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High-order layered self-assembled multicavity metal-organic capsules and anticooperative host-multi-guest chemistry

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1. General Procedures

All chemicals were purchased from commercial suppliers and used without further purification unless otherwise specified. Column chromatography was conducted using basic Al₂O₃ (Sinopharm Chemical Reagents Co. Ltd, 200-300 mesh) or SiO₂ (Qingdao Haiyang Chemical Co., Ltd, 200-300 mesh).

NMR spectra were recorded on a Bruker ADVANCE 400M, 500M and 600M NMR Spectrometer. ¹H NMR chemical shifts are reported in ppm downfield from tetramethylsilane (TMS) reference using the residual protonated solvent as an internal standard.

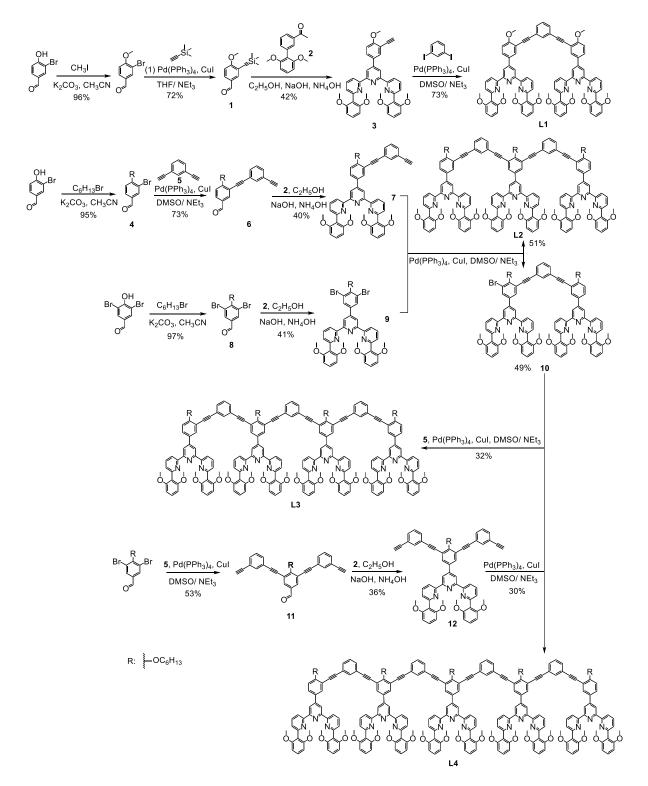
Mass spectra of complexes and ligands were determined on Waters Synapt G2 Mass Spectrometer with traveling wave ion mobility (TWIM) under the following conditions: ESI capillary voltage, 3.5 kV; cone voltage, 35 V; desolvation gas flow, 800 L/h. TWIM-MS was measured with IM traveling wave height, 25 V; and IM traveling wave velocity, 1000 m/s.

Atomic force microscopy (AFM) was conducted on a Bruker Dimension Icon AFM system with ScanAsyst and the data were processed by NanoScope Analysis version 1.5 (Bruker Software, Inc.). AFM samples were prepared by casting a sample solution $(1 \times 10^{-7} \text{ M})$ on a freshly cleaved mica surface.

Transmission electron microscope (TEM) images were recorded on a JEM 2100 transmission electron microscope operated at an accelerating voltage of 200 KV. TEM samples were prepared by drop-casting a sample solution $(1 \times 10^{-7} \text{ M})$ onto a carbon-coated copper grid and dried in vacuo for 24 h.

Molecular Modeling. Calculations were proceeded with Geometry Optimization and followed by Anneal in Forcite module of Materials Studio version 7.0 program (Accelrys Software, Inc.)., the counterions were omitted.

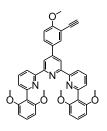
2. Synthesis of ligand L1-L5.



Scheme S1. Synthesis of ligands L1-L4.



Compound 1. Dissolved 3-bromo-4-methoxybenzaldehyde (2.15 g, 10 mmol) and ethynyltrimethylsilane (1.20 g, 12 mmol) in 50ml of anhydrous tetrahydrofuran, then added 30ml of anhydrous triethylamine, then added CuI 15 mg and Pd(PPh₃)₄ (177 mg, 0.15 mmol), and finally let the whole reaction under the atmosphere of Ar gas 85 °C reaction for 12h. After the reaction was cooled to room temperature, silica gel powder was added and decompression distillation was conducted to obtain the sample, The intermediate product was obtained by chromatography column method, dissolved in the mixed solvent of CHCl₃/MeOH (1:1, v/v) and added with NaOH, stirred at room temperature for 2h, and finally extracted to obtain compound 1. (1.60 g, 72%); ¹H NMR (400 MHz, CDCl₃) δ = 9.78 (s, 1H), 7.89 (s, 1H), 7.76-7.74 (d, J = 8 Hz, 1H), 6.91-6.89 (d, J = 8 Hz, 1H), 3.89 (s, 3H), 0.20 (s, 12H). ¹³C NMR (126 MHz, CDCl₃, ppm) δ = 190.12, 164.80, 136.09, 131.86, 129.54, 113.41, 110.78, 100.19, 99.62, 56.33.

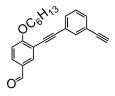


Compound 3. Compound **1** (2.20 g, 10 mmol), compound **2** (5.63 g, 22 mmol) and NaOH (1.60 g, 40 mmol) were mixed in 80ml EtOH and stirred at room temperature for 1 day. Then 40ml NH₃·H₂O was added to reflux for one day at 85 °C. After the reaction, it was cooled to room temperature and precipitation would occur. The solid was pumped out and recrystallized with CHCl₃/CH₃OH to give **3** as a white solid (2.67 g, 4.20 mmol) in 42% yield. ¹H NMR (500 MHz, 298 K, CDCl₃, ppm) δ = 8.65 (s, 2H), 8.62-8.61 (d, J = 5 Hz, 2H), 7.98 (s, 1H), 7.95-7.92 (t, 2H), 7.83-7.81 (d, 1H), 7.40-7.37 (t, 4H), 6.98-6.97 (d, J = 5 Hz, 1H), 6.74-6.73 (d, 4H), 3.96 (s, 3H), 3.80 (s, 12H), 3.34 (s, 1H). ¹³C NMR (126 MHz, 298 K, CDCl₃) δ = 160.93, 158.38, 156.45, 156.18, 153.86, 148.18, 136.42, 133.05, 131.41, 129.63, 129.39, 126.23, 119.82, 119.55, 118.72, 111.48, 110.83, 104.63, 81.35, 79.86, 56.19, 56.04.

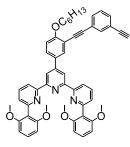


Compound 4. To a flask containing 3-bromo-4-hydroxybenzaldehyde (5.02 g, 25 mmol), 1-bromohexane (8.25 g, 50 mmol) and K₂CO₃ (6.90 g, 50 mmol), add 60ml CH₃CN as solvent. The system was pumped and backfilled with Ar. Then the mixture was refluxed for 1 days under Ar. After cooled to 25 °C, the K₂CO₃ in the reaction solution was filtered out, the filtrate was decompressed and distilled to obtain the colorless liquid compound **4** (6.77 g, 23.7 mmol) in 95% yield. ¹H NMR (400 MHz, 298 K, CDCl₃, ppm) δ = 9.83 (s, 1H), 8.08 (s, 1H), 7.81-7.78 (d, *J* = 12

Hz, 1H), 6.99-6.97(d, *J* = 8 Hz, 1H), 4.13-4.10 (t, 2H), 1.91-1.86 (m, 2H), 1.54-1.50 (m, 2H), 1.38-1.35 (m, 4H), 0.93-0.90 (m, 3H). ¹³C NMR (101 MHz, 298 K, CDCl₃, ppm) δ = 156.34, 155.98, 150.01, 149.19, 147.23, 136.80, 136.68, 131.57, 126.98, 123.75, 121.32, 119.14, 64.79.



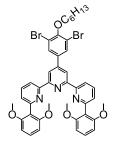
Compound 6. To a flask containing compound **4** (1.71 g, 6.0 mmol) and 1,3-diethynylbenzene (**5**) (1.50 g, 12 mmol), a mixed solvent (50 mL) of THF/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (212 mg, 0.18 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (SiO₂, CH₂Cl₂/hexane = 1:4) to give **6** as a white solid (1.44 g, 4.38 mmol) in 73% yield. ¹H NMR (400 MHz, 298 K, CDCl₃, ppm) δ = 9.90 (s, 1H), 8.03 (s, 1H), 7.87-7.85 (d, *J* = 8 Hz, 1H), 7.68 (s, 1H), 7.54-7.52 (d, *J* = 8 Hz, 1H), 7.49-7.48 (d, *J* = 4 Hz, 1H), 7.36-7.33 (t, 1H), 7.03-7.01 (d, *J* = 4 Hz, 1H), 4.18-4.14 (t, 2H), 3.13 (s, 1H), 1.94-1.91 (m, 2H), 1.60-1.57 (m, 2H), 1.43-1.36 (m, 4H), 0.94-0.91 (m, 3H). ¹³C NMR (101 MHz, 298 K, CDCl₃, ppm) δ = 135.94, 135.64, 132.86, 132.72, 132.38, 128.61, 128.44, 122.73, 122.50, 122.06, 82.59, 82.40, 80.76, 78.25, 77.96, 74.39.



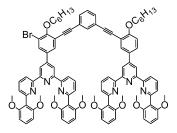
Compound 7. To a flask containing compound **6** (2.64 g, 8.0 mmol), compound **2** (4.42 g, 17.6 mmol) and NaOH (1.28 g, 32 mmol), add 80ml EtOH as solvent and stirred at room temperature for 1 day. Then 40ml NH₃·H₂O was added to reflux for one day at 85 °C. After the reaction, it was cooled to room temperature and precipitation would occur. The solid was pumped out and recrystallized with CHCl₃/CH₃OH to give 7 as a white solid (2.58 g, 4.20 mmol) in 40% yield. ¹H NMR (500 MHz, CDCl₃, ppm) δ = 8.65 (s, 2H), 8.60-8.59 (d, *J* = 4 Hz, 2H), 7.97-7.96 (d, *J* = 4 Hz, 1H), 7.93-7.89 (t, 2H), 7.77-7.75 (d, *J* = 4 Hz, 2H), 7.65 (s, 1H), 7.51-7.49 (d, *J* = 8 Hz, 1H), 7.44-7.42 (d, *J* = 8 Hz, 1H), 7.38-7.34 (t, 3H), 7.32-7.28 (t, 2H), 6.96-6.94 (d, *J* = 8 Hz, 1H), 6.72-6.70 (d, *J* = 8 Hz, 4H), 4.10-4.07 (t, 2H), 3.78 (s, 12H), 3.09 (s, 1H), 1.89-1.86 (m, 2H), 1.55-1.53 (m, 2H), 1.40-1.35 (m, 4H), 0.92-0.88 (m, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 164.99, 160.23, 158.40, 156.41, 153.79, 148.53, 136.49, 135.10, 132.21, 131.84, 131.58, 129.69, 128.37, 126.25, 124.12, 122.37, 119.60, 118.87, 112.84, 112.09, 104.68, 92.52, 86.63, 82.92, 68.88, 56.21, 31.64, 29.15, 25.81, 22.60, 14.07.



Compound 8. To a flask containing 3,5-dibromo-4-hydroxybenzaldehyde (5.60 g, 20 mmol), 1-bromohexane (6.60 g, 40 mmol) and K₂CO₃ (6.90 g, 50 mmol), add 60 ml CH₃CN as solvent. The system was pumped and backfilled with Ar. Then the mixture was refluxed for 1 days under Ar. After cooled to 25 °C, the K₂CO₃ in the reaction solution was filtered out, the filtrate was decompressed and distilled to obtain the colorless liquid compound **8** (7.06 g, 23.7 mmol) in 97% yield. ¹H NMR (400 MHz, CDCl₃) δ = 9.75 (s, 1H), 7.91 (s, 2H), 3.99-3.96 (t, 2H), 1.81-1.78 (m, 2H), 1.48-1.43 (m, 2H), 1.29-1.24 (m, 4H), 0.84-0.81 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 188.43, 158.52, 133.96, 133.89, 119.48, 74.13, 31.60, 30.01, 25.48, 22.62, 14.11.

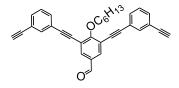


Compound 9. To a flask containing compound **8** (3.64 g, 10 mmol), compound **2** (5.52 g, 22 mmol) and NaOH (1.60 g, 40 mmol), add 80 ml EtOH as solvent and stirred at room temperature for 1 day. Then 30ml NH₃·H₂O was added to reflux for one day at 85 °C. After the reaction, it was cooled to room temperature and precipitation would occur. The solid was pumped out and recrystallized with CHCl₃/CH₃OH to give **9** as a white solid (3.44 g, 4.10 mmol) in 41% yield.; ¹H NMR (400 MHz, CDCl₃, ppm) δ = 8.60-8.58 (d, *J* = 8 Hz, 2H), 8.58 (s, 2H), 7.94 (s, 2H), 7.94-7.90 (t, 2H), 7.40-7.38 (d, *J* = 8 Hz, 2H), 7.39-7.35 (t, 2H), 6.73-6.71 (d, *J* = 8 Hz, 4H), 4.05-4.01 (t, 2H), 3.79 (s, 12H), 1.90-1.84 (m, 2H), 1.56-1.51 (m, 2H), 1.39-1.35 (m, 4H), 0.93-0.90 (m, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm): δ = 158.34, 156.70, 155.79, 153.92, 153.72, 146.53, 137.32, 136.46, 131.47, 129.72, 126.47, 119.59, 119.55, 118.90, 118.71, 104.63, 73.79, 56.16, 31.66, 30.01, 25.54, 22.63, 14.09.

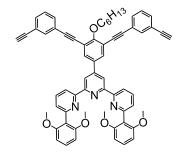


Compound 10. To a flask containing compound 7 (3.22 g, 4.0 mmol) and compound 9 (3.36 g, 4.0 mmol), a mixed solvent (50 mL) of DMSO/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (14 mg, 0.12 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 6 h under

Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:1) to give **10** as a white solid (3.20 g, 2.04 mmol) in 49% yield. ¹H NMR (400 MHz, CDCl₃) δ = 8.65 (s, 2H), 8.63 (s, 2H), 8.61-8.60 (d, *J* = 4 Hz, 2H), 8.59-8.58 (d, *J* = 4 Hz, 2H), 7.98-7.96 (d, *J* = 8 Hz, 2H), 7.92-7.88 (m, 5H), 7.77-7.75 (d, *J* = 8 Hz, 1H), 7.70 (s, 1H), 7.51-7.49 (d, *J* = 8 Hz, 1H), 7.47-7.45 (d, *J* = 8 Hz, 1H), 7.38-7.29 (m, 9H), 6.97-6.94 (d, *J* = 12 Hz, 1H), 6.69-6.66 (d, *J* = 8 Hz, 8H), 4.25-4.21 (t, 2H), 4.10-4.07 (t, 2H), 3.76 (s, 12H), 3.75 (s, 12H), 1.90-1.83 (m, 4H), 1.56-1.51 (m, 4H), 1.37-1.29 (m, 8H), 0.87-0.81 (m, 6H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 158.34, 158.32, 157.52, 156.68, 156.60, 155.85, 155.75, 153.90, 153.73, 147.24, 147.07, 146.53, 142.45, 139.83, 137.30, 136.42, 135.77, 134.47, 132.43, 132.22, 131.74, 131.65, 131.47, 131.00, 129.73, 128.42, 126.48, 126.41, 124.20, 123.19, 119.56, 119.53, 118.98, 118.90, 118.71, 118.05, 112.90, 112.11, 104.62, 99.98, 99.94, 93.74, 92.57, 86.59, 85.55, 74.47, 73.79, 68.89, 56.16, 56.14, 31.70, 31.66, 31.57, 30.32, 30.01, 29.33, 29.13, 25.83, 25.71, 25.54, 22.63, 14.09, 14.04, 14.01.

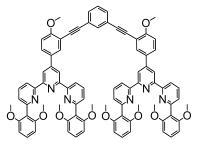


Compound 11. To a flask containing compound **8** (2.55 g, 7.0 mmol) and 1,3-diethynylbenzene (**5**) (2.65 g, 21 mmol), a mixed solvent (50 mL) of THF/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (212 mg, 0.18 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (SiO₂, CH₂Cl₂/hexane = 1:4) to give **6** as a white solid (1.69 g, 3.71 mmol) in 53% yield. ¹H NMR (400 MHz, CDCl₃, ppm) δ = 9.92 (s, 1H), 7.99 (s, 2H), 7.67 (s, 1H), 7.53-7.48 (t, 4H), 7.37-7.33 (t, 2H), 4.52-4.49 (t, 2H), 3.12 (s, 2H), 1.92-1.88 (m, 2H), 1.61-1.57 (m, 2H), 1.36-1.31 (m, 4H), 0.87-0.83 (m, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 188.64, 134.06, 133.98, 131.32, 130.76, 127.57, 122.09, 121.66, 116.70, 92.86, 84.10, 81.54, 77.05, 73.83, 30.70, 29.54, 24.90, 21.63, 13.02.

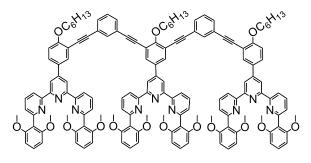


Compound 12. To a flask containing compound **11** (2.28 g, 5.0 mmol), compound **2** (2.76 g, 11 mmol) and NaOH (0.80 g, 20 mmol), add 50 ml EtOH as solvent and stirred at room temperature for 1 day. Then 20 ml NH₃·H₂O was added to reflux for one day at 85 °C. After the reaction, it was cooled to room temperature, the mixture was extracted

with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:2) to give **12** as a white solid (1.67 g, 1.80 mmol) in 36% yield. ¹H NMR (400 MHz, CDCl₃, ppm) δ = 8.66 (s, 2H), 8.62-8.60 (d, *J* = 8 Hz, 2H), 7.93 (s, 2H), 7.91-7.81 (m, 2H), 7.66 (s, 2H), 7.51-7.49 (d, *J* = 8 Hz, 2H), 7.47-7.45 (d, *J* = 8 Hz, 2H), 7.39-7.32 (m, 6H), 6.72-6.70 (d, *J* = 8 Hz, 4H), 4.38-4.35 (t, 2H), 3.78 (s, 12H), 3.11 (s, 2H), 1.91-1.86 (m, 2H), 1.63-1.56 (m, 2H), 1.35-1.29 (m, 4H), 0.86-0.83 (m, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 199.99, 160.51, 157.37, 154.95, 152.85, 146.73, 135.21, 134.00, 131.75, 130.94, 130.78, 128.98, 128.70, 127.45, 125.35, 122.60, 121.53, 118.58, 118.03, 116.59, 103.69, 91.86, 85.20, 81.72, 76.79, 73.76, 55.17, 30.78, 29.56, 25.03, 21.64, 13.03.

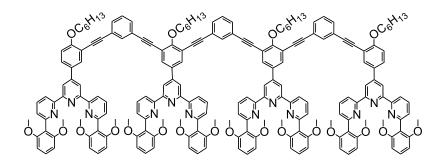


Compound L1. To a flask containing compound **3** (3.18 g, 5.0 mmol) and 1,3-diiodobenzene (750 mg, 2.27 mmol), a mixed solvent (50 mL) of THF/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (212 mg, 0.18 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:1) to give compound **L1** as a white solid (2.46 g, 1.66 mmol) in 73% yield. ¹H NMR (400 MHz, CDCl₃, ppm) $\delta = 8.66$ (s, 4H), 8.60-8.59 (d, J = 4 Hz, 4H), 7.99 (s, 2H), 7.92-7.88(s, 4H), 7.80 (s, 1H), 7.77-7.76 (d, J = 4 Hz, 2H), 7.51-7.49 (d, J = 8 Hz, 2H), 7.37-7.35 (d, J = 8 Hz, 4H), 7.33-7.29 (m, 5H), 6.98-6.96 (d, J = 8 Hz, 2H), 6.68-6.66 (d, J = 8 Hz, 8H), 3.94 (s, 6H), 3.75 (s, 24H). ¹³C NMR (101 MHz, CDCl₃, ppm) $\delta = 160.38$, 158.35, 156.45, 156.22, 153.88, 148.42, 136.43, 134.88, 132.59, 131.42, 131.29, 129.68, 129.02, 128.33, 126.24, 123.71, 119.72, 119.56, 118.81, 112.63, 110.91, 104.60, 92.92, 86.10, 56.18, 56.04.

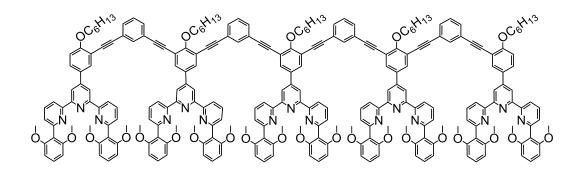


Compound L2. To a flask containing compound 7 (2.01 g, 2.50 mmol) and compound 9 (1.0 g, 1.19 mmol), a mixed solvent (50 mL) of DMSO/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (0.14 g, 0.12 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was

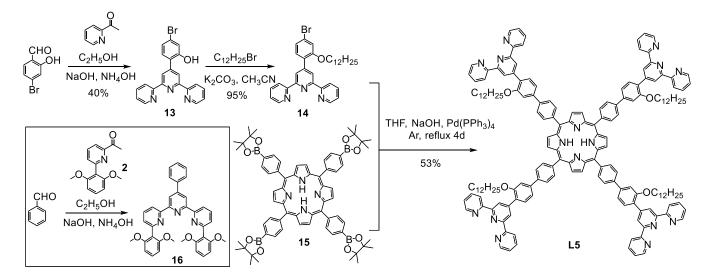
evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:1) to give compound **L2** as a white solid (1.39 g, 2.04 mmol) in 51% yield. ¹H NMR (400 MHz, CDCl₃, ppm) δ = 8.67 (s, 2H), 8.65 (s, 4H), 8.60-8.58 (d, *J* = 8 Hz, 6H), 7.97-7.96 (d, *J* = 4 Hz, 2H), 7.93 (s, 2H), 7.91-7.88 (t, 6H), 7.77-7.75 (d, *J* = 8 Hz, 2H), 7.71 (s, 2H), 7.51-7.47 (t, 4H), 7.36-7.24 (m, 16H), 6.96-6.94 (d, *J* = 8 Hz, 2H), 6.68-6.66 (d, *J* = 8 Hz, 8H), 6.65-6.63 (d, *J* = 8 Hz, 4H), 4.41-4.38 (t, 2H), 4.10-4.07 (t, 4H), 3.74 (s, 24H), 3.73 (s, 12H), 1.89-1.83 (m, 6H), 1.55-1.52 (m, 6H), 1.37-1.22 (m, 13H), 0.87-0.83 (t, 6H), 0.77-0.73 (t, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 161.55, 160.20, 158.34, 158.30, 156.54, 155.93, 153.92, 153.79, 148.63, 147.91, 136.54, 136.47, 136.41, 134.47, 134.42, 132.70, 132.21, 131.49, 131.17, 131.04, 129.71, 129.05, 128.38, 126.36, 126.28, 124.13, 123.48, 119.57, 119.50, 119.06, 118.84, 117.83, 112.94, 112.12, 104.62, 93.21, 92.67, 86.50, 85.96, 74.73, 68.89, 56.16, 56.13, 31.78, 31.56, 30.58, 29.13, 26.04, 25.71, 22.66, 22.63, 14.01.



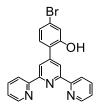
Compound L3. To a flask containing compound **10** (3.13 g, 2.0 mmol) and compound **5** (84 mg, 0.67 mmol), a mixed solvent (45 mL) of DMSO/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (0.14 g, 0.12 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:1) to give compound **L3** as a white solid (664 mg, 0.21 mmol) in 32% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.67 (s, 4H), 8.65 (s, 4H), 8.60 (d, 4H), 8.58 (d, 4H), 7.97-7.96 (d, *J* = 8 Hz, 2H), 7.93 (s, 4H), 7.91-7.87 (t, 8H), 7.76-7.73 (m, 3H), 7.71 (s, 2H), 7.52-7.47 (m, 6H), 7.35-7.23 (m, 19H), 6.96-6.94 (d, *J* = 8 Hz, 2H), 6.68-6.65 (d, *J* = 8 Hz, 8H), 6.64-6.62 (d, *J* = 8 Hz, 8H), 4.41-4.38 (t, 4H), 4.09-4.06 (t, 4H), 3.74 (s, 24H), 3.72 (s, 24H), 1.89-1.84 (m, 8H), 1.55-1.50 (m, 8H), 1.33-1.24 (m, 16H), 0.86-0.83 (t, 6H), 0.77-0.74 (t, 6H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 161.55, 160.17, 158.33, 158.28, 156.58, 156.40, 156.19, 155.95, 153.96, 153.87, 148.55, 147.90, 136.36, 134.49, 134.46, 132.79, 132.68, 132.21, 131.50, 131.41, 131.24, 131.04, 129.92, 129.89, 129.70, 129.64, 129.00, 128.51, 128.39, 126.34, 126.20, 124.14, 123.66, 123.46, 119.77, 119.61, 119.51, 119.49, 119.02, 118.77, 117.84, 117.77, 112.93, 112.11, 104.62, 104.59, 93.24, 93.01, 92.65, 86.53, 86.18, 85.93, 77.23, 74.74, 68.88, 56.14, 56.10, 31.78, 31.56, 30.58, 29.71, 29.32, 29.13, 26.04, 25.71, 22.66, 22.63, 14.01, 1.03.



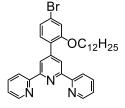
Compound L4. To a flask containing compound **10** (1.57 g, 1.0 mmol) and compound **12** (372 mg, 0.40 mmol), a mixed solvent (30 mL) of DMSO/Et₃N (3:2, v/v) was added. After Pd(PPh₃)₄ (0.14 g, 0.12 mmol) and CuI 15 mg was added into the mixture, the system was pumped and backfilled with Ar. Then the mixture was refluxed for 10 h under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 3:1) to give compound **L2** as a white solid (468 mg, 0.12 mmol) in 30% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.67 (s, 8H), 8.66 (s, 2H), 8.60-8.58 (d, *J* = 8 Hz, 10H), 7.97-7.96 (d, *J* = 4 Hz, 2H), 7.93 (s, 6H), 7.91-7.87 (m, 10H), 7.78-7.76 (d, *J* = 8 Hz, 2H), 7.73 (s, 2H), 7.71 (s, 2H), 7.52-7.47 (m, 10H), 7.37-7.23 (m, 24H), 6.96-6.94 (d, *J* = 8 Hz, 2H), 6.67-6.65 (d, 4H), 6.64-6.62 (d, 6H), 4.41-4.38 (t, 6H), 4.09-4.06 (t, 4H), 3.74 (s, 24H), 3.72 (s, 24H), 3.71 (s, 12H), 1.89-1.84 (m, 10H), 1.59-1.51 (m, 10H), 1.33-1.24 (m, 20H), 0.86-0.83 (t, 6H), 0.77-0.74 (t, 9H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 161.55, 160.20, 158.33, 158.28, 156.55, 155.92, 153.95, 153.82, 148.63, 147.91, 136.40, 134.46, 132.79, 132.68, 132.21, 131.50, 131.42, 131.17, 131.04, 129.73, 129.05, 128.52, 128.39, 126.37, 126.27, 124.14, 123.66, 123.64, 123.45, 119.57, 119.51, 119.05, 118.83, 117.84, 117.78, 112.93, 112.13, 104.62, 93.25, 93.06, 93.01, 92.65, 86.52, 86.19, 86.14, 85.93, 77.23, 74.74, 68.89, 56.15, 56.11, 40.98, 31.78, 31.56, 30.58, 29.78, 29.33, 29.13, 27.22, 26.04, 25.71, 22.66, 22.63, 14.03, 14.01.



Scheme S2. Synthesis of ligands L5.



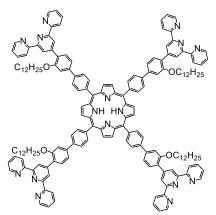
Compound 13. 4-bromo-2-hydroxybenzaldehyde (4.02 g, 20 mmol), 2-acetylpyridine (5.08 g, 42 mmol) and NaOH (3.2 g, 80 mmol) were mixed in 80 ml EtOH and stirred at room temperature for 1 day. Then 40 ml NH₃·H₂O was added to reflux for one day at 85 °C. After the reaction, it was cooled to room temperature and precipitation would occur. The yellow solid was pumped out and wash with a small amount of CH₃OH to give **13** as a yellow solid (3.23 g, 8.0 mmol) in 40% yield. ¹H NMR (400 MHz, DMSO) δ = 8.69-8.67 (d, *J* = 8 Hz, 2H), 8.67 (s, 2H), 8.63-8.61 (d, *J* = 8 Hz, 2H), 8.02-7.98 (t, 2H), 7.49-7.45 (m, 2H), 7.39-7.37 (d, *J* = 8 Hz, 2H), 7.06 (s, 1H), 6.93-6.90 (d, 1H). ¹³C NMR (101 MHz, DMSO) δ = 156.72, 154.56, 151.20, 149.59, 146.43, 141.10, 140.97, 137.55, 130.42, 124.21, 123.94, 123.57, 121.10, 119.82.



Compound 14. To a flask containing compound **13** (2.02 g, 5.0 mmol), 1-bromododecane (1.88 g, 7.5 mmol) and K₂CO₃ (1.73 g, 12.5 mmol), add 60 ml DMF as solvent. The system was pumped and backfilled with Ar. Then the mixture was refluxed for 1 days under Ar. After cooled to 25 °C, the mixture was extracted with CH₂Cl₂ and the combined organic extract was evaporated to dryness under reduced pressure. Then subjected to column chromatography (Al₂O₃, CH₂Cl₂/hexane = 1:10) to give compound **14** as a white solid (2.72 g, 23.7 mmol) in 95% yield. ¹H NMR (400 MHz, CDCl₃, ppm) δ = 8.73-8.72 (d, *J* = 4 Hz, 2H), 8.70-8.69 (d, *J* = 4 Hz, 4H), 7.91-7.88 (t, 2H), 7.47-7.45 (d, *J* = 8 Hz, 1H), 7.37-7.35 (t, *J* = 8 Hz, 2H), 7.23-7.21 (d, *J* = 8 Hz, 1H), 7.16 (s, 1H), 4.03-4.01 (t, *J* = 8 Hz, 2H), 1.76-1.71 (m, 2H), 1.43-1.36 (m, 2H), 1.28-1.09 (m, 16H), 0.91-0.88 (t, 3H). ¹³C NMR (101 MHz, CDCl₃, ppm) δ = 156.96, 156.20, 155.03, 148.97, 147.51, 137.00, 131.63, 127.25, 123.81, 123.72, 123.36, 121.72, 121.34, 115.72, 68.91, 31.93, 29.63, 29.62, 29.58, 29.40, 29.38, 29.37, 29.05, 26.12, 22.71, 14.16.

Compound 15 were synthesized according to the reported procedures. ^{S1}

Compound 16 were synthesized according to the reported procedures. ^{S2}



Compound L5. To a flask containing compound **15** (559 mg, 0.5 mmol), compound **14** (1.72 mg, 3.0 mmol) and K₂CO₃ (690 mg, 12.5 mmol) with 2 ml H₂O, then add 30 ml THF as solvent. After Pd(PPh₃)₄ (140 mg, 0.12 mmol) was added, the system was pumped and backfilled with Ar. Then the system was refluxed for 4 days under Ar. After cooled to 25 °C, the mixture was extracted with CHCl₃ and the combined organic extract was evaporated to dryness in vacuo to give a reside that was washed with MeOH, then subjected to column chromatography (Al₂O₃, CH₂Cl₂/MeOH = 100:1) and then recrystallized from a mixture of CHCl₃/MeOH to give **L5**, as a purple powder: (684 mg, 0.26 µmol), in 53% yield.. ¹H NMR (400 MHz, CDCl₃) δ = 9.06 (s, 8H), 8.90 (s, 8H), 8.78 (d, *J* = 4.5 Hz, 8H), 8.75 (d, *J* = 8.0 Hz, 8H), 8.39 (d, *J* = 7.9 Hz, 8H), 8.10 (d, *J* = 8.0 Hz, 8H), 7.92 (t, *J* = 8.6 Hz, 8H), 7.86 (d, *J* = 7.7 Hz, 4H), 7.65 (d, *J* = 8.7 Hz, 4H), 7.60 (s, 4H), 7.38 (m, 8H), 4.29 (t, *J* = 6.2 Hz, 8H), 1.88 (m, 8H), 1.53 (m, 8H), 1.25 (m, 64H), 0.90 (t, *J* = 7.0 Hz, 12H); ¹³C NMR (125 MHz, CDCl₃) δ = 157.07, 156.66, 155.23, 149.18, 148.20, 142.68, 141.60, 140.13, 136.79, 135.19, 131.25, 127.65, 125.52, 123.61, 121.92, 121.29, 119.96, 119.85, 111.39, 68.89, 31.95, 29.69, 29.66, 29.53, 29.49, 29.39, 26.32, 22.72, 14.16.

3. NMR and MS spectra of compounds

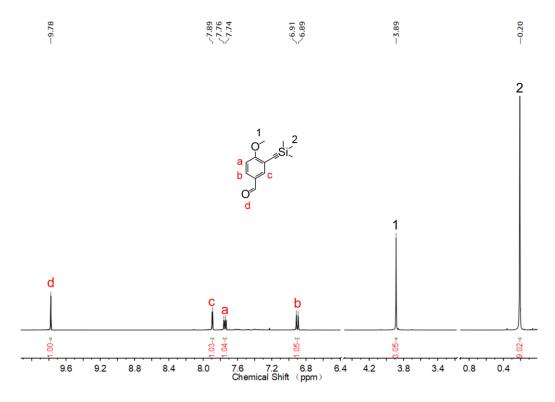


Figure S1. ¹H NMR spectrum of compound 1 in CDCl₃.

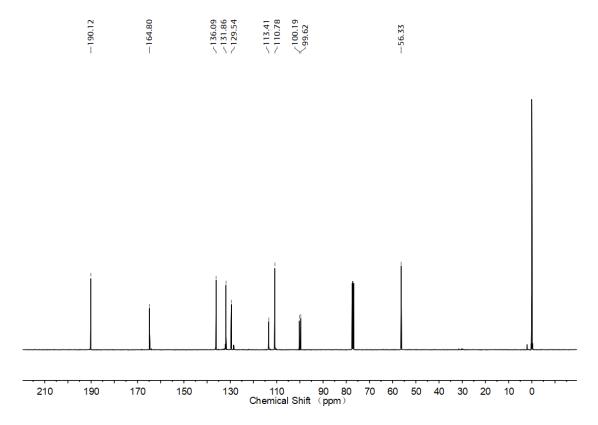


Figure S2. ¹³C NMR spectrum of compound 1 in CDCl₃.

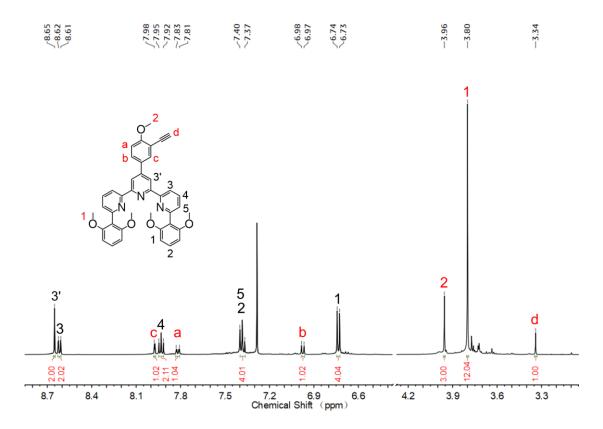


Figure S3. ¹H NMR spectrum of compound 3 in CDCl₃.

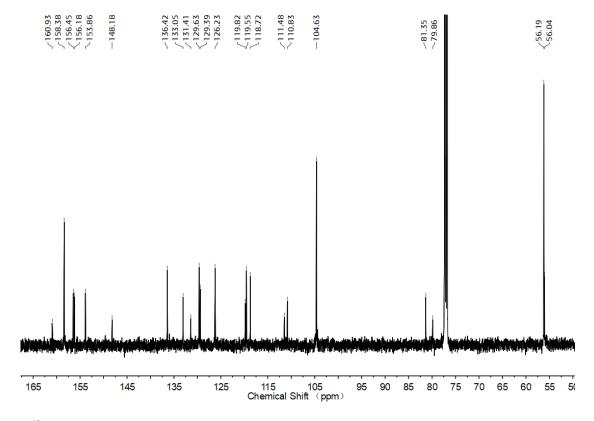


Figure S4. ¹³C NMR spectrum of compound 3 in CDCl₃.

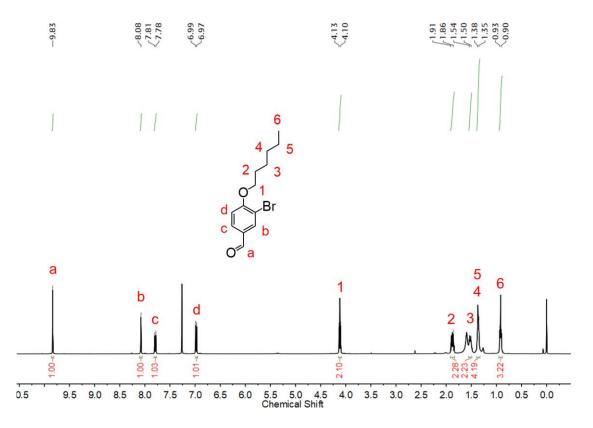


Figure S5. ¹H NMR spectrum of compound 4 in CDCl₃.

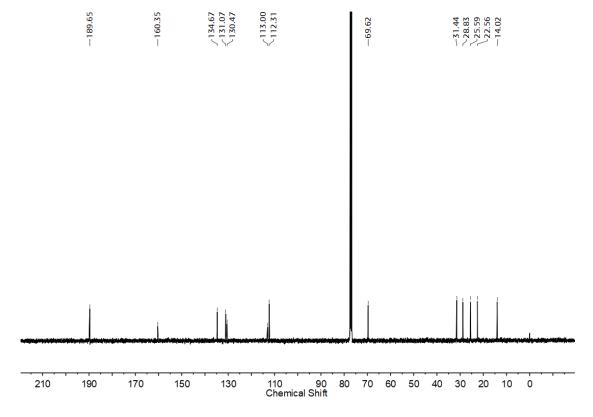


Figure S6. ¹³C NMR spectrum of compound 4 in CDCl₃.

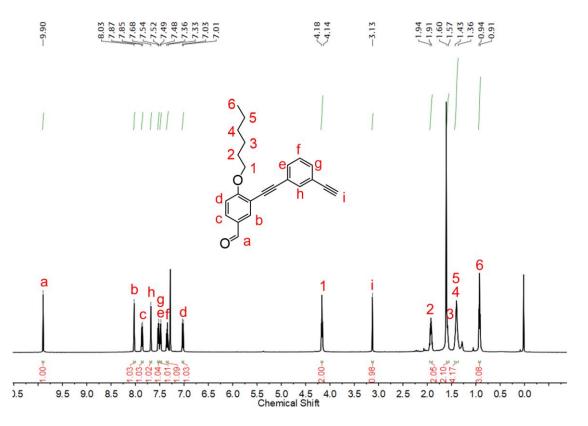


Figure S7. ¹H NMR spectrum of compound 6 in CDCl₃.

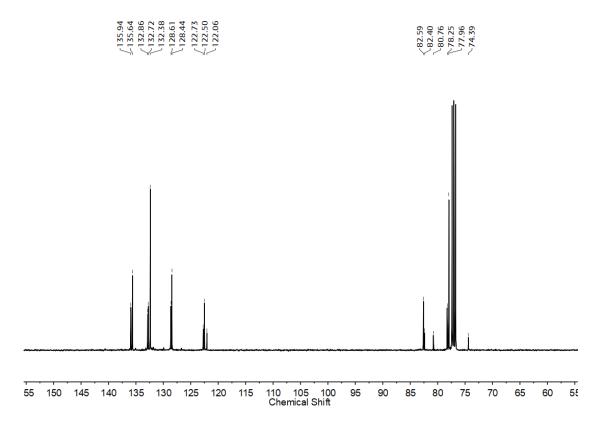


Figure S8. ¹³C NMR spectrum of compound 6 in CDCl₃.

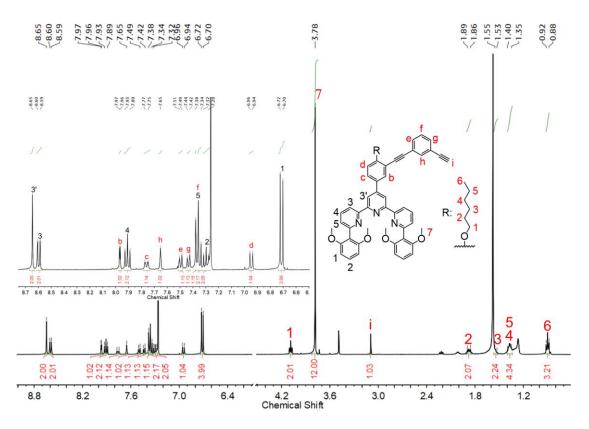


Figure S9. ¹H NMR spectrum of compound 7 in CDCl₃.

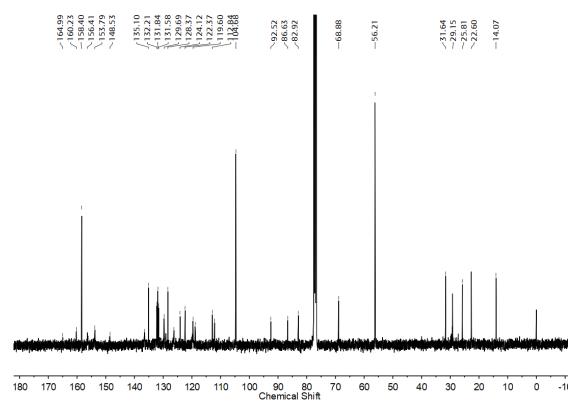


Figure S10. ¹³C NMR spectrum of compound 7 in CDCl₃.

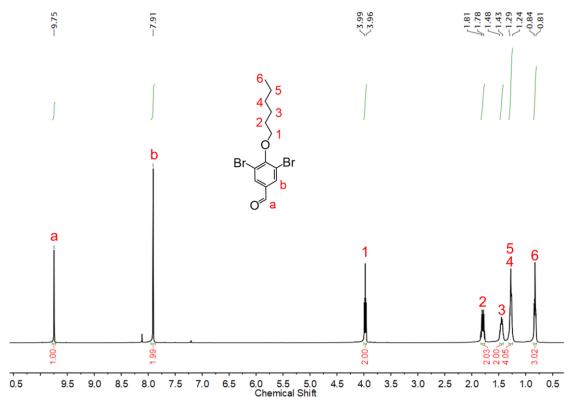


Figure S11. ¹H NMR spectrum of compound 8 in CDCl₃.

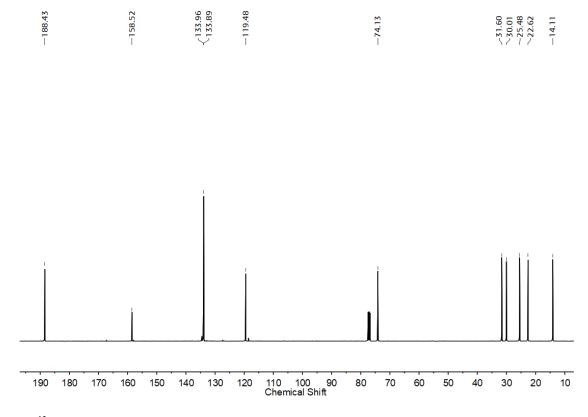


Figure S12. ¹³C NMR spectrum of compound 8 in CDCl₃.

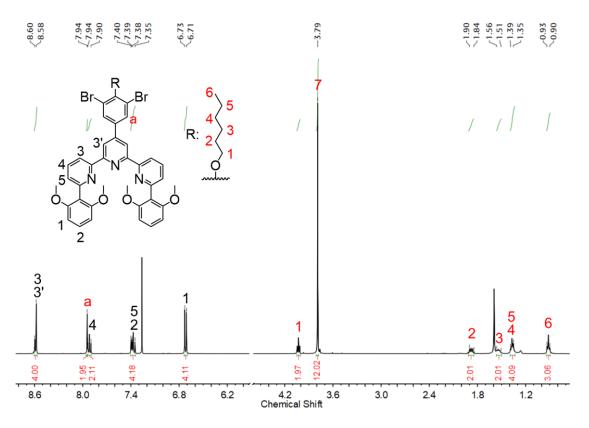


Figure S13. ¹H NMR spectrum of compound 9 in CDCl₃.

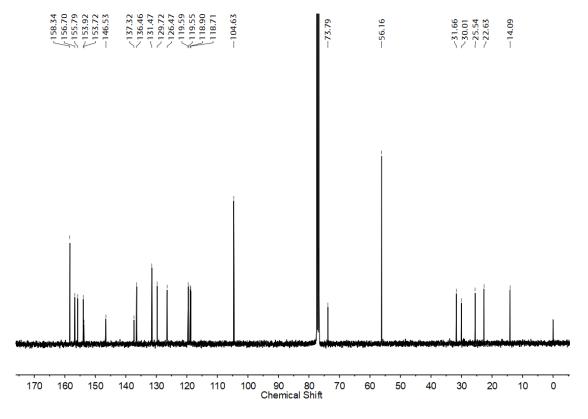


Figure S14. ¹³C NMR spectrum of compound 9 in CDCl₃.

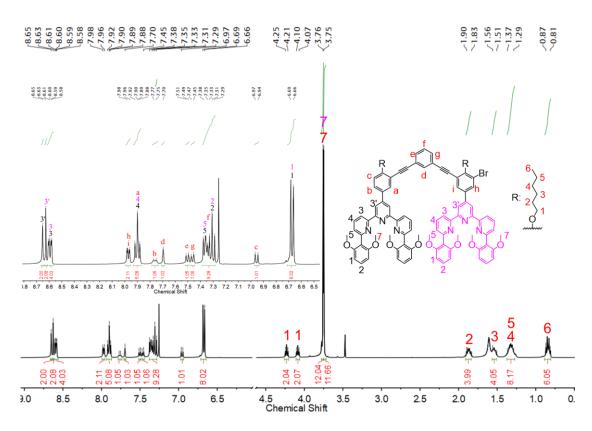


Figure S15. ¹H NMR spectrum of compound 10 in CDCl₃.

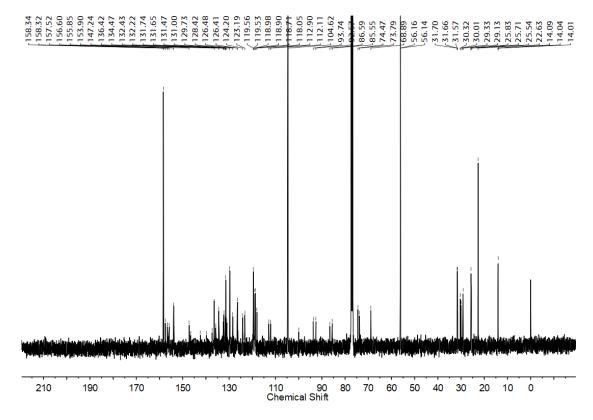


Figure S16. ¹³C NMR spectrum of compound 10 in CDCl₃.

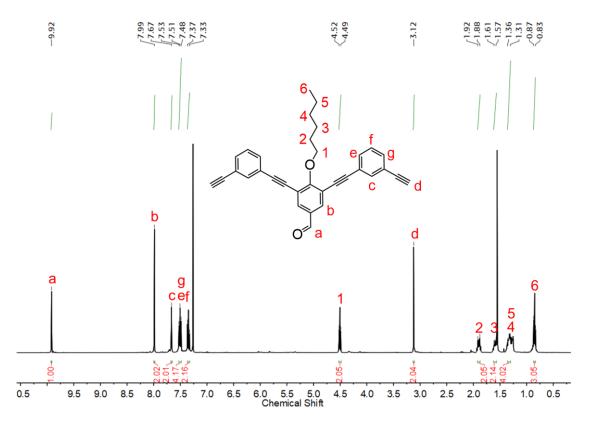


Figure S17. ¹H NMR spectrum of compound 11 in CDCl₃.

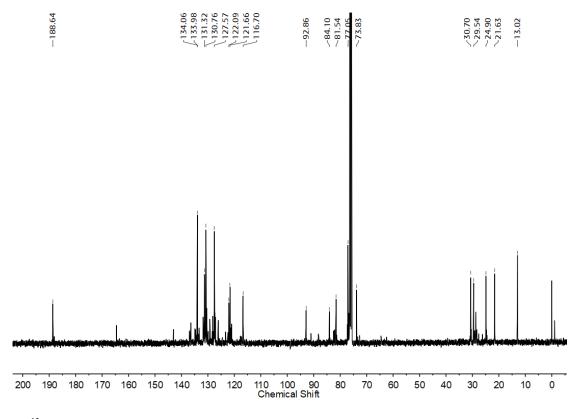


Figure S18. ¹³C NMR spectrum of compound 11 in CDCl₃.

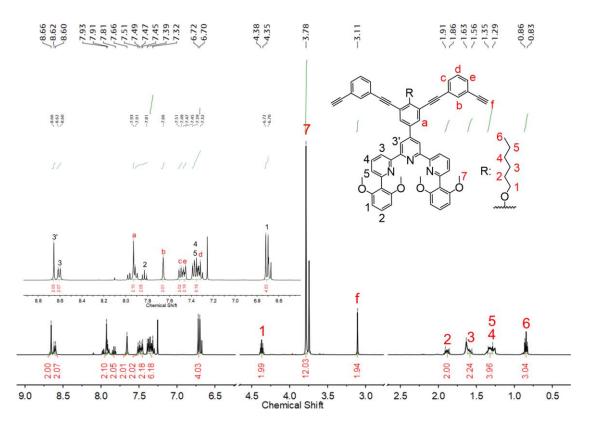


Figure S19. ¹H NMR spectrum of compound 12 in CDCl₃.

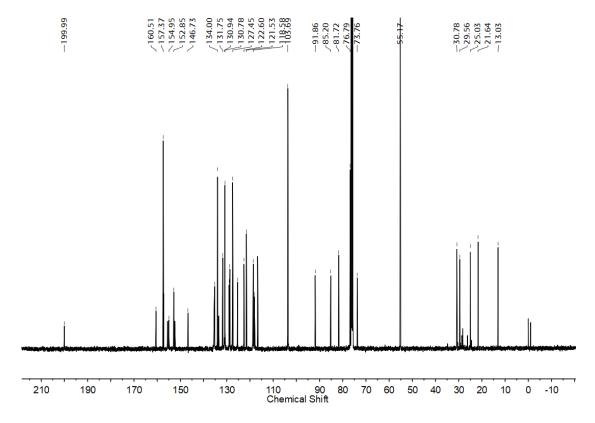


Figure S20. ¹³C NMR spectrum of compound 12 in CDCl₃.

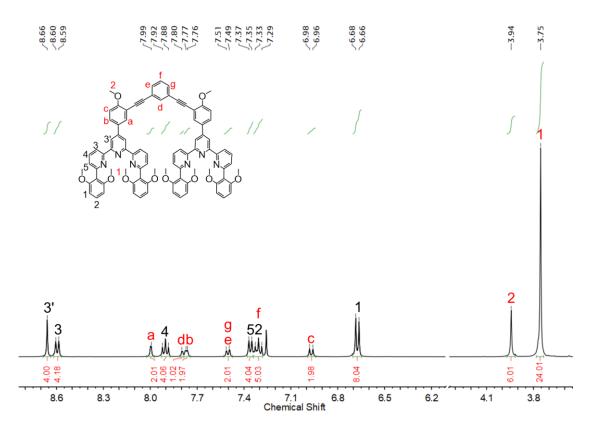


Figure S21. ¹H NMR spectrum of compound L1 in CDCl₃.

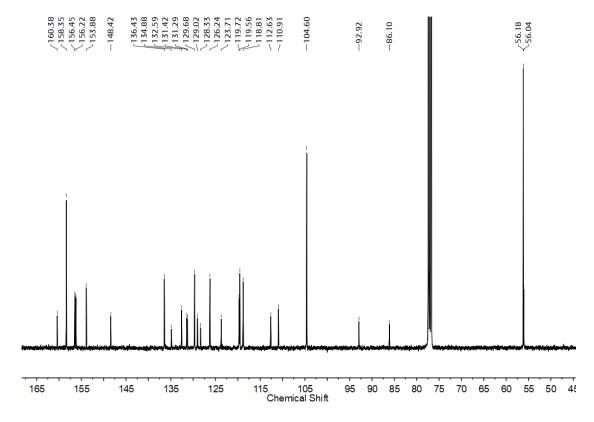


Figure S22. ¹³C NMR spectrum of compound L1 in CDCl₃.

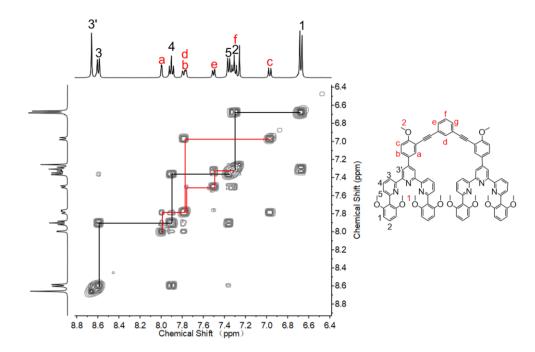


Figure S23. COSY spectrum of compound L1 in CDCl₃.

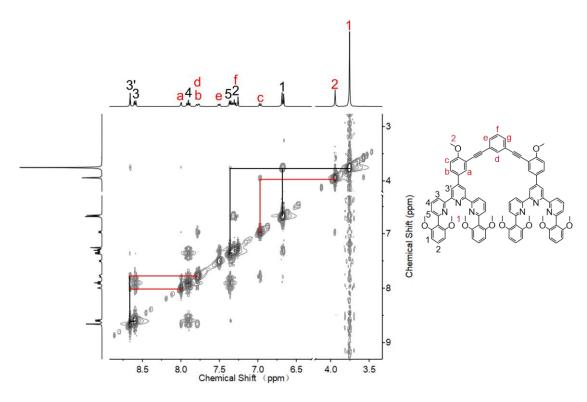


Figure S24. NOESY spectrum of compound L1 in CDCl₃.

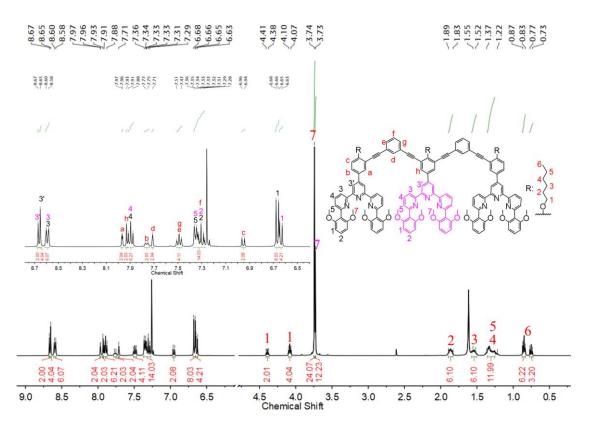


Figure S25. ¹H NMR spectrum of compound L2 in CDCl₃.

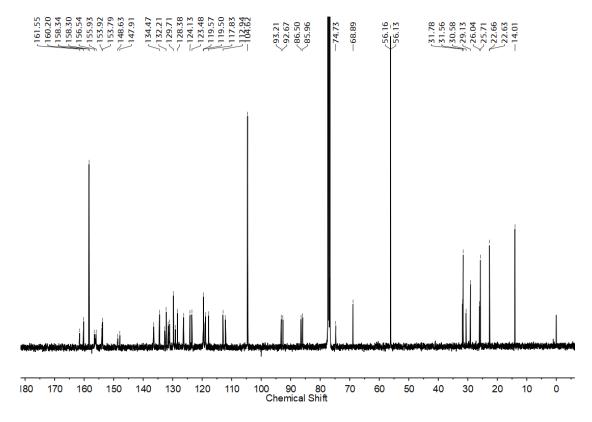


Figure S26. ¹³C NMR spectrum of compound L2 in CDCl₃.

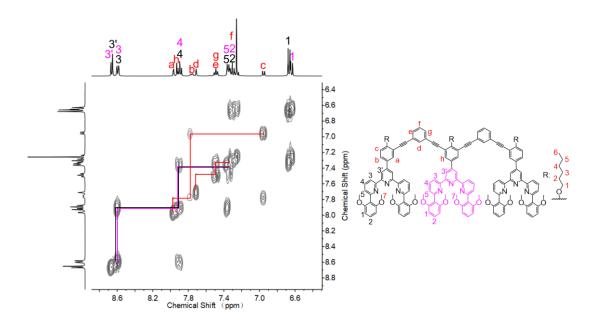


Figure S27. COSY spectrum of compound L2 in CDCl₃.

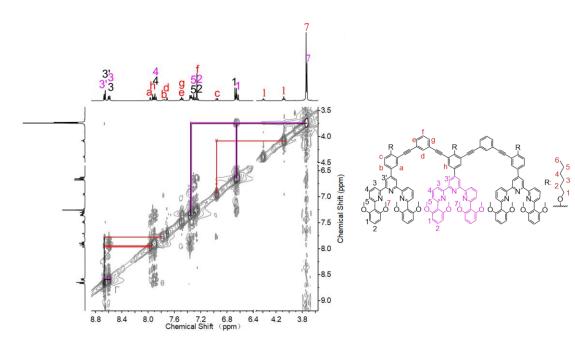


Figure S28. NOESY spectrum of compound L2 in CD₃CN.

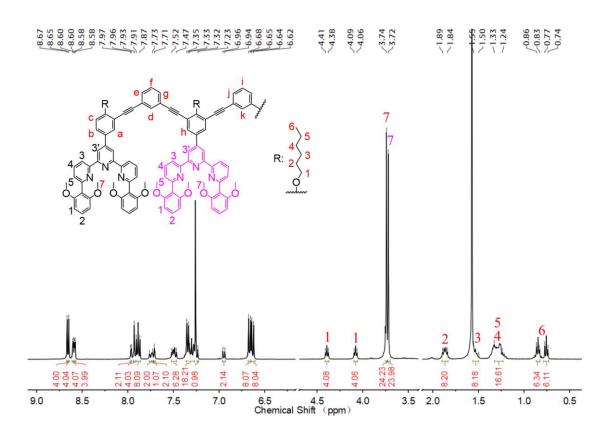


Figure S29. ¹H NMR spectrum of compound L3 in CDCl₃.

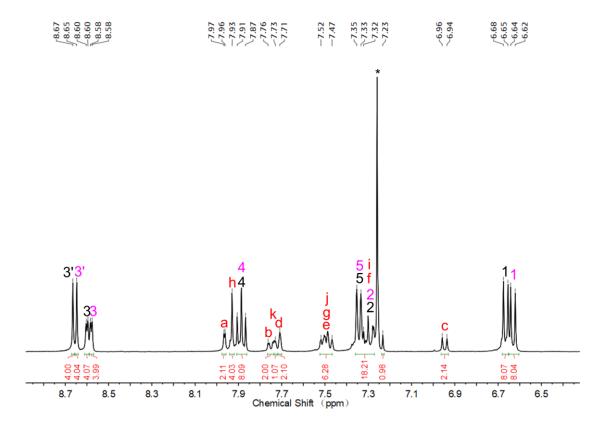


Figure S30. ¹H NMR spectrum of compound L3 in CDCl₃.

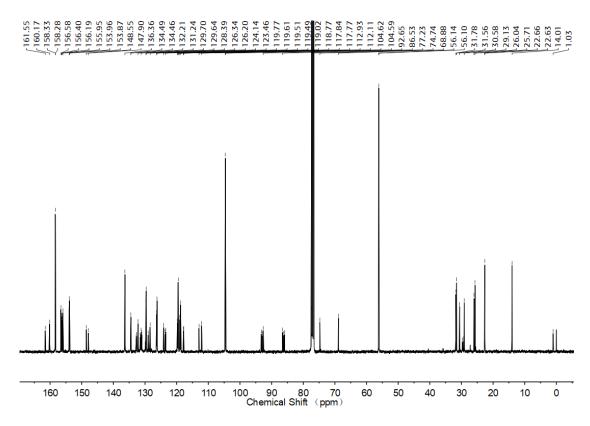


Figure S31. ¹³C NMR spectrum of compound L3 in CDCl₃.

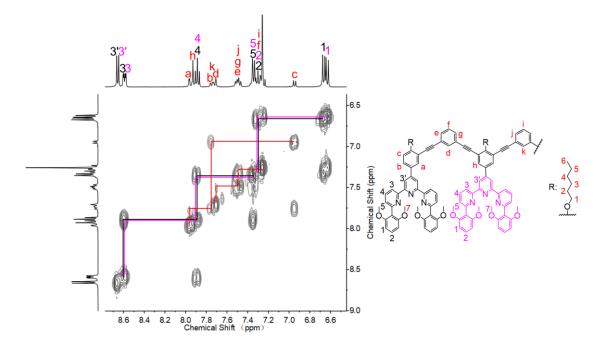
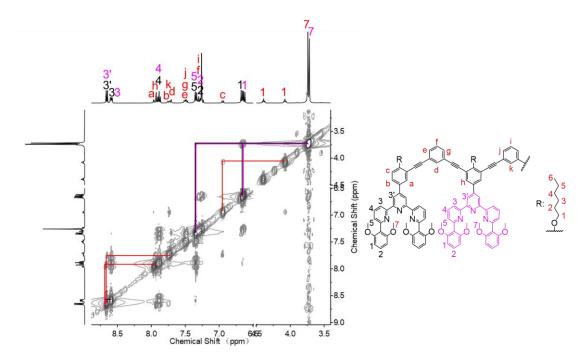
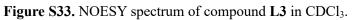


Figure S32. COSY spectrum of compound L3 in CDCl₃.





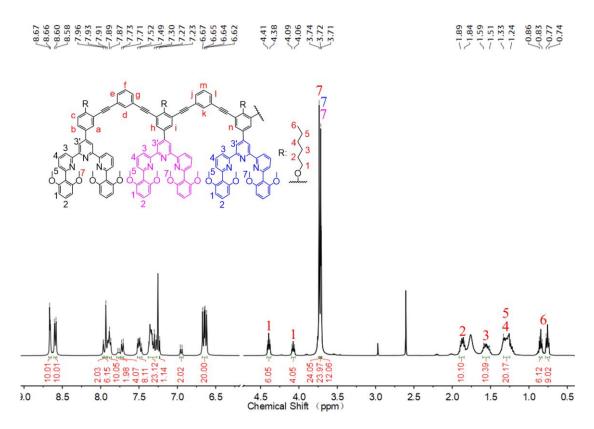


Figure S34. ¹H NMR spectrum of compound L4 in CDCl₃.



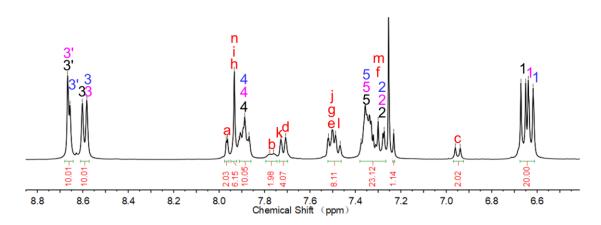


Figure S35. ¹H NMR spectrum of compound L4 in CDCl₃.

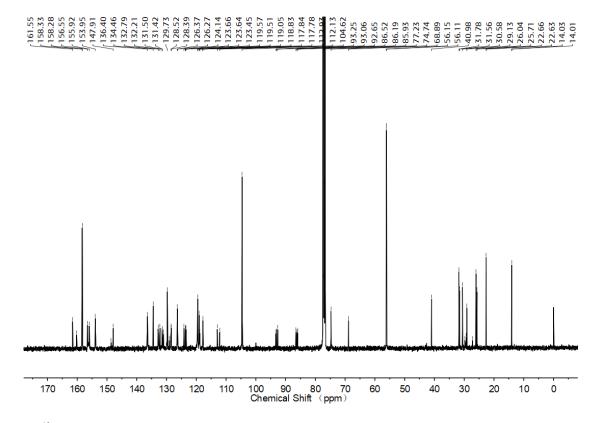


Figure S36. ¹³C NMR spectrum of compound L4 in CDCl₃.

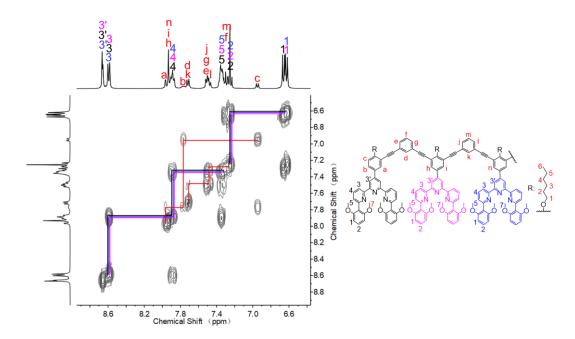


Figure S37. COSY spectrum of compound L4 in CDCl₃.

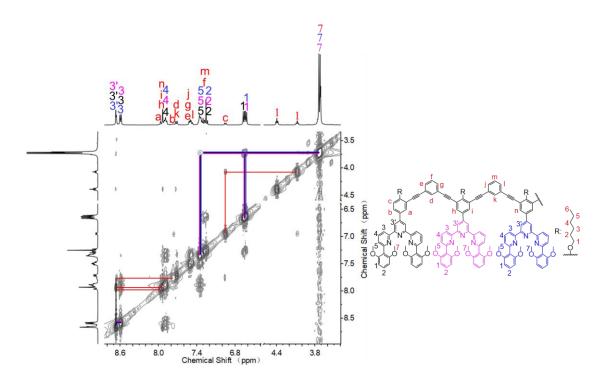


Figure S38. NOESY spectrum of compound L4 in CDCl₃.

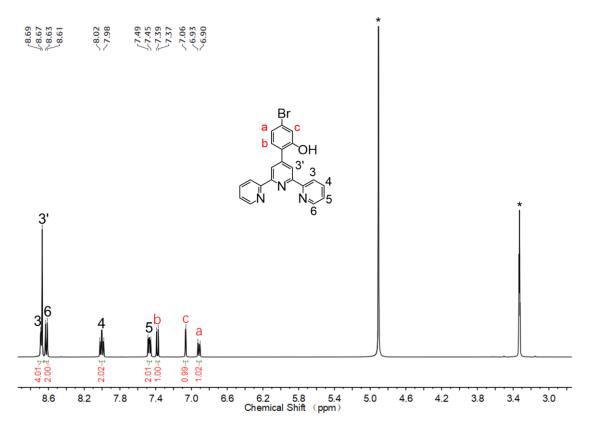


Figure S39. ¹H NMR spectrum of compound 13 in CDCl₃.

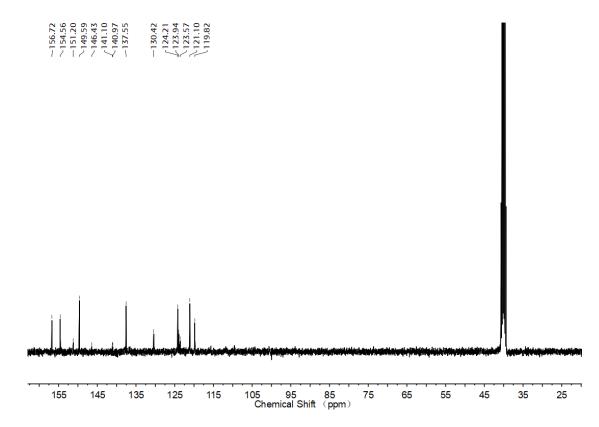
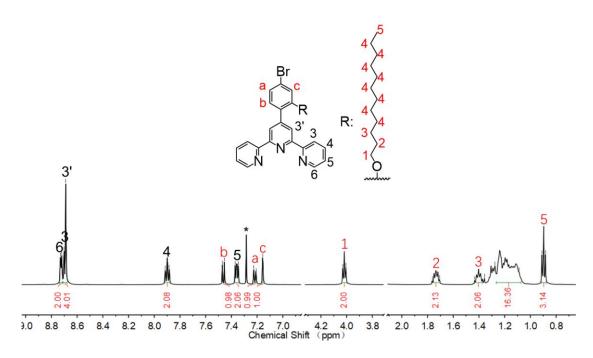
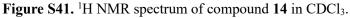


Figure S40. ¹³C NMR spectrum of compound 13 in CDCl₃.







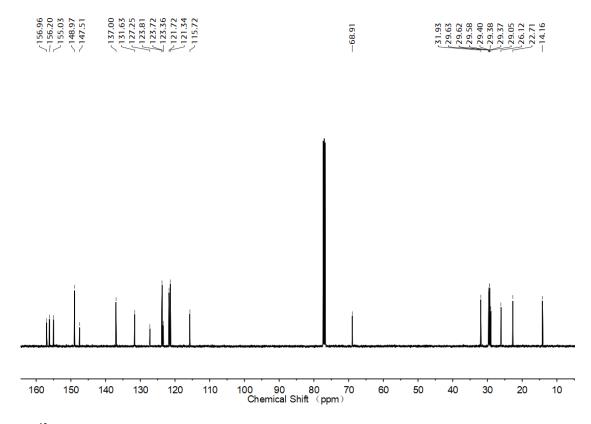


Figure S42. ¹³C NMR spectrum of compound 14 in CDCl₃.

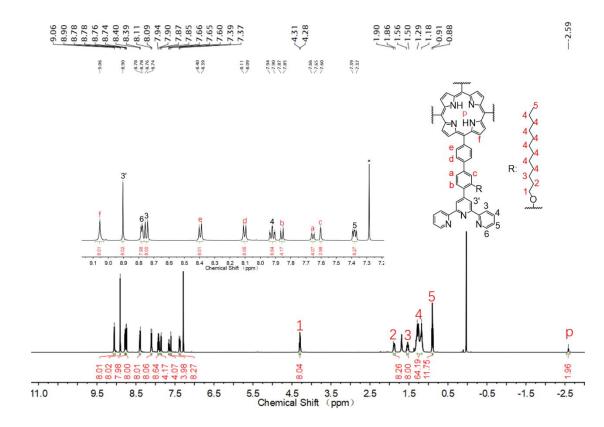


Figure S43. ¹H NMR spectrum of compound L5 in CDCl₃.

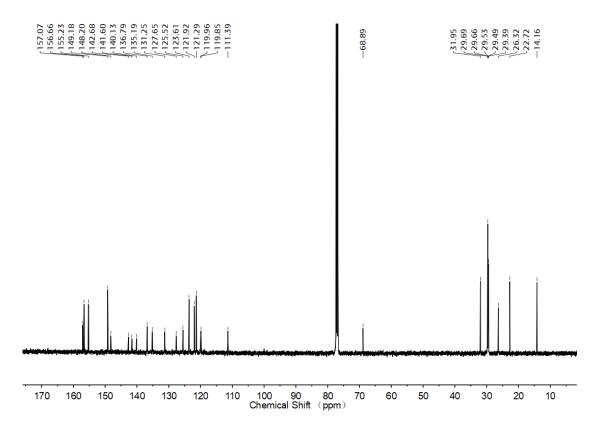


Figure S44. ¹³C NMR spectrum of compound L5 in CDCl₃.

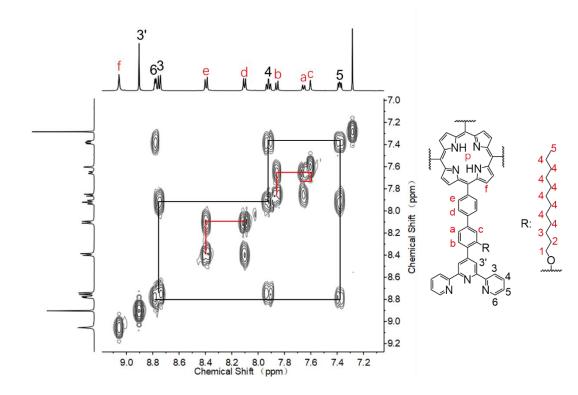


Figure S45. COSY spectrum of compound L5 in CDCl₃.

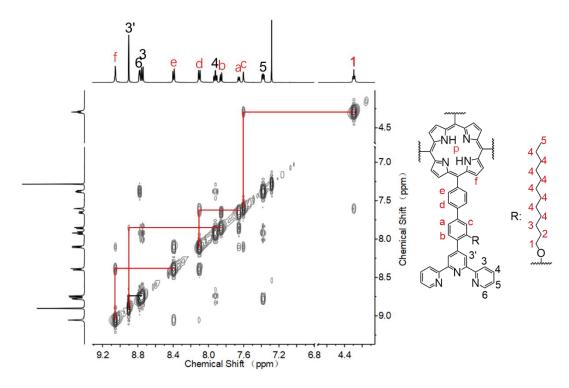


Figure S46. NOESY spectrum of compound L5 in CDCl₃.

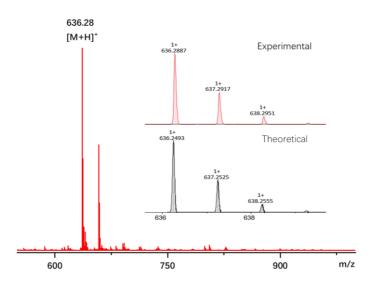


Figure S47. ESI-MS spectrum of compound 3.

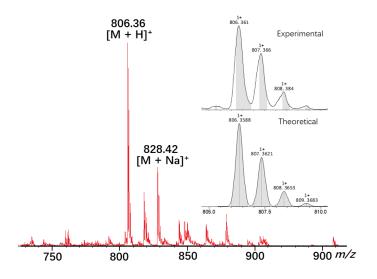


Figure S48. ESI-MS spectrum of compound 7.

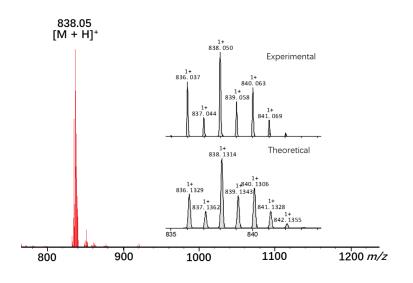


Figure S49. ESI-MS spectrum of compound 9.

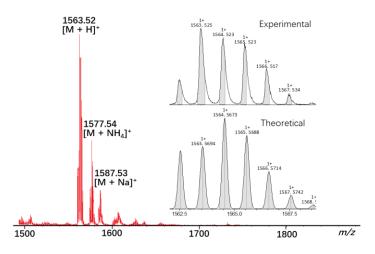


Figure S50. ESI-MS spectrum of compound 10.

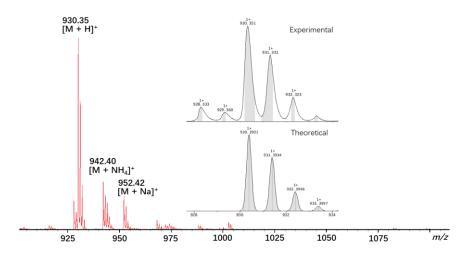


Figure S51. ESI-MS spectrum of compound 12.

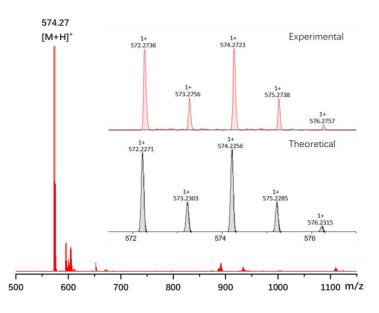


Figure S52. ESI-MS spectrum of compound 13.

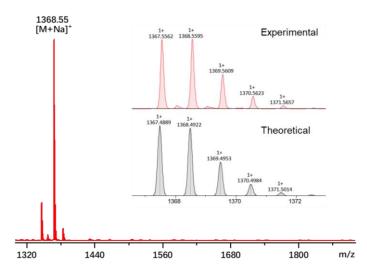


Figure S53. ESI-MS spectrum of compound L1.

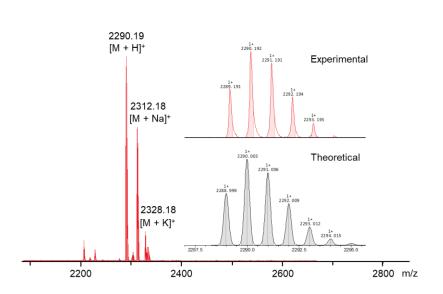


Figure S54. ESI-MS spectrum of compound L2.

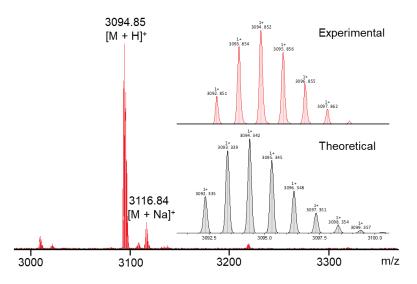


Figure S55. ESI-MS spectrum of compound L3.

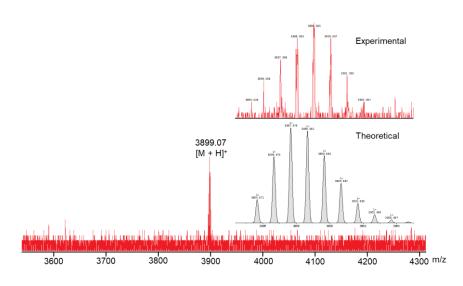


Figure S56. ESI-MS spectrum of compound L4.

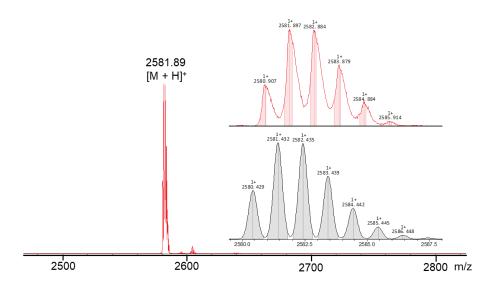
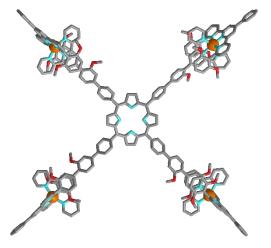


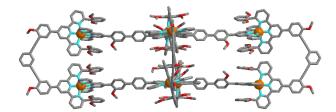
Figure S57. ESI-MS spectrum of compound L5.

4. Self-assembly of G0-G4.

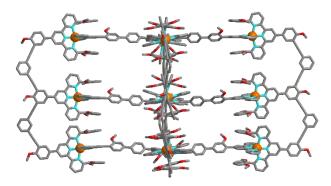
Dissolve ligands in 20ml of a mixed solvent of $CH_2Cl_2/MeOH = 2:1$, a solution of $Cd(NO_3)_2 \cdot 4H_2O$ in CH_3OH (2 mL) was added. After being stirred at 85 °C for 24 h, excess NH_4PF_6 was added into the solution to precipitate the complex, which was filtered, washed with H_2O , and then dried *in vacuo*.



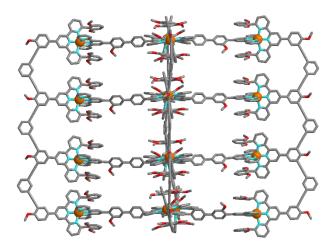
¹H NMR (400 MHz, CD₃CN) δ = 9.09 (s, 8H), 8.95 (s, 8H), 8.67-8.65 (d, 8H), 8.53 (s, 8H), 8.48-8.46 (d, 8H), 8.41-8.39 (d, 8H), 8.33-8.31 (d, 8H), 8.17-8.07 (m, 24H), 8.01-7.99 (d, 4H), 7.95-7.89 (m, 16H), 7.72-7.66 (m, 12H), 7.46-7.43 (t, 8H), 7.17-7.15 (d, 8H), 6.87-6.83 (t, 8H), 5.87-5.85 (d, 16H), 4.56-4.53 (t, 8H), 2.84 (s, 48H), 1.66-1.62 (m, 8H), 1.44-1.40 (m, 8H), 1.32-1.13 (m, 64H), 0.73-0.70 (m, 12H), -2.70 (s, 2H). ESI-MS (*m/z*): 669.79 [M-8 PF₆⁻]⁸⁺ (calcd. *m/z* = 669.75), 786.19 [M-7 PF₆⁻]⁷⁺ (calcd. *m/z* = 786.14), 941.4 [M-6 PF₆⁻]⁶⁺ (calcd. *m/z* = 941.5), 1158.7 [M-5 PF₆⁻]⁵⁺ (calcd. *m/z* = 1158.8).



¹H NMR (400 MHz, CD₃CN) δ = 9.09 (s, 16H), 9.03 (s, 16H), 8.78-8.76 (d, 16H), 8.60 (s, 8H), 8.53 (s, 16H), 8.49-8.47 (d, 8H), 8.44-8.40 (t, 24H), 8.36-8.32 (m, 24H), 8.16-8.13 (t, 16H), 8.02-7.94 (m, 56H), 7.74-7.72 (d, 12H), 7.65-7.61 (t, 4H), 7.50-7.46 (m, 24H), 6.95-6.94 (d, 16H), 6.88-6.84 (t, 16H), 5.83-5.81 (d, 32H), 4.63-4.60 (t, 16H), 4.17 (s, 24H), 2.77 (s, 96H), 1.74-1.69 (m, 24H), 1.52-1.47 (m, 24H), 1.41-1.16 (m, 128H), 0.75-0.72 (t, 24H), -2.93 (s, 4H). ESI-MS (*m*/*z*): 1106.35 [M-16 PF₆-]¹¹⁺ (calcd. *m*/*z* = 1106.35), 1231.49 [M-15 PF₆-]¹⁰⁺ (calcd. *m*/*z* = 1231.49), 1384.32 [M-14 PF₆-]⁹⁺ (calcd. *m*/*z* = 1384.32), 1575.48 [M-13 PF₆-]⁸⁺ (calcd. *m*/*z* = 1575.48), 1821.26 [M-12 PF₆-]⁷⁺ (calcd. *m*/*z* = 1821.25), 2148.96 [M-11 PF₆-]⁶⁺ (calcd. *m*/*z* = 2148.99), 2607.94 [M-10 PF₆-]⁵⁺ (calcd. *m*/*z* = 2609.96).

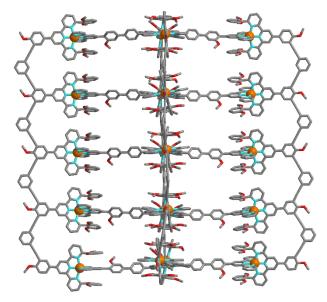


¹H NMR (400 MHz, CD₃CN) δ = 9.17 (s, 8H), 9.07 (s, 16H), 8.96 (s, 16H), 8.79 (s, 8H), 8.80-8.78 (d, 8H), 8.77-8.74 (d, 16H), 8.67 (s, 8H), 8.58 (s, 8H), 8.47 (s, 24H), 8.40-8.35 (m, 40H), 8.29-8.20 (m, 36H), 8.14-8.09 (m, 36H), 7.98-7.86 (m, 76H), 7.82-7.72 (m, 28H), 7.67-7.63 (m, 12H), 7.48-7.43(m, 32H), 6.88-6.87 (d, 16H), 6.83-6.79 (t, 16H), 6.76-6.72 (t, 8H), 6.54-6.52 (d, 8H), 5.78-5.75 (d, 32H), 5.64-5.62 (d, 16H), 4.75-4.72 (t, 8H), 4.62-4.51 (t, 24H), 4.38-4.35 (t, 16H), 2.72 (s, 96H), 2.57 (s, 48H), 2.09-2.01 (m, 48H), 1.77-1.64 (m, 48H), 1.58-1.44 (m, 72H), 1.38-1.11 (m, 168H), 1.04-1.00 (m, 36H), 0.74-0.67 (m, 36H), -3.01 (s, 4H), -3.27 (s, 2H). ESI-MS (*m/z*): 1213.64 [M-16 PF₆⁻]¹⁶⁺ (calcd. *m/z* = 1213.22), 1304.28 [M-15 PF₆⁻]¹⁵⁺ (calcd. *m/z* = 1303.76), 1407.79 [M-14 PF₆⁻]¹⁴⁺ (calcd. *m/z* = 1407.25), 1527.16 [M-13 PF₆⁻]¹³⁺ (calcd. *m/z* = 1526.65), 1666.55 [M-12 PF₆⁻]¹²⁺ (calcd. *m/z* = 1665.96), 1831.28 [M-11 PF₆⁻]¹¹⁺ (calcd. *m/z* = 1830.59), 2028.91 [M-10 PF₆⁻]¹⁰⁺ (calcd. *m/z* = 2028.15), 2270.59 [M-9 PF₆⁻]⁹⁺ (calcd. *m/z* = 2269.61), 2572.41 [M-8 PF₆⁻]⁸⁺ (calcd. *m/z* = 2571.43), 2960.61 [M-7 PF₆⁻]⁷⁺ (calcd. *m/z* = 2959.50).



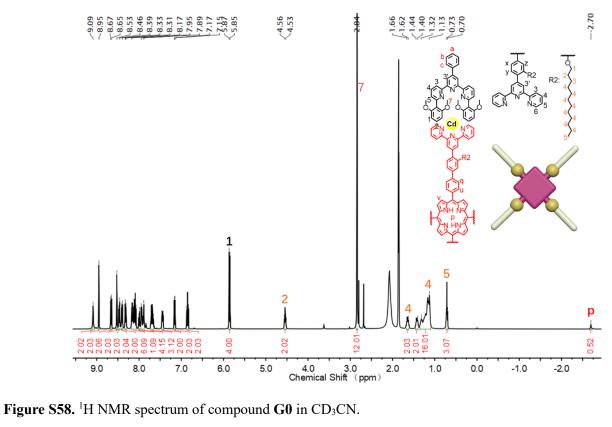
¹H NMR (400 MHz, CD₃CN) δ = 9.11 (s, 16H), 9.03 (s, 16H), 8.92 (s, 16H), 8.77-8.70 (m, 48H), 8.61 (s, 16H), 8.54 (s, 8H), 8.42 (s, 16H), 8.35-8.28 (m, 64H), 8.24-8.22 (d, 16H), 8.16-8.13 (m, 32H), 8.09-8.03 (m, 48H), 7.94-7.91 (m, 48H), 7.88-7.85 (m, 32H), 7.81-7.69 (m, 48H), 7.66-7.60 (m, 16H), 7.43-7.37 (m, 40H), 6.85-6.83 (d, 16H), 6.78-6.74 (t, 16H), 6.69-6.65 (t, 16H), 6.45-6.43 (d, 16H), 5.73-5.71 (d, 32H), 5.58-5.56 (d, 32H), 4.72-4.69 (t, 16H), 4.54-4.50 (t, 32H), 4.35-4.32 (t, 16H), 2.68 (s, 96H), 2.51 (s, 96H), 2.04-1.97 (m, 64H), 1.73-1.58 (m, 64H), 1.53-1.41 (m, 80H), 1.31-1.05 (m, 240H), 1.01-0.97 (m, 48H), 0.68-0.61 (m, 48H), -3.03 (s, 4H), -3.32 (s, 4H). ESI-MS (*m*/*z*): 1179.66 [M-22 PF₆⁻]²²⁺ (calcd. *m*/*z* = 1179.46), 1242.70 [M-21 PF₆⁻]²¹⁺ (calcd. *m*/*z* = 1242.53), 1312.12 [M-20 PF₆⁻]²⁰⁺ (calcd. *m*/*z* = 1311.91), 1388.84 [M-19 PF₆⁻]¹⁹⁺ (calcd. *m*/*z* = 1388.59), 1473.98 [M-18 PF₆⁻]¹⁸⁺ (calcd. *m*/*z* = 1473.79), 1569.27 [M-17 PF₆⁻]¹⁷⁺ (calcd. *m*/*z* = 1569.01), 1676.35 [M-16 PF₆⁻]¹⁶⁺ (calcd. *m*/*z*

= 1676.13), 1797.69 [M-15 PF₆⁻]¹⁵⁺ (calcd. m/z = 1797.54), 1936.71 [M-14 PF₆⁻]¹⁴⁺ (calcd. m/z = 1936.30), 2096.76 [M-13 PF₆⁻]¹³⁺ (calcd. m/z = 2096.40), 2283.66 [M-12 PF₆⁻]¹²⁺ (calcd. m/z = 2283.18), 2504.42 [M-11 PF₆⁻]¹¹⁺ (calcd. m/z = 2503.92), 2769.37 [M-10 PF₆⁻]¹⁰⁺ (calcd. m/z = 2768.81).



¹H NMR (400 MHz, CD₃CN) δ 9.11 (s, 24H), 9.03 (s, 16H), 8.91 (s, 16H), 8.75-8.71 (m, 64H), 8.62 (s, 24H), 8.54 (s, 8H), 8.41 (s, 16H), 8.35-8.27 (m, 72H), 8.23-8.21 (m, 20H), 8.16-8.12 (m, 40H), 8.07-8.02 (m, 60H), 7.93-7.84 (m, 116H), 7.79-7.59 (m, 88H), 7.41-7.38 (m, 48H), 6.84-6.82 (d, 16H), 6.77-6.73 (t, 16H), 6.66-6.63 (t, 24H), 6.43-6.41 (d, 16H), 6.37-6.36 (d, 8H), 5.72-5.70 (d, 32H), 5.55-5.51 (m, 48H), 4.72-4.69 (t, 24H), 4.53-4.46 (m, 40H), 4.35-4.30 (t, 16H), 2.67 (s, 96H), 2.50 (s, 96H), 2.47 (s, 48H), 1.87-1.70 (m, 80H), 1.68-1.58 (m, 80H), 1.31-1.06 (m, 360H), 1.00-0.97 (m, 60H), 0.67-0.59 (m, 60H), -3.04(s, 4H), -3.35(s, 6H). ESI-MS (*m/z*): 1516.28 [M-22 PF₆-]²²⁺ (calcd. *m/z* = 1516.13), 1595.35 [M-21 PF₆-]²¹⁺ (calcd. *m/z* = 1595.23), 1682.47 [M-20 PF₆-]²⁰⁺ (calcd. *m/z* = 1682.24), 1778.69 [M-19 PF₆-]¹⁹⁺ (calcd. *m/z* = 1778.41), 1885.41 [M-18 PF₆-]¹⁸⁺ (calcd. *m/z* = 1885.27), 2004.79 [M-17 PF₆-]¹⁷⁺ (calcd. *m/z* = 2004.69), 2139.33 [M-16 PF₆-]¹⁶⁺ (calcd. *m/z* = 2139.65), 2291.37 [M-15 PF₆-]¹⁵⁺ (calcd. *m/z* = 2291.32), 2465.53 [M-14 PF₆-]¹⁴⁺ (calcd. *m/z* = 2465.34), 2666.41 [M-13 PF₆-]¹³⁺ (calcd. *m/z* = 2666.14), 2900.82 [M-12 PF₆-]¹²⁺ (calcd. *m/z* = 2900.40).

5. NMR, MS, TWIM-MS spectra, AFM and TEM of G0-G4.



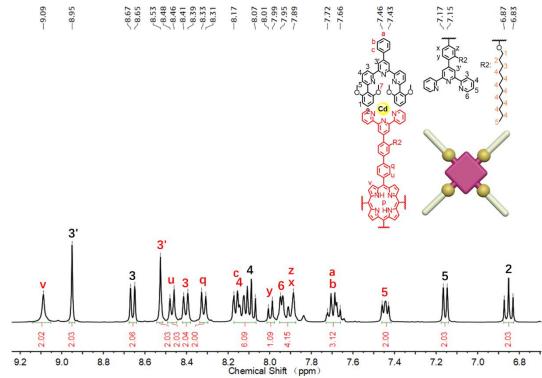


Figure S59. ¹H NMR spectrum of compound G0 in CD₃CN.

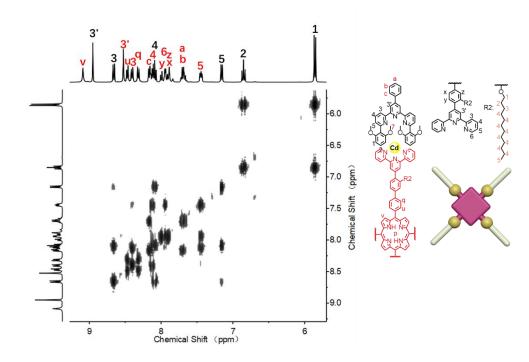


Figure S60. COSY spectrum of compound G0 in CD₃CN.

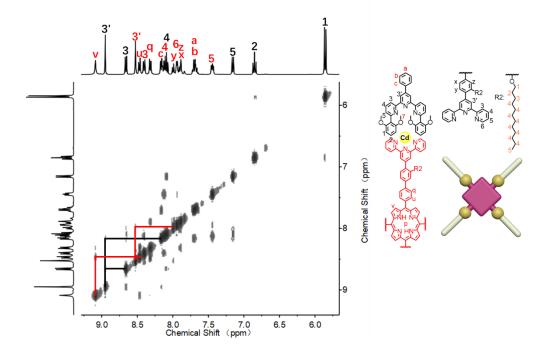


Figure S61. NOESY spectrum of compound G0 in CD₃CN.

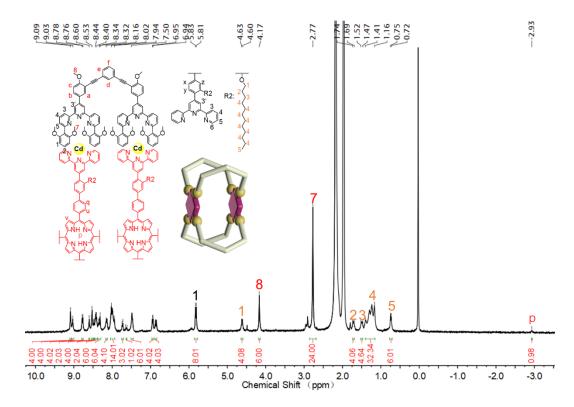


Figure S62. ¹H NMR spectrum of compound G1 in CD₃CN.

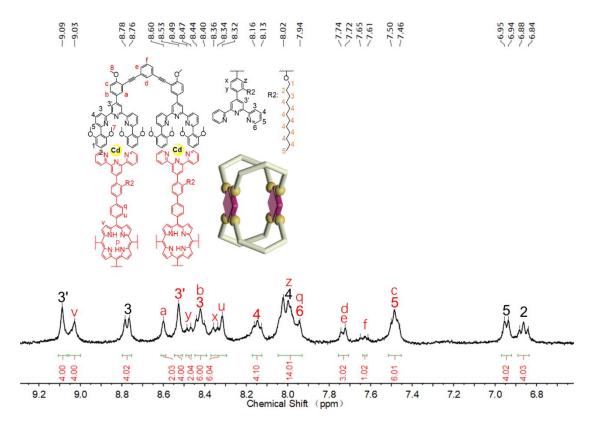


Figure S63. ¹H NMR spectrum of compound G1 in CD₃CN.

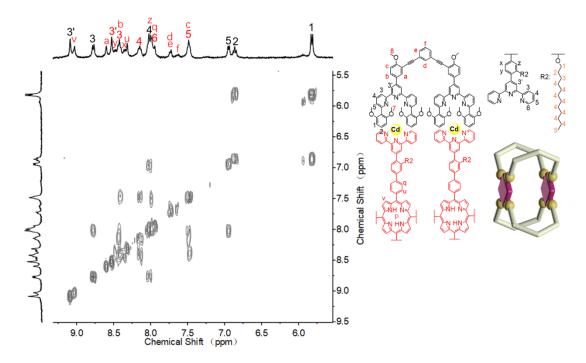


Figure S64. COSY spectrum of compound G1 in CD₃CN.

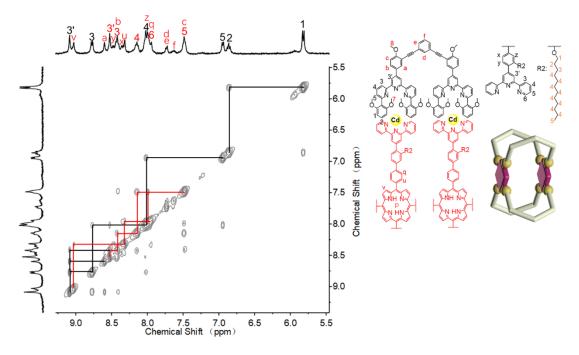


Figure S65. NOESY spectrum of compound G1 in CD₃CN.

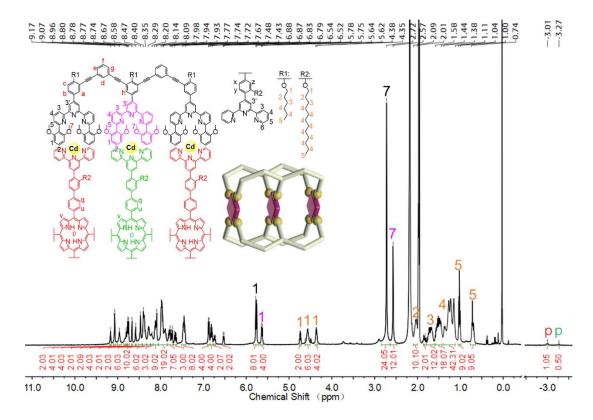


Figure S66. ¹H NMR spectrum of compound G2 in CD₃CN.

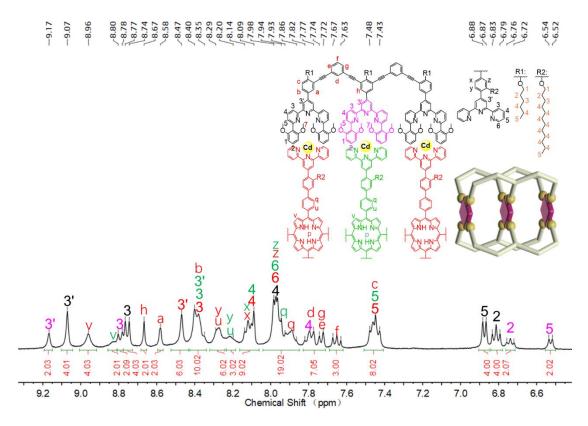


Figure S67. ¹H NMR spectrum of compound G2 in CD₃CN.

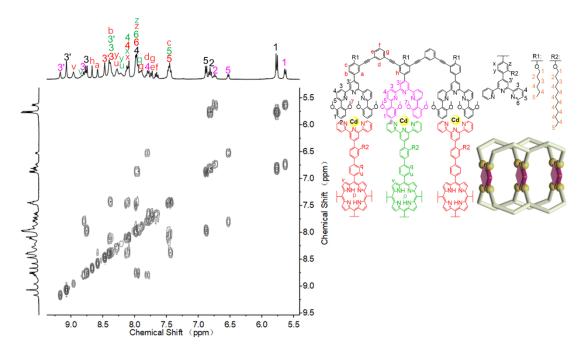


Figure S68. COSY spectrum of compound G2 in CD₃CN.

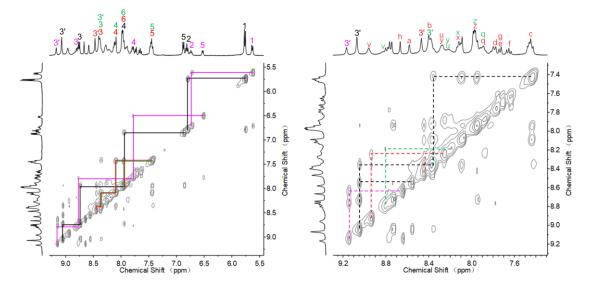


Figure S69. NOESY spectrum of compound G2 in CD₃CN.

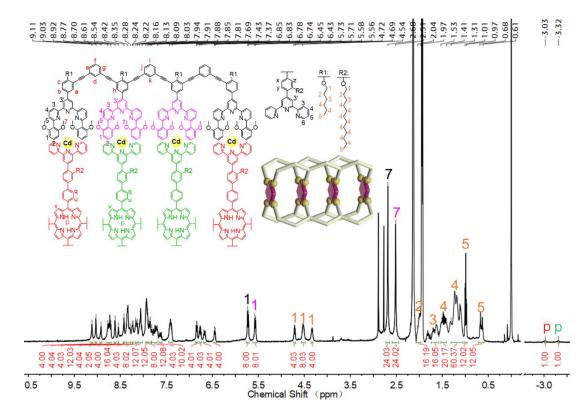


Figure S70. ¹H NMR spectrum of compound G3 in CD₃CN.

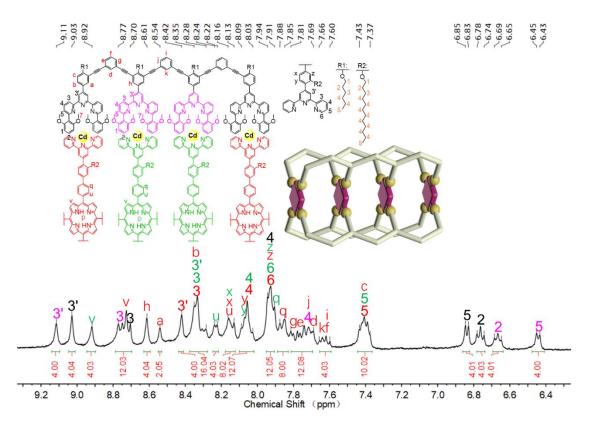


Figure S71. ¹H NMR spectrum of compound G3 in CD₃CN.

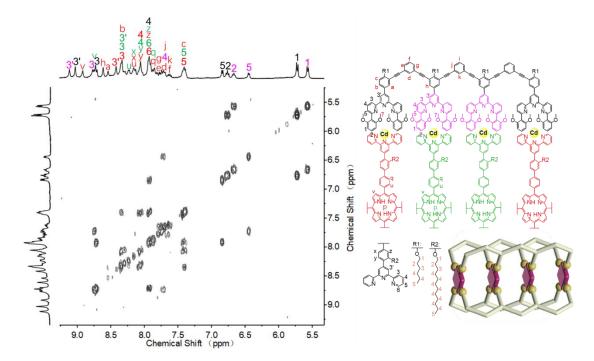


Figure S72. COSY spectrum of compound G3 in CD₃CN.

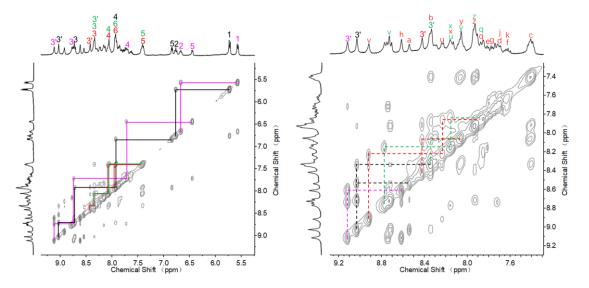


Figure S73. NOESY spectrum of compound G3 in CD₃CN.

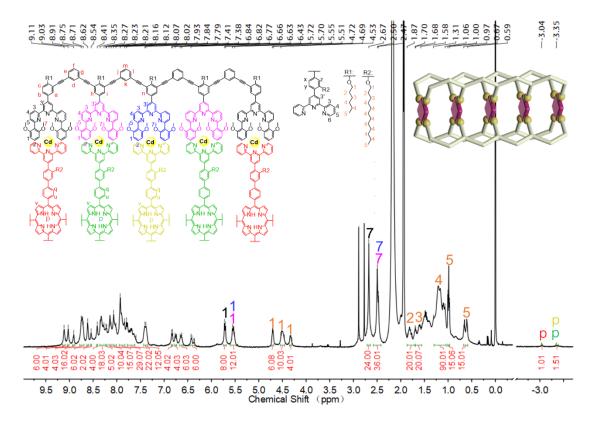


Figure S74. ¹H NMR spectrum of compound G4 in CD₃CN.

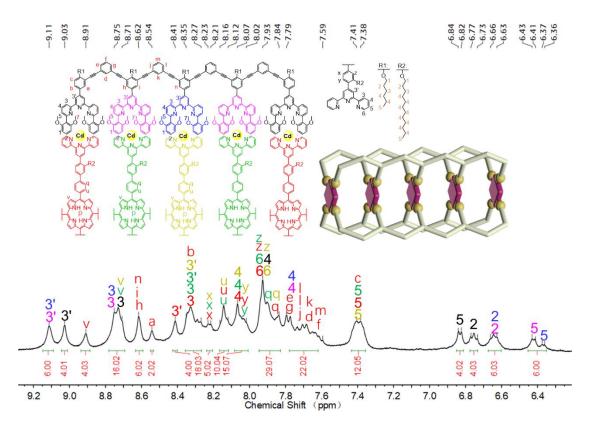


Figure S75. ¹H NMR spectrum of compound G4 in CD₃CN.

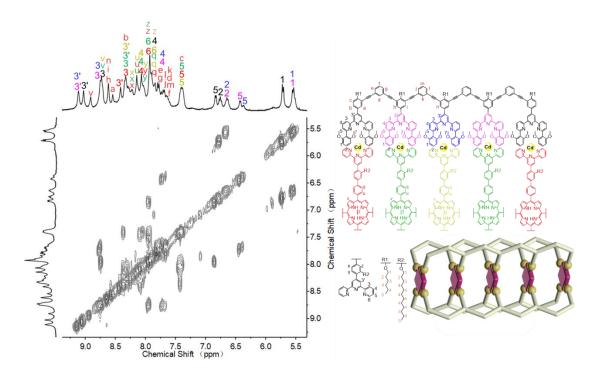


Figure S76. COSY spectrum of compound G4 in CD₃CN.

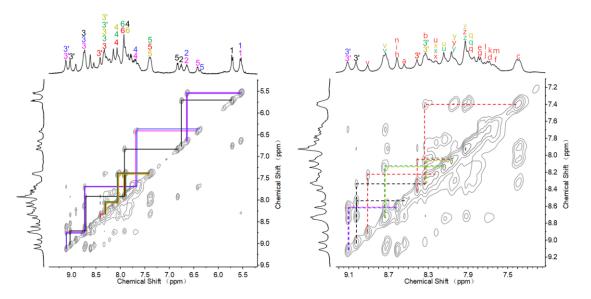


Figure S77. NOESY spectrum of compound G4 in CD₃CN.

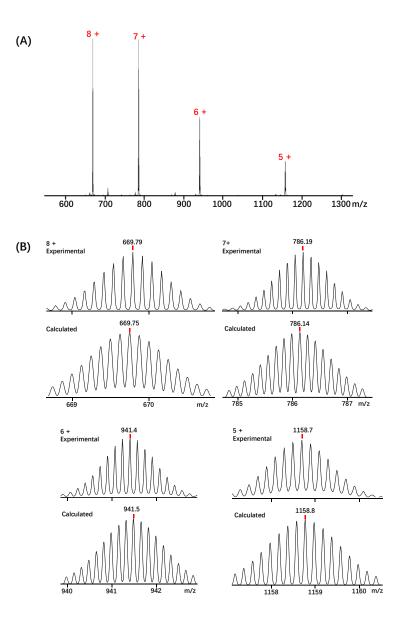


Figure S78: (A) ESI-MS spectrum of **G0**, (B) Calculated (bottom) and experimental (top) isotope patterns for the different charge states observed from **G0** (PF₆⁻ as counterion).

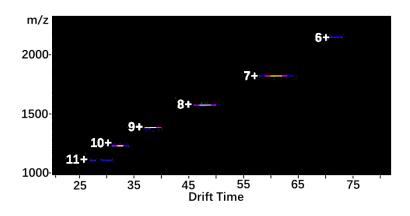


Figure S79. ESI-TWIM-MS plot of G1.

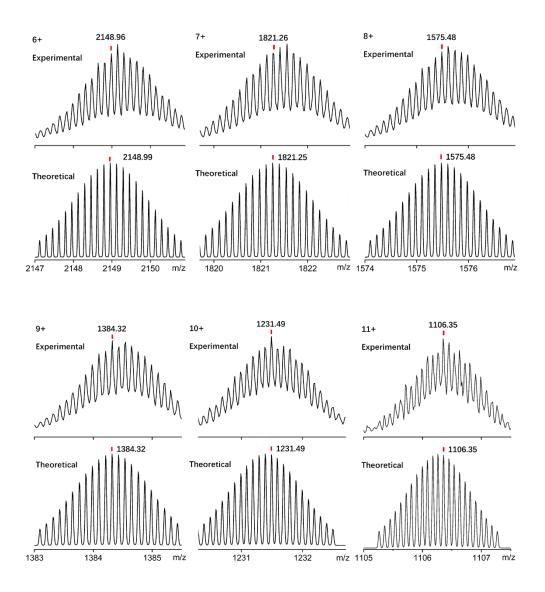


Figure S80. Theoretical (bottom) and experimental (top) isotope patterns for the different charge states observed from $G1(PF_6^-$ as counterion).

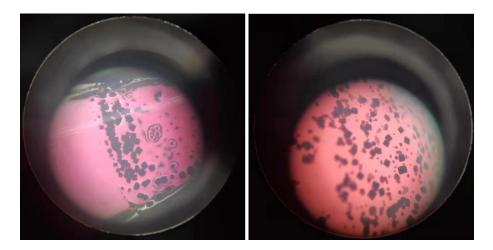


Figure S81. The image of G1 and C_{60} (@G1 crystals grown under slow vapor diffusion (10 days) of ethyl acetate into a DMF solution.

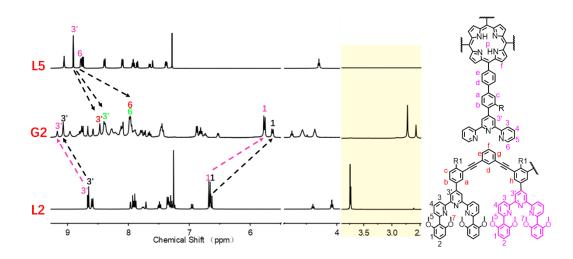


Figure S82. Comparison diagram of ¹H NMR spectra of G2 and its module ligands L2 and L5.

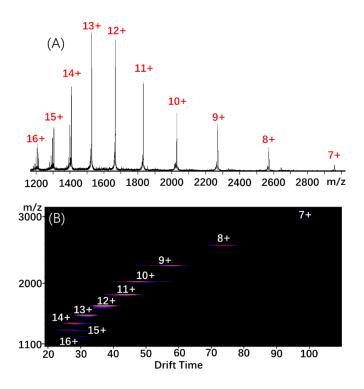


Figure S83. ESI-MS spectrum and ESI-TWIM-MS plot of G2.

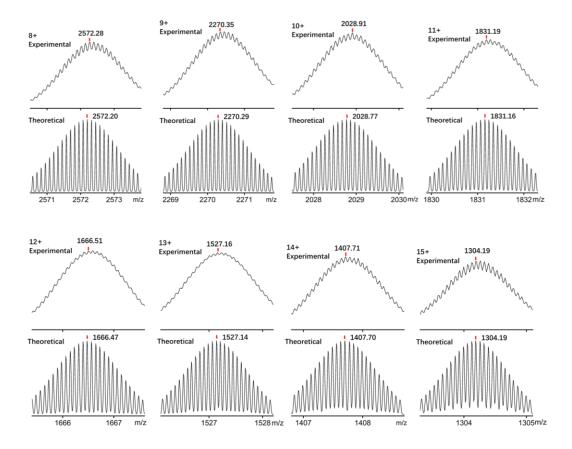


Figure S84: Theoretical (bottom) and experimental (top) isotope patterns for the different charge states observed from $G2(PF_6^-$ as counterion).

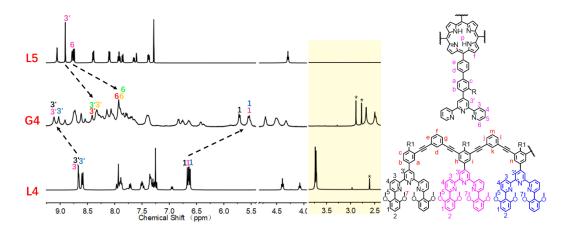


Figure S85. Comparison diagram of ¹H NMR spectra of G4 and its module ligands L4 and L5.

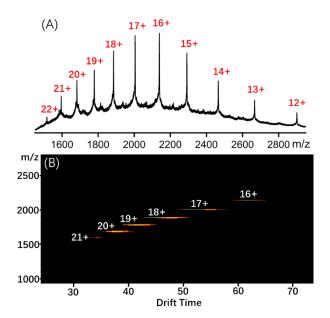


Figure S86. ESI-MS spectrum and ESI-TWIM-MS plot of G4.

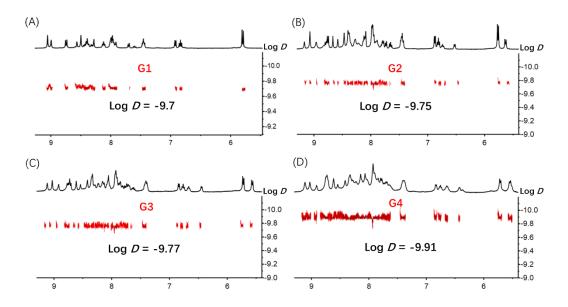


Figure S87. DOSY spectra (500 MHz, 298K) of G1-G4 in CD₃CN.

The hydrodynamic radius of **G1-G4** can be estimated, according to the Stokes-Einstein Equation. Where D is the diffusion constant, k is the Boltzmann's constant, T is the temperature, μ is the viscosity of solvents, and R is the radius:

$$D = kT/6\pi\mu R$$

D (G1) = 10^{-9.7} m² s⁻¹, D (G2) = 10^{-9.75} m² s⁻¹, D (G3) = 10^{-9.77} m² s⁻¹, D (G4) = 10^{-9.91} m² s⁻¹
k = 1.38×10⁻²³ N m K⁻¹,
T = 298 K,
 $\mu = 3.67 \times 10^{-4}$ N m⁻² s (CD₃CN, 298 K),
R(G1) = 3.0 nm, R(G2) = 3.3 nm, R(G3) = 3.5 nm, R(G4) = 4.8 nm.

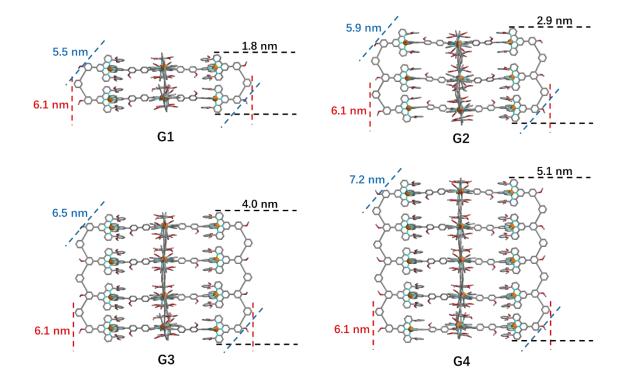


Figure S88. The molecular energy minimization diagram of **G1-G4** and the sizes of three different directions (Obtained through Materials Studio).

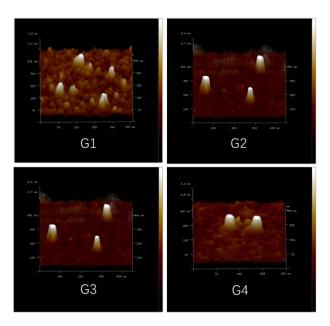


Figure S89. 3D AFM micrograph of G1-G4.

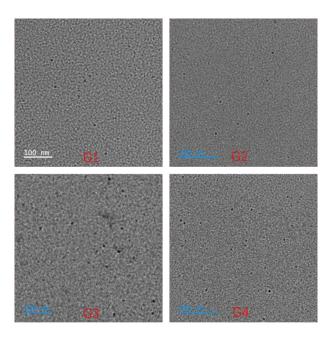


Figure S90. TEM micrograph of G1-G4.

6. The host-guest interaction between G0-G4 and C₆₀.

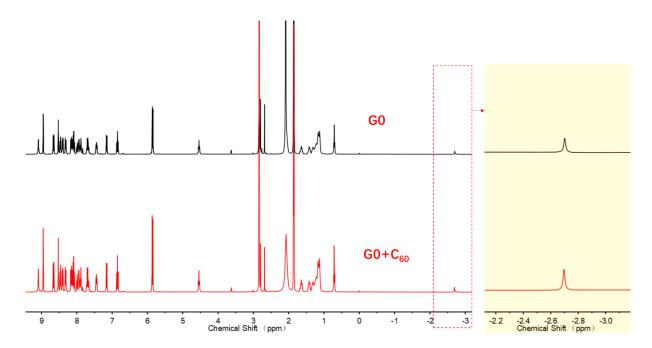


Figure S91. Comparison diagram of ¹H NMR spectra (600 M) of G0 and G0+C₆₀ (the pink part was enlarged).

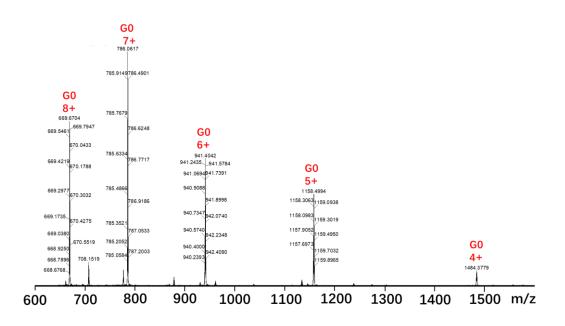


Figure S92: ESI-MS spectrum of G0+C₆₀.

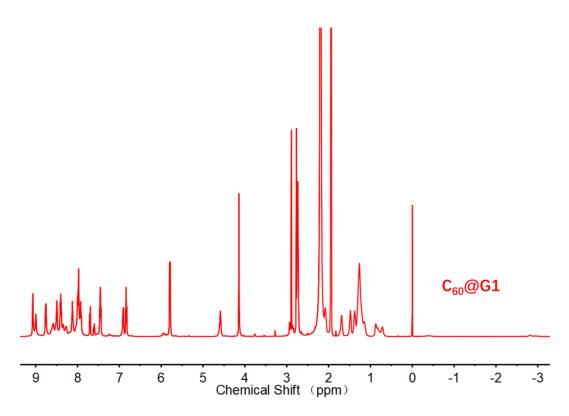


Figure S93. ¹H NMR spectrum of compound C₆₀@G1 in CD₃CN.

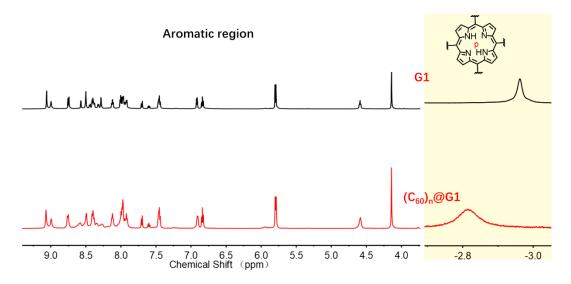


Figure S94. Comparison diagram of ¹H NMR spectra (600 M) of G1 and C_{60} (the pink part was enlarged).

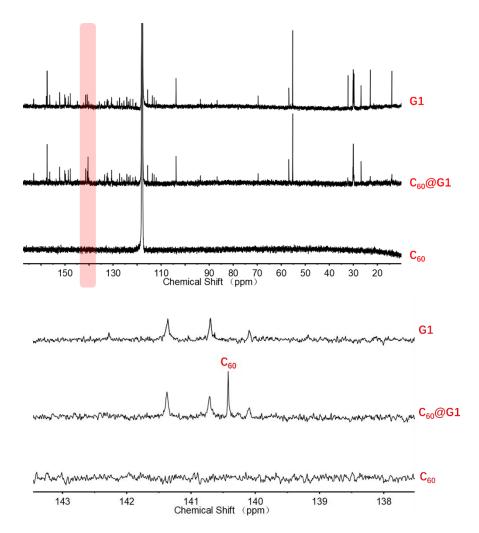


Figure S95. Comparison diagram of ¹³C NMR spectra (600 M) of G1, C₆₀@G1 and C₆₀ in CD₃CN.

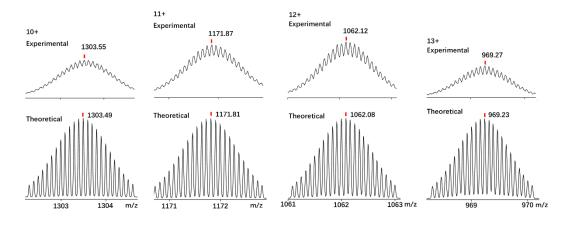


Figure S96. Theoretical (bottom) and experimental (top) isotope patterns for the different charge states observed from $C_{60}@G1(PF_6^-$ as counterion).

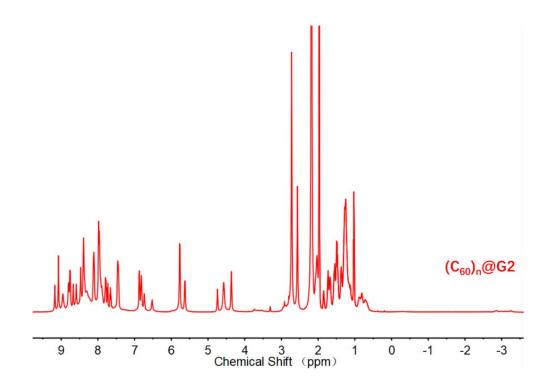


Figure S97. ¹H NMR spectrum of compound (C₆₀)_n@G2 in CD₃CN.

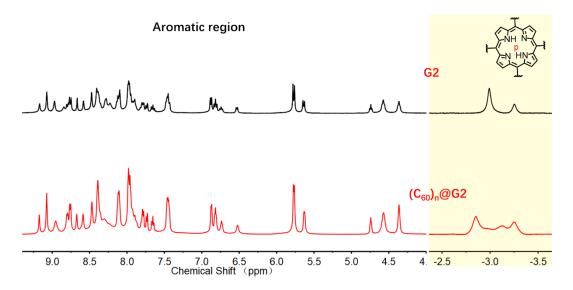


Figure S98. Comparison diagram of ¹H NMR spectra (600 M) of G2 and (C₆₀)_n@G2 (the pink part was enlarged).

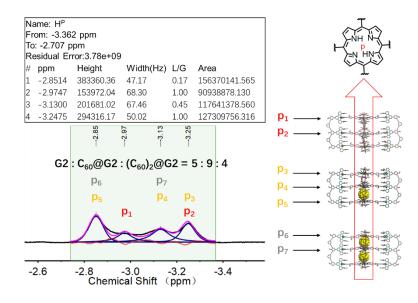


Figure S99. The ¹H NMR spectra integrations and attribution of H^P for $(C_{60})_n@G2$ and proportion of G2, $C_{60}@G2$ and $(C_{60})_2@G2$.

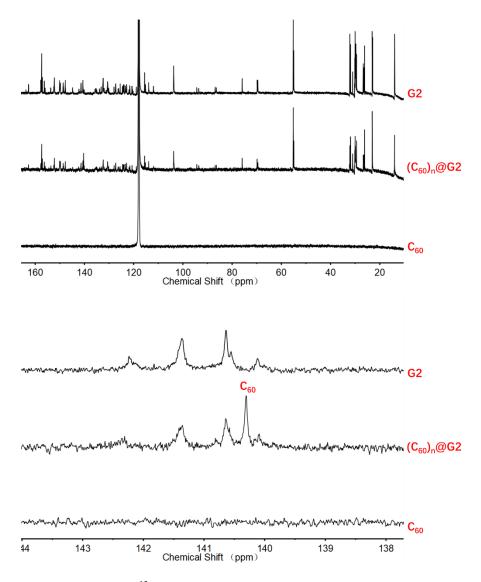


Figure S100. Comparison diagram of ¹³C NMR spectra (600 M) of G2, (C₆₀)_n@G2 and C₆₀ in CD₃CN.

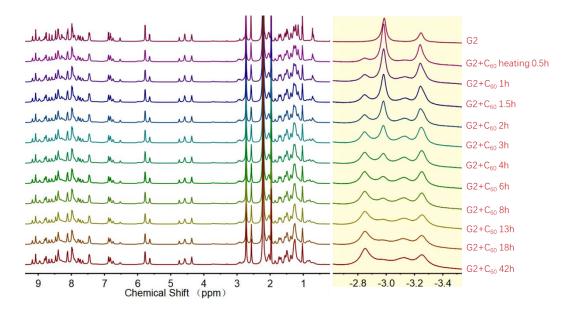


Figure S101. ¹H NMR spectra (600 M) monitoring of the process of G2 wrapping C_{60} under 353K heating conditions (the pink part was enlarged).

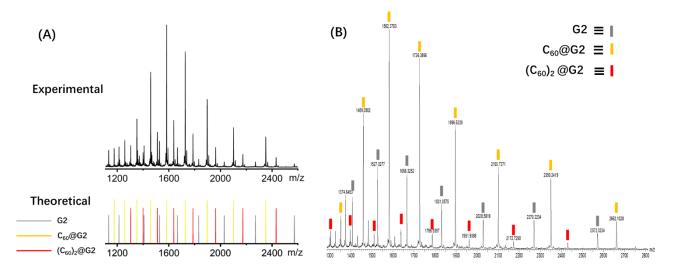


Figure S102. (A) Experimental and theoretical values of ESI-MS data in DMF for G2, $C_{60}@G2$ and $(C_{60})_2@G2$; (B) ESI-MS spectrum of $(C_{60})_n@G2$ in CH₃CN.

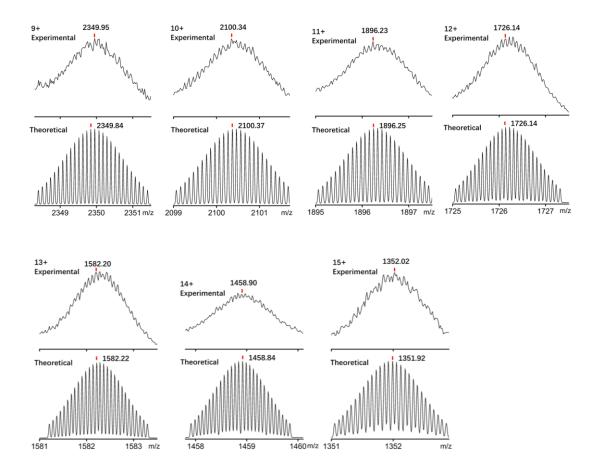


Figure S103. Theoretical (bottom) and experimental (top) isotope patterns for the different charge states observed from $C_{60}@G2$ in DMF (PF₆⁻ as counterion).

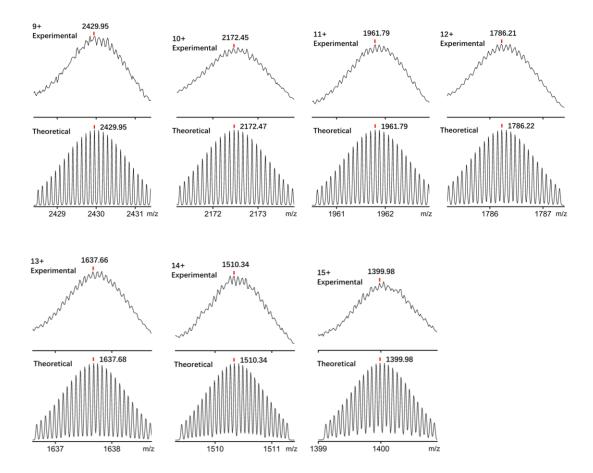


Figure S104. Theoretical (bottom) and experimental (top) isotope patterns for the different charge states observed from $(C_{60})_2$ (BG2 in DMF (PF_6^- as counterion).

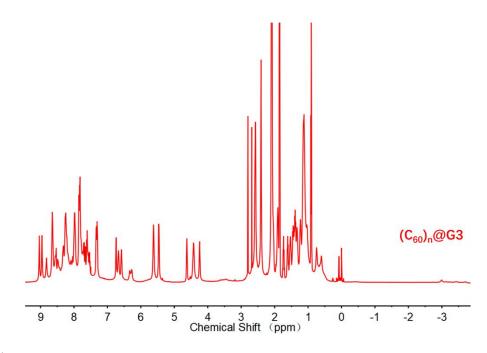


Figure S105. ¹H NMR spectrum of compound $(C_{60})_n@G3$ in CD₃CN.

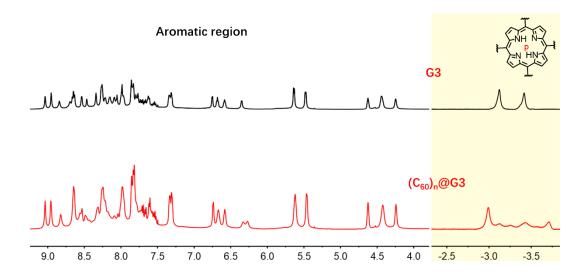


Figure S106. Comparison diagram of ¹H NMR spectra (600 M) of G3 and $(C_{60})_n @G3$ (the pink part was enlarged).

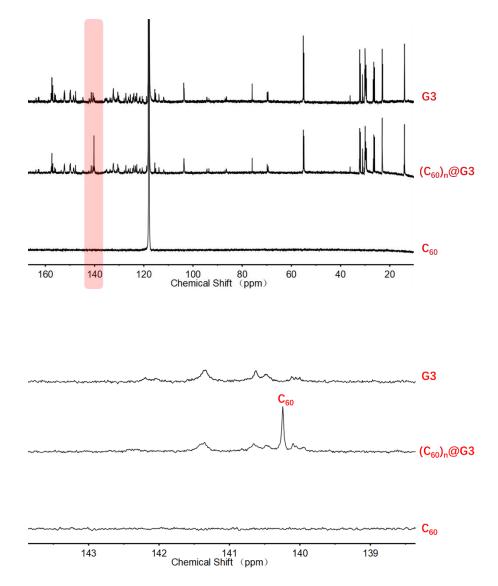


Figure S107. Comparison diagram of ¹³C NMR spectra (600 M) of G3, (C₆₀)_n@G3 and C₆₀ in CD₃CN.

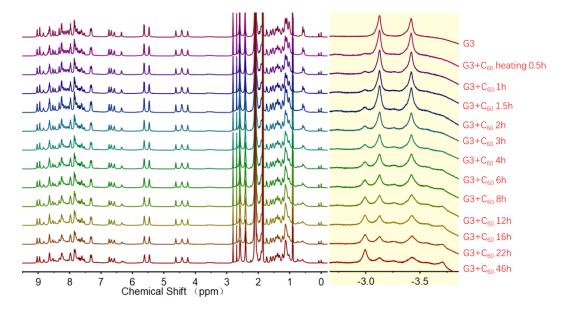


Figure S108. ¹H NMR spectra (600 M) monitoring of the process of G3 wrapping C_{60} under 353K heating conditions (the pink part was enlarged).

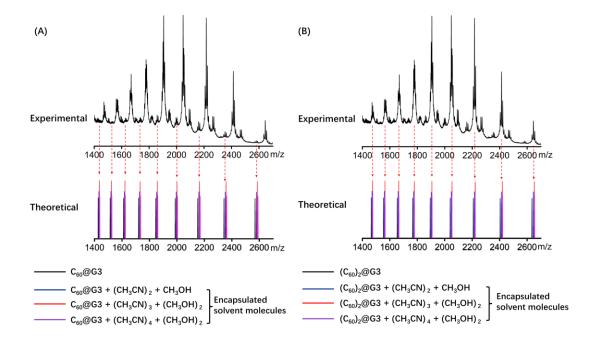


Figure S109. Experimental and theoretical values of mass spectrometry data for (A) part $C_{60}@G3$ and (B) part $(C_{60})_2@G3$.

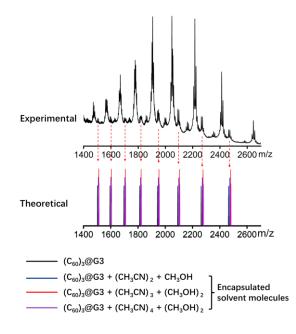


Figure S110. Experimental and theoretical values of mass spectrometry data for part $(C_{60})_3$ @G3.

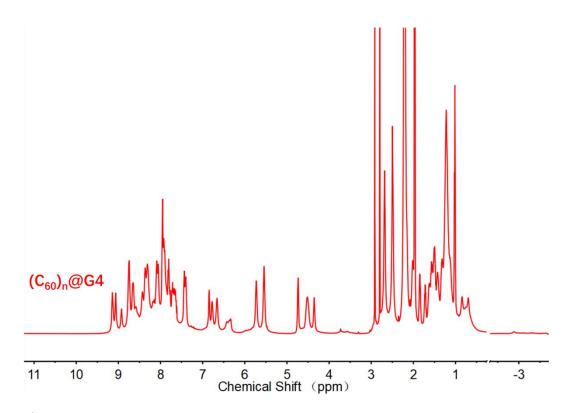


Figure S111. ¹H NMR spectrum of compound (C₆₀)_n@G4 in CD₃CN.

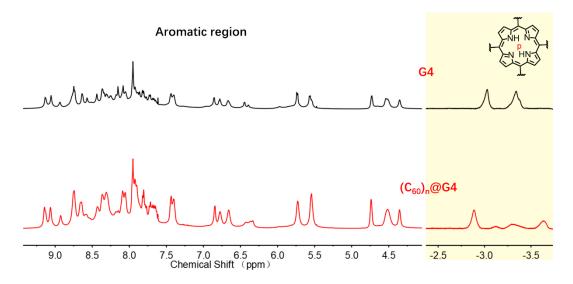


Figure S112. Comparison diagram of ¹H NMR spectra (600 M) of G4 and $(C_{60})_n@G4$ (the pink part was enlarged).

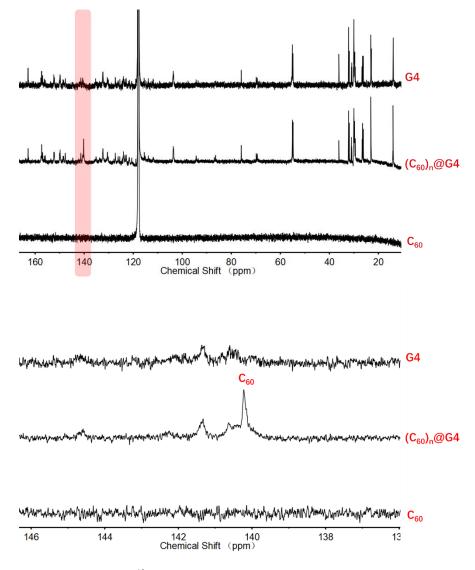


Figure S113. Comparison diagram of ¹³C NMR spectra (600 M) of G4, (C₆₀)_n@G4 and C₆₀ in CD₃CN.

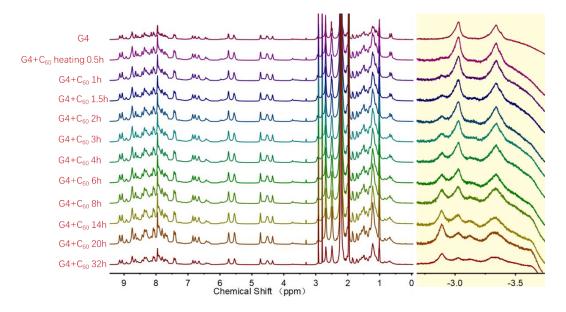


Figure S114. ¹H NMR spectra (600 M) monitoring of the process of G4 wrapping C_{60} under 353K heating conditions (the pink part was enlarged).

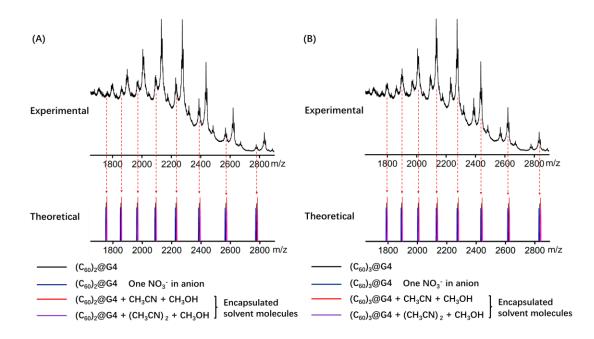


Figure S115. Experimental and theoretical values of mass spectrometry data for (A) part $(C_{60})_2@G4$ and (B) part $(C_{60})_3@G4$.

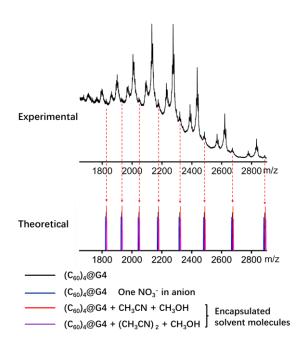


Figure S116: Experimental and theoretical values of mass spectrometry data for part $(C_{60})_4@G4$.

7. Calculation of binding constant (K_a), activation energy (E_a) and the energy state of host-guest complexes.

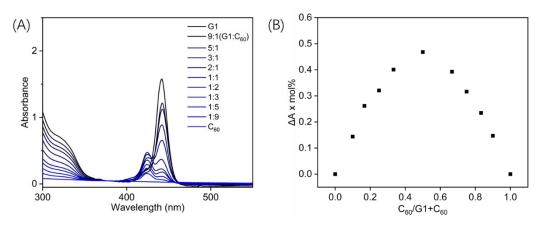


Figure S117. (A) UV-vis absorption of **G1** with guest C_{60} in different molar ratios (**G1** + $C_{60} = 4 \mu$ M). (B) Job's plot of the **G1** \supset C₆₀ in DMF, showing a 1:1 stoichiometry.

UV-Vis Titration of G1 and G2 with C₆₀

The binding constant (K_a) for the formation of C₆₀@G1, C₆₀@G2 and (C₆₀)₂@G2 in DMF was determined by UV titration on a Hitachi (model U-3010) UV–vis spectrophotometer in a 1-cm UV cuvette. The concentrated solution of C₆₀ (1 mM in PhMe) was added incrementally to the Metal-organic capsules solution (20 μ M in DMF). The UV-Vis spectra were recorded one after the other. From a global shift analysis of the titration data in absorption intensity at 401 nm, 433 nm and 466 nm using Bindfit ^{\$3,\$4} gave an association constant of (2.9 ± 0.6) × 10⁴ M⁻¹, (3.06 ± 0.5) × 10⁴ M⁻¹ and (1.99 ± 0.5) × 10⁴ M⁻¹ for C₆₀@G1, C₆₀@G2 and (C₆₀)₂@G2, respectively.

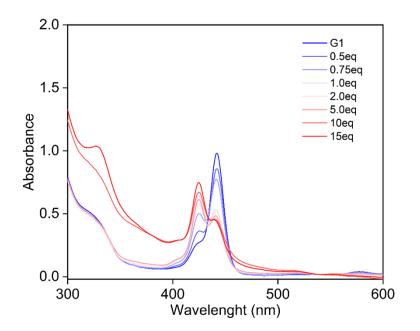


Figure S118. UV-Vis titration of C₆₀ into G1 in DMF.

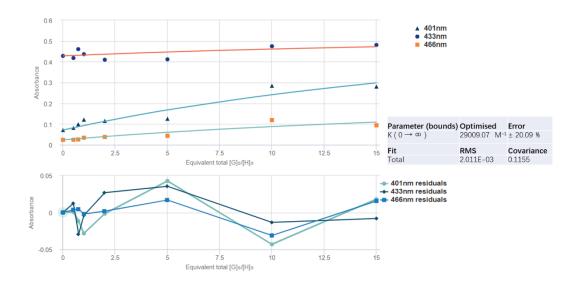


Figure S119. Binding isotherms (1:1 model) fitted to the absorbance shift of three bands vs. the equivalents of C_{60} added to determine the binding affinity of **G1** with C_{60} (top); and the residual plot from the fit (bottom).

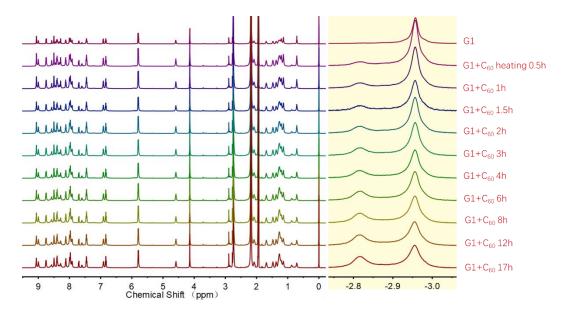


Figure S120. ¹H NMR spectra (600 M) monitoring of the process of G1 wrapping C_{60} under 333K heating conditions (the pink part was enlarged).

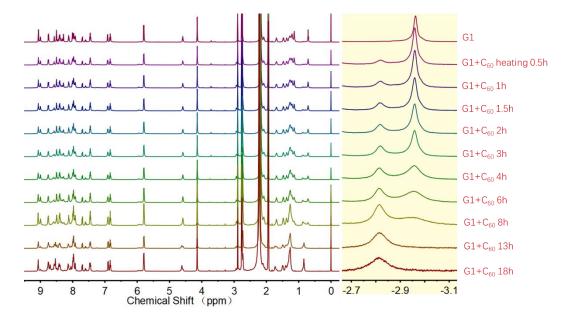


Figure S121. ¹H NMR spectra (600 M) monitoring of the process of G1 wrapping C_{60} under 353K heating conditions (the pink part was enlarged).

Table S1. The integral ratio (G1:C₆₀@G1) of proton (H^P) in porphyrin rings during ¹H NMR spectra (600 M) monitoring of the process of G1 wrapping C₆₀ under 333K and 353K heating conditions.

	Integral ratio	Time (h)										
		0.5	1	1.5	2	3	4	6	8	12	13	17
	G1 : C ₆₀ @G1 (333 K)	8.33	5.56	4.35	3.57	3.0	2.86	2.38	2.2	1.9	-	1.7
	G1 : C ₆₀ @G1 (353 K)	5.8	4.65	3.42	2.53	2.14	1.56	1.2	0.59	-	0	-

Table S2. According to the integration ratio in Table S1 and $c_0=0.581$ mM calculated lnc_t corresponding to different heating times at 333 K and 353 K.

Time (s)	1800	3600	5400	7200	10800	14400	21600
Inc _t (333 K)	-7.5636	-7.615	-7.6564	-7.6974	-7.7356	-7.7494	-7.8018
Inc _t (353 K)	-7.6089	-7.6459	-7.7063	-7.7824	-7.8341	-7.9462	-8.0566

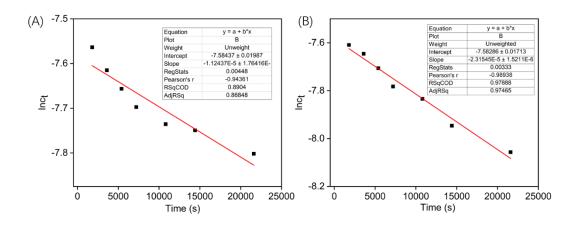


Figure S122. Based on the kinetic equations of the first order reaction ($\ln c_t = \ln c_0 - kt$), the rate constants (k) at the two temperatures were obtained by fitting straight lines. (A) Experiments conducted at 333K, (B) Experiments conducted at 353K.

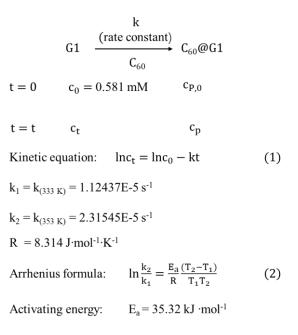


Figure S123. Calculation of Ea for G1 encapsulated C60 according to the Arrhenius formula.

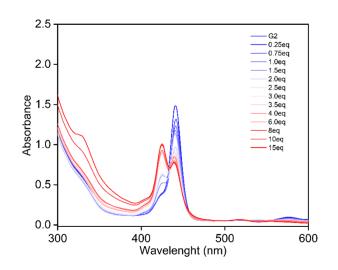
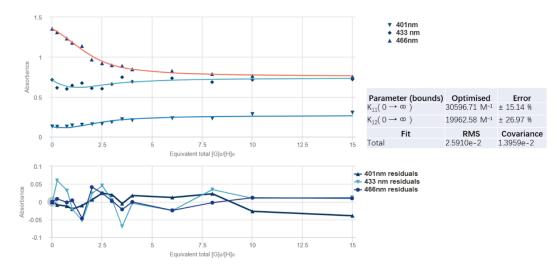


Figure S124. UV-Vis titration of C₆₀ into G2 in DMF.



The cooperativity parameter (α) of G2 wrapping C₆₀ is described as: $\alpha = K_2/K_1 = (3.06 \pm 0.5) \times 10^4/(1.99 \pm 0.5) \times 10^4 = 0.65 < 1$

Where $\alpha > 1$ is positive cooperativity and $\alpha < 1$ is negative cooperativity. ^{S5}

Figure S125. Binding isotherms (1:2 model) fitted to the absorbance shift of three bands vs. the equivalents of C_{60} added to determine the binding affinity (top); and the residual plot from the fit (bottom) as well as the calculation of cooperativity parameter (α).

Calculation of the energy state of the host-guest complexes

To obtain interaction between C₆₀ and supramolecules, we performed a systematic optimization protocol including forcefield and semiempirical calculations using xtb 6.5.0. ^{S6} First, the geometric structure for each cluster were preoptimized at the GFN-FF and GFN0-xTB method. ^{S7-S8} The obtained structure was further optimized using the semiempirical quantum mechanical GFN1-xTB method. ^{S9} (The saturated fatty chain in **G2-G4** was simplified to methoxy group).

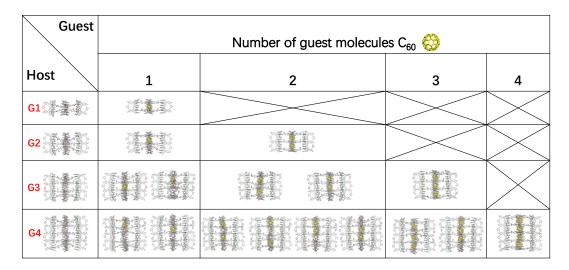


Figure S126. Possible structures after host-guest interactions between G1 to G4 and excessive C₆₀.

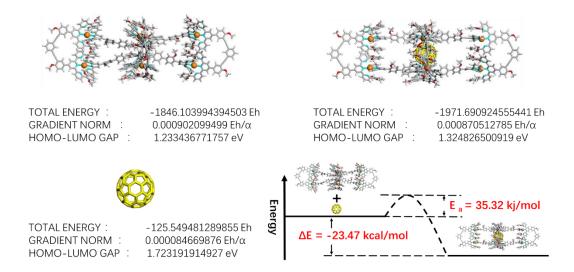
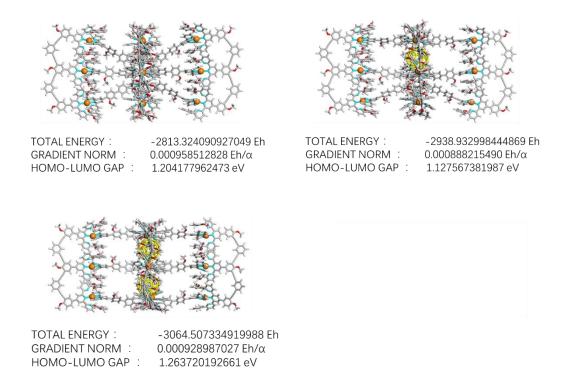
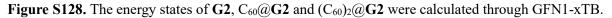
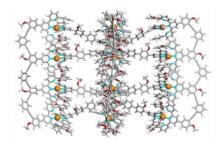


Figure S127. The total energy of G1, C_{60} @G1 and free C_{60} were calculated through GFN1-xTB, and the schematic diagram of total energy change (ΔE) in the process of G1 wrapping C_{60} . (1 Eh = 627.51 kcal/mol; activation energy Ea was calculated according to Figure S115 and S116)

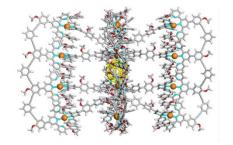






TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

-3780.534244146036 Eh 0.000965866563 Eh/α 1.226839006413 eV



TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

TOTAL ENERGY :

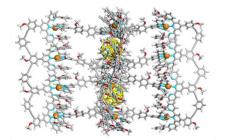
GRADIENT NORM :

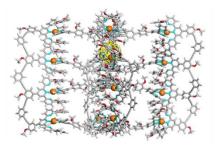
HOMO-LUMO GAP :

-3906.160044057117 Eh 0.000934230343 Eh/α 1.198578204357 eV

0.000943207022 Eh/α

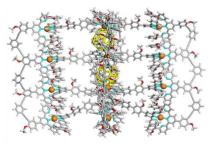
1.120770307700 eV





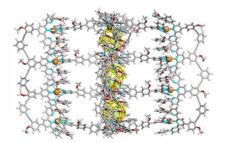
TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

-3906.129291778843 Eh 0.000673725244 Eh/α 1.153631490891 eV



TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

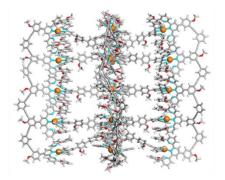
-4031.754725999013 Eh 0.000737533431 Eh/α 1.217912759524 eV



-4157.343145420004 Eh TOTAL ENERGY : -4031.751628181126 Eh GRADIENT NORM : 0.000777817993 Eh/α HOMO-LUMO GAP :

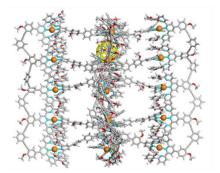
1.266380085963 eV

Figure S129. The energy states of G3, C₆₀@G3, (C₆₀)₂@G3, (C₆₀)₃@G3 and their isomers were calculated by GFN1-xTB.

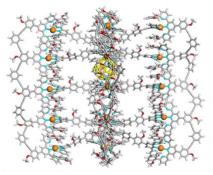


TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

-4747.720659267078 Eh 0.000917149387 Eh/α 1.188380806104 eV

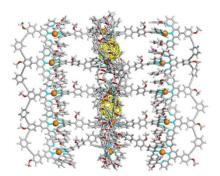


TOTAL ENERGY: -4873.341713829346 Eh 0.000979587293 Eh/α GRADIENT NORM : HOMO-LUMO GAP : 1.220356969356 eV

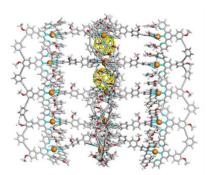


TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

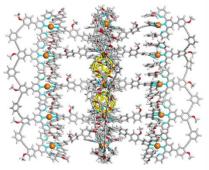
-4873.365396158342 Eh 0.000997319748 Eh/α 1.226305780489 eV



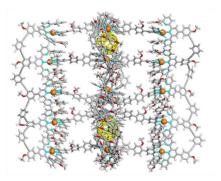
TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP : -4998.936285779753 Eh 0.000848218445 Eh/α 1.216063461570 eV



TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP : -4998.944138010906 Eh 0.000902564593 Eh/α 1.222442145196 eV

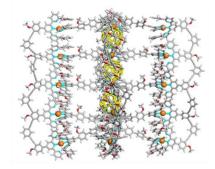


TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP : -4999.001964146004 Eh 0.000999867439 Eh/α 1.226251345097 eV



TOTAL ENERGY: GRADIENT NORM: HOMO-LUMO GAP:

-4998.942239875352 Eh 0.000747390472 Eh/α 1.144899376975 eV



TOTAL ENERGY : GRADIENT NORM : HOMO-LUMO GAP :

-5124. 585032104970 Eh 0.000936973429 Eh/α 1.213371919010 eV

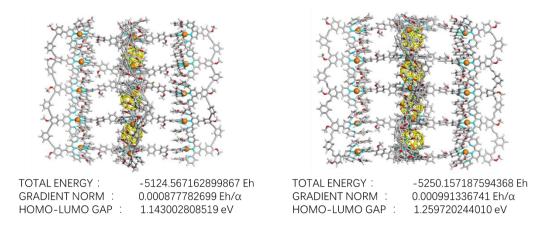


Figure S130. The energy states of G4, $C_{60}@G4$, $(C_{60})_2@G4$, $(C_{60})_3@G4$, $(C_{60})_4@G4$ and their isomers were calculated by GFN1-xTB.

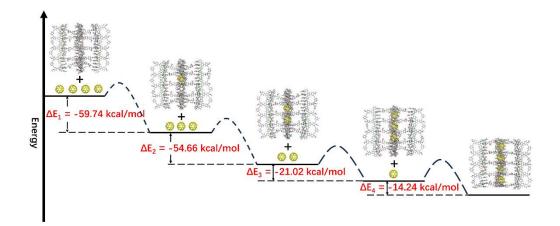


Figure S131. The energy schematic diagram of inferred main path and corresponding binding energy for **G4** wrapping C₆₀, respectively (the main path given by calculating the maximum energy difference).

8. Comparison of the properties of capsules loaded with C_{60} to produce ${}^{1}O_{2}$.

Capsules loaded with C_{60} were tested to generate singlet oxygen. 9,10-diphenylanthracene (DPHA) was used the ${}^{1}O_{2}$ scavenger monitors ${}^{1}O_{2}$ generation, capsules loaded with C_{60} complexes were used as a photocatalyst to dissolve in acetonitrile solution and detect the changes in absorption intensity of DPHA by UV at different irradiation times. This gradual decrease in absorption intensity can be ascribed to the formation of DPHA endoperoxide via singlet oxygen mediated oxidation of DPHA.

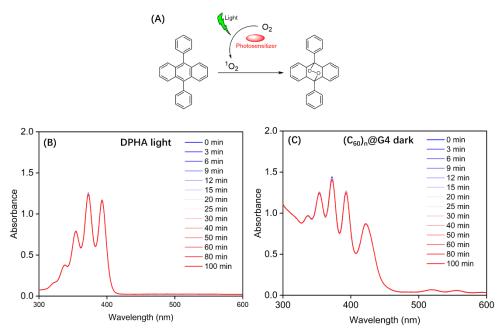


Figure S132. (A) The mechanism of 9,10-diphenylanthracene (DPHA) as the ${}^{1}O_{2}$ scavenger monitors singlet oxygen generation in the solution. (B) The absorption spectra of DPHA (106.0 μ M) after irradiation (405 nm, 20 mW/cm²) for different time no photosensitizer added. (C) The absorption spectra of DPHA (106.0 μ M) under dark conditions for different time in the presence of (C₆₀)_n@**G4** (1.0 μ M).

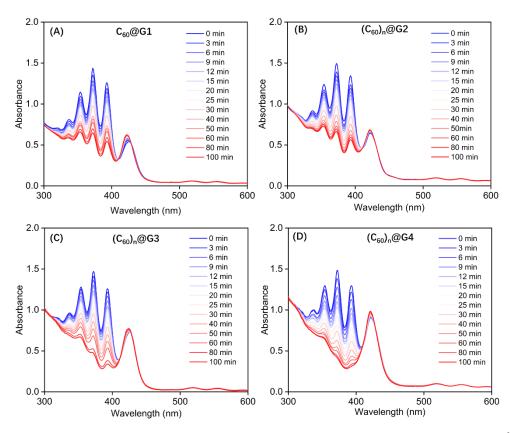


Figure S133. The absorption spectra of DPHA (106.0 μ M) after irradiation (405 nm, 20 mW/cm²) for different time in the presence of (A) C₆₀@G1 (1.0 μ M), (B) (C₆₀)_n@G2 (1.0 μ M), (C) (C₆₀)_n@G3 (1.0 μ M) and (D) (C₆₀)_n@G4 (1.0 μ M) in CH₃CN.

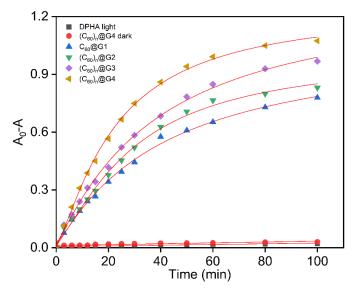


Figure S134. Two comparison samples and capsule encapsulated C_{60} complex $[C_{60}@G1, (C_{60})_n@G2, (C_{60})_n@G3$ and $(C_{60})_n@G4)]$ plots for the absorption decays of DPHA (106.0 μ M) at 373 nm.

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