Supplementary Information (SI) for RSC Advances. This journal is © The Royal Society of Chemistry 2024

Supporting Information

Fabrication of oxygen-releasing dextran microgels by droplet-based microfluidic method

Daisuke Tomioka, ^{†a} Shannon Anna Jung, ^{†b,c} Andrij Pich^{*b,c} and Michiya Matsusaki^{*a}

^aDepartment of Applied Chemistry, Graduate School of Engineering, Osaka University, 2-1 Yamadaoka, Suita, Osaka, 565-0871, Japan.

E-mail: m-matsus@chem.eng.osaka-u.ac.jp

^bDWI-Leibniz Institute for Interactive Materials, RWTH Aachen University, Forckenbeckstraße 50, 52074 Aachen, Germany

c Institute for Technical and Macromolecular Chemistry, RWTH Aachen University, Worringerweg 2, 52074 Aachen, Germany

Table of contents

- Fig. S1 Pictures of microfluidic synthesis
- Fig. S2 ¹H-NMR of dextran and Dex-MA
- Fig. S3 FT-IR spectra of dextran and Dex-MA
- Fig. S4 Elemental analysis on microgel to evaluate the leakage of CP nanoparticles
- Fig. S5 Cytocompatibility test of oxygen releasing microgels
- Fig. S6 SEM-EDX measurement of microgels
- Fig. S7 XRD measurement of microgels
- Fig. S8 Oxygen release behavior of ACC-CP microgel over time
- Fig. S9 Oxygen release behavior of microgels for the comparison
- Fig. S10 Stability of ACC-CP microgel
- Fig. S11 Oxygen release behavior of ACC-CP and CP nanoparticles

Fig. S1 Pictures of (a) microfluidic device and (b) microgel fabrication process.

Fig. S2¹H-NMR of dextran (top) and Dex-MA (bottom) in D₂O.

Fig. S3 FT-IR spectra of dextran and Dex-MA.

Fig. S4 EDX spectra of microgel containing 10 mg mL-1 CP nanoparticles (a) before and (b) after dispersing in ethanol for three times.

Fig. S5 Mitochondrial activities of NHDF after 1 day incubation with and without microgels containing 10 mg mL-1 CP nanoparticles in the presence of 1000 U mL-1 catalase under a hypoxic condition. Mitochondrial activity of each sample was standardized from the relative values as compared to 100 % for that of a cell culture under normoxic condition. All data are representative of four independent experiments, mean ± SD.

Fig. S6 (a) SEM image, (b) elemental mapping of calcium at the blue square area in (a) by EDX and (c) high magnification SEM image of the cross section at the crashed microgels containing 10 mg mL-1 CP nanoparticles. (d) SEM image, (e) elemental mapping of calcium at the blue square area in (d) by EDX and (f) high magnification SEM image of the cross section at the crashed ACC-CP microgels.

Fig. S7 XRD spectra of (a) microgel containing 10 mg mL-1 CP nanoparticles and (b) ACC-CP microgel.

Fig. S8 Dissolved oxygen concentration changes in 500 µL DMEM containing 100 U mL-1 catalase and 15 mg mL-1 Dex-MA microgel encapsulating ACC-CP under a hypoxic condition for 72 hours.

Fig. S9 Dissolved oxygen concentration changes in 500 µL DMEM containing 100 U mL-1 catalase and 15 mg mL-1 Dex-MA microgel encapsulating 5 and 10 mg mL-1 CP nanoparticles and ACC-CP under a hypoxic condition for the comparison.

Fig. S10 Bright field images of a freeze-dried ACC-CP microgel (a) soon after the dispersion in DMEM model solution (44 mM NaHCO₃ solution containing $0.1 \text{ mg} \text{ mL}^{-1} \text{ NaH}_2\text{PO}_4$) and (b) 1 day and (c) 7 days incubation in DMEM model solution at room temperature.

Fig. S11 (a) Dissolved oxygen concentration changes in 500 µL DMEM containing 100 U mL-1 catalase and 1 mg mL-1 CP nanoparticles and ACC-CP under a hypoxic condition. ACC-CP was fabricated by immersing 0.5 mg CP nanoparticles in $50 \mu L$ of $44 \text{ mM } \text{NaHCO}_3$ including 0.1 mg mL⁻¹ NaH₂PO₄ for 1 hour. SEM images of (b) prepared ACC-CP and (c) CP nanoparticles immersed in DMEM for 1 hour at a concentration of 1 mg mL-1 .