Electronic supplementary information

Photosensitizer-free singlet oxygen generation via charge transfer

transition involving molecular O² toward highly efficient oxidative

coupling of arylamines to azoaromatics

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Table of Contents

Experimental Section

1. Materials

Milli-Q water was obtained from a Millipore water purifier system (Milli-Q integral). *p*-chloroaniline, 5,5-dimethyl-1-pyrroline N-oxide (DMPO), *p*-toluidine, 2,2,6,6-tetramethylpiperidine (TEMP), and 2,4,6-trimethylaniline were procured from TCI. H₂O₂ test strips (MQuant test peroxide) were purchased from Supelco, Sigma-Aldrich. 1,4-diazabicyclo[2.2.2]octane (DABCO), and piperidine were purchased from SRL, India. Aniline hydrochloride, potassium phosphate (K3PO4), potassium carbonate (K2CO3) *p*-anisidine, acetonitrile (ACN), methanol (MeOH), benzene, mesitylene, 3,4-dimethylaniline, 2,4-dimethylaniline, and *p*-nitroaniline were purchased from Merck. 3-bromo-4-methylaniline, 3-fluoro-4-methylaniline, 3-chloro-4 methylaniline, 5,6,7,8-tetrahydro-2-naphthylamine, and 2-methoxyaniline hydrochloride were procured from BLD Pharmatech, India.

2. Methods

2.1. Procedure for synthesis of azo compounds

Initially, 0.05 mmol amine substrate was added to 5 ml Milli-Q water in a 15 ml glass vial. Subsequently, 2 equiv. K₃PO₄ was added to the above solution. The solution was then stirred for 3-5 minutes in the dark before illuminating the reaction mixture with 370 nm Kessil LED. The reaction mixture was irradiated for 90 min inside a homemade photocatalytic reactor equipped with Kessil LED and a portable fan under ambient conditions. The product was extracted with ethyl acetate, washed several times with water, and dried using anhydrous sodium sulfate. The solvent was evaporated using a rotary evaporator, and then the residue obtained was characterized using GC-MS. The obtained product was further purified using a silica gel column and was air-dried overnight for NMR analysis. For NMR analysis, the reaction was scaled up by three times.

2.2. Gas chromatography‐mass spectrometry measurements

Gas chromatography-mass spectrometry (GC-MS) spectra were recorded using a Shimadzu GC-MS, QP2010 mass spectrometer with a 30 m long Rxi-5Sil MS separation column with a 0.25 mm diameter and 0.25 µM thickness. Ethyl acetate was used as a solvent for the GC-MS analysis of the reaction mixture. The column oven temperature was set at 40 °C for 5 min, followed by a 20 °C/min ramp to 280 °C and held for 8 min.

2.3. Nuclear magnetic resonance measurements

AVANCE NEO Ascend 500 Bruker BioSpin, a 500 MHz spectrometer, was used to record ¹H and ¹³C nuclear magnetic resonance (NMR) spectra. Data for ¹H NMR spectra are reported as chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet), coupling constant (*J* Hz), and integration. The assignment data for ¹³C NMR spectra are reported as chemical shifts.

2.4. H2O² detection

The H2O² detection experiment was carried out under the standard optimized reaction conditions. Briefly, *p*-toluidine (0.05 mmol) and K₃PO₄ (2 equiv.) were added to 5 ml Milli-Q and stirred for 5 min. Subsequently, the above solution was irradiated with 370 nm Kessil LED for 90 min. Finally, 10-20 μl of the reaction mixture was dropped over the H₂O₂ test strips (MQuant test peroxide) to check the presence of H_2O_2 .

2.5. Scalability experiments

The scalability of the present synthesis approach was estimated by simply increasing the substrate (*p*-toluidine) concentration from 0.05 mmol to 5 mmol and 10 mmol. The 5 mmol reaction was carried out at standard reaction conditions under irradiation of one 370 nm Kessil LED (44 W), while to carry out the reaction with 10 mmol substrate, two 370 nm Kessil LEDs (44 W) were utilized.

2.6. Radical trapping experiment

Briefly, 0.05 mmol p-toluidine and 2 equiv. K₃PO₄ was added to a 1:1 mixture of methanol and water. The solution was then irradiated with a 370 nm Kessil LED and 1 equiv. 2,6-Di-tert-butyl-4-methylphenol (BHT) was added to the above reaction mixture to trap the radicals. The above mixture was irradiated for 90 min before conducting the HR-MS and GC-MS analysis.

2.7. Electron paramagnetic resonance experiment

Electron paramagnetic resonance (EPR) spectra were recorded on a JEOL EPR spectrometer (JES-FA200) to confirm the presence of reactive oxygen species (ROS). Initially, to check the presence of singlet oxygen in the aqueous solution, 5 ml Milli-Q was taken in a 15 ml glass vial and irradiated with a 370 nm Kessil LED. Subsequently, 5 equiv. TEMP was added to the above solution as a singlet oxygen scavenger. Next, 500 μL aliquot of this solution was collected after 30 min and transferred in a quartz tube for EPR analysis at 298 K.

Similarly, to check the presence of superoxide radicals in the reaction mixture, 5,5-dimethyl-1-pyrroline N-oxide (DMPO) was used as a radical scavenger. Briefly, 0.05 mmol p-toluidine and 2 equiv. K₃PO₄ was added to a 1:1 mixture of methanol and water. The solution was then irradiated with a 370 nm Kessil LED and 5 equiv. DMPO was added to the reaction mixture to trap the superoxide radical. Subsequently, 500 μL aliquot of this solution was collected after 30 min and transferred in a quartz tube for EPR analysis at 298 K.

2.8. Calculation of internal quantum yield for oxidative coupling of arylamines to azoaromatics

The internal quantum yield (ϕ _{int}) was calculated as per the formula given below,¹

$$
\varphi_{int} = \frac{moles\ of\ product\ formed}{moles\ of\ photons\ absorbed} \times 100
$$

The moles of the product formed were estimated using the conversion yield of *p*-toluidine to its corresponding azo compound while the moles of photons absorbed were estimated by multiplying photon flux (*qp*) with the total irradiation time (*t*). The photon flux was calculated using the following equation,

moles of photons absorbed =
$$
q_p \times t
$$
 (s) = $\frac{P_{abs} \times \lambda (m)}{hc \times (6.023 \times 10^{23})} \times t$ (s)

Where P_{abs} (J.s⁻¹) is the power of light absorbed by the reaction mixture, λ is the wavelength (m) of the LED light source, *h* is the Plank's constant (J.s), and c is the speed of light (m.s⁻¹).

The P_{abs} was calculated indirectly from the difference between incident light (P_0) and transmitted light (P_f) using the following Beers's law,

$$
A_{370} \times b = \log \frac{P_0}{P_f}
$$

$$
P_0 - P_f = P_{\text{abs}}
$$

Where '*b*' is the path length of the optical cell.

The calculated quantum yield is ~ 0.15 for the conversion of *p*-toluidine to its corresponding azo compound under standard reaction conditions.

Figure S1. UV-vis spectra of (A) mesitylene and (B) acetonitrile (ACN) under N₂ and O₂ atmospheres.

Figure S2. HR-MS spectrum of 9,10-diphenyl-9,10-epidioxyanthracene.

Figure S3. Time-dependent conversion yields for the oxidative coupling of 0.05 mmol *p*-toluidine in the presence of 2 equiv. K3PO⁴ in (A) methanol and (B) acetonitrile under the irradiation of 370 nm LED.

Figure S4. UV-vis spectrum of *p*-toluidine in water.

Figure S5. GC-MS spectrum for the oxidative coupling of *p***‐**toluidine and *p***‐**anisidine.

Figure S6. GC-MS spectrum for the oxidative coupling of p-toluidine and 3,4-dimethylaniline.

Figure S7. GC-MS spectrum for the oxidative coupling of p-toluidine and 2,4,6-trimethyaniline.

Figure S8. GC-MS spectrum for the oxidative coupling of *p***‐**toluidine and *p***‐**chloroaniline.

Figure S9. GC-MS spectrum for the oxidative coupling of *p***‐**chloroaniline and *p***‐**anisidine.

Figure S10. GC-MS spectrum for the oxidative coupling of p-toluidine and 5,6,7,8-tetrahydronaphthalene-1-amine.

Figure S11. GC-MS spectrum for the oxidative coupling of 3-fluoro-4-methylaniline and 2,4,6-trimethylaniline.

Figure S12. GC-MS spectrum for the oxidative coupling of 3,4-dimethylaniline and *p*-anisidine.

Figure S13. GC-MS spectrum for the oxidative coupling of aniline and *p***‐**toluidine.

Figure S14. GC-MS spectrum for the oxidative coupling of aniline and *p***‐**chloroaniline.

Figure S15. GC-MS spectrum for the oxidative coupling of *p***‐**chloroaniline and 2,4,6-trimethylaniline.

Figure S16. GC-MS spectrum for the oxidative coupling of 3-fluoro-4-methylaniline and 3,4-dimethylaniline.

Figure S17. GC-MS spectrum for the oxidative coupling of *p***‐**chloroaniline and 3,4-dimethylaniline.

Figure S18. GC-MS spectrum for the oxidative coupling of 2,4,6-trimethylaniline and *p*-anisidine.

Figure S19. GC-MS spectrum for the oxidative coupling of aniline and *p*-anisidine.

Figure S20. GC-MS spectrum for the oxidative coupling of 3-fluoro-4-methylaniline and *p*-anisidine.

Figure S21. GC-MS spectrum for the oxidative coupling of 3-fluoro-4-methylaniline and *p***‐**chloroaniline.

Figure S22. GC-MS spectrum for the oxidative coupling of 3-fluoro-4-methylaniline and p-toluidine.

Figure S23. GC-MS spectrum for the oxidative coupling of *p*-chloroaniline and 5,6,7,8-tetrahydronaphthalene-1-amine.

Figure S24. GC-MS spectrum for the oxidative coupling of 2,4,6-trimethylaniline and 3,4-dimethylaniline.

Figure S25. GC-MS spectrum for the oxidative coupling of 3,4-dimethylaniline and 5,6,7,8-tetrahydronaphthalene-1-amine.

Figure S26. GC-MS spectrum for the oxidative coupling of *p***‐**toluidine and *p***‐**nitroaniline.

Figure S27. GC-MS spectra of the oxidative coupling of 0.05 mmol *p*-toluidine under standard reaction conditions after (A)

15 and (B) 90 min of irradiation under 370 nm LED.

Figure S28. UV-vis spectra of different amine substrates (100 μM) in aqueous medium. (A) Aniline, (B) *p*-anisidine, (C) 2,4,6 trimethylaniline, (D) 3-chloro-4-methylaniline, (E) *p*-chloroaniline, (F) 5,6,7,8-tetrahydronapthalen-1-amine, and (G) *p*nitroaniline.

Figure S29. Changes in the UV-vis spectra of *p*-toluidine (100 μM) in the absence and presence of K₃PO₄ in aqueous medium.

Figure S30. HR-MS spectrum of BHT-*p*-toluidine adduct obtained after radical trap experiment with 1 equiv. BHT under standard reaction conditions in a 1:1 methanol-water mixture.

Figure S31. LC-MS spectrum of the hydrazobenzene intermediate obtained during the oxidative coupling of 0.05 mmol *p*toluidine under standard reaction conditions after 10 min of irradiation under 370 nm LED.

Figure S32. ¹H and ¹³C NMR spectra of 1,2-bis(4-methylphenyl)diazene.

Figure S33. ¹H and ¹³C NMR spectra of 1,2-bis(4-methoxyphenyl)diazene.

Figure S34. ¹H and ¹³C NMR spectra of 1,2-bis(2-methoxylphenyl)diazene.

Figure S35. ¹H and ¹³C NMR spectra of 1,2-bis(3,4-dimethylphenyl)diazene.

Figure S36. ¹H and ¹³C NMR spectra of 1,2-bis(2,4-dimethylphenyl)diazene.

Figure S37. ¹H and ¹³C NMR spectra of 1,2-bis(2,4,6-trimethylphenyl)diazene.

Figure S38. ¹H and ¹³C NMR spectra of azobenzene.

Figure S39. ¹H and ¹³C NMR spectra of 1,2-bis(4-chlorolphenyl)diazene.

Figure S40. ¹H and ¹³C NMR spectra of 1,2-bis(3-fluoro-4-methylphenyl)diazene.

Figure S41. ¹H and ¹³C NMR spectra of 1,2-bis(3-chloro-4-methylphenyl)diazene.

Figure S42. ¹H and ¹³C NMR spectra of 1,2-bis(3-bromo-4-methylphenyl)diazene.

Figure S43. ¹H and ¹³C NMR spectra of 1,2-bis(5,6,7,8-tetrahydronaphthalen-1-yl)diazene.

Table S1 Recent literatures on the synthesis of azo compounds

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