Supporting Information

Iridium-Catalyzed Asymmetric Hydrogenation of Tetrahydro-γ-Carboline: A Versatile Approach to Chiral *cis*-Hexahydro-γ-Carboline Derivatives Compatible with C6-Substituted Carbolines

Bowen Liu^a, Chun Zhang*,^a, Xiuxiu Li*,^b, Xumu Zhang*,^b

a Institute of Molecular Plus, Tianjin Key Laboratory of Molecular Optoelectronic Science, Department of Chemistry, School of Science, Tianjin University, Tianjin 300072, China
b Shenzhen Grubbs Institute, Department of Chemistry, Shenzhen Key Laboratory of Small Molecule Drug Discovery and Synthesis, and Medi-Pingshan, Southern University of Science and Technology, Shenzhen 518055, People's Republic of China

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I General Information

Unless extra indicated, the raw materials used in this article are commercially available. All reactions and manipulations involving air or humidity-sensitive compounds are carried out in the glove box. Unless otherwise specified, oil baths are used for heating in the reaction. Reactions were detected and analyzed by TLC, and fluorescence was observed with ultraviolet light (254 nm). In the NMR hydrogen spectroscopy, the chemical shift value of deuterated chloroform-*d* was used as a reference of 7.26 ppm to calibrate the chemical shift of the compounds. In the NMR carbon spectrum, the chemical shift of deuterated chloroform was used as a reference of 77.00 ppm or the chemical shift of deuterated DMSO-*d* of 39.50 ppm as a reference. The following letters indicate the splitting of multiple peaks: S-single peak, D-double peak, DD-double doublet, T-triplet peak, Q-quadruple peak, M-multiple peak.The determination of the enantiomer excess percentage of the compounds was performed using chiral HPLC analysis using Agilent. HPLC analysis of compounds was performed in OD-3, OD-H and AD-3 (polysaccharide derivative normal-phase coated chiral column) columns using hexane and isopropanol as eluents.

II Optimization of Reaction Conditions

N Br 1a	-OEt H ₂ (40 atm Rh-ZhaoPhos (1.0 DCM, rt, 24 TsOH (1.05 ed) D mol%) h quiv.) Br	O OEt
Entry	[Rh]-salt	Yield (%) ^b	ee (%) ^c
1	Rh(NBD) ₂ BF ₄	0	
2	[Rh(COD)Cl] ₂	0	
3	Rh(COD) ₂ BF ₄	0	
4	[RhOAc] ₂	0	

Table S1: The research of different [Rh]-salts ^a

^{*a*} **1a** (0.1 mmol, 1.0 equiv.), TsOH (0.105 mmol, 1.05 equiv.), [Rh]-salt (1 mol%), ZhaoPhos(1.05 mol%), DCM (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} ¹**H** NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC.

Table S2: The effect of different solvents ^{*a*}

O OF OF OF OF OF OF OF OF OF OF OF OF OF O	Et H ₂ (40 at Ir-ZhaoPhos (* Solvent, rt, TsOH (1.05	tm) 1.0 mol%) 24 h equiv.) Br 2	O O O E t A a
Entry	Solvent	Yield (%) ^b	ee (%) °
1	DCM	70	69
2	DCE	72	67
3	EtOAc	51	40
4	EtOH	Trace	
5	MeOH	Trace	
6	HFIP	0	
7	THF	Trace	
8^d	DCM	N.P.	

^{*a*} **1a** (0.1 mmol, 1.0 equiv.), TsOH (0.105 mmol, 1.05 equiv.), $[Ir(COD)CI]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), Solvent (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} ¹H NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC; ^{*d*} BINAP ligand was used instead of ZhaoPhos.

O N Br 1a	−OEt H ₂ (40 atm) Ir-ZhaoPhos (1.0 DCM, rt, 24 TsOH (1.05 eq) mol%) h uiv.) Br	O O O O Et N H Za
Entry	Concentration	Yield (%) ^b	ee (%) ^c
1	DCM (0.1 M)	71	69
2	DCM (0.05 M)	57	69
3 ^{<i>d</i>}	DCM (0.05 M)	83	49
4	DCM (0.2 M)	40	68

^{*a*} **1a** (0.1 mmol,1.0 equiv.), TsOH (0.105 mmol,1.05 equiv.), $[Ir(COD)CI]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), 25 °C, H₂ (40 atm),stir for 24 h; ^{*b*} ¹**H NMR** yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC; ^{*d*} N-Me-ZhaoPhos ligand was used instead of ZhaoPhos.

Table S4: The effect of salt additives ^{*a*}

O N Br 1a	-OEt H ₂ (40 atr Ir-ZhaoPhos (* additive (5 m DCM (0.1 M), TsOH (1.05	n) I mol%) nol%) rt, 24 h equiv.) Br 2	o OEt
Entry	Additive	Yield (%) ^b	ee (%) ^c
1	Fe(OTf) ₂	28	69
2	Sm(OTf) ₃	15	69
3	AgNO ₃	0	
4	AgSbF ₆	0	
5	AgOTf	0	
6	AgMeSO ₃	0	

^{*a*} **1a** (0.1 mmol,1.0 equiv.), TsOH (0.105 mmol,1.05 equiv.), $[Ir(COD)CI]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), additive (5 mol%), DCM (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} **1H NMR** yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC.

Table S5: The effect of acid additives ^a

O N Br 1a	OEt H ₂ (40 atr Ir-ZhaoPhos (1 DCM(0.1 M), Acid	n) .0 mol%) rt, 24 h Br	O O O Et
Entry	Acid	Yield (%) ^b	ee (%) °
1	no TsOH	0	
2	TsOH	84	65
3	TFA	27	72
4^d	TFA	24	75
5	TfOH	trace	
6	MsOH	47	57

^{*a*} **1a** (0.1 mmol,1.0 equiv.), Acid (0.15 mmol,1.50 equiv.), $[Ir(COD)CI]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), DCM (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} ¹**H** NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC; ^{*d*} TFA (0.105 mmol,1.05 equiv.).

Table S6: Preliminary attempts at chlorinated substrates (1b) ^a

O N CI 1b	OEt H ₂ (40 atm) Ir-ZhaoPhos (1.0 r DCM (0.1 M), rt, TFA (1.5 equir	nol%) 24 h /.) Cl	O O O Et N H 2b
Entry	Deviation	Yield (%) ^b	ee (%) ^c
1	none	30	84
2	TsOH (1.5 equiv.)	91	65
3^d	TsOH (1.5 equiv.)	96	52
4^e	DCM (0.15 M)	33	80
5 ^f	DCM (0.05 M)	17	78
6^g	TFA and TsOH	28	77

^{*a*} **1b** (0.1 mmol,1.0 equiv.), TFA (0.15 mmol,1.50 equiv.), [Ir(COD)Cl]₂ (0.5 mol%), ZhaoPhos (1.05 mol%), DCM (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} ¹**H** NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC; ^{*d*} N-Me-ZhaoPhos ligand was used instead of ZhaoPhos; ^{*e*} DCM (0.15 M); ^{*f*} DCM (0.05 M); ^{*g*} TFA (0.15 mmol,1.5 equiv.) and TsOH (0.15 mmol,1.5 equiv.) were involved in reaction at the same time.

	OEt H ₂ (40 atr Ir-ZhaoPhos (1. Solvent, rt, 1 TFA (1.5 ec	n) 0 mol%) 24 h quiv.) Cl	O OEt N N H 2b
Entry	Solvent	Yield (%) ^b	ee (%) ^c
1	DCE	17	85
2	THF	Trace	
3	1,4-dioxane	Trace	
4	EtOAc	Trace	
5	MeCN	0	

Table S7: Re-optimization of the reaction solvent ^a

^{*a*} **1b** (0.1 mmol,1.0 equiv.), TFA (0.15 mmol,1.50 equiv.), $[Ir(COD)Cl]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), Solvent (0.1 M), 25 °C, H₂ (40 atm), stir for 24 h; ^{*b*} ¹**H** NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC.

Table S8: The effect of temperature and pressure ^a

O OEt	H₂ Ir-ZhaoPhos (1.0 mol DCM, 30 °C, 24 h TFA (3.0 eq)	%) CI 2	O OEt
Entry	Pressure	Yield (%) ^b	ee (%) ^c
1	H ₂ (40 atm)	57	87
2	H ₂ (65 atm)	57	85
3	H ₂ (80 atm)	58	84
4^d	H ₂ (80 atm)	64	78

^{*a*} **1b** (0.1 mmol,1.0 equiv.), TFA (0.30 mmol,3.0 equiv.), $[Ir(COD)Cl]_2$ (0.5 mol%), ZhaoPhos (1.05 mol%), DCM (0.1 M), 25 °C, stir for 24 h; ^{*b*} ¹**H** NMR yield, Trimethoxybenzene was used as the internal standard; ^{*c*} The ee (%) was determined by HPLC; ^{*d*} The reaction temperature is 50 °C.

Table S9: Optimization of the Reaction Conditions of $1a^{a}$

N Br 1a	–OEt D	H ₂ (40 atm) (haoPhos(1.0 mol%) CM (0.1 M), rt, 24 h Acid (1.05 eq)	► Br	O OEt
entry	acid	deviation	$\operatorname{conv.}^{b}(\%)$	ee ^c (%)
1	TsOH	none	79	73
2	TsOH	Rh(NBD) ₂ BF ₄	N.R.	/
3	TsOH	[Rh(COD)Cl]2	N.R.	/
4	TsOH	DCE	72	67
5	TsOH	EtOAc	51	40
6	TsOH	THF	trace	/
7	TsOH	HFIP	N.R.	/
8	TsOH	Fe(OTf) ₂	28	69
9	TsOH	Sm(OTf) ₃	15	69
10	TsOH	AgOTf	trace	/
11	no TsOH	none	N.R.	/
12^{d}	TsOH	none	84	65
13^{d}	TFA	none	27	72
14^d	TfOH	none	trace	/
15^{d}	MsOH	none	47	57
16 ^e	TFA	none	45	84

^{*a*}Unless otherwise specified, the reactions were conducted using **1a** (0.1 mmol) in 1.0 mL of solvent, 100 μ L Ir-ZhaoPhos solution (0.01 M, [Ir(COD)Cl]₂/(S,R)-ZhaoPhos = 1/2.1), Acid (1.05 eq). ^{*b*}The conversions were determined by ¹H NMR. ^{*c*}The ee was determined by performing chiral HPLC. ^{*d*}1.5 eq acid was used. ^{*e*}3.0 eq TFA was used.

III Experimental Section

1. General procedure for synthesis of tetrahydro-y-carboline 1



Phenylhydrazine substrate 4 and piperidone 5 used are commercially available raw materials. Phenyl hydrochloride 4 (20 mmol, 1.0 equiv.) was added to a 100 mL dry three-mouth flask filled with stirred magnets, followed by a reflux tube mounted on the three-mouth flask, the equipment was replaced three times in an argon atmosphere, then absolute ethanol (0.6 M) was added to the reaction flask through a syringe, piperidone 5 (20 mmol, 1.0 equiv.) was added to the vial through a syringe under stirring conditions, and the reaction tube was then transferred to 90 °C and maintained at this temperature for one day. Until the white solid turbidity appears in the reaction bottle, the reaction is cooled to room temperature and slowly quenched with water. After washing the white solids with n-hexane, the organic phase was concentrated under vacuum. The residue was purified by silica gel chromatography (EtOAc /petroleum ether) to afford the product 1.

2. General procedure for hydrogenation of 1.

1.0 equivalent 1 was selected as the substrate, 0.5 mol% 1,5-cyclooctadiene iridium chloride dimer ([Ir(COD)Cl]₂) was used as the metal catalyst, 1.05 mol% 4-F-Ph-ZhaoPhos was used as the ligand, 3.0 equivalent trifluoroacetic acid (TFA) was used as the acid additive in the reaction system, and dichloromethane (DCM, 0.1 M) was selected as the reaction solvent. The reaction was carried out at room temperature for 24 hours in a hydrogen atmosphere at 40 atmospheres. After the reaction, slowly open the gas valve of the high-pressure reactor, carefully release the high-pressure hydrogen in the system, and after the pressure gauge pointer is zero, open the reactor and take out the ampoule containing the reactants. The reaction was quenched with a saturated sodium bicarbonate solution and extracted with ethyl acetate or methylene chloride, followed by drying with anhydrous sodium sulfate. The organic phase is concentrated by the rotary evaporator, and then the concentrated organic phase is added with dilute hydrochloric acid for pickling, the product is easy to become salt in the hydrochloric acid solution, the aqueous phase is transferred after pickling, and the pH of the system

is adjusted to alkaline by adding sodium hydroxide solution or saturated sodium bicarbonate solution, and the water is extracted with ethyl acetate or dichloromethane, and the product can be obtained after concentrating the organic phase. Due to the similarity and polarity of the raw material and product in the ethyl acetate/petroleum ether system, the product 2 can also be purified by silica gel chromatography separation (PE/EtOAc v/v = 2:1). The absolute configuration can be determined by comparing the optical rotation of compound 2j with the value reported in the literature¹. The configurations of the other chiral products were then assigned by analogy.

3. General procedure for synthesis of compound rac-2



1 (0.2 mmol, 1.0 equiv.) was added to a 25 mL dry Shrek reaction tube equipped with a stirred magnet, then the reaction tube was replaced three times under argon atmosphere, 1.0 mL of dichloromethane (DCM) was added to the reaction tube through a syringe, and then trifluoroacetic acid (2 mmol, 10.0 equiv.) was slowly added to the reaction tube through a syringe, stirred until the raw material was completely dissolved, and triethylsilane (0.6 mmol, 3.0 equiv.), place the reaction at room temperature for 12 h. After the reaction, trifluoroacetic acid was neutralized with saturated sodium bicarbonate aqueous solution, then organic phase was extracted by dichloromethane, the organic phase was dried with anhydrous sodium sulfate, and the residue was purified by silica gel chromatography (PE/EtOAc v/v = 5:1) to obtain racemic product *rac-2*. Some of the low-reactive substrates are hydrogenated using two equal amounts of enantiomer metal ligand complexes to obtain racemic products, so there may be several samples that are not completely racemic and two enantiomers are not equal (50/50).

4. General procedure for synthesis of Compound 3a



In argon atmosphere, to a solution of 2a (790 mg, 2 mmol) in acetone (15 mL) were added 2-chloro-N-methylacetamide (258 mg, 2.4 mmol), KI (166 mg, 1 mmol), and K₂CO₃ (413 mg, 3 mmol). The reaction mixture was heated at 70 °C for 24 h, cooled to room temperature, and then the solid was removed by filtration through celite, and the filtrate was concentrated under vacuum. The residue was dissolved in CH₂Cl₂, washed by saturated NaCl, and then was dried over anhydrous Na2SO4, and the filtrate was concentrated under vacuum to give crude product 3a. The residue was purified by silica gel flash column chromatography (EA) to give the compound **3a**. $[\alpha]_D^{25} + 21.3$ (c 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.28 (d, *J* = 8.0 Hz, 1H), 7.09 (d, *J* = 6.4 Hz, 1H), 6.73 (t, J = 7.6 Hz, 1H), 4.39 (d, J = 17.9 Hz, 1H), 4.09 (s, 2H), 3.99 -3.14 (m, 8H), 2.91 (d, J = 4.9 Hz, 3H), 2.02 – 1.90 (m, 1H), 1.89 – 1.77 (m, 1H), 1.23 (s, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-d) δ 171.5, 155.7, 134.3, 133.6, 123.4, 122.5, 104.9, 65.6, 61.5, 55.3, 43.5, 40.9, 39.9, 26.1, 14.6 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak AD-3 column (0.46×25 cm), Hexane/ⁱPrOH = 85:15, flow rate = 1.0 mL/min, λ = 254 nm, t_R: 7.941 min (S, R) (major), 11.994 min (R, S) (minor). HRMS (ESI/ion trap) m/z: $[M + H]^+$ calcd for $C_{17}H_{22}BrN_{3}O_{3}^{+}$: 396.0917, found: 396.0918.

5. Characterization date of 1, 2 and 3a



Ethyl 6-bromo-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**1a**)**:** The procedure was followed using 2-bromobenzazine hydrochloride (**4a**, 4.44 g, 20 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 3.0 mL, 20 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1a** as a white solid (5.41 g, 84% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.15 (s, 1H), 7.39 (d, J = 7.8 Hz, 1H), 7.30 (d, J = 7.6 Hz, 1H), 6.98 (t, J = 7.8 Hz, 1H), 4.67 (s, 2H), 4.21 (q, J = 7.1 Hz, 2H), 3.88 (s, 2H), 2.88 (s, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 156.1, 134.5, 132.8, 126.7, 123.9, 120.8, 116.7, 108.5, 104.3, 61.6, 41.1, 23.3, 14.7 ppm. HRMS (ESI/ion trap) m/z: [M + H]⁺ calcd for C₁₄H₁₇BrN₂O₂⁺ : 323.0390, found: 323.0388.



Ethyl 6-chloro-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**1b**)**:** The procedure was followed using 2-chlorobenzazine hydrochloride (**4b**, 3.56 g, 20 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 3.0 mL, 20 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1b** as a white solid (4.50 g, 81% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.19 (s, 1H), 7.35 (d, J = 7.8 Hz, 1H), 7.15 (d, J = 7.6 Hz, 1H), 7.03 (t, J = 7.8 Hz, 1H), 4.68 (s, 2H), 4.21 (q, J = 7.1 Hz, 2H), 3.88 (s, 2H), 2.88 (s, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³**C NMR** (151 MHz, Chloroform-*d*) δ 156.1, 133.1, 127.0, 121.0, 120.4, 116.2, 108.4, 61.6, 41.1, 23.4, 14.7 ppm. HRMS (ESI/ion trap) m/z: [M + H]⁺ calcd for C₁₄H₁₅ClN₂O₂⁺ : 279.0895, found: 279.0892.



Methyl 6-chloro-1,3,4,5-tetrahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (1c): The procedure was followed using 2-chlorobenzazine hydrochloride (4c, 3.56 g, 20 mmol, 1.0 equiv.) and N-methoxycarbonyl-4-piperidone (5c, 3.14 g, 20 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product 1c as a white solid (3.60 g, 70% yield). ¹H NMR (600 MHz, Chloroform-d) δ 8.13 (s, 1H), 7.34 (d, J = 7.6 Hz, 1H), 7.15 (d, J = 7.6 Hz, 1H), 7.04 (t, J = 7.8 Hz, 1H), 4.67 (s, 2H), 3.89 (s, 2H), 3.77 (s, 3H), 2.88 (s, 2H) ppm. ¹³C NMR (151 MHz, Chloroform-d) δ 156.5, 133.1, 126.9, 121.1, 120.5, 116.1, 52.8, 41.2, 23.3 ppm. HRMS (ESI/ion trap) m/z: [M + H]+ calcd for C13H13ClN2O2⁺ : 265.0738, found: 265.0736.



Ethyl 6-fluoro-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**1d**)**:** The procedure was followed using 2-fluorobenzazine hydrochloride (**4d**, 3.24 g, 20 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 3.0 mL, 20 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1d** as a white solid (2.83 g, 54% yield). ¹**H NMR** (400 MHz, Chloroform-*d*) δ 8.16 (s, 1H), 7.21 (d, J = 7.9 Hz, 1H), 7.01 (td, J = 7.9, 4.8 Hz, 1H), 6.87 (dd, J = 11.2, 7.9 Hz, 1H), 4.68 (s, 2H), 4.21 (q, J = 7.1 Hz, 2H), 3.87 (s, 2H), 2.87 (t, J = 5.8 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³**C NMR** (101 MHz, DMSO-*d*₆) δ 155.1, 150.1, 147.7, 134.0, 129.0, 123.3 (d, J = 13.1 Hz), 119.0 (d, J = 6.3 Hz), 113.5 (d, J = 2.2 Hz), 106.7, 105.7 (d, J = 16.3 Hz), 60.9, 40.9, 23.2, 14.6 ppm. ¹⁹**F NMR** (376 MHz, Chloroform-*d*) δ -135.12 ppm.



Ethyl 6-methyl-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate (1e):** The procedure was followed using *o*-tolylhydrazine hydrochloride (**4e**, 3.16 g, 20 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 3.0 mL, 20 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1e** as a white solid (4.79 g, 93% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.11 (s, 1H), 7.33 (d, J = 7.8 Hz, 1H), 7.05 (t, J = 7.5 Hz, 1H), 6.99 (d, J = 7.1 Hz, 1H), 4.72 (d, J = 8.3 Hz, 2H), 4.24 (q, J = 7.1 Hz, 2H), 3.89 (q, J = 10.9, 8.7 Hz, 2H), 2.87 (t, J = 5.6 Hz, 2H), 2.49 (s, 3H), 1.33 (t, J = 7.1 Hz, 3H) ppm. ¹³**C NMR** (151 MHz, Chloroform-*d*) δ 156.2, 135.3, 131.7, 124.9, 122.2, 119.7, 115.1, 107.5, 61.5, 41.3, 23.3, 16.6, 14.7 ppm. HRMS (ESI/ion trap) m/z: [M + H]⁺ calcd for C₁₅H₁₈N₂O₂⁺ : 259.1441, found: 259.1439.



Ethyl 6-methoxy-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate (1f):** The procedure was followed using 2-methoxybenzazine hydrochloride (**4f**, 1.74 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1f** as a white solid (1.86 g, 68% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.24 (s, 1H), 7.08 (d, J = 7.9 Hz, 1H), 7.03 (t, J = 7.8 Hz, 1H), 6.64 (d, J = 7.6 Hz, 1H), 4.69 (s, 2H), 4.22 (q, J = 7.1 Hz, 2H), 3.95 (s, 3H), 3.88 (s, 2H), 2.83 (t, J = 5.8 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³**C NMR** (151 MHz, Chloroform-*d*) δ 156.2, 145.7, 131.5, 126.7, 126.0, 120.0, 110.4, 107.5, 101.9, 61.5, 55.3, 41.3, 23.3, 14.7 ppm. HRMS (ESI/ion trap) *m/z*: [M + H]⁺ calcd for C₁₅H₁₈N₂O₃⁺ : 275.1390, found: 275.1387.



Ethyl 6-ethyl-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b***]indole-2-carboxylate (1g):** The procedure was followed using 2-ethylbenzazine hydrochloride (**4g**, 1.7 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1g** as a white solid (2.06 g, 76% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.46 (s, 1H), 7.36 (s, 1H), 7.13 (s, 1H), 7.07 (s, 1H), 4.77 (s, 2H), 4.28 (q, *J* = 7.1 Hz, 2H), 3.93 (s, 2H), 2.92 – 2.87 (m, 4H), 1.40 (t, *J* = 7.6 Hz, 3H), 1.37 (t, *J* = 7.1 Hz, 3H) ppm. ¹³**C NMR** (151 MHz, Chloroform-*d*) δ 156.2, 134.6, 131.6, 126.3, 125.1, 120.2, 119.7, 115.1, 107.3, 61.5, 41.3, 24.0, 23.3, 14.6, 13.9 ppm. HRMS (ESI/ion trap) *m/z*: [M + H]⁺ calcd for C₁₆H₂₀N₂O₂⁺ : 273.1598, found: 273.1595.



Ethyl 6-isopropyl-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate (1h):** The procedure was followed using 2-isopropylbenzazine hydrochloride (**4h**, 1.86 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1h** as a white solid (1.4 g, 49% yield). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 8.16 (s, 1H), 7.33 (d, J = 7.6 Hz, 1H), 7.12 (t, J = 6.9 Hz, 1H), 7.08 (d, J = 6.9 Hz, 1H), 4.72 (s, 2H), 4.23 (q, J = 7.1 Hz, 2H), 3.91 (s, 2H), 3.24 (p, J = 6.6 Hz, 1H), 2.89 (t, J = 5.1 Hz, 2H), 1.40 (d, J = 6.9 Hz, 6H), 1.33 (t, J = 7.1 Hz, 3H) ppm. ¹³C **NMR** (151 MHz, Chloroform-*d*) δ 156.2, 134.0, 131.5, 131.0, 125.3, 119.9, 117.7, 115.1, 107.6, 61.5, 41.3, 29.3, 23.4, 22.7, 14.7 ppm. HRMS (ESI/ion trap) m/z: [M + H]⁺ calcd for C₁₇H₂₂N₂O₂⁺ : 287.1754, found: 287.1751.



Ethyl 8-methyl-1,3,4,5-tetrahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (1i): The procedure was followed using *p*-tolylbenzazine hydrochloride (4i, 1.58 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (5, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product 1i as a white solid (2.45 g, 95% yield). ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.76 (s, 1H), 7.20 – 7.18 (m, 2H), 6.89 – 6.84 (m, 1H), 4.57 (s, 2H), 4.11 (q, *J* = 7.1 Hz, 2H), 3.75 (t, *J* = 5.5 Hz, 2H), 2.78 (t, *J* = 5.5 Hz, 2H), 2.36 (s, 3H), 1.23 (t, *J* = 7.1 Hz, 3H) ppm. ¹³C NMR (151 MHz, DMSO-*d*₆) δ 155.2, 134.2, 132.5, 127.0, 125.4, 122.1, 116.9, 110.6, 105.0, 60.8, 41.00, 22.9, 21.2, 14.7 ppm.



Ethyl 1,3,4,5-tetrahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (1j): The procedure was followed using benzazine hydrochloride (4j, 1.44 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (5, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product 1j as a white solid (2.14 g, 88% yield). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.87 (brs, 1H), 7.46 (d, J = 7.8 Hz, 1H), 7.31 (d, J = 8.0 Hz, 1H), 7.16 (t, J = 7.6 Hz, 1H), 7.11 (t, J = 7.4 Hz, 1H), 4.70 (s, 2H), 4.20 (q, J = 7.1 Hz, 2H), 3.88 (s, 2H), 2.85 (s, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 156.3, 134.7, 133.0, 126.8, 124.1, 120.9, 116.8, 108.5, 104.4, 61.8, 41.3, 23.5, 14.9 ppm.



Ethyl 8-fluoro-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**1k**)**:** The procedure was followed using 4-fluorobenzazine hydrochloride (**4k**, 1.62g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1k** as a white solid (2.38 g, 91% yield). **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.94 (s, 1H), 7.21 (dd, J = 8.8, 4.3 Hz, 1H), 7.09 (dd, J = 9.4, 2.5 Hz, 1H), 6.89 (td, J = 9.1, 2.5 Hz, 1H), 4.65 (s, 2H), 4.21 (q, J = 7.1 Hz, 2H), 3.86 (s, 2H), 2.84 (t, J = 5.8 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.7 (d, J = 231.0 Hz), 155.1, 134.8, 132.5, 125.3 (d, J = 10.1 Hz), 111.7 (d, J = 9.8 Hz), 108.4 (d, J = 25.8 Hz), 105.9, 102.3 (d, J = 23.4 Hz), 60.9, 40.9, 23.0, 14.6 ppm. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -124.38 ppm.



Ethyl 8-chloro-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate (11):** The procedure was followed using 4-chlorobenzazine hydrochloride (**4I**, 1.78 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1I** as a white solid (2.36 g, 85% yield). ¹**H NMR** (400 MHz, Chloroform-*d*) δ 8.03 (s, 1H), 7.41 (s, 1H), 7.21 (d, J = 8.6 Hz, 1H), 7.10 (dd, J = 8.6, 2.0 Hz, 1H), 4.64 (s, 2H), 4.20 (q, J = 7.1 Hz, 2H), 3.86 (s, 2H), 2.83 (t, J = 5.7 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. ¹³**C NMR** (101 MHz, DMSO-*d*₆) δ 155.1, 134.3, 126.2, 123.2, 120.4, 116.7, 112.3, 105.6, 60.8, 40.8, 22.8, 14.6 ppm.



Ethyl 8-bromo-1,3,4,5-tetrahydro-*2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**1m**): The procedure was followed using 4-bromobenzazine hydrochloride (**4m**, 2.21 g, 10 mmol, 1.0 equiv.) and N-ethoxycarbonyl-4-piperidone (**5**, 1.5 mL, 10 mmol, 1.0 equiv.). Purification using condition of silica gel chromatography afforded product **1m** as a white solid (2.16 g, 67% yield). **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.02 (s, 1H), 7.57 (s, 1H), 7.23 (dd, J = 8.6, 1.9 Hz, 1H), 7.17 (d, J = 8.6 Hz, 1H), 4.64 (s, 2H), 4.20 (q, J = 7.1 Hz, 2H), 3.86 (s, 2H), 2.83 (t, J = 5.8 Hz, 2H), 1.31 (t, J = 7.1 Hz, 3H) ppm. **¹³C NMR** (101 MHz, DMSO-*d*₆) δ 155.1, 134.5, 126.9, 123.0, 119.7, 112.8, 111.1, 105.5, 60.9, 40.8, 22.8, 14.6 ppm.



Ethyl (4a*S*,9b*R*)-6-bromo-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2a): The procedure was followed using 1a (32 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2a as an oily yellow solid (29.8 mg, 92% yield). [α]_D²⁵ + 55.0 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.19 (d, *J* = 8.0 Hz, 1H), 7.04 (d, *J* = 7.2 Hz, 1H), 6.59 (t, *J* = 7.6 Hz, 1H), 4.16 – 4.08 (m, 2H), 4.03 (q, *J* = 5.5 Hz, 1H), 4.00 – 3.72 (m, 2H), 3.61 – 3.53 (m, 1H), 3.48 – 3.21 (m, 3H), 1.95 – 1.87 (m, 1H), 1.83 – 1.74 (m, 1H), 1.25 (brs, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 149.3, 131.5, 130.5, 123.0, 120.1, 103.7, 61.3, 56.9, 43.6, 41.8, 39.6, 27.8, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, λ = 210 nm, t_{*R*}: 7.961 min (*S*, *R*) (major), 12.177 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₄H₁₇BrN₂O₂⁺ : 325.0546, found: 325.0545.



Ethyl (4aS,9bR)-6-chloro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2b): The procedure was followed using 1b (28 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2b as an oily yellow solid (24.3 mg, 87% yield). [α]_D²⁵ + 41.3 (*c* 0.3, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.04 (d, *J* = 8.1 Hz, 1H), 7.00 (d, *J* = 7.3 Hz, 1H), 6.66 (t, *J* = 7.7 Hz, 1H), 4.18 – 4.08 (m, 2H), 4.03 (dt, *J* = 9.6, 4.9 Hz, 1H), 3.94 – 3.17 (m, 6H), 1.96 – 1.88 (m, 1H), 1.83 – 1.74 (m, 1H), 1.25 (brs, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 147.8, 131.7, 127.7, 122.4, 119.7, 115.3, 61.3, 57.3, 43.6, 41.6, 39.6, 27.8, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, λ = 254 nm, t_R: 9.468 min (*S*, *R*) (major), 12.566 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₄H₁₇ClN₂O₂⁺ : 281.1051, found: 281.1048.



Methyl (4aS,9bR)-6-chloro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2c): The procedure was followed using 1c (26.4 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2c as an oily yellow solid (25 mg, 95% yield). [α]_D²⁵ + 66.5 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.05 (d, J = 8.1 Hz, 1H), 7.00 (s, 1H), 6.66 (t, J =7.6 Hz, 1H), 4.12 – 3.95 (m, 2H), 3.94 – 3.75 (m, 1H), 3.69 (s, 3H), 3.59 (s, 1H), 3.48 – 3.40 (m, 1H), 3.40 – 3.21 (m, 2H), 1.92 (s, 1H), 1.85 – 1.75 (m, 1H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 156.0, 147.8, 131.6, 127.8, 122.5, 119.8, 115.4, 57.2, 52.6, 43.8, 41.6, 39.7, 27.9 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, $\lambda = 254$ nm, t_{*R*}: 12.083 min (*S*, *R*) (major), 16.736 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₃H₁₅ClN₂O₂⁺ : 267.0895, found: 267.0893.



Ethyl (4aS,9bR)-6-fluoro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2d): The procedure was followed using 1d (26 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2d as an oily yellow solid (23.7 mg, 90% yield). $[\alpha]_D^{25}$ + 52.5 (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 6.92 (d, *J* = 7.3 Hz, 1H), 6.84 (t, 1H), 6.67 (td, *J* = 7.8, 4.6 Hz, 1H), 4.13 (qq, *J* = 7.2, 3.6 Hz, 2H), 4.04 (dt, *J* = 7.1, 4.9 Hz, 1H), 3.97 – 3.70 (m, 2H), 3.63 – 3.54 (m, 1H), 3.50 – 3.40 (m, 1H), 3.31 (brs, 2H), 1.92 (ddt, *J* = 14.2, 9.4, 4.6 Hz, 1H), 1.79 (dq, *J* = 14.4, 4.9 Hz, 1H), 1.29 – 1.23 (m, 3H) ppm. ¹³C NMR (101 MHz, Chloroform-*d*) δ 155.6, 150.5, 148.1, 137.6 (d, *J* = 12.7 Hz), 133.8, 119.6 (d, *J* = 19.8 Hz), 114.6 (d, *J* = 17.4 Hz), 61.2, 58.2, 43.6, 41.2, 39.6, 27.9, 14.7 ppm. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -135.4 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak AD-3 column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, λ = 254 nm, t_{*R*}: 6.770 min (*R*, *S*) (minor), 7.625 min (*S*, *R*) (major).



Ethyl (4aS,9bR)-6-methyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2e): The procedure was followed using 1e (25.8 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2e as an oily yellow solid (24.7 mg, 95% yield). [α]D²⁵ + 29.9 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 6.99 (d, J = 7.3 Hz, 1H), 6.90 (d, J = 7.5 Hz, 1H), 6.68 (t, J = 7.4 Hz, 1H), 4.19 – 4.08 (m, 2H), 3.98 (dt, J = 7.3, 4.9 Hz, 1H), 3.95 – 3.73 (m, 1H), 3.65 – 3.12 (m, 5H), 2.14 (s, 3H), 1.92 (td, J = 9.4, 4.7 Hz, 1H), 1.82 – 1.76 (m, 1H), 1.26 (s, 1H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 149.3, 129.4, 128.9, 121.6, 119.1, 61.2, 57.2, 43.9, 41.1, 39.7, 28.1, 16.7, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 90:10, flow rate = 1.0 mL/min, $\lambda = 210$ nm, t_{*R*}: 14.152 min (*S*, *R*) (major), 15.283 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₅H₁₈N₂O₂⁺ : 261.1598, found: 261.1595.



Ethyl (4aS,9bR)-6-methoxy-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2f): The procedure was followed using 1f (27 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2f as an oily yellow solid (20 mg, 73% yield). [α]D²⁵ + 7.9 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 6.80 (d, J = 7.3 Hz, 1H), 6.73 (t, J = 7.7 Hz, 1H), 6.69 (d, J = 7.7 Hz, 1H), 4.17 – 4.09 (m, 2H), 4.06 – 3.88 (m, 2H), 3.83 (s, 3H), 3.82 – 3.56 (m, 2H), 3.48 – 3.38 (m, 1H), 3.36 – 3.04 (m, 2H), 1.95 – 1.87 (m, 1H), 1.84 – 1.74 (m, 1H), 1.26 (brs, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 145.6, 139.6, 131.5, 119.7, 116.7, 109.8, 77.6 – 76.3 (m), 61.2, 57.9, 55.3, 44.0, 41.4, 39.7, 28.0, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, $\lambda = 210$ nm, t_{*R*}: 10.443 min (*S*, *R*) (major), 20.118 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₅H₂₀N₂O⁺: 277.1547, found: 277.1544.



Ethyl (4aS,9bR)-6-ethyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2g): The procedure was followed using 1g (27 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2g as an oily yellow solid (24.6 mg, 90% yield). [α]D²⁵ + 30.3 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.00 (d, J = 7.3 Hz, 1H), 6.94 (d, J = 7.5 Hz, 1H), 6.73 (t, J = 7.4 Hz, 1H), 4.18 – 4.08 (m, 2H), 3.98 (dt, J = 7.4, 5.0 Hz, 1H), 3.94 – 3.18 (m, 6H), 2.49 (q, J = 7.6 Hz, 2H), 1.91 (ddt, J = 14.0, 9.3, 4.7 Hz, 1H), 1.80 – 1.74 (m, 1H), 1.31 – 1.20 (m, 6H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 148.6, 129.4, 126.8, 125.3, 121.6, 119.1, 61.1, 57.2, 43.8, 41.1, 39.7, 28.1, 23.9, 14.7, 13.2 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 90:10, flow rate = 1.0 mL/min, $\lambda = 254$ nm, t_R: 10.727 min (*R*, *S*) (minor), 13.120 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₆H₂₂N₂O₂⁺ : 275.1754, found: 275.1751.



Ethyl (4aS,9bR)-6-isopropyl-1,3,4,4a,5,9b-hexahydro-*2H*-pyrido[4,3-*b*]indole-2carboxylate (2h): The procedure was followed using 1h (28 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2h as an oily yellow solid (22 mg, 77% yield). [α]_D²⁵ + 13.6 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.01 – 6.98 (m, 2H), 6.75 (t, *J* = 7.5 Hz, 1H), 4.13 (qq, *J* = 10.6, 7.1 Hz, 2H), 3.97 (dt, *J* = 7.3, 5.1 Hz, 1H), 3.94 – 3.20 (m, 6H), 2.81 (hept, *J* = 6.9 Hz, 1H), 1.95 – 1.87 (m, 1H), 1.80 – 1.73 (m, 1H), 1.30 – 1.22 (m, 9H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.7, 148.1, 130.1, 124.1, 121.6, 119.3, 61.2, 57.2, 43.8, 41.2, 39.9, 28.9, 28.2, 22.2, 22.2, 14.8 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/ⁱPrOH = 90:10, flow rate = 1.0 mL/min, λ = 254 nm, t_{*R*}: 6.447 min (*S*, *R*) (major), 7.545 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m*/*z*: [M + H]⁺ calcd for C₁₇H₂₄N₂O₂⁺ : 289.1911, found: 289.1907.



Ethyl (4aS,9bR)-8-methyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2i): The procedure was followed using 1i (26 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2i as an oily yellow solid (25.7 mg, 99% yield). [α]_D²⁵ + 53.9 (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 6.95 (s, 1H), 6.86 (d, *J* = 7.8 Hz, 1H), 6.58 (d, *J* = 7.8 Hz, 1H), 4.21 – 4.05 (m, 2H), 3.98 – 3.91 (m, 1H), 3.87 – 3.10 (m, 6H), 2.25 (s, 3H), 1.88 (tt, *J* = 9.3, 4.7 Hz, 1H), 1.82 – 1.67 (m, 1H), 1.32 – 1.22 (m, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.6, 148.3, 130.3, 128.3, 124.9, 109.9, 61.2, 57.6, 43.9, 40.9, 39.8, 28.0, 20.8, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, λ = 254 nm, t_R: 5.821 min (*S*, *R*) (major), 9.116 min (*R*, *S*) (minor).



Ethyl (4aS,9bR)-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (2j): The procedure was followed using 1j (24 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2j as an oily yellow solid (23 mg, 95% yield). $[\alpha]_D^{25}$ + 75.0 (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.13 (d, J = 7.2Hz, 1H), 7.05 (td, J = 7.6, 1.2 Hz, 1H), 6.73 (td, J = 7.4, 0.8 Hz, 1H), 6.66 (d, J = 7.8Hz, 1H), 4.20 – 4.06 (m, 2H), 3.97 (dt, J = 6.8, 4.9 Hz, 1H), 3.96 – 3.64 (m, 2H), 3.63 – 3.53 (m, 1H), 3.49 – 3.11 (m, 3H), 1.90 (ddt, J = 14.1, 9.3, 4.6 Hz, 1H), 1.76 (ddd, J = 14.4, 9.3, 5.3 Hz, 1H), 1.26 (t, J = 6.6 Hz, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.7, 150.9, 130.1, 128.1, 124.3, 119.1, 110.0, 61.3, 57.5, 43.9, 41.0, 39.8, 28.1, 14.8 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, λ = 210 nm, t_{*R*}: 10.812 min (*R*, *S*) (minor), 11.642 min (*S*, *R*) (major).



Ethyl (4aS,9bR)-8-fluoro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2k): The procedure was followed using 1k (26 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2k as an oily yellow solid (23 mg, 87% yield). [α]_D²⁵ + 60.0 (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 6.85 (dd, J = 8.2, 2.6 Hz, 1H), 6.74 (td, J = 8.9, 2.6 Hz, 1H), 6.56 (dd, J = 8.5, 4.3 Hz, 1H), 4.20 – 4.05 (m, 2H), 4.02 – 3.93 (m, 1H), 3.93 – 3.67 (m, 1H), 3.69 – 3.11 (m, 5H), 1.94 – 1.82 (m, 1H), 1.77 – 1.68 (m, 1H), 1.35 – 1.17 (m, 3H) ppm. ¹³C NMR (101 MHz, Chloroform-*d*) δ 157.0 (d, J = 23.0 Hz), 155.6, 146.7, 114.0 (d, J = 23.3 Hz), 111.7 (d, J = 24.0 Hz), 110.2 (d, J = 8.0 Hz), 61.3, 58.0, 43.6, 41.1, 39.7, 27.9, 14.7 ppm. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ -125.6 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/¹PrOH = 85:15, flow rate = 1.0 mL/min, $\lambda = 254$ nm, t_R: 9.008 min (*R*, *S*) (minor), 14.061 min (*S*, *R*) (major).



Ethyl (4aS,9bR)-8-chloro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2l): The procedure was followed using 1l (27.8 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2l as an oily yellow solid (27 mg, 98% yield). [α]_D²⁵ + 31.0 (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.07 (d, J = 2.1 Hz, 1H), 6.99 (dd, J = 8.3, 2.2 Hz, 1H), 6.55 (d, J = 8.2 Hz, 1H), 4.21 – 4.04 (m, 2H), 4.02 – 3.93 (m, 1H), 3.93 – 3.63 (m, 2H), 3.59 – 3.10 (m, 4H), 1.94 – 1.82 (m, 1H), 1.72 (s, 1H), 1.25 (s, 3H) ppm. ¹³C NMR (151 MHz, Chloroform-*d*) δ 155.5, 149.3, 131.6, 127.7, 124.4, 123.4, 110.6, 61.3, 57.7, 43.4, 41.0, 39.7, 27.8, 14.6 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 85:15, flow rate = 1.0 mL/min, $\lambda = 210$ nm, t_{*R*}: 9.429 min (*R*, *S*) (minor), 19.070 min (*S*, *R*) (major).



Ethyl (4aS,9bR)-8-bromo-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (2m): The procedure was followed using 1m (32 mg, 0.1 mmol, 1.0 equiv.) under H₂ (40 atm). Purification using condition of silica gel chromatography afforded product 2m as an oily yellow solid (28.7 mg, 89% yield). [α]_D²⁵ + 21.0 (*c* 1.0, CHCl₃). ¹H NMR (600 MHz, Chloroform-*d*) δ 7.21 (d, *J* = 2.0 Hz, 1H), 7.13 (dd, *J* = 8.2, 2.0 Hz, 1H), 6.52 (d, *J* = 8.2 Hz, 1H), 4.19 – 4.07 (m, 2H), 4.01 – 3.95 (m, 1H), 3.92 – 3.64 (m, 2H), 3.62 – 3.10 (m, 4H), 1.93 – 1.84 (m, 1H), 1.70 (s, 1H), 1.34 – 1.20 (m, 3H) ppm. ¹³C NMR (101 MHz, Chloroform-*d*) δ 155.5, 149.8, 132.2, 130.6, 127.2, 111.2, 110.4, 61.3, 57.6, 43.4, 41.0, 39.6, 27.8, 14.7 ppm. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralcel OD-H column (0.46 × 25 cm), Hexane/^{*i*}PrOH = 80:20, flow rate = 1.0 mL/min, λ = 254 nm, t_{*R*}: 7.589 min (*R*, *S*) (minor), 14.031 min (*S*, *R*) (major).

VI NMR Spectrum



-8.19 -8.19 -8.17 -7.15 7.15 7.15 7.03 7.03 7.02 7.03 -2.88 -2.88 -2.88 -2.88 -2.88 -0.00





































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V Chiral HPLC analysis





Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
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Use	Multiplier	&	Dilution	Factor	with	ISTD

Signal 1: DAD1 A, Sig=210,4 Ref=360,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	7.961	BB	0.4346	1.28227e4	384.99167	93.0155
2	12.177	MM	0.6223	962.84650	25.78707	6.9845





Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1 2	9.468	MM	0.3168	591.44482	31.11912	93.4760
	12.566	MM	0.4141	41.27860	1.66140	6.5240











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Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier 8	Dilution	Factor with	ISTDs

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	10.695	VB	0.3892	1.25151e4	491.31598	51.1826
2	13.001	BB	0.4708	1.19368e4	392.49207	48.8174





Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	6.537	BB	0.1908	5444.59473	444.97366	47.8712
2	7.630	BB	0.2244	5928.82080	410.85803	52.1288





Peak	RetTime Ty	pe Width	Area	Height	Area
#	[min]	[min]	[mAU*s]	[mAU]	%
1	5.821 BB 9.116 MM	0.1507	4708.04346 4858.10449	487.27863 279.16730	49.2157 50.7843







				C		
1	10.812	BV E	0.3071	362.35873	17.39060	2.5340
2	11.642	VB R	0.4737	1.39374e4	415.10941	97.4660

















VI References

1) Zheng, L.-S.; Yin, C.; Wang, F. Enantioselective synthesis of cis-hexahydro-γ-carboline derivatives via Ir-catalyzed asymmetric hydrogenation. *Chem. Commun.* 2022, **58**, 3286-3289.