Supplemental material for:

Total Synthesis of Bicyclomahanimbine by Cu(II)-Promoted Photoredox Process

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Materials and Methods

Unless otherwise stated, reactions were performed in oven-dried glassware fitted with rubber septa under an inert atmosphere and were stirred with Teflon-coated magnetic stirring bars. Liquid reagents and solvents were transferred via syringe using standard Schlenk techniques. Tetrahydrofuran (THF) and diethyl ether (Et₂O) were distilled over sodium/benzophenone ketyl. Dichloromethane (CH₂Cl₂), toluene, and benzene were distilled over calcium hydride. All other solvents and reagents were used as received unless otherwise noted. Reaction temperatures above 25 °C refer to oil bath temperature. Thin layer chromatography was performed using silica gel 60 F-254 precoated plates (0.25 mm) and visualized by UV irradiation, *p*-anisaldehyde stain, and other stains. Silica gel of particle size 100-200 mesh and 230-400 mess were used for flash column chromatography. Melting points were recorded on a digital melting point apparatus and are uncorrected. ¹H and ¹³C NMR spectra were recorded 400 and 500 MHz spectrometers with ¹³C operating frequencies of 100 and 125 MHz respectively. Chemical shifts (δ) are reported in ppm relative to the residual solvents, CDCl₃ signal ($\delta = 7.28$ for ¹H NMR and $\delta = 77.0$ for ¹³C NMR). Data for ¹H NMR spectra are reported as follows: chemical shift (multiplicity, coupling constants, and a number of hydrogen). Abbreviations are as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad). IR spectra were recorded on a FT-IR system (Spectrum BX) and are reported in the frequency of absorption (cm⁻¹). Only selected IR absorbencies are reported. High-Resolution Mass Spectrometry (HRMS) data were recorded on MicrOTOF-Q-II mass spectrometer using methanol as solvent.

Procedure for methylation of o-cresol:



To a solution of o-cresol (**22**) (4.14 g, 38.36 mmol, 1.00 equiv.) in acetone (42 mL), was added potassium carbonate (6.3 g, 44.03 mmol, 1.2 equiv.) followed by addition of dimethylsulphate (4.1 mL, 44.03 mmol, 1.2 equiv.) at 25 °C, and then the reaction mixture was warmed to 60 °C and vigorously stirred it for 2 h. After completion of reaction (monitored by TLC), then the reaction mixture was evaporated under lower pressure and the reaction mixture was diluted with ethyl acetate (55 mL) and water (42 mL). Then the biphasic layers were separated out and the remaining aqueous layer was further extracted with ethyl acetate (40 mL X 2). The combined organic layers were washed with brine (15 mL), dehydrated over Na₂SO₄, and evaporated under lowered pressure. The crude extract was purified by flash chromatography with n-hexane: EtOAc (49:1) to give 1-methoxy-2-methylbenzene (**23**).



1-Methoxy-2-methylbenzene (23): The compound (23) obtained as colorless oil (38.36 mmol; 4.3 g; 92%). $R_f = 0.7$ (in 2.5% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.33 (dd, *J* = 17.8, 7.8 Hz, 2H), 7.04 (t, *J* = 7.4 Hz, 1H), 6.98 (d, *J* = 8.2 Hz, 1H), 3.96 (s, 3H), 2.43 (s, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃): δ 157.8, 130.6, 126.8, 126.6, 120.3, 109.9, 55.2, 16.2.

IR (film): v_{max} 3089, 2854, 1512, 1461, 1232, 952 cm⁻¹.

Procedure for bromination of compound 23:



To a solution of 1-methoxy-2-methylbenzene (**23**) (4.1 g, 33.3 mmol, 1.00 equiv.) in acetonitrile (40 mL) was added *N*-bromo succinimide (NBS) (7.11 g, 39.96 mmol, 1.2 equiv.) at 25 °C. The reaction solution was stirred for 2 hours after it was quenched using saturated aqueous sodium thiosulfate (Na₂S₂O₃) (15 mL). The acetonitrile was evaporated and extracted with EtOAc (60 mL) and water (40 mL). The combined organic layers were washed with brine (15 mL), dehydrated over Na₂SO₄ and evaporated under lower pressure and the unrefined product was purified by flash chromatography with *n*-hexane: EtOAc (49:1) to give 4-bromo-1-methoxy-2-methylbenzene (**13**).



4-Bromo-1-methoxy-2-methylbenzene (13): Compound 13 obtained as a white solid (33.3 mmol; 5.7 g; 85%). $R_f = 0.45$ (in 5% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.29 (d, *J* = 7.1 Hz, 2H), 6.71 (d, *J* = 9.3 Hz, 1H), 3.83 (s, 3H), 2.23 (s, 3H).

¹³C{¹H} NMR (125 MHz, CDCl₃): δ 156.9, 133.2, 129.4, 129.0, 112.3, 111.5, 55.5, 16.1.

IR (film): v_{max} 3412, 3025, 1345, 1127, 957 cm⁻¹.

Procedure for preparation of boronic acid 11:



To a solution of 4-bromo-1-methoxy-2-methylbenzene (**13**) (5.6 g, 27.8 mmol, 1 equiv.) in dry THF (56 mL) was added ^{*n*}BuLi (12 mL, 30.6 mmol, 1.1 equiv.) dropwise at -78 °C. After stirring for 30 min trimethyl borate (B(OMe)₃) (7.8 mL, 69.5 mmol, 2.5 equiv.) was added dropwise. Then the reaction mixture was warmed to 25 °C. The reaction solution was stirred for 10 hours before it was quenched using 4(*N*) HCl. Then the reaction mixture was stirred for 4 hours and extracted with EtOAc (60 mL) and water (45 mL). The combined organic layers were washed with brine, dehydrated over Na₂SO₄ and evaporated under lower pressure. The crude solid product was washed with hexane to give (4-methoxy-3-methylphenyl) boronic acid, (**11**).



4-Methoxy-3-methylphenyl) boronic acid (11): Compound **11** obtained as white powder (27.8 mmol; 3.6 g; 78%). $R_f = 0.3$ (in 5% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, DMSO-d₆): δ 7.64 (d, *J* = 8.1 Hz, 1H), 7.57 (s, 1H), 6.88 (d, *J* = 8.2 Hz, 1H), 3.78 (s, 3H), 2.14 (s, 3H).

¹³C {¹H} NMR (126 MHz, DMSO-d₆): δ 159.5, 137.0, 134.2, 129.8, 124.5, 109.7, 55.4, 16.5.

IR (film): v_{max} 3058, 2845, 1425, 1329, 1175, 921 cm⁻¹.

Procedure for Suzuki reaction between compound 12 and 11:



To a solution of (4-methoxy-3-methylphenyl) boronic acid (**11**) (3.5 g, 21.0 mmol, 1.2 equiv.) in ethanol (14 mL) was added 2-bromonitrobenzene (3.5 g, 17.3 mmol, 1.0 equiv.) (**12**) in benzene (42 mL) followed by potassium carbonate (4.8 g, 34.6 mmol, 2.0 equiv.) dissolved in water (14 mL) and degassed the reaction mixture for 15 min. Next, tetrakis(triphenylphosphine)palladium [Pd (PPh₃)₄] (400 mg, 0.346 mmol, 0.02 equiv.) was added and the reaction mixture was warmed to 80 °C for 15 hours. When the reaction was completed (monitored by checking TLC), then the benzene was evaporated under lower pressure, and the crude product was purified by flash chromatography with *n*-hexane: EtOAc (35:1) to give **10**.



4'-Methoxy-3'-methyl-2-nitro-1,1'-biphenyl (10): Compound 10 obtained as yellow crystal solid (17.30 mmol; 3.5 g; 83%). $R_f = 0.46$ (in 2.5% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.83 – 7.80 (m, 1H), 7.62 – 7.58 (m, 1H), 7.45 (td, *J* = 7.4, 1.3 Hz, 2H), 7.17 – 7.13 (m, 2H), 6.89 (d, *J* = 8.1 Hz, 1H), 3.89 (s, 3H), 2.28 (s, 3H).

¹³C {¹H} NMR (126 MHz, CDCl₃): δ 157.9, 149.5, 136.1, 132.0, 131.9, 130.2, 129.0, 127.6, 127.1, 126.4, 123.9, 110.0, 55.4, 16.3.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₁₄H₁₄NO₃: 244.0974, found 244.0973.

IR (film): v_{max} 3075, 2814, 1345, 1334, 1129, 1074, 8768 cm⁻¹.



Procedure for the hydrogenation of compound 10:

In a clean and oven-dried round bottom flask, compound **10** (3.2 g, 13.25 mmol, 1.0 equiv.) was dissolved in methanol (25 mL) and degassed the reaction mixture for 15 min. Then, Pd-C (160 mg, 5 mol% (w/w)) was added to the reaction mixture and hydrogen gas was passed through the solution for 3 h at 25 °C. Upon completion of the reaction (monitored by TLC), methanol was evaporated and the crude product was purified by flash column chromatography with *n*-hexane: EtOAc (20:1) to **14**.



4'-Methoxy-3'-methyl -[1,1'-biphenyl]-2-amine (14): Compound **14** obtained as colorless liquid (11.6 mmol; 2.4 g; 85%). $R_f = 0.25$ (in 5% EtOAc in *n*-hexane).

¹**H** NMR (400 MHz, CDCl₃): δ 7.27 – 7.23 (m, 2H), 7.16 – 7.10 (m, 2H), 6.90 (d, *J* = 8.1 Hz, 1H), 6.82 (td, *J* = 7.4, 1.2 Hz, 1H), 6.77 (ddd, *J* = 7.8, 1.2, 0.6 Hz, 1H), 3.88 (s, 3H), 3.63 (d, *J* = 5.21 Hz, 2H), 2.28 (s, 3H).

¹³C {¹H} NMR (126 MHz, CDCl₃): δ 157.9, 149.5, 136.1, 132.0, 131.9, 130.2, 129.0, 127.6, 127.1, 126.4, 123.9, 110.0, 55.4, 16.3.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₁₄H₁₆NO: 214.1232, found 214.1235.

IR (film): v_{max} 2928, 1389, 1341, 1041, 951, 845 cm⁻¹.



Procedure for the acetylation of compound 14:

In a clean and oven-dried round bottom flask, compound **14** (2.1 g, 9.7 mmol, 1.0 equiv.) was dissolved in dichloromethane (25 mL) was added. Triethylamine (2 ml, 15.6 mmol, 1.5 equiv), acetic anhydride (1.1 mL, 11.6 mmol, 1.2 equiv) and 4-dimethylaminopyridine (237 mg, 1.94 mmol, 0.2 equiv) sequentially at 0 °C. Then, the reaction was warmed to 25 °C and stirred it for 3 h. Upon completion of the reaction (monitored by TLC), the reaction mixture was quenched with water (15 mL) and the biphasic layers were separated and the aqueous layer was further extracted with dichloromethane (30 mL X 2). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄ and concentrated under reduced pressure then the crude product was purified by flash column chromatography with *n*-hexane:EtOAc (7:1) to give **15**.



N-(4'-methoxy-3'-methyl-[1,1'-biphenyl]-2-yl) acetamide (15): Compound 15 obtained as white solid (9.7 mmol; 2.3 g; 91%). $R_f = 0.41$ (in 30% EtOAc in *n*-hexane).

¹**H NMR** (400 MHz, CDCl₃): δ 8.26 (d, *J* = 8.2 Hz, 1H), 7.34 – 7.30 (m, 1H), 7.25 (d, *J* = 0.5 Hz, 1H), 7.20 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.17 – 7.14 (m, 1H), 7.13 (d, *J* = 1.9 Hz, 2H), 6.91 (d, *J* = 8.2 Hz, 1H), 3.88 (s, 3H), 2.26 (s, 3H), 2.02 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃): δ 172.0, 157.6, 135.0, 131.7, 130.2, 129.9, 128.1, 127.7, 124.2, 121.3, 55.5, 16.4.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₁₆H₁₈NO₂: 256.1338, found 256.1329.

IR (film): v_{max} 3025, 2845, 1625, 1212, 1189, 914, 821 cm⁻¹.

Procedure for the synthesis of compound 16:



A 10 mL glass vial was poured with biaryl acetamide **15** (500 mg, 2.0 mmol, 1.0 equiv.) to which 1,1,1,3,3,3-hexfluoroisopropanol (5 mL) followed by sodium ethoxide (100 mg, 1.5 mmol, 0.75 equiv.) were added. A lid with graphite electrodes (52 x 5 mm) was attached and electrolysis was carried out with a constant current of 3 mA until 3.1 F/mol charge was passed (~ 9 h) at room temperature. Upon completion of the reaction (monitored by TLC), the reaction mixture was transferred to a 50 mL round bottom flask to which electrodes were washed with methanol. Solvents were evaporated under reduced pressure. Then the crude product was purified by flash column chromatography with *n*-hexane: EtOAc (10:1) to give **16**.



1-(2-Methoxy-3-methyl-9H-carbazol-9-yl) ethane-1-one (16): Compound **15** obtained as white solid (2.0 mmol; 277 mg; 73%). $R_f = 0.34$ (in 10% EtOAc in *n*-hexane).

¹**H** NMR (400 MHz, CDCl₃): δ 7.94 (d, *J* = 12.8 Hz, 2H), 7.87 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.69 (s, 1H), 7.35 (td, *J* = 7.2, 1.5 Hz, 2H), 3.94 (s, 3H), 2.87 (s, 3H), 2.35 (s, 3H).

¹³C {¹H} NMR (101 MHz, CDCl₃): δ 170.2, 158.1, 138.6, 138.1, 127.1, 125.5, 123.6, 123.6, 120.8, 119.2, 118.8, 115.5, 99.4, 55.8, 27.8, 16.7.

HRMS (ESI) m/z: $[M + H]^+$ calcd for $C_{16}H_{16}NO_2$: 254.1181, found 254.1171.

IR (film): v_{max} 2912, 1412, 1392, 1274, 1058, 1021 cm⁻¹.

Procedure for the hydrolysis of compound 16:



To a solution of compound **16** (813 mg, 3.21 mmol, 1 equiv.) in a mixed solvent system of MeOH: CHCl₃ (3:1) (12 mL) was added potassium carbonate (665 mg, 4.8 mmol, 1.5 eq.) at 25 °C and the reaction mixture was stirred for 3 h at the same temperature. After complete consumption of the starting material (monitored by TLC) the solution was diluted with CH_2Cl_2 (6 mL) and water (8 mL). The biphasic layers were separated and the aqueous layer was further extracted with dichloromethane (12 mL X 2). The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. The residual crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (7:1) to give **17**.



2-Methoxy-3-methyl-9H-carbazole (17): Compound 17 obtained as yellowish powder (3.21 mmol; 603 mg; 89%). $R_f = 0.2$ (in 10 % EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, DMSO-d₆): δ 10.99 (s, 1H), 7.94 (d, J = 7.4 Hz, 1H), 7.81 (d, J = 4.3 Hz, 1H), 7.40 (d, J = 7.3 Hz, 1H), 7.26 (dt, J = 6.5, 2.5 Hz, 1H), 7.09 (dt, J = 6.2, 3.0 Hz, 1H), 6.96 (dd, J = 5.9, 3.8 Hz, 1H), 3.88 (dt, J = 5.5, 3.3 Hz, 3H), 2.28 (dq, J = 7.3, 3.5 Hz, 3H).

¹³C{¹H} NMR (126 MHz, DMSO-d₆): δ 157.2, 140.0, 139.9, 124.2, 123.1, 121.6, 119.5, 118.8, 117.8, 115.7, 111.0, 93.1, 55.8, 17.1.

HRMS (ESI) m/z: $[M + H]^+$ calcd for $[C_{14}H_{14}NO + H]^+$: 212.1075, found 212.1075.

IR (film): v_{max} 3421, 3042, 2841, 1632, 1245, 1056 cm⁻¹.

Procedure for the demethylation of 24:



To a solution of compound **24** (538 mg, 2.55 mmol, 1.0 equiv.) in dichloromethane (8 mL) was added boron tribromide (725 μ l, 7.67 mmol, 3 equiv.) in dropwise manner at 0 °C. Then the reaction mixture was warmed to 25 °C and stirred it for 4 h. Upon complete consumption of starting materials (confirmed by checking TLC), the reaction mixture was quenched with water (8 mL) and the biphasic layers were separated and the aqueous layer was further extracted with dichloromethane (12 mL X 2). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and evaporated under lower pressure, then the crude product was purified by flash silica gel column chromatography with *n*-hexane: EtOAc (4:1) to give **9**.



2-Hydroxy-3-methylcarbazole (9): The compound (9) obtained as white solid (2.55 mmol; 357 mg; 71%). $R_f = 0.41$ (in 30% EtOAc in *n*-hexane).

¹**H NMR** (400 MHz, DMSO-D₆): δ 10.78 (d, *J* = 2.2 Hz, 1H), 9.38 (s, 1H), 7.84 (d, *J* = 7.7 Hz, 1H), 7.69 (s, 1H), 7.31 (d, *J* = 8.1 Hz, 1H), 7.20 – 7.14 (m, 1H), 7.01 (tt, *J* = 7.5, 1.3 Hz, 1H), 6.85 (d, *J* = 2.2 Hz, 1H), 2.22 (s, 3H).

¹³C {¹H} NMR (126 MHz, DMSO): δ 155.1, 140.1, 139.9, 123.9, 123.4, 121.6, 119.2, 118.6, 116.8, 115.4, 110.7, 96.5, 17.0.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₁₃H₁₂NO: 198.0919, found 198.0920.

IR (film): v_{max} 3425, 2912, 2856, 1621, 1187, 953, 824 cm⁻¹.

Procedure for the synthesis of mahanimbine (1):



In a clean and oven-dried round bottom flask, carbazole **9** (237 mg, 1.2 mmol, 1.0 equiv.) was dissolved in pyridine (5 mL). Citral (2.0 ml, 12 mmol, 10 equiv) was added to the reaction mixture

at 25 °C. Then the reaction mixture was warmed to 110 °C and stirred it for 10 h. Upon complete consumption of starting materials (confirmed by checking TLC), then the reaction mixture was evaporated under lower pressure and diluted with ethyl acetate (7 mL) and water (5 mL). The biphasic layers were separated and the aqueous layer was further extracted with ethyl acetate (7 mL X 2). The combined organic layers were dried over Na₂SO₄ and evaporated under vacuum. The crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (20:1) to give mahanimbine (**1**).



3,5-Dimethyl-3-(4-methylpent-3-en-1-yl)-3,11-dihydropyrano[**3,2-***a*]**carbazole**(**1**): Compound (**1**) obtained as yellowish liquid (1.2 mmol; 302 mg; 76%). $R_f = 0.45$ (in 10% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.94 (dd, J = 7.7, 1.1 Hz, 1H), 7.88 (s, 1H), 7.69 (s, 1H), 7.40 (dt, J = 8.1, 0.9 Hz, 1H), 7.35 – 7.31 (m, 1H), 7.21 (td, J = 7.4, 1.1 Hz, 1H), 6.67 (d, J = 9.8 Hz, 1H), 5.68 (d, J = 9.8 Hz, 1H), 5.15 (tt, J = 7.2, 1.5 Hz, 1H), 2.37 (s, 3H), 2.25 – 2.17 (m, 2H), 1.82 – 1.78 (m, 2H), 1.70 (s, 3H), 1.62 (s, 3H), 1.60 (s, 1H), 1.49 (s, 3H).

¹³C{¹H} NMR (100 MHz, CDCl₃): δ 150.0, 139.5, 135.0, 131.8, 128.6, 124.3, 124.3, 124.0, 121.3, 119.6, 119.4, 118.5, 117.6, 116.7, 110.5, 104.3, 78.3, 40.9, 25.9, 25.8, 22.9, 17.7, 16.2.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₃H₂₆NO: 332.2004, found 332.2014.

IR (film): v_{max} 3319, 2941, 2842, 1712, 1612, 1423, 1042 cm⁻¹.

Comparison of ¹H-NMR Data:

Ali's report ²³ on	Knolker's report ¹⁰ on	This report of mahanimbine [1]
mahanimbine [1]	mahanimbine [1]	(¹ H NMR, 500 MHz, CDCl ₃)
(¹ H NMR, 400 MHz, CDCl ₃)	(¹ H NMR, 500 MHz, CDCl ₃)	
7.91 (d, <i>J</i> = 7.8 Hz, 1H)	7.93 (d, <i>J</i> = 7.7 Hz, 1H)	7.94 (dd, <i>J</i> = 7.7, 1.1 Hz, 1H)
7.86 (s, 1H)	7.87 (brs, 1H)	7.88 (s, 1H)
7.66 (s, 1H)	7.68 (s, 1H)	7.69 (s, 1H)
7.37 (d, <i>J</i> = 8.0 Hz, 1H)	7.37 (d, <i>J</i> = 7.9 Hz, 1H)	7.40 (dt, <i>J</i> = 8.1, 0.9 Hz, 1H)
7.30 (t, <i>J</i> = 7.5 Hz, 1H)	7.32 (brt, $J = 7.5$ Hz, 1H)	7.35 – 7.31 (m, 1H)
7.17 (t, <i>J</i> = 7.4 Hz, 1H)	7.19 (brt, <i>J</i> = 7.3 Hz, 1H)	7.21 (td, <i>J</i> = 7.4, 1.1 Hz, 1H)
6.65 (d, <i>J</i> = 9.8 Hz, 1H)	6.63 (d, <i>J</i> = 9.8 Hz, 1H)	6.67 (d, <i>J</i> = 9.8 Hz, 1H)
5.66 (d, <i>J</i> = 9.8 Hz, 1H)	5.66 (d, <i>J</i> = 9.8 Hz, 1H)	5.68 (d, <i>J</i> = 9.8 Hz, 1H)
5.15 (t, <i>J</i> = 7.1 Hz, 1H)	5.14 (t, <i>J</i> = 7.1 Hz, 1H)	5.15 (tt, <i>J</i> = 7.2, 1.5 Hz, 1H)
2.33 (s, 3H)	2.37 (s, 3H)	2.37 (s, 3H)
2.19 – 2.14 (m, 2H)	2.19 (m, 2H)	2.25 – 2.17 (m, 2H)
1.79 – 1.74 (m, 2H)	1.79 (t, J = 8.3 Hz, 2H)	1.82 – 1.78 (m, 2H)
1.66 (s, 3H)	1.69 (s, 3H)	1.70 (s, 3H)
1.58 (s, 3H)	1.61 (s, 3H)	1.62 (s, 3H)
1.56 (s, 3H)	-	1.60 (s, 1H)
1.45 (s, 3H)	1.47 (s, 3H)	1.49 (s, 3H)

Comparison of ¹³C-NMR Data:

Ali's report ²³ on	Knolker's report ¹⁰ on	This report mahanimbine
mahanimbine [1]	mahanimbine [1]	[1]
(¹³ C NMR, 100 MHz,	(¹³ C NMR, 125 MHz,	(¹³ C NMR, 100 MHz,
CDCl ₃)	CDCl ₃)	CDCl ₃)
149.9	149.86	150.0
139.4	139.41	139.5
134.8	134.82	135.0

131.7	131.65	131.8
128.5	128.44	128.6
124.2	124.16	124.3
124.2		124.3
123.9	123.86	124.0
121.2	121.15	121.3
119.5	119.41	119.6
119.3	119.25	119.4
118.4	118.36	118.5
117.5	117.49	117.6
116.6	116.56	116.7
110.3	110.56	110.5
104.2	104.16	104.3
78.1	78.12	78.3
40.7	40.72	40.9
25.8	25.79	25.9
25.7	25.65	25.8
22.7	22.72	22.9
17.6	17.55	17.7
16.1	16.07	16.2



Procedure for the synthesis of bicyclomahanimbine (2):

To a solution of mahanimbine (1) (59 mg, 0.18 mmol, 1.0 equiv.) in diethyl ether (3 mL) was added copper triflate (7 mg, 0.018 mmol, 10 mol%) in presence of 500 W Na-lamp for 10 h. When the reaction was completed (confirmed by checking TLC), then the reaction mixture was extracted with EtOAc (6 mL X 3) and water (5 mL). The combined organic layers were dried over Na₂SO₄ and evaporated under vacuum. The crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (7:1) to give bicyclomahanimbine (**2**).



Bicyclomahanimbine (2): Compound **2** obtained as yellowish liquid (0.18 mmol; 52 mg; 87%). $R_f = 0.55$ (in 10% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.96 (dd, *J* = 7.8, 1.1 Hz, 1H), 7.71 (d, *J* = 0.8 Hz, 1H), 7.45 (s, 1H), 7.40 (dt, *J* = 8.0, 1.0 Hz, 1H), 7.33 – 7.31 (m, 1H), 7.21 – 7.18 (m, 1H), 3.32 (d, *J* = 9.5 Hz, 1H), 2.73 (dd, *J* = 9.5, 7.7 Hz, 1H), 2.54 (td, *J* = 8.3, 7.8 Hz, 1H), 2.38 (s, 3H), 2.11 – 2.07 (m, 1H), 1.76 (ddd, *J* = 7.1, 4.4, 1.8 Hz, 1H), 1.70 – 1.66 (m, 2H), 1.59 (s, 3H), 1.47 (s, 3H), 0.78 (s, 3H).

¹³C {¹H} NMR (125 MHz, CDCl₃): δ 150.4, 139.3, 137.8, 124.2, 123.9, 120.0, 119.3, 119.3, 119.3, 115.8, 110.3, 106.3, 83.5, 46.5, 39.3, 38.4, 37.9, 37.4, 35.1, 27.4, 25.6, 18.6, 16.7.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₃H₂₆NO :332.2014, found 332.2013. **IR** (film): v_{max} 3456, 2934, 1607, 1321, 1012, 951 cm⁻¹.

Comparison of ¹H-NMR Data:

Ali's report ²³ of	Knolker's report ¹⁰ of	Nafiah's report ²⁴ of	This report of report of
bicyclomahanimbine	bicyclomahanimbine [2]	bicyclomahanimbine [2]	bicyclomahanimbine [2]
[2]	(¹ H NMR, 500 MHz,	(¹ H NMR, 500 MHz,	(¹ H NMR, 500 MHz,
(¹ H NMR, 500 MHz,	CDCl ₃)	CDCl ₃)	CDCl ₃)
CDCl ₃)			
7.92 (d I - 7.7 Hz 1H)	7.94 (d, <i>J</i> = 7.7 Hz, 1H)	7.93 (d, <i>J</i> = 8.1 Hz, 1H)	7.96 (dd, <i>J</i> = 7.8, 1.1
7.52 (d, $3 - 7.7$ 112, 111)			Hz, 1H)
7.67 (s, 1H)	7.69 (s, 1H)	7.66 (s, 1H)	7.71 (d, $J = 0.8$ Hz, 1H)
7.41 (s, 1H)	7.43 (brs, 1H)	7.44 (brs, 1H)	7.45 (s, 1H)
7.36(d I - 8.0 Hz 1H)	7.38 (d, <i>J</i> = 8.0 Hz, 1H)	7.37 (d, <i>J</i> = 8.1 Hz, 1H)	7.40 (dt, $J = 8.0$, 1.0 Hz,
7.50 (u, J = 0.0 Hz, HI)			1H)
7.32 7.26 (m. 1H)	7.30 (brt, $J = 7.6$ Hz,	7.28 (td, <i>J</i> = 8.1, 1.0 Hz,	7.33 7.31 (m. 1H)
7.32 – 7.20 (m, m)	1H)	1H)	7.55 – 7.51 (m, 111)
7.20 7.14 (m 1H)	7.18 (brt, $J = 7.4$ Hz,	7.15 (td, <i>J</i> = 8.1, 1.0 Hz,	7.21 $7.18 (m. 1H)$
7.20 – 7.14 (III, 111)	1H)	1H)	7.21 – 7.18 (III, 111)
3.26 (d, J = 9.5 Hz, 1H)	3.30 (d, J = 9.5 Hz, 1H)	3.30 (d, J = 9.2 Hz, 1H)	3.32 (d, J = 9.5 Hz, 1H)
2.68 (dd, <i>J</i> = 9.3, 8.0	2.71 (dd, <i>J</i> = 9.3, 7.9	2.71 (t, <i>J</i> = 7.5 Hz, 1H)	2.73 (dd, <i>J</i> = 9.5, 7.7
Hz, 1H)	Hz, 1H)		Hz, 1H)
2.40 (t I - 6.2 Hz 1H)	2.51 (brt, $J = 6.6$ Hz,	2.50 (t, <i>J</i> = 7.4 Hz, 1H)	2.54 (td, <i>J</i> = 8.3, 7.8 Hz,
2.49 (t, $J = 0.2$ HZ, HI)	1H)		1H)
2.35 (s, 3H)	2.36 (s, 3H)	2.33 (s, 3H)	2.38 (s, 3H)
2.12 – 1.99 (m, 1H)	2.06 – 2.09 (m, 1H)	2.01 – 2.07 (m, 1H)	2.11 – 2.07 (m, 1H)
1.79.1.60 (m. 1H)	1.64.1.76 (m. 2H)	1.64 - 1.68 (m, 1H)	1.76 (ddd, <i>J</i> = 7.1, 4.4,
1./0-1.09 (III, 1П)	1.04-1.70 (III, 3 Π)		1.8 Hz, 1H)
1.69 – 1.61 (m, 2H)		-	1.70 – 1.66 (m, 2H)

1.54 (s, 3H)	1.57 (s, 3H)	1.57 (s, 3H)	1.59 (s, 3H)
-	-	1.55 - 1.57 (m, 1H)	-
-	-	1.53 - 1.57 (m, 1H)	-
1.44 (s, 3H)	1.45 (s, 3H)	1.43 (s, 3H)	1.47 (s, 3H)
0.74 (s, 3H)	0.75 (s, 3H)	0.74 (s, 3H)	0.78 (s, 3H)

Comparison of ¹³C-NMR Data:

Ali's report ²³ of	Knolker's report ¹⁰ of	Nafiah's report ²⁴ of	This report
bicyclomahanimbine	bicyclomahanimbine	bicyclomahanimbine	bicyclomahanimbine
[2]	[2]	[2]	[2]
(¹³ C NMR, 126	(¹³ C NMR, 125 MHz,	(¹³ C NMR, 125	(¹³ C NMR, 100 MHz,
MHz, CDCl ₃)	CDCl ₃)	MHz, CDCl ₃)	CDCl ₃)
150.33	150.35	150.2	150.4
139.20	139.21	139.3	139.3
137.74	137.75	137.8	137.8
124.17	124.16	124.2	124.2
123.86	123.87	124.0	123.9
119.98	120.00	120.1	120.0
119.30	119.31	119.4	119.3
119.28	119.28	119.4	119.3
119.24	119.26	119.2	119.3
115.73	115.74	115.8	115.8
110.26	110.25	110.4	110.3
106.30	106.30	106.5	106.3
83.49	83.51	83.5	83.5

46.38	46.40	46.5	46.5
39.24	39.27	39.5	39.3
39.29	38.31	38.4	38.4
37.75	37.78	37.9	37.9
37.30	37.34	37.5	37.4
35.03	35.07	35.2	35.1
27.40	27.41	27.5	27.4
25.58	25.60	25.7	25.6
18.56	18.58	18.6	18.6
16.70	16.70	16.8	16.7

Synthesis of Curruyanin (3):



In a clean and oven-dried round bottom flask, mahanimbine (1) (166 mg, 0.50 mmol, 1.0 equiv.) dissolved in toluene (5 mL). Camphor sulphonic acid (CSA) (174 mg, 0.75 mmol, 1.5 equiv) was added to the reaction mixture and was irradiated with light using a 500W Na-lamp for 24 h. Upon complete consumption of starting materials (monitored by TLC), volatiles were evaporated under lower pressure and extracted with ethyl acetate (7 mL X 3) and water (5 mL). The combined organic layers were dried over Na₂SO₄ and evaporated under vacuum. The crude product was purified by flash chromatography with *n*-hexane: EtOAc (50:1) to give curruyanin (**3**).



Curruyanin (3): Compound (3) obtained as white solid (0.50 mmol; 154 mg; 93%). $R_f = 0.65$ (in 10% EtOAc in *n*-hexane).

¹**H NMR** (500 MHz, CDCl₃): δ 7.91 (dq, *J* = 7.7, 0.9 Hz, 1H), 7.74 (s, 1H), 7.66 (t, *J* = 0.9 Hz, 1H), 7.34 (d, J = 7.9 Hz, 1H), 7.30 (d, *J* = 1.2 Hz, 1H), 7.17 (ddd, *J* = 7.9, 7.0, 1.1 Hz, 1H), 4.84 (s, 1H), 4.76 (s, 1H), 3.44 (d, *J* = 3.1 Hz, 1H), 2.65 – 2.57 (m, 1H), 2.36 (s, 3H), 2.12 (dt, *J* = 9.2, 2.4 Hz, 1H), 2.06 (dd, *J* = 12.9, 2.7 Hz, 1H), 1.94 – 1.90 (m, 1H), 1.68 – 1.63 (m, 2H), 1.53 (s, 3H), 1.46 (s, 3H).

¹³C {¹H} NMR (100 MHz, CDCl₃): δ 153.7, 150.2, 139.5, 138.4, 124.4, 123.6, 119.6, 119.2, 119.1, 117.7, 114.7, 112.1, 110.3, 105.3, 77.3, 74.0, 48.8, 40.0, 37.5, 36.3, 29.1, 23.2, 21.7, 16.8.

HRMS (ESI) m/z: $[M + H]^+$ calcd for C₂₃H₂₆NO :332.2007, found 332.2014.

IR (film): v_{max} 3421, 2912, 2815, 1458, 1378, 1229, 1105 cm⁻¹.

Comparison of ¹H-NMR Data:

Ali's report ²³ of curruyanin	Knolker's report ¹⁰ of curruyanin [3]	This report of curruyanin [3]
[3]	(¹ H NMR, 500 MHz, CDCl ₃)	(¹ H NMR, 500 MHz,
(¹ H NMR, 500 MHz, CDCl ₃)		CDCl ₃)
$7.88 (J I - 7.7 U_{7} 1U)$	7.90 (d, <i>J</i> = 7.7 Hz, 1H)	7.91 (dq, <i>J</i> = 7.7, 0.9 Hz,
7.00 (u, J - 7.7 112, 111)		1H)

7.68 (s, 1H)	7.73 (brs, 1H)	7.74 (s, 1H)
7.62 (s, 1H)	7.65 (s, 1H)	7.66 (t, <i>J</i> = 0.9 Hz, 1H)
7.29 (d, <i>J</i> = 8.0 Hz, 1H)	7.32 (d, $J = 8.0$ Hz, 1H)	7.34 (d, J = 7.9 Hz, 1H)
7.25 (t, <i>J</i> = 7.3 Hz, 1H)	7.27 (ddd, <i>J</i> = 8.1, 7.0, 1.1 Hz, 1H)	7.30 (d, <i>J</i> = 1.2 Hz, 1H)
7 13 (t I = 7 3Hz 1H)	7.15 (brt I – 7.4 Hz 1H)	7.17 (ddd, J = 7.9, 7.0, 1.1
/.15 (t, t = /.512, 111)	/.13 (010, 5 = /.1112, 111)	Hz, 1H)
4.77 (s, 1H)	4.82 (s, <i>J</i> = 1.6 Hz, 1H)	4.84 (s, 1H)
4.69 (s, 1H)	4.75 (s, 1H)	4.76 (s, 1H)
3.32 (s, 1H)	3.42 (d, <i>J</i> = 2.6 Hz, 1H)	3.44 (d, <i>J</i> = 3.1 Hz, 1H)
2.56 – 2.45 (m, 1H)	2.59 (m, 1H)	2.65 – 2.57 (m, 1H)
2.33 (s, 3H)	2.34 (s, 3H)	2.36 (s, 3H)
2.05 (d, J = 9.1 Hz, 1H)	2 10 (dt I = 91 21 Hz 1H)	2.12 (dt, <i>J</i> = 9.2, 2.4 Hz,
		1H)
-	2.03 (dd, <i>J</i> = 12.8, 2.6 Hz, 1H)	2.06 (dd, <i>J</i> = 12.9, 2.7 Hz,
		1H)
1.99 – 1.89 (m, 1H)	1.90 (dt, <i>J</i> = 12.8, 3.1 Hz, 1H)	1.94 – 1.90 (m, 1H)
1.87 – 1.77 (m, 1H)	-	-
1.65 – 1.51 (m, 1H)	1.59 – 1.68 (m, 2H)	1.68 – 1.63 (m, 2H)
1.45 (s, 3H)	1.51 (s, 3H)	1.53 (s, 3H)
1.40 (s, 3H).	1.44 (s, 3H).	1.46 (s, 3H).

Comparison of ¹³C-NMR Data:

Ali's report ²³ of curruyanin [3]	Knolker's report ¹⁰ of curruyanin [3]	This report of curruyanin [3]
(¹³ C NMR, 126 MHz, CDCl ₃)	(¹³ C NMR, 126 MHz, CDCl ₃)	(¹³ C NMR, 100 MHz, CDCl ₃)

153.59	153.61	153.7
149.97	150.06	150.2
139.39	139.34	139.5
138.29	138.38	138.4
124.21	124.24	124.4
123.46	123.47	123.6
119.45	119.47	119.6
119.07	119.08	119.2
119.00	119.02	119.1
117.52	117.58	117.7
114.54	114.57	114.7
111.97	112.02	112.1
110.15	110.15	110.3
105.17	105.19	105.3
73.87	73.91	74.0
48.60	48.69	48.8
39.84	39.90	40.0
37.27	37.35	37.5
36.12	36.22	36.3
28.95	28.98	29.1
23.03	23.08	23.2
21.54	21.59	21.7
16.78	16.76	16.8

Procedure for epoxidation of curryanin (3):



To a solution of curruyanin (3) (132 mg, 0.40 mmol, 1.0 equiv.) in dichloromethane (4 mL) was added meta-chloroperoxybenzoic acid (83 mg, 0.48 mmol, 1.2 equiv.) at 25 °C for 3 h. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with saturated aqueous solution of sodium bicarbonate (3 mL) and the biphasic layers were separated and the aqueous layer was further extracted with dichloromethane (5 mL X 2). The combined organic layers were washed with brine (3 mL), dried over Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (7:1) to give **21**.



(5,7-Dimethyl-2-2-methyloxiran-2-yl)-1,2,3,4,5,13-hexahydro-1,5-methanooxocino[3,2-a] carbazole (21): Compound 21 obtained as yellow solid (0.40 mmol; 128 mg; 92%). $R_f = 0.4$ (in 10% EtOAc in *n*-hexane).

¹**H** NMR (500 MHz, CDCl₃): δ 8.71 (s, 1H), 7.91 (dt, *J* = 7.8, 0.7 Hz, 1H), 7.68 (s, 1H), 7.42 (d, *J* = 8.0 Hz, 1H), 7.28 – 7.25 (m, 1H), 7.16 – 7.12 (m, 1H), 3.58 (d, *J* = 3.1 Hz, 1H), 2.71 (dd, *J* = 5.2, 0.8 Hz, 1H), 2.64 (d, *J* = 5.1 Hz, 1H), 2.34 (s, 3H), 2.11 – 2.07 (m, 1H), 1.85 – 1.83 (m, 2H), 1.73 (dt, *J* = 12.4, 3.2 Hz, 2H), 1.59 – 1.56 (m, 2H), 1.43 (s, 3H), 0.66 (s, 3H).

¹³C{¹H} NMR (126 MHz, CDCl₃): δ 153.6, 139.6, 138.1, 124.1, 123.6, 119.7, 119.0, 118.9, 117.5, 114.8, 110.6, 104.9, 74.2, 59.3, 58.5, 47.0, 39.7, 37.1, 30.7, 29.0, 22.6, 18.5, 16.8.

HRMS (ESI) m/z: [M + H]⁺calcd for C₂₃H₂₆NO₂: 348.1964, found 348.1964. **IR** (film): v_{max} 3121, 2932, 2842, 1523, 1234, 925 cm⁻¹.

Procedure for the Synthesis of Muruyazolinine (4):



In a clean and oven-dried round bottom flask, **21** (121 mg, 0.35 mmol, 1.0 equiv.) was dissolved in THF (4 mL). LiAlH₄ (33 mg, 0.88 mmol, 2.5 equiv.) was added portion wise over the period of 5 min at 0 °C and the reaction mixture was then warmed to 25 °C and stirred it for 18 h. After completion of the reaction (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of ammonium chloride (2 mL) and the solution was extracted with ethyl acetate (5 mL X 3). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄ and concentrated under reduced pressure. The residual crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (16:1) to give murruyazolinine (**4**)



5,7-Dimethyl-1,2,3,4,5,13-hexahydro-1,5-methanooxocino[3,2-a]carbazol-2-yl)propan-2-ol (4): Compound 4 obtained as white solid (0.35 mmol; 101 mg; 83%). $R_f = 0.25$ (in 10% EtOAc in *n*-hexane).

¹**H NMR** (400 MHz, CDCl₃): δ 7.90 (d, J = 7.8 Hz, 1H), 7.65 (d, J = 0.9 Hz, 1H), 7.37 (d, J = 8.0, 0.9 Hz, 1H), 7.28 – 7.25 (m, 1H), 7.13 (ddd, J = 8.1, 7.1, 1.1 Hz, 1H), 3.78 (q, J = 3.0 Hz, 1H), 2.33 (s, 3H), 2.13 – 2.07 (m, 1H), 1.92 – 1.87 (m, 1H), 1.87 – 1.83 (m, 2H), 1.65 – 1.57 (m, 1H), 1.44 (t, J = 3.6 Hz, 1H), 1.42 (s, 3H), 1.42 – 1.37 (m, 1H), 1.30 (s, 3H), 1.26 (d, J = 1.5 Hz, 1H), 0.54 (s, 3H).

¹³C {¹H} NMR (100 MHz, CDCl₃): δ 153.8, 139.8, 139.0, 124.5, 123.4, 119.5, 119.1, 118.8, 117.5, 114.8, 110.6, 105.9, 74.4, 74.4, 52.8, 40.5, 38.4, 33.4, 29.3, 29.0, 23.3, 23.0, 16.9.

HRMS (ESI) m/z: [M + H]⁺ calcd for C₂₃H₂₈NO₂: 350.2120, found 350.2111.

IR (film): v_{max} 3513, 3012, 2946, 1605, 1456, 1208, 1124 cm⁻¹.

Ali's report ²³ of	Knolker's report ¹⁰ of	Sarpong's report ²²	This report of
muruyazolinine (4)	muruyazolinine (4)	of muruyazolinine	muruyazolinine (4)
(¹ H NMR, 400 MHz,	(¹ H NMR, 600 MHz,	(4)	(¹ H NMR, 400
CDCl ₃)	CDCl ₃)	(¹ H NMR, 600	MHz, CDCl ₃)
		MHz, CDCl ₃)	
7.00(d I - 7.7 Hz 1H)	7.91 (d, <i>J</i> = 7.7 Hz,	7.90 (d, <i>J</i> = 7.7Hz,	7.90 (d, <i>J</i> = 7.8 Hz,
7.90 ($\mathbf{u}, \mathbf{J} = 7.7$ 112, 111)	1H)	1H)	1H)
7.64 (s. 111)	7.66 (d, $J = 0.5$ Hz,	7.66 (c. 111)	7.65 (d, $J = 0.9$ Hz,
7.04 (8, 111)	1H)	7.00 (8, 111)	1H)
7.27 (d I - 8.0 Hz 1H)	7.37 (d, <i>J</i> = 8.0 Hz,	7.37 (d, <i>J</i> = 7.9 Hz,	7.37 (d, $J = 8.0$ Hz,
1.57 (u, J - 0.0 HZ, 1 H)	1H)	1H)	1H)

Comparison of ¹H-NMR data:

7.27 (dd, <i>J</i> = 11.1, 4.0	7.26 (brt, $J = 7.5$ Hz,	7.26 (d, <i>J</i> = 15.3 Hz		
Hz, 1H)	1H)	1H)	7.28 – 7.25 (m, 1H)	
	7.13 (brt, $J = 7.4$ Hz,	7.13 (t, <i>J</i> = 7.4 Hz,	7.13 (ddd, $J = 8.1$,	
/.13 (t, J = /.4 Hz, 1H)	1H)	1H)	7.1, 1.1 Hz, 1H)	
266(4 I - 24 Hz 1H)	3.80 (brd, J = 2.7 Hz,	3.81 (d, J = 3.2 Hz,	3.78 (q, J = 3.0 Hz,	
3.00 (u, J - 2.4 HZ, 1H)	1H)	1H)	1H)	
2.33 (s, 3H)	2.34 (s, 3H)	2.33 (s, 3H)	2.33 (s, 3H)	
204(4 L - 122 Hz 1H)	2.10 (m. 1H)	2.10 (d, <i>J</i> = 13.5 Hz,	$212 \ 207 (m 1H)$	
2.04 (0, J = 15.5 HZ, 1H)	2.10 (III, 1H)	1H)	2.13 - 2.07 (III, 1H)	
	1.00 1.04 (m. 111)	2.01 (d, <i>J</i> = 6.6 Hz,	102 197 (m 111)	
1.77 ($1.7.9$ Hz 2H)	1.90 – 1.94 (III, 1 H)	1H)	1.92 – 1.87 (m, 1H)	
1.77 (u, j - 7.0 Hz, 2H)	1.88 (dd, <i>J</i> = 12.9, 2.8	1.86 (dd, <i>J</i> = 8.2, 3.6		
	Hz 1H)	Hz, 2H)	1.87 1.83 (m. 2H)	
	1.88 (dt, <i>J</i> = 12.9, 2.3	171 (c. 111)	1.07 1.03 (iii, 211)	
	Hz 1H)	1./1 (S, 1H)		
1.54 (m, 2H)	1.60 – 1.65 (m, 1H)	1.65 – 1.59 (m, 1H)	1.65 – 1.57 (m, 1H)	
	$1.41 1.45 \ (m 111)$	1.46 1.42 (m. 111)	1.44 (t, $J = 3.6$ Hz,	
	$1.41 - 1.43$ (III, 1 Π)	1.40 - 1.42 (III, 1H)	1H)	
1.42 -1.34 (m, 5H)	1.43 (s, 3H)	1.43 (s, 3H)	1.42 (s, 3H)	
			1.42 – 1.37 (m,	
	-	-	1H),	
1.19 (s, 3H)	1.32 (s, 3H)	1.33 (s, 3H)	1.30 (s, 3H)	
	1 29 1 21 (m 1H)	$1.21 1.29 \ (m 1H)$	1.26 (d, <i>J</i> = 1.5 Hz,	
-	1.20 – 1.31 (III, 1H)	1.51 – 1.28 (III, 1H)	1H)	
0.45 (s, 3H)	0.55 (s, 3H)	0.55 (s, 3H)	0.54 (s, 3H)	

Comparison of ¹³C-NMR Data:

Ali's report ²³ on muruyazolinine	Knolker's report ¹⁰ of	This report
(4)	muruyazolinine (4)	muruyazolinine (4)

$(^{13}C NMR 126 MHz CDCl_2)$	(¹³ C NMR, 150 MHz, CDCl ₃)	(¹³ C NMR, 100 MHz, CDCl ₂)
		00013)
153.79	153.64	153.8
139.83	139.69	139.8
139.08	138.91	139.0
124.47	124.35	124.5
123.46	123.30	123.4
119.51	119.41	119.5
119.14	118.99	119.1
118.80	118.65	118.8
117.52	117.40	117.5
114.76	114.71	114.8
110.76	110.51	110.6
106.04	105.82	105.9
74.41	74.32	74.4
_	74.26	74.4
52.70	52.76	52.8
40.43	40.41	40.5
38.28	38.28	38.4
33.20	33.36	33.4
29.25	29.21	29.3
28.99	28.87	29.0
23.19	23.22	23.3
22.91	22.86	23.0

17.01	16.80	16.9
17701	10100	1002

Procedure for synthesis of isocyclomahanimbine (5):



To a solution of muruyazolinine (**4**) (72 mg, 0.21 mmol, 1.0 equiv.) in benzene (3 mL) was added *para*-toluene sulfuric acid (*p*-TSA) (43 mg, 0.25 mmol, 1.2 equiv) at 25 °C. Then the reaction mixture was warmed to 80 °C and stirred for 2 h. After complete consumption of starting materials (monitored by TLC), then the volatile was evaporated under reduced pressure and extracted with EtOAc (6 mL) and water (4 mL). The combined organic layers were dried over Na₂SO₄ and evaporated under vacuum. The crude product was purified by silica gel column chromatography with *n*-hexane: EtOAc (20:1) to give isocyclomahanimbine (**5**).



Isocyclomahanimbine (5): Compound **5** obtained as white crystal solid (0.21 mmol; 61 mg; 89%). $R_f = 0.5$ (in 10% EtOAc in *n*-hexane). ¹**H NMR** (500 MHz, CDCl₃): δ 7.91 (d, J = 7.7 Hz, 1H), 7.65 (d, J = 0.9 Hz, 1H), 7.47 (s, 1H), 7.37 (dt, J = 8.0, 0.9 Hz, 1H), 7.30 – 7.29 (m, 1H), 7.19 – 7.15 (m, 1H), 4.29 (s, 1H), 2.43 (d, J = 8.8 Hz, 1H), 2.37 (s, 3H), 2.16 (d, J = 2.0 Hz, 3H), 2.10 – 2.07 (m, 1H), 2.04 – 2.01 (m, 1H), 1.95 (dd, J = 12.6, 2.7 Hz, 1H), 1.90 (s, 1H), 1.69 (d, J = 1.3 Hz, 3H), 1.63 (s, 1H), 1.46 (s, 3H).

¹³C {¹H} NMR (100 MHz, CDCl₃): δ 153.3, 139.3, 136.9, 132.4, 124.5, 123.6, 120.8, 119.3, 119.2, 119.1, 117.9, 114.9, 110.4, 107.1, 74.2, 40.8, 36.9, 32.1, 29.0, 23.4, 20.5, 20.4, 16.7.

HRMS (ESI) *m*/*z*: [M+H]⁺ Calcd for C₂₃H₂₆NO: 332.2014; Found 332.2012.

IR (film): v_{max} 3125, 1364, 1302, 1125, 1089, 921 cm⁻¹.

Comparison of ¹H-NMR Data:

Ali's report ²³ of	Knolker's report ¹⁰ of	This report of
isocyclomahanimbine [5]	isocyclomahanimbine [5]	isocyclomahanimbine [5]
(¹ H NMR, 500 MHz, CDCl ₃)	(¹ H NMR, 500 MHz,	(¹ H NMR, 500 MHz, CDCl ₃)
	CDCl ₃)	
7.86 (d, <i>J</i> = 7.6 Hz, 1H)	7.89 (d, <i>J</i> = 7.7 Hz, 1H)	7.91 (d, <i>J</i> = 7.7 Hz, 1H)
7.60 (s, 1H)	7.63 (s, 1H)	7.65 (d, <i>J</i> = 0.9 Hz, 1H)
7 30 (m. 2H)	7.45 (brs, 1H)	7.47 (s, 1H)
7.50 (m, 211)	7.35 (d, $J = 8.0$ Hz, 1H)	7.37 (dt, $J = 8.0, 0.9$ Hz, 1H)
7.28 - 7.20 (m. 1H)	7.27 (ddd, <i>J</i> = 8.0, 7.1, 1.0	7.30 - 7.29 (m. 1H)
7.20 7.20 (III, III)	Hz, 1H)	7.30 7.29 (iii, 111)
7.12 (m, 1H)	7.17 (brt, <i>J</i> = 7.4 Hz 1H)	7.19 – 7.15 (m, 1H)
3.99 (s, 1H)	4.27 (brs, 1H)	4.29 (s, 1H)

-	2.40 (dd, <i>J</i> = 14.5, 5.6 Hz, 1H)	2.43 (d, <i>J</i> = 8.8 Hz, 1H)
2.34 (s, 3H)	2.35 (d, <i>J</i> = 0.6 Hz, 3H)	2.37 (s, 3H)
2.31 (m, 1H)	2.14 (d, <i>J</i> = 2.0 Hz, 3H)	2.16 (d, <i>J</i> = 2.0 Hz, 3H)
-	2.05 – 2.09 (m, 1H)	2.10 – 2.07 (m, 1H)
1.99 (m, 1H)	2.01 (dt, <i>J</i> = 12.6, 3.2 Hz, 1H)	2.04 – 2.01 (m, 1H)
1.96 (s, 3H)	1.93 (dd, <i>J</i> = 12.6, 2.6 Hz, 1H)	1.95 (dd, <i>J</i> = 12.6, 2.7 Hz, 1H)
1.79 (m, 2H)	1.88 (m, 1H)	1.90 (s, 1H)
1.61 (s, 3H)	1.67 (d, $J = 0.9$ Hz, 3H)	1.69 (d, <i>J</i> = 1.3 Hz, 3H)
1.50 (m, 1H)	1.60 (td, <i>J</i> = 13.6, 5.7 Hz 1H)	1.63 (s, 1H)
1.38 (s, 3H).	1.44 (s, 3H).	1.46 (s, 3H).

Comparison of ¹³C-NMR Data:

Ali's report ²³ of isocyclomahanimbine [5]	Knolker's report ¹⁰ of isocyclomahanimbine [5]	This report of isocyclomahanimbine [5]
(¹³ C NMR, 126 MHz, CDCl ₃)	(¹³ C NMR, 125 MHz, CDCl ₃)	(¹³ C NMR, 100 MHz, CDCl ₃)
153.36	153.23	153.3
139.36	139.26	139.3
136.93	136.78	136.9
132.36	132.33	132.4
124.52	124.38	124.5

123.59	123.6
120.78	120.8
119.29	119.3
119.15	119.2
119.10	119.1
117.88	117.9
114.84	114.9
110.32	110.4
107.06	107.1
74.12	74.2
40.77	40.8
36.82	36.9
32.08	32.1
28.93	29.0
23.30	23.4
20.48	20.5
20.40	20.4
16.64	16.7
	123.59 120.78 119.29 119.15 119.10 117.88 114.84 110.32 107.06 74.12 40.77 36.82 32.08 28.93 23.30 20.48 20.40 16.64

Crystal Data and Structure Refinement of 5



Figure . Single crystal XRD structure of 5 ORTEP drawn at 50% probability level.

Solvent System for growing X-Ray Single Crystal:

Single crystals as Clear Colorless, Block and Crystal size/mm3: $0.4 \times 0.4 \times 0.4$ of **5**, were obtained by slow evaporation from methanol solvent after 11 days.

Instrument used for X-Ray Crystallography:

The X-ray data of **5** was collected at 300 K with Rigaku XtaLAB Synergy, Dual flex four-circle diffractometer with HyPix3000 detector and CuK α radiation ($\lambda = 1.54184$ Å). For **5**, the data was collected, reduced, and cell refinement was done in CrysAlis PRO (41_64.115a) software. The structure was solved by intrinsic phasing using SHELXT and implemented in the program system Olex2. All non-H atoms for **5**, were refined anisotropically against F2 for all reflections. The nonhydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atoms were included in calculated positions and treated as riding throughout the refinement.

CCDC Information for X-Ray Structure:

The cif file was deposited with the Cambridge Crystallographic Data Centre, and the following code was allocated: CCDC-2355365 for **5**.

Table 1 Crystal data and structure refinement for AB_DGJ_08SCM_auto.		
Identification code	2355365	
Empirical formula	$C_{46}H_{50}N_2O_2$	
Formula weight	662.88	
Temperature/K	100.00(13)	
Crystal system	triclinic	
Space group	P-1	
a/Å	9.8359(2)	
b/Å	13.2198(3)	
c/Å	15.2566(2)	
α/°	85.4510(10)	
β/°	82.6730(10)	
γ/°	69.423(2)	
Volume/Å ³	1840.79(7)	
Z	2	
$\rho_{calc}g/cm^3$	1.196	
μ/mm ⁻¹	0.557	
F(000)	712.0	
Crystal size/mm ³	0.4 imes 0.4 imes 0.4	
Radiation	$Cu K\alpha (\lambda = 1.54184)$	
2Θ range for data collection/°	5.844 to 136.41	
Index ranges	$-11 \le h \le 11, -15 \le k \le 15, -18 \le 1 \le 17$	

Reflections collected	35510
Independent reflections	6683 [R _{int} = 0.0463, R _{sigma} = 0.0290]
Data/restraints/parameters	6683/0/459
Goodness-of-fit on F ²	1.057
Final R indexes [I>=2σ (I)]	$R_1 = 0.0415, wR_2 = 0.1112$
Final R indexes [all data]	$R_1 = 0.0480, wR_2 = 0.1164$
Largest diff. peak/hole / e Å ⁻³	0.26/-0.19

checkCIF/PLATON report

Structure factors have been supplied for datablock(s) ab_dgj_08scm_auto

THIS REPORT IS FOR GUIDANCE ONLY. IF USED AS PART OF A REVIEW PROCEDURE FOR PUBLICATION, IT SHOULD NOT REPLACE THE EXPERTISE OF AN EXPERIENCED CRYSTALLOGRAPHIC REFEREE.

No syntax errors found. CIF dictionary Interpreting this report

Datablock: ab_dgj_08scm_auto

Bond precision:	C-C = 0.0020 A	Wavelength	=1.54184
Cell:	a=9.8359(2)	b=13.2198(3)	c=15.2566(2)
	alpha=85.451(1)	beta=82.673(1)	gamma=69.423(2)
Temperature:	100 K		-
	Calculated	Reported	
Volume	1840.79(7)	1840.79(7)
Space group	P -1	P -1	
Hall group	-P 1	-P 1	
Moiety formula	C23 H25 N O	2(C23 H25	N 0)
Sum formula	C23 H25 N O	C46 H50 N	12 02
Mr	331.44	662.88	
Dx,g cm-3	1.196	1.196	
Z	4	2	
Mu (mm-1)	0.557	0.557	
F000	712.0	712.0	
F000'	713.88		
h,k,lmax	11,15,18	11,15,18	
Nref	6731	6683	
Tmin,Tmax		0.565,1.0	000
Tmin'			
Correction meth AbsCorr = MULTI	od= # Reported T L -SCAN	imits: Tmin=0.565 Tm	max=1.000
Data completene	ss= 0.993	Theta(max) = 68.20	5
R(reflections)=	0.0415(5712)		wR2(reflections) =
0 1 050		450	0.1164(6683)
S = 1.058	Npar= 4	459	

The following ALERTS were generated. Each ALERT has the format test-name_ALERT_alert-type_alert-level. Click on the hyperlinks for more details of the test.

Alert level C

```
PLAT042_ALERT_1_C Calc. and Reported MoietyFormula Strings Differ
                                                                     Please Check
             Calc: C23 H25 N O
             Rep.: 2(C23 H25 N O)
PLAT053_ALERT_1_C Minimum Crystal Dimension Missing (or Error) ...
                                                                    Please Check
PLAT054_ALERT_1_C Medium Crystal Dimension Missing (or Error) ...
                                                                  Please Check
                                                                     Please Check
PLAT055_ALERT_1_C Maximum Crystal Dimension Missing (or Error) ...
PLAT230_ALERT_2_C Hirshfeld Test Diff for C29 --C30
                                                                      5.7 s.u.
                                                              .
PLAT250_ALERT_2_C Large U3/U1 Ratio for <U(i,j)> Tensor(Resd 2)
                                                                       2.1 Note
PLAT420_ALERT_2_C D-H Bond Without Acceptor N003 --H003 .
PLAT420_ALERT_2_C D-H Bond Without Acceptor N004 --H004 .
                                                                   Please Check
                                                                    Please Check
PLAT911_ALERT_3_C Missing FCF Refl Between Thmin & STh/L= 0.600
                                                                        21 Report
              -7-15 1, -2-14 7, 8 -6 7, 9 13 7, 11 3 8, -2-13 9,
              -7 -5 13, -3 8 13, -3 7 14, 3 11 14, 2 -7 15, 6 -2 15,
               3 10 15, 3 -5 16, 4 -4 16, 5 -2 16, 7 4 16,
                                                                     4 -2 17,
               2 -1 18,
                        0 2 18, 2 4 18,
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Alert level G
PLAT007_ALERT_5_G Number of Unrefined Donor-H Atoms .....
                                                                      2 Report
            H003 H004
PLAT045_ALERT_1_G Calculated and Reported Z Differ by a Factor ...
                                                                     2 Check
PLAT720_ALERT_4_G Number of Unusual/Non-Standard Labels .....
                                                                      6 Note
            0001 0002 N003 H003 N004 H004
PLAT912_ALERT_4_G Missing # of FCF Reflections Above STh/L= 0.600
                                                                    27 Note
PLAT969_ALERT_5_G The 'Henn et al.' R-Factor-gap value .....
                                                                 2.68 Note
            Predicted wR2: Based on SigI**2 4.35 or SHELX Weight 11.40
PLAT978_ALERT_2_G Number C-C Bonds with Positive Residual Density.
                                                                    16 Info
PLAT992_ALERT_5_G Repd & Actual _reflns_number_gt Values Differ by
                                                                     3 Check
```

```
0 ALERT level A = Most likely a serious problem - resolve or explain
0 ALERT level B = A potentially serious problem, consider carefully
9 ALERT level C = Check. Ensure it is not caused by an omission or oversight
7 ALERT level G = General information/check it is not something unexpected
5 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
5 ALERT type 2 Indicator that the structure model may be wrong or deficient
1 ALERT type 3 Indicator that the structure quality may be low
2 ALERT type 4 Improvement, methodology, query or suggestion
3 ALERT type 5 Informative message, check
```

It is advisable to attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the purpose of your study may justify the reported deviations and the more serious of these should normally be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. checkCIF was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

Publication of your CIF in IUCr journals

A basic structural check has been run on your CIF. These basic checks will be run on all CIFs submitted for publication in IUCr journals (*Acta Crystallographica, Journal of Applied Crystallography, Journal of Synchrotron Radiation*); however, if you intend to submit to *Acta Crystallographica Section C* or *E* or *IUCrData*, you should make sure that full publication checks are run on the final version of your CIF prior to submission.

Publication of your CIF in other journals

Please refer to the Notes for Authors of the relevant journal for any special instructions relating to CIF submission.

PLATON version of 06/01/2024; check.def file version of 05/01/2024

Datablock ab_dgj_08scm_auto - ellipsoid plot

