

## **SUPPORTING INFORMATION**

### **NeoMProbe: A New Class of Fluorescent Cellular and Tissue Membrane Probe**

Saurabh Anand,<sup>†</sup> Preeti Ravindra Bhoge,<sup>†</sup> Rakesh Raigawali,<sup>†</sup> Srinivas Vinod Saladi,<sup>± \*</sup> Raghavendra Kikkeri.<sup>\*†‡</sup>

<sup>†</sup>Indian Institute of Science Education and Research, Dr. Homi Bhabha Road, Pune 411008, India

<sup>±</sup>Department of Cell and Cancer Biology, University of Toledo, College of Medicine and Life Science, Toledo, OH 43614, USA

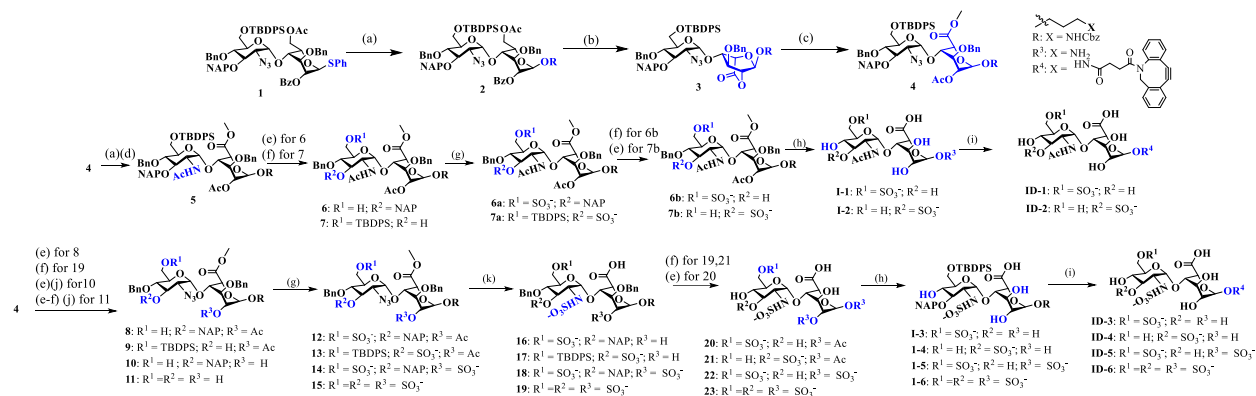
<sup>‡</sup>Department of CPAS, Jackson State University, Jackson, Mississippi, 39217, USA

Email. [rkikkeri@iiserpune.ac.in](mailto:rkikkeri@iiserpune.ac.in); [srinivas.saldi@utoledo.edu](mailto:srinivas.saldi@utoledo.edu)

### **1. General Information**

All chemicals were reagent grade and used without further purification unless otherwise noted. Reactions were carried out in anhydrous solvents under a nitrogen atmosphere. Reaction progress was monitored by analytical thin-layer chromatography (TLC) on Merck silica gel 60 F<sub>254</sub>. Spots on TLC plate were visualized under UV light or by dipping the TLC plate in CAM/ninhydrin solution followed by heating. Column chromatography was carried out using Fluka kiesel gel 60 (230-400 mesh). <sup>1</sup>H and <sup>13</sup>C NMR spectra of compounds were measured with Bruker 400 MHz, Bruker 600 MHz and Jeol 400 MHz using residual solvents as an internal reference (CDCl<sub>3</sub> δH 7.26 ppm, δC 77.3 ppm, CD<sub>3</sub>OD δH 3.31 ppm, δC 49.0 ppm, and D<sub>2</sub>O δH 4.79 ppm). The chemical shifts (δ) are reported in ppm and coupling constants (J) in Hz. UV-visible measurements were performed with Evolution 300 UV-visible spectrophotometer (Thermo Fisher Scientific, USA). Fluorescence spectra were measured with FluoroMax-4 spectrofluorometer (Horiba Scientific, U.S.A.). All microscopy images were captured using Leica SP8 confocal microscope and processed using Image J software.

## 2. Synthesis of Iduronic acid based Disaccharides



**Scheme S1. Synthesis of Iduronic acid based HS disaccharides** (a)  $R = \text{Linker}$ , NIS, TMSOTf, DCM,  $-10\text{ }^\circ\text{C}$ , 15 min; (b) NaOMe, MeOH, RT, 12 h; TEMPO, BAIB, DCM:H<sub>2</sub>O (1:1), RT; (c) 1 M LiOH, THF: H<sub>2</sub>O (1:1), RT, 2 h; MeI, K<sub>2</sub>CO<sub>3</sub>, DMF, 12 h; Ac<sub>2</sub>O, DCM:Py (1:1),  $0\text{ }^\circ\text{C}$ , 12 h; (d) Zn dust, THF:AcOH:Ac<sub>2</sub>O (3:2:1), RT, 12 h; (e) HF.Py, Py,  $0\text{ }^\circ\text{C}$ , 12 h; (f) DDQ, DCM:H<sub>2</sub>O (18:1), RT, 1 h; (g) SO<sub>3</sub>. Et<sub>3</sub>N, DMF,  $60\text{ }^\circ\text{C}$ , 24 h; (h) (i) LiOH.H<sub>2</sub>O, H<sub>2</sub>O, 12 h; (ii) H<sub>2</sub>, Pd(OH)<sub>2</sub>, MeOH, 48 h; (i) DBCO-NHS linker, Et<sub>3</sub>N, DMF, 24 h. (j) LiOH.H<sub>2</sub>O, H<sub>2</sub>O; (k) PMe<sub>3</sub>.THF, 24 h; (ii) SO<sub>3</sub>.Py. MeOH, 1N NaOH, 48 h.

### 1,6-Anhydro-[2-azido-4-O-benzyl-6-O-*tert*-butyldiphenylsilyl-3-O-(2 naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl]-(1 $\rightarrow$ 4)-O-2-O-benzoyl-3-O-benzyl- $\beta$ -L- idopyranose (1b)

To a solution of compound **1d** donor (1g, 1 eq) and acceptor (0.41 g, 0.8 eq) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was stirred under N<sub>2</sub> atmosphere in round bottom flask containing freshly dried 4 Å molecular sieves for 1 h. The mixture solution was cooled down to  $-78\text{ }^\circ\text{C}$  followed by the addition of NIS (0.46 g, 2.08 mmol) and TMSOTf (71  $\mu$ L, 0.39 mmol). The temperature was gradually increased to  $-20\text{ }^\circ\text{C}$  and reaction mixture was left for stirring for another 15 minutes. After 15 minutes reaction completion was monitored by TLC and quenched using few drops of Et<sub>3</sub>N. Molecular sieves were filtered using celite and organic layer was washed with aqueous Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and brine. The collected organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated and purified through silica gel column chromatography (ethyl acetate/ hexane= 1/12, v/v) to obtain compound **1e** in

65 % yield.  $^1\text{H}$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.08 – 8.04 (m, 2H), 7.83 (m, 4H), 7.70 – 7.67 (m, 4H), 7.62 – 7.54 (m, 2H), 7.51 – 7.42 (m, 7H), 7.42 – 7.35 (m, 5H), 7.31 – 7.27 (m, 3H), 7.24 – 7.18 (m, 3H), 7.17 – 7.13 (m, 2H), 5.52 (d,  $J$  = 1.8 Hz, 1H), 5.32 – 5.29 (m, 1H), 5.12 – 5.06 (m, 3H), 4.89 (t,  $J$  = 10.4 Hz, 2H), 4.76 (d,  $J$  = 10.8 Hz, 1H), 4.66 – 4.57 (m, 2H), 4.14 (d,  $J$  = 7.7 Hz, 1H), 4.10 – 3.99 (m, 3H), 3.88 (d,  $J$  = 1.8 Hz, 2H), 3.75 – 3.65 (m, 3H), 3.45 (dd,  $J$  = 10.3, 3.8 Hz, 1H), 1.07 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  165.8, 138.0, 137.5, 135.8, 135.7, 135.2, 133.4, 133.3, 133.2, 132.9, 129.9, 129.9, 129.8, 129.4, 128.6, 128.5, 128.4, 128.4, 128.1, 128.1, 128.0, 127.9, 127.8, 127.7, 127.0, 126.2, 126.1, 126.0, 99.7, 99.3, 80.0, 79.4, 78.9, 78.3, 77.4, 77.0, 76.7, 75.7, 75.5, 75.0, 74.2, 72.9, 65.7, 63.7, 62.6, 31.0, 26.9, 26.8, 25.3, 19.2. HR-ESI-MS ( $m/z$ ):  $[\text{M}+\text{Na}]^+$  calcd for  $\text{C}_{60}\text{H}_{61}\text{N}_3\text{O}_{10}\text{SiNa}$ , 1034.4024; found, 1034.4010.

**Phenyl-[2-azido-4-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl]-(1 $\rightarrow$ 4)-*O*-2-*O*-benzoyl-3-*O*-benzyl-6-*O*-acetyl-1-thio- $\alpha$ -L- idopyranoside (1)**

A solution of compound **1e** (1 g, 0.98 mmol, 1 eq) in  $\text{Ac}_2\text{O}$  (10 mL) was stirred at ice cold temperature for 15 minutes before the addition of  $\text{Cu}(\text{OTf})_2$  (0.035 g, 0.098 mmol, 0.1 eq). After 16 h, the reaction mixture was concentrated under reduced pressure and the residue was extracted with ethyl acetate,  $\text{NaHCO}_3$  and washed with brine. The combined organic layer was dried over  $\text{Na}_2\text{SO}_4$ , filtered, concentrated and preceded further without any purification. Next, the as obtained acetylated solution of compound (0.98 g, 0.88 mmol),  $\text{ZnI}_2$  (0.59 g, 1.84 mmol) and phenyl trimethylsilyl sulphide (0.50 g, 2.72 mmol) in  $\text{CH}_2\text{Cl}_2$  (15 mL) was stirred under  $\text{N}_2$  atmosphere for 2 h. Upon completion, the reaction mixture was filtered through celite, evaporated and purified through silica gel column chromatography (ethyl acetate/ hexane= 1/5, v/v) to obtain compound **1** in 85 % yield.  $^1\text{H}$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.20 – 8.16 (m, 2H), 7.81 – 7.74 (m, 3H), 7.70 (d,  $J$  = 1.5 Hz, 1H), 7.68 – 7.66 (m, 2H), 7.65 (d,  $J$  = 1.5 Hz, 1H), 7.58 – 7.55 (m, 3H), 7.51 – 7.46 (m, 4H), 7.43 – 7.37 (m, 5H), 7.36 – 7.27 (m, 14H), 7.19 – 7.16 (m, 2H), 5.62 (s, 1H), 5.40 (t,  $J$  = 2.5 Hz, 1H), 4.99 (d,  $J$  = 11.6 Hz, 1H), 4.90 - 4.88 (m, 1H), 4.82 – 4.68 (m, 3H), 4.63 (d,  $J$  = 3.7 Hz, 1H), 4.55 (d,  $J$  = 10.6 Hz, 1H), 4.41 (dd,  $J$  = 11.6, 8.1 Hz, 1H), 4.24 – 4.15 (m, 3H), 4.02 – 3.98 (m, 1H), 3.88 (d,  $J$  = 11.5, 1H), 3.81 – 3.65 (m, 4H), 3.36 (dd,  $J$  = 9.9, 3.7 Hz, 1H), 1.91 (s, 3H), 1.05 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz, CHLOROFORM-D)  $\delta$  170.5, 165.9, 138.1, 137.4, 136.0, 135.7, 135.6, 135.2, 133.6, 133.4, 133.1, 133.1, 131.8,

130.0, 130.0, 129.8, 129.8, 129.0, 128.7, 128.6, 128.5, 128.3, 128.2, 128.2, 128.1, 127.9, 127.9, 127.8, 127.8, 127.7, 127.6, 127.1, 126.2, 126.2, 126.1, 99.1, 86.0, 80.9, 77.9, 77.5, 77.4, 77.2, 76.8, 75.4, 75.2, 75.1, 73.3, 72.9, 72.0, 69.8, 66.5, 64.2, 63.2, 62.3, 27.0, 20.8, 19.4. HR-ESI-MS ( $m/z$ ):  $[M+Na]^+$  calcd for  $C_{68}H_{69}N_3O_{11}SSiNa$ , 1186.4320; found, 1186.4315.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[2-azido-4-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl]-(1 $\rightarrow$ 4)-*O*-2-*O*-benzoyl-3-*O*-benzyl-6-*O*-acetyl-1- $\alpha$ -L- idopyranoside (2)**

A solution of donor **1** (0.90 g, 0.42 mmol) and linker benzyl (3-hydroxypropyl) carbamate (0.097 g, 0.46 mmol) in  $CH_2Cl_2$  (15 mL) was stirred under  $N_2$  atmosphere in round bottom flask containing freshly dried 4 Å molecular sieves for 1 h. Next, NIS (0.15 g, 0.67 mmol) and TfOH (7.4  $\mu$ L, 0.084 mmol) was added at room temperature and reaction completion was monitored by TLC, quenched using few drops of  $NEt_3$ . Molecular sieves were filtered using celite and organic layer was washed with aqueous  $Na_2S_2O_3$  and brine. The collected organic layer was dried over  $Na_2SO_4$ , filtered, concentrated and purified through silica gel column chromatography (ethyl acetate/ hexane= 1/3, v/v) to obtain compound **2** in 87 % yield.  $^1H$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.17 – 8.09 (m, 2H), 7.84 – 7.72 (m, 3H), 7.69 – 7.63 (m, 4H), 7.58 (s, 1H), 7.50 – 7.45 (m, 2H), 7.43 – 7.38 (m, 2H), 7.37 – 7.27 (m, 20H), 7.20 – 7.14 (m, 2H), 5.64 (t,  $J = 6.0$  Hz, 1H), 5.07 (d,  $J = 6.9$  Hz, 3H), 4.94 (d,  $J = 1.4$  Hz, 1H), 4.81 (dd,  $J = 11.5, 8.4$  Hz, 2H), 4.67 (t,  $J = 11.3$  Hz, 2H), 4.57 (d,  $J = 10.6$  Hz, 1H), 4.43 – 4.21 (m, 4H), 4.07 (dd,  $J = 10.9, 4.1$  Hz, 1H), 4.04 – 3.95 (m, 2H), 3.84 (dd,  $J = 10.4, 5.7$  Hz, 2H), 3.78 – 3.65 (m, 3H), 3.58 – 3.40 (m, 3H), 3.30 – 3.18 (m, 2H), 1.92 (s, 3H), 1.85 (p,  $J = 6.9, 5.8$  Hz, 2H), 1.04 (s, 9H).  $^{13}C$  NMR (101 MHz, CHLOROFORM-D)  $\delta$  170.6, 165.8, 156.7, 138.1, 137.6, 136.8, 136.0, 135.7, 135.2, 133.6, 133.4, 133.3, 133.1, 130.0, 129.9, 129.8, 128.7, 128.5, 128.4, 128.4, 128.2, 128.1, 128.1, 127.9, 127.8, 127.7, 127.7, 127.1, 126.2, 126.1, 98.8, 98.2, 80.8, 77.9, 77.4, 77.1, 76.8, 75.4, 75.2, 74.6, 73.1, 72.6, 72.0, 68.7, 67.1, 66.7, 65.3, 64.2, 63.4, 62.3, 39.8, 29.5, 26.9, 20.8, 19.4. HR-ESI-MS ( $m/z$ ):  $[M+Na]^+$  calcd for  $C_{73}H_{78}N_4O_{14}SSiNa$ , 1285.5181; found, 1285.5174.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-azido-4-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-3-*O*-benzyl-1- $\alpha$ - L-idopyranosyluronate (3)**

A solution of compound **2** (1g, 1 eq) in DCM:MeOH (1:1, 14mL), NaOMe(1.2 eq) was added and kept it for stirring for 12 h at room temperature followed by quenching with IR Na<sup>+</sup> resin, filtered, evaporated and purified by silica gel column chromatography (ethyl acetate/ hexane= 1/2.5, v/v). After that compound was dissolved in DCM:H<sub>2</sub>O (1:1) followed by addition by of TEMPO (0.2 eq) and BAIB (3 eq) and kept it stirring for 12 h at room temperature followed by extraction with saturated aqueous solution NH<sub>4</sub>Cl. The organic layer was evaporated and purified by silica gel column chromatography. Then the compound was dissolved in THF:H<sub>2</sub>O (1:1) followed by addition of lithium hydroxide (10 eq) and kept it on stirring for 2 h at room temperature followed by evaporation and dried over vacuum. After that reaction mixture was dissolved in dry DMF and potassium carbonate (3 eq), methyl iodide (9 eq) was added and kept it on stirring for at room temperature. After 12 h extraction was done using 10 % HCl solution and organic layer was evaporated, purified using silica gel chromatography using ethyl acetate/hexane (2/3) to obtain compound **3** 84%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.84 – 7.68 (m, 6H), 7.68 – 7.61 (m, 2H), 7.50 – 7.27 (m, 19H), 7.26 – 7.13 (m, 5H), 5.56 (d, *J* = 6.0 Hz, 1H), 5.04 (s, 2H), 5.00 (d, *J* = 10.6 Hz, 3H), 4.93 – 4.88 (m, 2H), 4.80 (s, 1H), 4.76 (d, *J* = 11.4 Hz, 1H), 4.60 – 4.56 (m, 2H), 4.12 4.11 (m, 1H), 3.97 (dd, *J* = 11.6, 2.9 Hz, 1H), 3.93 – 3.80 (m, 5H), 3.71 - 3.67(m, 1H), 3.63 (s, 3H), 3.61 – 3.54 (m, 3H), 3.54 – 3.49 (m, 1H), 3.44 (d, *J* = 5.5 Hz, 1H), 1.90 – 1.77 (m, 2H), 1.08 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 169.8, 156.5, 138.3, 137.3, 136.6, 135.9, 135.6, 135.0, 133.4, 133.3, 133.1, 133.1, 129.8, 129.8, 128.7, 128.6, 128.4, 128.4, 128.3, 128.3, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.6, 127.2, 127.2, 127.1, 126.2, 126.2, 126.1, 126.0, 101.5, 94.9, 81.1, 77.6, 77.4, 77.3, 77.1, 76.7, 76.1, 74.8, 72.8, 71.9, 70.8, 70.7, 68.0, 67.0, 66.9, 66.6, 63.9, 62.1, 53.4, 52.4, 39.9, 29.5, 26.9, 19.4. HR-ESI-MS (*m/z*): [M+Na]<sup>+</sup> calcd for C<sub>65</sub>H<sub>72</sub>N<sub>4</sub>O<sub>13</sub>SiNa, 1167.4763; found, 1167.5478.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-azido-4-O-benzyl-6-O-tert-butyl)diphenylsilyl-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ - L-idopyranosyluronate (4)**

A solution of compound **3** (1g, 1 eq) in DCM:Py (1:1, 14mL), acetic anhydride (5 eq) and DMAP (0.2 eq) was added and kept it for stirring for 12 h at 0 °C followed by washing with 1N HCl 3 times and organic layer was evaporated and purified by silica gel column chromatography (ethyl acetate/ hexane= 1/4, v/v) to obtain **4**, 86 %. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.87 –

7.72 (m, 4H), 7.70 - 7.68 (m, 2H), 7.64 - 7.62 (m, 2H), 7.52 - 7.43 (m, 3H), 7.42 - 7.26 (m, 17H), 7.26 - 7.14 (m, 3H), 5.44 (t, J = 5.7 Hz, 1H), 5.05 (s, 2H), 5.02 - 4.91 (m, 3H), 4.90 (s, 1H), 4.87 - 4.80 (m, 2H), 4.79 - 4.71 (m, 2H), 4.65 (m, 2H), 4.02 - 3.93 (m, 3H), 3.91 - 3.81 (m, 4H), 3.76 - 3.68 (m, 1H), 3.60 - 3.52 (m, 4H), 3.43 - 3.31 (m, 2H), 3.28 - 3.20 (m, 1H), 2.07 (s, 3H), 1.86 - 1.73 (m, 2H), 1.06 (s, 8H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 170.1, 169.4, 156.5, 138.3, 137.3, 136.6, 135.8, 135.6, 135.1, 133.4, 133.3, 133.1, 133.1, 129.7, 128.5, 128.4, 128.4, 128.4, 128.3, 128.2, 128.0, 128.0, 128.0, 127.8, 127.7, 127.7, 127.6, 127.6, 127.5, 127.2, 127.1, 127.0, 126.1, 126.0, 126.0, 98.3, 97.3, 79.9, 77.9, 77.3, 77.0, 76.7, 75.5, 74.7, 72.6, 72.4, 72.0, 71.4, 68.0, 67.8, 67.6, 66.6, 63.6, 62.2, 52.2, 39.5, 29.4, 26.9, 26.9, 20.8, 20.7, 19.4. HR-ESI-MS (*m/z*): [M+Na]<sup>+</sup> calcd for C<sub>67</sub>H<sub>74</sub>N<sub>4</sub>O<sub>14</sub>SiNa, 1209.4868; found, 1209.4862.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-acetamido-4-O-benzyl-6-O-tert-butyl)diphenylsilyl-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (5)**

A solution of compound **4** (1g, 1 eq) in THF:CH<sub>3</sub>COOH:Ac<sub>2</sub>O (3:2:1, 18mL), zinc dust (30 eq) was added and kept it for stirring for 12 h at room temperature followed by celite filtration and organic layer was evaporated. After that compound was re-dissolved in ethyl acetate and washed with 10 % sodium bicarbonate 3 times followed by evaporation of organic layer, purification by silica gel column chromatography (ethyl acetate/ hexane= 1/2, v/v) to obtain **5**, 81 %. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.84 - 7.63 (m, 8H), 7.47 - 7.10 (m, 24H), 5.02 (s, 1H), 4.98 (d, J = 12.0 Hz, 1H), 4.92 (d, J = 11.2 Hz, 1H), 4.89 - 4.83 (m, 2H), 4.82 - 4.76 (m, 3H), 4.72 (d, J = 11.2 Hz, 1H), 4.48 (s, 1H), 4.37 (td, J = 10.1, 3.5 Hz, 1H), 4.05 (d, J = 2.9 Hz, 1H), 3.99 - 3.83 (m, 3H), 3.83 - 3.73 (m, 2H), 3.64 (s, 3H), 3.63 - 3.56 (m, 2H), 3.55 - 3.45 (m, 1H), 3.42 - 3.30 (m, 1H), 3.26 - 3.14 (m, 1H), 1.83 (s, 3H), 1.78 (m, 2H), 1.70 (s, 3H), 1.07 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 170.2, 169.5, 169.4, 156.5, 138.5, 136.9, 136.6, 135.9, 135.8, 135.6, 135.6, 133.4, 133.3, 133.1, 133.0, 129.8, 128.6, 128.5, 128.5, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.1, 128.0, 127.9, 127.9, 127.8, 127.7, 127.7, 127.6, 127.6, 127.4, 127.4, 127.4, 126.8, 126.2, 126.2, 126.1, 98.2, 95.3, 80.5, 77.9, 77.5, 77.2, 76.9, 75.2, 74.8, 73.1, 72.4, 68.0, 67.7, 67.7, 66.6, 62.6, 52.4, 52.2, 39.3, 29.4, 27.0, 26.9, 26.9, 23.3, 20.7, 19.4. HR-ESI-MS (*m/z*): [M+Na]<sup>+</sup> calcd for C<sub>69</sub>H<sub>78</sub>N<sub>2</sub>O<sub>15</sub>SiNa, 1225.5069; found, 1225.5059.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-acetamido-4-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-3-*O*-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-2-*O*-acetyl-3-*O*-benzyl-1- $\alpha$ -L-idopyranosyluronate (7a)**

To a solution of starting material **5** (1 mmol, 1 eq) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) and H<sub>2</sub>O (340  $\mu$ L) was added DDQ (5 mmol, 5 eq) in 4 portions over the interval of 20 min. After 1 h, the reaction mixture was quenched using NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub> and washed with brine. The collected organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated and purified through silica gel column chromatography. After that, compound was dissolved to 1 mmol in DMF (2 mL) was added SO<sub>3</sub>.NEt<sub>3</sub> complex (10 mmol per -OH) under N<sub>2</sub> atmosphere and stirred at 70 ° C for 3 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1/10 v/v) to obtain product **7a** in 71 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>)  $\delta$  7.82 – 7.66 (m, 4H), 7.50 – 7.24 (m, 21H), 5.39 – 5.22 (m, 2H), 5.14 – 5.02 (m, 3H), 5.00 – 4.89 (m, 2H), 4.82 – 4.59 (m, 4H), 4.23 – 4.09 (m, 2H), 4.04 – 3.81 (m, 6H), 3.71 (s, 3H), 3.57 (dd, J = 10.3, 5.4 Hz, 1H), 3.27 (dq, J = 19.7, 6.7 Hz, 2H), 2.17 (s, 3H), 2.03 (s, 3H), 1.89 – 1.73 (m, 2H), 1.12 (s, 9H). <sup>13</sup>C NMR (101 MHz, MeOD)  $\delta$  172.6, 170.6, 170.0, 157.3, 138.6, 137.6, 136.9, 135.6, 135.4, 133.3, 133.0, 129.6, 128.2, 128.1, 128.0, 128.0, 127.9, 127.7, 127.6, 127.5, 127.5, 127.5, 127.4, 127.2, 98.6, 96.4, 78.4, 76.1, 74.5, 73.0, 72.4, 72.2, 71.8, 68.3, 68.1, 66.5, 66.0, 62.7, 54.0, 53.8, 51.9, 48.5, 48.3, 48.1, 47.9, 47.7, 47.4, 47.2, 44.3, 38.0, 29.4, 29.3, 26.2, 22.0, 20.0, 18.8. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>58</sub>H<sub>69</sub>N<sub>2</sub>O<sub>18</sub>SSi<sup>-</sup>, 1141.4041; found, 1141.4019.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-azido-4-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-3-*O*-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-2-*O*-acetyl-3-*O*-benzyl-1- $\alpha$ -L-idopyranosyluronate (13)**

To a solution of starting material **4** (1 mmol, 1 eq) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) and H<sub>2</sub>O (340  $\mu$ L) was added DDQ (5 mmol, 1 eq) in 4 portions over the interval of 20 min. After 1 h, the reaction mixture was quenched using NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub> and washed with brine. The collected organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated and purified through silica gel column chromatography. After that, compound was dissolved to 1 mmol in DMF (2 mL) was added SO<sub>3</sub>.NEt<sub>3</sub> complex (10 mmol per -OH) under N<sub>2</sub> atmosphere and stirred at 70 ° C for 3

days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1/10 v/v) to obtain product **13** 77 % in yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.79 – 7.73 (m, 4H), 7.47 – 7.28 (m, 4H), 5.35 (d, J = 10.7 Hz, 1H), 5.21 – 5.05 (m, 4H), 5.03 – 4.91 (m, 3H), 4.82 – 4.63 (m, 4H), 4.29 – 4.15 (m, 2H), 4.01 – 3.77 (m, 5H), 3.73 (s, 3H), 3.60 (m, 1H), 3.39 – 3.17 (m, 4H), 2.24 (s, 3H), 1.84 (q, J = 5.9 Hz, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 170.8, 170.3, 157.3, 138.5, 137.6, 136.9, 135.6, 135.4, 133.3, 133.1, 129.7, 128.3, 128.2, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.5, 127.5, 127.3, 98.5, 96.9, 78.7, 76.4, 74.6, 72.6, 72.3, 71.2, 68.1, 68.0, 66.6, 66.1, 62.8, 62.4, 51.9, 48.5, 48.3, 48.1, 47.9, 47.7, 47.5, 47.3, 46.6, 38.1, 29.4, 29.3, 26.3, 19.9, 18.9. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>56</sub>H<sub>65</sub>N<sub>4</sub>O<sub>17</sub>SiS<sup>-</sup>, 1125.3840; found, 1125.3839.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-acetamido-4-O-benzyl-3-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1→4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ - L-idopyranosyluronate (7b)**

A solution of compound **7a** (1g, 1 eq) in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that in reaction mixture was evaporated followed by purification by silica gel column chromatography (MeOH/DCM = 1/10, v/v) to obtain **7b** in 85 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.55 – 7.44 (m, 2H), 7.41 – 7.25 (m, 13H), 5.28 – 5.15 (m, 2H), 5.07 (d, J = 2.0 Hz, 2H), 5.00 (d, J = 2.7 Hz, 1H), 4.91 (d, J = 3.3 Hz, 1H), 4.77 – 4.66 (m, 3H), 4.59 (d, J = 10.5 Hz, 1H), 4.15 (t, J = 3.6 Hz, 1H), 4.04 – 3.90 (m, 2H), 3.84 (s, 4H), 3.81 (d, J = 2.1 Hz, 2H), 3.79 – 3.71 (m, 1H), 3.66 (t, J = 9.2 Hz, 1H), 3.60 – 3.50 (m, 1H), 3.23 (m, 2H), 2.13 (s, 3H), 1.94 (s, 3H), 1.87 – 1.72 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 172.5, 170.7, 170.2, 157.4, 138.7, 137.8, 137.0, 128.4, 128.1, 127.8, 127.6, 127.5, 127.5, 127.5, 127.2, 98.7, 97.1, 77.8, 75.9, 74.5, 73.3, 72.9, 72.9, 72.2, 68.6, 68.3, 66.4, 66.0, 60.4, 53.9, 51.9, 48.6, 48.4, 48.2, 48.0, 47.8, 47.6, 47.3, 47.1, 37.9, 29.4, 29.3, 21.9, 21.8, 20.0. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>42</sub>H<sub>51</sub>N<sub>2</sub>O<sub>18</sub>S<sup>-</sup>, 903.2863; found, 903.2857.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-azido-4-O-benzyl-3-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1→4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ - L-idopyranosyluronate (17)**



A solution of compound **13** (1g, 1eq) in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that in reaction mixture was evaporated followed by purification by silica gel column chromatography (MeOH/DCM = 1/10, v/v) to obtain **17** in 85 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.51 – 7.45 (m, 2H), 7.40 – 7.25 (m, 13H), 5.24 (d, J = 10.4 Hz, 1H), 5.09 – 5.04 (m, 4H), 4.90 (d, J = 3.3 Hz, 1H), 4.84 (d, J = 1.8 Hz, 1H), 4.76 – 4.68 (m, 2H), 4.58 (d, J = 10.5 Hz, 2H), 4.20 (t, J = 3.6 Hz, 1H), 4.06 (t, J = 4.0 Hz, 1H), 3.81 (s, 3H), 3.81 – 3.76 (m, 3H), 3.74 – 3.50 (m, 3H), 3.30 – 3.16 (m, 3H), 2.17 (s, 3H), 1.84 – 1.73 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 171.0, 170.4, 157.4, 138.6, 137.7, 136.9, 128.4, 128.3, 128.2, 128.2, 128.1, 127.9, 127.8, 127.7, 127.7, 127.6, 127.5, 127.5, 127.3, 98.7, 97.8, 78.6, 76.2, 74.5, 72.9, 72.4, 72.4, 68.4, 68.1, 66.4, 66.1, 62.3, 60.3, 51.9, 48.6, 48.4, 48.2, 48.0, 47.7, 47.5, 47.3, 47.1, 38.0, 29.3, 19.9. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>40</sub>H<sub>47</sub>N<sub>4</sub>O<sub>17</sub>S<sup>-</sup>, 887.2668; found, 887.2657.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-acetamido-4-O-benzyl-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (6)**

A solution of compound **5** (1g, 1 eq) in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that in reaction mixture ethyl acetate was added and washed with 1N HCl 3 times followed by evaporation of organic layer, purification by silica gel column chromatography (ethyl acetate/ hexane= 2/3, v/v) to obtain **6**, 92 %. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.84 – 7.67 (m, 4H), 7.49 – 7.42 (m, 2H), 7.40 – 7.26 (m, 12H), 7.26 – 7.16 (m, 4H), 5.91 – 5.80 (m, 1H), 5.46 (t, J = 5.8 Hz, 1H), 5.11 – 5.01 (m, 2H), 4.97 – 4.83 (m, 3H), 4.81 – 4.64 (m, 5H), 4.63 – 4.46 (m, 2H), 4.25 (td, J = 9.8, 3.6 Hz, 1H), 3.96 (q, J = 3.4, 2.9 Hz, 1H), 3.85 – 3.79 (m, 1H), 3.77 (s, 3H), 3.73 – 3.62 (m, 3H), 3.61 – 3.53 (m, 3H), 3.46 – 3.34 (m, 1H), 3.27 – 3.18 (m, 1H), 1.87 – 1.73 (m, 5H), 1.65 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 169.9, 169.5, 169.4, 156.5, 138.1, 136.8, 136.6, 135.7, 133.2, 133.0, 128.7, 128.6, 128.5, 128.5, 128.5, 128.5, 128.4, 128.3, 128.3, 128.2, 128.1, 128.1, 128.0, 128.0, 127.9, 127.8, 127.7, 127.7, 127.6, 127.5, 126.6, 126.3, 126.1, 126.1, 126.0, 98.3, 95.7, 80.1, 77.8, 77.4, 77.1, 76.8, 75.0, 74.8, 72.6, 72.2, 70.3, 69.7, 68.0, 67.9, 67.5, 66.7, 61.5, 52.4, 52.3, 39.5, 29.4, 23.3, 20.7. HR-ESI-MS (*m/z*): [M+Na]<sup>+</sup> calcd for C<sub>53</sub>H<sub>60</sub>N<sub>2</sub>O<sub>15</sub>Na, 987.3891; found, 987.3875.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(2-azido-4-O-benzyl-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (8)**

A solution of compound **4** (1g, 1 eq) in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that reaction mixture was evaporated followed by purification by silica gel column chromatography (ethyl acetate/ hexane= 1/10, v/v) to obtain **10** in, 87 %. <sup>1</sup>H NMR (400 MHz, Chloroform-d)  $\delta$  7.84 – 7.70 (m, 4H), 7.48 – 7.41 (m, 3H), 7.39 – 7.26 (m, 11H), 7.25 (m, 3H), 5.57 (s, 1H), 5.05 (s, 1H), 5.01 (s, 1H), 4.97 (s, 2H), 4.87 – 4.82 (m, 2H), 4.69 – 4.59 (m, 2H), 4.50 (d, J = 12.3 Hz, 1H), 4.07 (s, 1H), 3.92 – 3.82 (m, 2H), 3.80 (s, 3H), 3.76 (d, J = 2.2 Hz, 1H), 3.74 – 3.64 (m, 3H), 3.64 – 3.57 (m, 2H), 3.54 – 3.52 (m, 2H), 3.49 – 3.43 (m, 2H), 3.30 – 3.20 (m, 1H), 1.90 – 1.77 (m, 2H). <sup>13</sup>C NMR (101 MHz, CHLOROFORM-D)  $\delta$  170.1, 156.6, 137.9, 137.3, 136.6, 135.0, 133.4, 133.1, 128.8, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.1, 128.0, 128.0, 127.8, 127.7, 127.1, 126.2, 126.1, 126.1, 101.7, 95.0, 81.1, 77.4, 77.3, 77.1, 76.8, 76.0, 74.8, 72.3, 71.4, 71.1, 68.1, 66.9, 66.8, 63.8, 61.3, 52.8, 40.0, 29.5. HR-ESI-MS (*m/z*): [M+Na] calcd for C<sub>42</sub>H<sub>48</sub>N<sub>4</sub>NaO<sub>13</sub>, 839.3116; found, 839.3110.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(methyl(2-acetamido-6-O-sulfonato-4-O-benzyl-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-2-O-acetyl-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (6a)**

To a solution of starting material **6** (1 mmol, 1 eq) in DMF (2 mL) was added SO<sub>3</sub>.NEt<sub>3</sub> complex (10 mmol per -OH, 10 eq) under N<sub>2</sub> atmosphere and stirred at 70 °C for 3 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1/10 v/v) to obtain product **6a** 78 %. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>)  $\delta$  7.86 – 7.78 (m, 2H), 7.78 – 7.71 (m, 1H), 7.52 – 7.45 (m, 2H), 7.44 – 7.26 (m, 16H), 5.08 (s, 2H), 4.99 – 4.96 (m, 3H), 4.94 – 4.93(m, 1H), 4.85 – 4.72 (m, 3H), 4.42 – 4.33 (m, 2H), 4.29 (dd, J = 10.2, 3.3 Hz, 1H), 4.24 (s, 1H), 4.01 (s, 1H), 3.92 (d, J = 8.7 Hz, 1H), 3.89 – 3.83 (m, 1H), 3.82 (s, 3H), 3.78 – 3.67 (m, 2H), 3.57 (m, 1H), 3.34 – 3.18 (m, 1H), 2.01 (s, 3H), 1.91 (s, 3H), 1.82 (m, 2 H). <sup>13</sup>C NMR (101 MHz, MeOD)  $\delta$  171.8, 170.2, 170.0, 157.4, 138.4, 137.6, 137.0, 135.9, 133.4, 133.1, 128.3, 128.1, 128.1, 127.9, 127.9, 127.7, 127.6, 127.6, 127.5, 127.5, 127.3, 127.3, 126.0, 125.8, 125.6, 125.5, 98.6, 94.8, 80.4, 77.8, 74.9, 74.6, 72.0, 70.8, 70.4, 69.2, 68.1, 67.7, 66.4, 66.0, 52.7, 51.7, 48.6, 48.4, 48.1,

47.9, 47.7, 47.5, 47.3, 47.1, 37.9, 29.3, 21.8, 19.7. HR-ESI-MS ( $m/z$ ):  $[M+Na]^+$  calcd for  $C_{53}H_{59}N_2O_{18}S^-$ , 1043.3489; found, 1043.3485.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-azido-4-*O*-benzyl-6-*O*-sulfonato-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl))]-( $1\rightarrow4$ )-*O*-2-*O*-acetyl-3-*O*-benzyl-1- $\alpha$ -L-idopyranosyluronate (12)**

A solution of compound **4** (1 mmol, 1 eq) in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that reaction mixture was evaporated followed by purification by silica gel column chromatography (ethyl acetate/ hexane= 1/10, v/v). After that, compound was dissolved to (1 mmol, 1 eq) in DMF (2 mL) was added  $SO_3.NEt_3$  complex (10 mmol per -OH) under  $N_2$  atmosphere and stirred at 70 °C for 3 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1/10 v/v) to obtain product **12** in 81% yield.  $^1H$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  7.88 – 7.72 (m, 4H), 7.51 – 7.44 (m, 3H), 7.41 – 7.32 (m, 10H), 7.35 – 7.26 (m, 5H), 5.08 (s, 2H), 5.03 (d,  $J = 11.3$  Hz, 1H), 5.01 – 4.93 (m, 3H), 4.95 – 4.86 (m, 2H), 4.82 (d,  $J = 10.8$  Hz, 1H), 4.79 – 4.65 (m, 2H), 4.41 – 4.29 (m, 2H), 4.16 (t,  $J = 3.1$  Hz, 1H), 3.98 (d,  $J = 3.2$  Hz, 1H), 3.97 – 3.87 (m, 2H), 3.83 (m, 1H), 3.80 (s, 3H), 3.79 – 3.72 (m, 1H), 3.58 – 3.52 (m, 1H), 3.32 – 3.18 (m, 2H), 2.05 (s, 3H), 1.80 (m, 2H).  $^{13}C$  NMR (101 MHz, MeOD)  $\delta$  170.3, 170.1, 157.4, 138.3, 137.6, 135.5, 133.3, 133.1, 128.1, 128.1, 127.9, 127.9, 127.8, 127.7, 127.6, 127.6, 127.5, 127.5, 127.4, 127.3, 127.3, 126.2, 125.8, 125.6, 125.6, 98.6, 97.5, 79.4, 77.7, 74.5, 74.5, 72.8, 72.0, 70.5, 68.0, 67.6, 66.3, 66.0, 65.5, 63.2, 51.7, 48.5, 48.3, 48.1, 47.9, 47.7, 47.4, 47.2, 47.0, 38.0, 29.2, 19.6. HR-ESI-MS ( $m/z$ ):  $[M]^-$  calcd for  $C_{51}H_{55}N_4O_{17}S^-$ , 1027.3288; found, 1027.3275.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl-(2-azido-4-*O*-benzyl-6-*O*-sulfonato-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl))]-( $1\rightarrow4$ )-*O*-2-*O*-sulfonato-3-*O*-benzyl-1- $\alpha$ -L-idopyranosyluronate (14)**

The compound **4** was dissolved to (1 mmol, 1eq) in DMF (2 mL) was added  $SO_3.NEt_3$  complex (10 mmol per -OH) under  $N_2$  atmosphere and stirred at 70 °C for 4 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1.3/10 v/v) to obtain product **14** in 76% yield.  $^1H$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  7.91 – 7.73 (m, 4H), 7.54 – 7.19 (m, 18H),

5.22 (s, 1H), 5.15 – 4.98 (m, 5H), 4.80 (d, J = 10.9 Hz, 2H), 4.67 (d, J = 11.8 Hz, 2H), 4.52 (s, 1H), 4.37 – 4.25 (m, 3H), 4.14 (t, J = 2.8 Hz, 1H), 4.03 – 3.98 (m, 1H), 3.87 – 3.80 (m, 2H), 3.78 (s, 3H), 3.73 – 3.56 (m, 3H), 3.40 (dd, J = 10.3, 3.4 Hz, 1H), 3.33 – 3.17 (m, 2H), 1.89 – 1.77 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 170.8, 157.5, 138.3, 137.7, 136.9, 135.8, 133.3, 133.1, 128.1, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.6, 127.6, 127.5, 127.3, 127.2, 126.3, 125.8, 125.7, 125.6, 99.4, 96.4, 79.8, 77.6, 74.9, 74.5, 71.8, 71.3, 71.1, 71.0, 70.5, 67.1, 66.4, 66.1, 65.7, 63.5, 53.9, 51.9, 48.4, 48.2, 47.9, 47.7, 47.5, 47.3, 47.1, 38.1, 29.3, 29.2. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>42</sub>H<sub>46</sub>N<sub>4</sub>O<sub>19</sub>S<sub>2</sub><sup>2-</sup>, 487.1104; found, 487.1098.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl-(2-azido-4-*O*-benzyl-6-*O*-sulfonato-3-*O*-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-2-*O*-sulfonato-3-*O*-benzyl-1- $\alpha$ -L-idopyranosyluronate (15)**

To a solution of starting material **4** (1 mmol, 1 eq) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) and H<sub>2</sub>O (340  $\mu$ L) was added DDQ (5 mmol) portion wise over the interval of 20 min. After 1 h, the reaction mixture was quenched using NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub> and washed with brine. The collected organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated and purified through silica gel column chromatography. After that, solution of product in dry pyridine (5 mL) 70% HF.Py complex (5 eq) was added at 0 °C and kept it for stirring for 12 h. After that in reaction mixture was evaporated followed by purification by silica gel column chromatography (ethyl acetate/hexane = 1/2, v/v). Then, compound was dissolved to 1 mmol in DMF (2 mL) was added SO<sub>3</sub>.NEt<sub>3</sub> complex (10 mmol per -OH) under N<sub>2</sub> atmosphere and stirred at 70 °C for 4 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1.5/10 v/v) to obtain product **15** in 62% yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.58 – 7.20 (m, 15H), 5.17 (s, 1H), 5.06 (dd, J = 6.5, 2.8 Hz, 2H), 4.92 (d, J = 10.6 Hz, 2H), 4.78 (d, J = 10.8 Hz, 2H), 4.72 – 4.60 (m, 2H), 4.46 (s, 1H), 4.27 – 4.24 (m, 2H), 4.17 – 4.08 (m, 1H), 4.03 – 3.86 (m, 1H), 3.89 – 3.72 (m, 2H), 3.75 (s, 3H), 3.32 – 3.11 (m, 3H), 1.85 – 1.80 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 170.4, 157.4, 138.6, 138.4, 137.8, 136.9, 128.8, 128.2, 128.1, 128.1, 128.0, 128.0, 127.9, 127.8, 127.8, 127.8, 127.7, 127.5, 127.5, 127.3, 99.4, 96.3, 77.7, 74.6, 71.8, 71.5, 70.2, 67.2, 66.0, 63.5, 53.9, 51.7, 48.6, 48.4, 48.1, 47.9, 47.7, 47.5, 47.3, 47.1, 44.4, 44.3, 29.2. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>30</sub>H<sub>37</sub>N<sub>4</sub>O<sub>20</sub>S<sub>3</sub><sup>3-</sup>, 289.7060; found, 289.7047.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(2-sulfonatamido-4-O-benzyl-6-O-sulfonato-3-O-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (16)**

A solution of compound **12** (0.6g, 1 eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction mixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v) to obtain compound **deacetylated product**. A solution of compound **deacetylated product** (0.6g, 1 eq) in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **aminated product** in 76 % yield. A solution of compound **aminated product** (0.2g, 1 eq) in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **39** in 64 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>)  $\delta$  7.86 – 7.74 (m, 4H), 7.60 – 7.58 (m, 1H), 7.43 – 7.36 (m, 4H), 7.32 – 7.24 (m, 9H), 7.22 – 7.19 (m, 3H), 5.38 (d, J = 3.9 Hz, 1H), 5.27 (d, J = 11.3 Hz, 1H), 5.03 (s, 2H), 4.93 (d, J = 10.1 Hz, 2H), 4.83 – 4.73 (m, 3H), 4.65 (q, J = 11.6 Hz, 2H), 4.43 – 4.23 (m, 1H), 4.25 (d, J = 10.8, 1H), 4.17 (d, J = 6.7 Hz, 2H), 3.94 – 3.86 (m, 2H), 3.84 – 3.73 (m, 2H), 3.73 – 3.65 (m, 1H), 3.63 – 3.48 (m, 2H), 3.29 – 3.15 (m, 2H), 1.82 – 1.74 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD)  $\delta$  172.9, 157.4, 138.5, 138.2, 136.9, 136.7, 133.4, 133.0, 128.1, 128.0, 128.0, 127.8, 127.7, 127.7, 127.6, 127.5, 127.4, 127.3, 127.2, 127.1, 126.2, 126.2, 125.5, 125.3, 101.2, 96.5, 80.6, 77.3, 75.0, 74.6, 73.1, 71.8, 69.9, 66.7, 66.1, 65.9, 58.2, 48.3, 48.1, 47.9, 47.7, 47.5, 47.3, 47.0, 38.3, 29.3. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>48</sub>H<sub>52</sub>N<sub>2</sub>O<sub>19</sub>S<sup>2-</sup>, 512.1308; found, 512.1297.

**N-benzyloxycarbonyl-3-aminopropyl-O-(((2-sulfonatamido-4-O-benzyl-3-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl)))-(1 $\rightarrow$ 4)-O-3-O-benzyl-1- $\alpha$ - L-idopyranosyluronate (17)**

A solution of compound **13** (0.6g, 1 eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction mixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v) to obtain deacetylated product. A solution of **deacetylated product** in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain **amine product** in 78 % yield. A solution of compound **amine product** (0.2g, 1 eq) in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **37** in 62 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>)  $\delta$  7.50 – 7.21 (m, 15H), 5.28 (d, J = 3.8 Hz, 1H), 5.04 (s, 2H), 4.96 (d, J = 11.0 Hz, 2H), 4.68 – 4.62 (m, 4H), 4.24 – 4.04 (m, 2H), 3.89 – 3.72 (m, 5H), 3.74 – 5.71 (m, 1H), 3.58 – 3.46 (m, 2H), 3.33 – 3.11 (m, 3H), 1.80 (p, J = 6.1 Hz, H). <sup>13</sup>C NMR (151 MHz, MeOD)  $\delta$  174.0, 157.4, 138.6, 138.2, 136.9, 128.4, 128.1, 127.9, 127.9, 127.9, 127.6, 127.6, 127.5, 127.5, 127.3, 127.2, 101.0, 95.9, 78.3, 74.3, 73.1, 72.2, 71.7, 71.6, 71.2, 67.2, 66.1, 60.7, 58.4, 53.9, 48.1, 48.0, 47.8, 47.7, 47.5, 47.4, 47.3, 38.4, 29.3. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>53</sub>H<sub>62</sub>N<sub>2</sub>O<sub>19</sub>SSi<sup>2-</sup>, 561.1584; found, 561.1579.

**N-benzyloxycarbonyl-3-aminopropyl-O-(((2-sulfonatamido-4-O-benzyl-6-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl)))-(1 $\rightarrow$ 4)-O-2-O-sulfonato-3-O-benzyl-1- $\alpha$ - L-idopyranosyluronate (18)**

A solution of compound **14** (0.2g, 1 eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin

and reaction mixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). A solution of compound 46a in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column. Then, a solution of compound 46b in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain **40** in 58 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.94 – 7.74 (m, 4H), 7.64 (d, J = 8.4 Hz, 1H), 7.53 – 7.14 (m, 17H), 5.36 (d, J = 11.6 Hz, 2H), 5.03 (s, 3H), 4.61 – 4.54 (m, 5H), 4.45 – 4.34 (m, 3H), 4.47 – 4.07 (m, 3H), 3.91 – 3.72 (m, 2H), 3.70 3.54(m, 4H), 3.24 – 3.15 (m, 2H), 1.80 – 1.74 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 174.6, 157.4, 138.3, 138.2, 136.9, 133.4, 133.0, 128.3, 128.1, 128.0, 127.9, 127.8, 127.6, 127.6, 127.5, 127.4, 127.3, 127.3, 127.2, 126.3, 126.3, 125.5, 125.4, 98.8, 98.7, 80.5, 77.3, 75.0, 74.4, 73.8, 71.7, 70.8, 69.7, 67.2, 66.0, 65.8, 53.9, 48.4, 48.2, 48.0, 47.7, 47.5, 47.3, 47.1, 38.0, 29.3. HR-ESI-MS (*m/z*): [M]<sup>3-</sup> calcd for C<sub>42</sub>H<sub>47</sub>N<sub>2</sub>O<sub>22</sub>S<sub>3</sub><sup>3-</sup>, 321.0495; found, 321.0484.

**N-benzyloxycarbonyl-3-aminopropyl-O-(((2-sulfonatamido-4-O-benzyl-6-O-sulfonato-3-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl)))-(1 $\rightarrow$ 4)-O-2-O-sulfonato-3-O-benzyl-1- $\alpha$ -L-idopyranosyluronate (19)**

A solution of compound **15** (0.2g, 1eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction mixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). A solution of compound **41a** in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using

(MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column. Then, a solution of compound **41b** in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **41** in 55 % yield. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.55 (d, J = 7.4 Hz, 2H), 7.46 (d, J = 7.6 Hz, 2H), 7.43 – 7.26 (m, 11H), 5.34 (d, J = 15.4 Hz, 2H), 5.10 (s, 2H), 4.77 (d, J = 6.3 Hz, 2H), 4.71 – 4.60 (m, 3H), 4.57 (s, 1H), 4.49 (d, J = 10.9 Hz, 1H), 4.44 – 4.18 (m, 4H), 4.10 (d, J = 10.1 Hz, 1H), 3.93 – 3.77 (m, 2H), 3.72 – 3.61 (m, 2H), 3.31 – 3.17 (m, 2H), 1.89 – 1.80 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 157.5, 138.4, 138.1, 136.9, 129.6, 129.5, 128.5, 128.1, 128.1, 127.9, 127.7, 127.5, 127.5, 127.4, 127.2, 98.6, 97.7, 78.1, 74.8, 73.4, 72.0, 71.7, 70.9, 69.5, 66.1, 65.9, 58.5, 48.4, 48.2, 47.9, 47.7, 47.5, 47.3, 47.1, 46.3, 38.1, 29.2. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>37</sub>H<sub>42</sub>N<sub>2</sub>O<sub>55</sub>S<sub>4</sub><sup>4-</sup>, 380.4864; found, 380.4858.

### **3-aminopropyl-*O*-[(2-acetamido-3-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1→4)-*O*- $\alpha$ -L-idopyranosiduronate (I-2)**

A solution of compound **7b** (60 mg, 1 eq) in H<sub>2</sub>O (2 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction fixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). The reaction mixture was dissolved in water and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction were pooled and lyophilized to yield fully deprotected 3-*O*-sulfated NAc-disaccharide. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.17 (d, J = 3.7 Hz, 1H), 4.91 (d, J = 2.7 Hz, 1H), 4.51 (d, J = 2.6 Hz, 1H), 4.44 (dd, J = 10.5, 9.0 Hz, 1H), 4.13 – 4.05 (m, 2H), 4.02 (t, J = 4.1 Hz, 1H), 3.95 – 3.89 (m, 1H), 3.89 – 3.82 (m, 3H), 3.73 (dt, J = 10.6, 5.6 Hz, 1H), 3.69 – 3.62 (m, 2H), 3.15 (td, J = 6.7, 1.4 Hz, 2H), 2.00 (s, 5H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 175.1, 174.3, 100.4, 94.9, 79.8, 73.7, 71.8, 68.7, 68.3, 68.2, 67.5, 66.4, 59.9, 52.1, 38.1, 26.3, 22.0. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>17</sub>H<sub>29</sub>N<sub>2</sub>O<sub>15</sub>S<sup>-</sup>, 533.1294; found, 533.1262.



**3-aminopropyl-*O*-[(2-acetamido-6-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1 $\rightarrow$ 4)-*O*- $\alpha$ -L-idopyranosiduronate (I-1)**

A solution of compound **19** (50 mg, 1 eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction fixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). The reaction mixture was dissolved in water and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected 6-*O*-sulfated NAc-disaccharide. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide)  $\delta$  5.06 (d, J = 3.7 Hz, 1H), 4.81 (d, J = 2.6 Hz, 1H), 4.44 (d, J = 2.6 Hz, 1H), 4.25 (dd, J = 11.1, 3.4 Hz, 1H), 4.12 (dd, J = 11.0, 2.1 Hz, 1H), 3.98 (t, J = 2.9 Hz, 1H), 3.92 (t, J = 4.0 Hz, 1H), 3.89 (d, J = 3.5 Hz, 1H), 3.86 (d, J = 3.6 Hz, 1H), 3.84 – 3.78 (m, 1H), 3.66 – 3.60 (m, 2H), 3.58 (dd, J = 4.7, 2.6 Hz, 1H), 3.48 (t, J = 9.6 Hz, 1H), 3.10 – 3.02 (m, 2H), 1.95 – 1.87 (m, 5H). <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  175.0, 174.3, 100.4, 94.3, 72.9, 71.0, 70.1, 69.1, 68.5, 68.2, 67.0, 66.5, 53.3, 38.3, 26.3, 21.9. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>17</sub>H<sub>29</sub>N<sub>2</sub>O<sub>15</sub>S<sup>-</sup>, 533.1294; found, 533.1291.

**3-aminopropyl-*O*-[(2-sulfonatomido-3-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1 $\rightarrow$ 4)-*O*- $\alpha$ -L-idopyranosiduronate (I-4)**

A solution of compound **38** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide. <sup>1</sup>H NMR (600 MHz, Deuterium Oxide)  $\delta$  5.33 (d, J = 3.7 Hz, 1H), 4.93 (s, 1H), 4.66 (d, J = 2.5 Hz, 1H), 4.20 (t, J = 3.5 Hz, 1H), 4.07 (d, J = 2.6 Hz, 1H), 3.95 – 3.88 (m, 1H), 3.84 – 3.74 (3, 2H), 3.75 (d, J = 3.7 Hz, 1H), 3.74 – 3.65 (m, 2H), 3.63 – 3.60 (m, 1H), 3.51 (t, J = 9.6 Hz, 1H), 3.24 – 3.12 (m, 3H), 2.03 – 1.96 (m, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O)  $\delta$  174.2, 100.5, 96.1, 74.1, 72.0, 71.2, 69.4, 67.8, 67.2, 66.6, 66.6, 60.0, 57.7, 55.4, 38.3, 26.2. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>15</sub>H<sub>26</sub>N<sub>2</sub>O<sub>17</sub>S<sup>2-</sup>, 285.0684; found, 285.0673.

**3-aminopropyl-*O*-[(2-sulfonatomido-6-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1 $\rightarrow$ 4)-*O*- $\alpha$ -L-idopyranosiduronate (I-3)**

A solution of compound **39** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.34 (d, J = 3.7 Hz, 1H), 4.97 – 4.85 (m, 1H), 4.60 (d, J = 2.4 Hz, 1H), 4.34 (dd, J = 11.0, 3.1 Hz, 1H), 4.27 – 4.11 (m, 2H), 4.08 (t, J = 2.8 Hz, 1H), 3.97 – 3.83 (m, 2H), 3.78 – 3.68 (m, 2H), 3.66 – 3.54 (m, 2H), 3.25 (dd, J = 10.0, 3.6 Hz, 1H), 3.21 – 3.11 (m, 2H), 2.04 – 1.98 (m, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.6, 100.4, 95.8, 74.2, 74.1, 71.1, 70.1, 69.1, 68.1, 68.0, 67.5, 67.5, 66.9, 66.8, 66.6, 66.5, 66.3, 57.6, 50.8, 43.2, 38.3, 26.2, 25.6. HR-ESI-MS (m/z): [M]<sup>2-</sup> calcd for C<sub>15</sub>H<sub>26</sub>N<sub>2</sub>O<sub>17</sub>S<sub>2</sub><sup>-</sup>, 285.0684; found, 285.0671.

**3-aminopropyl-*O*-[(2-sulfonatamido-6-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1→4)-*O*-2-*O*-sulfonato *O*- $\alpha$ -L-idopyranosiduronate (I-5)**

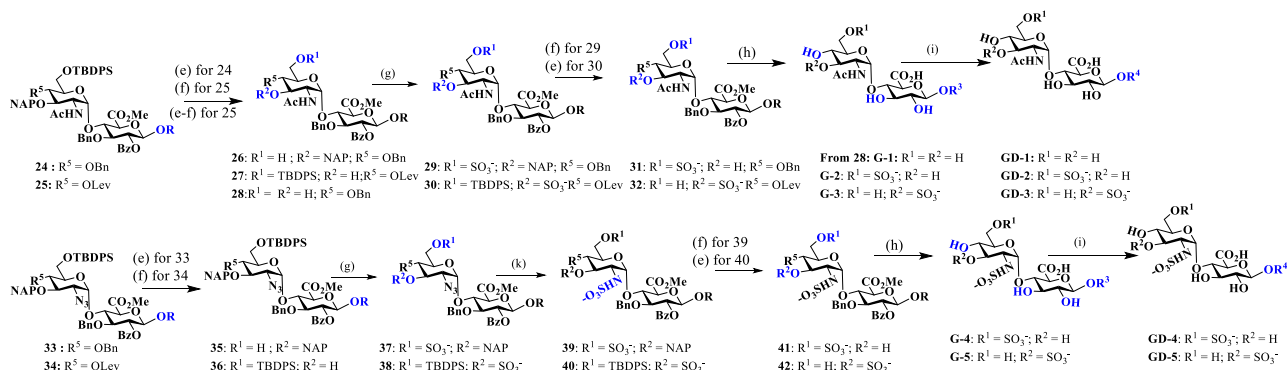
A solution of compound **40** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.32 (d, J = 3.5 Hz, 1H), 5.08 (d, J = 2.6 Hz, 1H), 4.48 (d, J = 2.7 Hz, 1H), 4.28 (dd, J = 11.0, 2.8 Hz, 1H), 4.24 – 4.11 (m, 3H), 4.03 (t, J = 3.1 Hz, 1H), 3.91 – 3.84 (m, 2H), 3.67 – 3.60 (m, 1H), 3.59 – 3.46 (m, 2H), 3.19 (dd, J = 10.2, 3.6 Hz, 1H), 3.10 (t, J = 6.5 Hz, 2H), 1.97 – 1.90 (m, 2H). <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O) δ 176.9, 101.2, 99.7, 78.4, 78.2, 73.5, 72.6, 71.7, 70.9, 70.8, 69.2, 69.0, 60.4, 40.9, 28.7. HR-ESI-MS (m/z): [M]<sup>2-</sup> calcd for C<sub>16</sub>H<sub>27</sub>N<sub>2</sub>O<sub>20</sub>S<sub>3</sub><sup>3-</sup>, 221.0112; found, 221.0195.

**3-aminopropyl-*O*-[(2-sulfonatamido-6-*O*-sulfonato-3-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)-(1→4)-*O*-2-*O*-sulfonato *O*- $\alpha$ -L-idopyranosiduronate (I-6)**

A solution of compound **41** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.34 (d, J = 3.7 Hz, 1H), 4.97 – 4.85 (m, 1H), 4.60 (d, J = 2.4 Hz, 1H), 4.34 (dd, J = 11.0, 3.1 Hz, 1H), 4.27 – 4.11 (m, 2H), 4.08 (t, J = 2.8 Hz, 1H), 3.97 – 3.83 (m, 2H), 3.78 – 3.68 (m, 2H), 3.66 – 3.54 (m, 2H), 3.25 (dd, J = 10.0, 3.6 Hz, 1H), 3.21 – 3.11 (m, 2H), 2.04 – 1.98 (m, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.6,

100.4, 95.8, 74.2, 74.1, 71.1, 70.1, 69.1, 68.1, 68.0, 67.5, 67.5, 66.9, 66.8, 66.6, 66.5, 66.3, 57.6, 50.8, 43.2, 38.3, 26.2, 25.6. HR-ESI-MS ( $m/z$ ):  $[M]^{2-}$  calcd for  $C_{15}H_{26}N_2O_{17}S^{2-}$ , 285.0684; found, 285.0671.

### 3. Synthesis of Glucuronic acid based Disaccharides



**Scheme S2. Synthesis of glucuronic acid based HS disaccharides** (a) R = Linker, NIS, TMSOTf, DCM, -10 °C, 15 min; (b) NaOMe, MeOH, RT, 12 h; TEMPO, BAIB, DCM:H<sub>2</sub>O (1:1), RT; (c) 1 M LiOH, THF: H<sub>2</sub>O (1:1), RT, 2 h; MeI, K<sub>2</sub>CO<sub>3</sub>, DMF, 12 h; Ac<sub>2</sub>O, DCM:Py (1:1), 0 °C, 12 h; (d) Zn dust, THF:AcOH:Ac<sub>2</sub>O (3:2:1), RT, 12 h; (e) HF.Py, Py, 0 °C, 12 h; (f) DDQ, DCM:H<sub>2</sub>O (18:1), RT, 1 h; (g) SO<sub>3</sub>. Et<sub>3</sub>N, DMF, 60 °C, 24 h; (h) (i) LiOH.H<sub>2</sub>O, H<sub>2</sub>O, 12 h; (ii) H<sub>2</sub>, Pd(OH)<sub>2</sub>, MeOH, 48 h; (i) DBCO-NHS linker, Et<sub>3</sub>N, DMF, 24 h. (j) LiOH.H<sub>2</sub>O, H<sub>2</sub>O; (k) PMe<sub>3</sub>.THF, 24 h; (ii) SO<sub>3</sub>.Py. MeOH, 1N NaOH, 48 h.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(2-acetamido-6-*O*-sulfonato-4-*O*-benzyl-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-*O*-3-*O*-benzyl-1- $\beta$ -D-glucopyranosiduronate (29)**

Compound **24** (1g, 1 eq) was dissolved in dry pyridine (5 mL), and a 70 % HF.Py complex (5 eq) was added at 0°C. The mixture was stirred for 12 hours. Subsequently, ethyl acetate was added to the reaction mixture and washed with 1 N HCl three times. The organic layer was then evaporated, and purification was carried out using silica gel column chromatography. Then sulfation reaction was performed as mentioned for compound **6** followed by ester hydrolysis was

performed using LiOH.H<sub>2</sub>O (20 eq) in MeOH:H<sub>2</sub>O (1:1) for 12 h to obtain compound **29**. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.18 – 6.88 (m, 22H), 5.26 (d, J = 3.5 Hz, 1H), 4.81 (s, 1H), 4.77 (s, 2H), 4.72 (d, J = 11.5 Hz, 1H), 4.66 – 4.58 (m, 2H), 4.49 (d, J = 10.2 Hz, 1H), 4.37 (d, J = 10.5 Hz, 1H), 4.24 (d, J = 10.1 Hz, 1H), 4.13 – 3.94 (m, 4H), 3.74 (q, J = 10.6, 10.0 Hz, 2H), 3.67 – 3.52 (m, 3H), 3.52 – 3.42 (m, 1H), 3.37 – 3.17 (m, 2H), 3.01 – 2.89 (m, 2H), 1.62 (s, 3H), 1.53 – 1.42 (m, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 175.1, 171.8, 157.8, 138.4, 138.4, 137.2, 136.9, 136.3, 133.4, 133.0, 130.3, 129.1, 128.3, 128.2, 128.1, 128.0, 127.8, 127.7, 127.6, 127.6, 127.5, 127.4, 126.0, 125.9, 125.7, 125.6, 102.9, 97.5, 84.9, 80.7, 78.1, 76.9, 75.3, 74.7, 74.3, 74.0, 69.9, 67.0, 66.3, 66.0, 53.1, 48.8, 48.6, 48.4, 48.2, 48.0, 47.8, 47.6, 47.3, 37.4, 29.7, 22.0. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>58</sub>H<sub>61</sub>N<sub>2</sub>O<sub>18</sub>S<sup>-</sup>, 1105.3646; found, 1105.3639.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(2-acetamido-6-O-*tert*-butyldiphenylsilyl-3-O-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-O-3-O-benzyl-1- $\beta$ -D-glucopyranosiduronate (28)**

A solution compound **24** containing (1 g, 1 eq) of starting material in 6 mL of CH<sub>2</sub>Cl<sub>2</sub> and 340  $\mu$ L of H<sub>2</sub>O was prepared, to which DDQ (5 mmol, 5 eq) was gradually added over a 20-minute period. After 1 hour, the reaction mixture was quenched with NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub>, and washed with brine. The organic layer obtained was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, concentrated, and subjected to purification through silica gel column chromatography. Subsequently, the compound was dissolved in 2 mL of DMF, and SO<sub>3</sub>.NEt<sub>3</sub> complex (10 mmol per -OH, 10 eq) was added under a N<sub>2</sub> atmosphere. The mixture was stirred at 70 °C for 3 days. After the reaction completion, the mixture was concentrated under reduced pressure, and the resulting product was purified using silica column chromatography (MeOH/DCM 1/10 v/v) to get product. Then, TBDPS was deprotected when the reaction mixture was stirred for 12 h in presence of 70 % HF.Py, in pyridine followed by ester hydrolysis was performed using LiOH.H<sub>2</sub>O (20 eq) in MeOH:H<sub>2</sub>O (1:1) for 12 h to obtain compound **28**. <sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 7.38 – 7.18 (m, 10H), 5.44 (d, J = 3.9 Hz, 1H), 5.04 (d, J = 15.7 Hz, 3H), 4.65 (d, J = 10.6 Hz, 1H), 4.50 (dd, J = 10.7, 8.9 Hz, 1H), 4.31 (d, J = 7.7 Hz, 1H), 4.06 (ddd, J = 10.6, 6.5, 3.8 Hz, 1H), 4.00 – 3.87 (m, 2H), 3.80 (dd, J = 21.6, 10.5 Hz, 3H), 3.68 (d, J = 8.8 Hz, 2H), 3.61 – 3.52 (m, 2H), 3.48 (dd, J = 8.9, 7.8 Hz, 1H), 3.24 (dt, J = 8.9, 6.6 Hz, 2H), 1.83 (s, 3H), 1.77 (q, J = 6.2 Hz, 2H). <sup>13</sup>C NMR (101 MHz, MeOD) δ 172.0, 157.6, 138.5, 128.1, 127.9,

127.6, 127.6, 127.4, 127.1, 103.0, 97.4, 84.5, 78.7, 75.5, 74.6, 74.0, 72.2, 69.7, 66.7, 66.0, 60.9, 52.1, 48.3, 48.0, 47.8, 47.6, 47.4, 47.2, 47.0, 37.3, 29.4, 21.5. HR-ESI-MS ( $m/z$ ):  $[M]^-$  calcd for  $C_{39}H_{47}N_2O_{17}S^-$ , 847.2601; found, 847.2598.

%

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-azido-6-*O*-sulfonato-4-*O*-levulinoyl-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-3-*O*-benzyl-1- $\beta$ -D-glucopyranosiduronate (37)**

The compound **33** was dissolved to (1 mmol, 1 eq) in DMF (2 mL) was added  $SO_3 \cdot NEt_3$  complex (10 mmol per -OH, 10 eq) under  $N_2$  atmosphere and stirred at 70 °C for 4 days. Upon completion of reaction, the reaction mixture was concentrated under reduced pressure and it was concentrated and purified using silica column chromatography (MeOH/DCM 1.3/10 v/v) to obtain product **37** in 78% yield.  $^1H$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  7.99 (dd,  $J = 8.3, 1.4$  Hz, 2H), 7.83 – 7.72 (m, 4H), 7.54 (t,  $J = 7.5$  Hz, 1H), 7.46 – 7.36 (m, 5H), 7.29 (t,  $J = 7.7$  Hz, 4H), 7.11 (d,  $J = 3.7$  Hz, 4H), 5.38 (d,  $J = 3.3$  Hz, 1H), 5.27 (t,  $J = 7.6$  Hz, 1H), 5.19 – 5.12 (m, 1H), 5.05 (d,  $J = 27.3$  Hz, 3H), 4.86 (d,  $J = 11.5$  Hz, 1H), 4.81 – 4.71 (m, 2H), 4.67 – 4.57 (m, 2H), 4.24 (t,  $J = 8.7$  Hz, 1H), 4.04 (dq,  $J = 14.5, 8.4, 7.5$  Hz, 2H), 3.89 (dt,  $J = 20.2, 9.4$  Hz, 3H), 3.75 (s, 3H), 3.64 (d,  $J = 9.8$  Hz, 1H), 3.45 (dd,  $J = 10.8, 6.1$  Hz, 1H), 3.37 – 3.27 (m, 1H), 3.07 (hept,  $J = 7.9$  Hz, 2H), 2.53 (t,  $J = 11.4$  Hz, 3H), 2.35 – 2.23 (m, 1H), 1.99 (s, 3H), 1.62 (dp,  $J = 13.9, 8.9, 8.1$  Hz, 2H).  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  209.0, 172.7, 169.2, 165.2, 156.6, 137.2, 136.9, 135.2, 133.5, 133.2, 133.0, 129.8, 129.4, 128.6, 128.5, 128.4, 128.1, 128.0, 127.9, 127.8, 127.7, 126.8, 126.1, 126.0, 100.8, 97.5, 82.0, 77.4, 77.3, 77.0, 76.7, 74.5, 73.4, 67.7, 66.4, 62.5, 53.5, 38.0, 37.5, 29.7, 29.4, 27.9. HR-ESI-MS ( $m/z$ ):  $[M]^-$  calcd for  $C_{54}H_{57}N_4O_{19}S^-$ , 1097.3343; found, 1097.3329.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(methyl(2-azido-6-*tert*-butyldiphenylsilyl-4-*O*-levulinoyl-3-*O*-sulfonato-2-deoxy- $\alpha$ -D-glucopyranosyl))-(1 $\rightarrow$ 4)-*O*-3-*O*-benzyl-1- $\beta$ -D-glucopyranosiduronate (38)**

A solution compound **34** containing (1 g, 1 eq) of starting material in 6 mL of  $CH_2Cl_2$  and 340  $\mu$ L of  $H_2O$  was prepared, to which DDQ (5 mmol) was gradually added over a 20-minute period. After 1 hour, the reaction mixture was quenched with  $NaHCO_3$ , extracted with  $CH_2Cl_2$ , and washed with brine. The organic layer obtained was dried over  $Na_2SO_4$ , filtered, concentrated,

and subjected to purification through silica gel column chromatography. Subsequently, the compound was dissolved in 2 mL of DMF, and  $\text{SO}_3\cdot\text{NEt}_3$  complex (10 mmol per -OH) was added under a  $\text{N}_2$  atmosphere. The mixture was stirred at 70 °C for 3 days. After the reaction completion, the mixture was concentrated under reduced pressure, and the resulting product was purified using silica column chromatography (MeOH/DCM 1/10 v/v) to obtain product **38** yielding 69 %.  $^1\text{H}$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.00 (d,  $J$  = 7.5 Hz, 2H), 7.67 – 7.58 (m, 4H), 7.52 (t,  $J$  = 7.4 Hz, 1H), 7.42 – 7.25 (m, 14H), 7.17 – 7.06 (m, 5H), 5.57 (d,  $J$  = 3.3 Hz, 1H), 5.37 – 5.29 (m, 1H), 5.22 (t,  $J$  = 9.6 Hz, 1H), 5.06 (d,  $J$  = 6.1 Hz, 1H), 5.02 (s, 2H), 4.80 (t,  $J$  = 9.8 Hz, 1H), 4.70 (q,  $J$  = 10.7 Hz, 2H), 4.61 (d,  $J$  = 7.1 Hz, 1H), 4.25 (t,  $J$  = 9.0 Hz, 1H), 4.13 – 3.99 (m, 2H), 3.85 (dt,  $J$  = 9.3, 5.8 Hz, 1H), 3.75 – 3.64 (m, 2H), 3.63 (d,  $J$  = 4.4 Hz, 1H), 3.60 (s, 3H), 3.49 (dq,  $J$  = 12.7, 6.3, 5.8 Hz, 1H), 3.26 (d,  $J$  = 10.4 Hz, 1H), 3.12 (ddt,  $J$  = 25.4, 12.5, 6.7 Hz, 2H), 2.88 – 2.68 (m, 4H), 2.58 (dt,  $J$  = 17.9, 5.9 Hz, 1H), 2.35 (dt,  $J$  = 18.6, 5.3 Hz, 1H), 2.15 (s, 3H), 1.66 (dt,  $J$  = 12.1, 7.0 Hz, 2H), 1.01 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  209.6, 172.8, 169.0, 165.2, 156.5, 137.2, 136.8, 135.7, 135.6, 133.5, 133.3, 132.9, 129.8, 129.3, 128.6, 128.5, 128.4, 128.0, 127.9, 127.8, 127.8, 127.7, 100.8, 97.3, 82.2, 77.4, 77.3, 77.1, 76.7, 76.3, 74.2, 73.5, 73.2, 70.7, 68.8, 67.3, 66.4, 61.8, 61.6, 52.8, 37.9, 37.6, 30.3, 30.1, 29.7, 29.4, 29.4, 28.1, 26.8, 19.3. HR-ESI-MS ( $m/z$ ):  $[\text{M}]^-$  calcd for  $\text{C}_{59}\text{H}_{67}\text{N}_4\text{O}_{19}\text{SiS}^-$ , 1195.3895; found, 1195.3881.

**N-benzyloxycarbonyl-3-aminopropyl-*O*-[(2-sulfonatamido-6-*O*-sulfonato-3-*O*-(2-naphthylmethyl)-2-deoxy- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-*O*-1- $\beta$ -D-glucopyranosiduronate (39)**

A solution of compound **37** (0.2g, 1 eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction fixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). A solution of compound 53a in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column. Then, a

solution of compound 53b in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **39** in 61 % yield. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 8.05 – 7.92 (m, 4H), 7.71 (dd, J = 8.5, 1.5 Hz, 1H), 7.56 (qd, J = 6.9, 3.4 Hz, 2H), 7.50 – 7.35 (m, 10H), 5.73 (d, J = 3.4 Hz, 1H), 5.10 (d, J = 11.3 Hz, 3H), 4.98 (d, J = 11.1 Hz, 1H), 4.92 – 4.84 (m, 3H), 4.46 (d, J = 8.1 Hz, 1H), 4.33 (dd, J = 11.0, 3.0 Hz, 1H), 4.20 (dd, J = 10.9, 1.8 Hz, 1H), 3.95 (dd, J = 10.3, 7.2 Hz, 2H), 3.89 – 3.80 (m, 3H), 3.73 – 3.63 (m, 3H), 3.53 (t, J = 8.4 Hz, 1H), 3.42 (dt, J = 6.3, 3.5 Hz, 1H), 3.25 (q, J = 6.3 Hz, 2H), 1.81 (p, J = 6.4 Hz, 2H). <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O) δ 175.0, 137.5, 135.7, 133.0, 132.8, 128.8, 128.8, 128.7, 128.4, 128.2, 128.1, 127.7, 127.6, 127.0, 126.4, 102.2, 96.3, 83.7, 79.3, 76.9, 74.8, 73.5, 73.1, 70.3, 68.9, 67.8, 66.9, 66.5, 57.4, 37.4, 28.8. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>41</sub>H<sub>46</sub>N<sub>2</sub>O<sub>19</sub>S<sub>2</sub><sup>2-</sup>, 467.1073; found, 467.1068.

**N-benzyloxycarbonyl-3-aminopropyl-O-[(2-sulfonatoimido-3-O-sulfonato-2-deoxy-α-D-glucopyranosyl)]-(1→4)-O-3-O-benzyl-1-β-D-glucopyranosiduronate (40)**

A solution of compound **38** (0.15g, 1eq) in THF:H<sub>2</sub>O (1:1, 5 mL) LiOH.H<sub>2</sub>O (20 eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using IR H<sup>+</sup> resin and reaction mixture was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 1/9 v/v). A solution of compound 54a in THF (3 mL) 1M PMe<sub>3</sub>.THF (8 eq) and 0.1M aqueous NaOH (10eq) was added and kept it on stirring for room temperature. After 12 h reaction was quenched using glacial acetic acid and reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using (MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column. Then, a solution of compound 54b in MeOH (3 mL) 0.1M NaOH (4 eq), triethylamine (8 eq), 10 eq of SO<sub>3</sub>.Py was added at an interval of 1 h for 6 times by maintain 9-10 pH of reaction mixture and kept it on stirring for room temperature. The progress of reaction was monitored using TLC plate (ethyl acetate/pyridine/water/acetic acid 8/5/3/1 v/v/v/v). After completion, reaction mixture solvent was evaporated and purified using reverse phase C-18 column chromatography using

(MeOH/H<sub>2</sub>O 2/3 v/v), followed by passing through DOWEX 50WX8 Na<sup>+</sup> resin column to obtain compound **40** in 57 % yield. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 7.55 – 7.50 (m, 2H), 7.49 – 7.35 (m, 8H), 5.74 (d, *J* = 3.3 Hz, 1H), 5.12 (s, 2H), 5.01 (d, *J* = 10.2 Hz, 1H), 4.90 (d, *J* = 10.3 Hz, 1H), 4.49 (d, *J* = 7.9 Hz, 1H), 4.40 (dd, *J* = 10.6, 8.2 Hz, 1H), 3.93 (s, 3H), 3.85 – 3.76 (m, 2H), 3.76 (d, *J* = 0.9 Hz, 1H), 3.72 – 3.61 (m, 3H), 3.48 (t, *J* = 8.2 Hz, 1H), 3.40 (dd, *J* = 10.6, 3.3 Hz, 1H), 3.25 – 3.24 (m, 2H), 1.81 (p, *J* = 6.2 Hz, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.4, 158.4, 137.4, 136.6, 129.0, 128.8, 128.6, 128.3, 128.3, 127.6, 102.1, 96.4, 83.7, 78.4, 74.3, 73.7, 72.2, 68.7, 67.8, 66.8, 60.3, 56.6, 55.4, 37.4, 28.7, 27.5, 19.1. HR-ESI-MS (*m/z*): [M]<sup>2-</sup> calcd for C<sub>30</sub>H<sub>38</sub>N<sub>2</sub>O<sub>19</sub>S<sub>2</sub><sup>2-</sup>, 397.0760; found, 397.0751.

**3-aminopropyl-*O*-[(2-acetamido-6-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)]-(1→4)-*O*- $\beta$ -D-glucopyranosiduronate (G-2)**

Compound **25** was dissolved in water, and Pd(OH)<sub>2</sub> was added, followed by stirring under an H<sub>2</sub> atmosphere for 48 hours. The mixture was then filtered, concentrated, and eluted through a Bond Elute C-18 column using H<sub>2</sub>O as the eluent. The combined water fractions were pooled and subjected to lyophilization, resulting in the fully deprotected GlA-NAc-disaccharide **50**. <sup>1</sup>H NMR (600 MHz, Deuterium Oxide) δ 5.43 (d, *J* = 3.8 Hz, 1H), 4.48 (dd, *J* = 8.0, 1.3 Hz, 1H), 4.37 – 4.30 (m, 1H), 4.18 (dt, *J* = 11.1, 1.8 Hz, 1H), 4.01 – 3.95 (m, 1H), 3.94 – 3.82 (m, 3H), 3.81 – 3.66 (m, 4H), 3.58 (dd, *J* = 10.4, 9.1, 1.3 Hz, 1H), 3.37 – 3.32 (m, 1H), 3.16 (q, *J* = 6.7 Hz, 2H), 2.05 (s, 3H), 1.99 (p, *J* = 6.4 Hz, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 175.0, 174.3, 102.2, 96.8, 76.6, 76.5, 75.5, 73.4, 70.6, 70.0, 69.0, 67.9, 66.3, 53.5, 37.6, 26.7, 21.9. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>17</sub>H<sub>29</sub>N<sub>2</sub>O<sub>15</sub>S<sup>-</sup>, 533.1294; found, 533.1281.

**3-aminopropyl-*O*-[(2-acetamido- $\alpha$ -D-glucopyranosyl)]-(1→4)-*O*- $\beta$ -D-glucopyranosiduronate (G-1)**

Compound **26** was dissolved in water, and Pd(OH)<sub>2</sub> was added, followed by stirring under an H<sub>2</sub> atmosphere for 48 hours. The mixture was then filtered, concentrated, and eluted through a Bond Elute C-18 column using H<sub>2</sub>O as the eluent. The combined water fractions were pooled and subjected to lyophilization, resulting in the fully deprotected GlA-NAc-disaccharide **51**. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.32 (d, *J* = 3.7 Hz, 1H), 4.38 (d, *J* = 8.0 Hz, 1H), 3.93 – 3.84 (m, 1H), 3.82 – 3.74 (m, 2H), 3.73 – 3.67 (m, 4H), 3.66 – 3.59 (m, 3H), 3.39 (dd, *J* = 10.1,



9.1 Hz, 1H), 3.28 – 3.19 (m, 1H), 2.01 – 1.84 (m, 5H). <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O) δ 175.2, 174.4, 100.4, 94.4, 73.1, 71.9, 71.1, 69.6, 68.7, 68.4, 67.3, 66.4, 60.1, 53.5, 38.2, 26.4, 21.9. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>17</sub>H<sub>30</sub>N<sub>2</sub>O<sub>12</sub>, 454.1799; found, 454.1793.

**3-aminopropyl-*O*-[(2-acetamido-3-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-*O*- $\beta$ -D-glucopyranosiduronate (G-3)**

Compound **27** was dissolved in water, and Pd(OH)<sub>2</sub> was added, followed by stirring under an H<sub>2</sub> atmosphere for 48 hours. The mixture was then filtered, concentrated, and eluted through a Bond Elute C-18 column using H<sub>2</sub>O as the eluent. The combined water fractions were pooled and subjected to lyophilization, resulting in the fully deprotected GlA-NAc-disaccharide **52**. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.36 (d, *J* = 3.6 Hz, 1H), 4.52 – 4.36 (m, 2H), 4.04 (dd, *J* = 10.6, 3.6 Hz, 1H), 3.94 (dt, *J* = 11.2, 5.8 Hz, 1H), 3.88 (d, *J* = 8.4 Hz, 1H), 3.84 – 3.77 (m, 2H), 3.74 (dd, *J* = 8.0, 5.6 Hz, 3H), 3.64 (t, *J* = 9.3 Hz, 1H), 3.43 – 3.25 (m, 2H), 3.15 (t, *J* = 7.2 Hz, 2H), 2.05 – 1.91 (m, 5H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.8, 174.3, 102.2, 102.2, 97.5, 79.6, 76.9, 76.8, 76.7, 76.2, 76.2, 73.3, 71.8, 68.2, 67.9, 67.7, 59.9, 52.3, 50.8, 42.5, 37.5, 26.7, 26.0, 22.1, 18.1, 18.1. HR-ESI-MS (*m/z*): [M]<sup>-</sup> calcd for C<sub>17</sub>H<sub>29</sub>N<sub>2</sub>O<sub>15</sub>S<sup>-</sup>, 533.1294; found, 533.1289.

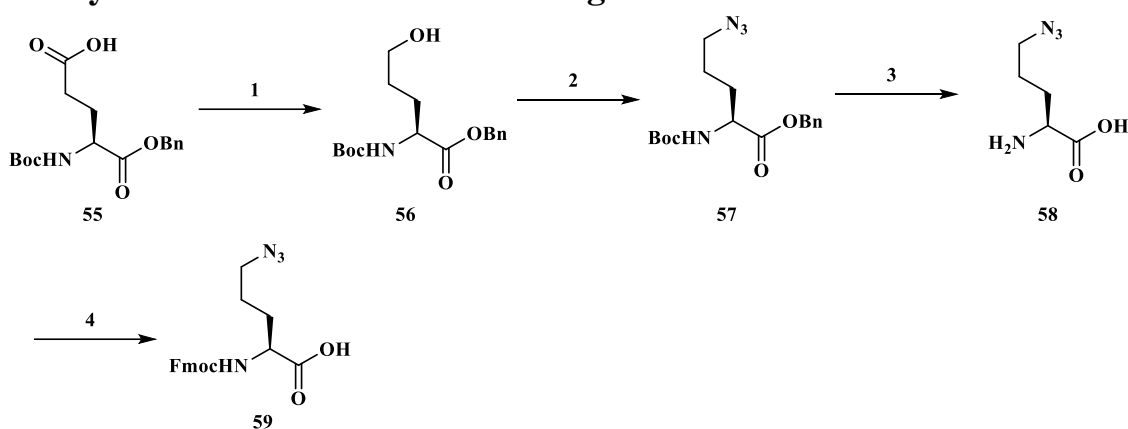
**3-aminopropyl-*O*-[(2-sulfonatamido-6-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-*O*-1- $\beta$ -D-glucopyranosiduronate (G-4)**

A solution of compound **42** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide **53**. <sup>1</sup>H NMR (600 MHz, Deuterium Oxide) δ 5.61 – 5.54 (m, 1H), 4.45 (dd, *J* = 7.9, 2.1 Hz, 1H), 4.25 (d, *J* = 11.0 Hz, 1H), 4.09 (d, *J* = 11.0 Hz, 1H), 3.92 (dt, *J* = 9.3, 5.6 Hz, 1H), 3.76 (tt, *J* = 16.9, 9.6 Hz, 5H), 3.55 – 3.46 (m, 2H), 3.30 (t, *J* = 7.9 Hz, 1H), 3.22 – 3.16 (m, 1H), 3.08 (s, 2H), 1.94 – 1.88 (m, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.3, 102.2, 97.4, 76.2, 76.2, 76.1, 75.9, 72.6, 71.0, 69.9, 69.0, 68.0, 66.4, 57.8, 37.6, 26.7.

**3-aminopropyl-*O*-[(2-sulfonatamido-3-*O*-sulfonato- $\alpha$ -D-glucopyranosyl)]-(1 $\rightarrow$ 4)-*O*-1- $\beta$ -D-glucopyranosiduronate (G-5)**

A solution of compound **43** in H<sub>2</sub>O and Pd(OH)<sub>2</sub> was added and stirred under H<sub>2</sub> atm. After 48 h, the mixture was filtered, concentrated and eluted through Bond Elute C-18 column using H<sub>2</sub>O as eluent. The combined H<sub>2</sub>O fraction was pooled and lyophilized to yield fully deprotected NS-disaccharide **54**. <sup>1</sup>H NMR (400 MHz, Deuterium Oxide) δ 5.59 (d, J = 3.5 Hz, 1H), 4.53 (d, J = 8.0 Hz, 1H), 4.36 (dd, J = 10.5, 9.0 Hz, 1H), 4.01 (dt, J = 11.5, 5.8 Hz, 1H), 3.92 – 3.75 (m, 7H), 3.68 (t, J = 9.2 Hz, 1H), 3.43 – 3.34 (m, 2H), 3.17 (p, J = 7.1 Hz, 2H), 2.00 (p, J = 6.4 Hz, 2H). <sup>13</sup>C NMR (151 MHz, D<sub>2</sub>O) δ 174.9, 102.2, 98.2, 78.6, 78.0, 76.6, 76.1, 72.5, 71.7, 68.3, 68.0, 59.9, 56.8, 37.6, 26.6.

### 1. Synthesis of Amino Acid Building Block



**Reagent and Yield:** (1) (a) NHS, DCC (b) NaBH<sub>4</sub>, 70 %, (2) (a) MsCl, NEt<sub>3</sub>, 90 %, (b) NaN<sub>3</sub>, 81 %, (3) (a) LiOH·H<sub>2</sub>O, 91 %, (b) 40 % TFA in DCM, 94 %, (4) Fmoc-Cl, Na<sub>2</sub>CO<sub>3</sub>, 62 %.

#### Benzyl (S)-2-((tert-butoxycarbonyl)amino)-5-hydroxypentanoate (**56**)

The commercially available Boc-NH-Glu (COOH)-O-tBu (4.54 g, 15 mmol) was dissolved in 30 mL of dry THF. The resulting mixture was cooled to 0 °C using a combination of salt and ice, and 2.06 mL (15 mmol) of triethylamine was added. After a 5-minute interval, N-hydroxysuccinamide (2 equivalents) and N,N'-Dicyclohexylcarbodiimide (5 equivalents) were introduced and stirred for 12 hours. Subsequently, NaBH<sub>4</sub> (3 equivalents) was added to the reaction mixture and stirred for an additional 30 minutes. The progress of the reaction was monitored using TLC. Upon completion of the reaction (approximately 30 minutes), the THF solvent was evaporated under reduced pressure, and any excess NaBH<sub>4</sub> was neutralized using a 10% HCl solution. The resulting mixture was then subjected to extraction with ethyl acetate (3 × 50 mL). The organic layer was washed with 10% HCl solution (3 × 50 mL), 10% Na<sub>2</sub>CO<sub>3</sub> (3 ×

50 mL), brine solution (3 × 50 mL), and finally dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The compound was purified using silica gel chromatography with ethyl acetate/hexane (1/3 v/v) to obtain compound **25**. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.39 – 7.30 (m, 5H), 5.12 (s, 2H), 4.84 – 4.78 (m, 1H), 3.63 (t, *J* = 6.3 Hz, 2H), 3.59 – 3.50 (m, 1H), 2.52 – 2.42 (m, 2H), 1.95 – 1.87 (m, 1H), 1.83 – 1.75 (m, 1H), 1.43 (s, 9H). <sup>13</sup>C NMR (101 MHz, CHLOROFORM-D) δ 173.6, 156.3, 135.8, 128.7, 128.4, 128.4, 79.8, 77.4, 77.1, 76.8, 66.6, 65.3, 52.3, 34.0, 31.0, 28.4, 26.3.

#### **Benzyl (S)-5-azido-2-((tert-butoxycarbonyl)amino)pentanoate (57)**

A solution of compound **56** comprising 1 equivalent of compound in dry dichloromethane (DCM) underwent treatment with 2 equivalents of triethylamine at 0 °C. After 5 minutes of stirring, 3 equivalents of methanesulfonyl chloride were introduced and the mixture was stirred for 45 minutes. Quenching of the reaction was achieved using 1N aqueous ammonium chloride (NH<sub>4</sub>Cl), and the resultant mixture was subjected to thrice washing with NH<sub>4</sub>Cl before undergoing drying over anhydrous sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>). Subsequent purification involved silica gel chromatography utilizing an ethyl acetate/hexane solvent system (1:3 v/v). Post-purification, the compound was dissolved in dry dimethylformamide (DMF), and 2 equivalents of sodium azide (NaN<sub>3</sub>) were incorporated, followed by stirring for 12 hours at 45 °C. To this resultant reaction mixture, 50 mL of ethyl acetate was added, followed by thrice washing with brine and final drying over Na<sub>2</sub>SO<sub>4</sub>. The compound underwent another purification step using silica gel chromatography with an ethyl acetate/hexane solvent system (2:3 v/v) to obtain **57**. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.41 – 7.28 (m, 5H), 5.13 (s, 2H), 4.62 (d, *J* = 8.1 Hz, 1H), 3.82 – 3.69 (m, 1H), 3.41 (qd, *J* = 12.3, 4.3 Hz, 2H), 2.45 (t, *J* = 7.4 Hz, 2H), 1.95 – 1.72 (m, 2H), 1.43 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 172.9, 155.3, 135.8, 128.6, 128.3, 128.3, 79.9, 77.4, 77.0, 76.7, 66.5, 54.9, 50.0, 30.8, 28.3, 27.4.

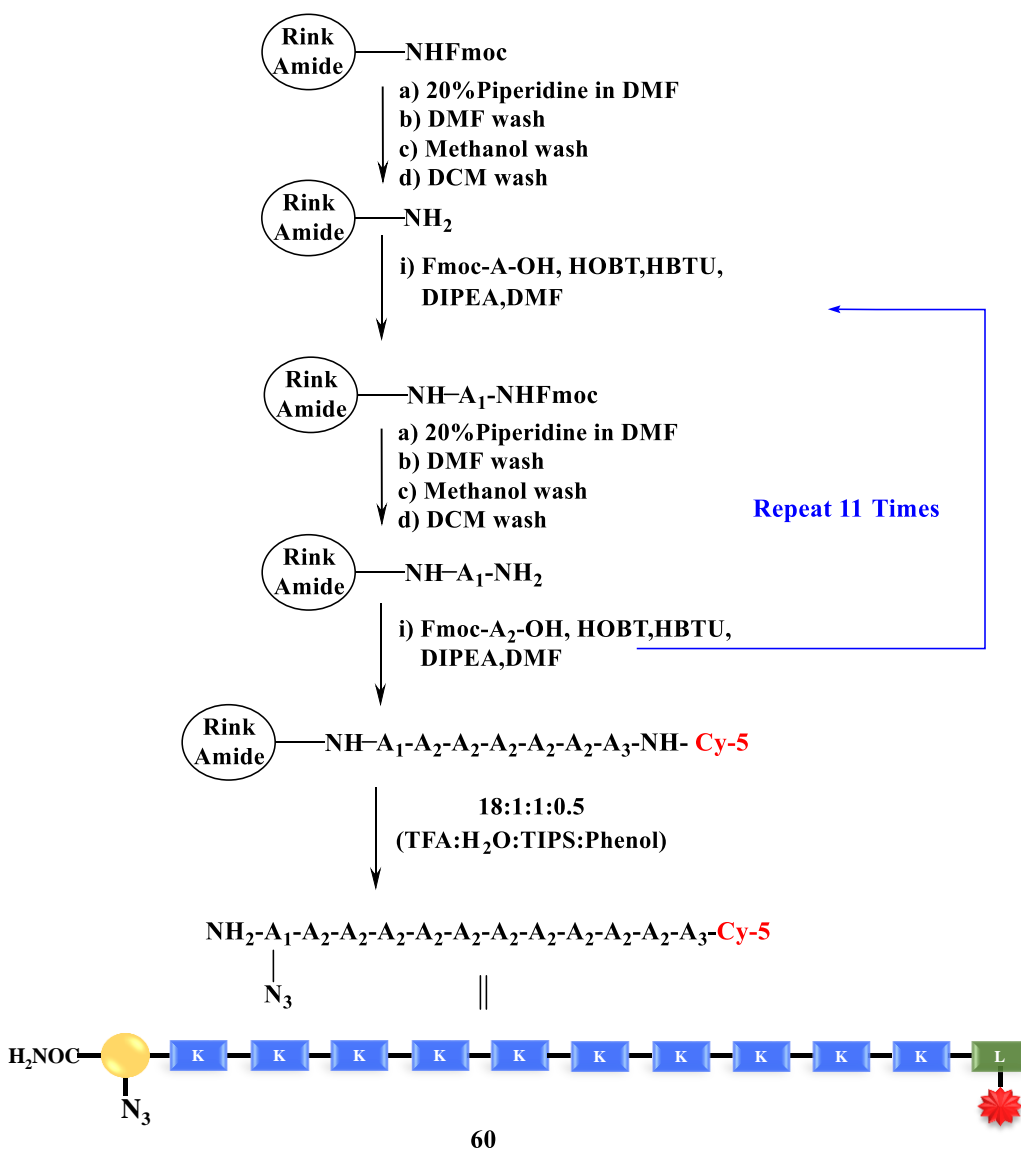
#### **(S)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-5-azidopentanoic acid (59)**

Compound **57** (1 equivalent) was dissolved in a mixture of MeOH:THF:H<sub>2</sub>O (1:1:1). Subsequently, 5 equivalents of LiOH·H<sub>2</sub>O were added, and the mixture was stirred for 2 hours at room temperature. After evaporation, ethyl acetate was added, followed by three washes with 1N HCl. The resulting solution was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Next, 40% TFA in DCM was added to the reaction mixture, and it was stirred for 2 hours at room temperature. After

evaporation and vacuum drying, Compound **58** (1 equivalent) was dissolved in 20 mL of H<sub>2</sub>O and cooled to 0 °C. To this solution, 10 mL of 10% Na<sub>2</sub>CO<sub>3</sub> was added, followed by Fmoc-Cl (2 equivalents), and the reaction mixture was stirred for approximately 4 hours. The progress of the reaction was monitored by TLC. Upon completion, H<sub>2</sub>O was evaporated under reduced pressure, and the resulting mixture was acidified with a 1N HCl solution. Extraction with ethyl acetate (3 × 50 mL) followed, and the combined organic layer was washed with a brine solution (3 × 50 mL). After drying over anhydrous Na<sub>2</sub>SO<sub>4</sub>, the organic layer was evaporated under reduced pressure. The compound was then purified using silica gel chromatography with ethyl acetate/hexane (2/3 v/v) to obtain compound **59**. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.57 (dd, *J* = 18.3, 7.0 Hz, 2H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.31 (t, *J* = 7.5 Hz, 2H), 5.38 (d, *J* = 8.1 Hz, 1H), 4.43 (t, *J* = 6.2 Hz, 2H), 4.22 (t, *J* = 6.6 Hz, 1H), 3.41 – 3.09 (m, 2H), 1.96 (s, 1H), 1.84 – 1.35 (m, 4H). <sup>13</sup>C NMR (101 MHz, CHLOROFORM-D) δ 176.4, 156.2, 143.9, 143.7, 141.4, 127.9, 127.2, 127.2, 125.1, 125.1, 120.1, 120.1, 77.5, 77.1, 76.8, 67.2, 53.3, 50.9, 47.2, 29.8, 29.7, 24.9.

## 5. Synthesis of Peptide Backbone

A 12mer peptide was manually synthesized via solid-phase peptide synthesis on rink amide resin at a 0.3 mmol scale, employing standard Fmoc conditions. The coupling reagents consisted of a combination of HBTU and HOBt, with DIPEA serving as the activator base. NMP was utilized as the coupling solvent, and 20% piperidine in DMF was applied for Fmoc group deprotection. To remove the peptides from the solid support, the resin underwent stirring for approximately 2 hours in a 10 mL cocktail containing trifluoroacetic acid (TFA)/triisopropylsilane/water/phenol (90:5:2.5:2.5). After filtration of the resin, the crude peptides in the filtrate were evaporated under reduced pressure, and the residue was triturated with 20 mL of cold diethyl ether. The resulting precipitate was separated through centrifugation. The crude peptides were subsequently dissolved in 10 mL of MeOH and subjected to purification on a C-18 column using reversed-phase HPLC. A MeOH and water gradient system containing 0.1% TFA at a 0.5 mL flow rate was employed for peptide purification. Confirmation of the purified peptides was accomplished through IR, NMR, and HR-ESI-MS.

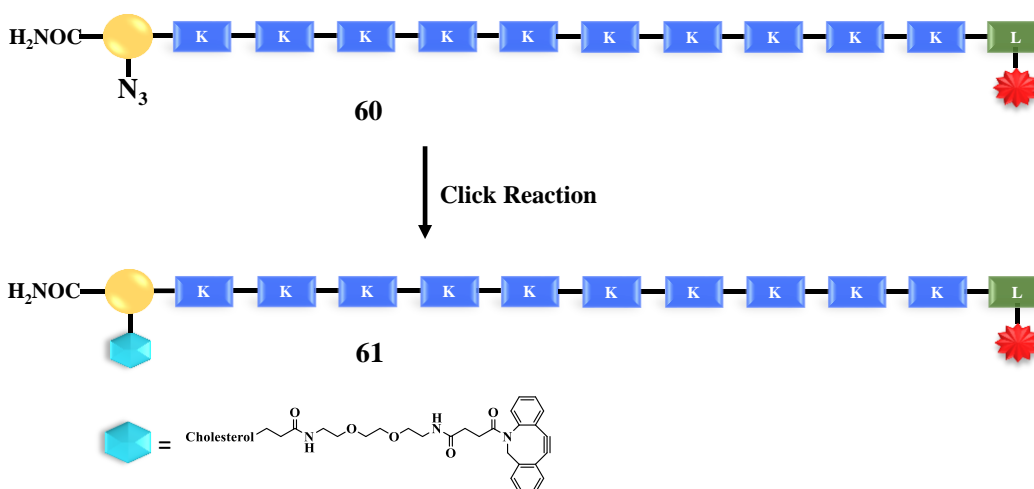


### 12mer Peptide Backbone

<sup>1</sup>H NMR (600 MHz, Methanol-*d*<sub>4</sub>) δ 8.13 (t, *J* = 11.3 Hz, 2H), 8.05 – 7.82 (m, 5H), 7.37 (dd, *J* = 7.6, 4.5 Hz, 2H), 7.30 (tt, *J* = 7.7, 1.9 Hz, 2H), 7.16 (dt, *J* = 14.6, 7.3 Hz, 4H), 6.55 – 6.46 (m, 1H), 6.16 (dd, *J* = 13.7, 4.0 Hz, 1H), 4.34 – 3.76 (m, 15H), 3.52 (s, 3H), 3.20 (dq, *J* = 4.4, 2.7, 2.2 Hz, 3H), 2.82 (h, *J* = 6.8 Hz, 23H), 2.23 (dt, *J* = 12.3, 6.1 Hz, 2H), 1.88 – 1.65 (m, 29H), 1.64 – 1.51 (m, 47H), 1.51 – 1.43 (m, 6H), 1.42 – 1.28 (m, 17H). <sup>13</sup>C NMR (151 MHz, MeOD) δ 176.4, 175.8, 175.7, 175.6, 175.2, 175.2, 175.1, 175.1, 175.0, 174.8, 174.6, 174.5, 174.1, 173.6, 173.4, 173.1, 161.3, 161.0, 158.0, 157.5, 157.2, 156.9, 154.2, 154.0, 142.8, 142.2, 141.2, 141.1, 128.6, 128.3, 125.2, 125.2, 124.9, 124.8, 122.0, 121.9, 119.6, 117.6, 117.5, 115.7, 115.7, 115.6,

114.3, 113.7, 111.8, 110.5, 110.5, 103.1, 102.8, 56.8, 56.7, 56.5, 56.3, 56.1, 55.4, 54.8, 54.7, 54.5, 53.8, 53.7, 53.4, 53.3, 53.1, 53.0, 52.8, 50.8, 50.7, 49.1, 49.1, 48.5, 48.1, 47.9, 47.8, 47.7, 47.5, 47.4, 47.2, 43.5, 39.2, 39.1, 38.9, 35.1, 30.2, 30.1, 29.9, 29.7, 29.5, 28.7, 27.5, 26.9, 26.7, 26.6, 26.4, 26.3, 25.4, 25.3, 25.1, 25.0, 23.1, 23.0, 22.9, 22.8, 22.7, 22.6, 22.3, 15.9. HR-ESI-MS Calcd for  $C_{100}H_{173}N_{28}O_{13}^+ [M]^+$  is 1974.3731, found is 1974.3710.

## 6. Synthesis Protocol for Cholesterol and Azide Linker Conjugation



### 6a. Procedure for conjugation of Cholesterol-DBCO

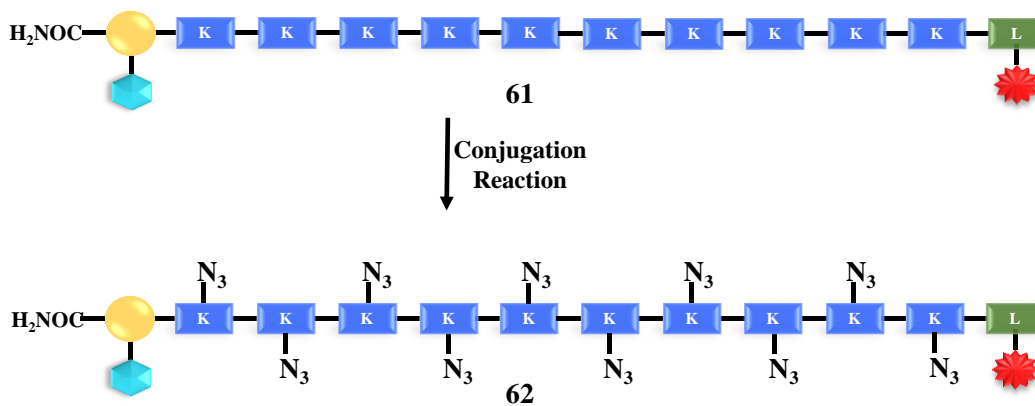
A 12-mer peptide (1 equivalent) was dissolved in 1 mL of MeOH, while cholesterol-DBCO (1 equivalent) was dissolved in 1 mL of DCM. The subsequent step involved the gradual addition of the cholesterol-DBCO solution into the peptide solution, with continuous stirring for 12 hours at room temperature. The progress of the reaction was tracked through MALDI-TOF/TOF analysis. Upon completion of the reaction (approximately 12 hours), the solvent was evaporated under reduced pressure. The crude compound underwent purification via C-18 bond elute column chromatography in a MeOH/H<sub>2</sub>O (6/10) solvent system, yielding the pure cholesterol-conjugated peptide compound. Confirmation was accomplished through IR, NMR, and HR-ESI-MS analyses.

### 12mer Peptide with Cholesterol

$^1H$  NMR (600 MHz, Methanol- $d_4$ )  $\delta$  8.31 – 8.19 (m, 2H), 7.94 (d,  $J = 77.2$  Hz, 2H), 7.71 – 7.52 (m, 4H), 7.50 (dd,  $J = 7.6, 2.2$  Hz, 3H), 7.45 – 7.36 (m, 5H), 7.33 – 7.25 (m, 6H), 6.63 (dd,  $J =$

13.9, 11.1 Hz, 1H), 6.32 – 6.26 (m, 2H), 5.37 – 5.32 (m, 1H), 4.31 – 3.97 (m, 17H), 3.64 (s, 3H), 3.62 – 3.48 (m, 11H), 2.97 – 2.93 (m, 28H), 2.46 – 2.27 (m, 9H), 2.00 – 1.78 (m, 36H), 1.73 (d,  $J = 3.3$  Hz, 17H), 1.62 – 1.67 (m, 28H), 1.63 – 1.36 (m, 48H), 1.29 (s, 3H), 1.21 – 1.01 (m, 10H), 1.00 (s, 3H), 0.94 (d,  $J = 6.5$  Hz, 4H), 0.89 (d,  $J = 2.2$  Hz, 4H), 0.88 (d,  $J = 2.2$  Hz, 4H), 0.71 (s, 3H).  $^{13}\text{C}$  NMR (151 MHz, MeOD)  $\delta$  174.2, 174.2, 174.2, 173.1, 172.9, 162.0, 161.8, 161.5, 161.3, 154.3, 154.0, 142.8, 142.2, 141.2, 141.1, 140.5, 131.1, 129.8, 129.8, 129.1, 128.4, 127.1, 126.8, 125.2, 125.0, 124.8, 122.1, 121.9, 121.4, 120.3, 119.7, 117.8, 115.9, 113.9, 110.5, 103.1, 102.7, 79.3, 69.9, 69.9, 69.8, 69.2, 69.2, 65.1, 63.7, 62.7, 56.8, 56.2, 50.3, 49.2, 49.1, 48.5, 48.1, 47.9, 47.8, 47.6, 47.5, 47.4, 47.2, 43.4, 42.1, 39.7, 39.3, 39.1, 39.0, 39.0, 38.8, 37.0, 36.6, 36.6, 36.0, 35.7, 35.1, 33.6, 30.4, 30.3, 30.2, 29.6, 29.4, 29.4, 29.3, 29.2, 29.1, 29.0, 28.8, 28.0, 27.9, 27.7, 26.9, 26.7, 26.6, 26.4, 26.3, 25.1, 24.6, 23.9, 23.5, 22.8, 22.7, 22.7, 22.6, 22.3, 21.8, 21.6, 20.8, 18.5, 17.9, 13.1, 10.9. HR-ESI-MS Calcd for  $\text{C}_{155}\text{H}_{252}\text{N}_{31}\text{O}_{19}^+$   $[\text{M}]^+$  is 2851.9700, found is 2852.9751 (M+H).

### 6b. Procedure for Conjugation of 3-Azidopropanoate Linker Conjugation



A 12-mer peptide, combined with cholesterol (1 equivalent), was dissolved in 1 mL of DMF. Subsequently, 40  $\mu\text{L}$  of DIPEA was added to the solution, followed by the gradual addition of a solution containing 2,5-dioxopyrrolidin-1-yl 3-azidopropanoate (10 equivalents per  $\text{NH}_2$ ) in 1 mL of DMF. The resulting peptide solution was stirred for 12 hours at room temperature. The progression of the reaction was monitored through MALDI-TOF/TOF analysis. After the reaction's completion (approximately 12 hours), the solvent was evaporated under reduced pressure. The crude compound underwent purification through LH – 20 size exclusion column

chromatography, utilizing MeOH as the solvent system. Additionally, it was passed through Amberlite® IR120, Na<sup>+</sup> form, to obtain the pure compound. Confirmation was done by IR, NMR, and MALDI-TOF/TOF analyses.

### **12mer Peptide with 3-Azidopropanoate Linker**

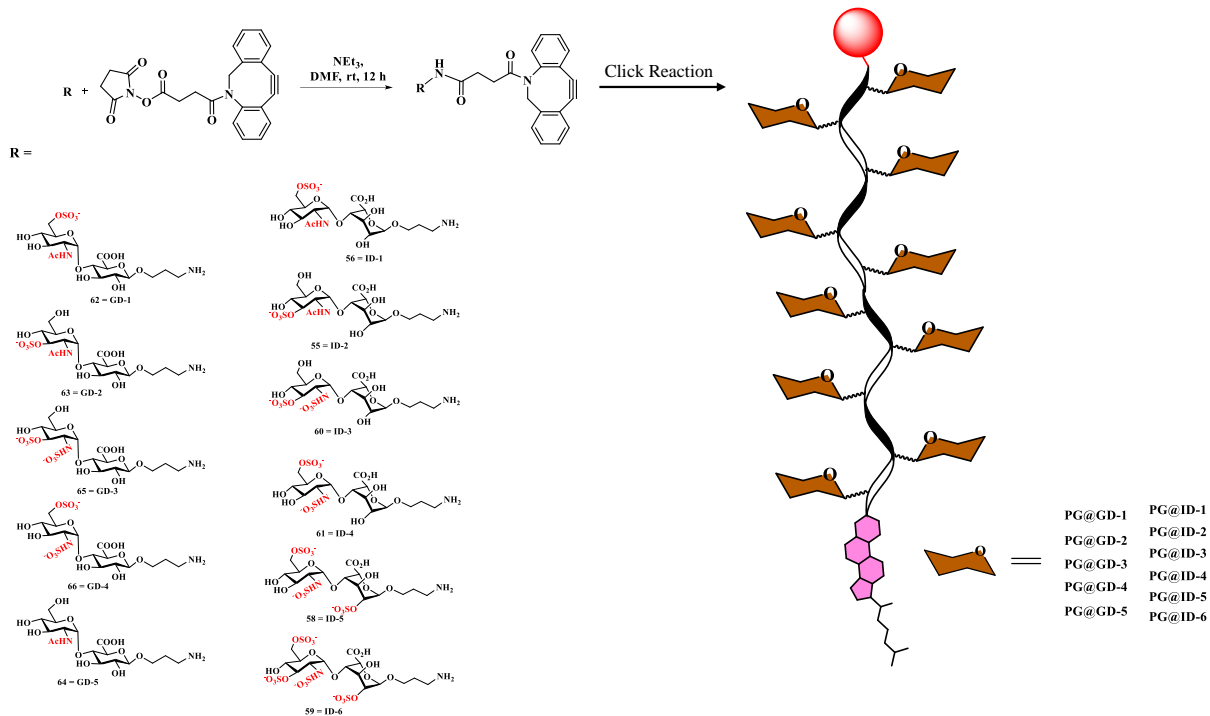
<sup>1</sup>H NMR (600 MHz, Methanol-d<sub>4</sub>) δ 7.71 (td, J = 14.7, 13.5, 8.4 Hz, 5H), 7.38 – 7.28 (m, 2H), 7.21 (d, J = 8.9 Hz, 1H), 7.14 (q, J = 7.4 Hz, 4H), 7.10 – 6.90 (m, 6H), 5.93 (dd, J = 21.7, 13.6 Hz, 1H), 5.05 (d, J = 5.0 Hz, 1H), 3.89 – 3.75 (m, 4H), 3.68 – 3.52 (m, 11H), 3.47 – 3.29 (m, 18H), 3.27 (dd, J = 11.4, 5.9 Hz, 28H), 2.99 – 2.79 (m, 24H), 2.62 – 2.47 (m, 7H), 2.21 – 2.12 (m, 23H), 1.69 – 1.48 (m, 25H), 1.45 (d, J = 3.6 Hz, 13H), 1.35 – 1.18 (m, 39H), 1.19 – 1.07 (m, 19H), 0.97 (s, 14H), 0.90 – 0.69 (m, 15H), 0.63 (d, J = 6.5 Hz, 4H), 0.57 (dd, J = 6.6, 2.6 Hz, 8H), 0.40 (s, 3H). <sup>13</sup>C NMR (151 MHz, MeOD) δ 175.8, 175.5, 175.4, 175.2, 174.5, 173.7, 173.2, 172.6, 171.0, 153.6, 153.4, 142.6, 142.0, 141.0, 140.8, 140.4, 131.2, 131.0, 129.4, 128.7, 127.9, 125.4, 125.1, 122.2, 122.1, 121.8, 110.8, 110.6, 103.1, 79.4, 77.7, 77.4, 77.2, 70.0, 69.8, 69.5, 65.2, 63.9, 63.0, 56.7, 56.1, 53.4, 51.5, 50.1, 49.3, 48.9, 48.7, 48.6, 48.4, 48.3, 48.1, 48.0, 47.4, 46.6, 44.0, 42.2, 39.7, 39.4, 39.3, 39.1, 38.9, 37.1, 36.9, 36.7, 36.1, 35.7, 35.3, 35.2, 34.0, 33.7, 32.2, 31.8, 31.8, 30.9, 29.6, 29.5, 29.5, 29.5, 29.3, 29.2, 29.1, 29.0, 28.9, 28.2, 28.1, 27.9, 27.6, 27.5, 27.3, 26.6, 25.4, 25.2, 24.8, 24.1, 23.7, 22.5, 22.5, 22.2, 21.0, 19.1, 18.4, 16.3, 13.7, 11.6. MALDI-TOF/TOF Calcd for C<sub>185</sub>H<sub>282</sub>N<sub>61</sub>O<sub>29</sub><sup>+</sup> [M] is 3822.2461, found is 3822.2245.

## **7. General Procedure for Conjugation of Sugar Molecules**

The sugar derivative was dissolved in 400 μL of dry DMF and stirred for 5 minutes. Next, NEt<sub>3</sub> (2 equivalents) was added, followed by dropwise addition of the solution of DBCO-NHS in 400 μL, while stirring at room temperature. After 12 hours, DMF was evaporated, and the crude product was purified using LH-20 size exclusion chromatography. Then, a compound with an azide linker (1 eq.) was dissolved in 500 μL of CHCl<sub>3</sub> and 500 μL of MeOH. Subsequently, a solution of sugar molecules (1.2 eq. per azide of peptide) in H<sub>2</sub>O (1 mL) was added, followed by the addition of another 500 μL of MeOH. The resulting solution was stirred for 7 – 8 days at room temperature. The progress of the reaction was monitored by infrared spectroscopy (IR). After the completion of the reaction (approximately 7 - 8 days), the solvent was evaporated under reduced pressure. The crude product was then purified using a 10 kDa centrifuge filter,



followed by a C-18 column with reversed-phase high-performance liquid chromatography (HPLC). A gradient system of (95% MeOH+5% H<sub>2</sub>O) and (95% water + 5% H<sub>2</sub>O) at a flow rate of 1 mL/min was employed for compound purification. Confirmation of the product was achieved through IR, NMR, and HRMS-ESI. When the reaction got completed, we can observe disappearance of azide peak at 2210 nm<sup>-1</sup> in IR spectrum.



### PG@ID-1

The compound was characterized using <sup>1</sup>H NMR, IR and HR-ESI-MS (*m/z*): [M]<sup>9-</sup>, calcd is 1336.6012; found, 1336.0099. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is ~ 19.5 min.

### PG@ID-2

The compound was characterized using <sup>1</sup>H NMR, IR and HR-ESI-MS (*m/z*): [M]<sup>9-</sup>, calcd is 1336.6012; found 1336.5524. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is ~ 19.5 min.

### PG@ID-3

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{19-}$  calcd is 652.3311; found, 652.2312. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 17.5$  min.

#### **PG@ID-4**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{19-}$  calcd is 652.3311; found, 652.2242. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 17.4$  min.

#### **PG@ID-5**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{29-}$  calcd is 454.1048; found, 454.0741. found, 1336.0099. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 16.1$  min.

#### **PG@ID-6**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{39-}$  calcd is 356.5128; found,  $[\text{M}]^{39-} + \text{Na} + 5\text{H}$  is 384.1110. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 15$  min.

#### **PG@GD-1**

The compound was characterized using  $^1\text{H}$  NMR, IR and MALDI-TOF/TOF. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 22.5$  min.

#### **PG@GD-2**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{19-}$  calcd is 1336.1012; found,  $[\text{M}]^{19-} + 6\text{H}$  is 1342.3002. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 19.5$  min.

#### **PG@GD-3**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{19-}$  calcd is 1336.1012; found, 1336.2484. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 19.5$  min.

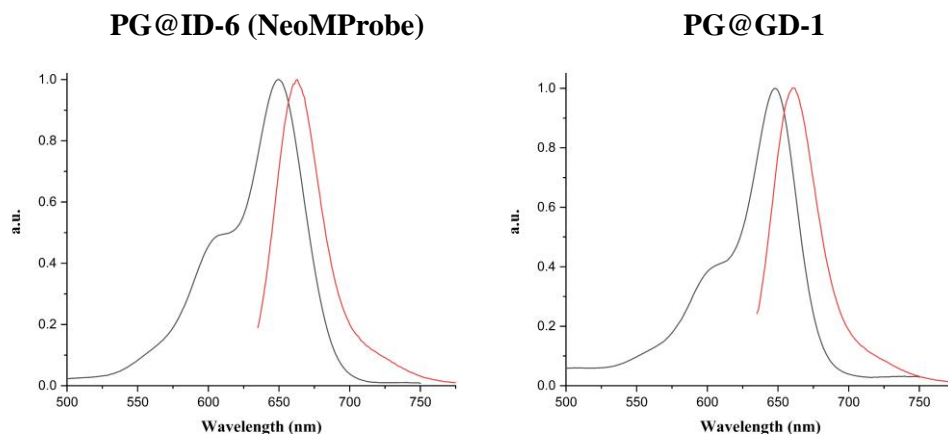
#### **PG@GD-4**

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{29-}$  calcd is 652.3311; found, 652.2471. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 17.3$  min.

### PG@GD-5

The compound was characterized using  $^1\text{H}$  NMR, IR and HR-ESI-MS ( $m/z$ ):  $[\text{M}]^{29-}$  calcd for 652.3311; found, 652.1973. The purity of compound was checked using reverse-phase high-performance liquid chromatography (HPLC) the elution time is  $\sim 17.3$  min.

## 8. Physical Characterization



Standard compound used for calculation of quantum yield is Atto 655.

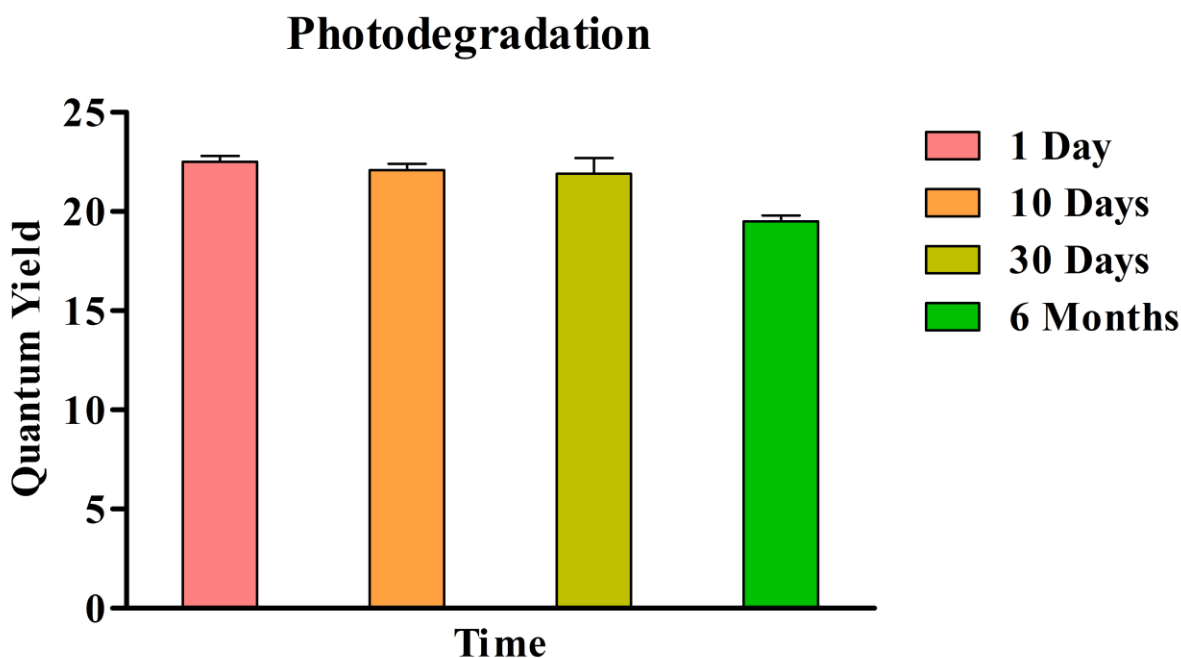
**Table S1:** Absorbance, emission and quantum yield of L-Iduronic acid-based molecules

S.No.	Compound	Absorbance (nm)	Emission (nm)	Quantum Yield
1.	PG@ID-1	648	660	22.8
2.	PG@ID-2	649	661	21.9
3.	PG@ID-3	649	662	22.3
4.	PG@ID-4	650	663	22.8
5.	PG@ID-5	648	662	22.2
6.	PG@ID-6	650	665	22.8

**Table S2:** Absorbance, emission and quantum yield of D-Glucuronic acid-based molecules

S.No.	Compound	Absorbance (nm)	Emission (nm)	Quantum Yield
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1.	PG@GD-1	649	658	21.8
2.	PG@GD-2	648	660	22.0
3.	PG@GD-3	649	662	22.3
4.	PG@GD-4	650	664	22.1
5.	PG@GD-5	651	663	22.5

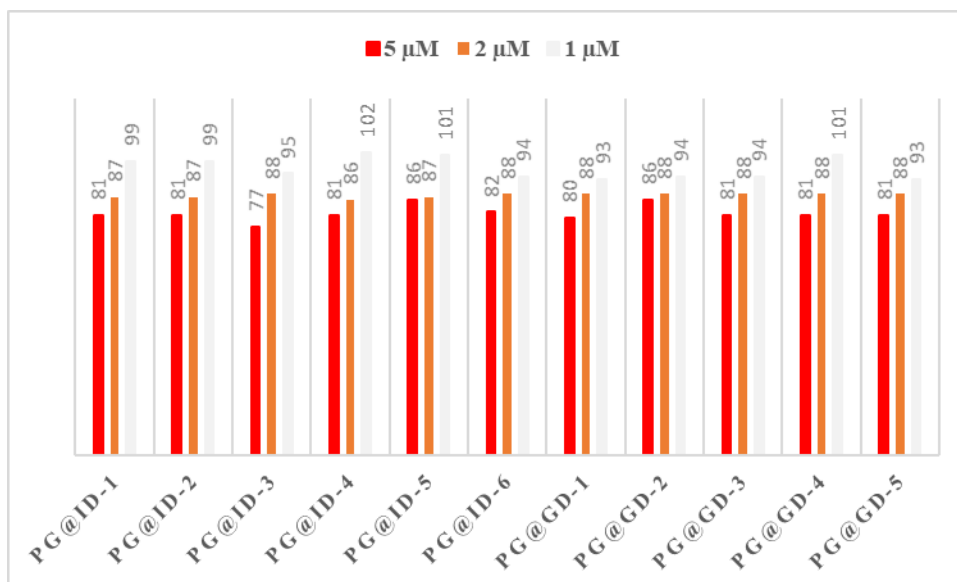


**Figure S1.** Quantum yield of the NeOMProbe at different time intervals.

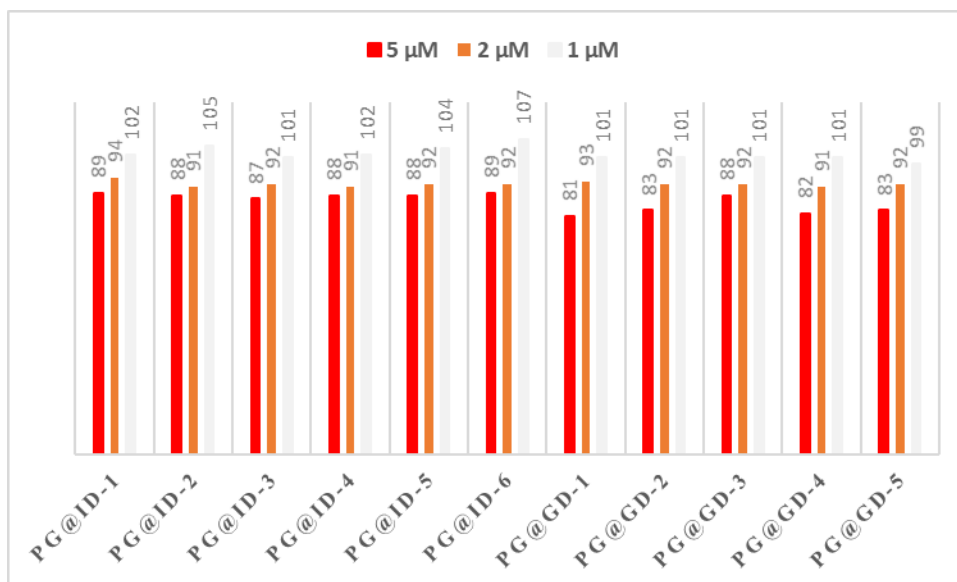
## 9. Cell Lines

MDA-MB-468 and NIH3T3 cells were cultured at 37 °C in a 5% CO<sub>2</sub> atmosphere in DMEM media supplemented with 10% fetal bovine serum (FBS) and 0.1% streptomycin.

**MTT Assay.** MTT assay was used to check toxicity of the molecules. MDA-MB-468 and NIH-3T3 cells ( $5 \times 10^3$  cells per well) were plated in 96 well plate in DMEM medium and incubated overnight followed by treatment different concentration of neoPGs and further incubation for 48 h. Afterward 10  $\mu$ l of MTT (5 mg/ml) reagent was added to each well and incubated plates further for 4h at 37 °C. Purple precipitate formed was dissolved by adding 100  $\mu$ l of DMSO and plate was read at 570 nm.

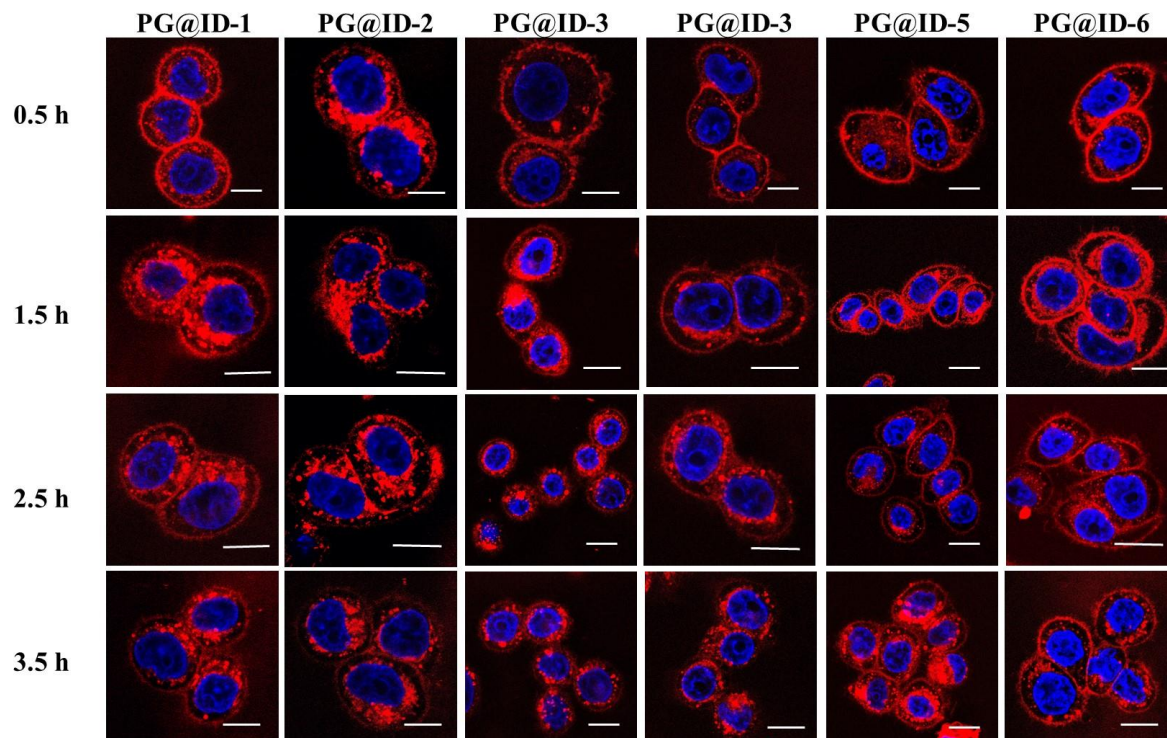


**Fig S2a.** MTT assay of NeoPGs in NIH-3T3 cells after 48 hours of incubation. The experiment represents the average of three independent experiments

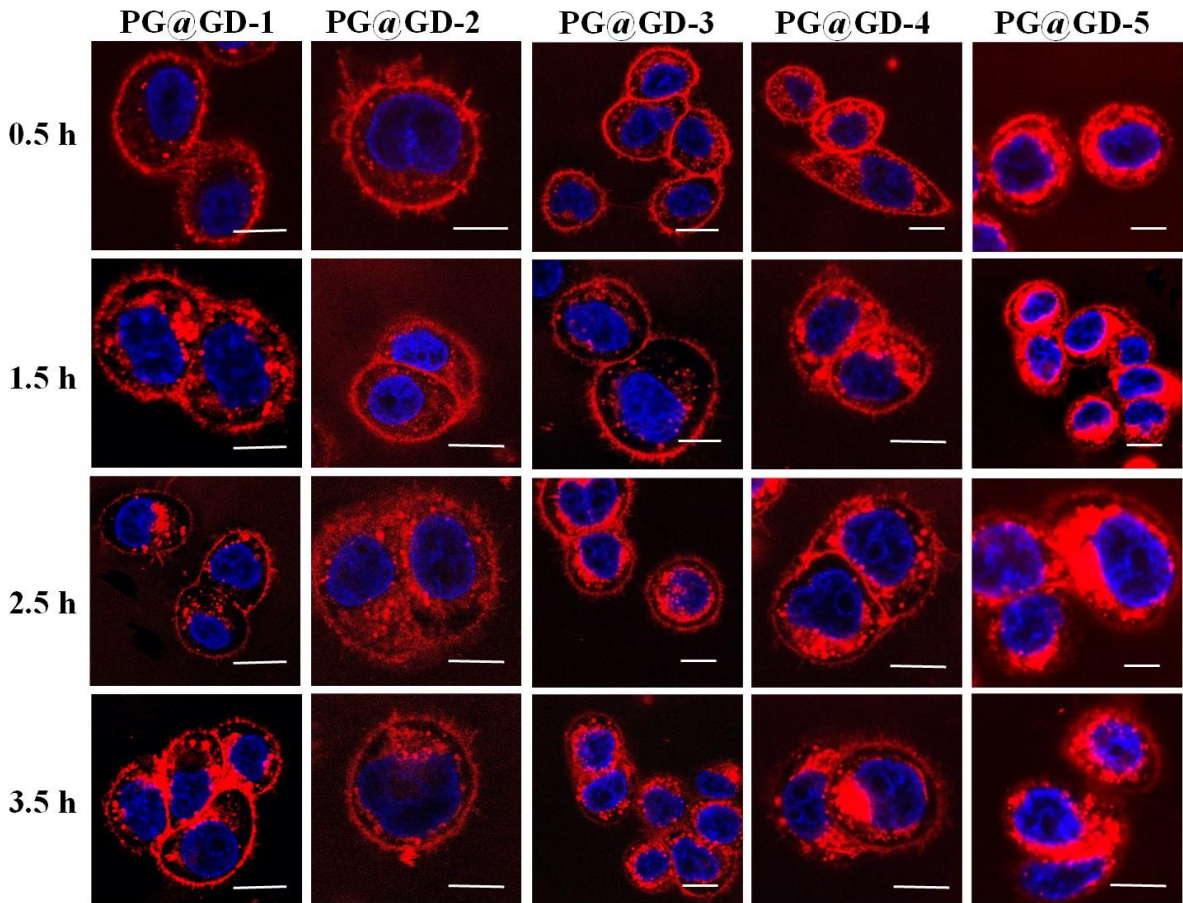


**Fig S2b.** MTT assay of NeoPGs in MDA-MB-468 cells after 48 hours of incubation. The experiment represents the average of three independent experiments

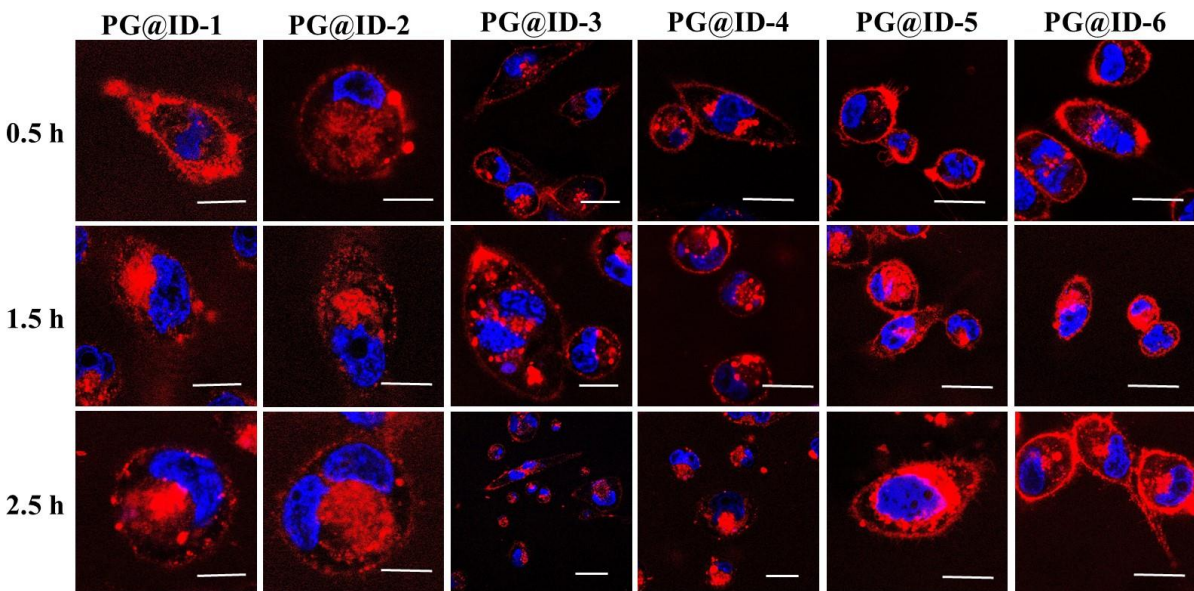
## 10. Confocal Imaging Study



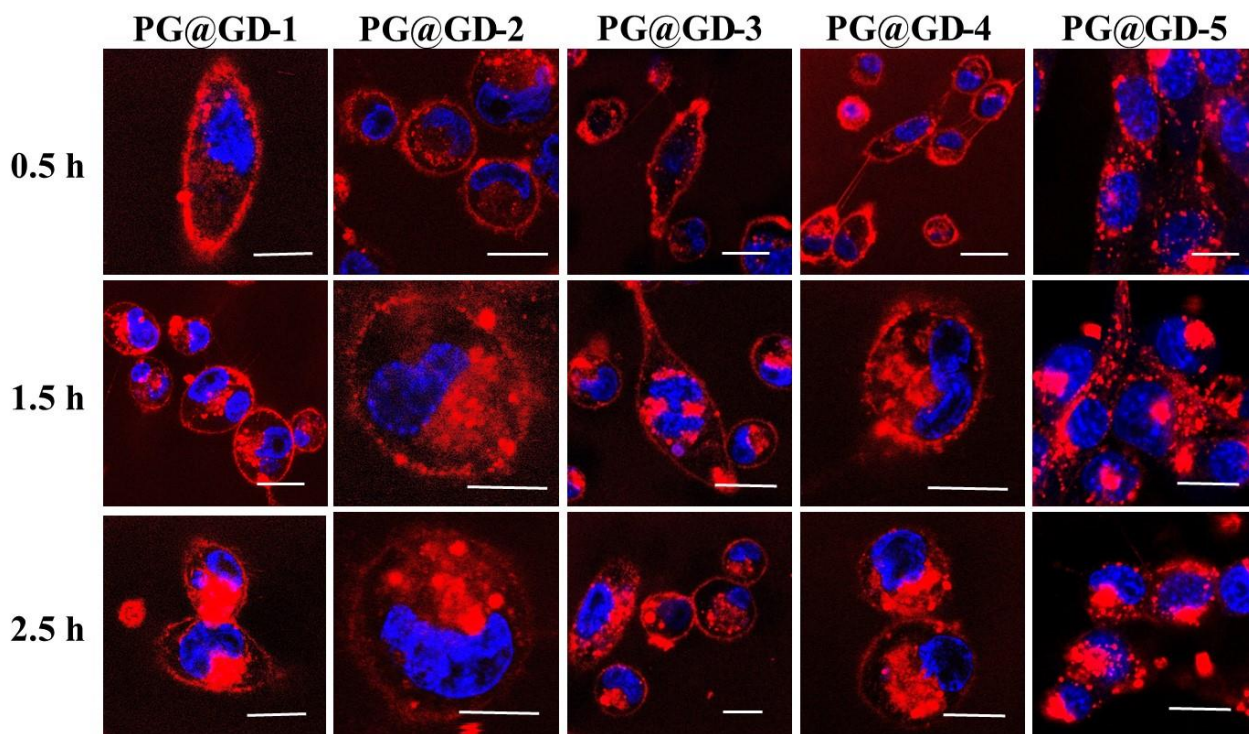
**Fig. S3:** Time dependent confocal images of MDA-MB-468 treated with 2  $\mu$ M of PG@ID's.



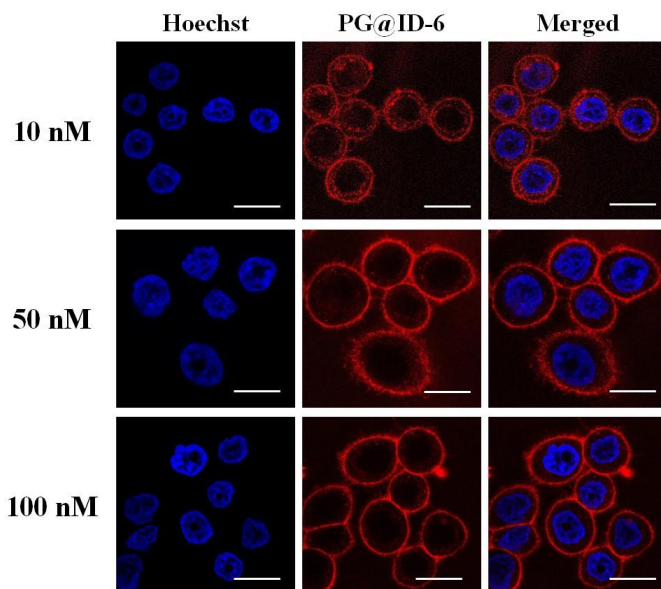
**Fig. S4:** Time dependent confocal images of MDA-MB-468 using 2  $\mu$ M of PG@GD's library.



**Fig. S5:** Time dependent confocal images of NIH3T3 treated with 2  $\mu$ M of PG@ID's library.



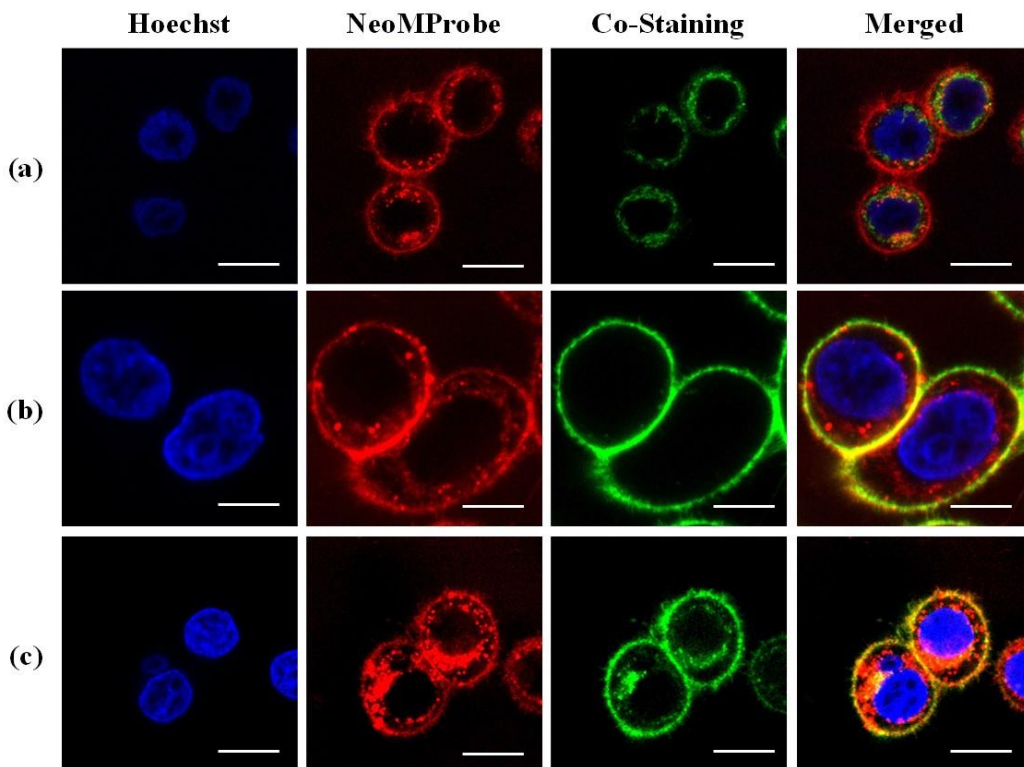
**Fig. S6:** Time dependent confocal images of NIH3T3 treated with 2  $\mu$ M of PG@GD's library.



**Fig. S7:** Confocal images of MDA-MB-468 treated with PG@ID- 6 from 10 nM to 100 nM concentration after 30 min of incubation.

## 11. Co-Localization Assay in Cellular Level

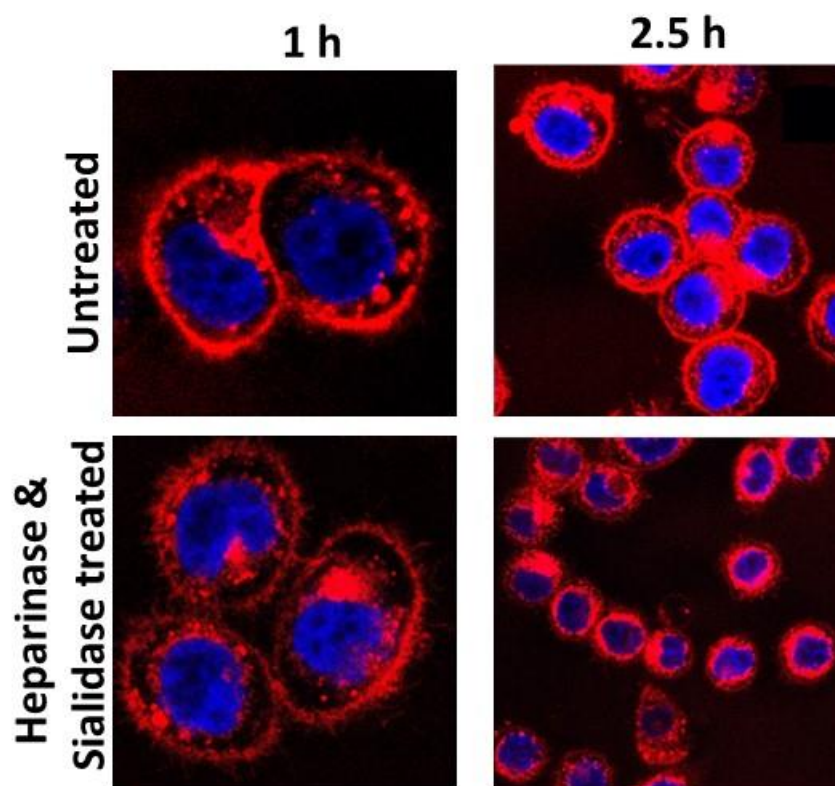




**Fig. S8:** Co-staining confocal images of MDA-MB-468 treated with NeoMProbe (PG@ID- 6) and (a) DiO (b) FITC-CD44 antibody and (c) FITC-E-cadherin antibody

## 12.Heparinase and sialidase Treated Assay

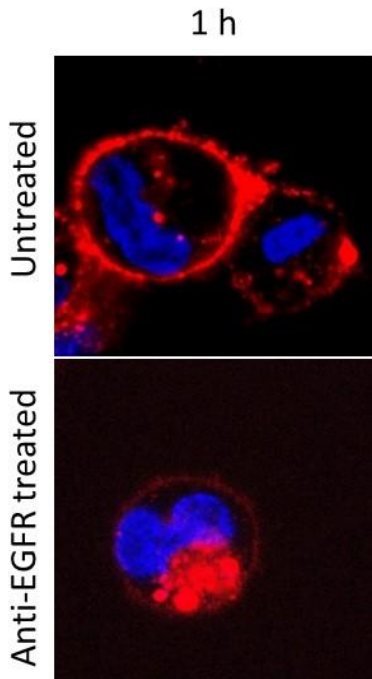
To investigate the glyocalyx anionic interaction modulating the activity, we first treated the cell with heparinase (0.03 IU/ml) for 1h at 37 °C, followed by sialidase (0.03 IU/ml) for 30 min at 4 °C respectively. Next cells incubated with **PG@ID-6 (NeoMProbe)** at 2  $\mu$ M concentration. Our results showed significant amount of molecules internalized after 1 h and 2.5 h respectively.



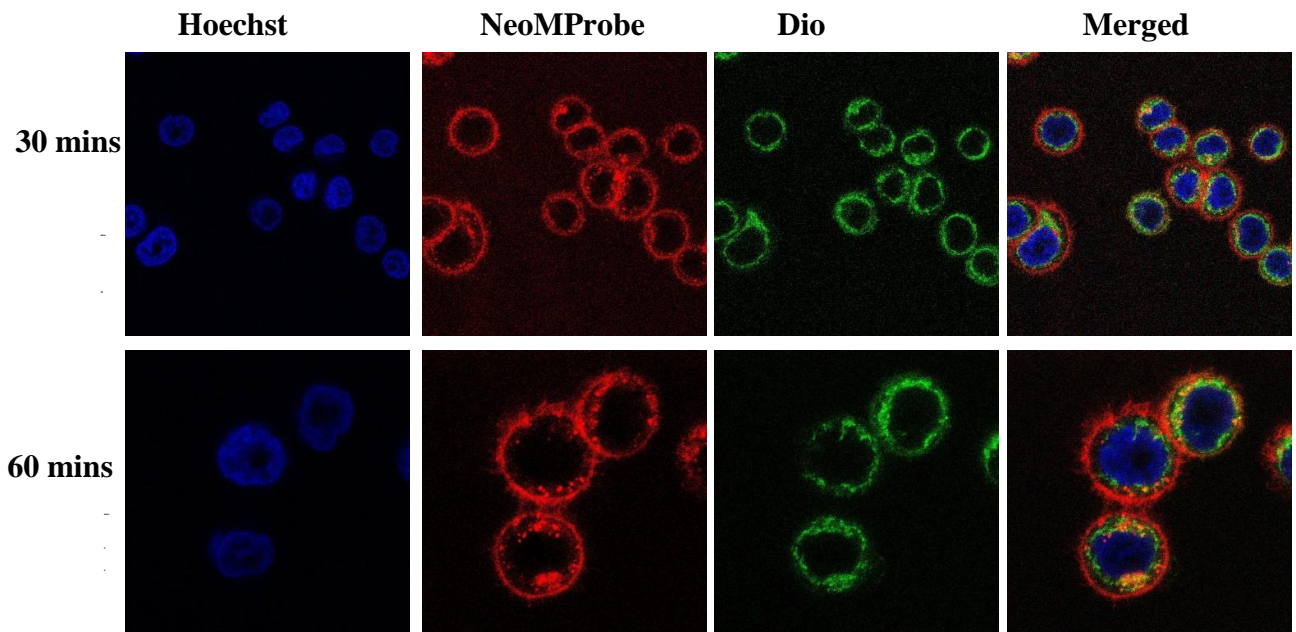
**Fig. S9:** NeoMProb plasma membrane stability in the presence and absence of Heparinase and sialidase.

### 13. Anti-EGFR treated assay.

To investigate the EGFR receptor stabilizing the PG@ID-6. We first treated the cells with mice IgG anti-EGFR antibody for (0.05 IU/ml) for 1 h and later incubated with PG@ID-6 for 1 h and imaged. Our results showed significant decrease in the plasma membrane decoration of PG@ID-6, indicating that the molecules bind to cell surface receptor and stabilize.



**Fig S10.** NeoMProbe plasma membrane stability in the presence and absence of anti-EGFR antibody.



**Figure S11.** Costaining images of Dio and NeoMProbe of MDA-MB-468 after 30 and 60 mins.

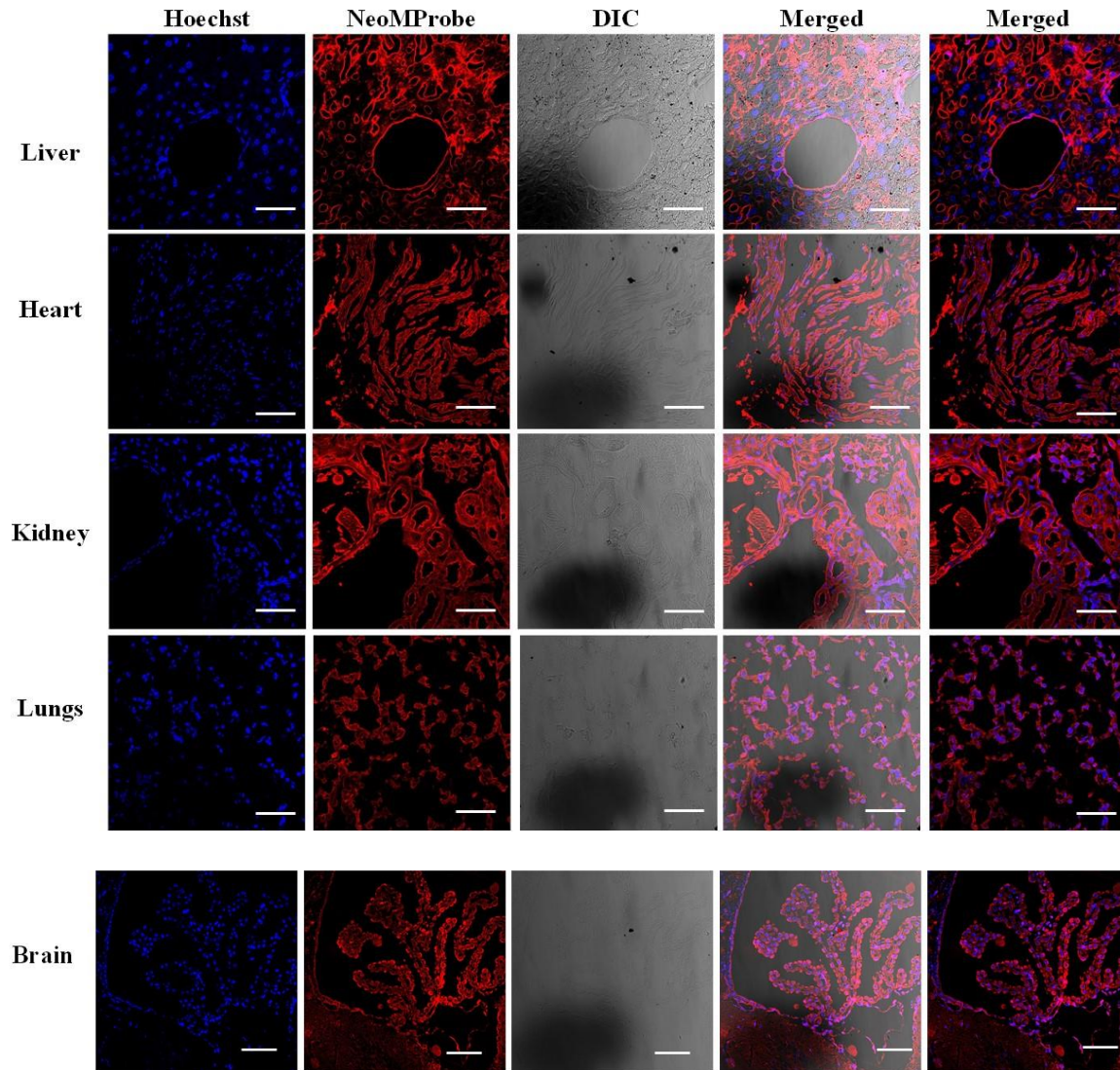
## **12. FACS Analysis**

Cells ( $5 \times 10^5$  cells per well) were seeded into a 24-well plate and incubated overnight. Subsequently, the cells were subjected to treatment with PG's mimetics for 1 hour. Followed by washing 3 times and re-suspended in FACS buffer for analysis.

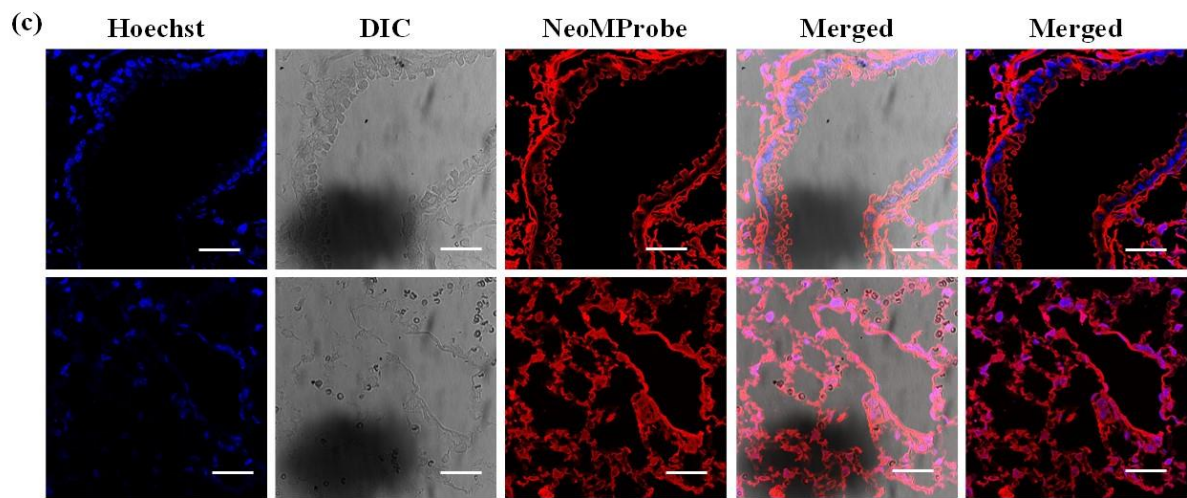
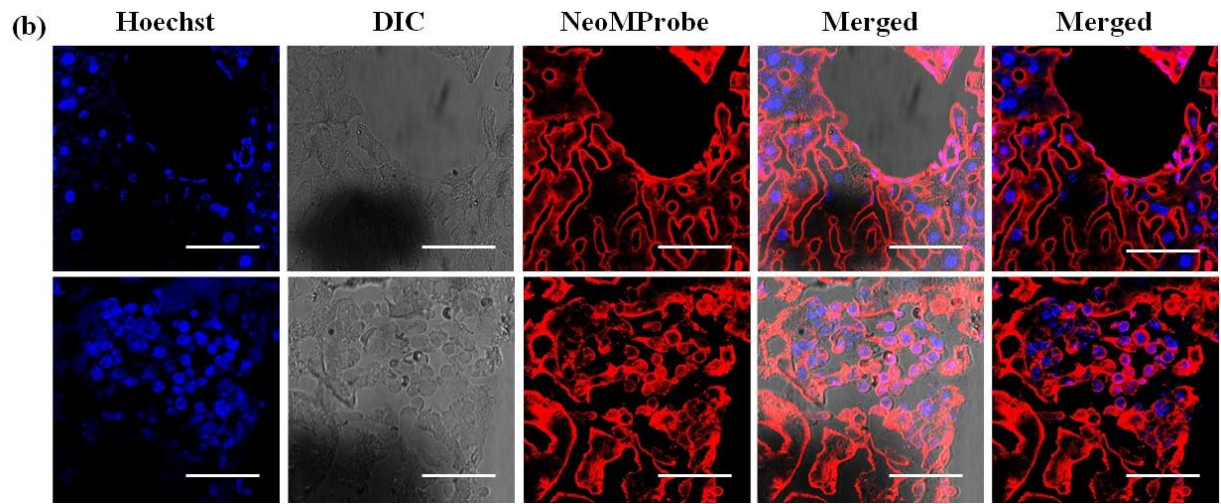
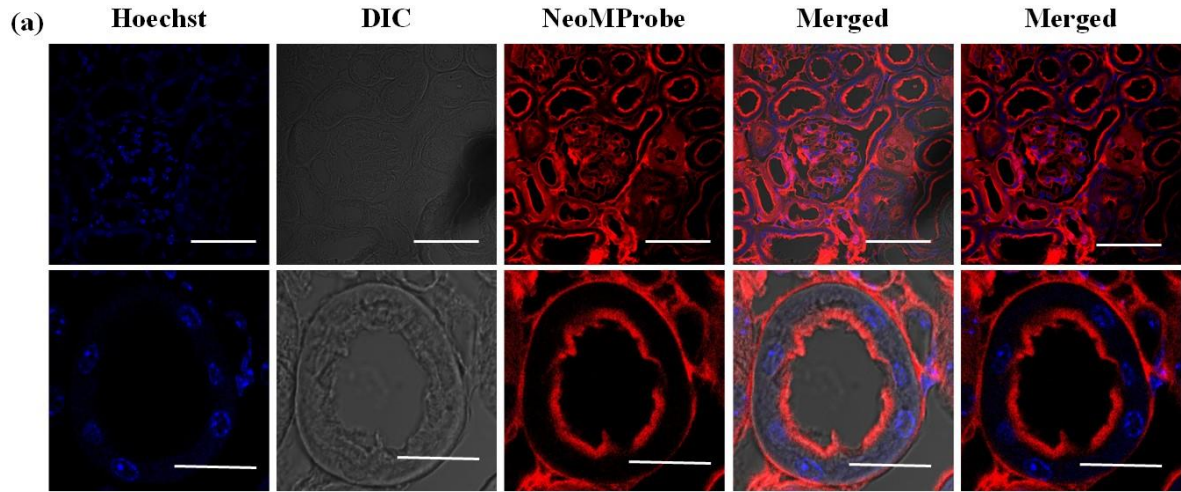
## **13. Tissue Section Images**

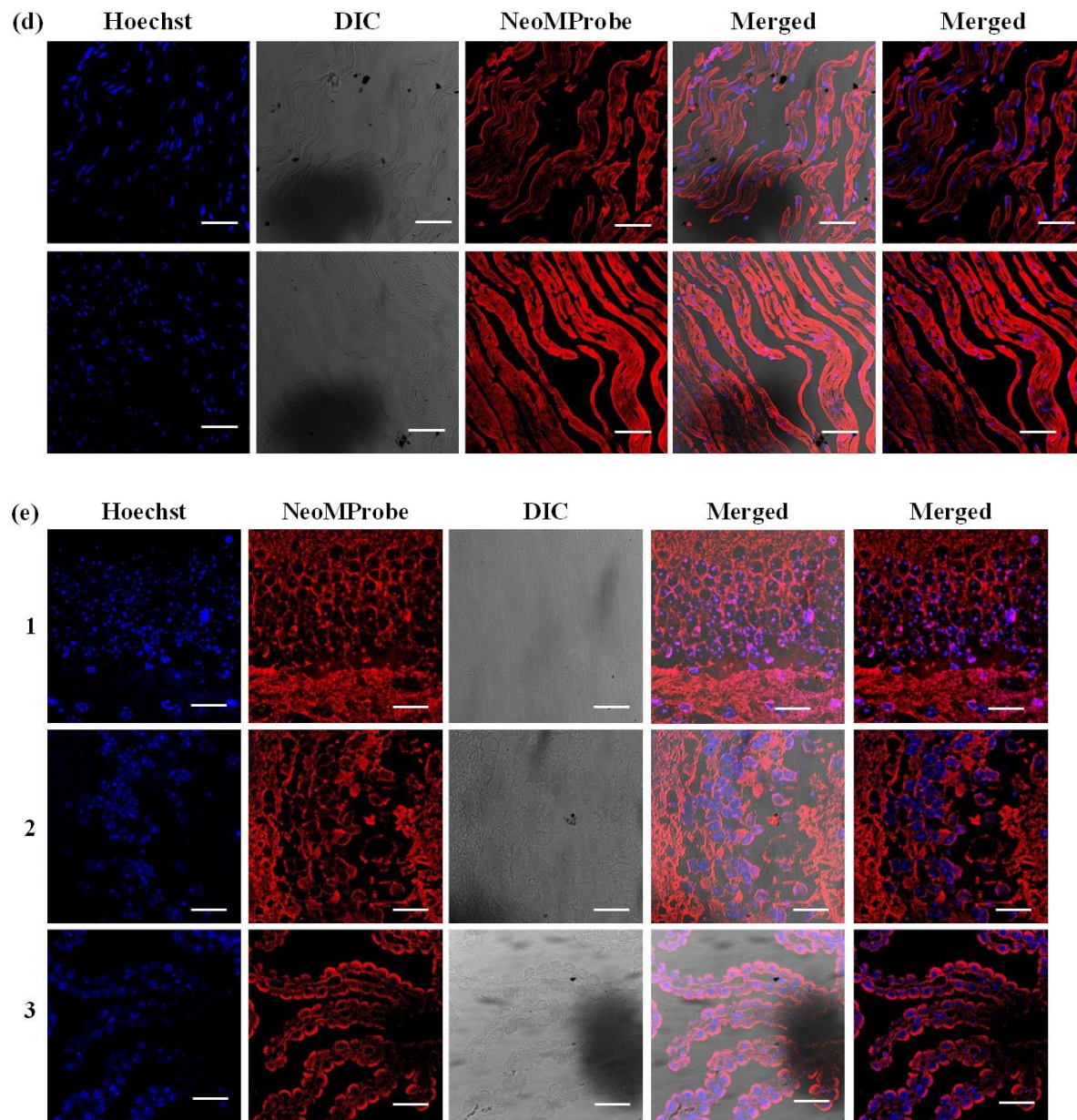
### **20a. *Ex vivo* tissue Section**

To illustrate the potential of the PG's mimetics as a tissue membrane staining, we stained the mouse tissue sections. For dewaxing of paraffin-embedded mouse tissue sections: Slides were initially heated at 60 °C for 15 minutes, then placed into glass slide holders and subjected to dewaxing as follows: Twice in 100% xylene wash for 5 minutes each, followed by immersion in 100% isopropanol with brief agitation (10–20 seconds) for 3 minutes, and finally twice in H<sub>2</sub>O with brief agitation. For rehydration, the glass slides were immersed in PBS for 30 minutes. Subsequently, tissue sections were treated with a 2 μM and 500 nM of **PG@ID-6** for 1 hour at rt, followed by PBS washing for 3 minutes. Finally, sections were treated with Hoechst for 20 minutes at rt, followed by PBS washing 2 times for 3 minutes, and then subjected to confocal imaging.

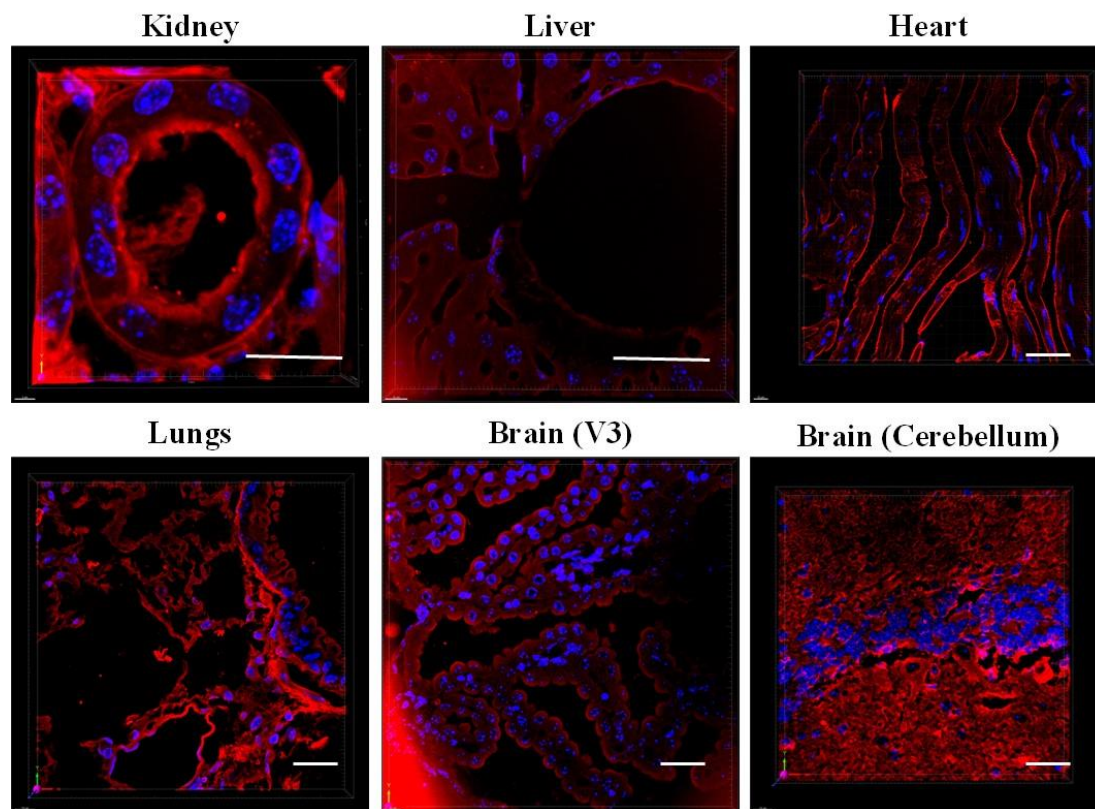


**Fig. S12:** Confocal images of tissue sections using NeoMProbe@500 nM concentration after 1 h of treatment





**Fig. S13:** Confocal images of tissue sections using NeoMProbe@2  $\mu$ M concentration after 1 h of treatment (a) Kidney Section (b) Liver Section (c) Lungs Section (d) Heart Section (e) Brain section (1. Hippocampus region 2. Cerebellum region 3. Third ventricle region (V3))



**Fig. S14:** Z stack images of tissue section using NeOMProbe@2  $\mu\text{M}$  concentration

#### 14. *Ex vivo* tissue Section Co-Staining

To illustrate the potential of the PG's mimetics as a tissue membrane staining, we stained the mouse tissue sections. For dewaxing of paraffin-embedded mouse tissue sections: Slides were initially heated at 60 °C for 15 minutes, then placed into glass slide holders and subjected to dewaxing as follows: Twice in 100% xylene wash for 5 minutes each, followed by immersion in 100% isopropanol with brief agitation (10–20 seconds) for 3 minutes, and finally twice in H<sub>2</sub>O with brief agitation. For rehydration, the glass side were immersed in PBS for 30 minutes:

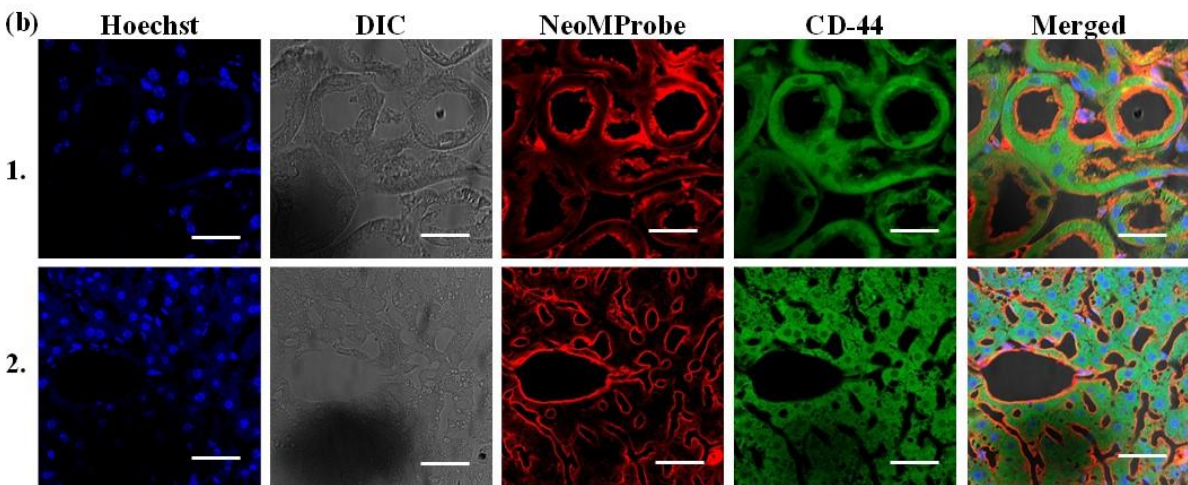
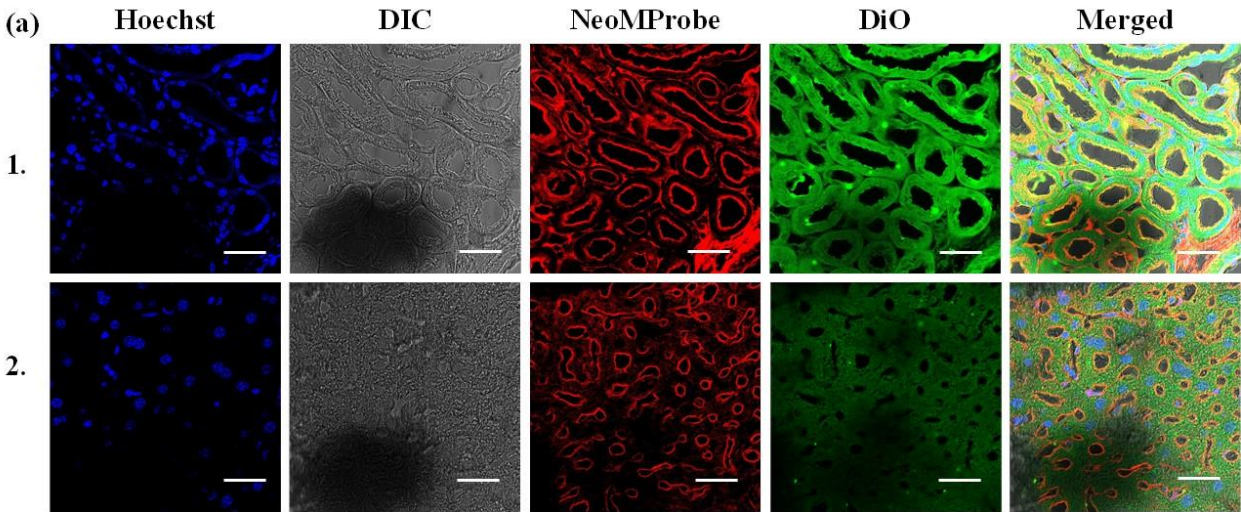
(a) Subsequently, tissue sections were treated with a 2  $\mu\text{M}$  of **PG@ID-6** and DiO ((3,3' - dioctadecyloxycarbocyanine, perchlorate) for 30 minutes at rt, followed by PBS washing for 3 minutes. Finally, sections were treated with Hoechst for 20 minutes at rt, followed by PBS washing 2 times for 3 minutes, and then subjected to confocal imaging.

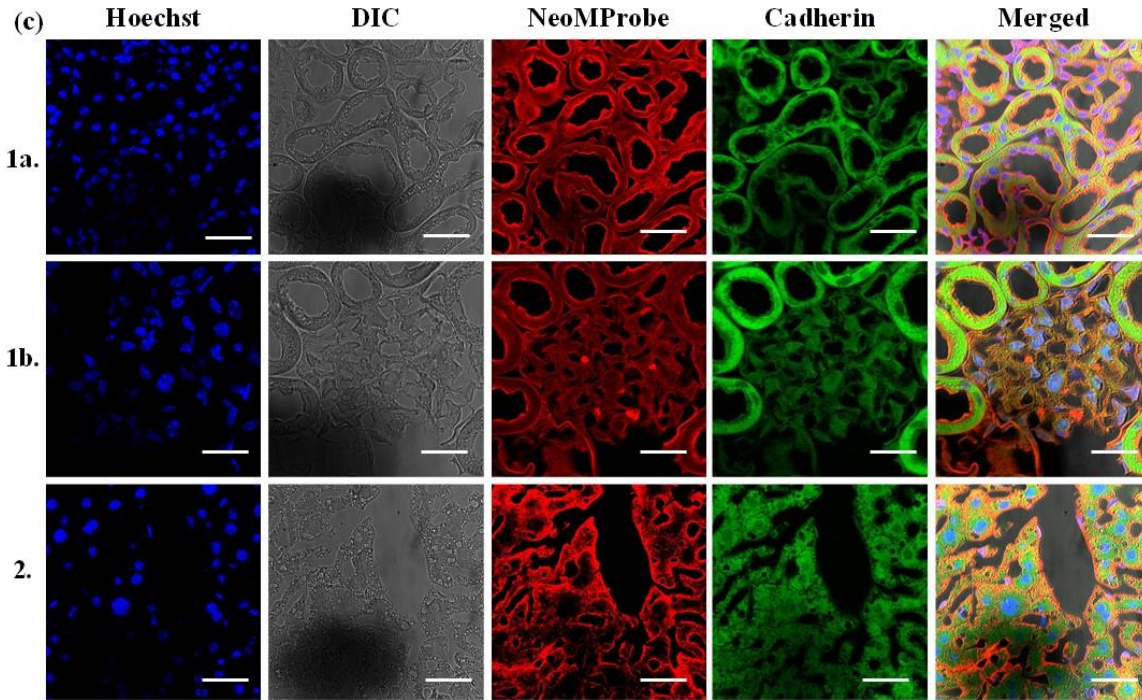
(b) Subsequently, it was treated with 0.1 % trypsin for 1 h followed by washing with PBS for 5 min 2 times. After that permeabilization was done using mixture of PBS and 0.25 % of triton X100 followed by washing with PBS for 5 min 2 times. Then, using 1% CD-44 anti-body in PBS



staining was done for 2 h at rt followed by washing with PBS for 5 min 2 times. Finally, tissue sections were treated with a 2  $\mu$ M of **PG@ID-6** and Hoechst for 30 minutes at rt, followed by PBS washing for 3 minutes 2 times, and then subjected to confocal imaging.

(c) Subsequently, it was treated with 0.1 % trypsin for 1 h followed by washing with PBS for 5 min 2 times. After that permeabilization was done using mixture of PBS, gelatin 1:1 and 0.25 % of triton X100 followed by washing with PBS for 5 min 2 times. Then, using 1% E-cadherin anti-body in PBS staining was done for 3 h at rt followed by washing with PBS for 5 min 2 times. Finally, tissue sections were treated with a 2  $\mu$ M of **PG@ID-6** and Hoechst for 30 minutes at rt, followed by PBS washing for 3 minutes 2 times, and then subjected to confocal imaging.





**Fig. S15:** Co-staining confocal images of **1** is kidney and **2** is liver using (a) DiO (b) CD-44 (c) E-Cadherin

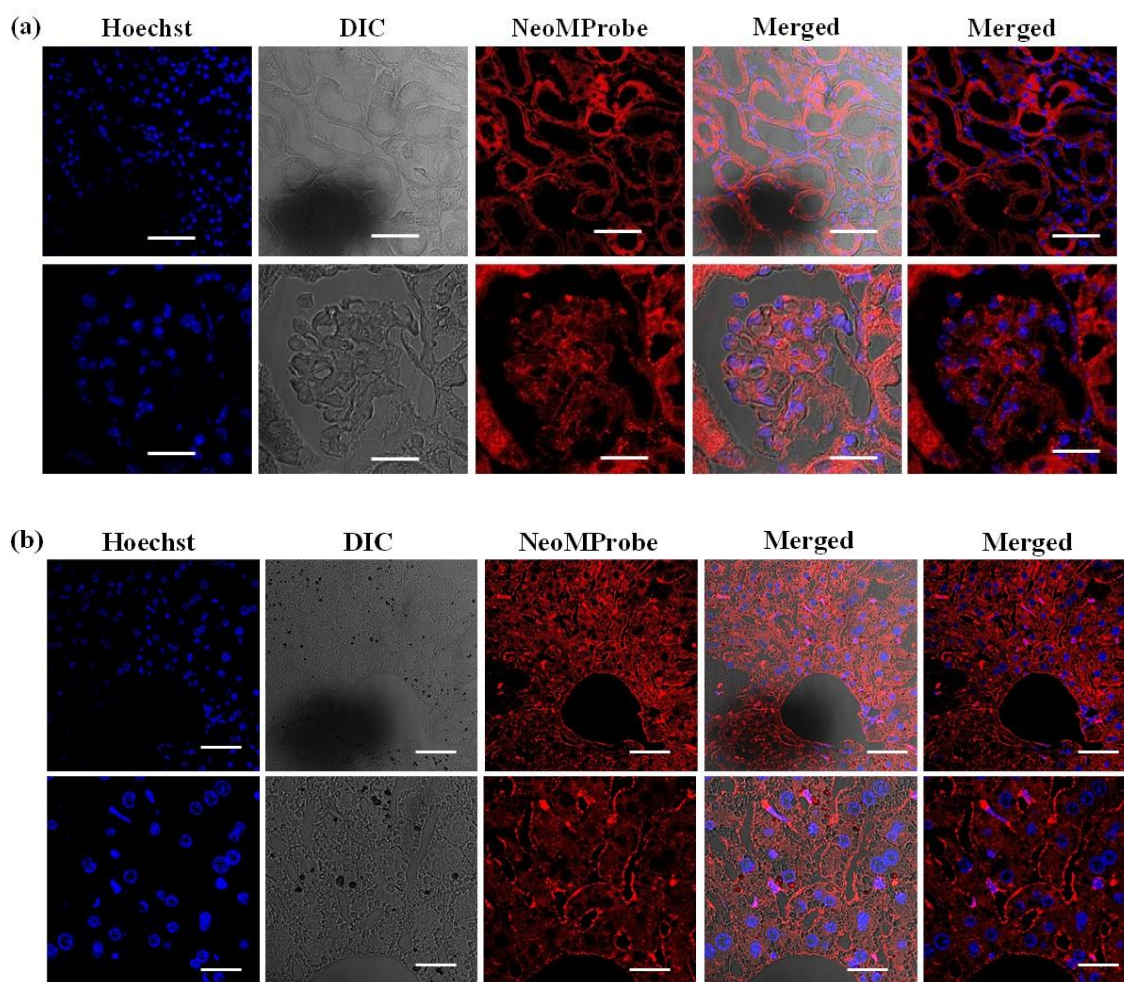
## 15. Animal Studies

### 15a. *In vivo* Studies

BALB/c 6-8 weeks old were taken from the National Facility for Gene Function in Health and Disease of IISER, Pune. Prior to experiments the mice were maintained in an animal house, with the proper amount of food and water. All experiments were performed in accordance with the relevant guidelines and regulations of the Institutional Animal Ethical Committee, set up by CPCSEA, Govt. of India. The mice (n = 3) were anesthetized by ketamine. After that **PG@ID-6 (NeoMProbe)** 1.8 mg/Kg were injected intraperitoneal. Then *in vivo* imaging was done using Quantum GX2 Micro-CT system (*excitation* = 640 nm and *emission* = 680 nm). First, before injection image was taken marked at 0 h, after injection at different time interval imaging was done upto 24 h. One mouse was sacrificed after 3 h. Subsequently, major organs including kidney and liver were collected and fixed with 4% paraformaldehyde. Organs were embedded in paraffin and 1 mm sections were cut using a microtome. The tissue sections were fixed on a poly- L -lysine-coated glass plates.

### 15b. Post Injection *in vivo* Tissue Section

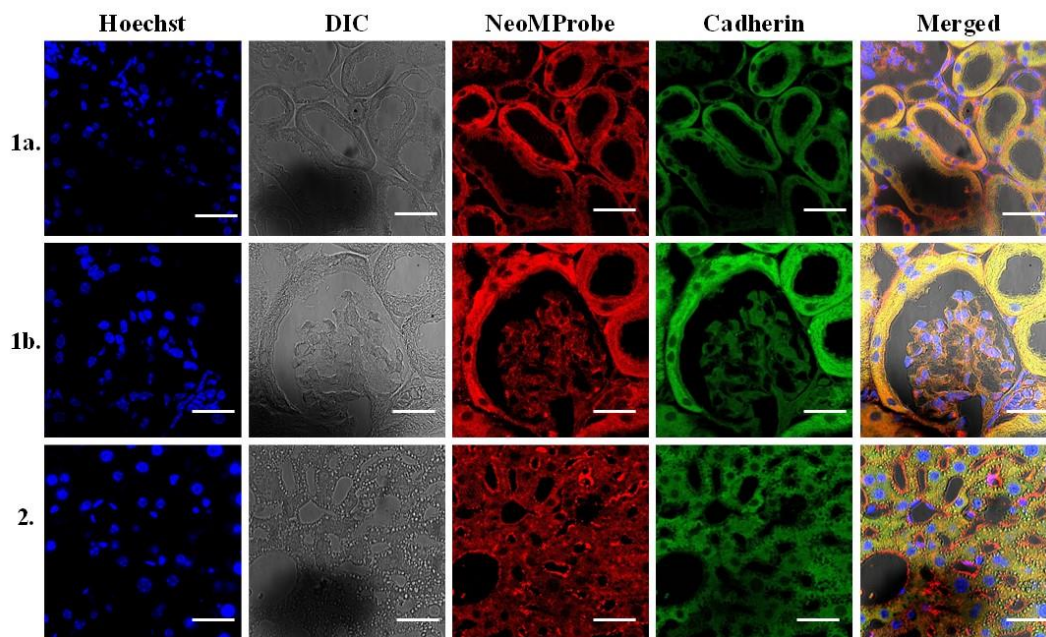
To illustrate the potential of the PG's mimetics as an *in vivo* tissue membrane stain, we stained the mouse tissue sections. For dewaxing of paraffin-embedded mouse tissue sections: Slides were initially heated at 60 °C for 15 minutes, then placed into glass slide holders and subjected to dewaxing as follows: Twice in 100% xylene wash for 5 minutes each, followed by immersion in 100% isopropanol with brief agitation (10–20 seconds) for 3 minutes, and finally twice in H<sub>2</sub>O with brief agitation. For rehydration, the glass slides were immersed in PBS for 30 minutes. Subsequently, tissue was treated with Hoechst for 20 minutes at rt, followed by PBS washing 2 times for 3 minutes, and then subjected to confocal imaging.



**Fig. S16:** Confocal images of tissue sections using NeoMProbe post injection (a) Kidney Section  
(b) Liver Section

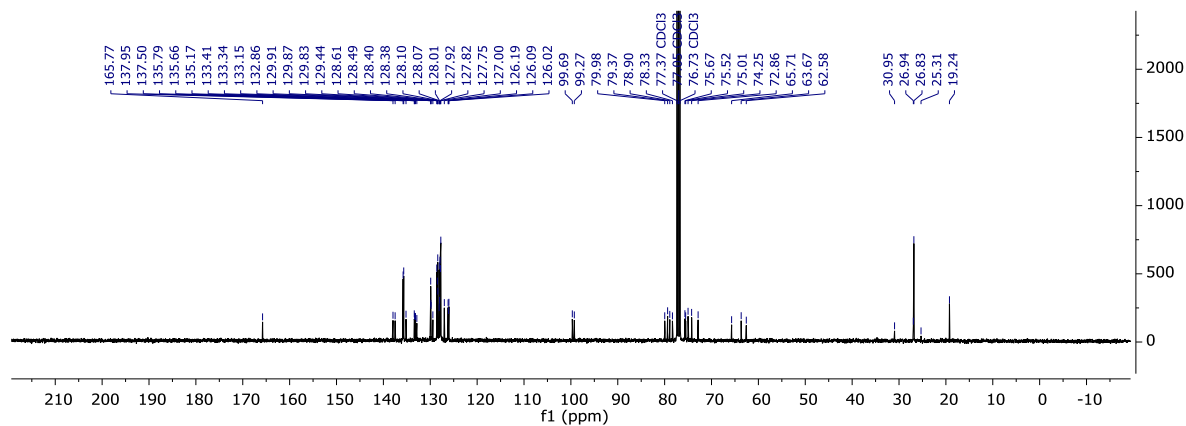
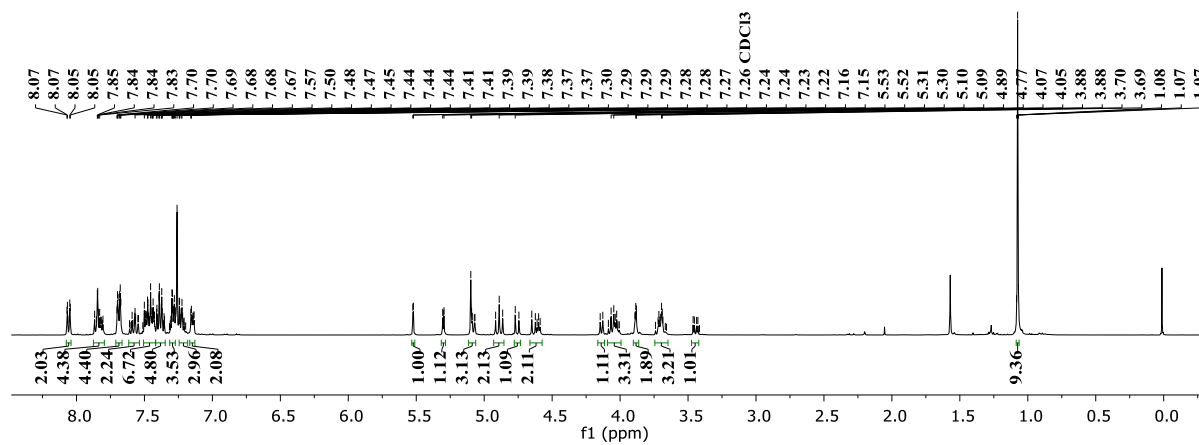
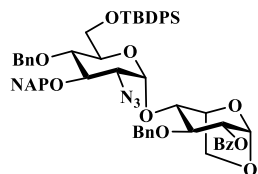
### 15c. Post Injection *in vivo* Tissue Section Co-Staining

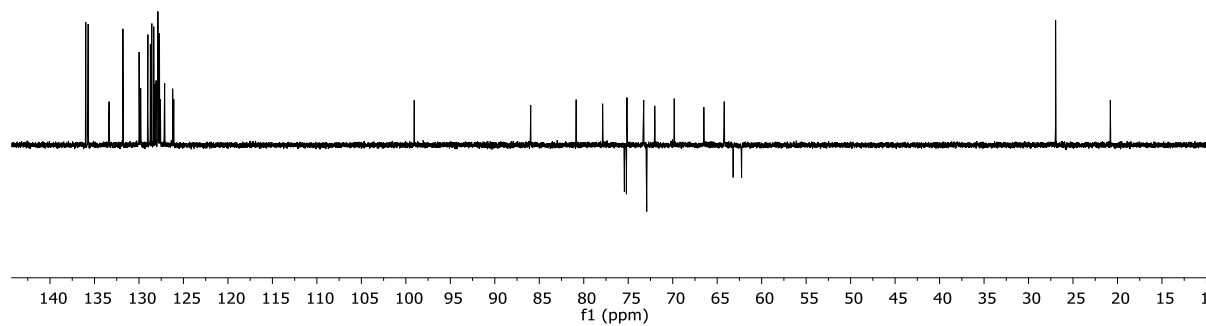
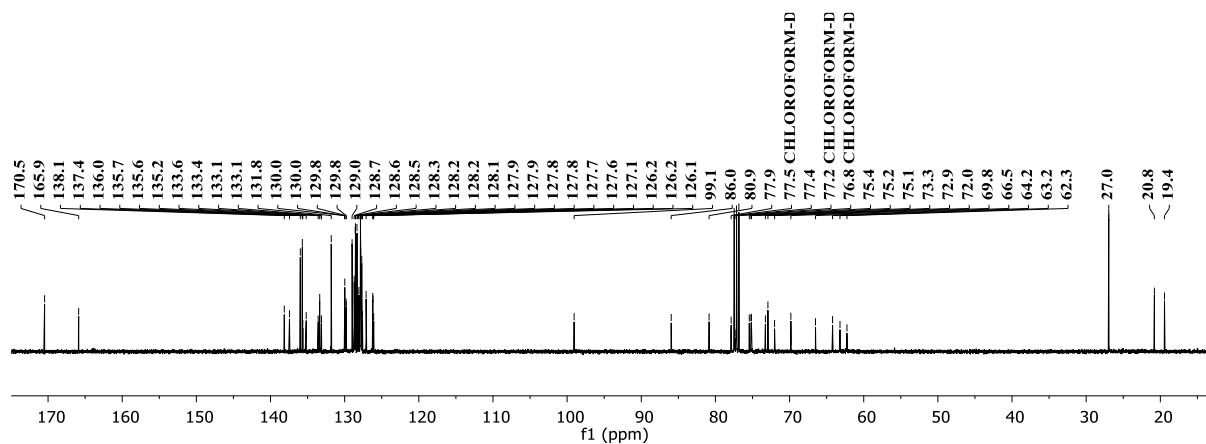
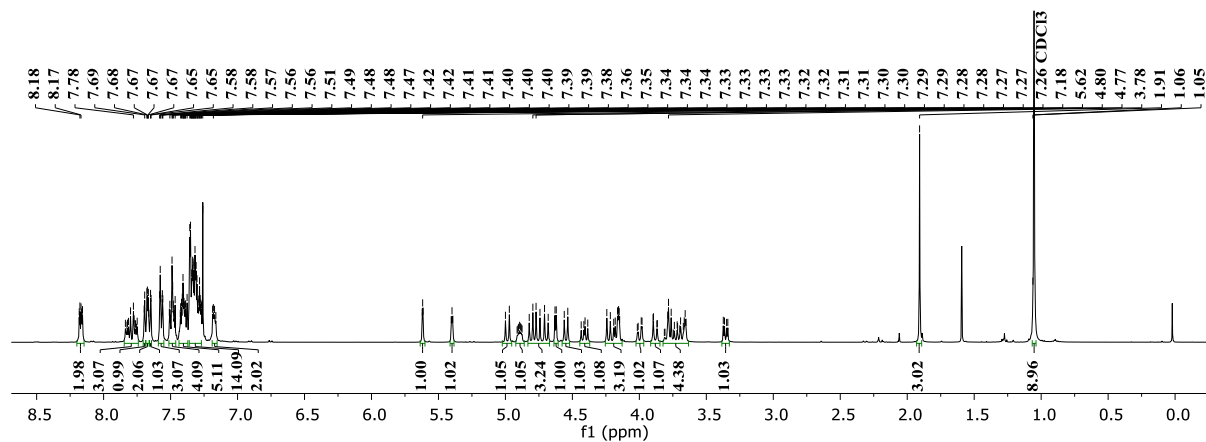
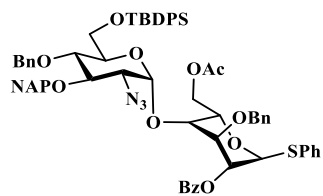
To illustrate the potential of the PG's mimetics as a tissue membrane staining, we stained the mouse tissue sections. For dewaxing of paraffin-embedded mouse tissue sections: Slides were initially heated at 60 °C for 15 minutes, then placed into glass slide holders and subjected to dewaxing as follows: Twice in 100% xylene wash for 5 minutes each, followed by immersion in 100% isopropanol with brief agitation (10–20 seconds) for 3 minutes, and finally twice in H<sub>2</sub>O with brief agitation. For rehydration, the glass slides were immersed in PBS for 30 minutes. Subsequently, it was treated with 0.1 % trypsin for 1 h followed by washing with PBS for 5 min 2 times. After that permeabilization was done using mixture of PBS, gelatin 1:1 and 0.25 % of triton X100 followed by washing with PBS for 5 min 2 times. Then, using 1% E-cadherin antibody in PBS staining was done for 3 h at rt followed by washing with PBS for 5 min 2 times. Finally, tissue sections were treated with a 2 μM of **PG@ID-6** and Hoechst for 30 minutes at rt, followed by PBS washing for 3 minutes 2 times, and then subjected to confocal imaging.



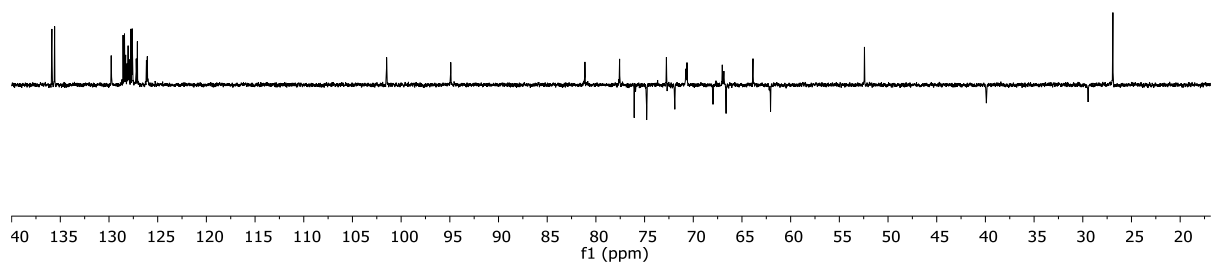
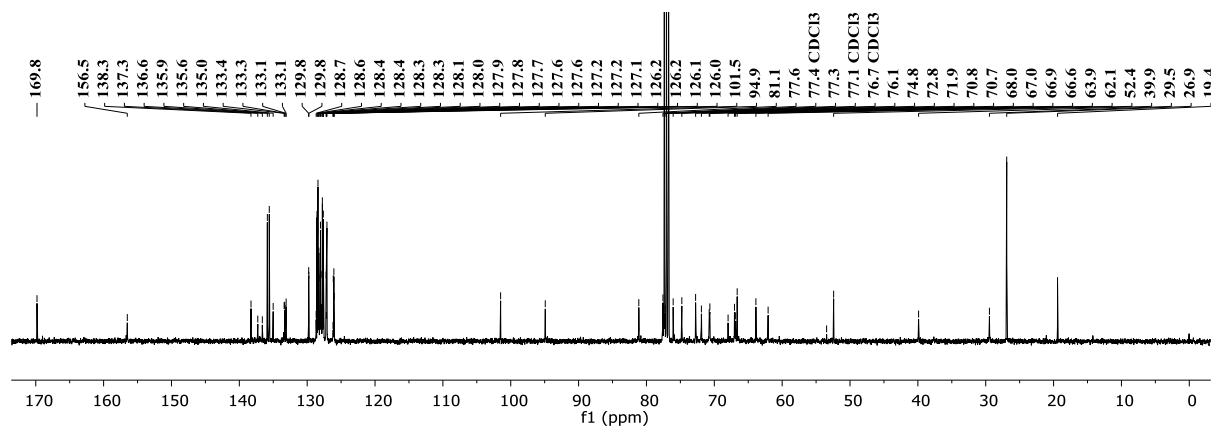
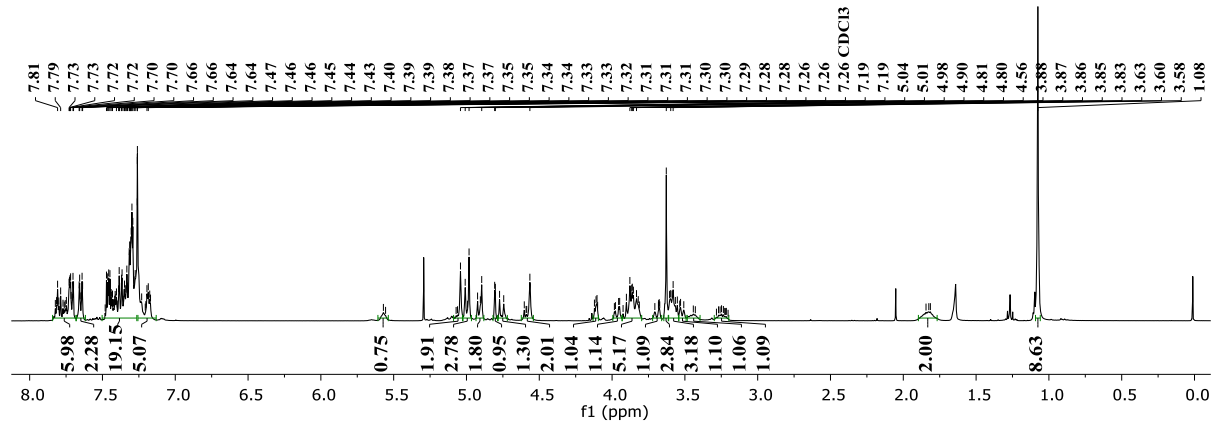
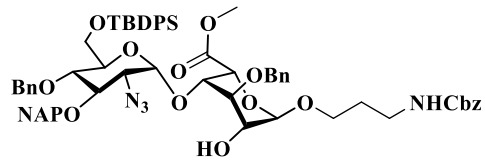
**Fig. S17:** Co-staining confocal images of **1a** and **1b** is kidney and **2** is liver using E-Cadherin

# 18. NMR and other Characterization Technique

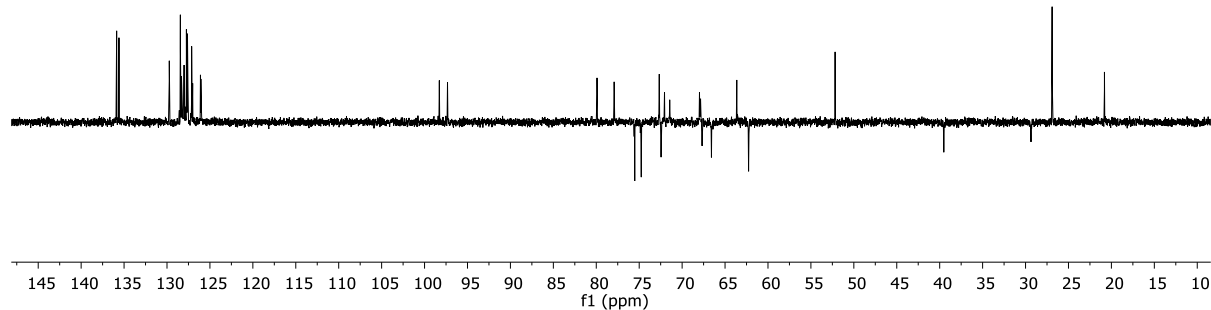
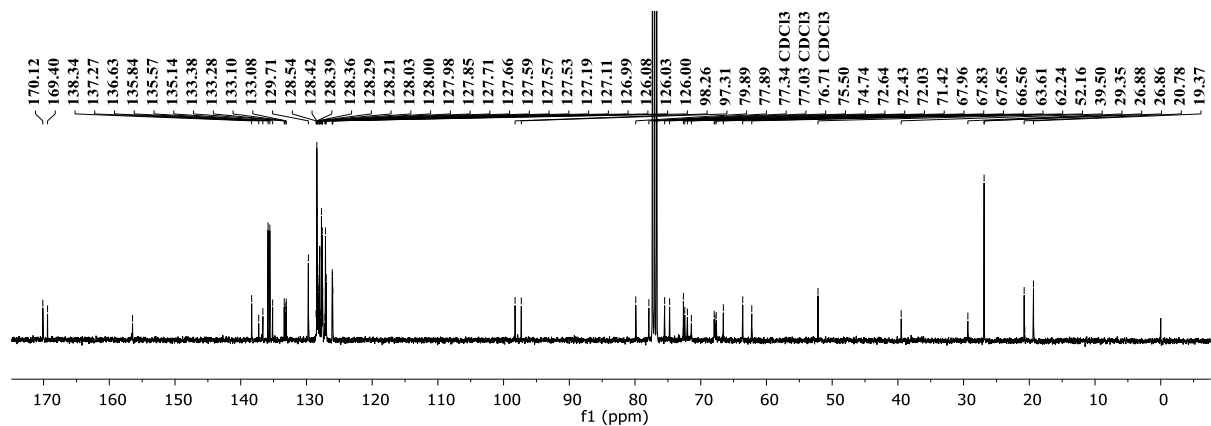
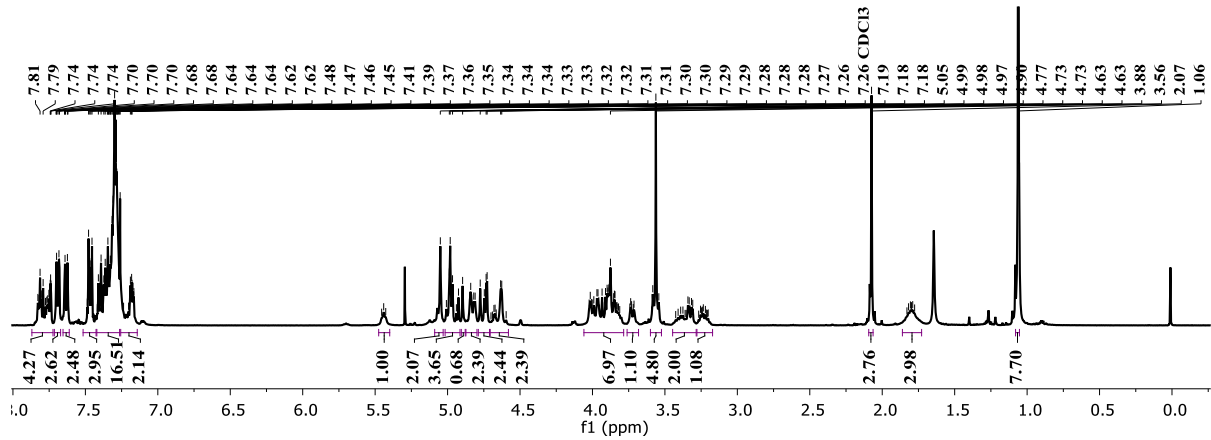
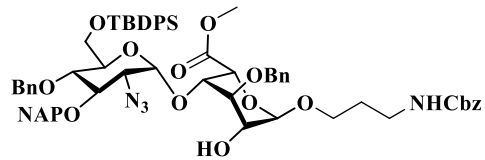


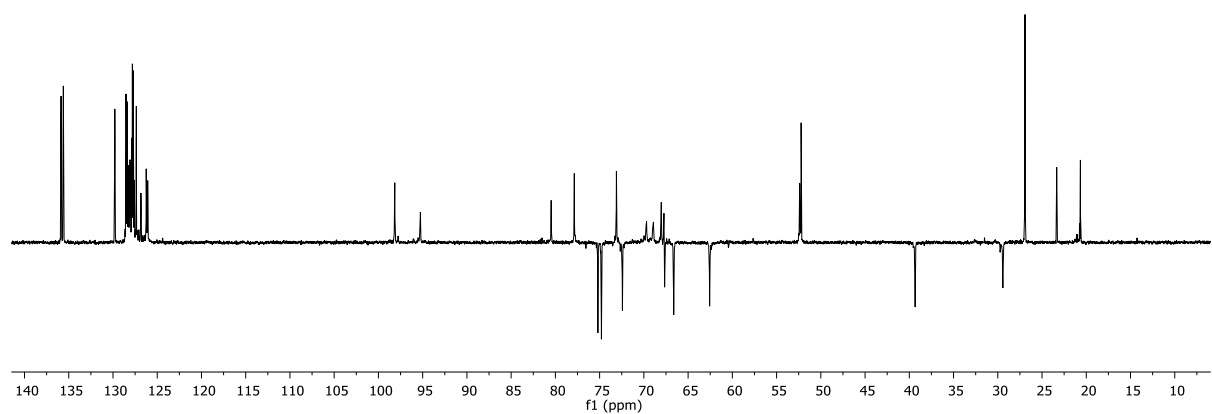
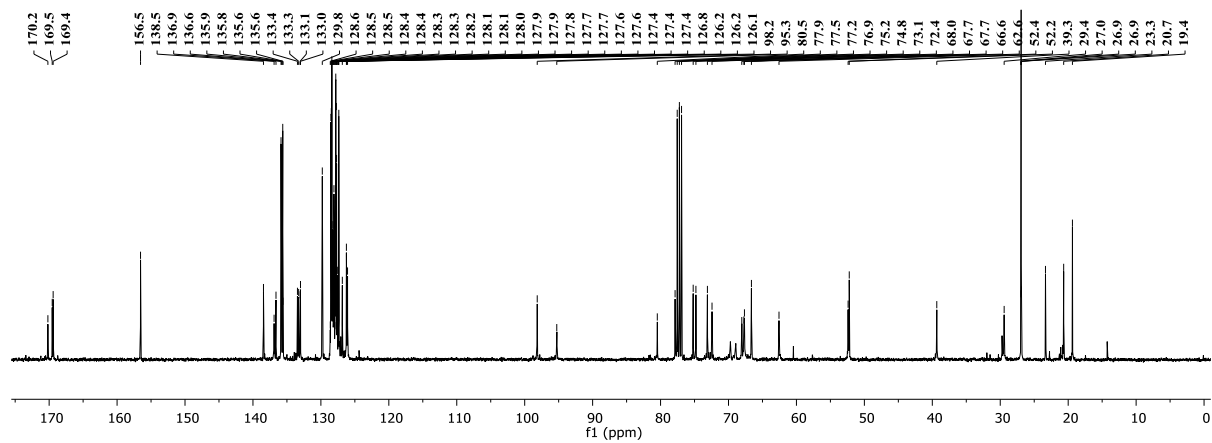
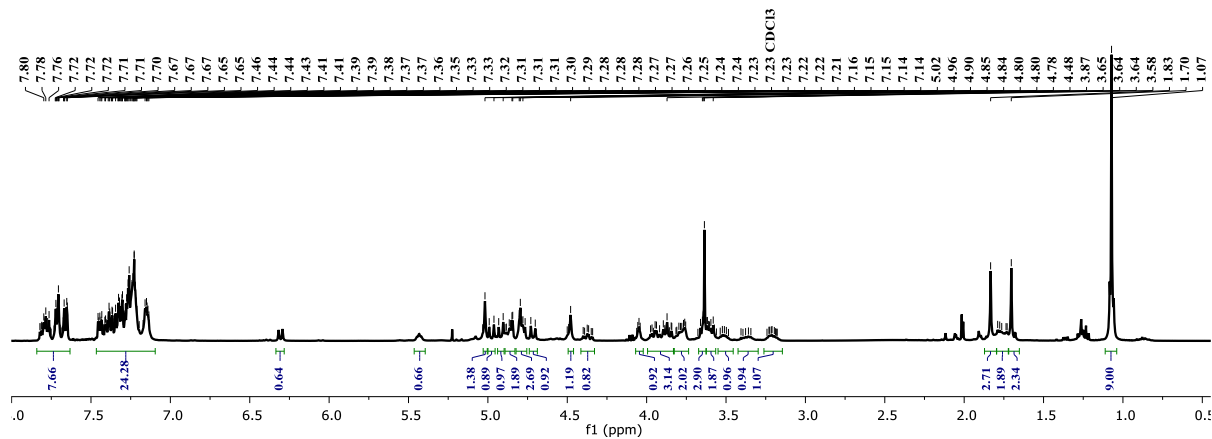
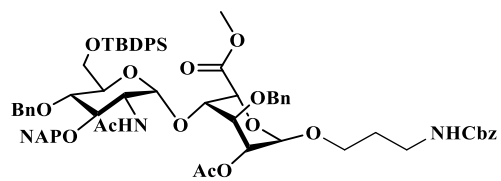


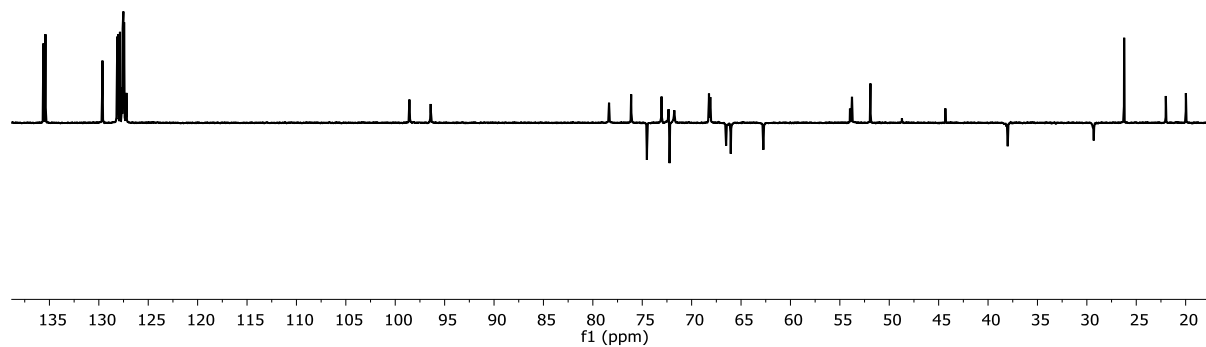
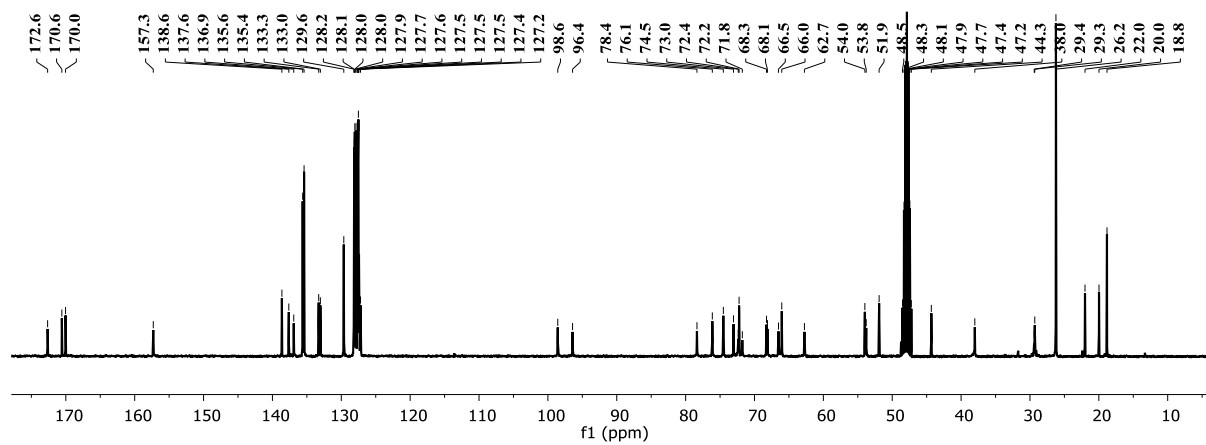
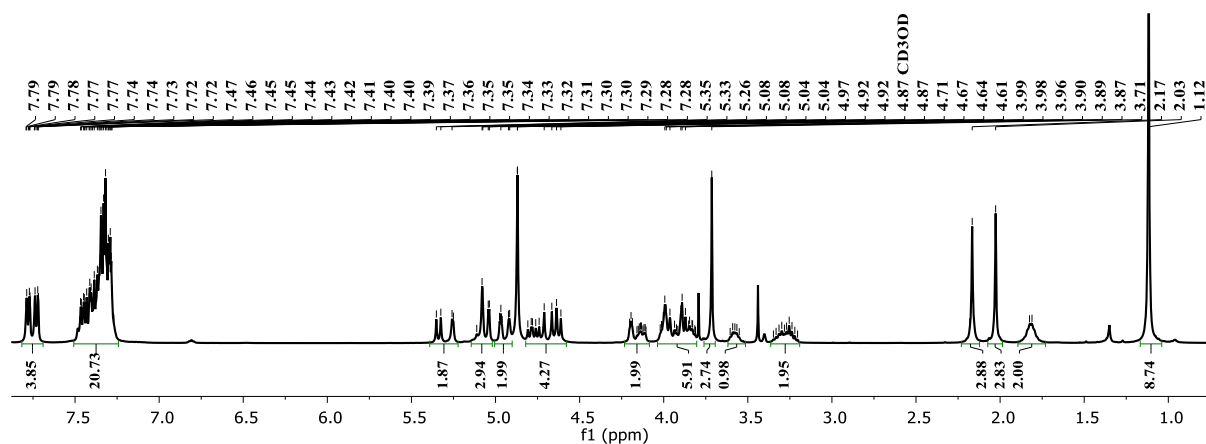
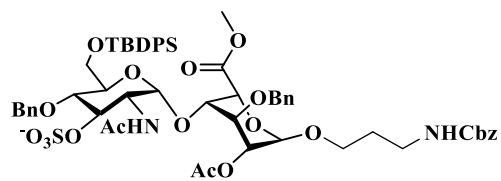


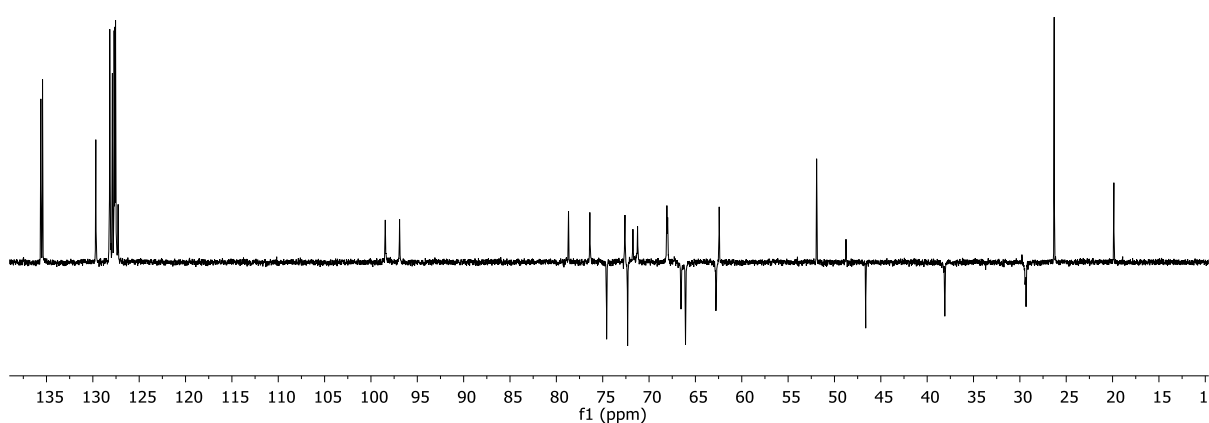
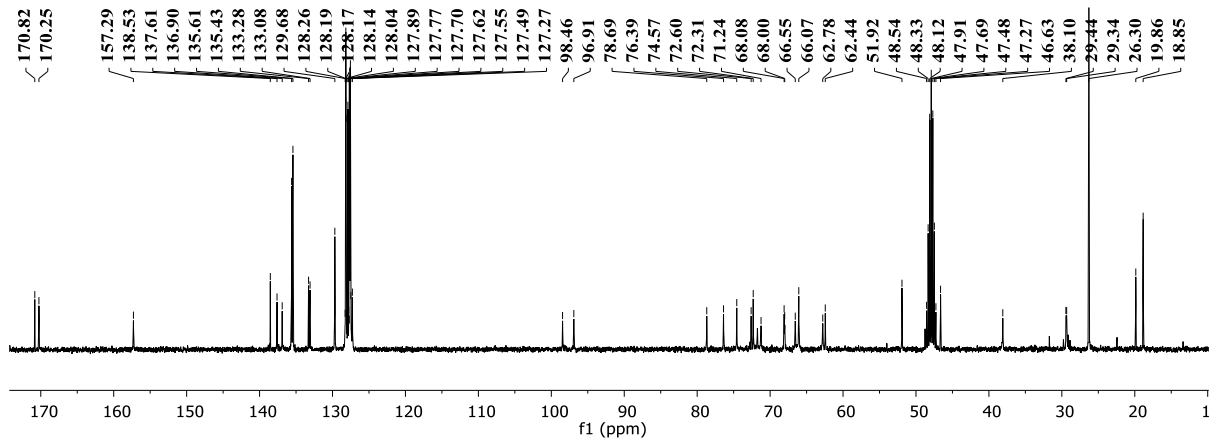
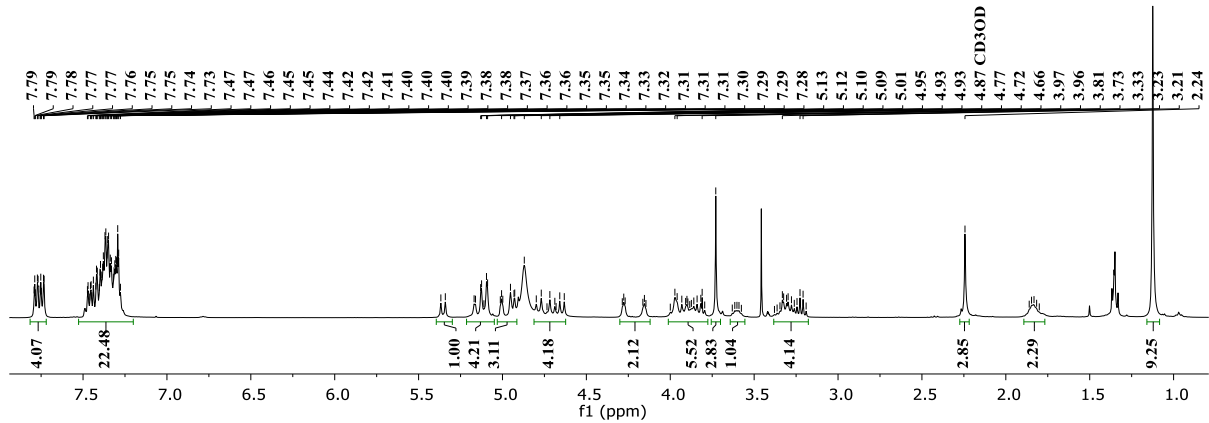
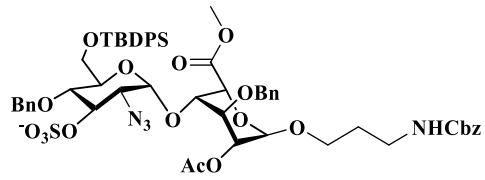


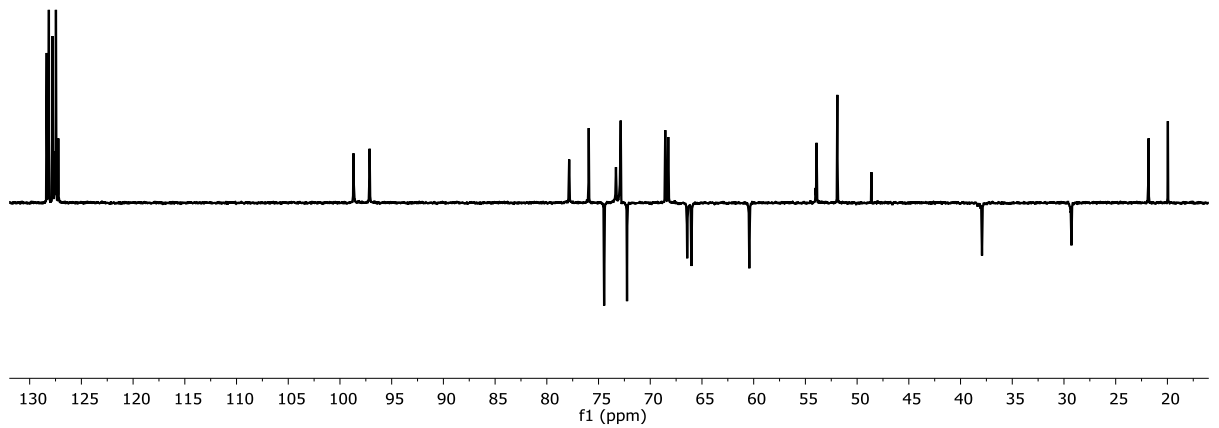
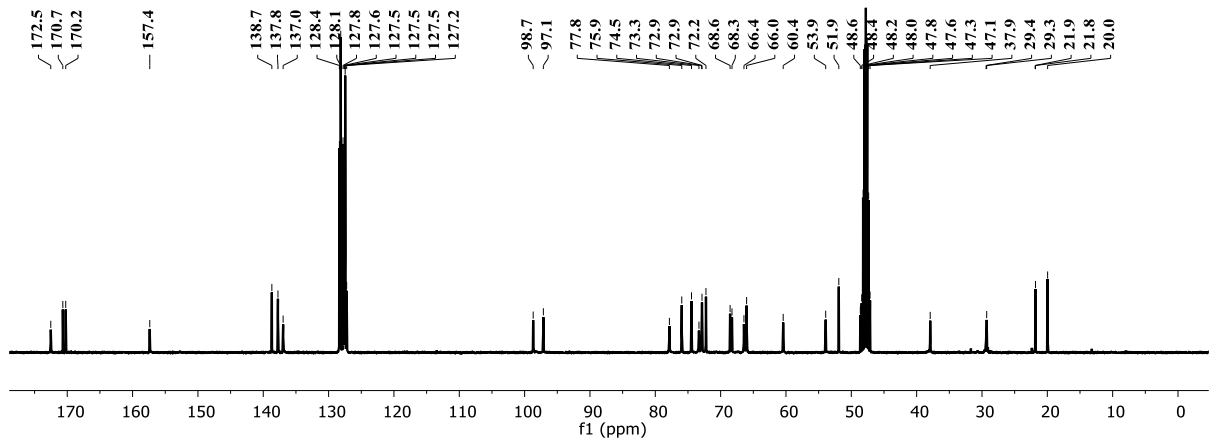
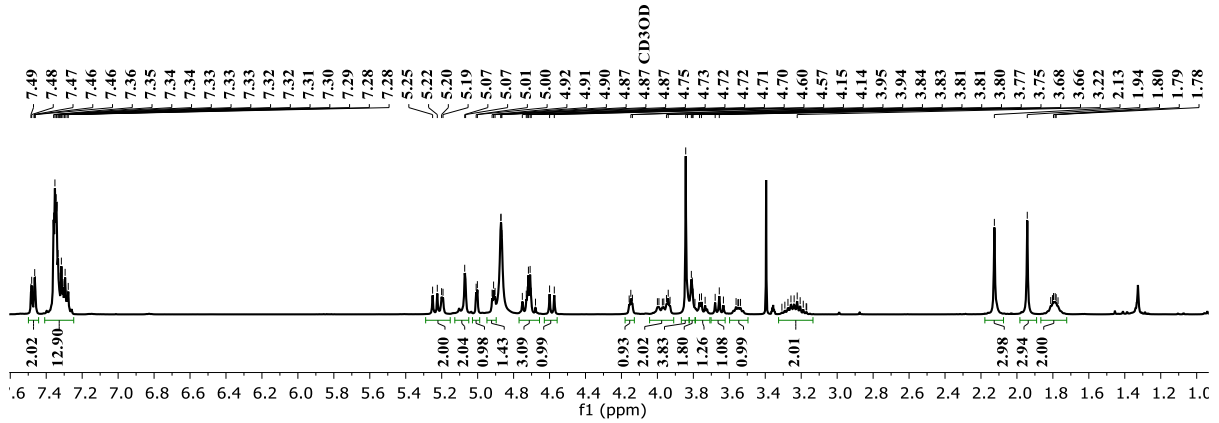
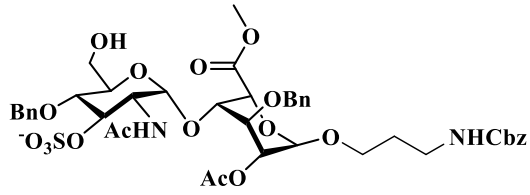


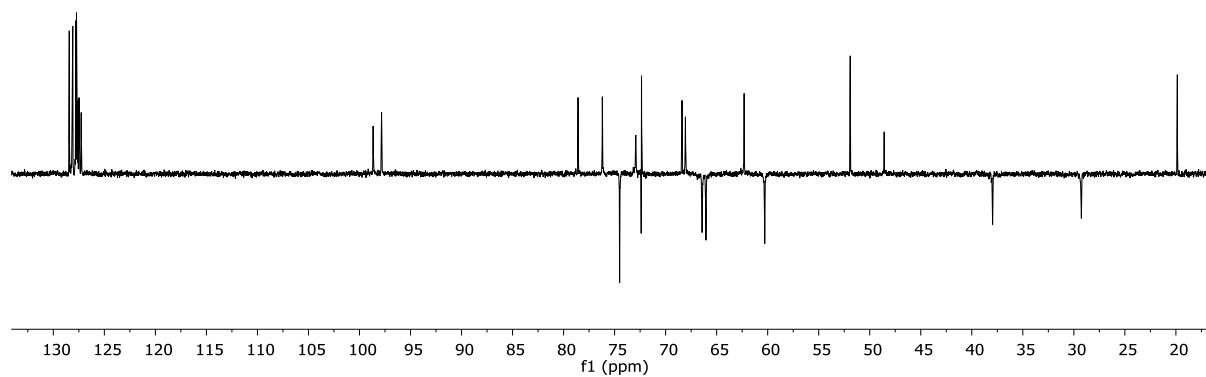
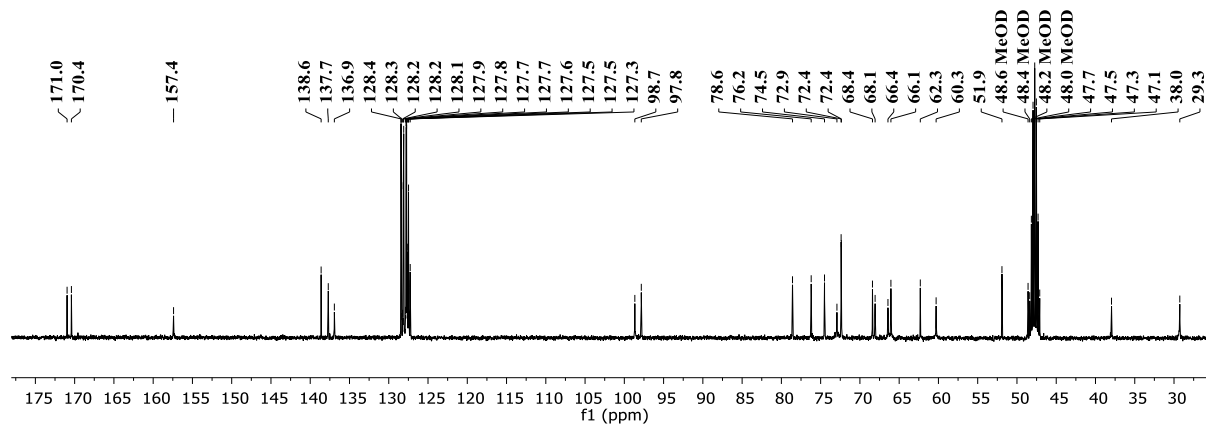
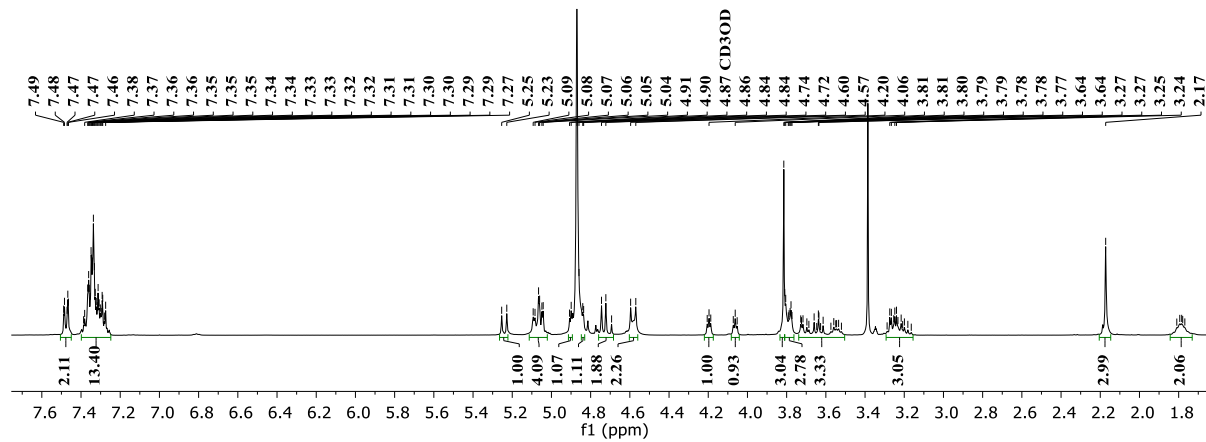
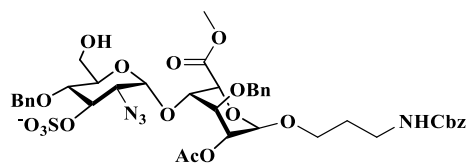


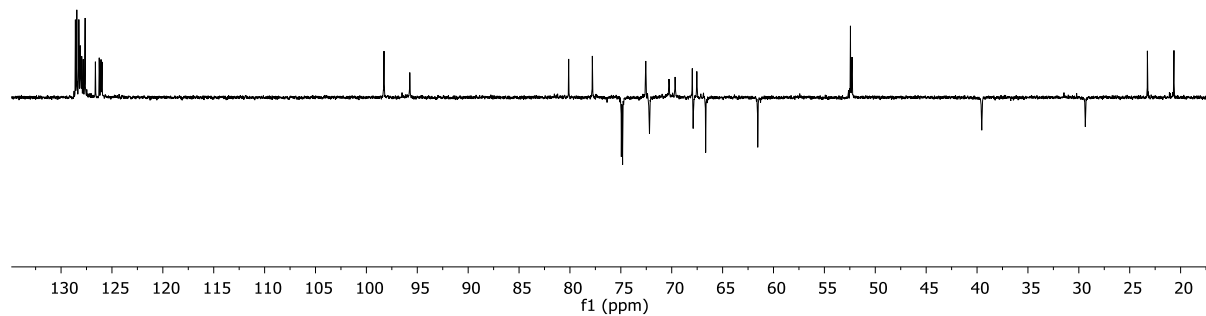
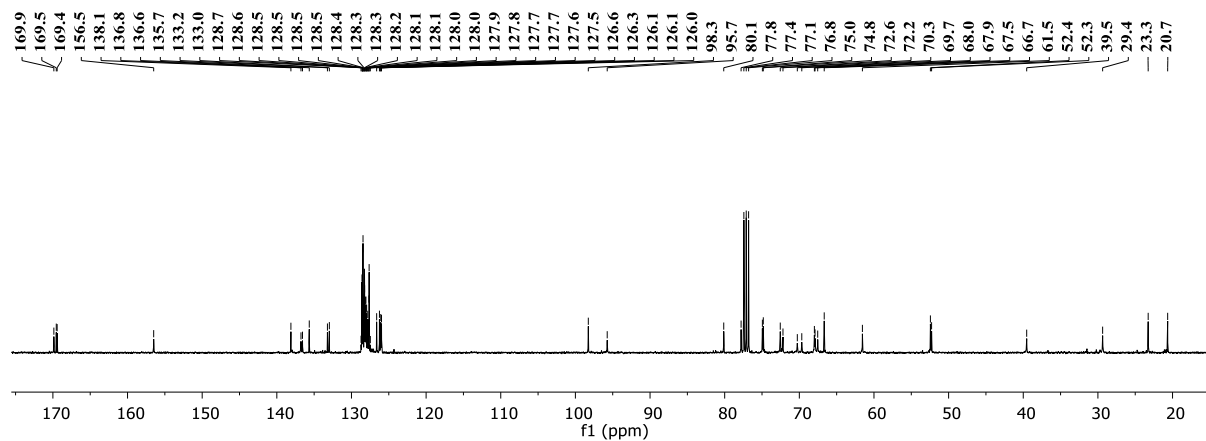
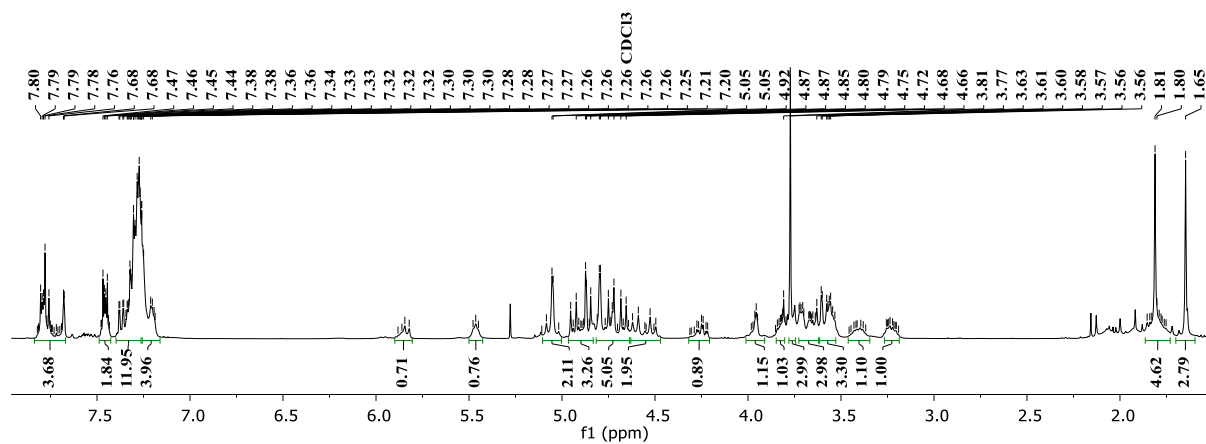
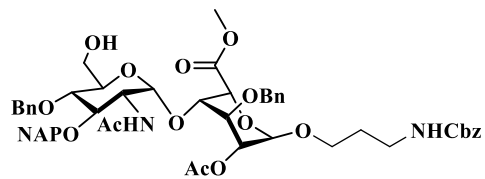


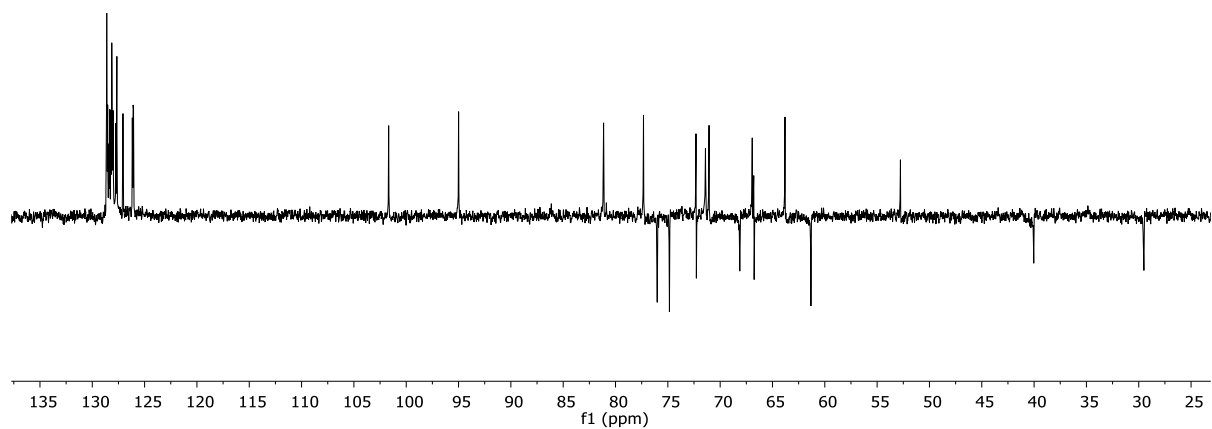
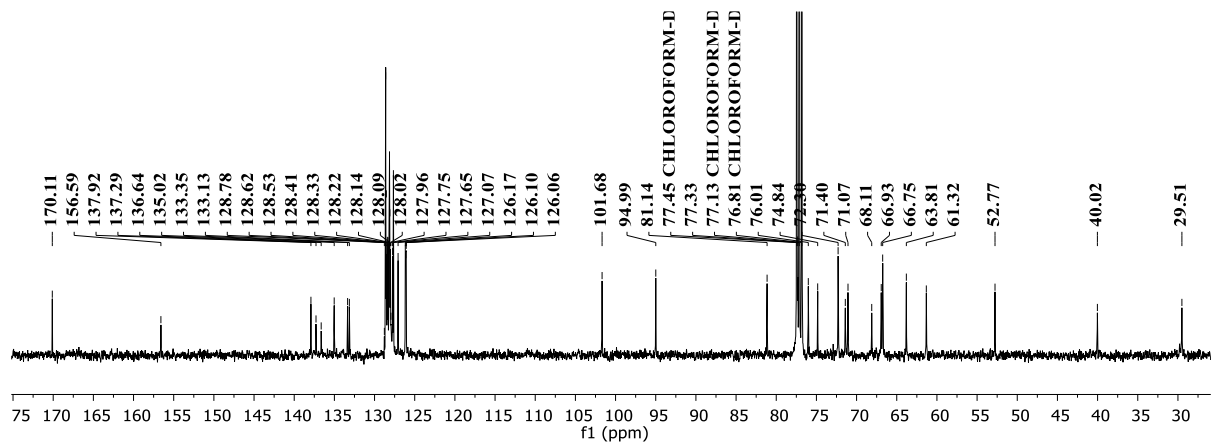
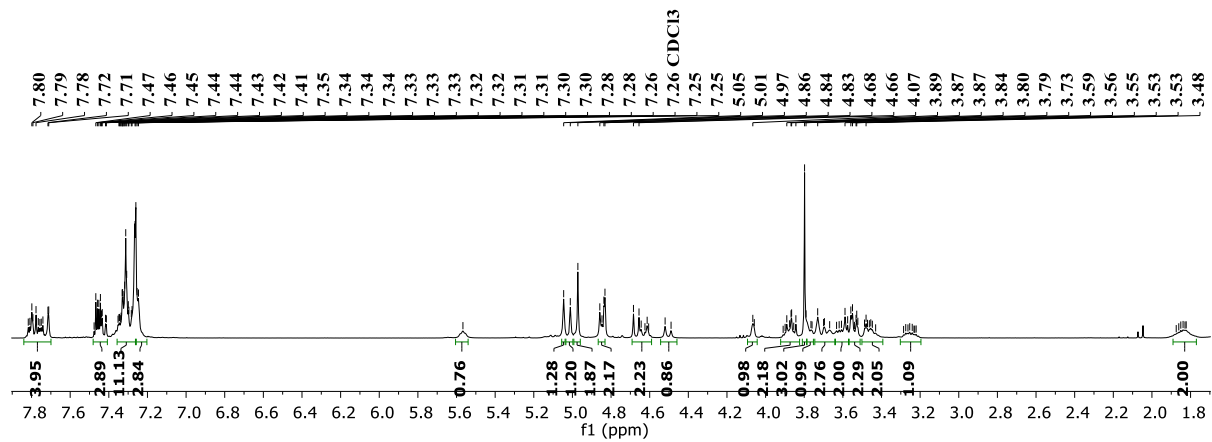
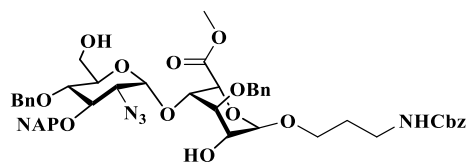




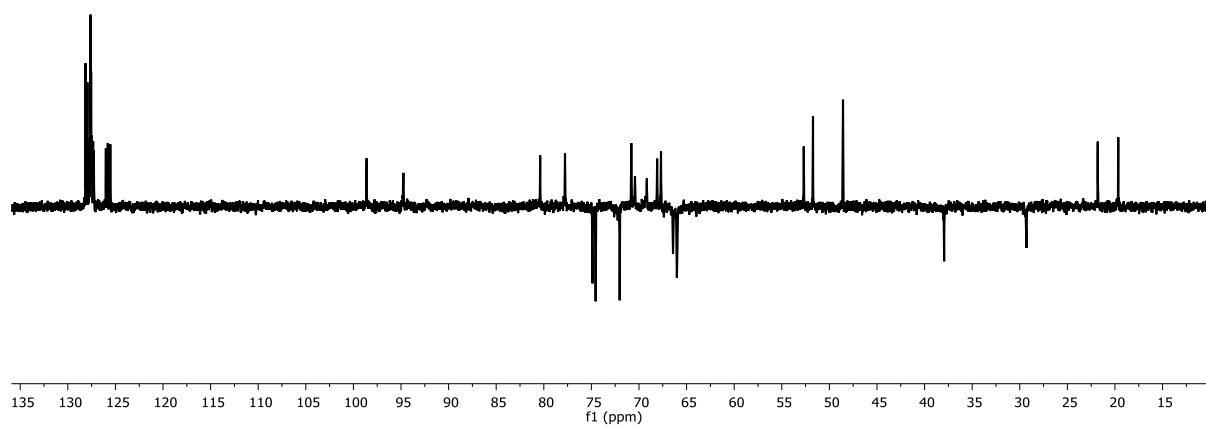
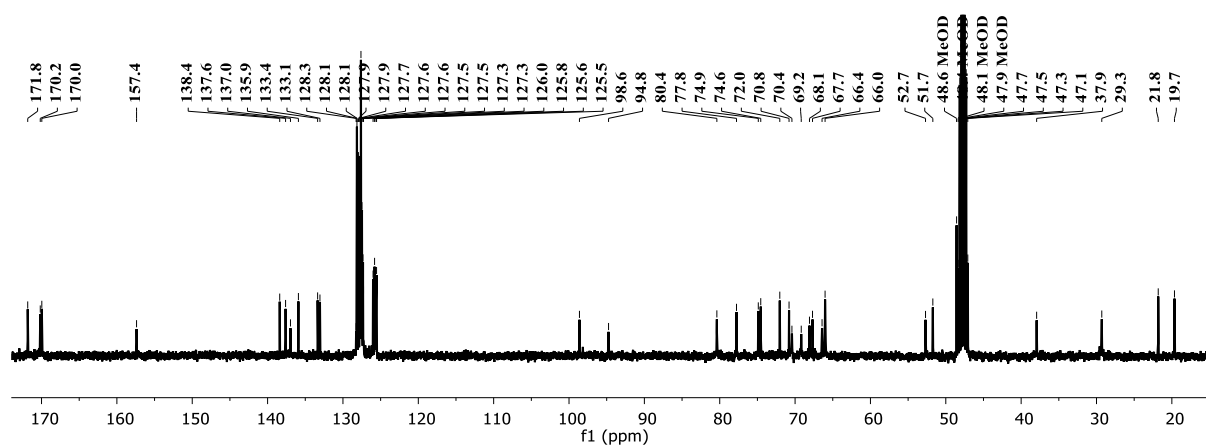
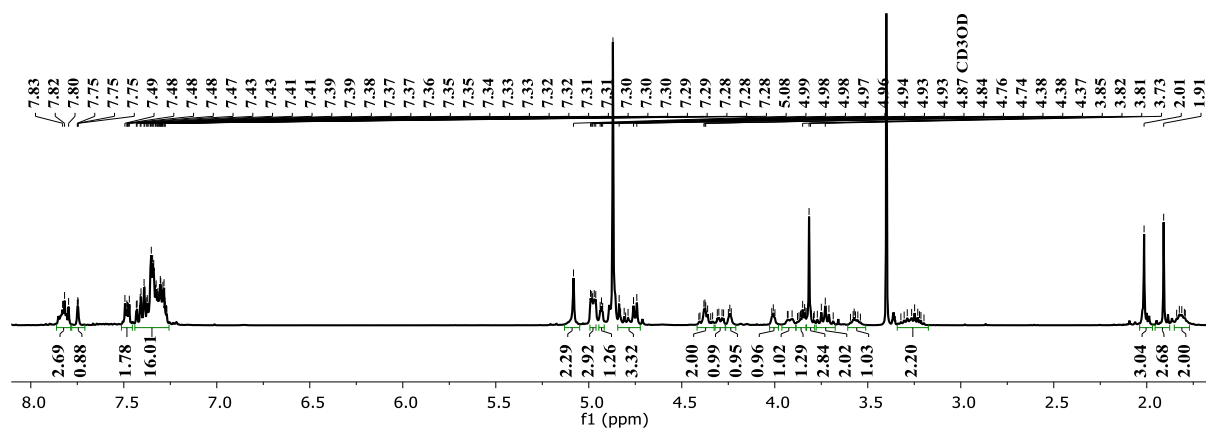
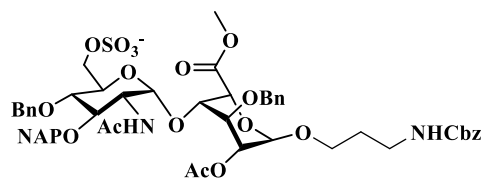


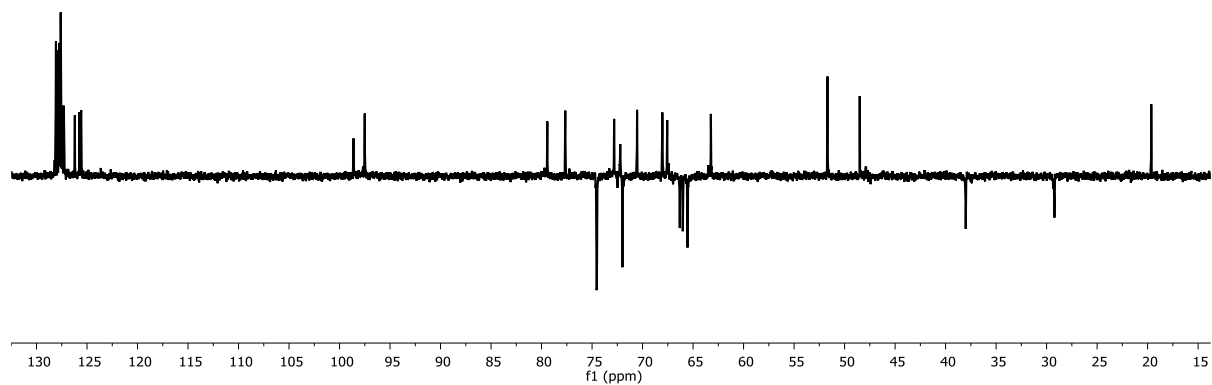
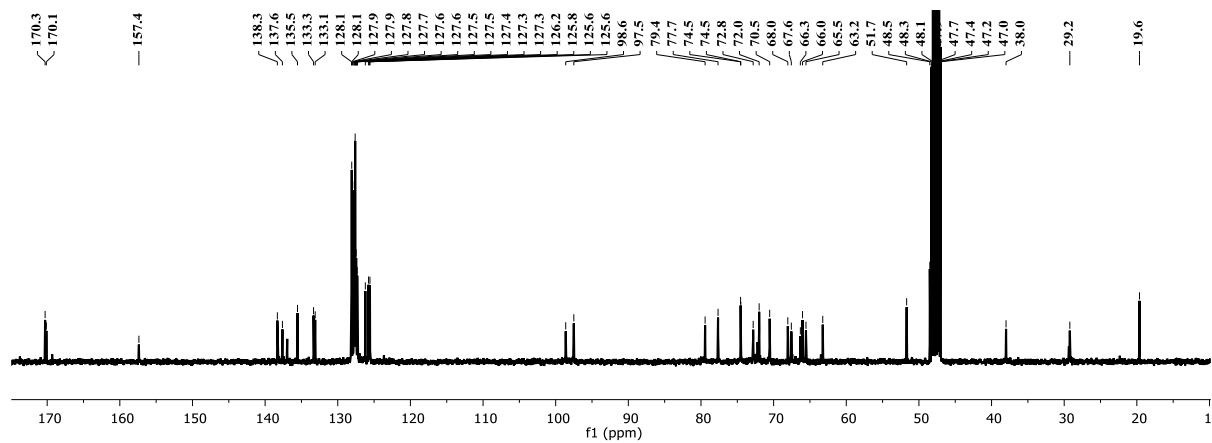
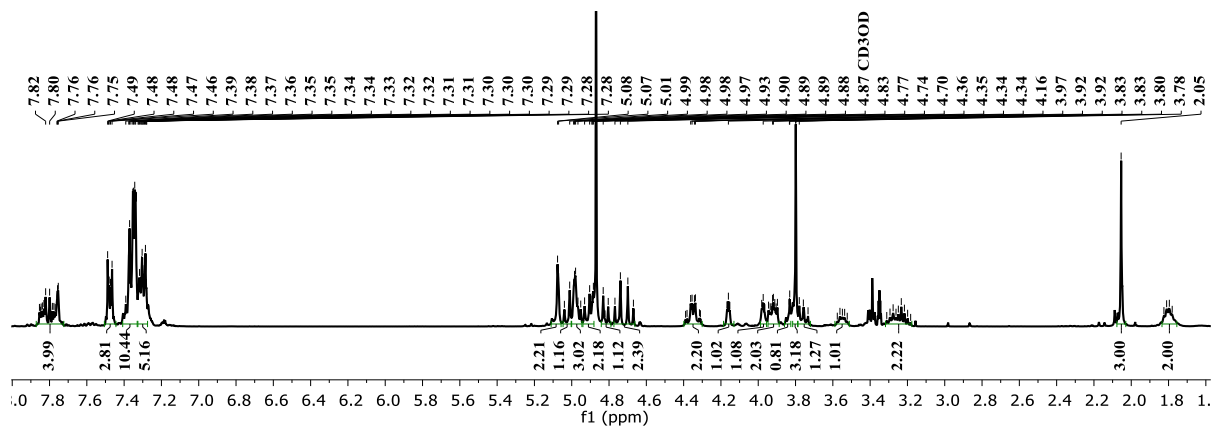
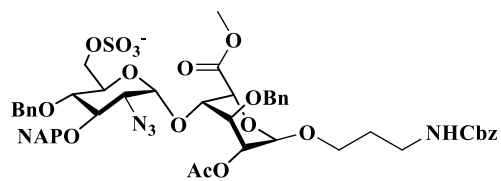


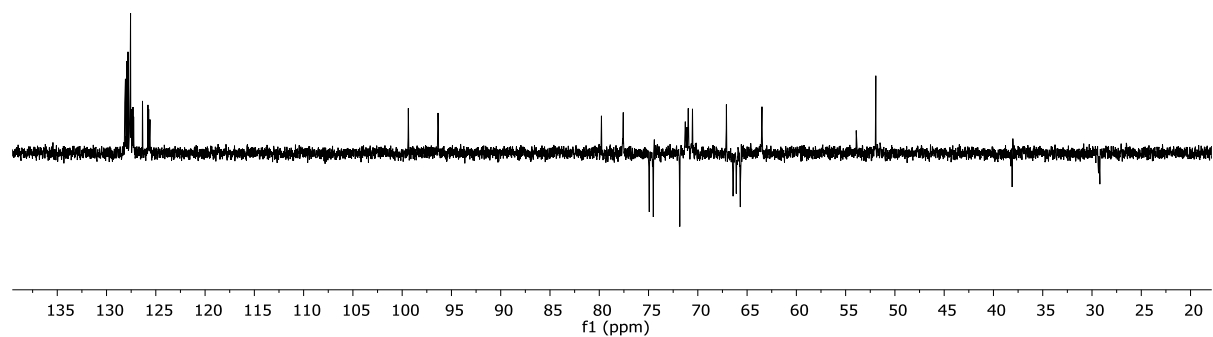
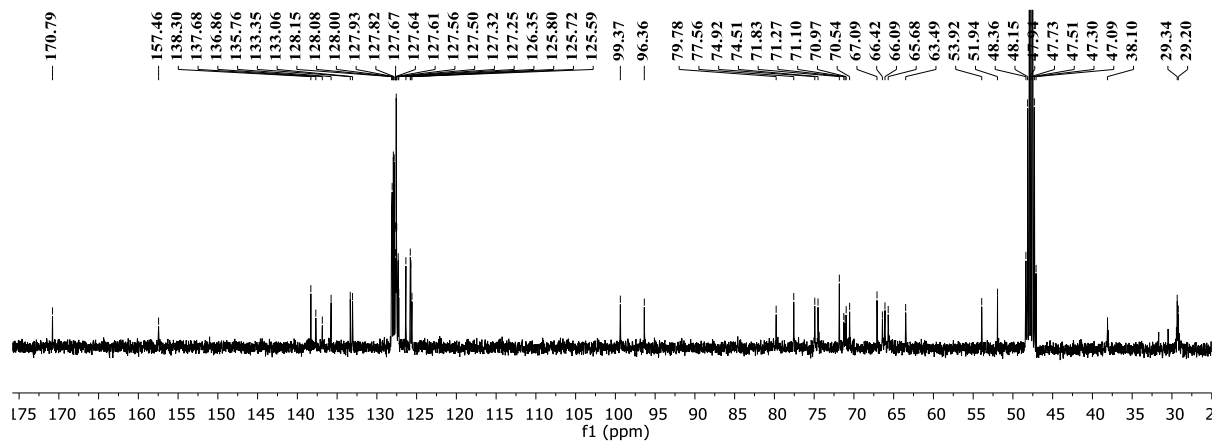
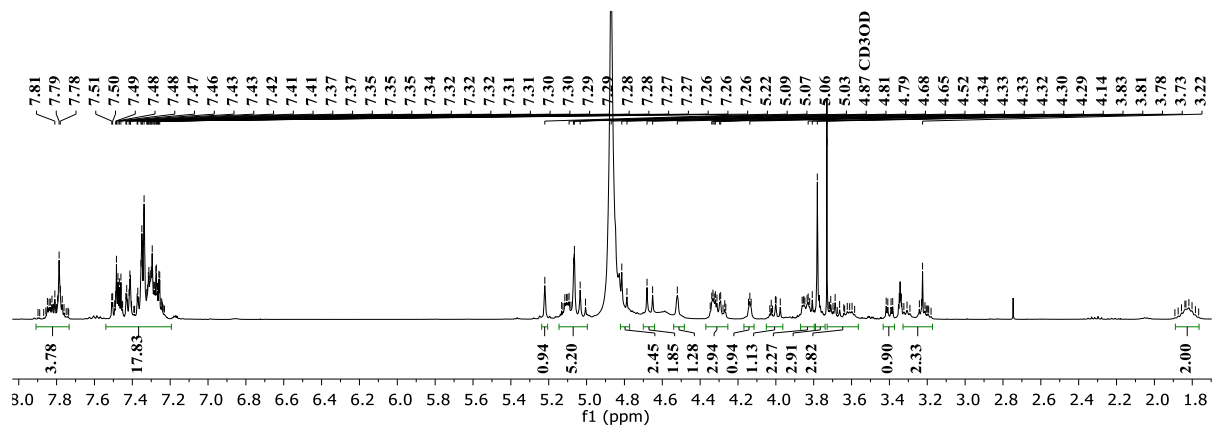
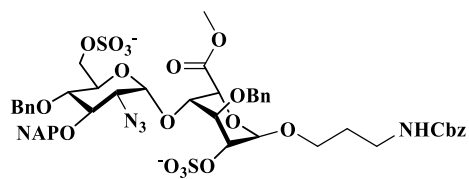


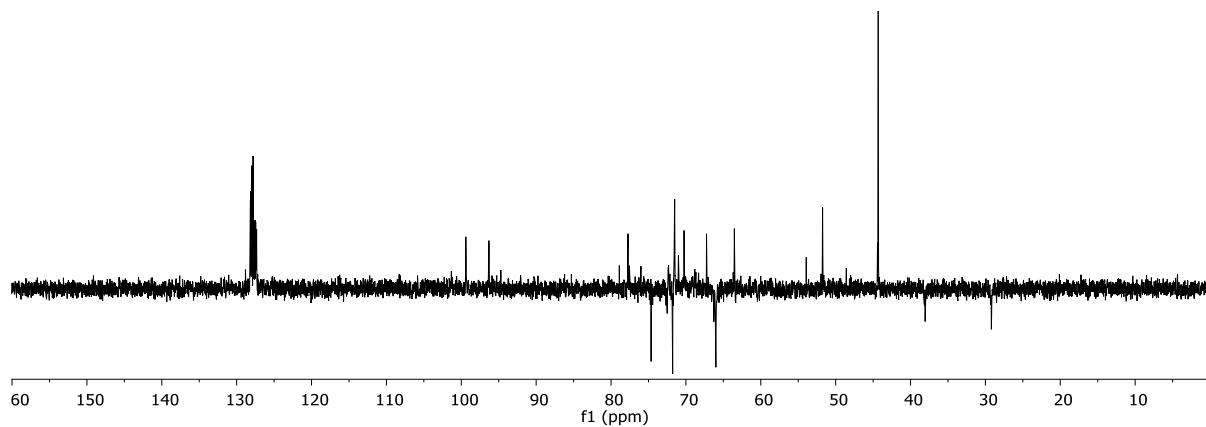
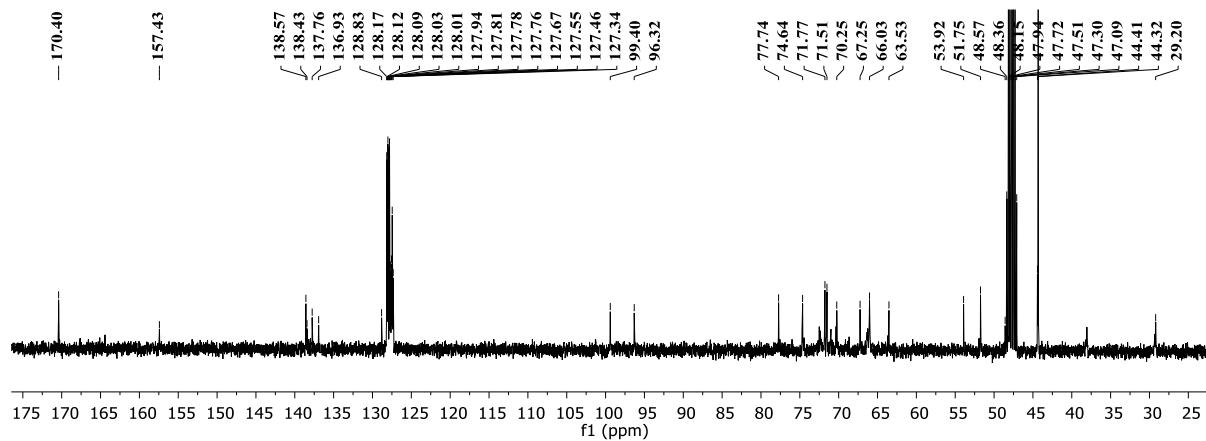
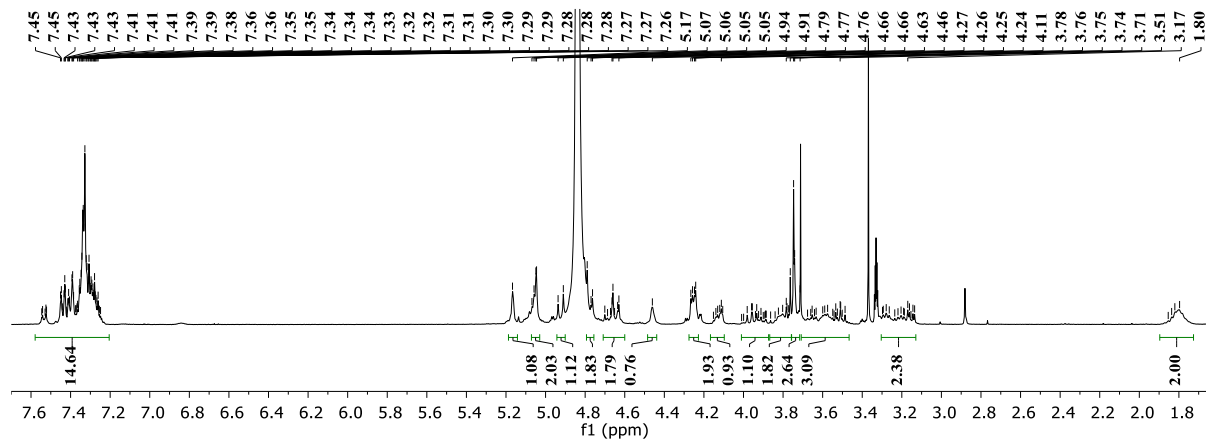
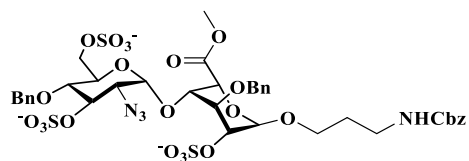


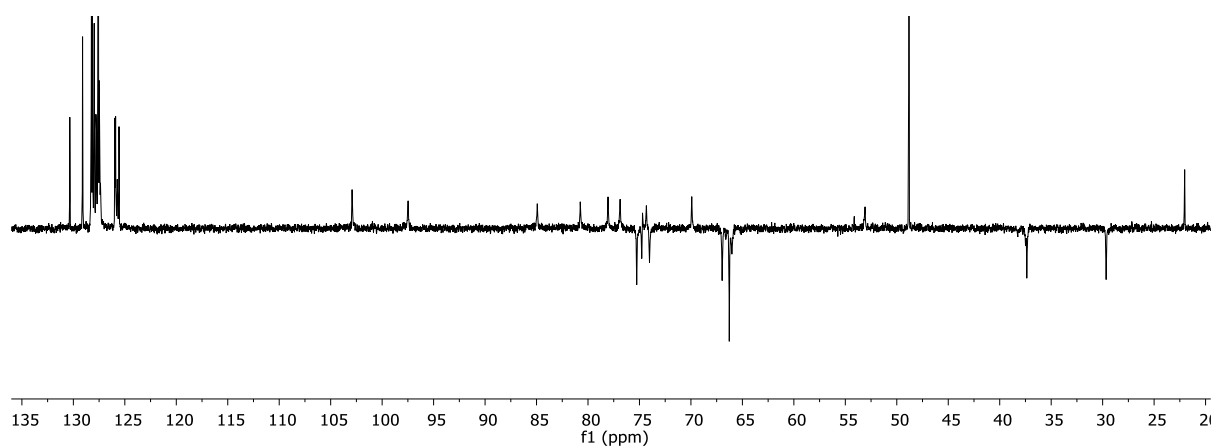
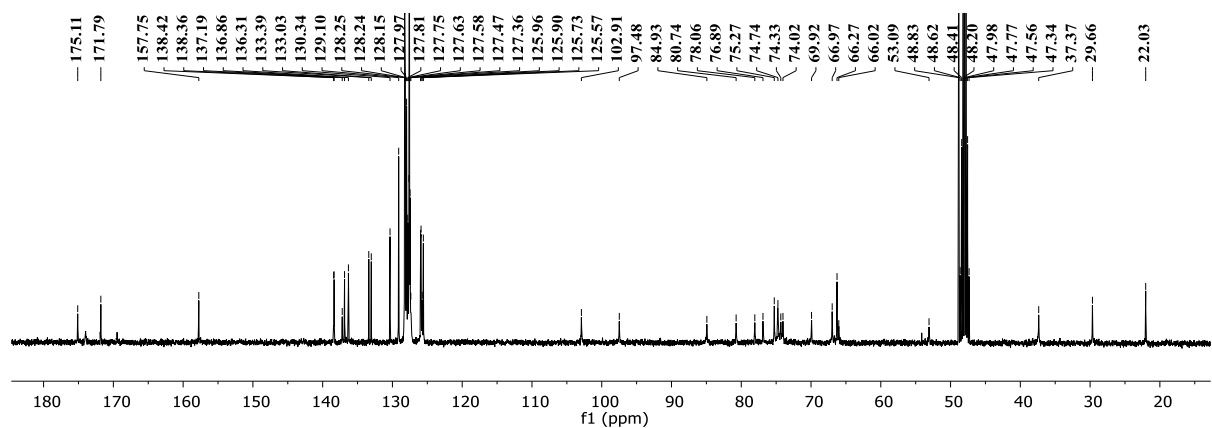
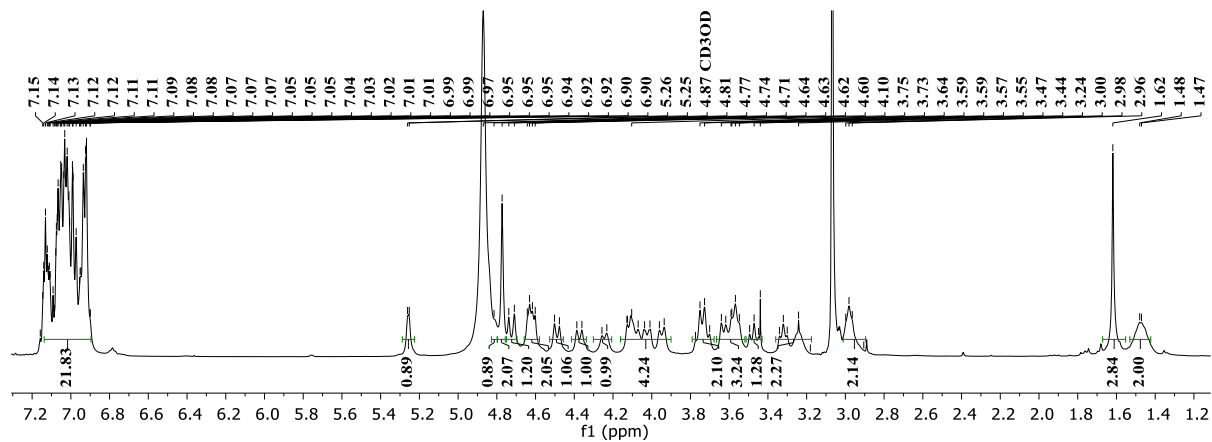
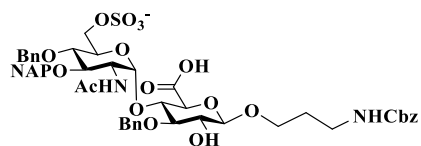


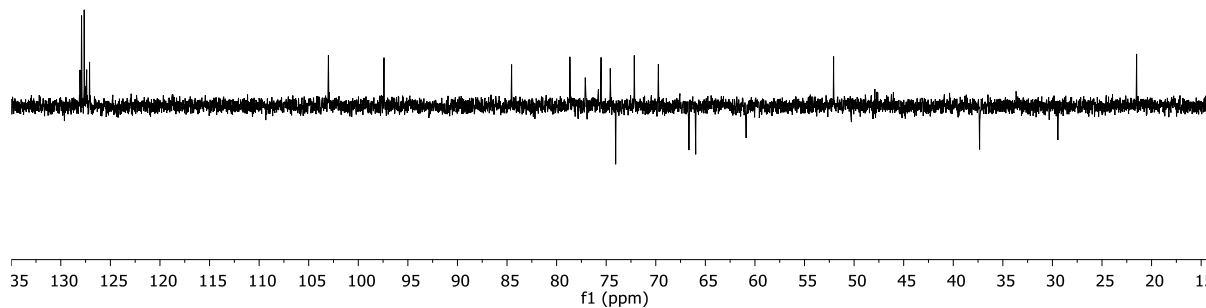
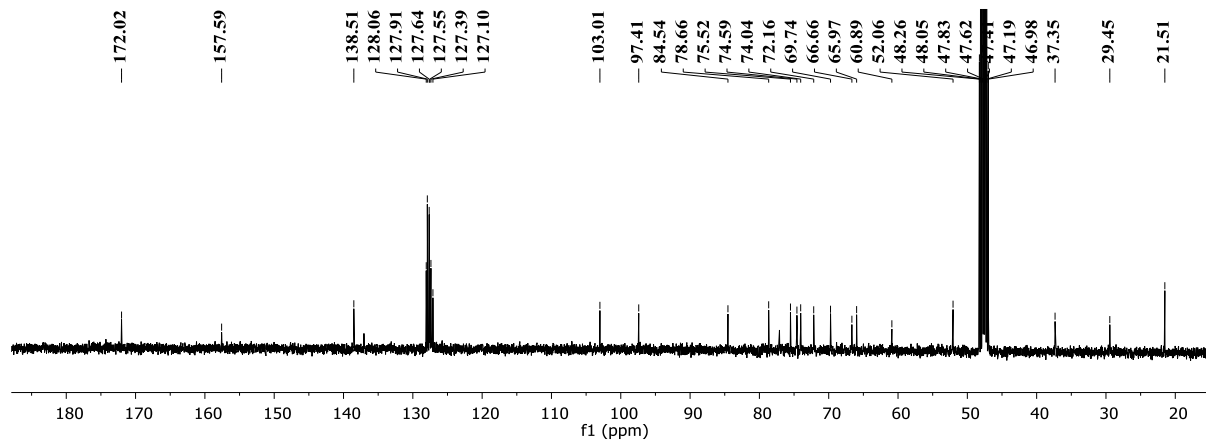
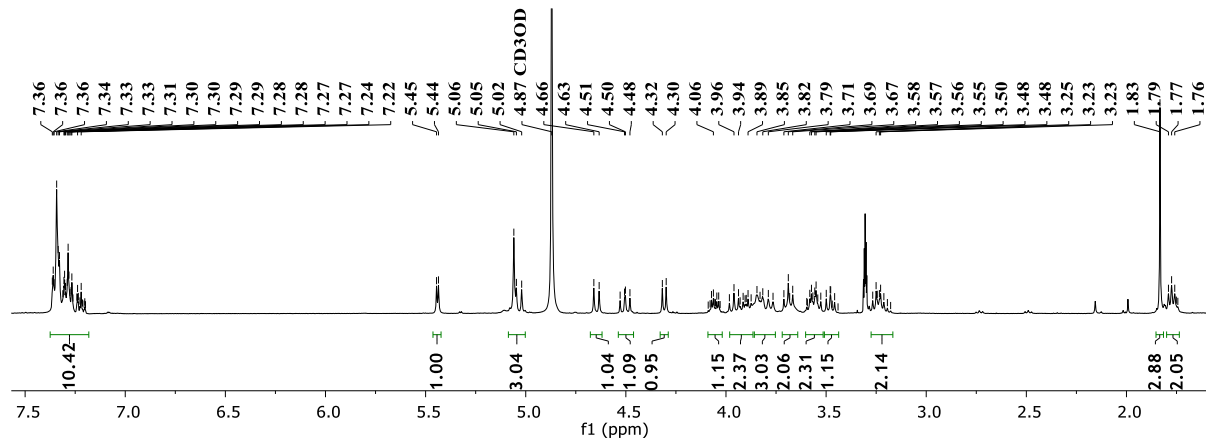
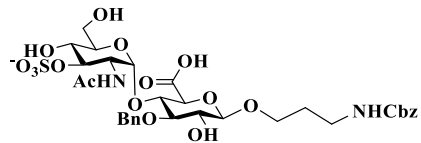


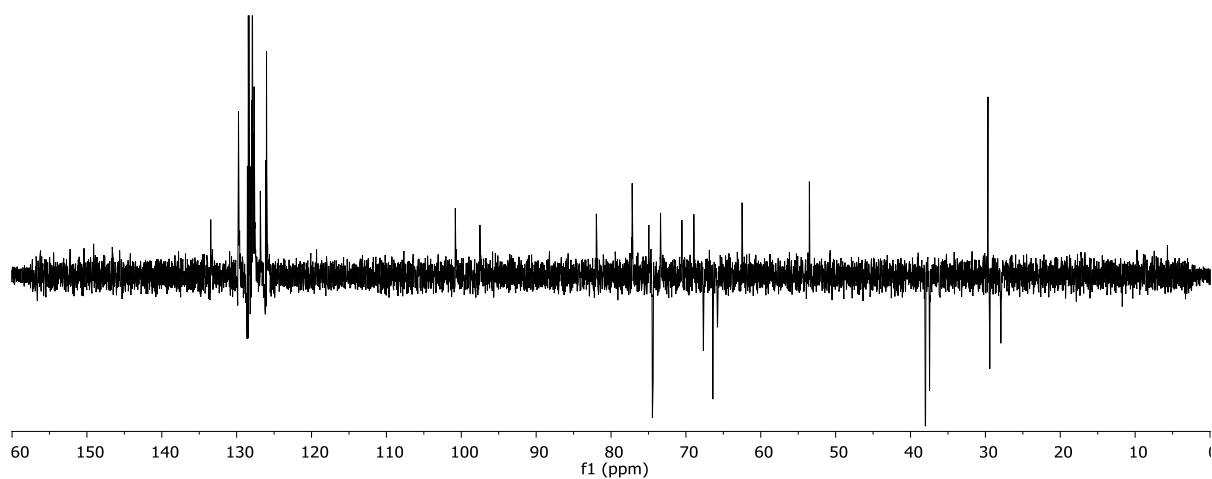
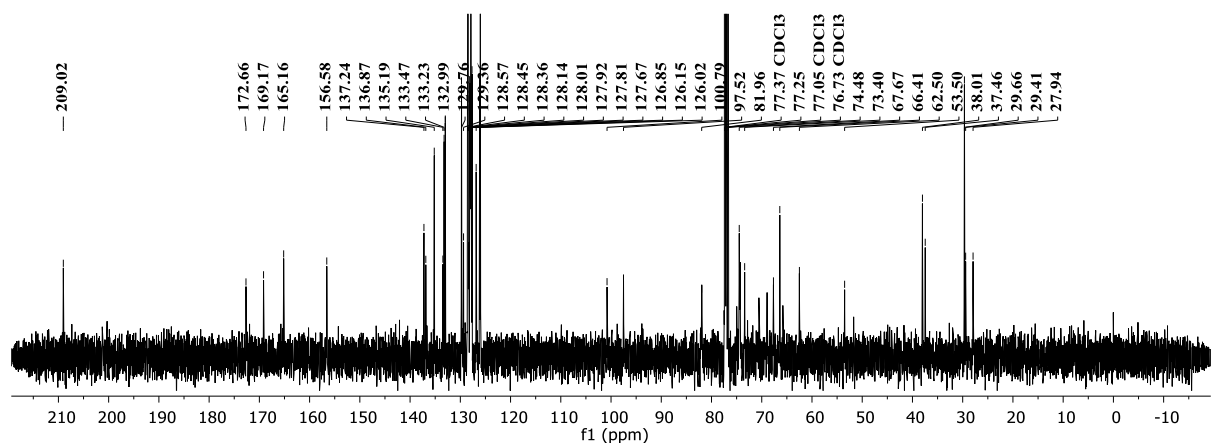
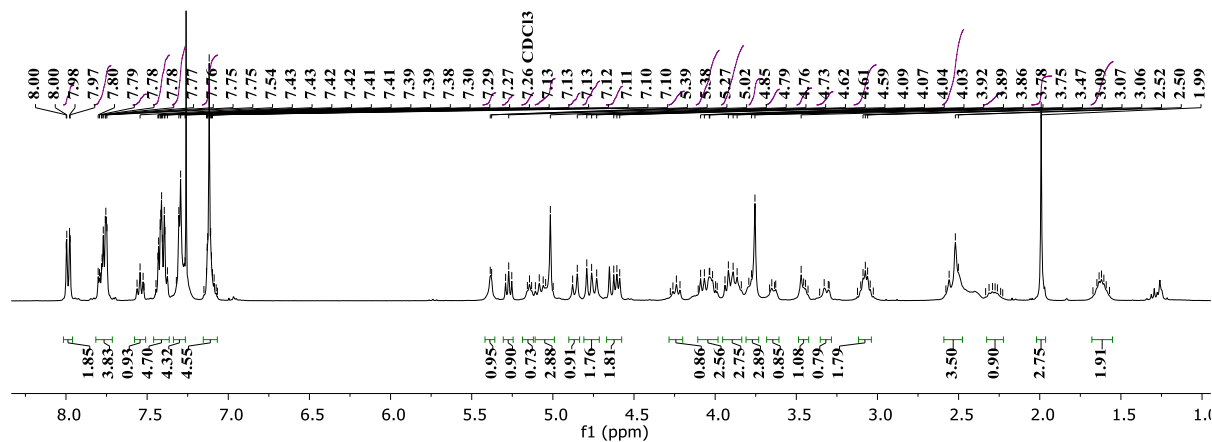
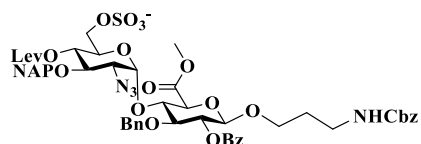


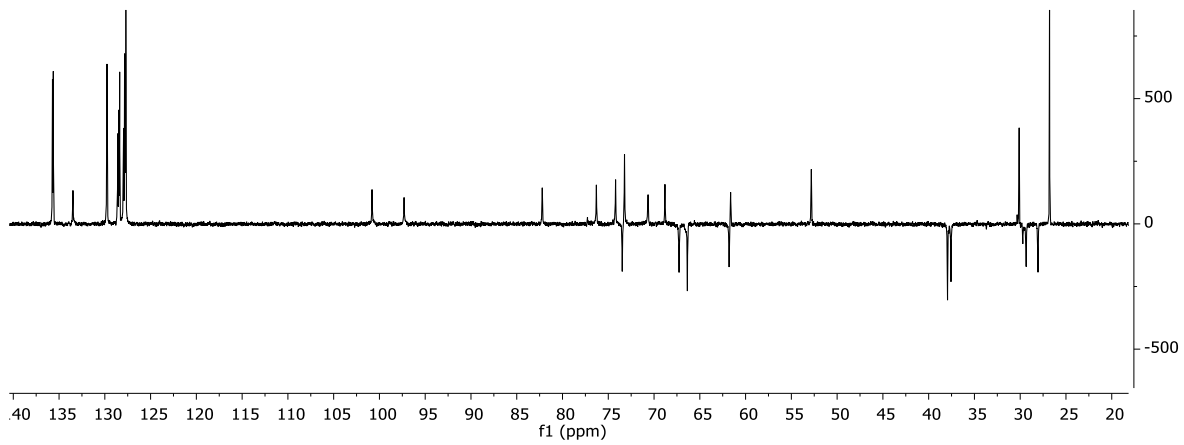
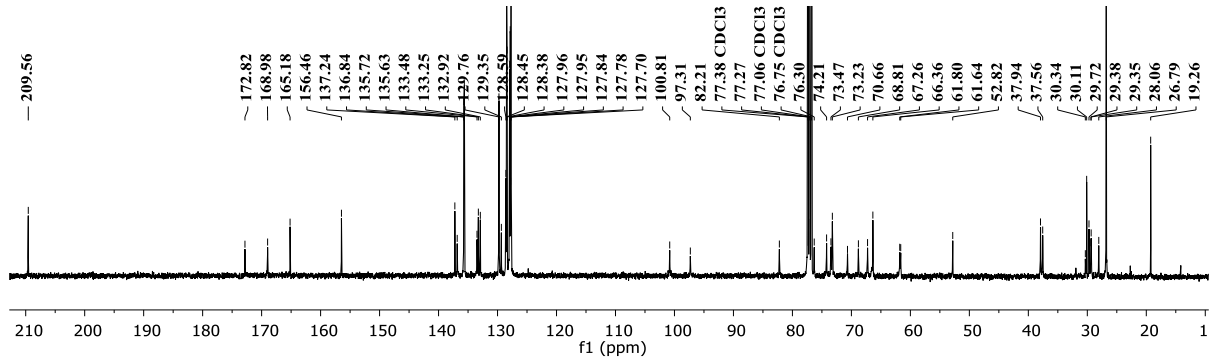
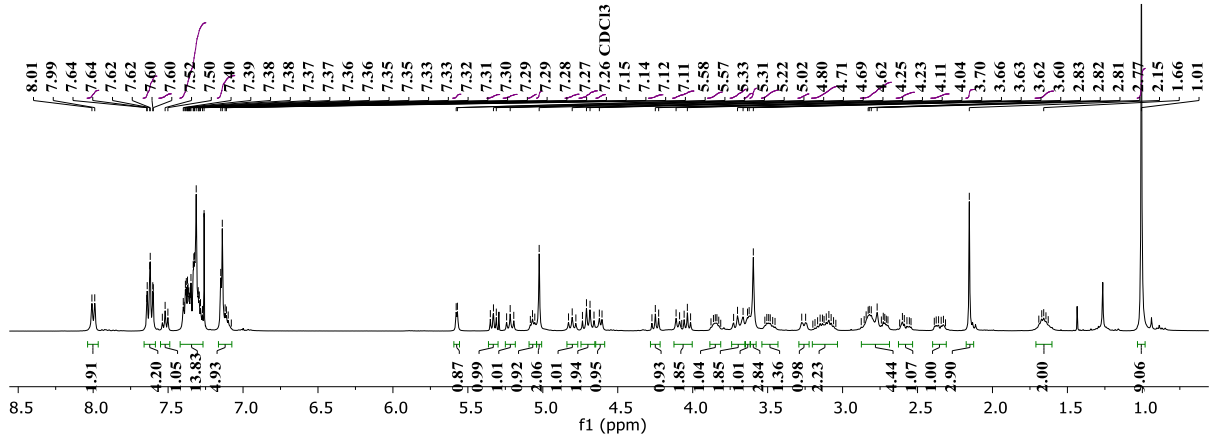
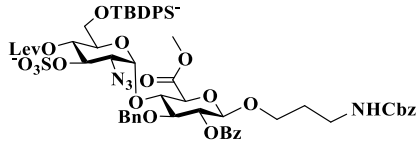




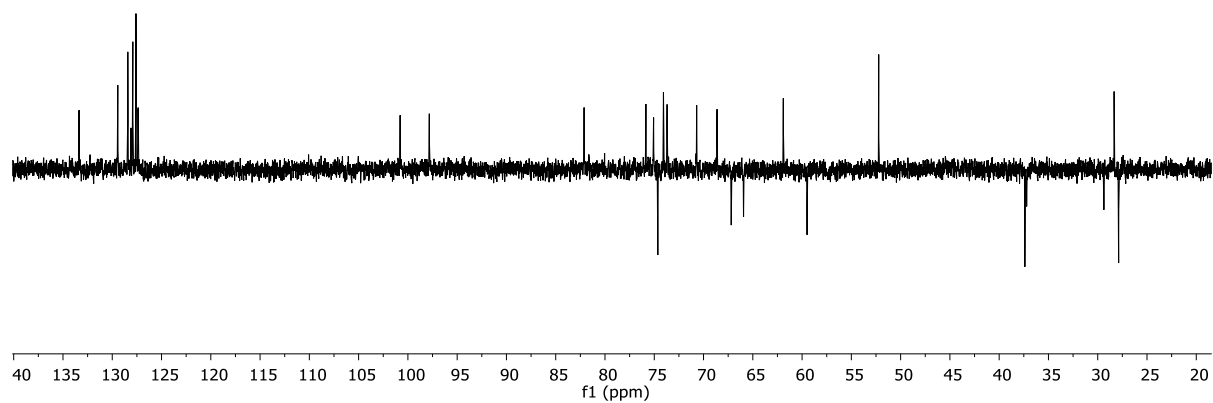
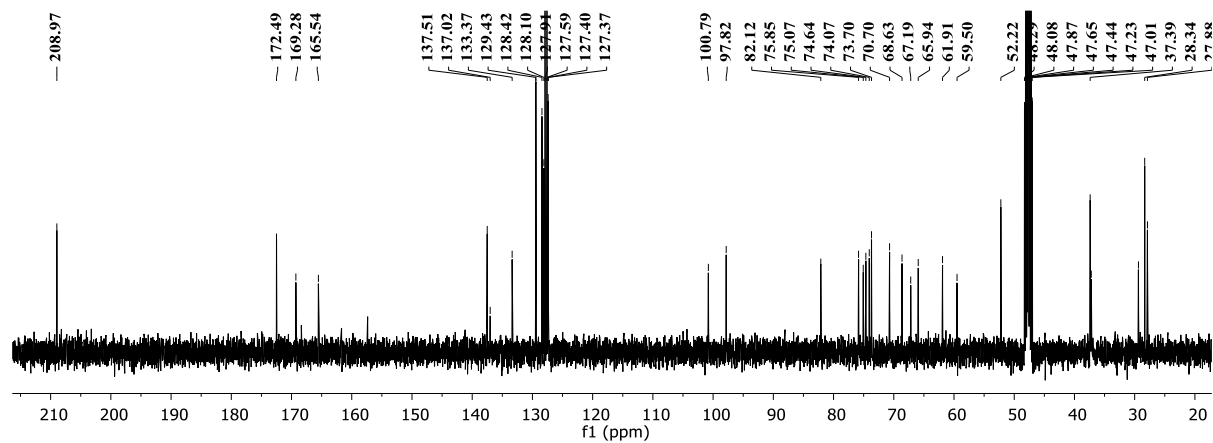
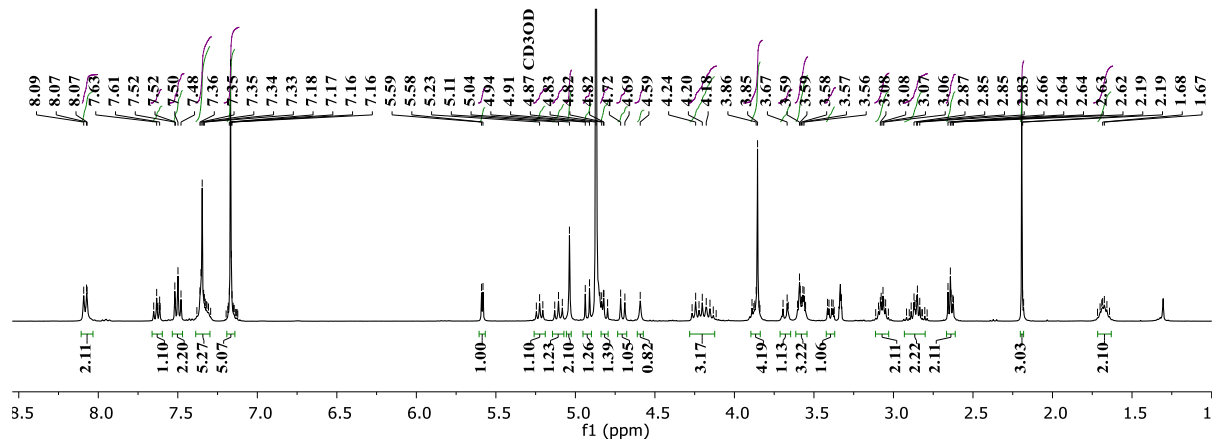
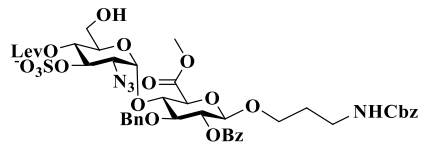


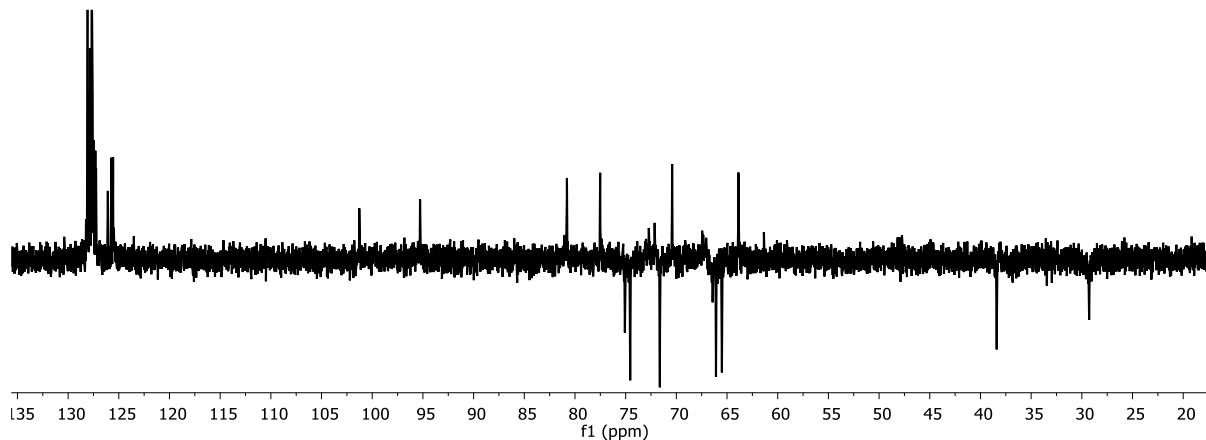
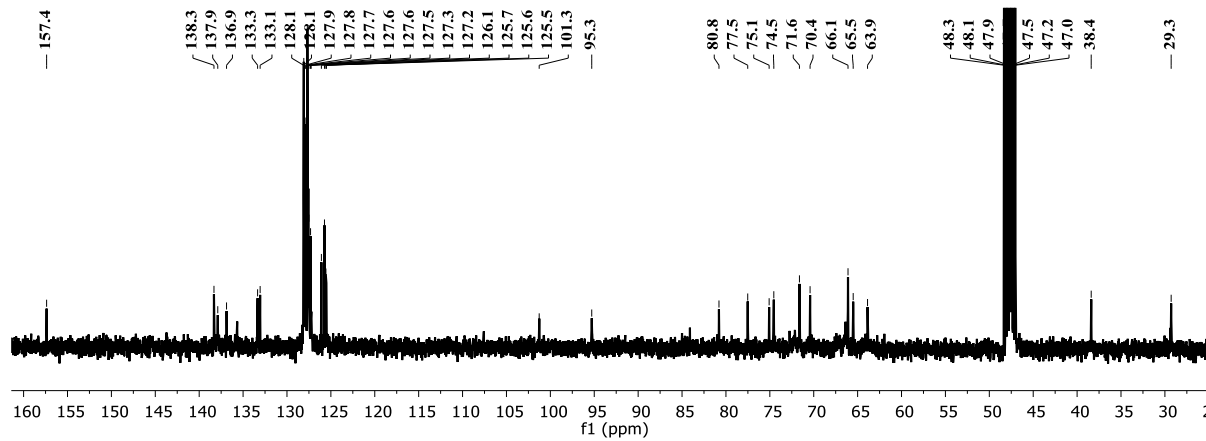
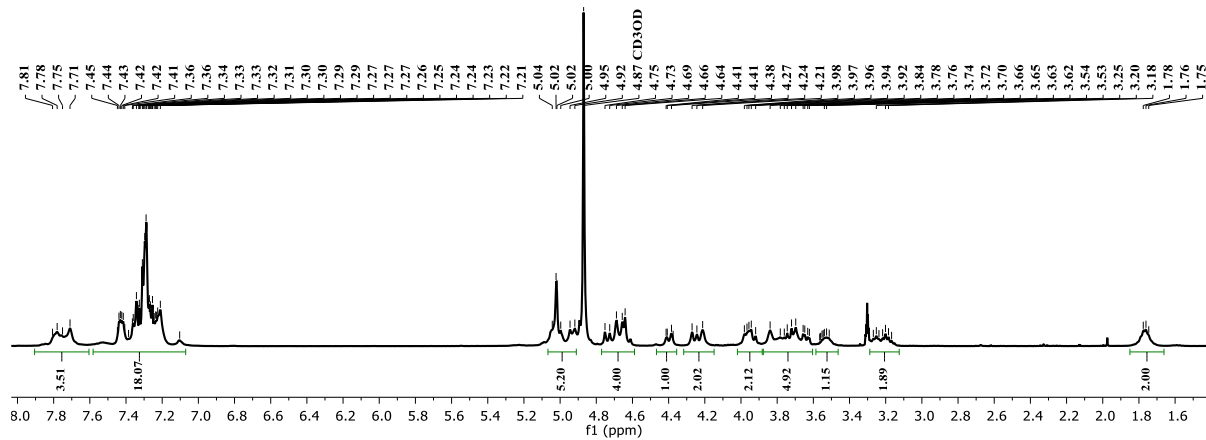
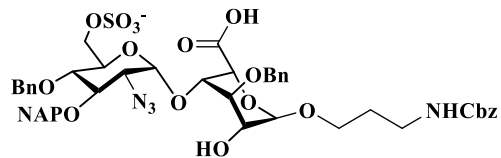


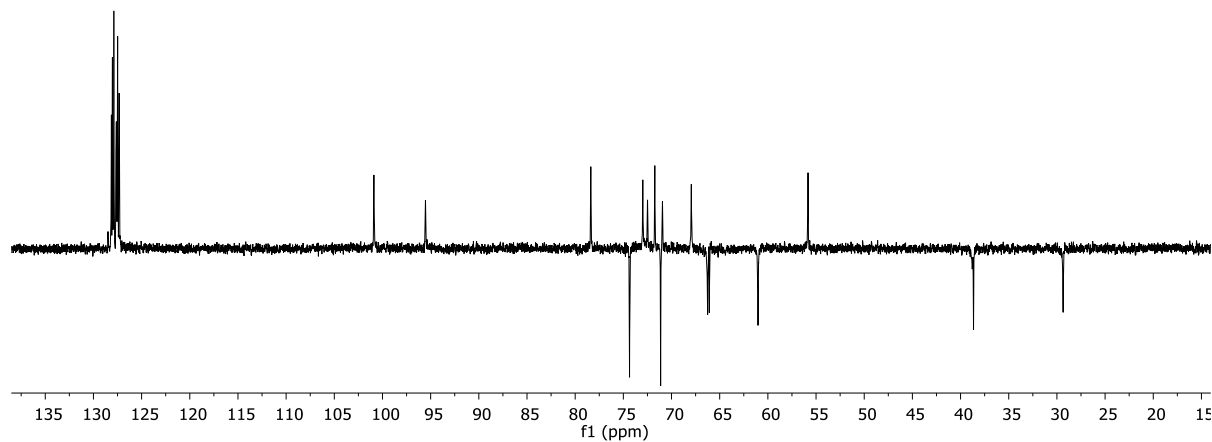
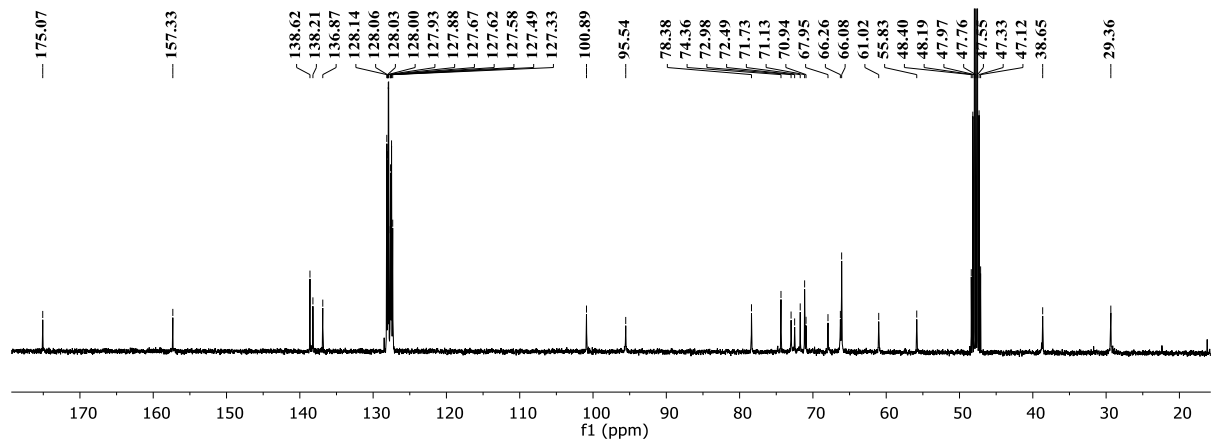
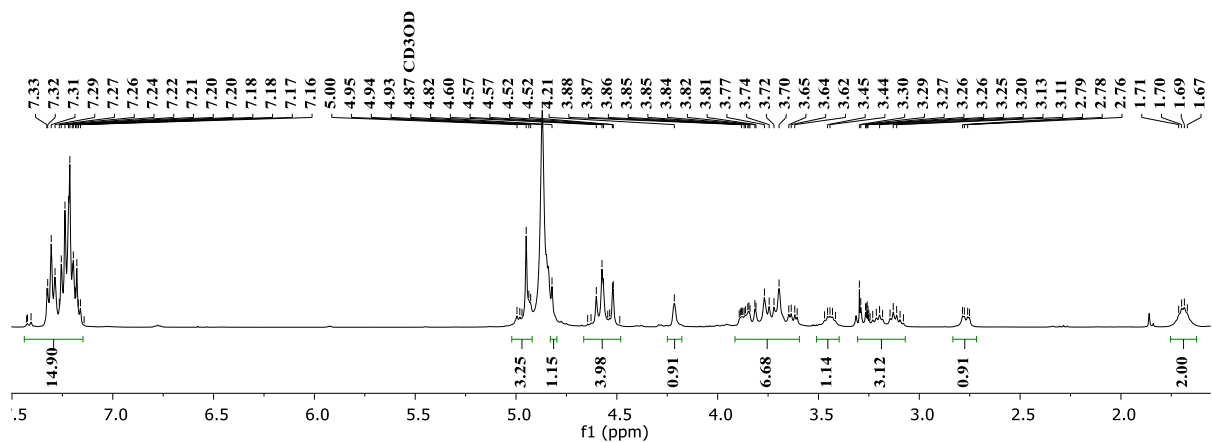
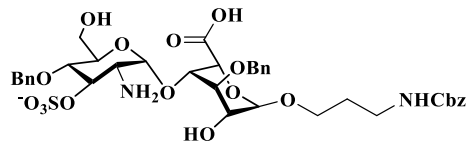


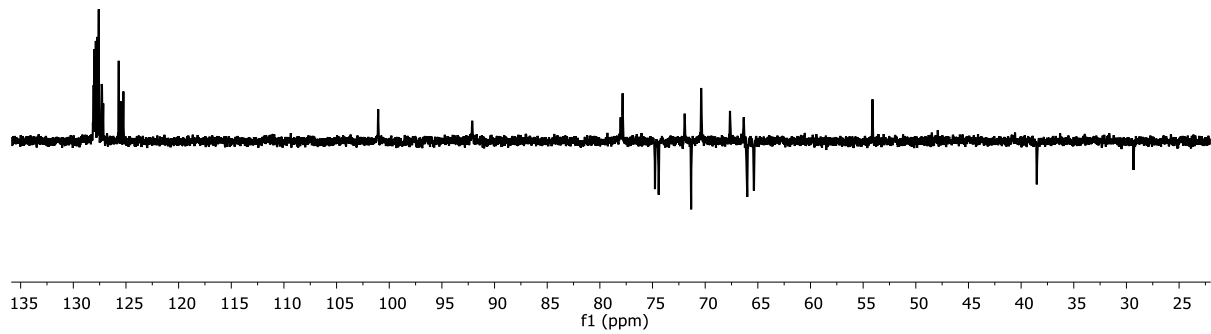
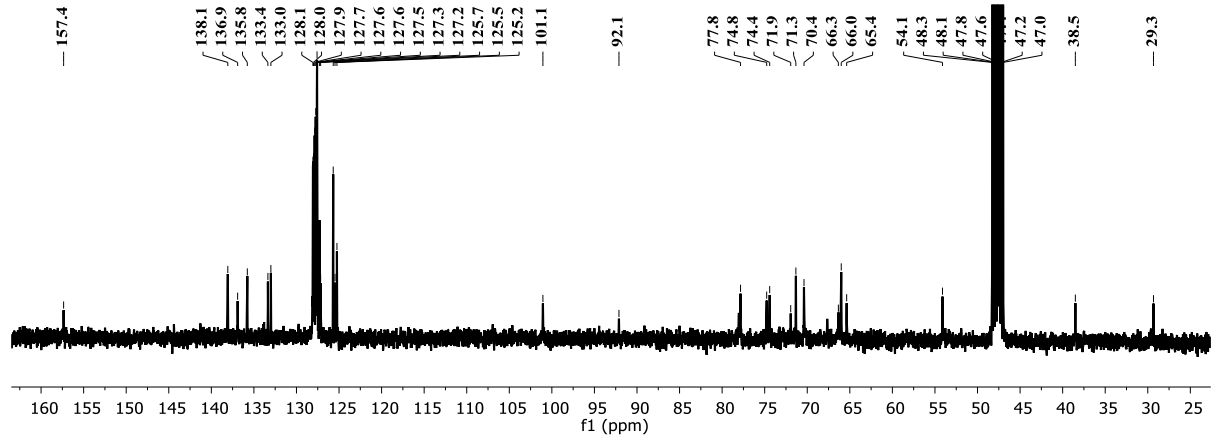
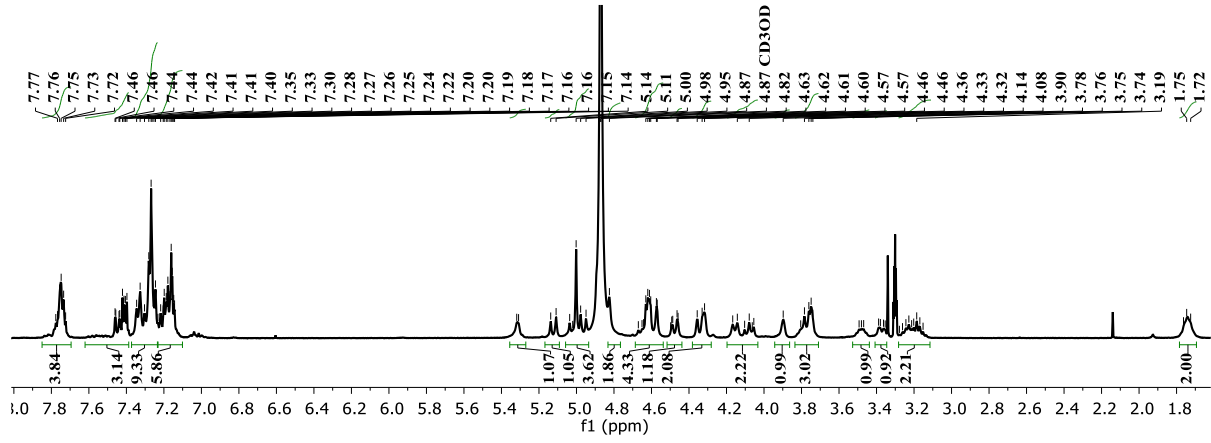
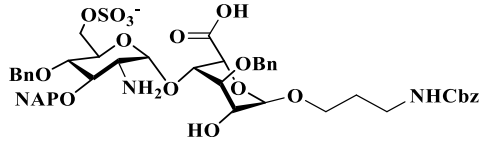


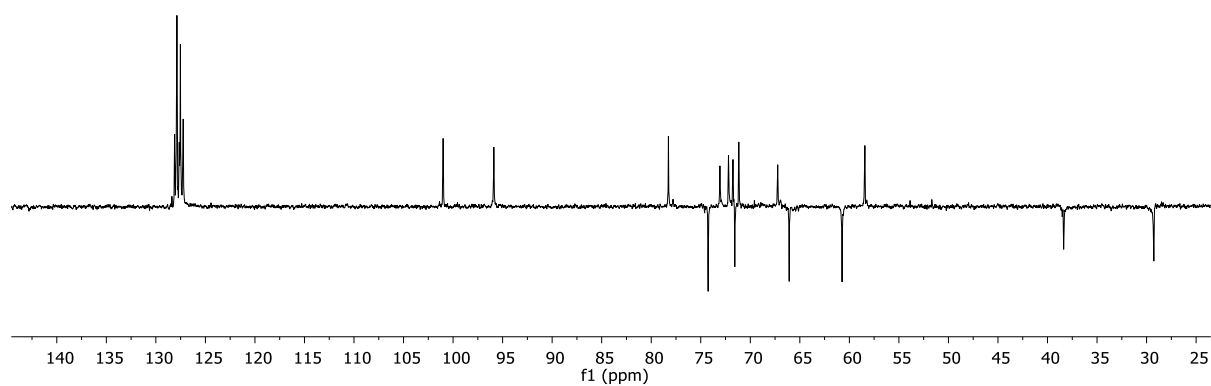
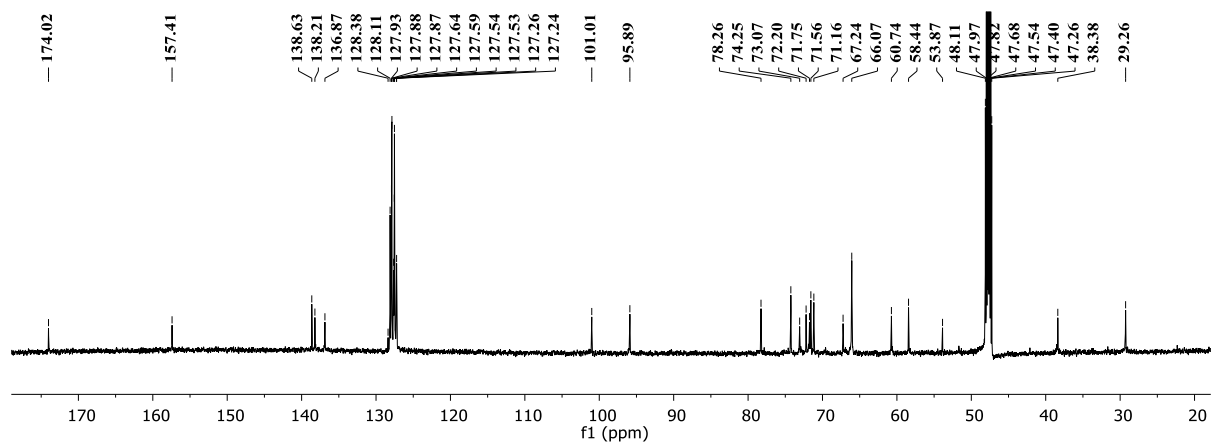
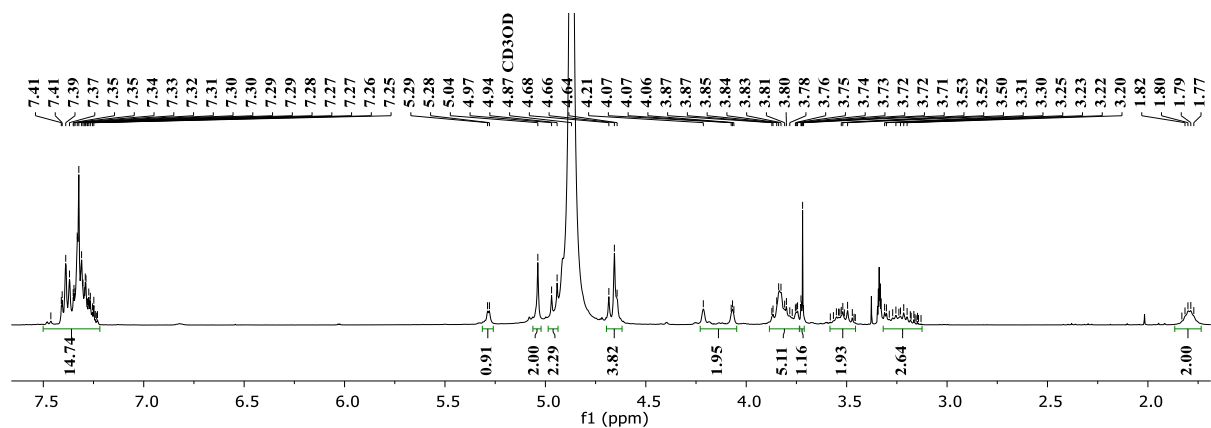
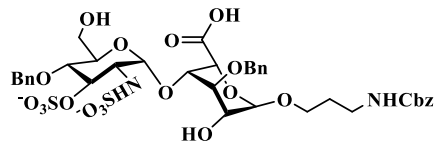


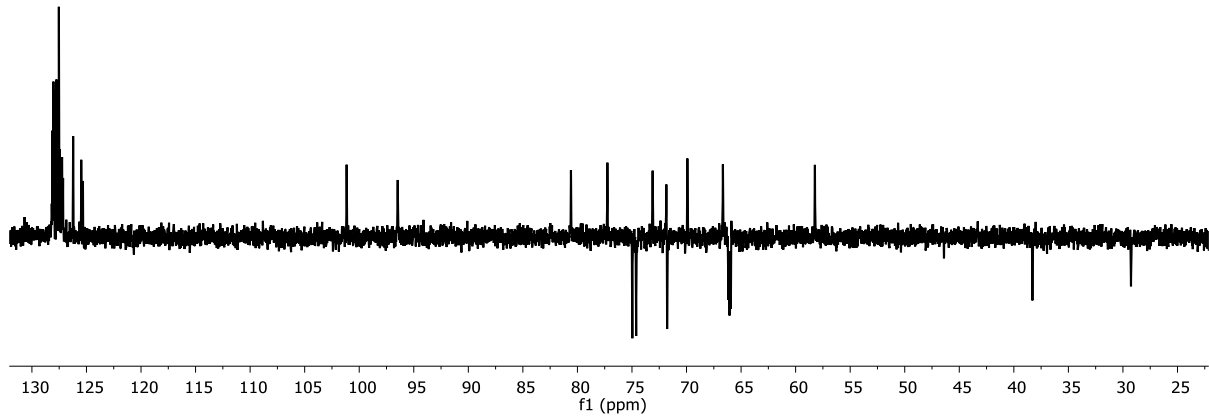
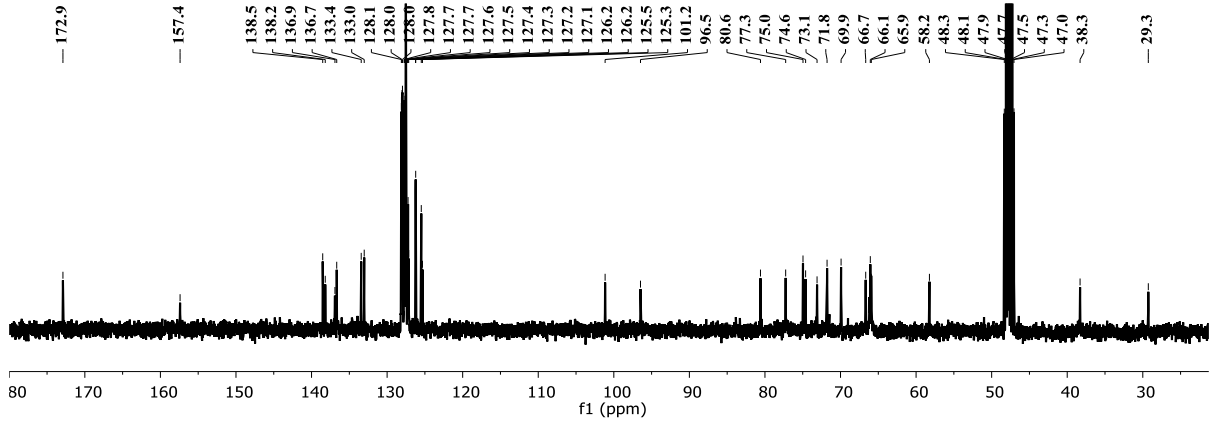
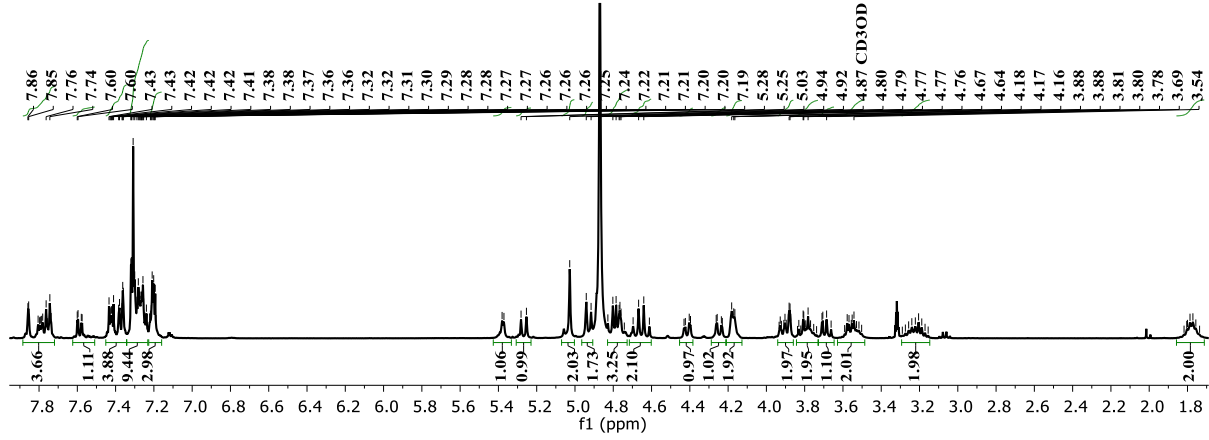
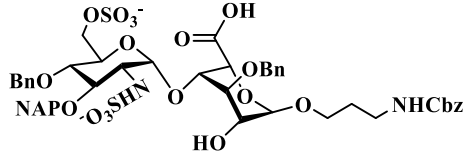


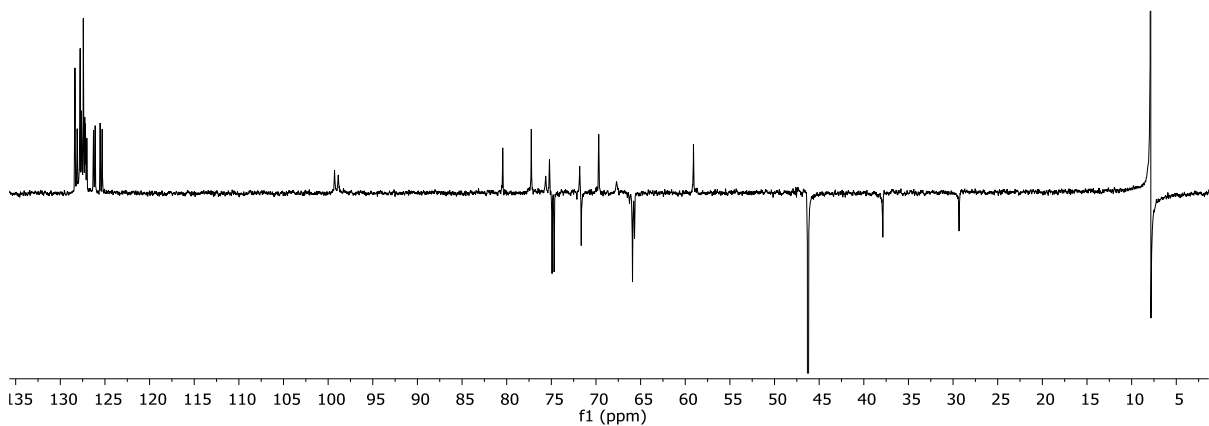
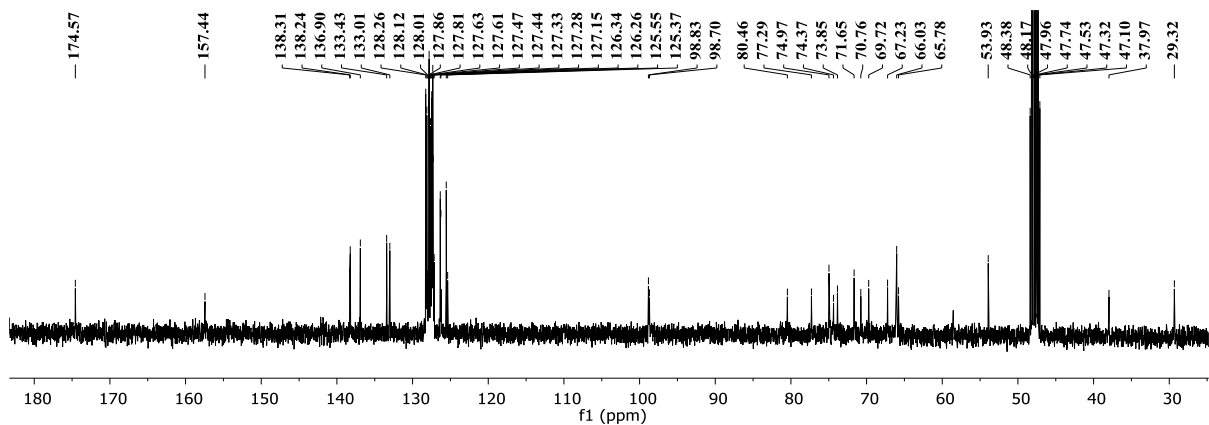
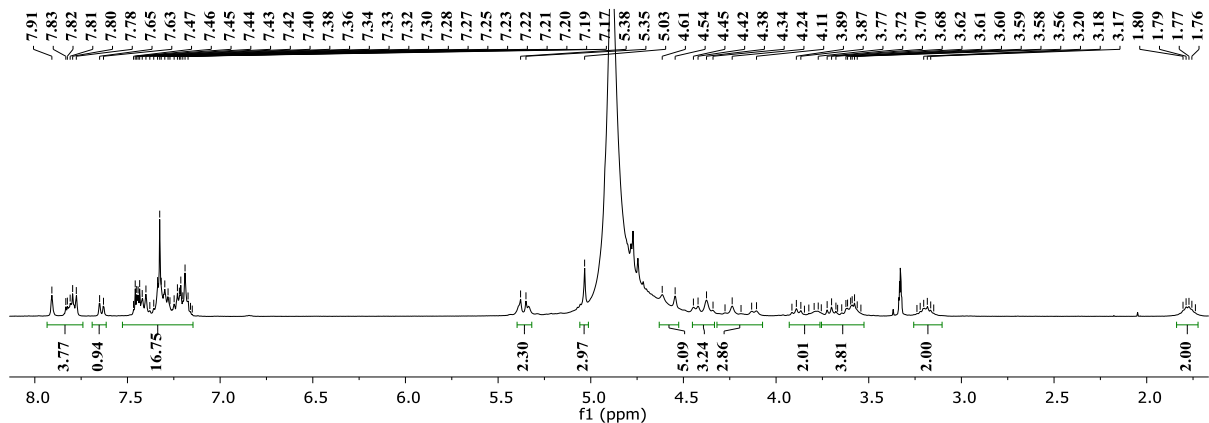
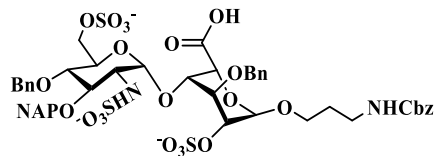


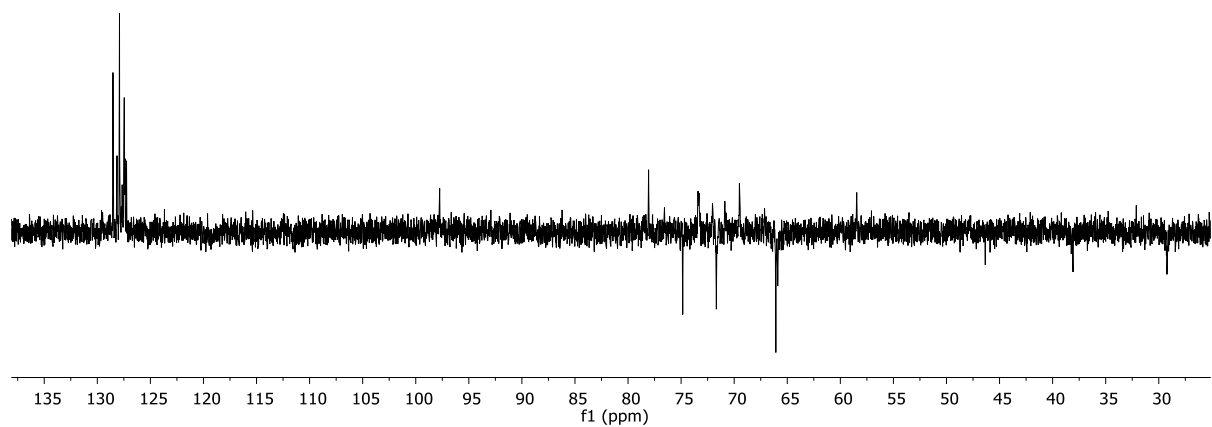
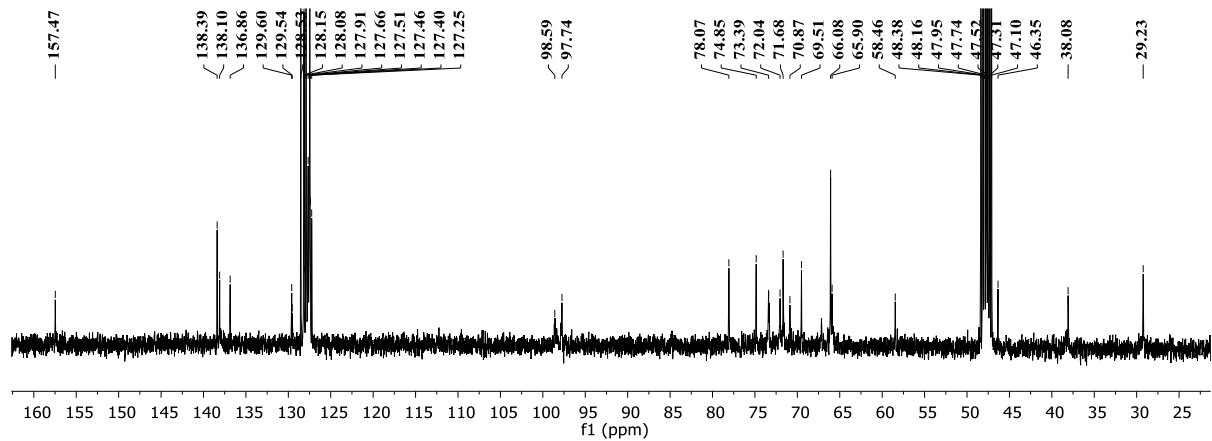
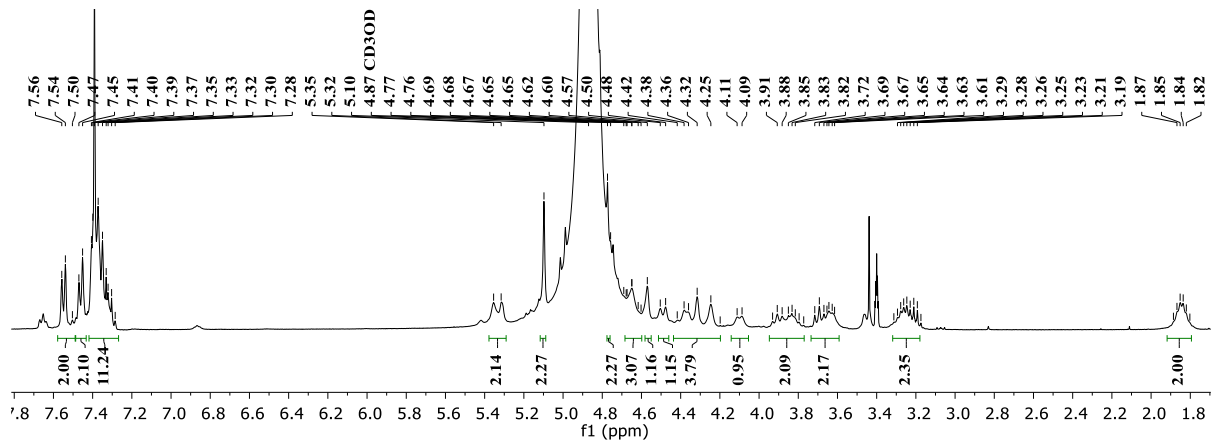
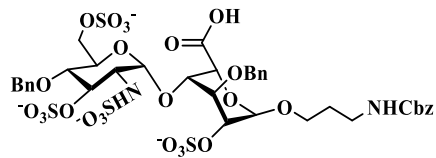




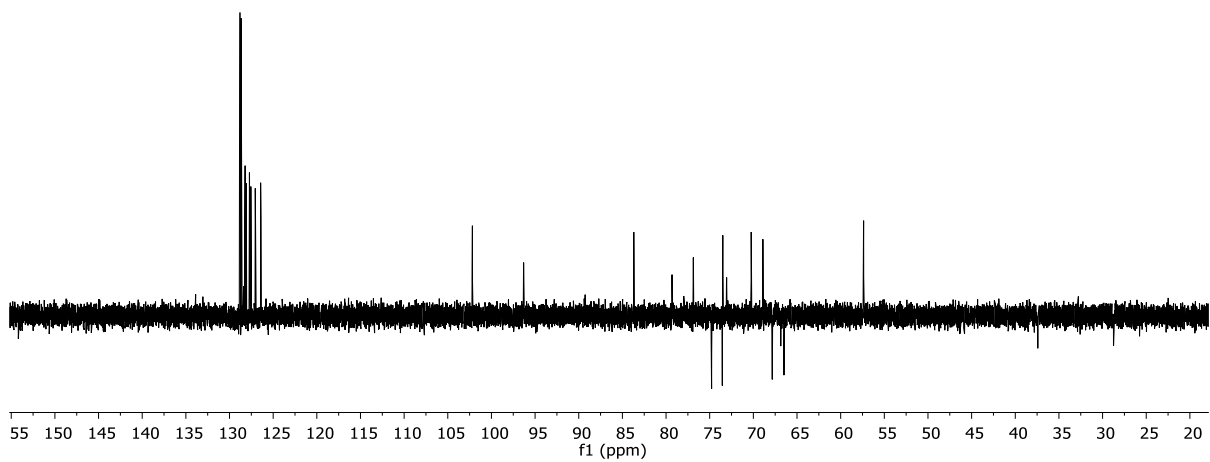
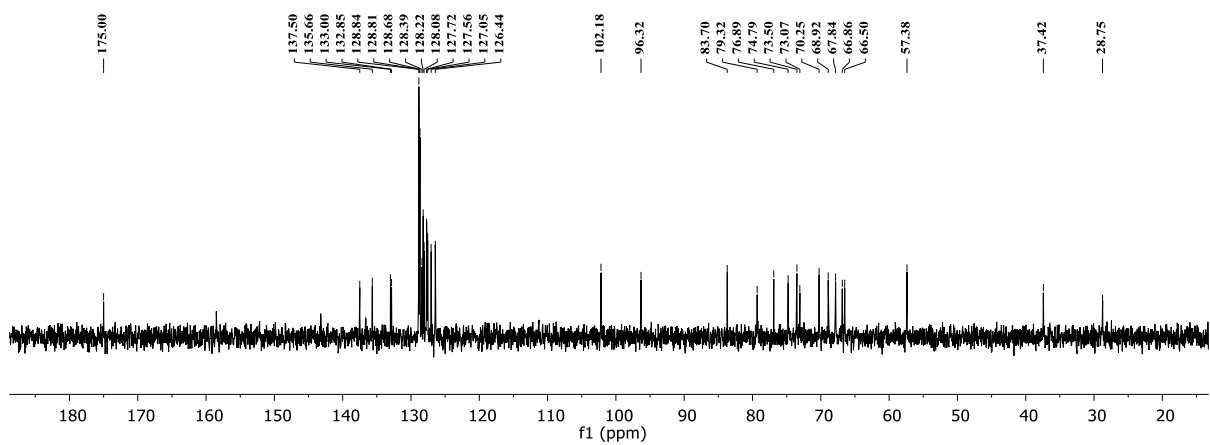
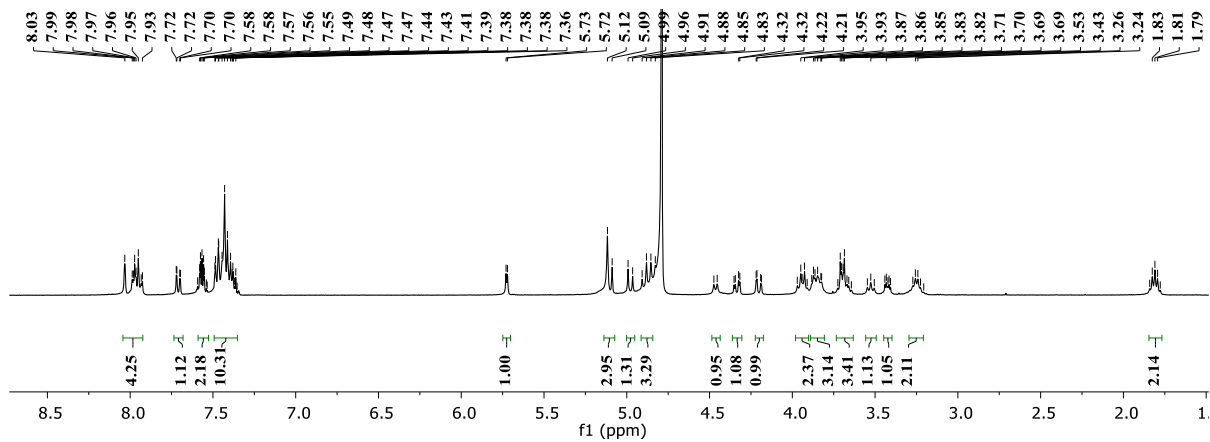
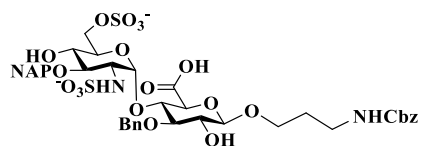


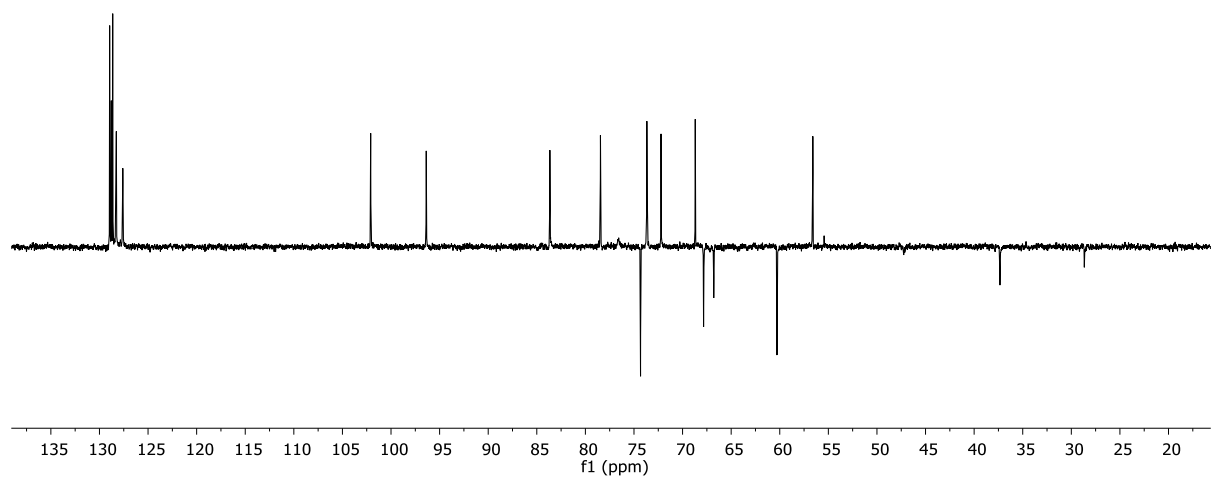
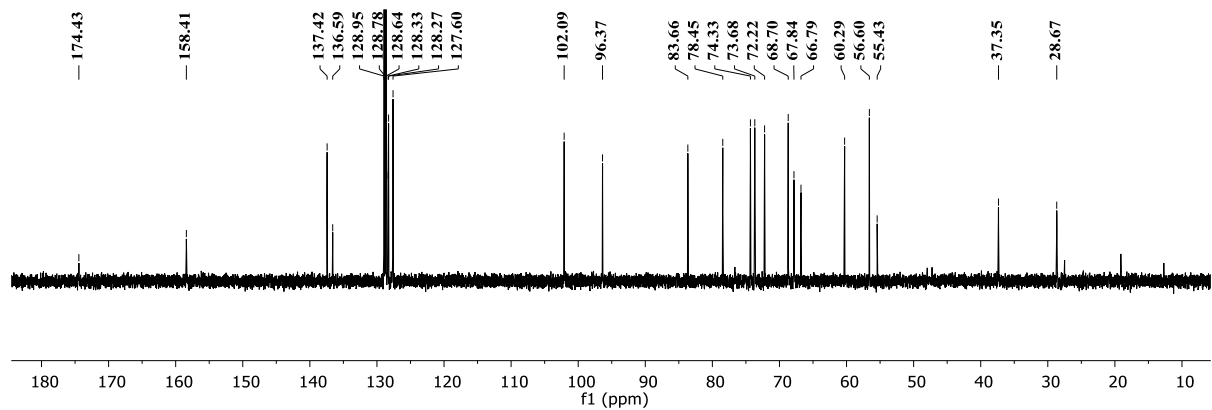
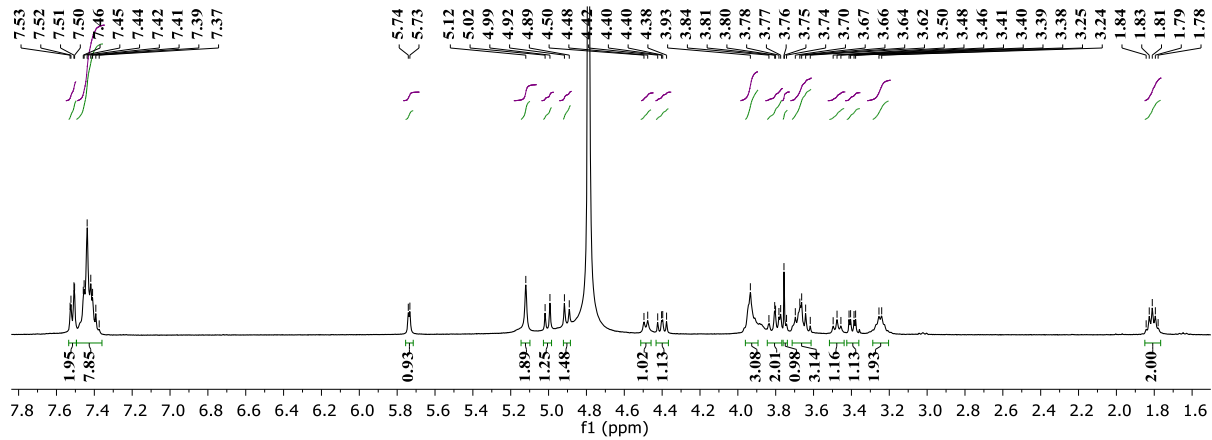
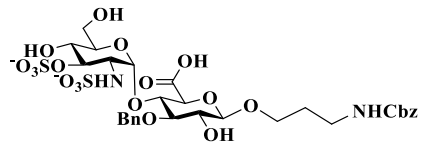


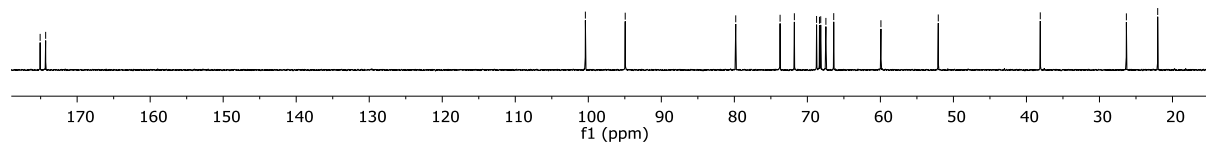
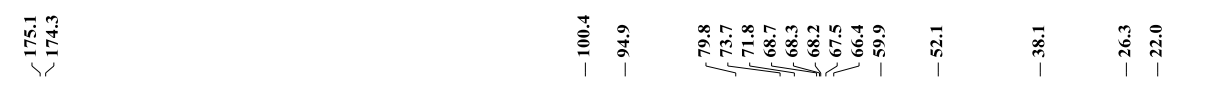
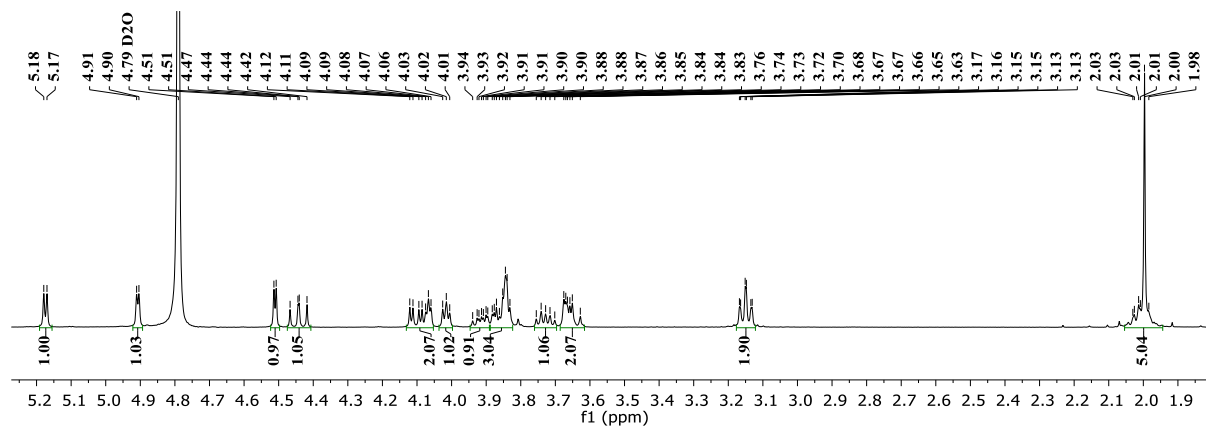
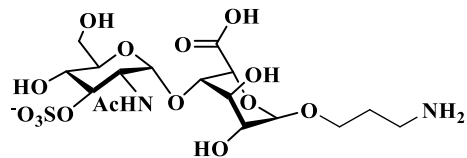


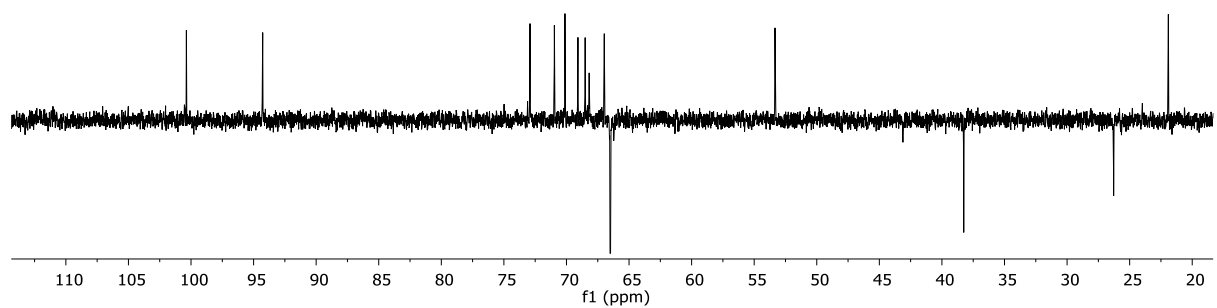
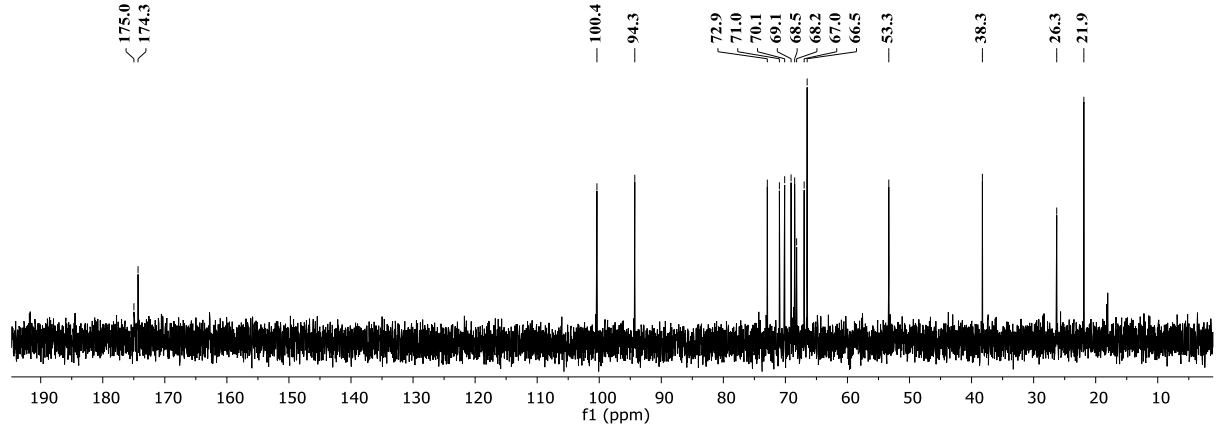
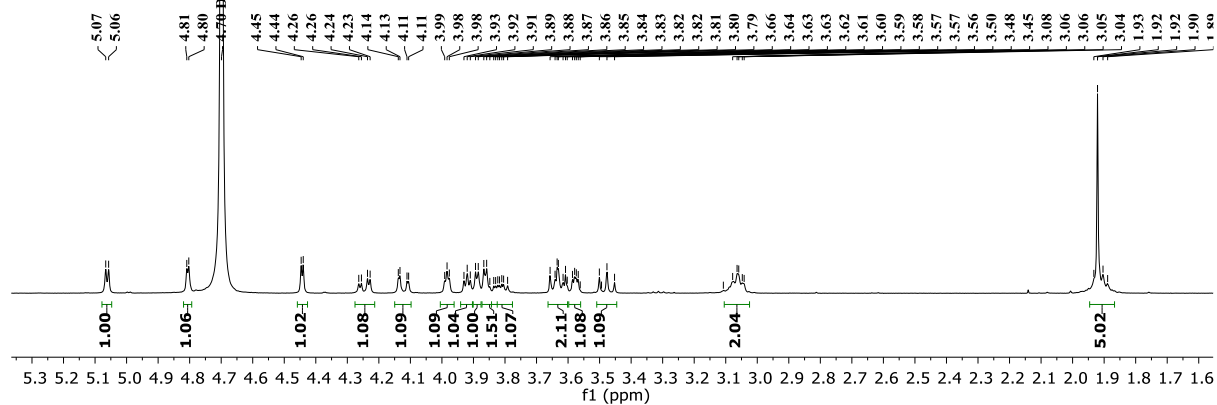
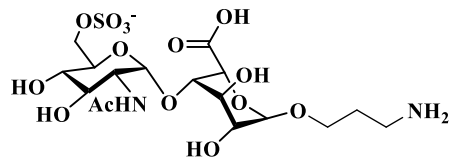


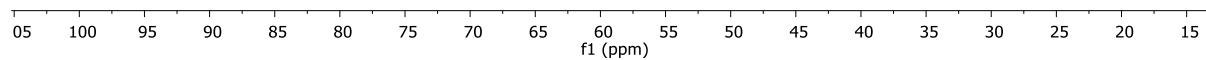
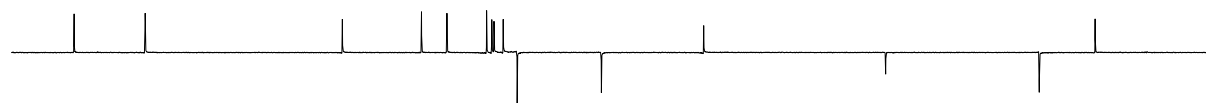
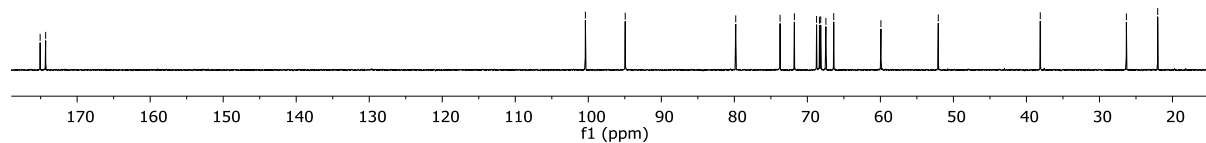
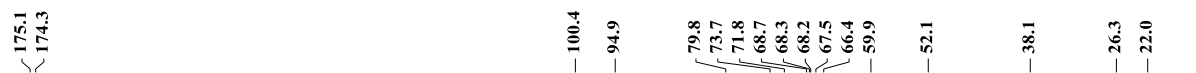
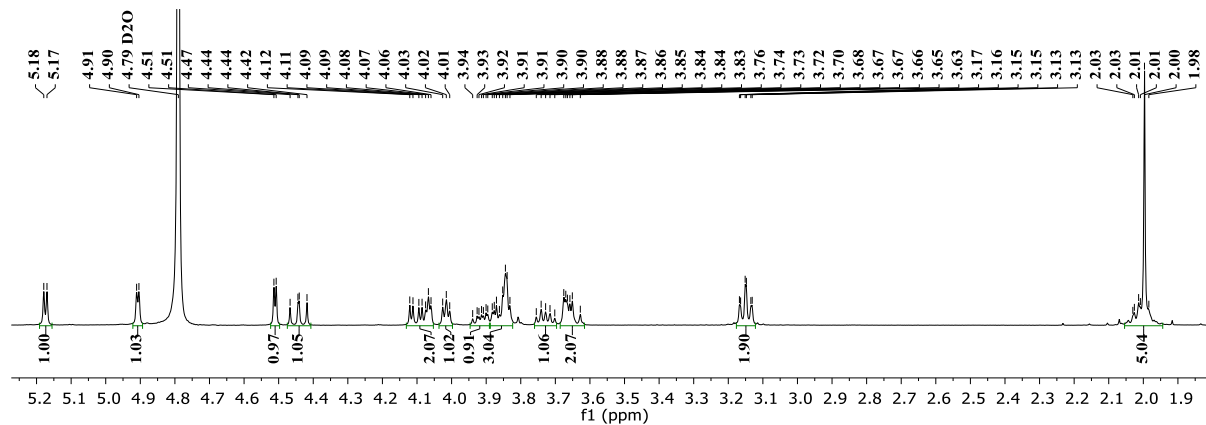
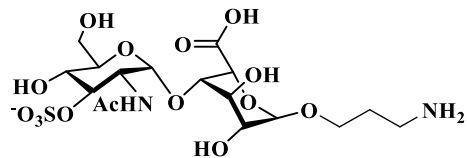


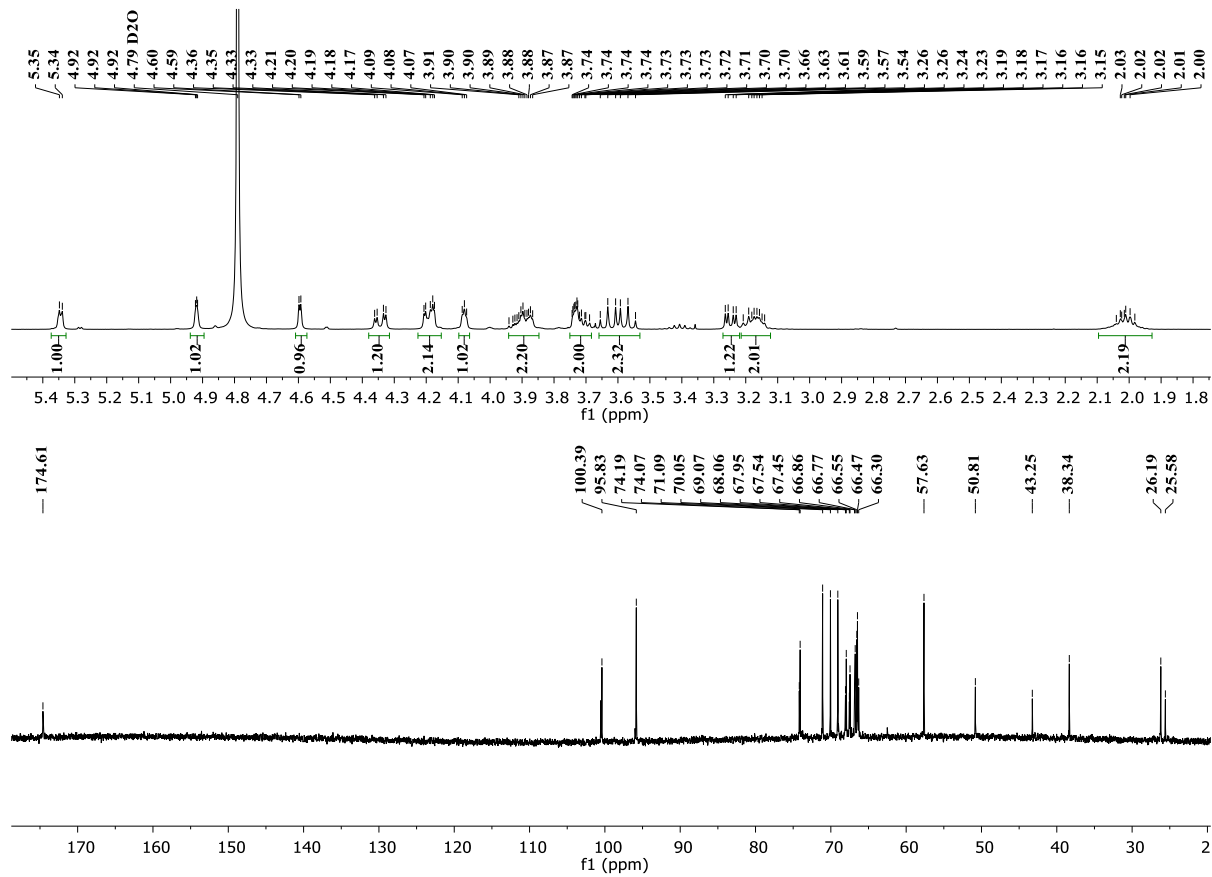
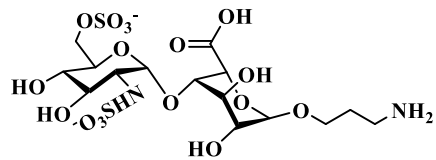




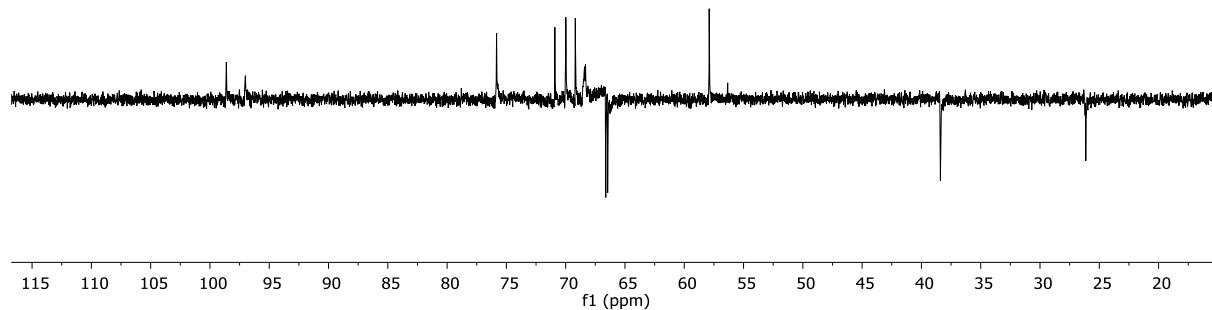
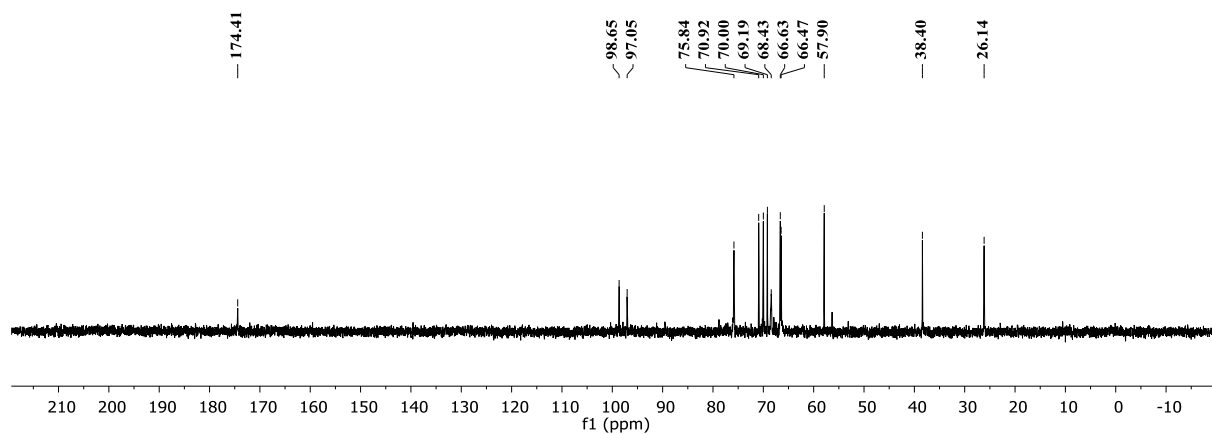
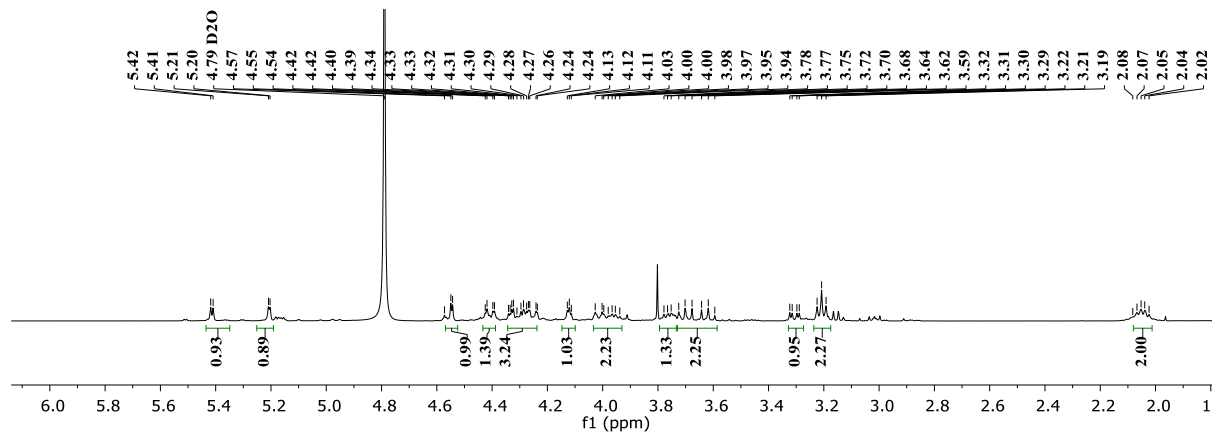
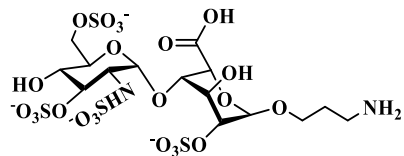




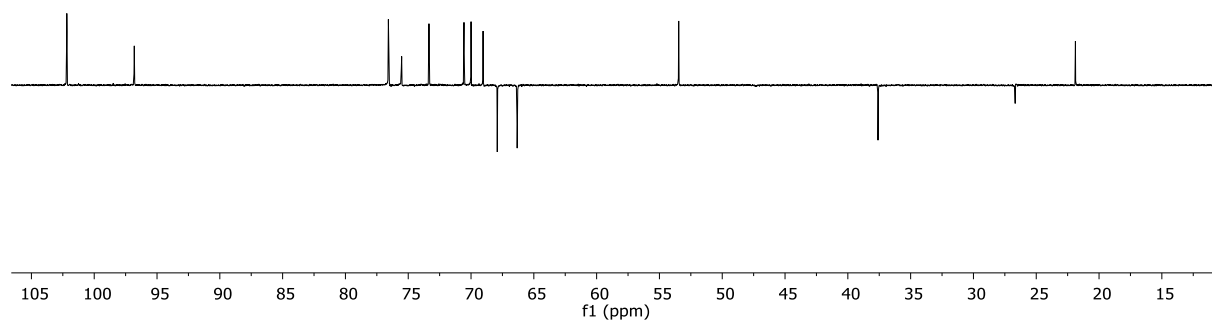
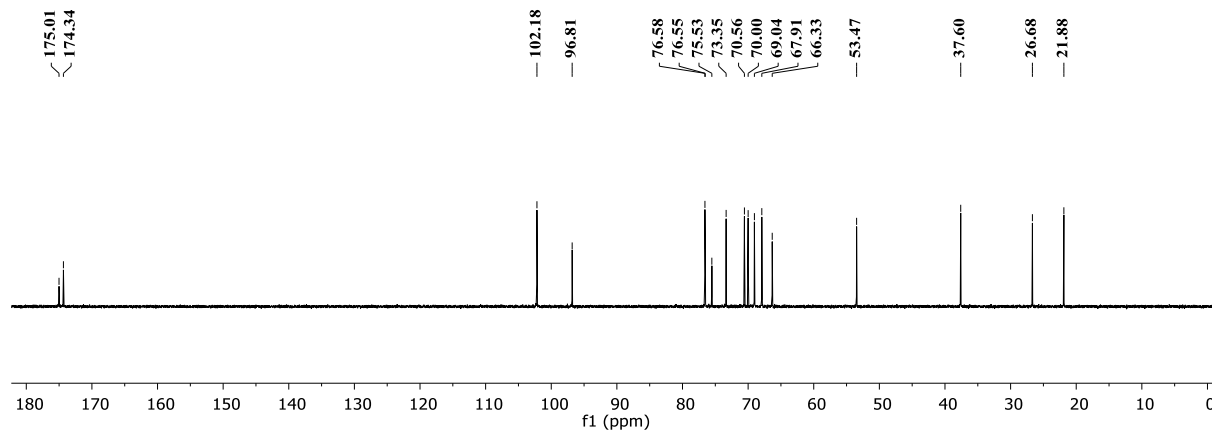
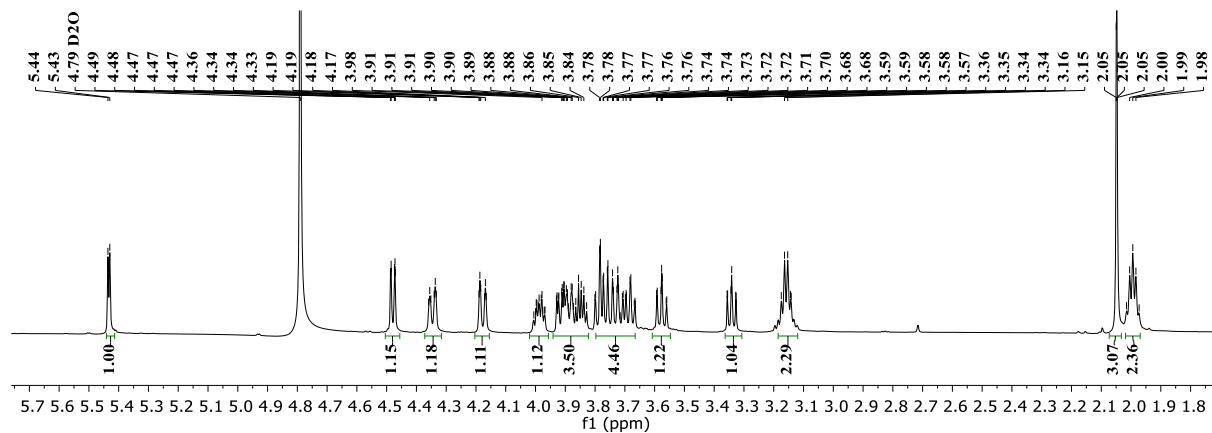
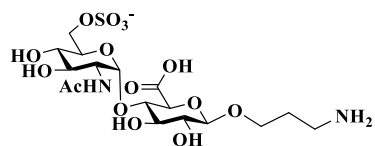


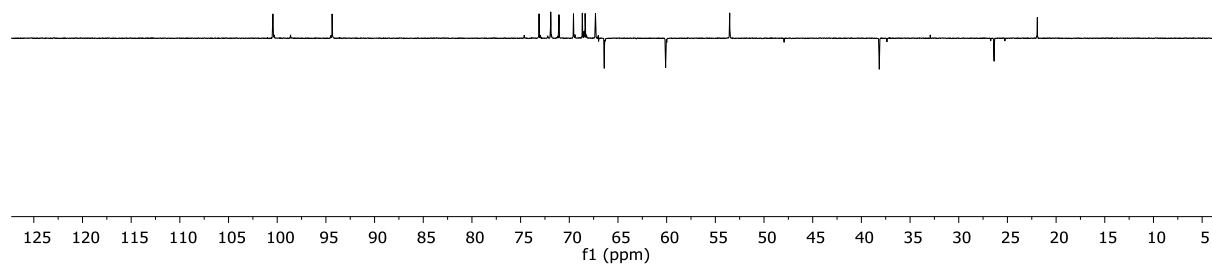
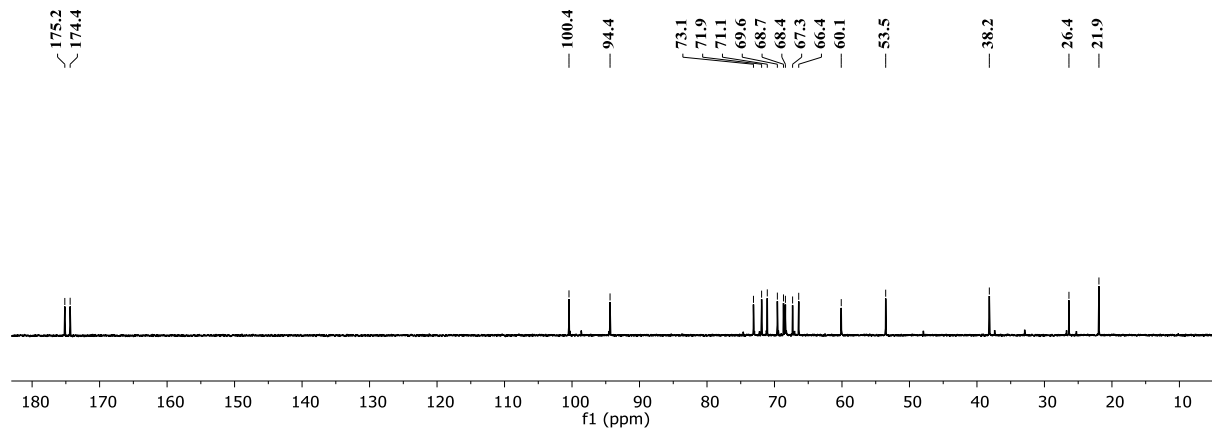
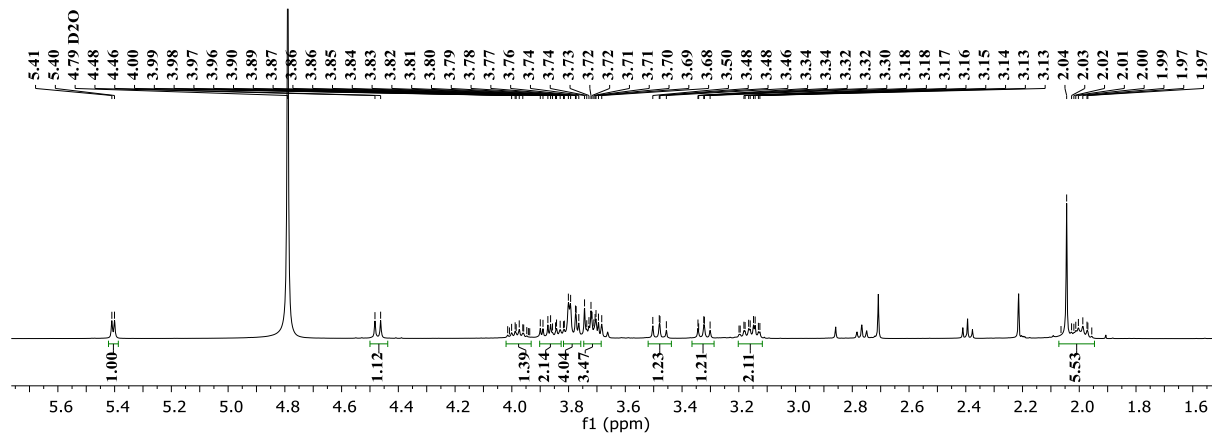
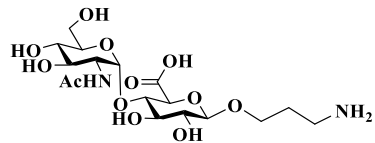


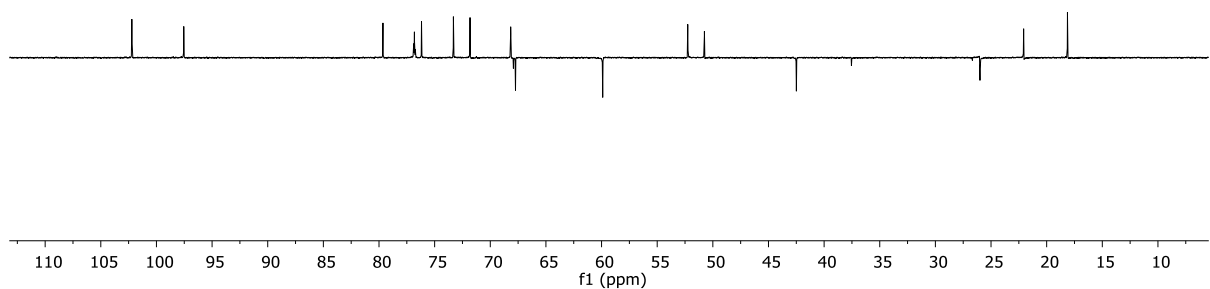
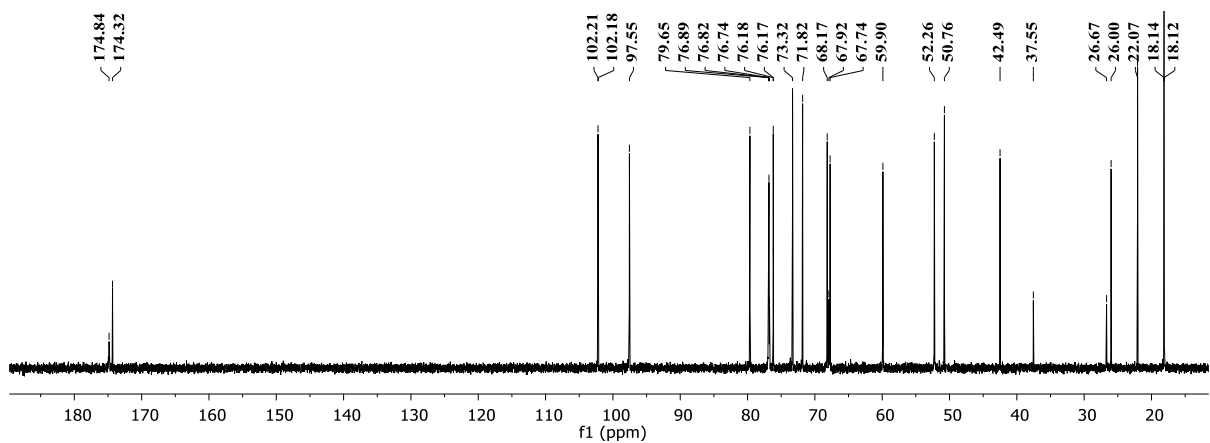
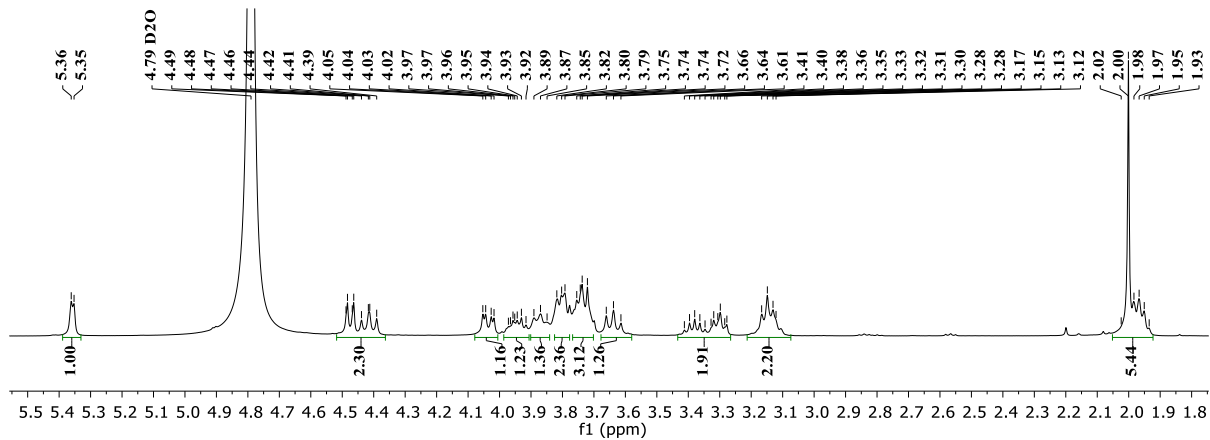
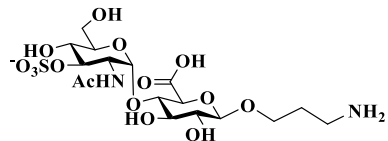


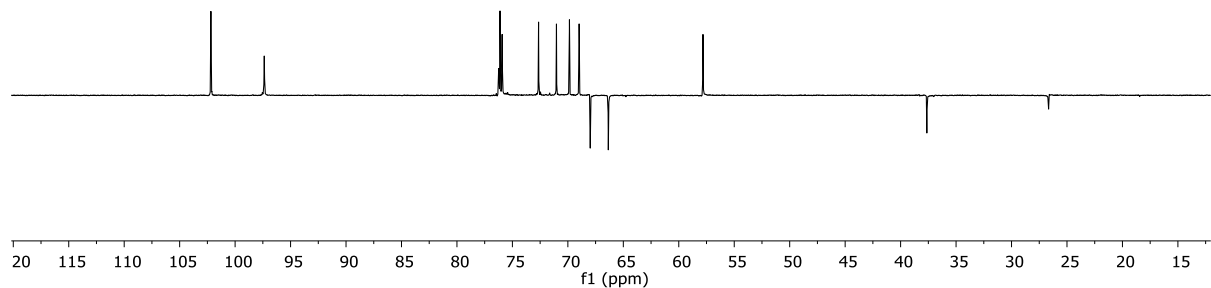
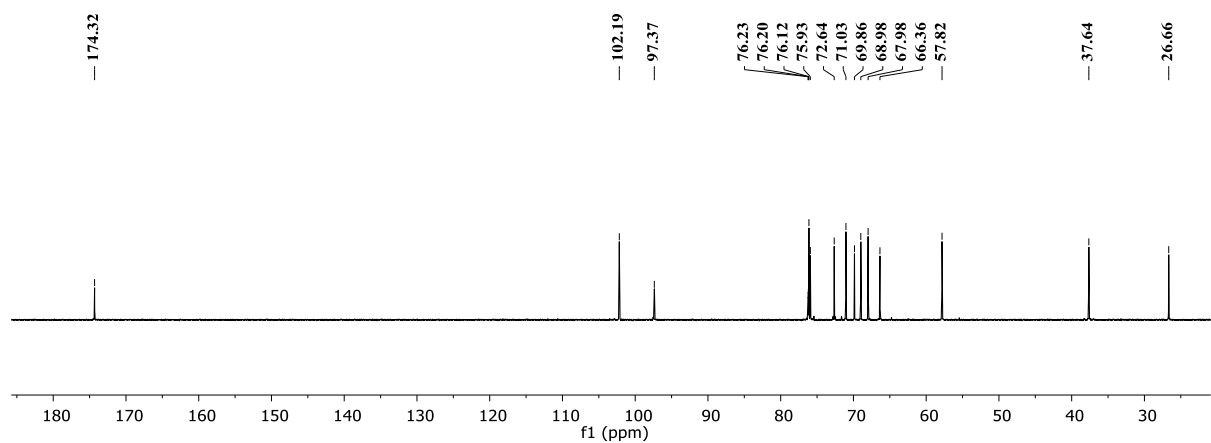
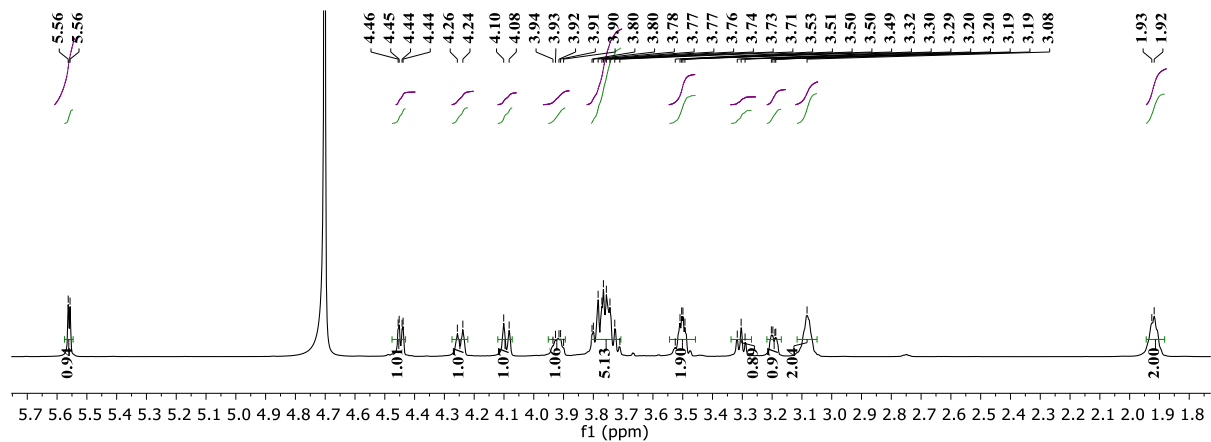
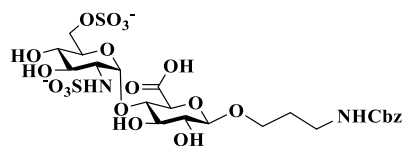


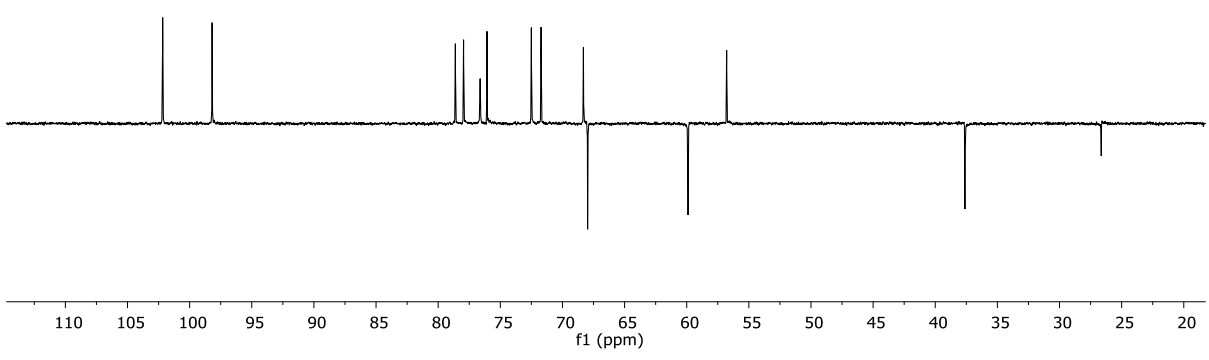
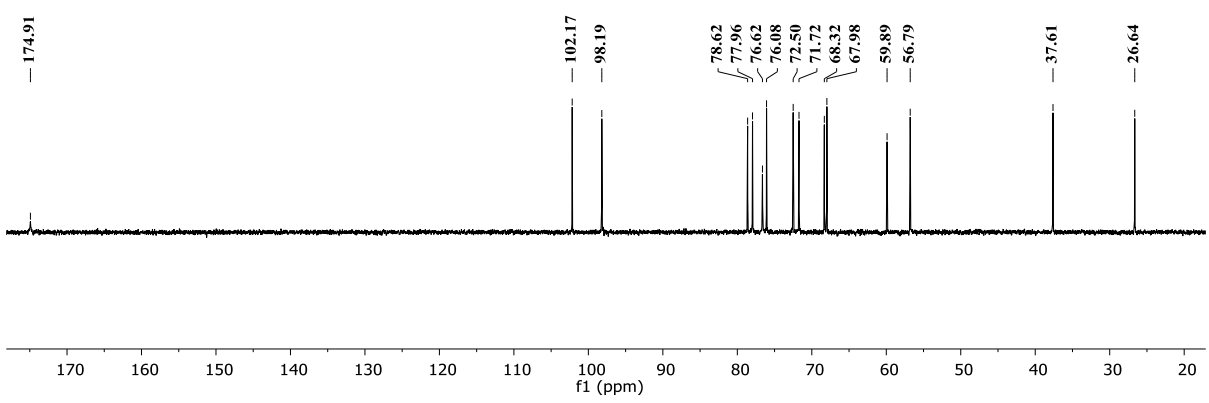
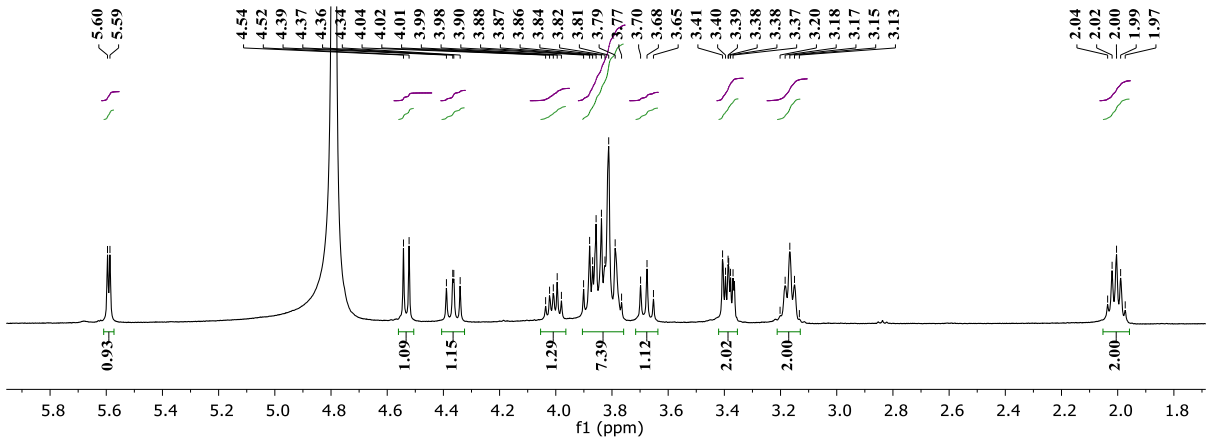
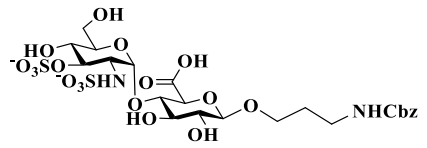


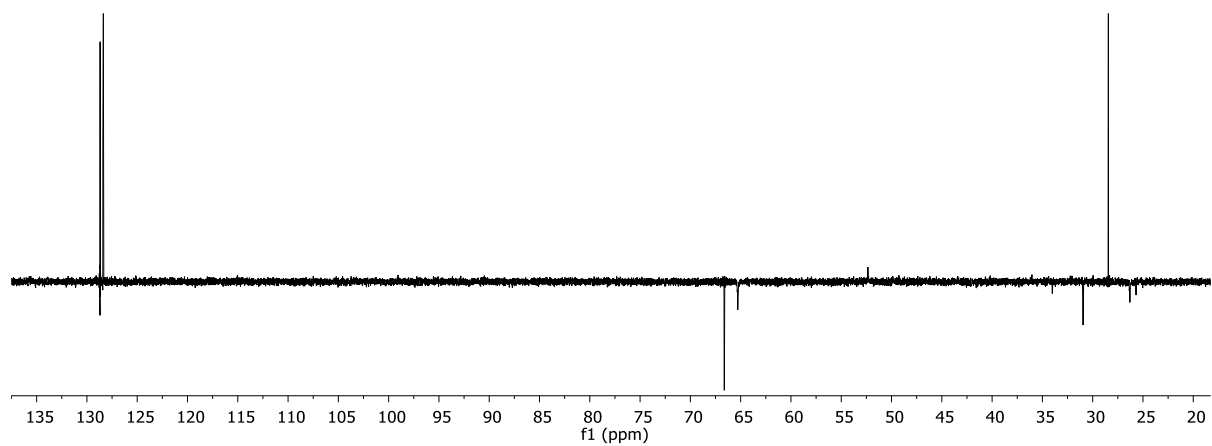
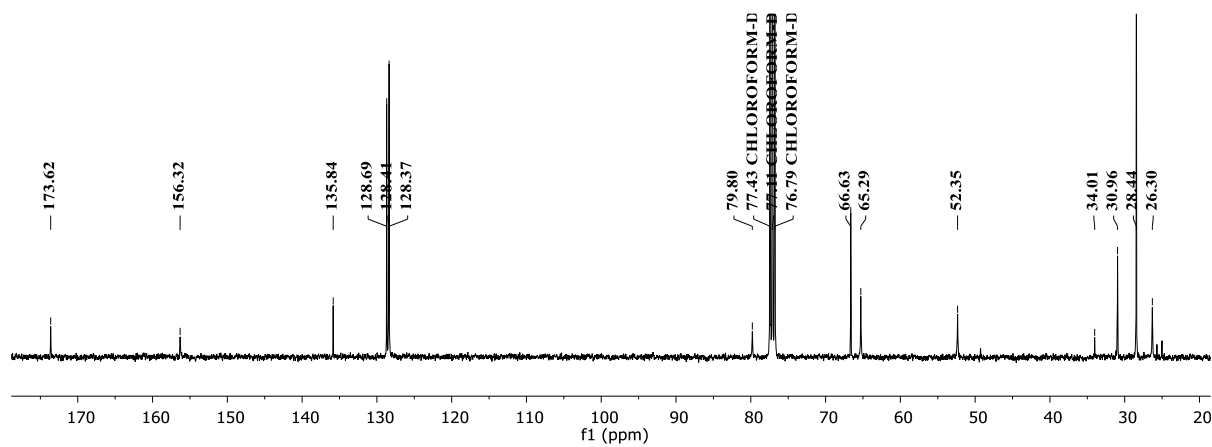
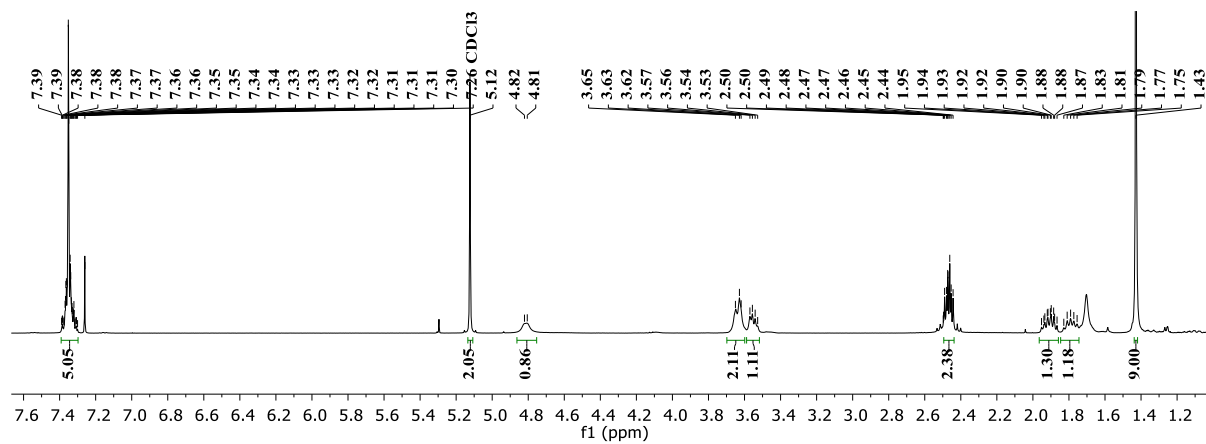
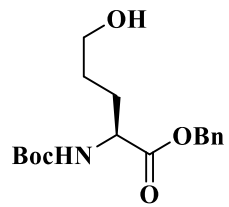


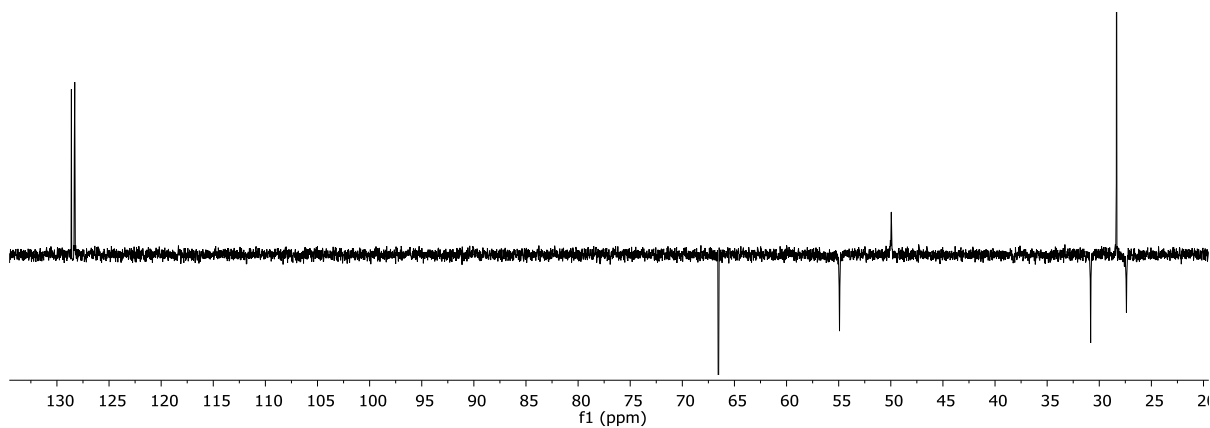
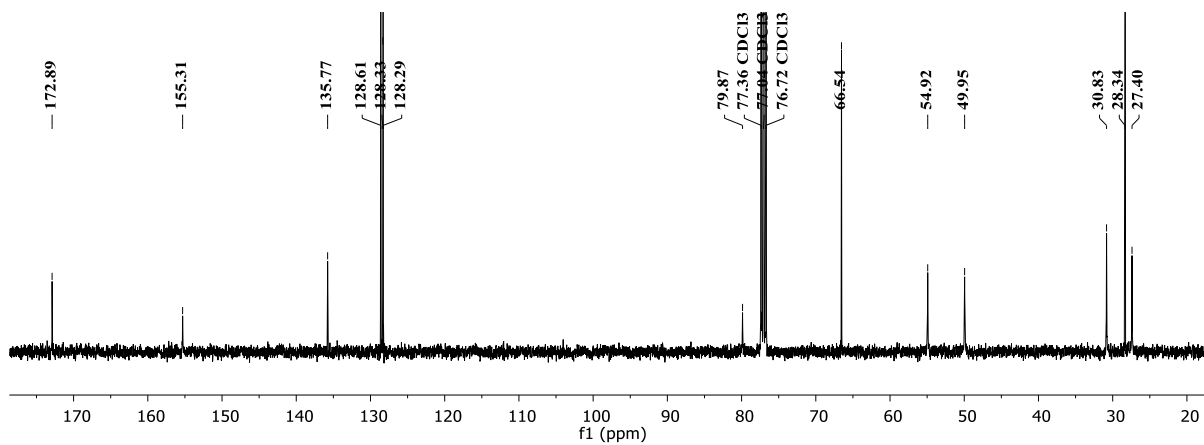
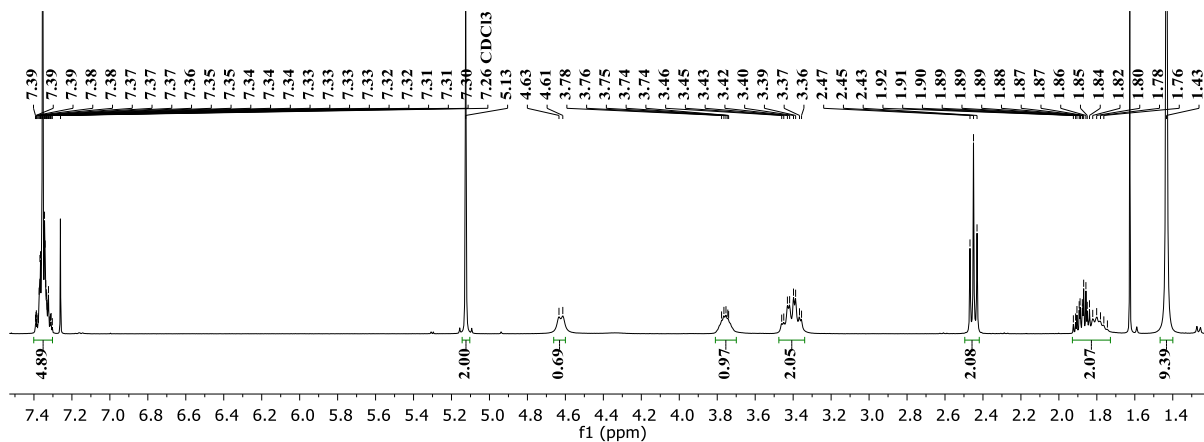
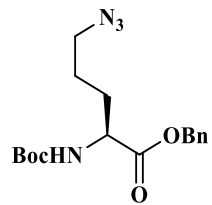








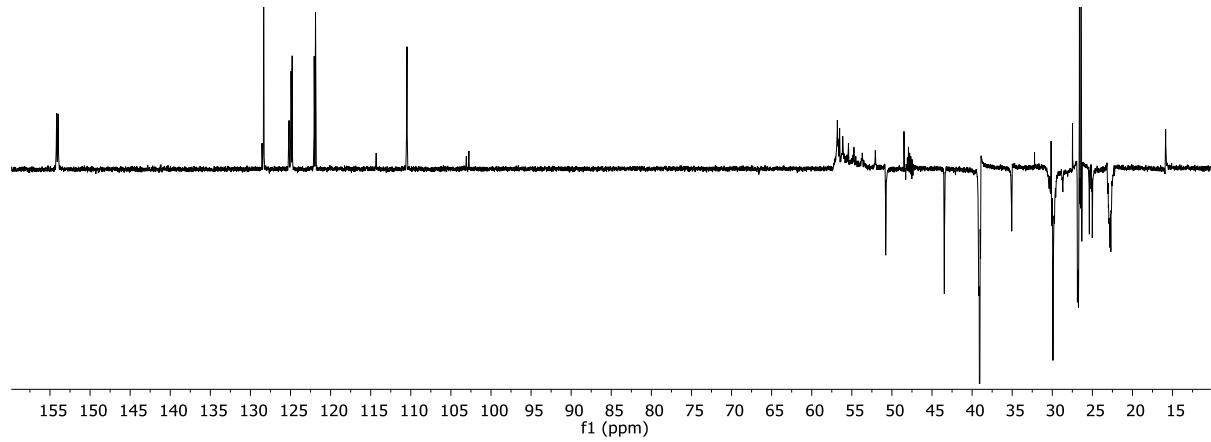
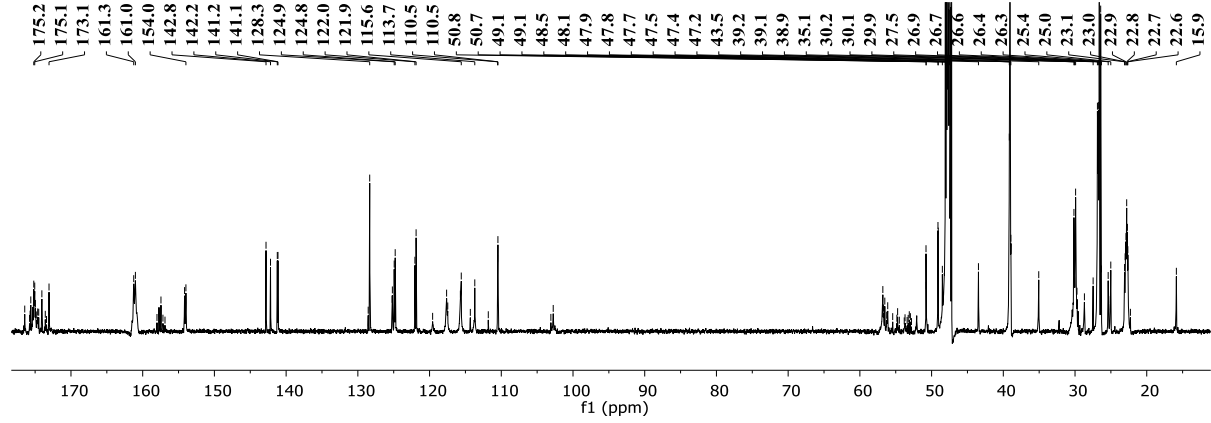
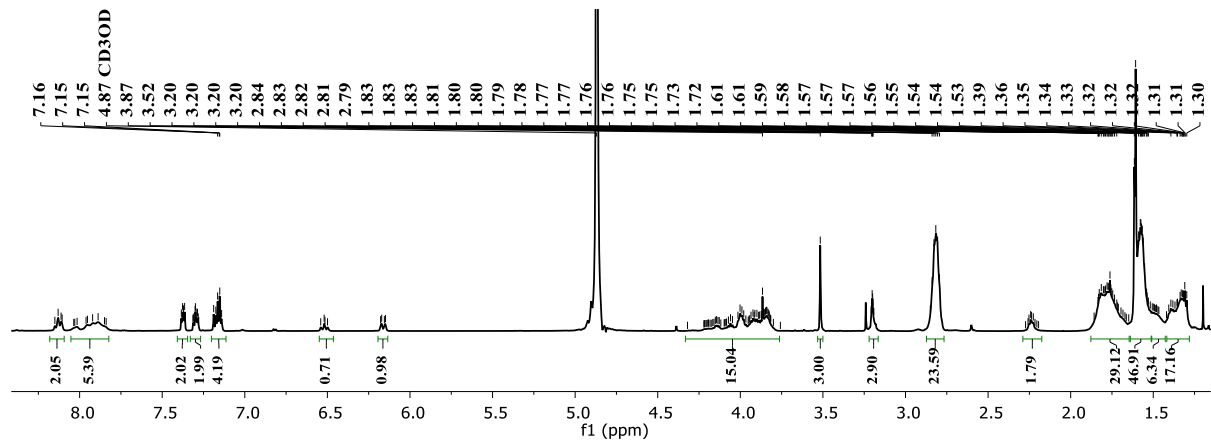






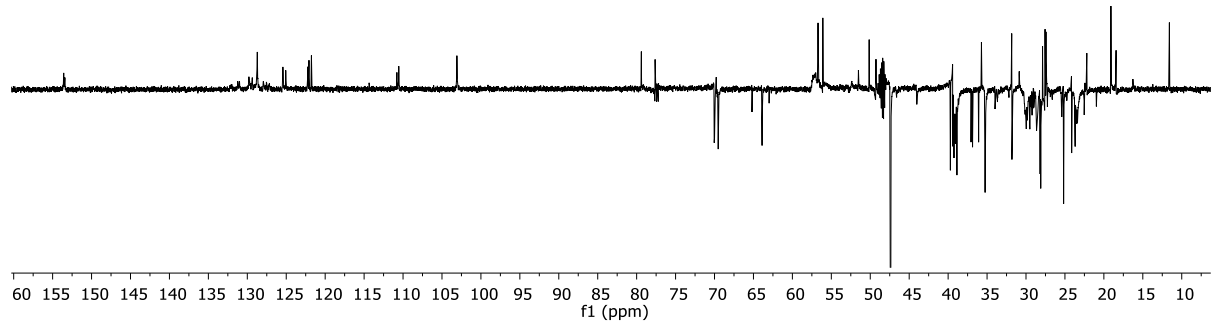
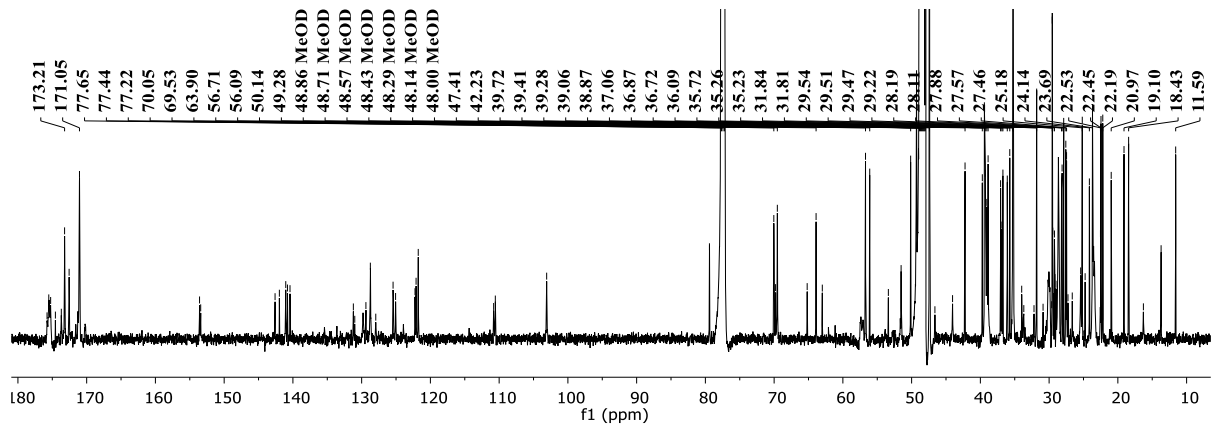
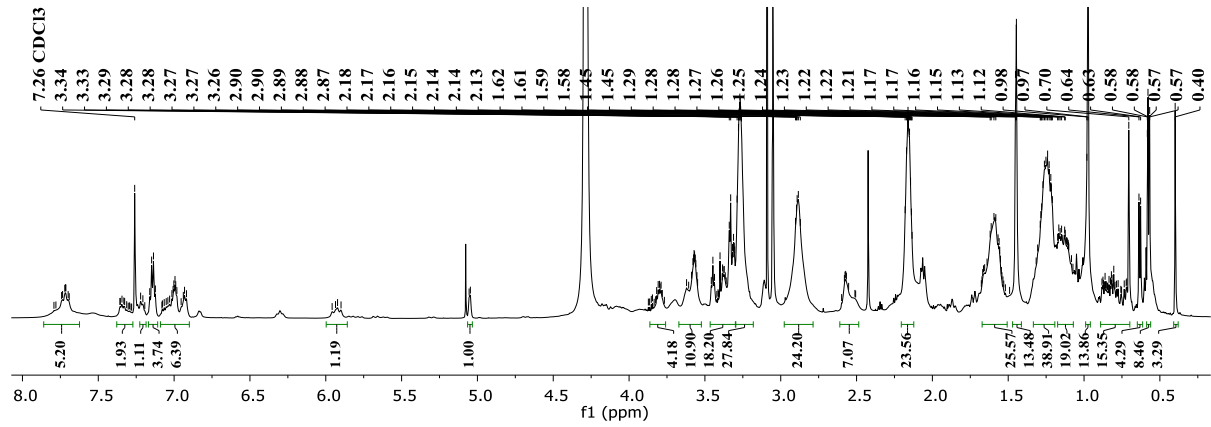


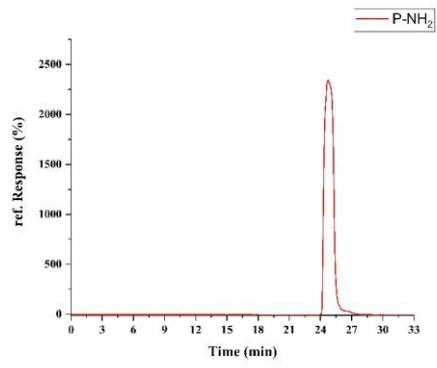
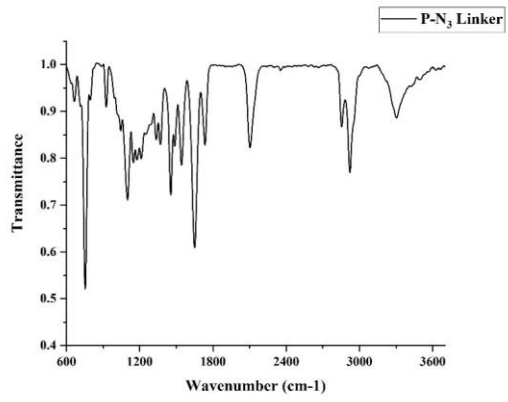
# 12mer Peptide NMR



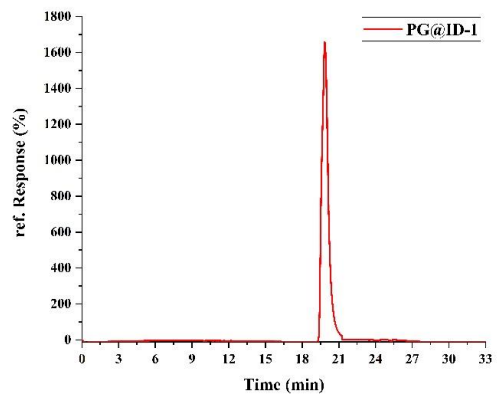
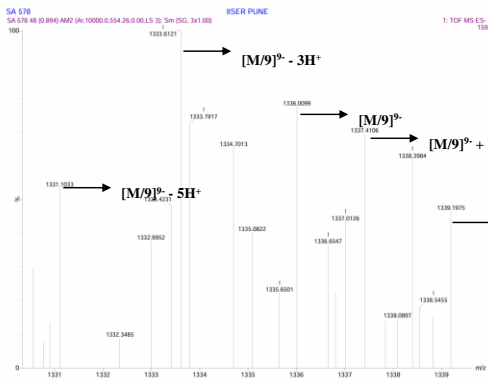
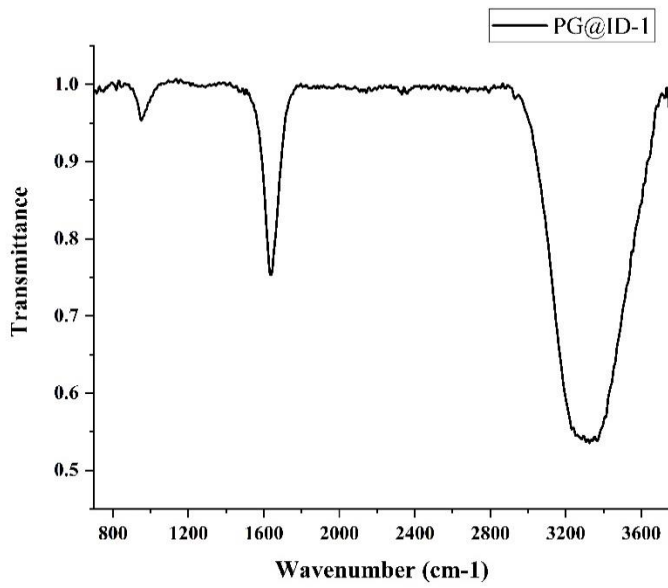
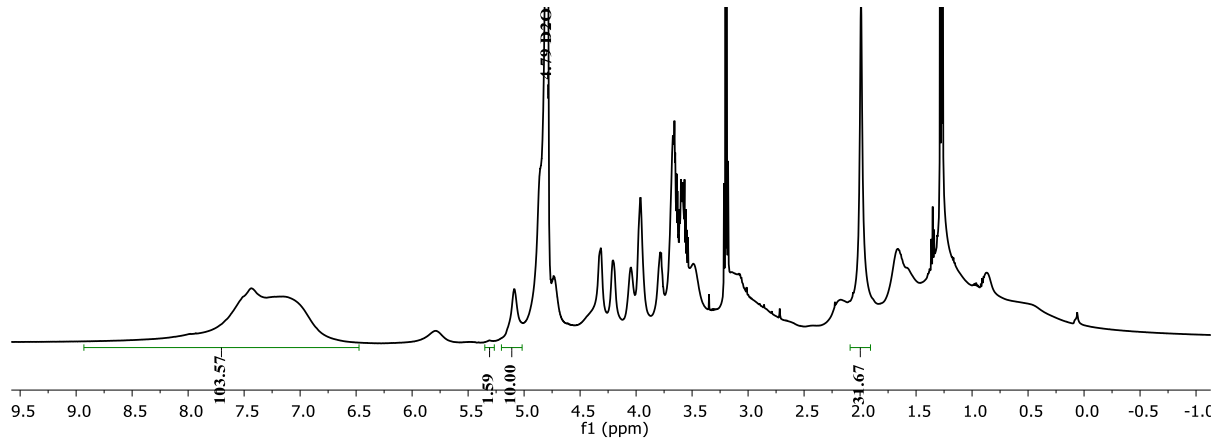


# 12mer Peptide with 3-Azidopropanoate Linker

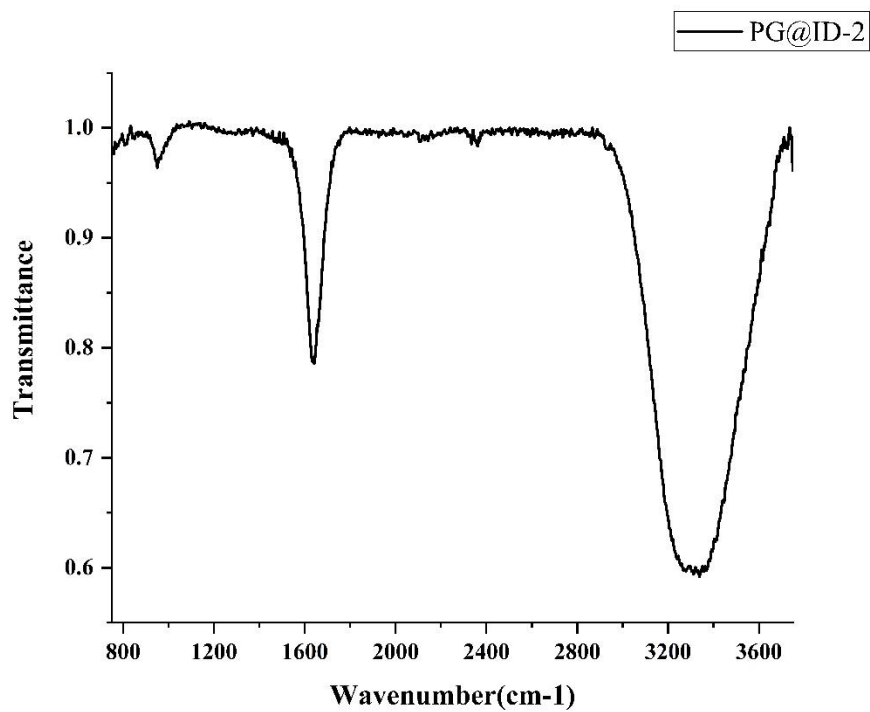
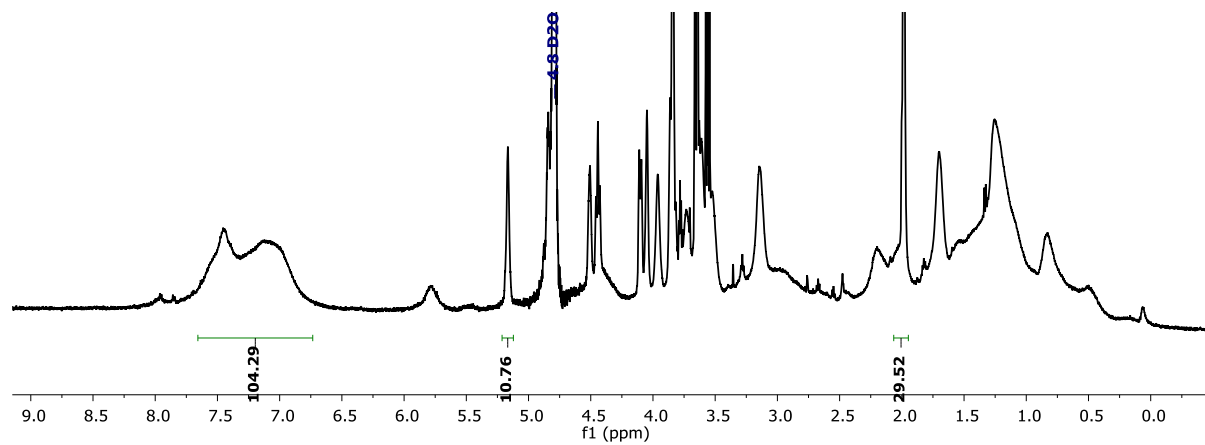


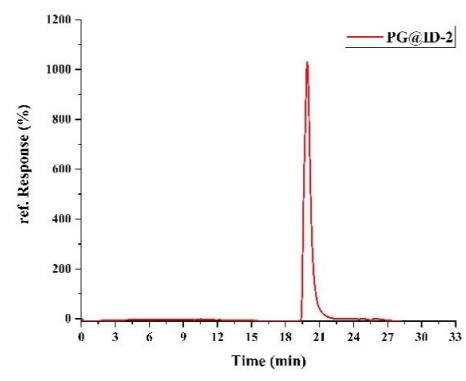
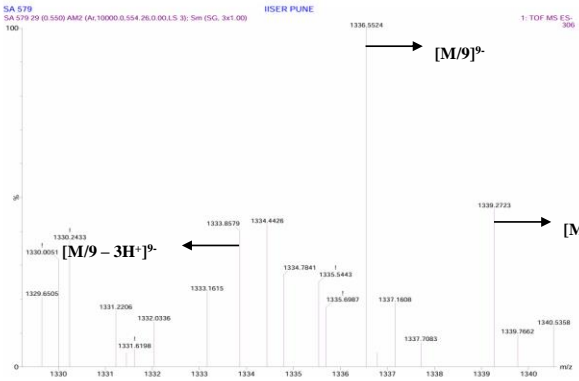


# PG@ID-1

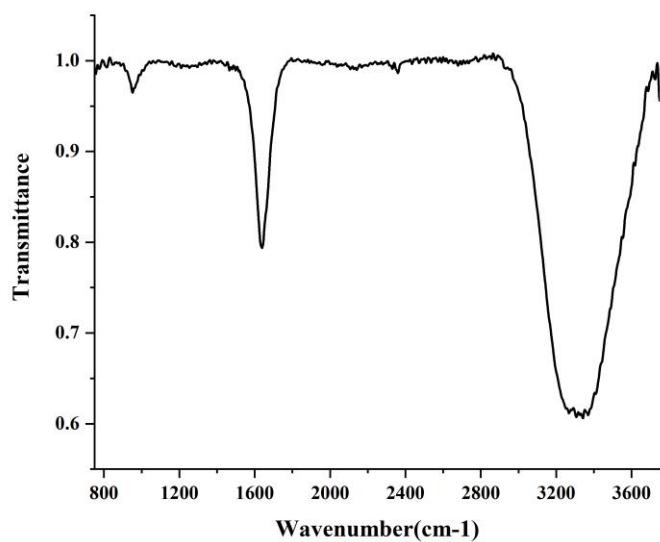
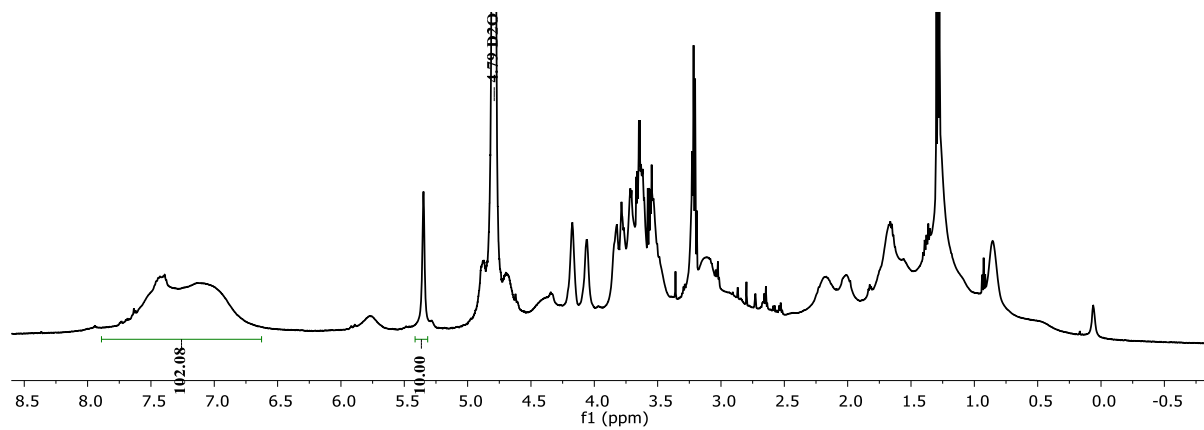


PG@ID-2

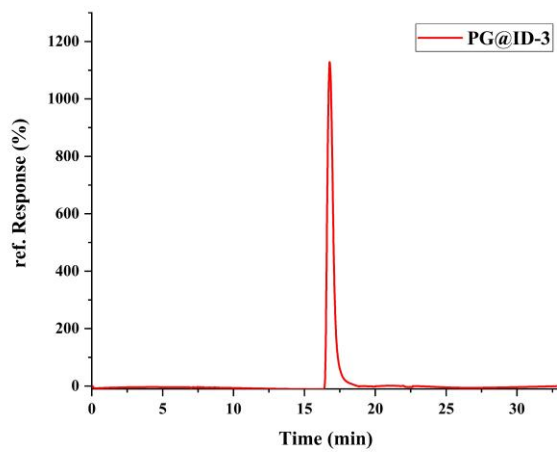
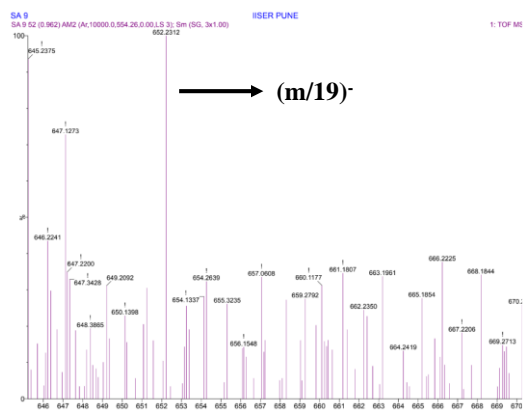




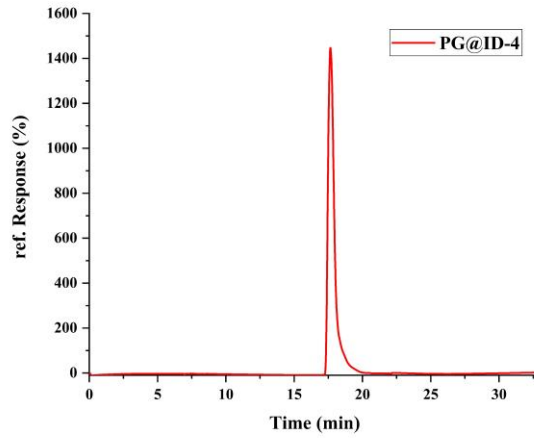
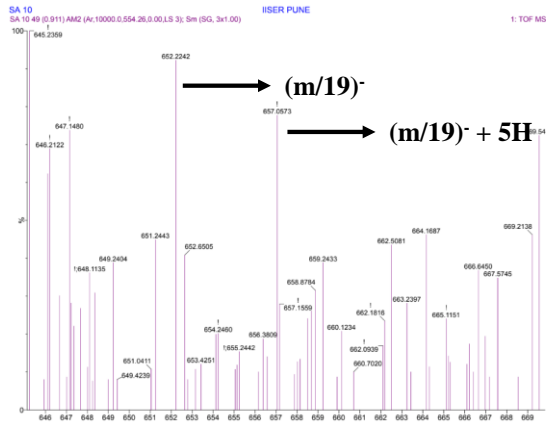
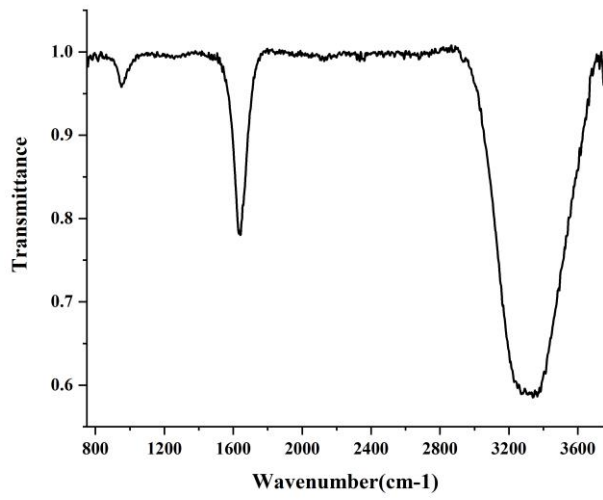
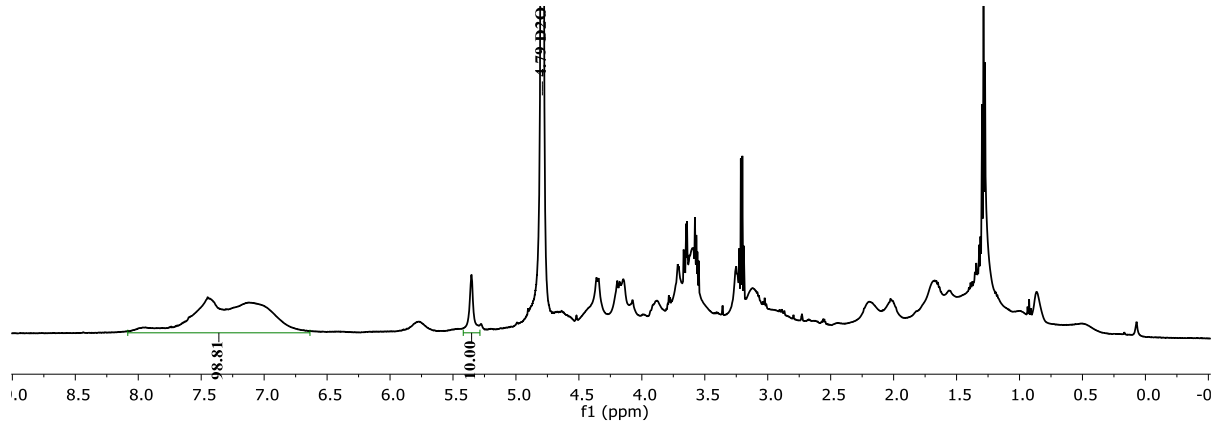
PG@ID-3



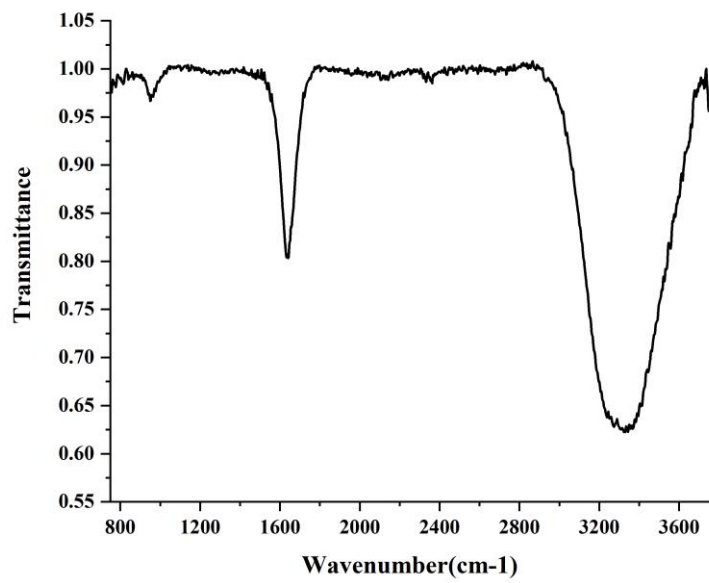
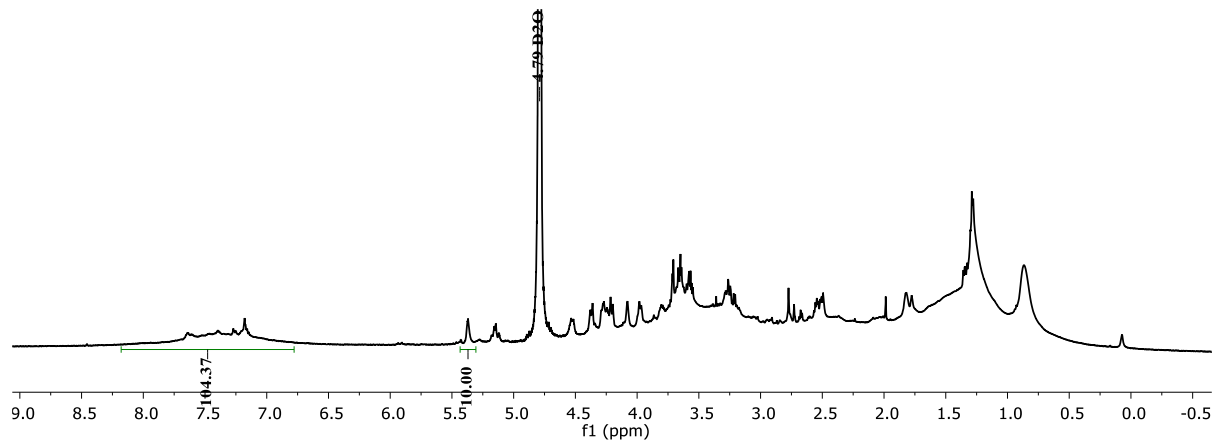


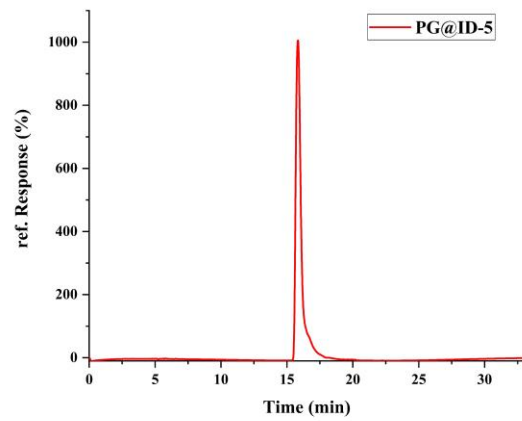
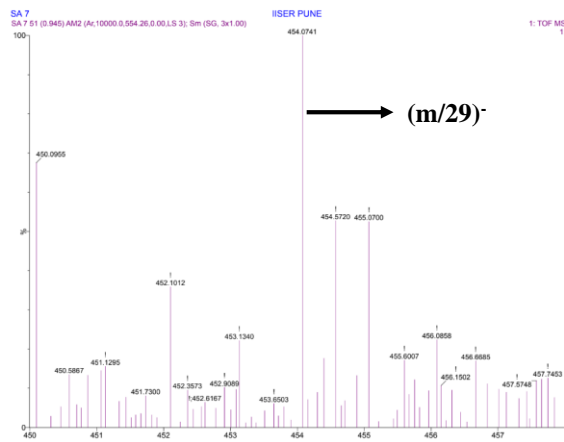


# PG@ID-4

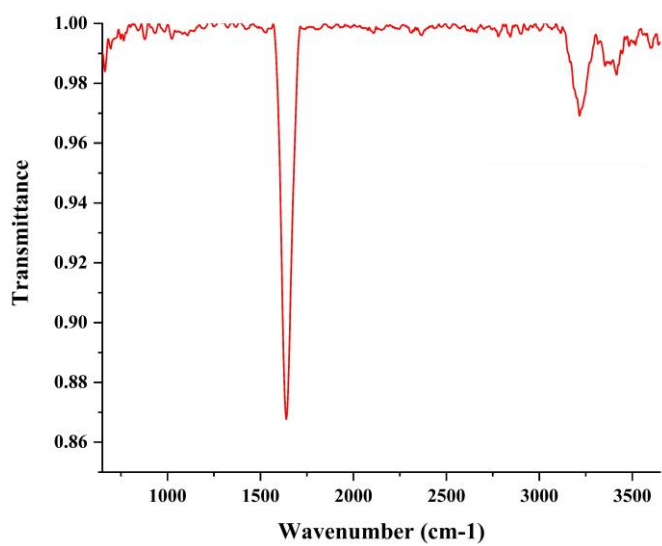
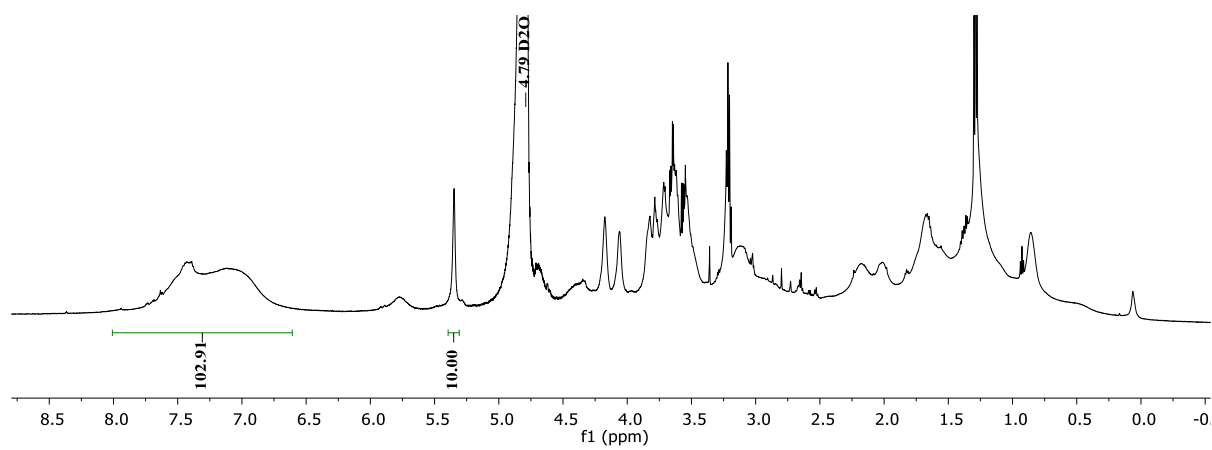


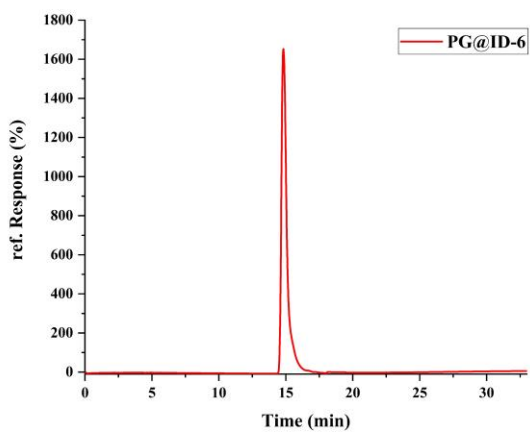
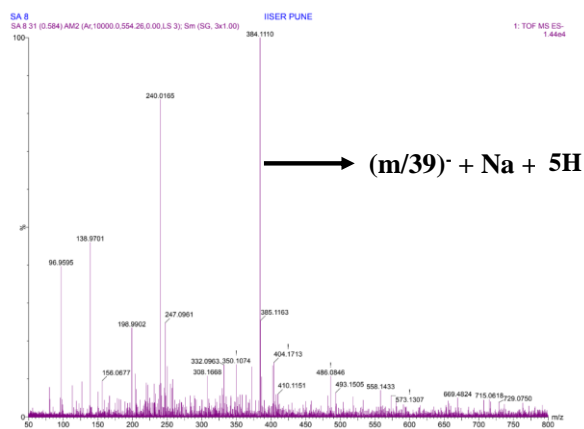
PG@ID-5



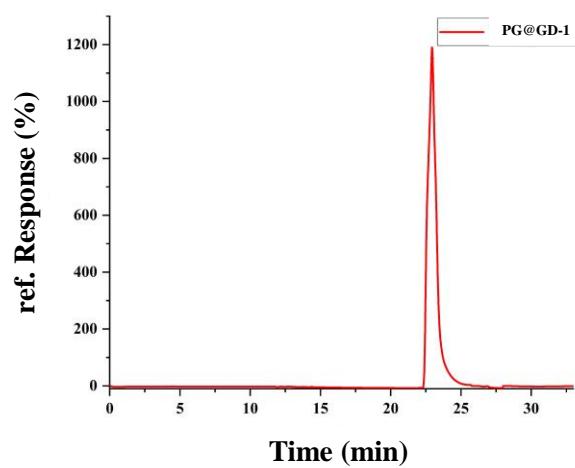
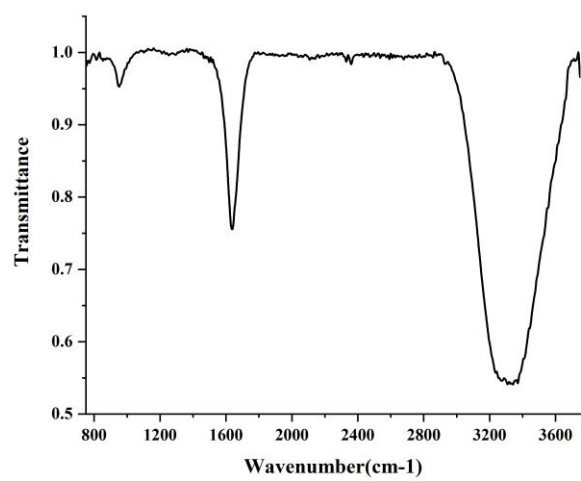
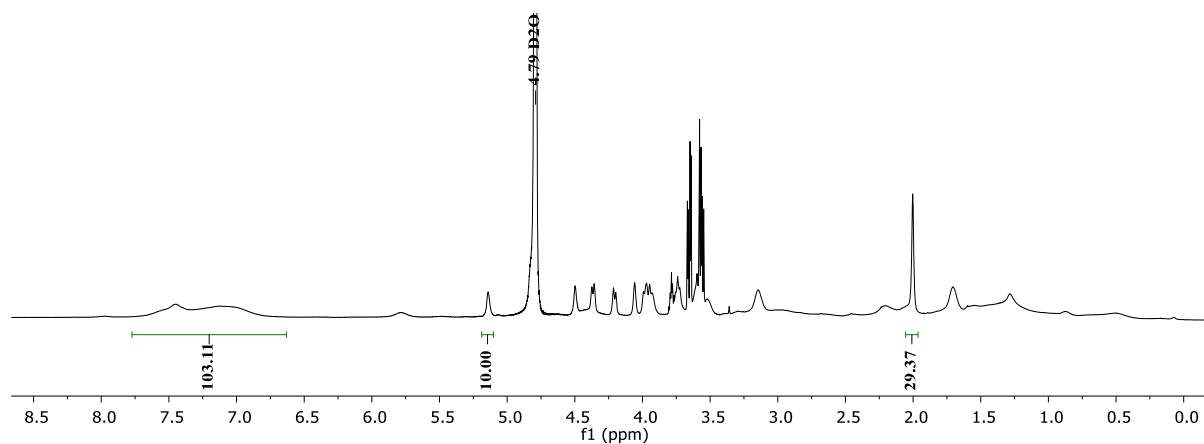


**PG@ID-6**

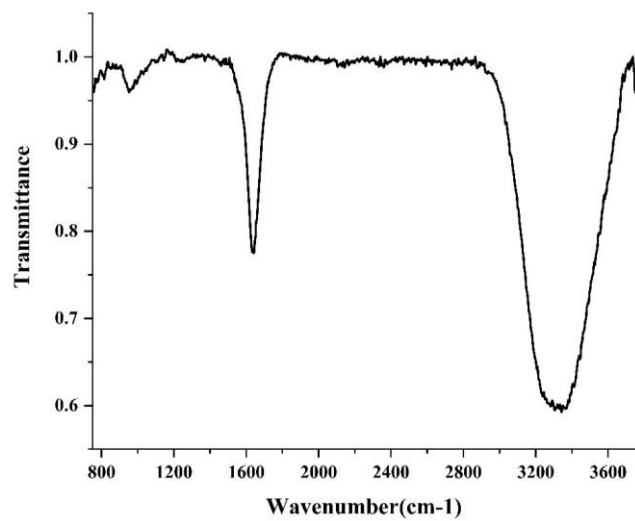
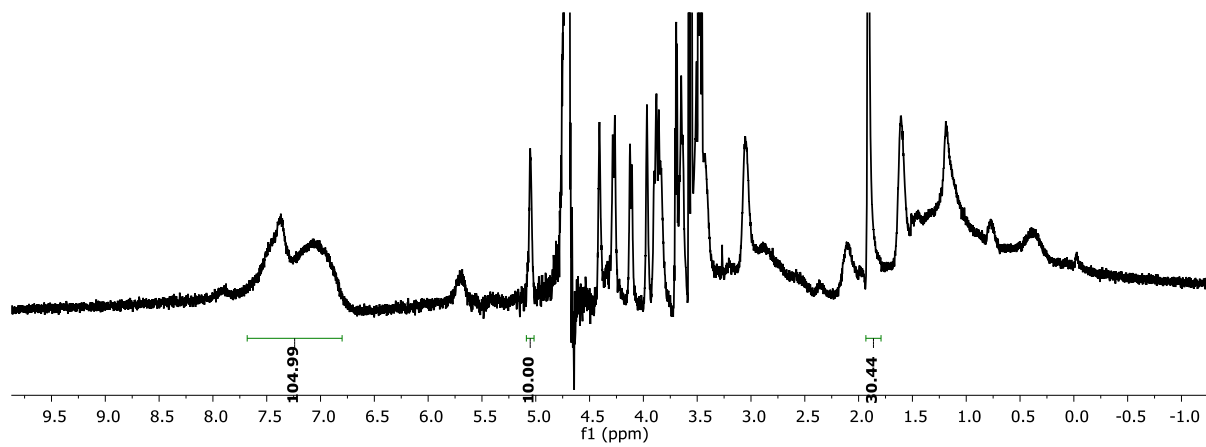




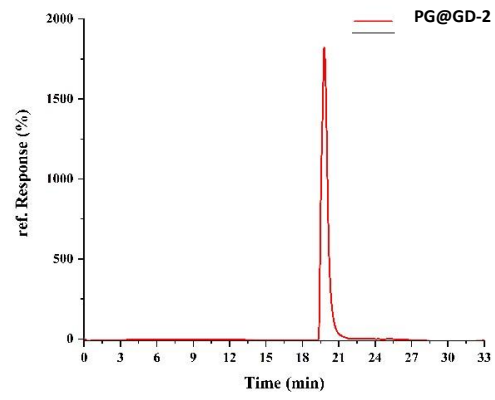
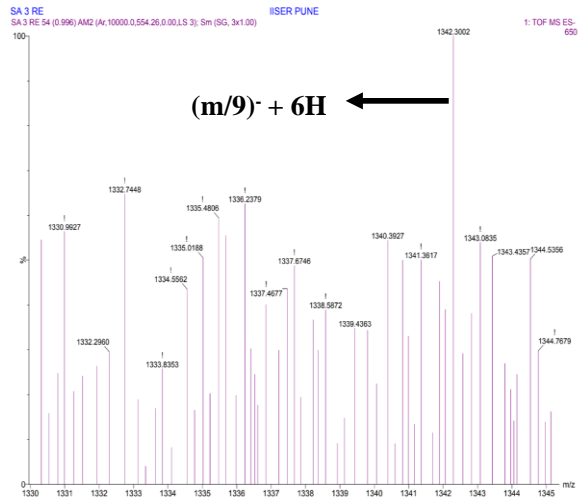
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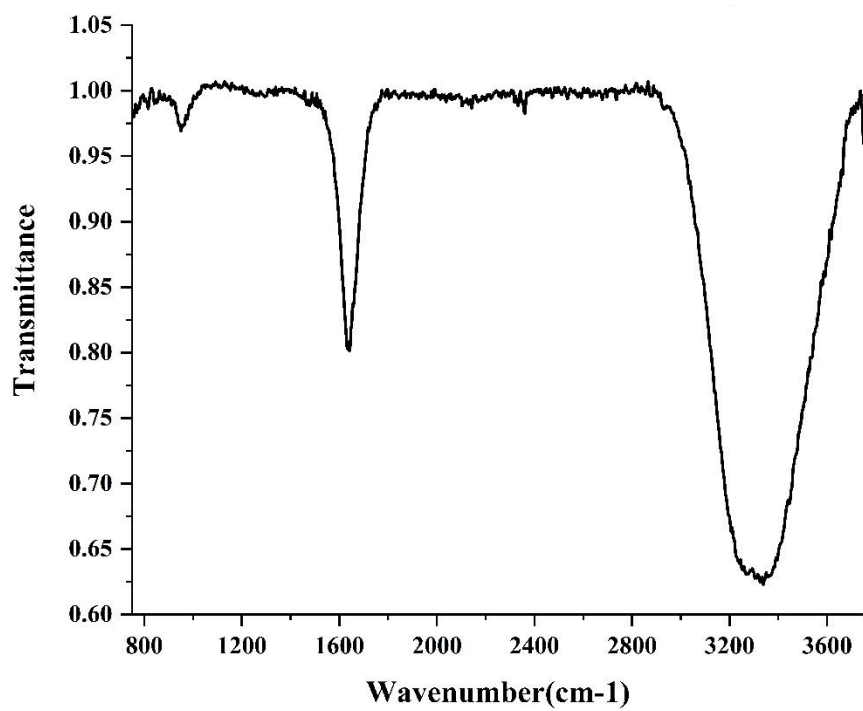
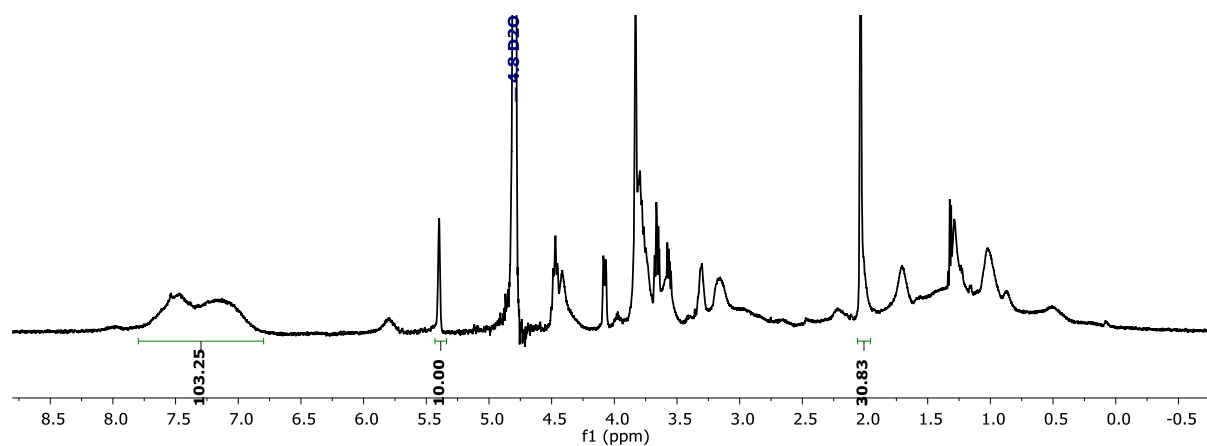
PG@GD-2

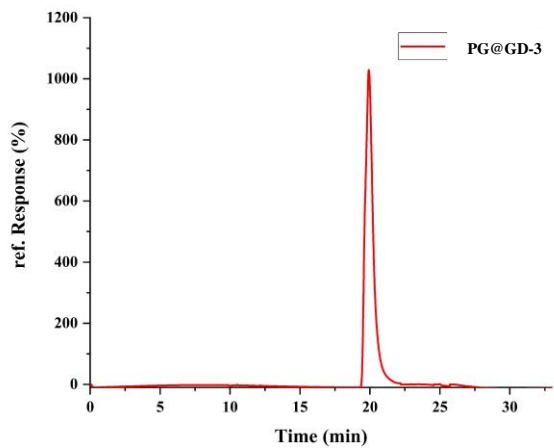
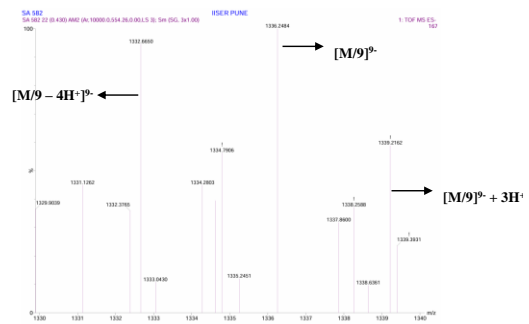




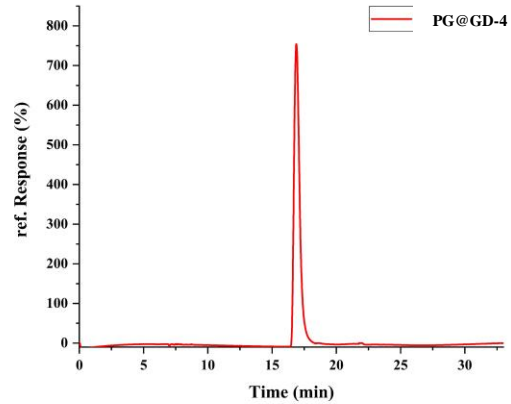
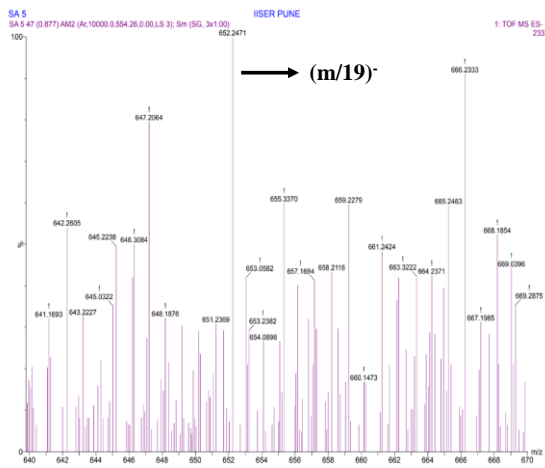
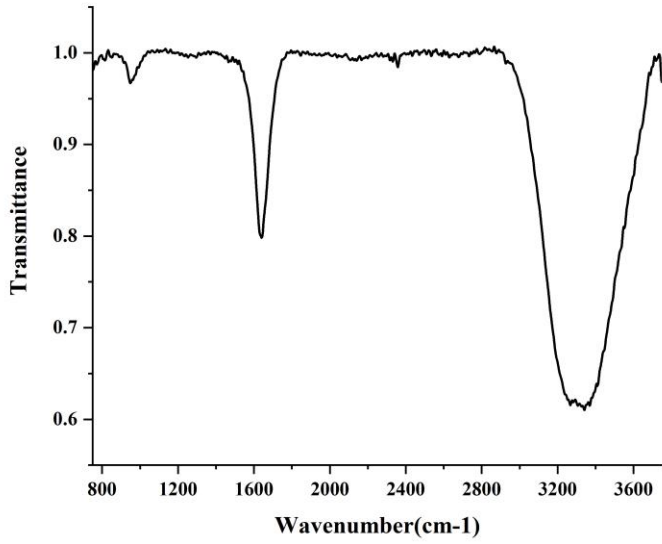
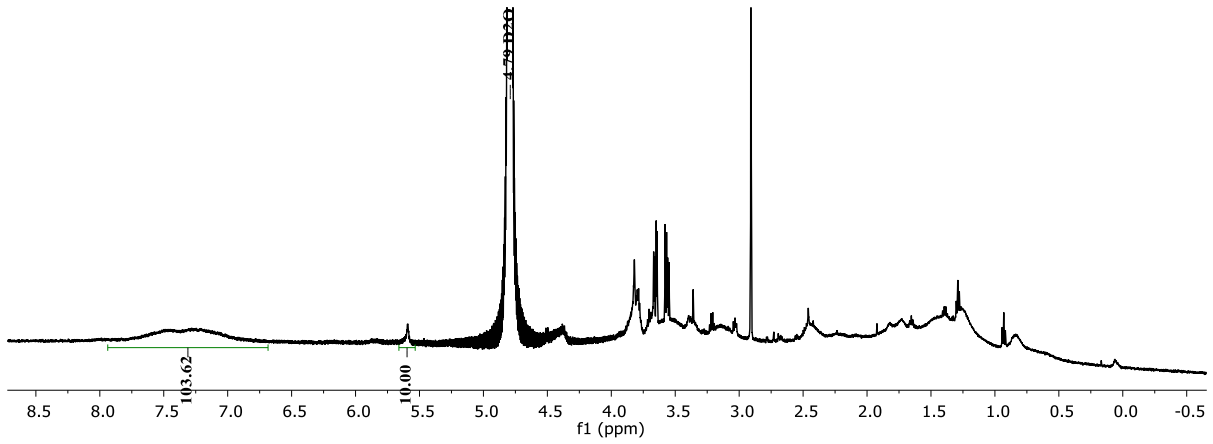


PG@GD-3





# PG@GD-4



PG@GD-5

