## **Electronic Supplementary Information (ESI)**

Novel polycyclic "turn on" and "turn off" pyrazoline and pyrazole fluorescent sensors selective for Fe<sup>3+</sup>/Fe<sup>2+</sup> in aqueous environments for real-world monitoring

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### **General Experimental**

Chemicals, solvents and reagents were purchased from commercial sources and used without further purification. PE refers to petroleum ether, bp 40-60 °C. Spectroscopy was performed with CHROMASOLV<sup>®</sup> gradient grade acetonitrile for HPLC,  $\geq$ 99.9%, from Sigma-Aldrich.

The metal complexes used in this study were: LiCl, NaCl, KCl, CaCl<sub>2</sub>, MgCl<sub>2</sub>, CuCl<sub>2</sub>, CuSO<sub>4</sub>, Cu(OAC)<sub>2</sub>, NiCl<sub>2</sub>, ZnCl<sub>2</sub>, CdCl<sub>2</sub>, RuCl<sub>3</sub>, CoCl<sub>2</sub>, MnCl<sub>2</sub>, PbCl<sub>2</sub>, ZnCl<sub>2</sub>, FeSO<sub>4</sub> and FeCl<sub>3</sub>.

TLCs were carried out on Merck Aluminium backed TLC plates Silica Gel 60 F254 and viewed using UV light of wavelength 254 nm. Merck Silica Gel (0.040-0.063 mm) was used for column chromatography. Compounds were loaded as an oil,  $CH_2Cl_2$  solution or dry loaded by adsorption onto silica.

NMR spectra were obtained on a Bruker Avance III (400 MHz) spectrometer and processed via TopSpin<sup>®</sup> software. The chemical shifts are recorded in parts per million (ppm) with reference to tetramethylsilane. The coupling constants J are quoted to the nearest 0.5 Hz and are not corrected.

High resolution Mass spectroscopy was performed on Bruker Quadrupole Time-of-Flight (qToF) mass spectrometer.

UV/Vis spectroscopy was performed on an Agilent Cary5000 in quartz cuvettes with a 1 cm pathlength using HPLC grade MeCN, 250-500 nm range with 0.2 sec dwell time. Detector switchover occurred at 350 nm.

FTIR spectroscopy was performed on a Bruker VERTEX 70 spectrometer.

Fluorescence spectroscopy was performed on an Edinburgh Instruments FLS1000 with a xenon excitation source, 2 nm bandwidths for both excitation and emission monochromator, scan speed of 1 nm and dwell time of 0.2 sec. Fluorescence quartz cuvettes with a 1 cm pathlength were used throughout with HPLC grade MeCN. Quantum yields were determined using the absolute method and use of a Edinburgh instruments integrating sphere, <u>https://www.edinst.com/wp-content/uploads/2016/02/FLS980-Series-Reference-Guide-Integrating-Sphere.pdf</u>

A 100 Watt 365 nm Analytikjena High intensity UV lamp was used to image the sensors in cuvettes with 5.0 equivalent indicated metal, sensor concentration was 20  $\mu$ M, solvent was MeCN.

All figures were plotted using SigmaPlot<sup>®</sup> 14.5 software.

#### General Synthesis (S1)

Synthesis of chalcones C1-C2



Using a method adapted from a previous synthesis (*RSC Adv.*, 2017, **7**, 44272), 5.0 mmol 2acetylpyridine was added to a stirred solution of 5.0 mmol aldehyde (1-naphthaldehyde or 9anthraldehyde) in MeOH followed by the addition of 5.0 mmol NaOH and stirring continued. After 24 hours the solvent was removed under reduced pressure and the residue was filtered, washed with copious amounts of cold  $H_2O$  and collected and dried to afford the desired chalcone without further purification.

#### Synthesis of 1-4



Using a method adapted from a previous synthesis (*RSC Adv.,* 2024, **14**, 3519), the required hydrazine ( $H_2NNHMe$  or  $H_2NNHPh$ ) 2.0 mmol was added to a stirred solution of the required chalcone (**C1** or **C2**) 1.0 mmol in MeOH at 60°C. After 24 hours the solvent was removed under reduced pressure to afford an oil which was extracted into 100 mL ethyl acetate and washed with 3 x 50 mL water. The ethyl acetate was removed under reduced pressure to afford an oil which was further purified by column chromatography using an 8:2 PE: ethyl acetate mixture to give the desired pyrazoline.

Synthesis of 5-7



Using a method from the literature (*New J. Chem.,* 2024, **48**, 13900), the required hydrazine ( $H_2NNHMe$  or  $H_2NNHPh$ ) 2.0 mmol was added to a stirred solution of required chalcone (**C1** or **C2**) 1.0 mmol in 50 mL MeOH at 60°C followed by the addition of CuCl<sub>2</sub> (1.0 mmol). After 24 hrs the solution was removed under reduced pressure, the residue resuspended in 100 mL of a saturated EDTA solution and extracted with ethyl acetate (3 x 50 mL). The ethyl acetate layers were combined, and solvent removed under reduced pressure. The residue was then purified by column chromatography using PE: ethyl acetate (80:20) to afford the required pyrazole.

Synthesis of (E)-3-(naphthalen-1-yl)-1-(pyridin-2-yl)prop-2-en-1-one (C1)



Yield 1.21g (93%);

Vmax (Solid)/cm<sup>-1</sup>1854, 1533, 1427and 1106;

<sup>1</sup>**H NMR**  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 7.52-7.64 (5 H, m, CH), 7.91-7.97 (3 H, m, CH), 8.09 (1 H, d, *J* = 7.2 Hz, CH), 8.27 (1 H, d, *J* = 8 Hz, CH), 8.37 (1 H, d, *J* = 8.4 Hz, CH), 8.43 (1 H, d, *J* = 16.0 Hz, CH), 8.79 (1 H, m, CH) and 8.82 (1 H, d, *J* = 16.0 Hz, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 121.8, 123.0, 123.3, 123.5, 125.1, 125.5, 125.6, 126.2, 126.9, 131.0, 132.0, 132.4, 133.8, 137.1, 141.4, 148.0, 154.3 and 189.4;

**HRMS m/z (qToF)** Found 260.1104 (M+H<sup>+</sup>). C<sub>18</sub>H<sub>14</sub>NO requires 260.1075.

Synthesis of (E)-3-(anthracen-9-yl)-1-(pyridin-2-yl)prop-2-en-1-one (C2)



Yield 1.15g (75%);

Vmax (Solid)/cm<sup>-1</sup>1717, 1531, 1426, 1381 and 1163;

<sup>1</sup>**H NMR**  $\delta_{\rm H}$  (400 MHz; CDCl<sub>3</sub>) 7.52-7.56 (5 H, m, CH), 7.94-7.95 (1 H, m, CH), 8.45-8.07 (2 H, m, CH), 8.29-8.33 (2 H, m, CH), 8.39-8.41 (2 H, m, CH), 8.50 (1 H, s, CH), 8.72-8.74 (1 H, m, CH) and 8.96 (1 H, d, *J* = 16.0 Hz, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 122.6, 123.1, 123.6, 125.1, 125.4, 125.5, 125.6, 126.7, 127.0, 127.3, 128.4, 128.5, 128.8, 129.1, 129.3, 129.8, 130.2, 130.4, 134.1, 136.8, 154.2 and 189.4;

**HRMS m/z (qToF)** Found 310.1255 (M+H<sup>+</sup>). C<sub>22</sub>H<sub>16</sub>NO requires 310.1232.

Above data consistent with spectra from RSC Adv., 2017, 7, 44272.

Synthesis of 2-(1-methyl-5-(naphthalen-1-yl)-4,5-dihydro-1H-pyrazol-3-yl)pyridine (1)



Yield 0.065g oil (23%);

**Vmax** (film)/cm<sup>-1</sup>1855, 1548, 1427, 1381 and 1166;

<sup>1</sup>**H NMR**  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 3.05-3.13 (4 H, m, CH<sub>3</sub>), 4.06 (1 H, m, CH), 4.95 (1 H, m, CH), 7.21 (1 H, m, CH), 7.53- 7.55 (3 H, m, CH), 7.68-7.07 (1 H, m, CH), 7.26-7.38 (2 H, m, CH), 7.85-7.93 (1 H, m, CH), 7.94-7.97 (1 H, m, CH), 8.07-8.09 (1 H, m, CH) and 8.57-8.59 (1 H, m, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 41.8, 120.5, 122.7, 123.4, 124.0, 125.4, 125.7, 126.1, 126.4, 128.1, 128.6, 128.9, 131.4, 134.0, 136.1, 136.2, 149.2, 150.2 and 152.1;

HRMS m/z (qToF) Found 288.1476 (M+H<sup>+</sup>). C<sub>19</sub>H<sub>18</sub>N3 requires 288.1501.

Synthesis of 2-(5-(naphthalen-1-yl)-1-phenyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (2)



Yield 0.093g oil (26%);

Vmax (film)/cm<sup>-1</sup>1704, 1565, 1443, 1237 and 1198;

<sup>1</sup>H NMR  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 3.34-3.40 (1 H, m, CH), 4.21-4.26 (1 H, m, CH), 6.05-6.84 (1 H, m, CH), 6.82-6.85 (1 H, m, CH), 7.10-7.18 (2 H, m, CH), 7.18-7.23 (3 H, m, CH), 7.35-7.38 (2 H, m, CH), 7.58-7.60 (3 H, m, CH), 7.07-7.74 (1 H, m, CH), 7.78-7.81 (1 H, m, CH), 8.12-8.14 (1 H, m, CH), 8.19-8.21 (1 H, m, CH) and 8.51-8.52 (1 H, m, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 42.2, 113.5, 119.6, 120.6, 122.7, 123.1, 125.8, 126.4, 128.0, 128.9, 129.0, 129.2, 129.9, 134.4, 136.0, 136.7, 144.3, 147.1, 148.6, 149.1, 152.1 and 152.1;

HRMS m/z (qToF) Found 338.1620 (M+H<sup>+</sup>). C<sub>23</sub>H<sub>20</sub>N<sub>3</sub> requires 338.1657.

Synthesis of 2-(5-(anthracen-9-yl)-1-methyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (3)



Yield 0.024g oil (7%);

Vmax (film)/cm<sup>-1</sup>1856, 1550, 1444 and 1192;

<sup>1</sup>H NMR  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 2.82 (3 H, s, CH<sub>3</sub>), 3.57- 3.65 (1 H, m, CH), 3.78-3.81 (1 H, m, CH), 5.66-5.72 (1 H, m, CH), 7.15-7.16 (1 H, m, CH), 7.33-7.46 (5 H, m, CH), 7.63-7.66 (1 H, m, CH), 7.93-7.99 (2 H, m, CH), 8.27-8.29 (1 H, m, CH), 8.40 (1 H, s, CH), 8.51- 8.52 (1 H, m, CH) and 8.73-8.74 (1 H, m, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 41.5, 68.0, 120.6, 122.5, 122.7, 124.8, 125.0, 125.1, 125.6, 126.1, 126.4, 126.7, 127.3, 128.5, 129.4, 129.5, 129.6, 129.8, 131.0, 131.2, 134.1, 136.1, 149.3, 149.6 and 152.3;

**HRMS m/z (qToF)** Found 350.1627 (M+H<sup>+</sup>). C<sub>24</sub>H<sub>2</sub>N<sub>3</sub> requires 350.1657.

Synthesis of 2-(5-(anthracen-9-yl)-1-phenyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (4)



Yield 0.061g oil (15%);

Vmax (film)/cm<sup>-1</sup>1854, 1546, 1427 and 1237;

<sup>1</sup>**H NMR**  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 3.66-3.74 (1 H, m, CH), 4.22-4.30 (1 H, m, CH), 6.70 (1 H, m, CH), 7.00 (3 H, m, CH), 7.36-7.39 (1 H, m, CH), 7.41-7.57 (2 H, m, CH), 7.60-7.66 (7 H, m, CH), 7.80 (1 H, m, CH), 8.01 (1 H, m, CH), 8.13 (2 H, m, CH), 8.31 (1 H, m, CH), 8.53 (2 H, m, CH) and 8.58 (1 H, m, CH);

<sup>13</sup>**C NMR**  $δ_c$  (400 MHz; CDCl<sub>3</sub>) 42.1, 61.3, 113.8, 119.9, 120.9, 122.3, 122.8, 124.2, 124.9, 125.1, 126.3, 127.0, 128.5, 128.8, 129.2, 129.4, 129.5, 129.0, 131.7, 132.2, 136.0, 145.3, 149.3 and 152.2;

HRMS m/z (qToF) Found 400.1774 (M+H<sup>+</sup>). C<sub>28</sub>H<sub>22</sub>N<sub>3</sub> requires 400.1814.

Synthesis of 2-(1-methyl-5-(naphthalen-1-yl)-1H-pyrazol-3-yl)pyridine (5)



Yield 0.014g oil (5%);

Vmax (film)/cm<sup>-1</sup>1711, 1547 and 1190;

<sup>1</sup>**H NMR**  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 4.03 (3 H, s, CH<sub>3</sub>), 6.28-6.30 (1 H, m, CH), 6.49-6.53 (1 H, m, CH), 6.70 (1 H, s, CH), 6.26-7.63 (6 H, m, CH), 7.92-7.95 (3 H, m, CH), 8.05-8.06 (1 H, m, CH), 8.21-8.22 (1 H, m, CH) and 8.92-8.94 (1 H, m, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 39.5, 63.7, 107.4, 122.5, 122.9, 123.7, 125.4, 125.8, 126.2, 126.3, 127.0, 128.3, 131.4, 134.0, 136.8, 142.1, 149.3, 149.8 and 150.0;

HRMS m/z (qToF) Found 286.1301 (M+H<sup>+</sup>). C<sub>19</sub>H<sub>16</sub>N<sub>3</sub> requires 286.1344.

Synthesis of 2-(5-(anthracen-9-yl)-1-methyl-1H-pyrazol-3-yl)pyridine (6)



Yield 0.024g oil (7%);

Vmax (film)/cm<sup>-1</sup>1714, 1559, 1443 and 1243;

<sup>1</sup>H NMR  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 3.95 (3 H, s, CH<sub>3</sub>), 6.09-6.11 (1 H, m, CH), 6.41-6.44 (1 H, m, CH), 6.69 (1 H, s, CH), 7.29-7.31 (2 H, m, CH), 7.38-7.49 (6 H, m, CH), 7.98-8.00 (2 H, m, CH) and 8.50 (1H, s, CH);

<sup>13</sup>**C NMR**  $δ_c$  (400 MHz; CDCl<sub>3</sub>); 59.4, 103.6, 110.5, 113.6, 115.7, 116.8, 119.9, 121.8, 125.4, 125.9, 126.3, 128.2, 128.7, 131.6, 131.9, 134.2 and 138.6;

HRMS m/z (qToF) Found 336.1490 (M+H<sup>+</sup>).  $C_{23}H_{18}N_3$  requires 336.1501.

Synthesis of 2-(5-(anthracen-9-yl)-1-phenyl-1H-pyrazol-3-yl)pyridine (7)



Yield 0.094g oil (24%);

Vmax (film)/cm<sup>-1</sup>1702. 1542, 1445 and 1167;

<sup>1</sup>H NMR  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 6.99-7.01 (3 H, m, CH), 7.19-7.21 (2 H, m, CH), 7.21-7.29 (1 H, m, CH), 7.31-7.47 (5 H, m, CH), 7.81-7.85 (3 H, m, CH), 8.02-8.04 (2 H, m, CH), 8.26-8.29 (1 H, m, CH), 8.56 (1 H, s, CH) and 8.73-8.74 (1 H, m, CH);

 $^{13}\textbf{C}$  NMR  $\delta_c$  (400 MHz; CDCl<sub>3</sub>) 110.0, 120.5, 122.8, 123.4, 124.6, 125.4, 125.8, 126.6, 127.1, 128.5, 128.6, 128.9, 131.1, 131.2, 136.7, 140.0, 140.9, 149.6, 152.2 and 152.4;

HRMS m/z (qToF) Found 398.1610 (M+H<sup>+</sup>). C<sub>28</sub>H<sub>20</sub>N<sub>3</sub> requires 398.1657.

## NMR Spectra (S2)

#### E)-3-(naphthalen-1-yl)-1-(pyridin-2-yl)prop-2-en-1-one (C1)





#### (E)-3-(anthracen-9-yl)-1-(pyridin-2-yl)prop-2-en-1-one (C2)



#### 2-(1-methyl-5-(naphthalen-1-yl)-4,5-dihydro-1H-pyrazol-3-yl)pyridine (1)



#### 2-(5-(naphthalen-1-yl)-1-phenyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (2)



#### 2-(5-(anthracen-9-yl)-1-methyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (3)



#### 2-(5-(anthracen-9-yl)-1-phenyl-4,5-dihydro-1H-pyrazol-3-yl)pyridine (4)



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ppm

 Final
 Crocessing parameters

 SI
 32768

 SF
 100.6127685 MHz

 MDW
 2200

 SLB
 0

 LB
 1.00 Hz

 GB
 0

 MPC
 1.40

Synthesis of 2-(1-methyl-5-(naphthalen-1-yl)-1H-pyrazol-3-yl)pyridine (5)



#### Synthesis of 2-(5-(anthracen-9-yl)-1-methyl-1H-pyrazol-3-yl)pyridine (6)



#### Synthesis of 2-(5-(anthracen-9-yl)-1-phenyl-1H-pyrazol-3-yl)pyridine (7)

## <sup>1</sup>H NMR Experiments (S3)



Partial <sup>1</sup>H NMR spectra for **1** only A (20  $\mu$ M, CDCl<sub>3</sub>) and B with 2.0 equilvalents Zn<sup>2+</sup>.



Partial <sup>1</sup>H NMR spectra for **2** only A (20  $\mu$ M, CDCl<sub>3</sub>) and B with 2.0 equilvalents Zn<sup>2+</sup>.



Partial <sup>1</sup>H NMR spectra for **3** only A (20  $\mu$ M, CDCl<sub>3</sub>) and B with 2.0 equilvalents Zn<sup>2+</sup>.



Partial <sup>1</sup>H NMR spectra for **4** only A (20  $\mu$ M, CDCl<sub>3</sub>) and B with 2.0 equilvalents Zn<sup>2+</sup>.



Partial <sup>1</sup>H NMR spectra for 7 only A (15  $\mu$ M, CDCl<sub>3</sub>) and B with 2.0 equilvalents Zn<sup>2</sup>.

## UV/Vis Spectroscopy (S4)



Wavelength (nm) UV/Vis study of 1 only (20  $\mu M$ , MeCN) and with 5 equivalents Zn^{2+}, Cd^{2+} and +Hg^{2+}. Arrow indicates change in bands.



UV/Vis study of **2** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.



UV/Vis study of **3** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.



UV/Vis study of **4** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.



UV/Vis study of **5** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.



UV/Vis study of **6** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.



UV/Vis study of **7** only (20  $\mu$ M, MeCN) and with 5 equivalents Zn<sup>2+</sup>, Cd<sup>2+</sup> and +Hg<sup>2+</sup>. Arrow indicates change in bands.

## Aqueous study of pyrazoline **1** in 7:3 MeCN:H<sub>2</sub>O solution on addition of Cd<sup>2+</sup> (0-10 equivalent), $\lambda_{ex}$ 280 nm (S5)



Aqueous study of sensor **2** (20  $\mu$ M, MeCN) with 5.0 equivalents indicated metal in 7:3 MeCN:H<sub>2</sub>O solution at  $\lambda_{ex}$  280 nm, cps is counts per second



Aqueous study of sensor **7** (20  $\mu$ M, MeCN) with 5.0 equivalents indicated metal in 7:3 MeCN:H<sub>2</sub>O solution at  $\lambda_{ex}$  250 nm, cps is counts per second



Real-world example of pyrazoline **2** in tap water panel A and mineral water panel B, solution was 7:3 MeCN:H<sub>2</sub>O, **2** concentration 20  $\mu$ M,  $\lambda_{ex}$  280 nm (S6)



Real-world example of pyrazoline **2** in pond water panel A and river water panel B, solution was 7:3 MeCN:H<sub>2</sub>O, **2** concentration 20  $\mu$ M,  $\lambda_{ex}$  280 nm (S7)



## Limit of Detection (LoD) Studies (S8)

The method reported by Lee *et al* was used to calculate limit of detection (LoD) for **2** ( $\lambda_{ex}$  280 nm) and **7** ( $\lambda_{ex}$  250 nm) in 7:3 MeCN:H<sub>2</sub>O with the average from two replicates used.

$$LoD = 3\sigma_{bi}/m$$

 $\sigma_{bi}$  = standard deviation of sensor only (n=10)

m = gradient of the slope, note the inverse sign was used as these are "turn off" sensors not "turn on"



B. P. Joshi, J. Park, W. I. Lee and K.-H. Lee, Talanta, 2009, 78, 903.

Analyte	Structure	Detection	Mode	LoD	Solvent	Reference
Fe <sup>3+</sup>	Pyrazoline	Fluorescence	"Turn off"	0.401 µM	THF:H <sub>2</sub> O = 9:1	J. Fluoresc., 2024, <b>34</b> , 159
Fe <sup>3+</sup>	Pyrazole	Fluorescence	"Turn off"	1.86 µM	EtOH:H <sub>2</sub> O= 7:3	Org. Biomol. Chem., 2023, <b>21</b> , 4482
Fe <sup>3+</sup>	Pyrazole	Fluorescence	"Turn off"	21 nM	DMSO:H <sub>2</sub> O= 9:1	J. Photochem. Photobiol., A, 2023, <b>437</b> , 114470
Fe <sup>3+</sup>	Pyrazole	Fluorescence	"Turn off"	45 nM	DMSO:H <sub>2</sub> O= 9:1	New J. Chem., 2023, <b>47</b> , 751
Fe <sup>3+</sup>	Pyrazole	Fluorescence	"Turn off"	21 nM	DMSO:H <sub>2</sub> O= 9:1	J. Photochem. Photobiol., A, 2023, <b>437</b> , 114470
Fe <sup>3+</sup>	Pyrazoline	Fluorescence	"Turn off"	0.12 μM	MeOH:H <sub>2</sub> O = 1:9	J. Fluoresc., 2022, <b>32</b> , 2319
Fe <sup>3+</sup>	Pyrazole	Fluorescence	"Turn on"	1.73 μM	MeCN:H <sub>2</sub> O = 7:3	Spectrochim. Acta, Part A, 2020, <b>230</b> , 117993
Fe <sup>3+</sup>	Quinazoline	Fluorescence	"Turn on"	3.5 µM	MeCN:H <sub>2</sub> O= 2:8	Analyst, 2012, <b>137</b> , 3335
Fe <sup>3+</sup>	Rhodamine– naphthalic anhydride	Fluorescence	"Turn on"	2.90 µM	MeOH:H <sub>2</sub> O= 6:4	<i>Dalton Trans.</i> , 2015, <b>44</b> , 11805

Selection of recently reported Fe<sup>3+</sup> LoD in a range of solvents

# Quantum Yield calculated using the absolute method and an integrating sphere (S9)

Sensor	Solvent		Quantum Yield φ
S2	MeCN	<b>S2</b> only	0.74
S2	MeCN	<b>S2</b> + Fe <sup>3+</sup>	<0.01
S2	MeCN:H <sub>2</sub> O 7:3	<b>S2</b> only	0.83
S2	MeCN:H <sub>2</sub> O 7:3	<b>S2</b> + Fe <sup>3+</sup>	0.07
S7	MeCN	<b>S7</b> only	0.33
S7	MeCN	<b>S7</b> + Fe <sup>3+</sup>	<0.01
S7	MeCN:H <sub>2</sub> O 7:3	S7 only	0.43
S7	MeCN:H <sub>2</sub> O 7:3	<b>S7</b> + Fe <sup>3+</sup>	0.18

## Recovery study for 2 and 7 (S10)

Sensor	Solvent	cps	% Fe <sup>3+</sup> Recovery	
Sensor <b>2</b> + 50 µM Fe <sup>3+</sup>	Mineral Water	2.309 x10 <sup>6</sup>	64%	- Sensor <b>2</b> with Fe <sup>3+</sup>
Sensor <b>2</b> + 100 μM Fe <sup>3+</sup>	Mineral Water	1.949 x10 <sup>6</sup>	40%	calibration curve
Sensor <b>2</b> + 50 μM Fe <sup>3+</sup>	Tap Water	2.314 x10 <sup>6</sup>	64%	s at
Sensor <b>2</b> + 100 µM Fe <sup>3+</sup>	Tap Water	2.080 ×10 <sup>6</sup>	38%	g 🔰 🔪
Sensor <b>2</b> + 50 µM Fe <sup>3+</sup>	Pond Water	1.870 ×10 <sup>6</sup>	80%	20 40 60 80 100
Sensor <b>2</b> + 100 µM Fe <sup>3+</sup>	Pond Water	$1.640 \times 10^{6}$	48%	Fe <sup>3+</sup> µM
Sensor <b>2</b> + 50 µM Fe <sup>3+</sup>	<b>River Water</b>	1.760 x 10 <sup>6</sup>	68%	
Sensor <b>2</b> + 100 µM Fe <sup>3+</sup>	River Water	$1.170 \times 10^{6}$	48%	

Sensor	Solvent	cps	% Fe <sup>3+</sup> Recovery	-
Sensor <b>7</b> + 50 µM Fe <sup>3+</sup>	Mineral Water	9.435 x10 <sup>6</sup>	92%	E Sensor 7 with Fe <sup>3</sup>
Sensor <b>7</b> + 100 µM Fe <sup>3+</sup>	Mineral Water	7.859 x10 <sup>6</sup>	66%	te
Sensor <b>7</b> + 50 μM Fe <sup>3+</sup>	Tap Water	9.561 x10 <sup>6</sup>	91%	cbs
Sensor <b>7</b> + 100 µM Fe <sup>3+</sup>	Tap Water	7.956 x10 <sup>6</sup>	65%	20 40 60 80 100
Sensor <b>7</b> + 50 µM Fe <sup>3+</sup>	Pond Water	6.920 x 10 <sup>6</sup>	125%	Fe <sup>3+</sup> µM
Sensor <b>7</b> + 100 µM Fe <sup>3+</sup>	Pond Water	5.750 x10 <sup>6</sup>	90%	
Sensor <b>7</b> + 50 μM Fe <sup>3+</sup>	River Water	5.980 x 10 <sup>6</sup>	145%	
Sensor <b>2</b> + 100 µM Fe <sup>3+</sup>	<b>River Water</b>	$5.360 \times 10^{6}$	97%	

## Job Plot for 7 (S11)



Job plot for **7** with Fe<sup>3+</sup>, [Fe<sup>3+</sup>] + [**7**] = 100  $\mu$ M, MeCN