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Supplementary Material

2 **Insight into the dynamic transformation properties of microplastic-**
3 **derived dissolved organic matter and its contribution to the**
4 **formation of chlorination disinfection by-products**

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12 **Characterization methods**

13 The UV absorbance of the PSMPs-DOM was measured in 1 cm quartz cuvettes
14 between 200 and 800 nm at a wavelength step size of 1 nm with ultrapure water as blank.
15 The absorption coefficient $\alpha(\lambda)$ (m^{-1}) was calculated using the equation (Helms et al.,
16 2008) :

$$17 \quad \alpha(\lambda) = 2.303A_{\lambda}/L$$

18 where A_{λ} is the absorbance at the position $\lambda(\text{nm})$, and L is the optical path diameter
19 (0.01m).

20 The spectral slope (S, nm^{-1}) was obtained by a non-linear exponential decay
21 model, and $S_{275-295}$ was the S value at wavelengths of 275-295 nm (Helms et al., 2008).
22 This value was used to characterize the relative molecular size of the DOM.

$$23 \quad \alpha_{\lambda} = \alpha_{\lambda_0} e^{-S(\lambda - \lambda_0)}$$

24 where α_{λ_0} is the absorption coefficient at the reference wavelength λ_0 , and S is the
25 spectral slope.

26 The PSMPs-DOM solution was freeze-dried using a freeze dryer (Alpha 1-4 LD
27 Plus) before the scanning of the Fourier transform infrared spectrometer. The scan
28 range of FTIR spectroscopy was 400-4000 cm^{-1} , and the scan resolution was 4 cm^{-1} .
29 Each FTIR spectrum was processed with PerkinElmer Spectrum software for baseline
30 correction.

31 Three-dimensional excitation emission matrix fluorescence spectra (3D-EEMs)
32 were taken with the excitation wavelengths ranging from 220 nm to 550 nm (scan
33 interval of 1nm), and the emission wavelengths ranging from 250 nm to 650 nm (scan
34 interval of 5nm) for PSMPs-DOM. In addition, the fluorescence spectrum of the

35 ultrapure water was measured and subtracted from the EEM spectrum of the DOM to
36 eliminate the Raman scattering effect (Christensen et al., 2005). The synchronous
37 fluorescence spectra were obtained in the wavelength range of 340-650 nm with a
38 constant offset ($\Delta\lambda = 120$ nm), and the scan interval was 1 nm and the scan speed was
39 600 nm/min⁻¹.

40 PARAFAC analysis was performed in MATLAB2021a (MathWorks, USA) using
41 the DOMFluor toolbox (Stedmon and Bro, 2008). Rayleigh and Raman scatters were
42 removed before PARAFAC analysis (Zepp et al., 2004). Split-half analysis, residual
43 analysis, and core consistency analysis were used to validate the model and the number
44 of fluorescent components (Stedmon and Bro, 2008). The values for each PARAFAC
45 fluorescent component was represented by the maximum fluorescence intensity (F_{\max})
46 (Stedmon and Markager, 2005).

47 The 2D-COS method was implemented to analyze the FTIR data, and the
48 concentration of chlorination was used as the perturbation variable. 2D-COS analysis
49 can extend the one-dimensional spectrum into two-dimensional spectra to catch the
50 signal peak variation (Noda et al., 2000). The order of change of the functional groups
51 was judged according to the positive/negative cross peaks of the synchronous and
52 asynchronous spectra. The 2D-COS was calculated using the 2D-shige software created
53 by Kwansei-Gakuin University, Japan (Noda, 2006).

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55 **The detailed extraction methods of THMs.**

56 After 24 h of chlorination reaction, DBPs were extracted from the samples through

57 liquid-liquid extraction method based on USEPA 551.1 with some modifications (Koh
58 et al., 2022). 3 mL methyl tert-butyl ether (MTBE) containing 4 g anhydrous sodium
59 sulfate was added to the samples, and the mixed solutions were shaken until the solids
60 were mostly dissolved. After 20 min of settling, 1 mL solution from the organic phase
61 was drawn out for analysis.

62 Reference

- 63 Christensen, J.H., Hansen, A.B., Mortensen, J., Andersen, O., 2005. Characterization
64 and matching of oil samples using fluorescence spectroscopy and parallel factor
65 analysis. *Anal. Chem.* 77, 2210–2217. <https://doi.org/10.1021/ac048213k>
- 66 Helms, J.R., Stubbins, A., Ritchie, J.D., Minor, E.C., Kieber, D.J., Mopper, K., 2008.
67 Absorption spectral slopes and slope ratios as indicators of molecular weight,
68 source, and photobleaching of chromophoric dissolved organic matter. *Limnol.*
69 *Oceanogr.* 53, 955–969. <https://doi.org/10.4319/lo.2008.53.3.0955>
- 70 Koh, K.Y., Chen, Z., Lin, S., Chandra Mohan, K., Luo, X., Chen, J.P., 2022. Leaching
71 of organic matters and formation of disinfection by-product as a result of
72 presence of microplastics in natural freshwaters. *Chemosphere* 299, 134300.
73 <https://doi.org/10.1016/j.chemosphere.2022.134300>
- 74 Noda, I., 2006. Progress in two-dimensional (2D) correlation spectroscopy. *J. Mol.*
75 *Struct.* 799, 2–15. <https://doi.org/10.1016/j.molstruc.2006.03.053>
- 76 Noda, I., Dowrey, A.E., Marcott, C., Story, G.M., Ozaki, Y., n.d. Generalized Two-
77 Dimensional Correlation Spectroscopy.
- 78 Stedmon, C.A., Bro, R., 2008. Characterizing dissolved organic matter fluorescence
79 with parallel factor analysis: a tutorial. *Limnol. Oceanogr. Methods* 6, 572–579.
80 <https://doi.org/10.4319/lom.2008.6.572>
- 81 Stedmon, C.A., Markager, S., 2005. Tracing the production and degradation of
82 autochthonous fractions of dissolved organic matter by fluorescence analysis.
83 *Limnol. Oceanogr.* 50, 1415–1426. <https://doi.org/10.4319/lo.2005.50.5.1415>
- 84 Zepp, R.G., Sheldon, W.M., Moran, M.A., 2004. Dissolved organic fluorophores in
85 southeastern US coastal waters: correction method for eliminating Rayleigh and
86 Raman scattering peaks in excitation–emission matrices. *Mar. Chem.* 89, 15–
87 36. <https://doi.org/10.1016/j.marchem.2004.02.006>

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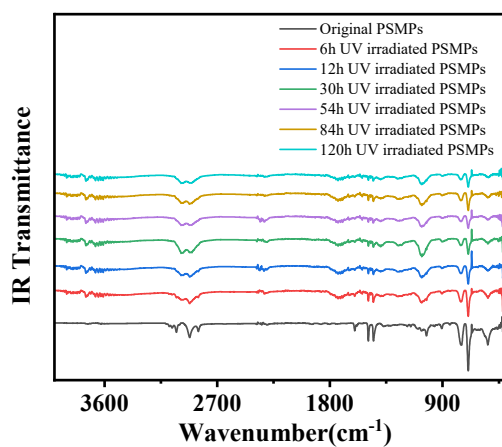
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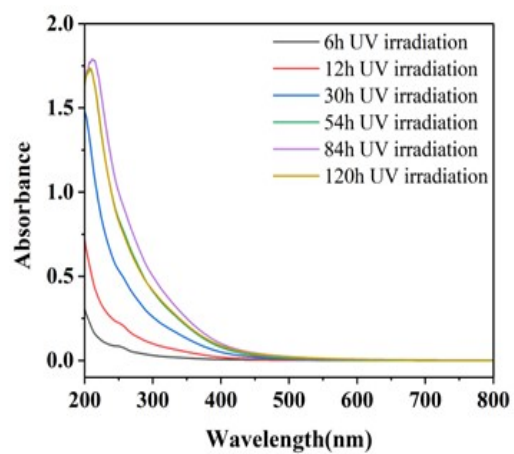
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Fig. S1 FTIR spectra of original PSMPs and PSMPs under different UV irradiation time

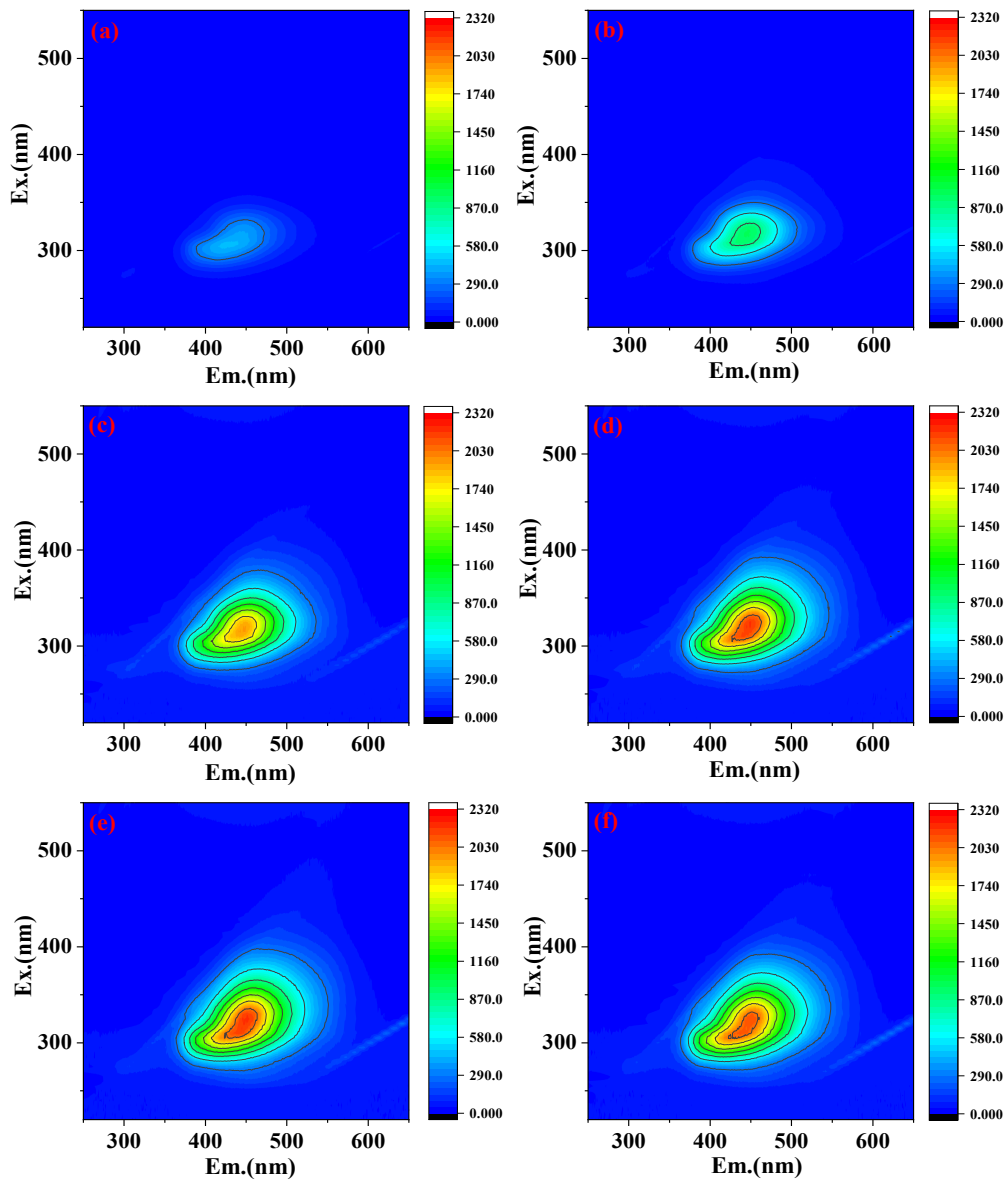


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Fig. S2 UV-vis spectra of PSMPs-DOM under and different UV irradiation time



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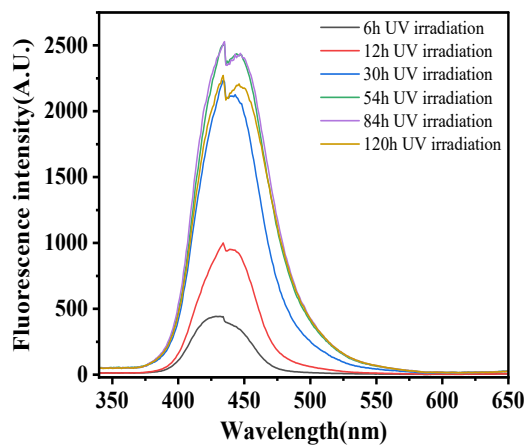
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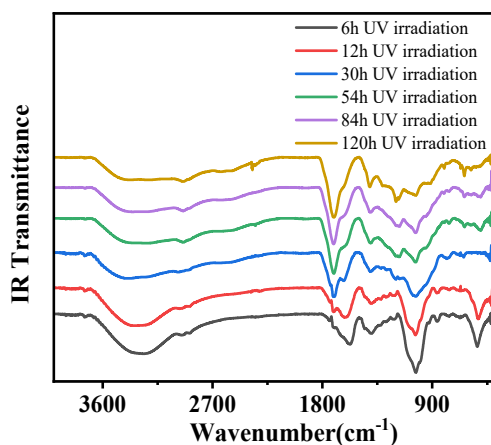
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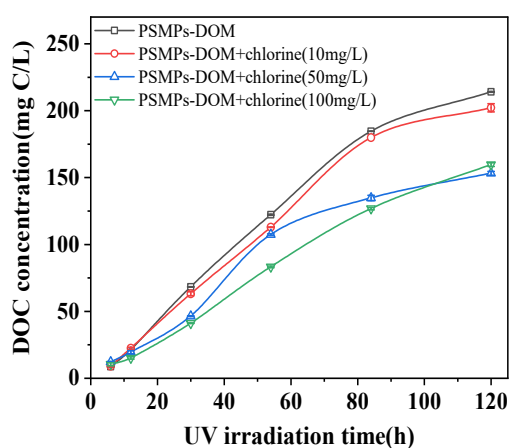
Fig. S3 EEM spectra of PSMPs-DOM under different UV irradiation time
((a)-(f), 6, 12, 30, 54, 84, 120 h)



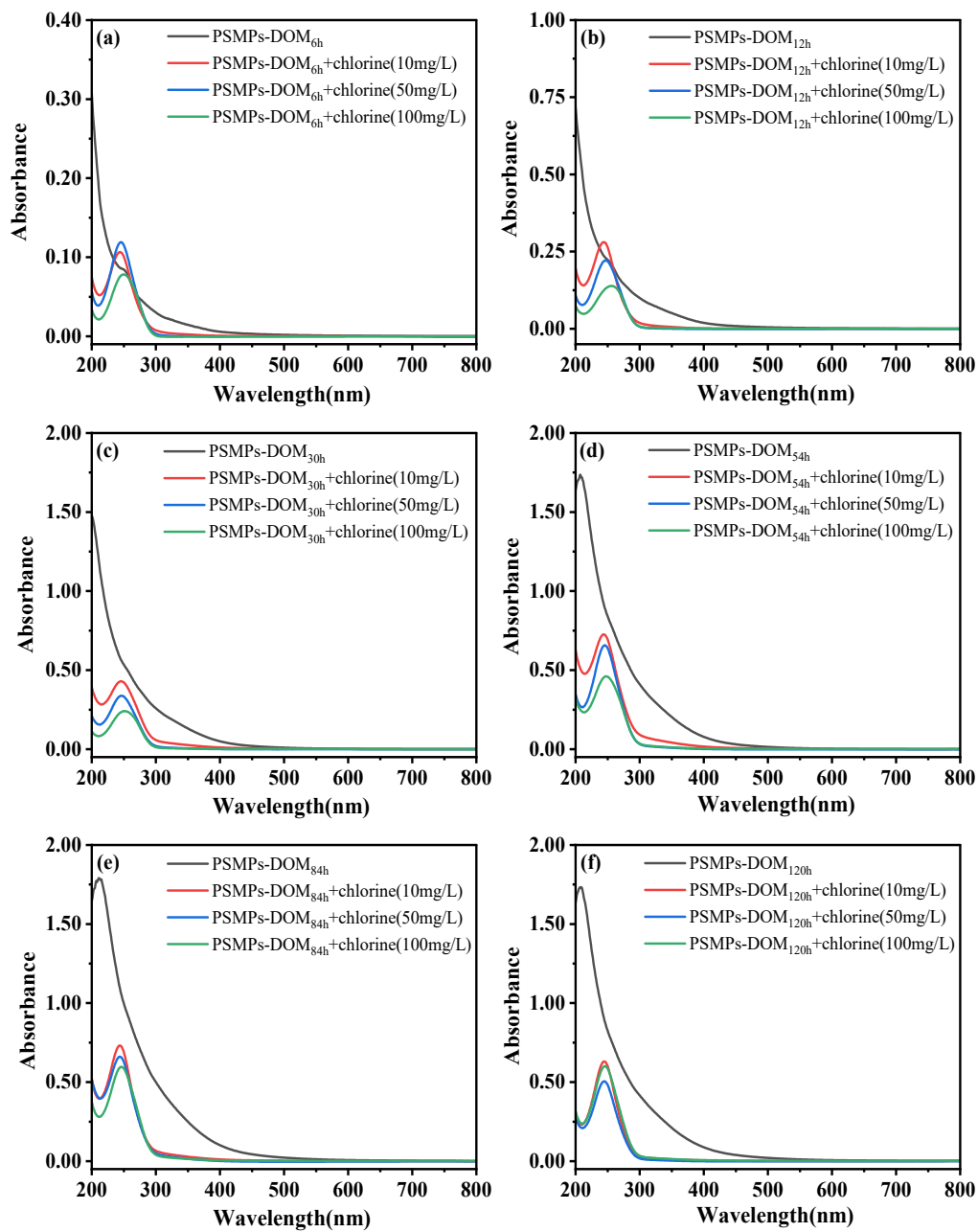
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 109 Fig. S4 Synchronous fluorescence spectra of PSMPs-DOM under different UV irradiation time
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 113 Fig. S5 FTIR spectra of PSMPs-DOM under different UV irradiation time
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 117 Fig. S6 DOC concentration of PSMPs-DOM before and after chlorination
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Fig. S7 UV-vis spectra of PSMPs-DOM before and after chlorination

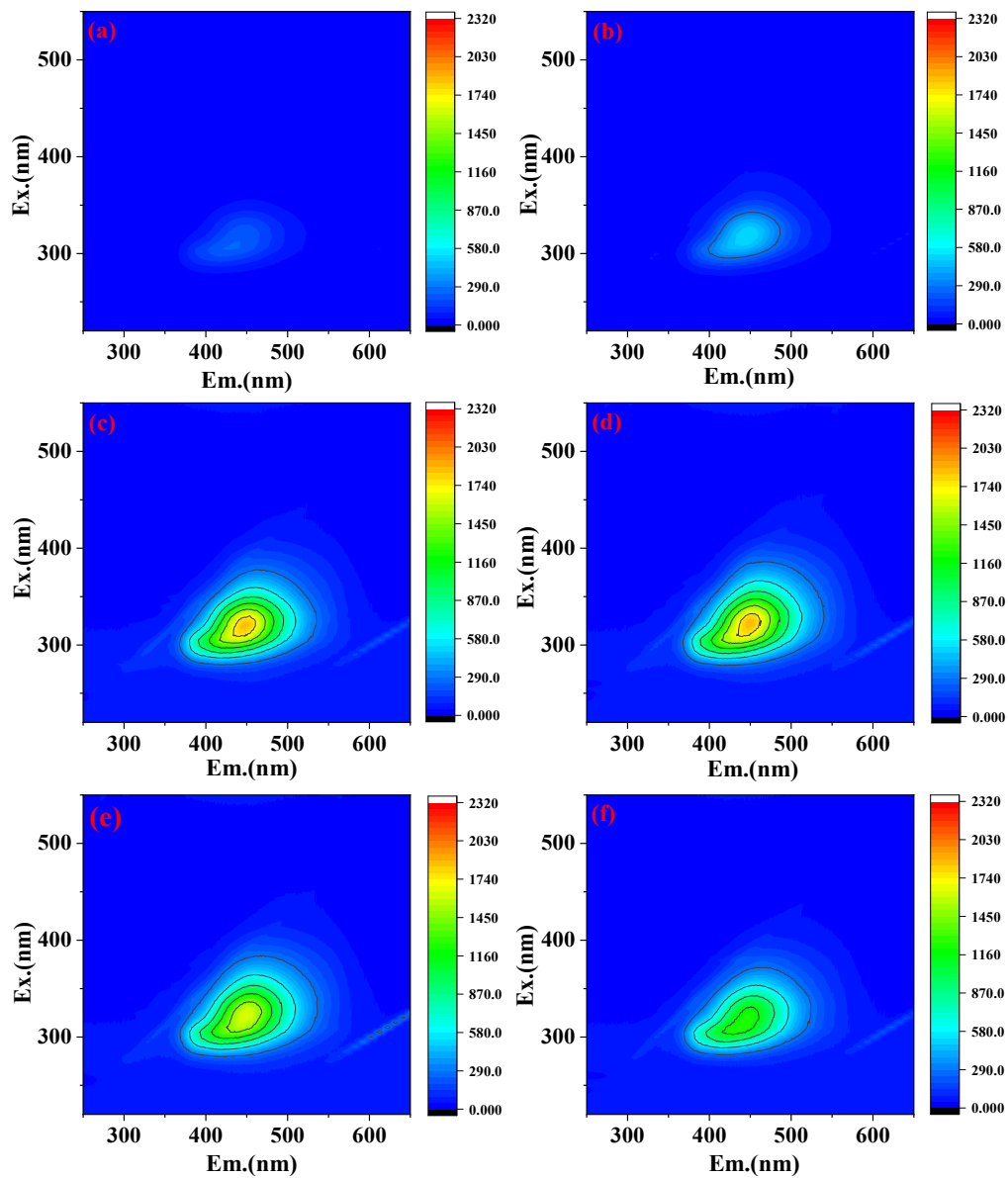


Fig. S8 EEM spectra of PSMPs-DOM_x under 10 mg/L chlorination
 ((a)-(f), x = 6, 12, 30, 54, 84, 120 h)

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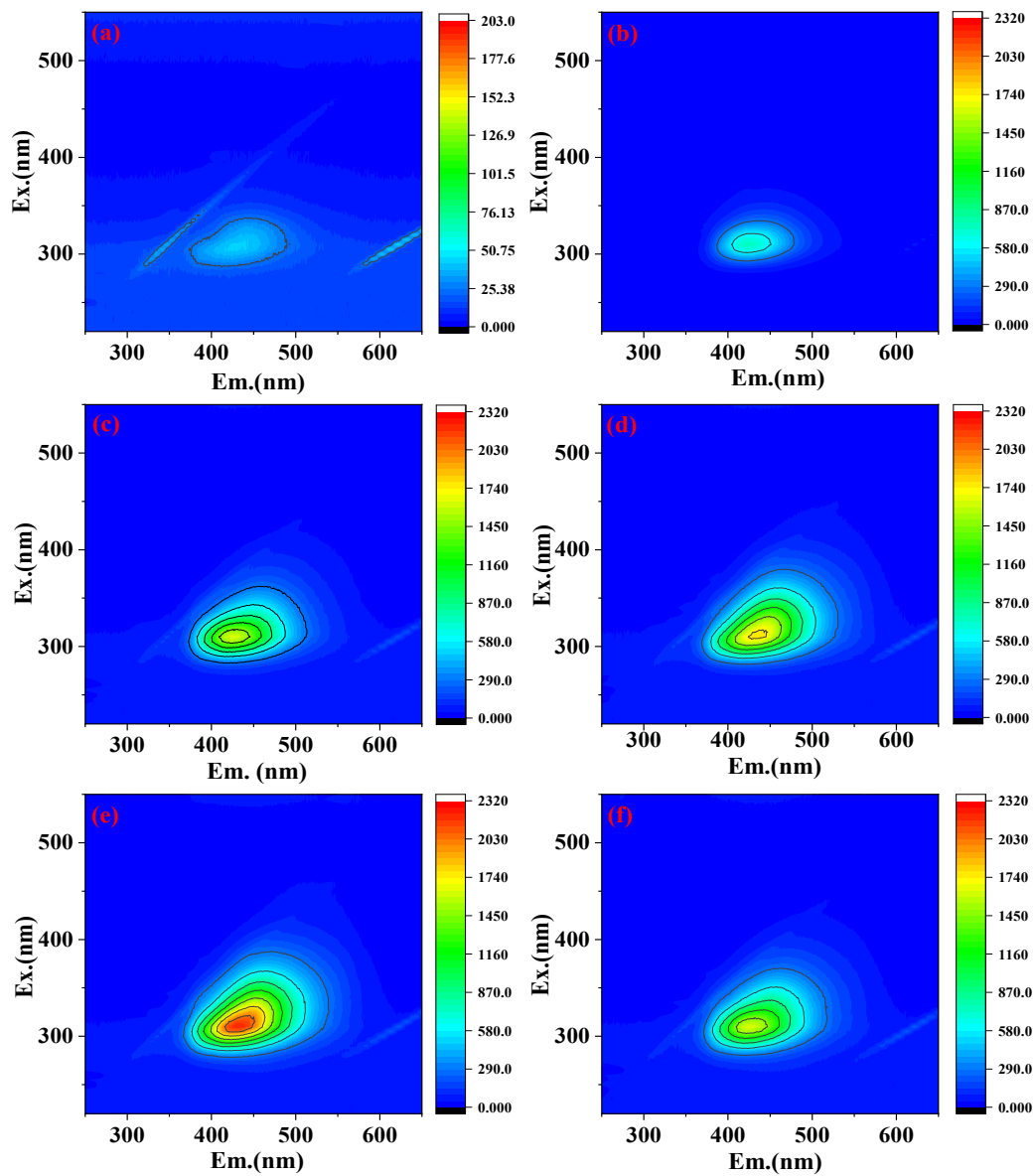
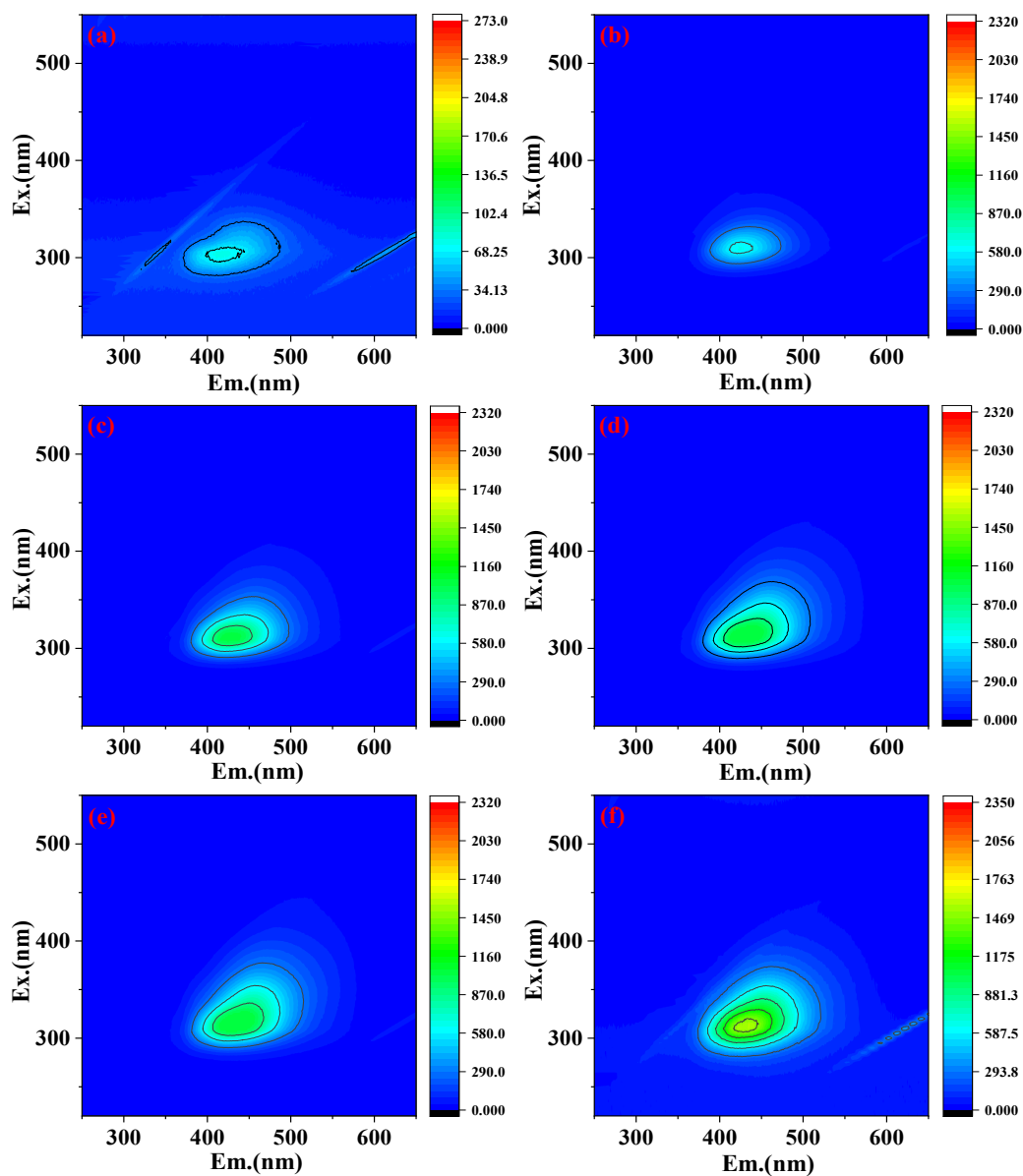


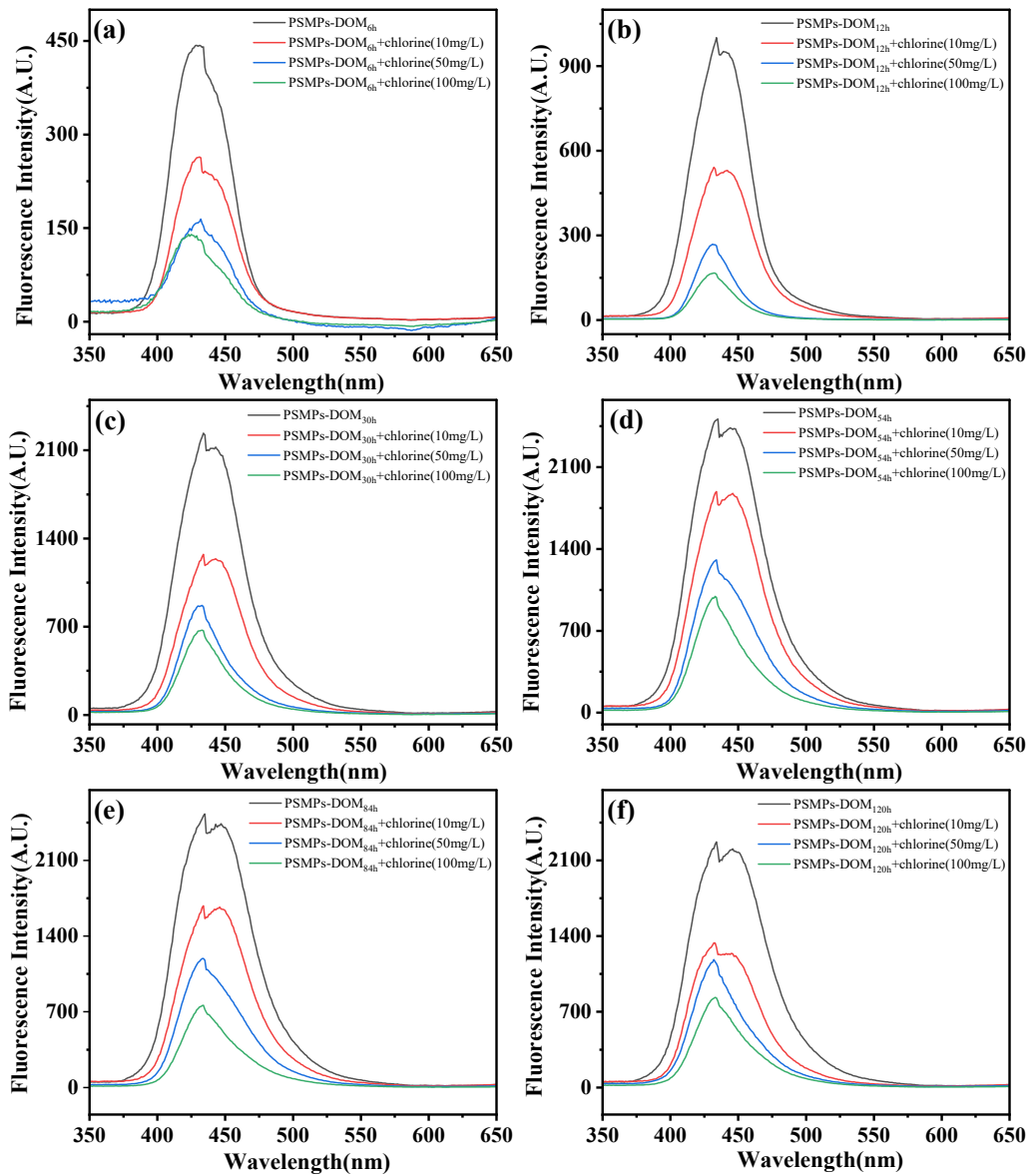
Fig. S9 EEM spectra of PSMPs-DOM_x under 50 mg/L chlorination ((a)-(f), x = 6, 12, 30, 54, 84, 120 h)

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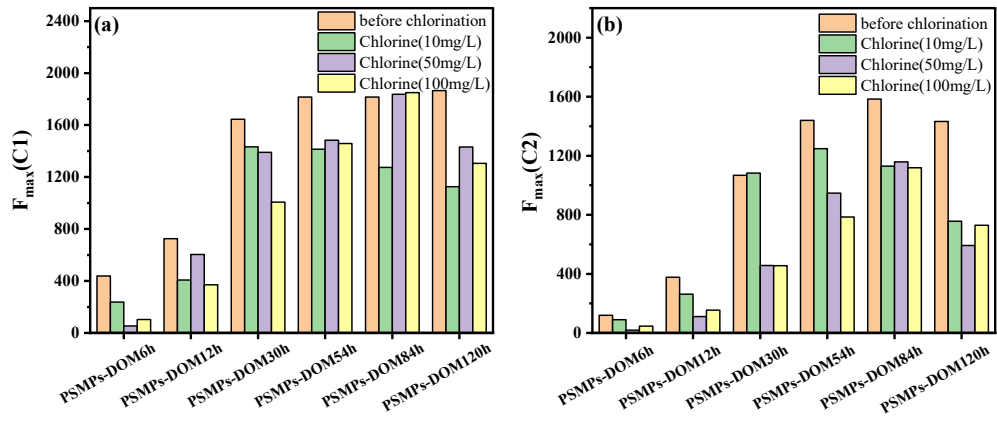
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Fig. S10 EEM spectra of PSMPs-DOM_x under 100 mg/L chlorination ((a)-(f), x = 6, 12, 30, 54, 84, 120 h)



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Fig. S11 Synchronous fluorescence spectra of PSMPs-DOM before and after chlorination



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Fig. S12 $F_{\max}(C1)$ (a) and $F_{\max}(C2)$ (b) of PSMPs-DOM before and after chlorination

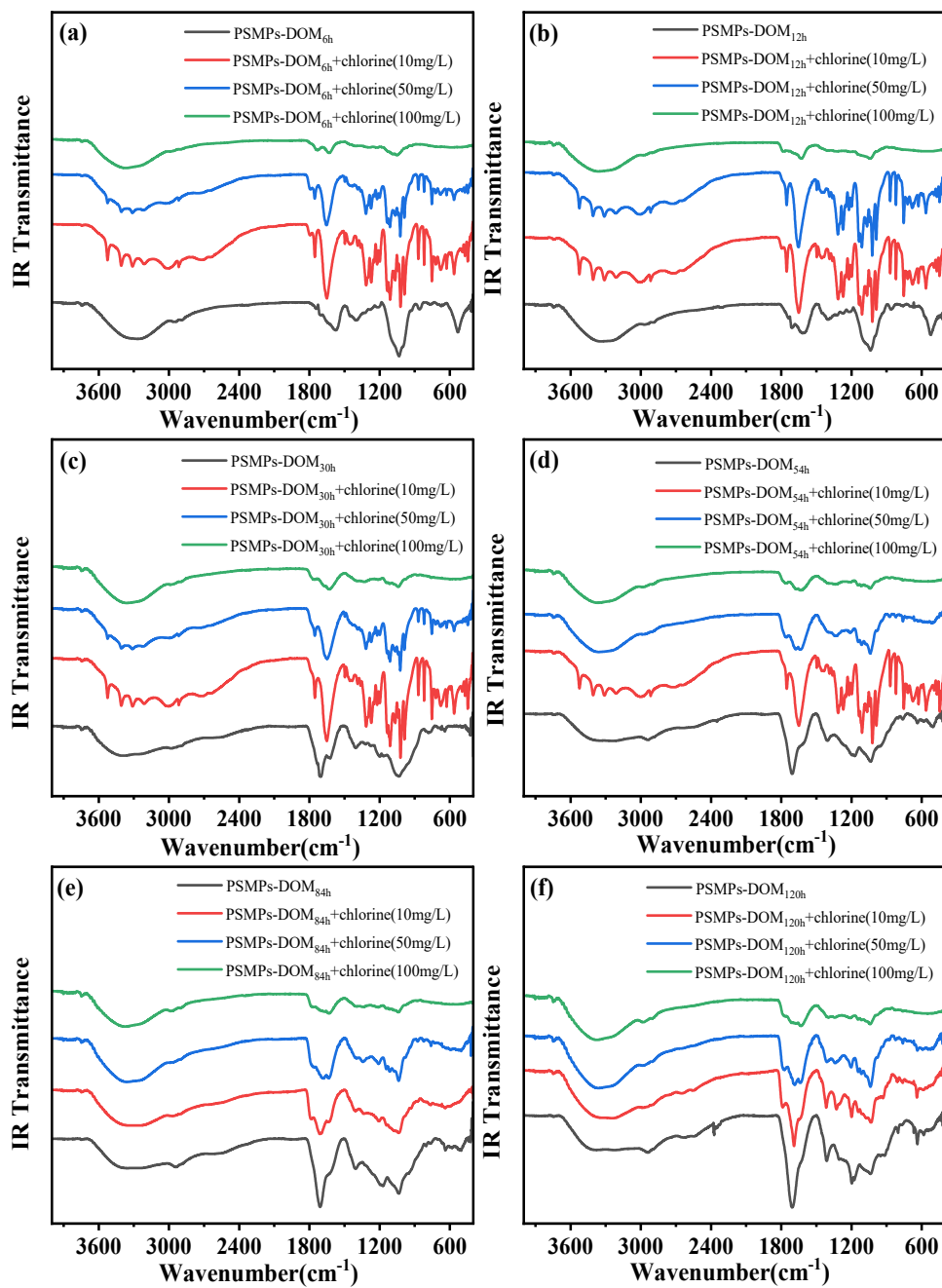


Fig. S13 FTIR spectra of PSMPs-DOM before and after chlorination

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Table S1. Analytical methods of THMs by GC-ECD

GC/ECD	Agilent Technologies 7820A, USA
Column types	HP-5, 30 m × 0.32 mm × 0.25 μm
Split ratio	Splitless
Injection volume	1 μL
Injection Temperature	200°C
Oven temperature and ramp	The initial temperature of 30°C for 10.5 min, then an increase of 14°C/min to 72°C, which was held for 1 min, and finally an increase of 40°C to 200°C, which was held for 2 min.
Detector temperature	300°C
Carrier gas	N ₂ , constant flow at 3 mL/min

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Table S2. The summary of Noda's rules for interpreting synchronous ($\Phi(v_1, v_2)$) and asynchronous ($\psi(v_1, v_2)$) cross peaks in 2D-COS analysis (Noda et al., 2000).

sign at $\Phi(v_1, v_2)$	sign at $\psi(v_1, v_2)$	interpretation
+		the intensity of v_1 and v_2 are changing in the same direction, i.e. increasing or decreasing together
-		the intensity of v_1 and v_2 are changing in the opposite direction
+	+	the change at v_1 is occurring predominantly before that at v_2
+	-	the change at v_1 is occurring predominantly after that at v_2
-	+	the change at v_1 is occurring predominantly after that at v_2
-	-	the change at v_1 is occurring predominantly before that at v_2

Table S3. Main absorption bands of PSMPs in the IR regions and their identifying assignment groups.

Wavenumber/ (cm ⁻¹)	Functional groups	References
3746	-OH	(Chen et al., 2020)
2973	-CH	(Zhang et al., 2020)
2902	-CH ₂	(Zhang et al., 2020)
1730	C=O	(Wang et al., 2023)
1066	C-O	(Leng, 2018)

Table S4. DOC concentration of PSMPs-DOM under different UV irradiation time

Samples	UV irradiation time/ (h)	DOC/ (mg C/L)
PSMPs-DOM	6	8.71±0.17
	12	21.31±0.17
	30	68.33±0.21
	54	122.29±0.48
	84	184.62±0.05
	120	214.05±0.32

Table S5. UV parameters of PSMPs-DOM under different UV irradiation time

Samples	UV irradiation time/ (h)	α_{254} / (m ⁻¹)	S ₂₇₅₋₂₉₅ / (nm ⁻¹)
PSMPs-DOM	6	18.60±0.13	0.018±0.002
	12	49.43±0.18	0.016±0.005
	30	117.59±0.25	0.015±0.007
	54	184.12±0.10	0.015±0.004
	84	218.33±0.44	0.015±0.001
	120	179.84±0.19	0.014±0.002

Table S6. F_{max} value of PSMPs-DOM under different UV irradiation time

Samples	UV irradiation time/ (h)	F _{max} (C1)	F _{max} (C2)
PSMPs-DOM	6	438	119
	12	726	377
	30	1644	1068
	54	1815	1440
	84	1815	1585
	120	1866	1433

Table S7. $\Delta\alpha_{254}$ and $\Delta S_{275-295}$ of PSMPs-DOM after chlorination

Chlorine dosage/ (mg/L)	Samples	$\Delta\alpha_{254}/ (\text{m}^{-1})$	$\Delta S_{275-295}/ (\text{nm}^{-1})$
10	PSMPs-DOM _{6h}	2.38±0.27	0.044±0.004
	PSMPs-DOM _{12h}	5.08±0.32	0.047±0.003
	PSMPs-DOM _{30h}	24.83±2.46	0.041±0.003
	PSMPs-DOM _{54h}	36.35±2.80	0.036±0.003
	PSMPs-DOM _{84h}	74.54±3.41	0.043±0.002
	PSMPs-DOM _{120h}	54.33±1.07	0.062±0.004
50	PSMPs-DOM _{6h}	5.86±0.26	0.071±0.004
	PSMPs-DOM _{12h}	1.81±0.33	0.074±0.006
	PSMPs-DOM _{30h}	44.96±0.30	0.061±0.005
	PSMPs-DOM _{54h}	47.21±0.83	0.064±0.003
	PSMPs-DOM _{84h}	86.03±0.91	0.050±0.002
	PSMPs-DOM _{120h}	78.71±1.05	0.069±0.002
100	PSMPs-DOM _{6h}	0.92±0.10	0.083±0.001
	PSMPs-DOM _{12h}	17.60±0.17	0.071±0.004
	PSMPs-DOM _{30h}	62.46±2.71	0.070±0.003
	PSMPs-DOM _{54h}	81.89±1.87	0.059±0.007
	PSMPs-DOM _{84h}	90.65±4.09	0.059±0.006
	PSMPs-DOM _{120h}	54.22±4.16	0.063±0.001

Table S8. Relative percentage of C1 and C2 in PSMPs-DOM before and after chlorination

Samples	Chlorine dosage/ (mg/L)	C1/ (%)	C2/ (%)
PSMPs-DOM _{6h}	0	78.61	21.39
	10	72.58	27.42
	50	73.83	26.17
	100	69.14	30.86
PSMPs-DOM _{12h}	0	65.80	34.20
	10	60.84	39.16
	50	84.51	15.49
	100	70.56	29.44
PSMPs-DOM _{30h}	0	60.62	39.38
	10	56.95	43.05
	50	75.27	24.73
	100	68.84	31.16
PSMPs-DOM _{54h}	0	55.76	44.24
	10	53.11	46.88
	50	61.02	38.98
	100	65.00	35.00
PSMPs-DOM _{84h}	0	53.39	46.61
	10	52.98	47.02
	50	61.33	38.67
	100	62.31	37.69
PSMPs-DOM _{120h}	0	56.57	43.43
	10	59.78	40.22
	50	70.74	29.26
	100	64.16	35.84

Table S9. Main absorption bands of PSMPs-DOM in the IR regions and their identifying assignment groups.

Samples	Wavenumber/ (cm ⁻¹)	Functional groups	References
PSMPs-DOM _x (x=6,12,30,54h)	1654	Aromatic C=C/C=O	(Lee et al., 2021)
	1314	C-OH	(Leng, 2018)
	1270	C-O	(Zhang et al., 2022)
	1139	C-O-C	(Gu et al., 2024)
	1022	Aromatic C-H	(Lee et al., 2021)
PSMPs-DOM _x (x=84,120h)	1721	O-C=O	(Liu et al., 2019)
	1139	C-O-C	(Gu et al., 2024)
	1022	Aromatic C-H	(Lee et al., 2021)

Table S10. 2D-FTIR-COS results of cross-peaks in synchronous and asynchronous maps

Group	Position/ (cm ⁻¹)	Sign						
PSMPs-DOM _{6h}		1754	1654	1300	1270	1111	1022	988
	1754	+	+(+)	+(+)	+(+)	+(+)	+(+)	+(+)
	1654		+	+(-)	+(+)	+(+)	+(+)	+(+)
	1300			+	+(+)	+(+)	+(+)	+(+)
	1270				+	+(+)	+(+)	+(+)
	1111					+	+(+)	+(+)
	1022						+	+(+)
	988							+
PSMPs-DOM _{12h}		1754	1654	1300	1270	1111	1022	988
	1754	+	+(-)	+(-)	+(-)	+(+)	+(+)	+(+)
	1654		+	+(-)	+(-)	+(+)	+(+)	+(+)
	1300			+	+(+)	+(+)	+(+)	+(+)
	1270				+	+(+)	+(+)	+(+)
	1111					+	+(+)	+(+)
	1022						+	+(-)
	988							+
PSMPs-DOM _{30h}		1754	1654	1300	1270	1111	1022	988
	1754	+	+(-)	+(-)	+(-)	+(-)	+(+)	+(+)
	1654		+	+(-)	+(+)	+(+)	+(+)	+(+)
	1300			+	+(+)	+(+)	+(+)	+(+)
	1270				+	+(-)	+(+)	+(+)
	1111					+	+(+)	+(+)
	1022						+	+(+)
	988							+
PSMPs-DOM _{54h}		1754	1654	1300	1270	1111	1022	988
	1754	+	+(+)	+(+)	+(+)	+(+)	+(+)	+(+)
	1654		+	+(+)	+(+)	+(+)	+(+)	+(+)
	1300			+	+(+)	+(-)	+(-)	+(+)
	1270				+	+(-)	+(-)	+(-)
	1111					+	+(+)	+(+)
	1022						+	+(+)
	988							+
PSMPs-DOM _{84h}		1709	1173	1032				
	1709	+	+(+)	+(-)				
	1173		+	+(-)				
	1032			+				
PSMPs-DOM _{120h}		1709	1173	1032				
	1709	+	+(+)	+(+)				
	1173		+	+(+)				
	1032			+				

Table S11. Formation potential of each THMs and total THMs in chlorinated PSMPs-DOM

Chlorine dosage/ (mg/L)	Samples	TCM/ ($\mu\text{g}/\text{mg C}$)	DCBM/ ($\mu\text{g}/\text{mg C}$)	DBCM/ ($\mu\text{g}/\text{mg C}$)	TBM/ ($\mu\text{g}/\text{mg C}$)	total THMs/ ($\mu\text{g}/\text{mg C}$)
10	PSMPs-DOM _{6h}	4.72±0.08	2.35±0.017	2.31±0.005	1.30±0.010	10.68±0.10
	PSMPs-DOM _{12h}	1.00±0.007	0.86±0.003	1.03±0.001	0.55±0.001	3.43±0.005
	PSMPs-DOM _{30h}	0.23±0.002	0.28±0.002	0.45±0.009	0.19±0.004	1.16±0.006
	PSMPs-DOM _{54h}	0.12±0.001	0.16±0.003	0.27±0.005	0.11±0.007	0.66±0.004
	PSMPs-DOM _{84h}	0.01±0.003	0.10±0.003	0.16±0.002	0.14±0.002	0.42±0.002
	PSMPs-DOM _{120h}	0.01±0.004	0.09±0.004	0.14±0.001	0.13±0.002	0.37±0.001
50	PSMPs-DOM _{6h}	15.99±0.01	2.10±0.003	1.55±0.003	0.94±0.003	20.58±0.02
	PSMPs-DOM _{12h}	7.95±0.08	0.98±0.007	0.95±0.001	0.60±0.007	10.48±0.09
	PSMPs-DOM _{30h}	0.93±0.02	0.39±0.001	0.43±0.001	0.28±0.002	2.02±0.02
	PSMPs-DOM _{54h}	0.28±0.004	0.17±0.002	0.27±0.004	0.13±0.008	0.85±0.01
	PSMPs-DOM _{84h}	0.17±0.005	0.13±0.001	0.20±0.002	0.13±0.005	0.63±0.006
	PSMPs-DOM _{120h}	0.12±0.003	0.12±0.001	0.15±0.003	0.14±0.007	0.53±0.008
100	PSMPs-DOM _{6h}	21.10±0.39	2.52±0.02	1.59±0.002	1.09±0.032	26.30±0.39
	PSMPs-DOM _{12h}	13.52±0.17	1.53±0.02	1.20±0.001	0.81±0.012	17.06±0.14
	PSMPs-DOM _{30h}	2.79±0.07	0.57±0.004	0.50±0.001	0.32±0.011	4.17±0.08
	PSMPs-DOM _{54h}	0.75±0.02	0.24±0.003	0.33±0.003	0.16±0.001	1.48±0.02
	PSMPs-DOM _{84h}	0.49±0.01	0.15±0.002	0.21±0.005	0.10±0.002	0.96±0.02
	PSMPs-DOM _{120h}	0.36±0.005	0.12±0.004	0.14±0.002	0.08±0.002	0.70±0.004

References:

- Chen, W., Ding, S., Lin, Z., Peng, Y., Ni, J., 2020. Different effects of N₂-flow and air-limited pyrolysis on bamboo-derived biochars' nitrogen and phosphorus release and sorption characteristics. *Science of The Total Environment* 711, 134828. <https://doi.org/10.1016/j.scitotenv.2019.134828>
- Gu, X., Chen, B., Liu, H., Feng, Y., Wang, B., He, S., Feng, M., Pan, G., Han, S., 2024. Photochemical behavior of dissolved organic matter derived from *Alternanthera philoxeroides* hydrochar: Insights from molecular transformation and photochemically reactive intermediates. *Journal of Hazardous Materials* 461, 132591. <https://doi.org/10.1016/j.jhazmat.2023.132591>
- Lee, Y.K., Hong, S., Hur, J., 2021. Copper-binding properties of microplastic-derived dissolved organic matter revealed by fluorescence spectroscopy and two-dimensional correlation spectroscopy. *Water Research* 190, 116775. <https://doi.org/10.1016/j.watres.2020.116775>
- Leng, E., 2018. In situ structural changes of crystalline and amorphous cellulose during slow pyrolysis at low temperatures.
- Liu, S., Feng, W., Song, F., Li, T., Guo, W., Wang, B., Wang, H., Wu, F., 2019. Photodegradation of algae and macrophyte-derived dissolved organic matter: A multi-method assessment of DOM transformation. *Limnologia* 77, 125683. <https://doi.org/10.1016/j.limno.2019.125683>
- Noda, I., Dowrey, A.E., Marcott, C., Story, G.M., Ozaki, Y., n.d. Generalized Two-Dimensional Correlation Spectroscopy.
- Wang, H., Zhu, J., He, Y., Wang, J., Zeng, N., Zhan, X., 2023. Photoaging process and mechanism of four commonly commercial microplastics. *Journal of Hazardous Materials* 451, 131151. <https://doi.org/10.1016/j.jhazmat.2023.131151>
- Zhang, H., Chen, W., Li, Q., Zhang, X., Wang, C., Yang, L., Wei, R., Ni, J., 2020. Difference in characteristics and nutrient retention between biochars produced in nitrogen-flow and air-limitation atmospheres. *J of Env Quality* 49, 1396–1407. <https://doi.org/10.1002/jeq2.20133>
- Zhang, H., Qian, W., Wu, L., Yu, S., Wei, R., Chen, W., Ni, J., 2022. Spectral characteristics of dissolved organic carbon (DOC) derived from biomass pyrolysis: Biochar-derived DOC versus smoke-derived DOC, and their differences from natural DOC. *Chemosphere* 302, 134869. <https://doi.org/10.1016/j.chemosphere.2022.134869>