Electronic Supporting Information

# Recyclable and air-stable colloidal manganese nanoparticles catalyzed hydrosilylation of alkene with tertiary silane

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### 1. General information

Gas chromatography (GC) was performed using a GC-2025 system (Shimadzu Co.) equipped with a flame ionization detector and a 0.22 mm  $\times$  25 m BP-5 capillary column. <sup>1</sup>H and <sup>13</sup>C nuclear magnetic resonance (NMR) spectra were acquired at 400 and 100 MHz, respectively, in CDCl<sub>3</sub> with tetramethylsilane as an internal standard (JNM-ECZ-400S, Japan Electron Optics Laboratory Co.). <sup>1</sup>H NMR chemical shifts were referenced to tetramethylsilane (0 ppm) or residual chloroform (7.26 ppm) as an internal standard. <sup>13</sup>C NMR chemical shifts were referenced to residual chloroform peaks (77 ppm). All substrates were commercially available and used without purification. Compounds 3b,<sup>1</sup> 3d,<sup>2</sup> 3g,<sup>3</sup> 3l,<sup>4</sup> 3m,<sup>5</sup> 3o,<sup>6</sup> 3q,<sup>7</sup> and 3r<sup>8</sup> were reported previously. The products were characterized by <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopies, and GCmass spectrometry (GC-MS, electron ionization). Compounds 3a, 3c, 3e, 3f, 3h, 3i, 3j, 3k, 3n, and 3p were also examined by Fourier transform infrared spectroscopy (FTIR, Shimadzu IRAffinity-1, Shimadzu Co.) and high-resolution MS (micrOTOF with ESI source, Bruker Inc.). GC-MS spectra were recorded with a GCMS-QP2010 SE instrument (Shimadzu Co.). The product yields were estimated from GC peak areas using the internal standard technique (nnonane as the internal standard). X-ray photoelectron spectroscopy (XPS) data were acquired using a PHI5000 VersaProbe instrument (ULVAC-PHI, Inc.) with an Al Ka X-ray source. Annular dark-field scanning transmission electron microscopy (ADF-STEM) images were obtained using a JEM-ARM200F instrument (Japan Electron Optics Laboratory, Co.) at an accelerating voltage of 200 kV. The measurement range was 300-800 nm. Photoluminescence (PL) spectra were recorded with a spectrofluorometer (RF-6000, Shimadzu Co.), with an excitation wavelength of 350 nm. Fluorescence spectra were recorded with a data interval of 0.5 nm, covering a fluorescence wavelength range of 360–600 nm, at a scanning rate of 6000 nm/min. Ultravioletvisible (UV-Vis) spectra were obtained using a UV-Vis spectrophotometer (UV-1900i, Shimadzu Co.). FTIR spectra were obtained using the liquid-film method on NaCl plates with an IRAffinity-1 FT-IR spectrometer (Shimadzu Co.) X-ray diffraction patterns were recorded using a MiniFlex600-C instrument (Rigaku). Mn K-edge X-ray absorption spectroscopy (XAS) was conducted in transmittance and fluorescence modes at the BL14B2 beamline (operated at 8 GeV) of SPring-8 (Hyogo, Japan) with a Si(111) double-crystal monochromator (proposal 2021A1615, 2022B1952).9 Data analysis was performed using Athena software, included in the Demeter package.<sup>10</sup> The Fourier transformation of the  $k^3$ -weighted extended X-ray absorption fine structure (EXAFS) was performed over the k range 3-12 Å<sup>-1</sup>.

#### 2. Experimental procedures

Typical procedure for Mn NPs-catalyzed hydrosilylation of 1a with 2a (Table 1, entry 4) Mn NPs (50  $\mu$ L, 0.01 mol%) were added to a Schlenk tube and the DMF was removed under vacuum. Then 1-dodecene (**1a**: 0.5 mmol), methyldiphenylsilane (**2a**: 2.5 mmol) and diglyme (1 mL) as the solvent were added and the reaction was carried out at 130 °C under Ar conditions for 24 h. The conversions and product yields were estimated from GC peak areas based on an internal standard.

The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1) and subsequently purified by distillation using a Kugelrohr apparatus (140 °C, 10 hPa, 1 h). The desired product was obtained as a colorless oil in 83% yield (152.1 mg).

Preparation of 0.1 M MnCl<sub>2</sub> solution



To a 20 mL vial bottle, MnCl<sub>2</sub>·4H<sub>2</sub>O (198 mg, 1 mmol) was dissolved in HCl aq (1 M, 10 mL). The solution was left under Ar in the dark overnight.

Preparation of 1.0 mM Mn NPs solution



All the preparative operations were carried out under open air. To a 300-mL three-necked round bottom flask, *N*,*N*-dimethylformamide (DMF) (50 mL) preheated for 10 min was added, and the mixture was stirred at 1500 rpm and 140 °C. Then, 0.1 M MnCl<sub>2</sub> solution (500  $\mu$ L) was added using micropipette, and the mixture was refluxed at 140 °C for 24 h. After reaction, the resulting solution was cooled to room temperature. The resulting solution was used as a 1.0 mM Mn NPs solution for catalytic reactions.

#### Preparation of samples for scanning tunnelling electron microscopy

For the before-reaction samples, DMF was removed from the Mn NP solution (1 mM, 1 mL) under vacuum, and the nanoparticles were dispersed in ethanol (2 mL). For the after-reaction samples, diglyme was removed by evaporation at 80°C and 30 hPa after the reaction. DMF (2

mL) was then added, and the reactants and products were extracted from the DMF layer using hexane (8 mL, five times). The hexane extracts were collected, and DMF was subsequently replaced with ethanol.

### Preparation of samples for FT-IR, X-ray photoelectron spectroscopy, and X-ray diffraction

The DMF in the Mn NPs suspension was removed by evaporator at 80 °C and 30 hPa. The Mn NPs samples were prepared by placing the Mn NPs on a 5 mm  $\times$  5 mm Ag plate. For XRD measurements, the dried and solidified Mn NPs were sealed in an airtight cell within an Ar-filled bag.

#### **Preparation of samples for XAS**

DMF was removed from the Mn NPs with an evaporator, and the final liquid samples were prepared by adding DMF to obtain the required concentration of 40 mM. The solution (40 mM based on Mn content) was transferred to a polytetrafluoroethylene (PTFE) cell and the cell was filled with the reaction solution. Unless otherwise noted, XAS experiments were performed with the PTFE cell by transmitting X-rays through holes in the PTFE cell sealed with Kapton tapes (light path = 3 mm).

### Preparation of samples for photoluminescence and UV-Vis spectroscopies

Samples (2 mL) of Mn NPs were prepared in DMF.

## 3. Results of Photoluminescence and UV-Vis spectroscopic analyses



**Fig. S1.** Photoluminescence spectra of Mn NPs under irradiation with 350 nm excitation light (red line: 1 mM, blue line: 0.1 mM). The decrease in fluorescence intensity at 1 mM is caused by concentration quenching.



Fig. S2. UV-Vis absorption spectrum for Mn NPs (0.02 mM).

## 4. Results of XPS analysis



Fig. S3. Wide-scan XPS spectrum of Mn NPs before reaction.



Fig. S4. XPS profile of the Mn  $2p_{3/2}$  region for Mn NPs before reaction.

Sample	Binding Energy (eV)	FWHM	Area (%)
	640.4	1.4	28.8
Mn NPs	641.4	1.4	20.1
Before reaction	642.2	1.4	14.4
$Mn 2n_{2}$	643.2	1.4	10.1
win 2p3/2	644.9	1.8	5.3
	646.5	3.0	21.3

Table S1. Mn  $2p_{3/2}$  spectral fitting parameters



**Fig. S5.** Wide-scan XPS spectrum of Mn NPs after reaction. The Si compounds on the surface of the Mn NPs sample could not be completely removed, which indicates unanalysable spectra in the Mn 2p peak area.

### 5. Results of XAS analysis

**Table S2.** Curve-fitting analysis for Mn K-edge extended X-ray absorption fine structures(EXAFS) analysis of Mn NPs before and after reaction.

Sample	Shell	CN <sup>a</sup>	$R$ (Å) $^{b}$	$\sigma^2 (\text{\AA}^2)^c$	$\mathrm{R_{f}}^{~d}$
Mn NPs before reaction	Mn-O	4.1	2.26	0.00111	0.016
Mn NPs after reaction	Mn-O	4.0	2.25	0.00070	0.022

<sup>a</sup>Coordination number. <sup>b</sup>Atomic distance. <sup>c</sup>Debye–Waller factor. <sup>d</sup>Residual factor.



Fig. S6. Mn K-edge XAS of Mn NPs\_before reaction (MnNPs\_br) and Mn NPs\_after reaction (MnNPs\_ar). (E<sub>0</sub>: MnNPs\_br=6545.2 eV, MnNPs\_ar=6545.2 eV).



Fig. S7. Mn K-edge XANES of Mn NPs, MnO and MnCl<sub>2</sub>

### 6. Recycling experiment procedure

Mn NPs (500  $\mu$ L, 0.1 mol%) were added to a Schlenk tube and the DMF removed under vacuum. Then, 1-dodecene (**1a**: 0.5 mmol), methyldiphenylsilane (**2a**: 2.5 mmol), and diglyme (1 mL) as the solvent were added and the reaction was carried out at 130 °C under Ar for 24 h. The conversions and product yields were estimated using GC from the peak areas based on an internal standard. After performing the reaction, diglyme was removed by evaporation at 80 °C under 30 hPa. Next, hexane (8 mL) and DMF (2 mL) were added, and the products were recovered from the reaction mixture with hexane (8 mL, five times; upper layer, hexane; lower layer, DMF). The Mn NPs remained well dispersed in the DMF layer, which could be observed by photoluminescence under UV (352 nm) irradiation. The DMF layer was then evaporated (80 °C, 30 hPa) and only the catalyst was recovered. Recovered Mn NPs were added to a Schlenk tube and 1-dodecene (**1a**: 0.5 mmol), methyldiphenylsilane (**2a**: 2.5 mmol), and diglyme (1 mL) as solvent were added, and the next reaction was carried out at 130 °C under Ar for 24 h.



Scheme S1. Catalyst recycling procedure.

## 7. Other information



**Fig. S8.** Energy-dispersive X-ray spectroscopic analysis of Mn peaks (left: before reaction; right: after reaction).



Fig. S9. FT-IR spectra of Mn NPs\_ar (blue), Mn NPs\_br (red), and DMF (black).



Fig. S10. XRD pattern for the Mn NPs before reaction.





**Fig. S11.** The HRMS results for radical trapping with methyldiphenylsilane. HRMS (ESI) m/z Calcd for C<sub>22</sub>H<sub>31</sub>NOSi [M+H]<sup>+</sup> 354.2248, found 354.2248.



Chemical Formula: C<sub>22</sub>H<sub>31</sub>NOSi *m*/z: 353 (100.0%), 354 (29.6%)



**Fig. S12.** The GC-MS results for radical trapping with methyldiphenylsilane. GC-MS (EI) m/z (relative intensity) 353(12) [M]<sup>+</sup>, 338(100), 137(55), 260(53).

Catalyst	Alkene	Silane	Temp. (°C)	Additive	Yield (%)	TON	Ref.
Mn NPs (0.005 mol %)	C <sub>10</sub> H <sub>21</sub>	HSiPh <sub>2</sub> Me	130	None	79	15800	This work
Cat 1 (0.5 mol %)	C <sub>6</sub> H <sub>13</sub>	HSi(OEt) <sub>3</sub>	25	<i>t</i> -BuONa (1.5 mol %)	>95	190	11
Cat 2 (0.5 mol %) [Mn] (1 mol %)	Myrcene	H₃SiPh	130	None	96	96	12
Cat 3 (5 mol %)		HSiMe₂Bn	60	None	95	19	13
Cat 4 (3 mol %) [Mn] (6 mol %)		HSiEt <sub>3</sub>	120	JackiePhos (6 mol %)	83	13.8	14
Cat 4 (1 mol %) [Mn] (2 mol %)	C <sub>6</sub> H <sub>13</sub>	HSiMe(OTMS) <sub>2</sub>	RT (UV)	None	85	42.5	15
Cat 3 (2 mol %)	Ph	HSiMe(OTMS) <sub>2</sub>	70	None	84	42	16
Cat 4 (2.5 mol %) [Mn] (5 mol %)	C <sub>6</sub> H <sub>13</sub>	HSi(OTMS) <sub>3</sub>	RT (White	HFIP (5 mol %)	94	18.8	17

 Table S3. Manganese catalysts for alkene hydrosilylation.



Ar

OC\_\_\_\_CO OC\_\_\_Mn\_\_CO | Br





Cat 2

Cat 3

Cat 4



Fig. S13. Unsuccessful substrates.



Fig. S14. Time course monitoring of hydrosilylation of 1-dodecene with methyldiphenylsilane.



Fig. S15. A plausible reaction mechanism of Mn NPs-catalyzed alkene hydrosilylation.

## 8. Screening information

Table S4. Screening of silane ration

		<sup>cat.</sup> Mn NPs (0.01 mol %)	)	SiPh <sub>2</sub> M
010H21 <b>1a</b> 0.5 mmol	+ HSIFII <sub>2</sub> Me – 2a X mmol	diglyme (1 mL) 130 °C, 24 h under Ar		<sub>0</sub> H <sub>21</sub> 3a
		Convers	ion (%)	GC Yield (%)
Entry	Silane amou	nt <b>1a</b>	2a	3a
1	0.5 mmol	>99	76	34
2	1.0 mmol	>99	50	55
3	1.5 mmol	97	34	59
4	2.0 mmol	94	34	62
	2.5 mm al	>00	23	86

### Table S5. Screening of catalyst amount

a 11 🔿		<sup>cat.</sup> Mn NPs ( <mark>X</mark> mol %	)	
C <sub>10</sub> H <sub>21</sub> <b>1a</b> 0.5 mmol	+ HSIPH <sub>2</sub> ivie - 2a 2.5 mmol	diglyme (1 mL) 130 °C, 24 h under Ar		C <sub>10</sub> H <sub>21</sub> 3a
		Conve	rsion (%)	GC Yield (%)
Entry	Catalyst amo	la la	2a	<b>3</b> a
1	0.01 mol %	% >99	23	86
2	0.02 mol %	% >99	44	79
3	0.05 mol %	‰	21	83
4	0.1 mol %	>99	25	79

Reaction conditions: 1-dodecene (0.5 mmol) reacted with methyldiphenylsilane (2.5 mmol) in the presence of Mn NPs (**X** mol %) in diglyme (1 mL) at 130 °C for 24 h.

### Table S6. Effect of radical scavenger

		<sup>cat.</sup> Mn NPs (0.01 mol % additive (X mmol)	) )	∽SiPh₂Me
C <sub>10</sub> H <sub>21</sub>	+ HSIPh <sub>2</sub> me – <b>2a</b> 2.5 mmol	diglyme (1 mL) 130 °C, 24 h under Ar		H <sub>21</sub> 3a
Entre		Convers	sion (%)	GC Yield (%)
Entry	Additive	1a	2a	<b>3</b> a
1	none	>99	23	86
2	TEMPO (2.5 m	mol) 24	>99	nd
3	TEMPO (0.2 m	mol) <1	23	nd
4	BHT (0.2 mm	ol) <1	_	nd

Reaction conditions: 1-dodecene (0.5 mmol) reacted with methyldiphenylsilane (2.5 mmol) in the presence of Mn NPs (0.01 mol %) and **additive** (**X** mmol) in diglyme (1 mL) at 130 °C for 24 h; nd: not detected.



Table S7. Screening of catalyst species

		USIDh Ma	catalyst	SiPh <sub>2</sub> Me
C <sub>10</sub> ⊓ <sub>21</sub> ×	+	HSIF H2IVIE	neat	$C_{10}H_{21} \sim 2$
1a		2a	130 °C, 24 h	3a
0.5 mmol		2.5 mmol	under Ar	

Entry		Convers	GC Yield (%)	
	Catalyst	<b>1</b> a	2a	3a
1	Mn NPs (0.01 mol %)	96	26	70
2	Mn <sub>2</sub> (CO) <sub>10</sub> (2.5 mol %)	92	-	59
3	HCl aq. (1 M, 50 μL, 10 mol %)	92	38	35
4	MnO (20 mol %)	<1	<1	nd
5	Mn NPs (0.01 mol %) <sup><i>a</i></sup>	-	-	32

Reaction conditions: 1-dodecene (0.5 mmol) reacted with methyldiphenylsilane (2.5 mmol) in the presence of **catalyst** (**X** mol %) at 130 °C for 24 h; nd: not detected. <sup>*a*</sup>Under air (balloon).

### 9. Characterization data

C<sub>10</sub>H<sub>21</sub> SiPh<sub>2</sub>Me

**3a**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1) and subsequently purified by distillation using a Kugelrohr apparatus (140 °C, 10 hPa, 1 h). The desired product was obtained as a colorless oil in 83% yield (152.1 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.53-7.49 (m, 4H), 7.39-7.32 (m, 6H), 1.39-1.16 (m, 20H), 1.05 (t, *J* = 8.0 Hz, 2H), 0.88 (t, *J* = 6.9 Hz, 3H), 0.53 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl3)  $\delta$ : 137.5 (C), 134.4 (CH), 129.0 (CH), 127.7 (CH), 33.6 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 23.7 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 14.1 (CH<sub>2</sub>), 14.1 (CH<sub>3</sub>), -4.4 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 698, 730, 786, 1112, 1250, 1427, 1466, 2853, 2924; GC-MS (EI) *m/z* (relative intensity) 289(5) [M-CH<sub>3</sub>]<sup>+</sup>, 135(100), 121(25), 127(14); HRMS (EI) *m/z* calcd for C<sub>25</sub>H<sub>38</sub>Si [M-CH<sub>3</sub>]<sup>+</sup>, 351.2508, found 351.2502.



**3b**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1) and subsequently purified by distillation using a Kugelrohr apparatus (130 °C, 0.3 mmHg, 1 h). The desired product was obtained as a colorless oil in 77% yield (130.3 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.53-7.51 (m, 4H), 7.39-7.32 (m, 6H), 1.34-1.24 (m, 16H), 1.07-1.05 (m, 2H), 0.88 (t, *J* = 6.9 Hz, 3H), 0.54 (s, 3H);<sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 137.5 (C), 134.4(CH), 129.0 (CH), 127.7 (CH), 33.6 (CH<sub>2</sub>), 31.8 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 23.7 CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 14.1 (CH<sub>3</sub>, CH<sub>2</sub>), -4.4 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 323(1) [M-CH<sub>3</sub>]<sup>+</sup>, 197(100), 198(20), 183(9).

The spectroscopic data were consistent with the reported values.<sup>1</sup>

Miura, H.; Doi, M.; Yasui, Y.; Masaki, Y.; Nishio, H.; Shishido, T. Diverse Alkyl-Silyl Cross-Coupling via Homolysis of Unactivated C(sp<sup>3</sup>)-O Bonds with the Cooperation of Gold Nanoparticles and Amphoteric Zirconium Oxides. *J. Am. Chem. Soc.* **2023**, *145*, 4613–4625.



**3c**: The desired product was isolated by column chromatography on silica gel using hexane as the eluent, followed by purification via distillation using a Kugelrohr apparatus (140 °C, 10 hPa, 1 h). The desired product was obtained as a colorless oil in 68% yield (104.9 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.49 (m, 4H), 7.38-7.31 (m, 6H), 1.71-1.64 (m, 5H), 1.28-1.03 (m, 8H), 0.86-0.82 (m, 2H), 0.53 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 137.5 (C), 134.4 (CH), 129.0 (CH), 127.7 (CH), 40.7 (CH), 32.9 (CH<sub>2</sub>), 31.1 (CH<sub>2</sub>), 26.7 (CH<sub>2</sub>), 26.4 (CH<sub>2</sub>), 11.0 (CH<sub>2</sub>), -4.5 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 698, 731, 788, 1112, 1250, 1427, 1449, 2849, 2920; HRMS (ESI) *m*/*z* Calcd for C<sub>21</sub>H<sub>29</sub>Si [M+H]<sup>+</sup> 309.2033, found 309.2032.



**3d**: The desired product was purified by distillation using a Kugelrohr apparatus (130°C, 0.3 mmHg, 1 h), and subsequently isolated by column chromatography on silica gel using hexane as the eluent. The desired product was obtained as a colorless oil in 65% yield (107.4 mg).

<sup>1</sup>H-NMR (400MHz; CDCl<sub>3</sub>)  $\delta$ : 7.51-7.49 (m, 4H), 7.38-7.31 (m, 6H), 7.25-7.23 (m, 2H), 7.17-7.11 (m, 3H), 2.57 (t, *J* = 7.8 Hz, 2H), 1.69-1.62 (m, 2H), 1.46-1.42 (m, 2H), 1.12-1.08 (m, 2H), 0.53 (s, 3H); <sup>13</sup>C-NMR (100MHz; CDCl<sub>3</sub>)  $\delta$ : 142.6 (C), 137.3 (C), 134.4 (CH), 129.0 (CH), 128.3 (CH), 128.2 (CH), 127.7 (CH), 125.5 (CH), 35.4 (CH<sub>2</sub>), 35.3 (CH<sub>2</sub>), 23.5 (CH<sub>2</sub>), 13.9 (CH<sub>2</sub>), -4.4 (CH<sub>3</sub>); GC-MS (EI) *m*/*z* (relative intensity) 315 (0.2) [M-CH<sub>3</sub>]<sup>+</sup> 197 (100) 252 (36) 198 (19). The spectroscopic data were consistent with the reported values.<sup>2</sup>



**3e**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1). The desired product was obtained as a colorless oil in 66% yield (104.4 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.50-7.46 (4H, m), 7.37-7.32 (6H, m), 7.26-7.24 (2H, m), 7.17-7.12 (3H, m), 2.64 (2H, t, *J* = 7.6 Hz), 1.72-1.68 (2H, m), 1.12-1.08 (2H, m), 0.53 (3H, s); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 142.3 (C), 137.1 (C), 134.4 (CH), 129.0 (CH), 128.5 (CH), 128.2 (CH), 127.7 (CH), 125.6 (CH), 39.7 (CH<sub>2</sub>), 25.8 (CH<sub>2</sub>), 13.8 (CH<sub>2</sub>), -4.4 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 698, 731, 789, 1112, 1251, 1427, 1453, 2858, 2926, 3024, 3068; GC-MS (EI) *m/z* (relative intensity) 289(5) [M-CH<sub>3</sub>]<sup>+</sup>, 135(100), 121(25), 127(14); HRMS (ESI) *m/z* Calcd for C<sub>22</sub>H<sub>24</sub>SiNa [M+Na]<sup>+</sup> 339.1539, found 339.1540.



**3f**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1). The desired product was obtained as a colorless oil in 77% yield (133.4 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.46 (4H, m), 7.38-7.30 (6H, m), 7.03 (2H, d, *J* = 8.7 Hz), 6.80 (2H, d, *J* = 8.7 Hz), 3.77 (3H, s), 2.58 (2H, t, *J* = 7.5 Hz), 1.67-1.65 (2H, m), 1.10-1.07 (2H, m), 0.53 (3H, s); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 157.6 (C), 137.2 (C), 134.4 (CH), 129.3 (CH), 129.0 (CH), 127.8 (C), 127.7 (CH), 113.5 (CH), 55.2 (CH<sub>3</sub>), 38.7 (CH<sub>2</sub>), 25.9 (CH<sub>2</sub>), 13.7 (CH<sub>2</sub>), -4.4 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 700, 730, 788, 1037, 1111, 1248, 1427, 1512, 2927, 3068; HRMS (ESI) *m/z* Calcd for C<sub>23</sub>H<sub>26</sub>OSiNa [M+Na]<sup>+</sup> 369.1645, found 369.1652.



**3g**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 8:2). The desired product was obtained as a colorless oil in 50% yield (74.1 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.51-7.49 (4H, m), 7.38-7.33 (6H, m), 2.40 (2H, t, *J* = 7.4 Hz), 2.10 (3H, s), 1.66-1.59 (2H, m), 1.42-1.34 (2H, m), 1.07 (2H, t, *J* = 8.5 Hz), 0.54 (3H, s); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 209.2 (C), 137.1 (C), 134.3 (CH), 129.1 (CH), 127.8 (CH), 43.4 (CH<sub>2</sub>), 29.8 (CH<sub>3</sub>), 27.6 (CH<sub>2</sub>), 23.4 (CH<sub>2</sub>), 14.0 (CH<sub>2</sub>), -4.8 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 239(2) [M]<sup>+</sup>-C<sub>3</sub>H<sub>5</sub>O, 197(100), 219(56), 137(28).



3h: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 99:1) and subsequently purified by distillation using a Kugelrohr apparatus (200 °C, 10 hPa, 0.5 h). The desired product was obtained as a colorless oil in 59% yield (117.0 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.49 (4H, m), 7.37-7.32 (6H, m), 3.66 (3H, s), 2.29 (2H, t, *J* = 7.5 Hz), 1.60 (2H, t, *J* = 7.3 Hz), 1.34-1.26 (14H, m), 1.06-1.04 (2H, m), 0.53 (3H, s). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 174.3 (C), 137.4 (C), 134.4 (CH), 129.0 (CH), 127.7 (CH), 51.4 (CH<sub>3</sub>), 34.0 (CH<sub>2</sub>), 33.6 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 29.4 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 29.1 (CH<sub>2</sub>), 29.1 (CH<sub>2</sub>), 24.9 (CH<sub>2</sub>), 23.7 (CH<sub>2</sub>), 14.1 (CH<sub>2</sub>), -4.4 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 699, 787, 1112, 1250, 1427, 1740, 2851, 2922; HRMS (ESI) *m/z* Calcd for C<sub>25</sub>H<sub>36</sub>O<sub>2</sub>SiNa [M+Na]<sup>+</sup> 419.2377, found 419.2379.



**3i**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 95:5). The desired product was obtained as a colorless oil in 60% yield (97.9 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>,)  $\delta$ : 7.52-7.31 (m, 10H), 4.01 (t, J = 6.7 Hz, 2H), 2.01 (s, 3H), 1.60 (t, J = 6.9 Hz, 2H), 1.40-1.39 (m, 4H), 1.09-1.01 (m, 2H), 0.53 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 171.1 (C), 137.2 (C), 134.3 (CH), 129.0 (CH), 127.7 (CH), 64.5 (CH<sub>2</sub>), 29.8 (CH<sub>2</sub>), 28.1 (CH<sub>2</sub>), 23.5 (CH<sub>2</sub>), 20.9 (CH<sub>3</sub>), 14.1 (CH<sub>2</sub>), -4.5 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 311 (1) [M-CH<sub>3</sub>] <sup>+</sup> 197 (100) 179 (33) 249 (22); HRMS (ESI) *m/z* Calcd for C<sub>20</sub>H<sub>26</sub>O<sub>2</sub>SiNa [M+Na]<sup>+</sup> 350.1621, found 350.1629.



**3j**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 9:1). The desired product was obtained as a colorless oil in 73% yield (108.2 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.51 (m, 4H), 7.40-7.33 (m, 6H), 2.87-2.85 (m, 1H), 2.71 (dd, J = 5.0, 4.0 Hz, 1H), 2.42 (dd, J = 5.0, 2.7 Hz, 1H), 1.58-1.41 (m, 6H), 1.11-1.07 (m, 2H), 0.55 (s, 3H) ; <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 137.2 (C), 134.4 (CH), 129.1 (CH), 127.7 (CH), 52.2 (CH), 47.0 (CH<sub>2</sub>), 32.0 (CH<sub>2</sub>), 29.7 (CH<sub>2</sub>), 23.6 (CH<sub>2</sub>), 14.1 (CH<sub>2</sub>), -4.5 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 699, 732, 787, 1112, 1251, 1427, 2924; HRMS (ESI) *m/z* Calcd for C<sub>19</sub>H<sub>24</sub>OSiNa [M+Na]<sup>+</sup> 319.1489, found 319.1474.



**3k**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 9:1). The desired product was obtained as a colorless oil in 77% yield (120.3 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.50 (4H, m), 7.39-7.32 (6H, m), 3.66 (1H, dd, *J* = 11.5, 3.1 Hz), 3.48-3.43 (2H, m), 3.33 (1H, dd, *J* = 11.5, 5.9 Hz), 3.14-3.10 (1H, m), 2.78 (1H, dd, *J* = 5.0, 4.1 Hz), 2.58 (1H, dd, *J* = 5.0, 2.8 Hz), 1.71-1.63 (2H, m), 1.10-1.06 (2H, m), 0.55 (3H, s); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 136.9 (C), 134.4 (CH), 129.1 (CH), 127.8 (CH), 74.1 (CH<sub>2</sub>), 71.3 (CH<sub>2</sub>), 50.8 (CH), 44.3 (CH<sub>2</sub>), 24.0 (CH<sub>2</sub>), 10.1 (CH<sub>2</sub>), -4.5 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 700, 732, 786, 1112, 1251, 1427, 2869, 2932; HRMS (ESI) *m/z* calcd for C<sub>19</sub>H<sub>24</sub>O<sub>2</sub>SiNa [M+Na]<sup>+</sup>, 335.1438, found 335.1445.



**31**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 95:5). The desired product was obtained as a colorless oil in 64% yield (83.3 mg).

<sup>1</sup>H-NMR (400 MHz,CDCl<sub>3</sub>) δ: 7.50-7.48 (m, 2H), 7.35-7.34 (m, 3H), 3.16-3.10 (m, 2H), 2.17-2.13 (m, 1H), 2.07-1.93 (m, 1H), 1.83-1.60 (m, 1H), 1.52-0.79 (m, 6H), 0.73-0.67 (m, 2H), 0.24 (s, 6H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>) δ: 139.3 (C), 133.4 (CH), 128.7 (CH), 127.6 (CH), 53.2 (CH), 52.7 (CH), 51.9 (CH), 51.9 (CH), 35.4 (CH), 32.4 (CH), 31.5 (CH<sub>2</sub>), 30.6 (CH<sub>2</sub>), 30.3 (CH<sub>2</sub>), 30.1 (CH<sub>2</sub>), 26.6 (CH<sub>2</sub>), 25.3 (CH<sub>2</sub>), 23.9 (CH<sub>2</sub>), 23.5 (CH<sub>2</sub>), 12.5 (CH<sub>2</sub>), 12.4 (CH<sub>2</sub>), -3.1 (CH<sub>3</sub>), -3.17 (CH<sub>3</sub>), -3.21 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 245 (1) [M-CH<sub>3</sub>]<sup>+</sup> 135 (100) 137 (34) 136 (14).



3m: The desired product was isolated by distillation using a Kugelrohr apparatus (160 °C, 10 hPa, 1 h). The desired product was obtained as a colorless oil in 73% yield (141.8 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.51-7.50 (m, 2H), 7.34-7.34 (m, 3H), 3.80-3.78 (m, 6H), 1.24-1.18 (m, 9H), 0.79-0.76 (m, 2H), 0.56-0.54 (m, 2H), 0.26 (s, 6H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 139.0 (C), 133.6 (CH), 128.7 (CH), 127.6 (CH), 58.3 (CH<sub>2</sub>), 18.2 (CH<sub>3</sub>), 6.7 (CH<sub>2</sub>), 2.4 (CH<sub>2</sub>), -3.7 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 311 (45) [M-CH<sub>3</sub>]<sup>+</sup> 135 (100) 205 (53) 311 (45).

The spectroscopic data were consistent with the reported values.<sup>5</sup>



**3n**: The desired product was isolated by column chromatography (silica gel, hexane/ethyl acetate, 8:2). The desired product was obtained as a colorless oil in 69% yield (102.9 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.52-7.50 (4H, m), 7.39-7.32 (6H, m), 3.61 (2H, t, *J* = 6.6 Hz), 1.53-1.51 (2H, m), 1.38-1.34 (6H, m), 1.21 (1H, br), 1.08-1.06 (2H, m), 0.54 (3H, s); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 137.3 (C), 134.4 (CH), 129.0 (CH), 127.7 (CH), 62.9 (CH<sub>2</sub>), 33.2 (CH<sub>2</sub>), 32.6 (CH<sub>2</sub>), 25.2 (CH<sub>2</sub>), 23.7 (CH<sub>2</sub>), 14.0 (CH<sub>2</sub>), -4.5 (CH<sub>3</sub>); IR (neat, cm<sup>-1</sup>) 699, 731, 787, 1112, 1250, 1427, 2926, 3333; HRMS (ESI) *m*/*z* Calcd for C<sub>19</sub>H<sub>26</sub>OSiNa [M+Na]<sup>+</sup> 321.1645, found 321.1642.



**30**: The desired product was isolated by column chromatography (silica gel, hexane 100%). The desired product was obtained as a colorless oil in 33% yield (36.0 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.58-7.56 (2H, m), 7.37-7.34 (3H, m), 4.19 (2H, d, *J* = 3.3 Hz), 1.84-1.43 (14H, m), 1.29-1.25 (1H, m); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 135.5 (CH), 132.6 (C), 129.4 (CH), 127.8 (CH), 28.6 (CH<sub>2</sub>), 27.3 (CH<sub>2</sub>), 26.8 (CH<sub>2</sub>), 26.5 (CH<sub>2</sub>), 20.1 (CH); GC-MS (EI) *m/z* (relative intensity) 218 (0.8) [M]<sup>+</sup> 140 (100) 107 (92) 112 (60).

The spectroscopic data were consistent with the reported values.<sup>6</sup>



**3p**: The desired product was isolated by column chromatography (silica gel, hexane 100%) and subsequently purified by distillation using a Kugelrohr apparatus (150 °C, 0.3 mmHg, 30 min). The desired product was obtained as a colorless oil in 63% yield (95.9 mg).

<sup>1</sup>H-NMR (400 MHz ; CDCl<sub>3</sub>)  $\delta$ : 7.52-7.33 (m, 5H), 1.30-1.26 (m, 20H), 0.88 (t, *J* = 6.9 Hz, 3H), 0.73 (t, *J* = 7.8 Hz, 2H), 0.25 (s, 6H); <sup>13</sup>C-NMR (100 MHz ; CDCl<sub>3</sub>)  $\delta$ : 139.7 (C), 133.5 (CH), 128.6 (CH), 127.6 (CH), 33.6 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 31.7 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 23.8 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 15.6 (CH<sub>2</sub>), 14.1 (CH<sub>3</sub>), -3.0 (CH<sub>3</sub>); GC-MS (EI) *m/z* (relative intensity) 289 (4) [M-CH<sub>3</sub>]<sup>+</sup> 135 (100) 121 (26) 127 (15).



**3q**: The desired product was isolated by column chromatography (silica gel, hexane 100%) and subsequently purified by distillation using a Kugelrohr apparatus (110 °C, 0.3 mmHg, 30 min). The desired product was obtained as a colorless oil in 82% yield (144.5 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.56-7.54 (m, 4H), 7.38-7.35 (m, 6H), 4.84 (t, *J* = 3.7 Hz, 1H), 1.47-1.11 (m, 22H), 0.88 (t, *J* = 6.9 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 135.1 (CH), 134.7 (C), 129.4 (CH), 127.9 (CH), 33.1 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 24.3 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 14.1 (CH<sub>3</sub>), 12.1 (CH<sub>2</sub>); GC-MS (EI) *m/z* (relative intensity) 275 (1) [M-C<sub>6</sub>H<sub>5</sub>] <sup>+</sup> 183 (100) 175 (32) 105 (19).

The spectroscopic data were consistent with the reported values.<sup>7</sup>



**3r**: The desired product was isolated by column chromatography (silica gel, hexane 100%). The desired product was obtained as a colorless oil in 82% yield (113.3 mg).

<sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.58-7.55 (m, 2H), 7.38-7.34 (m, 3H), 4.28 (t, *J* = 3.7 Hz, 2H), 1.47-1.43 (m, 2H), 1.36-1.25 (m, 18H), 0.95-0.92 (m, 2H), 0.88 (t, *J* = 6.9 Hz, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 135.1 (CH), 132.8 (C), 129.4 (CH), 127.9 (CH), 32.8 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 29.5 (CH<sub>2</sub>), 29.3 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 25.0 (CH<sub>2</sub>), 22.6 (CH<sub>2</sub>), 14.2 (CH<sub>3</sub>), 9.9 (CH<sub>2</sub>); GC-MS (EI) *m/z* (relative intensity) 275 (0.03) [M-H] + 99 (100) 107 (54) 113 (38).

The spectroscopic data were consistent with the reported values.8

# 10. NMR-Spectra

## 3a <sup>1</sup>H-NMR





## **3b** <sup>1</sup>H-NMR





## 3c<sup>1</sup>H-NMR





## 3d <sup>1</sup>H-NMR





## **3e** <sup>1</sup>H-NMR





### 3f<sup>1</sup>H-NMR





## **3h** <sup>1</sup>H-NMR





### 3i<sup>1</sup>H-NMR





### 3l<sup>1</sup>H-NMR





## 3m<sup>1</sup>H-NMR





## **3n** <sup>1</sup>H-NMR





## **30** <sup>1</sup>H-NMR





## **3p** <sup>1</sup>H-NMR





# **3q** <sup>1</sup>H-NMR





## 3r<sup>1</sup>H-NMR





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