

# Real-time Observation of sub-100-fs Charge and Energy Transfer Processes in DNA Dinucleotides

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## Electronic Supplementary Information

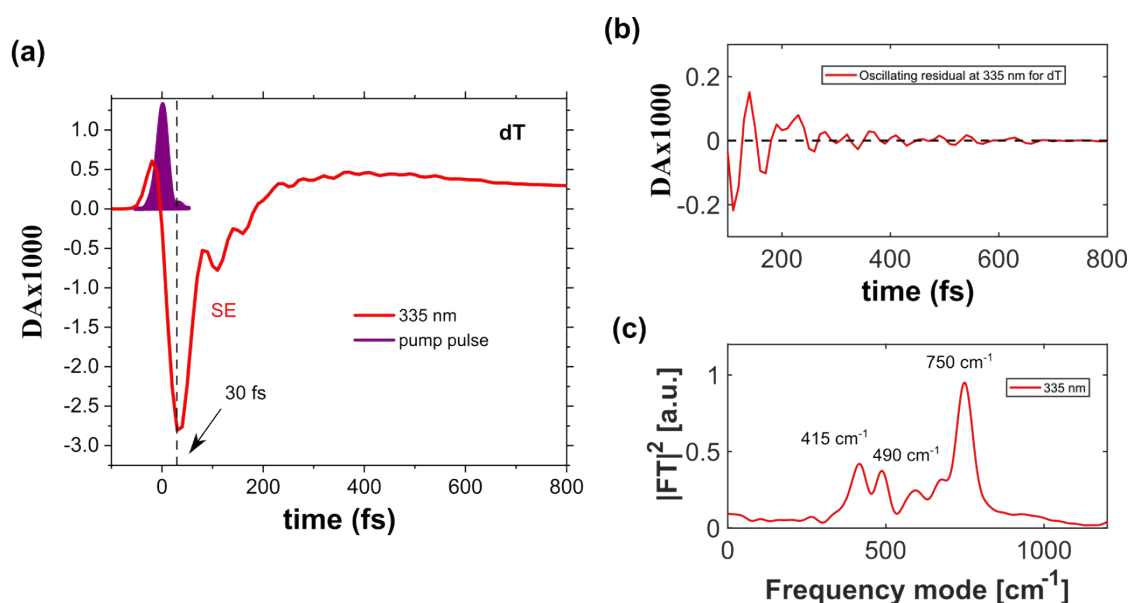
- 1. Temporal resolution of the TA experiments**
- 2. Additional Figures**
- 3. Additional Tables**

## 1. Temporal resolution of the transient absorption experiments

As described in the main text, the 266-nm UV pump pulses are generated as the second harmonic of a broadband visible NOPA. The pulses are compressed to a 24-fs duration (FWHM) with the aid of a prism pair prior to their comprehensive temporal and spectral characterization by two-dimensional spectral interferometry. Details of the experimental setup can be found in reference 1. The transient absorption (*TA*) signal is recorded as a function of the time delay between the pump pulse and a chirped broadband probe pulse generated as a white light continuum (WLC) in a 3-mm-thick CaF<sub>2</sub> plate. We have previously demonstrated that the chirp of the WLC probe has a minimal effect on the overall “effective” time resolution<sup>2</sup> of the *TA* experiment, provided that broadband detection of the *TA* signal with sufficiently high temporal resolution is adopted, as in our case. Chirp correction on the experimental datasets is performed using a custom-made Matlab code, aligning the sharp cross-phase modulation (XPM) responses to mark the zero-time arrival between the pump and probe pulses.

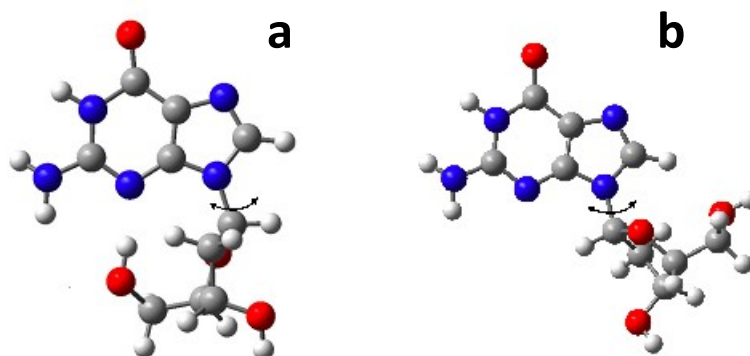
UV *TA* experiments aiming for high temporal resolution are challenging due to the co-presence of pulse dispersion and XPM, caused by the interaction of the pump pulse with the cuvette front and back glass surfaces, as well as XPM and group velocity mismatch between the pump and probe pulses in the sample. We bypass these problems, as previously demonstrated, by using high concentration solutions in 1-mm-pathlength cuvettes.<sup>3</sup> The high concentrations of the samples (dG, dT, equimolar mixture of dT and dG, and **TG**) are crucial because the *TA* signal is only recorded when there is an overlap of the pump and probe pulses. If the first 200 or 300  $\mu\text{m}$  of the 1-mm cuvette pathlength absorbs almost all the pump light, the total thickness of the sample becomes irrelevant, and only the effective thickness matters. In this case the pump pulse intensity on the cuvette back window, which is 1 mm away from the front window, is low enough that the generated XPM artefact is negligible. The dispersion caused by the entrance front glass window of the cuvette on the pump pulses can be pre-compensated by adjusting the negative dispersion introduced by the prism compressor. Additionally, self-absorption of the probe pulse can be safely excluded, as the ground state absorption of the studied molecules (extending at wavelengths shorter than 300 nm) does not overlap with our probe spectrum (320-650 nm).

To confirm the high temporal resolution of our setup, we performed TA experiments on dT under the same conditions used for the TG dinucleotide, the results of which are reported in Fig. S1. We observe the immediate rise of the stimulated emission (SE) signal from the  $\pi\pi^*$ T state populated by the pump pulse (Fig. S1a). We can estimate the instrumental response function (IRF) of the setup by the time that the SE signal takes to rise from 10% to 90% of its maximum, which is 27 fs. We also observe, superimposed to the SE dynamics, an oscillatory signal which is due to impulsively excited coherent molecular vibrations (Figure S1(b)). The oscillatory signal arises from the wave packet motion on the excited state potential energy surface<sup>4</sup>. Performing a Fourier transform on the oscillating residuals (Fig. S1c), we observe peaks at 415  $\text{cm}^{-1}$ , 490  $\text{cm}^{-1}$  and 750  $\text{cm}^{-1}$ . The 750  $\text{cm}^{-1}$  peak (which corresponds to a 42-fs oscillation period) is typical of a breathing mode of the aromatic ring, characterized by a large amplitude of the N1C6C5 angle.<sup>5</sup> To experimentally resolve such high-frequency modulation, an IRF significantly shorter than the periods of the observed vibrations is required.

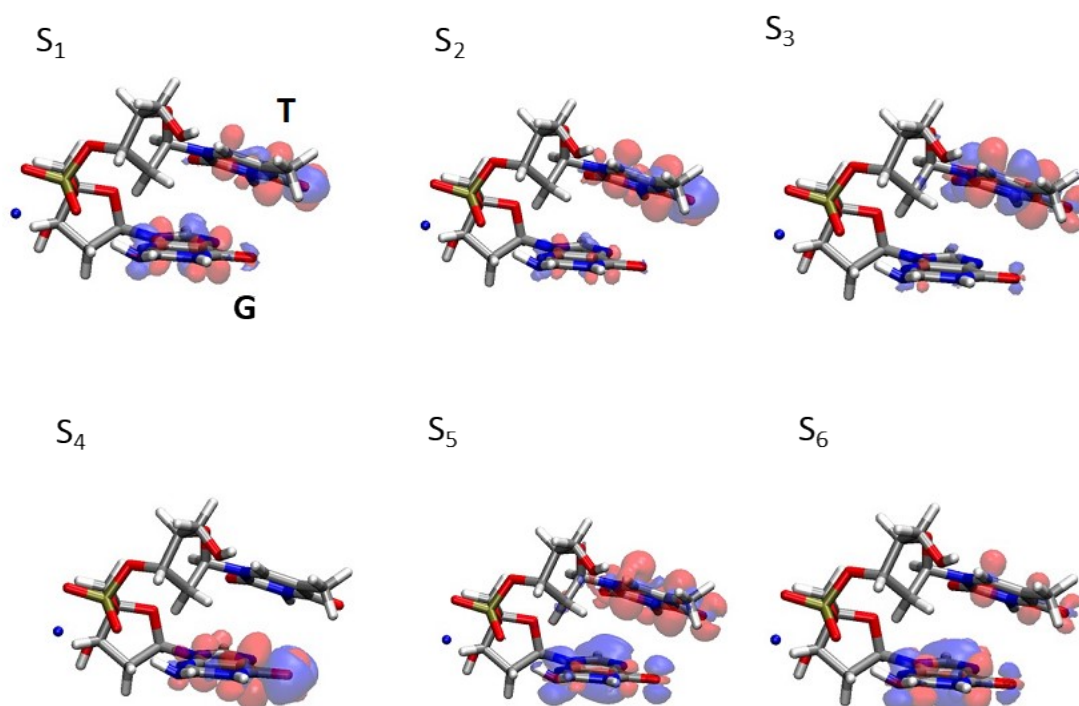


**Fig. S1:** (a) TA dynamics for the monomeric dT at 335 nm probe wavelength. One observes an instantaneous (within the 27-fs IRF) appearance of the SE. The SE possesses oscillating modulation due to the impulsively generated coherent molecular vibrations. (b) Isolation of the oscillating residual at 335 nm. (c) Fourier Transform (FT) power spectrum obtained after performing FT of the oscillating residuals in order to retrieve the vibrational content of the molecular response.

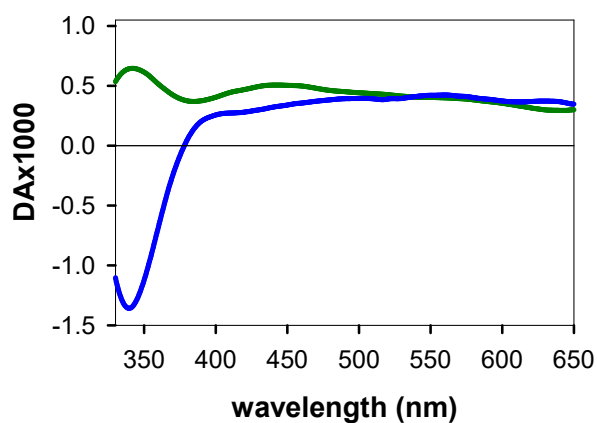
## 2. Additional Figures



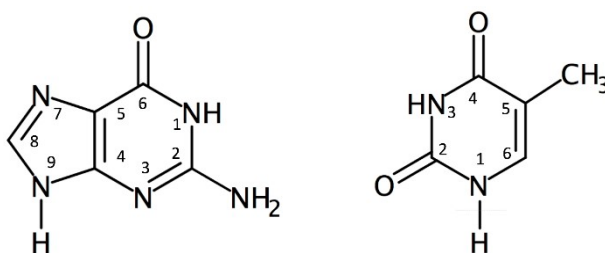
**Fig. S2.** The *anti-anti* (a) and *anti-syn* (b) conformers of **TG**, encountered, respectively, in B-form duplexes and in G-Quadruplexes; the arrows indicate rotation around the glycosidic bond.



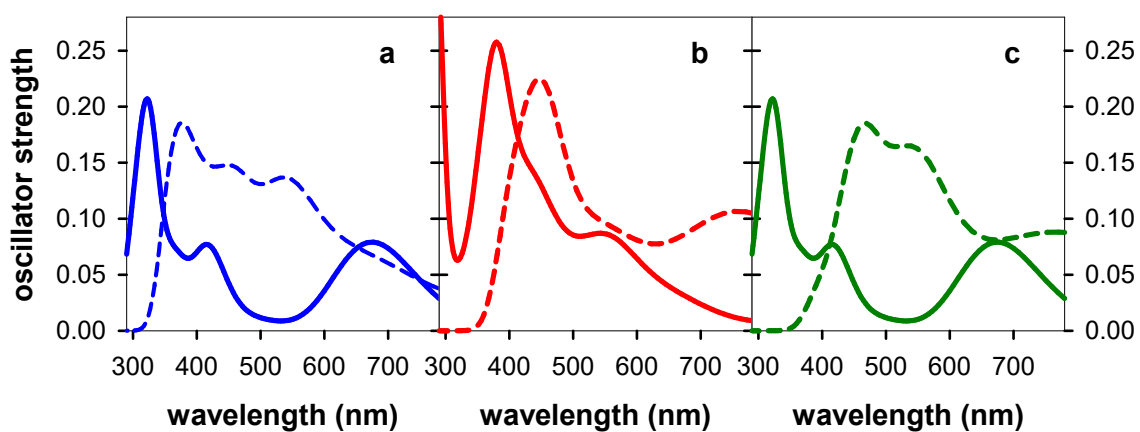
**Fig. S3.** Excited state-ground state density difference determined for the 6 lowest Franck-Condon states of *anti-syn* **TG**. A decrease in the electronic density is illustrated in blue, while an increase in red.



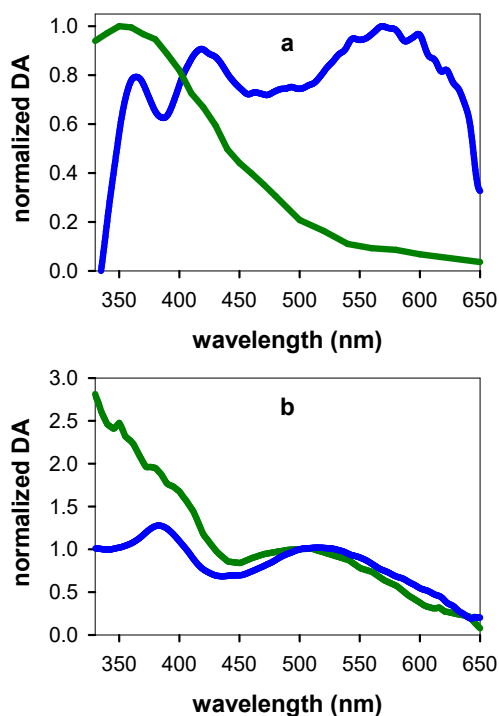
**Fig. S4.** TAS measured for dT (blue) and dG (green) at 50 fs.



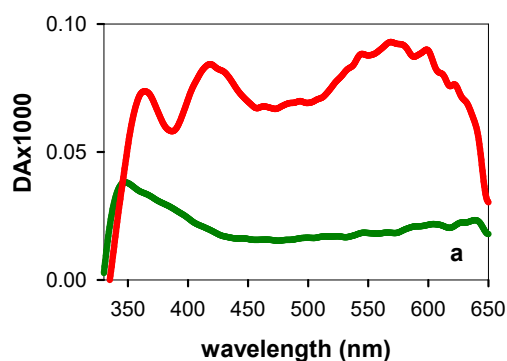
**Fig. S5.** Atom labelling for the guanine and thymine chromophores.



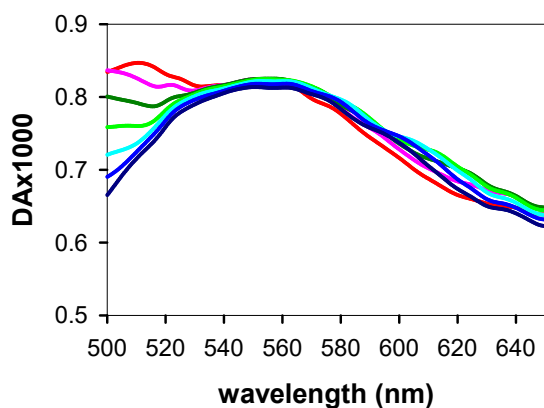
**Fig. S6.** TAS computed for selected Franck-Condon states (dashed lines) and the corresponding minima (solid lines) of *anti-syn* TG:  $S_3 \rightarrow \text{min-}\pi\pi^*G$  (a),  $S_5 \rightarrow \text{min-}CT$  (b) and  $S_6 \rightarrow \text{min-}\pi\pi^*G$  (c).



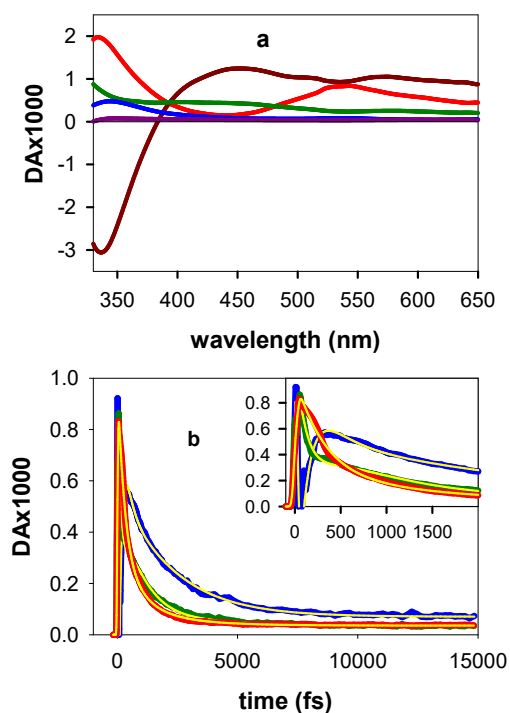
**Fig. S7.** In blue: *TAS* recorded for TG (blue) at 40 ps (a) and 5 ps (b). In green: spectrum of the triplet state determined for thymidine monophosphate by flash photolysis (a);<sup>6</sup> spectrum corresponding to an equimolar mixture of the guanosine radical cation<sup>7</sup> and the thymidine radical anion (b).<sup>8</sup> We note that the anion spectrum, obtained by pulse radiolysis is not accurate in the UV region because this method generates additional transient species.



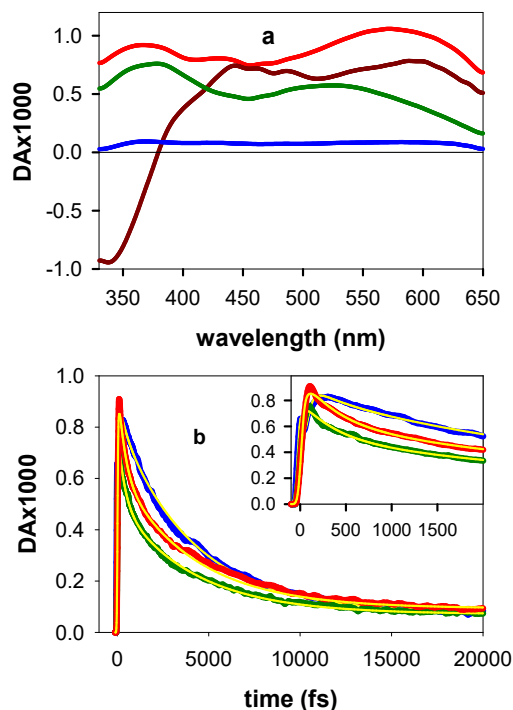
**Fig. S8.** Comparison of the *TAS* recorded at 40 ps for TG (red) and an equimolar mixture of dT and dG (green).



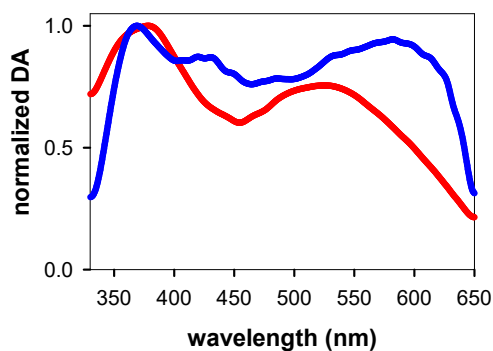
**Fig. S9.** TAS recorded for an equimolar mixture of dT and dG from 40 to 100 fs with 10 fs steps.



**Fig. S10.** (a) EAS components retrieved from global analysis of the TA data for an equimolar mixture of dG and dT and corresponding to time constants of 120 fs (dark red), 250 fs (red), 800 fs (green), 2.1 ps (blue) and 2 ns (violet). (b) Kinetic traces at 365 nm (blue), 445 nm (green) and 520 nm (red) shown together with the fitted functions (yellow).

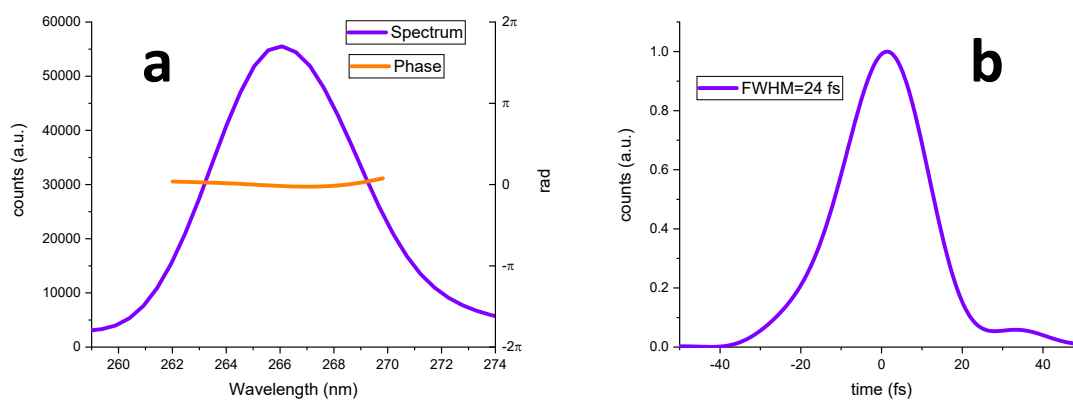


**Fig. S11.** (a) *EAS* components retrieved from a global analysis of the *TA* data for **TG** and corresponding to time constants of 50 fs (dark red), 380 fs (red), 4.1 ps (green) and 2 ns (blue). (b) Kinetic traces at 365 nm (blue), 445 nm (green) and 520 nm (red) shown together with the fitted functions (yellow).

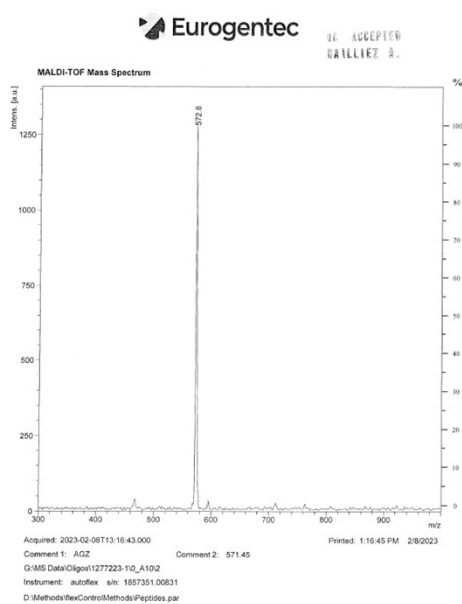


**Fig. S12.** Normalized *EAS* corresponding to the time constants of 4.1 ps (red) and 2 ns (blue) retrieved from the global analysis performed for the *TA* data of **TG**.





**Fig. S13.** Full spectral and temporal characterization of the exciting laser pulses.



**Fig. S14.** MALDI-TOF spectrum provided by Eurogentec for TG.

## Additional Tables

**Table S1.** Relative energies of the different possible **TG** conformers with respect to the *anti-anti* one, computed at the PCM/M052X/6-31G(d) level of theory.

conformer	$\Delta E$ in eV (kcal/mol)	$\Delta G$ in eV (kcal/mol)
<i>anti-anti</i>	0.0 (0.0)	0.0 (0.0)
<i>anti-syn</i>	-0.16 (-3.7)	-0.06 (-1.4)
<i>syn-anti</i>	0.11 (2.5)	0.13 (3.1)
<i>syn-syn</i>	0.03 (0.7)	0.11 (2.6)

**Table S2.** Properties of the Franck-Condon states determined for *anti-anti TG* at the PCM/M052X/6-31G(d) level of theory. VAE: vertical absorption energy; f: oscillator strength;  $\delta$ : charge transfer character.<sup>9</sup>

	character	VAE (eV)	f	$\delta$ (a.u.)
S <sub>1</sub>	$n\pi^*(T)$	5.21	0.001	0.02
S <sub>2</sub>	$\pi\pi^*G(La)$	5.29	0.162	0.04
S <sub>3</sub>	$\pi\pi^*T$	5.37	0.193	0.02
S <sub>4</sub>	$n\pi^*G + \pi\pi^*T + CT$	5.63	0.127	0.20
S <sub>5</sub>	$n\pi^*G + \pi\pi^*T + CT$	5.63	0.170	0.23
S <sub>10</sub>	$G \rightarrow T CT$	6.78	0.002	0.61

**Table S3.** Properties of the Franck-Condon states determined for *syn-anti TG* at the PCM/M052X/6-31G(d) level of theory. VAE: vertical absorption energy; f: oscillator strength;  $\delta$ : charge transfer character.

	character	VAE (eV)	f	$\delta$ (a.u.)
S <sub>1</sub>	$n\pi^*(T)$	5.18	0.006	0.02
S <sub>2</sub>	$\pi\pi^*G(La) + CT$	5.24	0.072	0.37
S <sub>3</sub>	$\pi\pi^*T + CT$	5.36	0.067	0.13
S <sub>4</sub>	$\pi\pi^*T + CT$	5.47	0.126	0.35
S <sub>5</sub>	$n\pi^*G$	5.62	0.001	0.01
S <sub>10</sub>	$G \rightarrow T CT$	6.70	0.019	0.57

**Table S4.** Properties of the Franck-Condon states determined for *syn-syn TG* at the PCM/M052X/6-31G(d) level of theory. VAE: vertical absorption energy; f: oscillator strength;  $\delta$ : charge transfer character.

	character	VAE (eV)	f	$\delta$ (a.u.)
S <sub>1</sub>	$\pi\pi^*G(La)$	5.19	0.040	0.11
S <sub>2</sub>	$n\pi^*(T)$	5.23	0.001	0.02
S <sub>3</sub>	$\pi\pi^*T + CT$	5.38	0.309	0.20
S <sub>4</sub>	$n\pi^*G$	5.62	0.040	0.02
S <sub>5</sub>	$\pi\pi^* G (Lb)$	5.68	0.317	0.08
S <sub>6</sub>	$G \rightarrow T CT$	5.72	0.047	0.52

**Table S5.** Properties of the minima located on the *PES* of the first excited state determined for *anti-syn TG* at the PCM/M052X/6-31G(d) level of theory. ADE: adiabatic (with respect to the S<sub>0</sub> at the FC region) energies; VEE: vertical (with respect to the S<sub>0</sub> at the corresponding minima) emission energies; f: oscillator strengths; initial state optimized and main reaction coordinate.

	ADE (eV)	VEE (eV)	f	initial state	Reaction coordinate
min- $\pi\pi^*G (La)$	4.36	2.06	0.0153	S <sub>1</sub> , S <sub>3</sub> , S <sub>6</sub>	C1-C2-NH2
min- <i>CT</i>	4.50	3.23	0.088	S <sub>5</sub>	Interbase distance + Ring T
min- $n\pi^*T$	4.77	3.91	0.0014	S <sub>2</sub> , S <sub>4</sub>	C4-O4

**Table S6.** Properties of the minima located on the *PES* of the first excited state determined for *anti-anti TG* at the PCM/M052X/6-31G(d) level of theory. ADE: adiabatic (with respect to the  $S_0$  at the FC region) energies; VEE: vertical (with respect to the  $S_0$  at the corresponding minima) emission energies; f: oscillator strengths; initial state optimized.  $S_4$  arrives directly to a  $S_1/S_0$  degeneracy region.<sup>9</sup> For completeness we have also characterized  $S_5$  that arrives to a CT minimum but being the *anti-syn TG* one and requiring a very large rearrangement of the bases.

	ADE (eV)	VEE (eV)	f	initial state
min- $\pi\pi^*G$ (La)	4.46	1.98	0.018	$S_2$
min- $\pi\pi^*T$	4.29	3.27	0.126	$S_3, S_{10}$
min- $n\pi^*T$	4.58	3.80	0.004	$S_1$

**Table S7.** Properties of the minima located on the *PES* of the first excited state determined for *syn-anti TG* at the PCM/M052X/6-31G(d) level of theory. ADE: adiabatic (with respect to the  $S_0$  at the FC region) energies; VEE: vertical (with respect to the  $S_0$  at the corresponding minima) emission energies; f: oscillator strengths; initial state optimized and main reaction coordinate.

	ADE (eV)	VEE (eV)	f	initial state	Reaction coordinate
min- $n\pi^*G$ (La)	4.55	1.17	0.003	$S_3$	C6-O6
min- <i>CT</i>	4.31	3.32	0.024	$S_2, S_{10}$	Interbase distance + Ring T
min- $n\pi^*T$	4.76	3.78	0.002	$S_1, S_4, S_5$	C4-O4

**Table S8.** Properties of the minima located on the *PES* of the first excited state determined for *syn-syn TG* at the PCM/M052X/6-31G(d) level of theory. ADE: adiabatic (with respect to the  $S_0$  at the FC region) energies; VEE: vertical (with respect to the  $S_0$  at the corresponding minima) emission energies; f: oscillator strengths; initial state optimized and main reaction coordinate.  $S_2 S_4$  arrives directly to  $S_1/S_0$  degeneracy regions.

	ADE (eV)	VEE (eV)	f	initial state	Reaction coordinate
min- $\pi\pi^*$ G (La)	4.44	2.08	0.0164	$S_3$	C1-C2-NH2
CI-CT				$S_2, S_5$ and $S_6$	Dimerization C5T-C8G
min- $n\pi^*$ T	4.80	3.90	0.0009	$S_1$	C4-O4

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