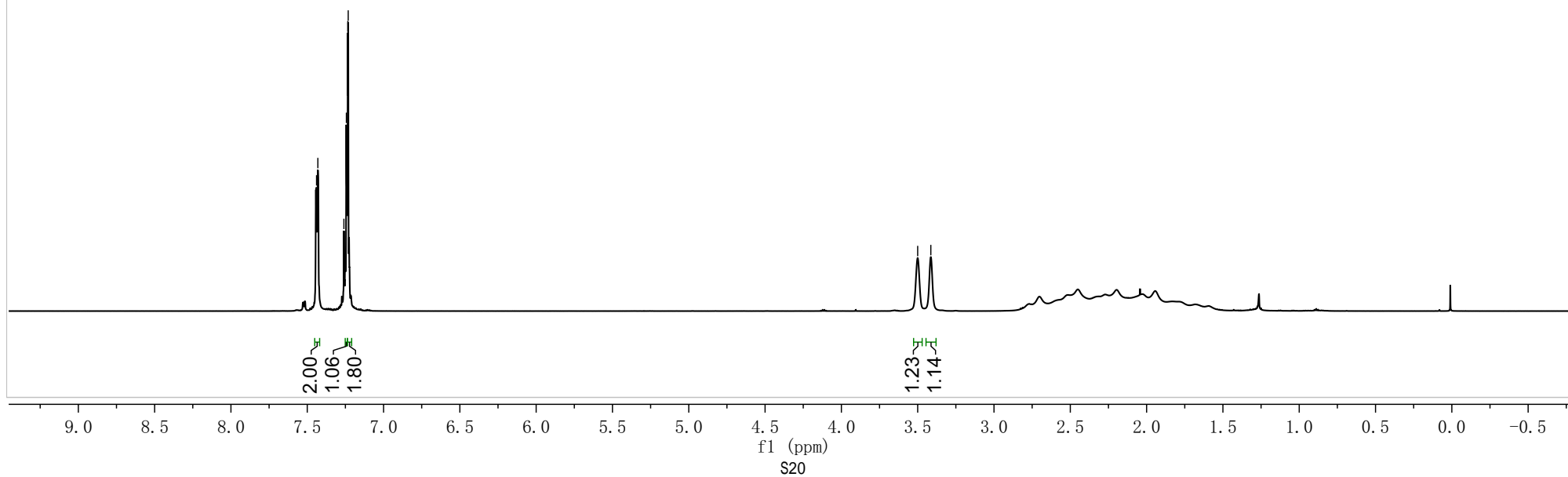
7.44
7.43
7.26
7.24
7.23
7.23

3.50
3.41



1a

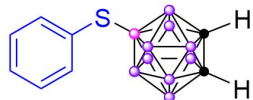
¹H NMR (600 MHz, CDCl₃)



7.44
7.43
7.26
7.24
7.23

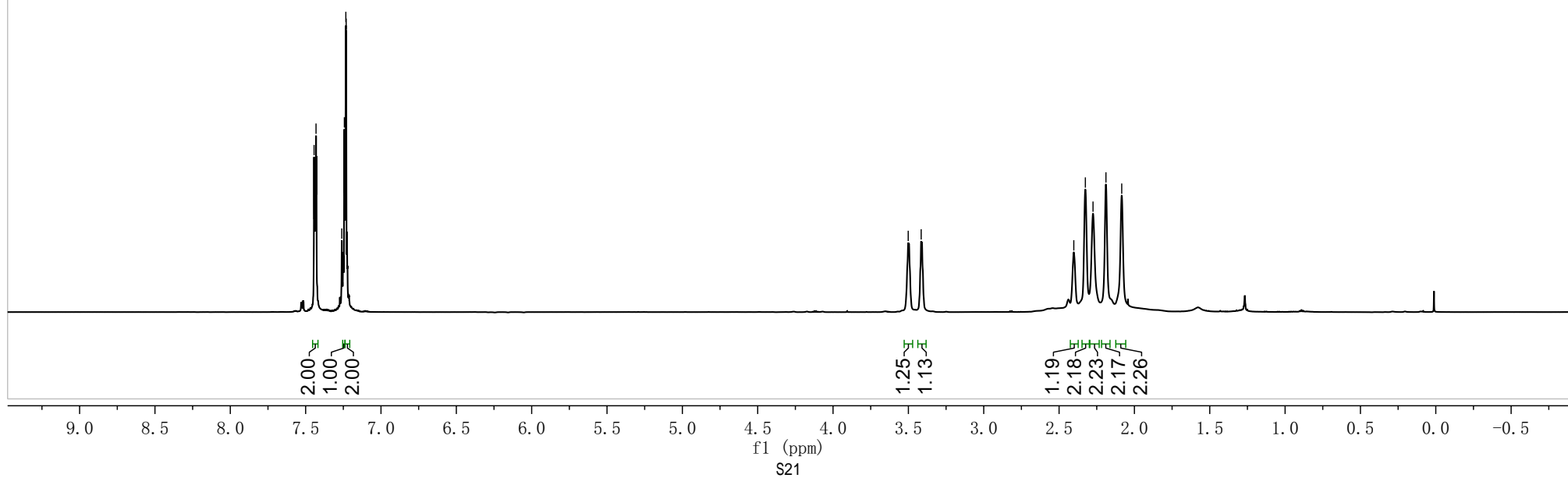
3.50
3.42

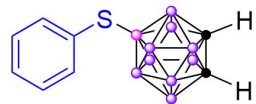
2.40
2.33
2.27
2.19
2.08



1a

$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





1a

^{11}B NMR (193 MHz, CDCl_3)

—7.45

—1.87

—2.65

—8.23

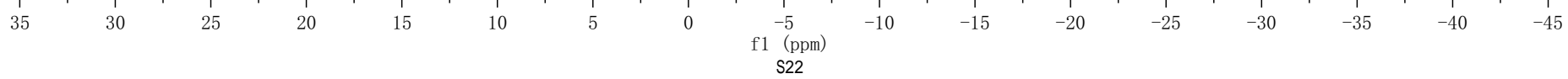
—9.02

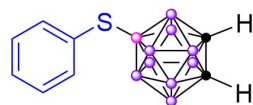
—13.48

—14.22

—15.03

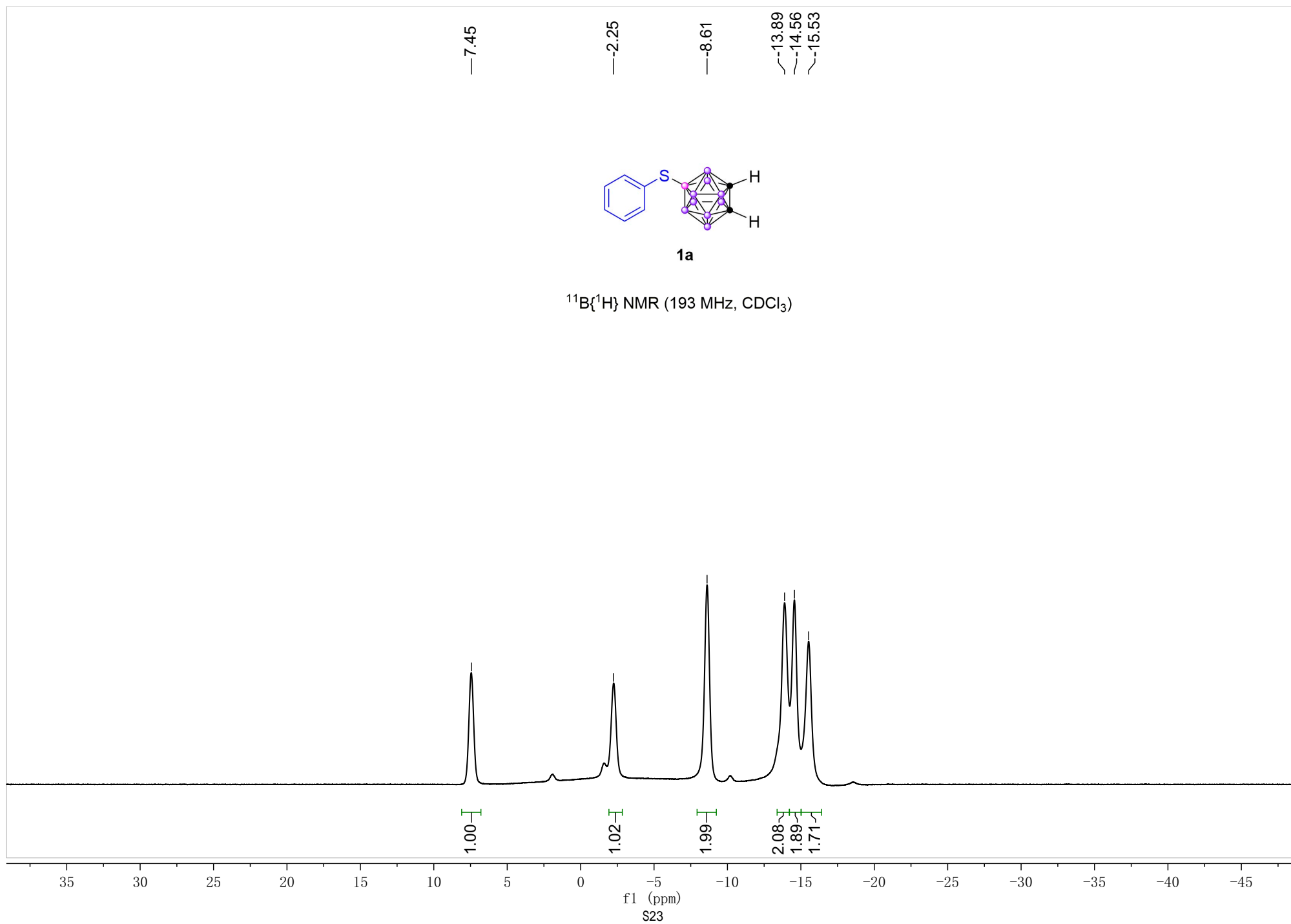
—16.01

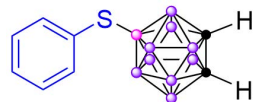




1a

$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)





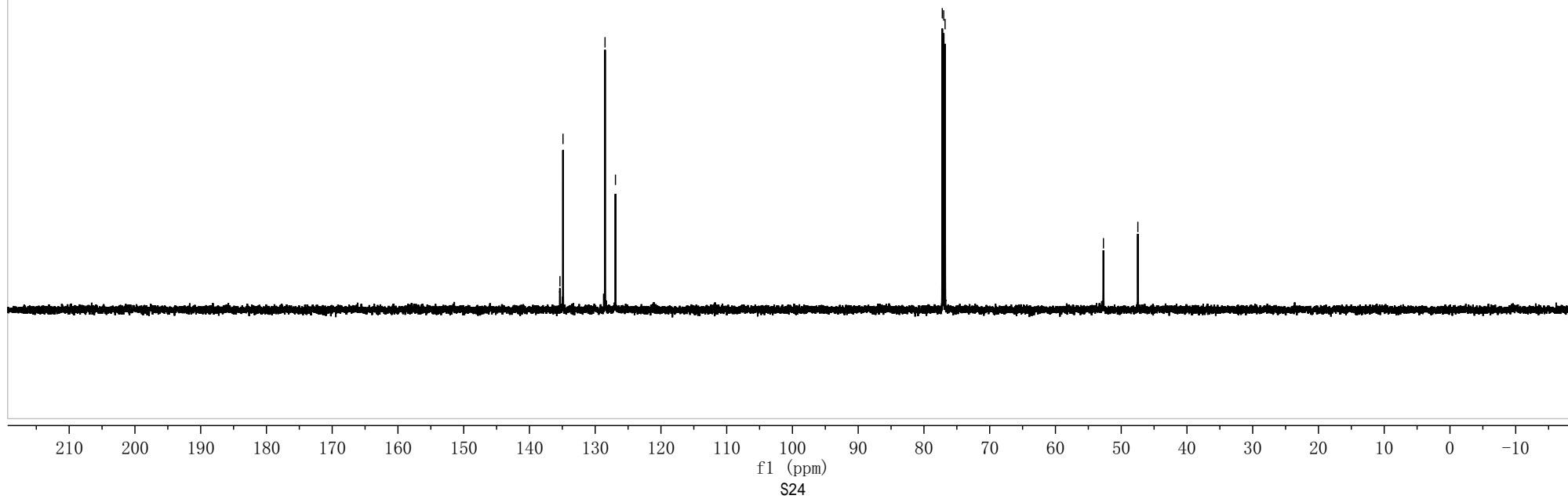
1a

^{13}C NMR (151 MHz, CDCl_3)

135.36
134.90
128.51
126.92

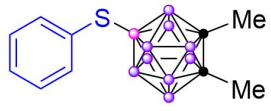
77.21
77.00
76.79

52.70
47.47



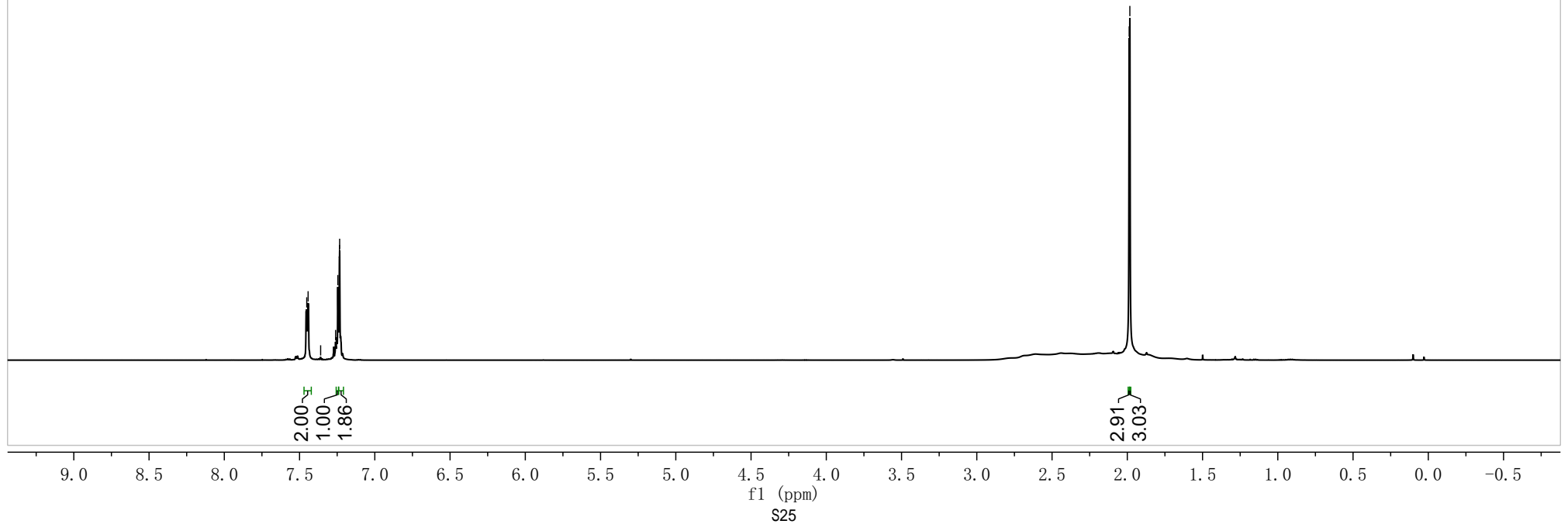
7.45
7.44
7.36
7.26
7.25
7.23
7.23

1.99
1.98



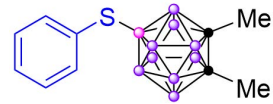
1b

¹H NMR (600 MHz, CDCl₃)



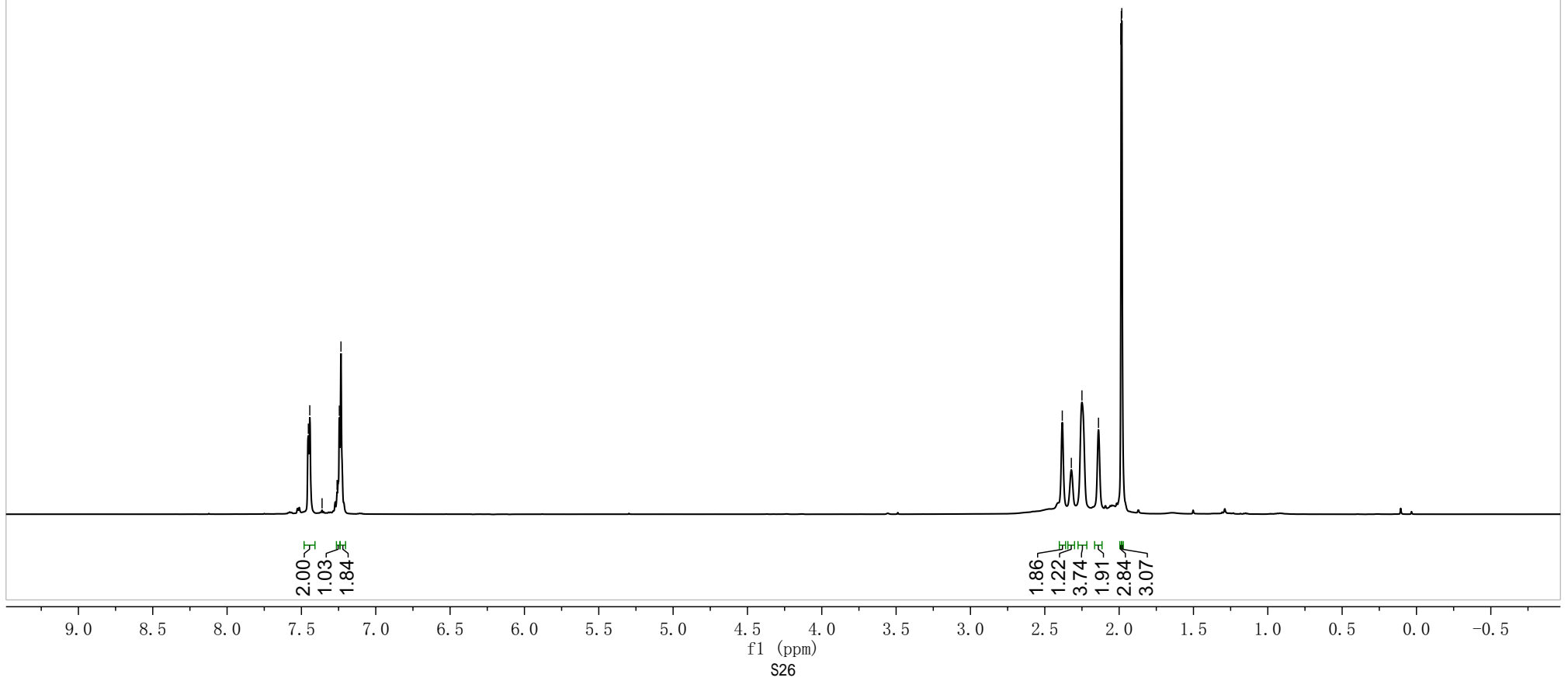
7.45
7.44
7.36
7.26
7.25
7.23

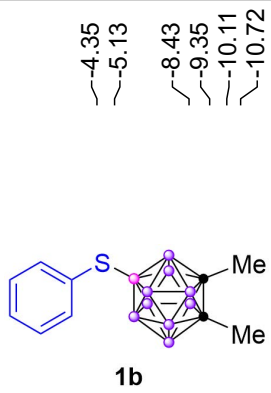
2.38
2.32
2.25
2.14
1.99
1.98



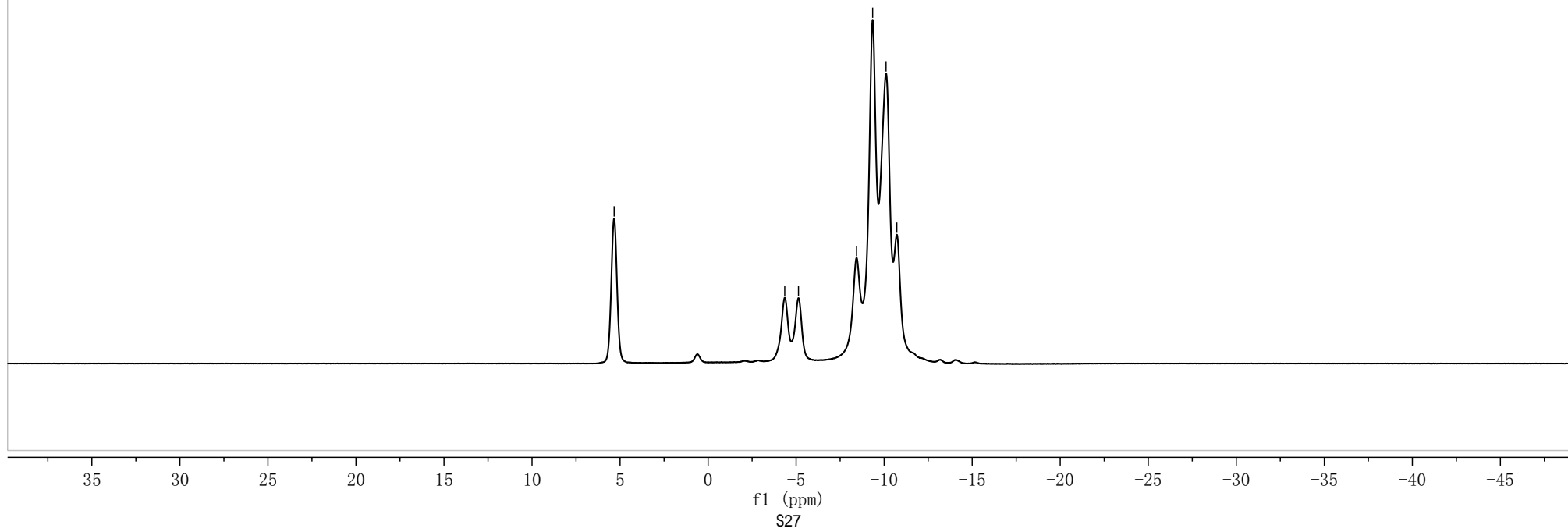
1b

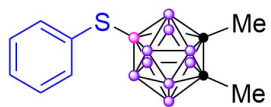
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





^{11}B NMR (193 MHz, CDCl_3)





1b

$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

—5.33

—4.75

—8.88

—9.75

—10.28

1.00

1.13

1.91

4.02

1.98

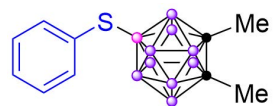
35 30 25 20 15 10 5 0 f1 (ppm) -5 -10 -15 -20 -25 -30 -35 -40 -45

S28

135.75
134.81
128.38
126.71

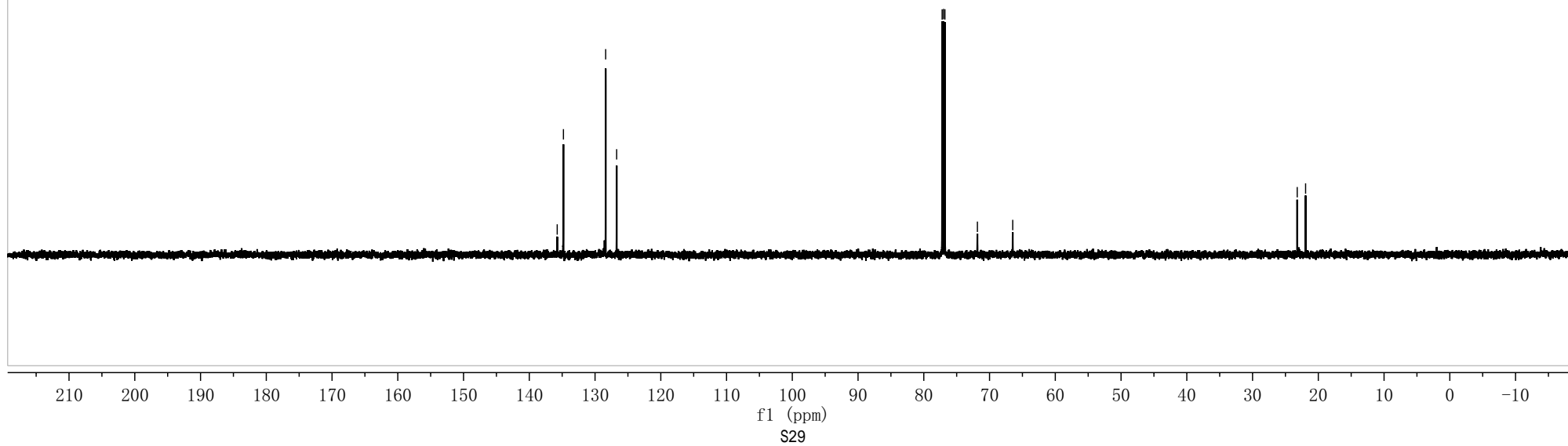
77.21
77.00
76.79
71.85
66.47

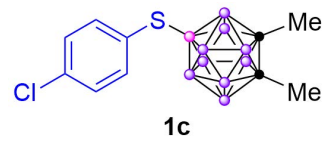
23.21
21.93



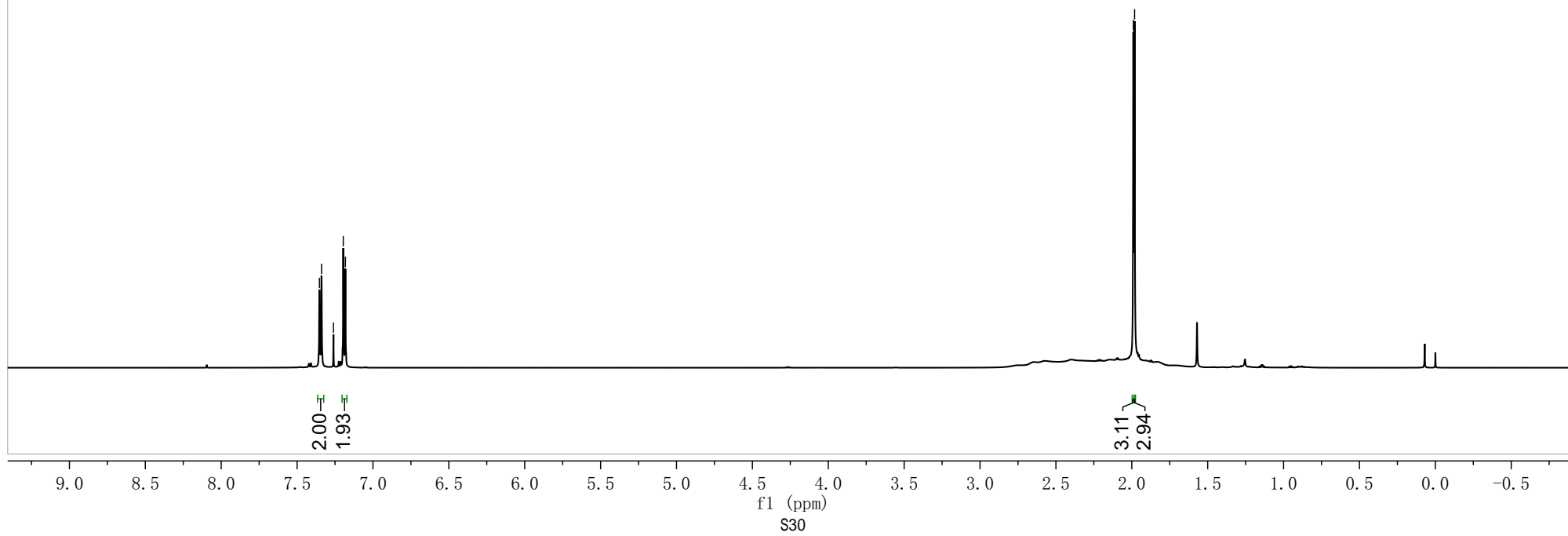
1b

¹³C NMR (151 MHz, CDCl₃)



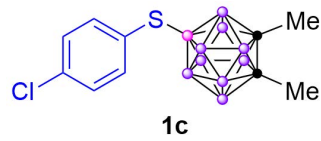


^1H NMR (600 MHz, CDCl_3)

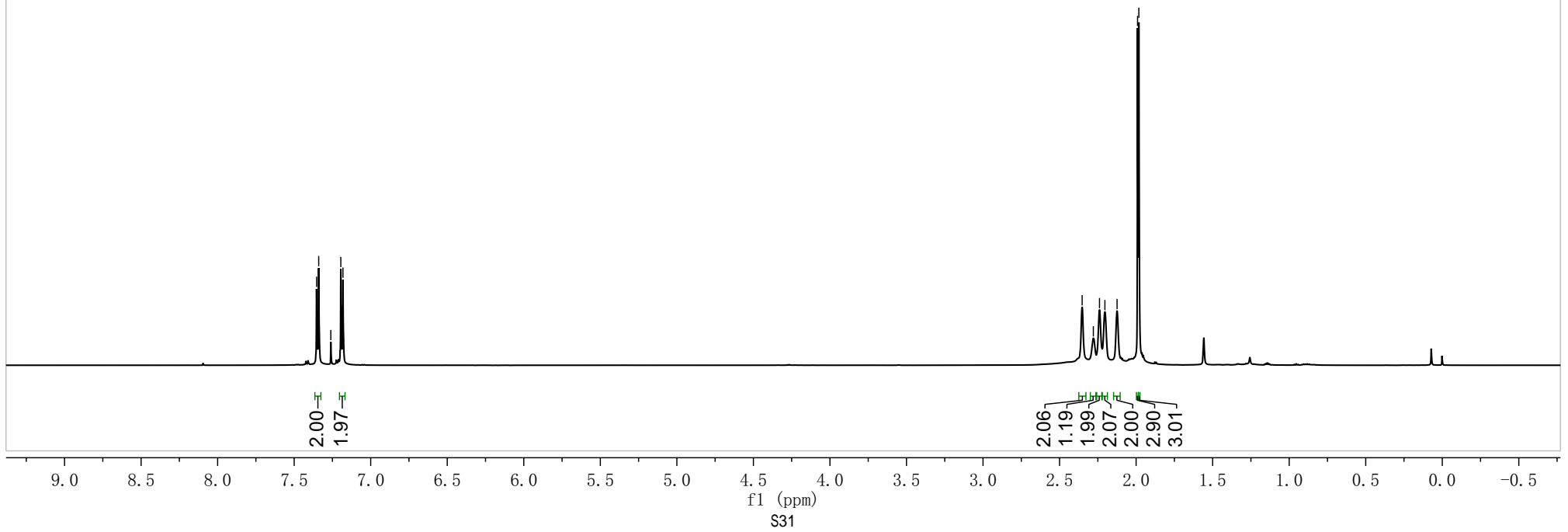


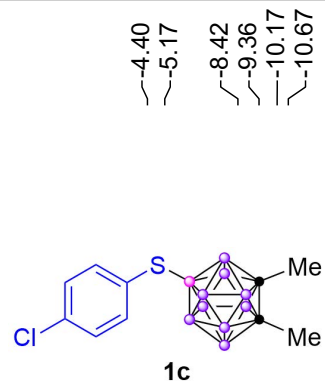
7.35
7.34
7.26
7.20
7.18

2.35
2.28
2.24
2.20
2.12
1.99
1.98

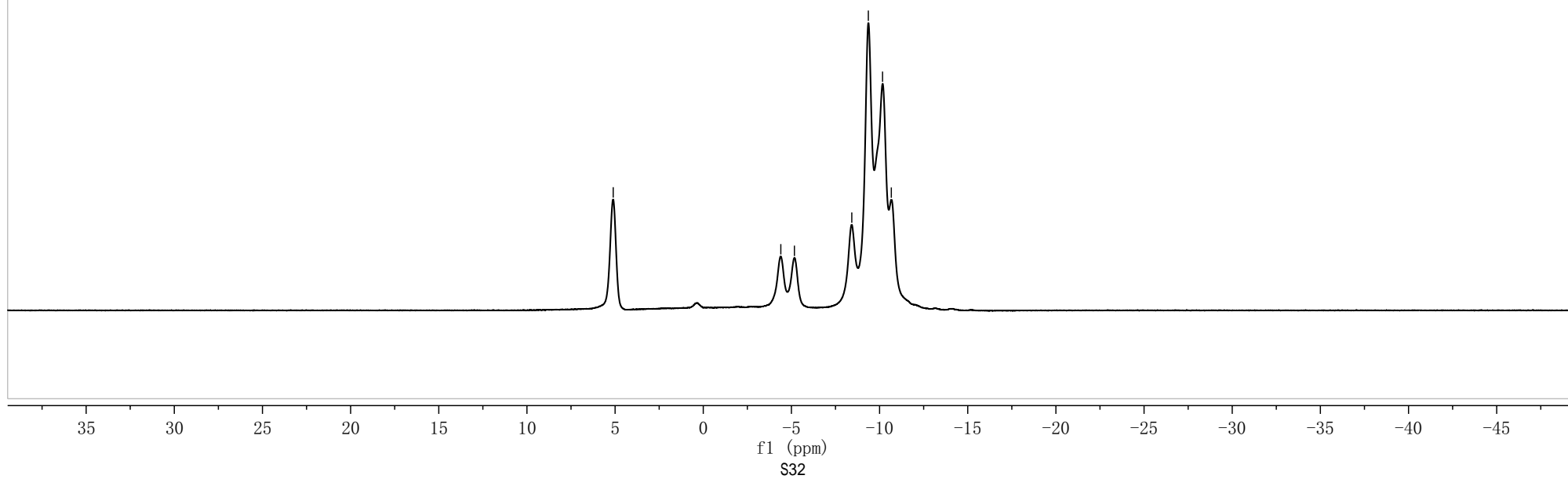


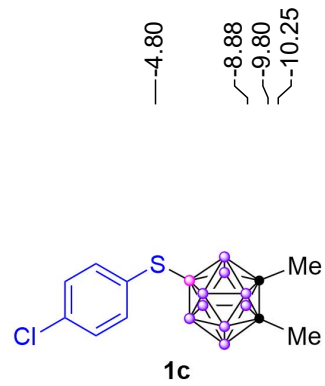
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)



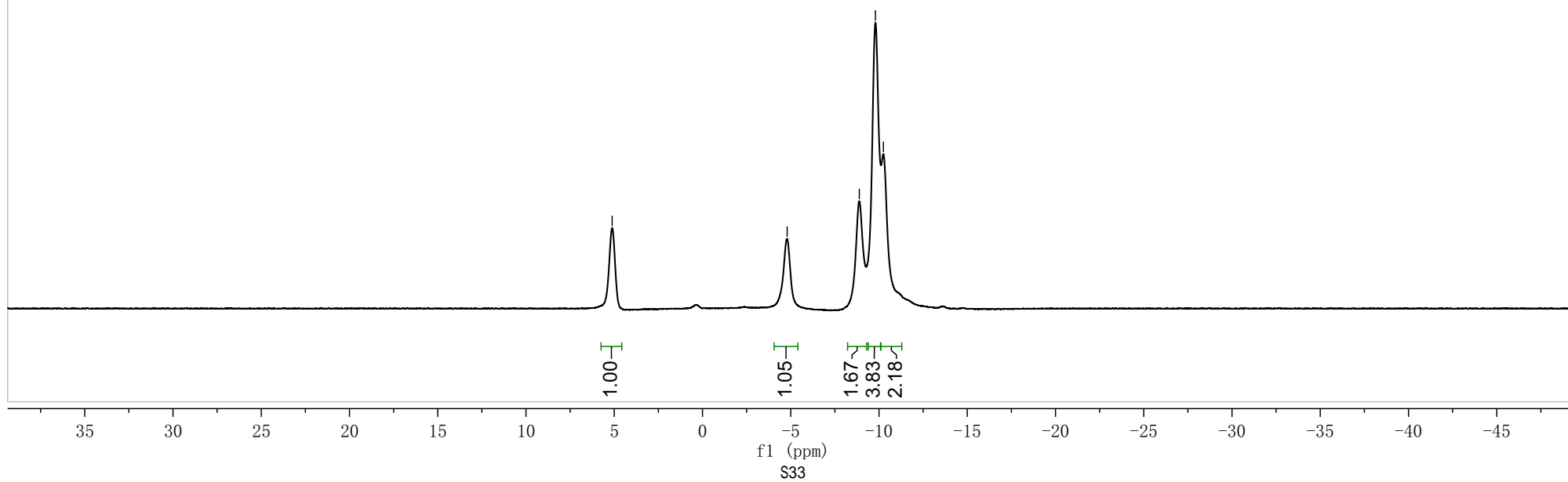


¹¹B NMR (193 MHz, CDCl₃)





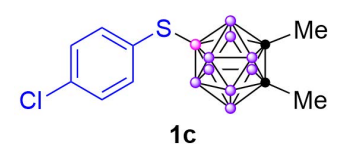
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)



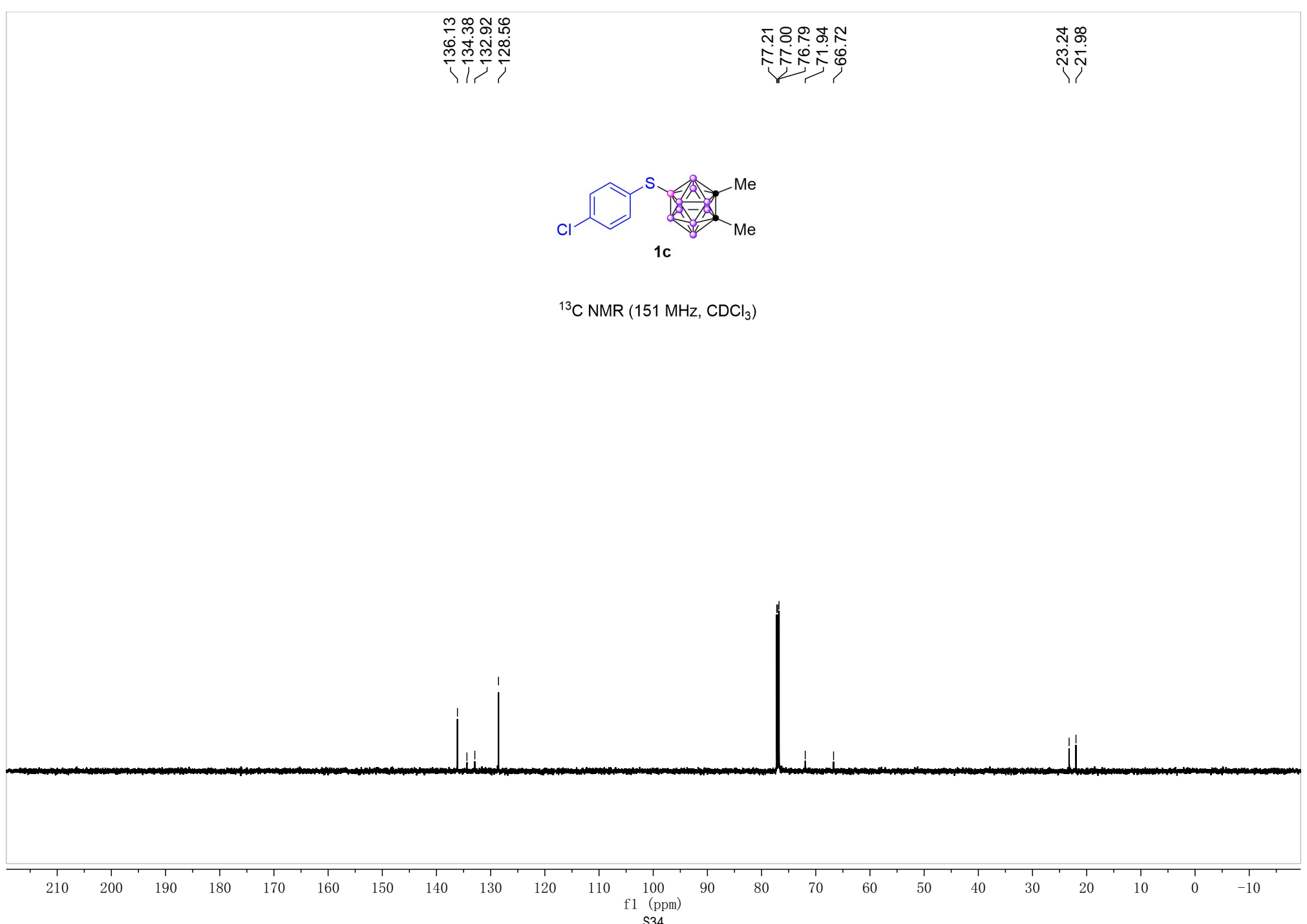
136.13
134.38
132.92
128.56

77.21
77.00
76.79
71.94
66.72

23.24
21.98

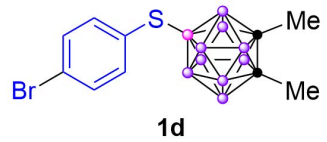


¹³C NMR (151 MHz, CDCl₃)

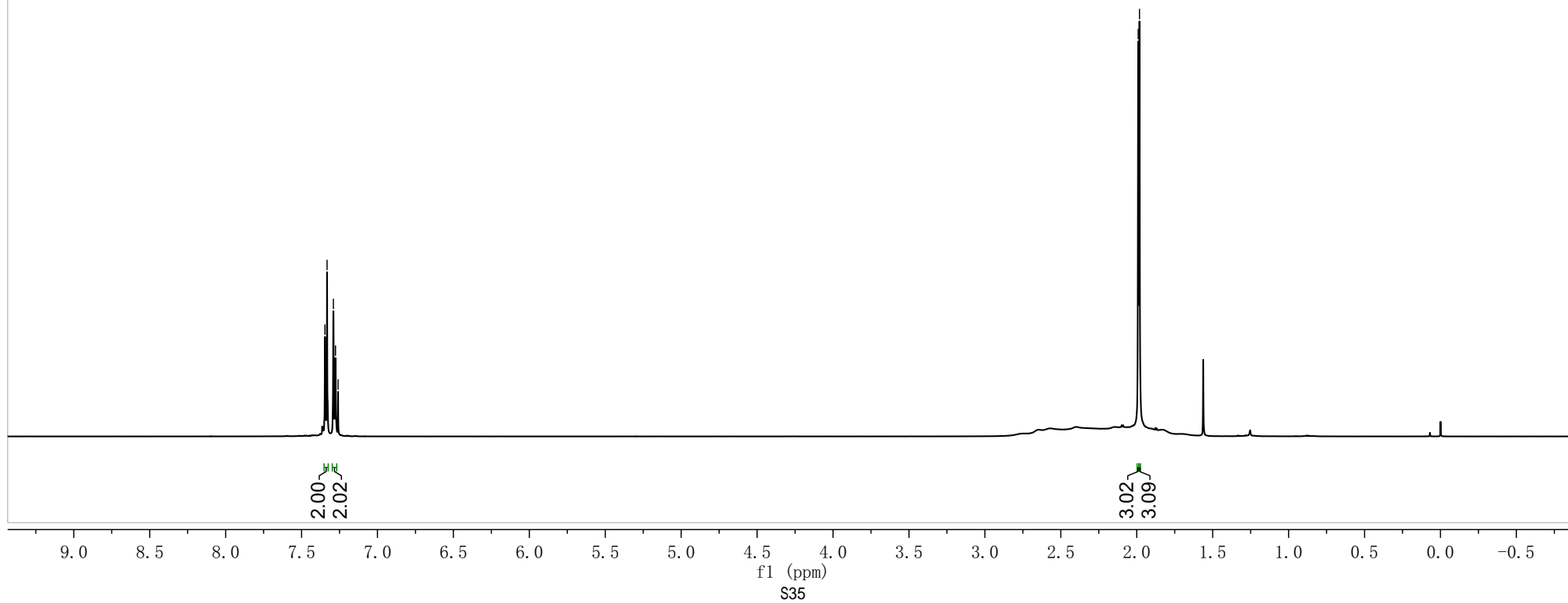


7.35
7.33
7.29
7.28
7.26

1.99
1.98

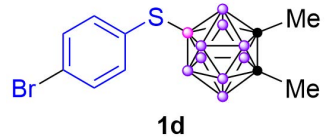


¹H NMR (600 MHz, CDCl₃)

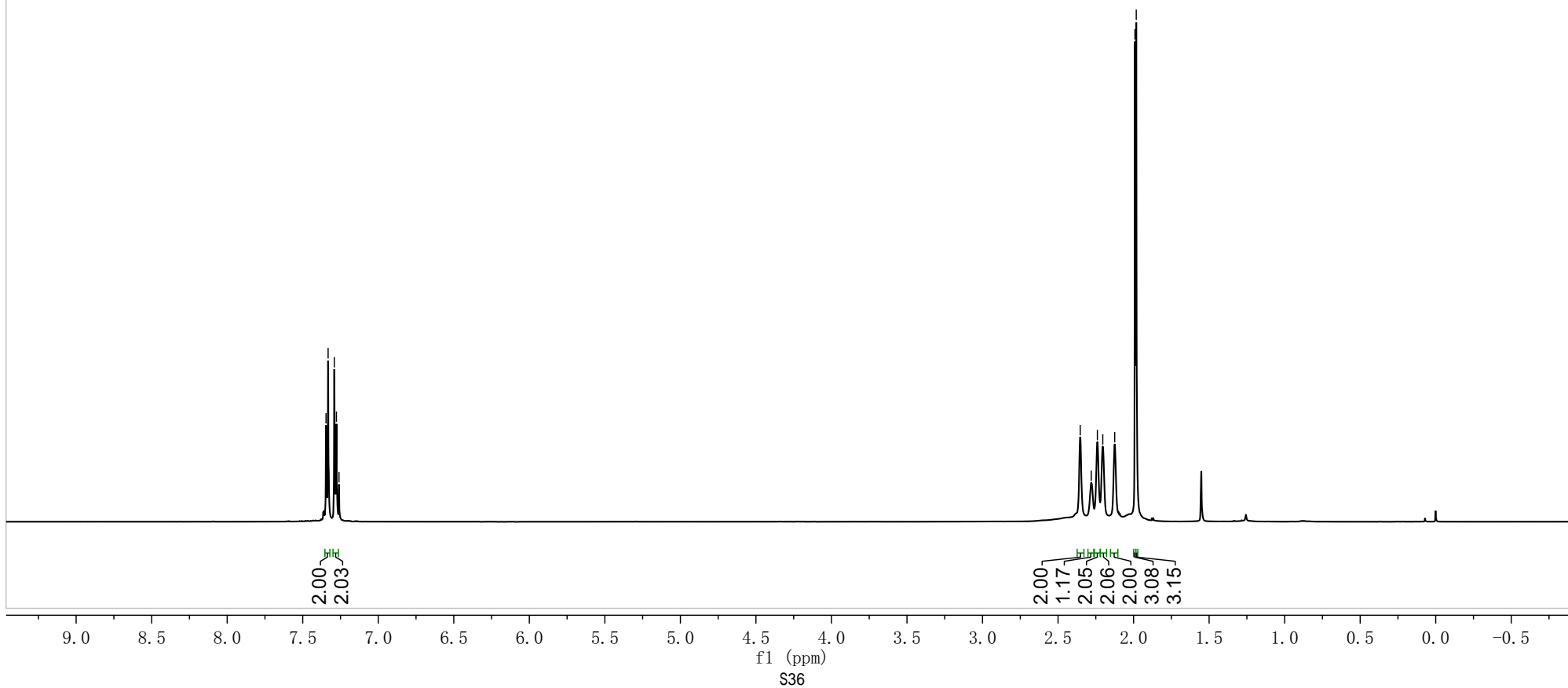


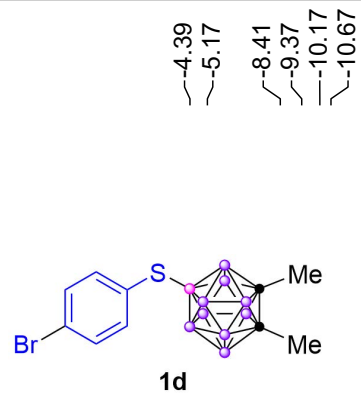
7.35
7.33
7.29
7.28
7.26

2.35
2.28
2.24
2.20
2.12
1.99
1.98

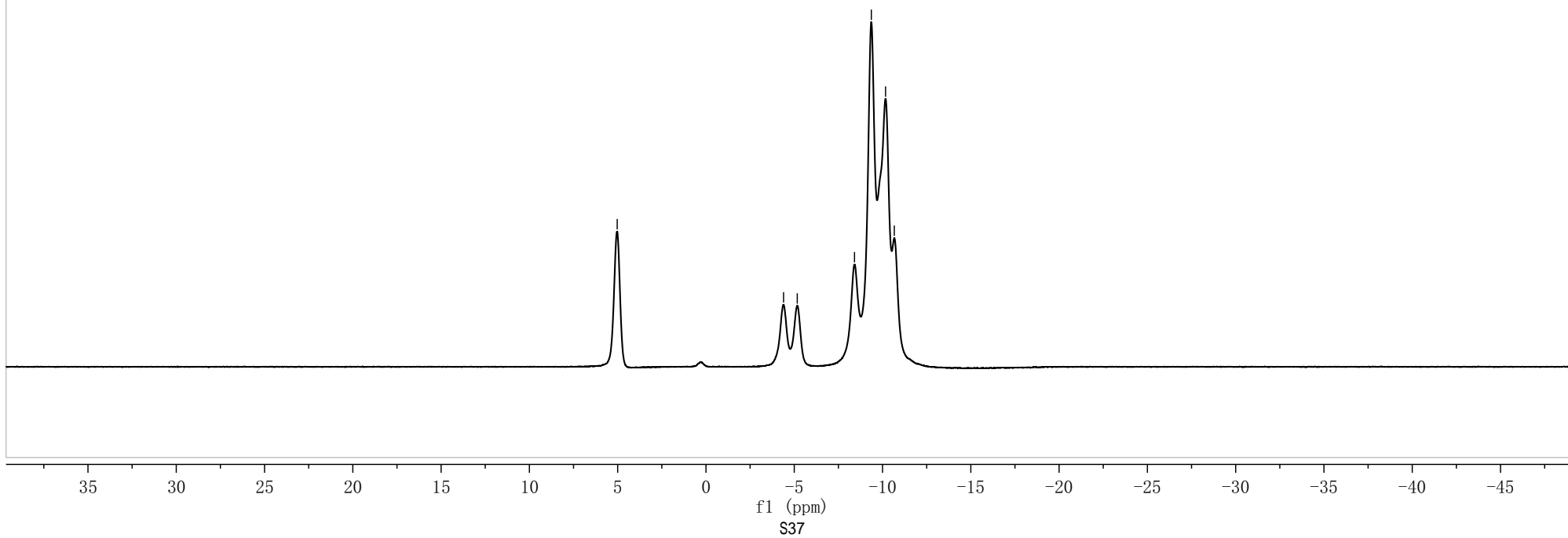


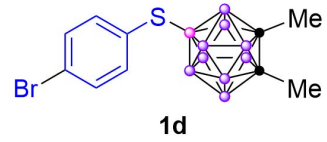
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





^{11}B NMR (193 MHz, CDCl_3)



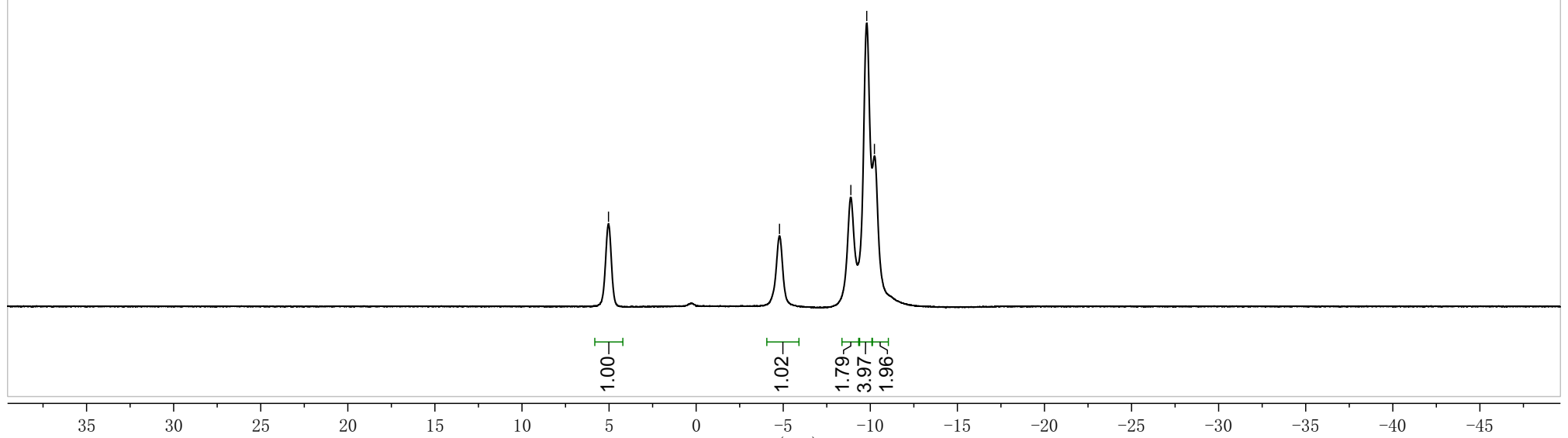


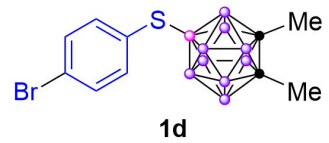
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

5.03
4.79
8.88
9.80
10.24

1.00
1.02
1.79
3.97
1.96

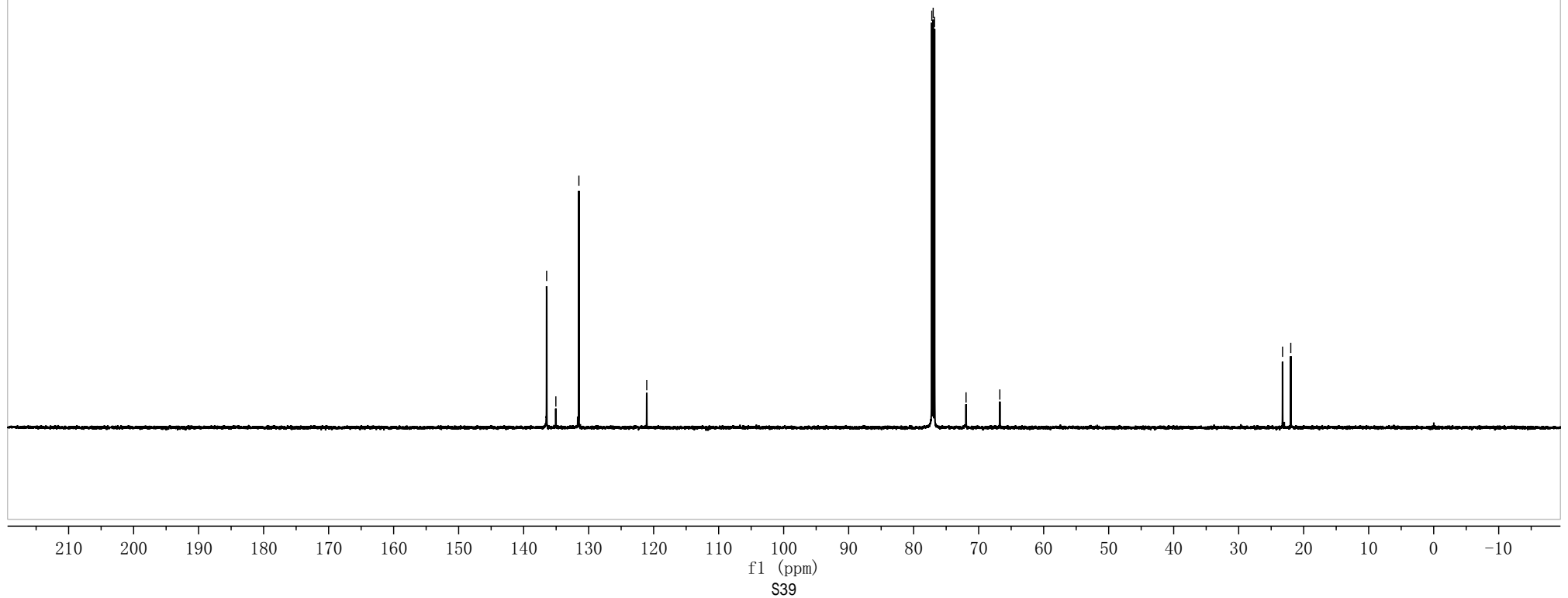
f1 (ppm)
S38

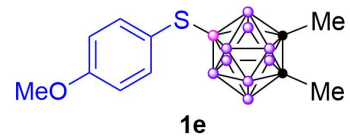




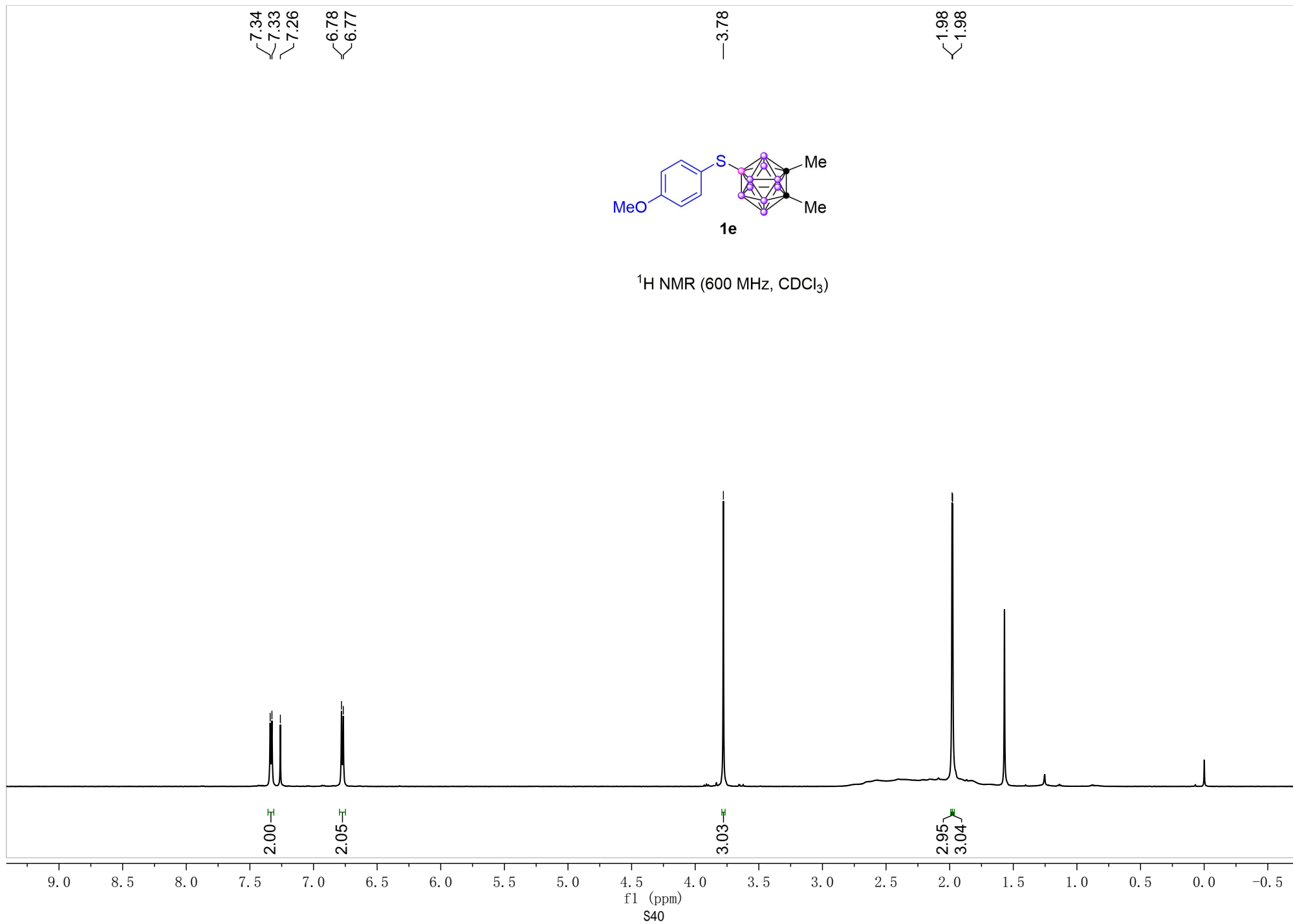
¹³C NMR (151 MHz, CDCl₃)

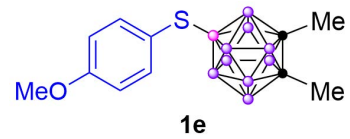
136.46
135.04
131.50
121.06
77.21
77.00
76.79
71.94
66.74
23.25
21.99



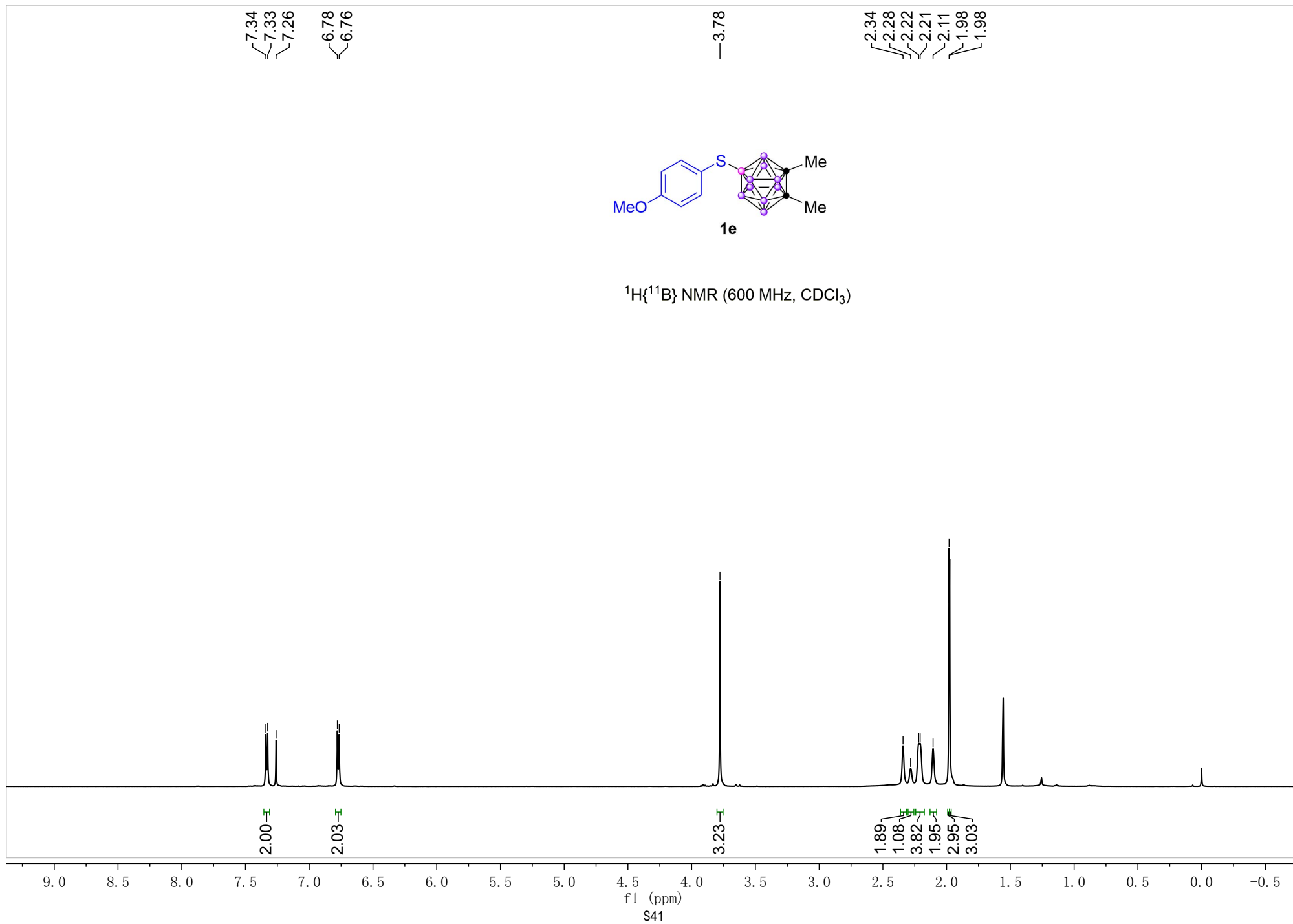


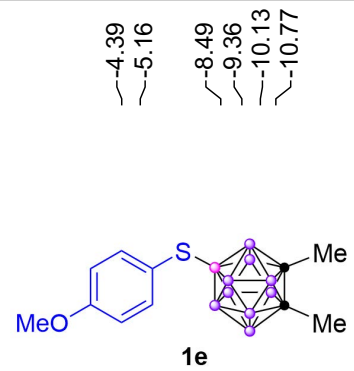
¹H NMR (600 MHz, CDCl₃)



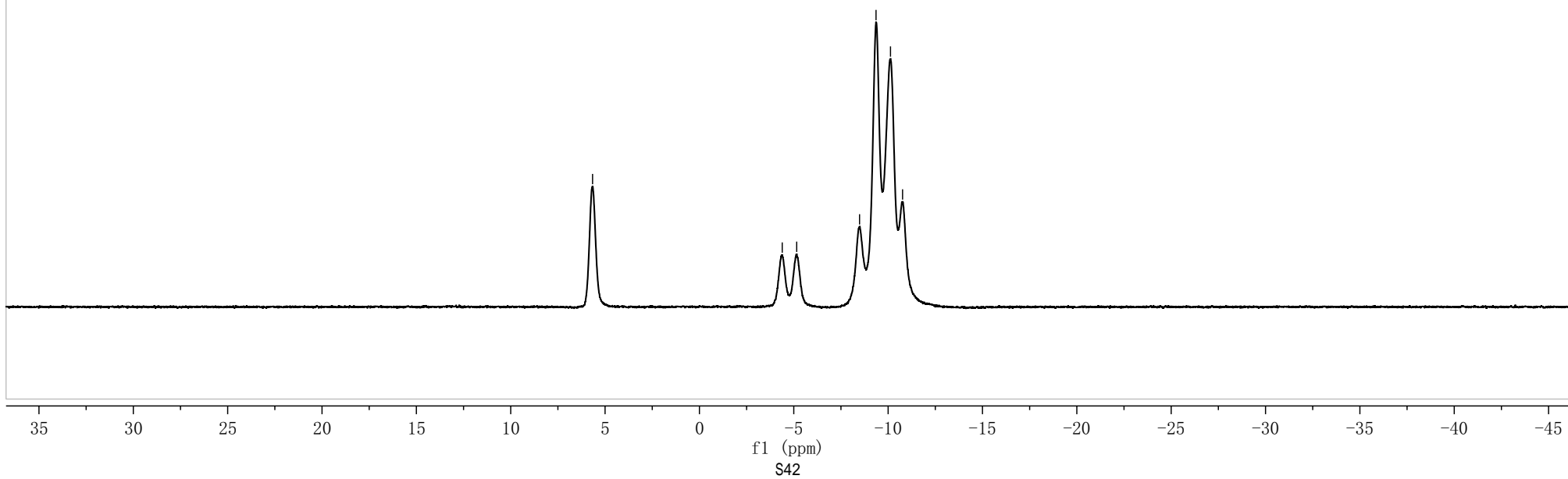


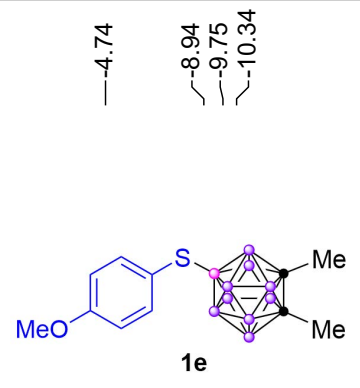
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)



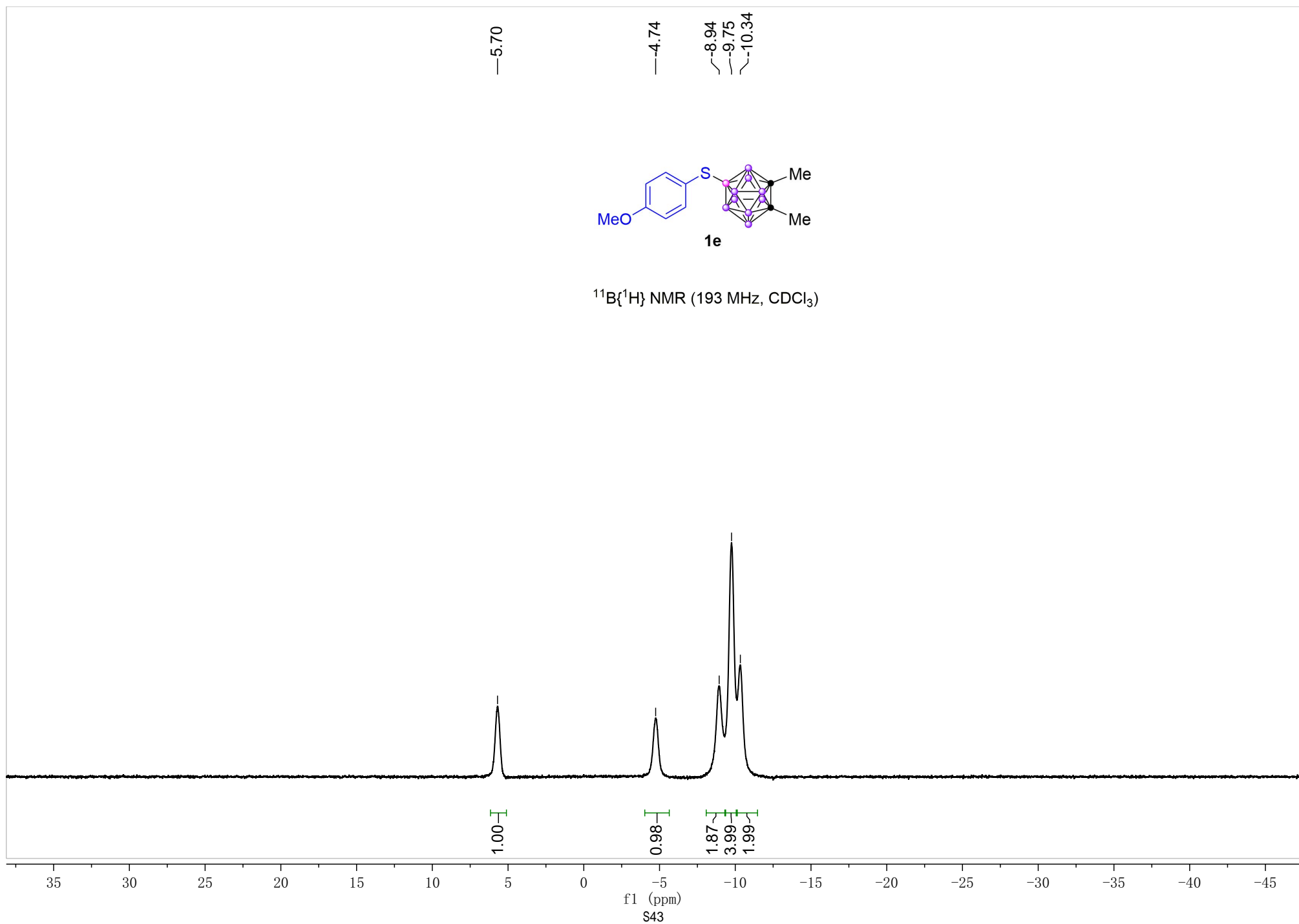


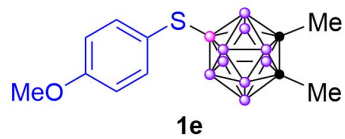
^{11}B NMR (193 MHz, CDCl_3)



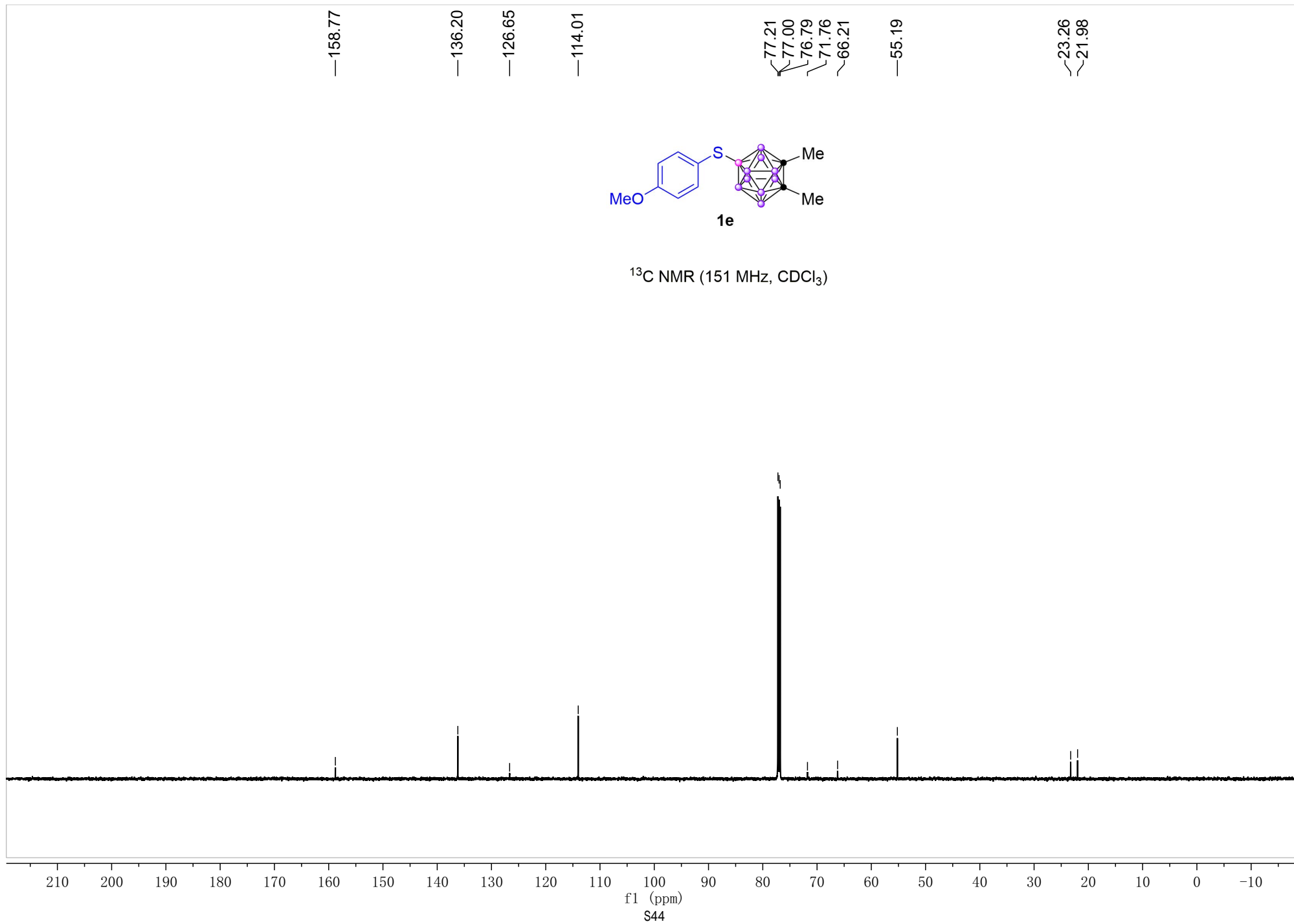


¹¹B{¹H} NMR (193 MHz, CDCl₃)



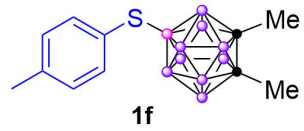


^{13}C NMR (151 MHz, CDCl_3)



7.32
7.31
7.26
7.05
7.03

2.31
1.98
1.98



¹H NMR (600 MHz, CDCl₃)

2.00

1.99

3.05

3.08

3.09

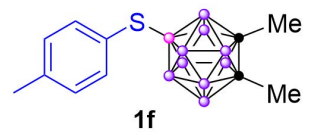
9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5

f1 (ppm)

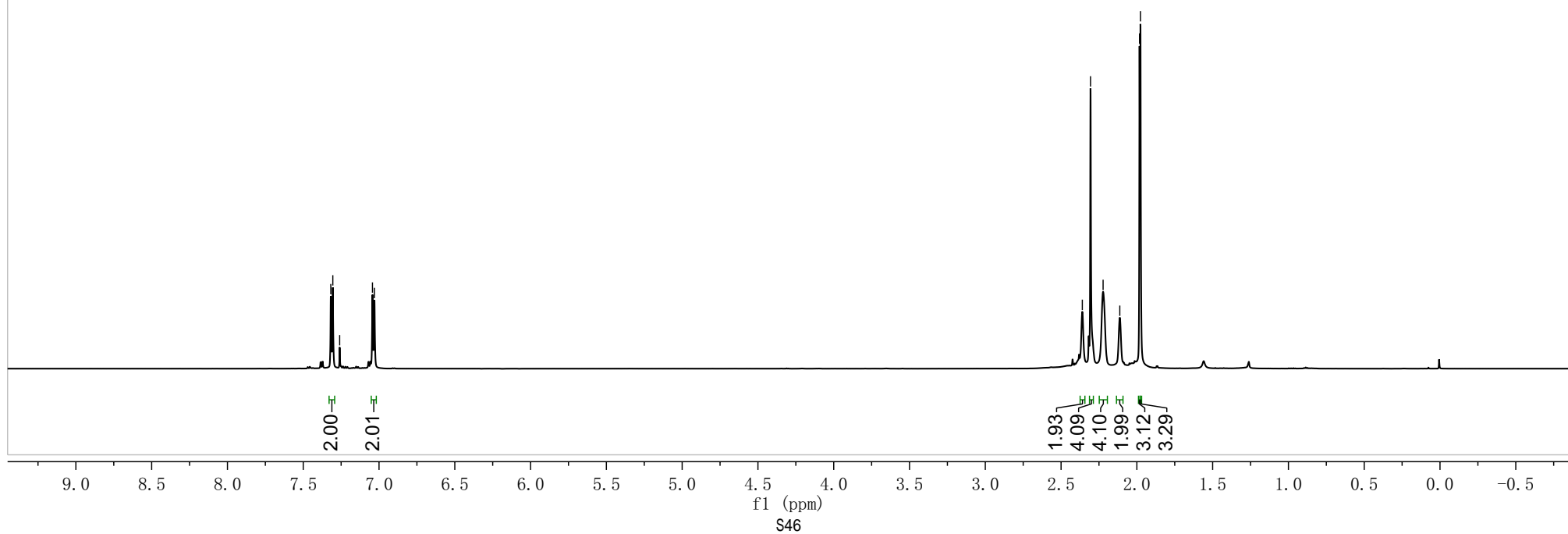
S45

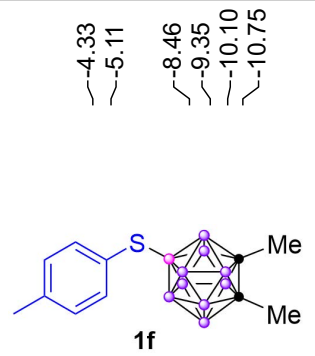
7.32
7.30
7.26
7.04
7.03

2.36
2.31
2.22
2.11
1.98
1.98

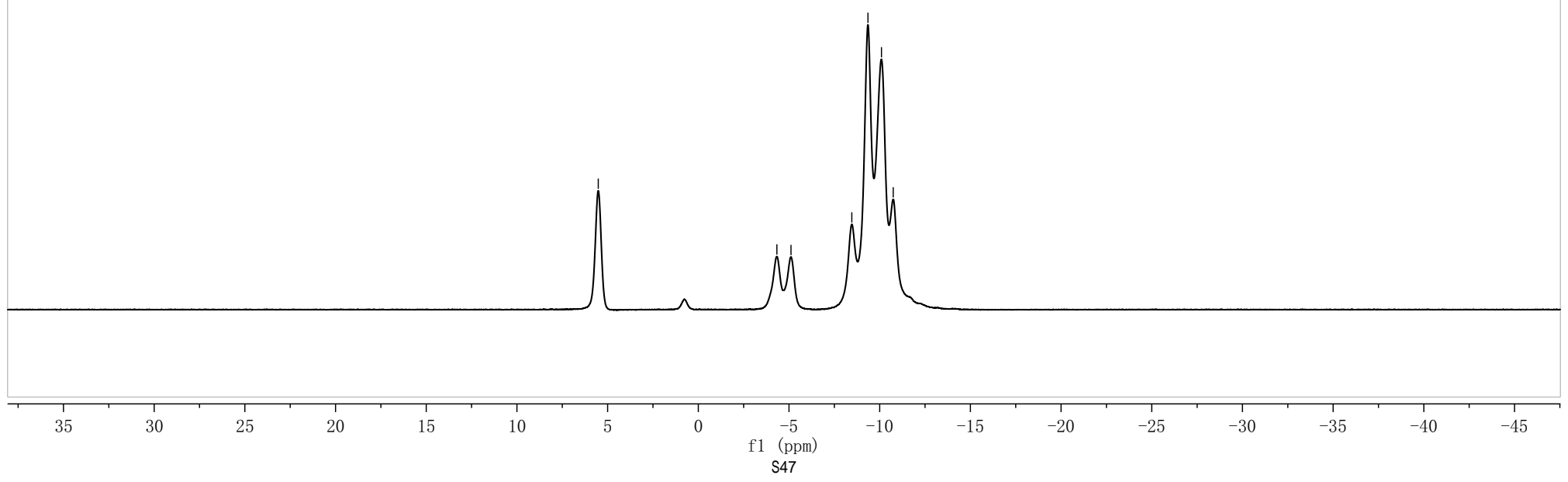


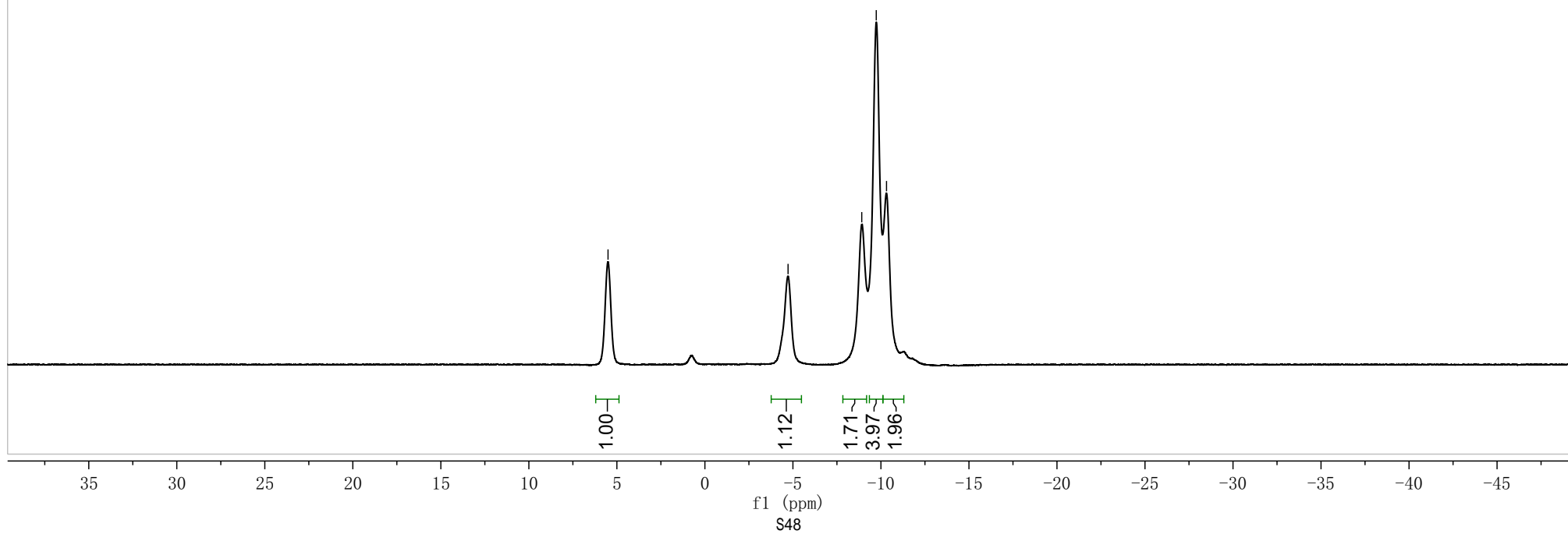
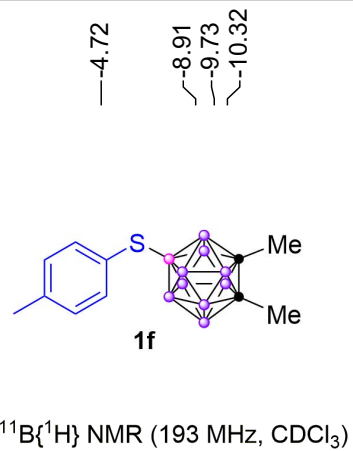
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





^{11}B NMR (193 MHz, CDCl_3)

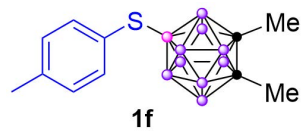




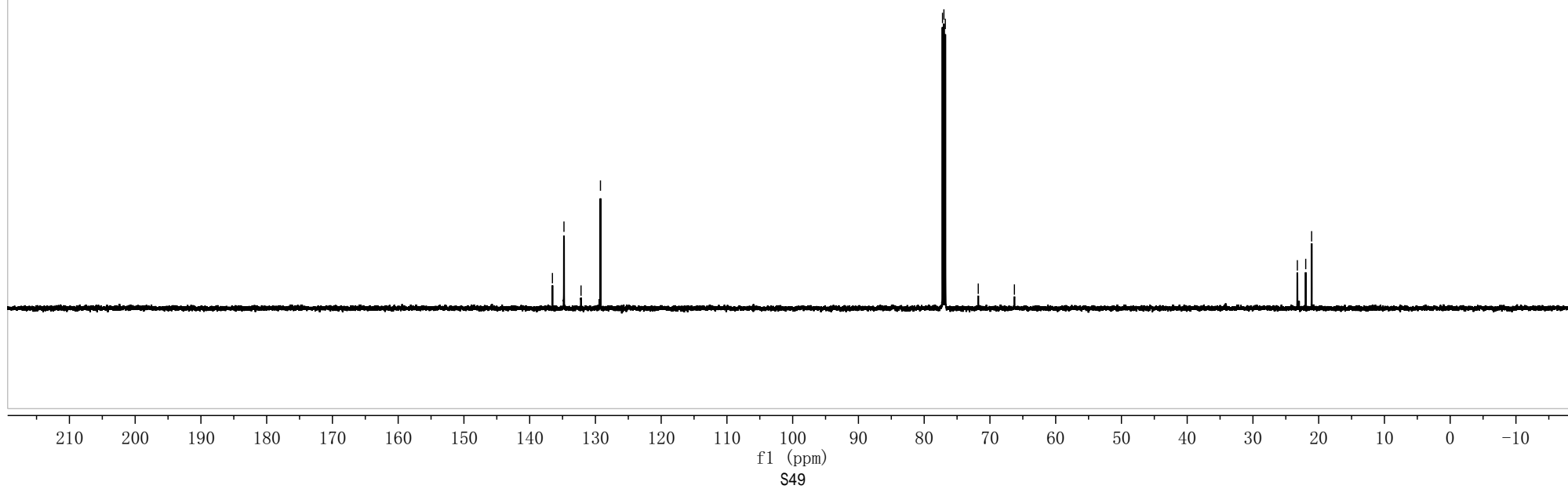
136.54
134.78
132.19
129.24

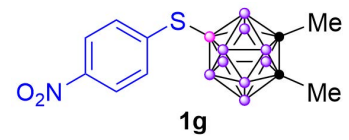
77.21
77.00
76.79
71.78
66.28

23.24
21.97
21.07

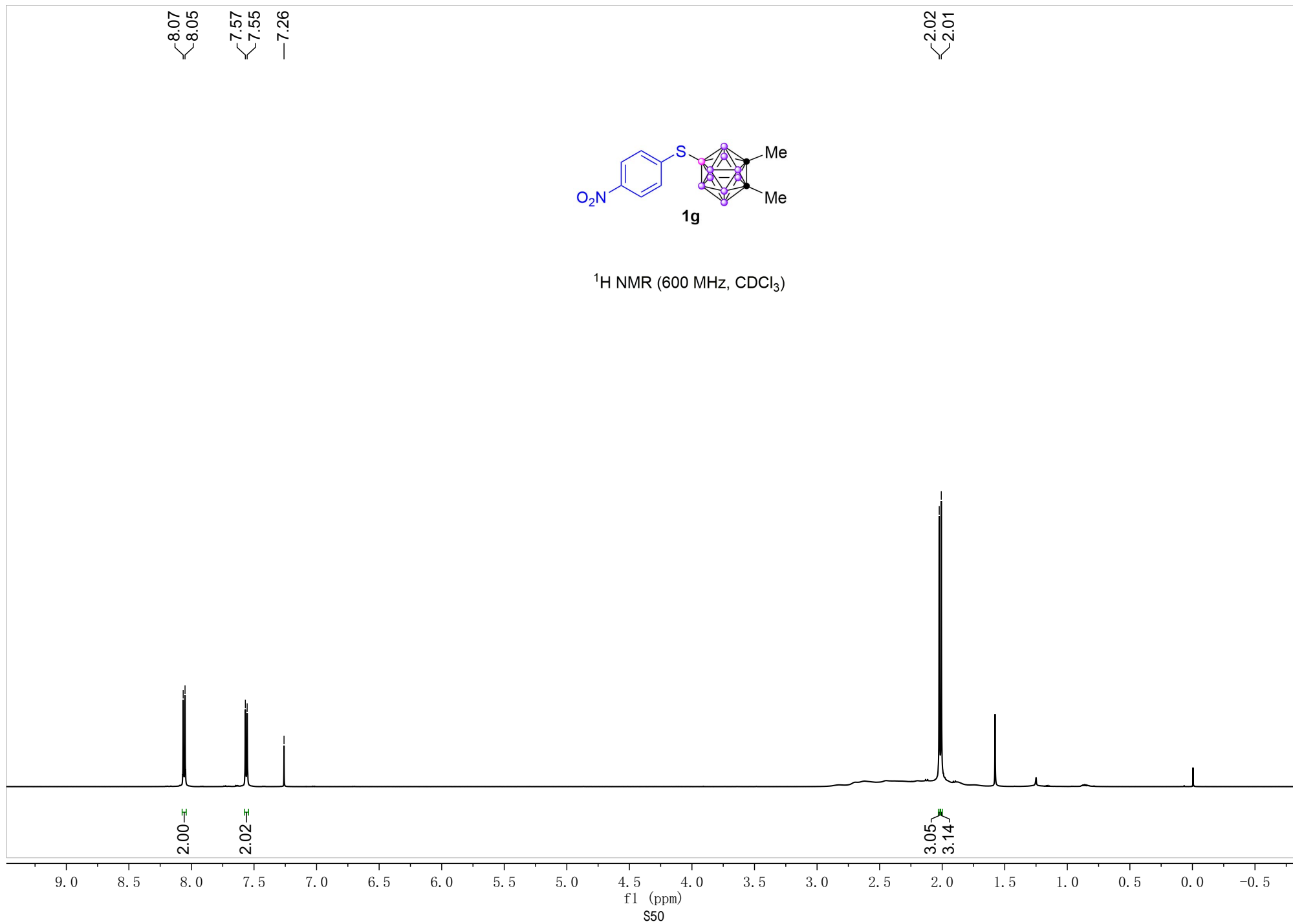


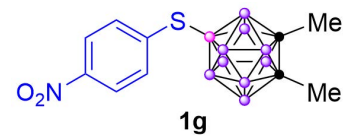
^{13}C NMR (151 MHz, CDCl_3)



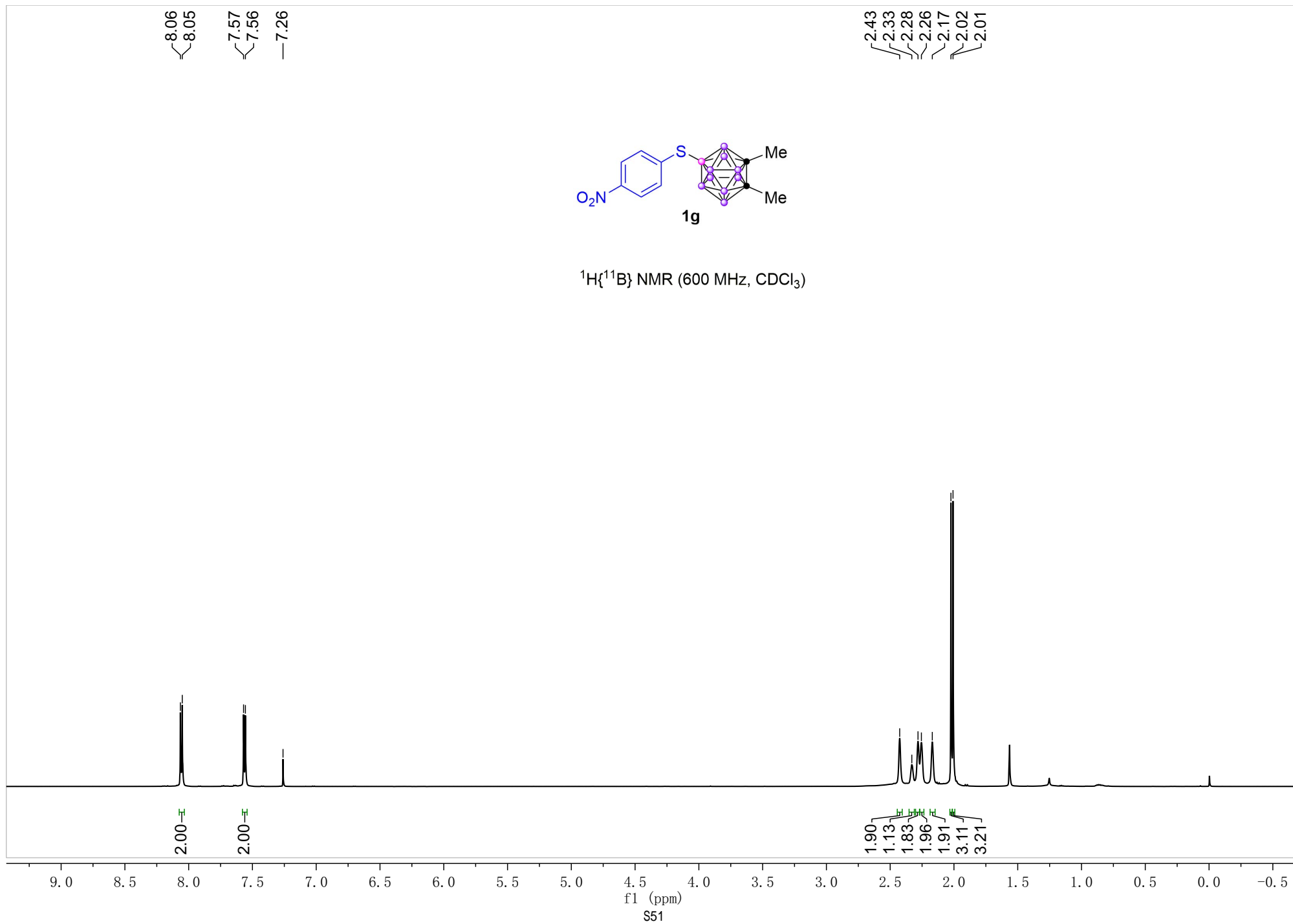


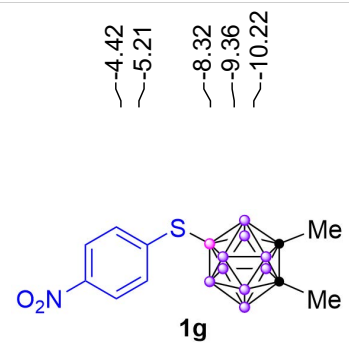
^1H NMR (600 MHz, CDCl_3)



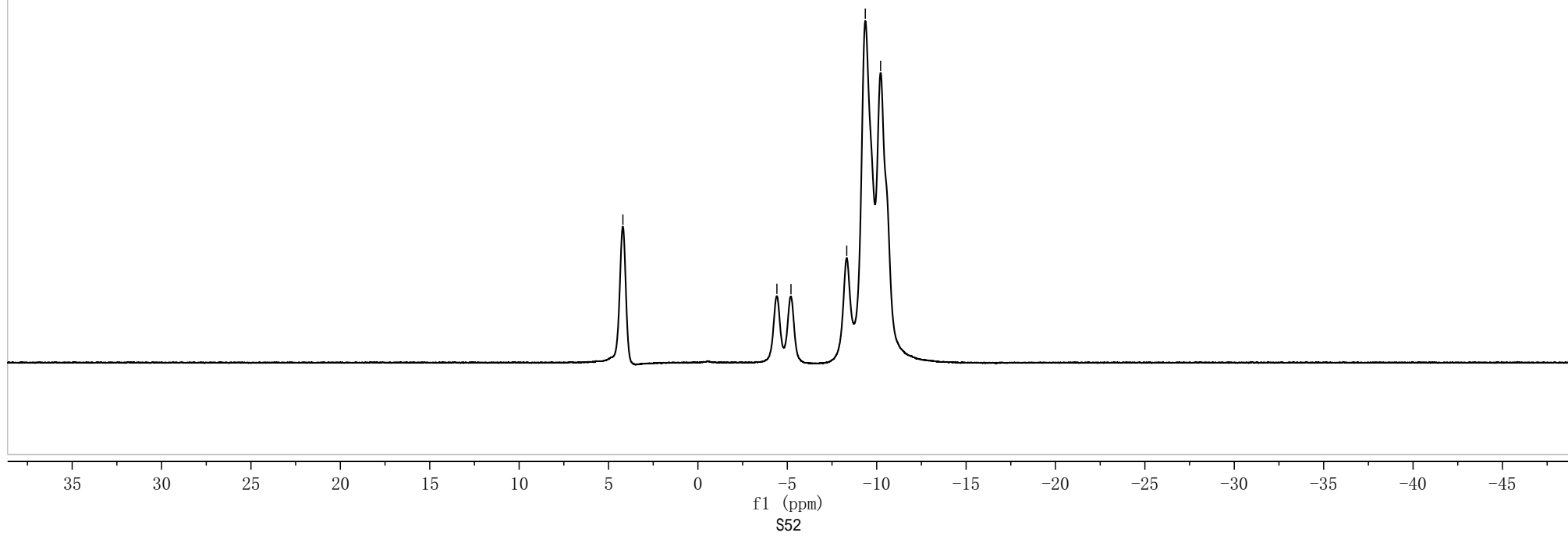


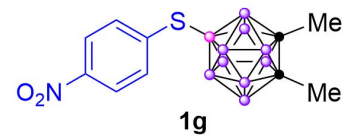
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





^{11}B NMR (193 MHz, CDCl_3)



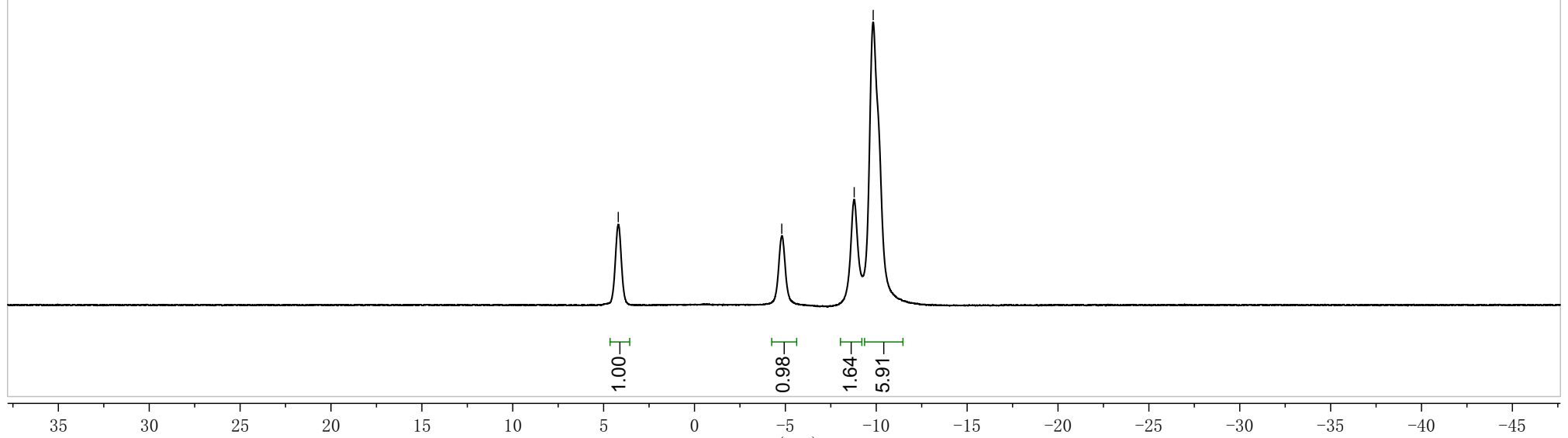


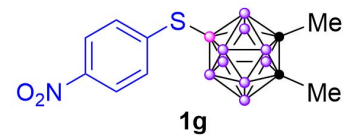
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

—4.19
—4.80
—8.79
—9.83

1.00
0.98
1.64
5.91

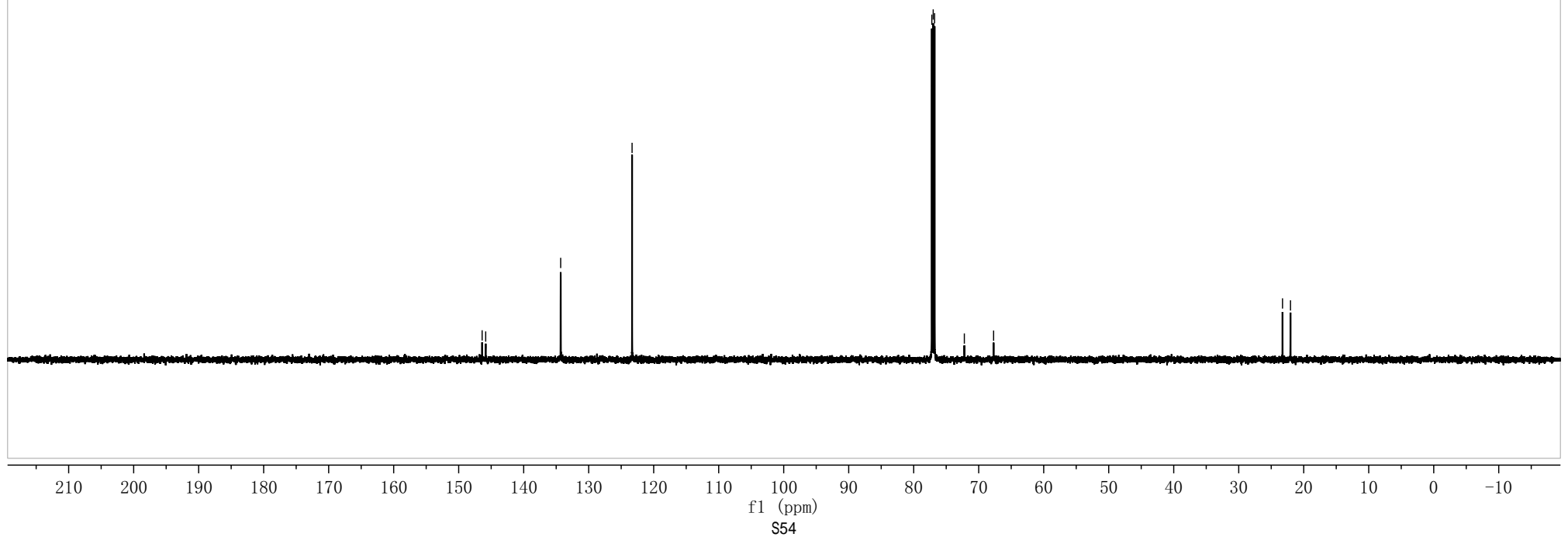
f1 (ppm)
S53





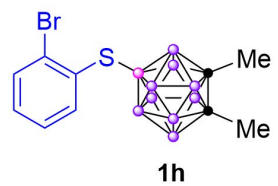
¹³C NMR (151 MHz, CDCl₃)

146.39
145.84
134.31
123.32
77.21
77.00
76.79
72.21
67.71
23.26
22.03

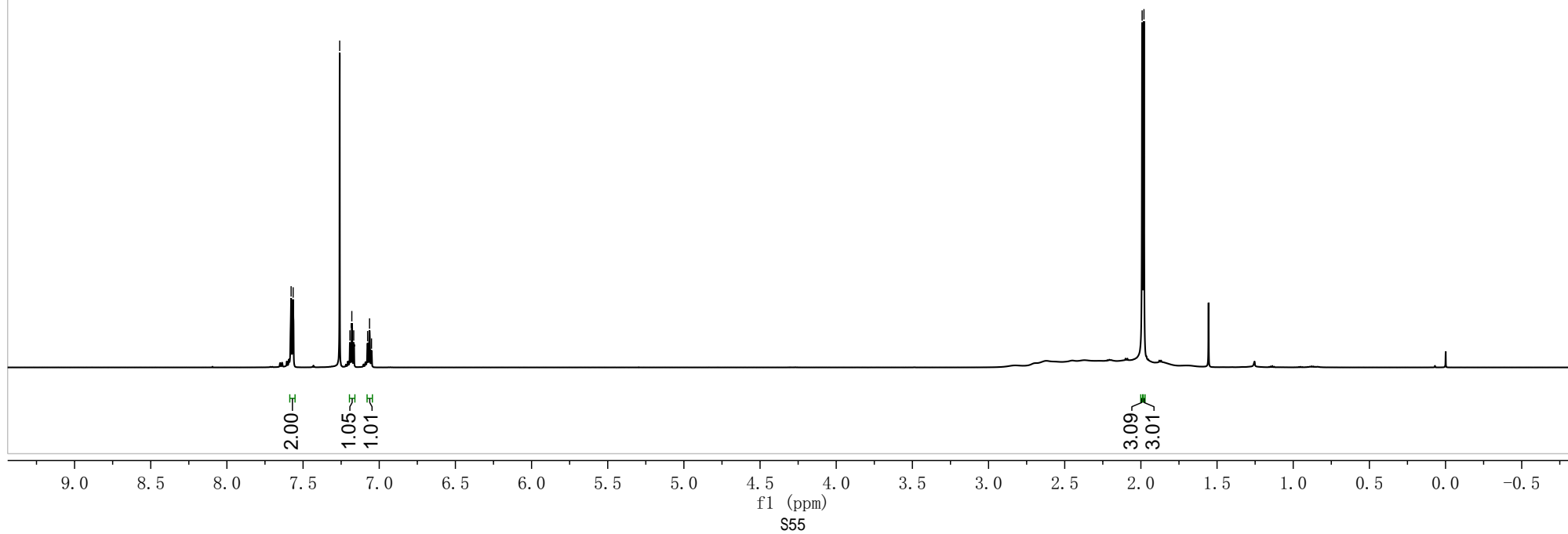


7.58
7.57
7.26
7.19
7.18
7.17
7.08
7.06
7.05

1.99
1.98

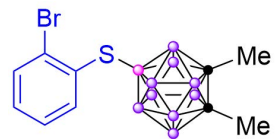


$^1\text{H NMR}$ (600 MHz, CDCl_3)



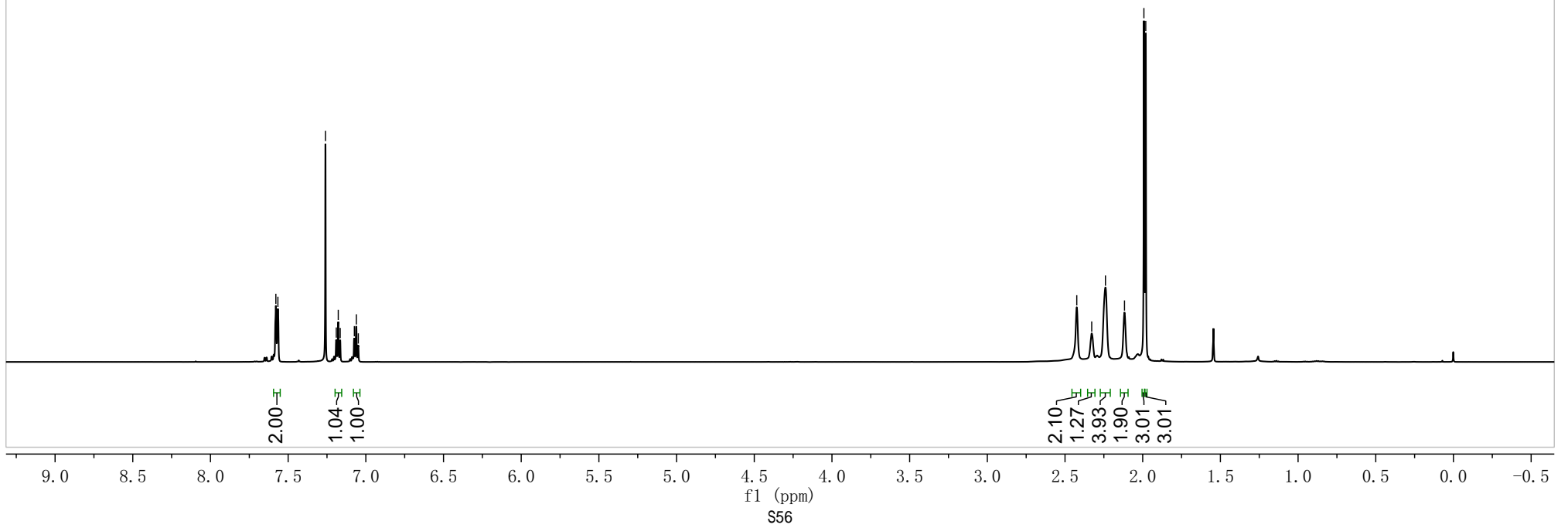
7.58
7.57
7.26
7.19
7.18
7.16
7.07
7.06
7.05

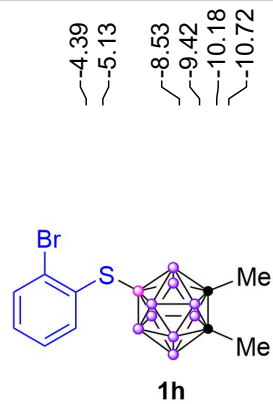
2.42
2.33
2.24
2.12
1.99
1.98



1h

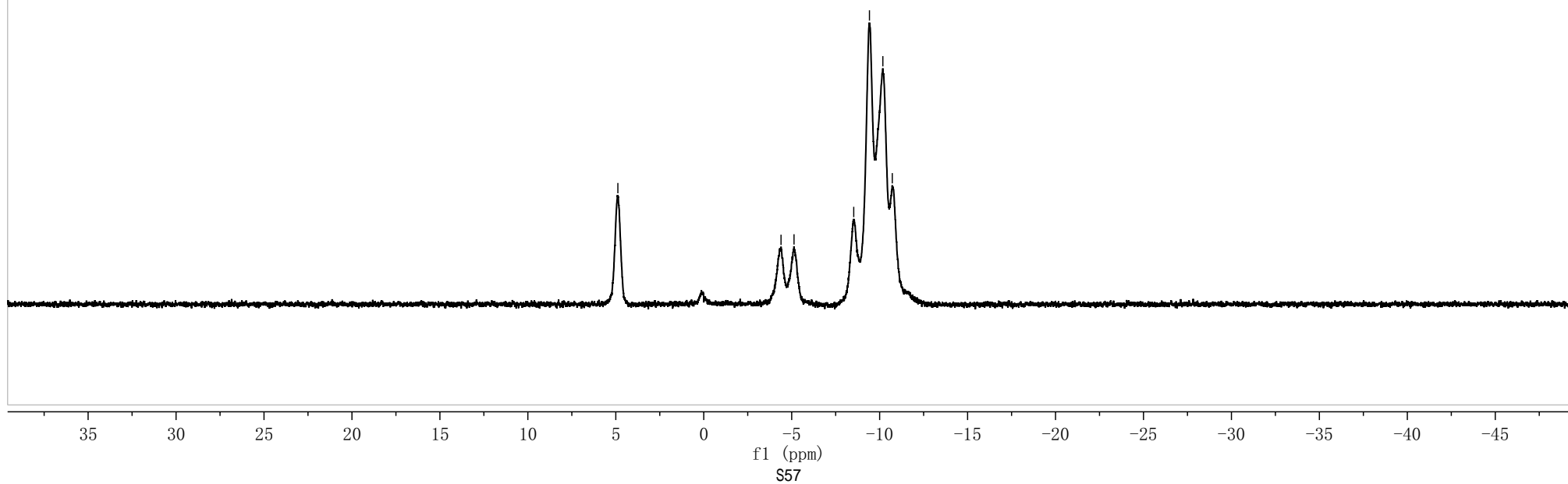
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)

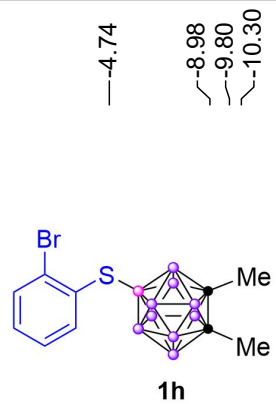




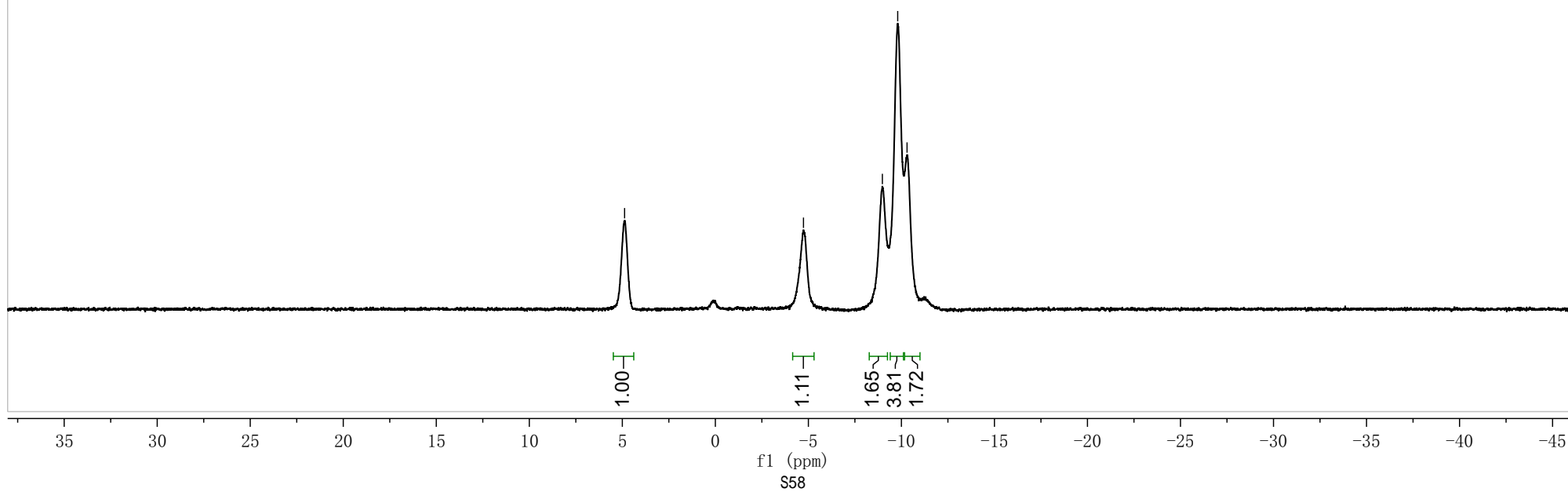
¹¹B NMR (193 MHz, CDCl₃)

Chemical shift values (ppm):
-4.88
-4.39
-5.13
-8.53
-9.42
-10.18
-10.72





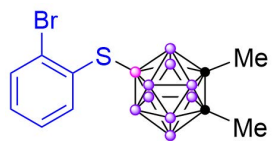
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)



137.63
137.16
132.85
130.27
128.42
127.23

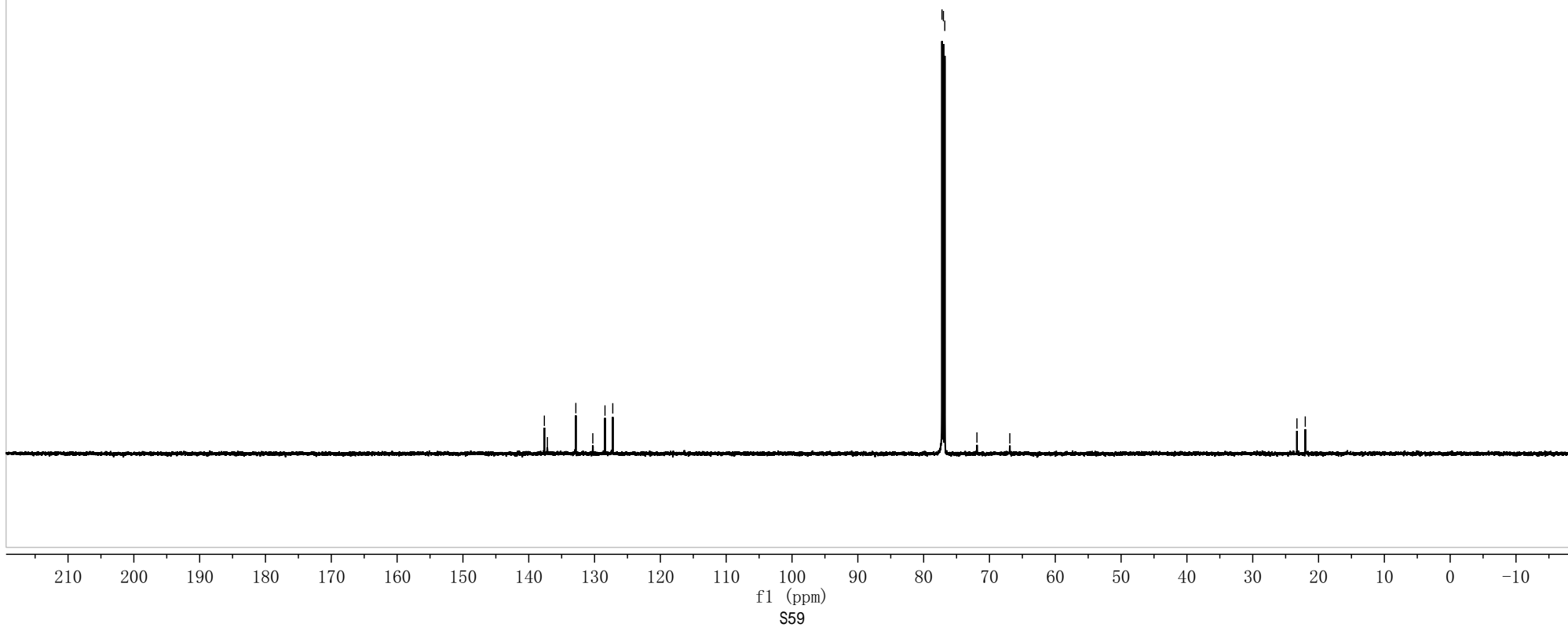
77.21
77.00
76.79
71.90
66.91

23.28
22.02



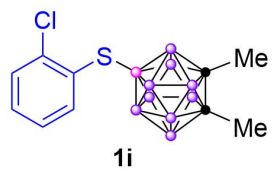
1h

¹³C NMR (151 MHz, CDCl₃)

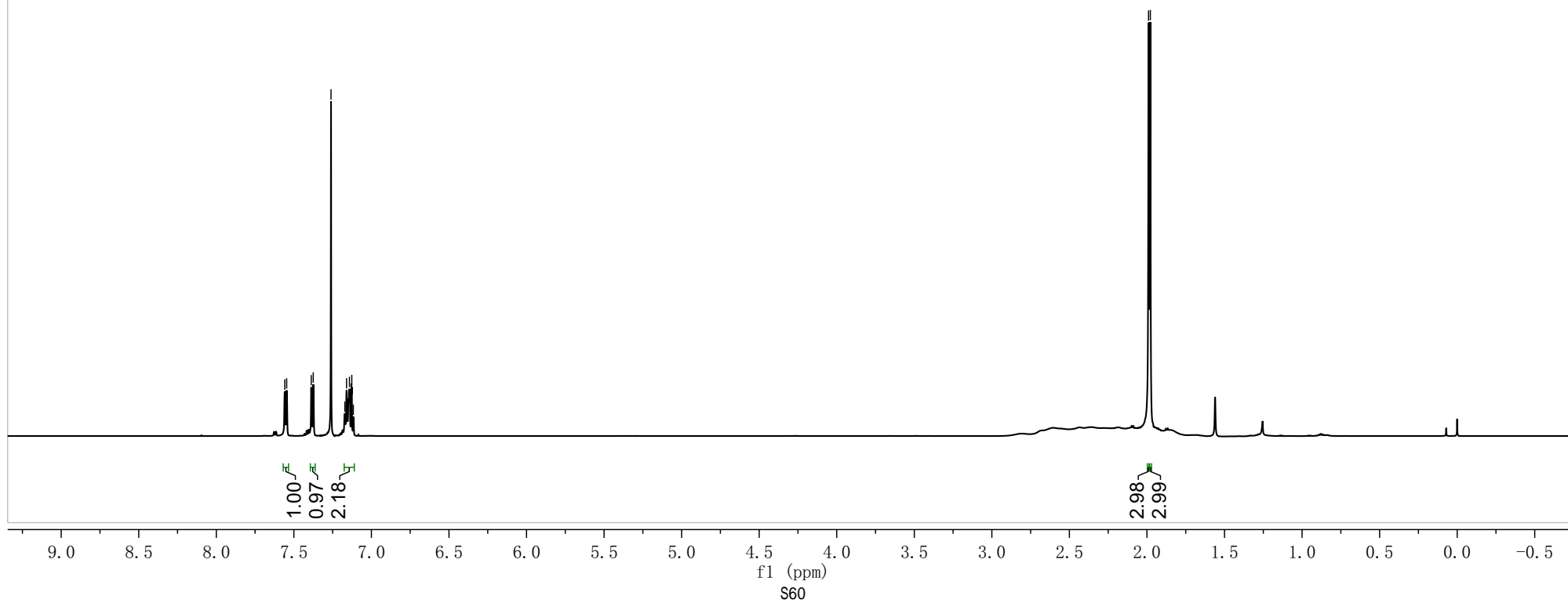


7.56
7.55
7.39
7.38
7.26
7.17
7.16
7.14
7.13
7.12

1.99
1.98

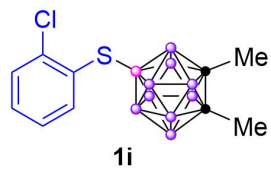


$^1\text{H NMR}$ (600 MHz, CDCl_3)

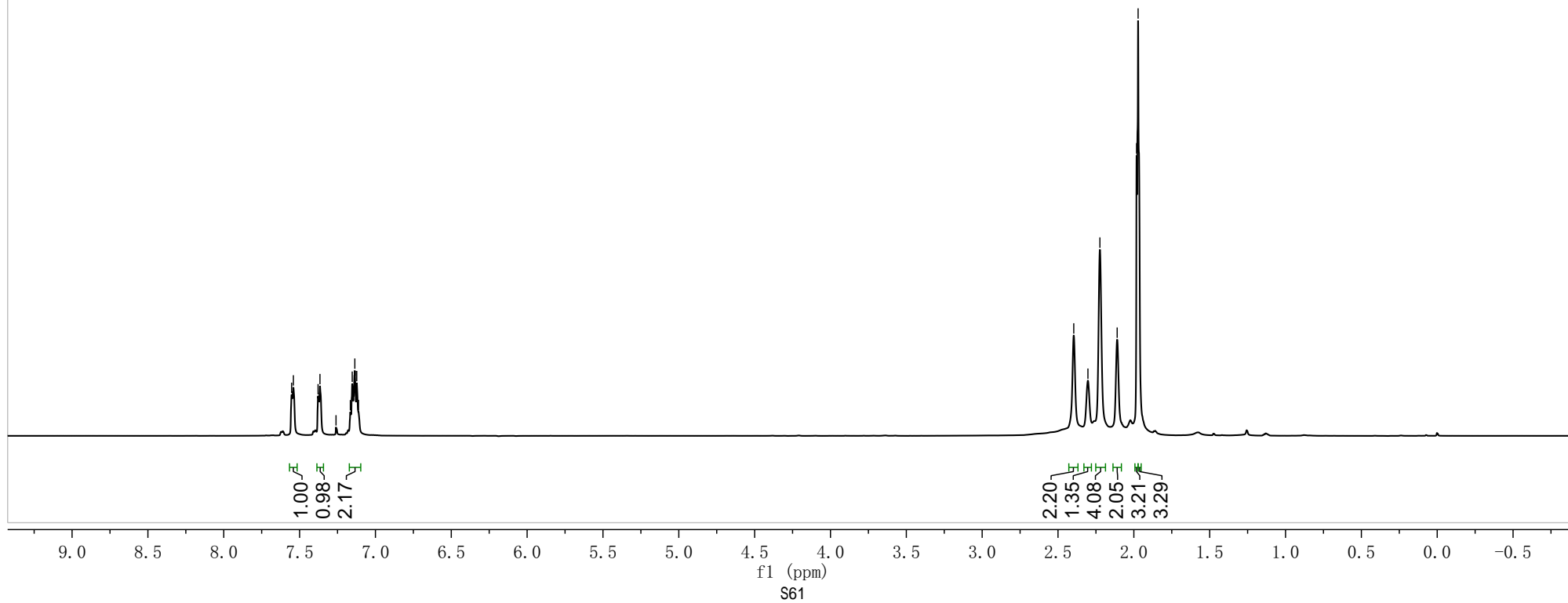


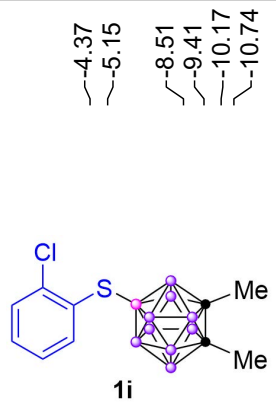
7.55
7.54
7.38
7.37
7.26
7.17
7.15
7.14
7.12
7.11

2.40
2.30
2.22
2.11
1.98
1.97



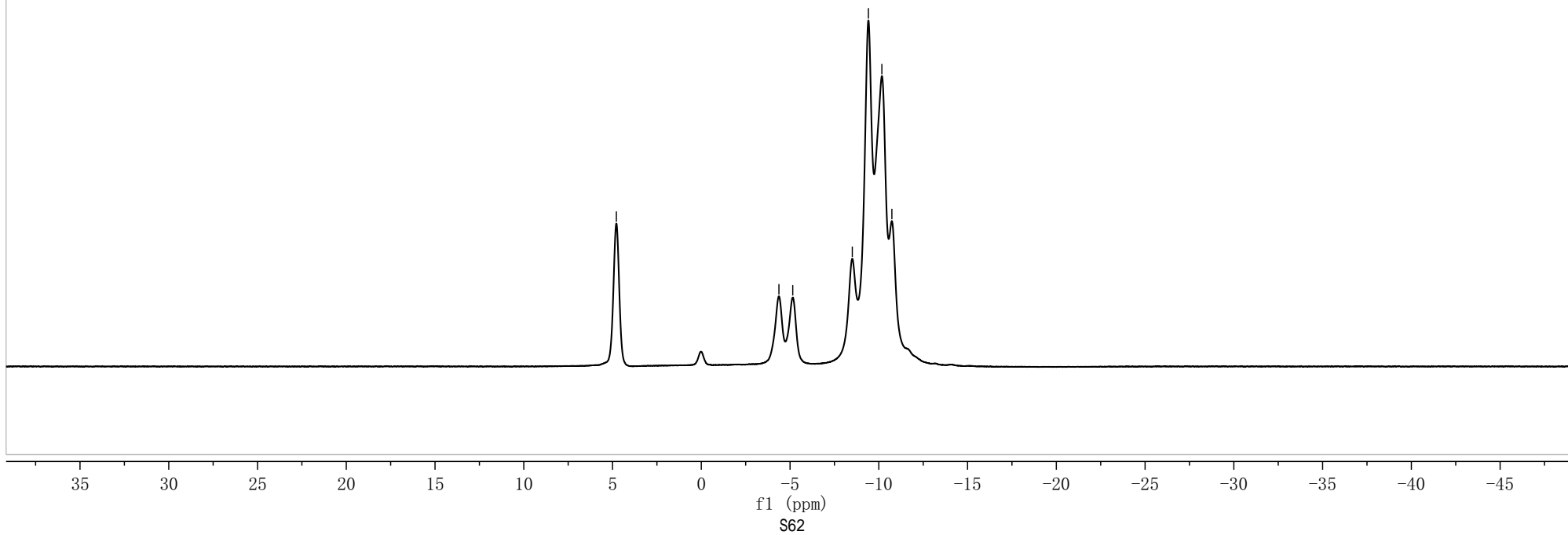
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)

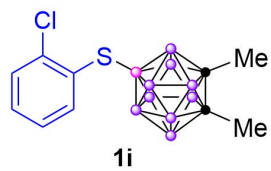




¹¹B NMR (193 MHz, CDCl₃)

Chemical shift values (ppm):
-4.79
-4.37
-5.15
-8.51
-9.41
-10.17
-10.74





$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

—4.78
—4.77
—8.98
—9.82
—10.30

1.00
1.13
1.58
3.58
1.81

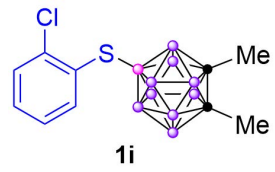
f1 (ppm)
S63

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45

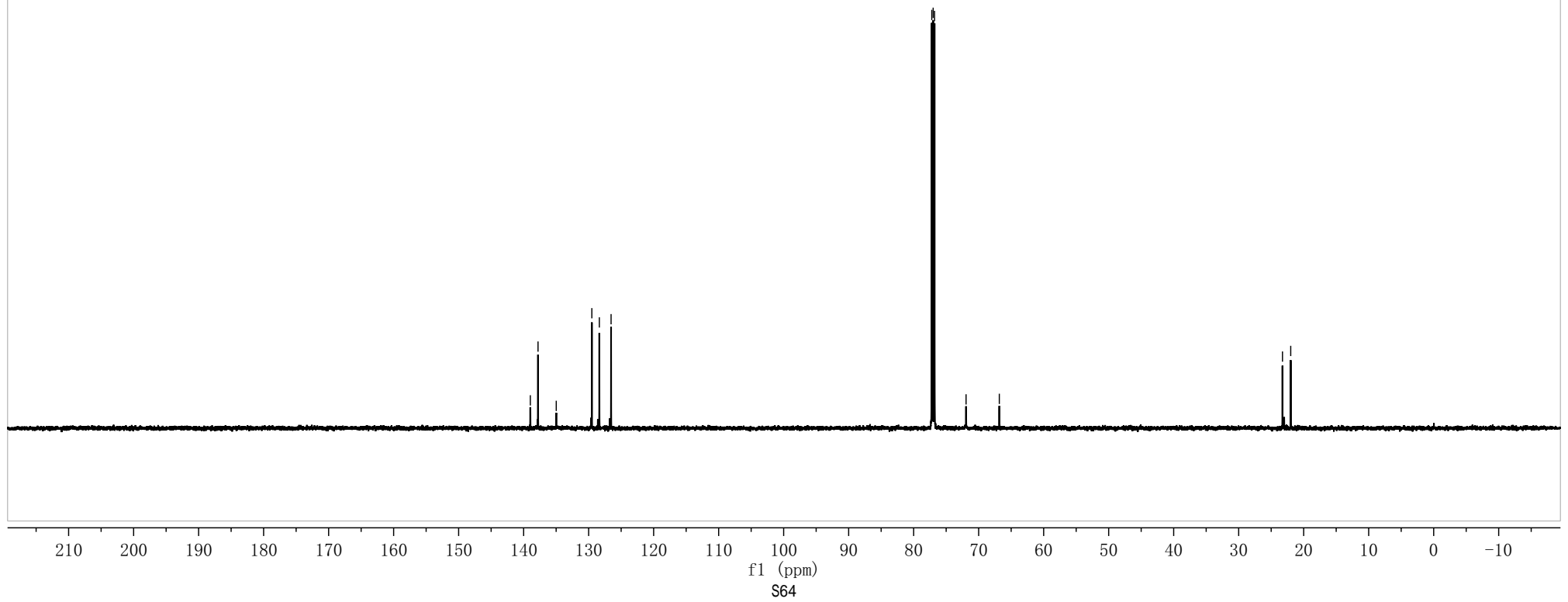
138.97
137.79
134.98
129.51
128.35
126.55

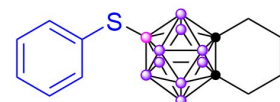
77.21
77.00
76.79
71.94
66.82

23.26
21.99



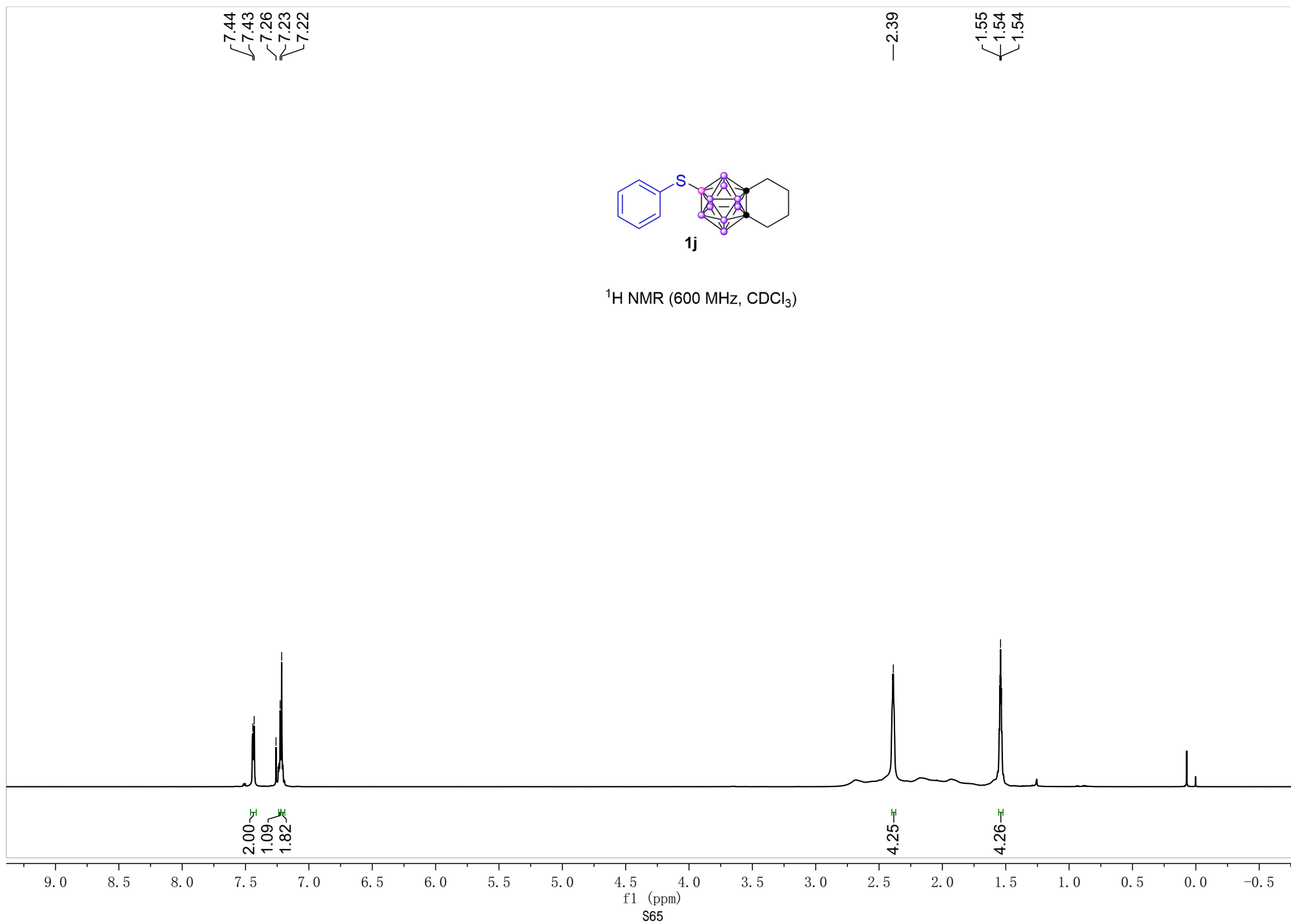
^{13}C NMR (151 MHz, CDCl_3)

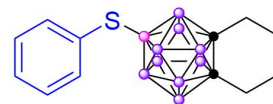




1j

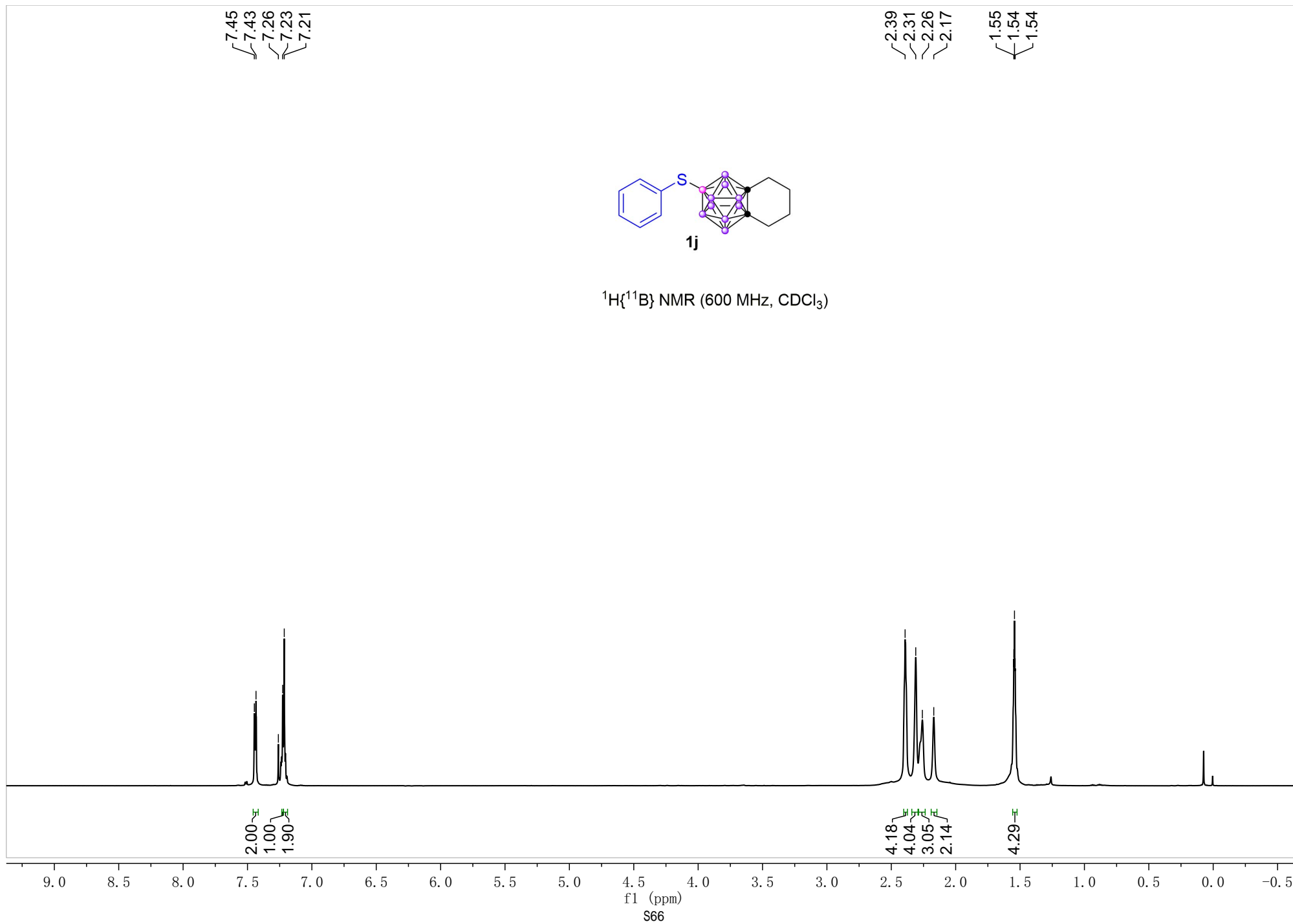
^1H NMR (600 MHz, CDCl_3)

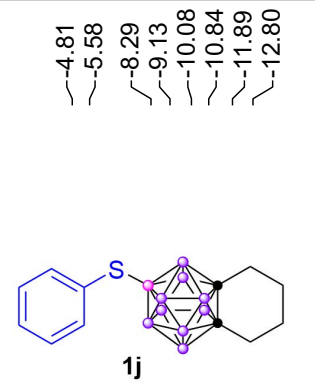




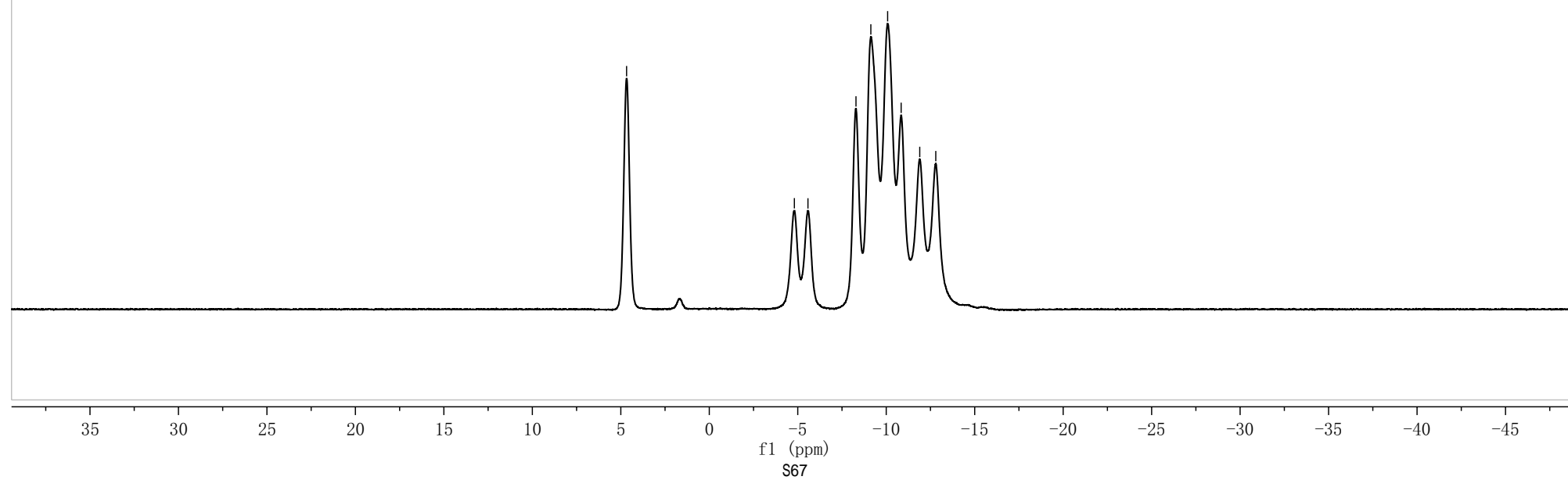
1j

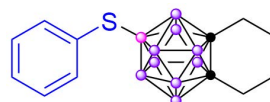
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)





^{11}B NMR (193 MHz, CDCl_3)





1j

$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

—4.67

—5.17

—8.68

—9.82

—10.42

—12.33

1.00

1.03

1.90

1.83

1.78

1.75

35

30

25

20

15

10

5

0

f1 (ppm)

S68

-5

-10

-15

-20

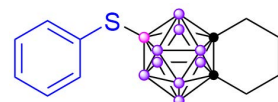
-25

-30

-35

-40

-45



1j

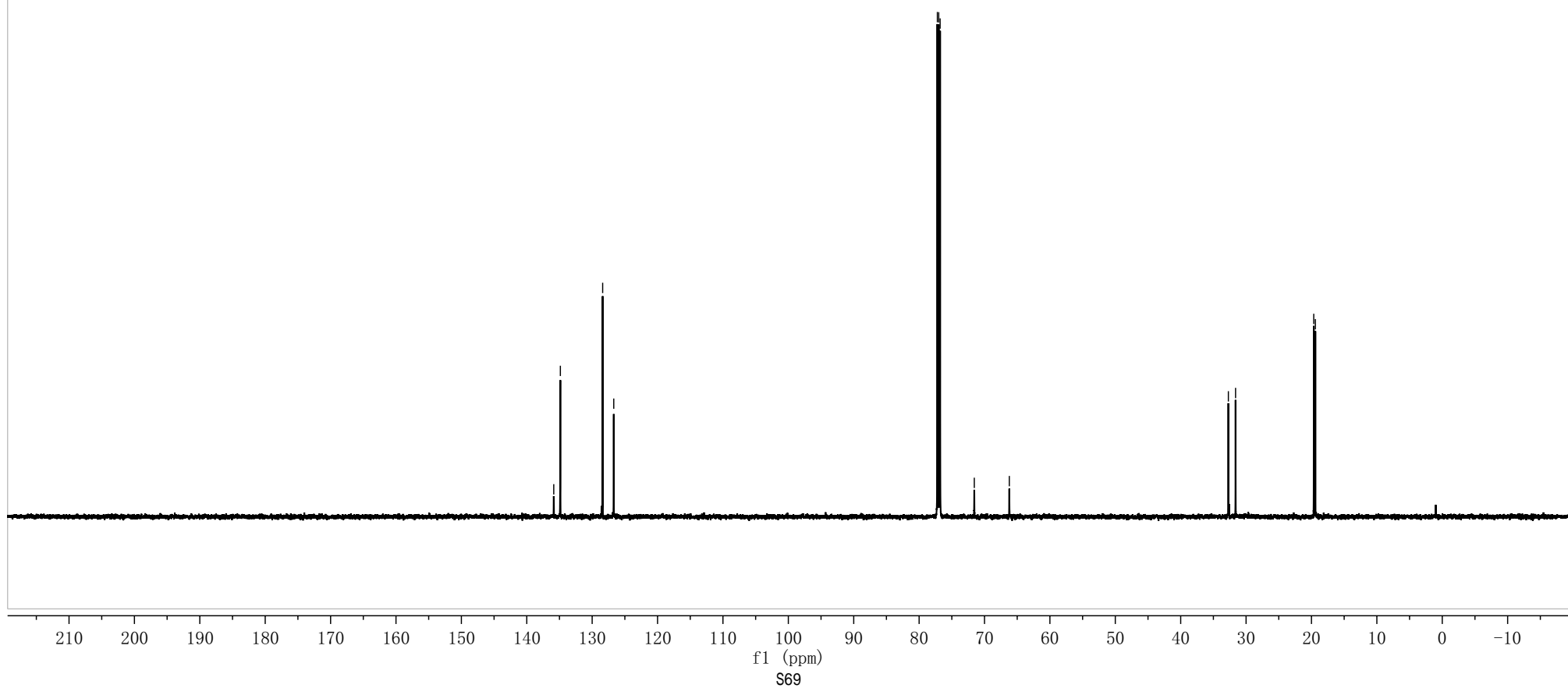
¹³C NMR (151 MHz, CDCl₃)

135.87
134.86
128.39
126.70

77.21
77.00
76.79
71.56
66.21

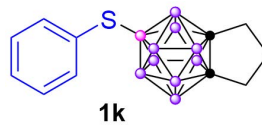
32.71
31.61

19.65
19.40

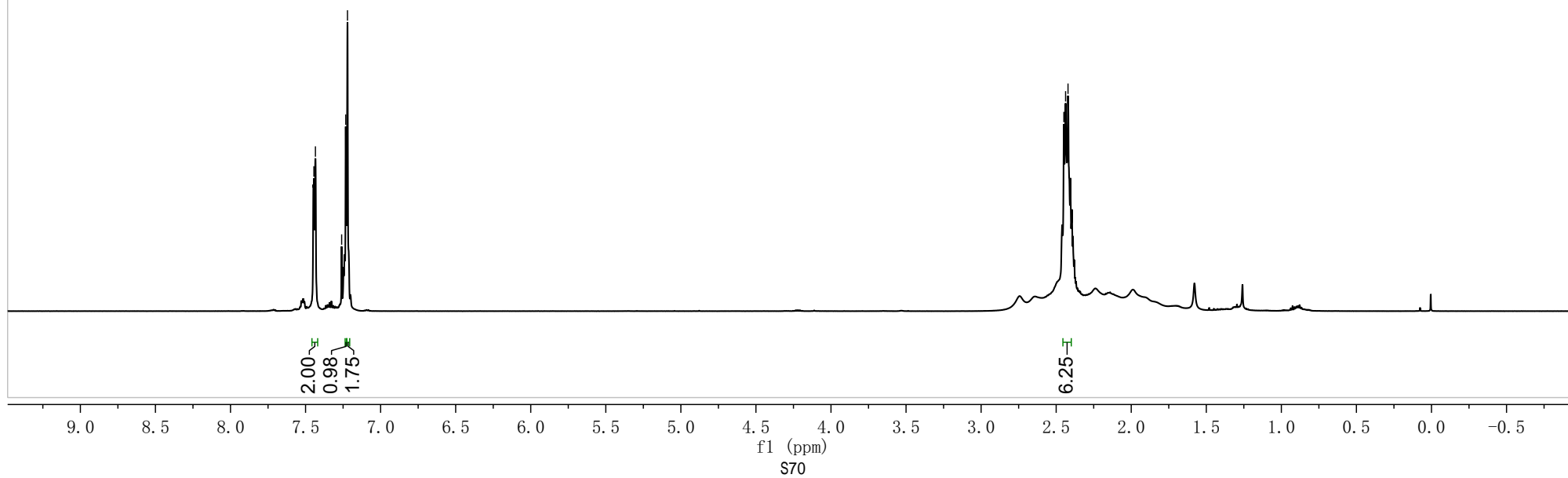


7.44
7.43
7.26
7.23
7.22

2.45
2.44
2.42

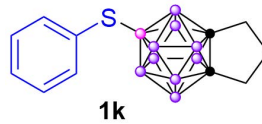


^1H NMR (600 MHz, CDCl_3)

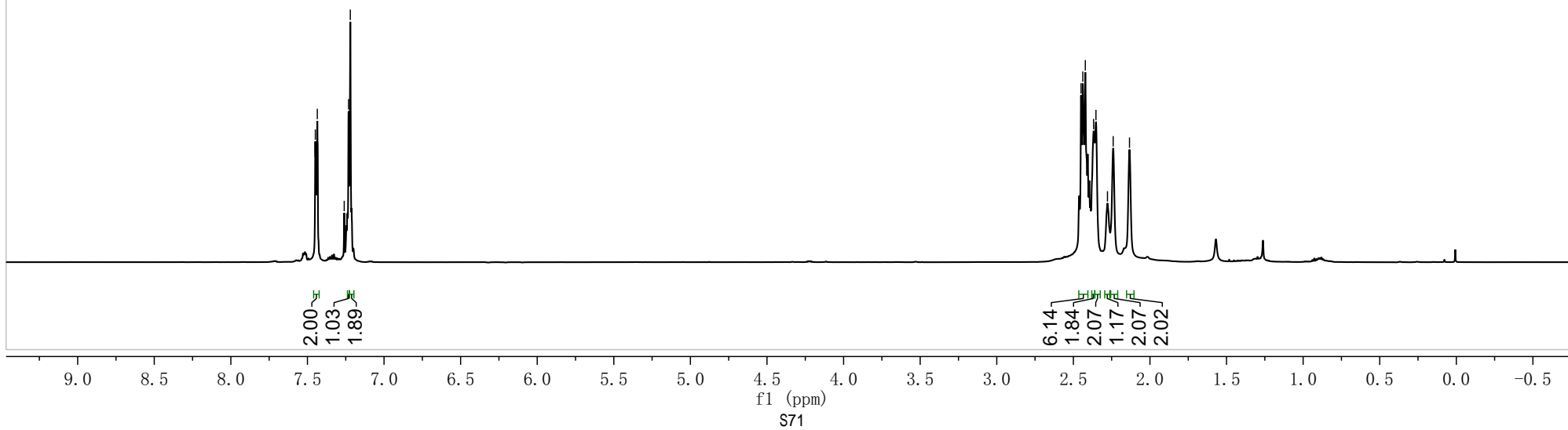


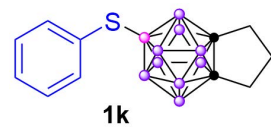
7.45
7.44
7.26
7.23
7.22

2.45
2.44
2.42
2.37
2.35
2.28
2.24
2.13



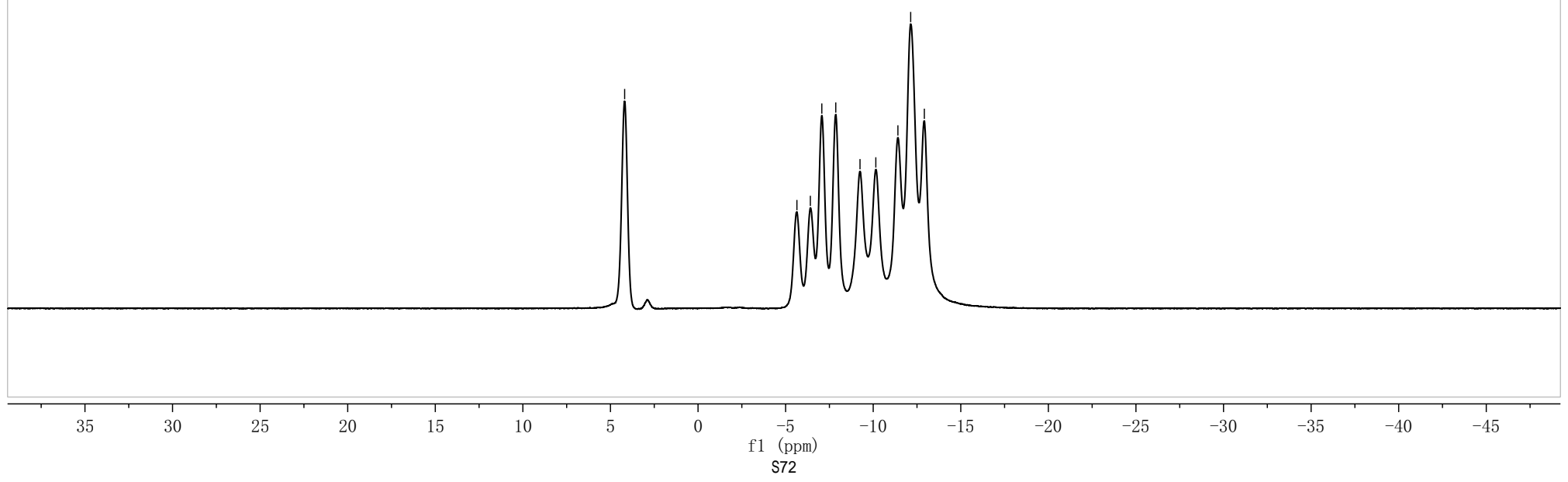
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)

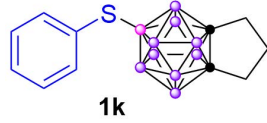




^{11}B NMR (193 MHz, CDCl_3)

—4.19
—5.65
—6.41
—7.07
—7.86
—9.25
—10.15
—11.41
—12.14
—12.92





$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

—4.19

-6.03
-7.47
-9.70
-11.84
-12.49

1.00

1.01

1.86

1.76

1.79

1.90

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45

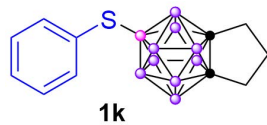
f1 (ppm)

S73

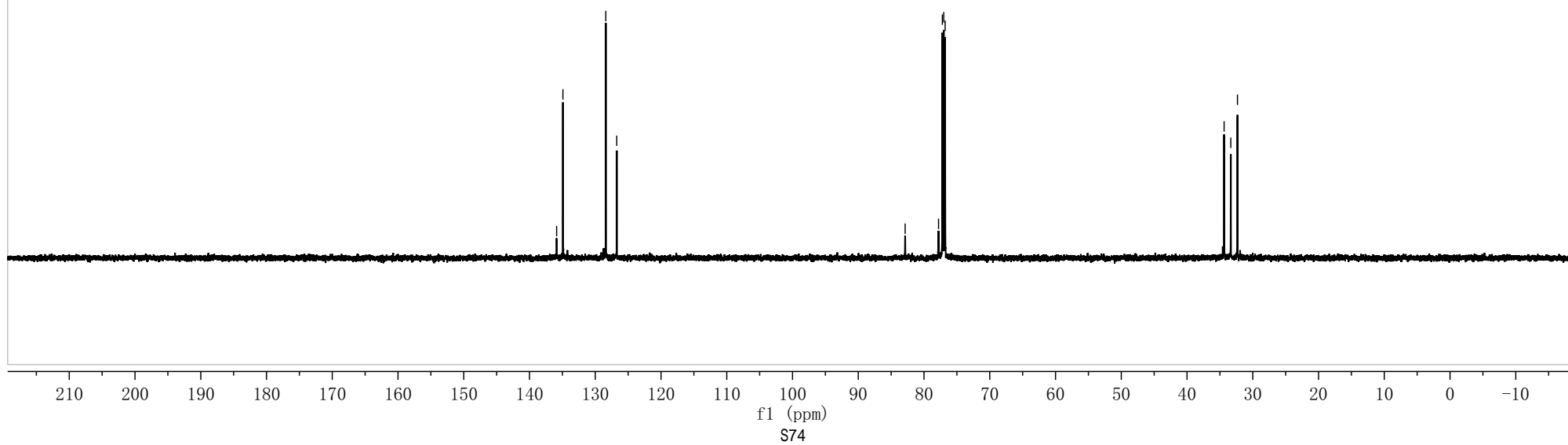
135.87
134.92
128.40
126.73

82.87
77.79
77.21
77.00
76.79

34.36
33.34
32.33

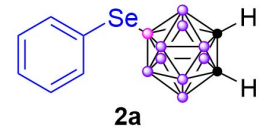


^{13}C NMR (151 MHz, CDCl_3)

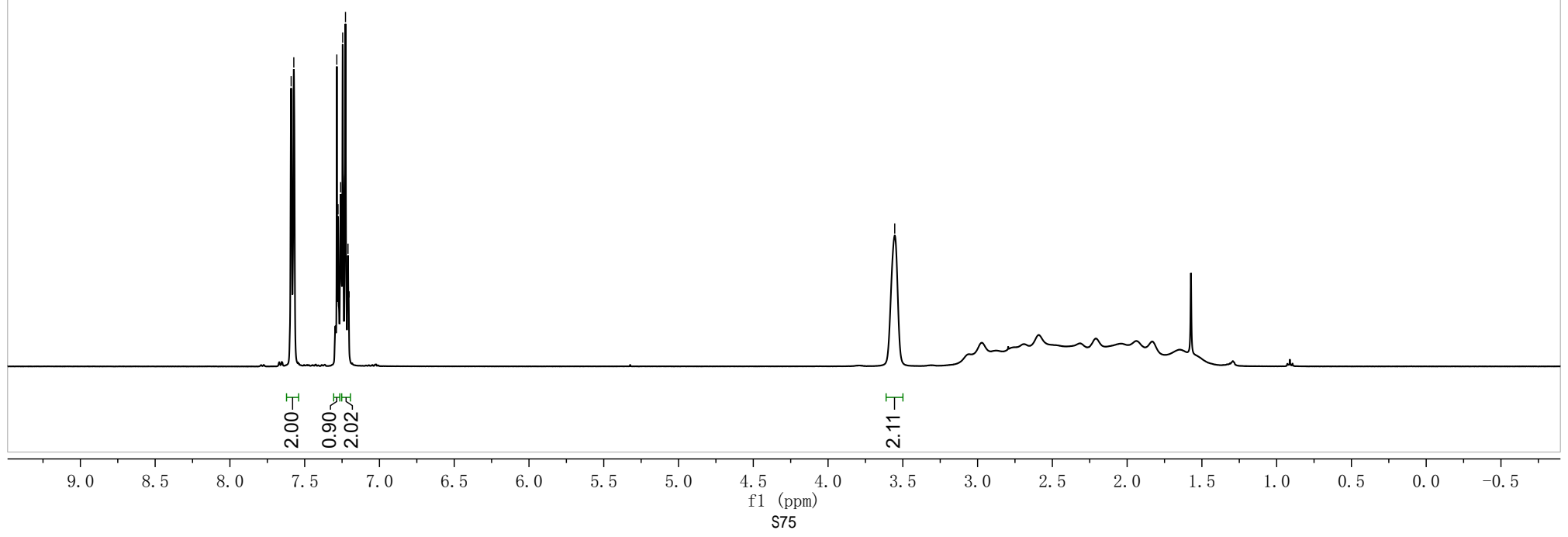


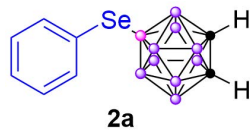
7.59
7.57
7.29
7.28
7.26
7.25
7.23
7.21

3.55

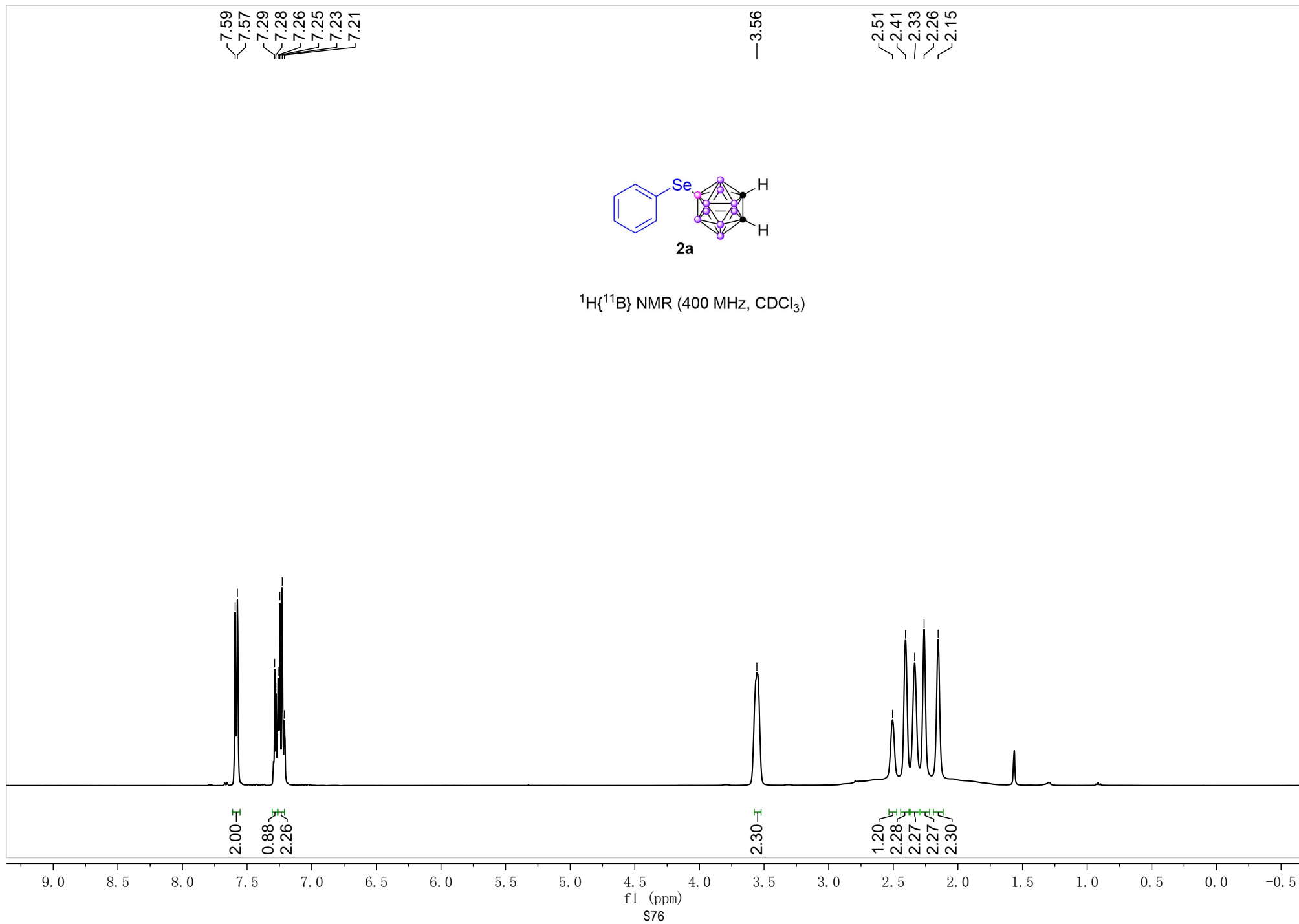


¹H NMR (400 MHz, CDCl₃)

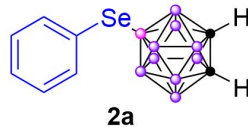




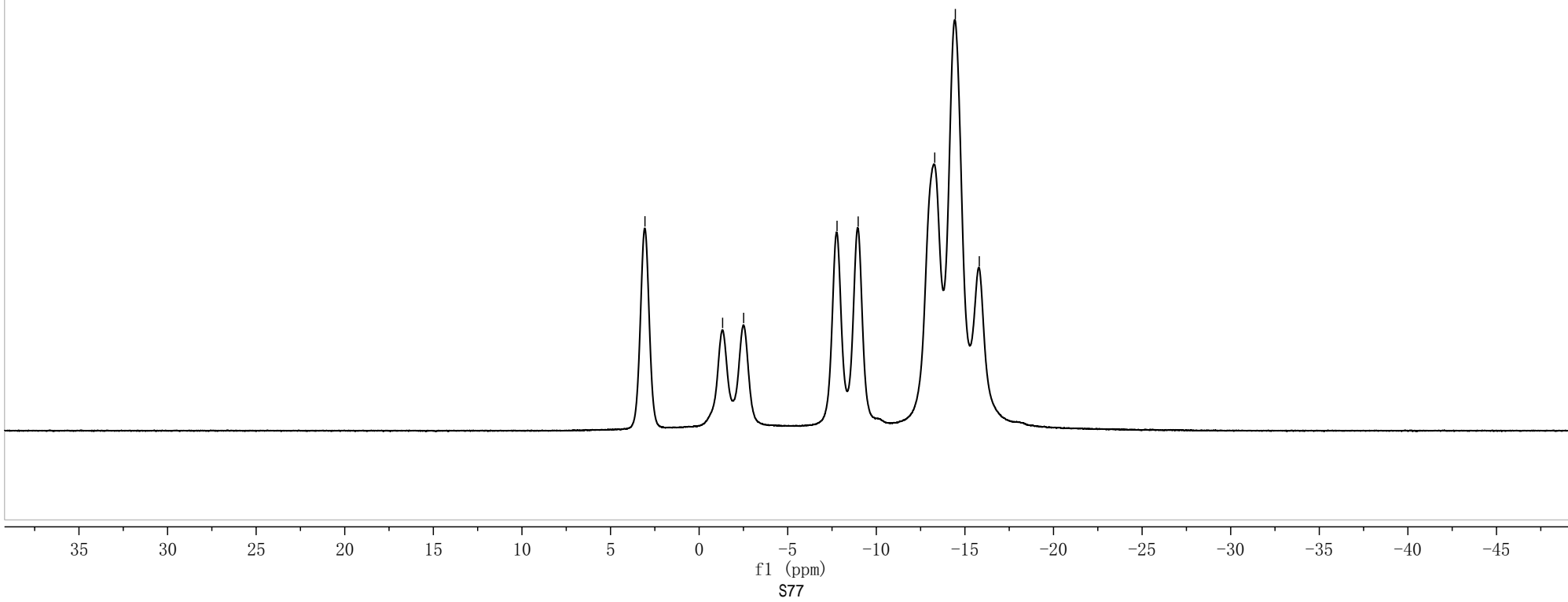
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)

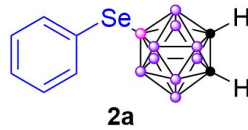


—3.06
—1.33
—2.51
—7.78
—8.98
—13.30
—14.46
—15.81



^{11}B NMR (128 MHz, CDCl_3)



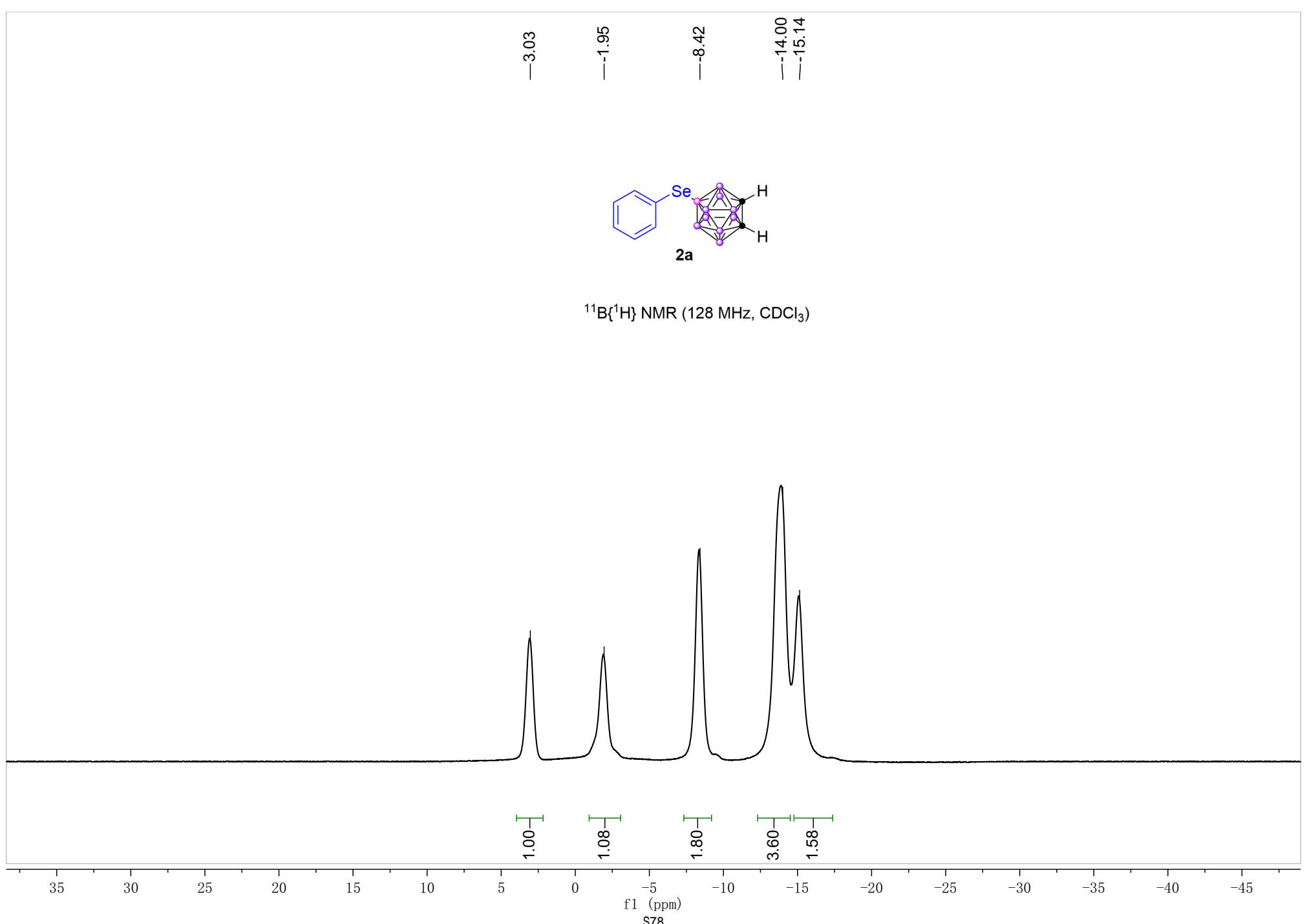


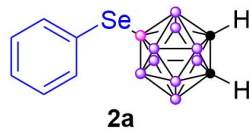
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

—3.03
—1.95
—8.42
—14.00
—15.14

1.00
1.08
1.80
3.60
1.58

f1 (ppm)
S78



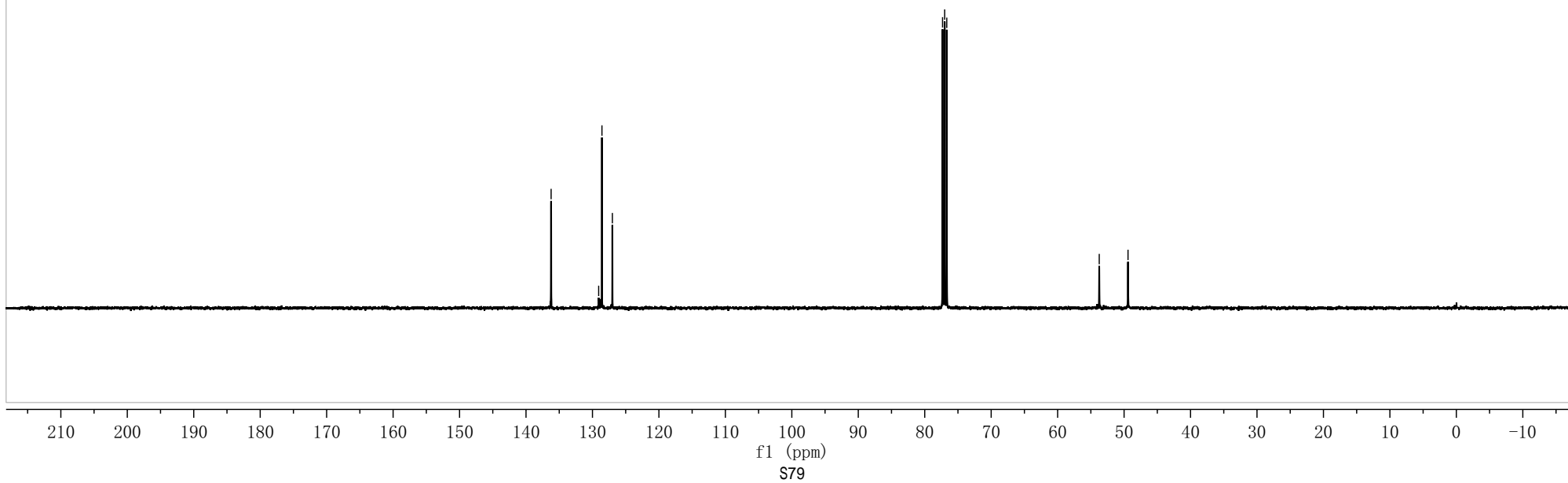


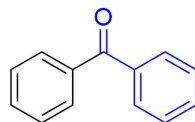
^{13}C NMR (101 MHz, CDCl_3)

—136.22
—129.06
—128.57
—127.00

—77.32
—77.00
—76.68

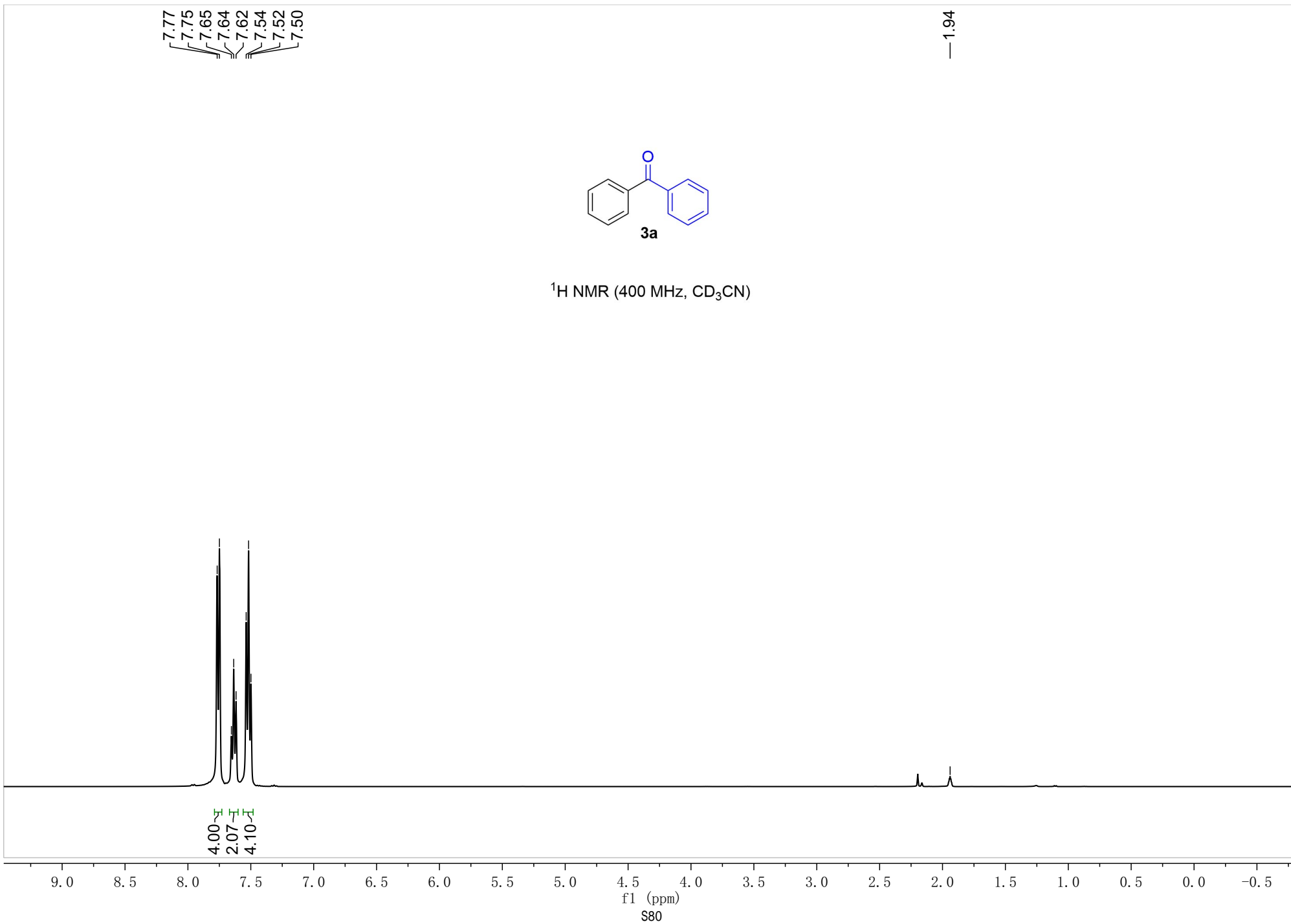
—53.74
—49.41

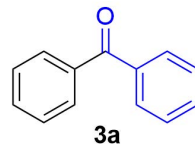




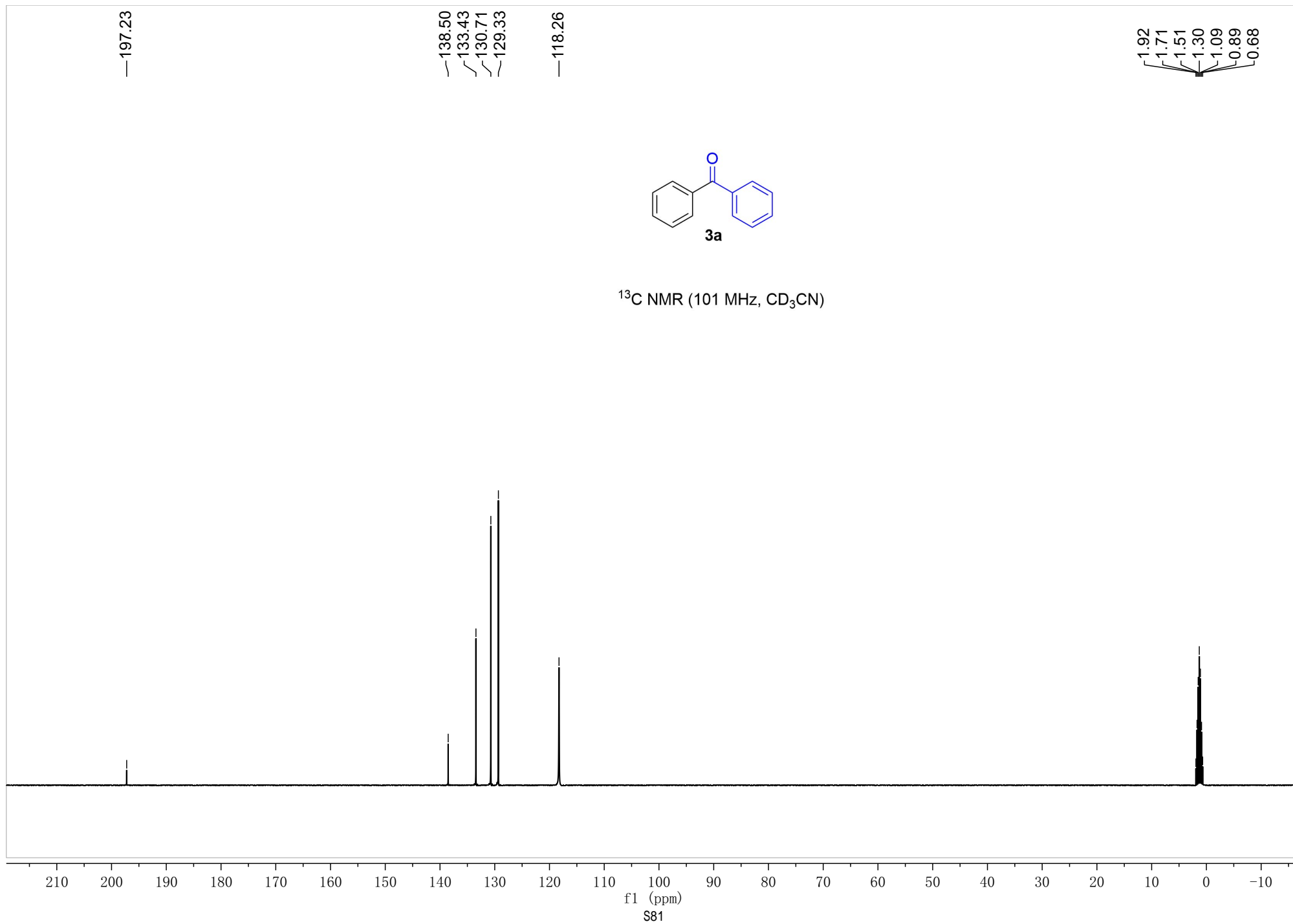
3a

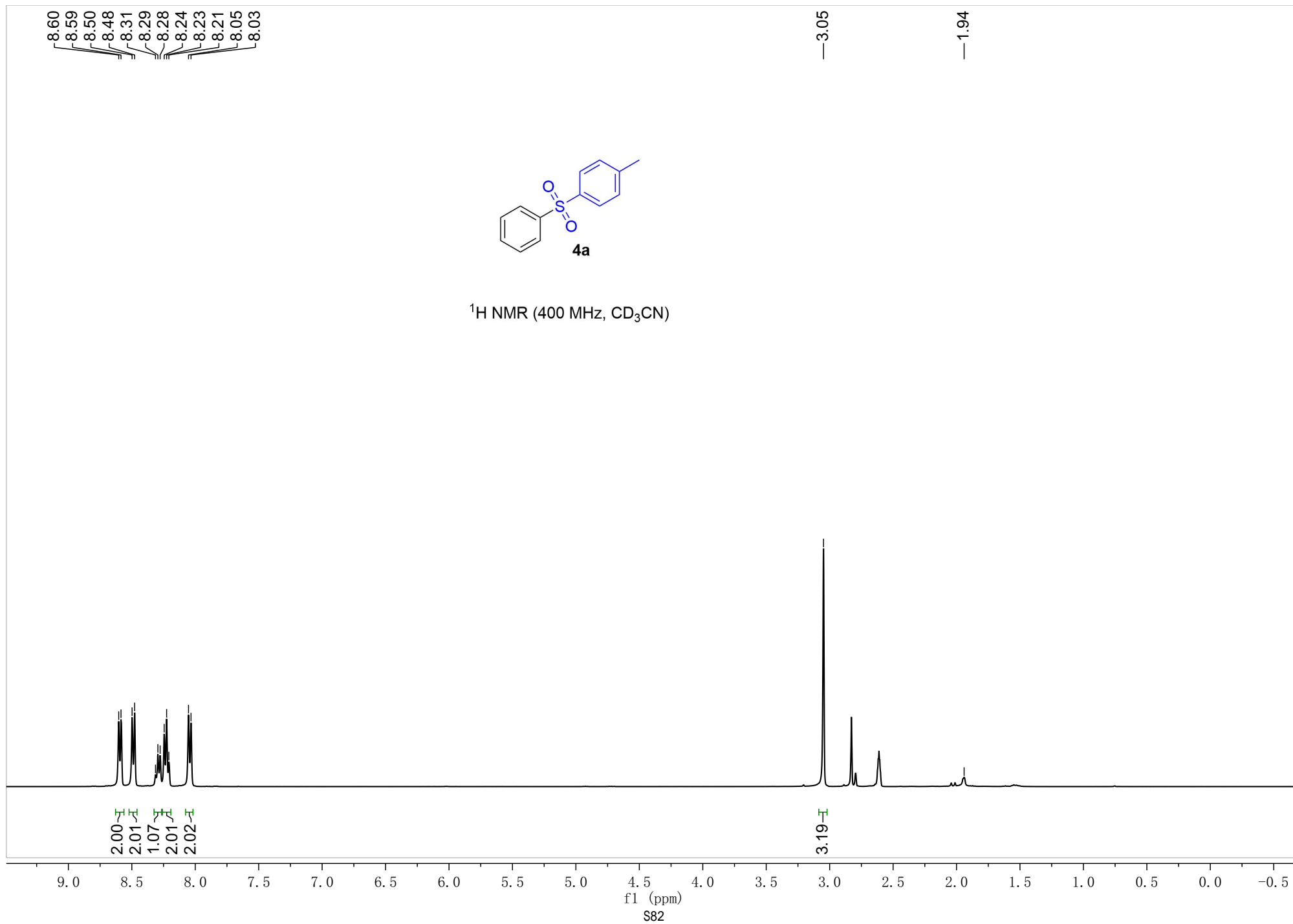
¹H NMR (400 MHz, CD₃CN)

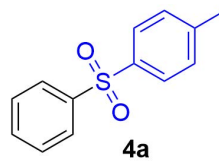




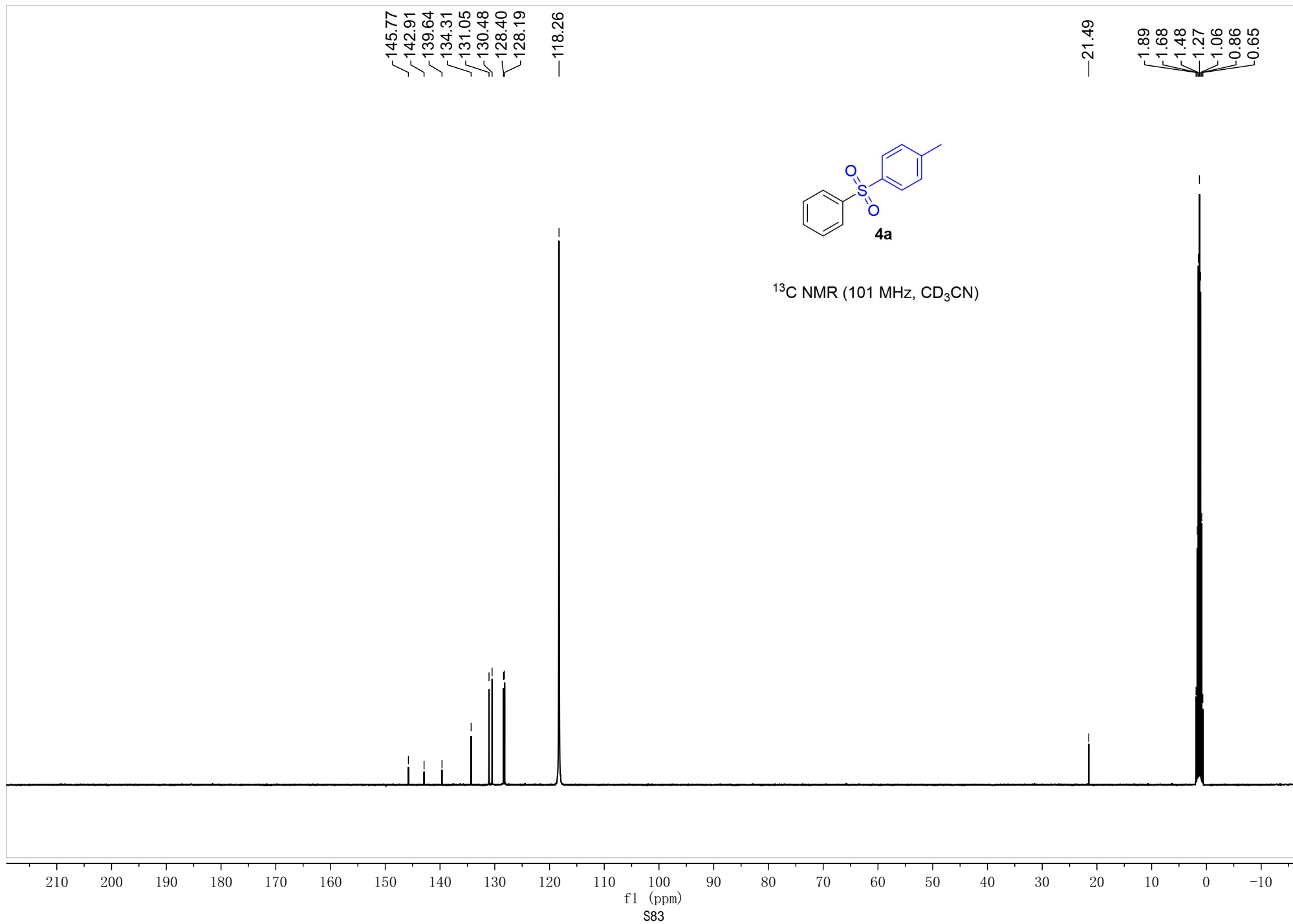
¹³C NMR (101 MHz, CD₃CN)





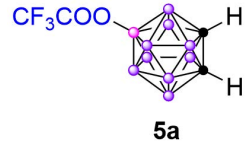


¹³C NMR (101 MHz, CD₃CN)

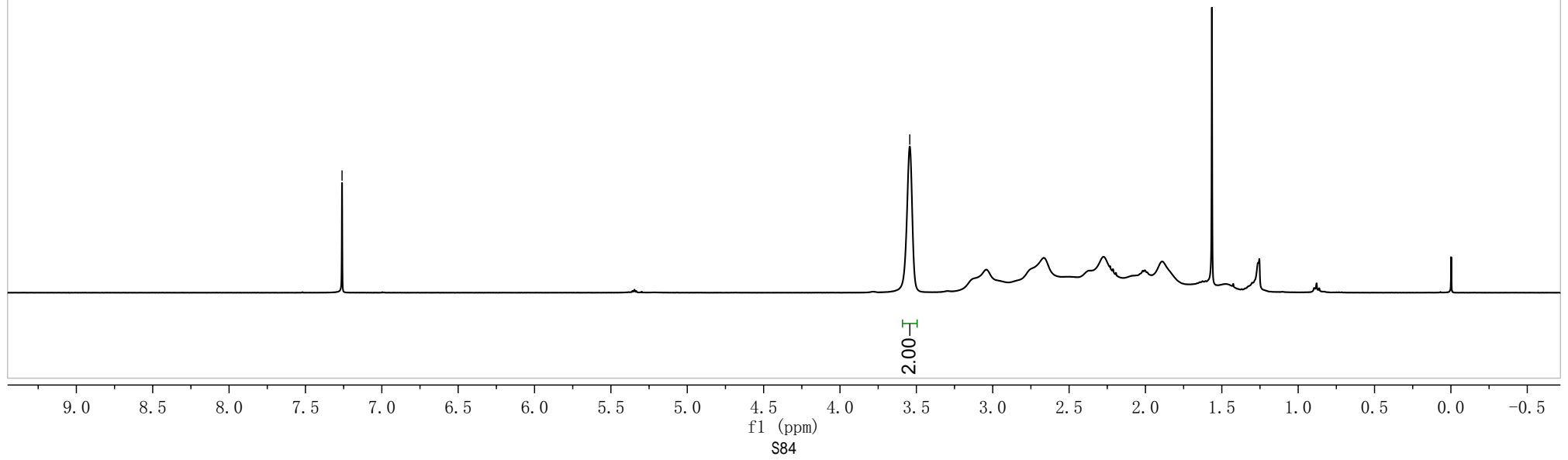


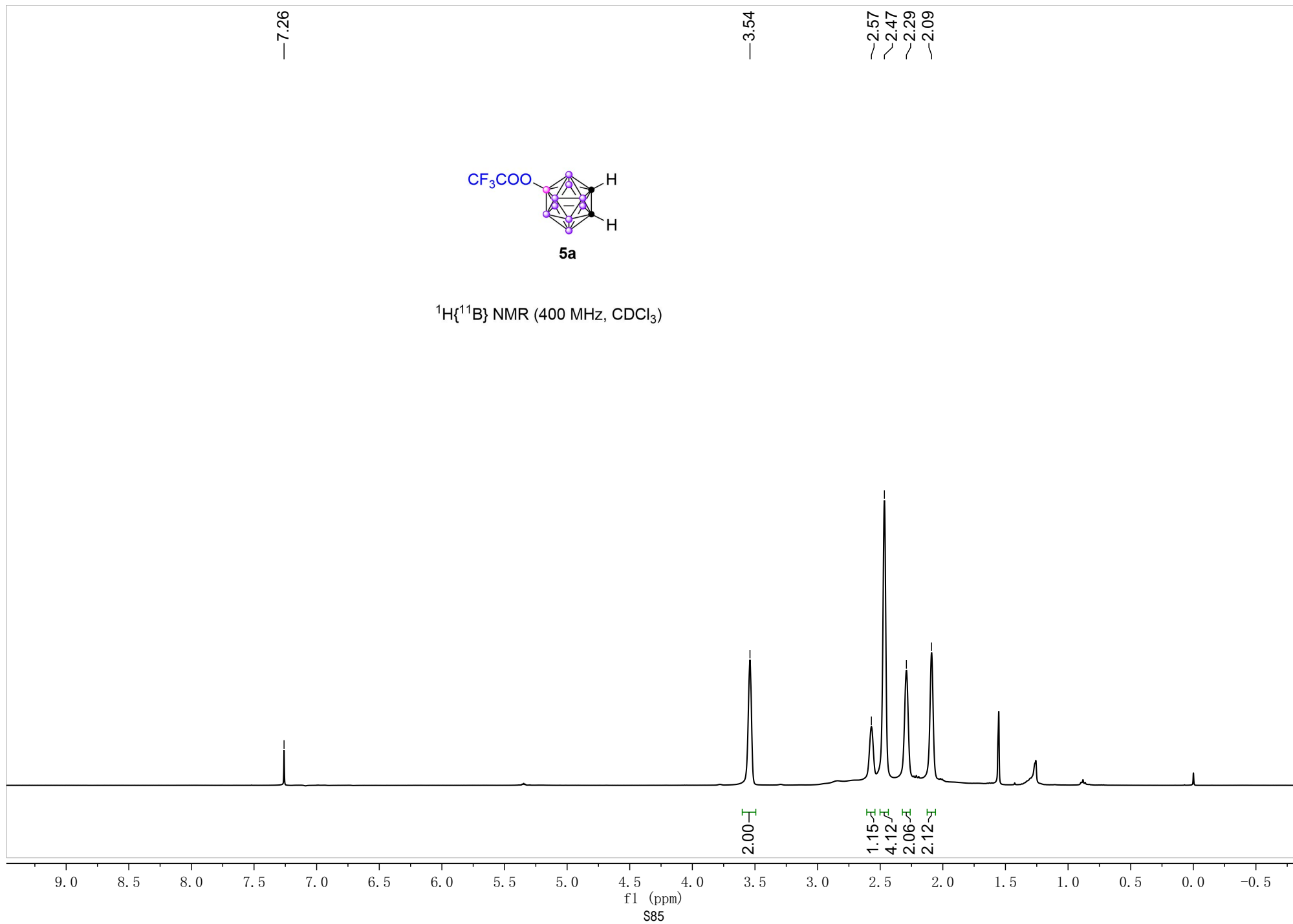
—7.26

—3.54

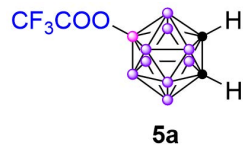


¹H NMR (400 MHz, CDCl₃)

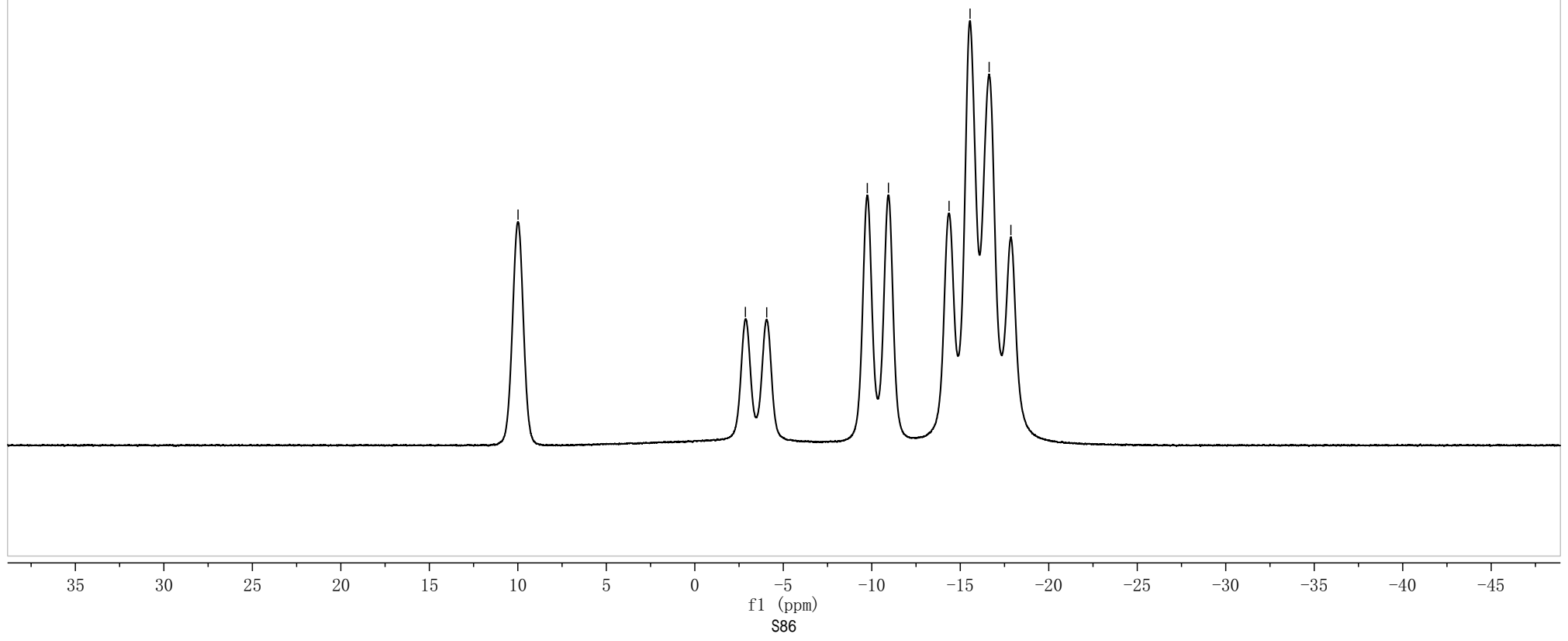


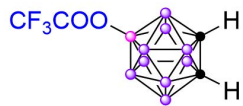


—9.99
—2.86
—4.06
—9.75
—10.94
—14.37
—15.55
—16.63
—17.86



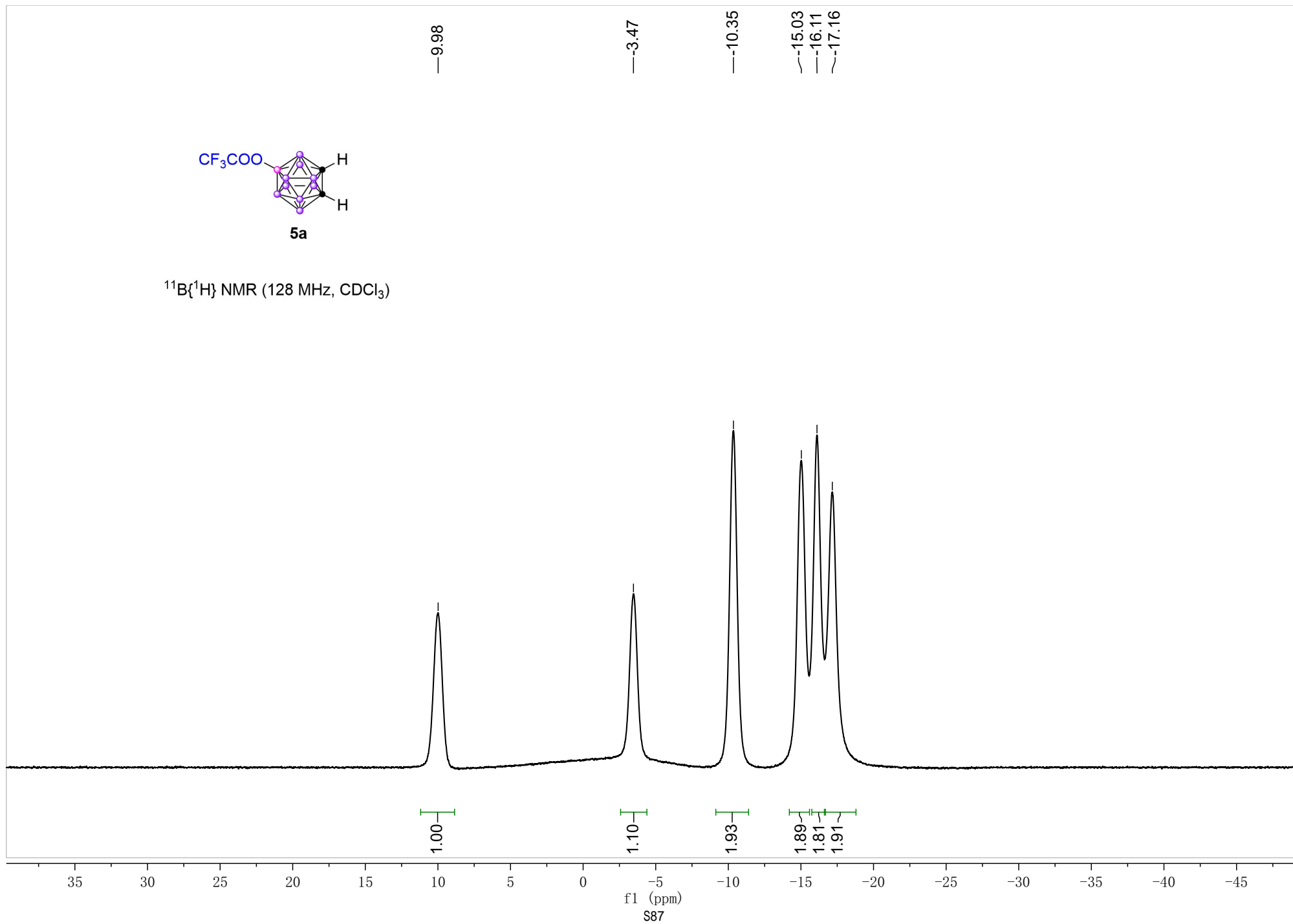
¹¹B NMR (128 MHz, CDCl₃)

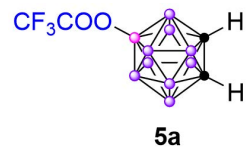




5a

¹¹B{¹H} NMR (128 MHz, CDCl₃)





^{19}F NMR (377 MHz, CDCl_3)

---75.88



10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210

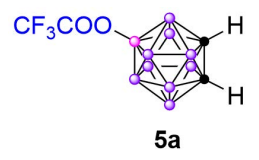
f1 (ppm)
S88

156.41
155.99
155.58
155.16

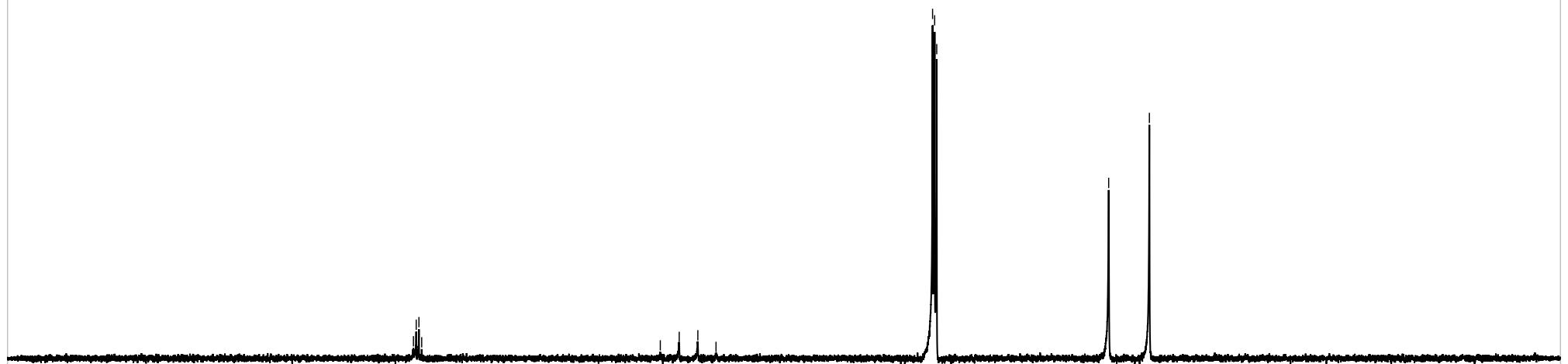
118.80
115.92
113.08
110.31

77.32
77.00
76.68

50.48
44.28

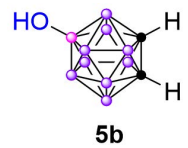


¹³C NMR (101 MHz, CDCl₃)



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm)
S89

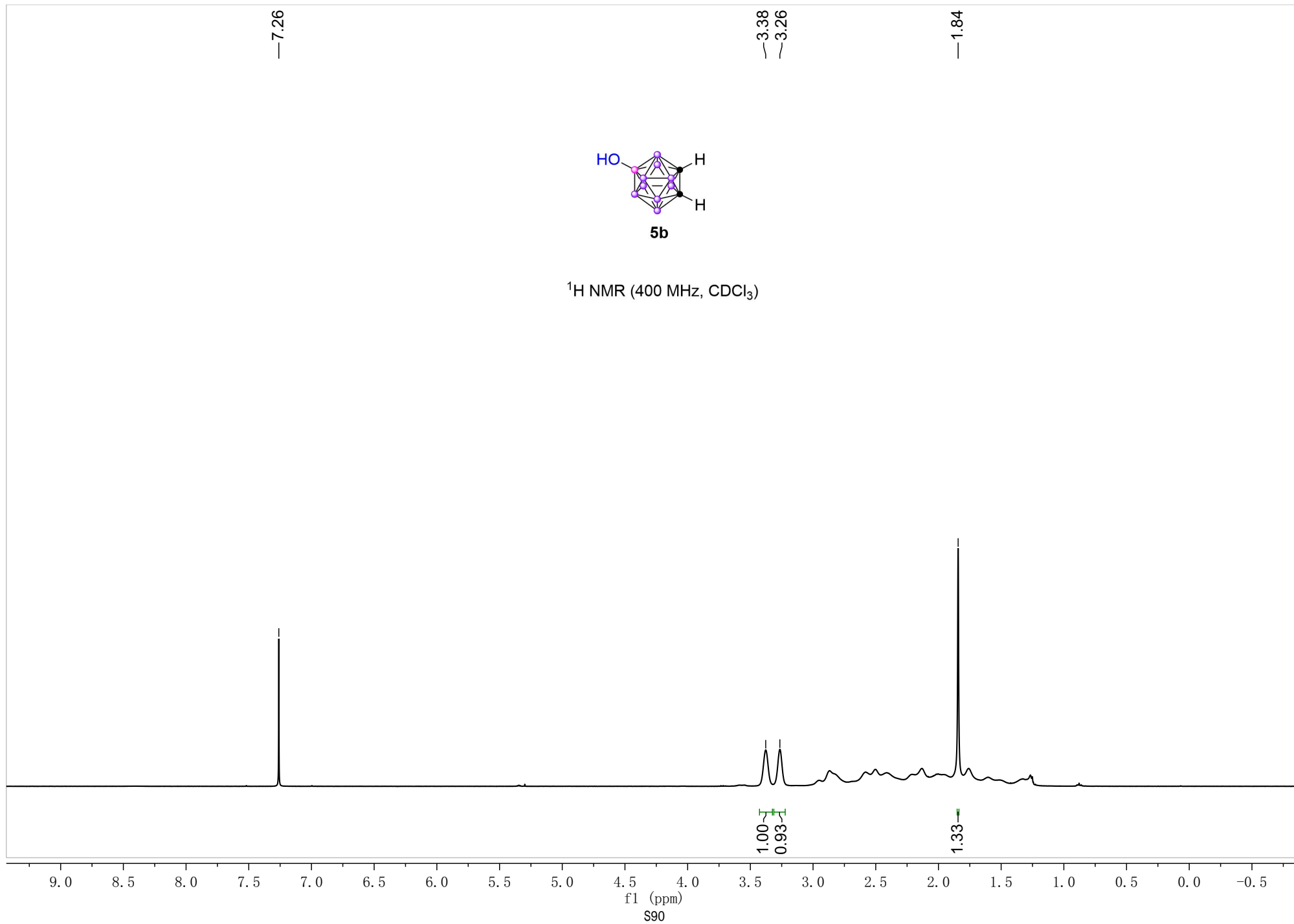


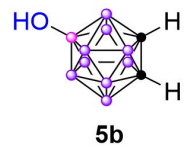
¹H NMR (400 MHz, CDCl₃)

—7.26

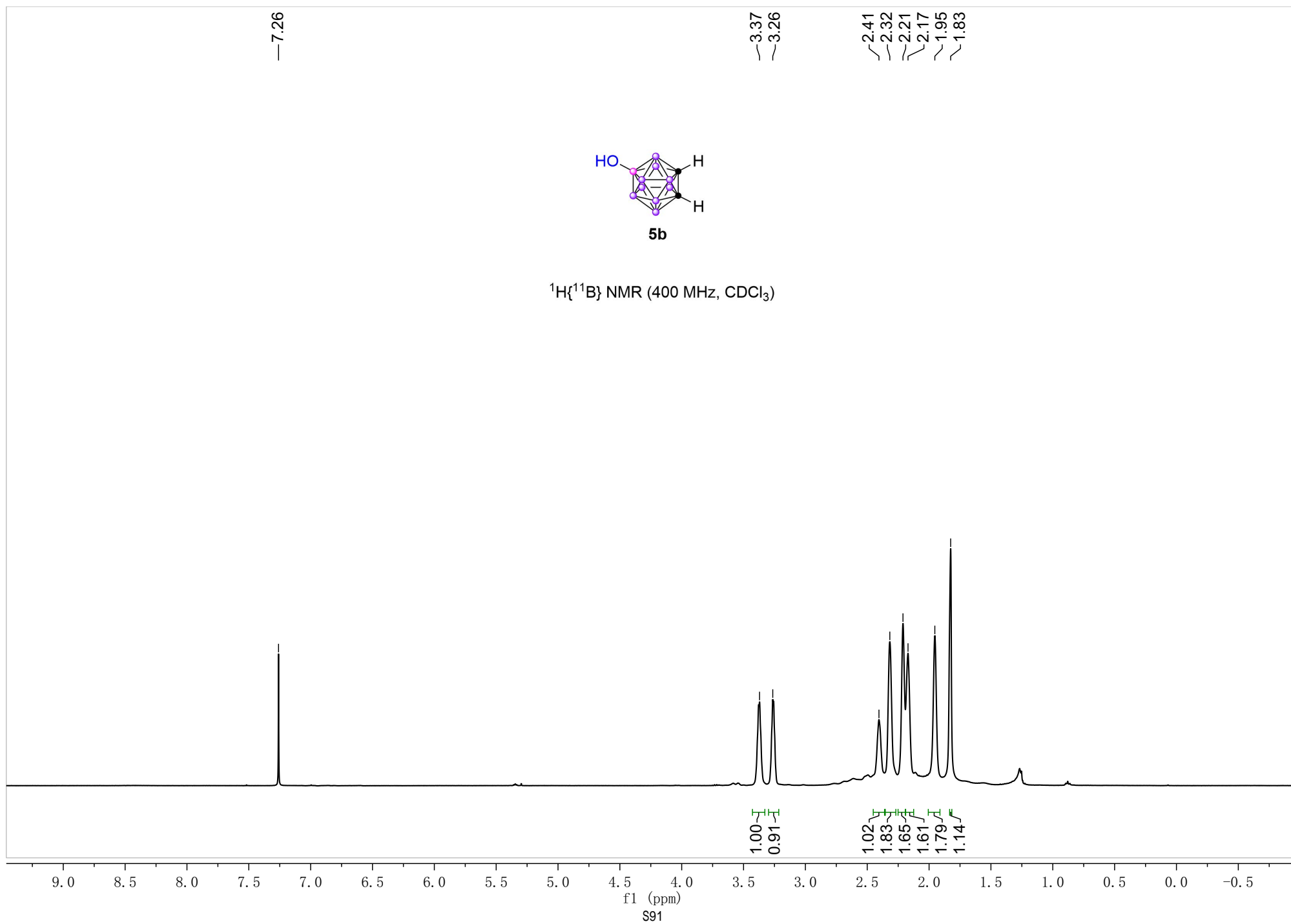
—3.38
—3.26

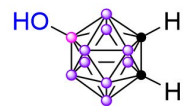
—1.84





$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)





5b

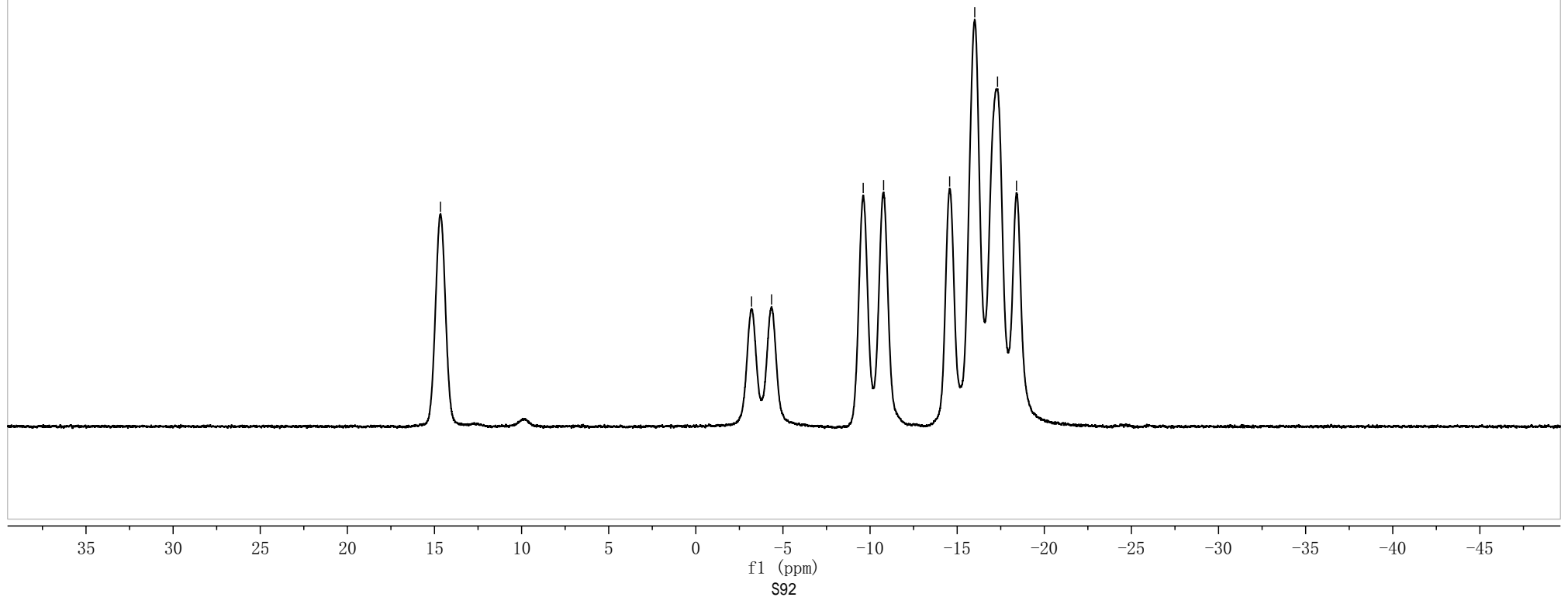
^{11}B NMR (128 MHz, CDCl_3)

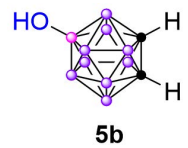
—14.65

—3.20
—4.35

—9.61
—10.78

—14.57
—16.01
—17.32
—18.40





$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

—14.68

—-3.76

—-10.18

—-15.21

—-16.75

—-17.72

1.00

1.17

2.07

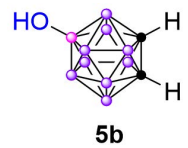
2.09

2.07

1.89

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45

f1 (ppm)
S93

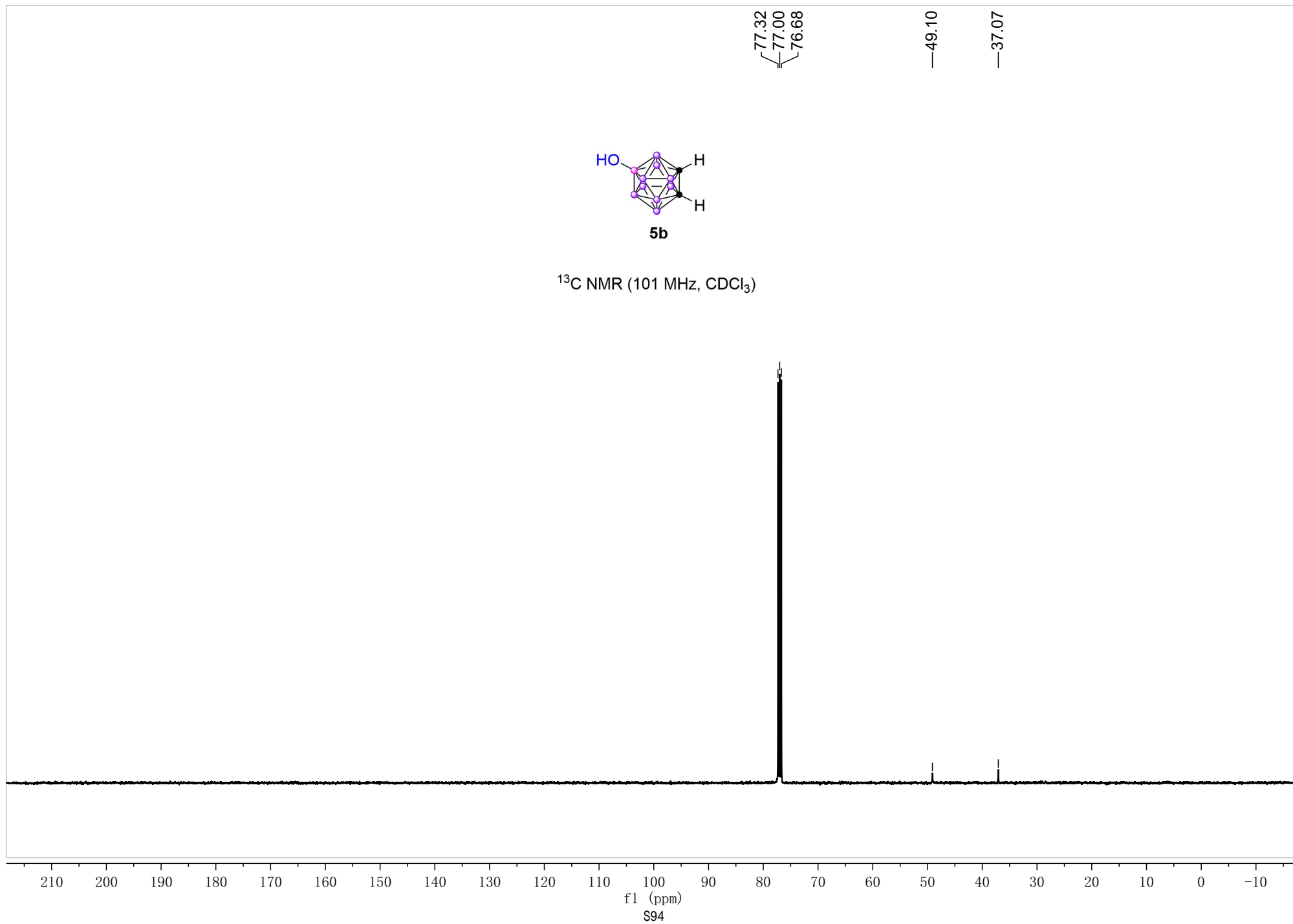


^{13}C NMR (101 MHz, CDCl_3)

77.32
77.00
76.68

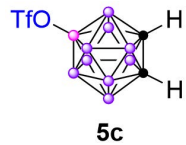
49.10

37.07

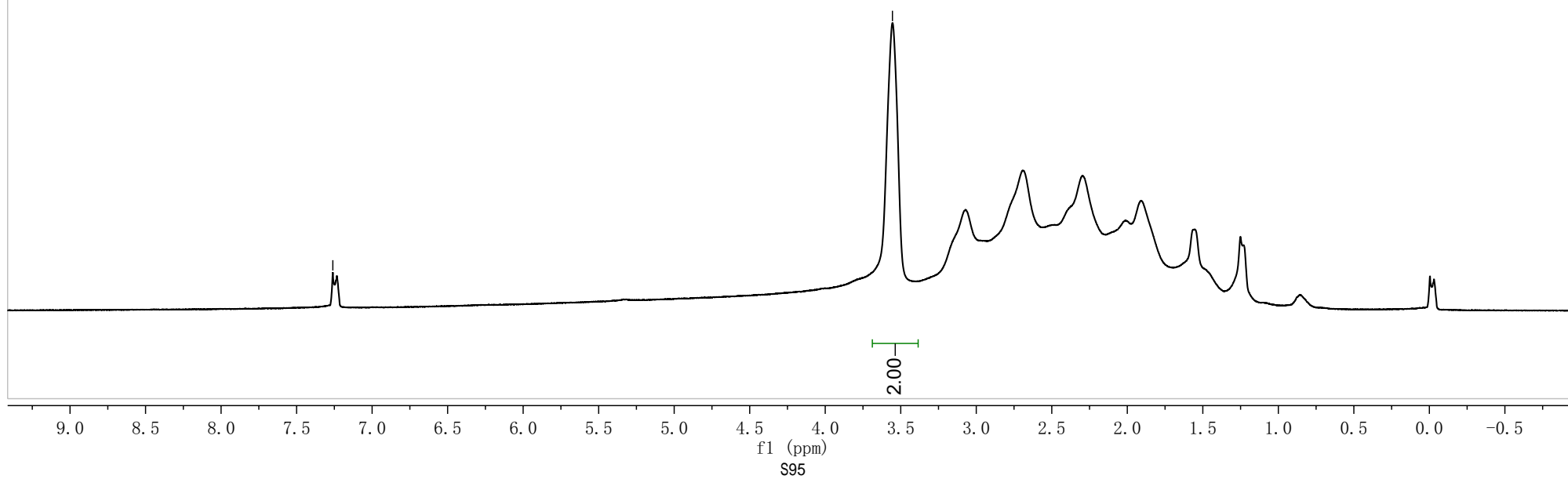


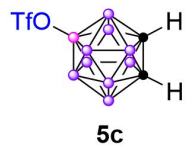
—7.26

—3.55

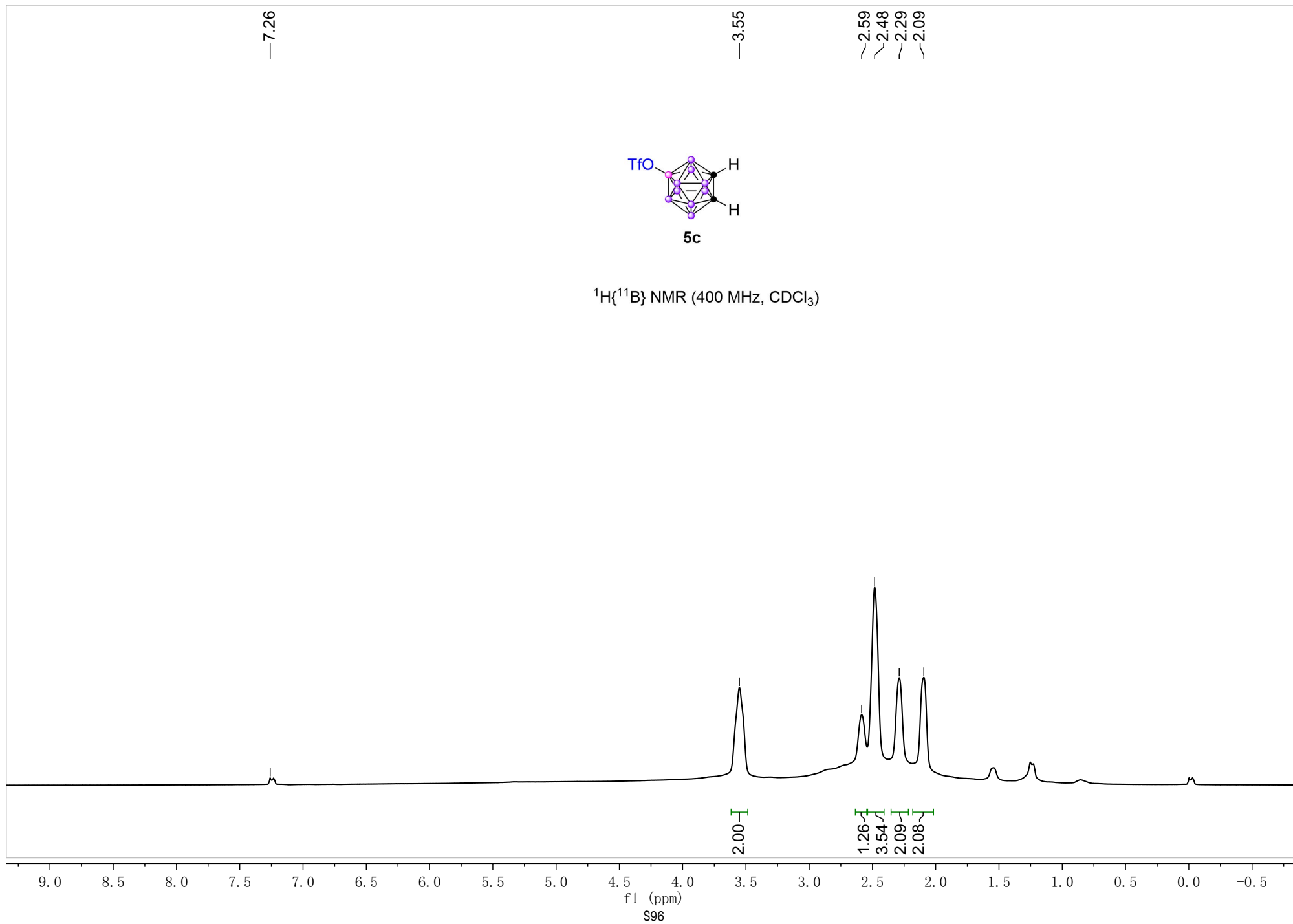


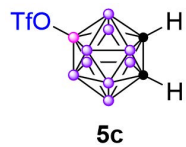
^1H NMR (400 MHz, CDCl_3)





$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)





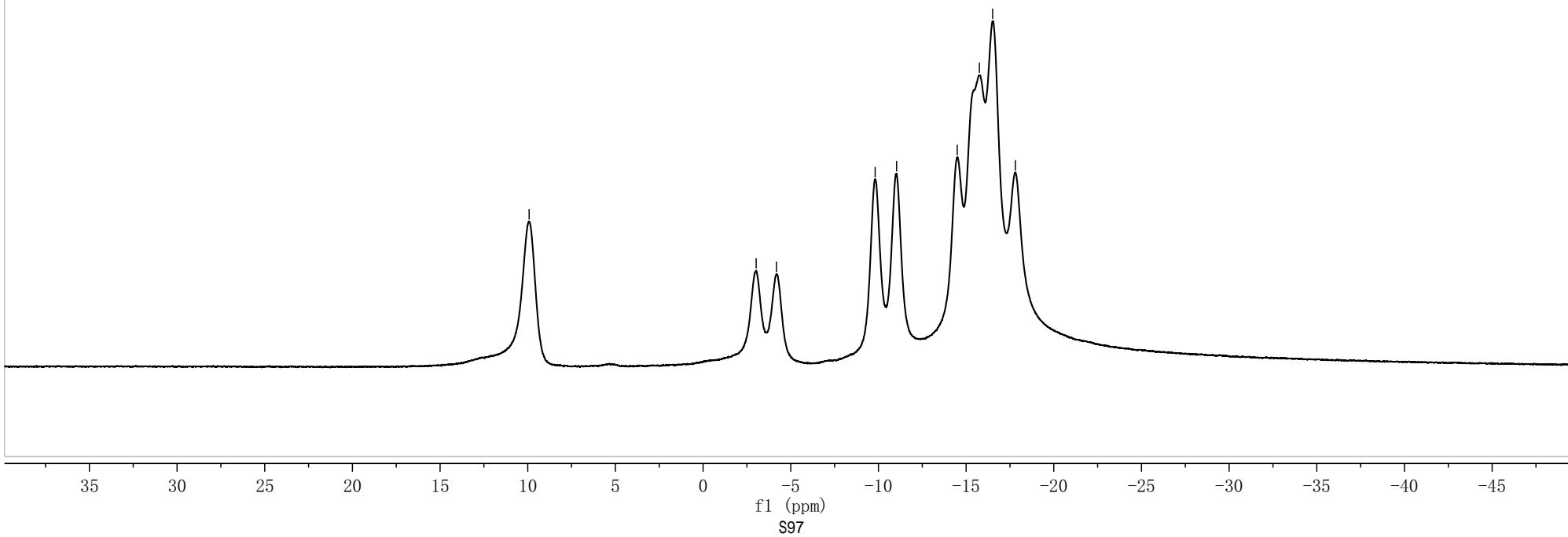
^{11}B NMR (128 MHz, CDCl_3)

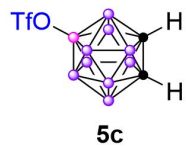
—9.92

—3.03
—4.18

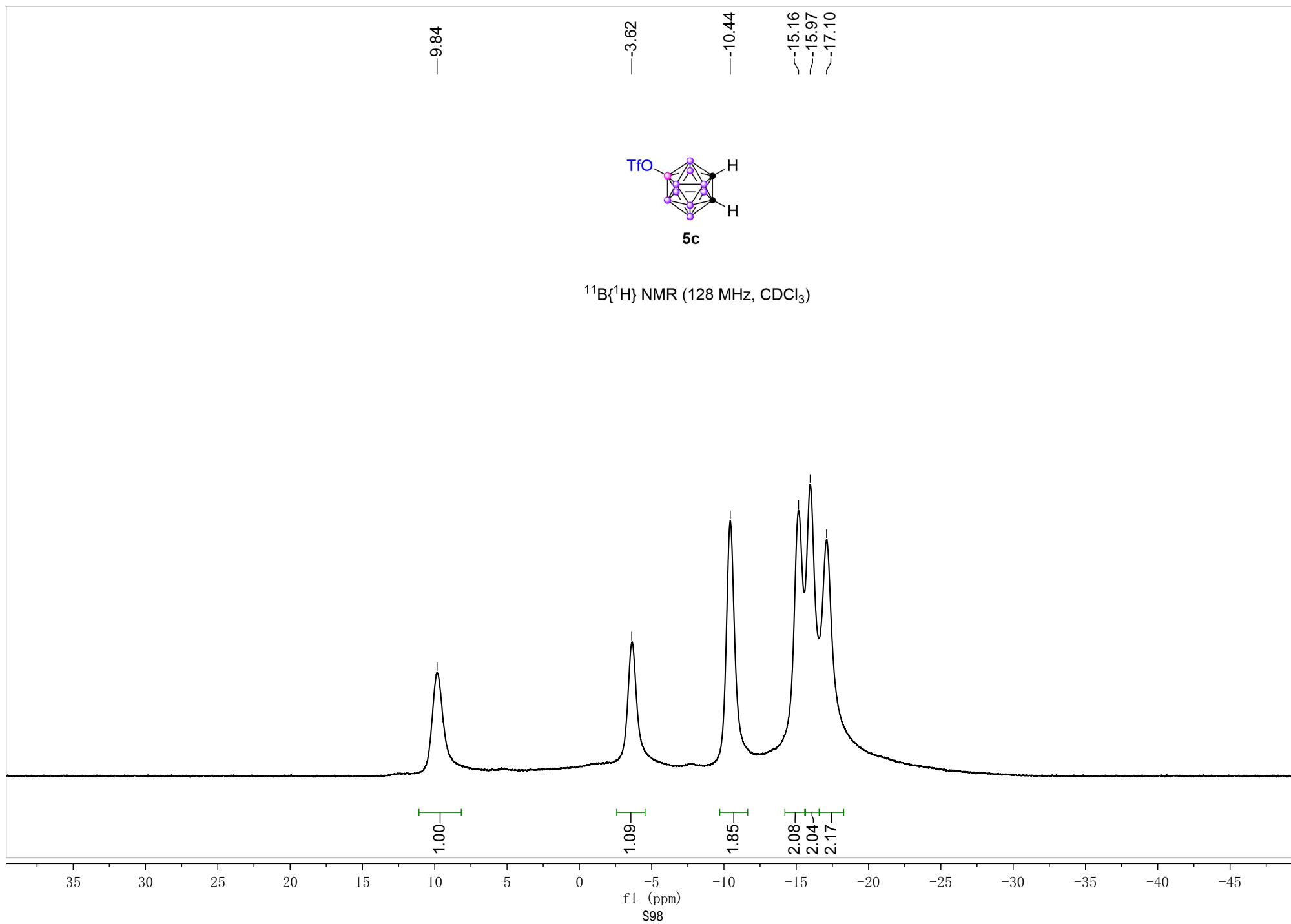
—9.81
—11.04

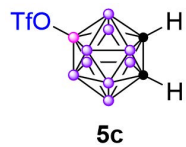
—14.49
—15.76
—16.52
—17.81





$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)



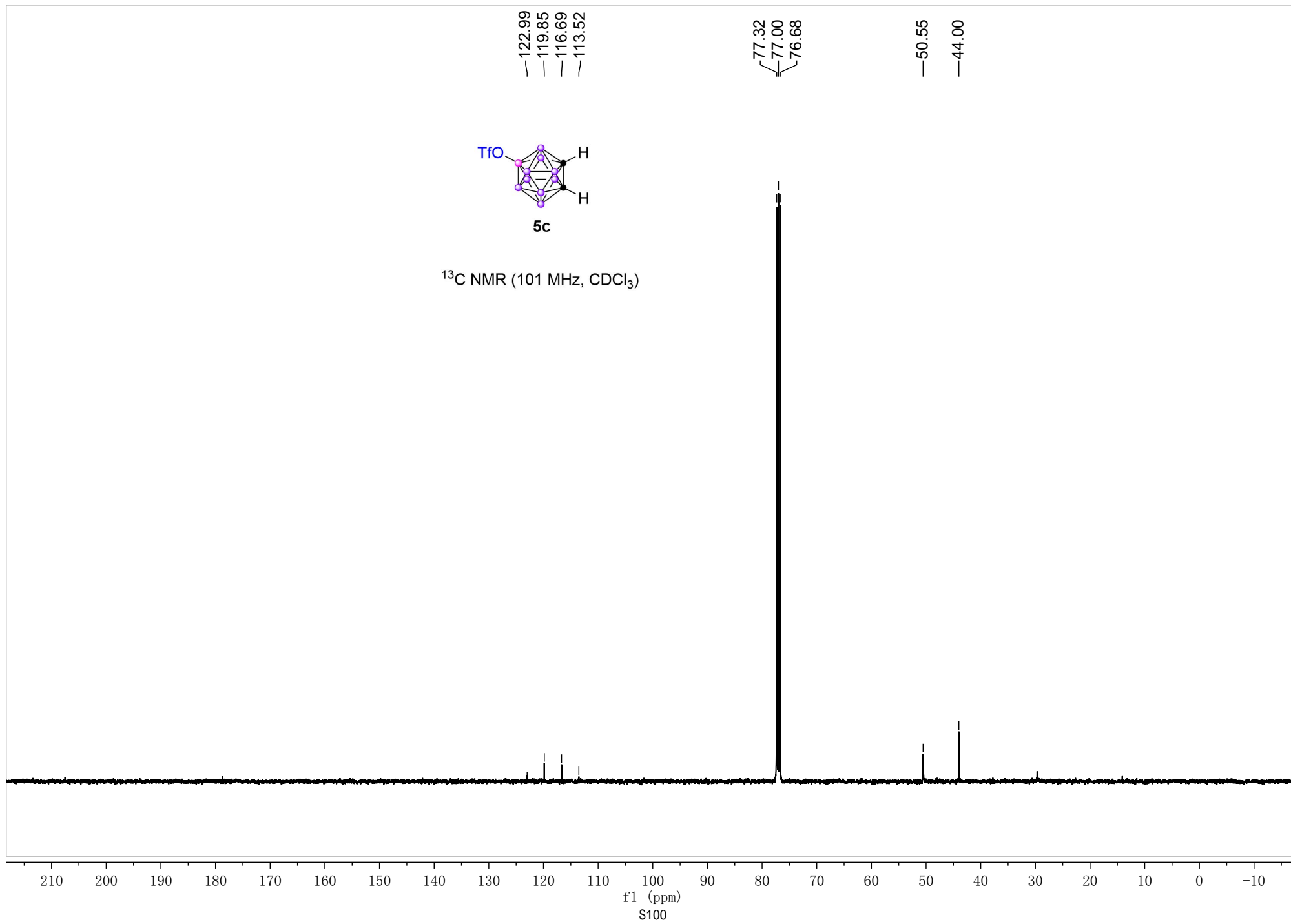


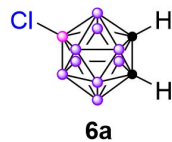
^{19}F NMR (377 MHz, CDCl_3)

---75.91



20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220
f1 (ppm)
S99



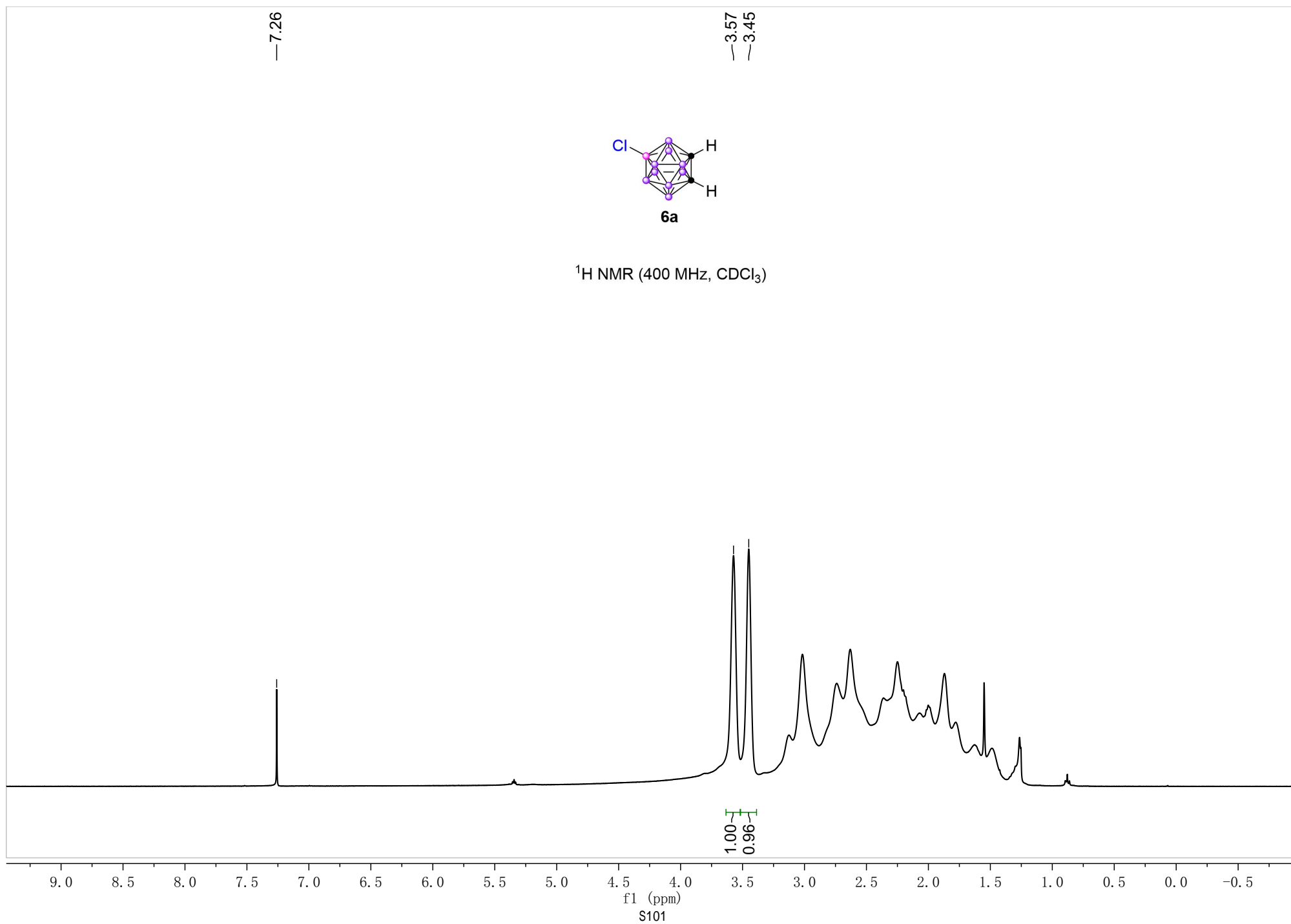


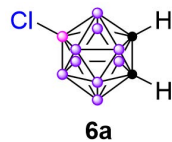
^1H NMR (400 MHz, CDCl_3)

—7.26

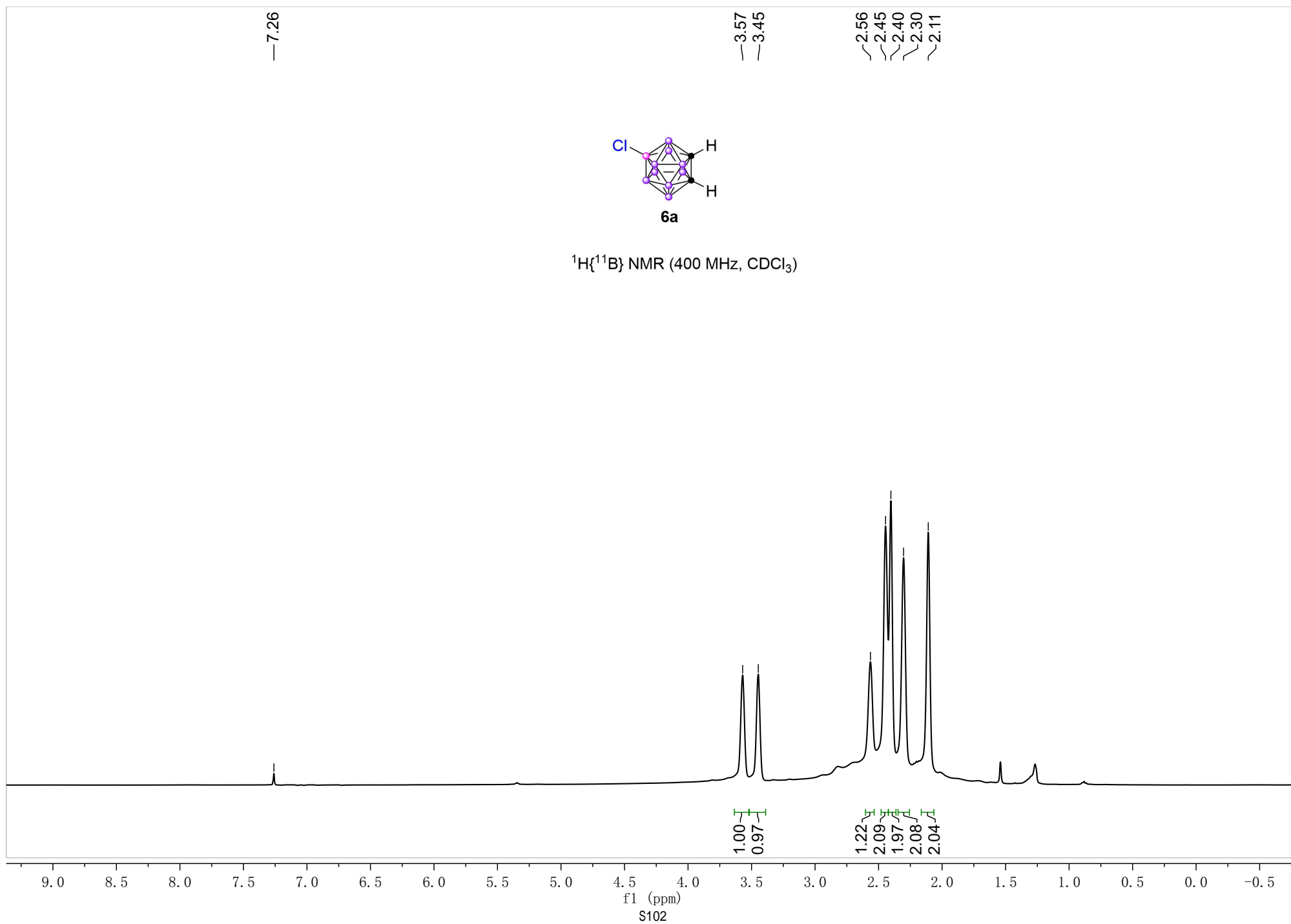
—3.57
—3.45

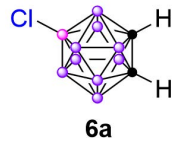
1.00
0.96





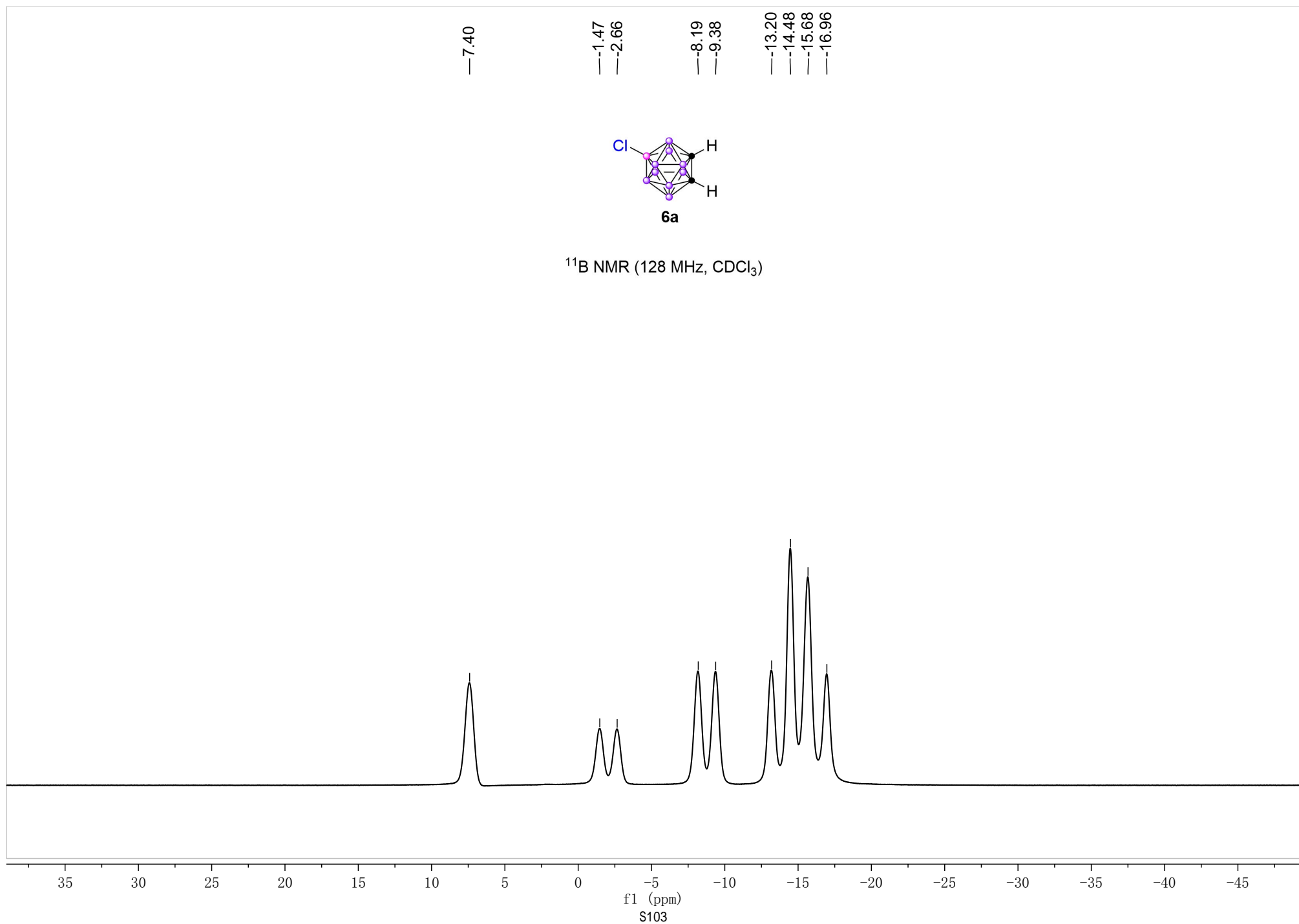
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



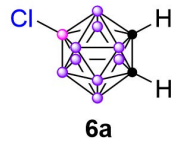


¹¹B NMR (128 MHz, CDCl₃)

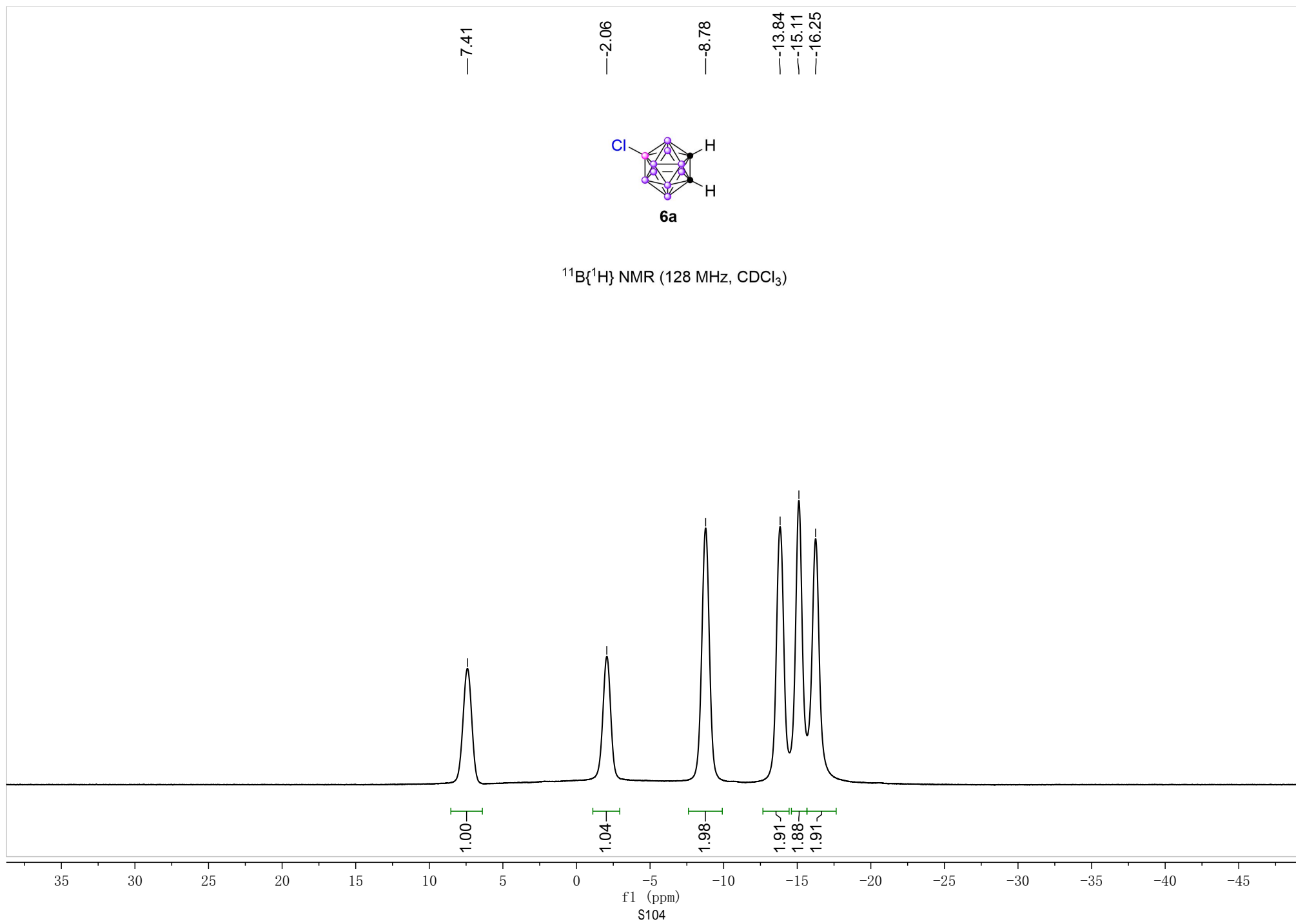
7.40
-1.47
-2.66
-8.19
-9.38
-13.20
-14.48
-15.68
-16.96

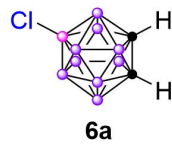


35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45
f1 (ppm)
S103



$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)



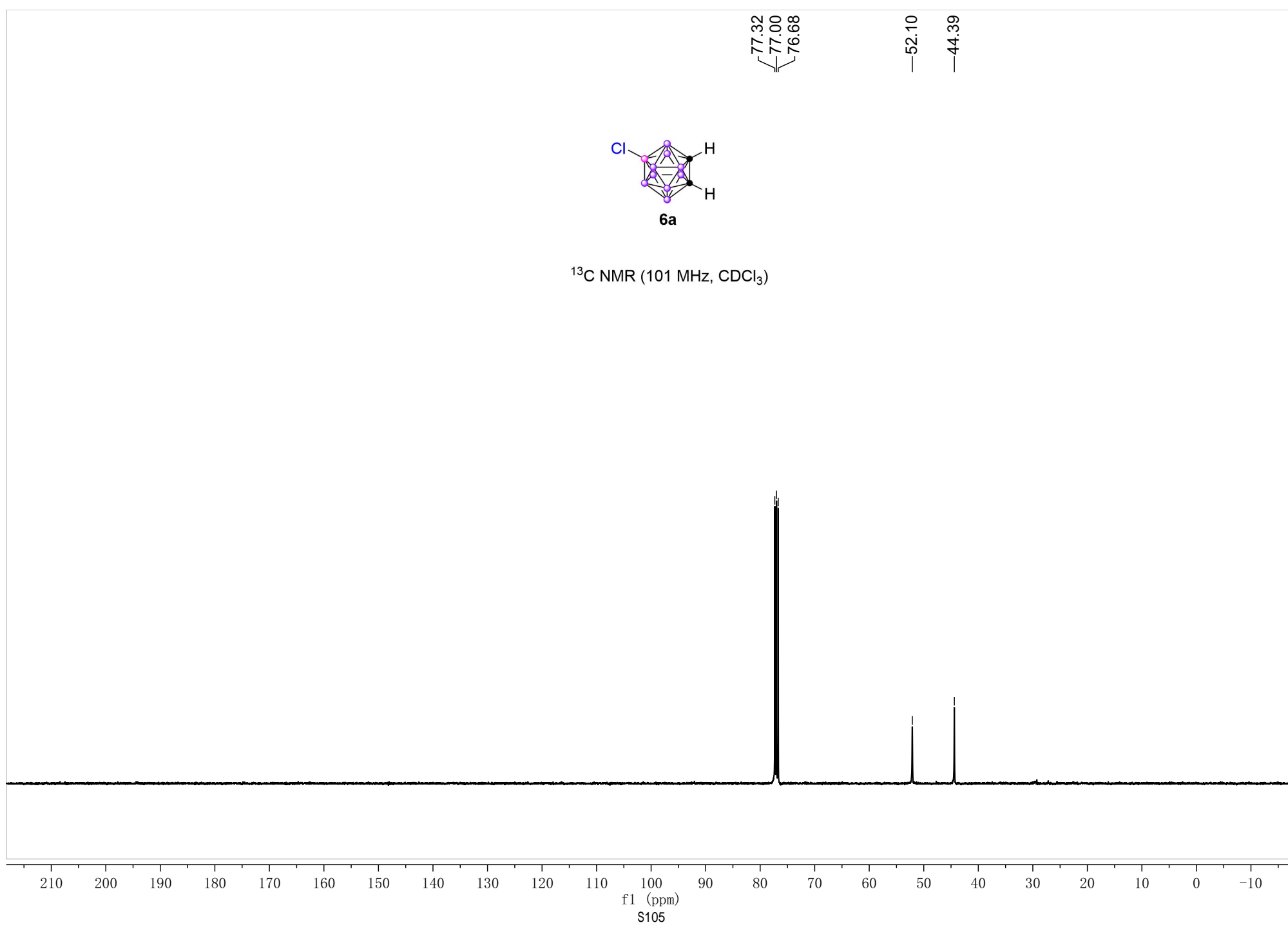


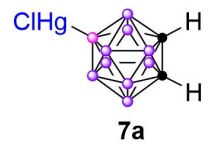
^{13}C NMR (101 MHz, CDCl_3)

77.32
77.00
76.68

52.10

44.39

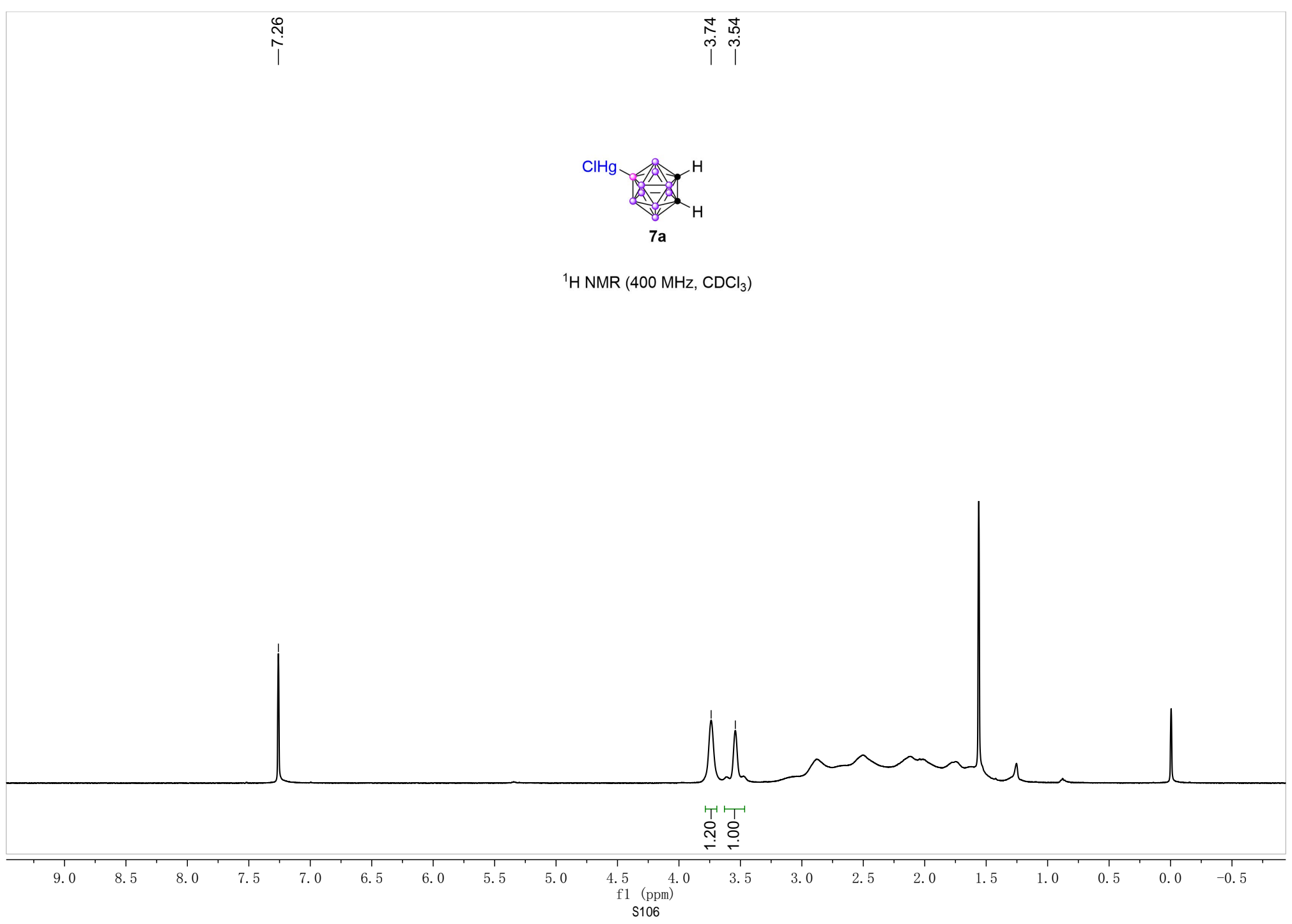


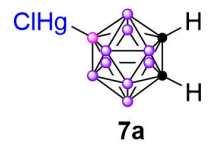


¹H NMR (400 MHz, CDCl₃)

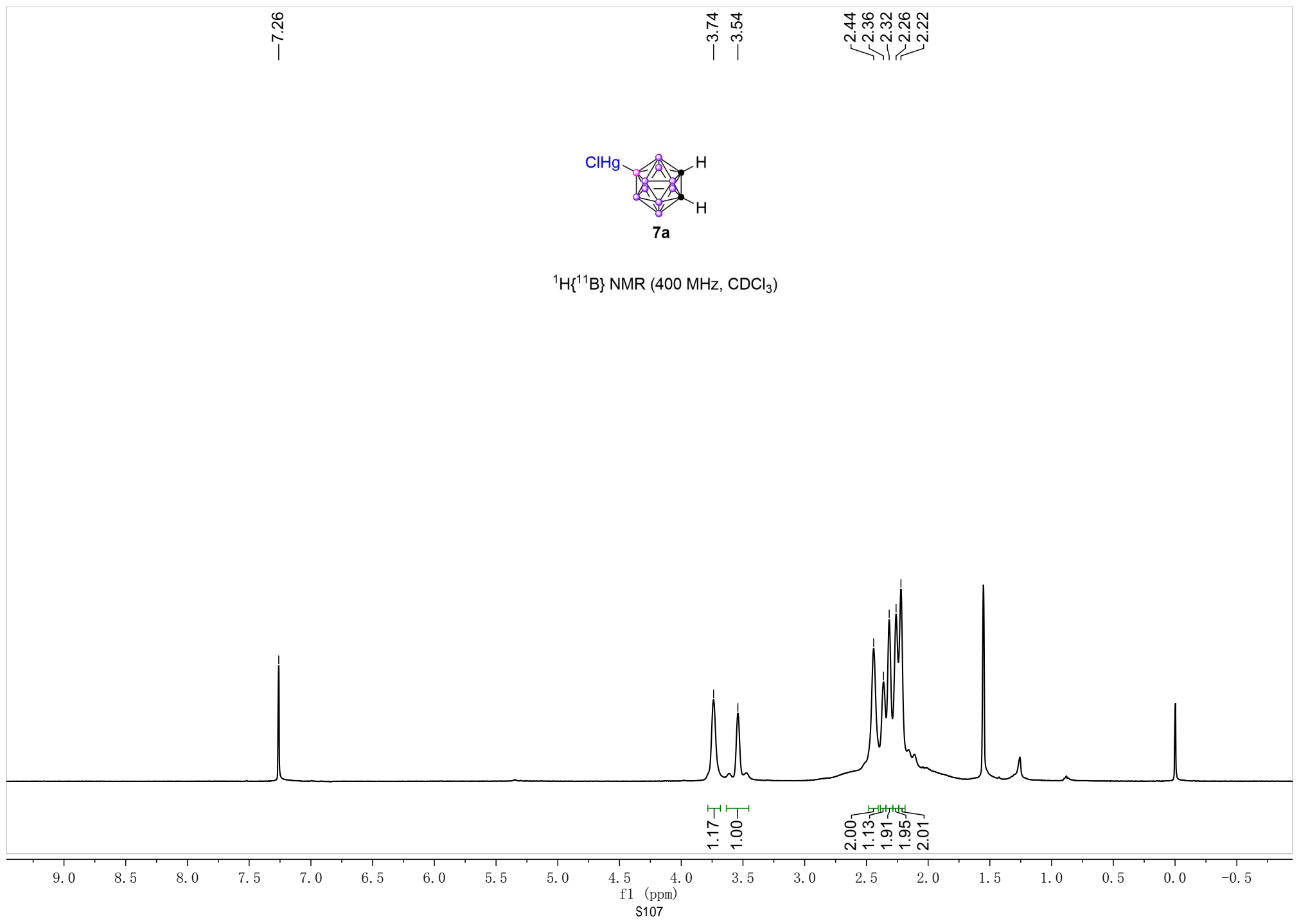
—7.26

—3.74
—3.54

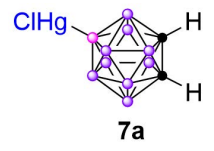




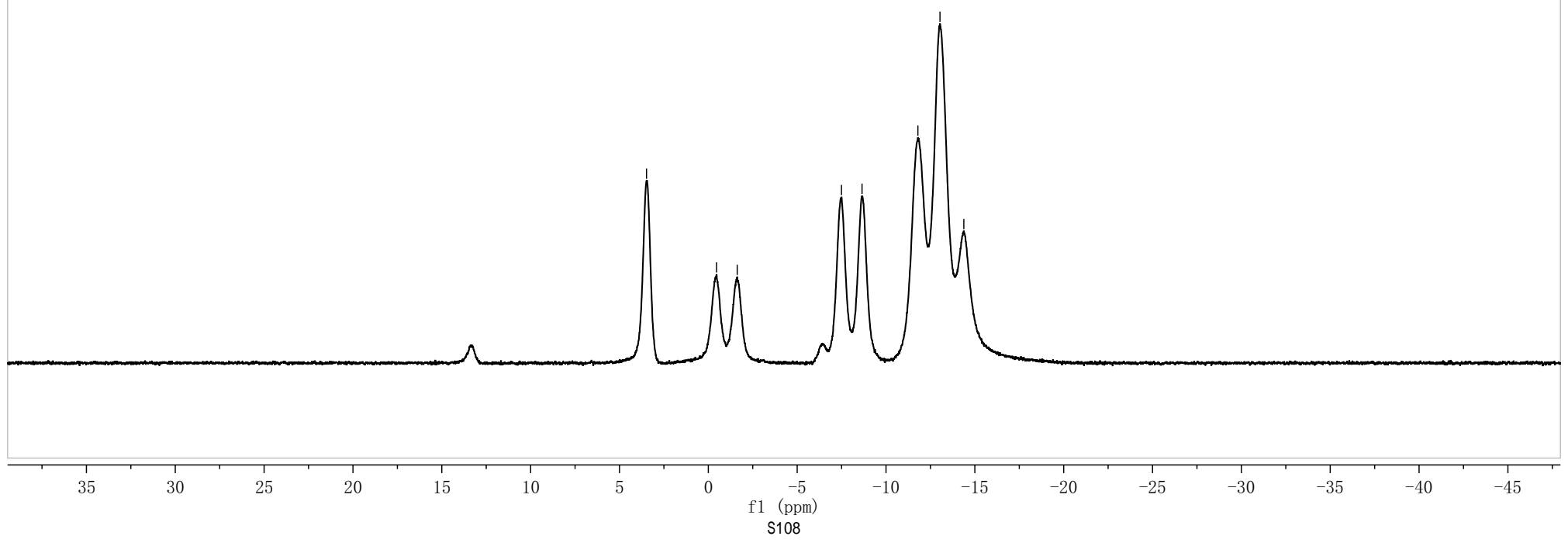
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)

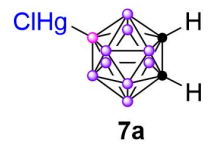


— 3.48
— 0.46
— 1.63
— 7.49
— 8.65
— 11.79
— 13.04
— 14.38



¹¹B NMR (128 MHz, CDCl₃)



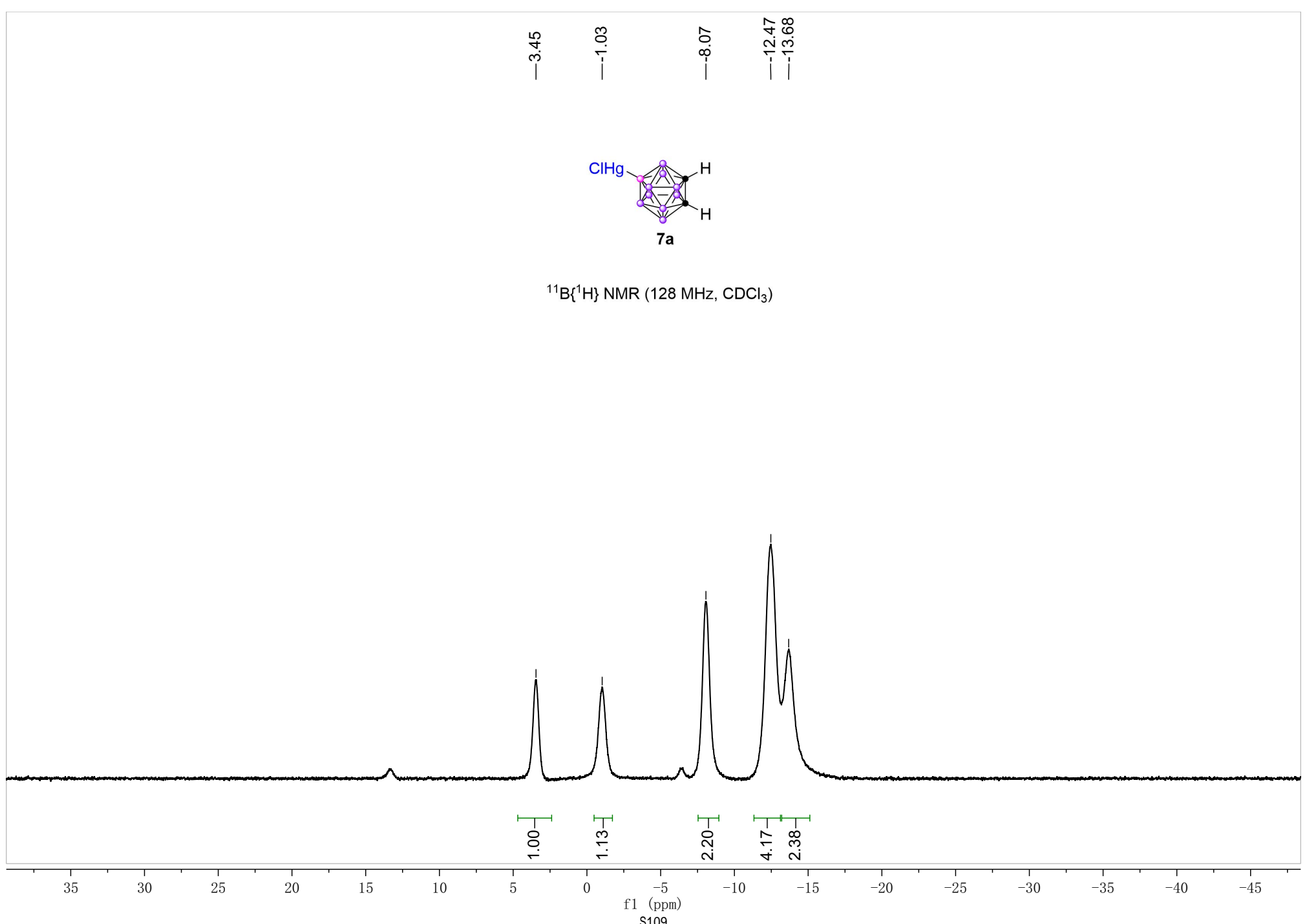


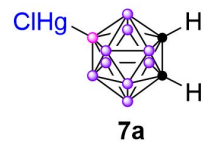
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

—3.45
—-1.03
—-8.07
—-12.47
—-13.68

1.00
1.13
2.20
4.17
2.38

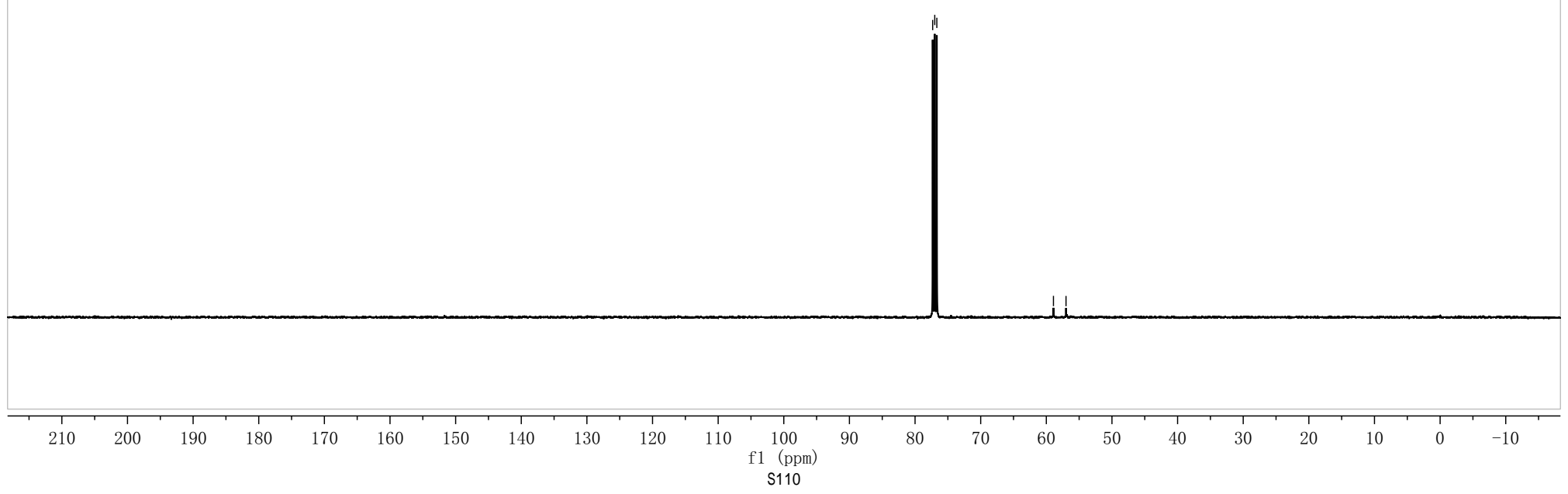
f1 (ppm)
S109

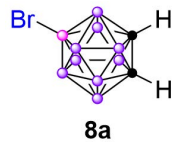




¹³C NMR (101 MHz, CDCl₃)

77.32
77.00
76.68
58.91
57.00

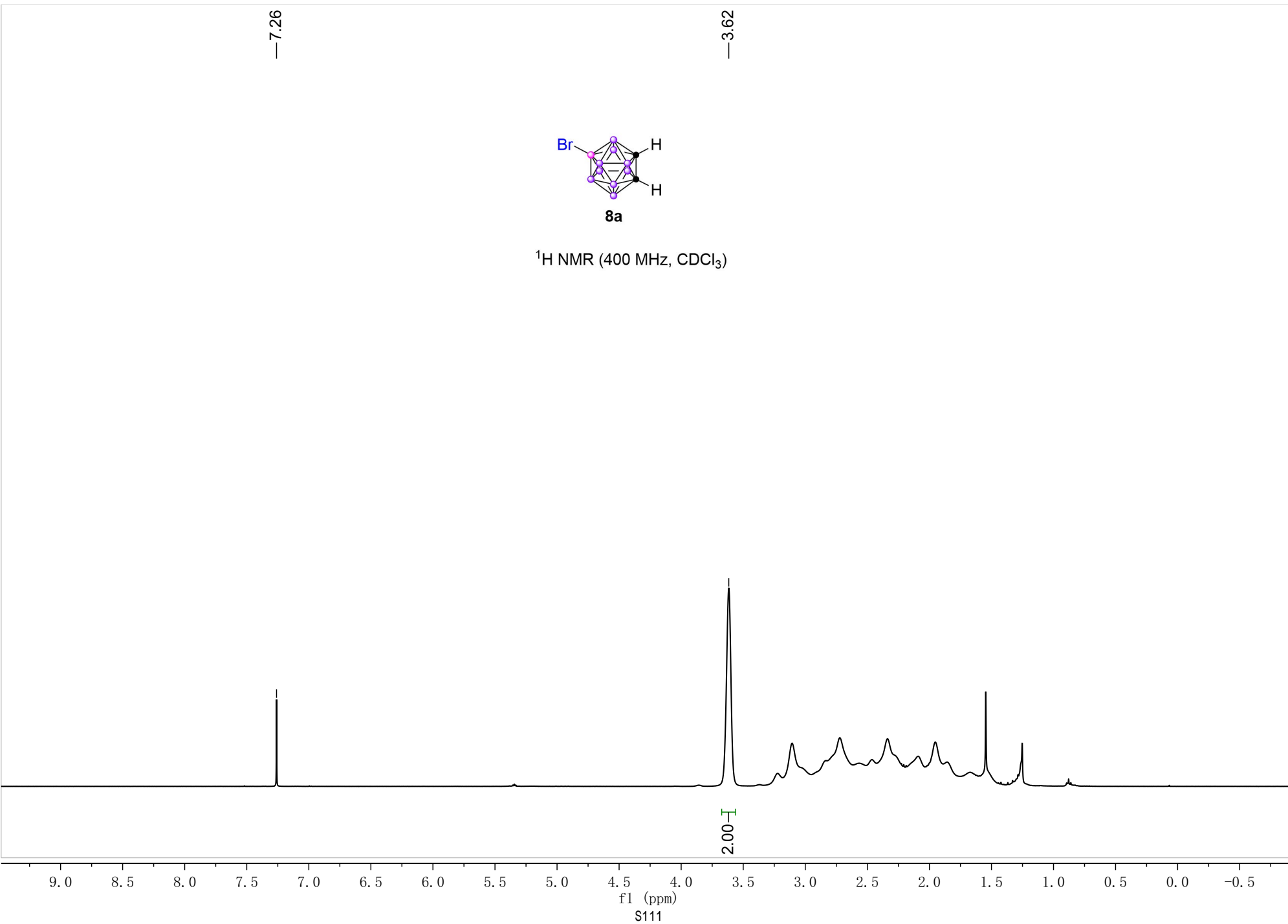


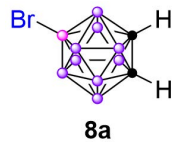


^1H NMR (400 MHz, CDCl_3)

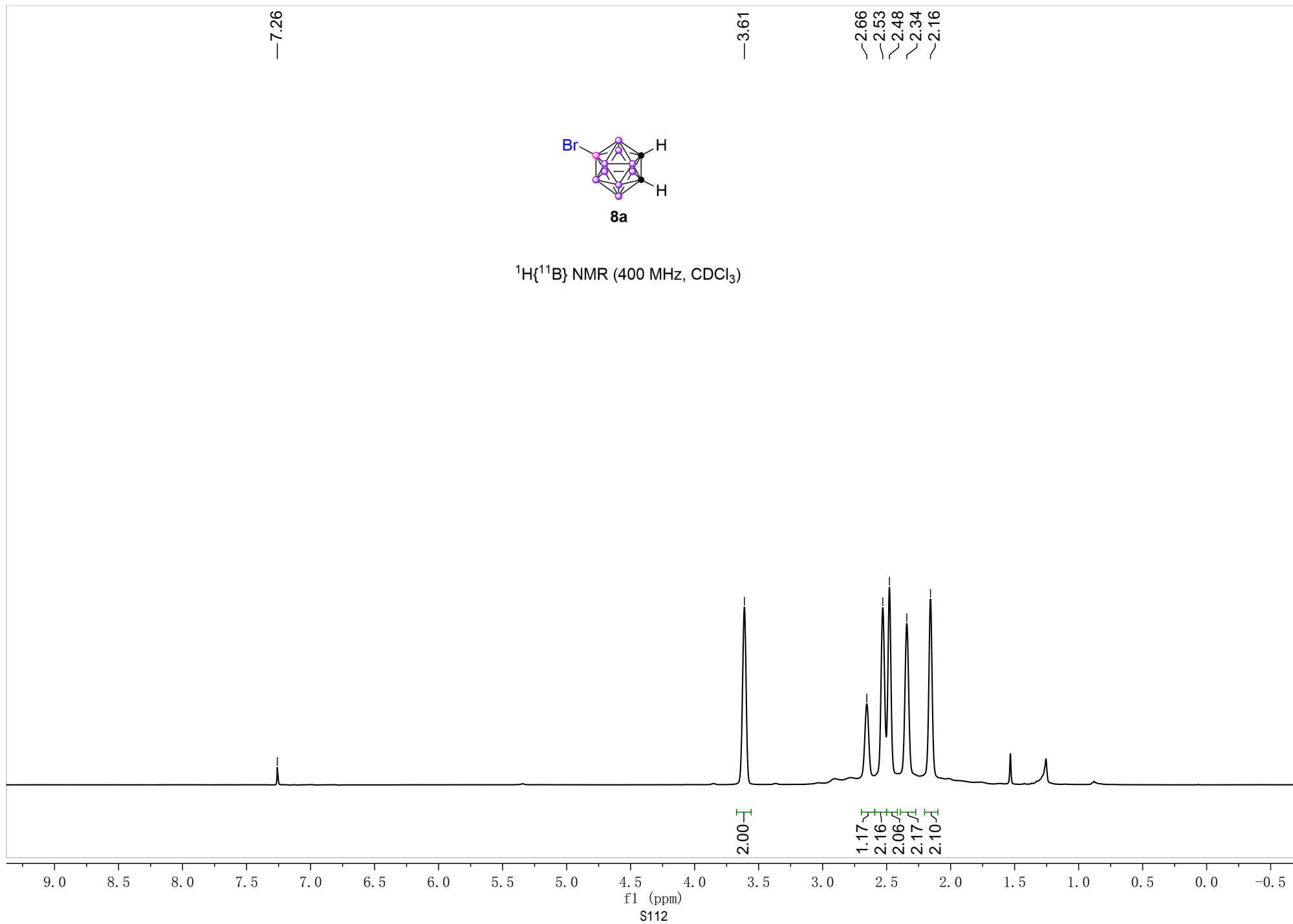
—7.26

—3.62





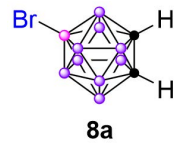
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



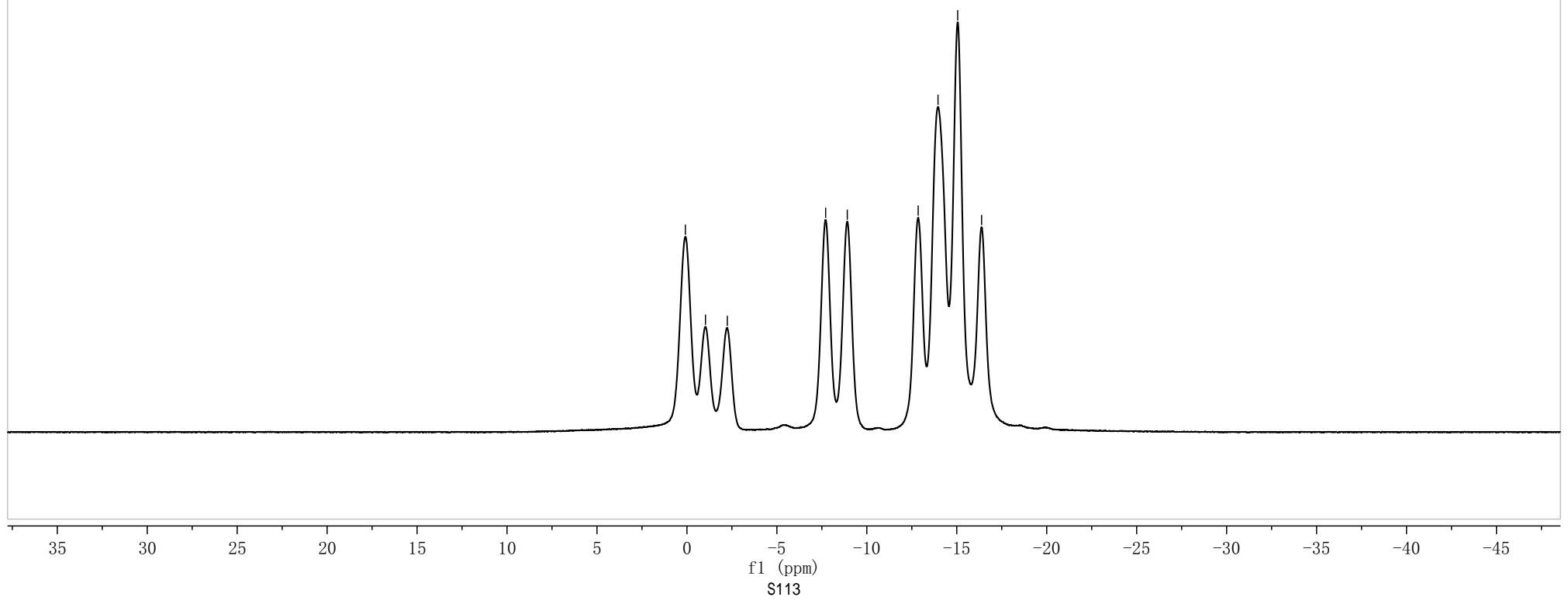
-0.09
-1.03
-2.24

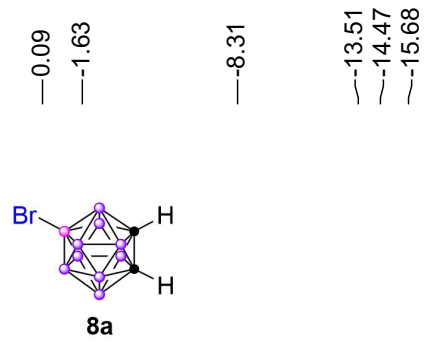
-7.71
-8.91

-12.85
-13.96
-15.05
-16.38

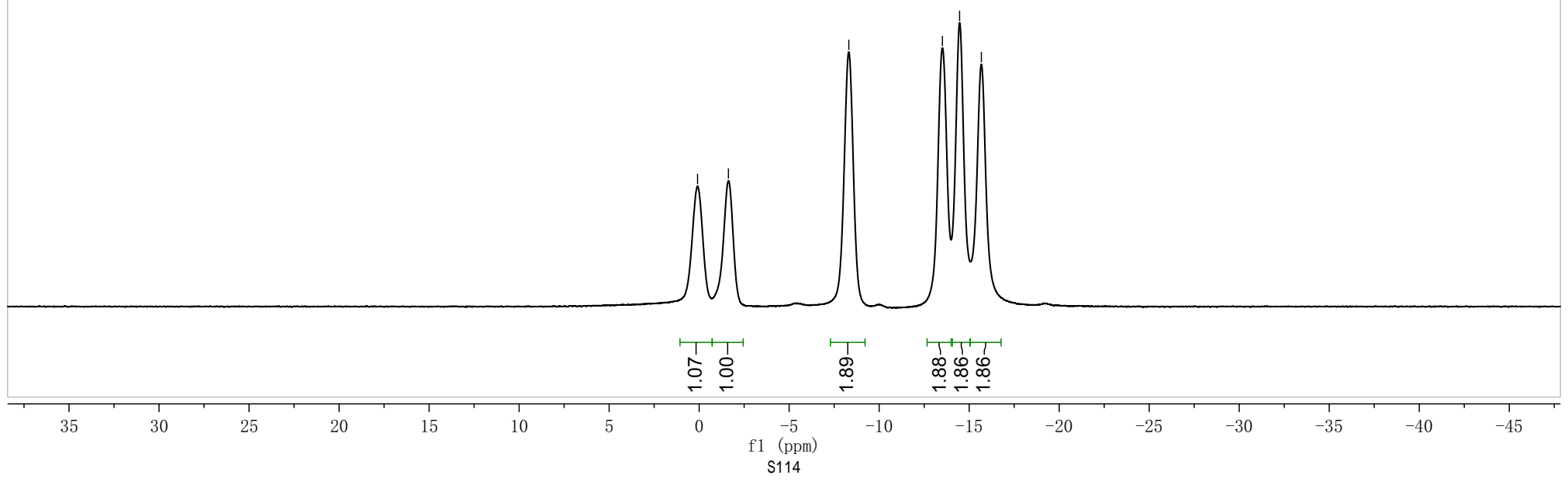


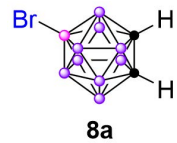
^{11}B NMR (128 MHz, CDCl_3)





¹¹B{¹H} NMR (128 MHz, CDCl₃)

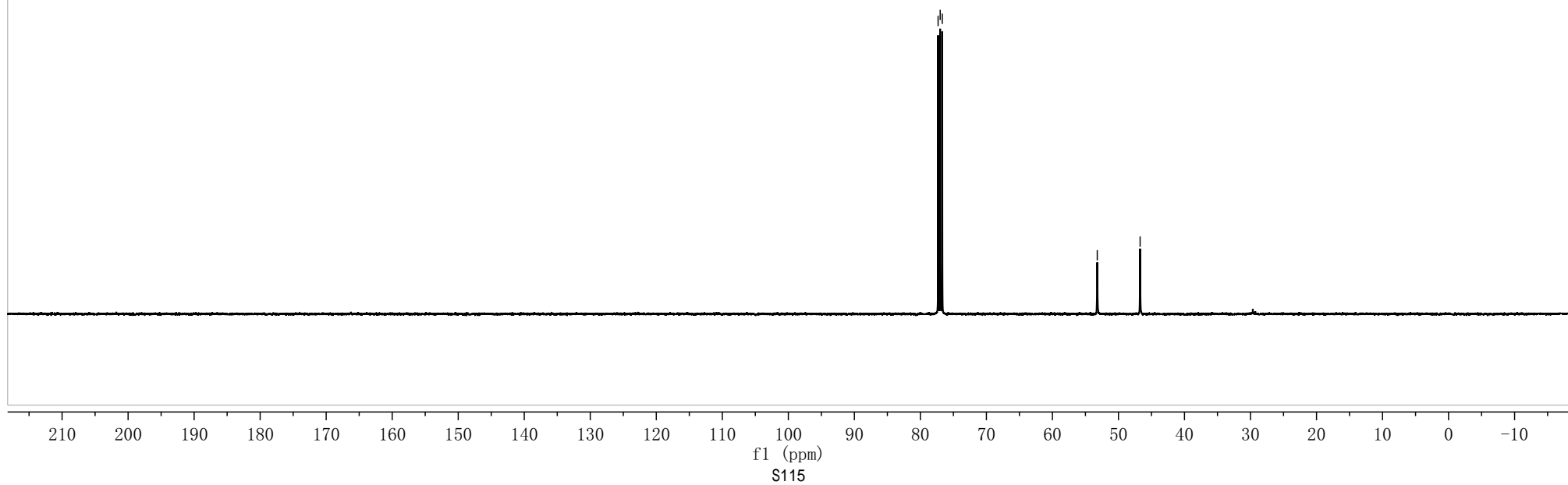


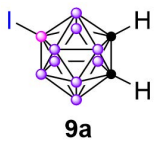


^{13}C NMR (101 MHz, CDCl_3)

77.32
77.00
76.68

53.21
46.72



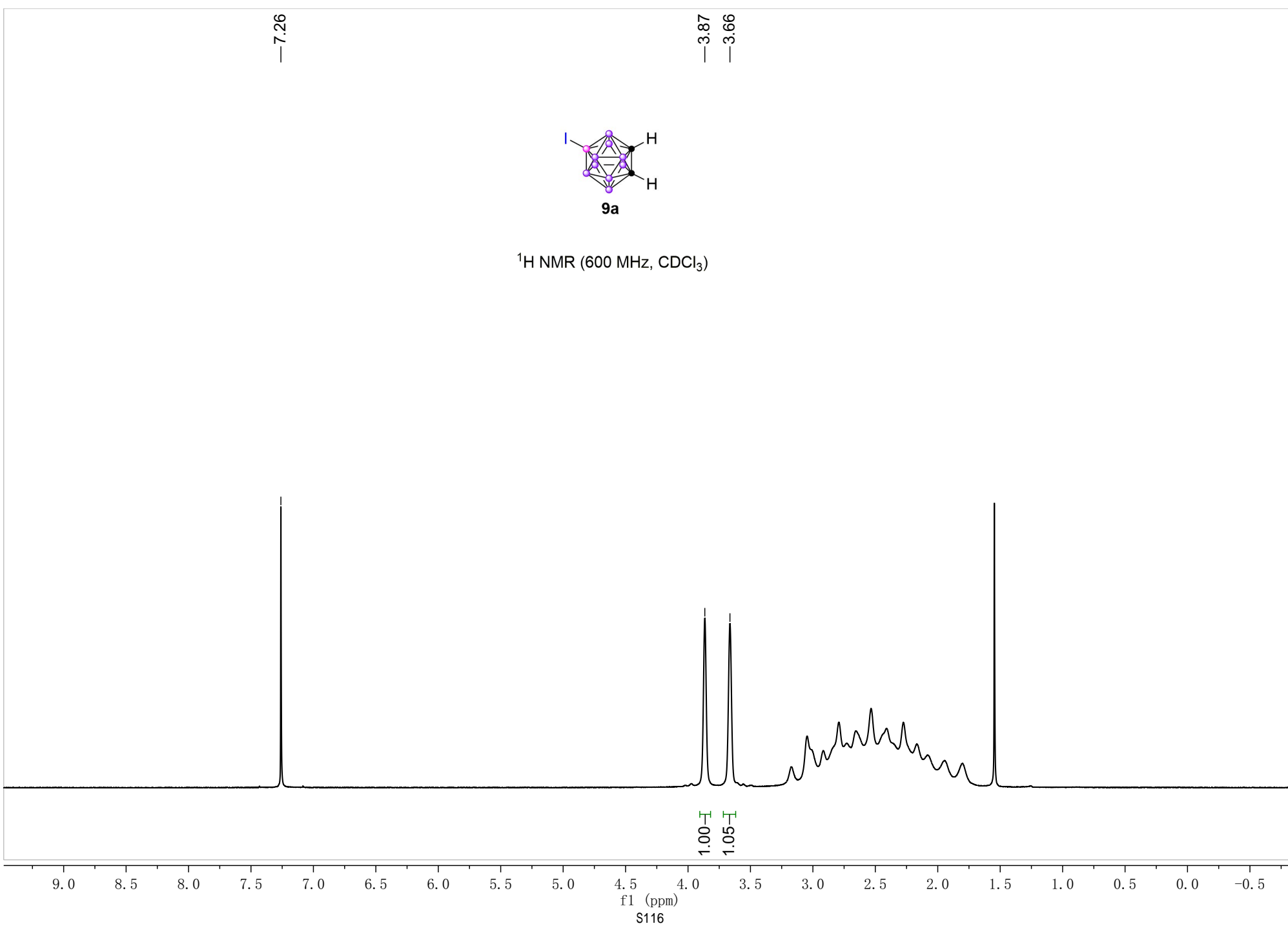


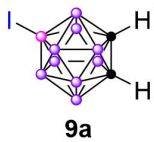
^1H NMR (600 MHz, CDCl_3)

—7.26

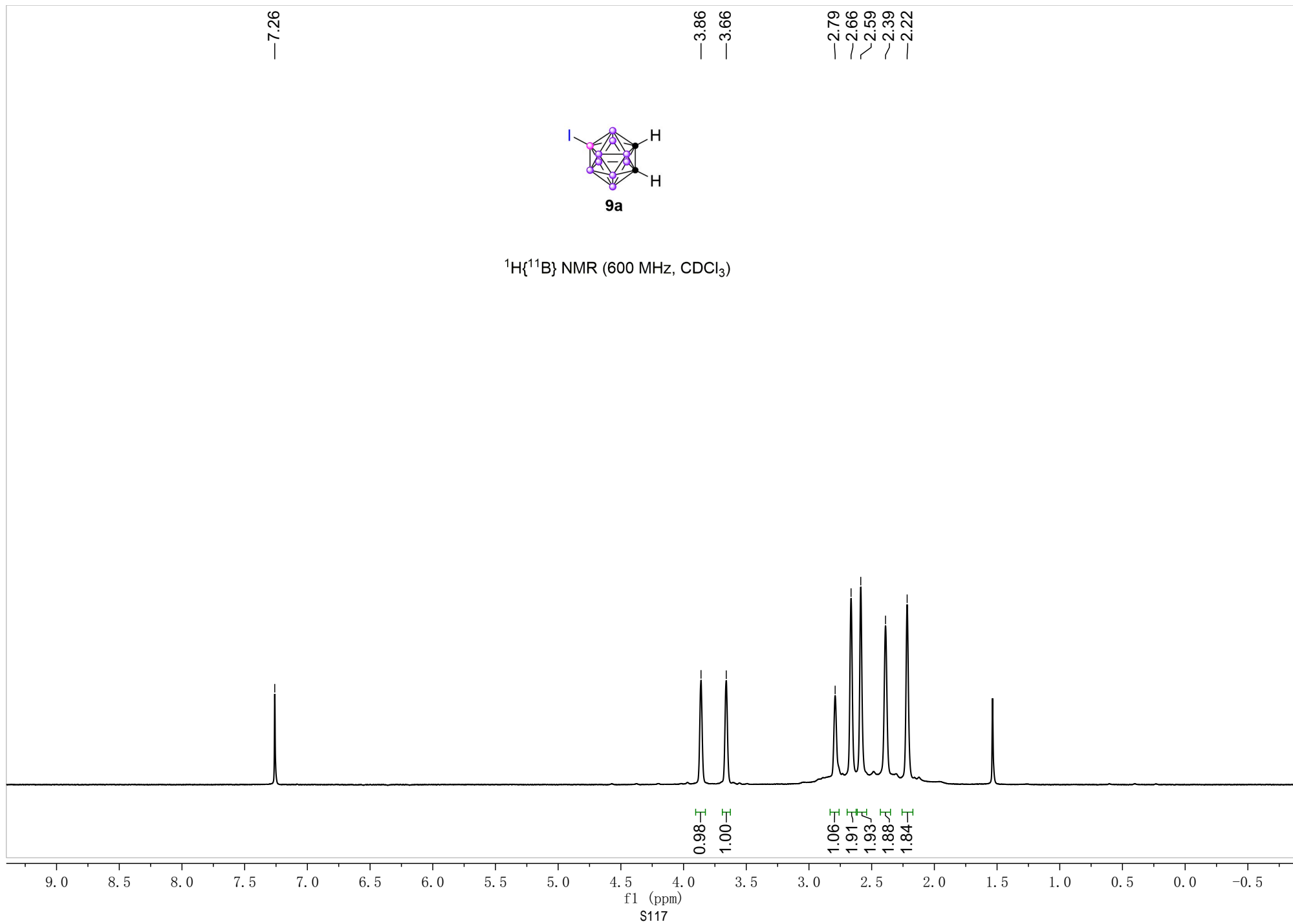
—3.87

—3.66

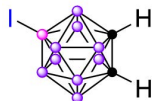




$^1\text{H}\{^1\text{B}\}$ NMR (600 MHz, CDCl_3)

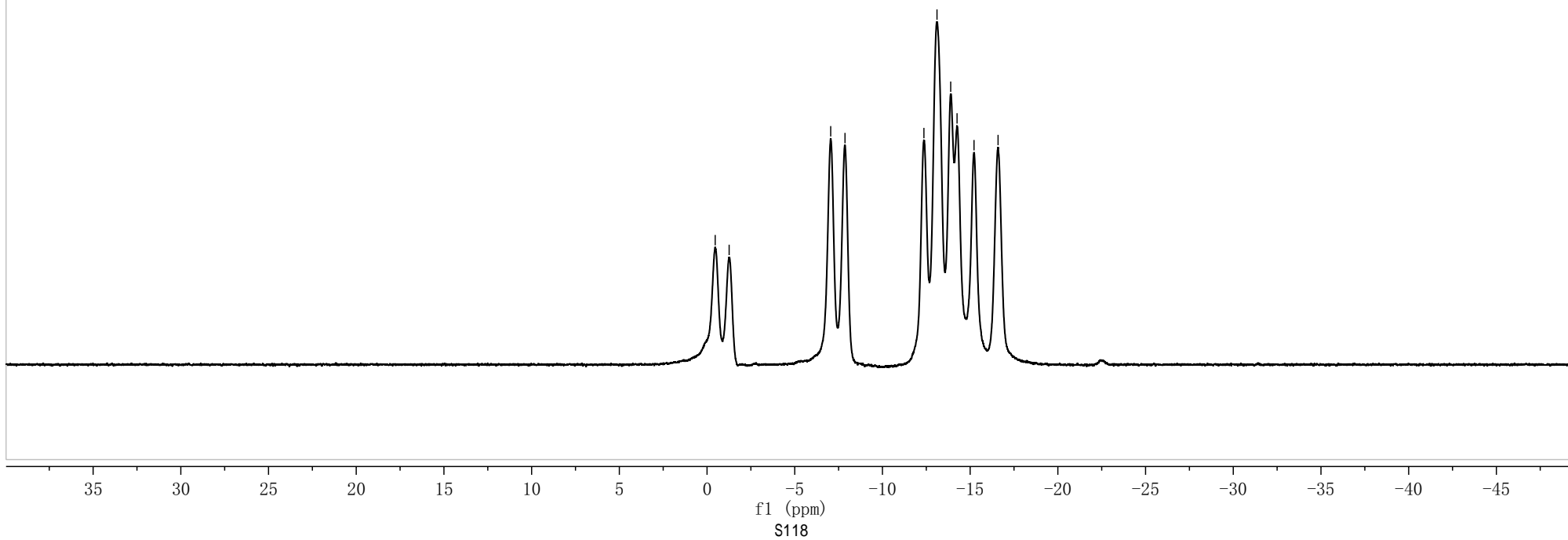


-0.46
-1.26
-7.05
-7.87
-12.36
-13.11
-13.89
-14.25
-15.22
-16.59

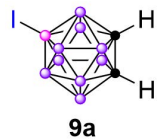


9a

^{11}B NMR (193 MHz, CDCl_3)



---0.87
---7.45
---12.80
---13.44
---14.75
---16.57

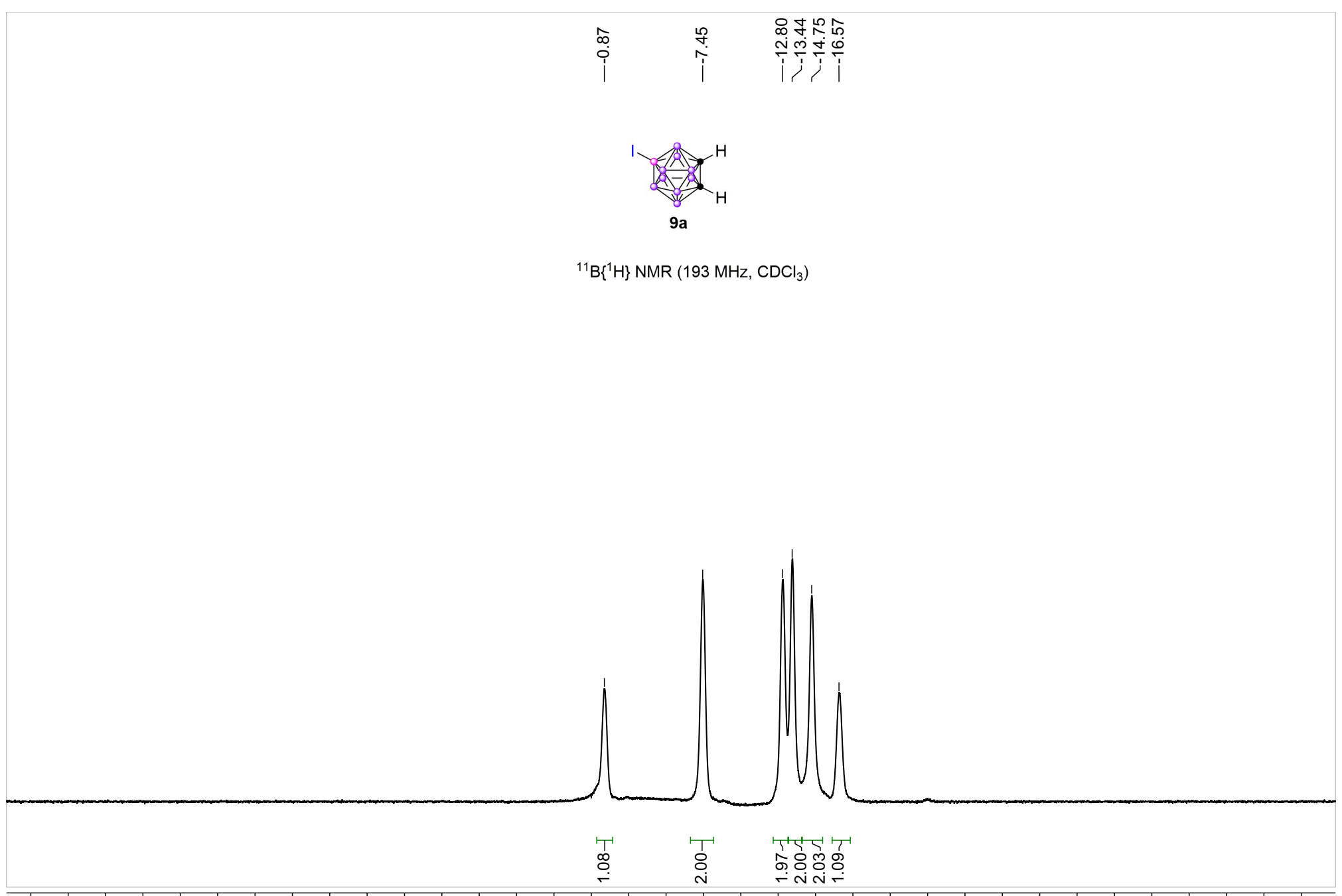


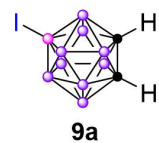
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

1.08
2.00
1.97
2.00
2.03
1.09

f1 (ppm)
S119

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45

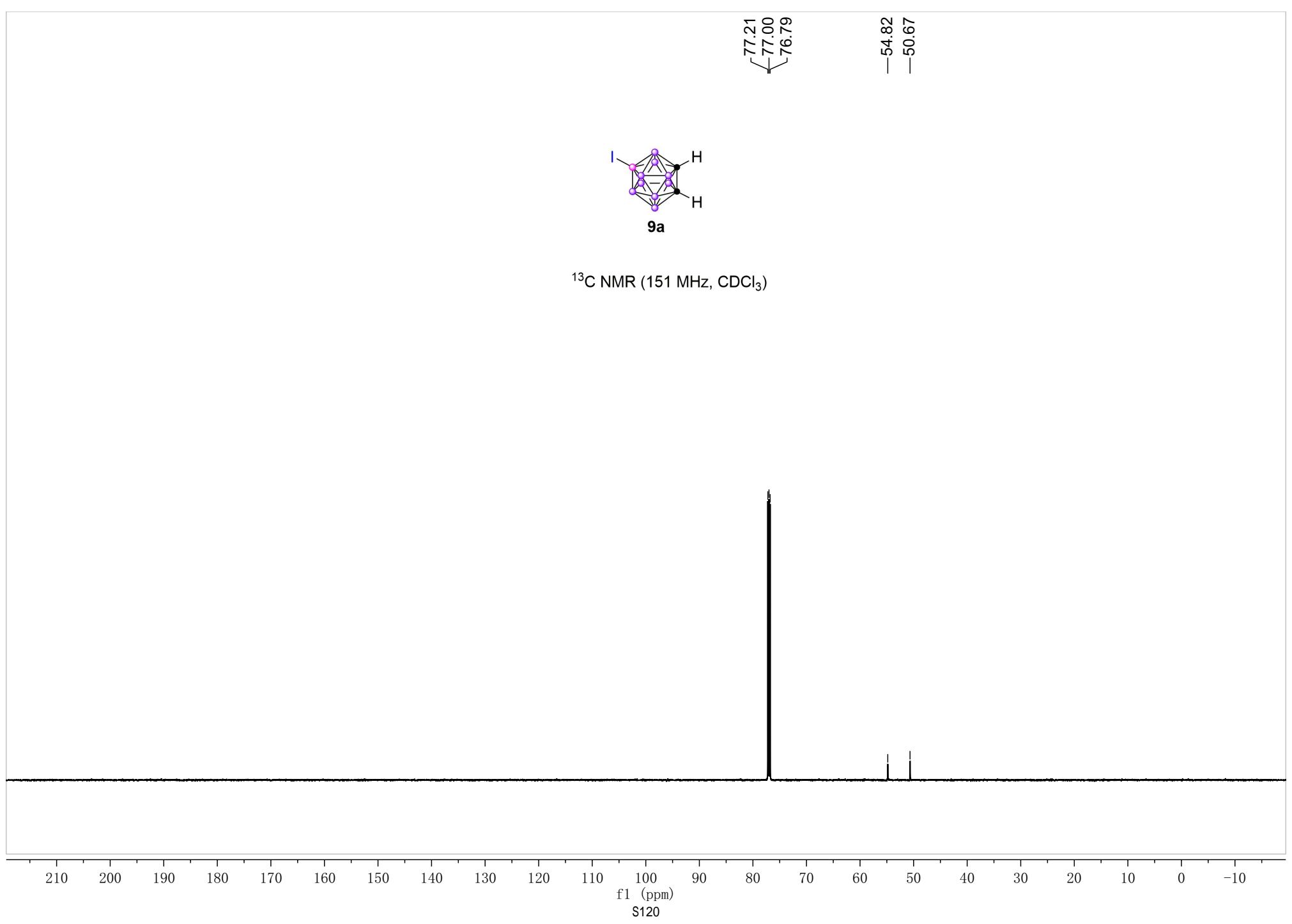


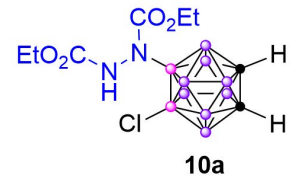


^{13}C NMR (151 MHz, CDCl_3)

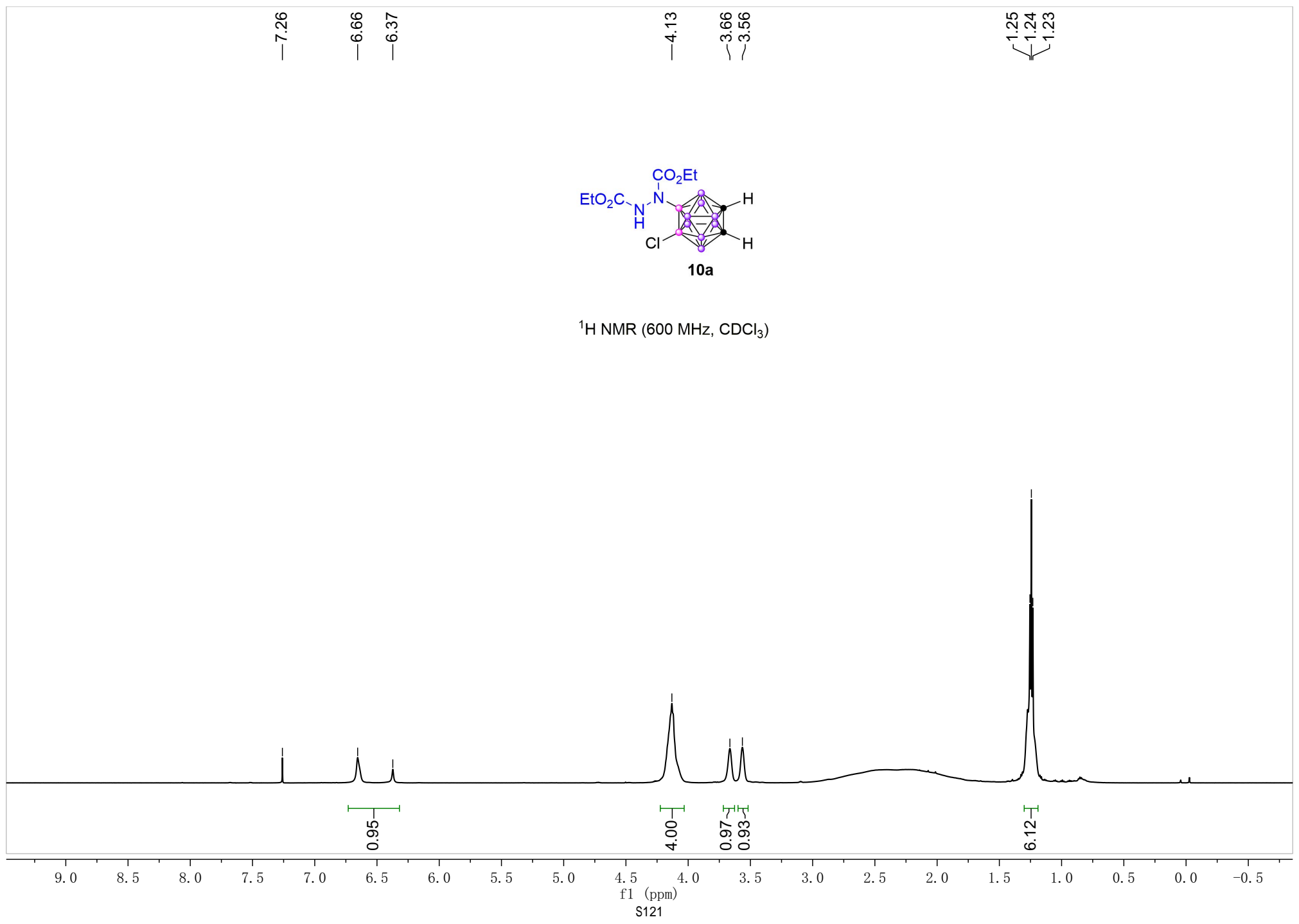
77.21
77.00
76.79

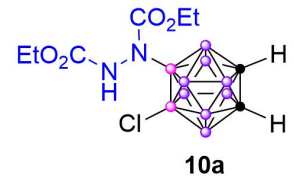
54.82
50.67



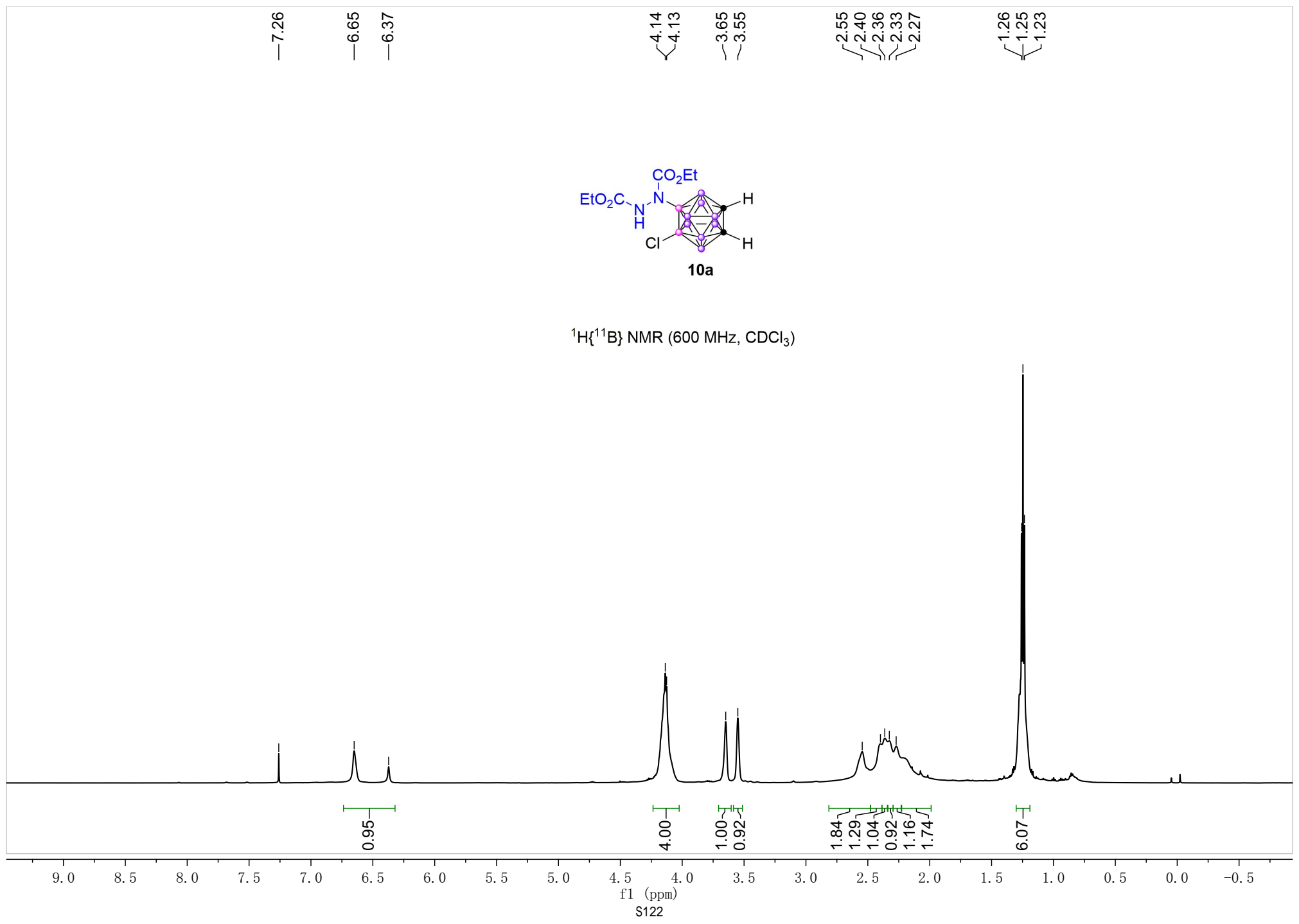


¹H NMR (600 MHz, CDCl₃)





$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)



—7.26

—6.65

—6.37

4.14

4.13

3.65

3.55

2.55

2.40

2.36

2.33

2.27

1.26

1.25

1.23

0.95

4.00

1.00

0.92

1.84

1.29

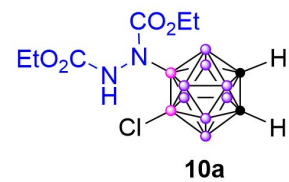
1.04

0.92

1.16

1.74

6.07

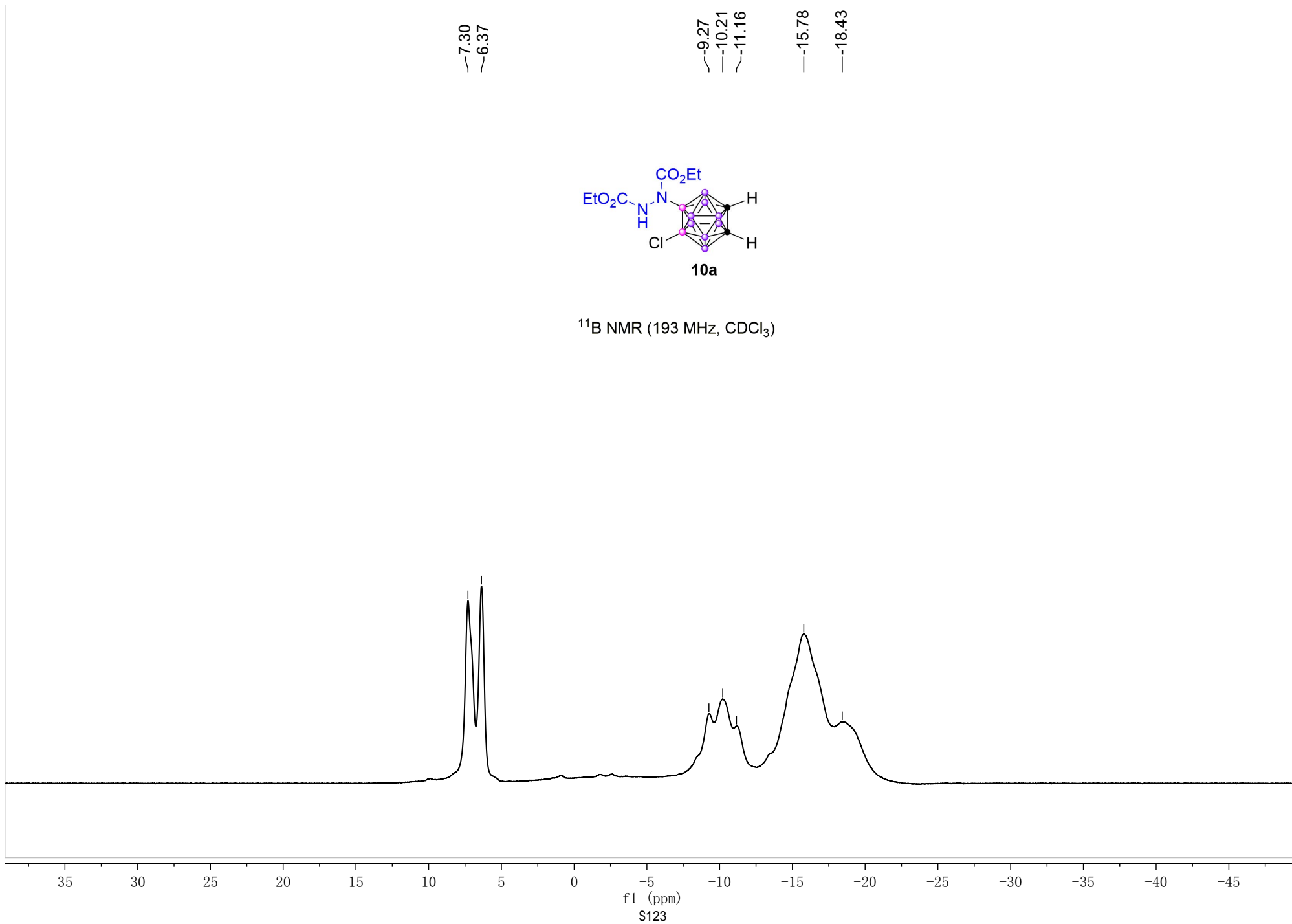


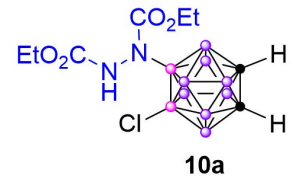
^{11}B NMR (193 MHz, CDCl_3)

7.30
6.37

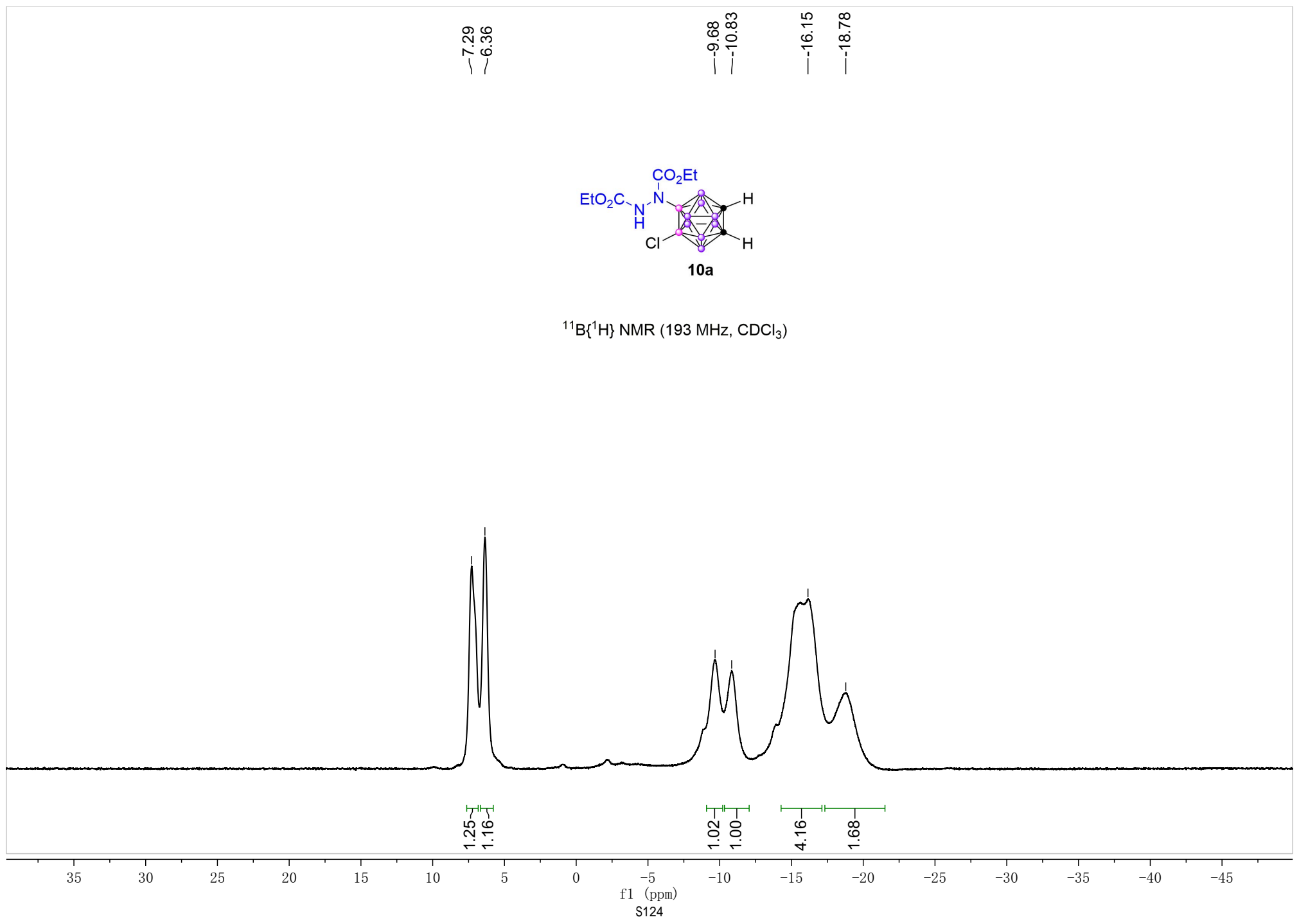
9.27
10.21
11.16

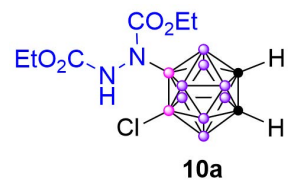
15.78
18.43





$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)





¹³C NMR (151 MHz, CDCl₃)

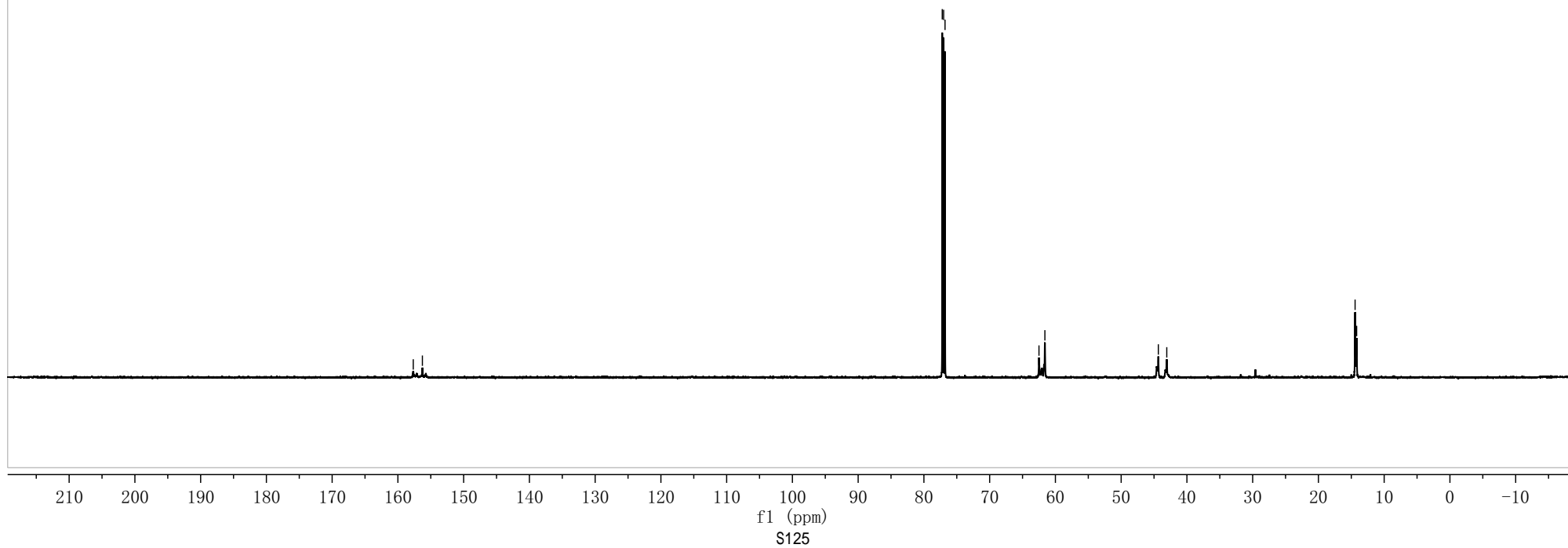
157.66
156.27

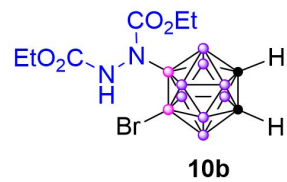
77.21
77.00
76.79

62.50
61.60

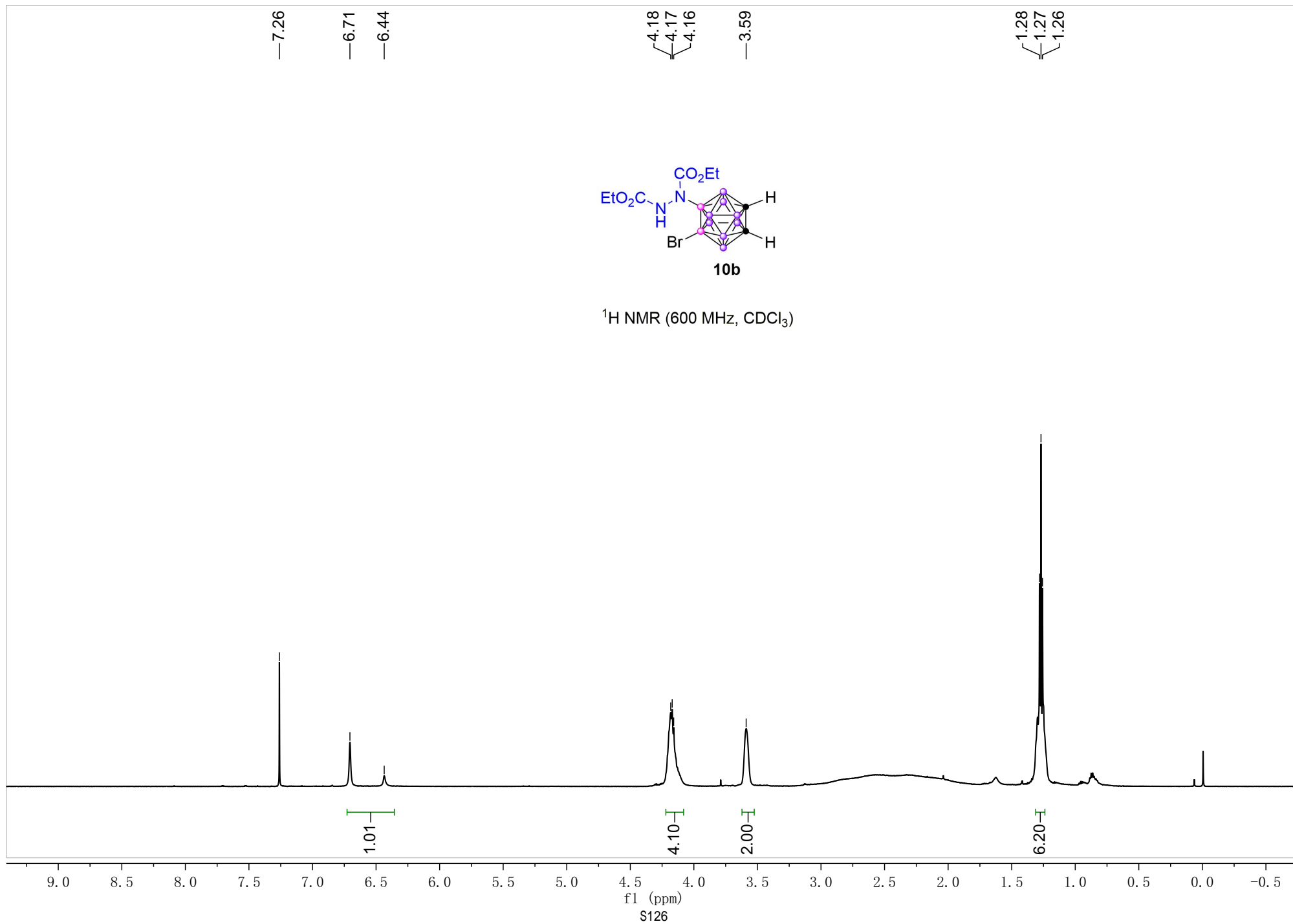
44.35
43.06

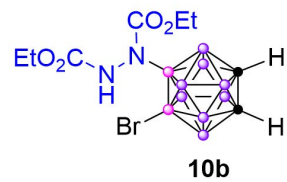
14.42
14.18



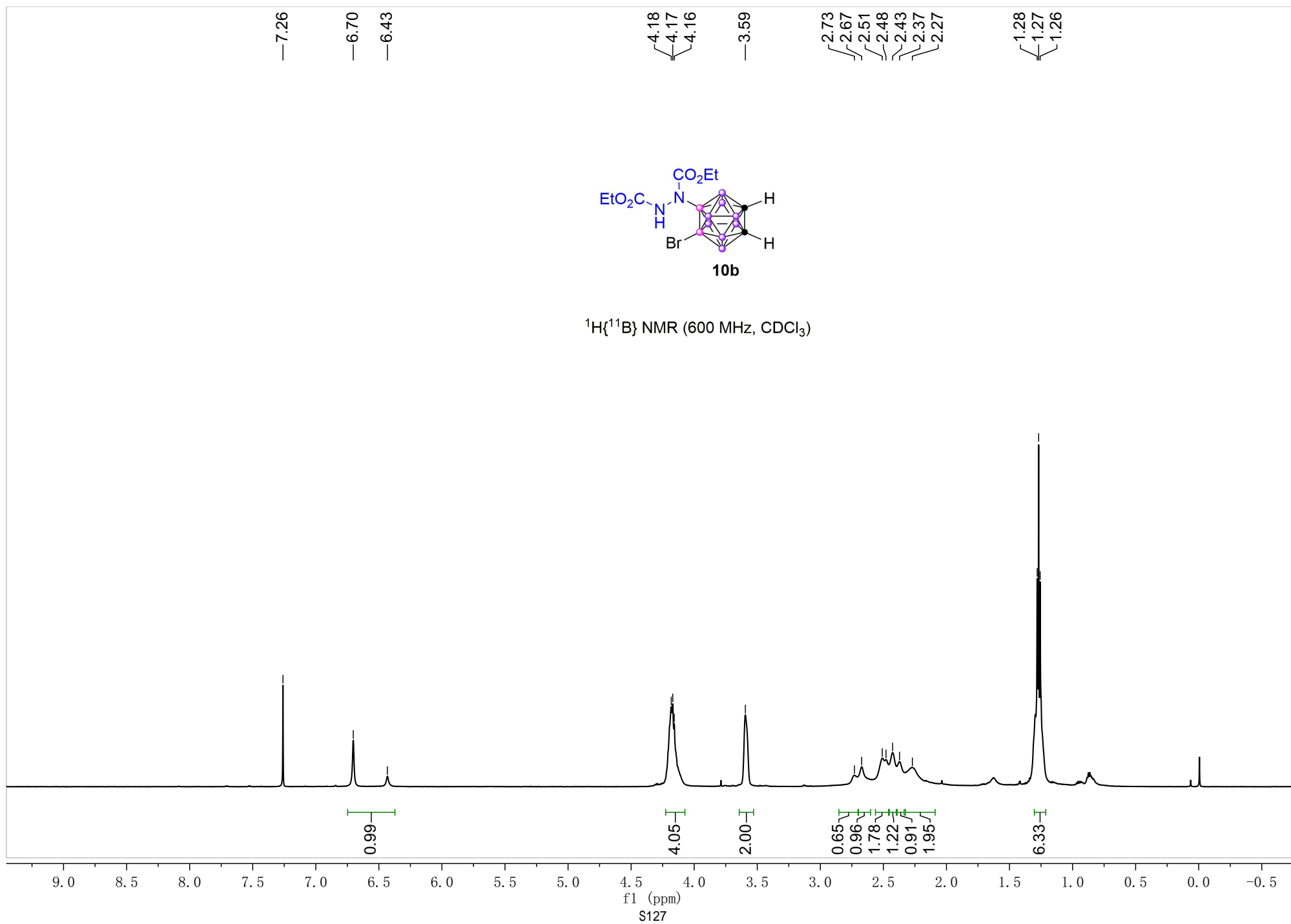


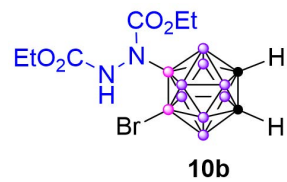
¹H NMR (600 MHz, CDCl₃)



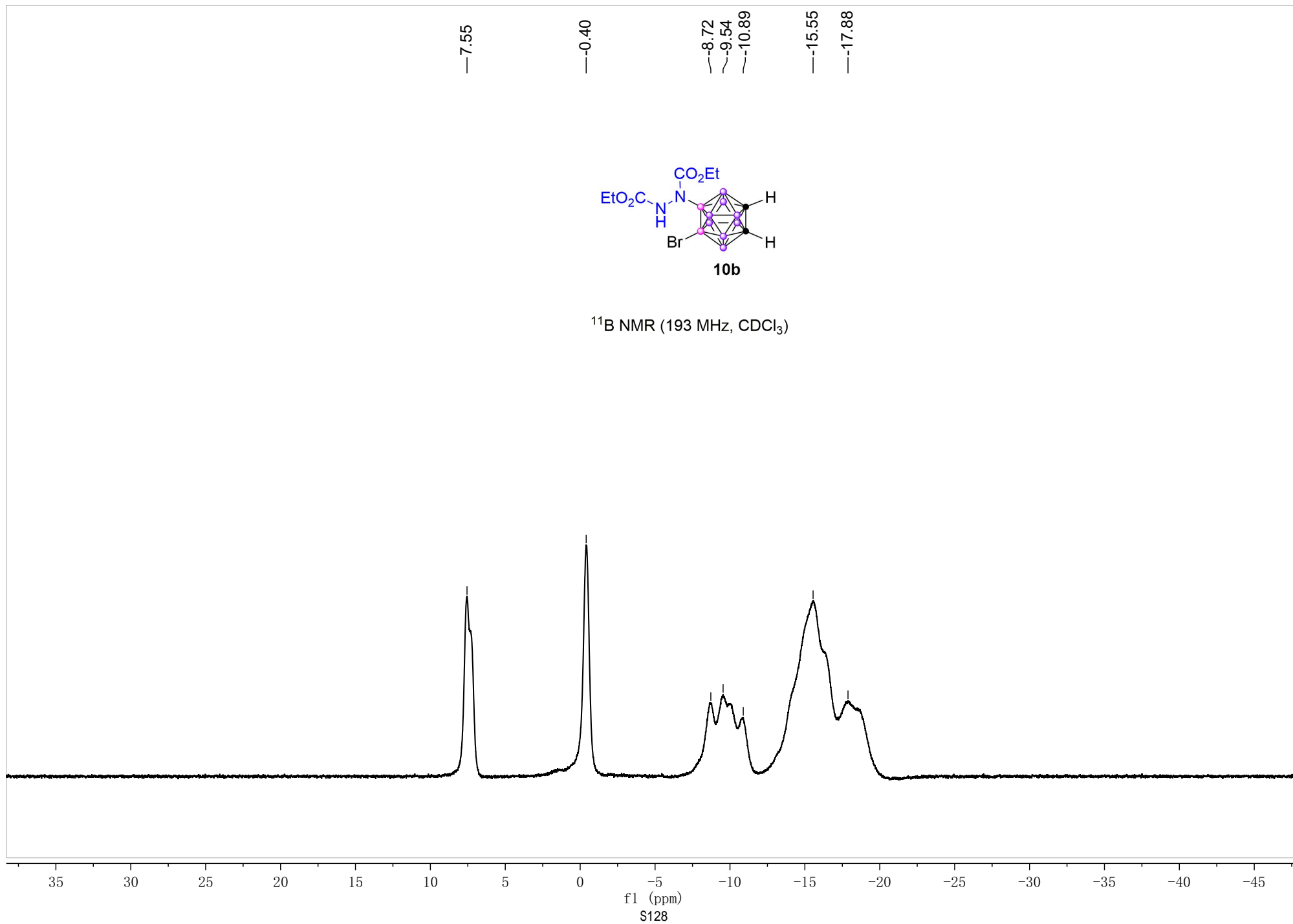


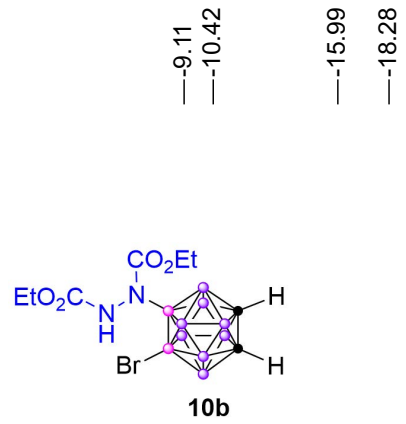
¹H{¹¹B} NMR (600 MHz, CDCl₃)



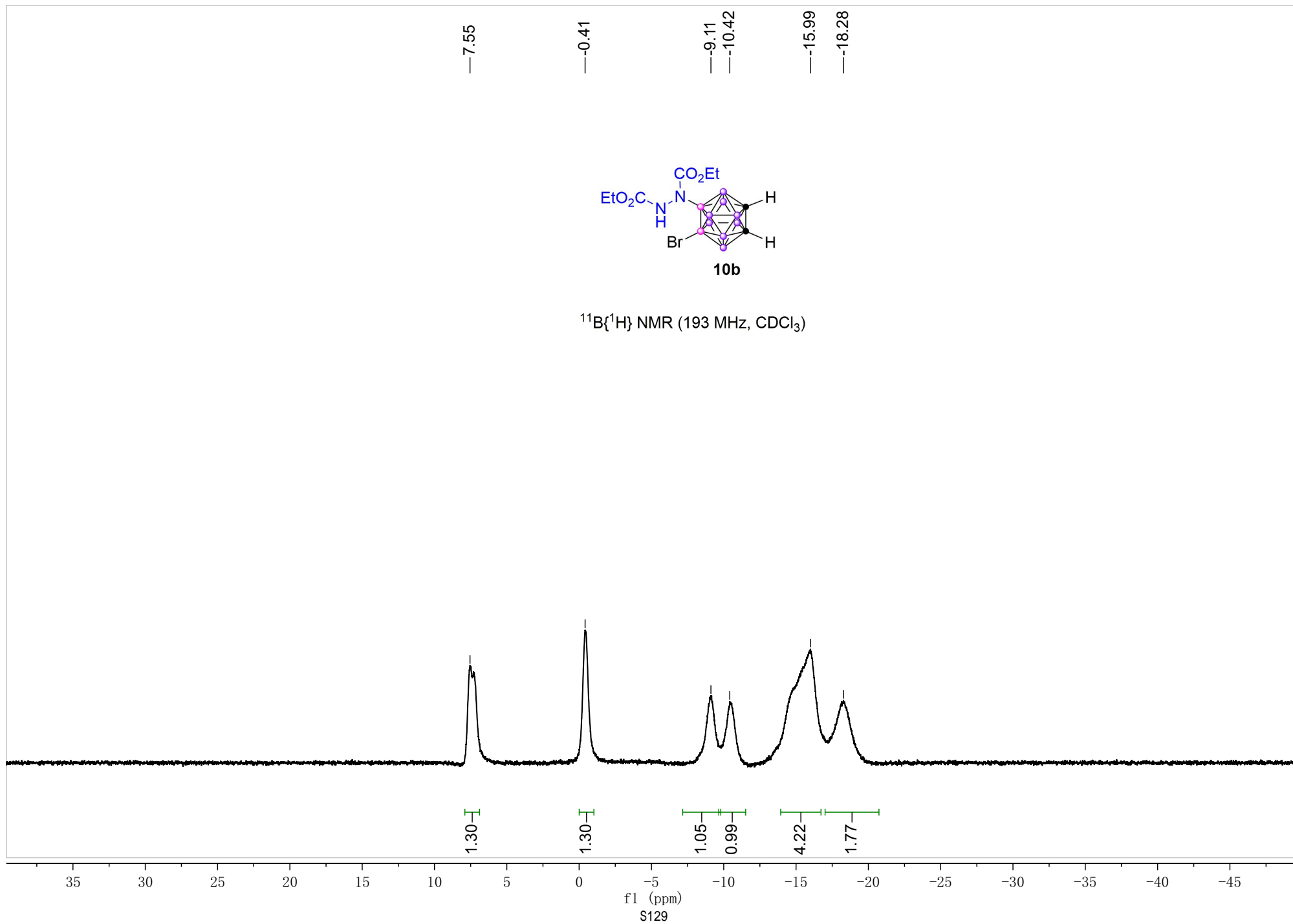


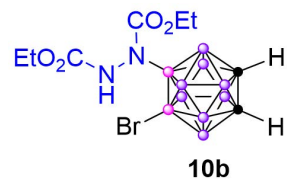
^{11}B NMR (193 MHz, CDCl_3)





¹¹B{¹H} NMR (193 MHz, CDCl₃)





^{13}C NMR (151 MHz, CDCl_3)

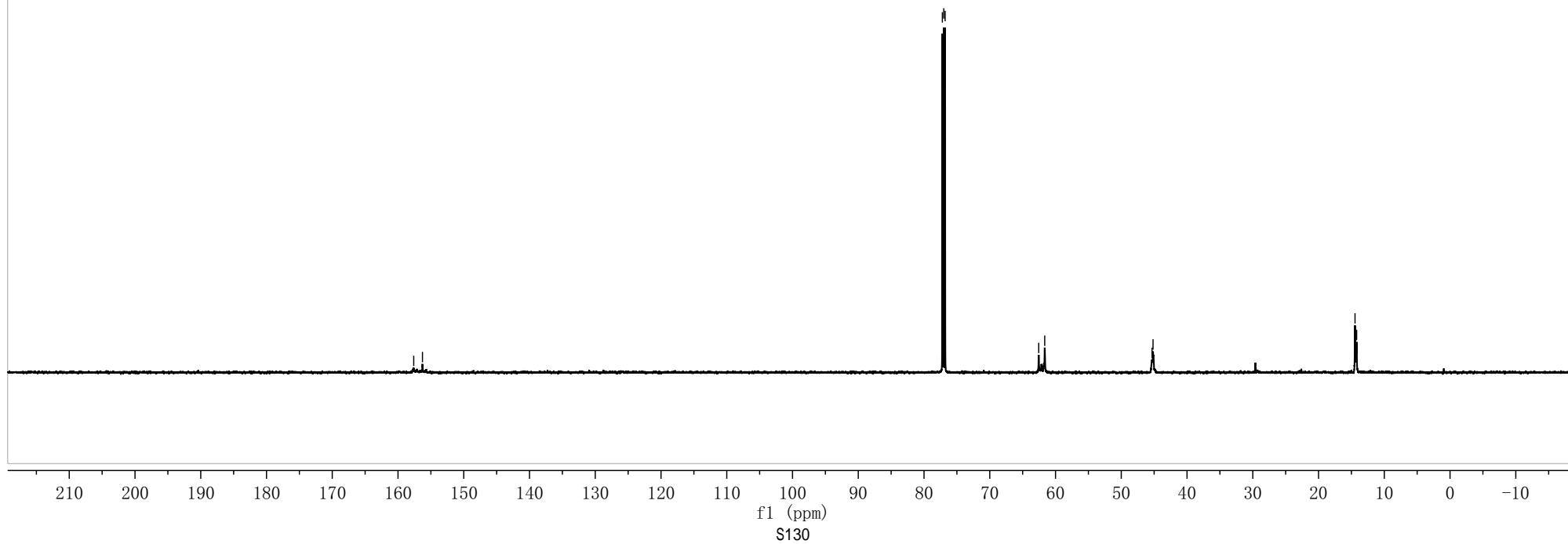
157.61
156.28

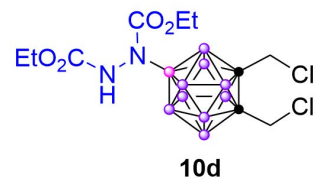
77.21
77.00
76.79

62.56
61.64

45.32
45.17

14.45
14.20





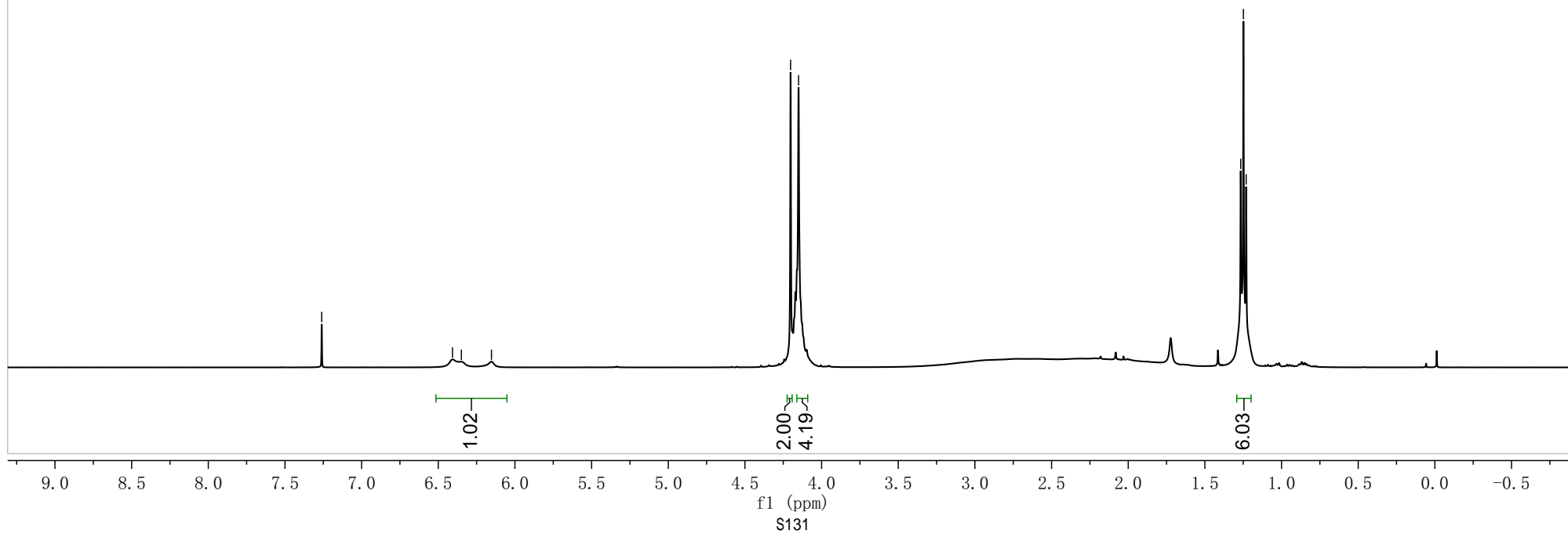
¹H NMR (400 MHz, CDCl₃)

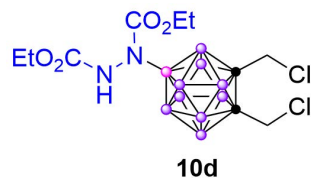
7.26

6.41
6.35
6.15

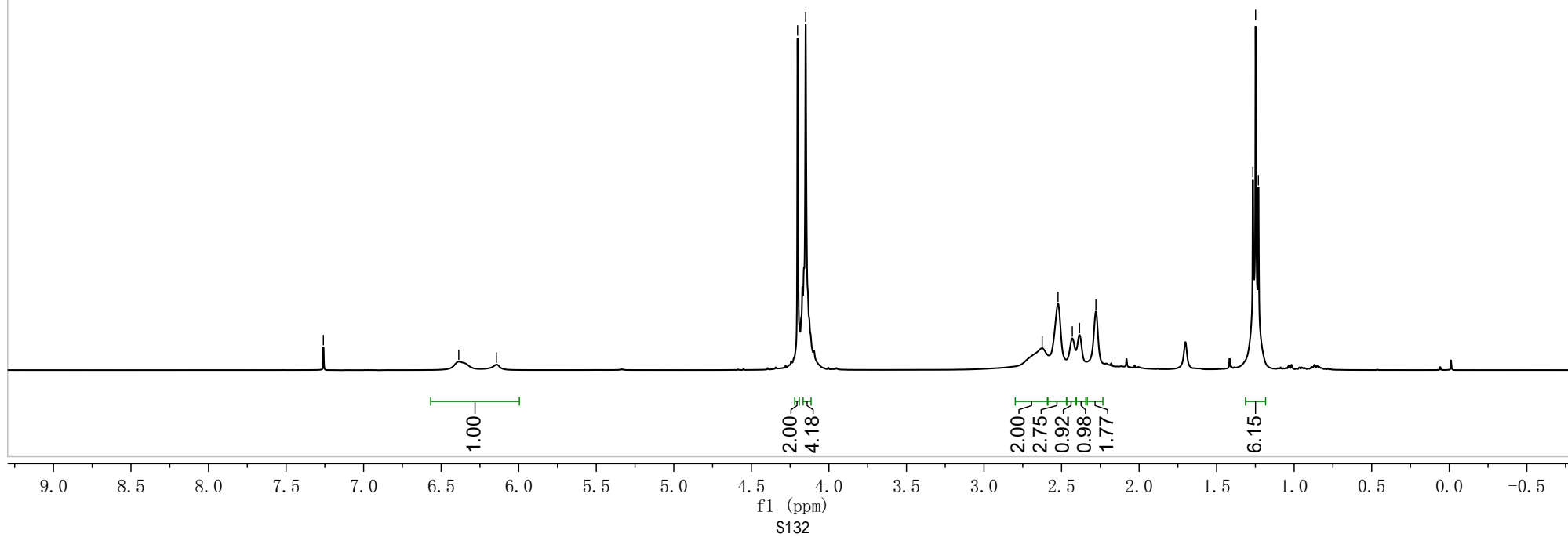
4.20
4.15

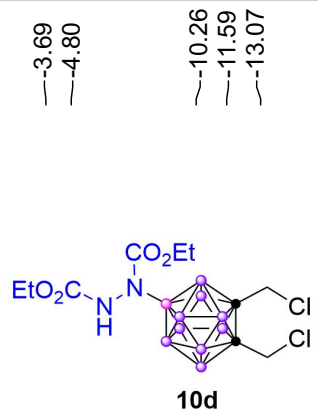
1.27
1.25
1.23





$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



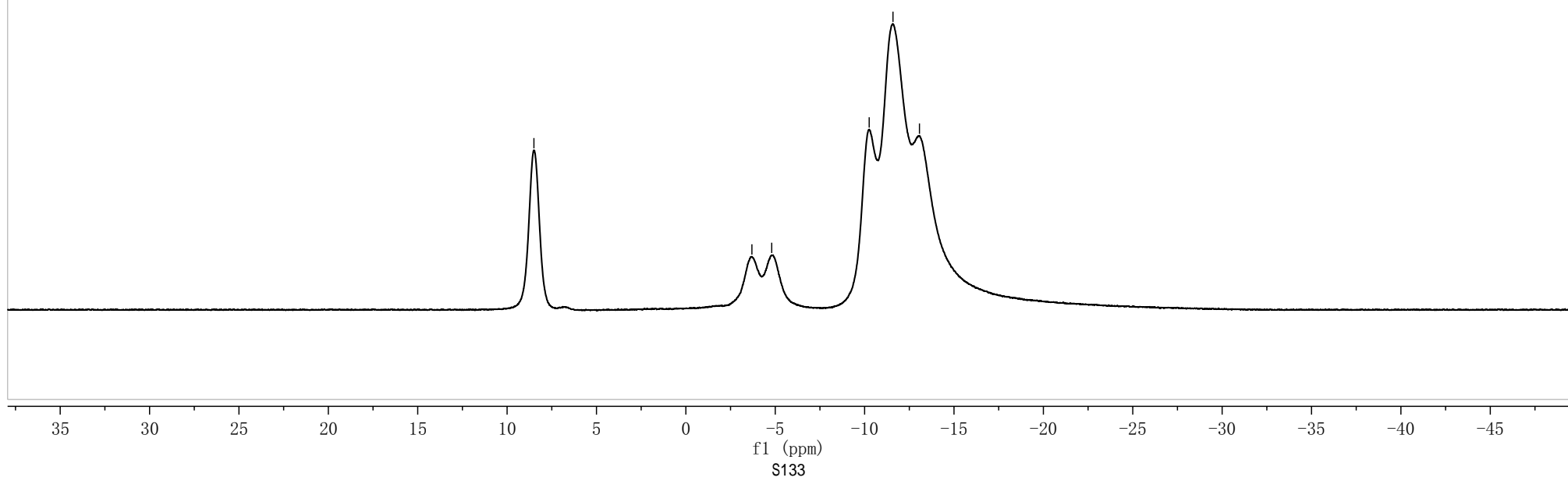


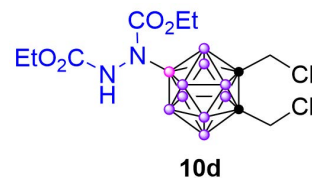
¹¹B NMR (128 MHz, CDCl₃)

—8.50

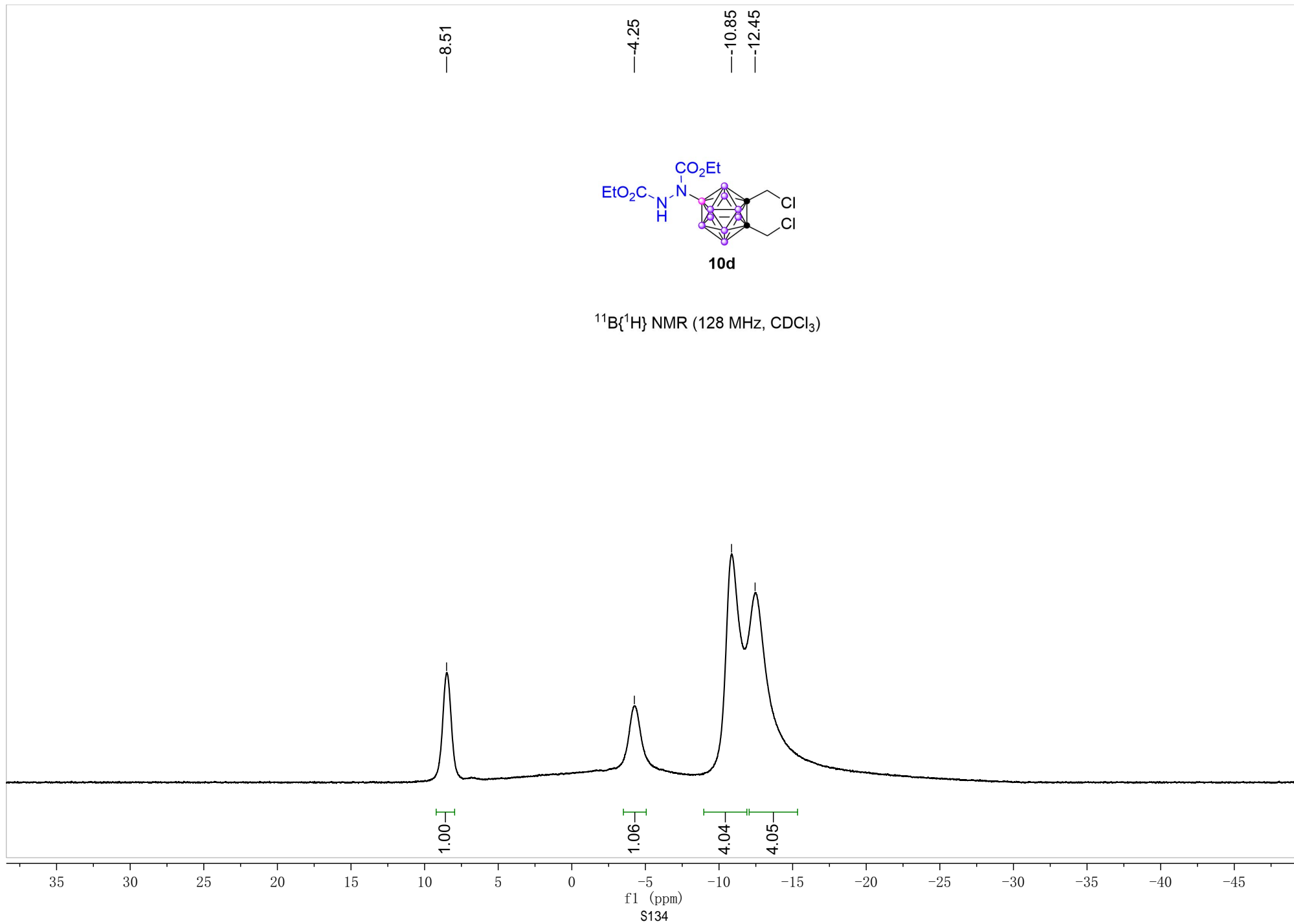
—3.69
—4.80

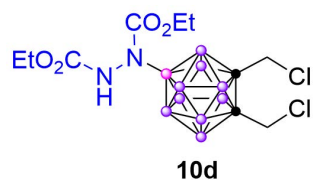
—10.26
—11.59
—13.07





$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)





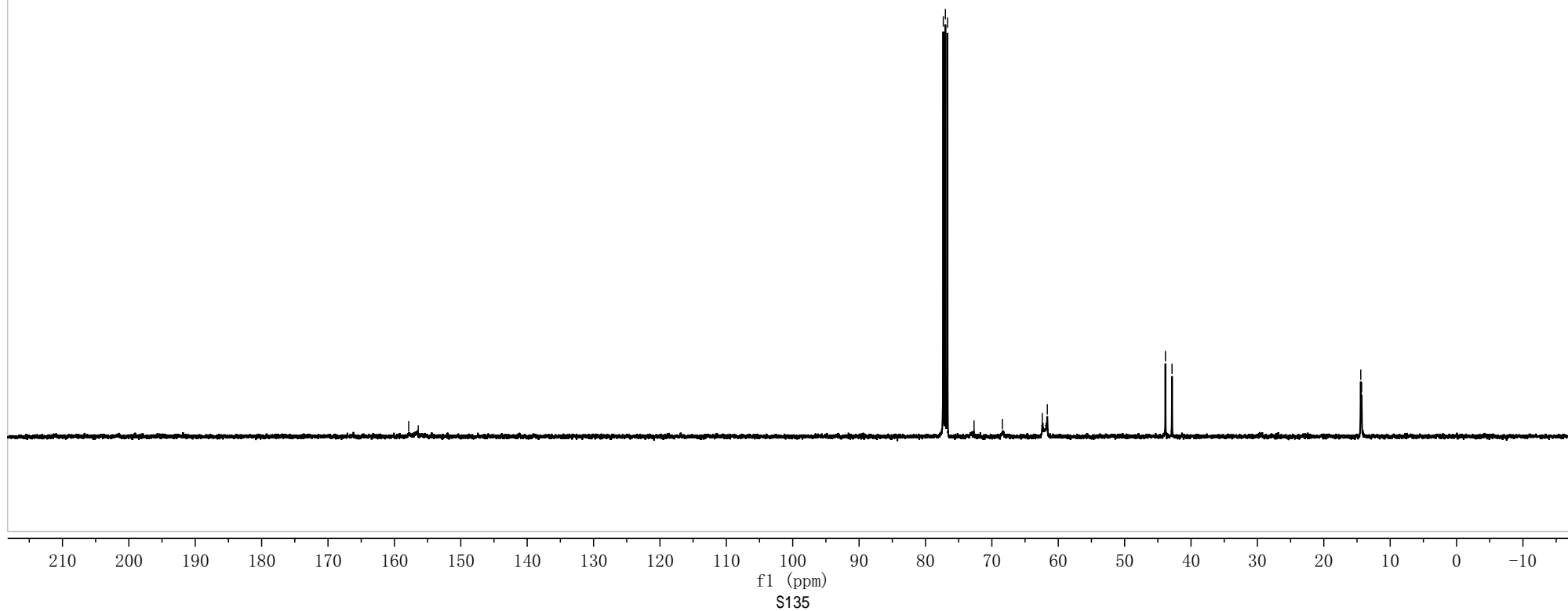
^{13}C NMR (101 MHz, CDCl_3)

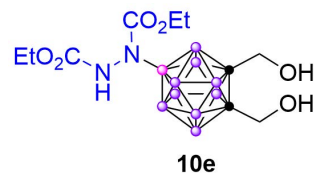
157.85
156.41

77.32
77.00
76.68
72.68
68.41
62.41
61.65

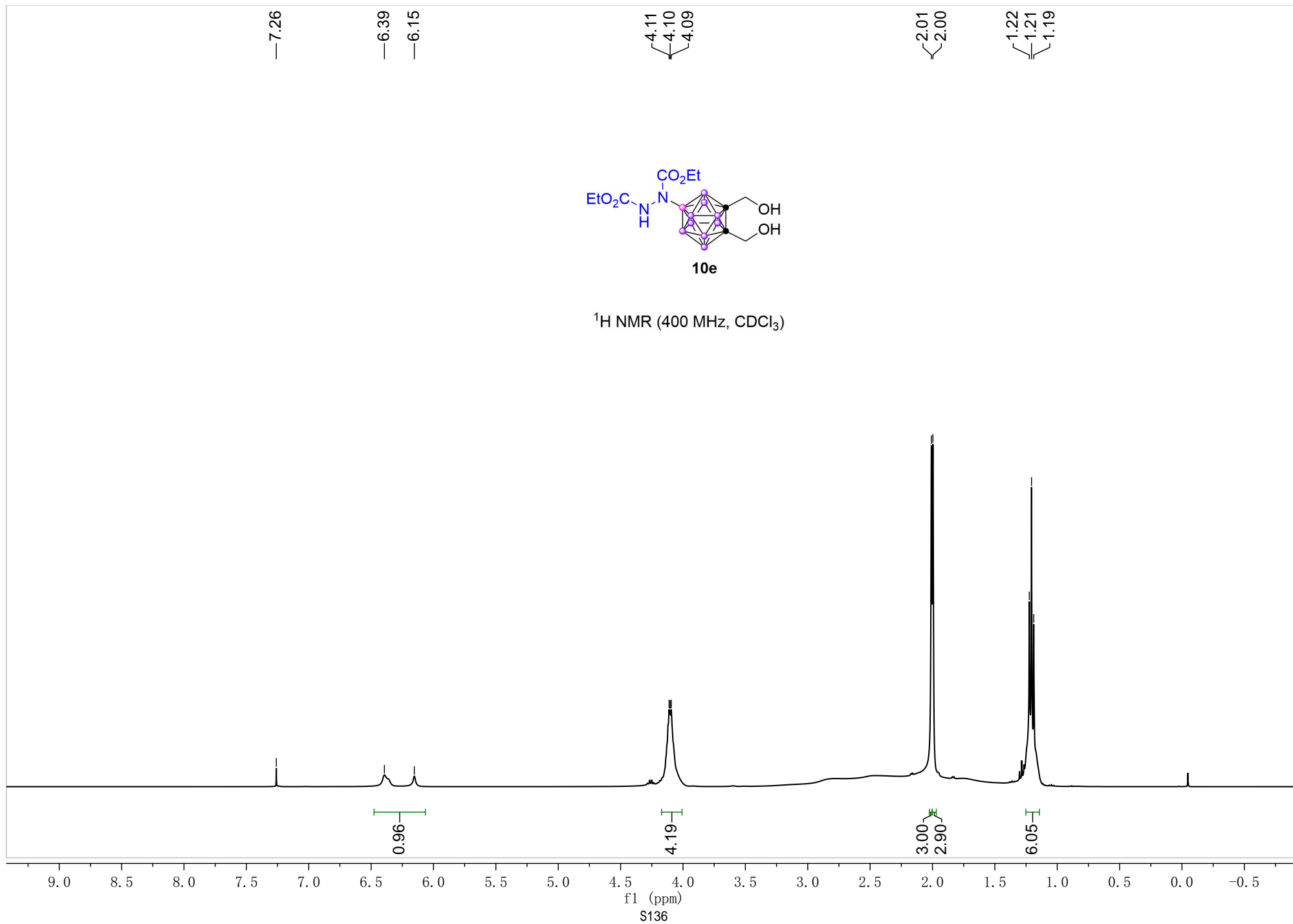
43.85
42.86

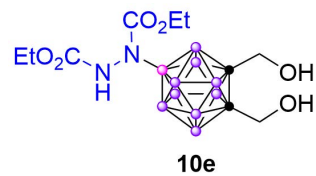
14.43
14.30



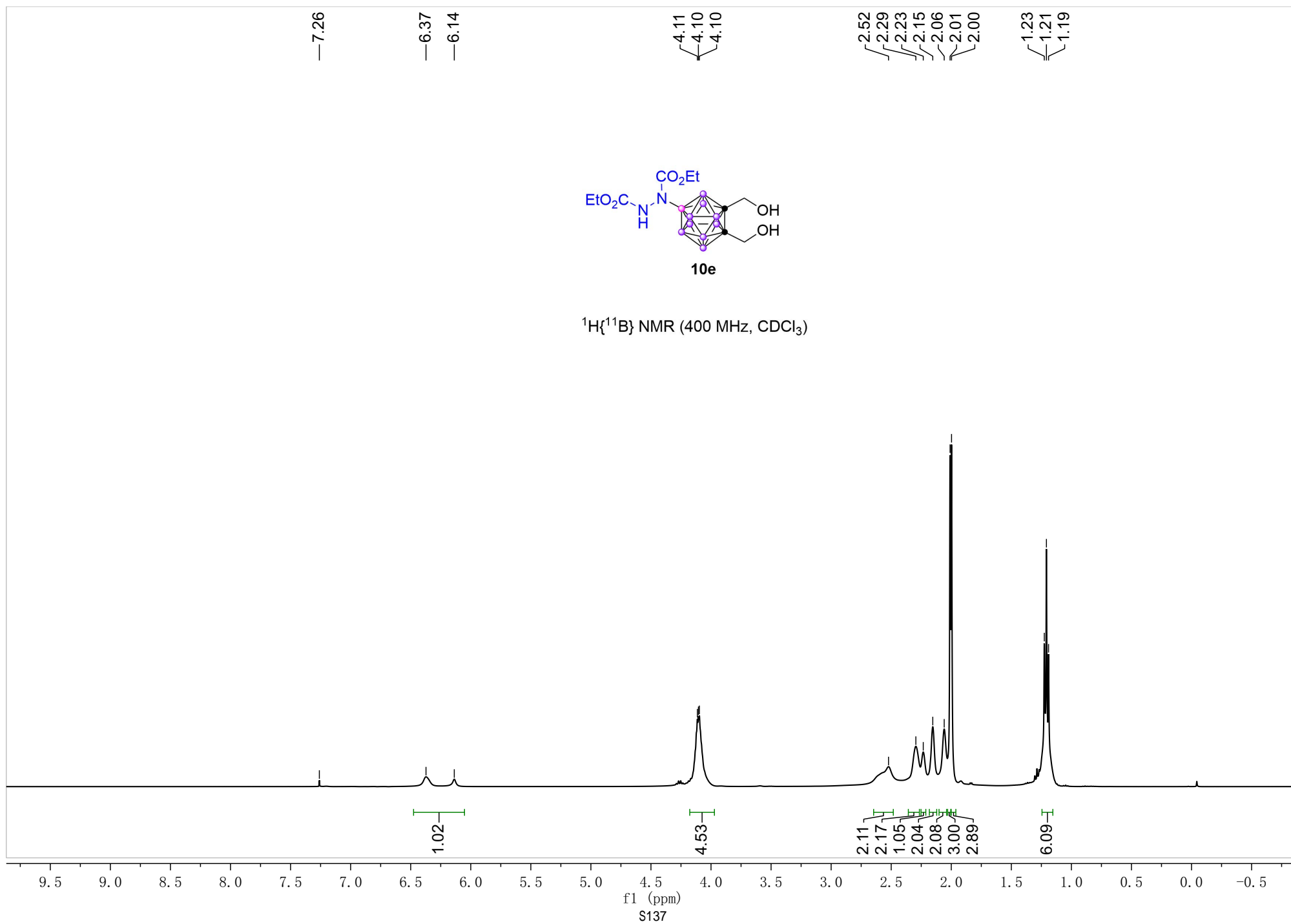


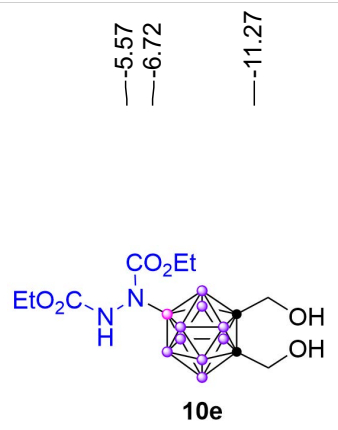
¹H NMR (400 MHz, CDCl₃)





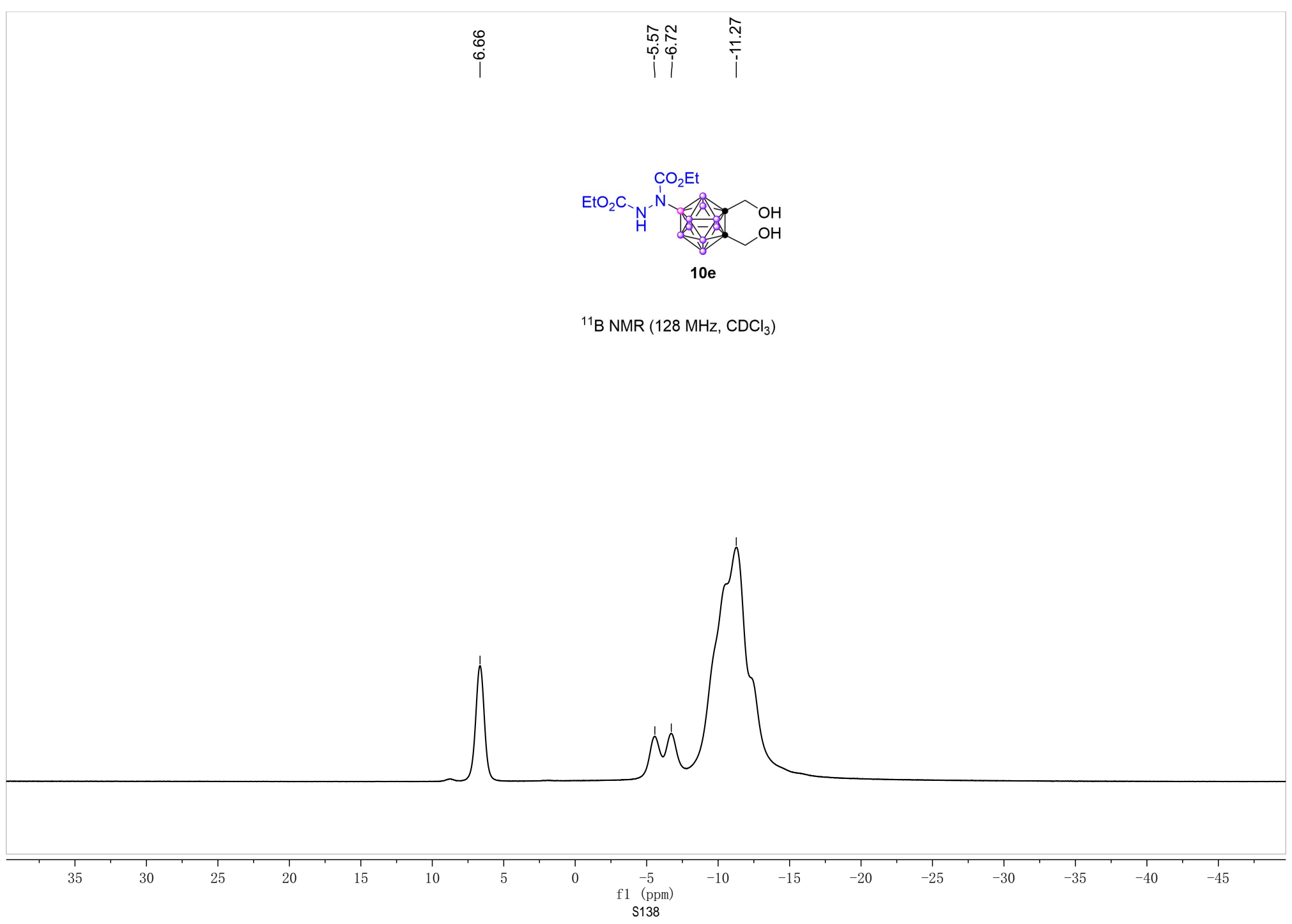
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



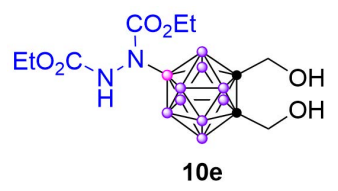


¹¹B NMR (128 MHz, CDCl₃)

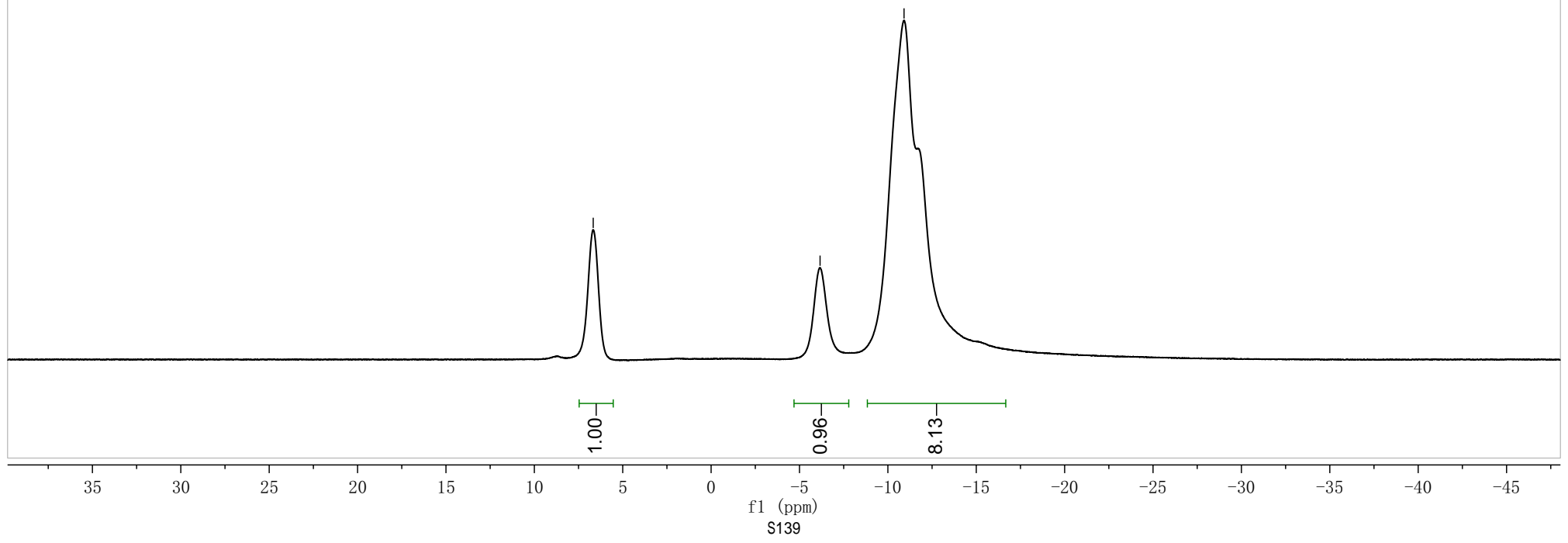
6.66
5.57
6.72
11.27

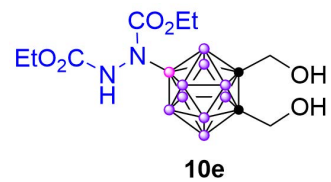


—6.67
—6.16
—10.91



$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)



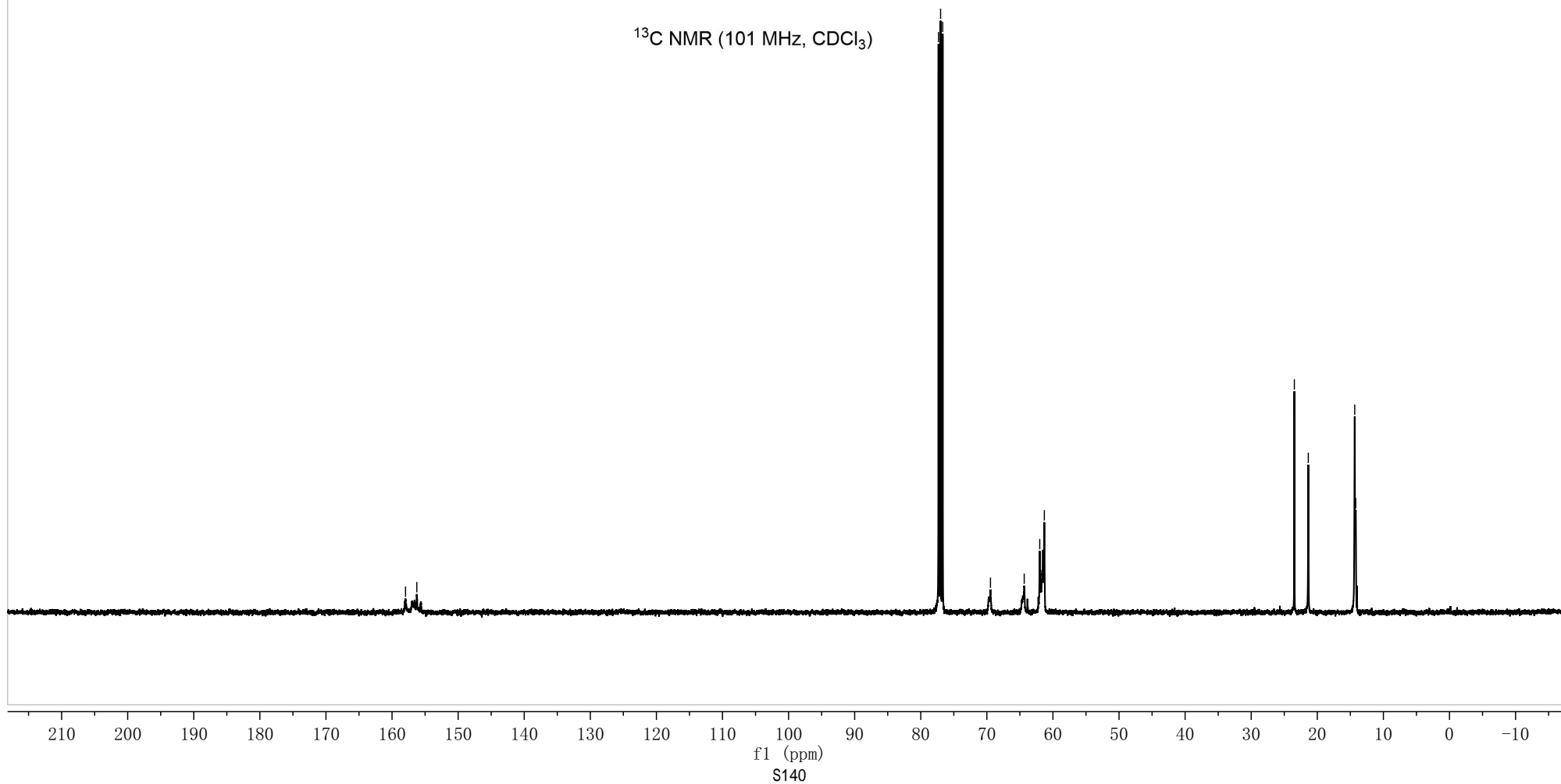


¹³C NMR (101 MHz, CDCl₃)

~157.96
~156.25

77.32
77.00
76.68
69.45
64.35
62.00
61.30

~23.47
~21.36
14.34
14.18

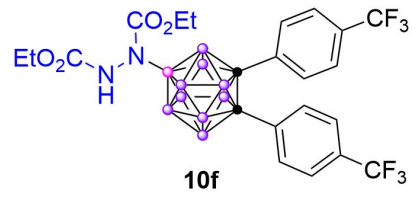


7.58
7.56
7.54
7.45
7.43
7.26

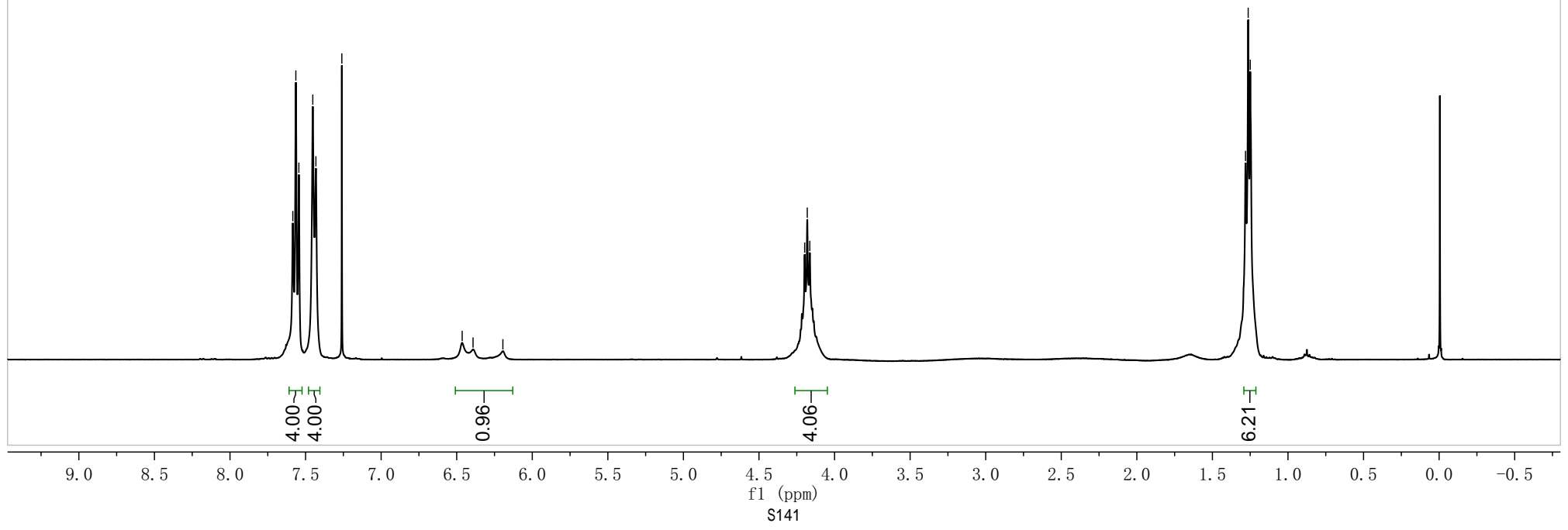
6.46
6.39
6.19

4.20
4.18
4.16

1.28
1.26
1.25



¹H NMR (400 MHz, CDCl₃)



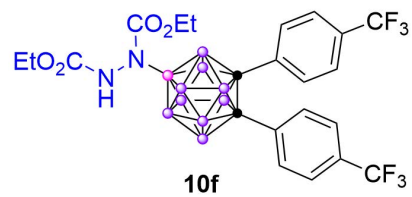
7.58
7.56
7.54
7.45
7.43
7.26

6.46
6.39
6.19

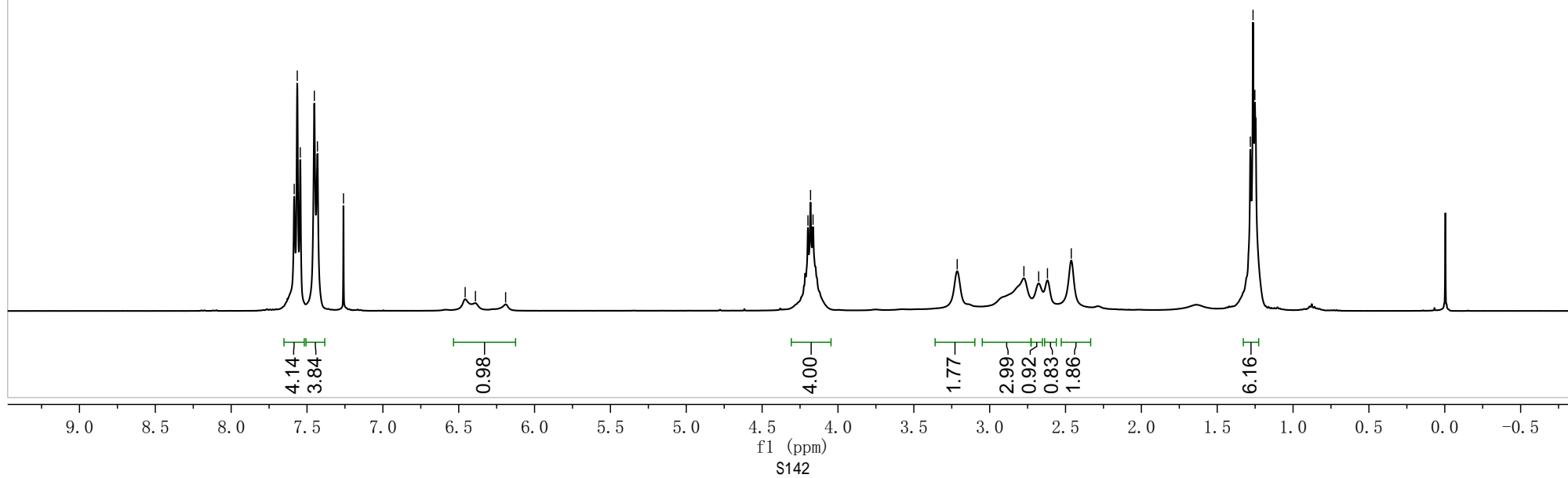
4.20
4.18
4.16

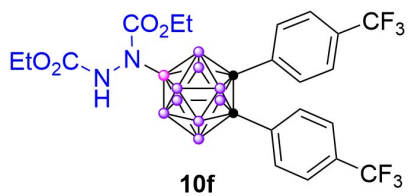
3.21
2.77
2.68
2.62
2.46

1.28
1.26
1.25



$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



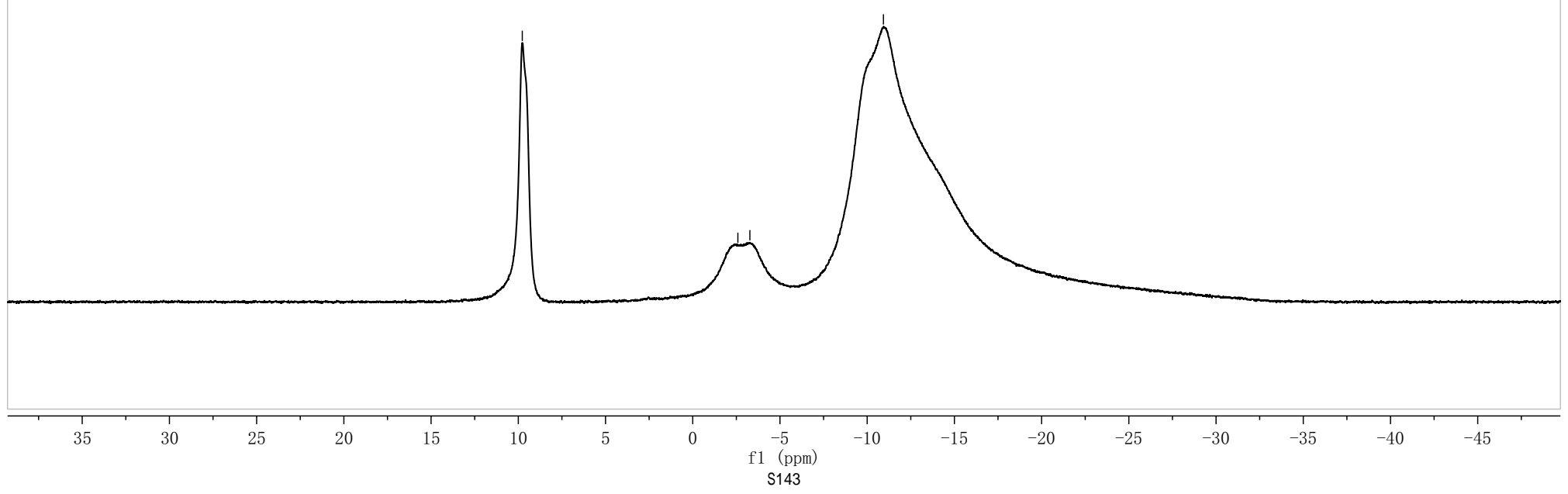


^{11}B NMR (128 MHz, CDCl_3)

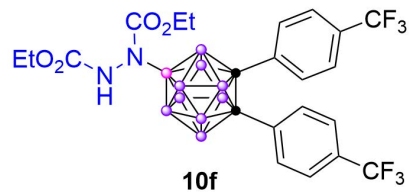
—9.77

—2.59
—3.28

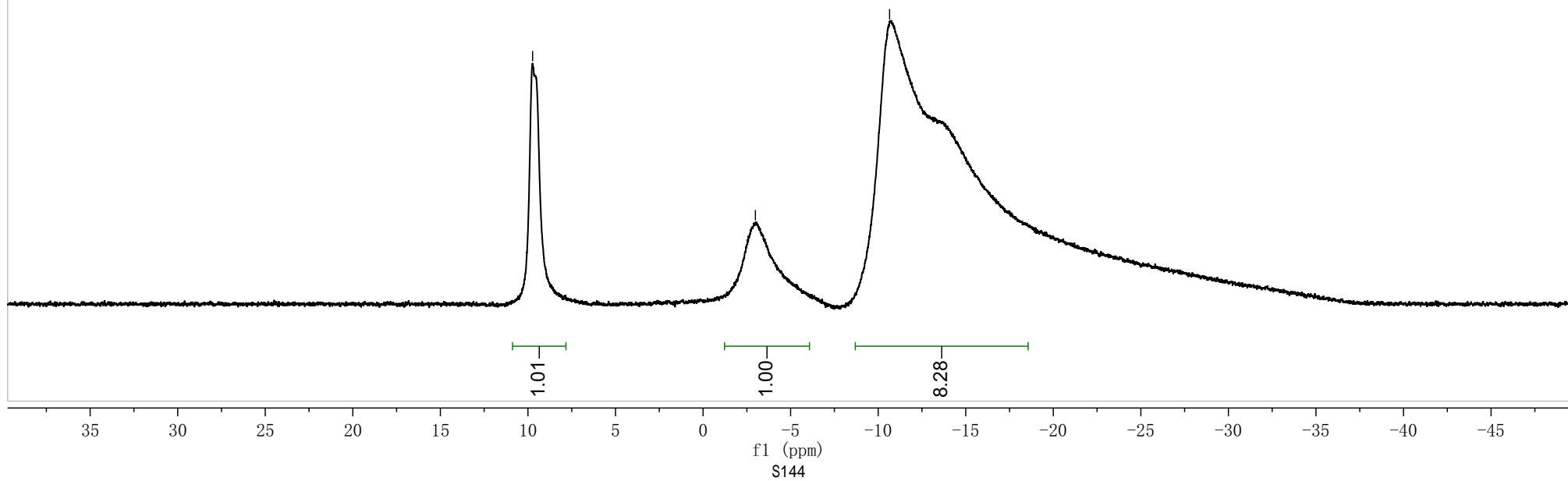
—10.93

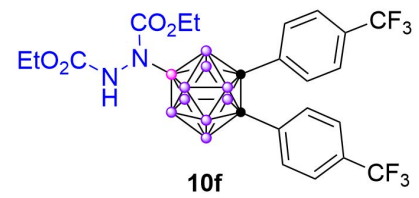


9.73
-2.99
-10.65



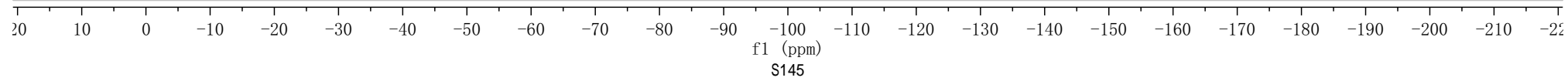
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)





^{19}F NMR (377 MHz, CDCl_3)

-63.22
-63.23

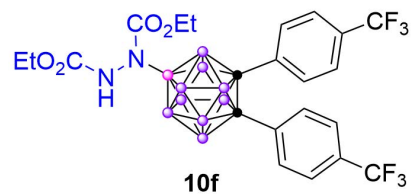


157.89
156.64

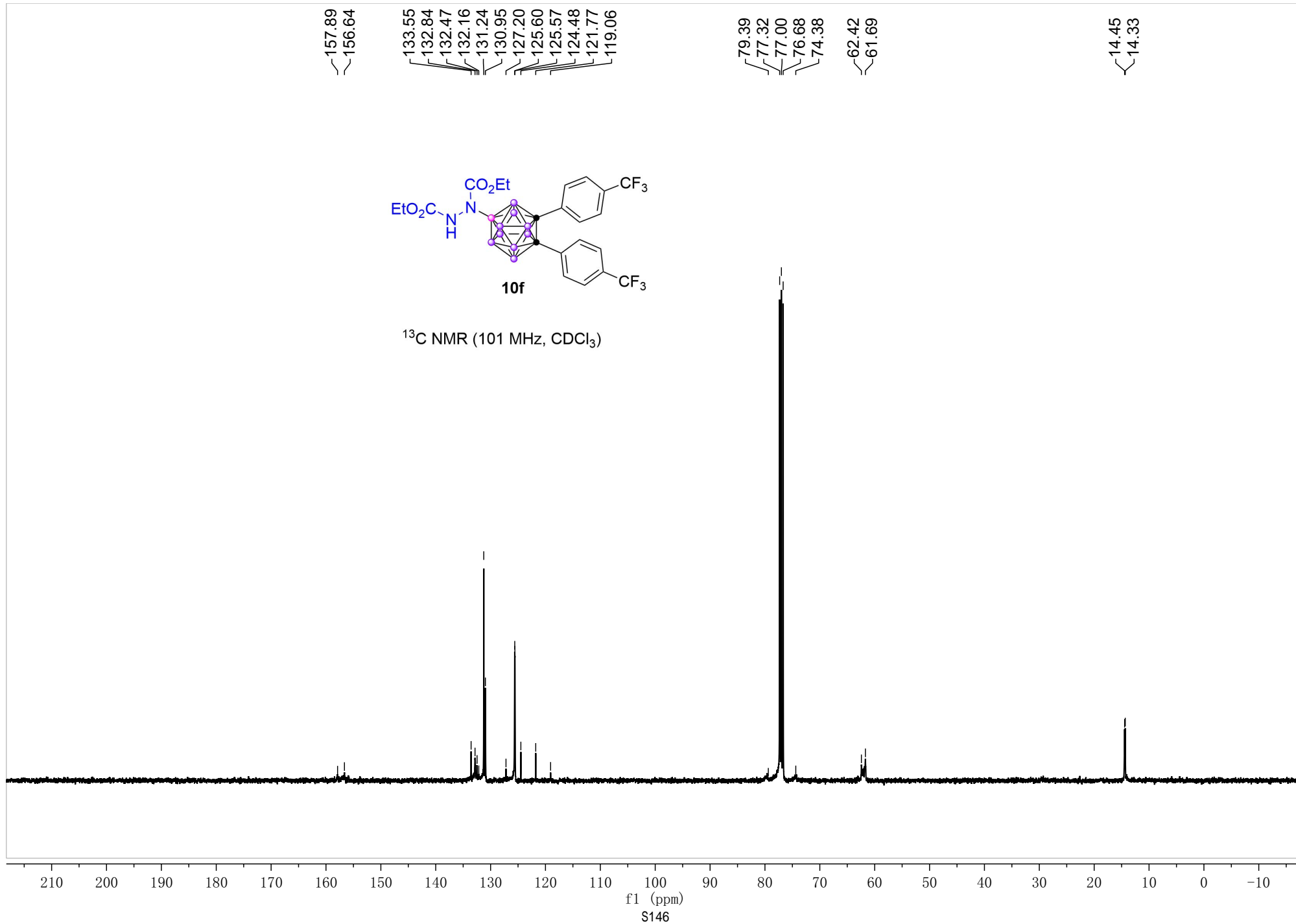
133.55
132.84
132.47
132.16
131.24
130.95
127.20
125.60
125.57
124.48
121.77
119.06

79.39
77.32
77.00
76.68
74.38
62.42
61.69

14.45
14.33



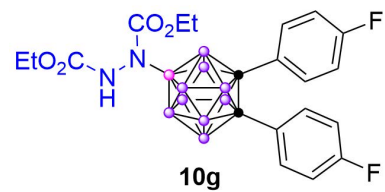
^{13}C NMR (101 MHz, CDCl_3)



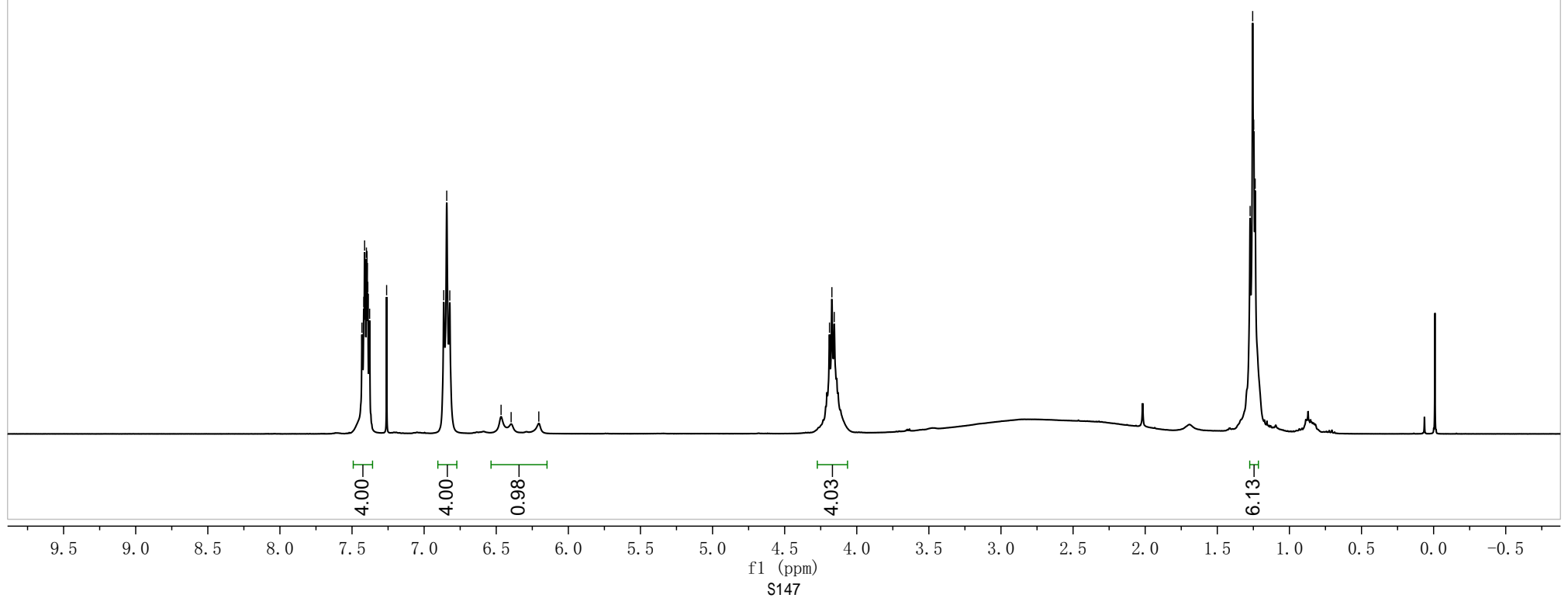
7.43
7.42
7.41
7.41
7.40
7.40
7.39
7.38
7.38
7.26
6.86
6.84
6.82
6.47
6.40
6.20

4.19
4.17
4.16

1.27
1.25
1.25
1.24



^1H NMR (400 MHz, CDCl_3)

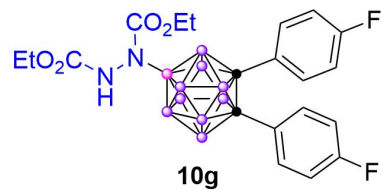


7.43
7.41
7.40
7.38
7.26
6.86
6.84
6.82
6.45
6.39
6.20

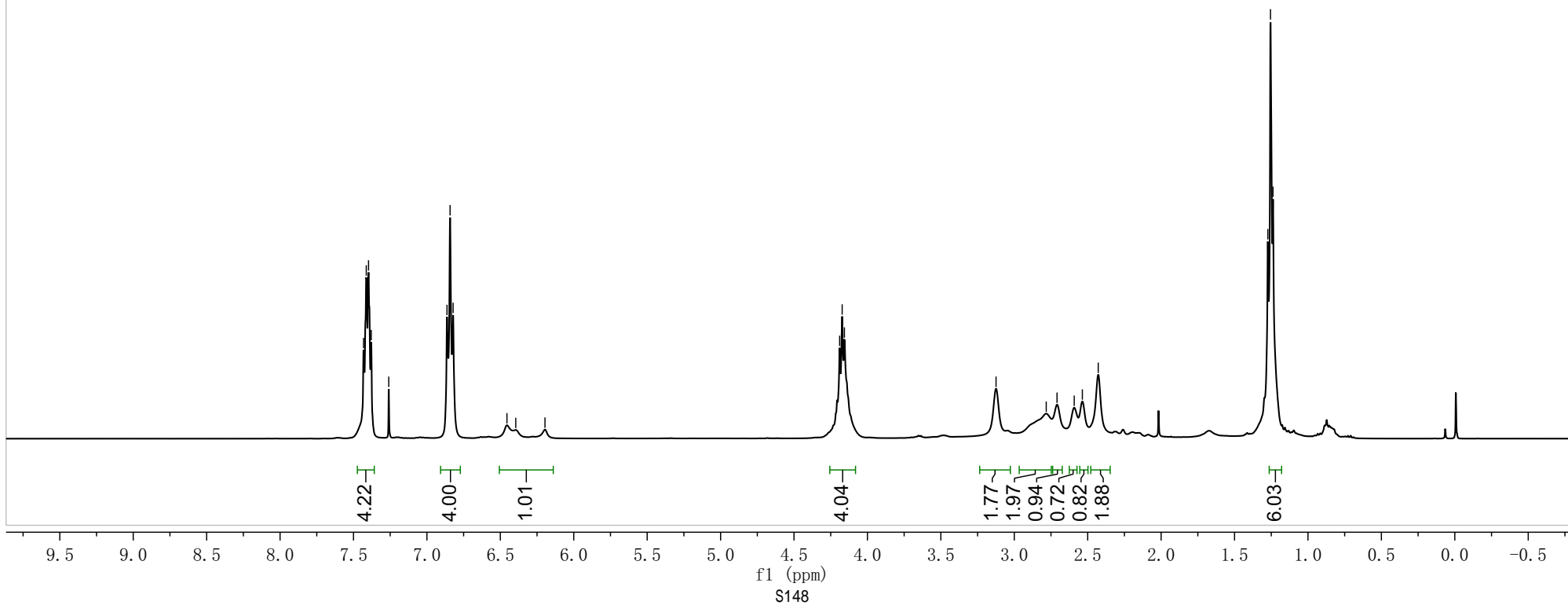
4.19
4.17
4.16

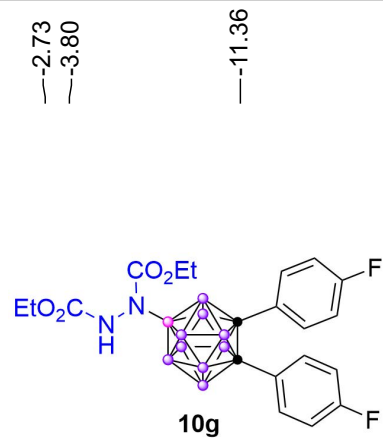
3.12
2.78
2.71
2.59
2.54
2.43

1.27
1.26
1.24



$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



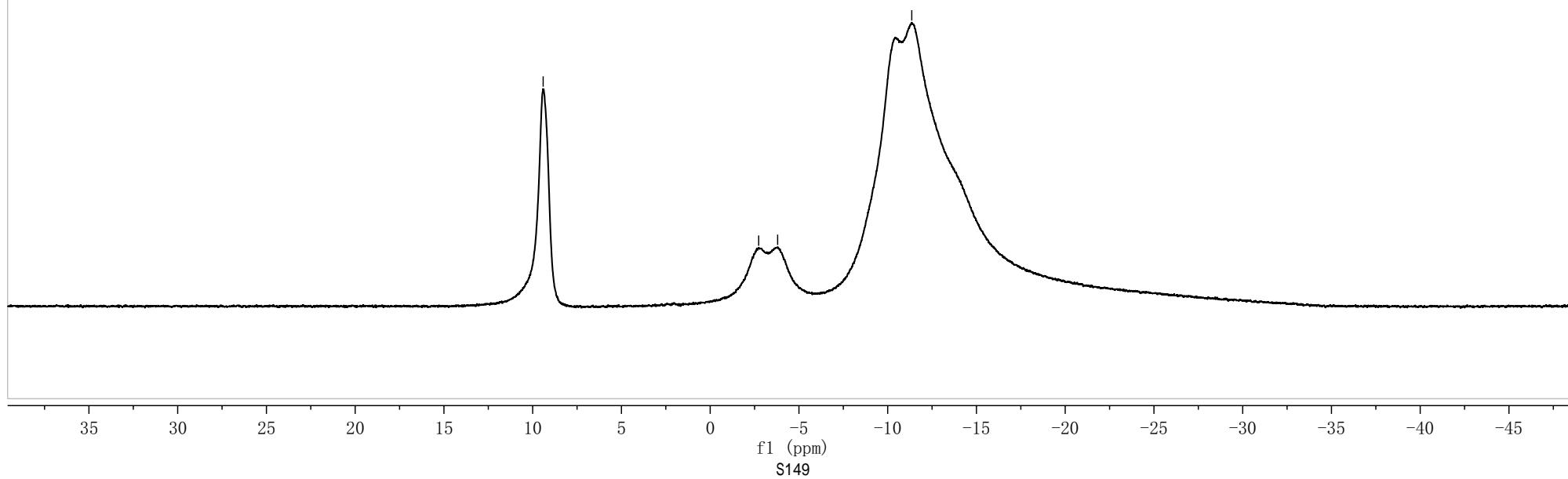


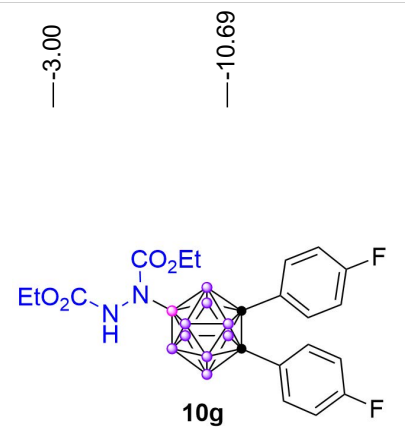
¹¹B NMR (128 MHz, CDCl₃)

—9.41

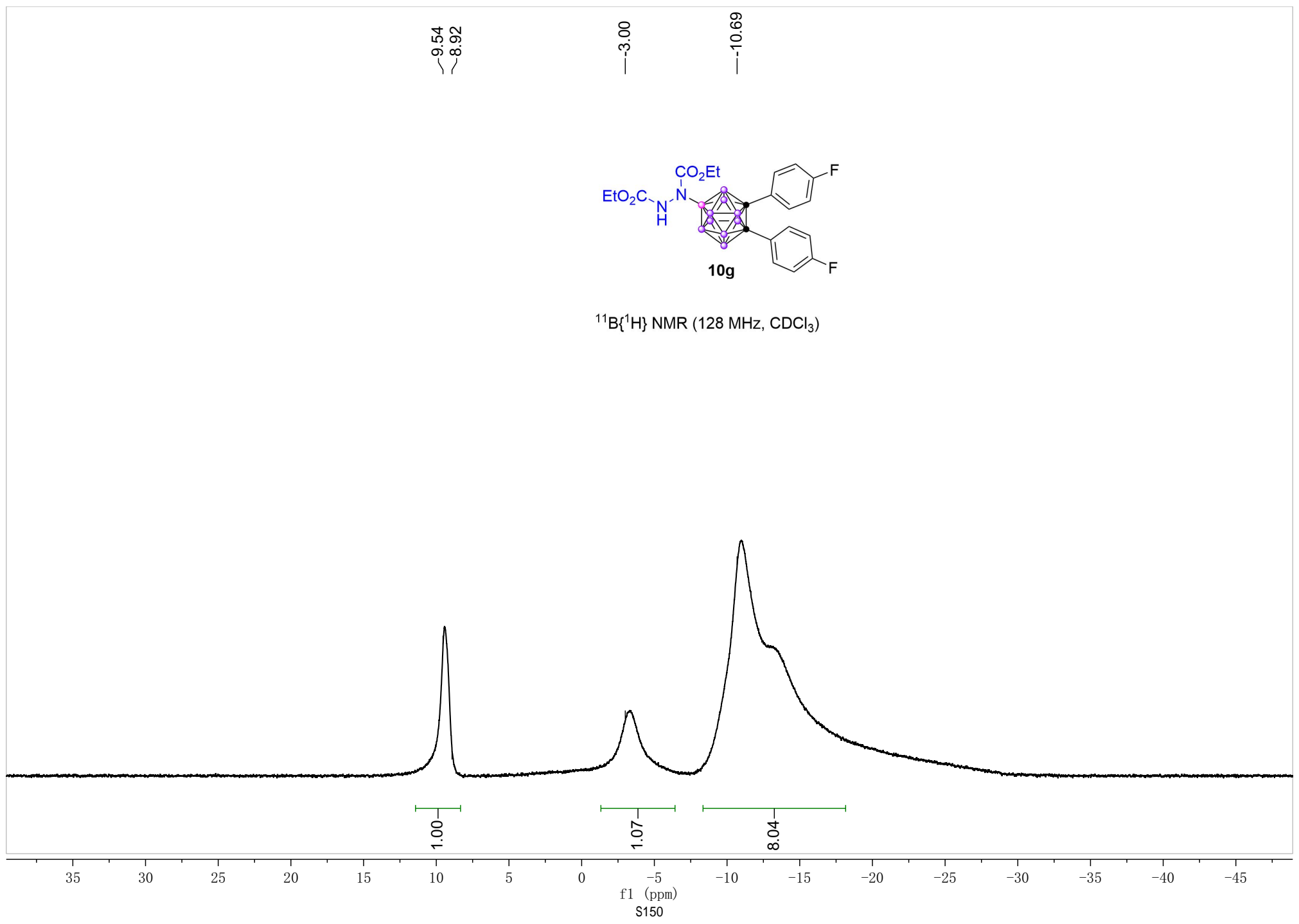
—2.73
—3.80

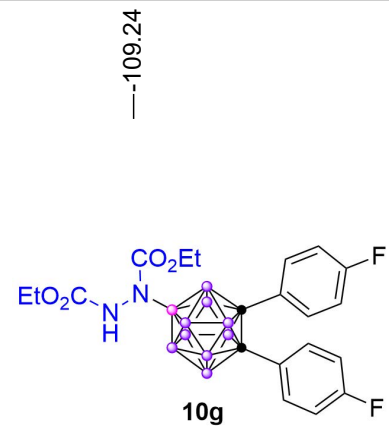
—11.36



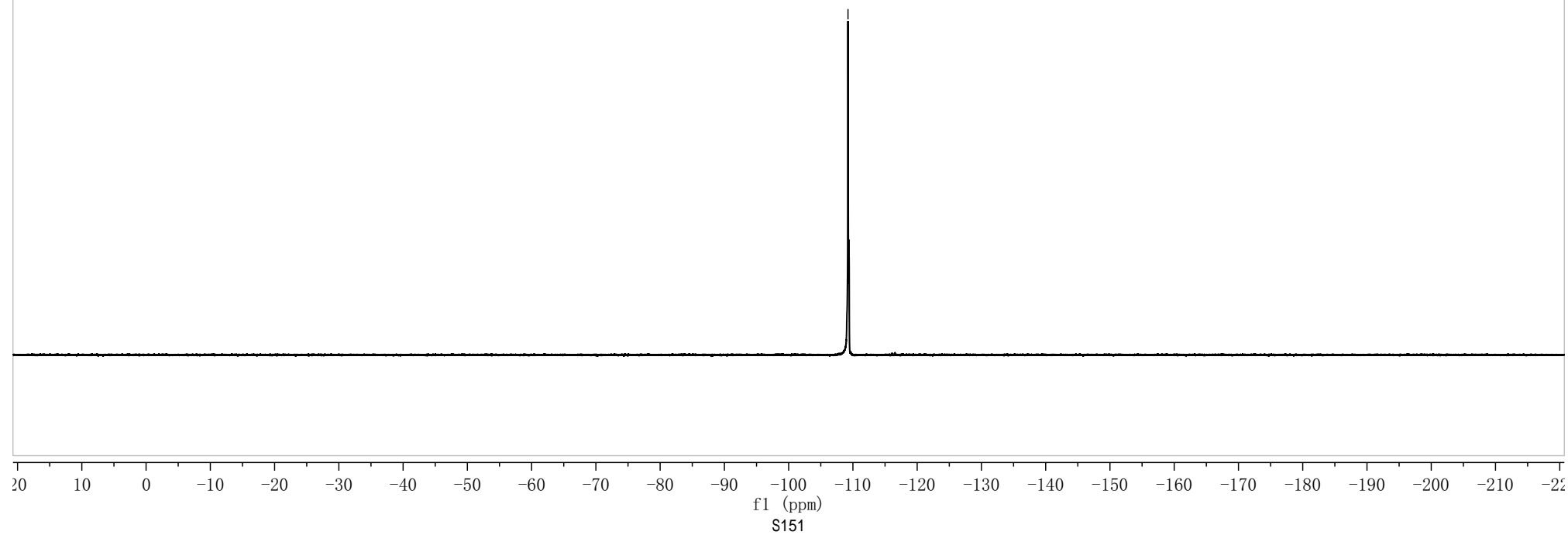


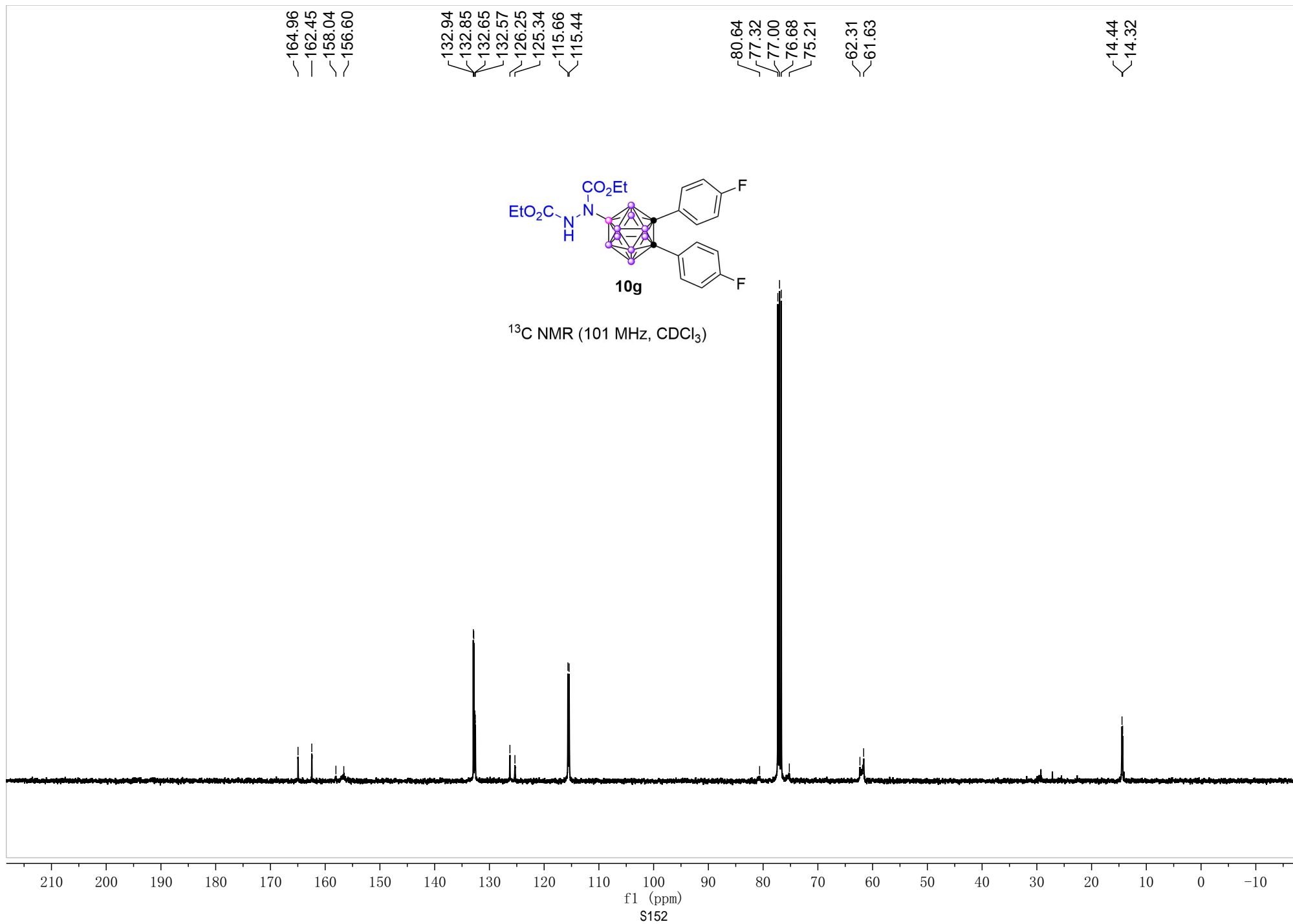
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

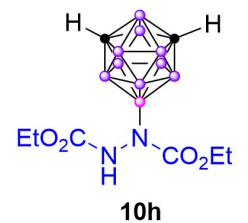




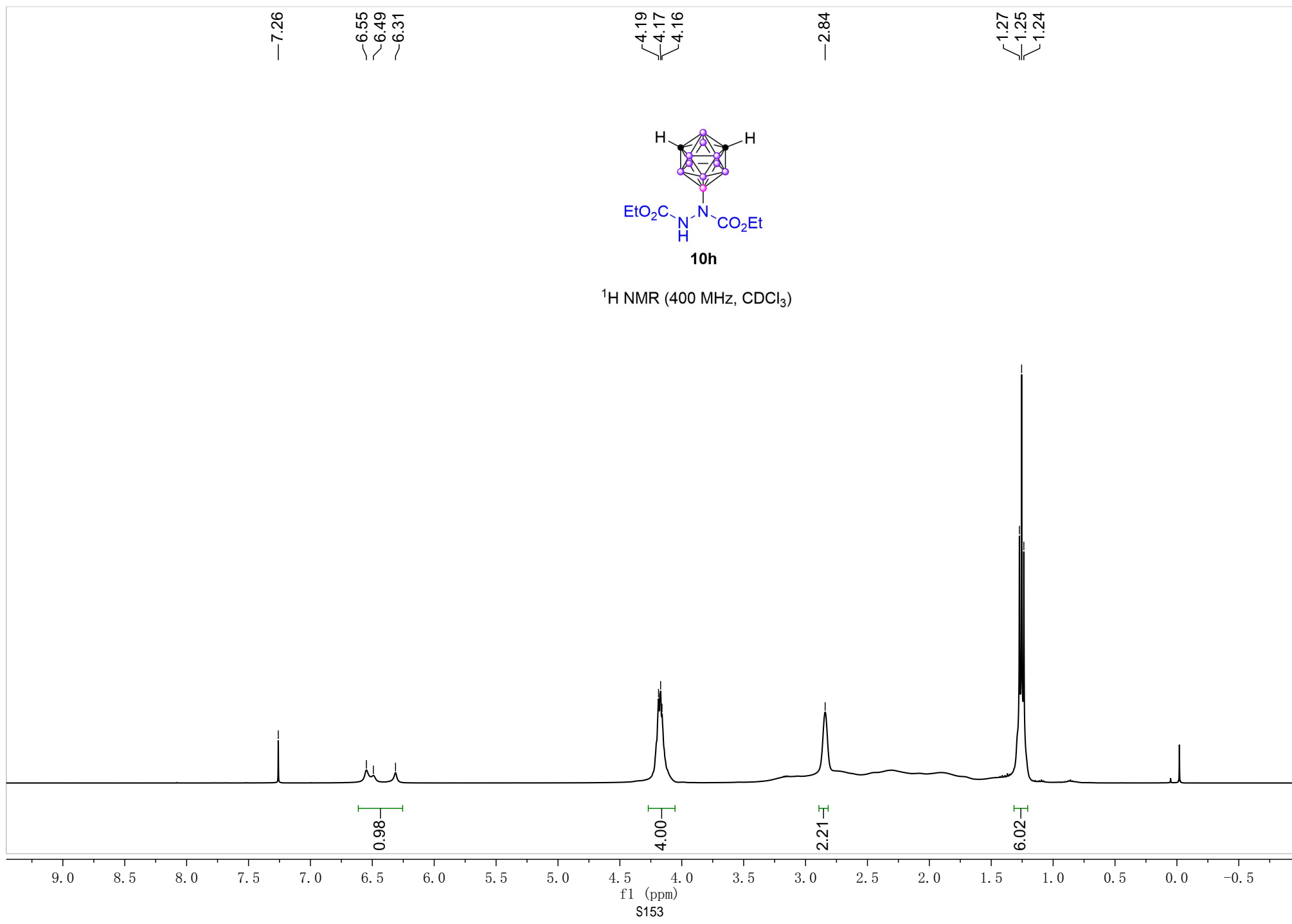
¹⁹F NMR (377 MHz, CDCl₃)

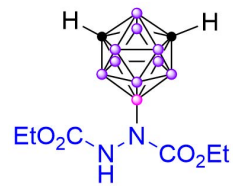






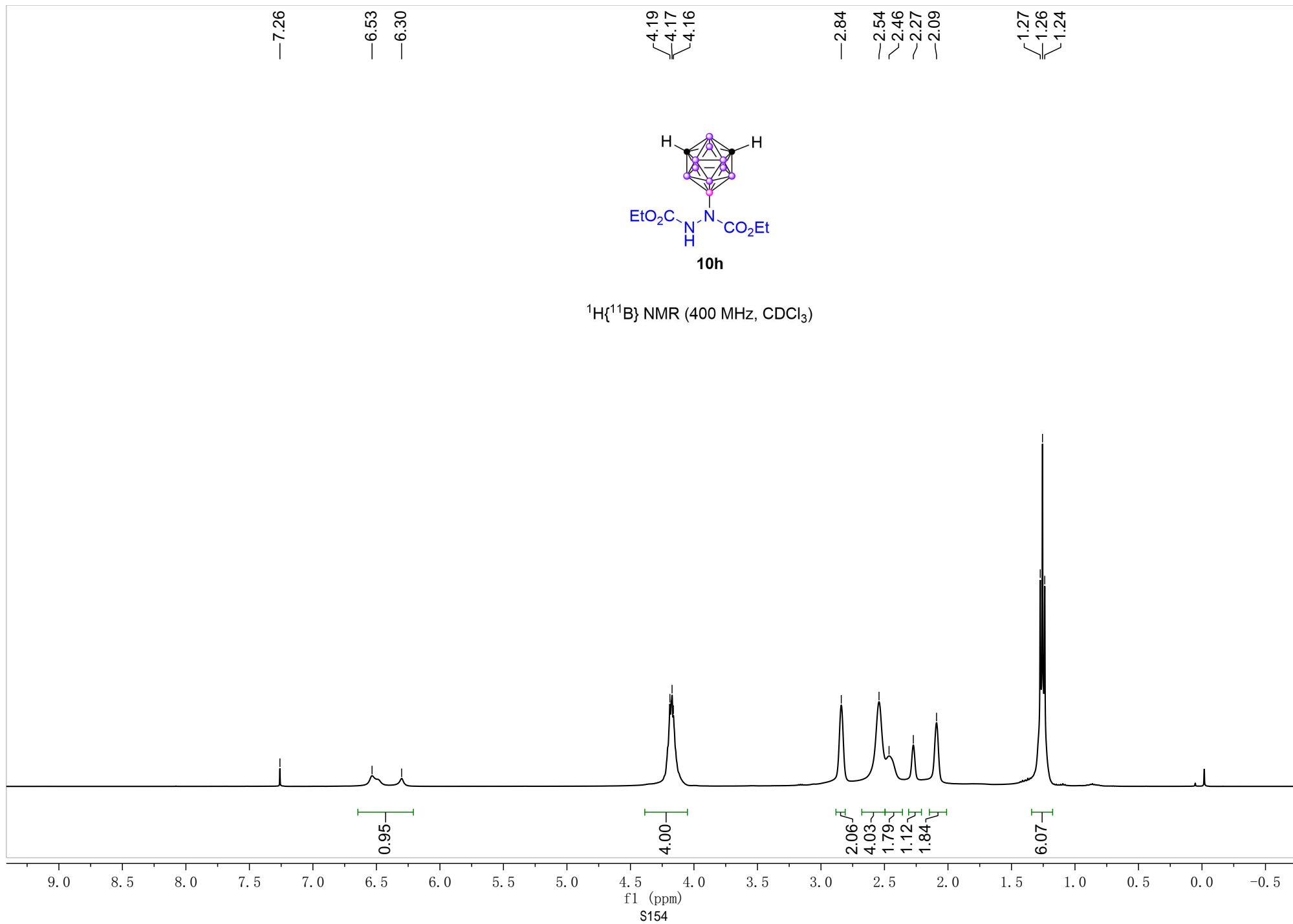
¹H NMR (400 MHz, CDCl₃)





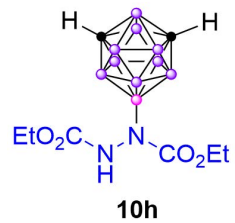
10h

$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)

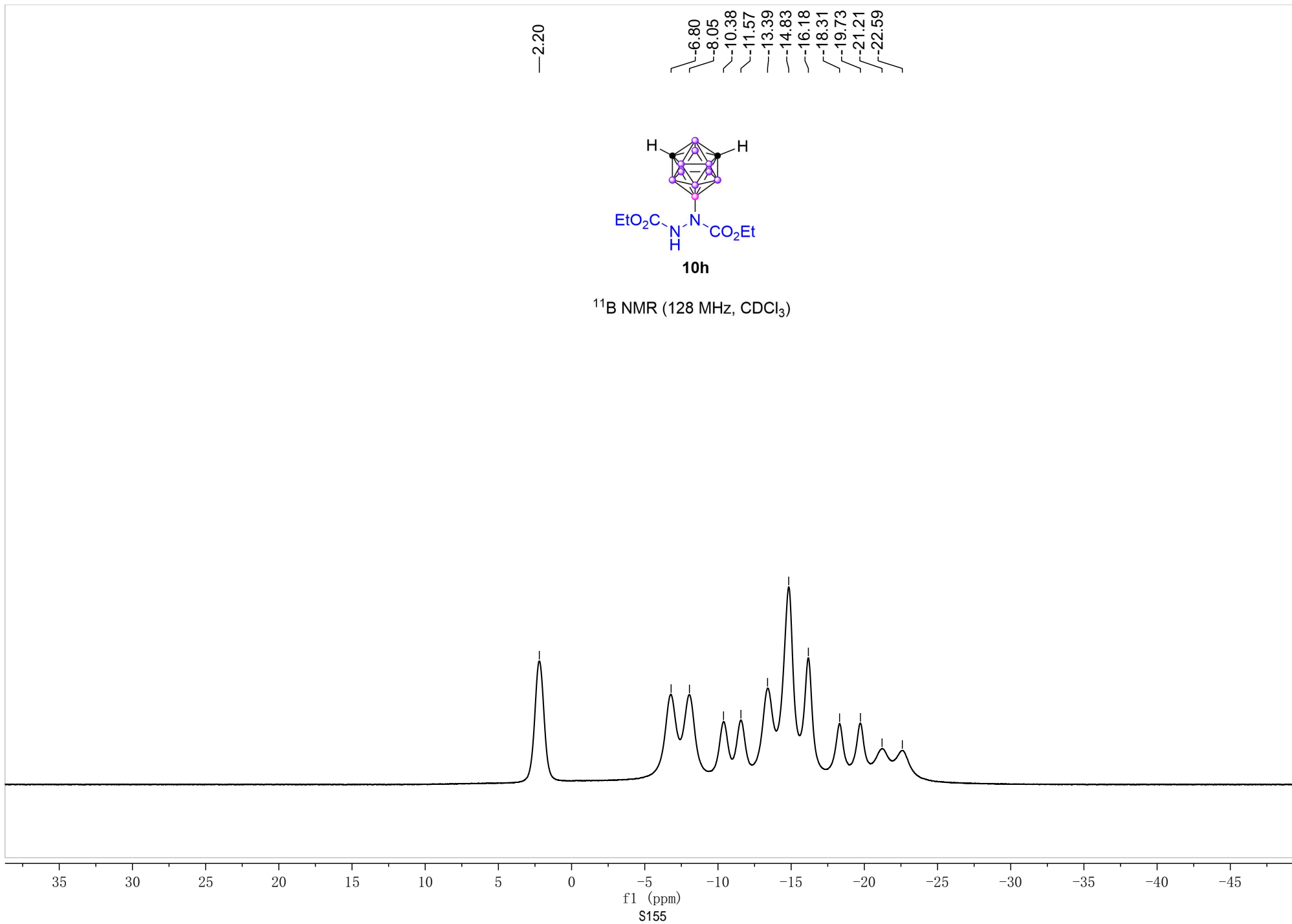


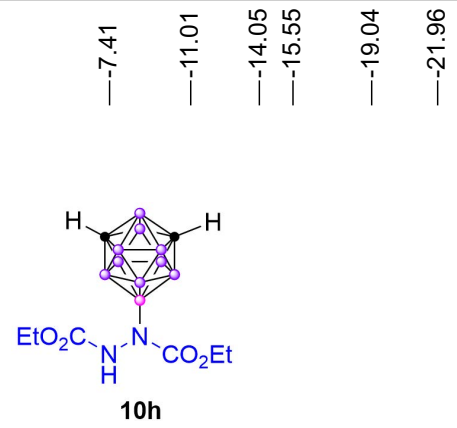
-2.20

-6.80
-8.05
-10.38
-11.57
-13.39
-14.83
-16.18
-18.31
-19.73
-21.21
-22.59



¹¹B NMR (128 MHz, CDCl₃)





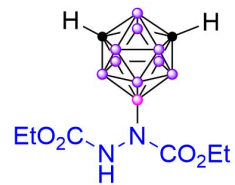
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

—2.22
—7.41
—11.01
—14.05
—15.55
—19.04
—21.96

1.10
2.00
1.00
1.88
2.10
1.04
1.01

f1 (ppm)
S156

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45



10h

¹³C NMR (101 MHz, CDCl₃)

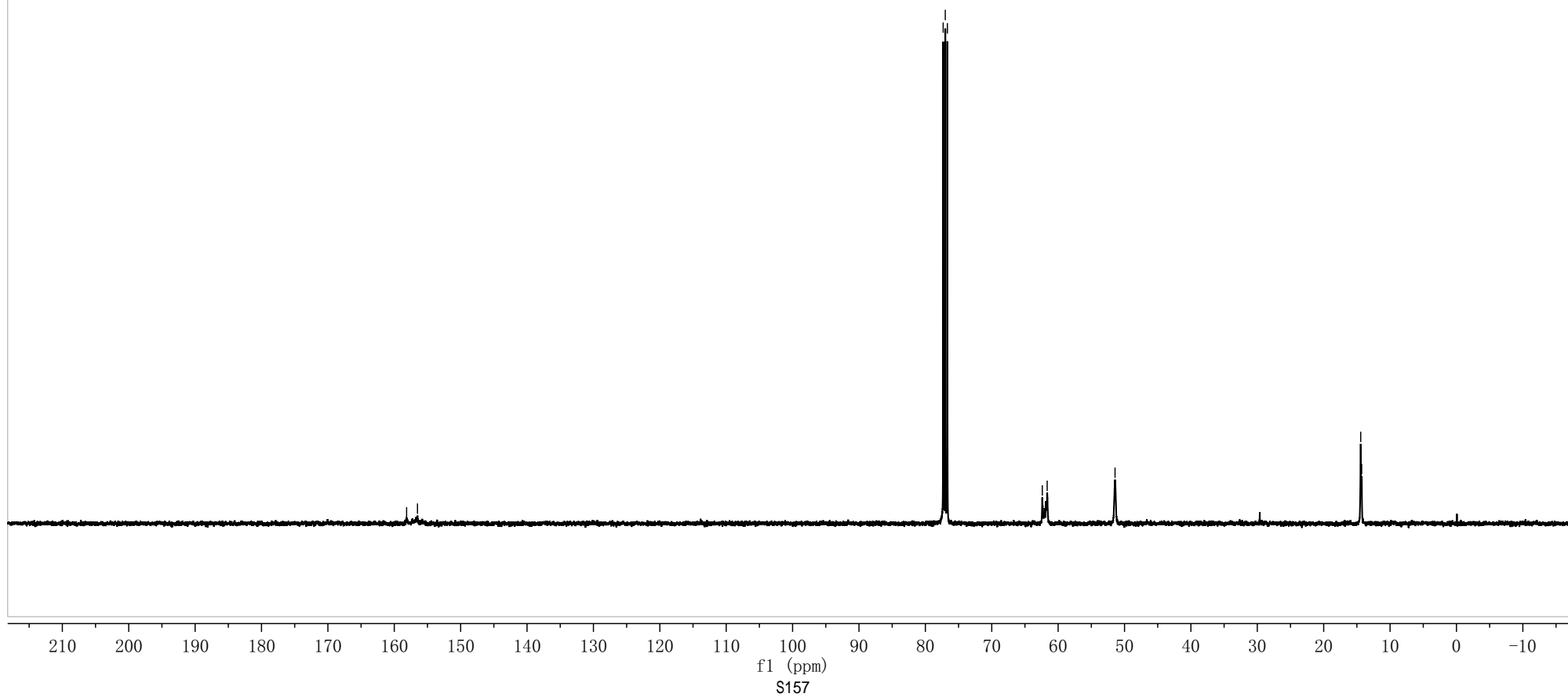
158.14
156.50

77.32
77.00
76.68

62.38
61.65

51.43

14.42
14.28

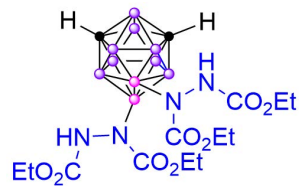


7.61
7.51
7.26
7.09
6.84
6.74

4.17

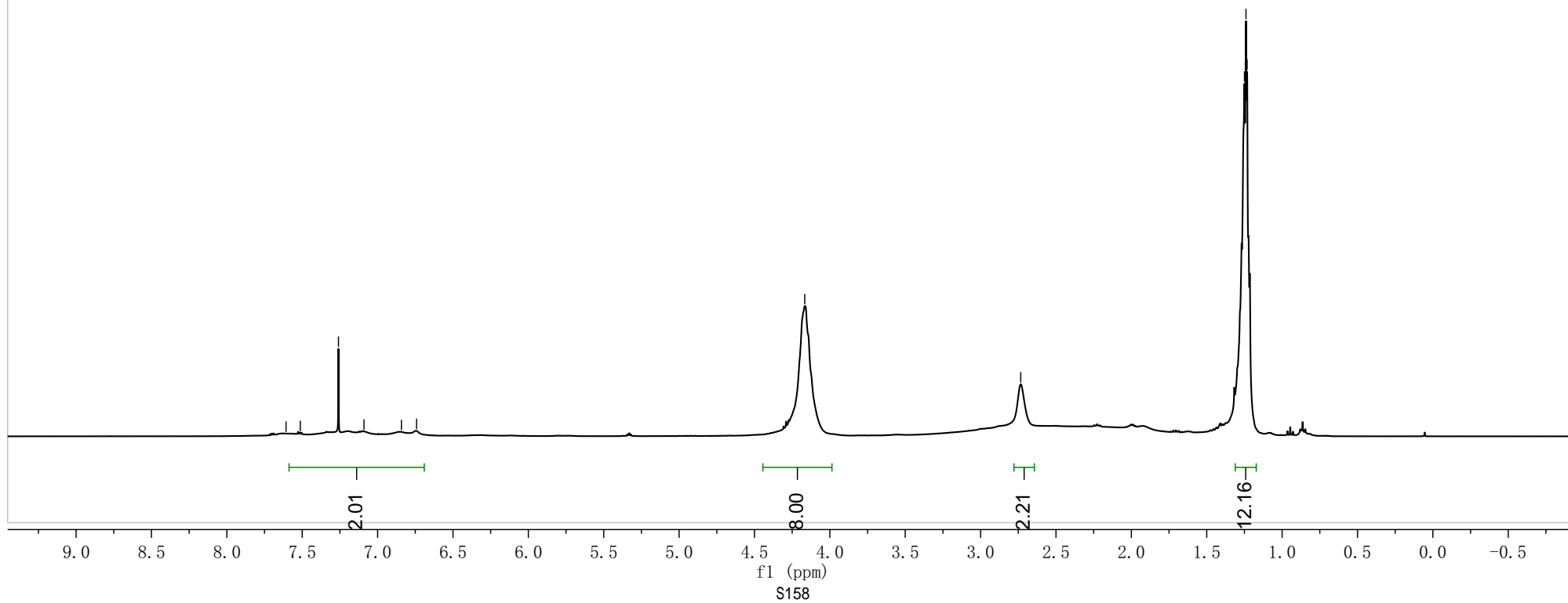
2.73

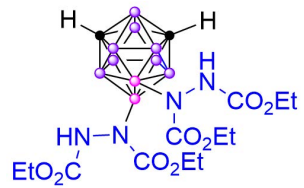
1.25
1.24
1.23



10h'

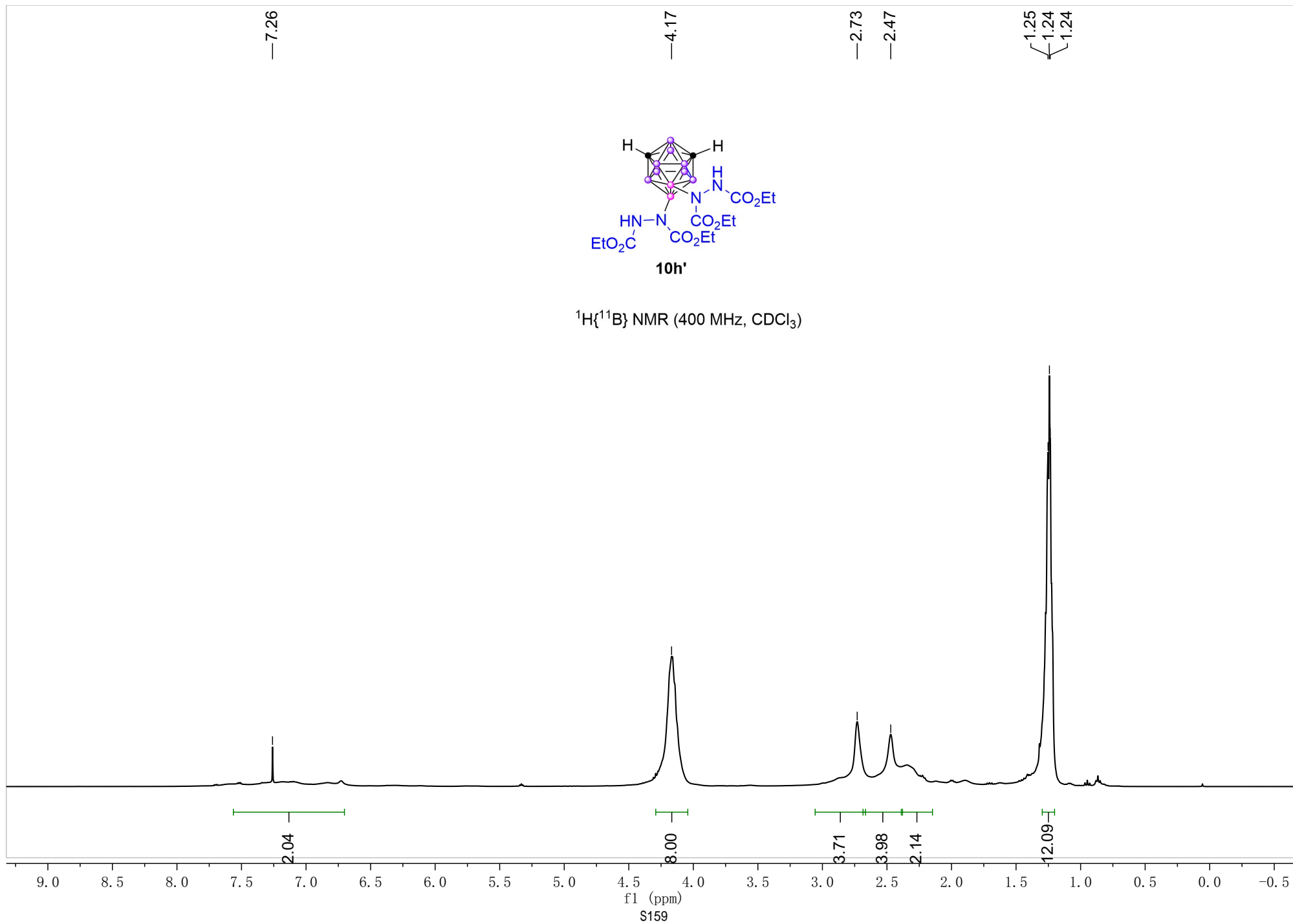
¹H NMR (400 MHz, CDCl₃)



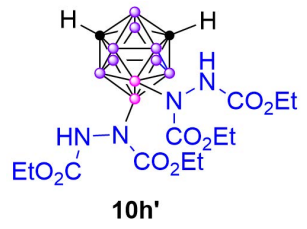


10h'

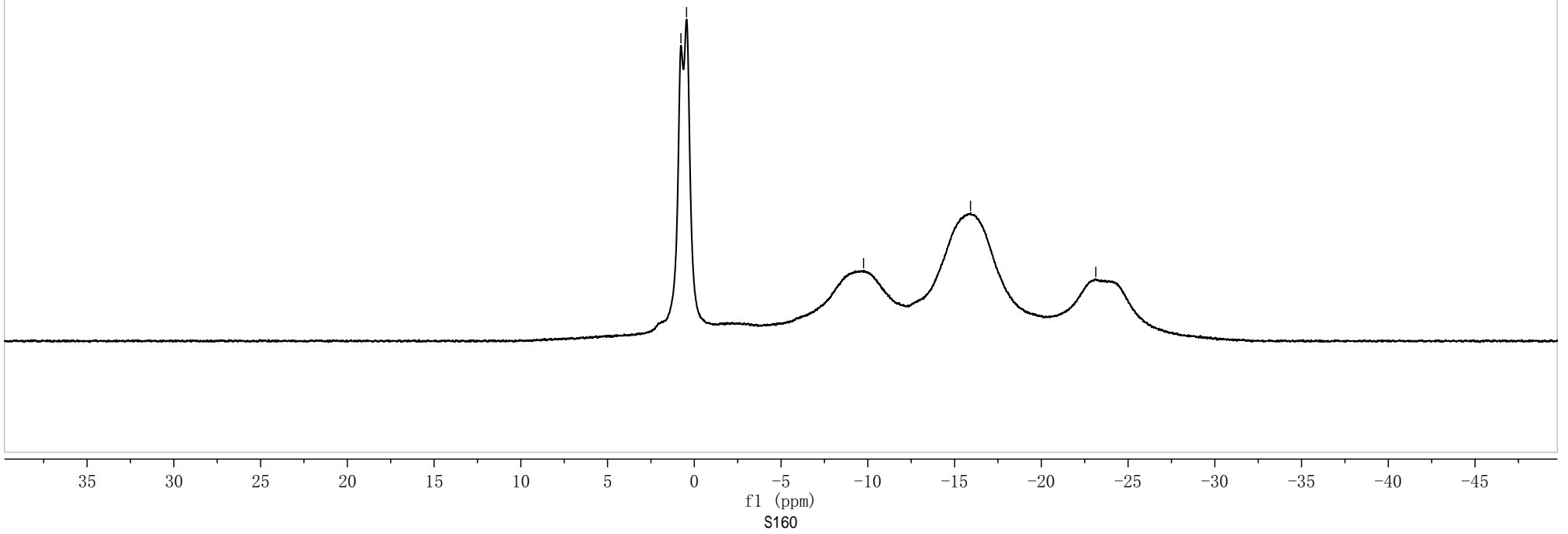
$^1\text{H}\{^{11}\text{B}\}$ NMR (400 MHz, CDCl_3)



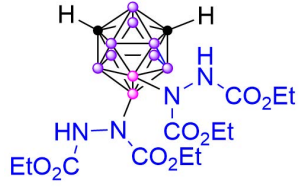
0.77
0.45
-9.75
-15.92
-23.14



¹¹B NMR (128 MHz, CDCl₃)

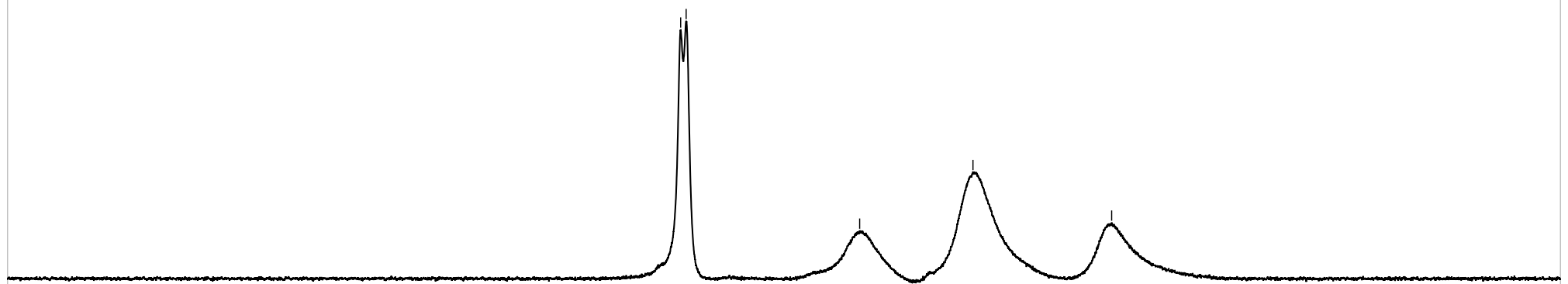


0.77
0.46
-9.52
-16.03
-24.00



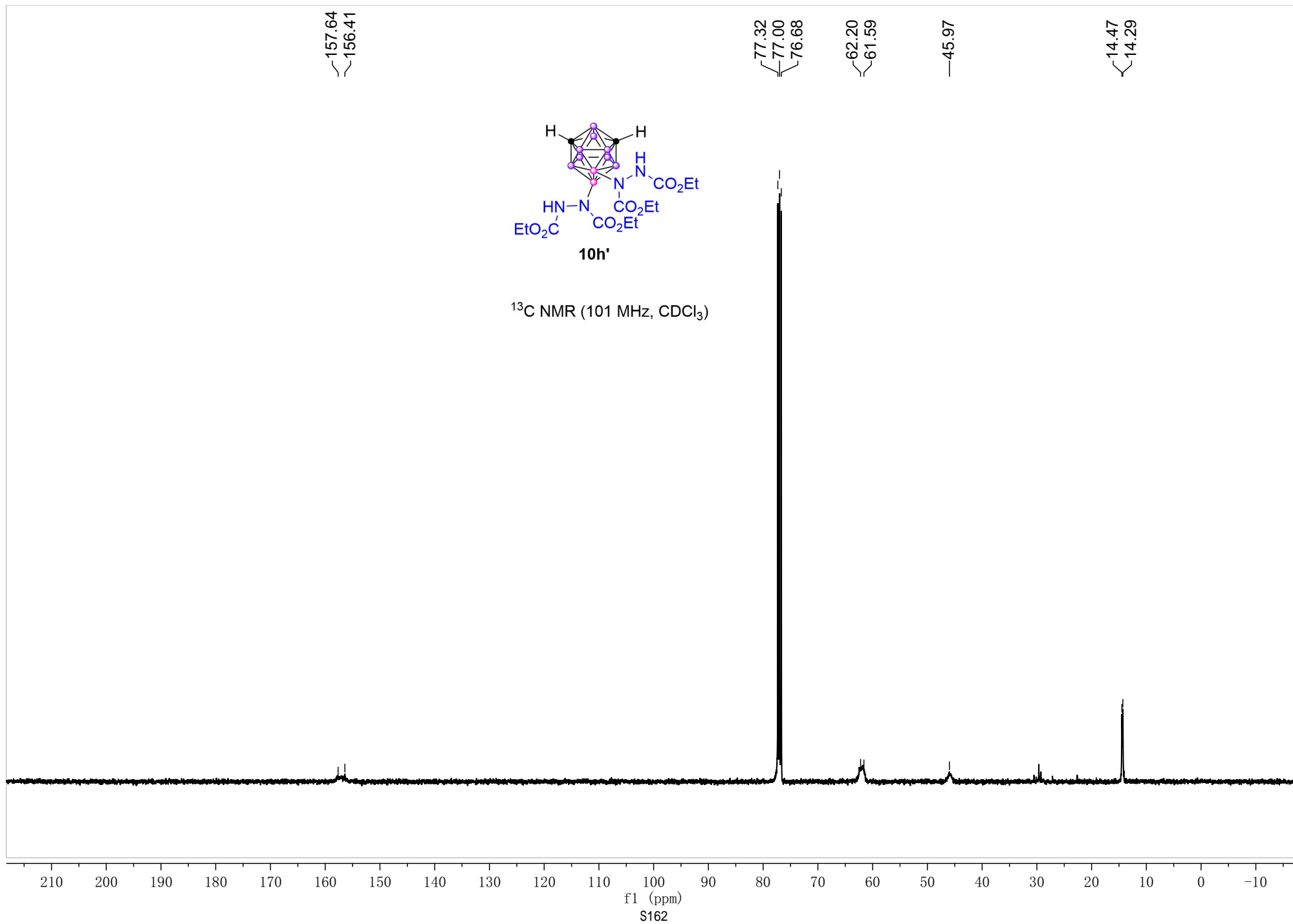
10h'

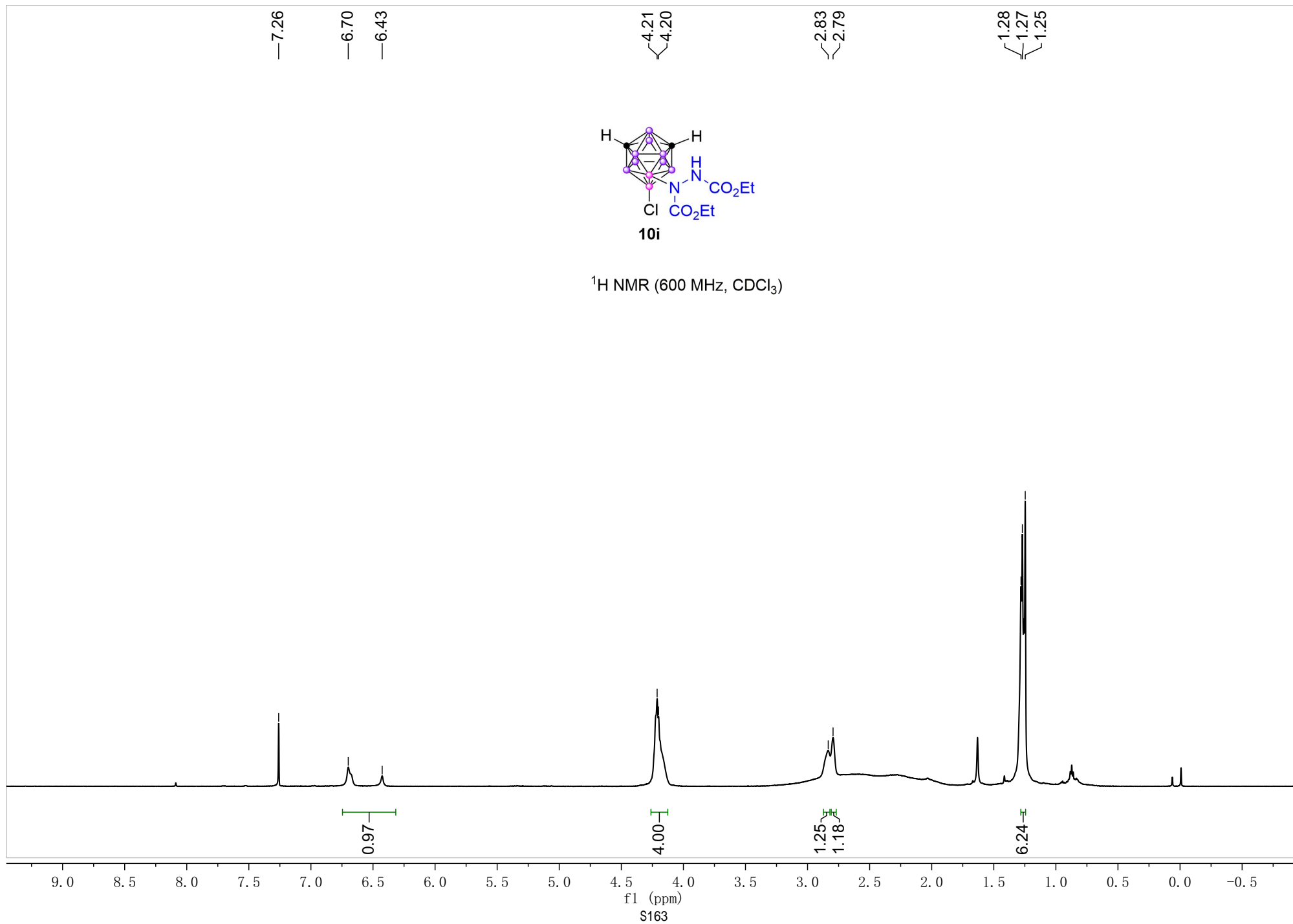
$^{11}\text{B}\{^1\text{H}\}$ NMR (128 MHz, CDCl_3)

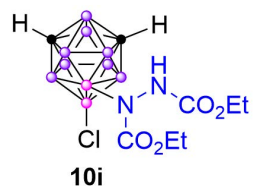


1.06
1.15
1.62
4.00
2.05

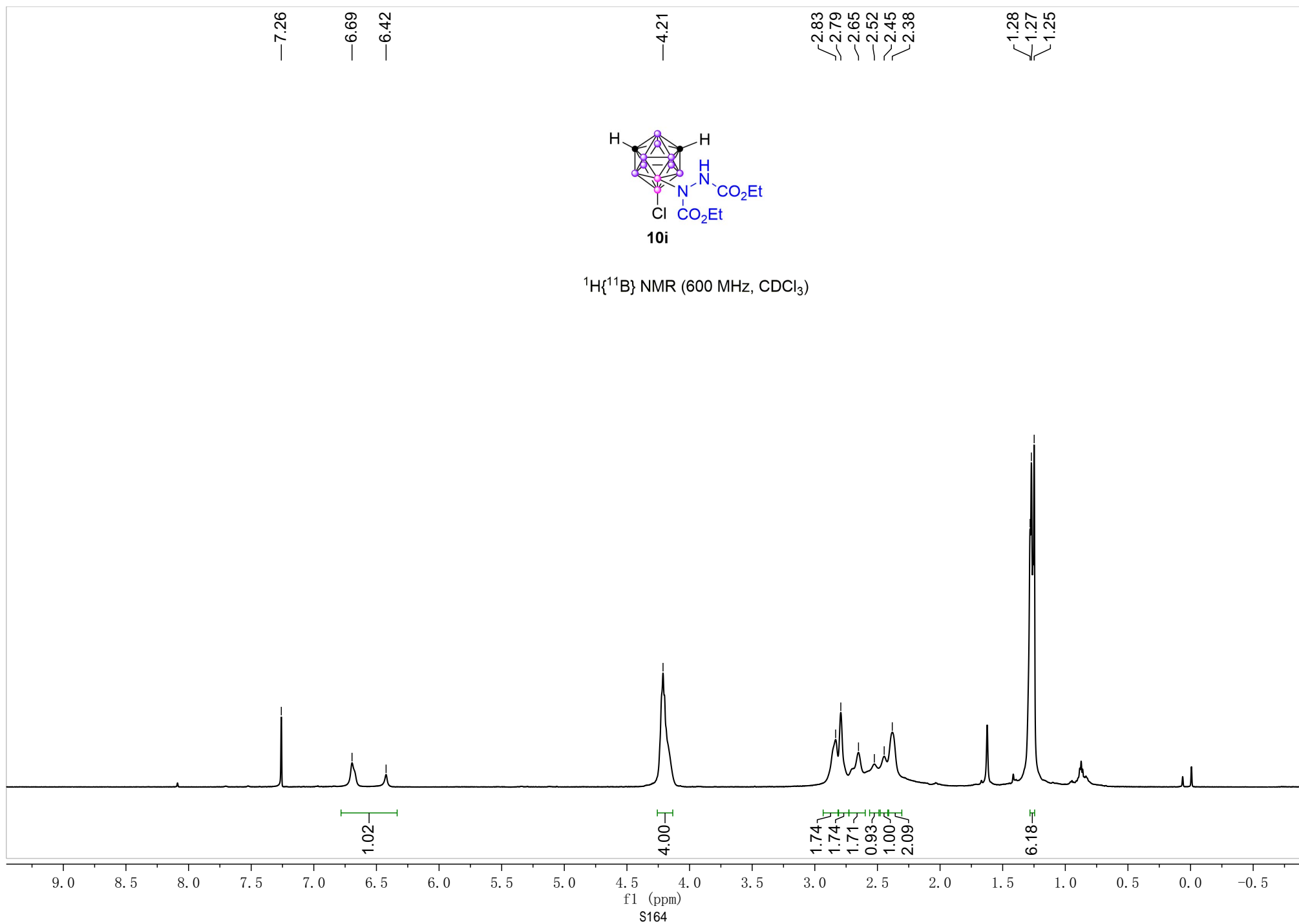
35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45
f1 (ppm)
S161



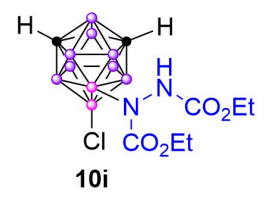




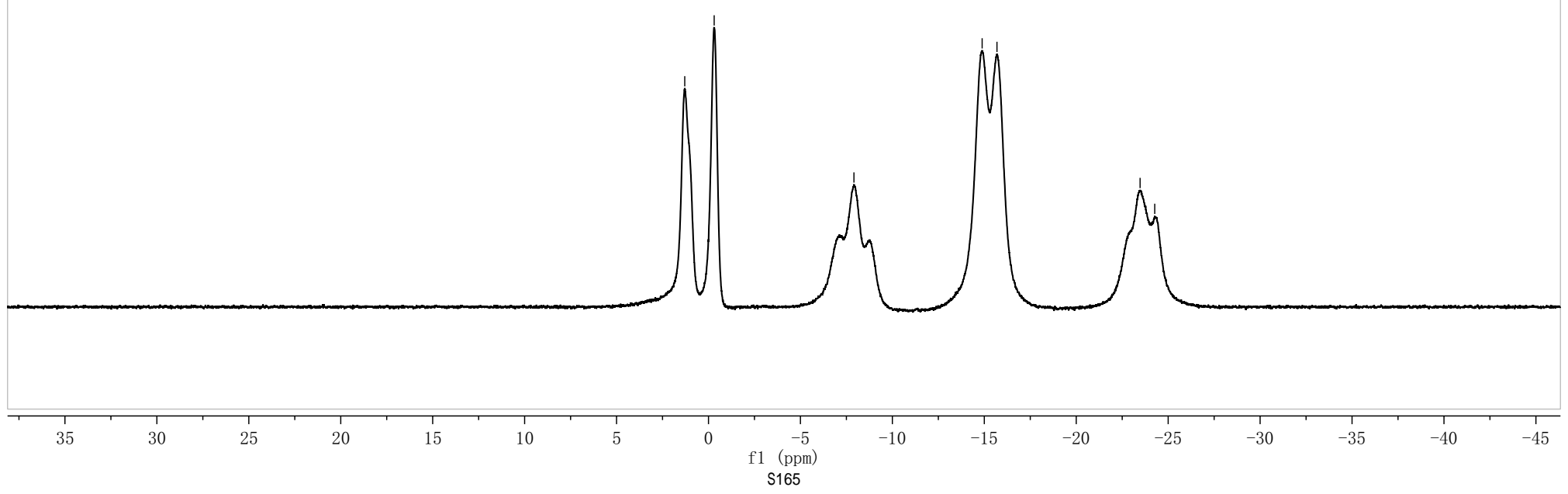
$^1\text{H}\{^{11}\text{B}\}$ NMR (600 MHz, CDCl_3)



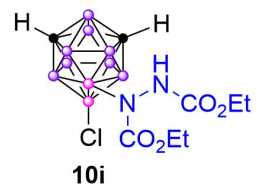
—1.29
—0.31
—7.91
—14.88
—15.69
—23.47
—24.27



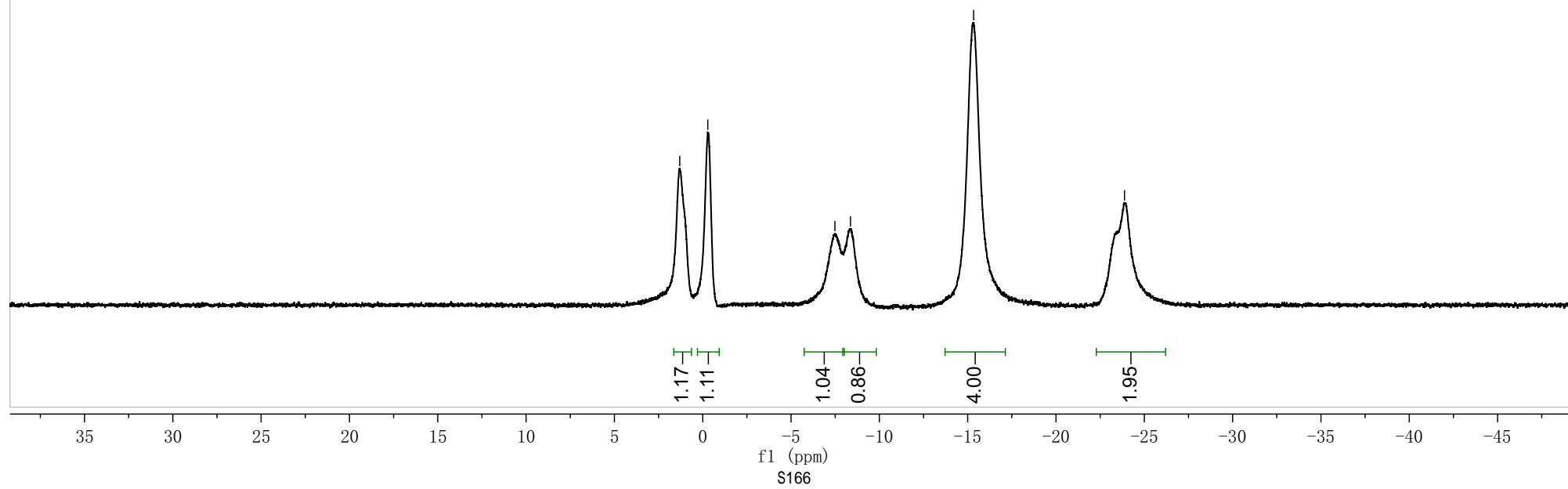
¹¹B NMR (193 MHz, CDCl₃)

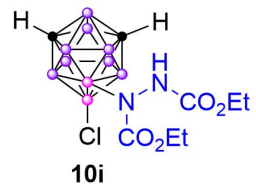


— 1.30
— -0.29
— 7.48
— 8.36
— -15.34
— -23.88

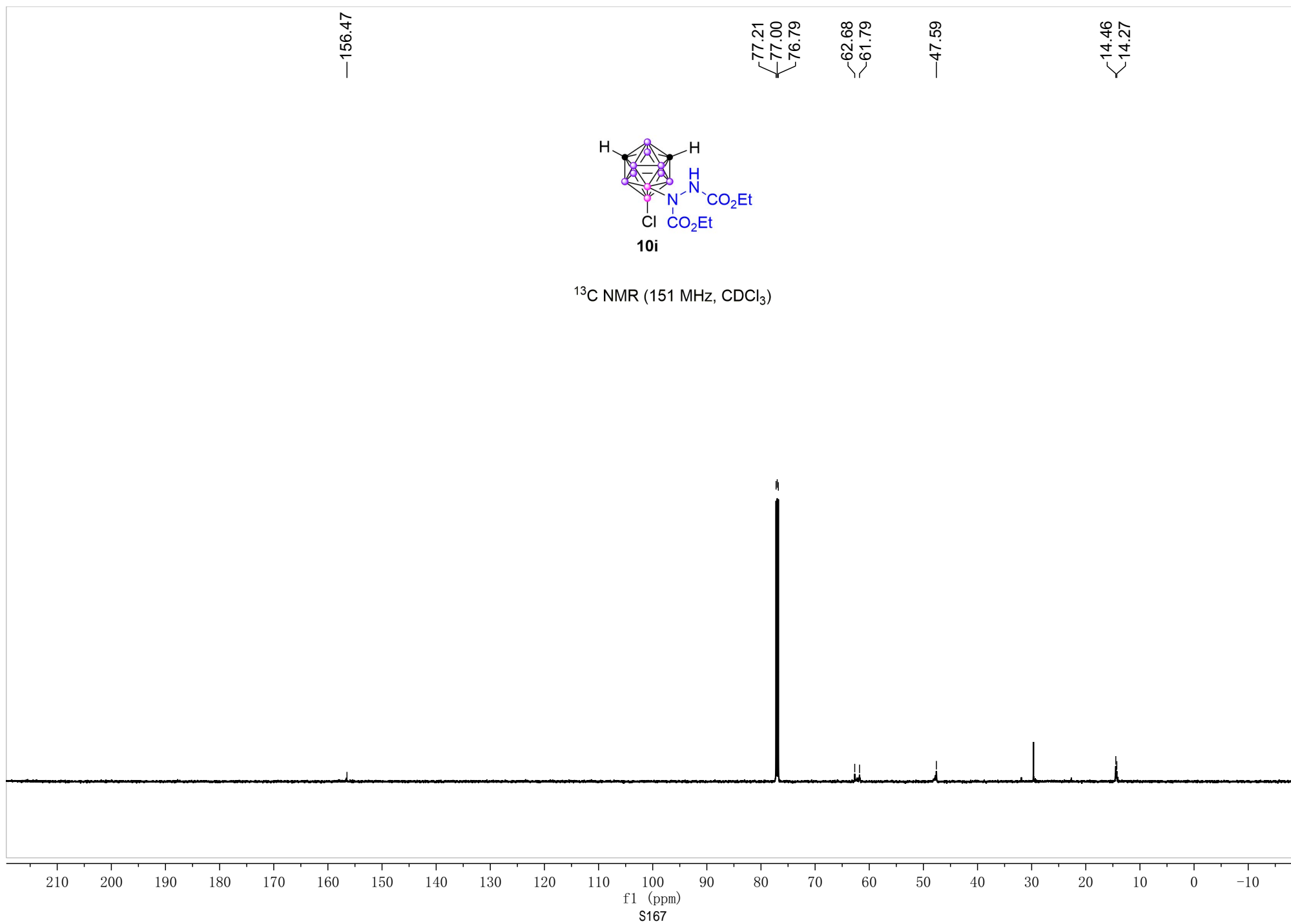


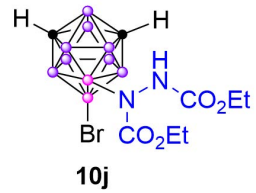
$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)



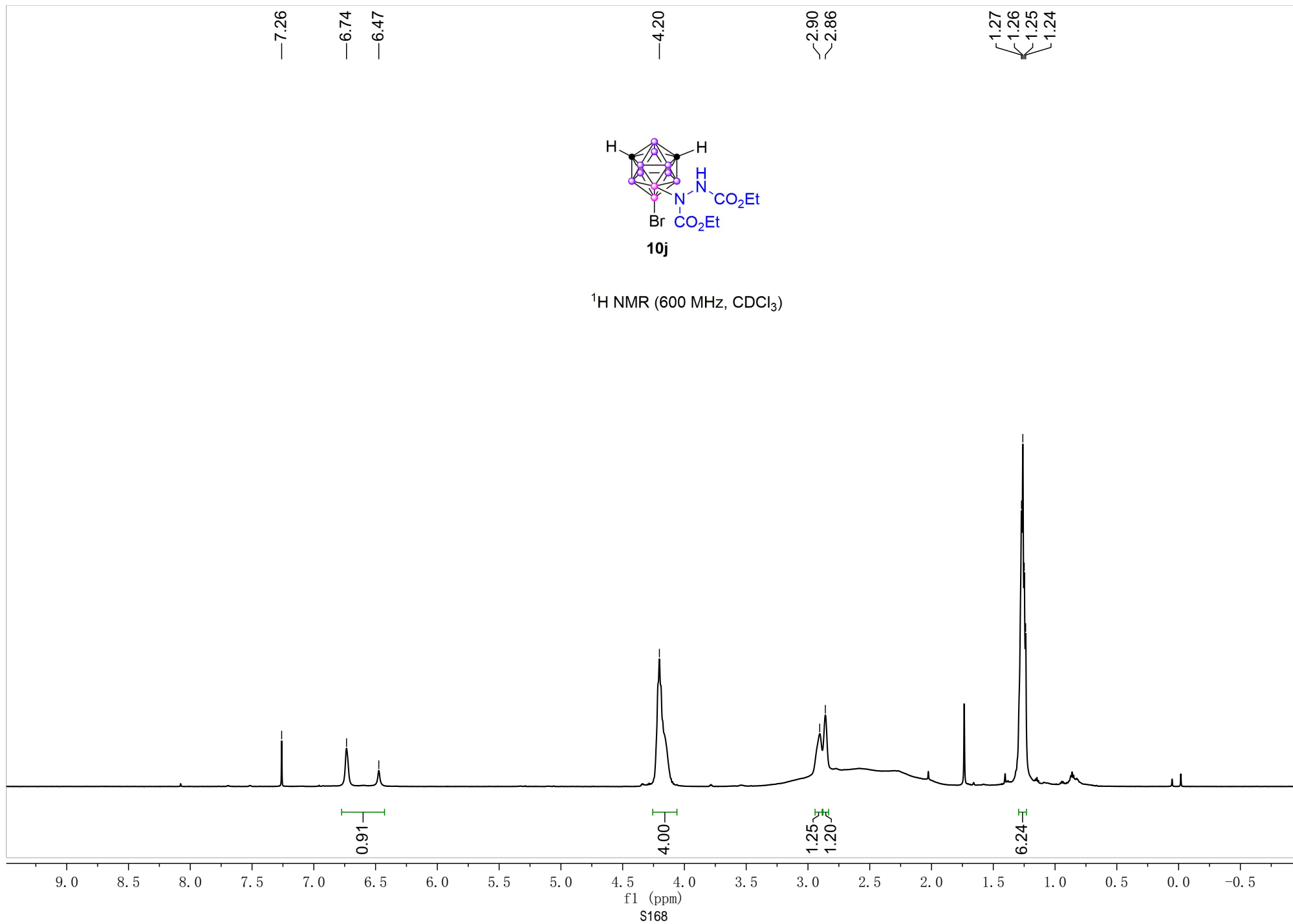


¹³C NMR (151 MHz, CDCl₃)





¹H NMR (600 MHz, CDCl₃)



7.26

6.73

6.47

4.20

2.91

2.86

2.82

2.69

2.57

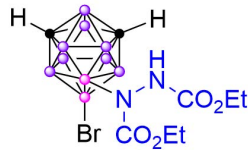
2.48

2.43

1.27

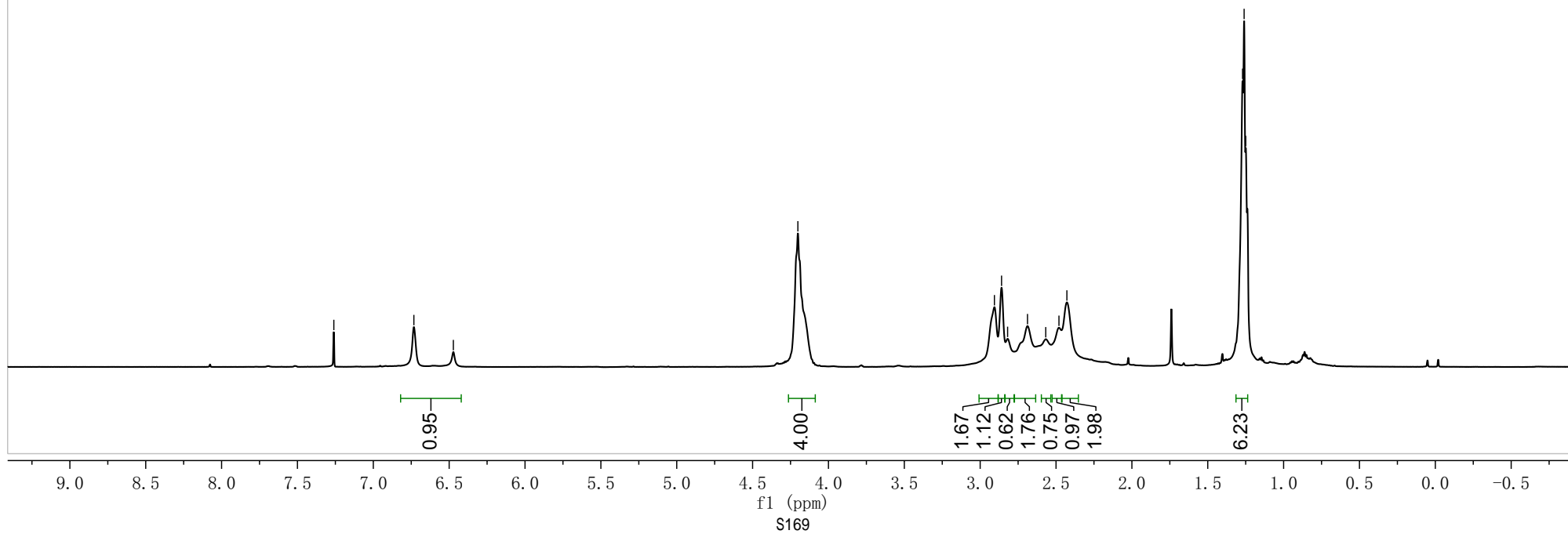
1.26

1.25

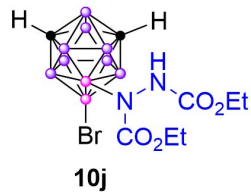


10j

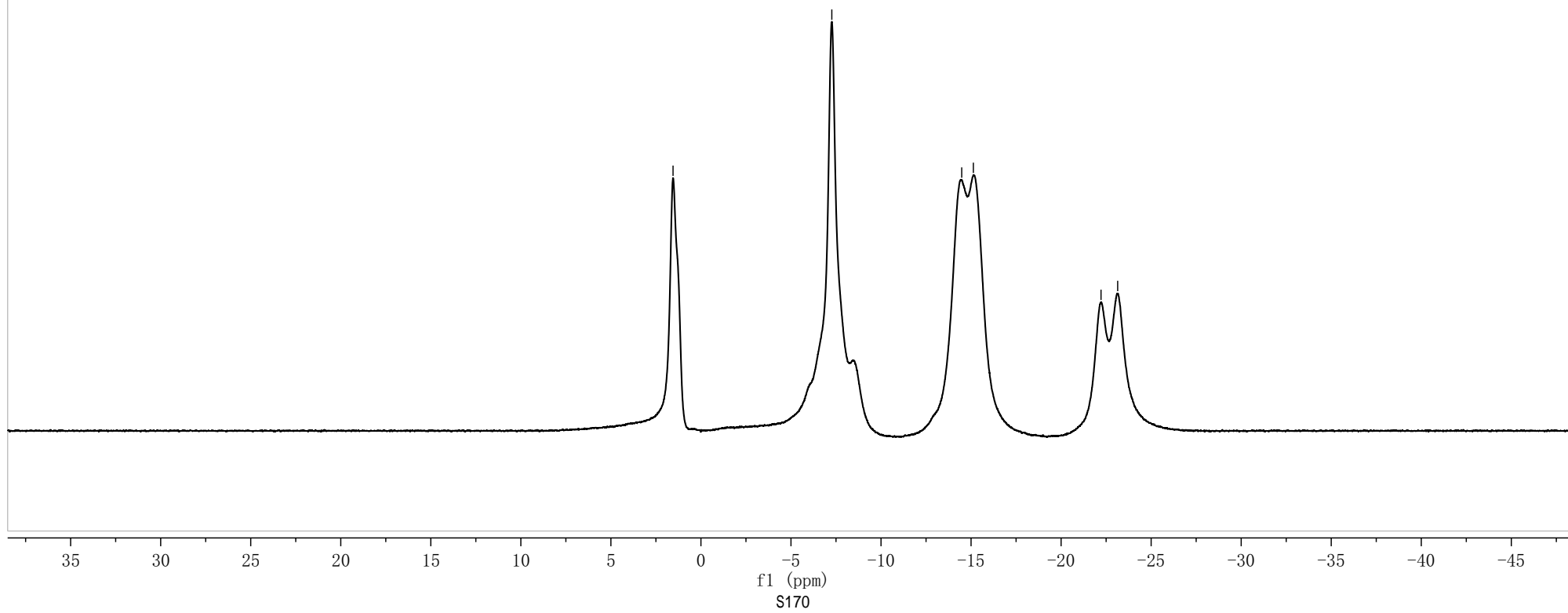
¹H{¹¹B} NMR (600 MHz, CDCl₃)



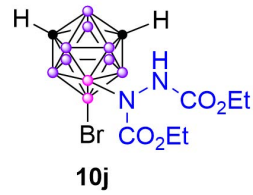
—1.55
—7.27
—14.48
—15.13
—22.22
—23.14



¹¹B NMR (193 MHz, CDCl₃)



—1.56 —7.22 —14.76 —22.66

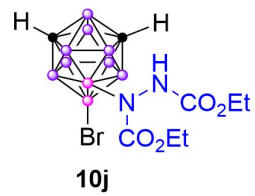


$^{11}\text{B}\{^1\text{H}\}$ NMR (193 MHz, CDCl_3)

1.18 2.99 4.00 1.95

f1 (ppm)
S171

35 30 25 20 15 10 5 0 -5 -10 -15 -20 -25 -30 -35 -40 -45



¹³C NMR (151 MHz, CDCl₃)

157.78
156.55

77.21
77.00
76.79

62.67
61.75

49.24
48.94

14.43
14.24

