

## Supporting Information

### **Copper-Doped Cherry Blossom Carbon Dots with Peroxidase-Like Activity for Antibacterial Applications**

*Yitong Wang<sup>a</sup>, Tianliang Li<sup>b</sup>, Lixing Lin<sup>b</sup>, Dong Wang<sup>b</sup>, Lingyan Feng<sup>a,b,c\*</sup>*

[a] Y. Wang, Prof. L. Feng,  
QianWeichang College, Shanghai University,  
Shanghai 200444, China.

\*E-mail: [lingyanfeng@t.shu.edu.cn](mailto:lingyanfeng@t.shu.edu.cn)

[b] Dr. T. Li, L. Lin, D. Wang, Dr. Y. Chen, Prof. L. Feng,  
Materials Genome Institute, Shanghai Engineering Research Center for Integrated  
Circuits and Advanced Display Materials, and Shanghai Engineering Research Center  
of Organ Repair, Shanghai University,  
Shanghai 200444, China.

[c] Prof. L. Feng,  
Joint International Research Laboratory of Biomaterials and Biotechnology in Organ  
Repair, Ministry of Education,  
Shanghai 200444, China.

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## **1. Supplemental Methods**

### **1.1 Electron Spin Resonance**

Electron Spin Resonance (ESR) is a detection technique that employs molecular probes to capture target products. In this method, DMPO, a commonly used trapping agent, can react with hydroxyl radicals ( $\bullet\text{OH}$ ) to form a complex that generates characteristic ESR signals (DMPO-OH) when subjected to an applied magnetic field. The peak-to-peak height of the resulting ESR spectrum is indicative of the concentration of  $\bullet\text{OH}$  radicals. In the experiment outlined, the concentration of DMPO utilized was 100 mM.

### **1.2 Bacterial culture**

*E. coli* and *S. aureus* were individually inoculated into Luria Bertani (LB) broth, containing 25 g/L of the medium, and Tryptone Soy Broth (TSB), containing 30 g/L, respectively. These cultures were then placed into a shaking incubator set at 37.0°C with a shaking speed of 220 rpm for a duration of 12 hours. Subsequently, 100  $\mu\text{L}$  of each bacterial culture was transferred to a 96-well plate to measure the optical density (OD) at a wavelength of 600 nm. The bacterial concentration was determined from the  $\text{OD}_{600}$  readings and then adjusted to the desired concentration by dilution with the respective growth medium. The diluted cultures were subsequently stored at a temperature of 4°C.

### **1.3 Cell culture**

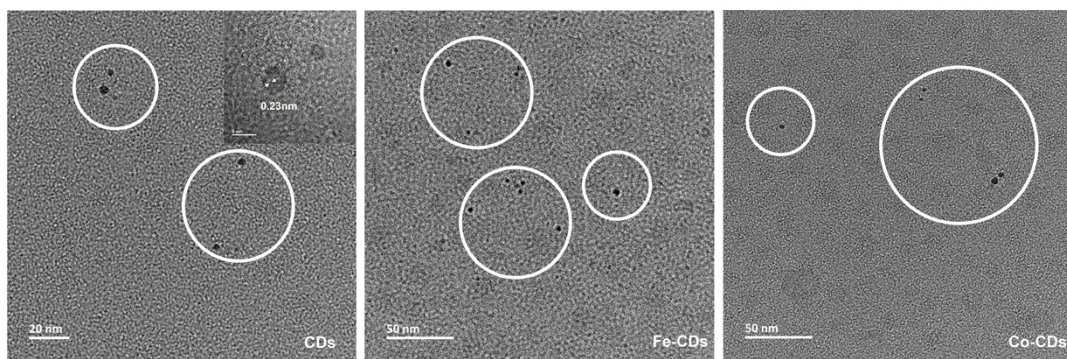
Mouse fibroblast cells (L929) were procured from Sangon Biotech Co., Ltd. These L929 cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% Fetal Bovine Serum (FBS) and 1% Penicillin/Streptomycin (PS). The incubation was conducted at a temperature of 37°C within an atmosphere containing 5%  $\text{CO}_2$ , providing an optimal environment for cell growth and maintenance.

### **1.4 Exploration of antibacterial concentrations**

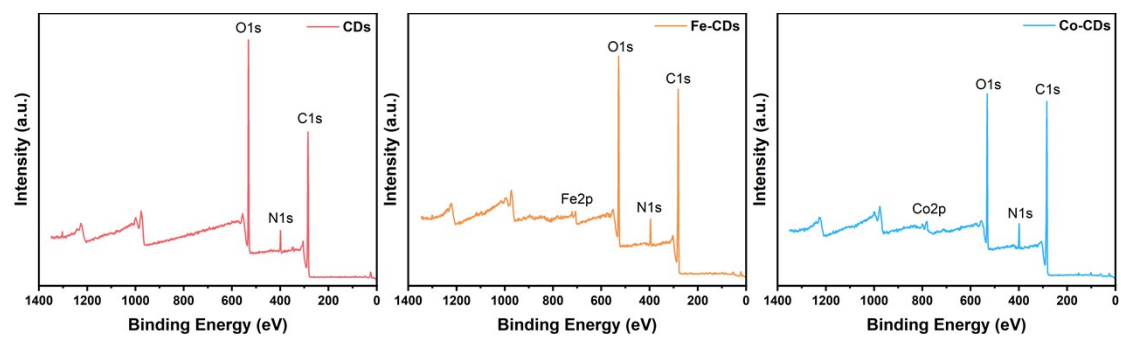
*E. coli* ( $10^8$  CFU  $\text{mL}^{-1}$ ) was mixed separately with different concentrations of  $\text{H}_2\text{O}_2$  ( $10^{-1}$ ,  $10^{-2}$ ,  $10^{-3}$ ,  $10^{-4}$ ,  $10^{-5}$ ,  $10^{-6}$  M) and Cu-CDs (15.625, 31.25, 62.5, 125, 250, 500  $\mu\text{g}$   $\text{mL}^{-1}$ ) for 0.5 h. Next, 50  $\mu\text{L}$  of the mixture was evenly spread onto the surface of an agar dish and incubated at 37°C for 12 hours.

### **1.5 Antibacterial Ring Test**

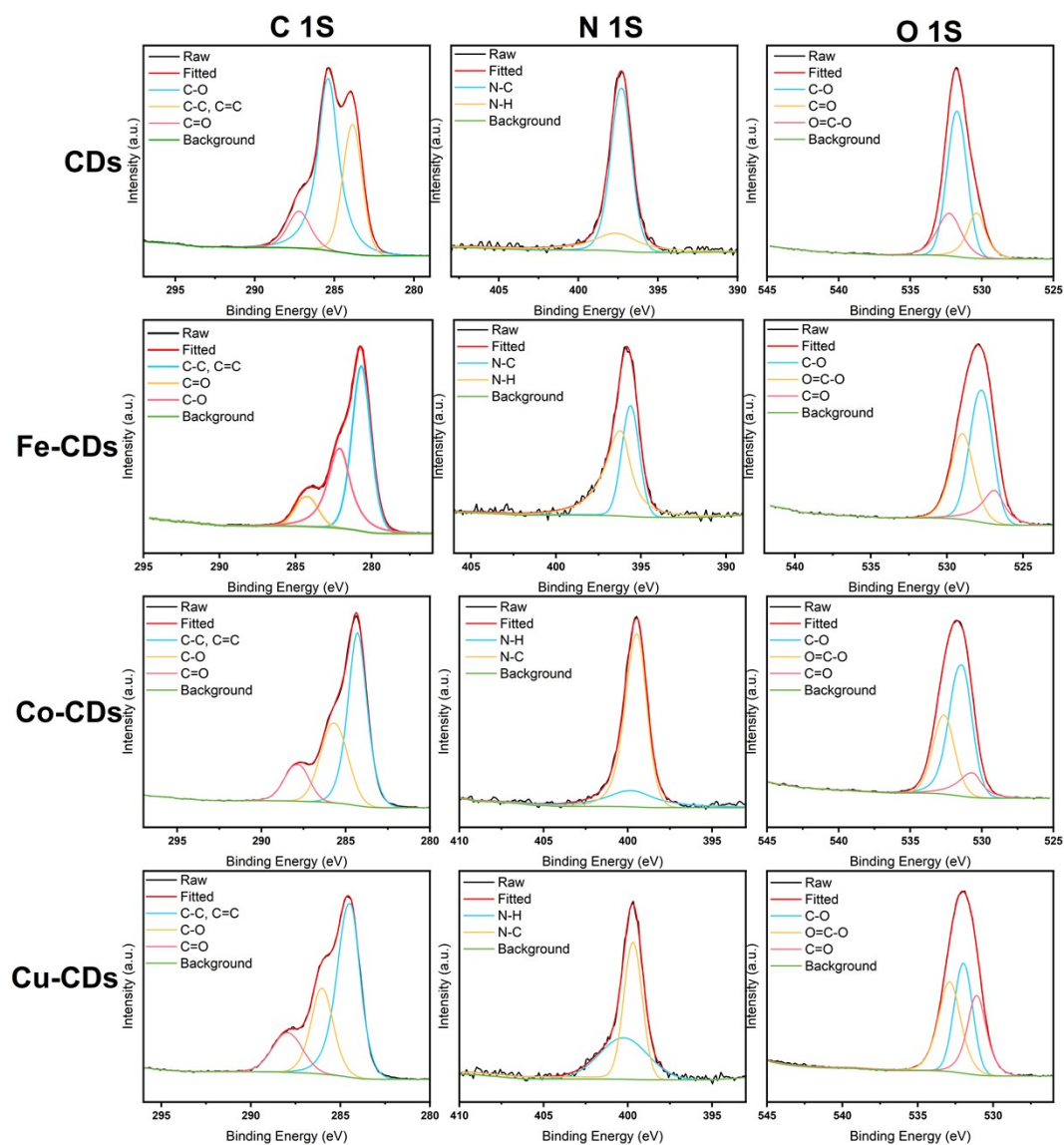
During the experimental procedure, the optical density of the pure bacterial culture is meticulously adjusted to an  $OD_{600}$  of 0.1, equating to a concentration of  $10^8$  CFU  $mL^{-1}$ . This suspension is uniformly spread on the agar plate to ensure an even distribution of bacteria. Following this, paper discs with a radius of 2.5 mm, which have been saturated with a solution of carbon dots and allowed to dry, are carefully placed on the agar. Post a 12-hour incubation period, the radius of the formed inhibition zone is meticulously measured, providing a quantitative assessment of the antibacterial effect.



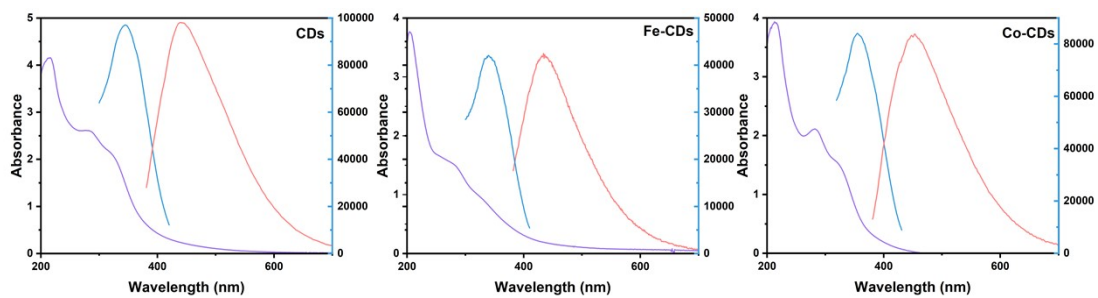
**Fig. S1** TEM, HRTEM images of CDs and TEM images of Fe-CDs, Co-CDs.



**Fig. S2** XPS full spectrum of CDs, Fe-CDs and Co-CDs.

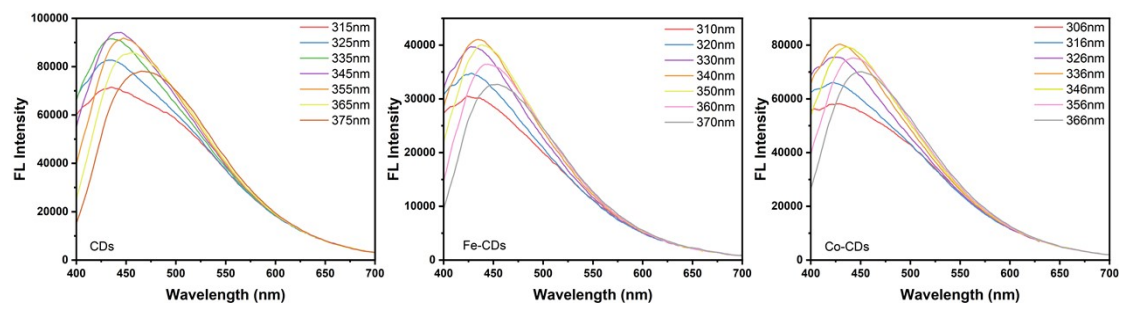


**Fig. S3** High-resolution XPS spectrum of C 1s, N 1s, and O 1s for all CDs.

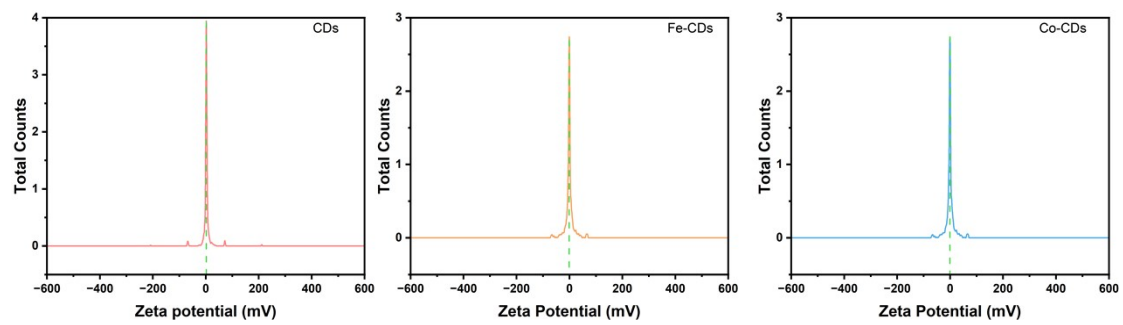


**Fig. S4** UV–Vis absorption spectra, the emission spectra and the excitation spectra XPS full spectrum of CDs, Fe-CDs and Co-CDs.

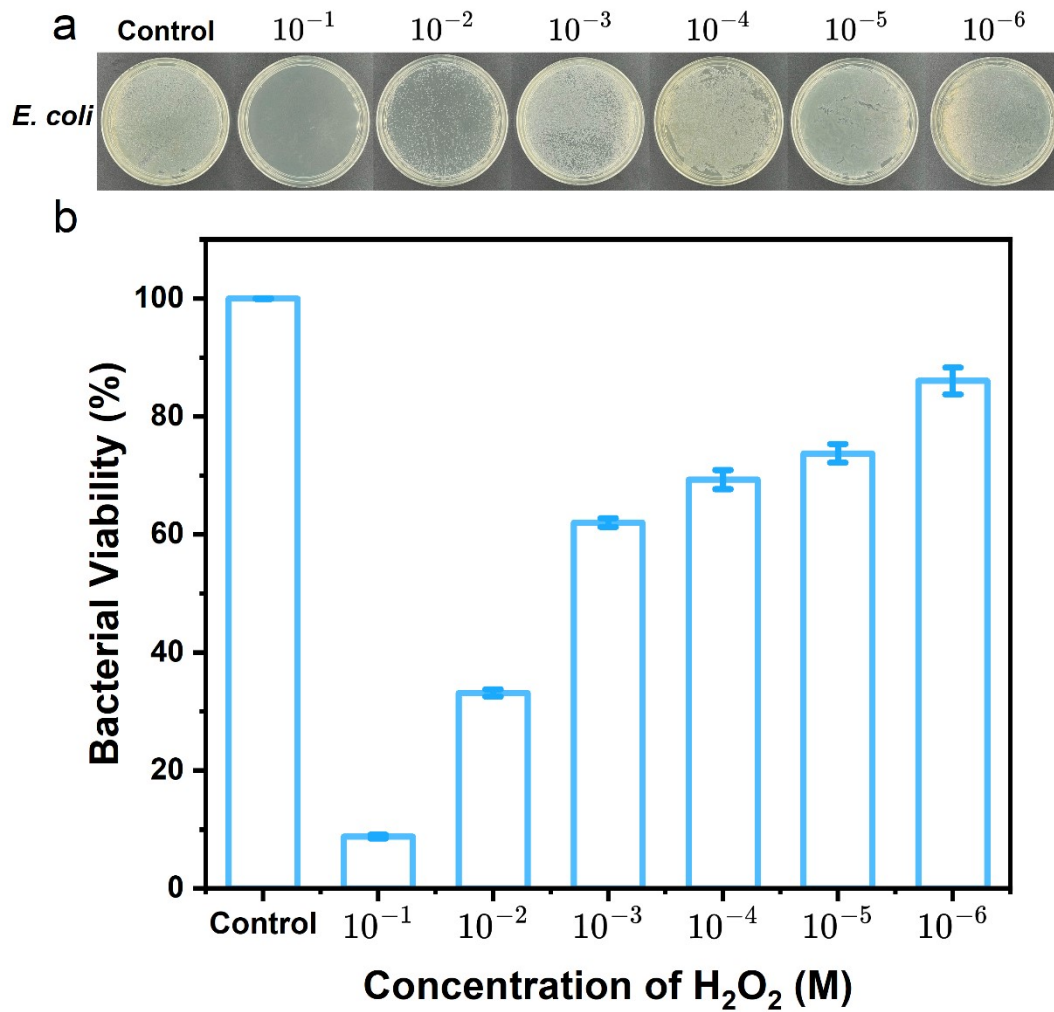




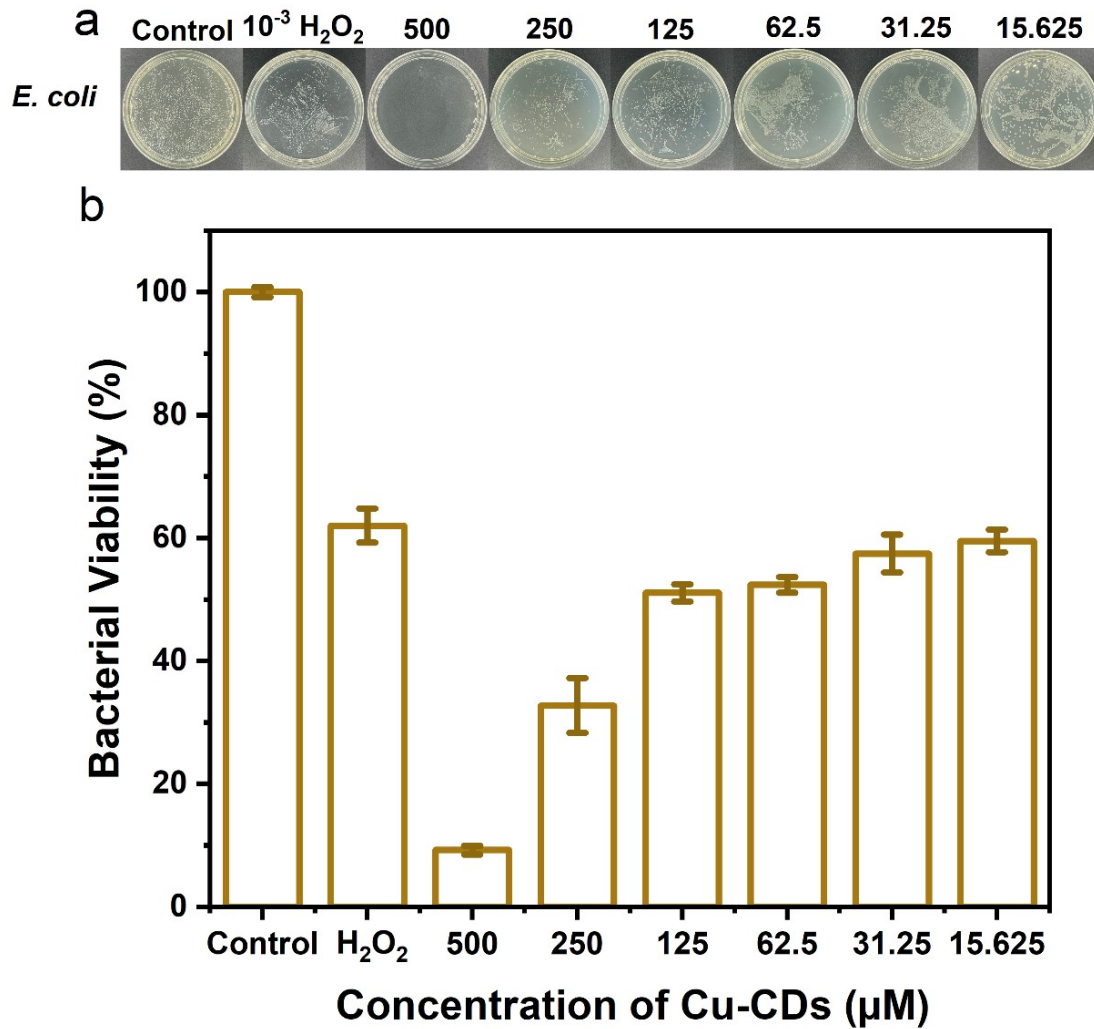
**Fig. S5** Fluorescence spectra under different excitation wavelength of CDs, Fe-CDs and Co-CDs.



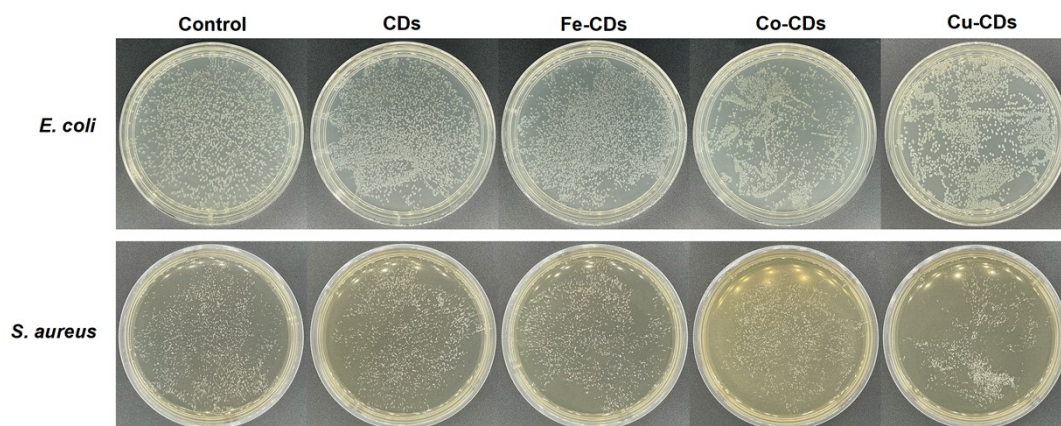
**Fig. S6** Zeta potential distribution of CDs, Fe-CDs and Co-CDs.



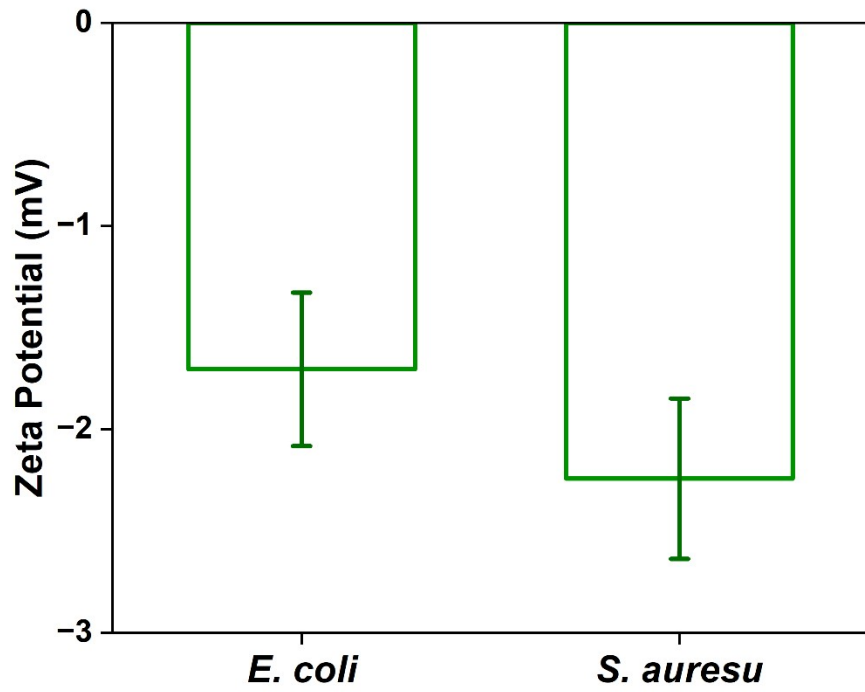
**Fig. S7** The MIC of H<sub>2</sub>O<sub>2</sub> to *E. coli*. (a) The images of agar plate of bacterial colonies of *E. coli* with concentrations of H<sub>2</sub>O<sub>2</sub>. (b) The bacterial viability of *E. coli* with different concentrations of H<sub>2</sub>O<sub>2</sub>. Data are presented as mean  $\pm$  SD (n = 3), ns:  $p > 0.05$ , \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$



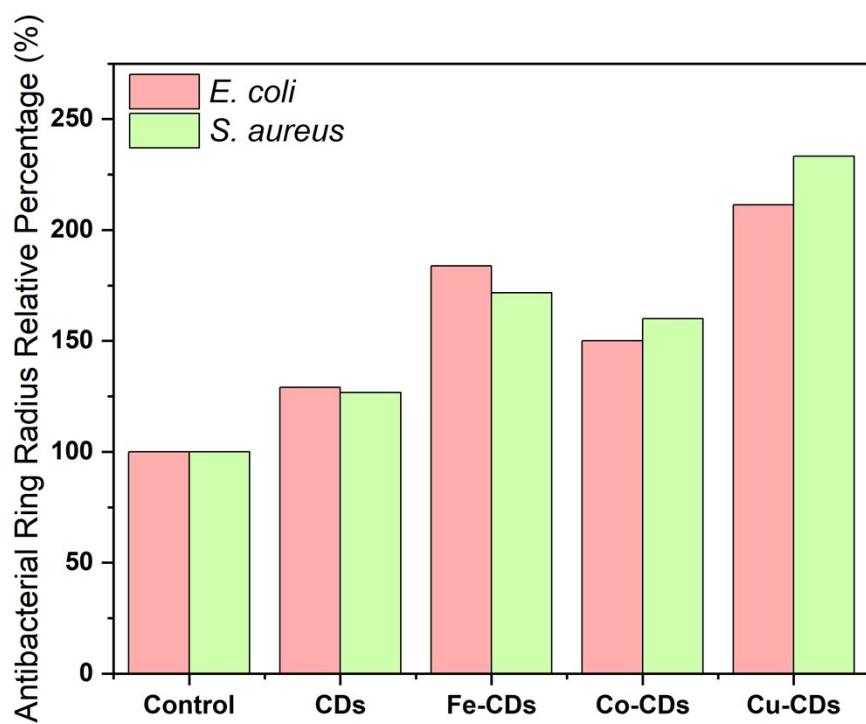
**Fig. S8** The MIC of Cu-CDs with  $H_2O_2$  to *E. coli*. (a) The images of agar plate of bacterial colonies of *E. coli* with concentrations of  $H_2O_2$ . (b) The bacterial viability of *E. coli* with different concentrations ( $\mu\text{g mL}^{-1}$ ) of Cu-CDs. Data are presented as mean  $\pm$  SD ( $n = 3$ ), ns:  $p > 0.05$ , \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$



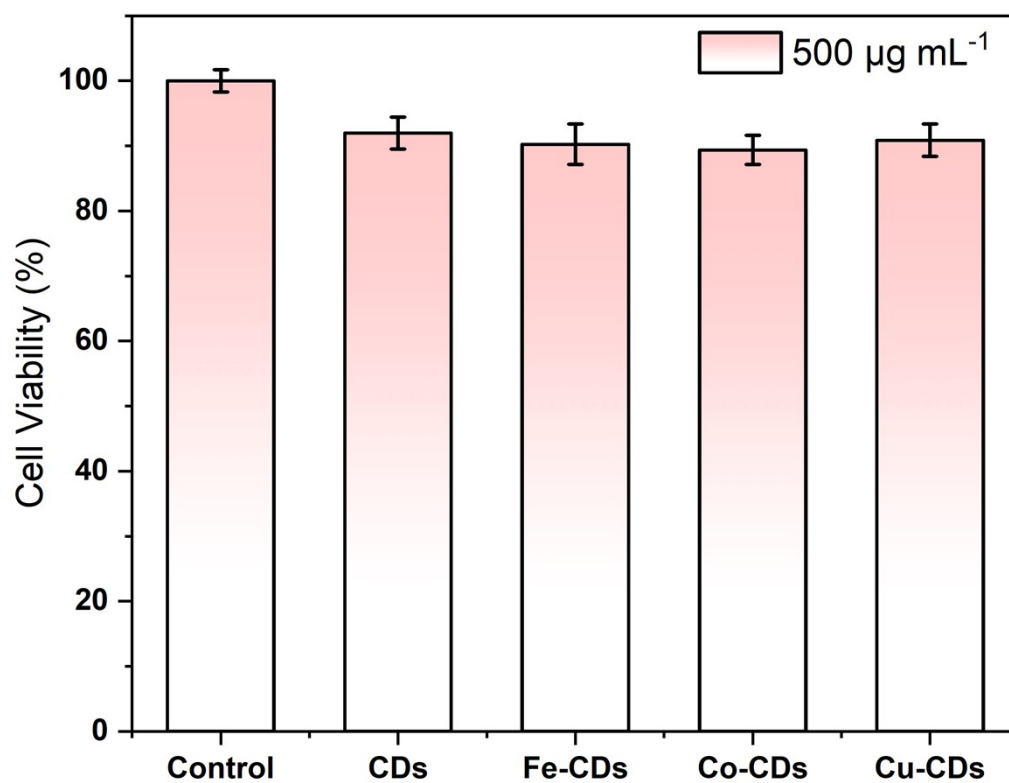
**Fig. S9** Images of *E. coli* and *S. aureus* treated with various CDs nanozymes without  $H_2O_2$ .



**Fig. S10** Zeta potential of *E. coli* and *S. aureus* in the corresponding liquid culture medium.



**Fig. S11** Antibacterial ring radius relative percentage of CDs, Fe-CDs, Co-CDs, and Cu-CDs against *E. coli* and *S. aureus*.



**Fig. S12** Cell viability of HSF cells in response to CDs at concentration of 500 µg mL<sup>-1</sup>.