

Supplementary Information

Potent inhibitors of the human RNA ligase Rlig1 highlights its role in RNA integrity maintenance under oxidative cellular stress

Lisa A. Schlor^{a,b}, Maya Peußner^a, Silke Müller^{c,d}, Andreas Marx^{a,b}

a Department of Chemistry, University of Konstanz, Universitätsstraße 10, 78457 Konstanz, Germany. E-mail: andreas.marx@uni-konstanz.de

b Konstanz Research School Chemical Biology, University of Konstanz, Universitätsstraße 10, 78457 Konstanz, Germany.

c Department of Biology, University of Konstanz, Universitätsstraße 10, 78457 Konstanz, Germany.

d Screening Center, University of Konstanz, Universitätsstraße 10, 78457 Konstanz, Germany.

Table of contents

1	Supplementary figures	4
2	Supplementary tables	10
3	Methods	15
3.1	Biochemical experiments	15
	Plasmid construction	15
	Expression and purification of recombinant proteins	16
	High-throughput screening of inhibitors for Rlig1-AMP	17
	Preparation of 5' ³² P-labelled oligonucleotides	18
	Radioactive ligation assays	18
	Urea-PAGE and autoradiographic imaging	19
3.2	Cell culture	20
	Cell viability assay of HEK293 cells treated with inhibitors	20
	Cell viability assay of HEK293 cells treated with inhibitors and menadione	20
	Light-microscopy of inhibitor- and menadione-treated HEK293 cells	21
	RNA integrity analysis of HEK293 cells treated with inhibitors and different concentrations of menadione	21
3.3	Chemical synthesis	22
	General experimental details	22
	Analytical methods	22
	General procedures	23
	General procedure A: Boc-protection of aryl amines	23
	General procedure B: Buchwald-Hartwig amination	24
	General procedure C: reduction of nitroaryls	36
	General procedure D: ester hydrolysis	43
	General procedure E: amide coupling	46
	Synthesis of truncated SGI-1027 structures	64
4	Supplementary references	67
5	Appendix	67

Amino acid sequence of expressed proteins	67
Sequence of plasmids used in this study.....	67
NMR Spectra	70

1 Supplementary figures

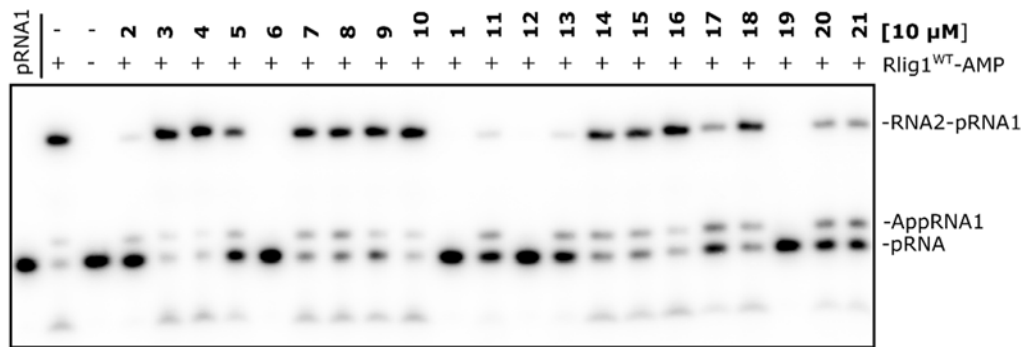


Fig. S1: Hit validation of 21 compounds derived from high-throughput screening in a radioactive RNA ligation assay. PAGE analysis of reaction products after incubation with Rlig1 and 10 μM of each compound at 37°C for 20 minutes (representative image of n = 2).

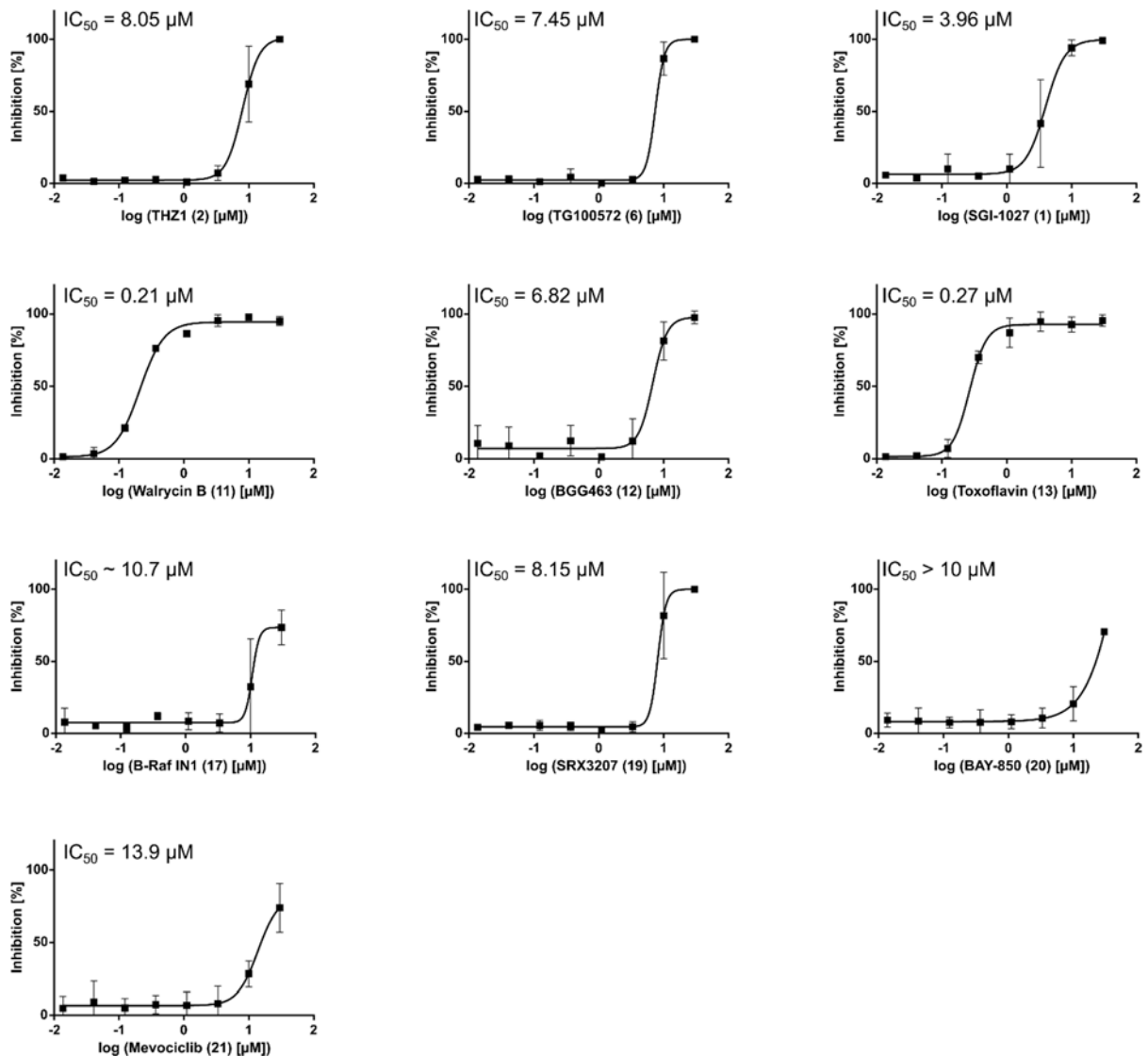


Fig. S2: Dose-response curves of active hits *in vitro* against Rlig1-AMP. The IC₅₀ values were calculated by a non-linear curve fit/dose-response curve in GraphPad Prism. All data represent the mean value for three biological replicates. Error bars represent ± SEM. Source data are provided as a Source Data file.

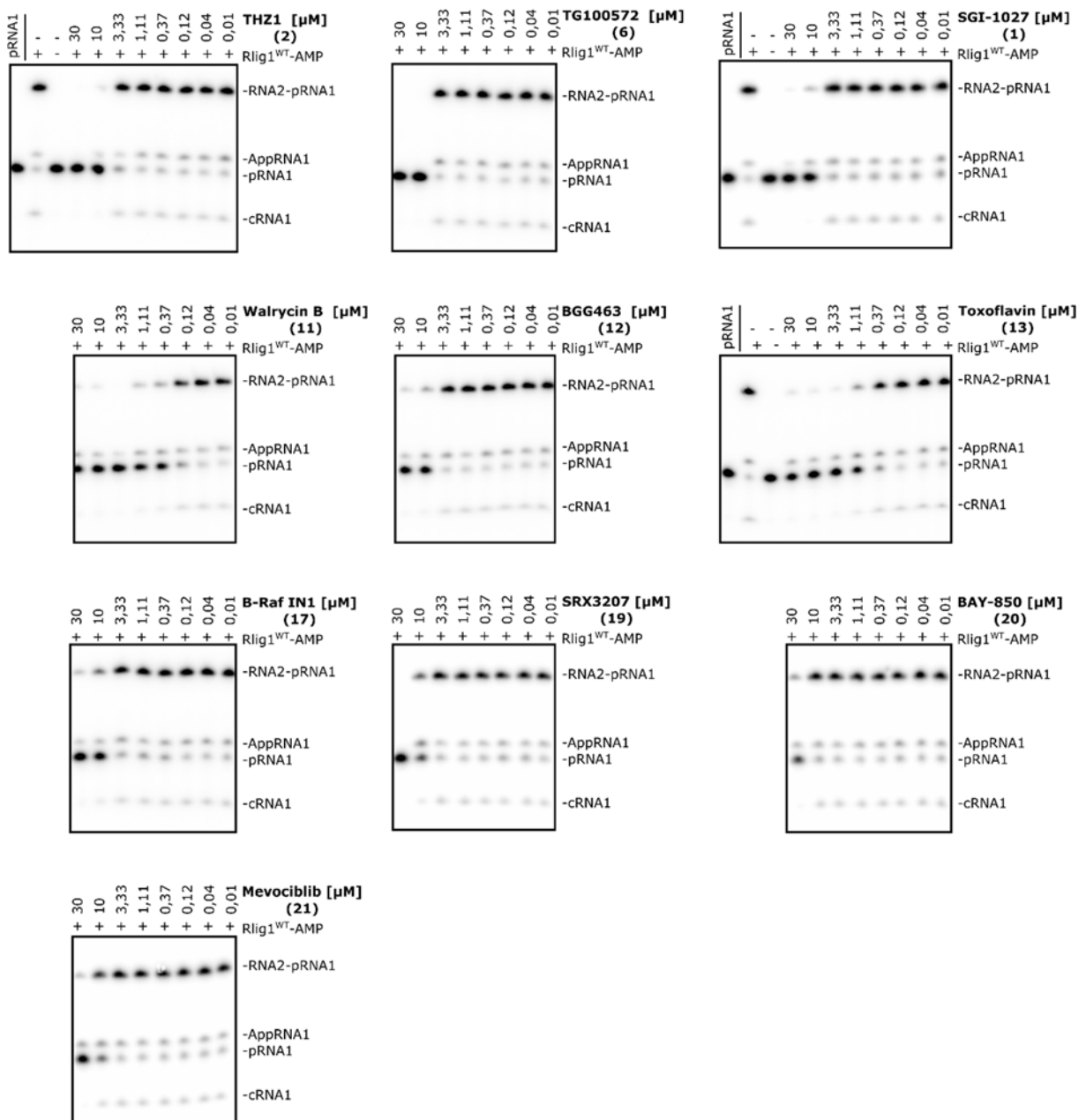


Fig. S3: Urea-PAGE analysis of dose-response experiment for active MCE hits (representative image of $n = 3$). RNA ligation reactions were performed with 500 nM Rlig1-AMP and 50 nM pRNA for 20 min at 37 °C at varying concentrations of compounds (30 – 0.01 μM). The rate of RNA-pRNA formation was quantified by the intensities of the bands for each chamber and normalized to the positive control.

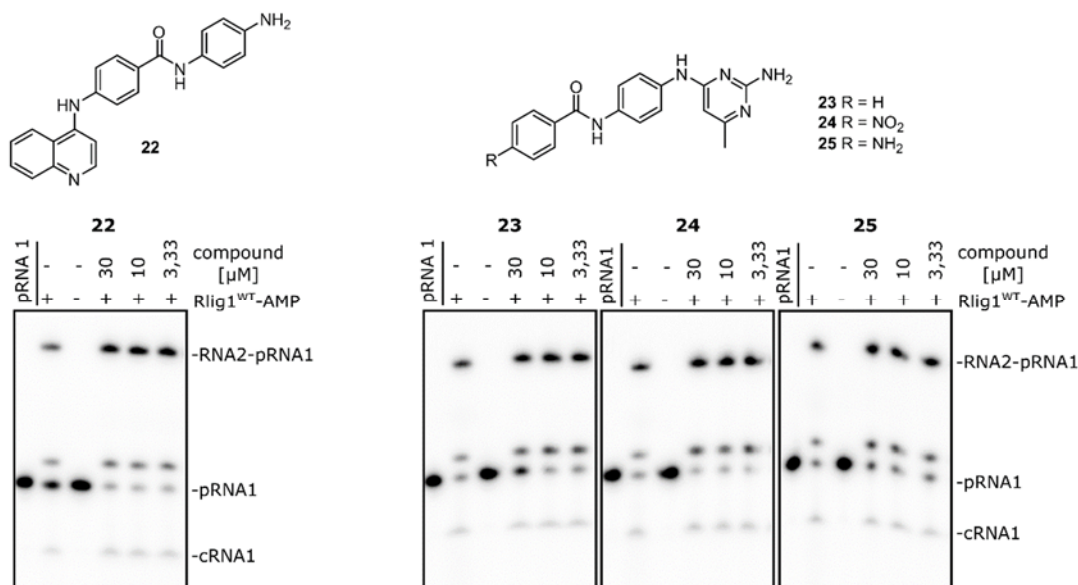


Fig. S4: Structure of the synthesized truncated SGI-1027 derivatives **22-25** and biological evaluation by urea-PAGE analysis. RNA ligation reactions were performed with 500 nM Rlig1-AMP and 50 nM pRNA1 for 20 min at 37 °C at varying concentrations of compounds (30, 10, 3.33 μM).

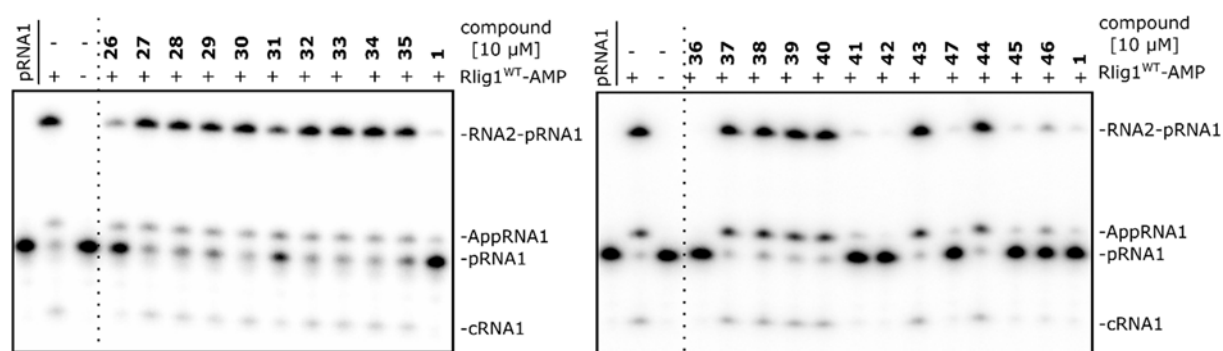


Fig. S5: Non-cropped PAGE analysis depicted in **Fig. 3B**. RNA ligation reactions were performed with 500 nM Rlig1-AMP and 50 nM pRNA1 for 20 min at 37 °C at a constant compound concentration of 10 μM.

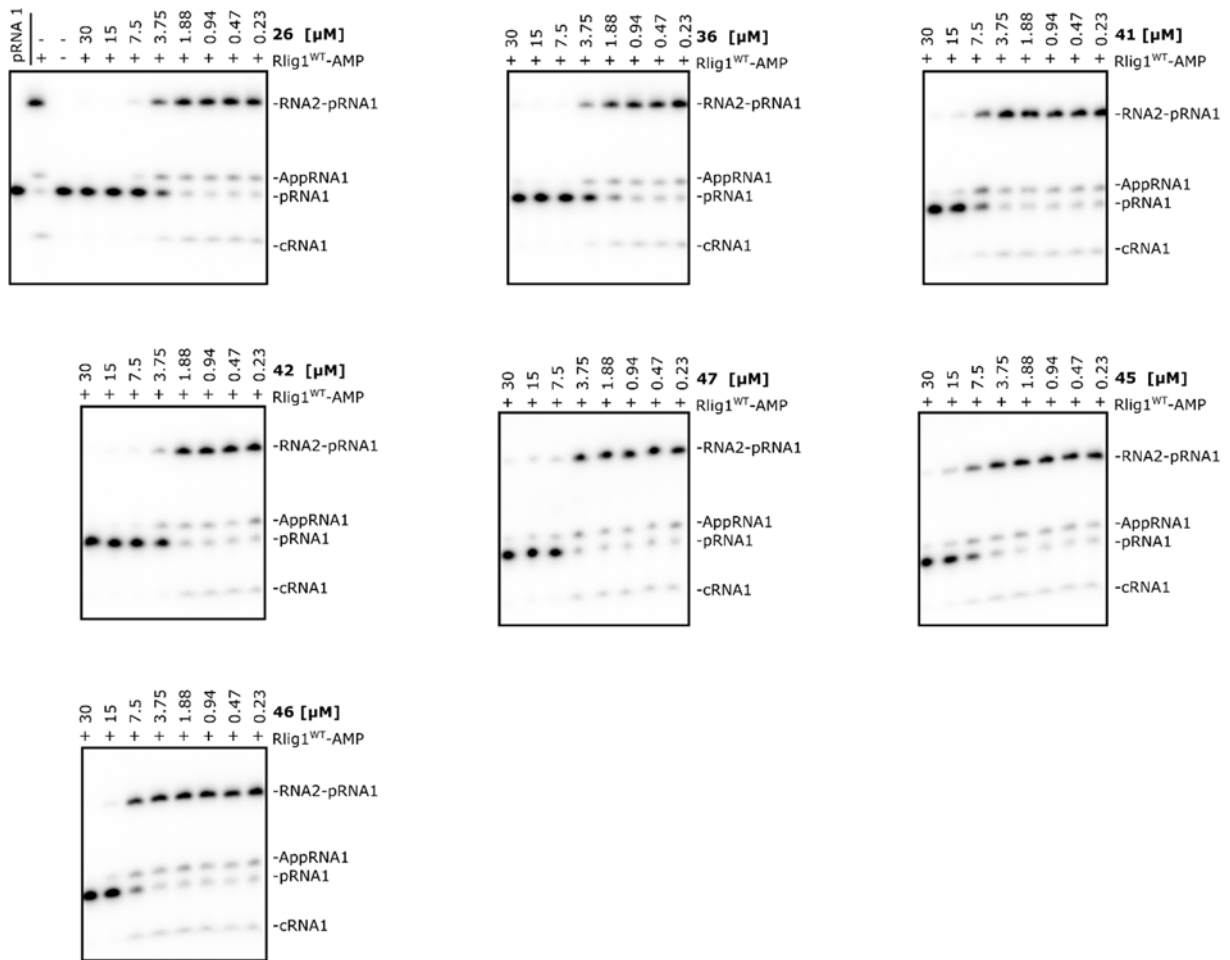


Fig. S6: Urea-PAGE analysis of dose-response experiment for active SGI-1027 derivatives (representative image of $n = 3$). RNA ligation reactions were performed with 500 nM Rlig1-AMP and 50 nM pRNA1 for 20 min at 37 °C at varying concentrations of compounds (30 – 0.23 μM). The rate of RNA2-pRNA1 formation was quantified by the intensities of the bands for each chamber and normalized to the positive control. Source data are provided as a Source Data file.

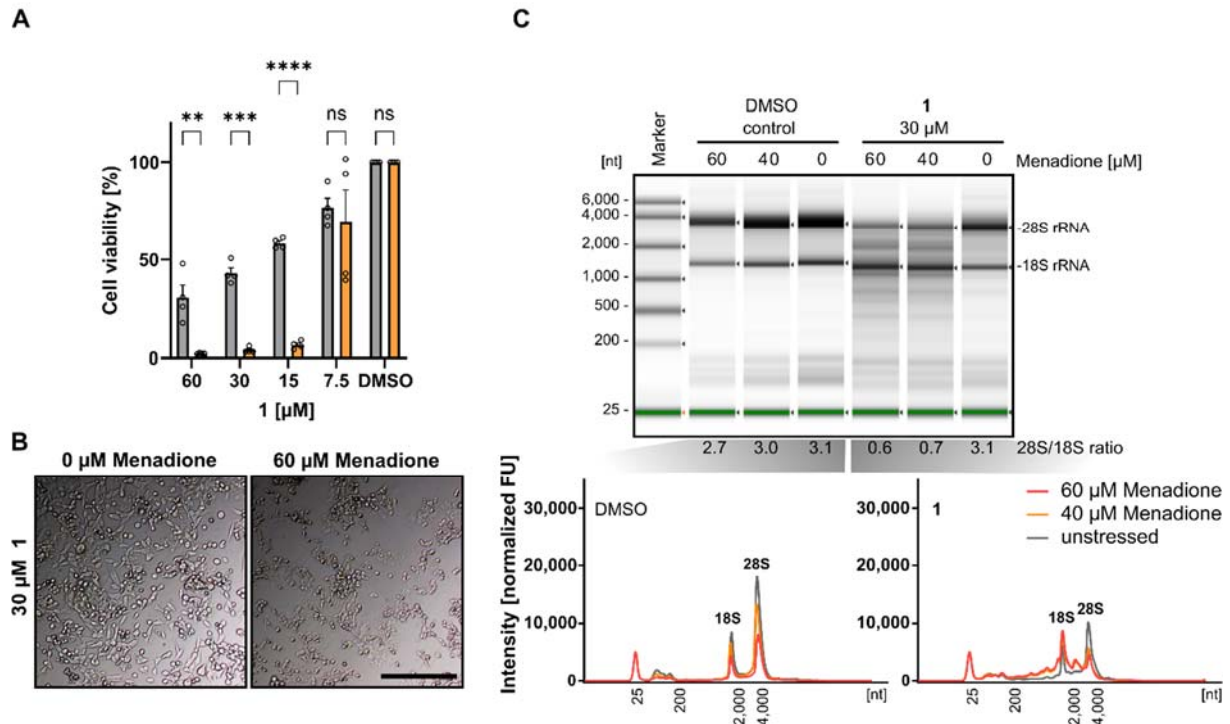


Fig. S7: A) Comparison of cell viability of HEK293 cells treated with compound **1** with and without 5-hour treatment with 40 μM menadione. Percentage cell viability was normalized to HEK293 cells treated without inhibitor or menadione. Hollow circles represent individual data points for n = 4 biological replicates. Error bars represent ± SEM. Statistical significance was calculated using two-way ANOVA with Sidak's multiple comparisons test: ^{ns}P > 0.05; *P ≤ 0.05; **P ≤ 0.01, ***P ≤ 0.001, ****P < 0.0001. B) Light microscopy of HEK293 treated with 30 μM **1**, followed by 5-hour treatment of 60 μM menadione. Scale bar is 500 μm. C) Analysis of the total RNA from cell extract of HEK293 cells treated with compound **1** for 24 h, followed by treatment with 40 μM and 60 μM menadione. RNA was isolated and analyzed using TapeStation (version 4.1.1). (top) Separation profile of analyzed samples. The ratio of 28S and 18S rRNA intensities is listed below the profile. (bottom) Electropherograms of total RNA from cells treated with DMSO as control and compound **1** at different menadione concentrations. The 18S/28S peaks are annotated. Experiments were conducted in duplicate. Source data are provided as a Source Data file.

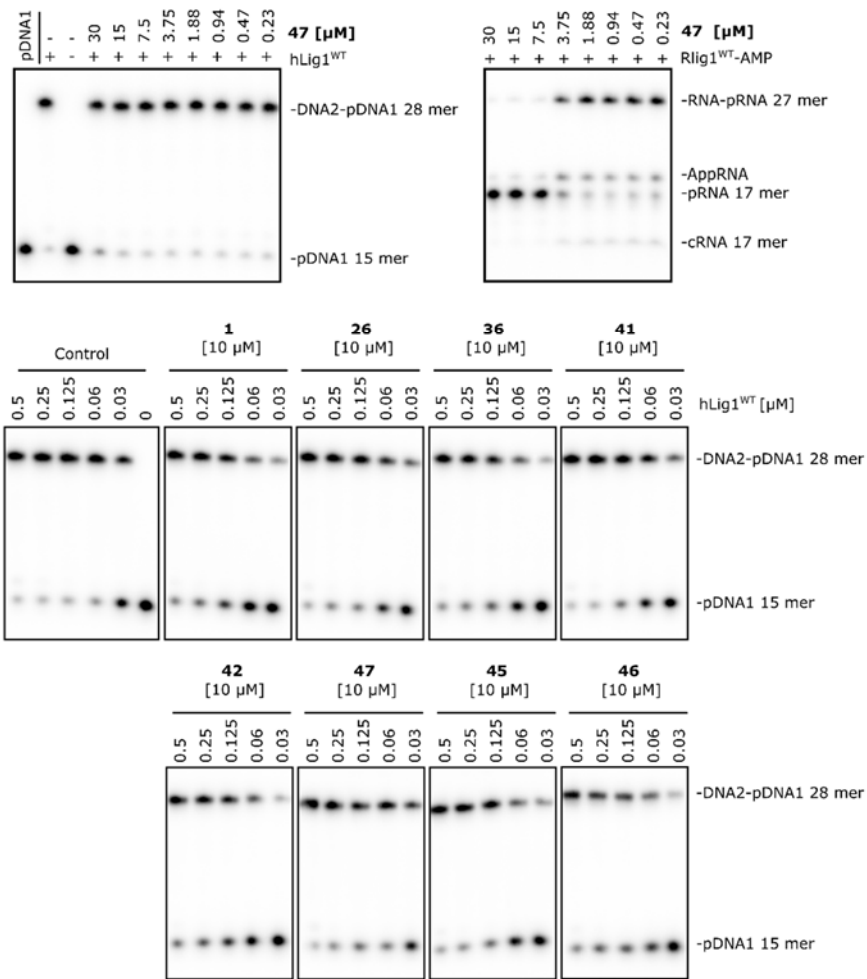


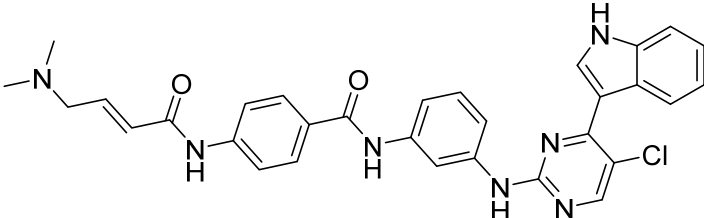
Fig. S8: (top) Urea-PAGE analysis of comparison of DNA ligation with hLig1 and RNA ligation with Rlig1 across various concentrations of compound **47**. Reactions were performed for 20 min at 37 °C. (bottom) Urea-PAGE analysis of DNA ligation turnover at varying hLig1 concentrations with a constant 10 μM concentration of all active SGI-1027 derivatives. Reactions were performed for 30 min at 37 °C (representative images, n = 3).

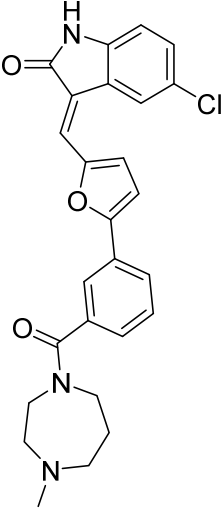
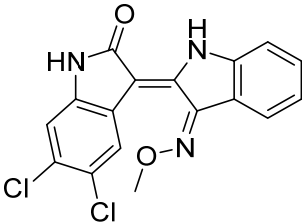
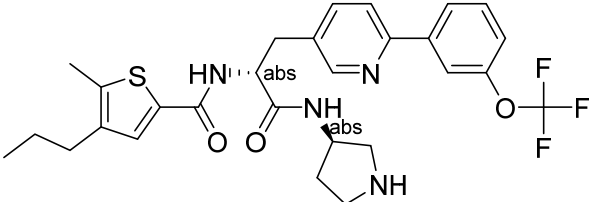
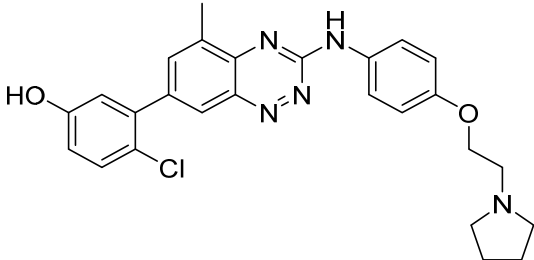
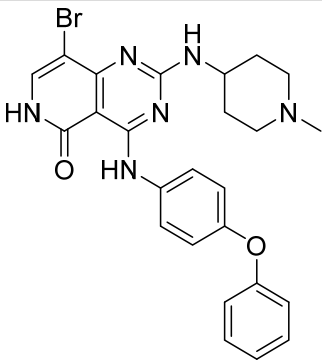
2 Supplementary tables

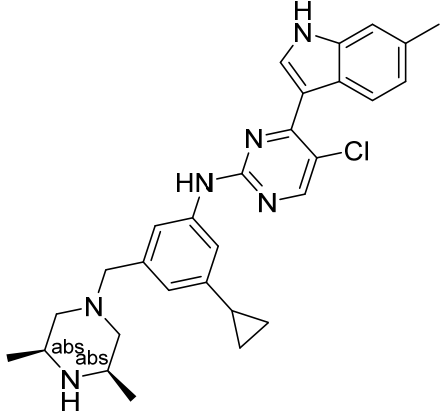
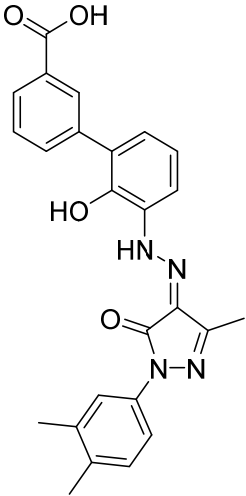
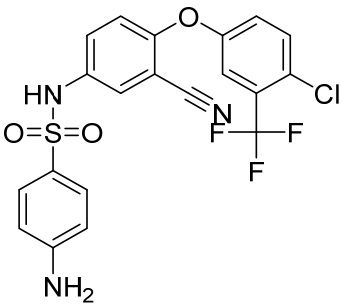
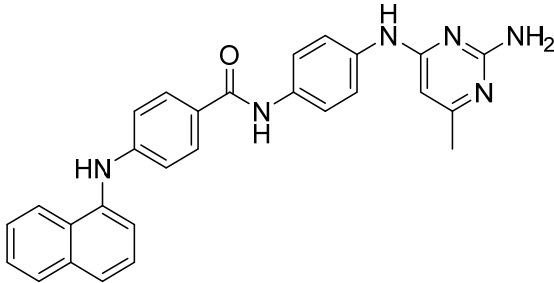
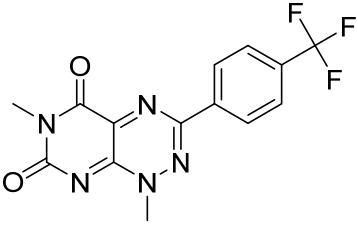
Table S1: Overview of all compound plates that were tested in HTS with corresponding Z' values.

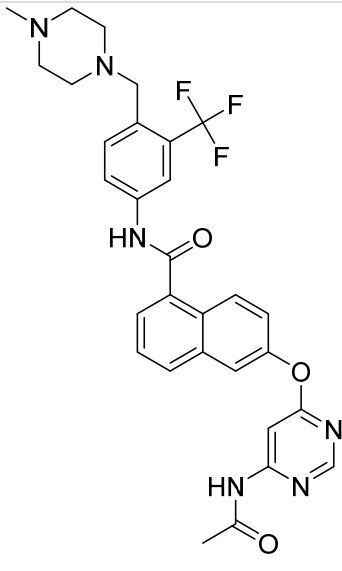
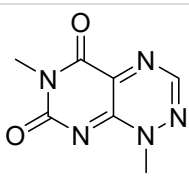
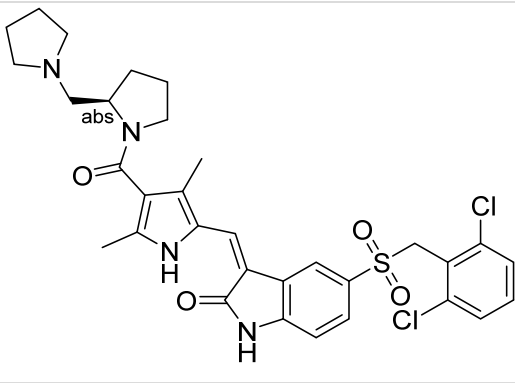
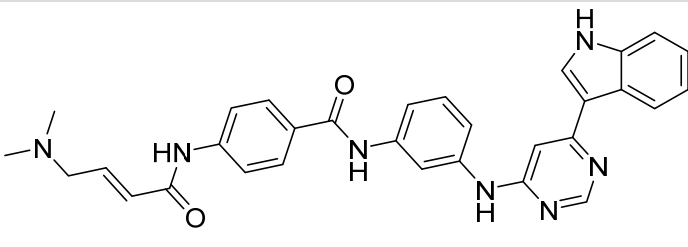
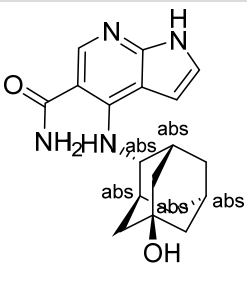
Plates	Filename-timepoint	Z'-Factor	Plates	Filename-timepoint	Z'-Factor
1	220704023HYCPK27668b-2	0.719	24	220714035HYCPK27691b-2	0.767
2	220704025HYCPK27669b-2	0.711	25	220714037HYCPK27692b-2	0.758
3	220704027HYCPK27670b-2	0.716	26	220714039HYCPK27693b-2	0.738
4	220704029HYCPK27671b-2	0.817	27	220714041HYCPK27694b-2	0.731
5	220704031HYCPK27672b-2	0.727	28	220714042HYCPK27695b-2	0.825
6	220704033HYCPK27673b-2	0.692	29	220718025HYCPK27696b-2	0.609
7	220704035HYCPK27674b-2	0.778	30	220718027HYCPK27697b-2	0.692
8	220704036HYCPK27675b-2	0.708	31	220718029HYCPK27698b-2	0.666
9	220707025HYCPK27676b-2	0.678	32	220718031HYCPK27699b-2	0.685
10	220707027HYCPK27677b-2	0.718	33	220718033HYCPK27700b-2	0.730
11	220707029HYCPK27678b-2	0.731	34	220718035HYCPK27701b-2	0.745
12	220707031HYCPK27679b-2	0.725	35	220718037HYCPK27702b-2	0.736
13	220707033HYCPK27680b-2	0.740	36	220718039HYCPK27703b-2	0.670
14	220707035HYCPK27681b-2	0.685	37	220718041HYCPK27704b-2	0.719
15	220707037HYCPK27682b-2	0.697	38	220718042HYCPK27705b-2	0.732
16	220707039HYCPK27683b-2	0.712	39	220720023HYCPK27706b-2	0.698
17	220707041HYCPK27684b-2	0.711	40	220720025HYCPK27707b-2	0.722
18	220707042HYCPK27685b-2	0.682	41	220720027HYCPK27712b-2	0.790
19	220714025HYCPK27686b-2	0.799	42	220720029HYCPK27713b-2	0.754
20	220714027HYCPK27687b-2	0.673	43	220720031HYCPK27714b-2	0.787
21	220714029HYCPK27688b-2	0.717	44	220720033HYCPK27715b-2	0.720
22	220714031HYCPK27689b-2	0.741	45	220720035HYCPK27716b-2	0.743
23	220714033HYCPK27690b-2	0.742	46	220725017HYCPK27667b-2	0.681

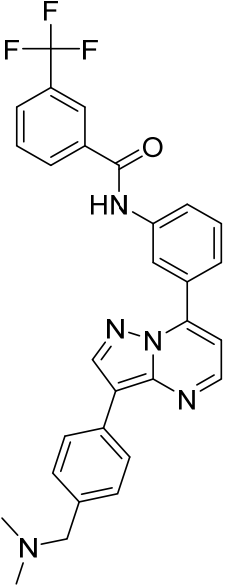
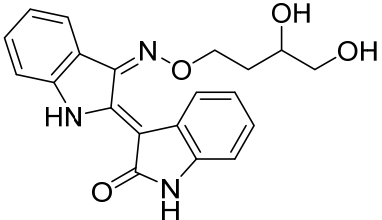
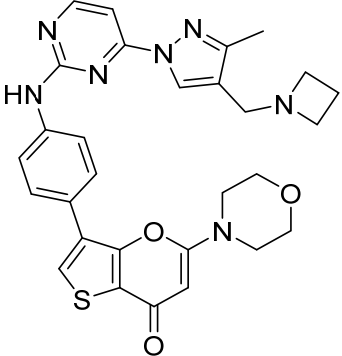
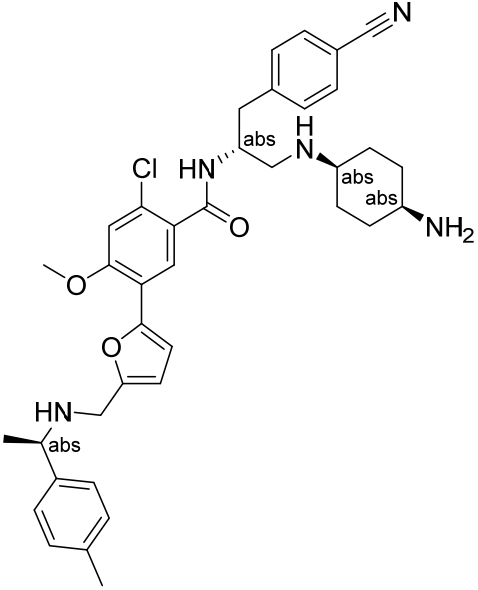
Table S2: List of initial hits from the high-throughput screening with their biological targets.

Compound	Name	Structure	Targets
2	THZ1 (Hydrochloride)		CDK

3	CX-6258		Pim
4	KY19382		GSK-3; Wnt; β -catenin
5	AZ82		Kinesin
6	TG 100572 (Hydrochloride)		FGFR; PDGFR; Src; VEGFR HCl
7	G-749		Apoptosis; FLT3

8	HM43239		Apoptosis; FLT3
9	Eltrombopag		Bacterial; Thrombopoietin Receptor
10	Lp-PLA2-IN-3		Phospholipase
1	SGI-1027		Apoptosis; DNA Methyltransferase
11	Walrycin B		Antibiotic; Bacterial

12	BGG463		CDK
13	Toxoflavin		Antibiotic; Bacterial; β-catenin
14	PHA-665752		Apoptosis; Autophagy; c- Met/HGFR
15	THZ-P1-2		Autophagy
16	Peficitinib		JAK

17	B-Raf IN 1	 <p>The structure shows a central pyridine ring substituted at the 2-position with a 1,2,4-triazole ring. The 4-position of the pyridine is substituted with a 4-(dimethylamino)phenyl group. The 5-position of the pyridine is substituted with an amide group (-NH-CO-), which is further linked to a 4-(trifluoromethyl)phenyl group.</p>	Raf
18	Indirubin Derivative E804	 <p>The structure is a dimeric indirubin derivative. It features two indole-like rings fused together. One ring has a carbonyl group and an NH group. The other ring has a nitrogen atom connected to a propyl chain with two hydroxyl groups (-OH).</p>	IGF-1R
19	SRX3207	 <p>The structure is a complex heterocyclic molecule. It features a central benzothiazine-like core with a sulfur atom and a carbonyl group. It is substituted with a morpholine ring, a pyridine ring, and a 1,2,4-triazole ring. A methyl group and a pyrrolidine ring are also present.</p>	PI3K; Syk
20	BAY-850	 <p>The structure is a complex molecule with multiple stereocenters. It features a central benzothiazine-like core with a sulfur atom and a carbonyl group. It is substituted with a morpholine ring, a pyridine ring, and a 1,2,4-triazole ring. A methyl group and a pyrrolidine ring are also present. The stereocenters are labeled 'abs'.</p>	Epigenetic Reader Domain

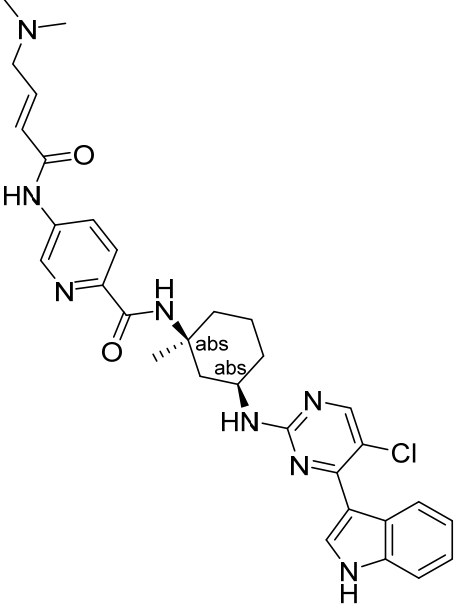
21	Mevociclib		CDK
----	------------	--	-----

Table S3: List of oligonucleotide sequences used in this study. All oligonucleotides were obtained from Biomers or Integrated DNA Technologies (IDT).

Oligonucleotide	Sequence 5' - 3'
RNA1	GGC ACU CAG ACU CAG AG
RNA2	CUC UGA GUA A
RNA1-BHQ1	GGC ACU CAG ACU CAG AG-BHQ1
RNA2-FAM	FAM-CUC UGA GUA A
DNA1	GTC GGA CTG ATT CGG
DNA2	GTG CTG ATG CGT C
DNA3	CCG AAT CAG TCC GAC GAC GCA TCA GCA C

3 Methods

3.1 Biochemical experiments

Biological assays were adapted from previously described procedures.¹

Plasmid construction

The plasmid construct of pET15b-Rlig1 was obtained from Yizhi Yuan and transformed into *Escherichia coli* (*E. coli*) BL21 (DE3) competent cells. The gene fragment of hLig1 bearing 5'-NdeI and 3'-XhoI restriction cleavage sites was synthesized by Integrated DNA technologies. The gene fragment was ligated into pJET1.2/blunt vector using CloneJET PCR Cloning Kit for amplification. The NdeI and XhoI restriction digested gene fragments and pET28a vector were isolated and ligated. The plasmid construct of pET28a-hLig1 was

transformed in Rosetta™ BL21 (DE3) *E. coli* cells. Sequence information is listed in the appendix.

Expression and purification of recombinant proteins

Rlig1-AMP

Rlig1-AMP was expressed and purified from a pET15b vector in BL21 (D3) *E. coli* cells with an N-terminal His-tag according to the protocol developed by Yuan *et al.*¹ Cells were cultured in LB with 100 mg/L carbenicillin overnight at 37°C and 220 rpm. 1 L of LB_{carb} was inoculated with the overnight culture to an OD₆₀₀ of 0.1 and incubated at 37°C and 220 rpm until an OD₆₀₀ of 0.6 was reached. Gene expression was induced with 1 mM IPTG and incubated at 20 °C for 18 hours and 220 rpm. The cells were harvested by centrifugation (4000 rpm, 30 min, 4 °C), resuspended in lysis buffer (50 mM Tris-HCl pH 8.0, 150 mM NaCl, 20 mM imidazole, 1 mM DTT, 0.1% Triton-X100, 1 mg/mL Pefabloc, 1 µg/µL Aprotinin/Leupeptin) and lysed by sonication (5 cycles, 20% intensity, 45 sec) on ice. The lysate was centrifuged (18000 rpm, 30 min, 4 °C) and the supernatant was filtered through a 0.45 µm syringe filter. The AMPylated His₆-Rlig1 was purified using a 5 mL HisTrap™ FF crude column with a step gradient of 10% buffer B (buffer A: 50 mM Tris-HCl pH 8.0, 150 mM NaCl, 20 mM imidazole, 1 mM DTT; buffer B: 50 mM Tris-HCl pH 8.0, 150 mM NaCl, 500 mM imidazole, 1 mM DTT). Fractions were analyzed by SDS-PAGE and fractions containing AMPylated His₆-Rlig1 were pooled and dialyzed against 50 mM Tris-HCl pH 8.0, 100 mM NaCl, 1mM DTT at 4 °C overnight. The protein was further purified using a 5 mL HiTrap™ Q HP column (buffer A: 50 mM Tris-HCl pH 8.0, 100 mM NaCl, and 1.0 mM DTT; buffer B: 50 mM Tris-HCl, 1000 mM NaCl, and 1.0 mM DTT). Fractions were analyzed by SDS-PAGE, and those containing only Rlig1-AMP were pooled, concentrated, and stored at -20 °C in storage buffer containing 50 mM Tris-HCl pH 8.0, 100 mM NaCl, 1 mM DTT and 50% (v/v) glycerol.

hLig1

hLig1 was expressed and purified from a pET28a vector in Rosetta™ BL21 (DE3) *E. coli* cells with an N-terminal His-tag adapted from the protocol of Pascal *et al.*² Cells were cultured in LB with 100 mg/mL kanamycin overnight at 37°C and 220 rpm. 1 L of LB_{kana} was inoculated with the overnight culture to an OD₆₀₀ of 0.1 and incubated at 37°C and 220 rpm until an OD₆₀₀ of 0.6 was reached. Gene expression was induced by the addition of 0.5 mM IPTG and the culture was incubated at 30°C for four hours and 220 rpm. The cells were harvested by centrifugation (4000 rpm, 30 min, 4°C), resuspended in 20 mL lysis buffer (50 mM Tris-HOAc [pH 7.5], 100 mM NaCl, 10% glycerol, 1 mM EDTA, 0.1% Triton-X100, 1mM DTT, 10 mM imidazole, 25µL/mL lysozyme, 1 mg/mL Pefabloc, 1 µg/µL Aprotinin/Leupeptin) and lysed by sonication (5 cycles, 18% intensity, 45 sec) on ice. The lysate was centrifuged (18000 rpm,

30 min, 4°C) and the supernatant was filtered through a 0.45 µm syringe filter. His₆-hLig1 was purified using a 5 mL HisTrap™ FF crude column with a gradient of 0% to 100% buffer B (buffer A: 50 mM Tris-HCl pH 8.0, 150 mM NaCl, 20 mM imidazole, 1 mM DTT; buffer B: 50 mM Tris-HCl pH 8.0, 150 mM NaCl, 500 mM imidazole, 1 mM DTT). Fractions containing His₆-hLig1 were pooled, concentrated and rebuffered to SEC buffer (50 mM Tris-HCl [pH 8], 100 mM NaCl, 1 mM DTT). The protein was further purified with SEC (HiLoad 16/600 Superdex 200 pg, GE Healthcare) using SEC buffer. Fractions containing His₆-hLig1 were pooled, concentrated and stored at -80°C in 25 mM Tris-HCl [pH 8.0], 50 mM NaCl, 0.5 mM DTT and 50% (v/v) glycerol.

High-throughput screening of inhibitors for Rlig1-AMP

The bioactive chemical library was purchased from MedChemExpress (Art.-Nu.: HY-L001) and screened using a fully automated Freedom evo screening unit from Tecan. His₆-Rlig-AMP, which was recombinantly expressed in *E. coli*, was diluted to 750 nM in 1x RNA ligation buffer (50 mM Tris-HCl [pH 7.1], 5 mM MgCl₂ and 1 mM DTT). 40 µL of the cooled enzyme solution was dispensed into all wells of columns 1-23 of a 384-well plate (white, non-binding, Greiner) using a multichannel arm (MCA) and an MCA 384 adapter for disposable tips. Column 24 was filled with 40 µL of the same buffer without the enzyme using the four-channel liquid handling arm (LiHa). The library compounds were transferred in a final concentration of 10 µM into all wells of columns 1-24 of the plate using the MCA with a 384 fixed tips tool. The plate was incubated at 37 °C for 10 min with shaking at 8.5 Hz. 5'-Phosphorylated RNA1-BHQ1 (225 nM) and RNA2-FAM (150 nM) were dissolved in 1x RNA ligation buffer and subsequently cooled to 4°C. The BHQ1-labelled RNA was used in excess to ensure efficient quenching of the FAM-labelled strand. A 20 µL aliquot of this solution was dispensed into each well using a Multidrop 384 reagent dispenser (Thermo Scientific). The plate was incubated at 37 °C for 20 minutes. The reactions were stopped by adding 20 µL of an 80 mM EDTA solution in buffer containing 50 mM Tris-HCl (pH 7.1) and incubated at 37 °C for an additional 10 min. Fluorescence intensity was immediately measured using a Spark microplate reader (Tecan) with excitation at 495 nm and detection of fluorescence emission at 520 nm. The compounds were also treated under the same conditions without enzyme, and relative enzymatic activity was calculated as the difference in fluorescence intensity readouts between the two conditions and the controls. Compounds were considered hits if the inhibition was above 70%. Z'-factors were calculated for each plate as quality control, using the mean (µ) and standard deviation (σ) of the positive (c +, 100% activity) and negative (c -, 0% activity) controls. Data analysis was performed using the KNIME Analytics platform.

$$Z' = 1 - \frac{3\sigma_{c+} + 3\sigma_{c-}}{|\mu_{c+} - \mu_{c-}|}$$

Preparation of 5' ³²P-labelled oligonucleotides

RNA or DNA oligonucleotides were radioactively labelled by 5'-phosphorylation with [γ -³²P]-ATP. 1.0 μ M oligonucleotides were incubated with 0.4 u/ μ L of T4 polynucleotide kinase (PNK) and 200 μ M 0.555 MBq γ -³²P-ATP (185 TBq/mmol, Hartmann Analytic, SRP-401) in 1x T4 PNK reaction buffer at 37 °C for 1 h in a total volume of up to 50 μ L. The reaction was stopped by heating to 95 °C for 2 min. Unreacted ATP was removed by gel filtration using columns packed with 750 μ L Superdex™ G-25 Superfine (GE Healthcare) and centrifuged at 3300 rpm for 2 min. The gel filtration material was prepared by suspending 10 g Superdex™ G-25 Superfine in 75 mL H₂O containing 60 mg NaN₃. Prior to filtration, the packed gel filtration columns were washed with 400 μ L H₂O, to remove NaN₃.

Radioactive ligation assays

General RNA ligation assay

50 nM 5' ³²P-labelled RNA1 and 250 nM unlabelled RNA2 were incubated with 1 μ M or 500 nM Rlig1-AMP and 200 μ M ATP in 1x RNA ligation buffer (50 mM Tris-HOAc pH 7.0, 5 mM MgCl₂, and 1 mM DTT) at 37 °C for 20 min in a total volume of 10 μ L. The reactions were stopped by the addition of 90 μ L stopping solution (80% (v/v) formamide, 20 mM EDTA, 0.025% (w/v) bromophenol blue, and 0.025% (w/v) xylene cyanol) and heated at 95 °C for 2 min. 1 μ L samples were resolved by urea-PAGE and analyzed by autoradiographic imaging.

Hit validation assay and determination of IC₅₀ values

Hits from the HTS were purchased from MedChemExpress at a concentration of 10 mM in DMSO. For the hit validation assay, a 1 mM stock solution of compound dissolved in DMSO was used. For the concentration-dependent assay, the compounds were diluted in 1:2 steps, starting at 3 mM and ending at 0.02 mM in DMSO. Before the reaction, 1 μ L of the compound in DMSO was diluted in 19 μ L Milli-Q water. A 4 μ L sample of Mastermix (1.25 μ M His₆-Rlig1-AMP in 1.5x RNA ligation buffer with 200 μ M ATP) was mixed with 2 μ L of diluted compound or 5% DMSO as control. The mixture was preincubated at 37 °C for 10 min. Then, 4 μ L of Mastermix (125 nM RNA1, 625 nM RNA2 in 1x RNA ligation buffer) was added, and the mixture was incubated at 37 °C for 20 min. The reactions were stopped by the addition of stopping solution, resulting in 0.005 μ M 5'-³²P-RNA1, and heated at 95°C for 2 min. The samples were resolved using urea-PAGE and analyzed by autoradiographic imaging. Ligation turnover was quantified by ImageLab (Version 6.0.1) and the IC₅₀ value was calculated using a non-linear curve fit/ dose-response curve in GraphPad Prism (Version 10.2.3). All data represent the mean and standard error of triplicates.

DNA ligation assay with hLig1 and 47

Compound **47** was diluted in 1:2 steps, starting at 3 mM and ending at 0.02 mM in DMSO. Before the reaction, 1 μ L of the dilution series in DMSO was transferred into 19 μ L Milli-Q water. A 4 μ L sample of Mastermix [1.25 μ M hLig1, 500 μ M ATP in 1x DNA ligation buffer (50 mM Tris-HOAc [pH 7.5], 50 mM NaCl, 5 mM MgCl₂, 0.05 mg/mL BSA and 1 mM DTT)] and 2 μ L of compound dilution were mixed and preincubated at 37 °C for 10 min. 4 μ L of DNA Mastermix (125 nM pDNA1 , 250 nM DNA2, 250 nM DNA3 in 1x DNA ligation buffer) was added to the samples, followed by incubation at 37°C for 20 min. The reactions were stopped by adding 90 μ L stopping solution and heating at 95°C for 2 min. 1 μ L samples were resolved via urea-PAGE and analyzed by autoradiographic imaging. Ligation turnover was quantified using ImageLab, and product formation was plotted against the logarithmic compound concentration in GraphPad Prism.

DNA ligation assay with various hLig1 concentration at constant inhibitor concentration

Compounds dissolved in DMSO were 1:20 diluted in Milli-Q water to a concentration of 50 μ M. 6 μ L of Mastermix with various hLig1 concentration (5, 2.5, 1.25, and 0.625 μ M in 1 x DNA ligation buffer) and 2 μ L of Mastermix (16.6 μ M compound, 333 μ M ATP in 1.25x DNA ligation buffer) were incubated at 37 °C for 30 min. Following preincubation, 2 μ L DNA Mastermix (500 nM DNA1, 1.00 μ M DNA2, 1.00 μ M DNA3 in Milli-Q water) was added, and the samples were incubated at 37 °C for 30 min. The reactions were stopped by adding 90 μ L stopping solution and heating at 95°C for 2 min. 1 μ L samples were resolved via urea-PAGE and analyzed by autoradiographic imaging. Ligation turnover was quantified using ImageLab, and product formation at the indicated compound concentrations was visualized in GraphPad Prism.

Urea-PAGE and autoradiographic imaging

For the separation of oligonucleotide samples, a denaturing 12 % urea-PAGE sequencing gel was used. 1 μ L of the denatured sample was loaded onto a pre-warmed urea-PAGE sequencing gel. Electrophoresis was performed in 1x TBE buffer (90 mM Tris-HCl [pH 8.0], 90 mM boric acid, 2.0 mM EDTA) at 100 W and a maximal temperature of 45 °C for 45 min. The gel was transferred onto a Whatman filter paper and dried at 80 °C for 2 h under vacuum. After cooling to room temperature, the gel was exposed to a storage phosphor screen overnight. The autoradiographic imaging was performed by Typhoon FLA-9500 (GE Healthcare) in phosphorimaging mode. Gels were further analyzed using ImageLab.

3.2 Cell culture

Multipipetting was performed using Integra Viaflo or Integra Voyager pipettes. For automated multipipetting, an Integra Assist Plus pipetting robot was employed.

HEK293 cells were maintained in DMEM GlutaMAX™ medium (Gibco™) supplemented with 10% (v/v) FCS. Cells were cultured at 37 °C in an humidified incubator with 7.5% CO₂.

Cell viability assay of HEK293 cells treated with inhibitors

For the cell viability assay, cells were seeded at a density of 1×10^4 cells per well in a black 384-well plate (μ CLEAR®, cell culture, Greiner) in a total volume of 40 μ L medium. The plate was incubated at 37 °C, 7.5% CO₂, and 95% humidity for 24 h. Stock solutions of the compounds were prepared in DMSO at a concentration of 10 mM and 1:2 diluted in DMSO in a 96-well plate. Prior to addition to the cells, the compounds were further diluted in medium. Then, 20 μ L of the compound dilution was added to the corresponding wells of the cell plate, resulting in a final volume of 60 μ L per well and 0.9% DMSO. The plate was incubated for an additional 24 h at 37 °C. Cell viability was assessed using the CellTiter-Glo® 2.0 Cell Viability Assay (Promega). Upon addition of 15 μ L of CellTiterGlo® reagent per well using the Multidrop 384 reagent dispenser (Thermo Scientific), the plate was centrifuged for 1 min at 800 rpm and incubated for 40 min on a shaker at 25°C and 800 rpm. The luminescence was read out at 25 °C with a Spark microplate reader (Tecan). The luminescence values of the treated cells were compared to the value of the cells treated with DMSO as control (equals 100% viability). The EC₅₀ values were determined using GraphPad Prism software by fitting the data using a non-linear curve fit/ dose-response curve (fitting with a variable slope with four parameters logistic). All data represent mean \pm standard error of three independent experiments consisting of four technical replicates each.

Cell viability assay of HEK293 cells treated with inhibitors and menadione

1000x menadione stock solutions were prepared in DMSO and frozen in aliquots. Prior to treatment, aliquots were thawed and diluted in medium to 4-fold concentration. Upon 24 h treatment with the inhibitors at the indicated concentrations, 20 μ L of menadione in medium was added to a final menadione concentration of 40 μ M or 60 μ M in 80 μ L total volume. For the control wells with and without compound, 20 μ L of DMSO in medium were added. The plate was incubated at 37 °C, 7.5% CO₂, and 95% humidity for 5 h. Subsequently, 40 μ L was removed from each well. Cell viability was assessed using the CellTiter-Glo® 2.0 Cell Viability Assay (Promega) as described before. The percentual cell viability was normalized to the control wells with and without menadione and graphs were plotted using GraphPad Prism software. All data represent mean \pm standard error of four independent experiments consisting of four technical replicates each.

Light-microscopy of inhibitor- and menadione-treated HEK293 cells

5 x 10⁵ cells were seeded in 1.5 mL DMEM GlutaMAX™ medium supplemented with 10% (v/v) FCS in a 6-well plate (Sarstedt). The plate was incubated at 37 °C, 7.5% CO₂, and 95% humidity for 24 h. The cells were treated with 250 µL medium with either DMSO (control) or 30 µM compound **1**, **47** or **26**. After 24 h, 250 µL medium with either DMSO (control) or menadione in DMSO was added to a final concentration of 60 µM. After 5 h, pictures were taken with a light microscope using a 10x objective with 25 mm field of view.

RNA integrity analysis of HEK293 cells treated with inhibitors and various concentrations of menadione

5.0 x 10⁵ cells were seeded in 1.5 mL DMEM GlutaMAX™ medium supplemented with 10% (v/v) FCS in a 6-well plate (Sarstedt). The plate was incubated at 37 °C, 7.5% CO₂, and 95% humidity for 24 h. Compound stocks at 10 mM in DMSO were pre-diluted in medium before being added to the cells. The cells were treated with 250 µL of the compound dilution to a final concentration of 30 µM. DMSO was used as a control. The plate was incubated for an additional 24 h at 37 °C. 1000x menadione stock solution in DMSO was diluted in medium and 250 µL were added to a final menadione concentration of 40 µM or 60 µM. After 5 h, the cells were detached from the surface by vigorously pipetting up and down. The cell suspensions were then transferred to a 2 mL Eppendorf tube and centrifuged (350 g, 5 min, 4 °C). The cell pellets were washed with 1 mL DPBS (Gibco™), centrifuged again (350 g, 5 min, 4 °C) and snap-frozen in liquid nitrogen. For the RNA extraction, 600 µL TRIzol Reagent (Invitrogen) were added, and the pellets were resuspended by vortexing for 10 min at r.t. After 5 min incubation at r.t., 150 µL chloroform were added, the samples were vortexed for 1 min and centrifuged (12000 g, 15 min, 4 °C). The upper aqueous phases were transferred into new Eppendorf tubes and 150 µL chloroform were added. The samples were vortexed for 1 min, incubated for 5 min at r.t. and centrifuged (12000 g, 15 min, 4 °C). The upper clear phases were transferred into new Eppendorf tubes and 360 µL ice-cold isopropanol were added. The samples were mixed by shaking and stored for 24 h at -80 °C. The samples were centrifuged (20700 g, 30 min, 4 °C) and the supernatant was discarded. The RNA pellet was resuspended in 600 µL ice-cold ethanol and centrifuged (20700 g, 10 min, 4 °C). The supernatant was discarded and the ethanol washing step was repeated once again. The pellet was dried on air for 10 min and then dissolved in 40 µL Milli-Q. RNA concentration was determined by Nanodrop (Thermo Scientific) and 250 ng/µL RNA were analyzed according to the manufacturer instructions on an Agilent 4150 TapeStation system. Experiments with DMSO and 40 µM menadione were conducted in triplicate, while those with 60 µM menadione were performed in duplicate.

3.3 Chemical synthesis

General experimental details

Commercially available starting materials, reagents and anhydrous solvents were obtained from commercial suppliers (Sigma Aldrich, Merck, VWR chemicals, Carl Roth, Thermo Fisher Scientific, Fluka, TCI, ABCR and aablocks) and used without further purification if not stated otherwise. Dry solvents were purchased. For chemical synthesis desalted water was used. All air or water sensitive reactions were carried out under nitrogen atmosphere in dried reaction flasks.

The final compounds were further purified by precipitation. Therefore, the compound was diluted in a minimal amount of iso-propanol (2-5 mL) under reflux. The solution was transferred to a 50 mL falcon and 15-20 mL ice-cold water were added. The falcon was centrifuged (4000 rpm, 20 min, 4 °C), the precipitate was washed with 15 mL water and centrifuged again (4000 rpm, 20 min, 4 °C). The compound was dried under reduced pressure.

Analytical methods

Analytical thin layer chromatography (TLC)

Analytical thin layer chromatography was recorded on silica gel plates (aluminum) from Merck (Type 60-F254, 0.2 mm silica). For visualization, TLC plates were exposed to UV-light ($\lambda = 254$ nm), and/or stained with KMnO_4 (1.50 g KMnO_4 , 10.0 g K_2CO_3 and 1.25 mL 10% NaOH in 200 mL water) or ninhydrin (1 g ninhydrin in 100 mL EtOH) with subsequent heat treatment (ca. 250 °C).

Nuclear magnetic resonance spectroscopy (NMR)

NMR spectra were recorded at room temperature on Bruker Avance III 400 (400 MHz), Bruker Avance III 500 (500 MHz) or Bruker Avance III 600 (600 MHz) spectrometers. The chemical shifts (δ) are reported in parts per million (ppm) and ^1H , ^{13}C and ^{19}F spectra are referenced to residual proton and carbon signals of the deuterated solvents MeOD and DMSO- d_6 . Multiplicities are denoted as s = singlet, d = doublet, t = triplet, q = quartet, p = quintet, dd = doublet of doublet, dt = doublet of triplet, m = multiplet, br = broad. NMR data were processed by MestReNova (version 14.2.0-26256). The coupling constants, J, are reported in Hertz (Hz). Since some signals could not be assigned clearly, they were marked with an asterisk (*); the corresponding assignments are interchangeable.

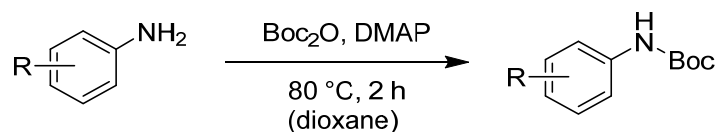
Mass-spectrometry

High resolution mass-spectrometry (HR-MS) were performed by QTOF 6546 (ESI-TOF) from Agilent or TQ-Orbitrap Discovery mass spectrometer from Thermo Scientific.

General procedures

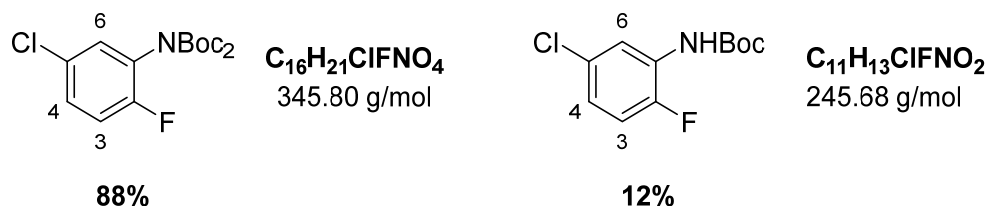
The procedures were adapted from published protocols.³⁻⁴

General procedure A: Boc-protection of aryl amines



To a stirred solution of aryl amine (1.00 eq) in anhydrous dioxane was added a solution of di-*tert*-butyl dicarbonate (1.00 or 2.00 eq) in 1 mL anhydrous dioxane and 4-dimethylaminopyridine (0.100 eq). The reaction mixture was stirred at 80 °C for 2-4 hours. Upon cooling to room temperature, the reaction was stopped with sat. NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). The combined organic layer was dried over MgSO₄, filtered and concentrated. Each residue was purified by silica gel column chromatography to afford the Boc protected aryl amine.

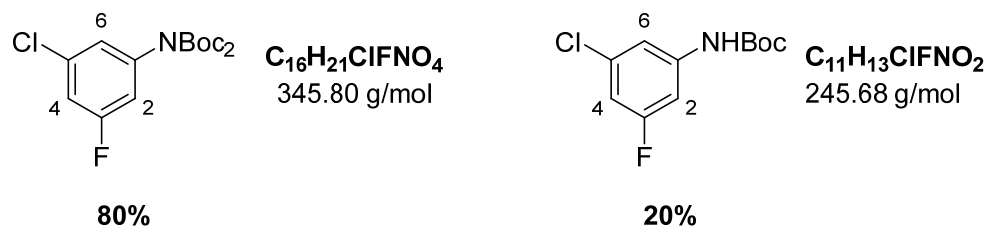
***tert*-Butyl (tert-butoxycarbonyl)(5-chloro-2-fluorophenyl)carbamate & *tert*-butyl (5-chloro-2-fluorophenyl)carbamate (48)**



Following general procedure A, 5-chloro-2-fluoroaniline (350 mg, 2.10 mmol, 1.00 eq), Boc₂O (916 mg, 4.20 mmol, 2.00 eq) and DMAP (24.0 mg, 196 μmol, 0.10 eq) in 5 mL dioxane was stirred at 80 °C for 4 h. The reaction mixture was purified by extraction and silica gel column chromatography [10% → 100% EtOAc /pentane] to provide **48** (485 mg, 1.46 mmol, 70%) as a yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 7.60 (dd, J = 6.7, 2.7 Hz, 1H, H-6), 7.52 – 7.44 (m, 1H, H-3*), 7.42 – 7.32 (m, 1H, H-4*), 1.37 (s, 18H, Boc₂).

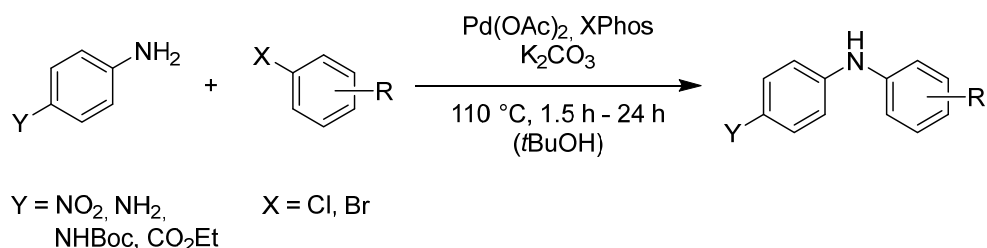
***tert*-Butyl (tert-butoxycarbonyl)(5-chloro-3-fluorophenyl)carbamate & *tert*-butyl (5-chloro-3-fluorophenyl)carbamate (49)**



Following general procedure A, 3-chloro-5-fluoroaniline (340 mg, 2.34 mmol, 1.00 eq), Boc₂O (1.00 g, 4.60 mmol, 2.00 eq) and DMAP (28.5 mg, 233 μmol, 0.10 eq) in 5 mL dioxane was stirred at 80 °C for 2 h. The reaction mixture was purified by silica gel column chromatography [10% EtOAc /pentane] to provide **49** (655 mg, 2.05 mmol, 88%) as a yellow solid.

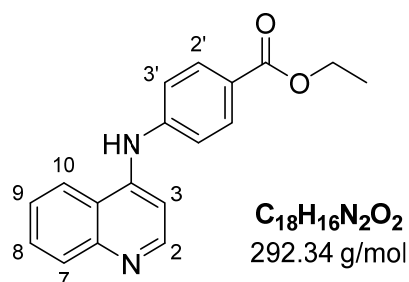
¹H-NMR (DMSO, 400 MHz) δ = 7.44 (dt, J = 8.7, 2.1 Hz, 1H, H-6), 7.31 – 7.28 (m, 2H, H-2, H-4), 1.39 (s, 18H, Boc₂).

General procedure B: Buchwald-Hartwig amination



An oven-dried 25 mL flask with Pd(OAc)₂ (2 mol%) and XPhos (6 mol%) was evacuated and flushed with nitrogen thrice. Degassed *tert*-butanol (*t*BuOH) and degassed water (2 mol%) were added and the suspension was heated to 80 °C for one minute. Subsequently, the aryl halide (1.00 eq), the aryl amine (1.20 eq) and K₂CO₃ (1.45 eq) were added, and the reaction mixture was stirred at 110 °C for 1.5 – 24 hours. Completion of the reaction was monitored by TLC. Upon cooling to room temperature, the suspension was diluted with EtOAc and sat. NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). The combined organic layer was dried over MgSO₄, filtered and concentrated. Each residue was purified by silica gel column chromatography to afford the coupled aryl amine compound.

Ethyl 4-(quinolin-4-ylamino)benzoate (50)



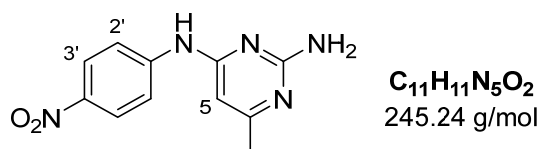
Following general procedure B, a mixture of ethyl-4-aminobenzoate (500 mg, 3.03 mmol, 1.24 eq), 4-chloroquinoline (320 μL, 400 mg, 2.44 mmol, 1.00 eq), Pd(OAc)₂ (12.0 mg, 50.0 μmol, 0.02 eq), XPhos (59.0 mg, 120 μmol, 0.05 eq), K₂CO₃ (476 mg, 3.44 mmol, 1.41 eq) in 4 mL *t*BuOH and 3 μL water was stirred at 110 °C for three hours. The product **50** was isolated by silica column chromatography [DCM, 1% TEA → DCM, 0.5% MeOH 1% TEA] as a light yellow solid (669 mg, 2.29 mmol, 94%).

TLC: *R_f* = 0.38 (5% MeOH/1%TEA/ DCM) [UV].

¹H-NMR (DMSO-*d*₆, 400 MHz) δ = 9.32 (s, 1H, NH), 8.60 (d, *J* = 5.14 Hz, 1H, H-2), 8.34 (d, *J* = 8.39 Hz, 1H, H-10), 7.94 (d, *J* = 8.44 Hz, 2H, H-2'), 7.93 (t, *J* = 8.26 Hz, 1H, H-7), 7.74 (t, *J* = 7.63 Hz, 1H, H-8), 7.58 (d, *J* = 7.63 Hz, 1H, H-9), 7.44 (d, *J* = 8.44 Hz, 2H, H-3'), 7.27 (d, *J* = 5.14 Hz, 1H, H-3), 4.29 (q, *J* = 7.11 Hz, 2H, CH₂), 1.32 (t, *J* = 7.11 Hz, 3H, CH₃).

These data are in accordance with previous literature reports.³

6-Methyl-*N*⁴-(4-nitrophenyl)pyrimidine-2,4-diamine (**51**)



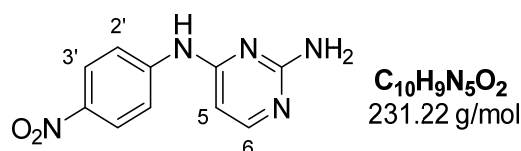
Following general procedure B, 4-nitroaniline (2.35 g, 17.0 mmol, 1.22 eq), 2-amino-4-chloro-6-methyl pyrimidine (2.00 g, 13.9 mmol, 1.00 eq), Pd(OAc)₂ (40.0 mg, 180 μmol, 0.013 eq), XPhos (213 mg, 450 μmol, 0.032 eq) and K₂CO₃ (2.80 g, 20.3 mmol, 1.45 eq) in 28 mL *t*BuOH and 9 μL water was stirred at 110 °C for 90 minutes. The product was isolated by silica column chromatography [0.5% MeOH/ DCM, 1% TEA → DCM, 5% MeOH 1% TEA] as a yellow solid (1.78 g, 7.23 mmol, 52%).

TLC: *R*_f = 0.66 (5% MeOH/1%TEA/ DCM) [UV].

¹H-NMR (DMSO-*d*₆, 400 MHz) δ = 9.78 (s, 1H, NH), 8.13 (d, *J* = 9.30 Hz, 2H, H-3'), 8.01 (d, *J* = 9.30 Hz, 2H, H-2'), 6.40 (s, 2H, NH₂), 5.99 (s, 1H, H-5), 2.14 (s, 3H, CH₃).

These data are in accordance with previous literature reports.³

*N*⁴-(4-Nitrophenyl)pyrimidine-2,4-diamine (**52**)



Following general procedure B, 4-nitroaniline (320 mg, 2.32 mmol, 1.22 eq), 2-amino-4-chloropyrimidine (250 mg, 1.93 mmol, 1.00 eq) Pd(OAc)₂ (5.60 mg, 25.0 μmol, 0.013 eq), XPhos (29 mg, 61.0 μmol, 0.032 eq) and K₂CO₃ (386 mg, 2.79 mmol, 1.45 eq) in 4 mL *t*BuOH and 2 μL water was stirred at 110 °C for 90 minutes. Purification by silica gel column chromatography afforded [0.5% MeOH/1% TEA/ DCM → 3% MeOH/1% TEA / DCM] compound **52** (228 mg, 98.4 μmol, 51%) as a yellow solid.

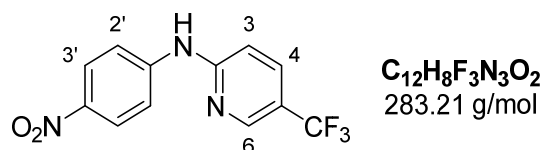
TLC: *R*_f = 0.38 (5% MeOH/1%TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.89 (s, 1H, NH), 8.14 (d, *J* 9.34 Hz, 2H, H-3'), 8.03 (d, *J* = 9.34 Hz, 2H, H-2'), 7.95 (d, *J* = 5.62 Hz, 1H, H-6), 6.12 (d, *J* = 5.62 Hz, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 162.84, 160.15, 157.39, 147.34, 140.18, 124.85, 118.08, 97.66.

HR-MS *m/z* calcd. for C₁₀H₉N₅O₂⁺ (*M* + 1)⁺ 232.0834, found 232.0841.

***N*-(4-Nitrophenyl)-5-(trifluoromethyl)pyridin-2-amine (53)**



Following general procedure B, 4-nitroaniline (232 mg, 1.68 mmol, 1.22 eq), 2-chloro-5-(trifluoromethyl)pyridine (250 mg, 1.38 mmol, 1.00 eq), Pd(OAc)₂ (4.00 mg, 18.0 μmol, 0.013 eq), XPhos (21.0 mg, 44.0 μmol, 0.032 eq) and K₂CO₃ (276 mg, 2.00 mmol, 1.45 eq) in 5 mL *t*BuOH and 4 μL water was stirred at 110 °C for three hours. The reaction mixture was quenched with 10 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [0.5% TEA/ DCM] afforded **53** (123 mg, 441 μmol, 32%) as yellow solid.

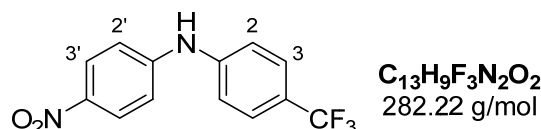
TLC: R_f = 0.49 (1%TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.53 (s, 1H, H-6), 8.18 (d, J = 9.28 Hz, 2H, H-3'), 7.95 (d, J = 9.28 Hz, 2H, H-2'), 7.86 (dd, J = 2.54, 8.78 Hz, 1H, H-4), 7.00 (d, J = 8.78 Hz, 1H, H-3).

¹³C-NMR (DMSO, 101 MHz) δ = 158.74, 148.39, 146.15, 142.59, 135.68, 125.95, 118.84, 112.82.

HR-MS m/z calcd. for $C_{12}H_9F_3N_3O_2^+$ ($M + 1$)⁺ 284.0641, found 284.0649.

4-Nitro-*N*-(4-(trifluoromethyl)phenyl)aniline (54)



Following general procedure B, 4-nitroaniline (229 mg, 1.66 mmol, 1.20 eq), 1-chloro-4-(trifluoromethyl)benzene (250 mg, 1.38 mmol, 1.00 eq), Pd(OAc)₂ (7.00 mg, 30.0 μmol, 0.02 eq), XPhos (38.0 mg, 80.0 μmol, 0.06 eq) and K₂CO₃ (276 mg, 2.00 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μL water was stirred at 110 °C for three hours. The reaction mixture was quenched with 10 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [5%EtOAc/1%TEA/pentane → 15%EtOAc/1%TEA/pentane] afforded **54** (321 mg, 1.15 mmol, 83%) as yellow solid.

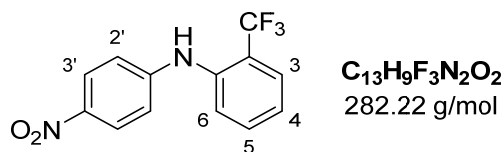
TLC: R_f = 0.55 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.63 (s, 1H, NH), 8.15 (d, J = 9.24 Hz, 2H, H-3'), 7.68 (d, J = 8.45 Hz, 2H, H-3), 7.39 (d, J = 8.45 Hz, 2H, H-2), 7.24 (d, J = 9.24 Hz, 2H, H-2').

¹³C-NMR (DMSO, 101 MHz) δ = 149.02, 144.34, 139.33, 126.74, 125.99, 118.85, 115.20.

HR-MS m/z calcd. for $C_{13}H_{10}F_3N_2O_2^+$ ($M + 1$)⁺ 283.0689, found 283.0688.

***N*-(4-Nitrophenyl)-2-(trifluoromethyl)aniline (55)**



Following general procedure B, 4-nitroaniline (229 mg, 1.66 mmol, 1.20 eq), 1-chloro-2-(trifluoromethyl)benzene (186 μ L, 250 mg, 1.38 mmol, 1.00 eq), Pd(OAc)₂ (7.00 mg, 30.0 μ mol, 0.02 eq), XPhos (38.0 mg, 80.0 μ mol, 0.06 eq) and K₂CO₃ (276 mg, 2.00 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μ L water was stirred at 110 °C for two hours. The reaction mixture was quenched with 10 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% TEA/pentane → 20% EtOAc/1% TEA/pentane] afforded **55** (366 mg, 1.30 mmol, 94%) as yellow, crystalline solid.

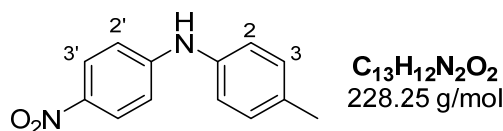
TLC: R_f = 0.86 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.87 (s, 1H, NH), 8.09 – 8.00 (m, 2H, H-3'), 7.83 (d, J = 7.7 Hz, 1H, H-3), 7.74 (t, J = 7.8 Hz, 1H, H-5), 7.54 (d, J = 7.8 Hz, 1H, H-6), 7.50 (t, J = 7.6 Hz, 1H, H-4), 6.84 – 6.76 (m, 2H, H-2').

¹³C-NMR (DMSO, 101 MHz) δ = 152.82, 138.04, 137.98, 134.09, 129.00, 127.25, 126.69, 126.05, 113.19.

HR-MS m/z calcd. for C₁₃H₁₀F₃N₂O₂⁺ ($M + 1$)⁺ 283.0689, found 283.0691.

4-Methyl-*N*-(4-nitrophenyl)aniline (56)



Following general procedure B, 4-nitroaniline (327 mg, 2.37 mmol, 1.20 eq), 4-chlorotoluene (250 mg, 1.97 mmol, 1.00 eq), Pd(OAc)₂ (9.00 mg, 40.0 μ mol, 0.02 eq), XPhos (57.0 mg, 120 μ mol, 0.06 eq) and K₂CO₃ (395 mg, 2.86 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μ L water was stirred at 110 °C for 3 hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [5% EtOAc/1% TEA/pentane → 10% EtOAc/1% TEA/pentane] afforded **56** (369 mg, 1.61 mmol, 82%) as orange solid.

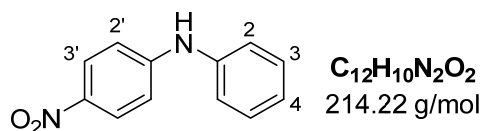
TLC: R_f = 0.66 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.21 (s, 1H, NH), 8.06 (d, J = 9.3 Hz, 2H, H-3'), 7.20 (d, J = 8.4 Hz, 2H, H-3), 7.13 (d, J = 8.4 Hz, 2H, H-2), 6.98 (d, J = 9.3 Hz, 2H, C-2'), 2.29 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 151.34, 137.50, 137.30, 132.91, 129.96, 126.23, 121.46, 112.89, 20.47.

HR-MS m/z calcd. for C₁₃H₁₃N₂O₂⁺ ($M + 1$)⁺ 229.0972, found 229.0974.

4-Nitro-*N*-phenylaniline (**57**)



Following general procedure B, 4-nitroaniline (368 mg, 2.66 mmol, 1.20 eq), chlorobenzene (250 mg, 2.22 mmol, 1.00 eq), Pd(OAc)₂ (10.0 mg, 44.0 μmol, 0.02 eq), XPhos (63.0 mg, 133 μmol, 0.06 eq) and K₂CO₃ (445 mg, 3.22 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μL water was stirred at 110 °C for 2.5 hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% TEA/pentane → 20% EtOAc/1% TEA/pentane] afforded **57** (347 mg, 1.62 mmol, 73%) as yellow crystals.

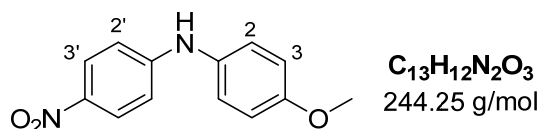
TLC: *R*_f = 0.59 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.30 (s, 1H, NH), 8.09 (d, *J* = 9.3 Hz, 2H, H-3'), 7.39 (dd, *J* = 8.5, 7.3 Hz, 2H, H-3), 7.24 (d, *J* = 8.5, 1.2 Hz, 2H, H-2), 7.10 (t, *J* = 7.3 Hz, 1H, H-4), 7.06 (d, *J* = 9.3 Hz, 2H, H-2').

¹³C-NMR (DMSO, 101 MHz) δ = 150.81, 140.05, 137.90, 129.53, 126.20, 123.45, 120.85, 113.35.

HR-MS *m/z* calcd. for C₁₂H₁₁N₂O₂⁺ (*M* + 1)⁺ 215.0815, found 215.0818.

4-Methoxy-*N*-(4-nitrophenyl)aniline (**58**)



Following general procedure B, 4-nitroaniline (290 mg, 2.10 mmol, 1.20 eq), 4-chloroanisole (250 mg, 1.73 mmol, 1.00 eq), Pd(OAc)₂ (8.00 mg, 34.6 μmol, 0.02 eq), XPhos (50.0 mg, 104 μmol, 0.06 eq) and K₂CO₃ (346 mg, 2.50 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μL water was stirred at 110 °C for two hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [5% EtOAc/1% TEA/pentane → 15% EtOAc/1% TEA/pentane] afforded **58** (300 mg, 1.22 mmol, 71%) as orange solid.

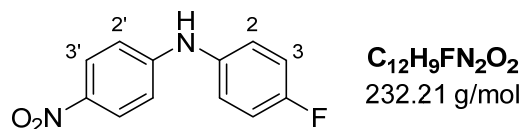
TLC: *R*_f = 0.33 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.11 (s, 1H, NH), 8.04 (d, *J* = 9.3 Hz, 2H, H-3'), 7.18 (d, *J* = 8.9 Hz, 2H, H-2), 6.98 (d, *J* = 8.9 Hz, 2H, H-3), 6.88 (d, *J* = 9.3 Hz, 2H, H-2'), 3.76 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 156.15, 152.14, 137.12, 132.51, 126.31, 124.02, 114.80, 112.29, 55.29.

These data are in accordance with previous literature reports. Literature spectra were recorded in deuterated CDCl₃.⁵

4-Fluoro-*N*-(4-nitrophenyl)aniline (**59**)



Following general procedure B, 4-nitroaniline (316 mg, 2.29 mmol, 1.20 eq), 1-chloro-4-fluorobenzene (204 μ L, 250 mg, 1.91 mmol, 1.00 eq), Pd(OAc)₂ (9.00 mg, 38.2 μ mol, 0.02 eq), XPhos (55.0 mg, 115 μ mol, 0.06 eq) and K₂CO₃ (381 mg, 2.76 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μ L water was stirred at 110 °C for two hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [7.5% EtOAc/1% TEA/pentane \rightarrow 10% EtOAc/1% TEA/pentane] afforded **59** (288 mg, 1.24 mmol, 65%) as orange solid.

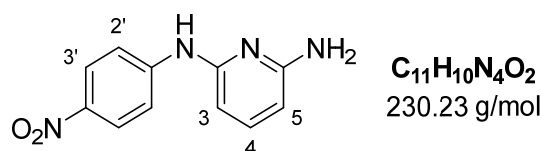
TLC: R_f = 0.60 (20% EtOAc/80% pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.25 (s, 1H, NH), 8.08 (d, J = 9.3 Hz, 2H, H-3'), 7.35 – 7.14 (m, 4H, H-2, H-3), 6.98 (d, J = 9.3 Hz, 2H, H-2').

¹³C-NMR (DMSO, 101 MHz) δ = 151.22, 137.82, 136.30, 126.23, 123.49, 123.41, 116.33, 116.11, 112.95.

HR-MS m/z calcd. for C₁₂H₁₀FN₂O₂⁺ ($M + 1$)⁺ 233.0721, found 233.0722.

N'-(4-Nitrophenyl)pyridine-2,6-diamine (**60**)



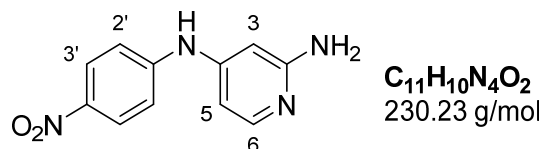
Following general procedure B, 4-nitroaniline (383 mg, 2.77 mmol, 1.20 eq), 2-amino-6-bromopyridine (400 mg, 2.31 mmol, 1.00 eq), Pd(OAc)₂ (10.0 mg, 46.0 μ mol, 0.02 eq), XPhos (66.0 mg, 139 μ mol, 0.06 eq) and K₂CO₃ (463 mg, 3.35 mmol, 1.45 eq) in 5 mL *t*BuOH and 3.5 μ L water was stirred at 110 °C for four hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [20% EtOAc \rightarrow 50% EtOAc/1% TEA/pentane] afforded **60** (222 mg, 1.18 mmol, 51%) as orange solid.

TLC: R_f = 0.52 (50% EtOAc/1% TEA/ pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.56 (s, 1H, NH), 8.08 (d, J = 9.4 Hz, 2H, H-3'), 7.91 (d, J = 9.4 Hz, 1H, H-2'), 7.30 (t, J = 7.9 Hz, 1H, H-4), 6.11 (d, J = 7.7 Hz, 1H, H-3), 6.02 (d, J = 7.9 Hz, 1H, H-5), 5.93 (s, 2H, NH₂).

$^{13}\text{C-NMR}$ (DMSO, 101 MHz) δ = 158.47, 153.27, 148.96, 138.79, 138.50, 125.15, 116.16, 99.93, 99.13.

***N*'-(4-Nitrophenyl)pyridine-2,4-diamine (61)**



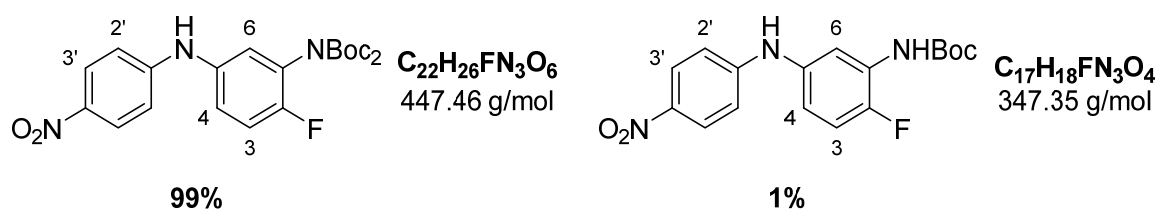
Following general procedure B, 4-nitroaniline (516 mg, 3.73 mmol, 1.20 eq), 4-chloropyridine-2-amine (400 mg, 3.11 mmol, 1.00 eq), $\text{Pd}(\text{OAc})_2$ (14.0 mg, 60.0 μmol , 0.02 eq), XPhos (91.0 mg, 190 μmol , 0.06 eq) and K_2CO_3 (622 mg, 4.50 mmol, 1.45 eq) in 5 mL *t*BuOH and 3.5 μL water was stirred at 110 $^\circ\text{C}$ for four hours. The reaction mixture was quenched with 15 mL NaHCO_3 solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% MeOH \rightarrow 5% MeOH/1% TEA/ DCM] afforded **61** (257 mg, 1.12 mmol, 36%) as yellow solid.

TLC: R_f = 0.30 (5% MeOH/1% TEA/ DCM) [UV].

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 9.34 (s, 1H, NH), 8.15 (d, J = 9.2 Hz, 2H, H-3'), 7.78 (d, J = 5.7 Hz, 1H, H-6), 7.23 (d, J = 9.3 Hz, 2H, H-2'), 6.34 (dd, J = 5.8, 2.0 Hz, 1H, H-5), 6.28 (d, J = 2.0 Hz, 1H, H-3), 5.84 (s, 2H, NH_2).

$^{13}\text{C-NMR}$ (DMSO, 101 MHz) δ = 160.76, 148.74, 148.40, 139.37, 125.74, 116.15, 103.11, 94.61.

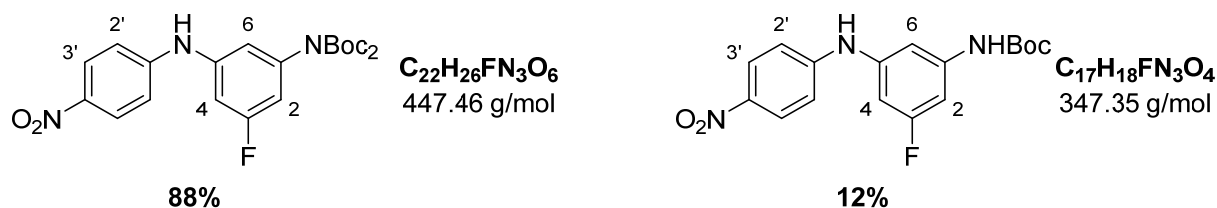
***tert*-Butyl (*tert*-butoxycarbonyl)(2-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate (62)**



Following general procedure B, 4-nitroaniline (134 mg, 980 μmol , 1.20 eq), 4-chloropyridine-2-amine (200 mg, 810 μmol , 1.00 eq), $\text{Pd}(\text{OAc})_2$ (5.00 mg, 20.0 μmol , 0.02 eq), XPhos (23.2 mg, 50.0 μmol , 0.06 eq) and K_2CO_3 (162 mg, 1.17 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μL water was stirred at 110 $^\circ\text{C}$ for four hours. The reaction mixture was quenched with 15 mL sat. NaHCO_3 solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% TEA/ DCM] afforded **62** (175 mg, 410 μmol , 50%) as yellow solid.

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 9.30 (s, 1H, NH), 8.09 (d, J = 9.2 Hz, 2H, H-3'), 7.33 (t, J = 9.2 Hz, 1H, H-6), 7.31 – 7.17 (m, 2H, H-3, H-4), 6.97 (d, J = 9.3 Hz, 2H, H-2'), 1.40 (s, 18H, Boc_2).

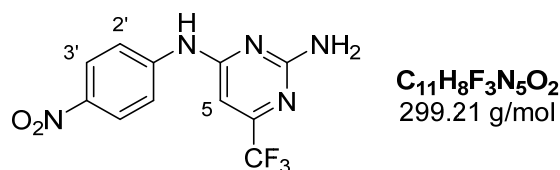
tert-Butyl (tert-butoxycarbonyl)(3-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate & tert-butyl (3-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate (63)



Following general procedure B, 4-nitroaniline (134 mg, 980 μ mol, 1.20 eq), 4-chloropyridine-2-amine (200 mg, 810 μ mol, 1.00 eq), Pd(OAc)₂ (5.00 mg, 20.0 μ mol, 0.02 eq), XPhos (23.2 mg, 50 μ mol, 0.06 eq) and K₂CO₃ (162 mg, 1.17 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μ L water was stirred at 110 °C for 5.5 hours. The reaction mixture was quenched with 15 mL sat. NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% TEA/ DCM] afforded **63** (161 mg, 380 μ mol, 46%) as yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 9.49 (s, 1H, NH), 8.14 (d, J = 9.2 Hz, 2H, H-3'), 7.14 (d, J = 9.3 Hz, 2H, H-2'), 7.06 (dt, J = 10.6, 2.2 Hz, 1H, H-6*), 6.93 – 6.80 (m, 2H, H-2*, H-4*), 1.42 (s, 18H, Boc₂).

N⁴-(4-Nitrophenyl)-6-(trifluoromethyl)pyrimidine-2,4-diamine (64)



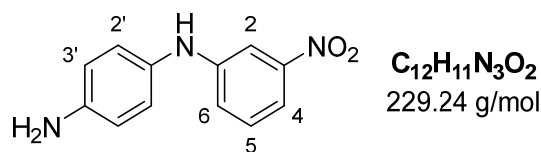
Following general procedure B, 4-nitroaniline (334 mg, 2.42 mmol, 1.20 eq), 4-chloropyridine-2-amine (400 mg, 2.02 mmol, 1.00 eq), Pd(OAc)₂ (9.00 mg, 40.0 μ mol, 0.02 eq), XPhos (57.0 mg, 120 μ mol, 0.06 eq) and K₂CO₃ (404 mg, 2.93 mmol, 1.45 eq) in 5 mL *t*BuOH and 3.5 μ L water was stirred at 110 °C for 5 hours. The reaction mixture was quenched with 15 mL sat. NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [10% EtOAc → 30% EtOAc/1% TEA/ pentane] afforded **64** (238 mg, 788 μ mol, 39%) as yellow solid.

TLC: R_f = 0.37 (20% EtOAc/1% TEA/ pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.28 (s, 1H, NH), 8.18 (d, J = 9.3 Hz, 2H, H-3'), 8.05 (d, J = 9.3 Hz, 2H, H-2'), 7.13 (s, 2H, NH₂), 6.46 (s, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 163.32, 161.24, 154.41, 154.07, 146.32, 141.05, 124.88, 118.80, 93.54.

N'-(3-Nitrophenyl)benzene-1,4-diamine (**65**)



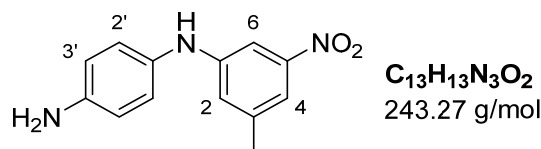
Following general procedure B, benzene-1,4-diamine (321 mg, 2.97 mmol, 1.20 eq), 1-bromo-3-nitrobenzene (400 mg, 1.98 mmol, 1.00 eq), Pd(OAc)₂ (9.00 mg, 40.0 μmol, 0.02 eq), XPhos (57.0 mg, 120 μmol, 0.06 eq) and K₂CO₃ (397 mg, 2.87 mmol, 1.45 eq) in 5 mL *t*BuOH and 3.5 μL water was stirred at 110 °C for 4 hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% MeOH → 5% MeOH/1% TEA/ DCM] afforded **65** (257 mg, 1.13 mmol, 57%) as red oil.

TLC: *R*_f = 0.50 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.10 (s, 1H, NH), 7.46 (s, 1H, H-2), 7.41 – 7.37 (m, 1H, H-6), 7.33 (t, *J* = 8.0 Hz, 1H, H-4), 7.15 – 7.07 (m, 1H H-5), 6.88 (d, *J* = 8.5 Hz, 2H, H-2'), 6.59 (d, *J* = 8.6 Hz, 2H, H-3'), 4.96 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 148.82, 148.59, 145.45, 130.16, 129.33, 124.22, 119.27, 114.76, 110.79, 106.07, 40.15, 39.94, 39.73, 39.52, 39.31, 39.10, 38.89, 24.03.

N'-(3-Methyl-5-nitrophenyl)benzene-1,4-diamine (**66**)



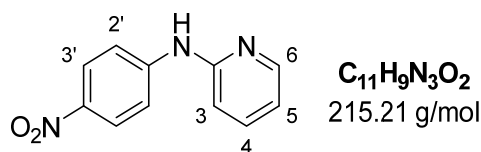
Following general procedure B, benzene-1,4-diamine (346 mg, 3.20 mmol, 1.20 eq), 1-chloro-3-methyl-5-nitrobenzene (400 mg, 2.13 mmol, 1.00 eq), Pd(OAc)₂ (10.0 mg, 43.0 μmol, 0.02 eq), XPhos (61.0 mg, 123 μmol, 0.06 eq) and K₂CO₃ (427 mg, 3.08 mmol, 1.45 eq) in 5 mL *t*BuOH and 3.5 μL water was stirred at 110 °C for 5 hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [2% MeOH/1% TEA/ DCM] afforded **66** (321 mg, 1.32 mmol, 62%) as red oil.

TLC: *R*_f = 0.33 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.01 (s, 1H, NH), 7.28 (s, 1H, H-6), 7.24 (s, 1H, H-4), 6.91 (s, 1H, H-2), 6.86 (d, *J* = 8.6 Hz, 2H, H-2'), 6.59 (d, *J* = 8.5 Hz, 2H, H-3'), 4.95 (s, 2H, NH₂), 2.28 (s, 1H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 148.80, 148.46, 145.42, 140.04, 129.39, 124.34, 119.48, 114.71, 111.46, 103.85, 20.98.

***N*-(4-Nitrophenyl)pyridin-2-amine (67)**

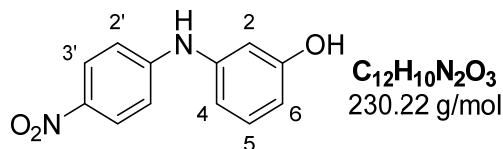


Following general procedure B, 4-nitroaniline (419 mg, 3.04 mmol, 1.20 eq), 4-bromopyridine (400 mg, 2.53 mmol, 1.00 eq), Pd(OAc)₂ (11.0 mg, 50.0 μmol, 0.02 eq), XPhos (72.0 mg, 150 μmol, 0.06 eq) and K₂CO₃ (507 mg, 3.67 mmol, 1.45 eq) in 7 mL *t*BuOH and 3.5 μL water was stirred at 110 °C for 5 hours. The reaction mixture was quenched with 15 mL sat. NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [1% TEA/ DCM] afforded **67** (512 mg, 2.38 mmol, 94%) as yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 8.33 – 8.22 (m, 1H, H-6), 8.17 (d, J = 9.3 Hz, 1H, H-3'), 7.93 (d, J = 9.3 Hz, 2H, H-2'), 7.78 – 7.58 (m, 1H, H-4), 6.98 (d, J = 8.4 Hz, 1H, H-3), 6.97 – 6.92 (m, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 154.46, 148.25, 147.32, 139.28, 137.99, 125.31, 116.62, 116.45, 112.49.

3-((4-Nitrophenyl)amino)phenol (68)

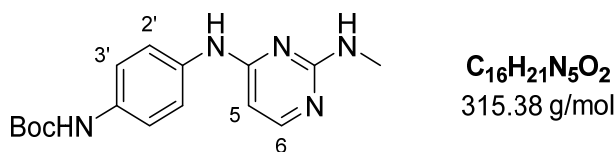


Following general procedure B, 4-nitroaniline (516 mg, 3.73 mmol, 1.20 eq), 3-chlorophenol (400 mg, 3.11 mmol, 1.00 eq), Pd(OAc)₂ (16.0 mg, 70.0 μmol, 0.02 eq), XPhos (100.0 mg, 210 μmol, 0.06 eq) and K₂CO₃ (705 mg, 4.50 mmol, 1.45 eq) in 4 mL *t*BuOH and 3 μL water was stirred at 110 °C for 16 hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [3% MeOH/1% TEA/DCM] afforded **68** (704 mg, 3.06 mmol, 98%) as yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 9.54 (br. s, 1H, OH), 9.20 (s, 1H, NH), 8.08 (d, J = 9.3 Hz, 2H, H-3'), 7.15 (t, J = 8.0 Hz, 1H, H-5), 7.05 (d, J = 9.3 Hz, 2H, H-2'), 6.70 – 6.60 (m, 2H, H-2, H-4), 6.50 (ddd, J = 8.1, 2.3, 0.9 Hz, 1H, H-6).

¹³C-NMR (DMSO, 101 MHz) δ = 158.36, 150.82, 141.16, 137.83, 130.24, 126.19, 113.57, 111.40, 110.71, 107.57.

***tert*-Butyl (4-((2-(ethylamino)pyrimidin-4-yl)amino)phenyl)carbamate (69)**

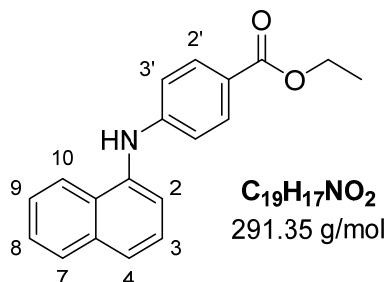


Following general procedure B, *tert*-butyl (4-aminophenyl)carbamate (872 mg, 4.19 mmol, 1.20 eq), 4-chloro-*N*-methylpyrimidin-2-amine (400 mg, 3.49 mmol, 1.00 eq), Pd(OAc)₂ (16.0 mg, 70.0 μmol, 0.02 eq), XPhos (100.0 mg, 210 μmol, 0.06 eq) and K₂CO₃ (699 mg, 5.00 mmol, 1.45 eq) in 3 mL *t*BuOH and 2 μL water was stirred at 110 °C for five hours. The reaction mixture was quenched with 15 mL NaHCO₃ solution and extracted with EtOAc (3 x 100 mL). Purification by silica gel column chromatography [2% → 5% MeOH/1% TEA/DCM] afforded **69** (523 mg, 1.82 mmol, 52%) as white solid.

¹H-NMR (DMSO, 400 MHz) δ = 9.16 (s, 1H, NH) 8.96 (s, 1H, NH), 7.79 (d, *J* = 5.7 Hz, 1H, H-6), 7.57 (d, *J* = 8.8 Hz, 2H, H-3'), 7.34 (d, *J* = 8.8 Hz, 1H, H-2'), 5.92 (d, *J* = 5.7 Hz, 1H, H-5), 2.77 (d, *J* = 4.6 Hz, 3H, CH₃), 1.47 (s, 9H, Boc).

The impure compound was used without further purification in the next step.

Ethyl 4-(naphthalen-1-ylamino)benzoate (70)



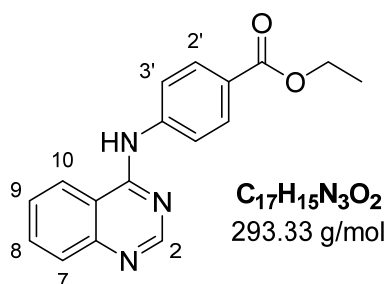
Following general procedure B, a mixture of ethyl-4-aminobenzoate (487 mg, 2.95 mmol, 1.20 eq), 1-chloronaphthalene (400 mg, 2.46 mmol, 1.00 eq), Pd(OAc)₂ (11 mg, 49.1 μmol, 0.02 eq), XPhos (70.3 mg, 148 μmol, 0.06 eq), K₂CO₃ (493 mg, 3.57 mmol, 1.45 eq) in 4 mL *t*BuOH and 3 μL water was stirred at 110 °C for 3 hours. The product was isolated by silica column chromatography [15% EtOAc/ 1% TEA /pentane] as light yellow solid (628 mg, 2.26 mmol, 87%).

TLC: *R*_f = 0.50 (15% EtOAc/ 1% TEA /pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.83 (s, 1H, NH), 8.06 – 8.02 (m, 1H, H-10), 7.97 – 7.93 (m, 1H, H-7), 7.77 (d, *J* = 8.8 Hz, 2H, H-2'), 7.73 (d, *J* = 8.0 Hz, 1H, H-4), 7.57 – 7.48 (m, 3H, H-3, H-8, H-9), 7.45 (d, *J* = 7.5, 1.3 Hz, 1H, H-2), 6.93 (d, *J* = 8.8 Hz, 2H, H-3'), 4.23 (q, *J* = 7.1 Hz, 2H, CH₂), 1.28 (t, *J* = 7.1 Hz, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 165.62, 150.63, 136.93, 134.41, 130.95, 128.40, 128.29, 126.31, 126.09, 125.75, 124.21, 122.91, 119.19, 118.86, 113.78, 59.82, 14.33.

Ethyl 4-(quinazolin-4-ylamino)benzoate (71)

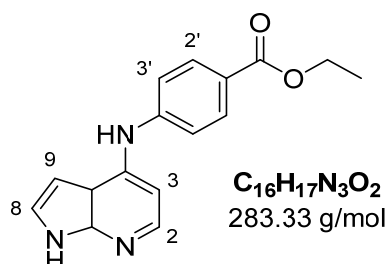


Following general procedure B, a mixture of ethyl-4-aminobenzoate (482 mg, 2.92 mmol, 1.20 eq), 4-chloroquinazoline (400 mg, 2.43 mmol, 1.00 eq), Pd(OAc)₂ (11.0 mg, 48.6 μmol, 0.02 eq), XPhos (70.0 mg, 146 μmol, 0.06 eq), K₂CO₃ (487 mg, 3.52 mmol, 1.45 eq) in 4 mL *t*BuOH and 3 μL water was stirred at 110 °C for 6 hours. The product was isolated by silica column chromatography [30% EtOAc/ 1% TEA /pentane] as light yellow solid (530 mg, 1.81 mmol, 35%).

TLC: *R*_f = 0.18 (30% EtOAc/ 1% TEA /pentane) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.05 (s, 1H, NH), 8.71 (s, 1H, H-2), 8.61 (d, *J* = 8.3 Hz, 1H, H-7), 8.13 (d, *J* = 8.7 Hz, 2H, H-2'), 7.99 (d, *J* = 8.7 Hz, 2H, H-3'), 7.91 (t, *J* = 7.6 Hz, 1H, H-8), 7.84 (d, *J* = 8.5 Hz, 1H, H-10), 7.69 (t, *J* = 8.3 Hz, 1H, H-9), 4.31 (q, *J* = 7.1 Hz, 2H, CH₂), 1.33 (t, *J* = 7.1 Hz, 3H, CH₃).

Ethyl 4-((3a,7a-dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzoate (72)

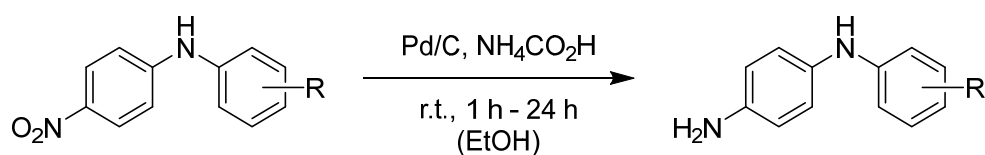


Following general procedure B, a mixture of ethyl-4-aminobenzoate (515 mg, 3.12 mmol, 1.20 eq), 4-chloro-7H-pyrrolo[2,3-d]pyrimidine (400 mg, 2.46 mmol, 1.00 eq), Pd(OAc)₂ (12.0 mg, 52.4 μmol, 0.02 eq), XPhos (75.0 mg, 157 μmol, 0.06 eq), K₂CO₃ (525 mg, 3.80 mmol, 1.45 eq) in 4 mL *t*BuOH and 3 μL water was stirred at 110 °C for 6 hours. The product **72** was isolated by silica column chromatography [30% EtOAc/ 1% TEA /pentane] as light yellow solid (259 mg, 921 μmol, 35%).

¹H-NMR (DMSO, 400 MHz) δ = 11.48 (s, 1H, NH), 9.08 (s, 1H, NH), 8.01 (d, *J* = 5.4 Hz, 1H, H-2), 7.90 (d, *J* = 8.8 Hz, 2H, H-2'), 7.34 (d, *J* = 8.8 Hz, 2H, H-3'), 7.28 (dd, *J* = 3.4, 2.1 Hz, 1H, H-8), 6.89 (d, *J* = 5.4 Hz, 1H, H-3), 6.57 (dd, *J* = 3.5, 1.6 Hz, 1H, H-9), 4.28 (q, *J* = 7.1 Hz, 2H, CH₂), 1.31 (t, *J* = 7.1 Hz, 3H, CH₃).

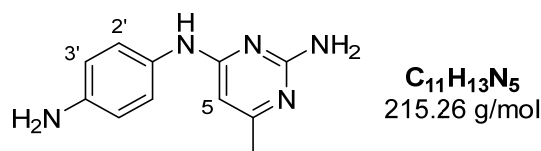
¹³C-NMR (DMSO, 101 MHz) δ = 165.46, 150.00, 146.50, 143.74, 141.37, 130.67, 123.09, 121.64, 117.46, 110.11, 101.27, 98.33, 60.17, 14.29.

General procedure C: reduction of nitroaryls



To a stirred solution of nitro compound (1.00 eq) in EtOH was added 10 wt% palladium on carbon (1.10 eq) and ammonium formate (10.0 eq). The reaction mixture was stirred at room temperature for 1 – 24 hours. Upon completion of the reaction, the suspension was filtered over Celite and washed with EtOAc or DCM. The filtrate was concentrated and dissolved in EtOAc and sat. NaHCO₃ solution. The aqueous phase was washed with EtOAc (3 x 100 mL) and the combined organic layer was dried over MgSO₄, filtered and concentrated.

N⁴-(4-Aminophenyl)-6-methylpyrimidine-2,4-diamine (73)



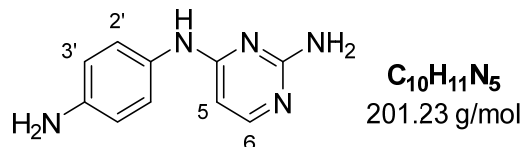
Following general procedure C, **51** (591 mg, 2.40 mmol, 1.00 eq), ammonium formate (1.73 g, 27.4 mmol, 11.4 eq) and 10 wt% Pd/C (450 mg, 4.23 mmol, 1.76 eq) in 25 mL ethanol was stirred at room temperature for 4.5 hours. The residue was filtrated, washed with DCM and concentrated. Extraction with EtOAc afforded pure compound **73** as brown-orange solid (536 mg, 2.11 mmol, 88%).

TLC: $R_f = 0.48$ (5% MeOH/1% TEA/DCM) [UV].

¹H-NMR (DMSO-*d*₆, 400 MHz) $\delta = 8.36$ (s, 1H, NH), 7.14 (d, $J = 8.61$ Hz, 2H, H-2'), 6.51 (d, $J = 8.61$ Hz, 2H, H-3'), 5.87 (s, 2H, NH₂), 5.67 (s, 1H, H-5), 4.81 (s, 2H, NH₂), 2.01 (s, 3H, CH₃).

These data are in accordance with previous literature reports.³

N⁴-(4-Aminophenyl)pyrimidine-2,4-diamine (74)



Following general procedure C, **52** (226 mg, 977 μ mol, 1.00 eq), ammonium formate (702 mg, 11.1 mmol, 11.4 eq) and 10 wt% Pd/C (182 mg, 1.71 mmol, 1.76 eq) were stirred in 10 mL ethanol at room temperature for 5 hours. The residue was filtrated, washed with EtOAc and concentrated. Extraction with EtOAc afforded pure compound **74** as brown-orange solid (154 mg, 762 μ mol, 78%).

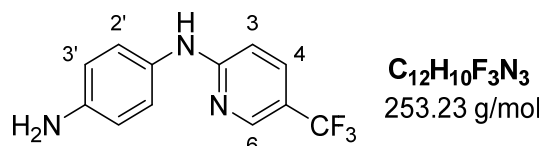
TLC: $R_f = 0.20$ (5% MeOH/1% TEA/DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.51 (s, 1H, NH), 7.67 (d, J = 5.80 Hz, 1H, H-6), 7.17 (d, J = 8.60 Hz, 2H, H-2'), 6.51 (d, J = 8.60 Hz, 2H, H-3'), 5.93 (s, 2H, NH₂), 5.80 (d, J = 5.80 Hz, 1H, H-5), 4.81 (bs, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 163.13, 161.54, 155.80, 144.48, 129.08, 122.80, 114.14, 95.39.

HR-MS m/z calcd. for C₁₀H₁₂N₆⁺ (M + 1)⁺ 202.1093, found 202.1095.

N¹-(5-(Trifluoromethyl)pyridin-2-yl)benzene-1,4-diamine (75)



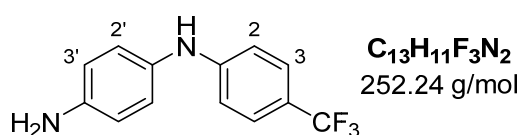
Following general procedure C, **53** (63.0 mg, 222 μ mol, 1.00 eq), ammonium formate (160 mg, 2.53 mmol, 11.4 eq) and 10 wt% Pd/C (41 mg, 387 μ mol, 1.76 eq) in 5 mL ethanol were stirred at room temperature overnight. The residue was filtrated, washed with EtOAc and concentrated. Extraction with EtOAc afforded pure compound **75** as brown solid (55.0 mg, 218 μ mol, 98%).

¹H-NMR (MeOD, 400 MHz) δ = 8.25 (s, 1H, H-6), 7.64 (dd, J = 8.92 Hz, 1H, H-4), 7.19 (d, J = 8.65 Hz, 2H, H-2'), 6.75 (d, J = 8.65 Hz, 2H, H-3'), 6.70 (d, J = 8.92 Hz, 1H, H-3).

¹³C-NMR (MeOD, 101 MHz) δ = 161.03, 146.36, 145.38, 135.36, 131.75, 124.92, 117.24, 109.46.

HR-MS m/z calcd. for C₁₂H₁₁H₃N₃⁺ (M + 1)⁺ 254.0900, found 254.0902.

N¹-(4-(Trifluoromethyl)phenyl)benzene-1,4-diamine (76)



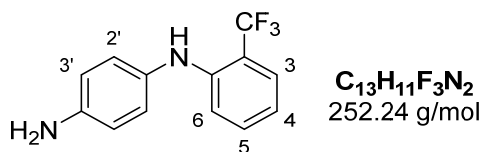
Following general procedure C, **54** (317 mg, 1.12 mmol, 1.00 eq), ammonium formate (777 mg, 1.90 mmol, 11.0 eq) and 10 wt% Pd/C (202 mg, 1.90 mmol, 1.76 eq) were stirred in 10 mL ethanol at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. Compound **76** was obtained as brown solid (269 mg, 1.06 mmol, 95%).

¹H-NMR (DMSO, 400 MHz) δ = 8.08 (s, 1H, NH), 7.38 (d, J = 8.6 Hz, 2H, H-3), 6.86 (d, J = 8.6 Hz, 2H, C-2'), 6.80 (d, J = 8.5 Hz, 2H, C-2), 6.57 (d, J = 8.6 Hz, 2H, C-3'), 4.93 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 150.87, 145.78, 129.82, 126.76, 124.67, 115.10, 112.81.

HR-MS m/z calcd. for C₁₃H₁₂F₃N₂⁺ (M + 1)⁺ 253.0947, found 253.0984.

***N'*-(2-(Trifluoromethyl)phenyl)benzene-1,4-diamine (77)**



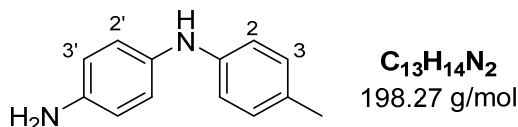
Following general procedure C, **55** (360 mg, 1.27 mmol, 1.00 eq), ammonium formate (880 mg, 14.0 mmol, 11.0 eq) and 10 wt% Pd/C (162 mg, 1.53 mmol, 1.20 eq) in 10 mL ethanol were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. Compound **77** (306 mg, 1.22 mmol, 96%) was obtained as brown liquid.

¹H-NMR (DMSO, 400 MHz) δ = 7.45 (d, *J* = 6.1 Hz, 1H, H-3), 7.29 (t, *J* = 7.8 Hz, 1H, H-4), 6.92 (s, 1H, NH), 6.85 (d, *J* = 8.5 Hz, 2H, H-2'), 6.80 – 6.67 (m, 2H, H-5, H-6), 6.57 (d, *J* = 8.5 Hz, 2H, H-3'), 4.98 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 145.88, 145.44, 133.06, 129.41, 126.41, 126.35, 126.15, 116.60, 115.03, 114.56.

HR-MS *m/z* calcd. for C₁₃H₁₂F₃N₂⁺ (*M* + 1)⁺ 253.0947, found 253.0949.

***N'*-(*p*-Tolyl)benzene-1,4-diamine (78)**



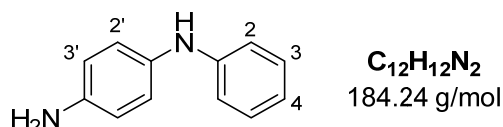
Following general procedure C, **56** (365 mg, 1.60 mmol, 1.00 eq), 10 wt% Pd/C (187 mg, 1.76 mmol, 1.10 eq) and ammonium formate (1.00 g, 16.0 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **78** (216 mg, 1.09 mmol, 68%) as light brown solid.

¹H-NMR (DMSO, 400 MHz) δ = 8.25 (s, 1H, NH), 7.28 (s, 1H, NH₂), 6.90 (d, *J* = 8.4 Hz, 2H, H-2'), 6.78 (d, *J* = 8.6 Hz, 2H), 6.70 (d, *J* = 8.4 Hz, 2H), 6.51 (d, *J* = 8.6 Hz, 1H), 2.16 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 167.91, 163.99, 144.09, 143.33, 132.36, 129.36, 125.62, 121.69, 114.75, 114.14, 20.14.

HR-MS *m/z* calcd. for C₁₃H₁₅N₂⁺ (*M* + 1)⁺ 199.1230, found 199.1234.

***N'*-Phenylbenzene-1,4-diamine (79)**



Following general procedure C, **57** (344 mg, 1.61 mmol, 1.00 eq), 10 wt% Pd/C (188 mg, 1.77 mmol, 1.10 eq), and ammonium formate (1.02 g, 16.1 mmol, 10.0 eq) in 13 mL EtOH

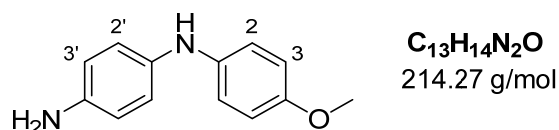
were stirred at room temperature overnight. The reaction mixture was filtrated, washed with DCM and concentrated to obtain **79** (279 mg, 1.50 mmol, 93%) as brown oil.

¹H-NMR (DMSO, 400 MHz) δ = 7.44 (s, 1H, NH), 7.21 – 6.96 (m, 2H, H-3), 6.82 (d, J = 8.6 Hz, 2H, H-2'), 6.76 (d, J = 7.5 Hz, 2H, H-2), 6.59 (t, J = 7.2 Hz, 1H, H-4), 6.53 (d, J = 8.5 Hz, 2H, H-3'), 4.75 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 146.72, 143.87, 131.55, 128.94, 122.55, 116.96, 114.71, 113.60.

HR-MS m/z calcd. for C₁₂H₁₃N₂⁺ (M + 1)⁺ 185.1073, found 185.1075.

N¹-(4-Methoxyphenyl)benzene-1,4-diamine (80)



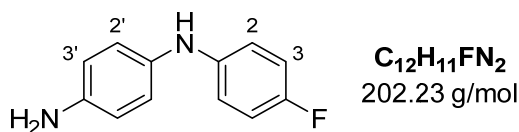
Following general procedure C, **58** (296 mg, 1.21 mmol, 1.00 eq), 10 wt% Pd/C (142 mg, 1.33 mmol, 1.10 eq), and ammonium formate (762 mg, 12.1 mmol, 10.0 eq) in 15 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **80** (249 mg, 1.21 mmol, 96%) as brown solid.

¹H-NMR (DMSO, 400 MHz) δ = 8.18 (s, 1H, NH), 7.13 (s, 2H, NH₂), 6.82 – 6.64 (m, 6H, H-2', H-2, H-3), 6.50 (d, J = 8.6 Hz, 2H, H-3'), 3.65 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 163.42, 151.84, 142.75, 139.94, 133.49, 120.56, 116.10, 114.87, 114.50, 55.25.

HR-MS m/z calcd. for C₁₃H₁₅N₂O⁺ (M + 1)⁺ 215.1179, found 215.1184.

N¹-(4-Fluorophenyl)benzene-1,4-diamine (81)



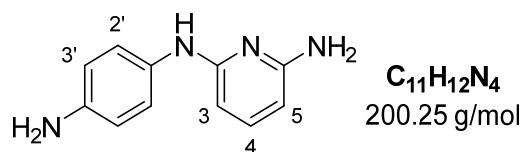
Following general procedure C, **59** (285 mg, 1.23 mmol, 1.00 eq), 10 wt% Pd/C (144 mg, 1.35 mmol, 1.10 eq), and ammonium formate (775 mg, 12.3 mmol, 10.0 eq) in 10 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **81** (193 mg, 1.23 mmol, 77%) as brown solid.

¹H-NMR (DMSO, 400 MHz) δ = 7.40 (s, 1H, NH), 6.93 (t, J = 8.9 Hz, 2H, H-2), 6.79 (d, J = 8.6 Hz, 2H, H-2'), 6.77 – 6.70 (m, 2H, H-3), 6.53 (d, J = 8.6 Hz, 2H, H-3').

¹³C-NMR (DMSO, 101 MHz) δ = 163.51, 156.00, 153.69, 143.79, 143.29, 132.01, 122.14, 115.45, 115.23, 114.83, 114.75.

HR-MS m/z calcd. for C₁₂H₁₂FN₂⁺ (M + 1)⁺ 203.0979, found 203.0982.

N'-(4-Aminophenyl)pyridine-2,6-diamine (**82**)



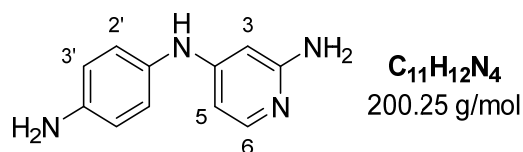
Following general procedure C, **60** (195 mg, 850 μ mol, 1.00 eq), 10 wt% Pd/C (99.0 mg, 930 μ mol, 1.10 eq), and ammonium formate (536 mg, 8.50 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. Extraction with EtOAc and sat. $NaHCO_3$ solution afforded **82** (156 mg, 773 μ mol, 91%) as dark red solid.

TLC: R_f = 0.52 (5% MeOH/1% TEA/ DCM) [UV].

1H -NMR (DMSO, 400 MHz) δ = 7.80 (s, 1H, NH), 7.14 (d, J = 8.6 Hz, 2H, H-2'), 7.06 (t, J = 7.8 Hz, 1H, H-4), 6.49 (d, J = 8.6 Hz, 1H, H-3'), 5.78 (d, J = 7.9, 1H, H-3), 5.70 (d, J = 7.7 Hz, 1H, H-5), 5.44 (s, 2H, NH_2), 4.68 (s, 2H, NH_2).

^{13}C -NMR (DMSO, 101 MHz) δ = 158.98, 156.82, 143.51, 138.50, 131.82, 122.07, 114.73, 96.40, 95.88.

N'-(4-Aminophenyl)pyridine-2,4-diamine (**83**)

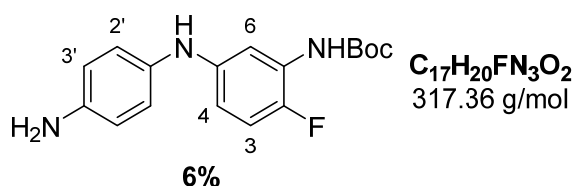
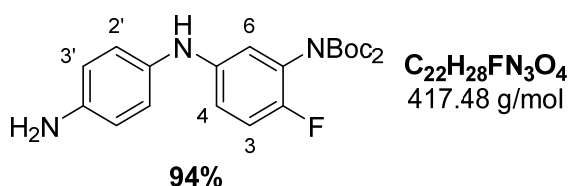


Following general procedure C, **61** (257 mg, 1.11 mmol, 1.00 eq), 10 wt% Pd/C (131 mg, 1.23 mmol, 1.10 eq), and ammonium formate (700 mg, 11.1 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **83** (206 mg, 1.02 mmol, 92%) as brown solid.

1H -NMR (DMSO, 400 MHz) δ = 7.78 (s, 1H, NH), 7.48 (d, J = 5.8 Hz, 1H, H-6), 6.81 (d, J = 8.5 Hz, 2H, H-2'), 6.55 (d, J = 8.5 Hz, 2H, H-3'), 5.93 (dd, J = 5.9, 2.1 Hz, 1H, H-5), 5.74 (d, J = 2.0 Hz, 1H, H-3), 5.38 (s, 2H, NH_2), 4.89 (s, 2H, NH_2).

^{13}C -NMR (DMSO, 101 MHz) δ = 160.36, 153.53, 147.37, 145.14, 129.27, 124.45, 114.49, 100.59, 89.02.

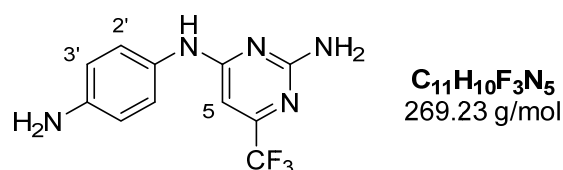
tert-Butyl (5-((4-aminophenyl)amino)-2-fluorophenyl)(*tert*-butoxycarbonyl)carbamate & *tert*-butyl (5-((4-aminophenyl)amino)-2-fluorophenyl)carbamate (**84**)



Following general procedure C, **62** (175 mg, 410 μmol , 1.00 eq), 10 wt% Pd/C (52.3 mg, 49.0 μmol , 1.10 eq), and ammonium formate (259 mg, 4.11 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **84** (158 mg, 377 μmol , 92%) as brown solid.

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 7.49 (s, 1H, NH), 7.02 (t, J = 9.4 Hz, 1H, H-6), 6.78 (d, J = 8.6 Hz, 2H, H-2'), 6.74 – 6.68 (m, 1H, H-3), 6.60 – 6.51 (m, 3H, H-4, H-3'), 4.80 (s, 2H, NH_2), 1.38 (s, 18H, Boc_2).

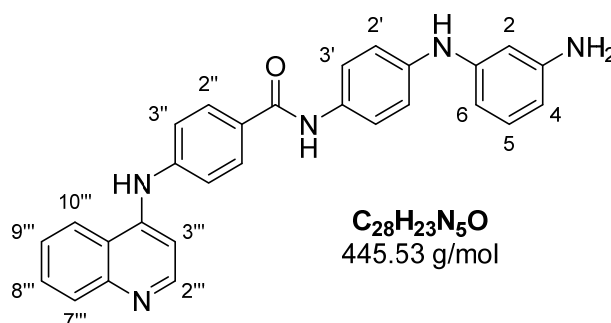
***N*⁴-(4-Aminophenyl)-6-(trifluoromethyl)pyrimidine-2,4-diamine (85)**



Following general procedure C, **64** (235 mg, 790 μmol , 1.00 eq), 10 wt% Pd/C (92.0 mg, 860 μmol , 1.10 eq), and ammonium formate (497 mg, 7.90 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **85** (175 mg, 648 μmol , 82%) as grey solid.

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 9.08 (s, 1H, NH), 7.23 (br. s, 2H, H-2'), 6.60 (s, 2H, NH_2), 6.53 (d, J = 8.7 Hz, 2H, H-3'), 6.15 (br.s, 1H, H-5), 4.91 (br.s, 2H, NH_2).

***N*-(4-((3-Aminophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (39)**



Following general procedure C, **86** (320 mg, 670 μmol , 1.00 eq), 10 wt% Pd/C (78.0 mg, 737 μmol , 1.10 eq), and ammonium formate (422 mg, 6.70 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. The crude product was purified by silica gel column chromatography [2% MeOH \rightarrow 5% MeOH/1% TEA/DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to obtain **39** (71.0 mg, 154 μmol , 23%) as yellowish solid.

TLC: R_f = 0.28 (5% MeOH/1% TEA/ DCM) [UV].

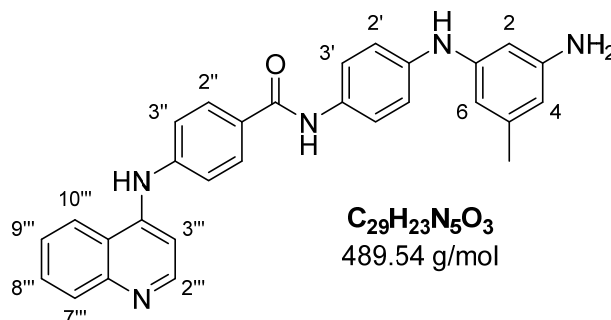
$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 9.97 (s, 1H, NH), 9.23 (s, 1H, NH), 8.57 (d, J = 5.2 Hz, 1H, H-2''), 8.39 (d, J = 8.4 Hz, 1H, H-10'''), 8.00 (d, J = 8.6 Hz, 2H, H-3'''), 7.93 (d, J = 8.4 Hz, 1H, H-7'''), 7.75 (s, 1H, NH), 7.72 (t, J = 8.0 Hz, 1H, H-8'''), 7.61 (d, J = 8.8 Hz, 2H, H-3'), 7.57 (t, J =

8.4 Hz, 1H, H-9'''), 7.47 (d, J = 8.4 Hz, 2H, H-2''), 7.19 (d, J = 5.2 Hz, 1H, H-3'''), 7.03 (d, J = 8.9 Hz, 2H, H-2'), 6.85 (t, J = 7.9 Hz, 1H, H-5), 6.33 (s, 1H, H-2), 6.22 (d, J = 7.8 Hz, 1H, H-6), 6.04 (d, J = 7.9 Hz, 1H, H-4), 4.95 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 164.29, 149.43, 144.53, 139.74, 131.45, 129.46, 129.35, 128.98, 128.87, 124.98, 122.29, 121.64, 120.36, 119.76, 117.37, 106.00, 104.84, 101.74.

HR-MS m/z calcd. for C₂₈H₂₄N₅O⁺ (M + 1)⁺ 446.1975, found 446.2001.

N-(4-((3-Amino-5-methylphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (43)



Following general procedure C, **87** (149 mg, 300 μmol, 1.00 eq), 10 wt% Pd/C (36.0 mg, 330 μmol, 1.10 eq), and ammonium formate (189 mg, 3.00 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. The crude product was purified by silica gel column chromatography [2% MeOH → 3% MeOH/1% TEA/DCM] to obtain **43** (67.0 mg, 144 μmol, 48%) as yellowish solid.

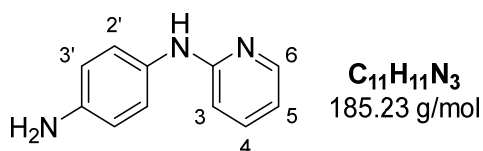
TLC: R_f = 0.33 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.97 (s, 1H, NH), 9.22 (s, 1H, NH), 8.58 (d, J = 5.3 Hz, 1H, H-2'''), 8.39 (d, J = 8.4 Hz, 1H, H-10'''), 8.00 (d, J = 8.6 Hz, 2H, H-3'''), 7.93 (d, J = 8.4 Hz, 1H, H-7'''), 7.73 (t, J = 7.6 Hz, 1H, H-8'''), 7.68 (s, 1H, NH), 7.59 (d, J = 8.9 Hz, 2H, H-3'), 7.56 (t, J = 8.4 Hz, 1H, H-9''') 7.47 (d, J = 8.3 Hz, 2H, H-2''), 7.19 (d, J = 5.2 Hz, 1H, H-3'''), 7.02 (d, J = 8.9 Hz, 2H, H-2'), 6.15 (s, 1H, H-2), 6.05 (s, 1H, H-6*), 5.89 (s, 1H, H-4*), 4.84 (s, 2H, NH₂), 2.09 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 164.30, 150.70, 149.24, 149.01, 146.39, 144.46, 139.84, 138.26, 131.38, 129.44, 129.25, 128.98, 128.85, 124.97, 122.28, 121.64, 120.38, 119.74, 117.47, 106.94, 105.82, 103.56, 99.27, 21.43.

HR-MS m/z calcd. for C₂₉H₂₄N₅O₃⁺ (M + 1)⁺ 460.2131, found 460.2157.

N¹-(Pyridin-2-yl)benzene-1,4-diamine (88)

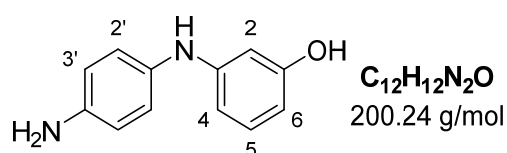


Following general procedure C, **67** (506 mg, 2.35 mmol, 1.00 eq), 10 wt% Pd/C (300 mg, 2.82 mmol, 1.20 eq), and ammonium formate (1.48 g, 23.5 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature for three hours. The reaction mixture was filtrated, washed with EtOAc and concentrated to obtain **88** (445 mg, 2.35 mmol, 100%) as solid.

¹H-NMR (DMSO, 400 MHz) δ = 8.37 (s, 1H, NH), 8.08 – 7.98 (m, 1H, H-6), 7.46 – 7.34 (m, 1H, H-4), 7.20 (d, J = 8.7 Hz, 2H, H-2'), 6.62 (d, J = 8.4 Hz, 1H, H-3), 6.59 – 6.55 (m, 1H, H-5), 6.53 (d, J = 8.7 Hz, 2H, H-3').

¹³C-NMR (DMSO, 101 MHz) δ = 157.09, 147.42, 143.51, 136.83, 130.60, 121.58, 114.24, 112.67, 108.70, 40.15, 39.99, 39.94, 39.78, 39.73, 39.52, 39.31, 39.10, 38.89.

3-((4-Aminophenyl)amino)phenol (**89**)

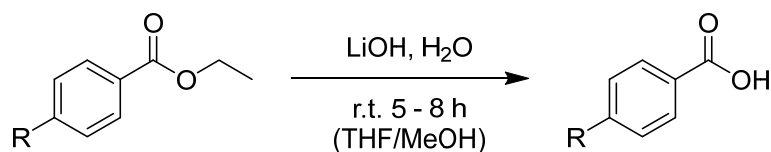


Following general procedure C, **68** (704 mg, 3.06 mmol, 1.00 eq), 10 wt% Pd/C (390 mg, 3.67 mmol, 1.20 eq), and ammonium formate (1.92 g, 30.6 mmol, 10.0 eq) in 20 mL EtOH were stirred at room temperature overnight. The reaction mixture was filtrated, washed with EtOAc and concentrated. The crude product was further purified by precipitation with isopropanol/ water to obtain **89** (250 mg, 1.25 mmol, 40%) as brown solid.

¹H-NMR (DMSO, 400 MHz) δ = 8.93 (s, 1H, OH), 7.32 (s, 1H, NH), 6.85 (t, J = 8.2 Hz, 1H, H-5), 6.80 (d, J = 8.6 Hz, 2H, H-2'), 6.52 (d, J = 8.5 Hz, 2H, H-3'), 6.24 – 6.13 (m, 2H, H-2, H-4), 6.05 – 5.91 (m, 1H, H-6), 4.74 (s, 2H, NH₂).

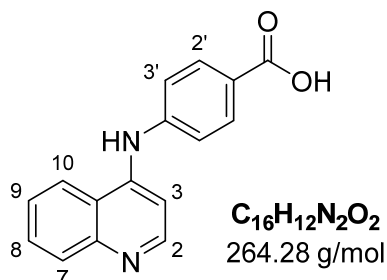
¹³C-NMR (DMSO, 101 MHz) δ = 158.12, 148.05, 143.83, 131.65, 129.52, 122.83, 114.63, 105.18, 104.46, 100.41.

General procedure D: ester hydrolysis



Ethyl benzoate compound (1.00 eq) was dissolved in 7 mL anhydrous THF/MeOH (1:1 v/v). Lithium hydroxide (10.0 eq) and 8.5 mL water were added, and the reaction mixture was stirred at room temperature for 5 – 8 hours. The mixture was acidified with 1 M hydrochloric acid below pH 4. The obtained precipitate was filtered, washed with cold water and dried at 50 °C under reduced pressure.

4-(Quinolin-4-ylamino)benzoic acid (**90**)

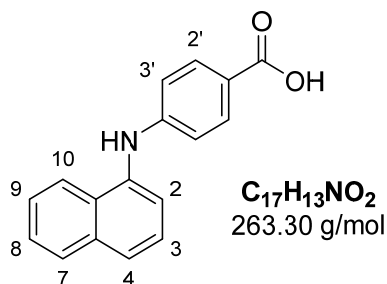


Following general procedure D, a mixture of ethyl 4-(quinolin-4-ylamino)benzoate (**50**) (669 mg, 2.29 mmol, 1.00 eq), LiOH (548 mg, 22.9 mmol, 10.0 eq) in 20 mL THF/MeOH (1:1 v/v) and 23 mL water were stirred at room temperature for five hours. Precipitation with 1 M hydrochloric acid solution afforded product **90** as a yellowish solid (571 mg, 2.15 mmol, 94%).
TLC: $R_f = 0.05$ (1% MeOH/1% TEA/DCM) [UV].

¹H-NMR (DMSO-*d*₆, 400 MHz) $\delta = 11.02$ (br.s, 1H, OH), 8.81 (d, $J = 8.49$ Hz, 1H, H-2), 8.59 (d, $J = 6.9$ Hz, 1H, H-10), 8.11 (d, $J = 8.52$ Hz, 2H, H-2'), 8.10 (d, $J = 8.41$ Hz, 1H, H-7), 8.04 (t, $J = 8.41$ Hz, 1H, H-8), 7.83 (t, $J = 7.70$ Hz, 1H, H-9), 7.63 (d, $J = 8.52$ Hz, 2H, H-3'), 7.06 (d, $J = 6.9$ Hz, 1H, H-3).

These data are in accordance with previous literature reports.³

4-(Naphthalen-1-ylamino)benzoic acid (**91**)

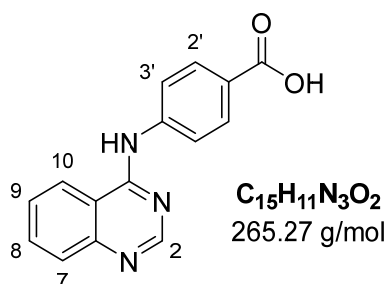


Following general procedure D, a mixture of ethyl 4-(quinazolin-4-ylamino)benzoate (**70**) (596 mg, 2.04 mmol, 1.00 eq), LiOH (487 mg, 20.4 mmol, 10.0 eq) in 16 mL THF/MeOH (1:1 v/v) and 23 mL water were stirred at room temperature for five hours. Precipitation with 1 M hydrochloric acid solution afforded product **91** as a white solid (430 mg, 1.63 mmol, 80%).

¹H-NMR (DMSO-*d*₆, 400 MHz) $\delta = 9.18$ (s, 1H, NH), 8.48 (d, $J = 7.5$ Hz, 1H, H-10), 8.36 (d, $J = 7.3$ Hz, 1H, H-7), 8.17 (d, $J = 8.7$ Hz, 2H, H-2'), 8.13 (d, $J = 7.9$ Hz, 1H, H-4), 7.99 – 7.89 (m, 3H, H-3, H-8, H-9), 7.87 (d, $J = 7.5$, 1H, H-2), 7.34 (d, $J = 8.7$ Hz, 2H, H-3').

These data are in accordance with previous literature reports.⁶

4-(Quinazolin-4-ylamino)benzoic acid (**92**)

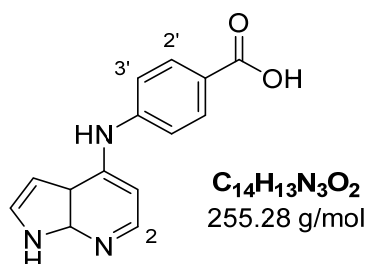


Following general procedure D, a mixture of ethyl 4-(quinazolin-4-ylamino)benzoate (**71**) (530 mg, 1.81 mmol, 1.00 eq), LiOH (433 mg, 18.1 mmol, 10.0 eq) in 12 mL THF/MeOH (1:1 v/v) and 10 mL water were stirred at room temperature for five hours. Precipitation with 1 M hydrochloric acid solution afforded product **92** as a white solid (460 mg, 1.73 mmol, 96%).

¹H-NMR (DMSO-*d*₆, 400 MHz) δ = 11.94 (s, 1H, OH), 9.08 (d, *J* = 8.4 Hz, 1H, H-2), 9.00 (s, 1H, NH), 8.12 (t, *J* = 7.1 Hz, 1H, H-10), 8.07 – 8.01 (m, 3H, H-10, H-2'), 7.96 (d, *J* = 7.7 Hz, 2H, H-3'), 7.87 (t, *J* = 7.7 Hz, 1H, H-9).

¹³C-NMR (DMSO, 101 MHz) δ = 166.68, 159.82, 151.13, 140.88, 136.26, 129.89, 128.65, 128.22, 124.93, 124.11, 120.22, 113.77.

4-((3a,7a-Dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzoic acid (**93**)

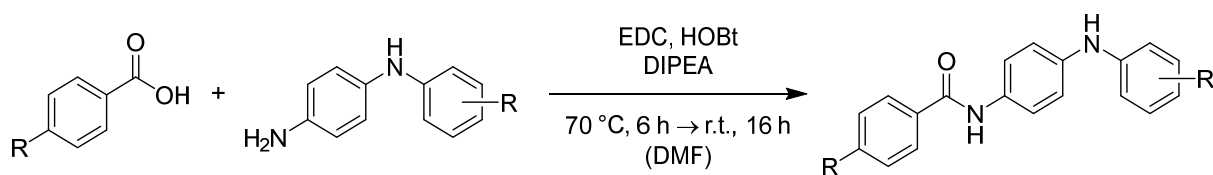


Following general procedure D, a mixture of ethyl ethyl 4-((3a,7a-dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzoate (**72**) (259 mg, 921 μ mol, 1.00 eq), LiOH (216 mg, 9.03 mmol, 10.0 eq) in 8 mL THF/MeOH (1:1 v/v) and 8 mL water were stirred at room temperature for five hours. Precipitation with 1 M hydrochloric acid solution afforded product **93** as a white solid (201 mg, 79.0 μ mol, 88%).

¹H-NMR (DMSO, 400 MHz) δ = 12.61 (s, 1H, NH), 10.66 (s, 1H, NH), 8.08 (d, *J* = 6.9 Hz, 1H, H-2), 8.04 (d, *J* = 8.5 Hz, 2H, H-2'), 7.54 (d, *J* = 8.6 Hz, 2H, H-3'), 7.42 (dd, *J* = 3.6, 2.3 Hz, 1H, H-8), 7.06 – 7.01 (m, 1H, H-3), 6.94 (d, *J* = 6.9 Hz, 1H, H-9).

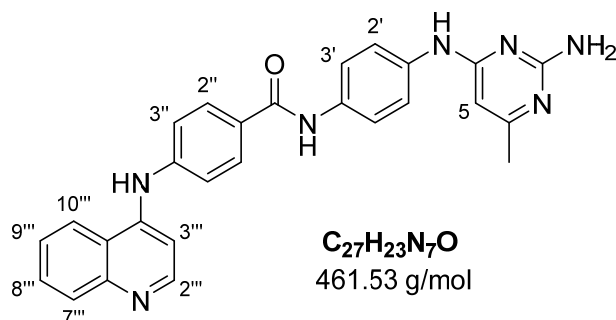
¹³C-NMR (DMSO, 101 MHz) δ = 166.70, 149.26, 142.37, 138.12, 134.97, 130.85, 127.38, 124.45, 122.73, 122.64, 110.22, 101.16, 98.85.

General procedure E: amide coupling



Benzoic acid (1.00 eq) and aryl amine (1.00 eq) were dissolved in 5 mL anhydrous DMF. DIPEA (2.50 eq), EDC hydrochloride (2.00 eq) and HOBT (1.20 eq) were added, and the reaction mixture was stirred at 70 °C for six hours and at room temperature overnight. The solvent was removed under reduced pressure and crude product was purified by silica gel column chromatography. Unless otherwise stated, final compounds were further purified by precipitation in an isopropanol/water mixture. Therefore, the product was dissolved in isopropanol (1 - 2 mL) under heat exposure and subsequently placed on ice. The pure product was precipitated by the addition of 15 – 20 mL water. The suspension was centrifuged (20 min, 4000 rpm, 4 °C), washed once with 10 mL water and finally dried under reduced pressure.

N-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (1)



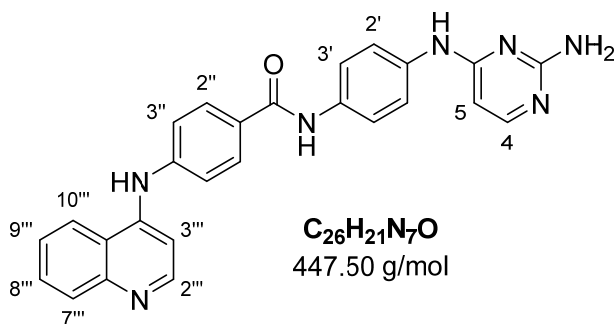
Following general procedure E, **90** (49.0 mg, 185 μ mol, 1.00 eq), **73** (40.0 mg, 185 μ mol, 1.00 eq), EDC hydrochloride (71.0 mg, 370 μ mol, 2.00 eq), HOBT (30.0 mg, 222 μ mol, 1.20 eq), and TEA (85.0 μ L, 62.5 mg, 463 μ mol, 2.50 eq) in 2 mL DMF were stirred at 70 °C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [2% MeOH/1% TEA/DCM \rightarrow 10% MeOH/1% TEA/DCM] to yield **1** (42.0 mg, 90.7 μ mol, 49%) as a light orange solid.

¹H-NMR (DMSO, 400 MHz) δ = 10.08 (s, 1H, NH), 9.26 (s, 1H, NH), 8.98 (s, 1H, NH), 8.57 (d, J = 5.22 Hz, 1H, N-2'''), 8.37 (d, J = 8.30 Hz, 1H, H-10'''), 8.00 (d, J = 8.71 Hz, 2H, H-3'''), 7.93 (d, J = 8.5 Hz, 1H, H-7'''), 7.74 (t, J = 7.35 Hz, 1H, H-8'''), 7.66 (s, 4H, H-2', H-3'), 7.58 (t, J = 7.35 Hz, 1H, H-9'), 7.47 (d, J = 8.71 Hz, 2H, H-2''), 7.20 (d, J = 5.22 Hz, 1H, H-3'''), 6.11 (s, 2H, NH₂), 5.86 (s, 1H, H-5), 2.08 (s, 3H, CH₃).

HR-MS m/z calcd. for C₂₇H₂₄N₇O⁺ (M + 1)⁺ 462.2023, found 462.2045.

These data are in accordance with previous literature reports.³

***N*-4-((2-Aminopyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (26)**



Following general procedure E, **90** (92.0 mg, 347 μ mol, 1.00 eq), **74** (70.0 mg, 347 μ mol, 1.00 eq), TEA (120 μ L, 88.0 mg, 868 μ mol, 2.50 eq), EDC hydrochloride (133 mg, 694 μ mol, 2.00 eq), and HOBt (56.0 mg, 416 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/DCM \rightarrow 10% MeOH/1% TEA/DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **26** (42.0 mg, 90.7 μ mol, 49%) as a light yellow solid.

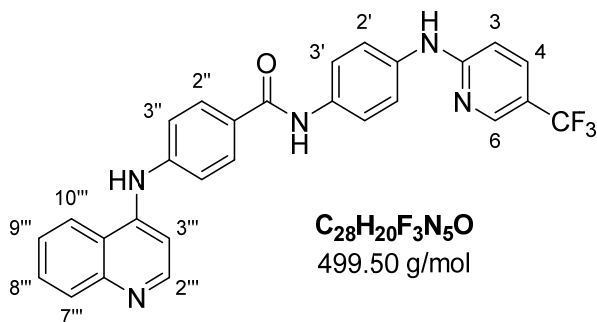
TLC: R_f = 0.37 (10% MeOH/1% TEA/DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.06 (s, 1H, NH), 9.23 (s, 1H, NH), 9.05 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.38 (d, J = 8.4 Hz, 1H, H-10'''), 8.01 (d, J = 8.3 Hz, 2H, H-3'''), 7.93 (d, J = 8.3 Hz, 1H, H-7'''), 7.79 (d, J = 5.7 Hz, 1H, H-4), 7.74 (t, J = 7.7 Hz, 1H, H-8'''), 7.69 (s, 4H, H-3', H-2'), 7.58 (t, J = 7.6 Hz, 1H, H-9'''), 7.48 (d, J = 8.3 Hz, 2H, H-2''), 7.20 (d, J = 5.2 Hz, 1H, H-3'''), 6.18 (s, 2H, NH₂), 5.98 (d, J = 5.7 Hz, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 164.52, 163.04, 160.79, 156.20, 150.76, 149.08, 146.33, 144.22, 136.38, 133.37, 129.46, 129.31, 129.08, 128.72, 125.02, 122.30, 120.82, 120.41, 119.69, 119.66, 103.67, 96.44.

HR-MS m/z calcd. for C₂₆H₂₂N₇O⁺ ($M + 1$)⁺ 448.1880, found 448.1884.

4-(Quinolin-4-ylamino)-*N*-4-((5-(trifluoromethyl)pyridin-2-yl)amino)phenyl)benzamide (27)



Following general procedure E, **90** (52.2 mg, 197 μ mol, 1.00 eq), **75** (50.0 mg, 197 μ mol, 1.00 eq), TEA (68.0 μ L, 50.0 mg, 493 μ mol, 2.50 eq), EDC hydrochloride (76.0 mg, 394 μ mol, 2.00 eq), and HOBt (32.0 mg, 236 μ mol, 1.20 eq) in 3 mL DMF were heated to 70°C for 6 hours

and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/DCM → 4% MeOH/1% TEA/DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **27** (7.00 mg, 14.1 μmol, 7%) as a light yellow solid.

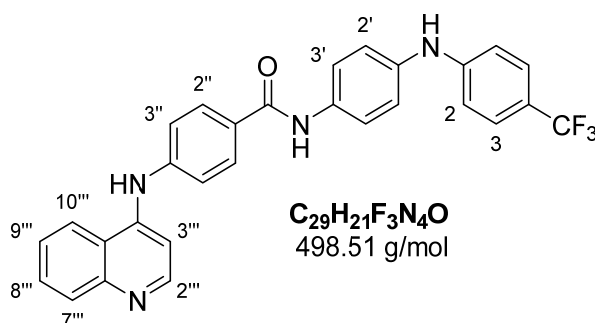
TLC: R_f = 0.24 (5% MeOH/1% TEA/DCM) [UV].

¹H-NMR (MeOD, 400 MHz) δ = 8.49 (d, J = 5.7 Hz, 1H, H-2''), 8.38 (s, 1H, H-6), 8.35 (s, 1H, H-10'''), 8.04 (d, J = 8.6 Hz, 2H, H-3''), 7.94 (dd, J = 8.5, 1.2 Hz, 1H, H-7'''), 7.85 – 7.77 (m, 1H, H-8'''), 7.73 (dd, J = 8.9, 2.6 Hz, 1H, H-4), 7.69 – 7.58 (m, 5H, H-2', H-3', H-9'''), 7.53 (d, J = 8.7 Hz, 2H, H-2''), 7.23 (d, J = 5.7 Hz, 1H, H-3'''), 6.87 (d, J = 8.9 Hz, 1H, H-3).

¹³C-NMR (MeOD, 101 MHz) δ = 168.11, 160.19, 151.09, 150.12, 148.18, 146.55, 145.18, 138.46, 135.46, 135.00, 132.15, 131.80, 130.50, 127.95, 127.35, 123.30, 123.28, 122.82, 121.83, 121.63, 111.04, 104.11.

HR-MS m/z calcd. for $C_{28}H_{21}F_3N_5O^+$ ($M + 1$)⁺ 500.1693, found 500.1694.

4-(Quinolin-4-ylamino)-*N*-(4-((4-(trifluoromethyl)phenyl)amino)phenyl)benzamide (**28**)



Following general procedure E, **90** (73.0 mg, 277 μmol, 1.00 eq), **76** (70.0 mg, 277 μmol, 1.00 eq), TEA (95.0 μL, 70.0 mg, 690 μmol, 2.50 eq), EDC hydrochloride (106 mg, 554 μmol, 2.00 eq), and HOBt (45.0 mg, 330 μmol, 1.20 eq) in 3 mL DMF were heated to 70°C for 4 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/DCM → 4% MeOH/1% TEA/DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **28** (11.0 mg, 22.1 μmol, 8%) as a light yellow solid.

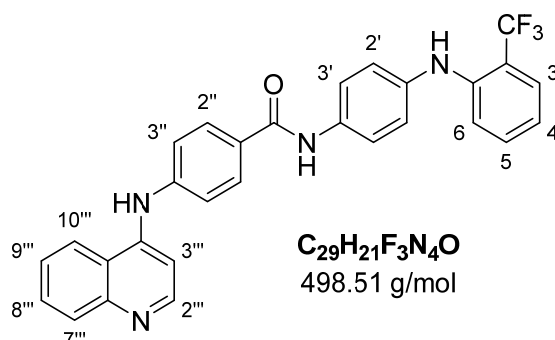
TLC: R_f = 0.33 (5% MeOH/1% TEA/DCM) [UV].

¹H-NMR (MeOD, 400 MHz) δ = 8.49 (d, J = 5.7 Hz, 1H, H-2'''), 8.37 (d, J = 8.5 Hz, 1H, H-10'''), 8.04 (d, J = 8.7 Hz, 2H, H-3'''), 7.94 (d, J = 8.5 Hz, 1H, H-7'''), 7.87 – 7.77 (m, 1H, H-8'''), 7.74 – 7.59 (m, 3H, H-3, H-9'''), 7.53 (d, J = 8.6 Hz, 2H, H-2''), 7.44 (d, J = 8.5 Hz, 2H, H-3'), 7.26 – 7.16 (m, 3H, H-2, H-3''), 7.10 (d, J = 8.6 Hz, 2H, H-2').

¹³C-NMR (MeOD, 101 MHz) δ = 158.41, 141.54, 140.27, 139.93, 138.27, 135.40, 130.50, 124.74, 122.55, 122.16, 120.81, 118.09, 118.02, 117.98, 117.70, 114.24, 113.63, 113.21, 111.87, 111.69, 106.11, 94.35.

HR-MS m/z calcd. for $C_{29}H_{22}F_3N_4O^+$ ($M + 1$)⁺ 499.1740, found 499.1743.

4-(Quinolin-4-ylamino)-*N*-(4-((2-(trifluoromethyl)phenyl)amino)phenyl)benzamide (29)



Following general procedure E, **90** (73.0 mg, 277 μ mol, 1.00 eq), **77** (70.0 mg, 277 μ mol, 1.00 eq), DIPEA (117 μ L, 89.0 mg, 690 μ mol, 2.50 eq), EDC hydrochloride (106 mg, 554 μ mol, 2.00 eq), and HOBt (45.0 mg, 330 μ mol, 1.20 eq) in 3 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH \rightarrow 2% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **29** (96.0 mg, 216 μ mol, 78%) as a light yellow solid.

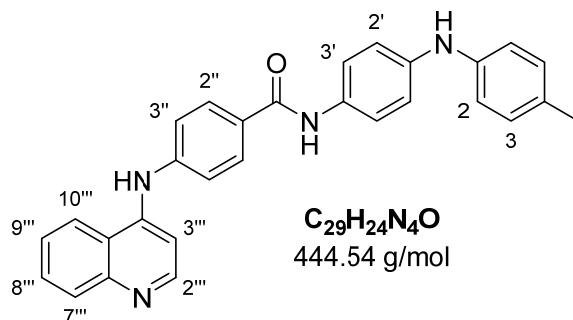
TLC: R_f = 0.32 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.09 (s, 1H, NH), 9.25 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.39 (d, J = 8.5 Hz, 1H, H-10'''), 8.02 (d, J = 8.7 Hz, 2H, H-3'''), 7.94 (d, J = 8.4 Hz, 1H, H-7'''), 7.74 (t, J = 8.5, 1H, H-9'''), 7.70 (d, J = 8.8 Hz, 2H, H-3'), 7.64 – 7.54 (m, 2H, H-H-8''', H-4*), 7.48 (d, J = 8.7 Hz, 2H, H-2'''), 7.46 (d, 1H, H-3), 7.38 (s, 1H, NH), 7.23 (d, J = 8.5 Hz, 1H, H-6), 7.21 (d, J = 5.2 Hz, 1H, H-3'''), 7.09 (d, J = 8.9 Hz, 2H, H-2'), 7.02 (t, J = 7.6 Hz, 1H, H-5*).

¹³C-NMR (DMSO, 101 MHz) δ = 164.51, 150.67, 148.97, 146.42, 142.83, 138.66, 133.62, 133.31, 129.49, 129.07, 128.68, 126.81, 125.86, 125.02, 123.15, 122.32, 121.53, 121.43, 120.31, 120.11, 119.72, 119.62, 117.75, 117.46, 103.62.

HR-MS m/z calcd. for $C_{29}H_{22}F_3N_4O^+$ ($M + 1$)⁺ 499.1740, found 499.1739.

4-(Quinolin-4-ylamino)-*N*-(4-(*p*-tolylamino)phenyl)benzamide (30)



Following general procedure E, **90** (93.3 mg, 353 μ mol, 1.00 eq), **78** (70.0 mg, 353 μ mol, 1.00 eq), DIPEA (150 μ L, 114 mg, 880 μ mol, 2.50 eq), EDC hydrochloride (135 mg, 706 μ mol,

2.00 eq), and HOBt (56.8 mg, 330 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [0.5% MeOH \rightarrow 1% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **30** (62.0 mg, 137 μmol , 39%) as a light yellow solid.

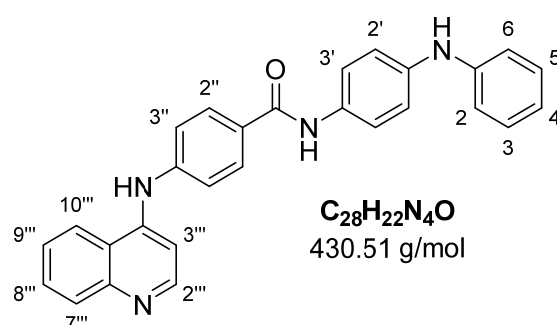
TLC: R_f = 0.33 (5% MeOH/1% TEA/ DCM) [UV].

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 10.03 (s, 1H, NH), 8.58 (d, J = 5.5 Hz, 1H, H-2'''), 8.45 (d, J = 8.6 Hz, 1H, H-10'''), 8.03 (d, J = 8.7 Hz, 2H, H-3'''), 7.99 – 7.88 (m, 2H, NH, H-7'''), 7.87 – 7.73 (m, 1H, H-8'''), 7.69 – 7.57 (m, 3H, H-3', H-9'''), 7.50 (d, J = 8.7 Hz, 2H, H-2''), 7.16 (d, J = 5.5 Hz, 1H, H-3'''), 7.04 (d, J = 8.4 Hz, 2H, H-3), 7.02 (d, J = 8.7 Hz, 2H, H-2'), 6.96 (d, J = 8.4 Hz, 2H, H-2), 2.22 (s, 3H, CH_3).

$^{13}\text{C-NMR}$ (DMSO, 101 MHz) δ = 164.26, 149.46, 147.73, 143.48, 141.27, 139.90, 131.43, 130.20, 129.58, 129.05, 128.09, 127.81, 125.36, 122.52, 121.81, 120.52, 119.93, 116.72, 116.61, 103.05, 20.28.

HR-MS m/z calcd. for $\text{C}_{29}\text{H}_{25}\text{N}_4\text{O}^+$ ($M + 1$)⁺ 445.2020, found 445.2029.

N-(4-(Phenylamino)phenyl)-4-(quinolin-4-ylamino)benzamide (31)



Following general procedure E, **90** (100 mg, 380 μmol , 1.00 eq), **79** (70.0 mg, 380 μmol , 1.00 eq), DIPEA (161 μL , 123 mg, 950 μmol , 2.50 eq), EDC hydrochloride (146 mg, 760 μmol , 2.00 eq), and HOBt (62.0 mg, 460 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **31** (127 mg, 296 μmol , 78%) as a white solid.

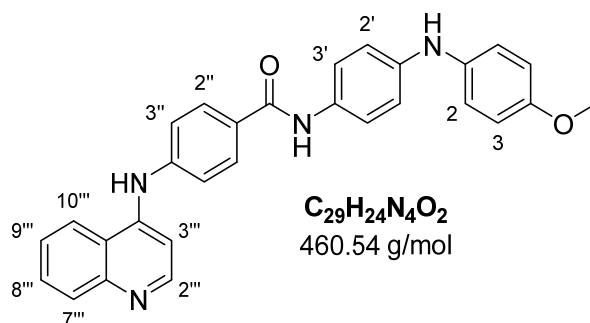
TLC: R_f = 0.32 (5% MeOH/1% TEA/ DCM) [UV].

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 10.03 (s, 1H, NH), 9.24 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.39 (d, J = 8.5, 1.4 Hz, 1H, H-10'''), 8.08 (s, 1H, NH), 8.01 (d, J = 8.7 Hz, 2H, H-3'''), 7.93 (d, J = 8.5 Hz, 1H, H-7'''), 7.74 (t, J = 8.3 Hz, 1H, H-8'''), 7.65 (d, J = 8.9 Hz, 2H, H-3'), 7.58 (t, J = 7.6 Hz, 1H, H-9'''), 7.48 (d, J = 8.7 Hz, 2H, H-2''), 7.23 (d, J = 8.7 Hz, 1H, H-3'''), 7.20 (d, J = 7.5 Hz, 2H, H-2, H-6), 7.08 (d, J = 8.9 Hz, 2H, H-2'), 7.04 (d, J = 7.5 Hz, 2H, H-3, H-5), 6.78 (t, J = 7.3 Hz, 1H, H-4).

¹³C-NMR (DMSO, 101 MHz) δ = 164.38, 150.70, 144.03, 139.09, 132.09, 129.48, 129.16, 129.03, 128.80, 125.01, 122.31, 121.73, 120.39, 119.75, 119.05, 117.57, 115.88, 103.58.

HR-MS m/z calcd. for C₂₈H₂₃N₄O⁺ (M + 1)⁺ 431.1866, found 431.1869.

N-(4-((4-Methoxyphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (32)



Following general procedure E, **90** (86.0 mg, 326 μmol, 1.00 eq), **80** (70.0 mg, 326 μmol, 1.00 eq), DIPEA (138 μL, 105 mg, 815 μmol, 2.50 eq), EDC hydrochloride (125 mg, 654 μmol, 2.00 eq), and HOBt (53.0 mg, 390 μmol, 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **32** (82.0 mg, 179 μmol, 55%) as a white solid.

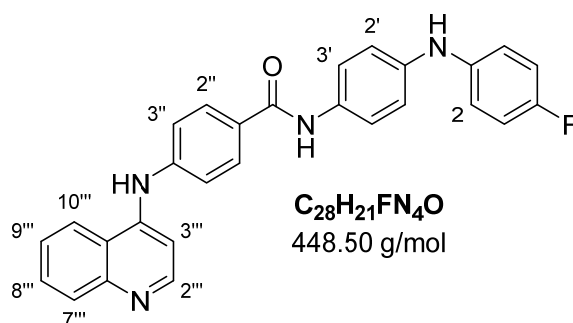
TLC: R_f = 0.29 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.96 (s, 1H, NH), 9.23 (s, 1H, NH), 8.57 (d, J = 5.2 Hz, 1H, H-2'''), 8.39 (d, J = 8.5, 1H, H-10'''), 8.00 (d, J = 8.6 Hz, 2H, H-3'''), 7.93 (d, J = 8.4 Hz, 1H, H-7'''), 7.78 (s, 1H, NH), 7.73 (t, J = 7.6 Hz, 1H, H-8'''), 7.65 – 7.53 (m, 3H, H-3', H-9'''), 7.47 (d, J = 8.6 Hz, 2H, H-2'''), 7.19 (d, J = 5.1 Hz, 1H, H-3'''), 7.03 (d, J = 9.0 Hz, 2H, H-2), 6.94 (d, J = 9.0 Hz, 2H, H-2'), 6.86 (d, J = 8.9 Hz, 1H, H-3), 3.71 (s, 1H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 164.27, 153.41, 150.73, 149.04, 146.41, 144.06, 141.00, 136.81, 130.84, 129.47, 129.27, 128.99, 128.88, 125.00, 122.31, 121.94, 120.40, 119.76, 119.40, 115.41, 114.58, 103.56, 55.24.

HR-MS m/z calcd. for C₂₉H₂₅N₄O₂⁺ (M + 1)⁺ 461.1972, found 461.1973.

N-(4-((4-Fluorophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (33)



Following general procedure E, **90** (91.0 mg, 346 μmol , 1.00 eq), **81** (70.0 mg, 346 μmol , 1.00 eq), DIPEA (148 μL , 112 mg, 870 μmol , 2.50 eq), EDC hydrochloride (133 mg, 692 μmol , 2.00 eq), and HOBt (57.0 mg, 420 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **33** (101 mg, 225 μmol , 65%) as a yellow solid.

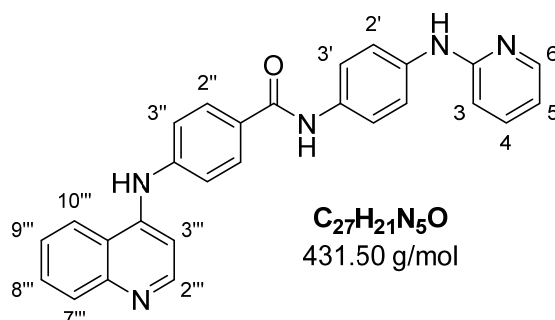
TLC: R_f = 0.37 (5% MeOH/1% TEA/ DCM) [UV].

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 10.14 (s, 1H, NH), 8.62 (d, J = 8.6 Hz, 1H, H-10'''), 8.58 (d, J = 6.0 Hz, 1H, H-2'''), 8.09 (d, J = 5.4 Hz, 2H, H-3''), 8.07 (s, 1H, NH), 8.01 (dd, J = 8.5, 1.3 Hz, 1H, H-7'''), 7.89 (t, J = 8.3 Hz, 1H, H-8'''), 7.70 (t, J = 7.8 Hz, 1H, H-9'''), 7.65 (d, J = 9.0 Hz, 2H, H-3'), 7.56 (d, J = 8.6 Hz, 2H, H-2''), 7.09 (d, J = 6.0 Hz, 1H, H-3'''), 7.07 – 7.02 (m, 6H, H-2', H-2, H-3).

$^{13}\text{C-NMR}$ (DMSO, 101 MHz) δ = 164.21, 157.20, 154.86, 150.38, 146.97, 143.84, 142.19, 140.36, 140.34, 139.75, 131.73, 131.68, 131.03, 129.16, 126.08, 124.94, 123.04, 122.12, 121.87, 118.99, 117.91, 117.84, 116.82, 115.77, 115.54, 102.01.

HR-MS m/z calcd. for $\text{C}_{28}\text{H}_{22}\text{FN}_4\text{O}^+$ ($M + 1$)⁺ 449.1772, found 449.1777.

N-4-(**Pyridin-2-ylamino**)phenyl)-4-(**quinolin-4-ylamino**)benzamide (**34**)



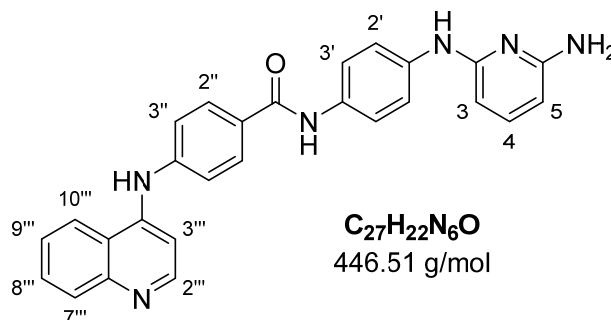
Following general procedure E, **90** (100 mg, 377 μmol , 1.00 eq), **88** (70.0 mg, 377 μmol , 1.00 eq), DIPEA (160 μL , 121 mg, 940 μmol , 2.50 eq), EDC hydrochloride (145 mg, 754 μmol , 2.00 eq), and HOBt (61.0 mg, 452 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH \rightarrow 5% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **34** (73.0 mg, 170 μmol , 45%) as a white solid.

TLC: R_f = 0.23 (5% MeOH/1% TEA/ DCM) [UV].

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 10.04 (s, 1H, NH), 9.23 (s, 1H, NH), 8.97 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.39 (d, J = 8.4 Hz, 1H, H-10'''), 8.14 (dd, J = 5.1, 2.0 Hz, 1H, H-6), 8.02 (d, J = 8.5 Hz, 2H, H-3''), 7.93 (d, J = 8.3 Hz, 1H, H-7'''), 7.73 (t, J = 8.3, 1H, H-8'''), 7.66 (d, J = 2.8 Hz, 4H, H-2', H-3'), 7.61 – 7.51 (m, 2H, H-4, H-9'''), 7.48 (d, J = 8.4 Hz, 2H, H-2'), 7.21 (d, J = 5.2 Hz, 1H, H-3'''), 6.81 (d, J = 8.4 Hz, 1H, H-3), 6.75 – 6.67 (m, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 164.40, 155.99, 150.76, 149.08, 147.28, 146.35, 144.15, 137.62, 137.15, 132.39, 129.45, 129.30, 129.05, 128.78, 125.01, 122.30, 121.11, 120.41, 119.71, 118.23, 113.94, 110.38, 103.63, 40.15, 39.99, 39.94, 39.73, 39.52, 39.31, 39.10, 38.89.
HR-MS m/z calcd. for C₂₇H₂₂N₅O⁺ (M + 1)⁺ 432.1819, found 432.1820.

N-(4-((6-Aminopyridin-2-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (35)



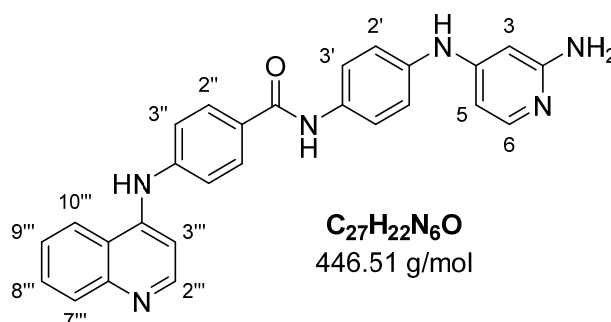
Following general procedure E, **90** (198 mg, 750 μ mol, 1.00 eq), **82** (150 mg, 750 μ mol, 1.00 eq), DIPEA (318 μ L, 242 mg, 1.87 mmol, 2.50 eq), EDC hydrochloride (287 mg, 1.50 mmol, 2.00 eq), and HOBT (121 mg, 900 μ mol, 1.20 eq) in 6 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [0.5% MeOH \rightarrow 5% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **35** (89.0 mg, 203 μ mol, 27%) as a light brown solid.

TLC: R_f = 0.14 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 9.99 (s, 1H, NH), 9.23 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.50 (s, 1H, NH), 8.38 (d, J = 8.6 Hz, 1H, H-10'''), 8.01 (d, J = 8.7 Hz, 2H, H-3'''), 7.93 (d, J = 8.3 Hz, 1H, H-7'''), 7.74 (t, J = 7.0 Hz, 1H, H-9'''), 7.67 – 7.52 (m, 5H, H-2', H-3', H-8'''), 7.48 (d, J = 8.3 Hz, 2H, H-2''), 7.26 – 7.13 (m, 2H, H-4, H-3'''), 5.98 (d, J = 7.8 Hz, 1H, H-3), 5.82 (d, J = 7.8 Hz, 1H, H-5), 5.65 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 164.37, 158.45, 155.10, 150.77, 149.07, 146.40, 144.09, 138.42, 138.25, 131.69, 129.48, 129.30, 129.03, 128.91, 125.03, 122.31, 121.05, 120.41, 119.76, 117.93, 103.60, 97.11, 96.96.

N-(4-((2-Aminopyridin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (36)



Following general procedure E, **90** (92.0 mg, 350 μmol , 1.00 eq), **83** (70.0 mg, 350 μmol , 1.00 eq), DIPEA (152 μL , 113 mg, 875 μmol , 2.50 eq), EDC hydrochloride (137 mg, 700 μmol , 2.00 eq), and HOBt (57.0 mg, 420 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for 6 hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [3% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **36** (65.0 mg, 147 μmol , 42%) as a light yellow solid.

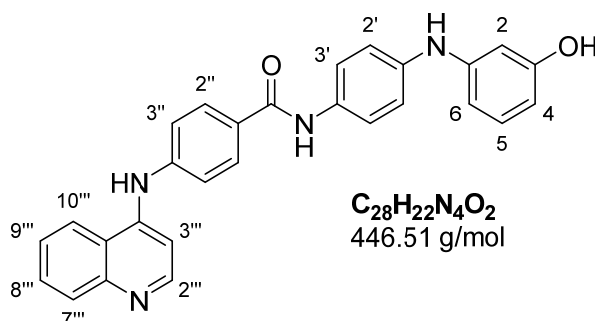
TLC: R_f = 0.38 (10% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.10 (s, 1H, NH), 9.24 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.38 (d, J = 8.5 Hz, 1H, H-10'''), 8.30 (s, 1H, NH), 8.01 (d, J = 8.7 Hz, 2H, H-3''), 7.93 (d, J = 8.3 Hz, 1H, H-7'''), 7.73 (t, 1H, H-8'''), 7.72 (d, J = 8.9 Hz, 2H, H-3'), 7.59 (d, 1H, H-6), 7.57 (t, 1H, H-9'''), 7.48 (d, J = 8.3 Hz, 2H, H-2''), 7.21 (d, J = 5.2 Hz, 1H, H-3'''), 7.13 (d, J = 8.9 Hz, 2H, H-2'), 6.12 (dd, J = 5.8, 2.0 Hz, 1H, H-5), 6.04 (d, J = 2.0 Hz, 1H, H-3), 5.52 (s, 2H, NH₂).

¹³C-NMR (DMSO, 101 MHz) δ = 164.57, 160.72, 151.44, 150.76, 149.07, 148.07, 146.33, 144.27, 136.86, 133.78, 129.47, 129.30, 129.10, 128.64, 125.02, 122.31, 121.38, 120.61, 120.43, 119.68, 103.69, 101.15, 90.31.

HR-MS m/z calcd. for C₂₇H₂₃N₆O⁺ ($M + 1$)⁺ 447.1928, found 447.1958.

N-4-((3-Hydroxyphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (**40**)



Following general procedure E, **90** (91.0 mg, 340 μmol , 1.00 eq), **89** (69.0 mg, 340 μmol , 1.00 eq), DIPEA (148 μL , 110 mg, 850 μmol , 2.50 eq), EDC hydrochloride (130 mg, 680 μmol , 2.00 eq), and HOBt (55.0 mg, 408 μmol , 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [2% MeOH \rightarrow 5% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **40** (50.0 mg, 112 μmol , 33%) as light yellow crystals.

TLC: R_f = 0.22 (5% MeOH/1% TEA/ DCM) [UV].

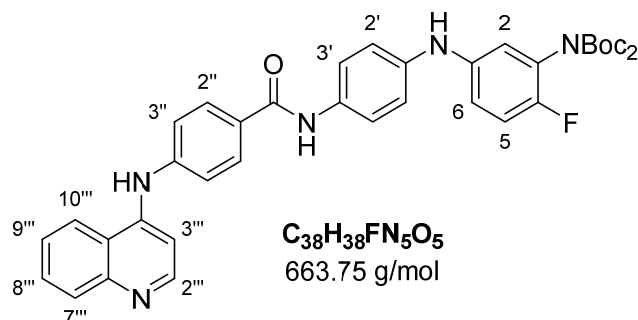
¹H-NMR (MeOD, 400 MHz) δ = 8.45 (d, J = 5.6 Hz, 1H, H-2'''), 8.27 (d, J = 8.4 Hz, 1H, H-10'''), 7.96 (d, J = 8.6 Hz, 1H, H-3'), 7.91 (d, J = 8.5 Hz, 1H, H-7'''), 7.71 (t, 1H, H-8'''), 7.57 – 7.48 (m, 3H, H-3', H-9'''), 7.44 (d, 2H, J = 8.51 Hz, H-2''), 7.22 – 7.16 (m, 1H, H-3'), 7.08 (d, J = 8.8

Hz, 2H, H-2'), 7.00 (t, J = 8.0 Hz, 1H, H-5), 6.60 – 6.56 (m, 1H, H-2), 6.53 (dd, J = 8.0, 2.1 Hz, 1H, H-6), 6.29 (dd, J = 7.7, 1.7 Hz, 1H, H-4).

¹³C-NMR (MeOD, 101 MHz) δ = 167.92, 159.26, 151.24, 149.70, 149.39, 146.71, 145.54, 142.06, 132.35, 131.07, 130.91, 130.80, 130.72, 130.26, 130.13, 129.13, 126.67, 123.83, 122.97, 122.81, 121.85, 121.76, 121.61, 119.12, 109.65, 108.09, 104.63, 104.32.

HR-MS m/z calcd. for C₂₈H₂₃N₄O₂⁺ (M + 1)⁺ 447.1816, found 447.1843.

tert-Butyl (tert-butoxycarbonyl)(2-fluoro-5-((4-(4-(quinolin-4-ylamino)benzamido)phenyl)amino)phenyl)carbamate (94)



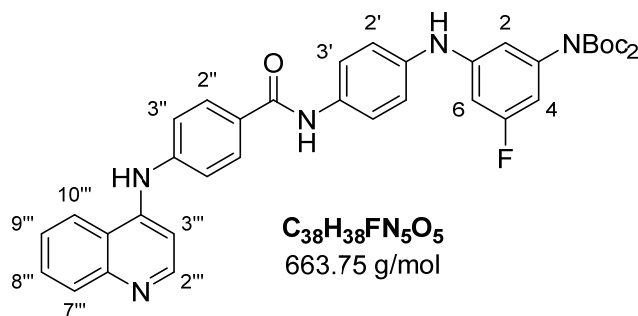
Following general procedure E, **90** (53.0 mg, 198 μmol, 1.00 eq), **84** (83.0 mg, 198 μmol, 1.00 eq), DIPEA (86.0 μL, 64.0 mg, 495 μmol, 2.50 eq), EDC hydrochloride (76.0 mg, 396 μmol, 2.00 eq), and HOBt (32.0 mg, 238 μmol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [2% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **94** (82.0 mg, 124 μmol, 63%) as light yellow crystals.

TLC: R_f = 0.23 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (MeOD, 400 MHz) δ = 8.49 (d, J = 5.4 Hz, 1H, H-2'''), 8.31 (dd, J = 8.7, 1.3 Hz, 1H, H-10'''), 8.00 (d, J = 8.7 Hz, 1H, H-3'''), 7.94 (dd, J = 8.1, 1.0 Hz, 1H, H-7'''), 7.75 (ddd, J = 8.4, 6.8, 1.3 Hz, 1H, H-8'''), 7.62 – 7.54 (m, 3H, H-3', H-9'''), 7.50 (d, J = 8.7 Hz, 2H, H-2''), 7.24 (d, J = 5.5 Hz, 1H, H-3'''), 7.12 – 6.96 (m, 4H, H-5, H-6, H-2'), 6.95 – 6.88 (m, 1H, H-2), 1.43 (s, 18H, Boc₂).

¹³C-NMR (MeOD, 101 MHz) δ = 167.95, 154.42, 152.55, 152.03, 151.19, 149.65, 149.60, 145.61, 142.21, 142.03, 132.78, 131.18, 130.82, 130.17, 129.08, 128.57, 128.42, 126.74, 123.95, 122.87, 121.89, 119.55, 119.45, 118.56, 118.32, 117.09, 116.88, 104.34, 84.43, 49.71, 49.64, 49.50, 49.43, 49.21, 49.00, 48.79, 48.57, 48.36, 28.07.

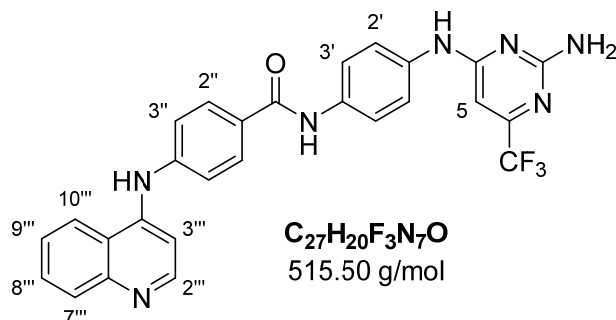
***tert*-Butyl (*tert*-butoxycarbonyl)(3-fluoro-5-((4-(4-(quinolin-4-ylamino)benzamido)phenyl)amino)phenyl)carbamate (**95**)**



Compound **63** (161 mg, 380 μ mol, 1.00 eq) was stirred with 10 wt% Pd/C (49.0 mg, 460 μ mol, 1.20 eq), and ammonium formate (240 mg, 3.80 mmol, 10.0 eq) in 10 mL EtOH overnight. After filtration and washing with EtOAc, the crude intermediate was directly subjected to amide coupling using general procedure F. To a solution of **90** (70.0 mg, 265 μ mol, 1.00 eq) and the crude intermediate (84.0 mg, 265 μ mol, 1.00 eq) in 4 mL DMF were added NEt₃ (92.0 μ L, 67.0 mg, 663 μ mol, 2.50 eq), EDC hydrochloride (101 mg, 530 μ mol, 2.00 eq), and HOBt (44.0 mg, 318 μ mol, 1.20 eq). The mixture was heated to 70°C for six hours and left at room temperature overnight. The crude product was purified by silica gel column chromatography [2% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **95** (80.0 mg, 121 μ mol, 52% over two steps) as light yellow crystals.

¹H-NMR (DMSO, 400 MHz) δ = 10.09 (s, 1H, NH), 9.23 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.44 (s, 1H, NH), 8.38 (d, J = 8.4 Hz, 1H, H-10'''), 8.01 (d, J = 8.6 Hz, 2H, H-3'''), 7.93 (d, J = 8.4 Hz, 1H, H-7'''), 7.72 (m, 3H, H-3', H-8'''), 7.58 (t, J = 7.6 Hz, 1H, H-9'''), 7.48 (d, J = 8.5 Hz, 2H, H-2''), 7.21 (d, J = 5.1 Hz, 1H, H-3'''), 7.11 (d, J = 8.9 Hz, 2H, H-2'), 6.70 (dt, J = 11.4, 2.2 Hz, 1H, H-6), 6.57 (s, 1H, H-4), 6.46 (dt, J = 9.4, 2.1 Hz, 1H, H-2), 1.42 (s, 18H, Boc₂).

***N*-(4-((2-Amino-6-(trifluoromethyl)pyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (**41**)**



Following general procedure E, **90** (69.0 mg, 260 μ mol, 1.00 eq), **85** (70.0 mg, 260 μ mol, 1.00 eq), DIPEA (113 μ L, 84.0 mg, 650 μ mol, 2.50 eq), EDC hydrochloride (100 mg, 520 μ mol, 2.00 eq), and HOBt (42.0 mg, 310 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column

chromatography [3% MeOH → 5% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **41** (24.0 mg, 46.8 μmol, 18%) as a light yellow solid.

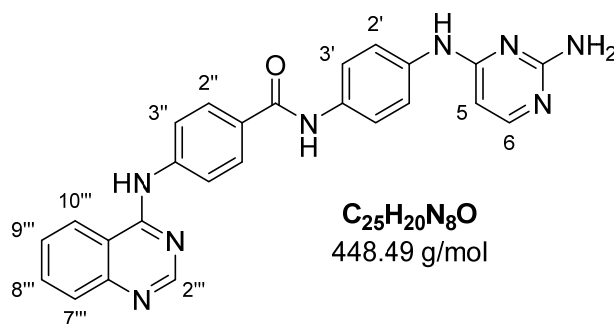
¹H-NMR (DMSO, 400 MHz) δ = 10.20 (s, 1H, NH), 9.62 (s, 1H, NH), 8.59 (d, J = 5.8 Hz, 1H, H-2^{'''}), 8.54 (d, J = 8.4 Hz, 1H, H-10^{'''}), 8.07 (d, J = 8.3 Hz, 2H, H-3^{''}), 7.98 (d, J = 8.4 Hz, 1H, H-7^{'''}), 7.85 (t, J = 7.7 Hz, 1H, H-8^{'''}), 7.75 (s, 4H, H-2', H-3'), 7.68 (t, J = 8.0 Hz, 1H, H-9^{'''}), 7.55 (d, J = 8.4 Hz, 2H, H-2^{''}), 7.13 (d, J = 5.8 Hz, 1H, H-3^{'''}), 6.83 (s, 2H, NH₂), 6.36 (s, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 164.49, 163.48, 161.46, 147.96, 142.81, 135.49, 134.14, 131.11, 130.31, 129.20, 127.21, 126.05, 125.81, 124.43, 122.77, 121.44, 120.82, 120.18, 119.35, 119.09, 109.64, 102.47, 40.15, 39.99, 39.94, 39.73, 39.52, 39.31, 39.10, 38.89.

¹⁹F-NMR (DMSO, 376 MHz) δ = -69.88.

HR-MS m/z calcd. for C₂₇H₂₁F₃N₇O⁺ (M + 1)⁺ 516.1754, found 516.1791.

N-(4-((2-Aminopyrimidin-4-yl)amino)phenyl)-4-(quinazolin-4-ylamino)benzamide (47)



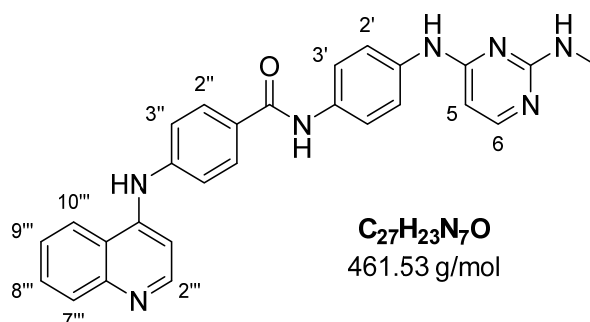
Following general procedure E, **92** (210 mg, 790 μmol, 1.00 eq), **74** (170 mg, 790 μmol, 1.00 eq), DIPEA (346 μL, 256 mg, 1.98 mmol, 2.50 eq), EDC hydrochloride (302 mg, 1.58 mmol, 2.00 eq), and HOBT (128 mg, 948 μmol, 1.20 eq) in 8 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [5% MeOH → 10% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **47** (136 mg, 300 μmol, 38%) as a light yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 10.08 (s, 1H, NH), 10.00 (s, 1H, NH), 9.05 (s, 1H, NH), 8.70 (s, 1H, H-2^{'''}), 8.62 (d, J = 8.3 Hz, 1H, H-10^{'''}), 8.09 (d, J = 8.8 Hz, 2H, H-3^{''}), 8.01 (d, J = 8.8 Hz, 2H, H-2^{''}), 7.90 (t, J = 7.6 Hz, 1H, H-8^{'''}), 7.84 (d, J = 8.3 Hz, 1H, H-7^{'''}), 7.79 (d, J = 5.7 Hz, 1H, H-6), 7.71 – 7.67 (m, 1H, H-9^{'''}), 7.69 (s, 4H, H-2', H-3'), 6.17 (s, 2H, NH₂), 5.99 (d, J = 5.8 Hz, 1H, H-5).

¹³C-NMR (DMSO, 101 MHz) δ = 164.64, 163.02, 160.78, 157.56, 156.16, 154.31, 149.79, 142.21, 136.39, 133.33, 133.22, 129.58, 128.13, 127.91, 126.48, 123.03, 121.07, 120.79, 119.65, 115.26, 96.42.

HR-MS m/z calcd. for C₂₅H₂₁N₈O⁺ (M + 1)⁺ 449.1833, found 449.1855.

***N*-4-((2-(Methylamino)pyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (42)**



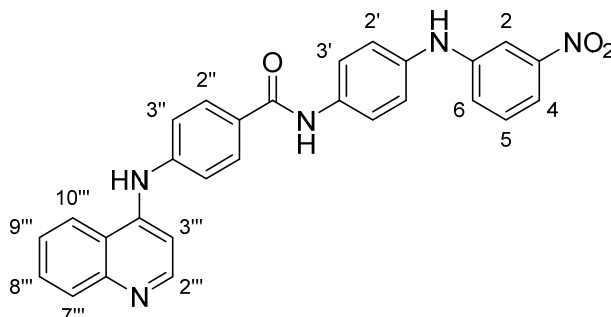
Following general procedure E, **90** (109 mg, 410 μ mol, 1.00 eq), **96** (89.0 mg, 410 μ mol, 1.00 eq), DIPEA (179 μ L, 133 mg, 1.03 mmol, 2.50 eq), EDC hydrochloride (157 mg, 820 μ mol, 2.00 eq), and HOBt (67.0 mg, 492 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [2% MeOH \rightarrow 7% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield **42** (100 mg, 217 μ mol, 53%) as light yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 10.06 (s, 1H, NH), 9.23 (s, 1H, NH), 9.10 (s, 1H, NH), 8.58 (d, J = 5.2 Hz, 1H, H-2'''), 8.39 (d, J = 8.4 Hz, 1H, H-10'''), 8.01 (d, J = 8.4 Hz, 2H, H-3''), 7.93 (d, J = 8.4 Hz, 1H, H-7'''), 7.83 (d, J = 5.7 Hz, 1H, H-6), 7.77 – 7.71 (m, 1H, H-9'''), 7.70 (s, 4H, H-2', H-3'), 7.64 – 7.53 (m, 1H, H-8'''), 7.48 (d, J = 8.3 Hz, 2H, H-2''), 7.20 (d, J = 5.2 Hz, 1H, H-3'''), 6.62 (d, J = 5.2 Hz, 1H, NH), 5.98 (d, J = 5.7 Hz, 1H, H-5), 2.80 (d, J = 4.7 Hz, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 164.50, 162.49, 160.47, 150.66, 148.99, 146.38, 144.25, 136.39, 133.30, 129.44, 129.21, 129.05, 128.71, 124.98, 122.29, 120.82, 120.41, 119.69, 119.54, 103.63, 27.92.

HR-MS m/z calcd. for C₂₇H₂₄N₇O⁺ (M + 1)⁺ 462.2037, found 462.2062.

***N*-4-((3-Nitrophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (86)**



Following general procedure E, **90** (161 mg, 610 μ mol, 1.00 eq), **65** (140 mg, 610 μ mol, 1.00 eq), DIPEA (266 μ L, 197 mg, 1.53 mmol, 2.50 eq), EDC hydrochloride (233 mg, 1.22 μ mol, 2.00 eq), and HOBt (99.0 mg, 730 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel

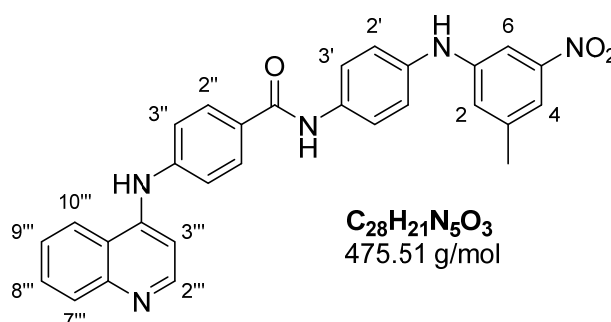
column chromatography [1% MeOH → 3% MeOH/1% TEA/ DCM] to yield **86** (208 mg, 439 μmol, 72%) as dark red solid.

TLC: R_f = 0.47 (10% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.15 (s, 1H, NH), 9.35 (s, 1H, NH), 8.70 (s, 1H, NH), 8.58 (d, J = 5.3 Hz, 1H, H-2'''), 8.42 (d, J = 8.4 Hz, 1H, H-10'''), 8.03 (d, J = 8.7 Hz, 2H, H-3''), 7.94 (d, J = 8.5 Hz, 1H, H-7'''), 7.81 – 7.70 (m, 4H, H-3', H-8''', H-2*), 7.64 – 7.57 (m, 1H, H-9'''), 7.57 – 7.53 (m, 1H, H-6*), 7.50 (d, J = 8.8 Hz, 2H, H-2''), 7.46 (t, J = 8.1 Hz, 1H, H-5*), 7.43 – 7.35 (m, 1H, H-4*), 7.22 – 7.14 (m, 3H, H-2', H-3''').

¹³C-NMR (DMSO, 101 MHz) δ = 164.54, 148.78, 146.09, 136.98, 134.02, 130.43, 129.60, 129.08, 128.74, 125.05, 122.41, 121.67, 120.76, 120.32, 119.87, 119.83, 112.51, 107.98, 103.53.

***N*-(4-((3-Methyl-5-nitrophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (87)**



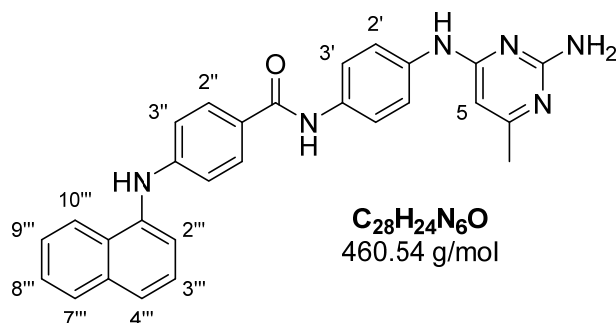
Following general procedure E, **90** (146 mg, 550 μmol, 1.00 eq), **66** (135 mg, 550 μmol, 1.00 eq), DIPEA (228 μL, 178 mg, 1.38 mmol, 2.50 eq), EDC hydrochloride (211 mg, 1.10 mmol, 2.00 eq), and HOBt (89.0 mg, 660 μmol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [1% MeOH → 3% MeOH/1% TEA/ DCM] to yield **87** (149 mg, 259 μmol, 47%) as a dark red solid.

TLC: R_f = 0.32 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.15 (s, 1H, NH), 8.60 (s, 1H, NH), 8.58 (d, J = 5.3 Hz, 1H, H-2'''), 8.44 (d, J = 8.5 Hz, 1H, H-10'''), 8.03 (d, J = 8.7 Hz, 2H, H-3''), 7.94 (d, J = 8.5 Hz, 1H, H-7'''), 7.81 – 7.69 (m, 3H, H-3', H-8'''), 7.60 (t, J = 7.7 Hz, 1H, H-9'''), 7.56 (t, J = 2.2 Hz, 1H, H-6*), 7.50 (d, J = 8.7 Hz, 2H, H-2''), 7.40 (s, 1H, H-4*), 7.21 – 7.15 (m, 3H, H-2', H-2*), 2.35 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 164.53, 148.77, 145.91, 140.45, 137.11, 133.92, 129.09, 125.16, 122.46, 121.68, 121.07, 120.17, 120.08, 119.93, 113.17, 105.71, 103.37, 21.00.

***N*-4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(naphthalen-1-ylamino)benzamide (44)**

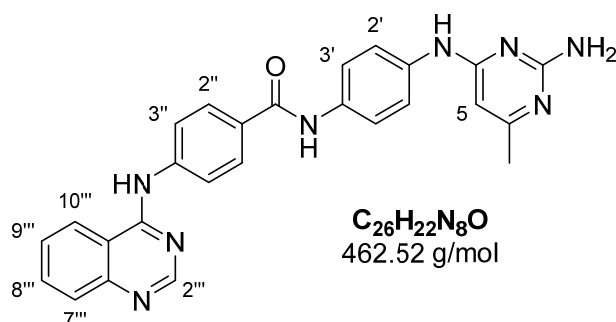


Following general procedure E, **70** (70.0 mg, 252 μ mol, 1.00 eq), **73** (54.0 mg, 252 μ mol, 1.00 eq), DIPEA (110 μ L, 631 μ mol, 2.50 eq), EDC hydrochloride (97.0 mg, 505 μ mol, 2.00 eq), and HOBt (41.0 mg, 303 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [5% MeOH/1% TEA/ DCM] to yield **44** (35.0 mg, 76.0 μ mol, 30%) as light yellow solid.

¹H-NMR (DMSO, 600 MHz) δ = 9.84 (s, 1H, NH), 8.96 (s, 1H, NH), 8.68 (s, 1H, NH), 8.10 (d, J = 8.0 Hz, 1H, H-10'''), 7.95 (d, J = 9.6 Hz, 1H, H-7'''), 7.84 (d, J = 8.8 Hz, 2H, H-2''), 7.69 (d, J = 8.1 Hz, 1H, H-4'''), 7.67 – 7.60 (m, 4H, H-2', H-3'), 7.58 – 7.48 (m, 3H, H-3''', H-8''', H-9'''), 7.45 (d, J = 7.6 Hz, 1H, H-2'''), 7.00 (d, J = 8.8 Hz, 2H, H-3'''), 6.16 (s, 2H, NH₂), 5.86 (s, 1H, H-5), 2.09 (s, 1H, CH₃).

¹³C-NMR (DMSO, 151 MHz) δ = 164.73, 162.59, 161.41, 148.89, 137.61, 136.14, 134.44, 133.64, 129.16, 128.28, 128.04, 128.00, 126.27, 126.14, 125.57, 124.44, 123.49, 123.47, 122.92, 120.71, 120.60, 119.81, 117.83, 114.19, 94.61, 23.29.

***N*-4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(quinazolin-4-ylamino)benzamide (45)**



Following general procedure E, **92** (70.0 mg, 264 μ mol, 1.00 eq), **73** (56.8 mg, 264 μ mol, 1.00 eq), DIPEA (115 μ L, 659 μ mol, 2.50 eq), EDC hydrochloride (101 mg, 528 μ mol, 2.00 eq), and HOBt (43.0 mg, 317 μ mol, 1.20 eq) in 4 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [5% MeOH/1% TEA/ DCM] and RP-HPLC (NUCLEODUR® C18 Pyramid,

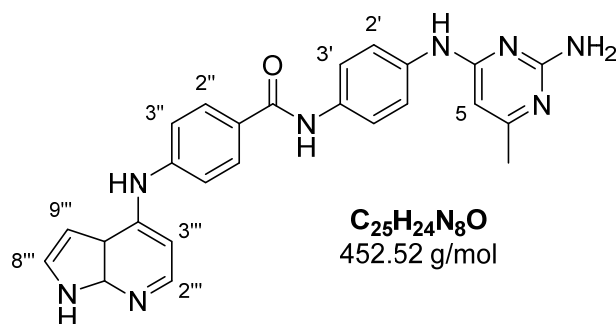
buffer A: 50 mM ammonium acetate in water [pH 8.5], buffer B: 98% MeOH and 2% buffer A) to yield **45** (8.00 mg, 17.3 μ mol, 7%) as light yellow solid.

¹H-NMR (DMSO, 600 MHz) δ = 10.08 (s, 1H, NH), 10.01 (s, 1H, NH), 8.95 (s, 1H, NH), 8.69 (s, 1H, H-2'''), 8.62 (dd, J = 8.6, 1.4 Hz, 1H, H-7'''), 8.09 (d, J = 8.8 Hz, 1H, H-2''), 8.01 (d, J = 8.8 Hz, 1H, H-3''), 7.91 (t, J = 7.0 Hz, 1H, H-9'''), 7.84 (d, J = 8.4 Hz, 1H, H-10'''), 7.73 – 7.58 (m, 5H, H-8''', H-2', H-2'), 6.13 (s, 2H, NH₂), 5.86 (s, 1H, H-5), 2.08 (s, 1H, CH₃).

¹³C-NMR (DMSO, 151 MHz) δ = 164.77, 164.64, 162.83, 161.40, 157.58, 154.33, 149.81, 142.22, 136.59, 133.27, 133.22, 129.59, 128.15, 127.94, 126.52, 123.06, 121.09, 120.81, 119.69, 115.28, 94.60, 23.45.

HR-MS m/z calcd. for C₂₆H₂₃N₈O⁺ (M + 1)⁺ 463.1989, found 463.2008.

***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-((3a,7a-dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzamide (46)**

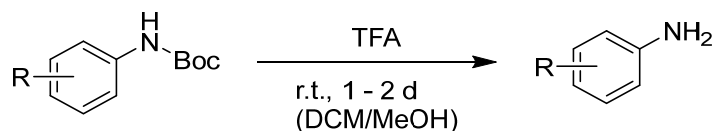


Following general procedure E, **93** (70.0 mg, 267 μ mol, 1.00 eq), **73** (59.4 mg, 267 μ mol, 1.00 eq), DIPEA (117 μ L, 690 μ mol, 2.50 eq), EDC hydrochloride (106 mg, 550 μ mol, 2.00 eq), and HOBT (44.8 mg, 332 μ mol, 1.20 eq) in 5 mL DMF were heated to 70°C for six hours and at room temperature overnight. The crude product was purified by silica gel column chromatography [5% MeOH/1% TEA/ DCM] to yield **46** (58.1 mg, 129 μ mol, 47%) as light yellow solid.

¹H-NMR (DMSO, 400 MHz) δ = 11.44 (s, 1H, NH), 9.98 (s, 1H, NH), 8.96 (s, 1H, NH), 8.90 (s, 1H, NH), 8.00 (d, J = 5.4 Hz, 1H, H-2'''), 7.96 (d, J = 8.7 Hz, 2H, H-2''), 7.66 (d, J = 2.9 Hz, 4H, H-2', H-3'), 7.37 (d, J = 8.7 Hz, 2H, H-3''), 7.27 (dd, J = 3.4, 2.2 Hz, 1H, H-8'''), 6.86 (d, J = 5.4 Hz, 1H, H-3'''), 6.60 (dd, J = 3.5, 1.7 Hz, 1H, H-9'''), 6.09 (s, 2H, NH₂), 5.86 (s, 1H, H-5), 2.08 (s, 3H, CH₃).

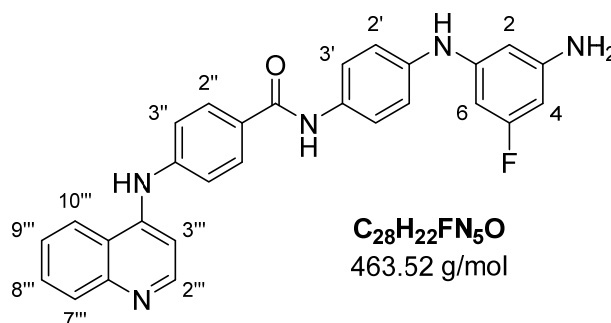
¹³C-NMR (DMSO, 101 MHz) δ = 164.90, 164.56, 162.91, 161.40, 149.95, 144.72, 143.80, 142.00, 136.46, 133.32, 128.93, 127.29, 122.77, 120.78, 119.70, 117.97, 109.69, 100.38, 98.31, 94.54, 23.49.

General procedure F: Boc-deprotection



To a solution of boc-protected compound (1.00 eq) in 4 mL DCM and 1 mL methanol was added 2 mL trifluoroacetic acid. The reaction mixture was stirred at room temperature for two days. Completion of deprotection was monitored by TLC. The solvent was removed under reduced pressure and the residue was dissolved in EtOAc and sat. NaHCO₃ solution. The aqueous phase was alkalinized with 10% aqueous NaOH solution and extracted with EtOAc (3 x 50 mL). The combined organic layer was dried over MgSO₄, filtered and concentrated. Each residue was purified by silica gel column chromatography and final compounds were further purified by precipitation as stated above.

***N*-(4-((3-Amino-5-fluorophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (38)**



Following general procedure F, **95** (64.0 mg, 96.4 μmol, 1.00 eq) and 2 mL TFA in DCM /MeOH (4 mL/1 mL) were stirred at room temperature for two days. The crude product was purified by silica gel column chromatography [3% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) to yield deprotected **38** (29.0 mg, 55.0 μmol, 57%) as light yellow solid.

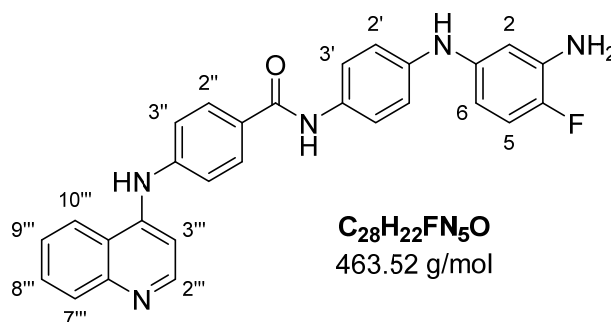
TLC: *R_f* = 0.31 (5% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (MeOD, 400 MHz) δ = 8.49 (d, *J* = 5.5 Hz, 1H, H-2'''), 8.30 (d, *J* = 8.4 Hz, 1H, H-10'''), 7.99 (d, *J* = 8.6 Hz 2H, H-3'''), 7.93 (d, *J* = 8.5, 1H, H-7'''), 7.74 (t, *J* = 8.0, 1H, H-8'''), 7.61 – 7.51 (m, 3H, H-9''', H-3'), 7.49 (d, *J* = 7.6 Hz, 2H, H-2''), 7.23 (d, *J* = 5.5 Hz, 1H, H-3'''), 7.11 (d, *J* = 8.8 Hz, 2H, H-2'), 6.24 (t, *J* = 1.9 Hz, 1H, H-2*), 6.09 (dt, *J* = 11.4, 2.1 Hz, 1H, H-6*), 5.92 (dt, *J* = 10.9, 2.1 Hz, 1H, H-4*).

¹³C-NMR (MeOD, 101 MHz) δ = 167.97, 167.33, 164.97, 151.42, 151.29, 149.78, 149.48, 147.91, 147.78, 145.63, 141.42, 132.96, 131.10, 130.77, 130.16, 129.18, 126.70, 123.75, 122.84, 121.90, 121.82, 120.01, 104.37, 99.46, 94.89, 94.64, 94.10, 93.85.

HR-MS *m/z* calcd. for C₂₈H₂₃FN₅O⁺ (*M* + 1)⁺ 464.1881, found 464.1914.

***N*-(4-((3-Amino-4-fluorophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (37)**



Following general procedure F, 2 mL TFA was slowly added to a solution of **94** (82.0 mg, 123 μ mol, 1.00 eq) in DCM/MeOH (4 mL/1 mL) and the reaction mixture was stirred at room temperature for two days. The reaction mixture was quenched with aqueous NaHCO₃ solution and extracted with EtOAc (3 x 50 mL). Purification by silica gel column chromatography [2% MeOH/1% TEA/ DCM] and precipitated with isopropanol (2 mL) and water (15 mL) afforded **37** (37 mg, 80.0 μ mol, 65%) as yellowish solid.

TLC: R_f = 0.29 (5% MeOH/1% TEA/ DCM) [UV].

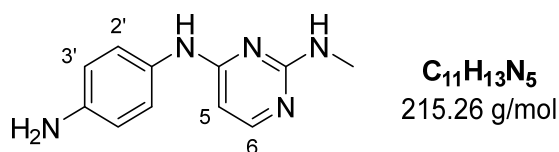
¹H-NMR (MeOD, 400 MHz) δ = 8.50 (d, J = 5.5 Hz, 1H, H-2'''), 8.33 (dd, J = 8.6, 1.3 Hz, 1H, H-10'''), 8.00 (d, J = 8.7 Hz, 2H, H-3'''), 7.94 (d, J = 9.0 Hz, 1H, H-7'''), 7.77 (ddd, J = 8.4, 6.8, 1.4 Hz, 1H, H-8'''), 7.59 (ddd, J = 8.3, 6.9, 1.3 Hz, 1H, H-9'''), 7.50 (dd, J = 8.8, 3.8 Hz, 4H, H-3', H-2''), 7.24 (d, J = 5.5 Hz, 1H, H-3'''), 7.03 (d, J = 8.8 Hz, 2H, H-2'), 6.83 (dd, J = 11.0, 8.7 Hz, 1H, H-5), 6.63 (dd, J = 7.9, 2.7 Hz, 1H, H-2), 6.37 (ddd, J = 8.7, 3.8, 2.7 Hz, 1H, H-6).

¹³C-NMR (MeOD, 126 MHz) δ = 151.19, 149.68, 143.20, 141.76, 131.20, 130.97, 130.16, 129.07, 126.76, 123.96, 122.88, 121.97, 118.05, 116.20, 116.04, 108.93, 108.07, 104.33.

¹⁹F-NMR (MeOD, 471 MHz, DMSO) δ = -146.58.

HR-MS m/z calcd. for C₂₈H₂₃FN₅O⁺ ($M + 1$)⁺ 464.1881, found 464.1919.

***N*⁴-(4-Aminophenyl)-*N*²-methylpyrimidine-2,4-diamine (96)**



Following general procedure F, 2 mL TFA was slowly added to a solution of **69** (588 mg, 1.86 mmol, 1.00 eq) in DCM/MeOH (4 mL/1 mL) and the reaction mixture was stirred at room temperature for two days. The reaction mixture was quenched with aqueous NaHCO₃ solution and extracted with EtOAc (3 x 50 mL). Purification by silica gel column chromatography [2% MeOH/1% TEA/ DCM] afforded **96** (94 mg, 428 μ mol, 23%) as yellowish solid.

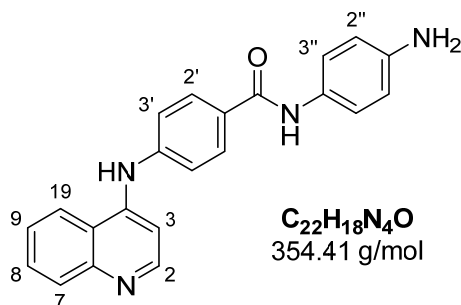
TLC: R_f = 0.35 (10% MeOH/1% TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 8.60 (s, 1H, NH), 7.71 (d, J = 5.7 Hz, 1H, H-6), 7.22 (d, J = 8.2 Hz, 2H, H-2'), 6.51 (d, J = 8.7 Hz, 2H, H-3'), 6.42 (s, 1H, NH), 5.81 (d, J = 5.8 Hz, 1H, H-5), 4.89 (s, 2H, NH₂), 2.74 (d, J = 4.7 Hz, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 162.40, 161.01, 155.19, 144.22, 129.15, 122.34, 114.00, 27.86.

Synthesis of truncated SGI-1027 structures

N-(4-Aminophenyl)-4-(quinolin-4-ylamino)benzamide (22)



Following general procedure E, benzene-1,4-diamine (20.0 mg, 189 μ mol, 1.00 eq), 4-(quinolin-4-ylamino)benzoic acid (**90**) (49.0 mg, 189 μ mol, 1.00 eq), DIPEA (80.0 μ L, 61.0 mg, 473 μ mol, 2.50 eq), EDC hydrochloride (43.0 mg, 227 μ mol, 1.20 eq) and HOBt (30.0 mg, 227 μ mol, 1.20 eq) in 2 mL DMF were stirred at 70 °C for two hours. The crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM \rightarrow 10% MeOH/1% TEA / DCM]. Recrystallization in 50 mL isopropanol afforded the final compound **22** as a light yellow crystals (10.0 mg, 28.4 μ mol, 15%).

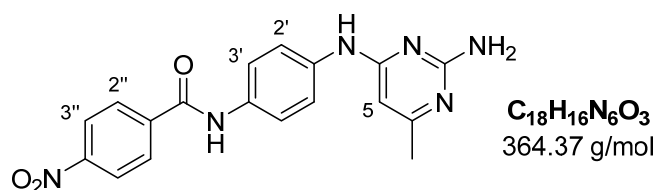
TLC: R_f = 0.46 (5% MeOH/1%TEA/ DCM) [UV].

¹H-NMR (MeOD, 400 MHz) δ = 8.50 (d, J = 5.55 Hz, 1H, H-2), 8.34 (d, J = 8.40 Hz, 1H, H-10), 8.00 (d, J = 8.64 Hz, 2H, H-3'), 7.94 (d, J = 8.0 Hz, 1H, H-7), 7.84 – 7.74 (m, 1H, H-8), 7.66 – 7.56 (m, 1H, H-9), 7.51 (d, J = 8.64 Hz, 2H, H-2'), 7.39 (d, J = 8.69 Hz, 2H, H-3''), 7.23 (d, J = 5.55 Hz, 1H, H-3), 6.76 (d, J = 8.69 Hz, 2H, H-2'').

HR-MS m/z calcd. for C₂₂H₁₉N₅O⁺ (M + 1)⁺ 355.1559, found 355.1560.

These data are in accordance with previous literature reports. Literature spectra were recorded in deuterated DMSO.⁴

N-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-nitrobenzamide (24)



To a solution of 4-nitrobenzoyl chloride (66.0 mg, 357 μ mol, 1.00 eq) and **73** (100 mg, 465 μ mol, 1.30 eq) in 10 mL anhydrous DMF was added Cs₂CO₃ (348 mg, 1.07 mmol,

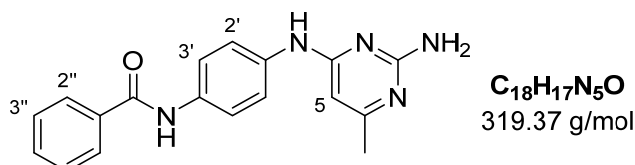
3.00 eq). The reaction mixture was stirred at room temperature for 4 hours. The solvent was removed under reduced pressure and the crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM → 10% MeOH/1% TEA / DCM] and RP-HPLC (NUCLEODUR® C18, 50 mM ammonium acetate in water/ MeCN) to afford **24** (9.00 mg, 25.0 μmol, 7%) as orange solid.

¹H-NMR (DMSO, 500 MHz) δ = 10.46 (s, 1H, NH), 8.98 (s, 1H, NH), 8.36 (d, J = 8.70 Hz, 2H, H-3''), 8.17 (d, J = 8.70 Hz, 2H, H-2''), 7.68 (dd, J = 9.05 Hz, 4H, H-2', H-3'), 6.10 (s, 2H, NH₂), 5.86 (s, 1H, H-5), 2.08 (s, 3H, CH₃).

HR-MS m/z calcd. for C₁₈H₁₇N₆O₃⁺ (M + 1)⁺ 365.1362, found 365.1364.

These data are in accordance with previous literature reports.³

***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)benzamide (23)**



To a solution of *N*⁴-(4-aminophenyl)-6-methylpyrimidine-2,4-diamine (**73**) (100 mg, 465 μmol, 1.10 eq) in 10 mL anhydrous THF were added TEA (147 μL, 107 mg, 1.06 mmol, 2.50 eq) and benzoyl chloride (50.0 μL, 60.0 mg, 423 μmol, 1.00 eq). The reaction mixture was stirred at 80 °C for 2 hours and at room temperature overnight. The solvent was removed under reduced pressure and the crude product was purified by silica gel column chromatography [1% MeOH/1% TEA/ DCM → 5% MeOH/1% TEA/ DCM]. **23** (69.3 mg, 216 μmol, 51%) was obtained after further purification by extraction with EtOAc (3 x 50 mL) and sat. NaHCO₃ solution as light brown solid.

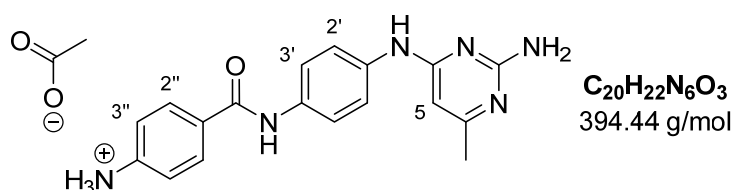
TLC: *R_f* = 0.33 (10% MeOH/1%TEA/ DCM) [UV].

¹H-NMR (DMSO, 400 MHz) δ = 10.14 (s, 1H, NH), 8.93 (s, 1H, NH), 7.94 (d, J = 6.7 Hz, 2H, H-2''), 7.66 (s, 4H, H-2', H-3'), 7.60 – 7.41 (m, 3H, H-3'', H-4''), 6.11 (s, 2H, NH₂), 5.85 (s, 1H, H-5), 2.08 (s, 3H, CH₃).

¹³C-NMR (DMSO, 101 MHz) δ = 165.14, 164.91, 162.90, 161.39, 136.75, 135.13, 133.02, 131.38, 128.35, 127.57, 120.85, 119.64, 94.59, 23.50.

HR-MS m/z calcd. for C₁₈H₁₈N₅O⁺ (M + 1)⁺ 320.1506, found 320.1512.

4-((4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)carbamoyl)benzenaminium (25)



To a solution of **73** (100 mg, 465 μmol , 1.10 eq) in 10 mL anhydrous THF were added TEA (147 μL , 107 mg, 1.06 mmol, 2.50 eq) and 4-nitrobenzoyl chloride (78.0 mg, 423 μmol , 1.00 eq). The reaction mixture was stirred at 80 °C for 5 hours. The solvent was removed under reduced pressure and the residue was extracted with EtOAc (3 x 100 mL) and sat. NaHCO_3 -solution. The combined organic layer was dried over MgSO_4 , filtered and concentrated. In the second step, the 4-nitrobenzamide intermediate, 10 wt% Pd/C (45.0 mg, 423 μmol , 1.00 eq) and ammonium formate (243 mg, 3.85 mmol, 11.0 eq) were suspended in 10 mL EtOH and stirred at room temperature overnight. The suspension was filtered over Celite and washed with EtOAc. The filtrate was concentrated and purified via RP-HPLC (NUCLEODUR® C18, 50 mM ammonium acetate in water/ MeCN) to obtain **25** (24.0 mg, 59.2 μmol , 14%) as light yellow acetate salt.

$^1\text{H-NMR}$ (DMSO, 400 MHz) δ = 9.65 (s, 1H, NH), 8.87 (s, 1H, NH), 7.70 (d, J = 8.7 Hz, 2H, H-2''), 7.67 – 7.49 (m, 4H, H-2', H-3'), 6.59 (d, J = 8.7 Hz, 2H, H-3''), 6.09 (s, 2H, NH_2), 5.84 (s, 1H, H-5), 5.71 (s, 2H, NH_2), 2.07 (s, 3H, CH_3), 1.90 (s, 3H, CH_3COO^-).

$^{13}\text{C-NMR}$ (DMSO, 101 MHz) δ = 172.08, 164.97, 162.88, 161.44, 151.96, 136.02, 133.78, 129.21, 121.30, 120.62, 119.76, 112.54, 94.49, 23.46, 21.12.

HR-MS m/z calcd. for $\text{C}_{18}\text{H}_{19}\text{N}_6\text{O}^+$ ($M + 1$)⁺ 335.1615, found 335.1615.

These data are in accordance with previous literature reports.³

4 Supplementary references

- [1] Y. Yuan, F. M. Stumpf, L. A. Schlor, O. P. Schmidt, P. Saumer, L. B. Huber, M. Frese, E. Höllmüller, M. Scheffner, F. Stengel, K. Diederichs, A. Marx, *Nat. Commun.* **2023**, *14*.
- [2] J. M. Pascal, P. J. O'Brien, A. E. Tomkinson, T. Ellenberger, *Nature* **2004**, *432*, 473-478.
- [3] P. García-Domínguez, C. Dell'aversana, R. Alvarez, L. Altucci, A. R. Lera, *Bioorganic Med. Chem. Lett.* **2013**, *23*, 1631-1635.
- [4] S. Valente, Y. Liu, M. Schnekenburger, C. Zwergel, S. Cosconati, C. Gros, M. Tardugno, D. Labella, C. Florean, S. Minden, H. Hashimoto, Y. Chang, X. Zhang, G. Kirsch, E. Novellino, P. B. Arimondo, E. Miele, E. Ferretti, A. Gulino, M. Diederich, X. Cheng, A. Mai, *J. Med. Chem.* **2014**, *57*, 701-713.
- [5] M. Huang, Z. Yi, Y. Wan, X. Zhu, *Synthesis* **2018**, *50*, 3911-3920.
- [6] R. Di Santo, R. Costi, G. Cuzzucoli Crucitti, L. Pescatori, F. Rosi, L. Scipione, D. Celona, M. Vertechy, O. Ghirardi, P. Piovesan, M. Marzi, S. Caccia, G. Guiso, F. Giorgi, P. Minetti, *J. Med. Chem.* **2012**, *55*, 8538-8548.

5 Appendix

Amino acid sequence of expressed proteins

Expressed amino acid sequence from pET15b-Rlig1:

MGSSHHHHHHSSGLVPR↓GSHMKRLGVSQRKMPVFTVEVKEEPSSKREHQPFKVLATETVSHKALDADIYSAIP
TEKVDGTCCYVTTYKDQPYLWARLDRKPNKQAEKRFKNFLHSHKENPKEFFWNVEEDFKPAPECWIPAKETEQING
NPVPDENGHIPGWVPEKNNKQYCWHSVVNYEFEIALVLKHHPPDDSGLLEISAVPLSDLLEQTLELIGNINGNPY
GLGSKKHPLHLLIPHGAFQIRNPLSLKHNDLVSWFEDCKEKGIEGIVWHCSGCLIKVHRHHLGLCWPIPDTYM
NSRPVIINMNLNKDSAFDIKCLFNHFLKIDNQKFVRLKDIIFDV*

Expressed amino acid sequence from pET28a-hLig1:

MQLSHHHHHHSSGENLYFQ↓GHMMQRSIMSFFHPKKEGKAKKPEKEASNSSRETEPPPKAALKEWNGVWVSESD
SPVKRPRGKAARVLGSEGEEDALSPAKGQKPALDCSQVSPRRPATSPENNASLSDTSPMDSSPSGIPKRRTAR
KQLPKRTIQEVLEEQSEDEDREAKRKKEEEEETPKESLTAEVATEKEGEDGDQPTTPPKPLKTSKAETPTESVS
EPEVATKQELQEEEEQTKPPRRAPKTLSSFFTPRKPAAKKEVKEEPPGAPGKEGAAEGPLDPSGYNPAKNNYHPV
EDACWKPGQKVPLYAVARTFEKIEEVSARLRMVETLSNLLRSVVALSPPDLLPVLYLSLNLHGLPQQGLELGVGDG
VLLKAVAQATGRQLESVRAEAAEKGDVGLVAENSRSTQRMLMPPPLTASGVFSKFRDIARLTGSASTAKKIDIIKGL
FVACRHSEARFIARSLSGRLRLGLAEQSVLAALSQAVSLTTPPGQEFPAMVDAGKGTAEARKTWLEEQGMILKQ
TFCEVPDLRIIPVLLHGLERLPEHCKLSPGIPLKPLAHPTRGISEVLKRFEEAAFTCEYKYDQRAQIHALEGG
EVKIFSRNQEDNTGKYPDIISRIPIKLPVSTFILDTEAVAWDREKKIQPFQVLTRKRKEVDASEIQVQVCLYAFD
LIYLNESLVREPLSRRRQLLRENFVETEGEFVFATSLDTKDIEQIAEFLEQSVKDSCEGLMVKTLDVDATYEIAKRS
HNWLKLLKDYLDVGGDTLDLVVIGAYLGRGKRAGRYGGFLLASYDEDSEELQAICKLGTGFSDEELEEHHQSLKA
LVLPSRPYVRIDGAVIPDHWLDPSAVWEVKCADLSLPIYPAARGLVSDKGISLRFPRFIRVREDKQPEQATTS
QVACLYRKQSQIQNQGGEDSGSDPEDTY*

Sequence of plasmids used in this study

pET28a-hLig1

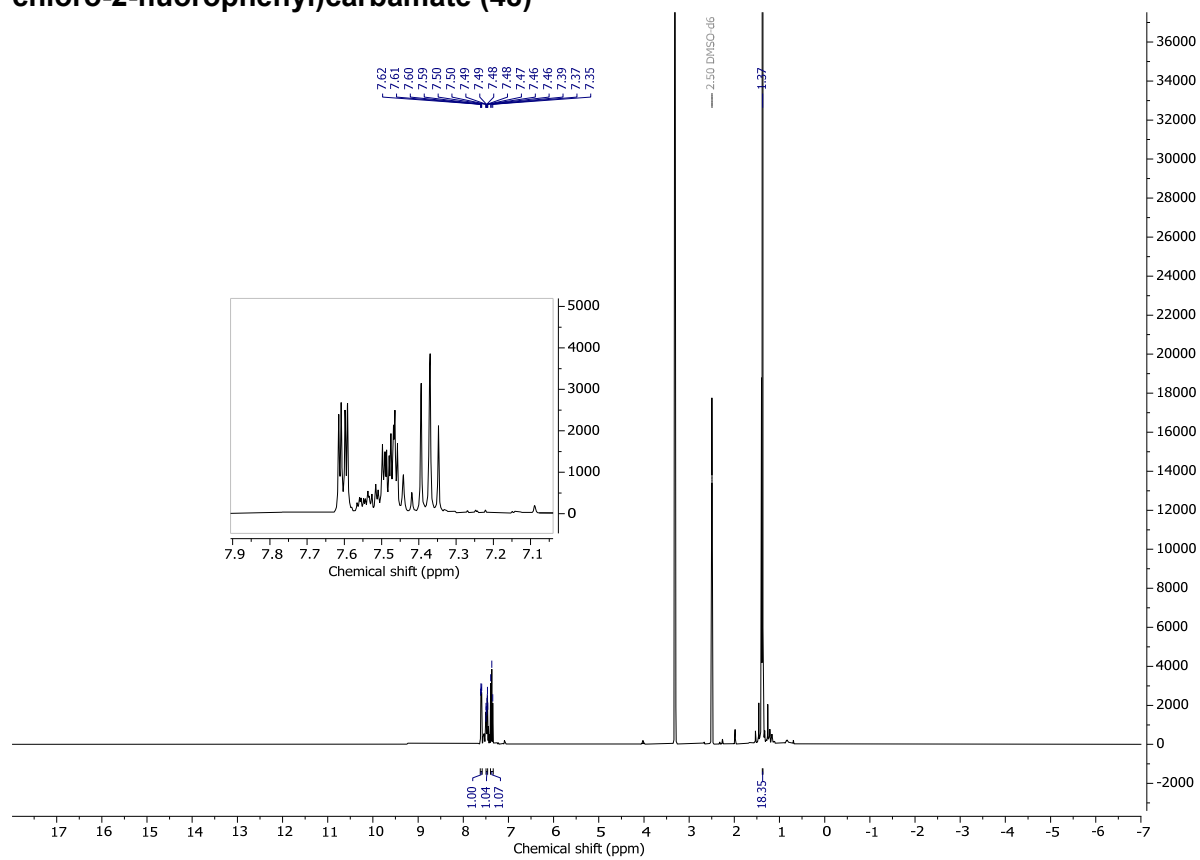
TGGCGAATGGGACGCGCCCTGTAGCGGCGCATTAAAGCGCGGCGGGTGTGGTGGTTACGCGCAGCGTGACC
GCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTTTTCGCTTTCTTCCCTTCTTCTCGCCACGTTCCGCCGC
TTTCCCCGTCAAGCTCTAAATCGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTTTACGGCACCTCGACCCCA
AAAACTTGATTAGGGTGATGGTTCACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCCCTTTGACGTT
GGAGTCCACGTTCTTTAATAGTGACTCTTGTTCCAAACACTGGAACAACACTCAACCCTATCTCGGTCTATTCTT
TTGATTTATAAGGATTTTCCGATTTCCGGCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTTAACGCGAA
TTTTAACAAAAATTAACGCTTACAATTTAGGTGGCACTTTTCGGGGAAATGTGCGCGGAACCCCTATTTGTTTA
TTTTTCTAAATACATTCAAATATGTATCCGCTCATGAATTAATCTTAGAAAAACTCATCGAGCATCAAATGAACT
GCAATTTATTCATATCAGGATTTCAATACCATATTTTTGAAAAAGCCGTTTCTGTAATGAAGGAGAAAACTCACC

GAGGCAGTTCATAGGATGGCAAGATCCTGGTATCGGTCTGCGATTCCGACTCGTCCAACATCAATACAACCT
ATTAATTTCCCCTCGTCAAAAATAAGGTTATCAAGTGAGAAATCACCATGAGTGACGACTGAATCCGGTGAGAA
TGGCAAAAAGTTTATGCATTTCTTCCAGACTTGCTCAACAGGCCAGCCATTACGCTCGTCAAAAATCACTCG
CATCAACCAAACCGTTATTCATTCTGATTGCGCCTGAGCGAGACGAAATACGCGATCGCTGTAAAAGGACA
ATTACAAACAGGAATCGAATGCAACCGGCCGAGGAACACTGCCAGCGCATCAACAATATTTTACCTGAATCA
GGATATTCTTCTAATACCTGGAATGCTGTTTTCCCGGGGATCGCAGTGGTGAGTAACCATGCATCATCAGGAGT
ACGGATAAAATGCTTGATGGTCGGAAGAGGCATAAATCCGTCAGCCAGTTTAGTCTGACCATCTCATCTGTAA
CATCATTGGCAACGCTACCTTTGCCATGTTTCAGAAACAACCTCTGGCGCATCGGGCTTCCCATACAATCGATAG
ATTGTGCGACCTGATTGCCCGACATTATCGCGAGCCATTTATACCCATATAAATCAGCATCCATGTTGGAATTT
AATCGCGGCCTAGAGCAAGACGTTTTCCCGTTGAATATGGCTCATAACACCCCTTGTATTACTGTTTATGTAAGC
AGACAGTTTTATTGTTTCATGACCAAAATCCCTAACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCGTAGAA
AAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAACCACCGCT
ACCAGCGGTGGTTTGTGGCCGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAACCTGGCTTCAGCAGAGC
GCAGATACCAATACTGTCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACTCTGTAGCACCCGCTA
CATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCTTACCGGGTTGGA
CTCAAGACGATAGTTACCGGATAAGGCCGAGCGGTGGGGTGAACGGGGGTTCTGTCACACAGCCAGCT
TGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTATTAGAAAGCGCCACGCTTCCCGAAG
GGAGAAAAGGCCGACAGGTATCCGGTAAGCGGCAGGGTGGAAACAGGAGAGCGCACGAGGGGAGCTTCCAGG
GGGAAACGCCTGGTATCTTTATAGTCTGTGGGTTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGATGC
TCGTGAGGGGGGGCGGAGCCTATGGAAAACGCCAGCAACGCGGCCTTTTTACGGTTCTTGGCCTTTTGCTG
GCCTTTTGCTCATATGTTCTTCTGCGTTATCCCTGATTCTGTGGATAACCGTATTACCGCCTTTGAGTGAG
CTGATACCGCTCGCCGACGCCAACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAGAGCGCCT
GATGCGGTATTTCTCCTTACGCATCTGTGCGGTATTTACACCCGCAATGGTGCACCTCTCAGTACAATCTGCTC
TGATGCCGCATAGTTAAGCCAGTATACACTCCGCTATCGCTACGTGACTGGGTGATGGCTGCGCCCCGACACC
CGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAGACAAGCTGTGACC
GTCTCCGGGAGCTGCATGTGTGAGAGTTTTACCGTATCACCGAAACGCGCGAGGCAGCTGCGGTAAAG
CTCATCAGCGTGGTCTGAAGCGATTACAGATGTCTGCCTGTTTCATCCGCGTCCAGCTCGTTGAGTTTTCTCC
AGAAGCGTTAATGTCTGGCTTCTGATAAAGCGGGCCATGTTAAGGGCGGTTTTTCTGTTTGGTCACTGATG
CCTCCGTGTAAGGGGGATTTCTGTTTCATGGGGTAATGATACCGATGAAACGAGAGAGGATGCTCACGATACG
GGTACTGATGATGAACATGCCCGTTACTGGAACGTTGTGAGGGTAAACAACCTGGCGGTATGGATGCGGGC
GGACCAGAGAAAAATCACTCAGGGTCAATGCCAGCGCTTCTGTTAATACAGATGTAGGTGTTCCACAGGGTAGC
CAGCAGCATCCTGCGATGCAGATCCGGAACATAATGGTGCAGGGCGCTGACTTCCGCGTTTTCCAGACTTTAC
GAAACACGGAACCGAAGACCATTATGTTGTTGCTCAGGTGCGAGACGTTTTGCAGCAGCAGTGCCTTAC
GTTGCTGCGTATCGGTGATTCTGCTAACCAGTAAGGCAACCCCGCCAGCCTAGCCGGTCTCAAC
GACAGGAGCACGATCATGCGCACCCGTGGGGCCGCTGCGGCGATAATGGCCTGCTTCTGCGCGAAACG
TTTGGTGGCGGGACCAGTGACGAAGGCTTGAGCGAGGGCGTGCAAGATTCCGAATACCGCAAGCGACAGGC
CGATCATCGTCCGCTCCAGCGAAAGCGGTCTCGCCGAAAAATGACCCAGAGCGCTGCCGGCACCTGTCT
ACGAGTTGCATGATAAAGAAGACAGTATAAGTGGCGGACGATAGTCATGCCCCGCGCCACCCGGAAGGAG
CTGACTGGGTTGAAGGCTCTCAAGGGCATCGGTGCGAGATCCCGGTGCCTAATGAGTGAGCTAACTTACATTA
TTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTGTCGAGCTGCATTAATGAATCGGCCAAC
GCGCGGGGAGGCGGTTTTGCGTATTGGCGCCAGGGTGGTTTTCTTTTACCAGTGAGACGGGCAACAG
CTGATTGCCCTTACCAGCTGGCCCTGAGAGAGTTGACAGCAAGCGGTCCACGCTGGTTTGCCCCAGCAGGC
GAAAATCCTGTTTGGTGGTTAACGGCGGGATATAACATGAGCTGTCTTCCGATCGTCTGATCCCCTACC
GAGATATCCGCACCAACGCGCAGCCCGACTCGGTAATGGCGCGCATTGCGCCAGCGCCATCTGATCGTT
GGCAACCAGCATCGCAGTGGGAACGATGCCCTAATTCAGCATTGTCATGGTTTGTGAAAACCGGACATGGC
ACTCCAGTCCCTTCCCGTTCCGCTATCGGCTGAATTTGATTGCGAGTGAGATATTTATGCCAGCCAGCCAGA
CGCAGACGCGCCGAGACAGAACTTAATGGGCCCGCTAACAGCGCGATTTGCTGGTGACCCAATGCGACCAG
ATGCTCCACGCCAGTCCGCTACCGTCTCATGGGAGAAAATAATACTGTTGATGGGTGTCTGGTCAGAGACA
TCAAGAAAATAACGCCGGAACATTAGTGCAGGCAGCTTCCACAGCAATGGCATCCTGGTATCCAGCGGATAGT
TAATGATCAGCCACTGACGCGTTGCGCGAGAAGATTGTGCACCGCCGCTTTACAGGCTTCGACGCGGCTTC
GTTCTACCATCGACACCACGCTGGCACCCAGTTGATCGGCGGAGATTAATCGCCGCGACAATTTGCG
ACGGCGCGTGCAGGGCCAGACTGGAGGTGGCAACGCCAATCAGCAACGACTGTTTGCCCGCCAGTTGTTGT
GCCACGCGGTTGGGAATGTAATTCAGCTCCGCCATCGCCGCTTCCACTTTTTTCCCGGTTTTTGCAGAAACG
TGGCTGGCCTGGTTACCACGCGGGAAACGGTCTGATAAGAGACACCGGCATACTCTGCGACATCGTATAAC
GTTACTGGTTTTACATTCACCACCCTGAATGACTCTCTTCCGGGCGCTATCATGCCATAACCGGAAAGGTTTT
GCGCCATTGATGGTGTCCGGGATCTCGACGCTCTCCCTTATGCGACTCCTGCATTAGGAAGCAGCCAGTA
GTAGGTTGAGGCCGTTGAGCACCGCCGCGCAAGGAATGGTGCATGCAAGGAGATGGCGCCCAACAGTCCC
CCGGCCACGGGGCCTGCCACCATACCCACGCCGAAACAAGCGCTCATGAGCCGAAGTGGCGAGCCCGAT
CTTCCCATCGGTGATGTGGCGATATAGGCCGAGCAACCGCACCTGTGGCGCCGGTGTGCGGGCCACG

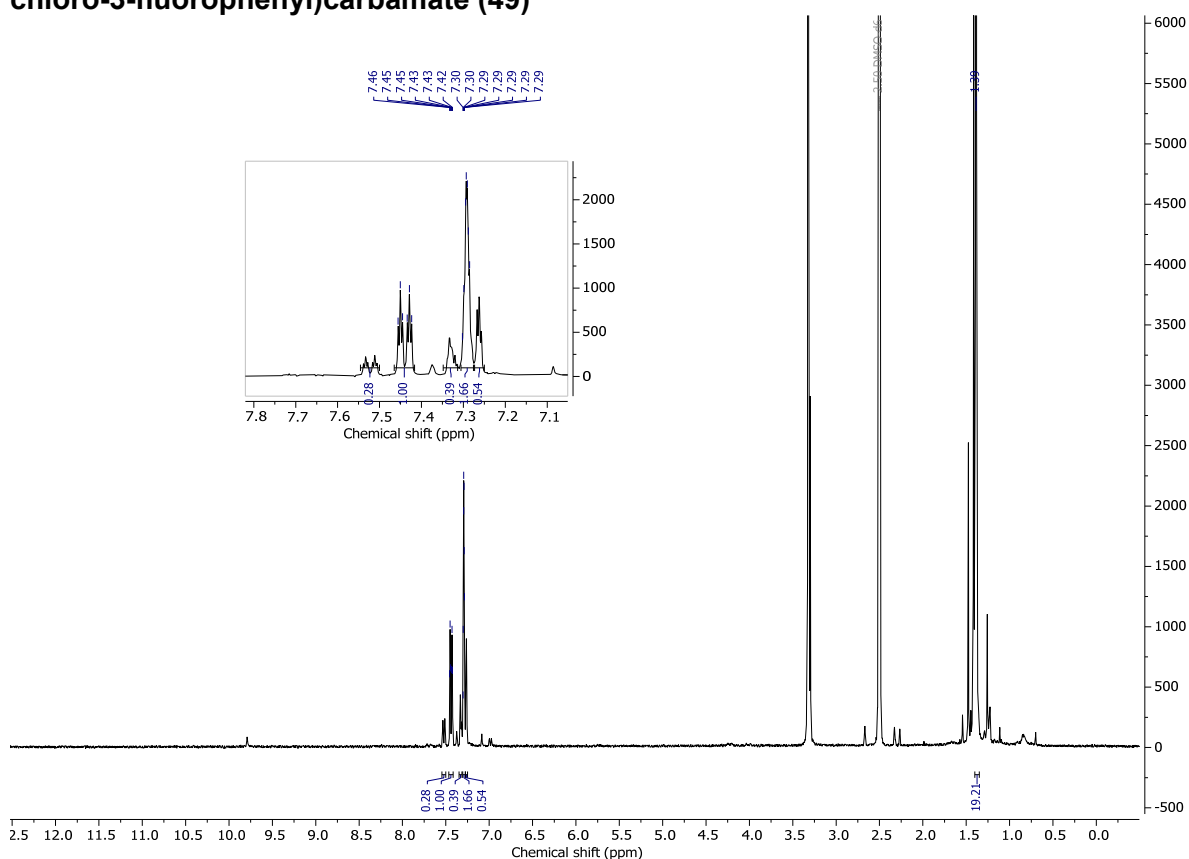
ATGCGTCCGCGCTAGAGGATCGAGATCTCGATCCCGCGAAATTAATACGACTCACTATAGGGAGAGGAATTGT
GAGCGGATAACAATTCCCCTCTAGAAATAATTTTGTTTAACTTTAAGAAGGAGAGCAGCTATGCAGCTTAGCCAT
CATCATCATCACAGCAGCGGCGAAAACCTGTATTTTCAGGGCCATATGATGCAACGCTCGATTATGAGCTT
CTTTCACCCCTAAAAAGAGGGGAAAGCTAAAAAGCCAGAGAAGGAAGCATCTAACTCTTCACGTGAGACTGA
GCCGCCTCCCAAGGCTGCGTTGAAAGAGTGAATGGCGTTGTAAGTAAAAGTGACTCTCCCGTCAAACGCC
CCGGGCGCAAGGCTGCACGTGTATTAGGTTGAGAAGGAGAGGAGGAAGACGAAGCCTTGTGCGCTGCTAAA
GGGCAAAAGCCCGCGCTTGACTGCTCTCAAGTGTCCCTCCTCGTCTGCGACTAGCCCAGAGAATAATGCA
AGTCTGTCCGATAACCACTTATGGATTGTCCTTAGCGGGATCCCAAAGCGCCGTAAGTGCACGTAAGCAG
CTTCCCAAACGTACCAATTCAGGAAGTCTTGAAGAGCAATCGGAAGATGAGGATCGTGAAGCAAAACGCAAA
AAGGAGGAGGAGGAAGAAGAACTCCGAAGGAGAGCCTGACTGAAGCTGAGGTGCGAACCCGAGAAGGAGG
GTGAGGATGGCGATCAGCCAACAACGCCACCGAAGCCACTGAAGACCTCAAAGGCAGAGACGCCAACCGAG
AGCGTGAGCGAACCCGAGGTTGCTACAAAGCAAGAGTTGCAGGAGGAGGAAGAGCAGACTAAACCCCCACG
TCGCGCACCGAAAACATTATCGTCTTCTTTACCCCGCGTAAACCCGCGAGTAAAAAGGAGGTCAAGGAAGA
AGAGCCGGGCGCACCAAGTAAGGAAGGTGCAGCGGAAGGACCTTTGATCCAAGTGGGTATAACCCCGCAA
AAAATAACTATCACCCCGTAGAGGATGCCTGTTGGAAGCCGGGGCAGAAAAGTCCCCTACTTAGCGGTGCGTC
GTACGTTGCAAAAAATTGAAGAAGTATCGGCGCGTCTGCGTATGGTGGAGACACTTTCCAATCTGCTGCGCTC
TGTTGTTGCCCTGTGCGCTCCCGATTTGTTGCCAGTTTTGTACCTGAGCTTGAACCACCTGGGCCCTCCTCAA
CAAGGCTTAGAATTGGGGGTAGGCGATGGTGTATTGTTGAAAGCAGTAGCGCAAGCCACGGGCCGTCAACTG
GAGTCCGTTGTCGAGAGGCAGCGGAAAAGGTGATGTAGGGTTAGTCGCTGAAAACAGCCGAGTACGCA
GCGTTTGATGTTGCCCGCCACCCCTGACGGCCTCCGGAGTGTCTCAAAGTTCGCGATATCGCCCGTTT
AACCGGGTCAGCATCTACAGCGAAAAGATTGACATCATCAAAGGCCTGTTGTTGCTTGTGCGCCACAGCGA
GGCTCGTTTTATTGCGCGCTCTTATCCGGGCGCCTTCGTCTGGGCTTAGCCGAACAGTCAGTGTGTCAGC
ATTGTACAGGCCGTTTCGCTTACTCCCCAGGGCAGGAGTTCCCCCTGCCATGGTAGACGCTGGCAAAG
GGAAAAGTGCAGGAGGCCGTAAGACATGGTTGGAGGAGCAAGGTATGATCCTTAAGCAGACCTTTTGCAGG
TCCCAGACCTGGACCGCATCATCCCTGTGTTGCTGGAGCATGGCTTAGAACGCCTTCCGGAGCATTGCAAGC
TGTCCCCTGGGATTCCGTTAAAGCCGATGTTAGCCCATCCTACCCGTGGTATTTTCAGAGGTTCTGAAGCGCTT
CGAAGAGGCTGCCTTACATGTGAATATAAATACGACGGGCAACGCGCACAAATCCACGCTTTGGAGGGGG
CGAGGTCAAGATCTTTTCGCGTAATCAAGAGGACAACACGGGCAAATACCCCGACATTATTTGCGCGATCCCT
AAAATTAAGTGCAGGTTACGAGCTTCAATTTAGATACAGAAGCGGTGGCTTGGGACCGCGAGAAAAAGC
AAATTCAGCCCTTCCAGGTCTTGACCACGCGTAAACGTAAGGAGGTTGACGCGTCGGAAATCCAAGTACAAG
TCTGTCTGTACGCTTCGATTTAATCTATTTAAATGGTAAAAGTTTGGTGCCTGAACCATTATCCCGCCGCGT
CAGCTGCTTCGTGAAAACCTCGTTGAAACCGAGGGCGAATTTGTTTTGCGACTTCACTGGATACAAAAGACA
TCGAACAAATCGCGGAGTTCCTTGAGCAATCCGTCAAAGACAGTTGCGAGGGACTGATGGTCAAGACCTTAG
ACGTTGATGCCACTTATGAGATCGCTAAACGCTCCCATTAATTGTTGAAACTGAAAAAGGACTACCTTGATGGC
GTGGGTGATACACTTGACCTTGTGGTCATCGGAGCCTATCTGGGACGTGGTAAGCGCGCAGGACGCTACGGT
GGTTTCTTGTTAGCCTCCTATGACGAGGATAGTGAGGAATTGCAGGCCATTTGTAAGCTGGGTACGGGGTTCT
CTGACGAAGAGTTGGAGGAACACCACCAATCACTTAAAGCATTGGTCTTGCACAGTCCCTGCTCCCTACGTCC
GTATCGACGGGGCAGTAATTCGGATCACTGGCTTGACCCTAGCGCGGTCTGGGAAGTTAAATGCGCTGATC
TGTCCTTAAGTCCAATCTATCCCGCGGCACGCGGGCTTGTGATTCTGACAAAGGGATCTCGTTCGTTTTCC
CCGTTTCATCCGTGTGCGCGAGGACAAACAACCCGAACAAGCCACGACGAGTGCACAGGTGGCGTCCCTGT
ACCGCAACAATCGCAGATTCAGAATCAGCAAGGCGAAGATTCAGGGAGCGACCCGGAGGACACGTATTAAT
GAGGATCCCTCGAGCACCACCACCACCCTGAGATCCGGCTGCTAACAAAGCCCCGAAAGGAAGCTGAG
TTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCCTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTT
TGCTGAAAGGAGGAACTATATCCGGAT

NMR Spectra

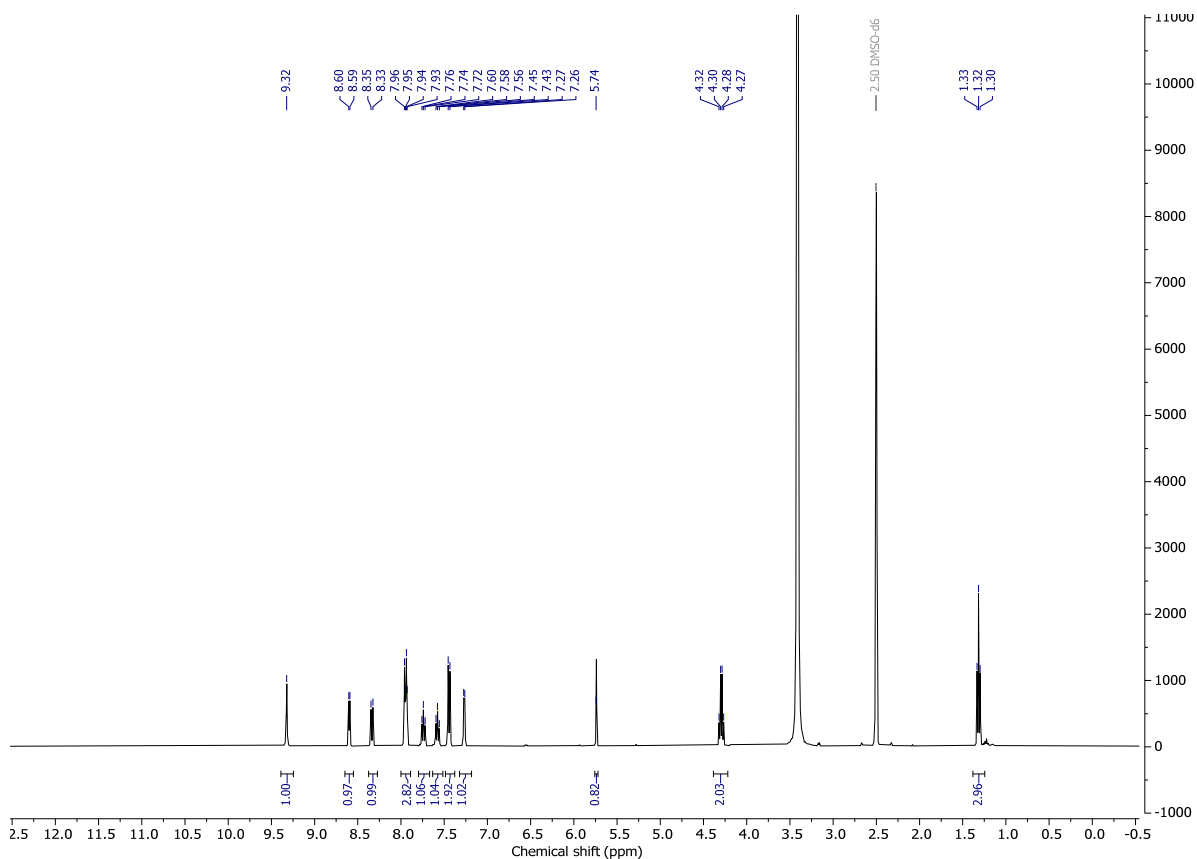
tert-Butyl (tert-butoxycarbonyl)(5-chloro-2-fluorophenyl)carbamate & *tert*-butyl (5-chloro-2-fluorophenyl)carbamate (48)



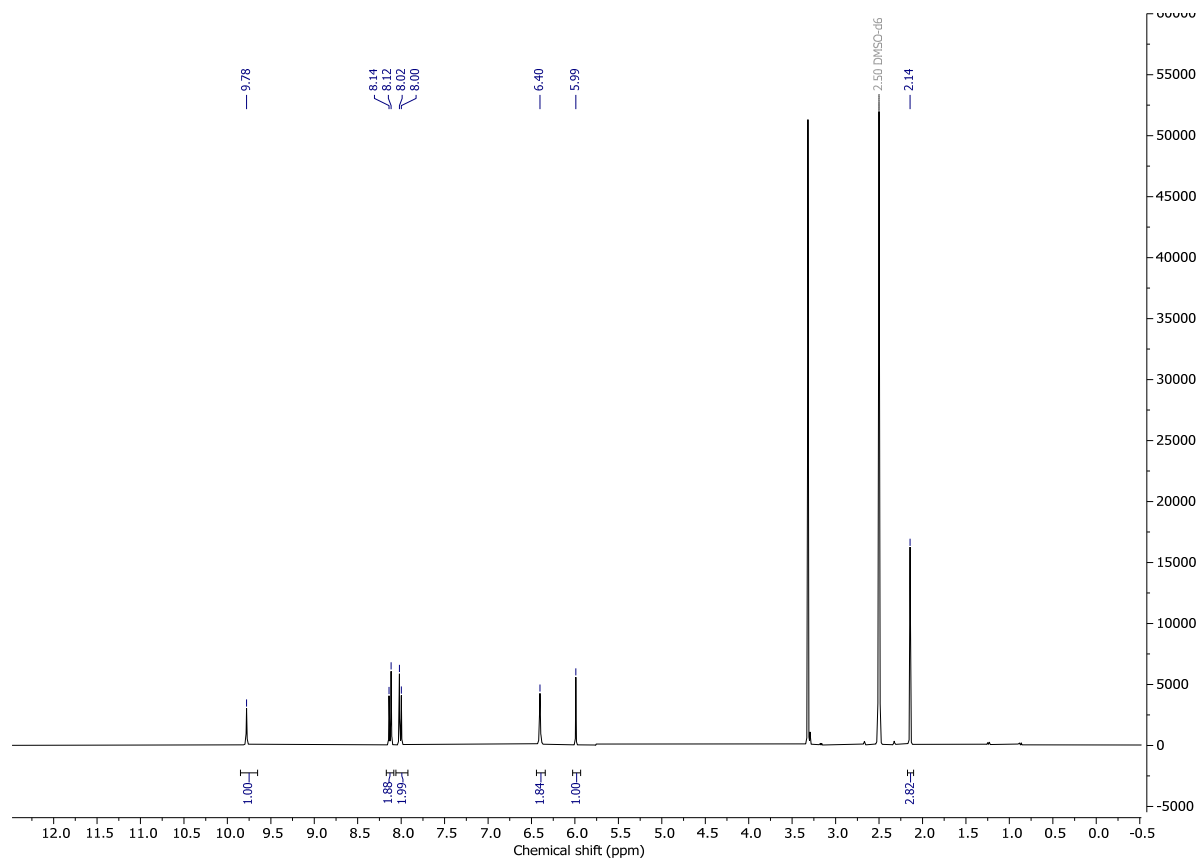
tert-Butyl (tert-butoxycarbonyl)(5-chloro-3-fluorophenyl)carbamate & tert-butyl (5-chloro-3-fluorophenyl)carbamate (49)



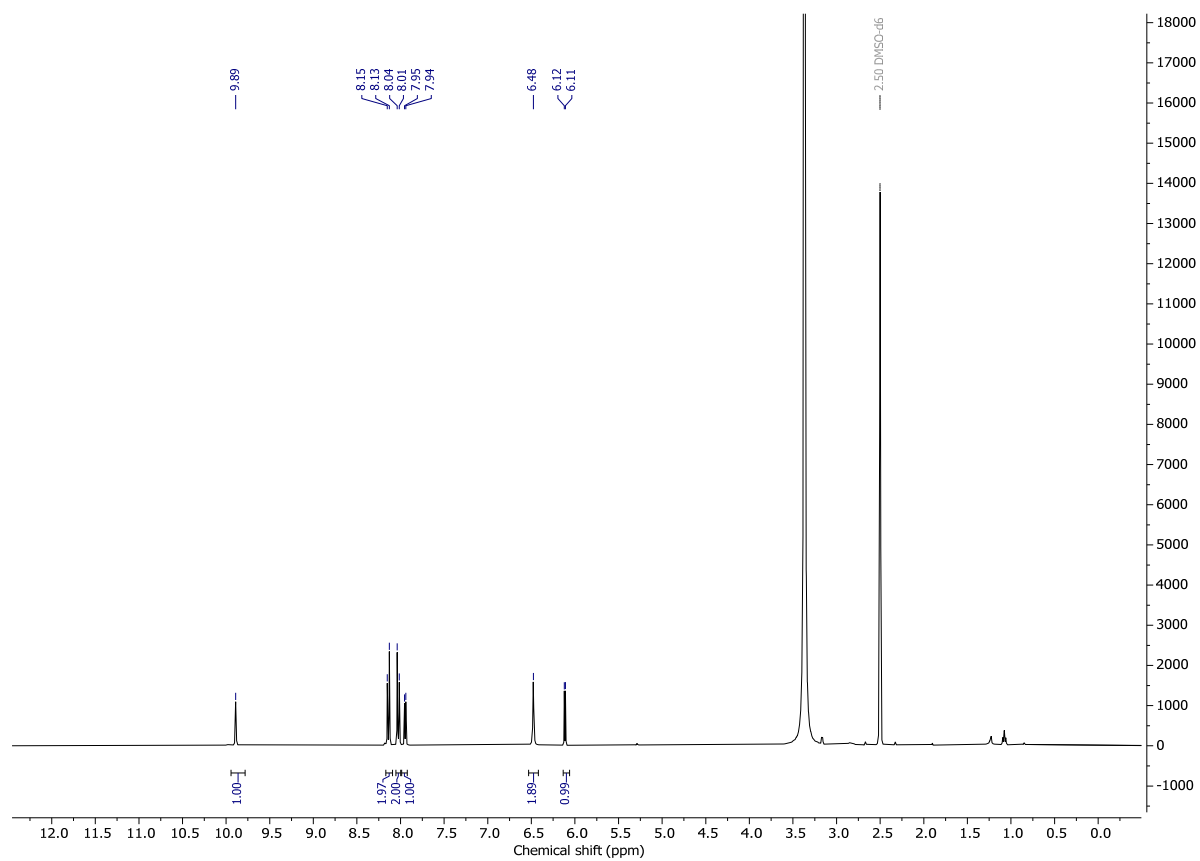
Ethyl 4-(quinolin-4-ylamino)benzoate (50)

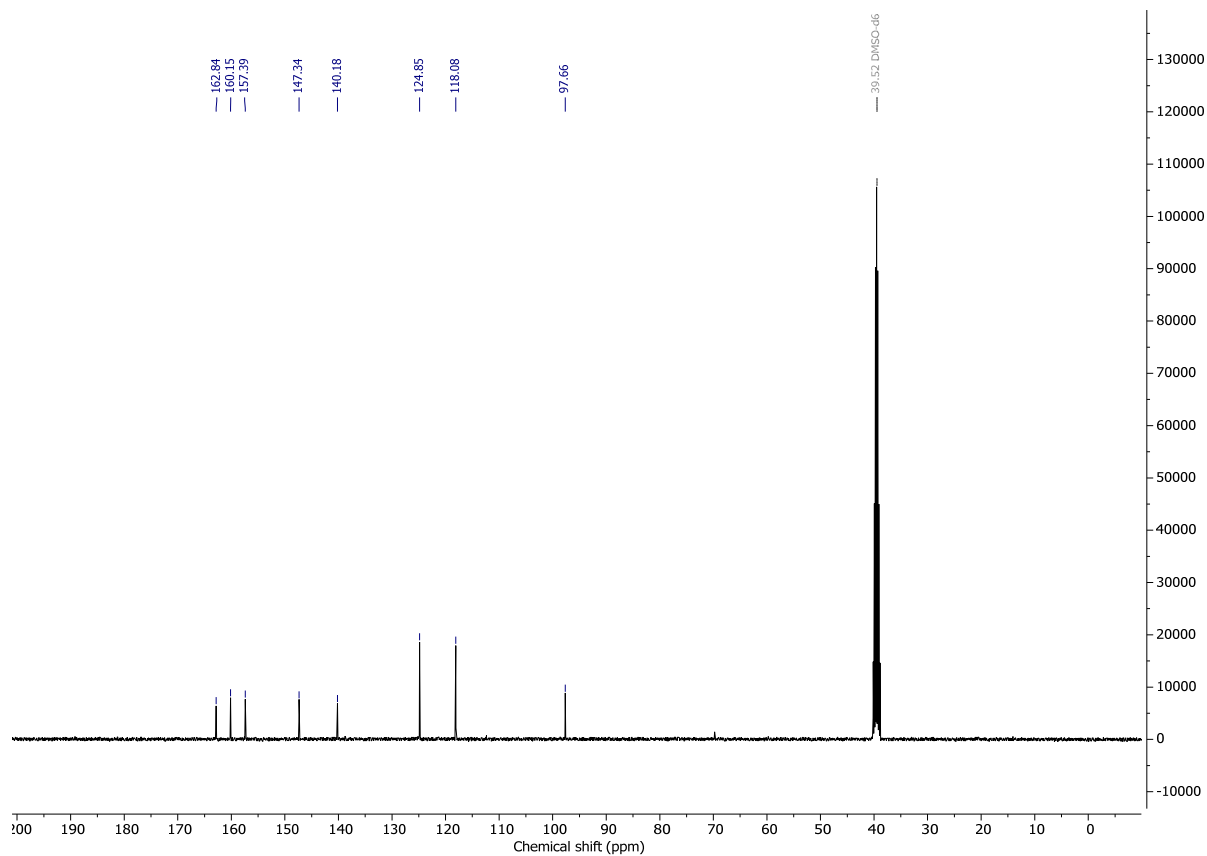


6-Methyl-*N*⁴-(4-nitrophenyl)pyrimidine-2,4-diamine (51)

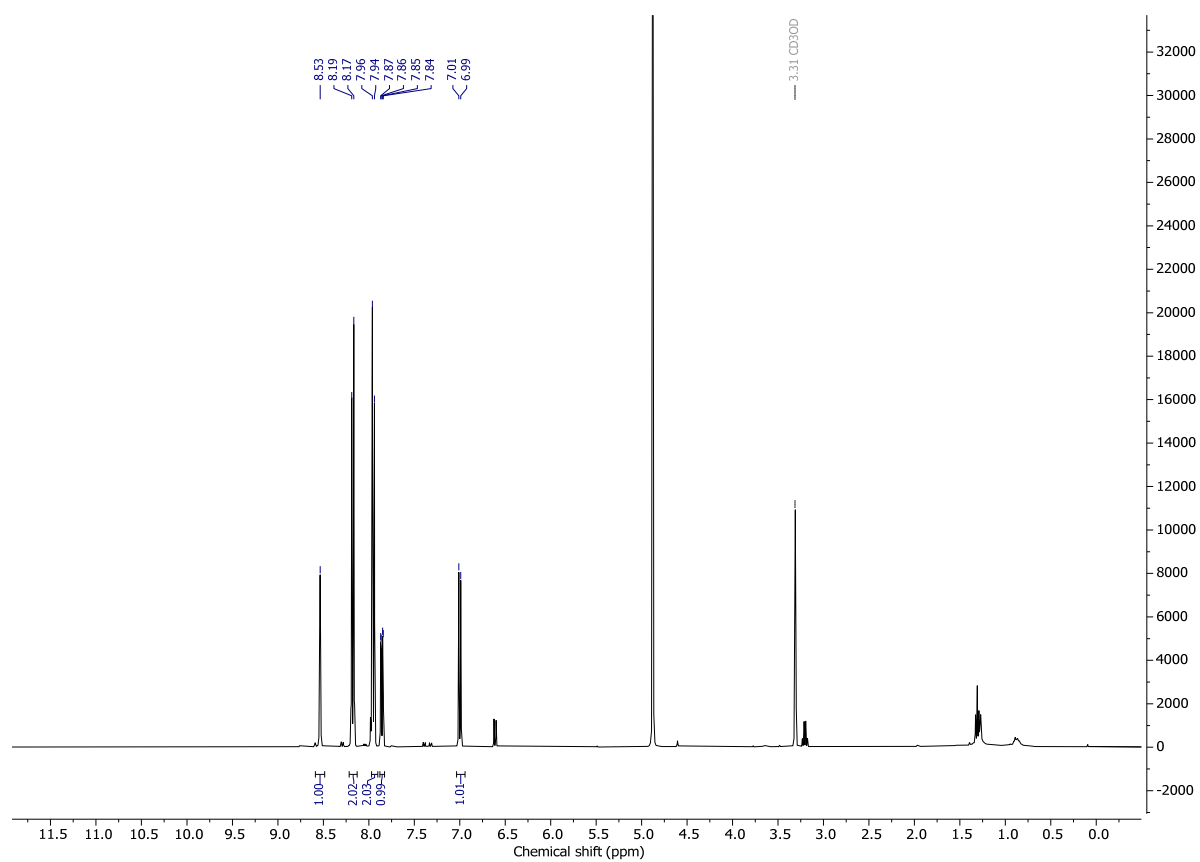


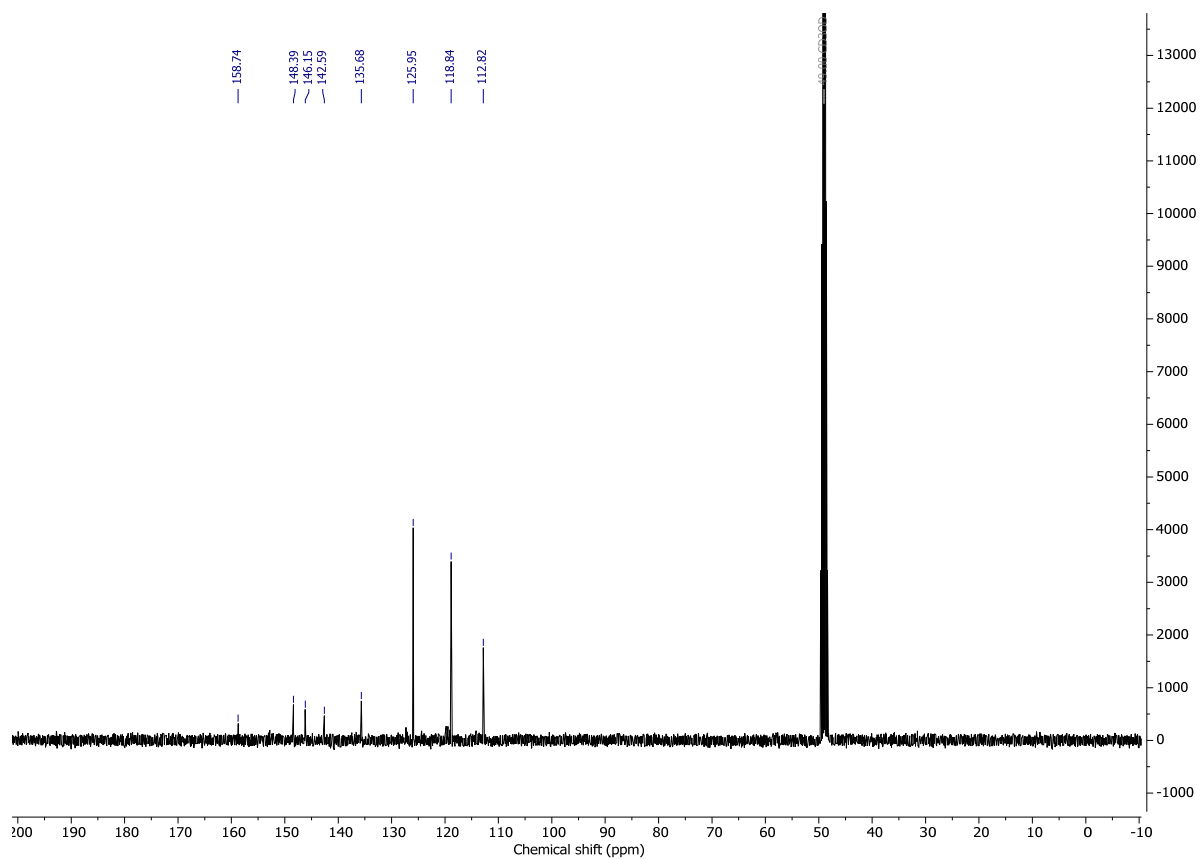
*N*⁴-(4-Nitrophenyl)pyrimidine-2,4-diamine (52)



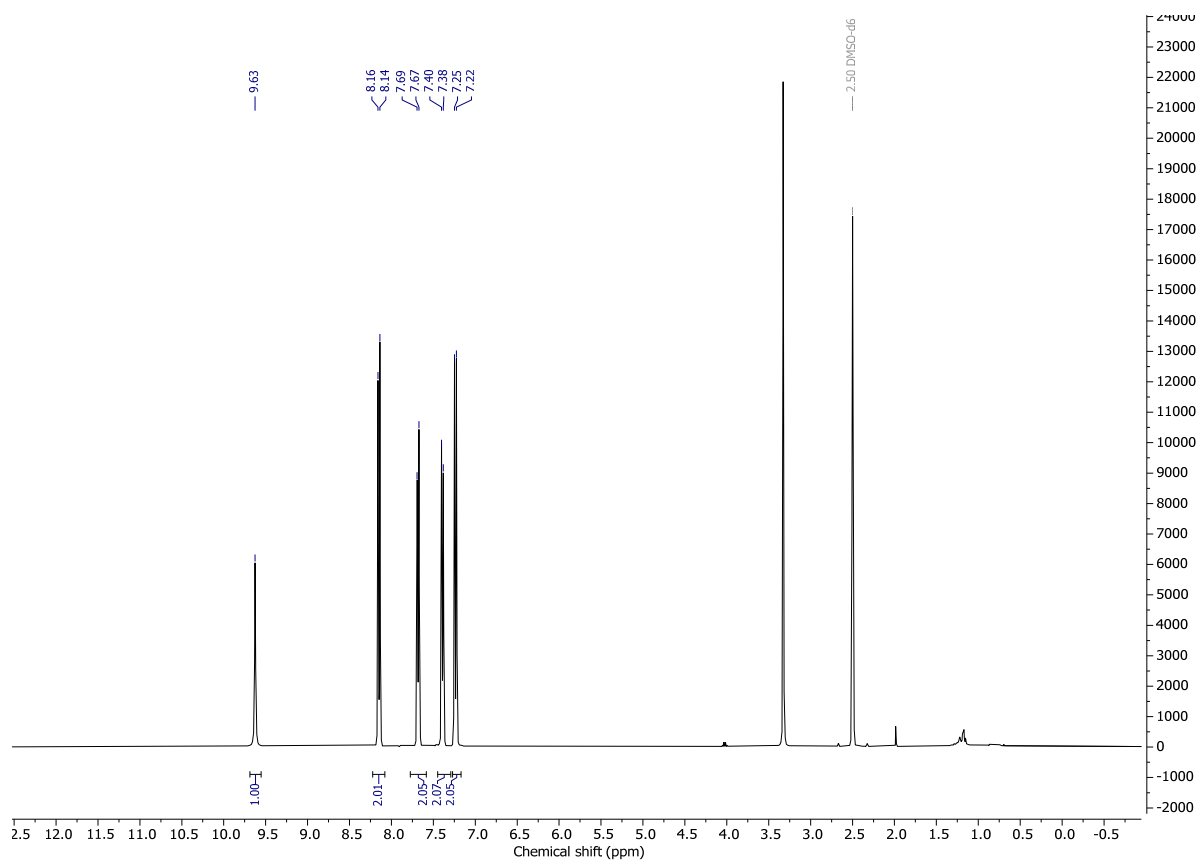


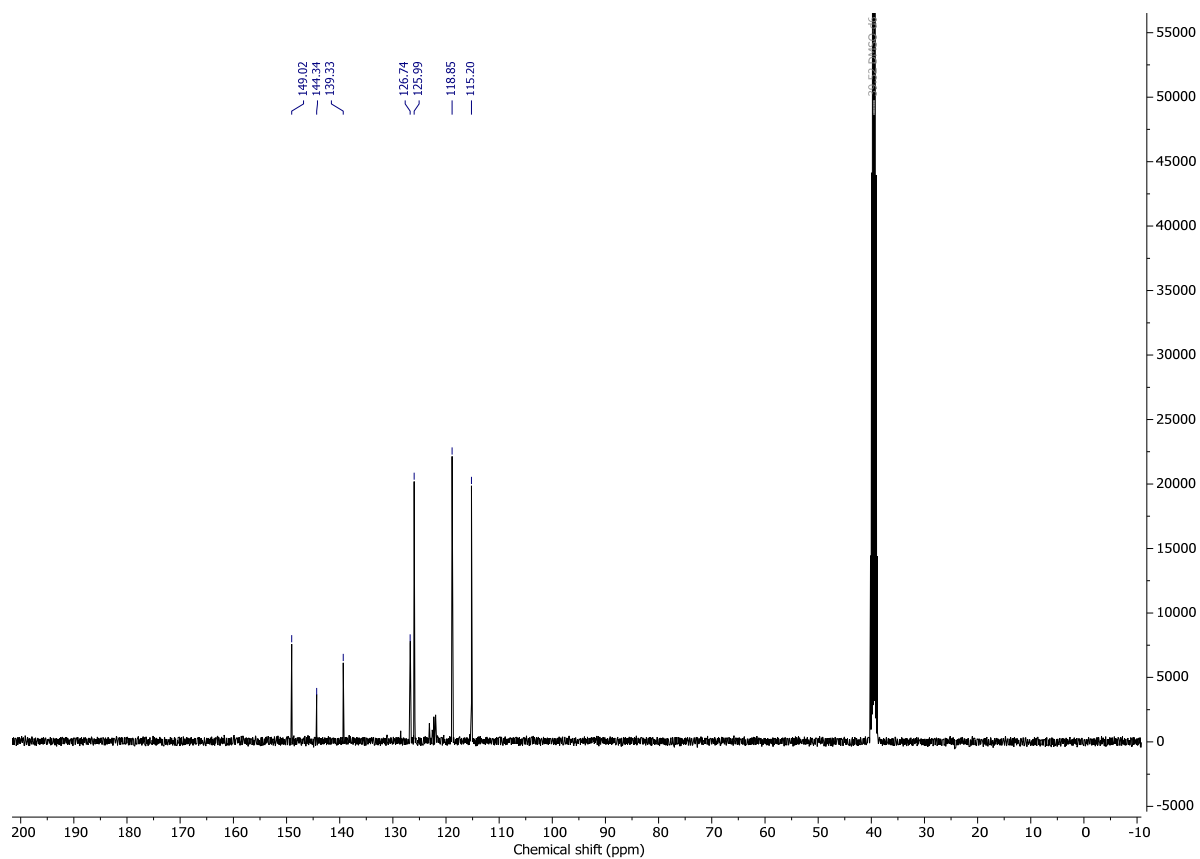
***N*-(4-Nitrophenyl)-5-(trifluoromethyl)pyridin-2-amine (53)**



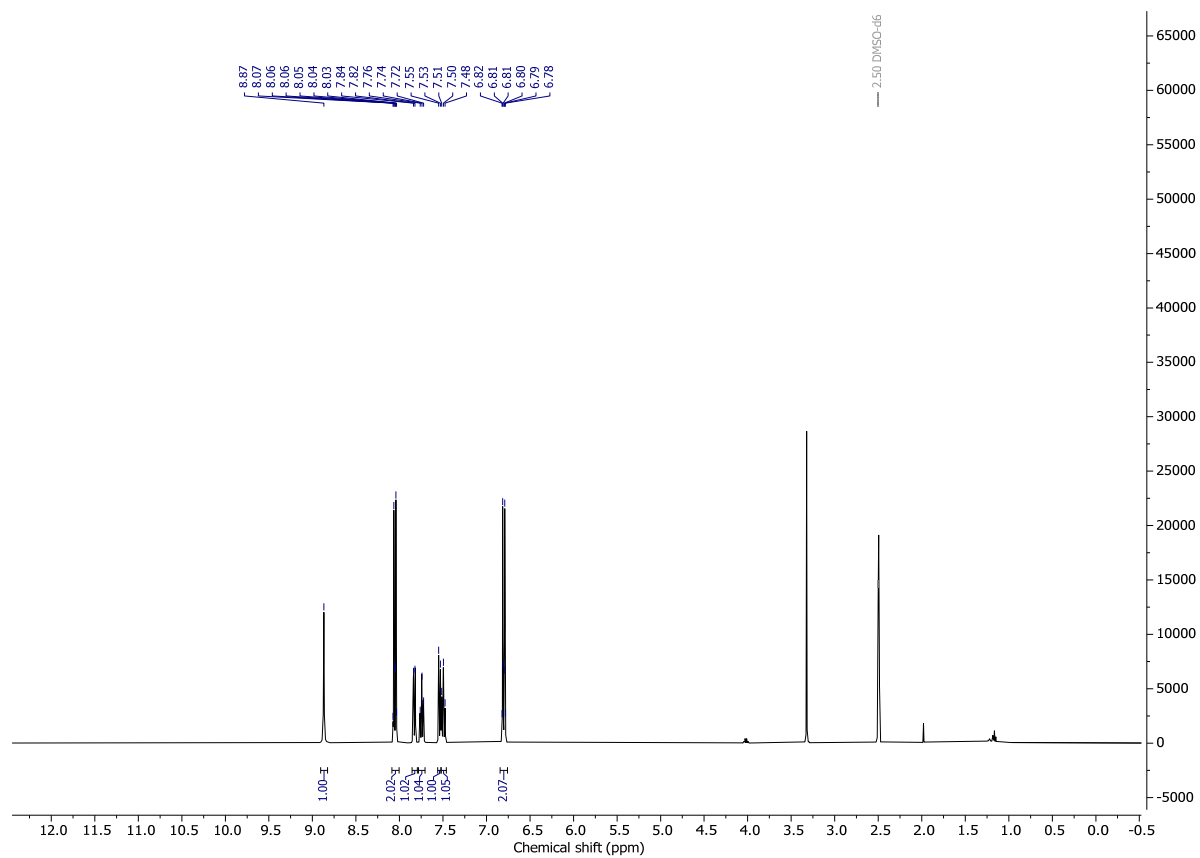


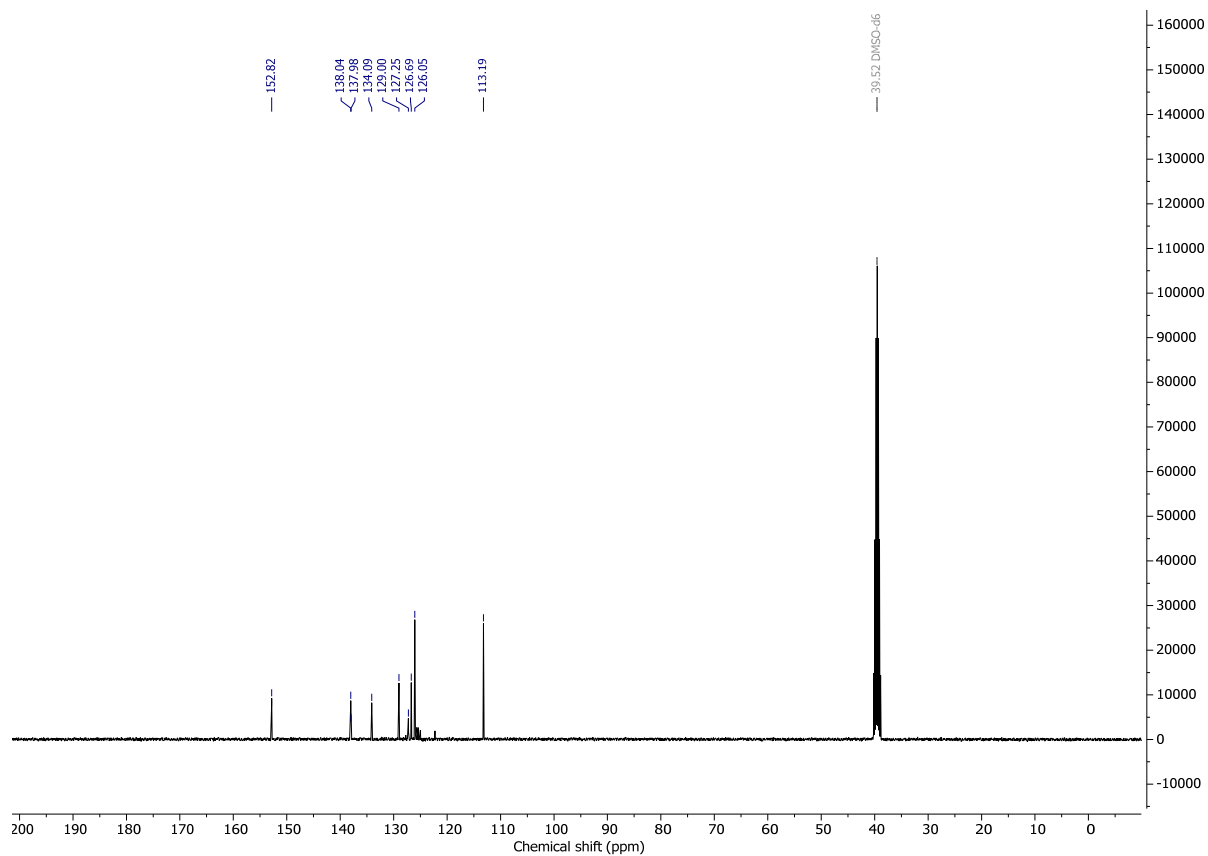
4-Nitro-N-(4-(trifluoromethyl)phenyl)aniline (54)



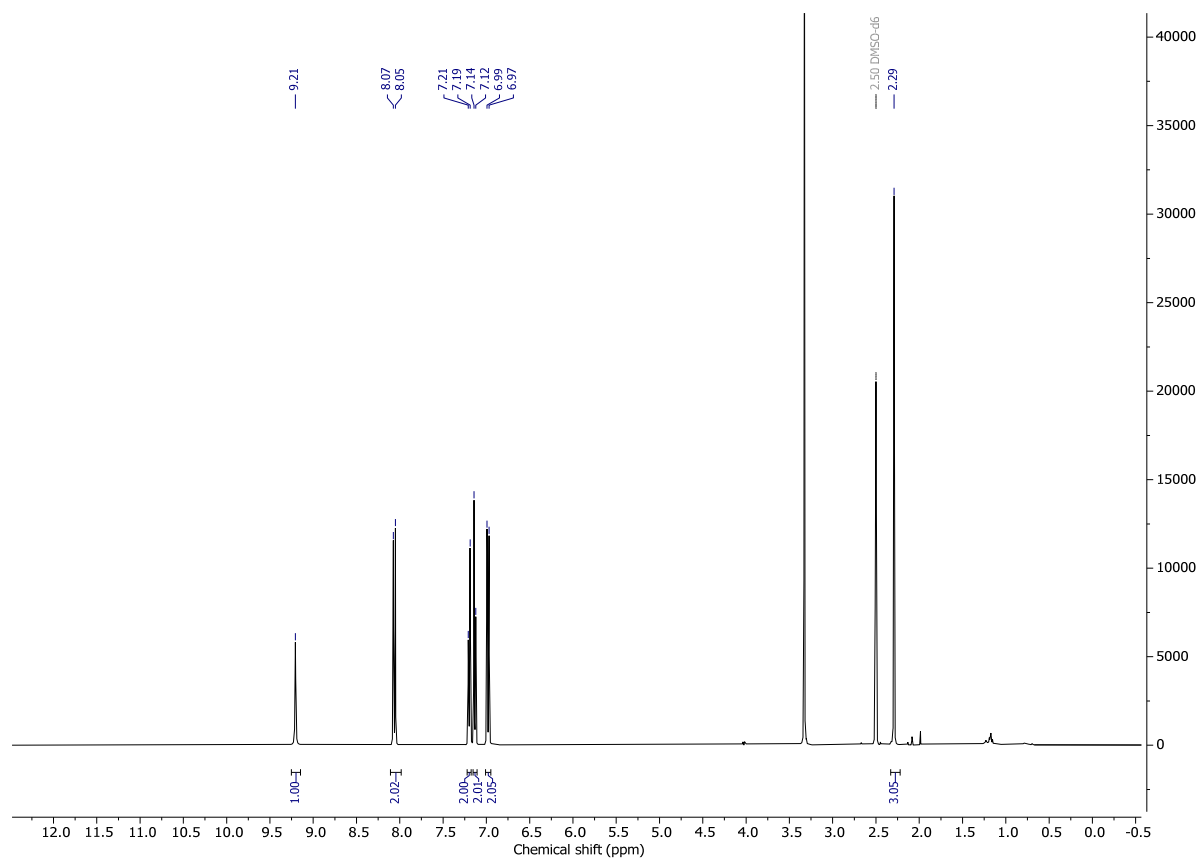


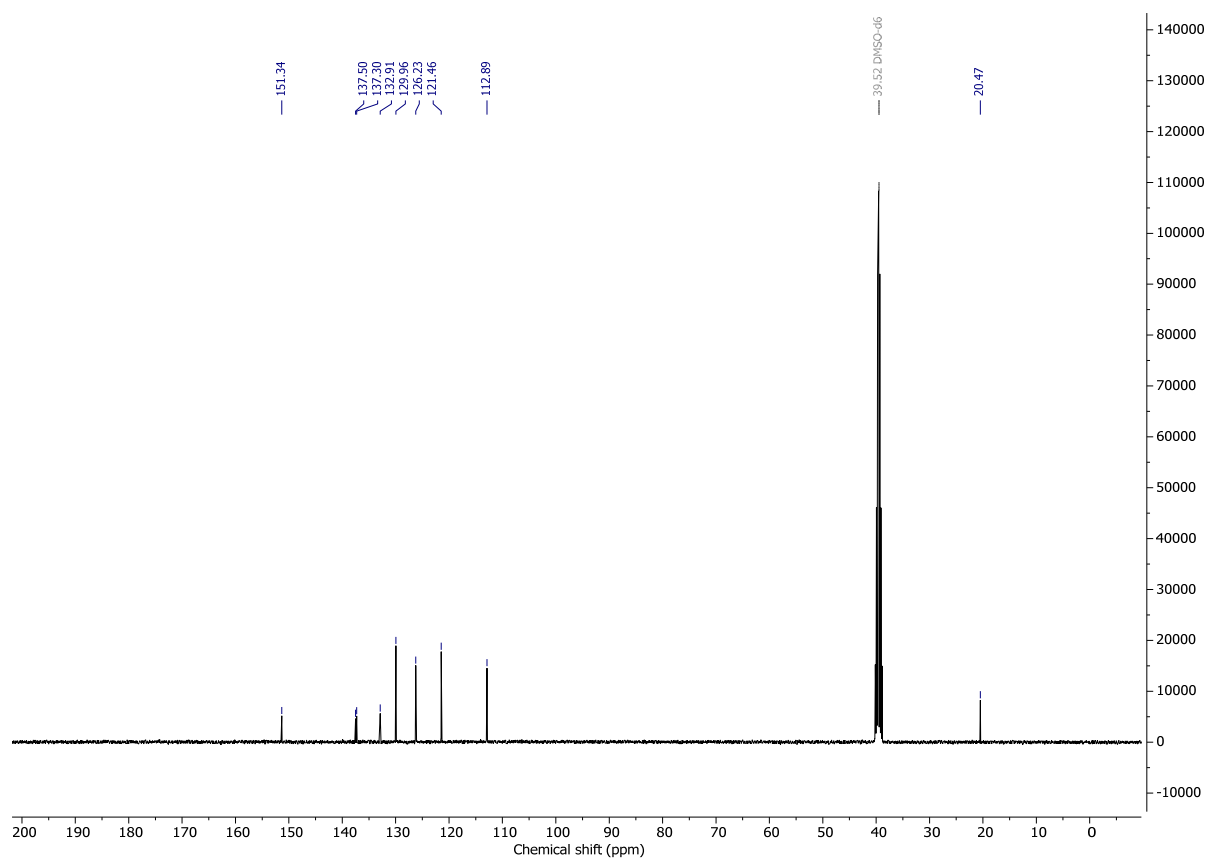
***N*-(4-Nitrophenyl)-2-(trifluoromethyl)aniline (55)**



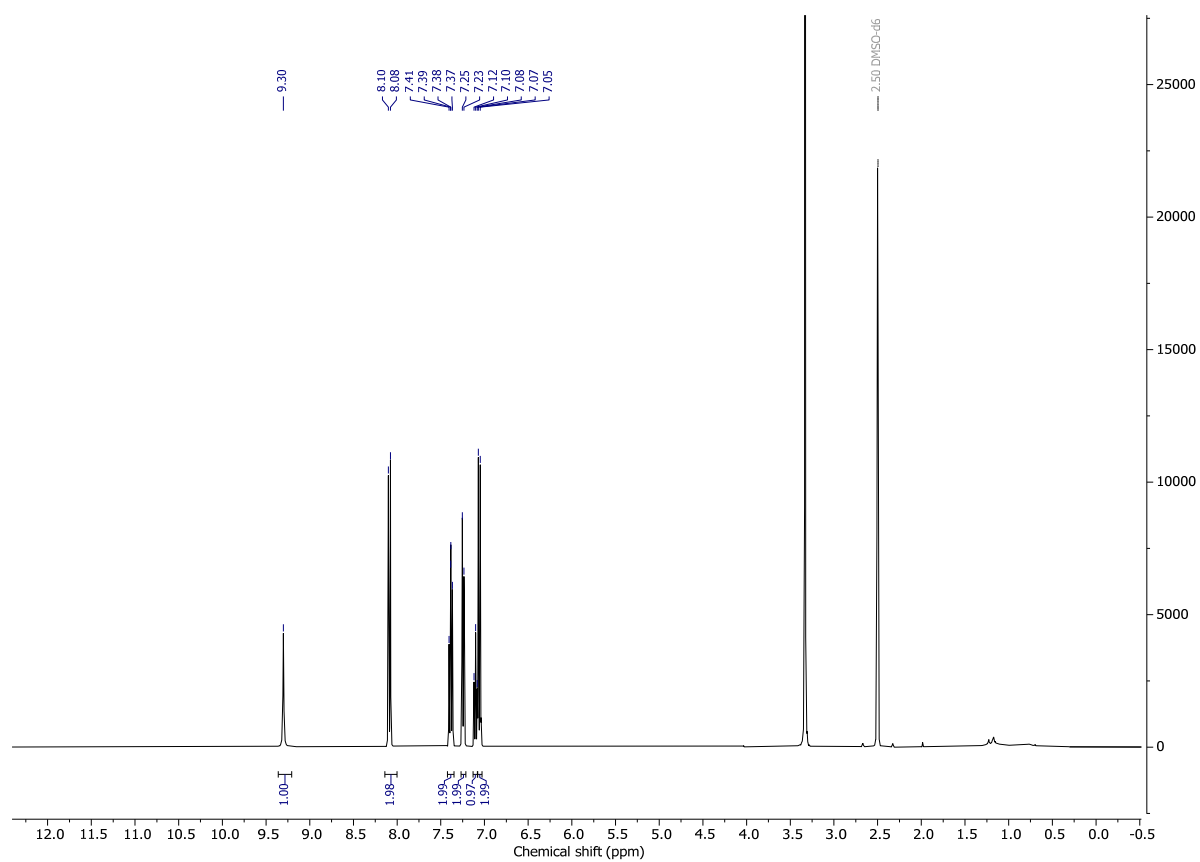


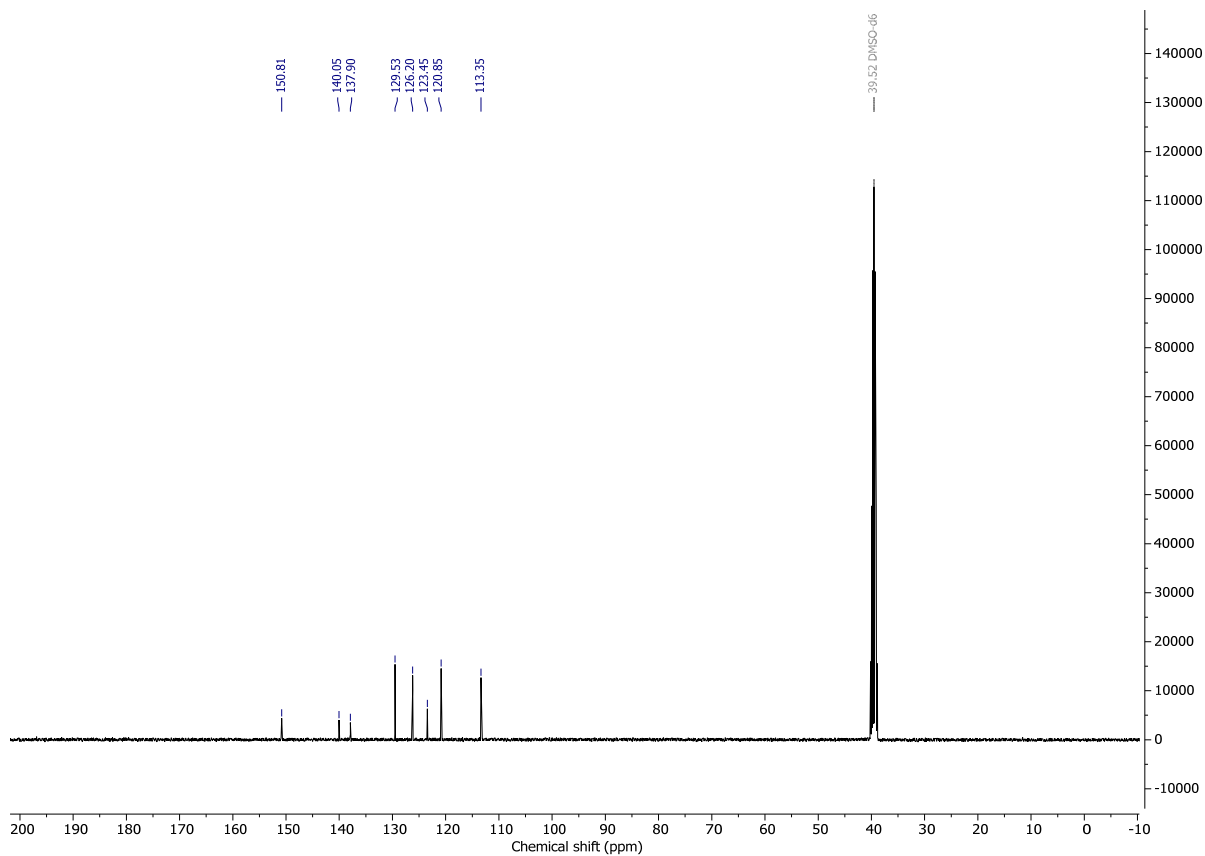
4-Methyl-N-(4-nitrophenyl)aniline (56)



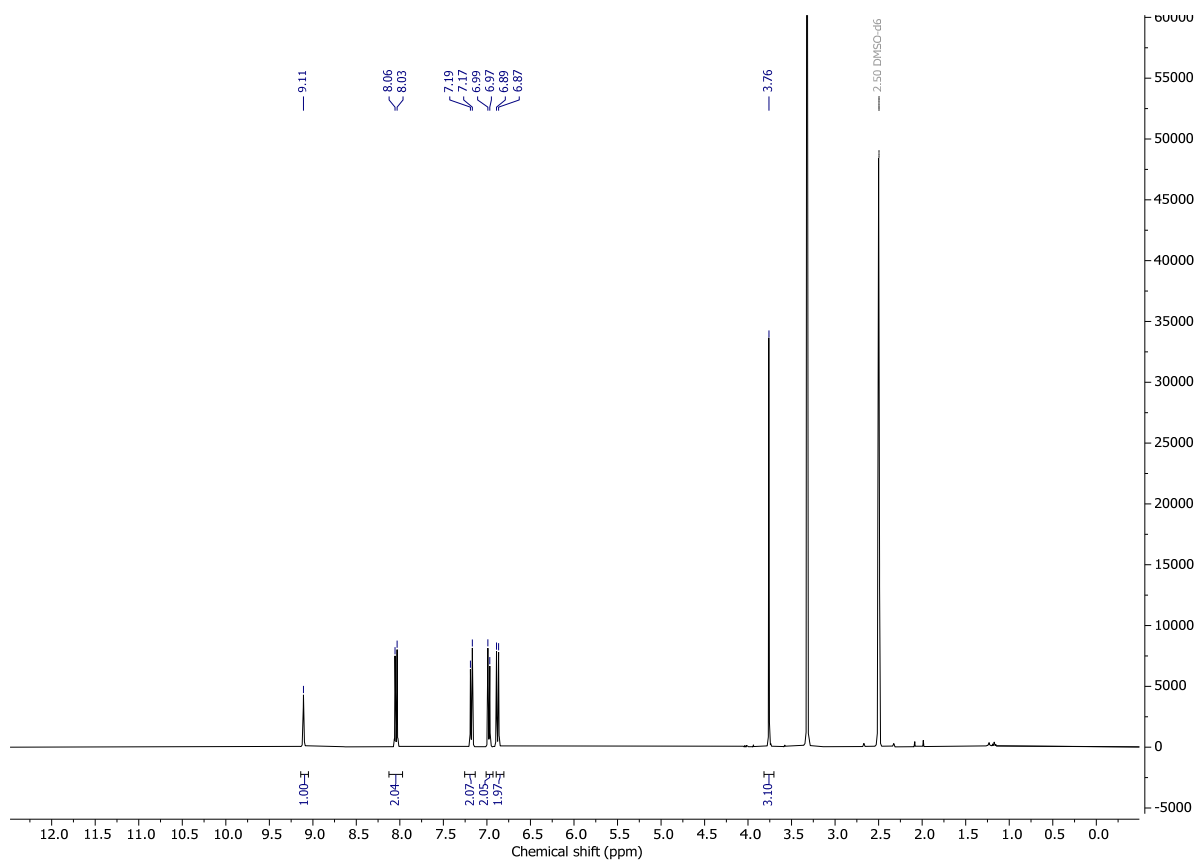


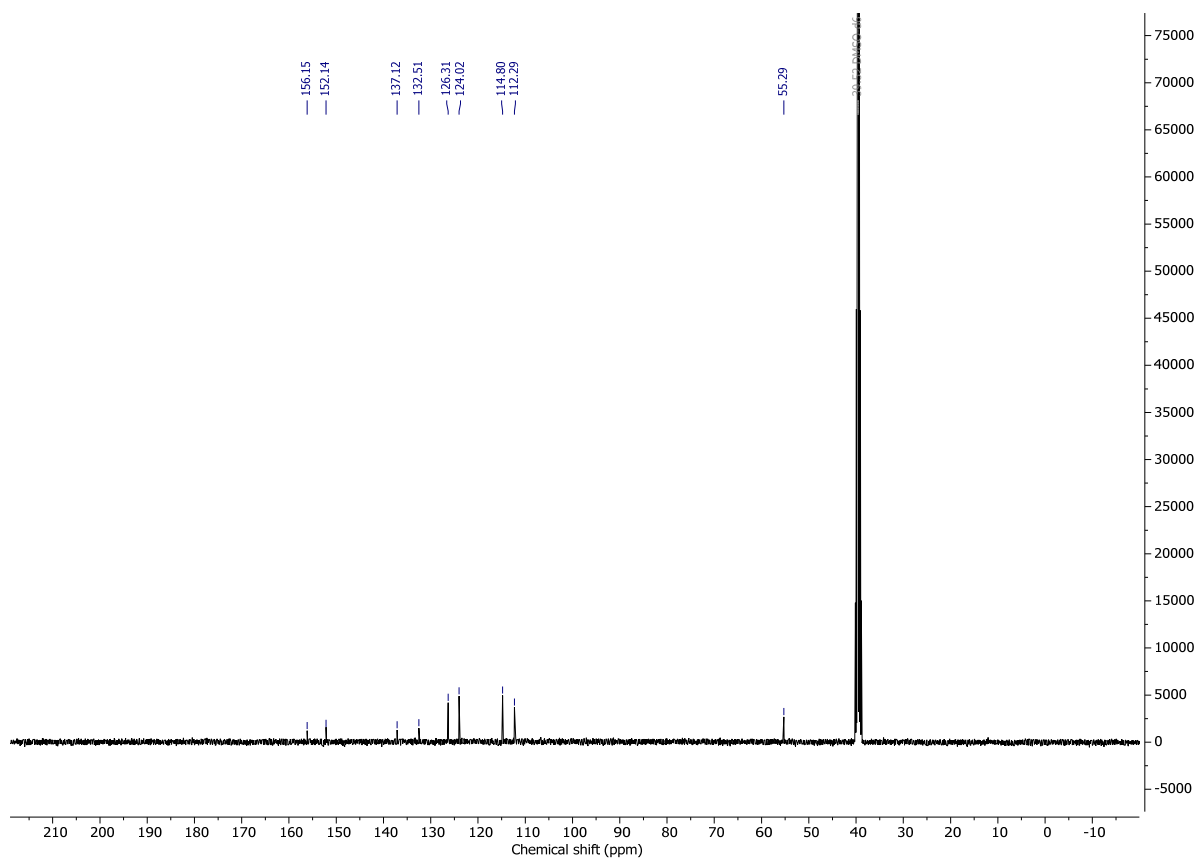
4-Nitro-N-phenylaniline (57)



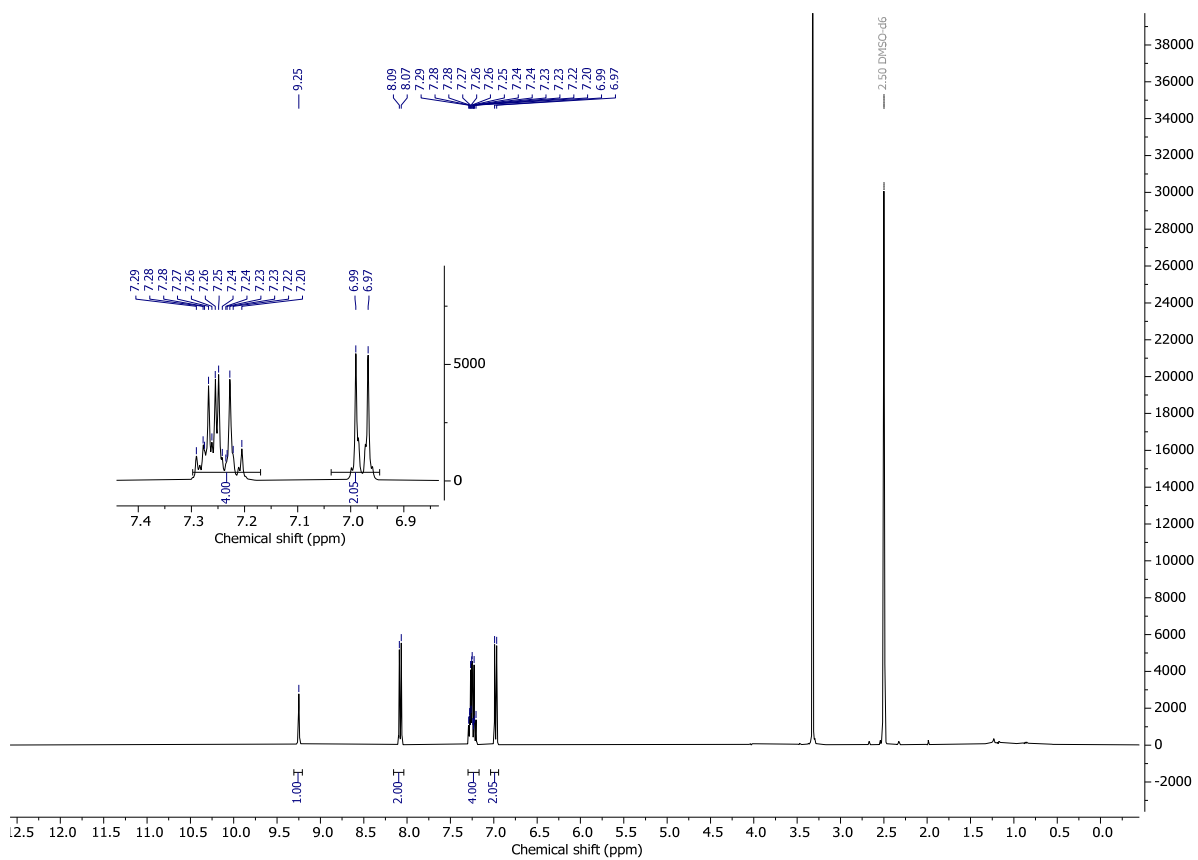


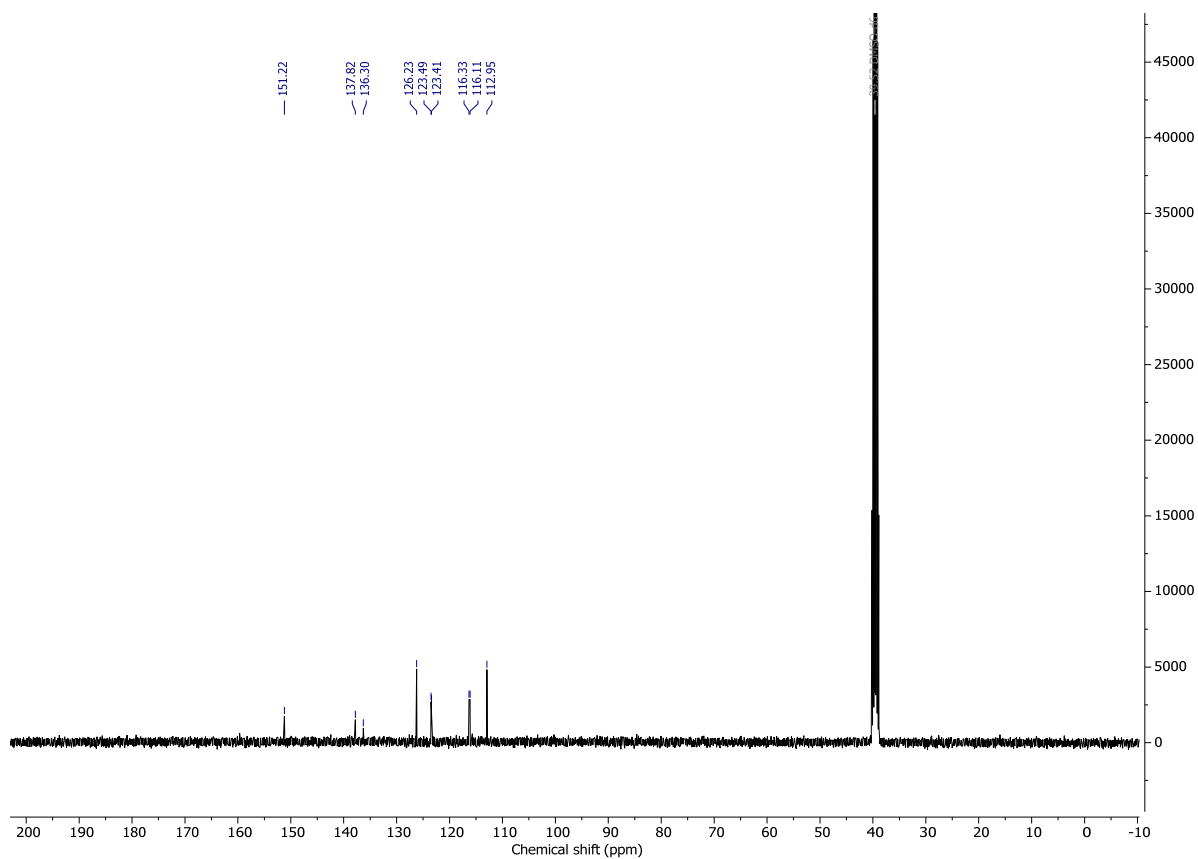
4-Methoxy-N-(4-nitrophenyl)aniline (58)



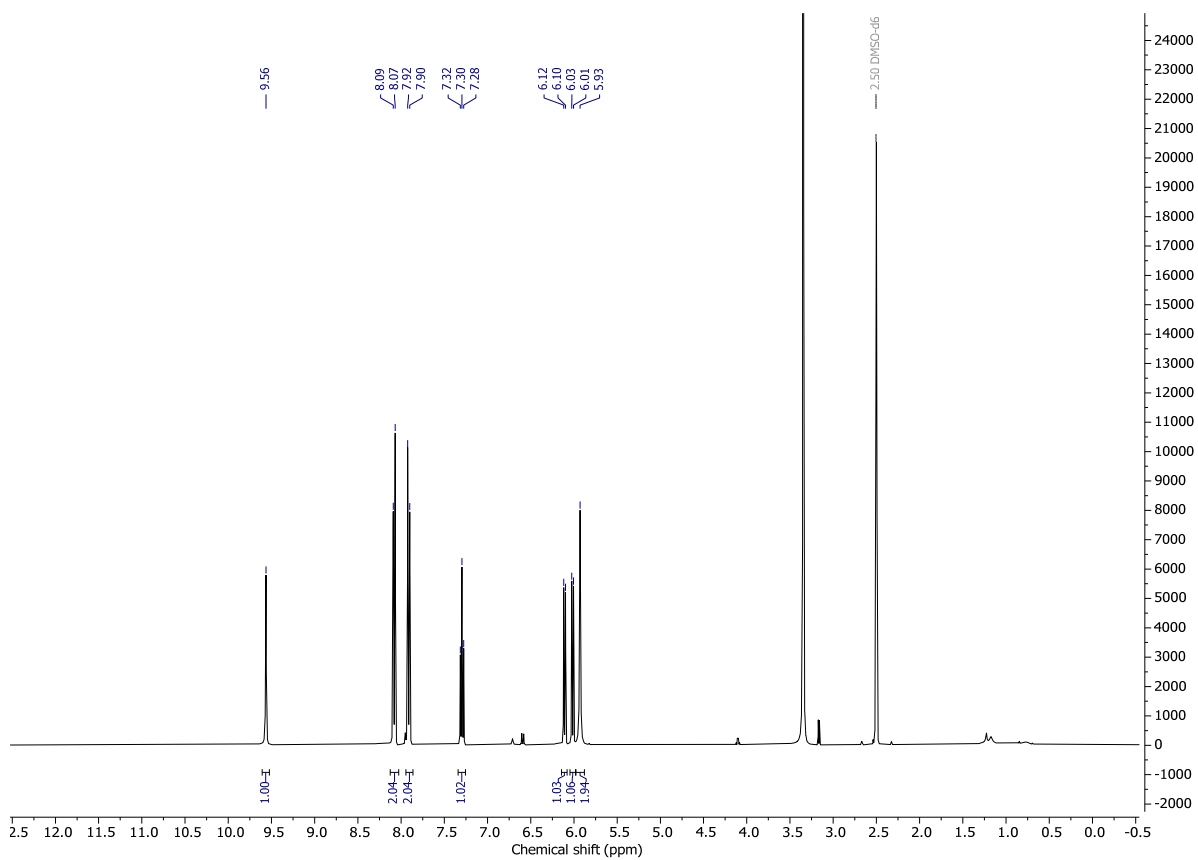


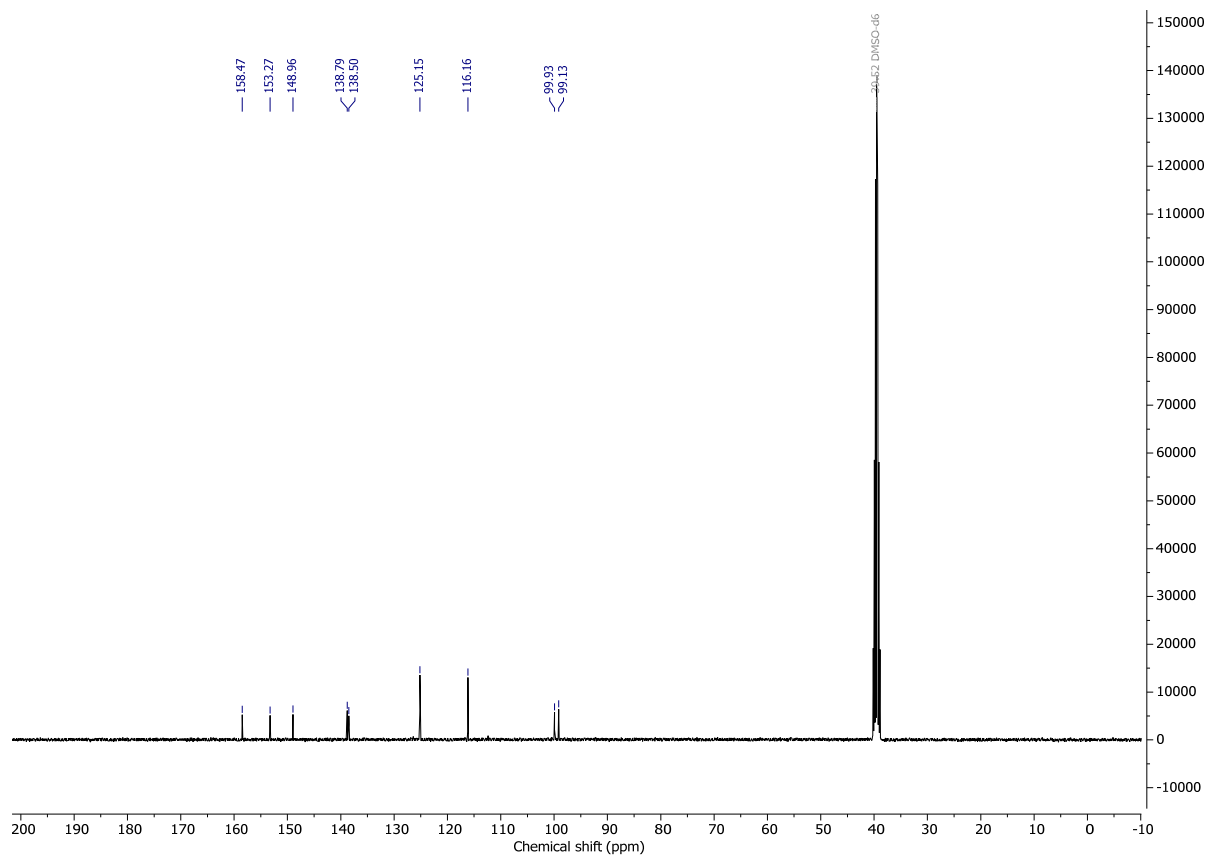
4-Fluoro-N-(4-nitrophenyl)aniline (59)



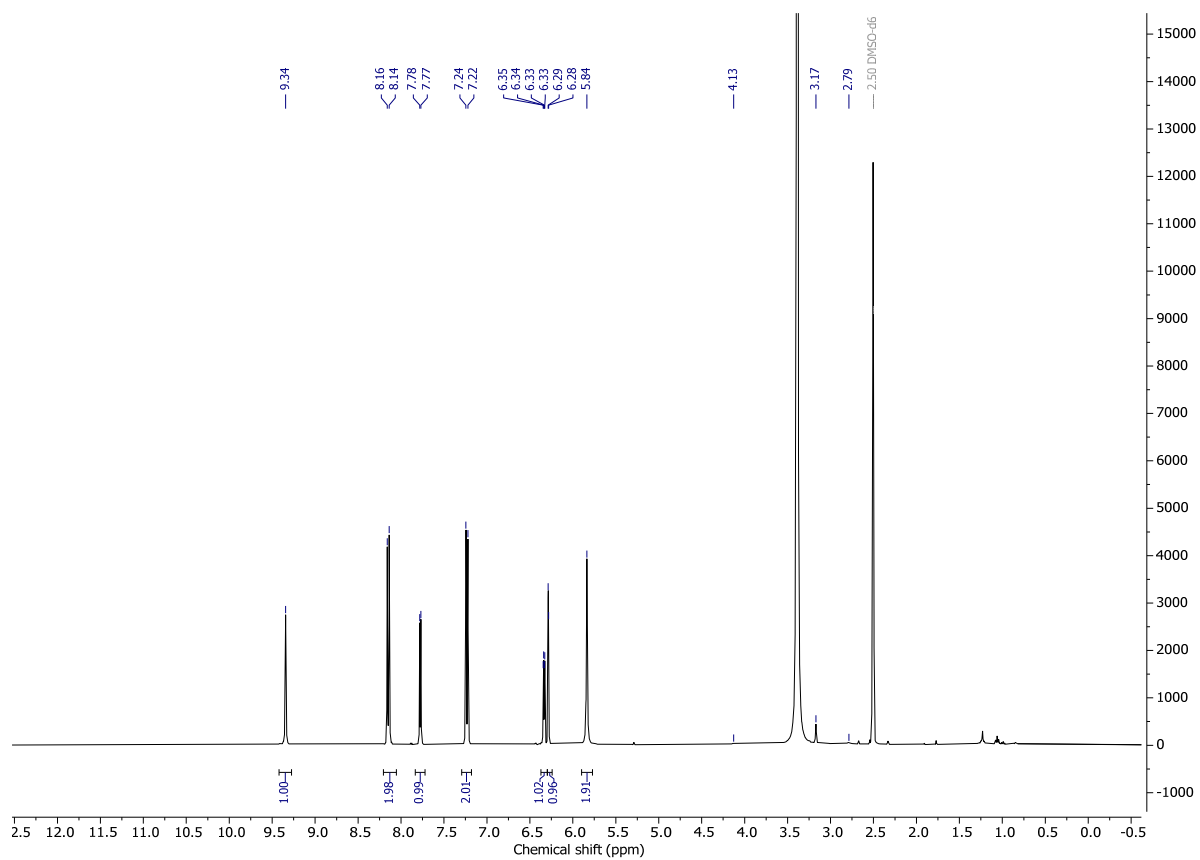


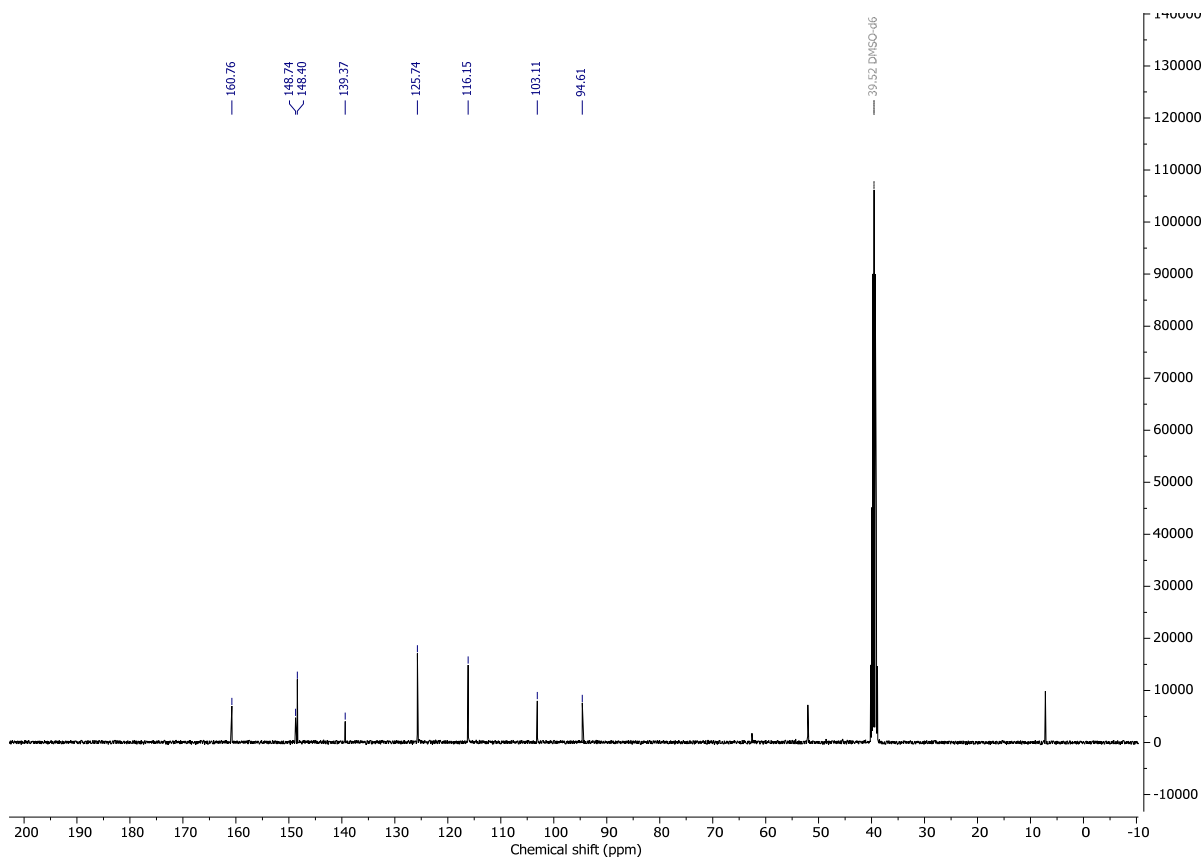
***N*²-(4-Nitrophenyl)pyridine-2,6-diamine (60)**



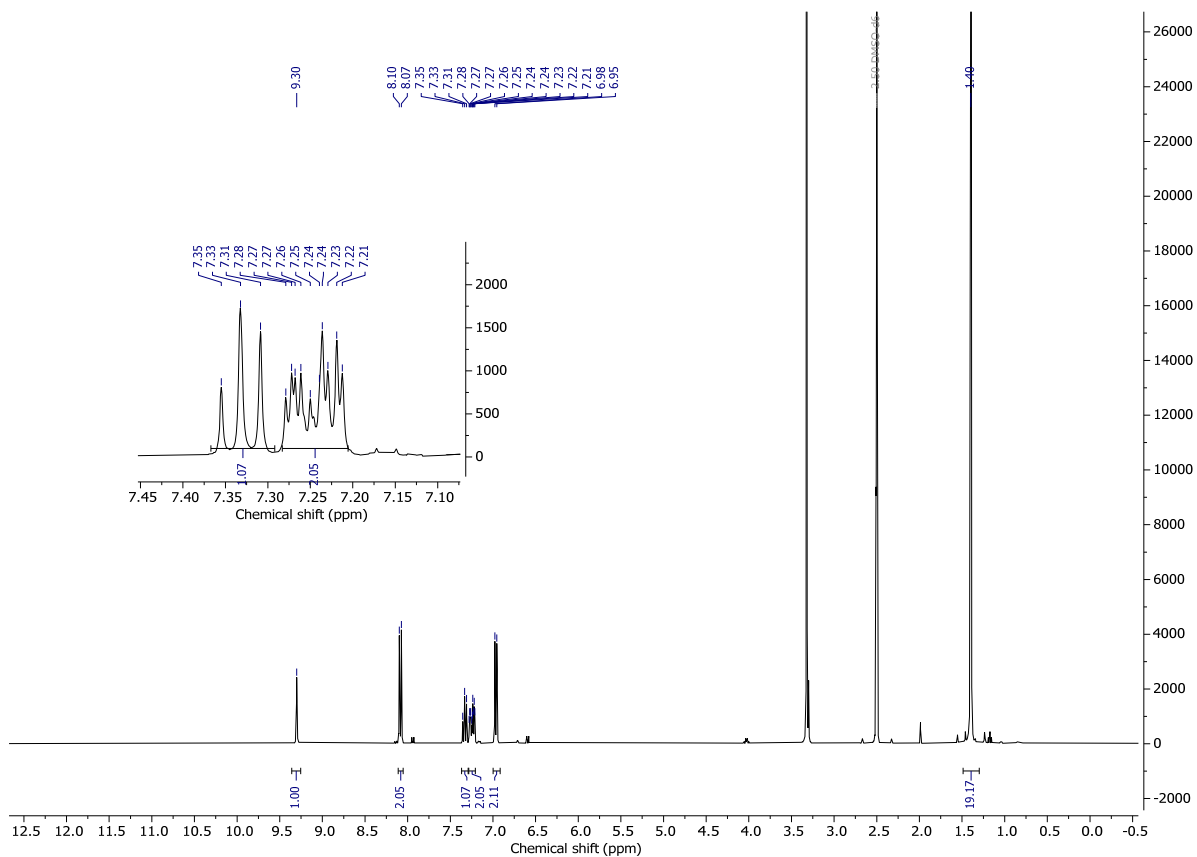


***N*⁴-(4-Nitrophenyl)pyridine-2,4-diamine (61)**

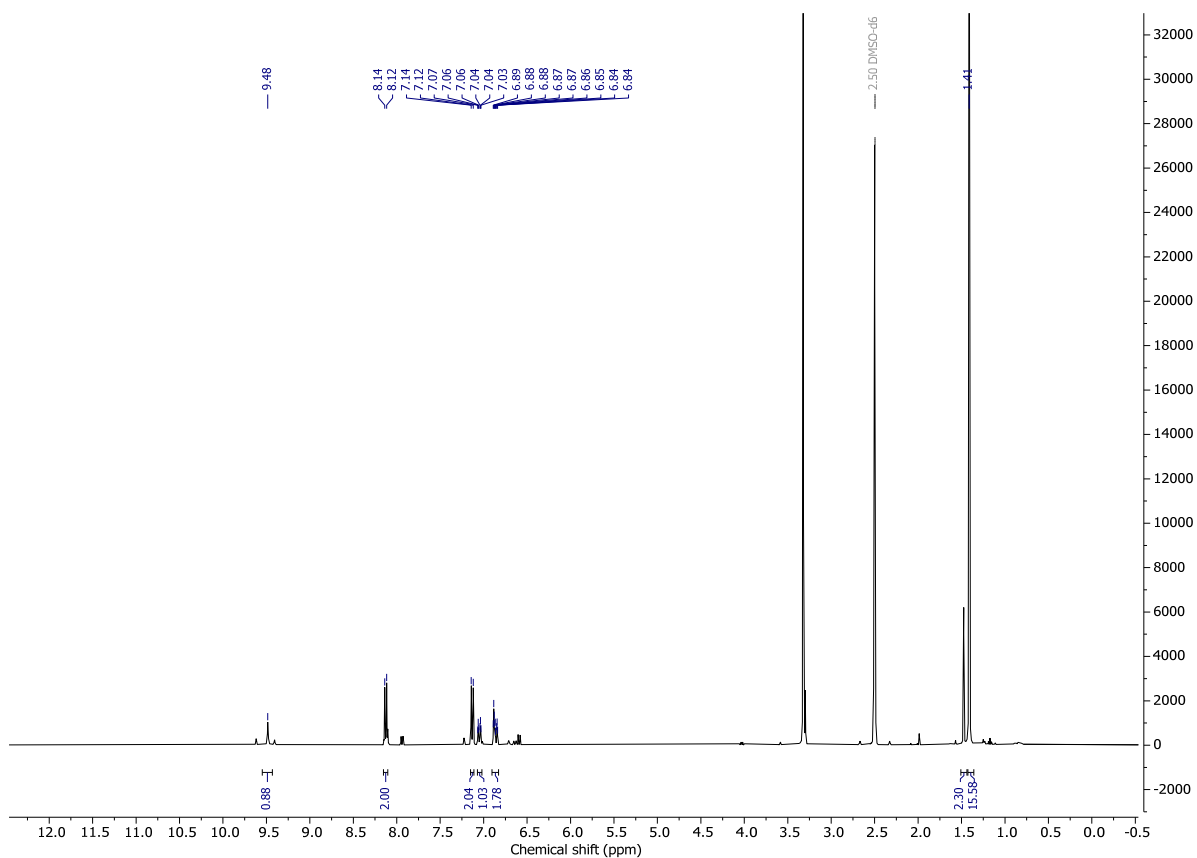




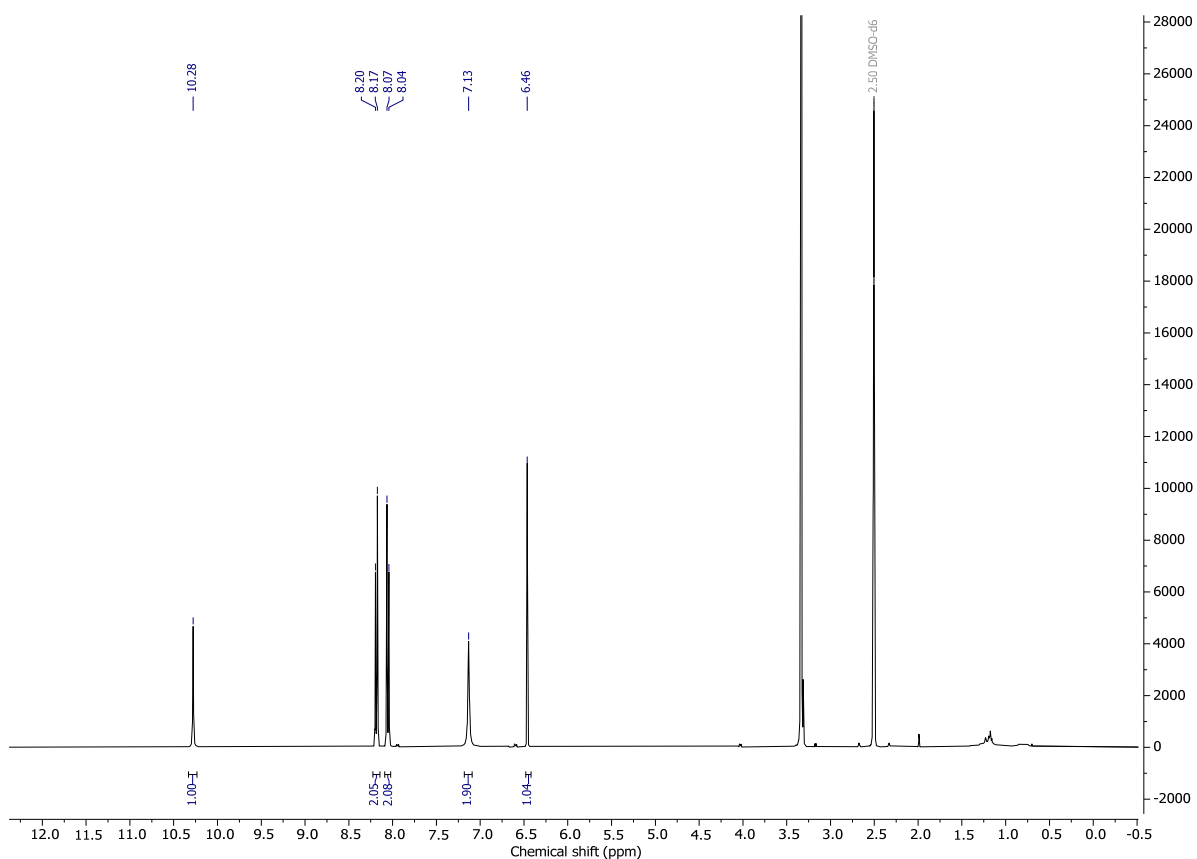
***tert*-Butyl (*tert*-butoxycarbonyl)(2-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate (62)**

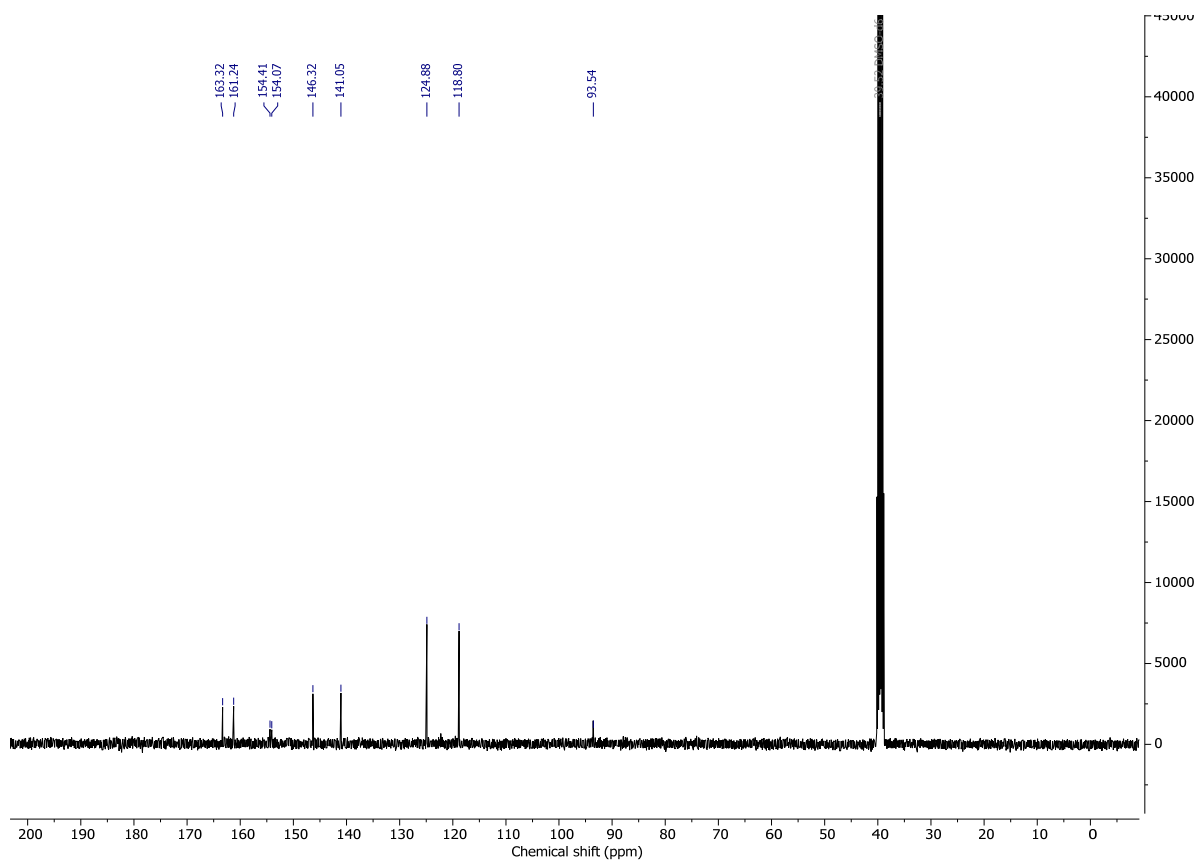


***tert*-Butyl (*tert*-butoxycarbonyl)(3-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate & *tert*-butyl (3-fluoro-5-((4-nitrophenyl)amino)phenyl)carbamate (63)**

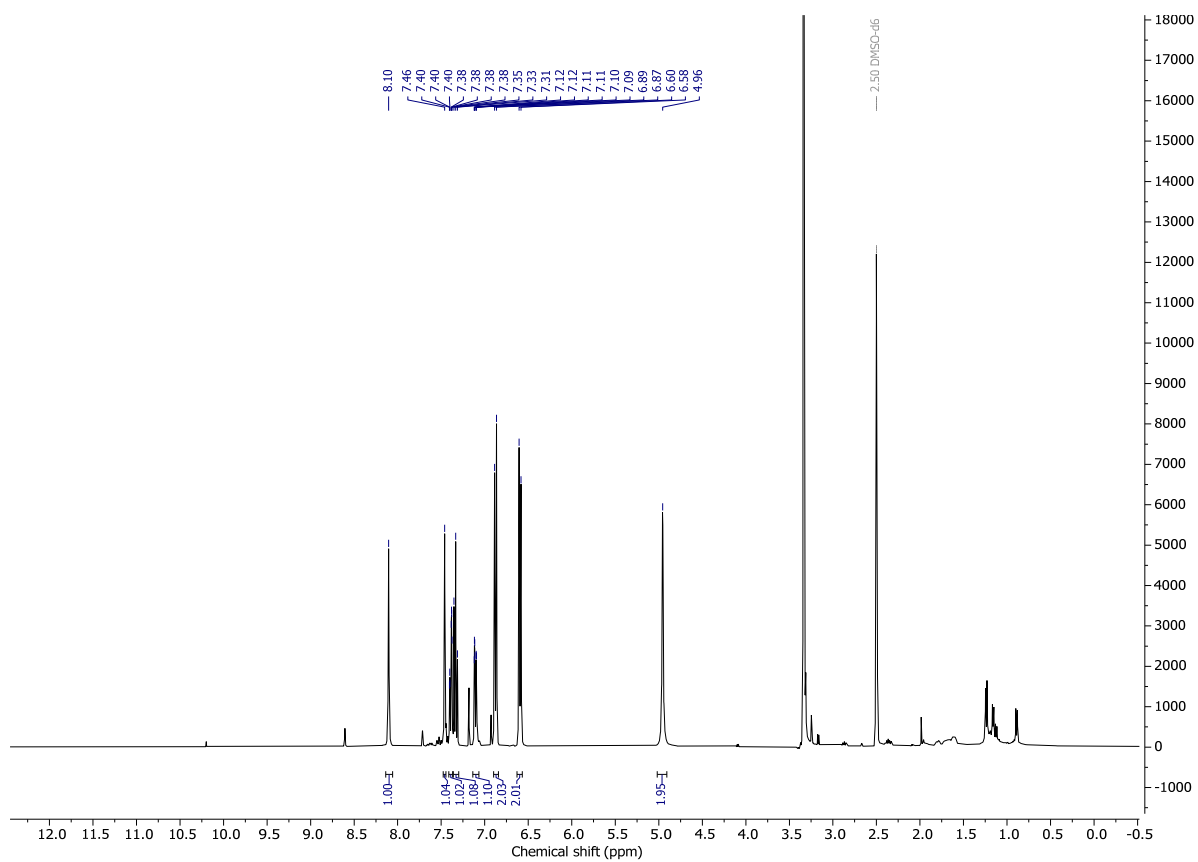


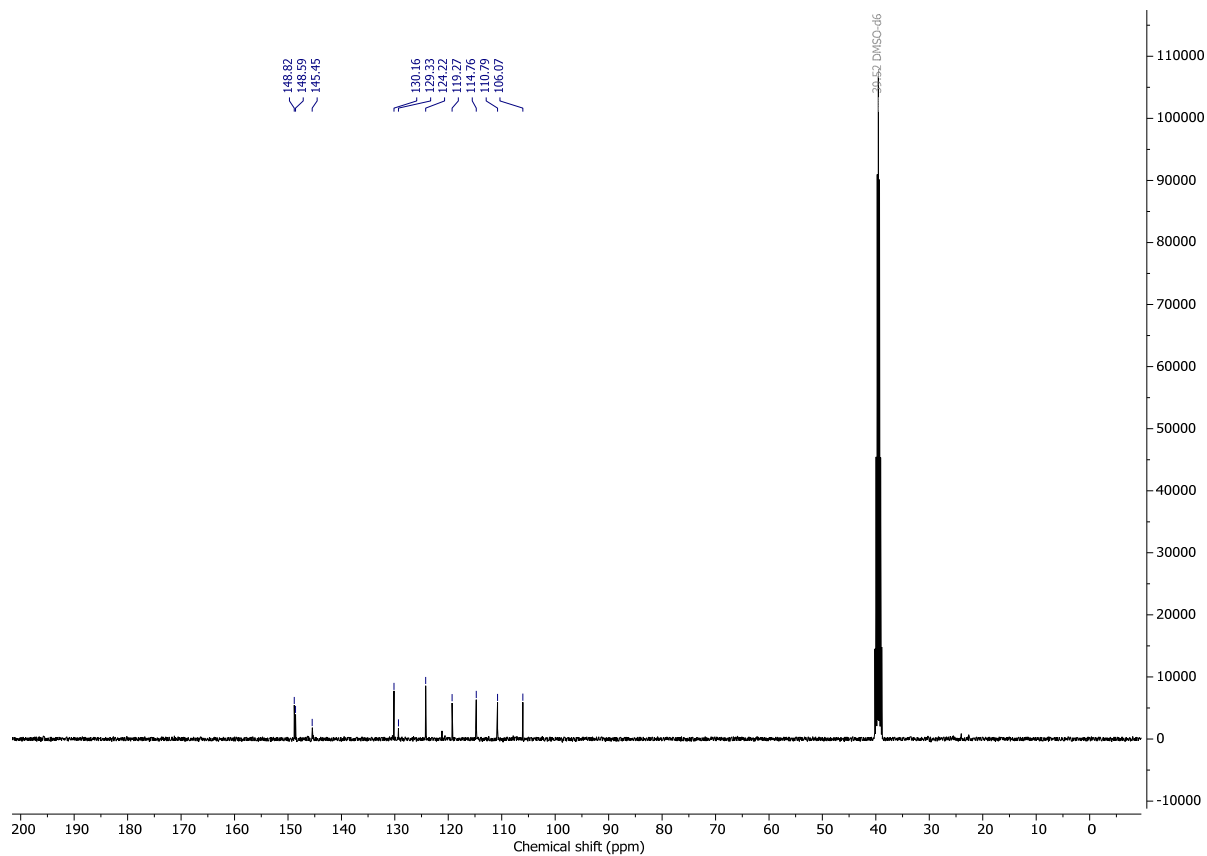
***N*⁴-(4-Nitrophenyl)-6-(trifluoromethyl)pyrimidine-2,4-diamine (64)**



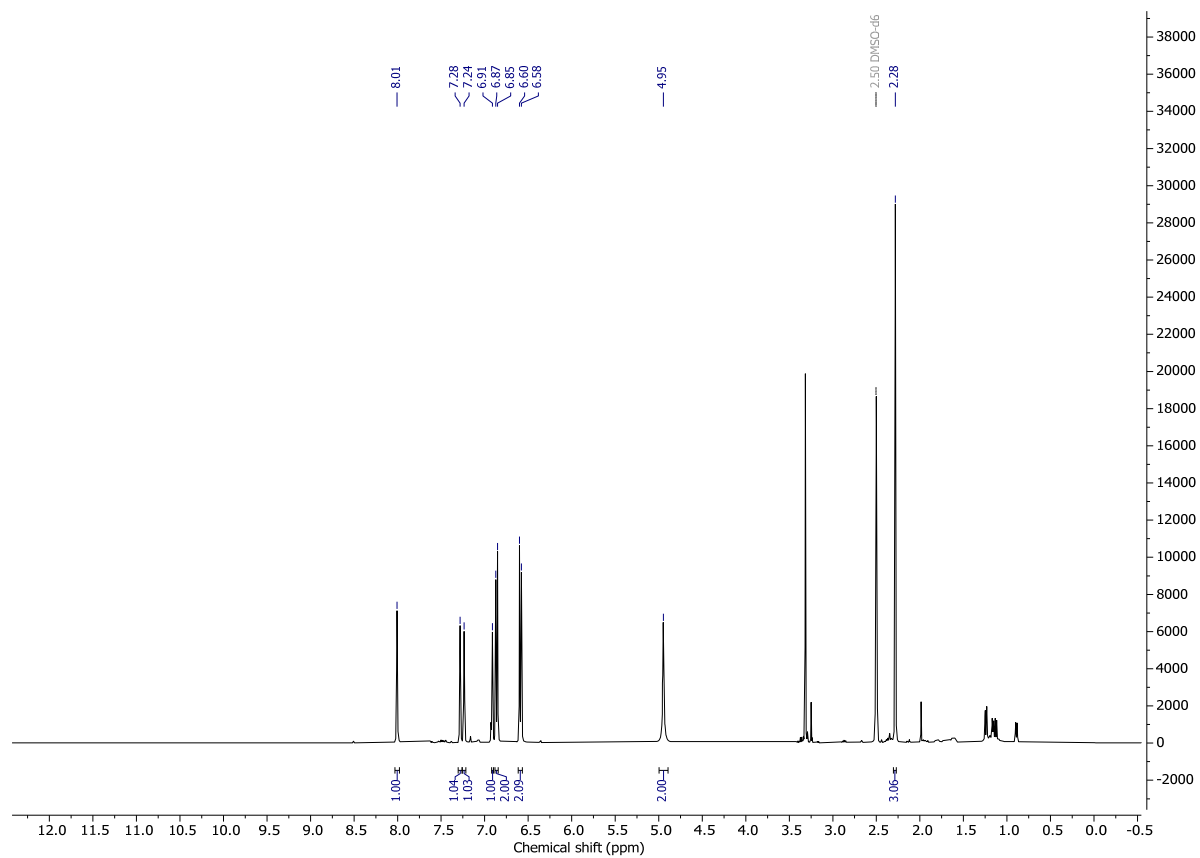


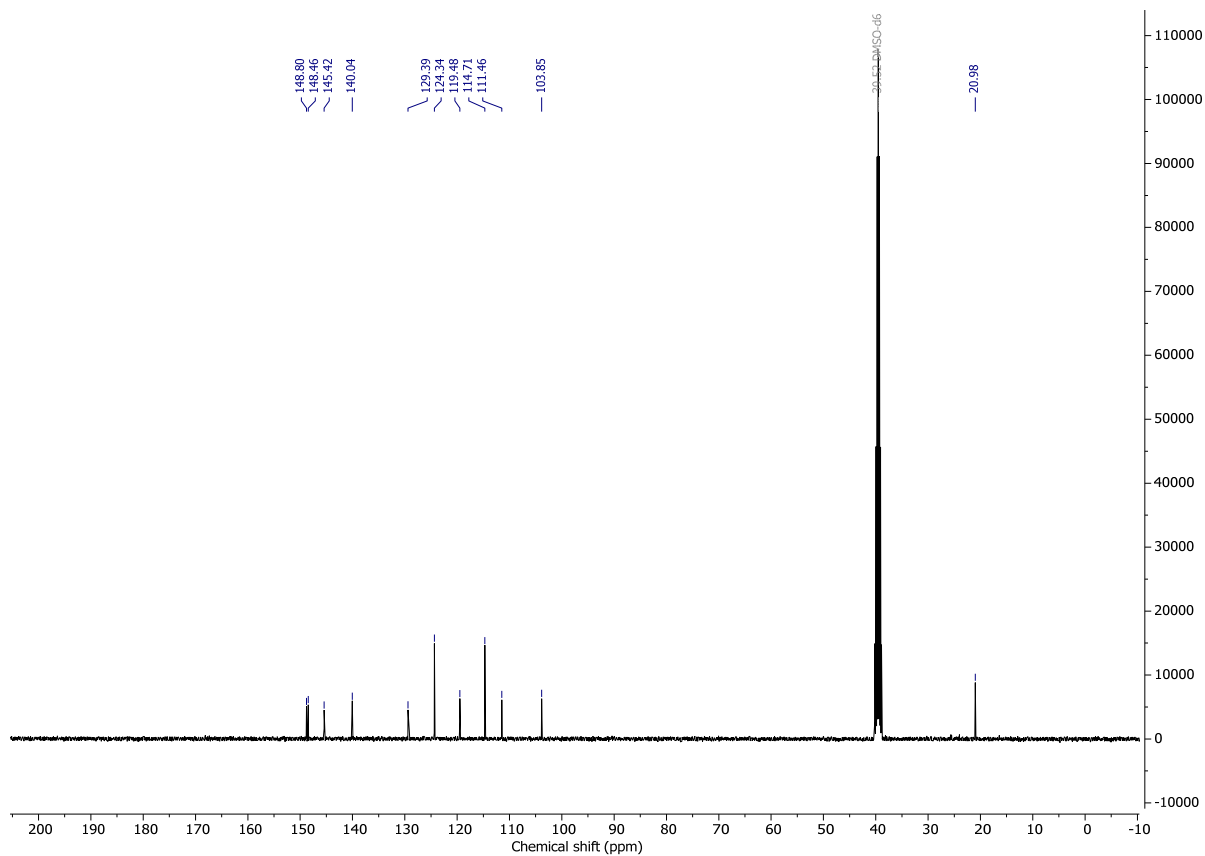
***N'*-(3-Nitrophenyl)benzene-1,4-diamine (65)**



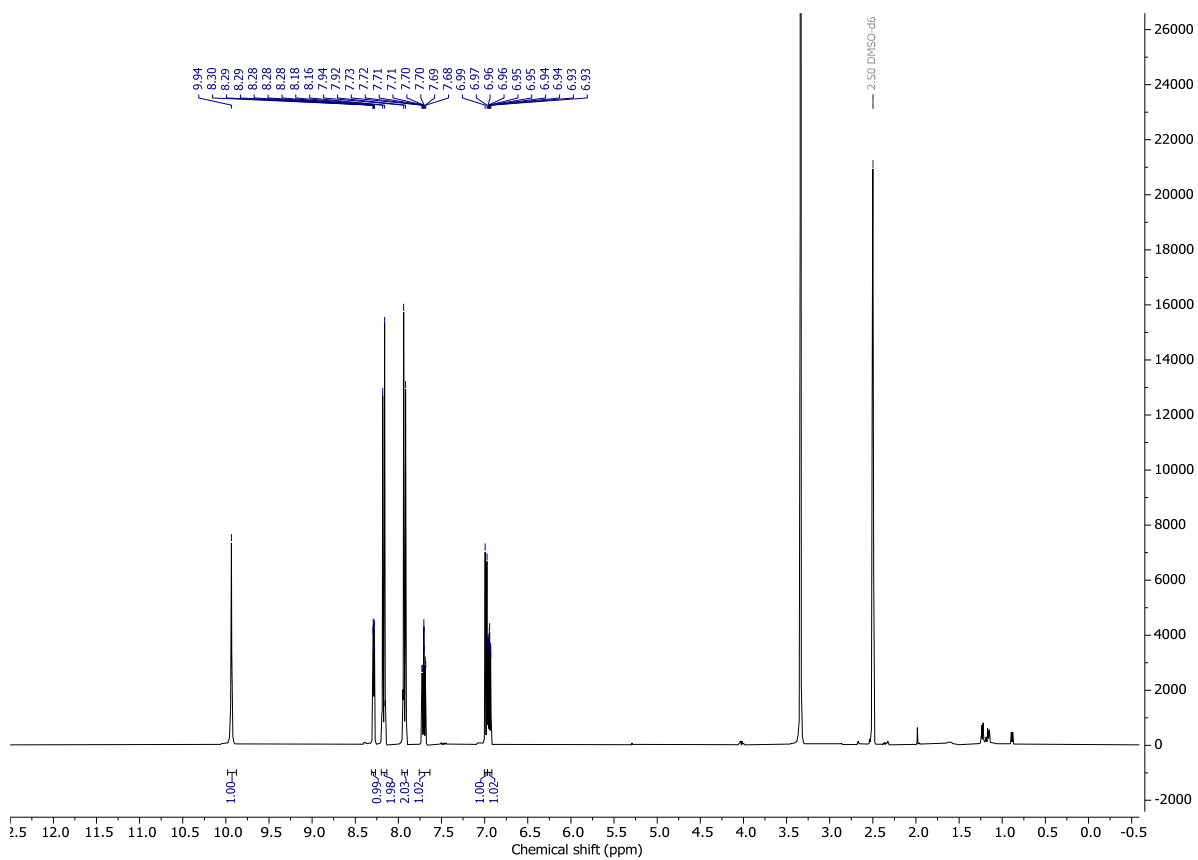


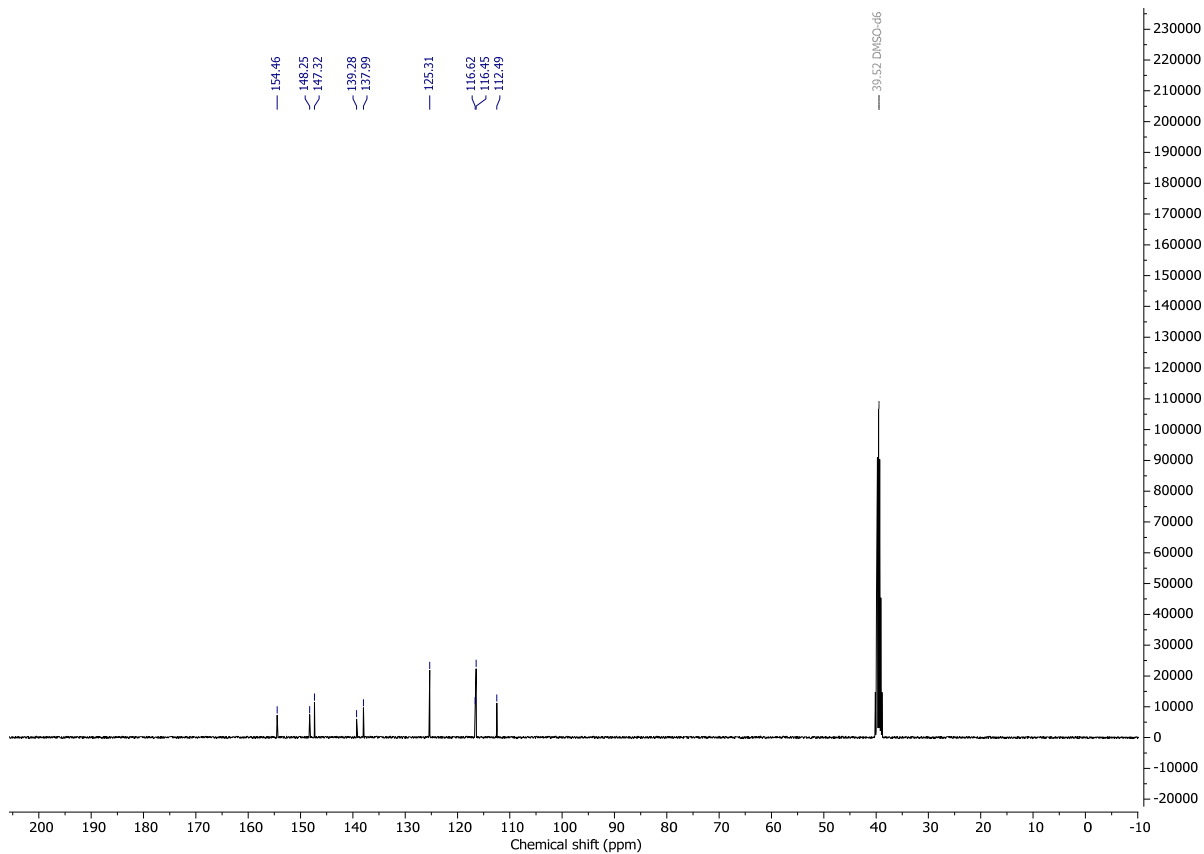
***N*¹-(3-Methyl-5-nitrophenyl)benzene-1,4-diamine (66)**



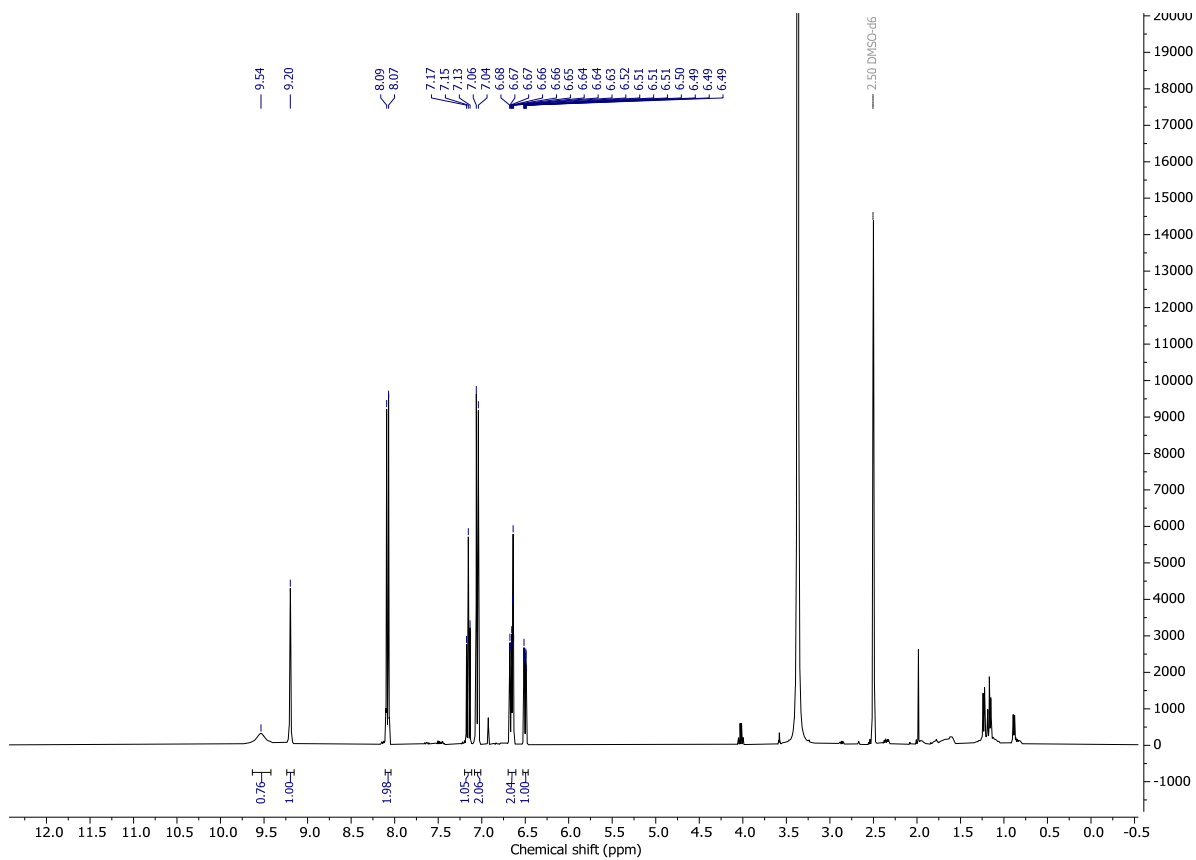


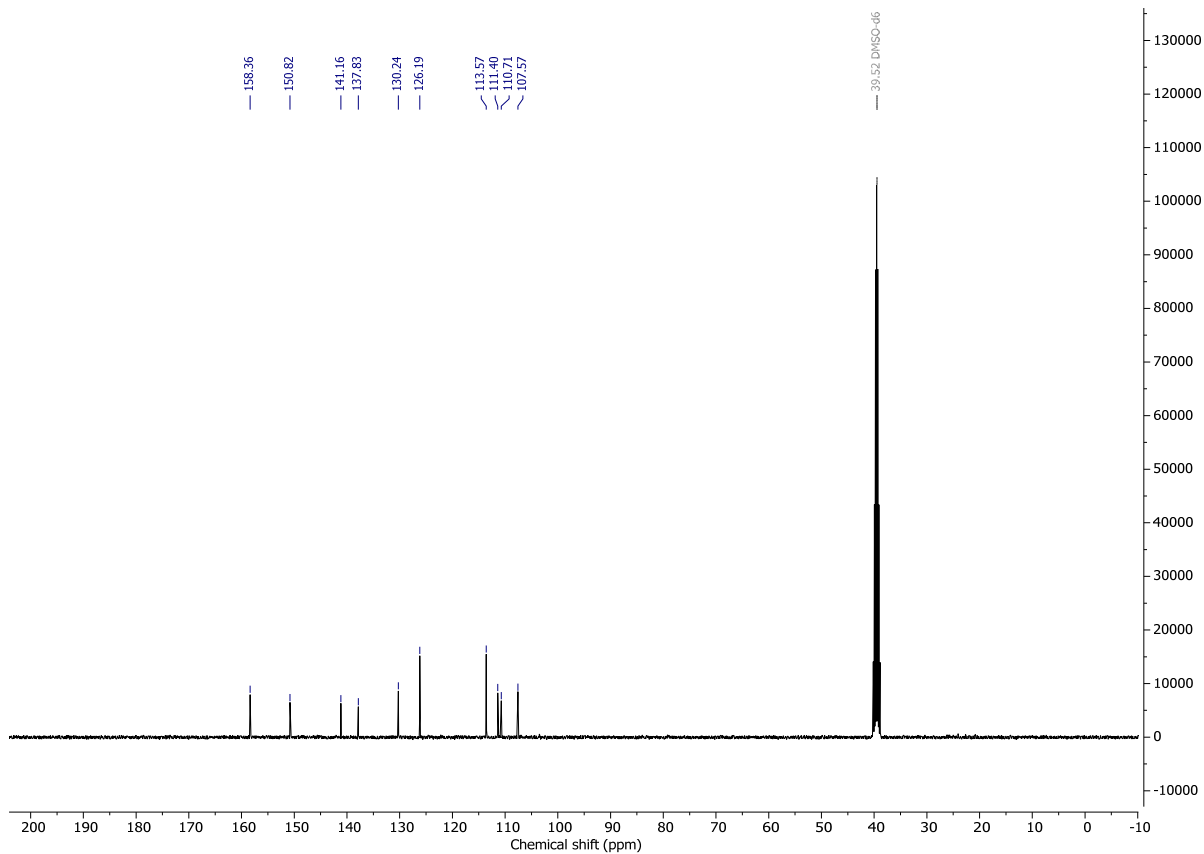
N-(4-Nitrophenyl)pyridin-2-amine (67)



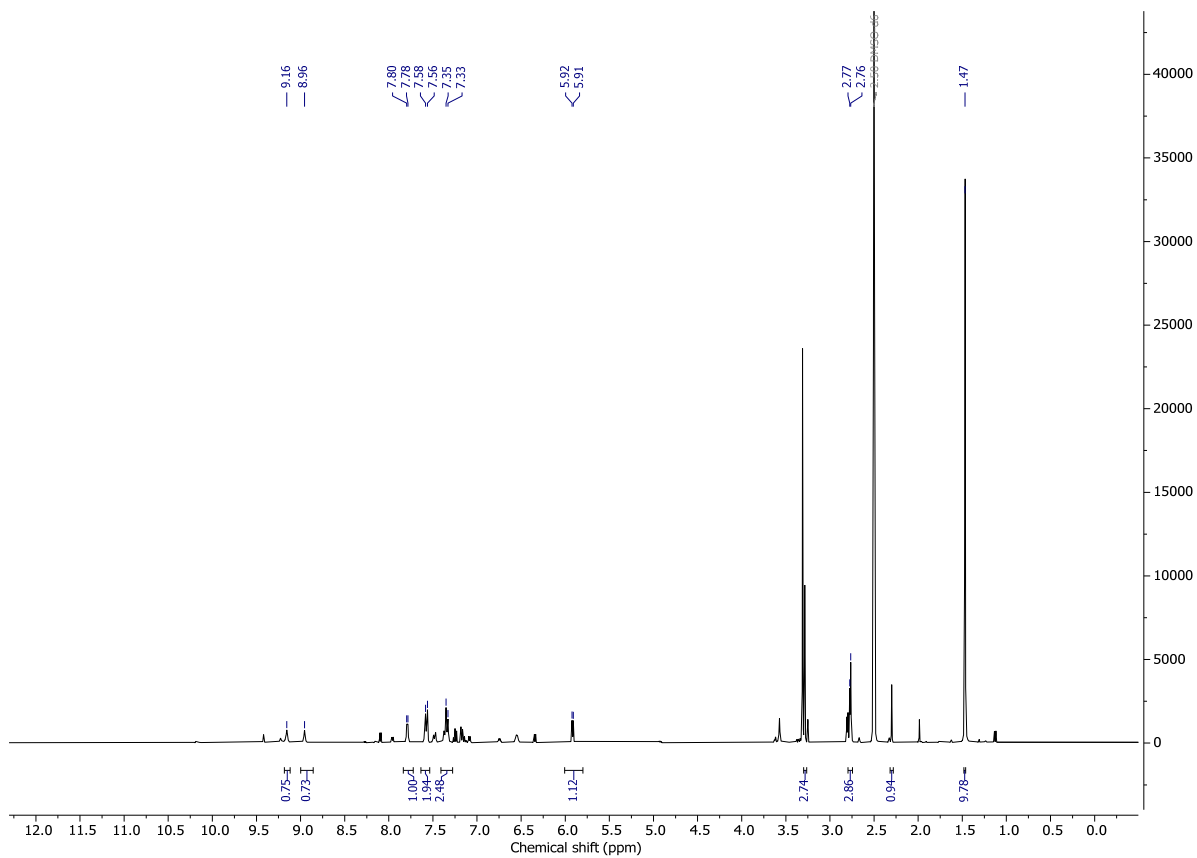


3-((4-Nitrophenyl)amino)phenol (68)

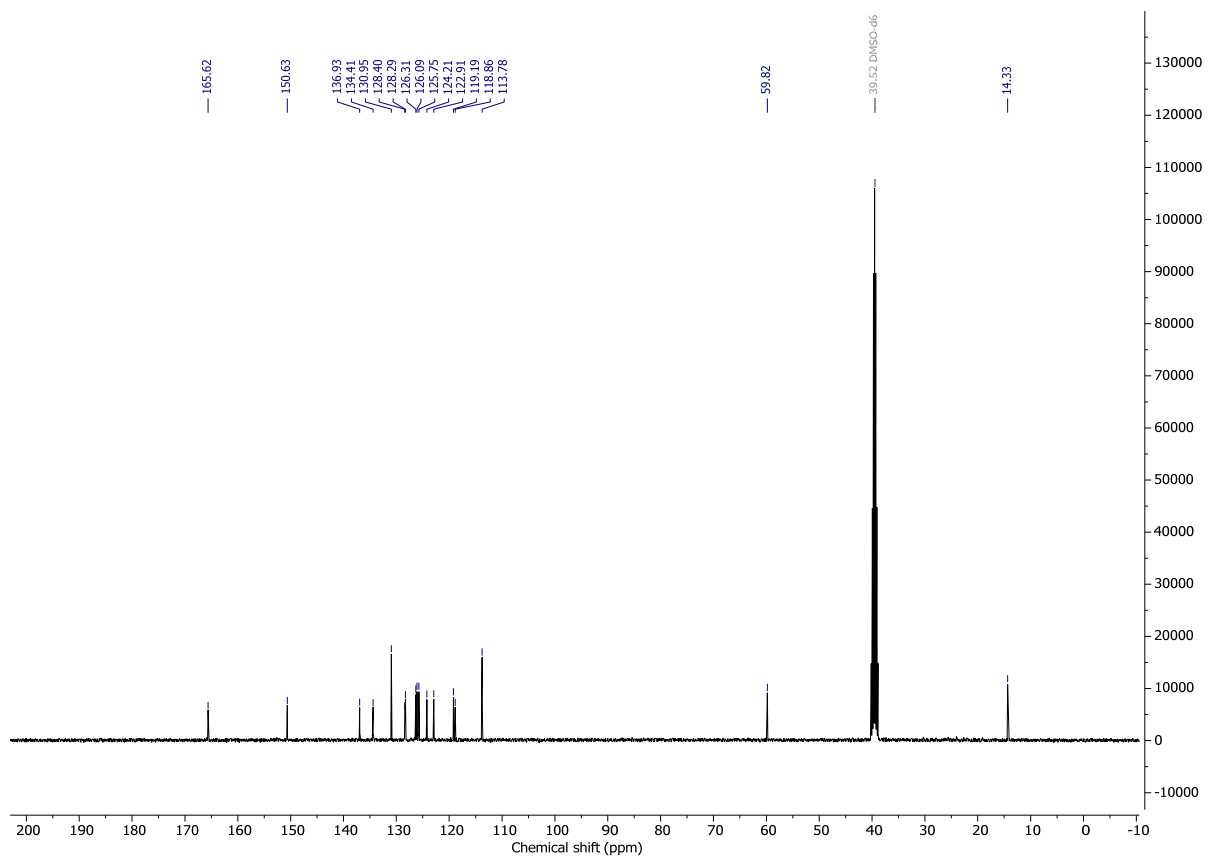
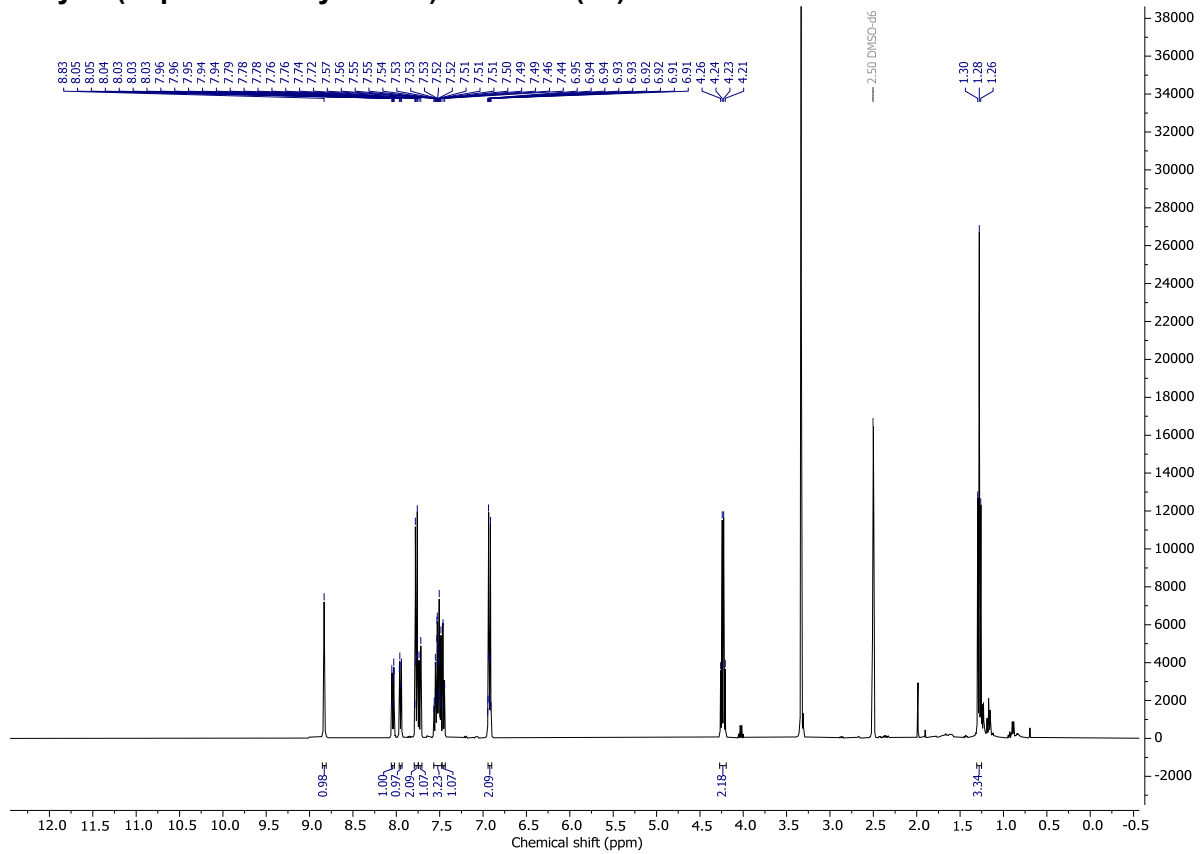




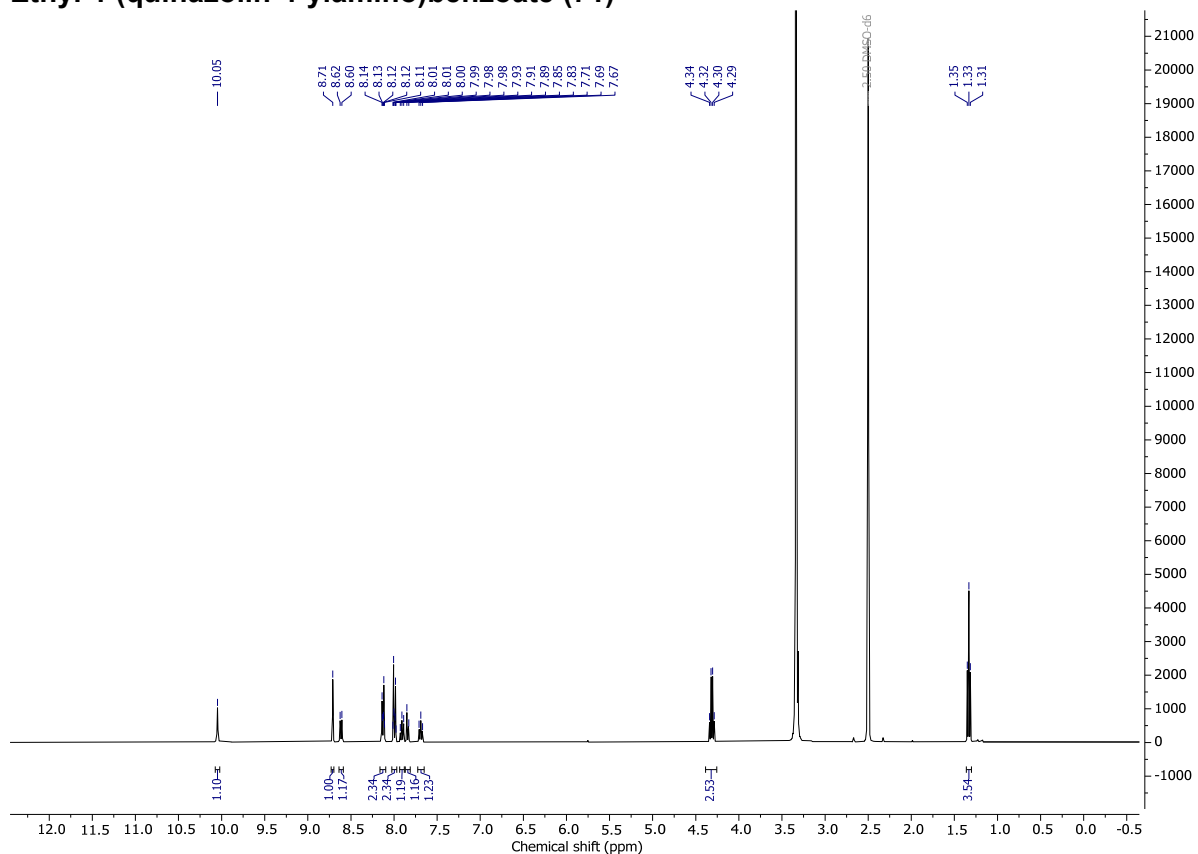
***tert*-Butyl (4-((2-(ethylamino)pyrimidin-4-yl)amino)phenyl)carbamate (69)**



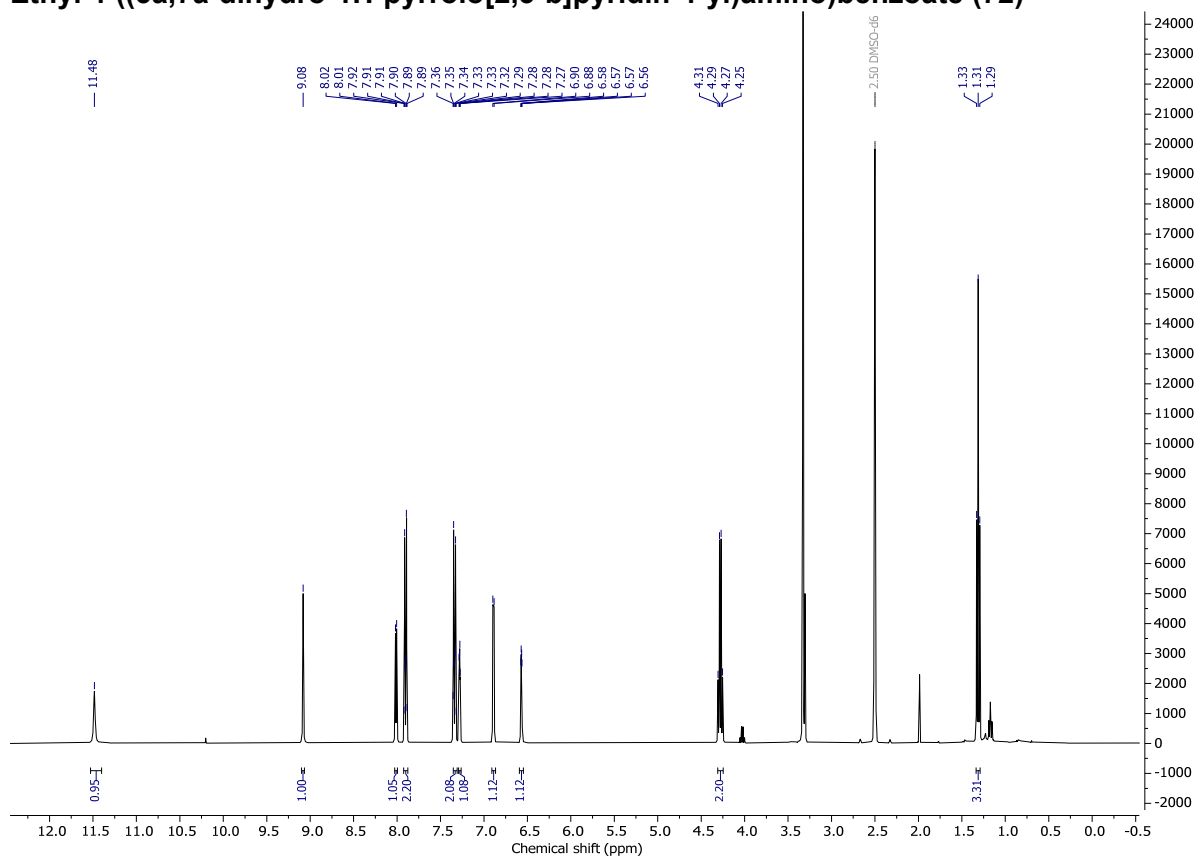
Ethyl 4-(naphthalen-1-ylamino)benzoate (70)

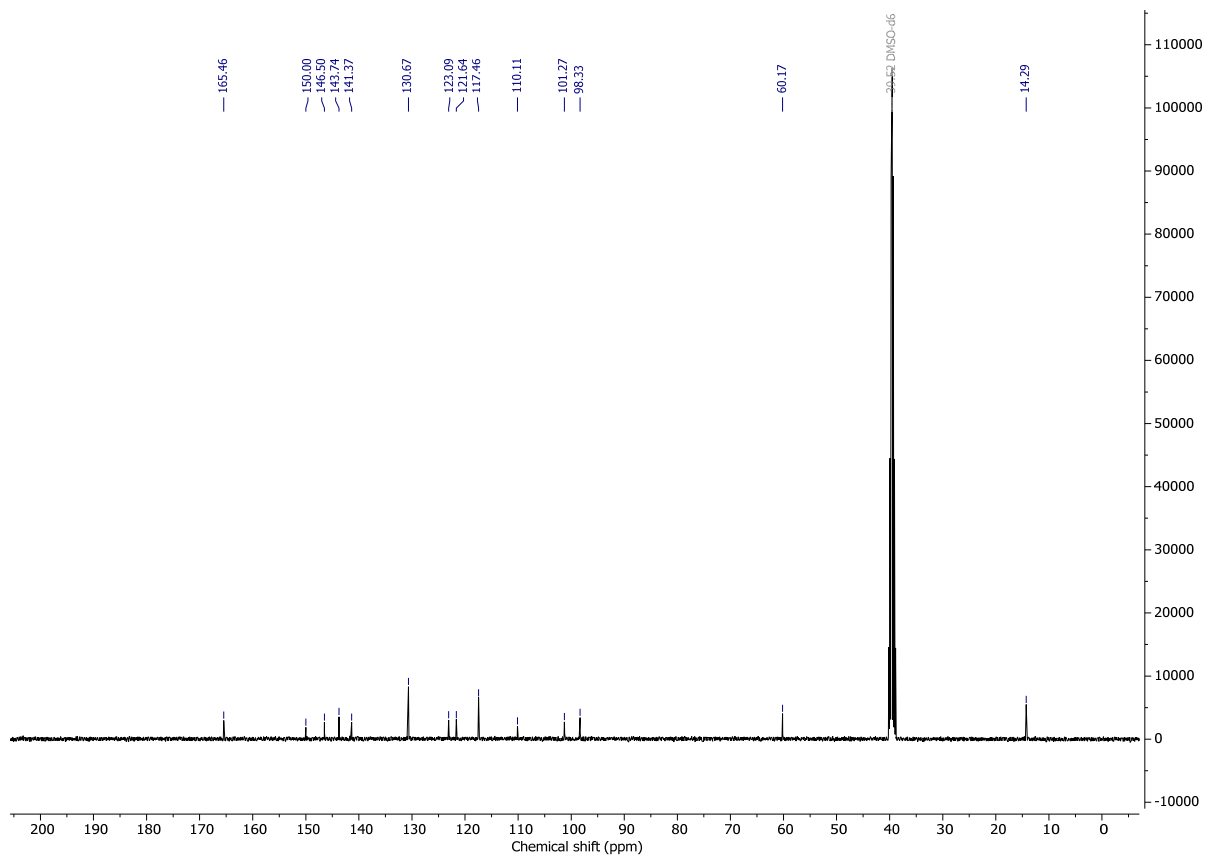


Ethyl 4-(quinazolin-4-ylamino)benzoate (71)

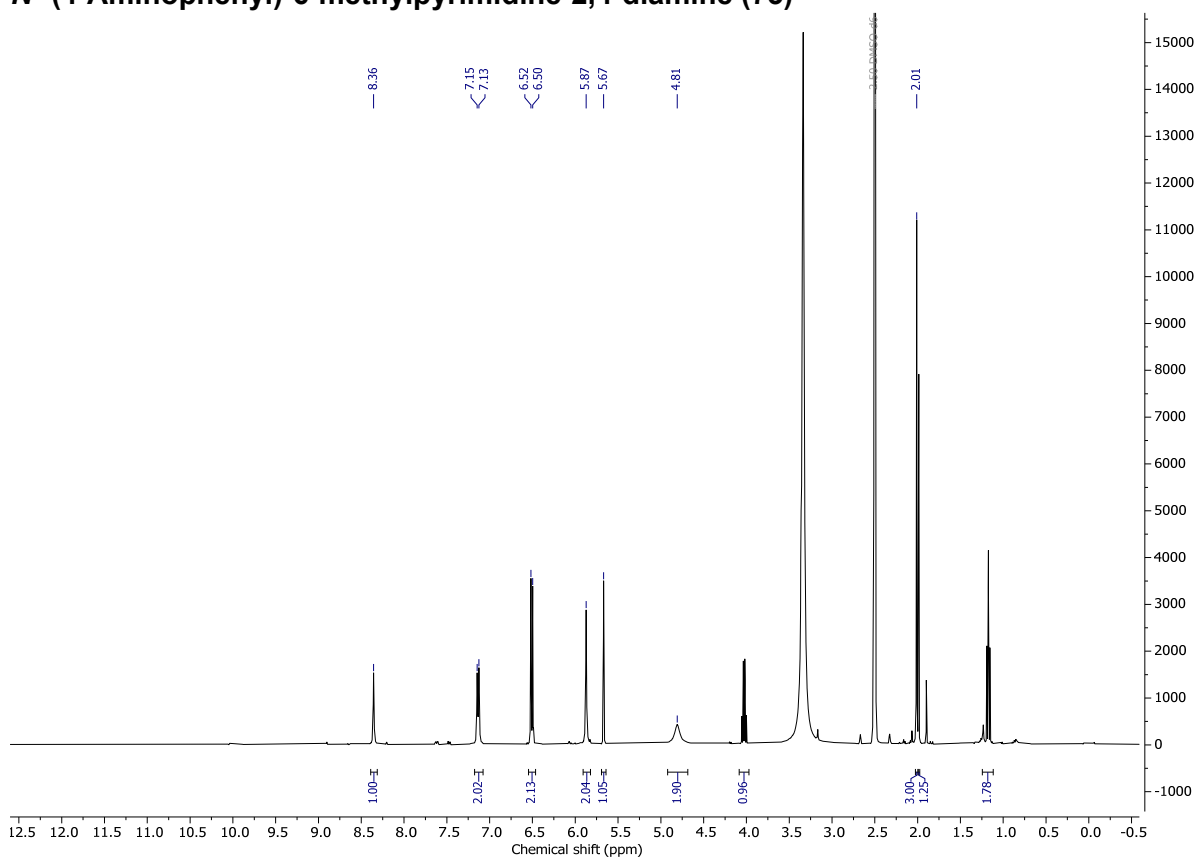


Ethyl 4-((3a,7a-dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzoate (72)

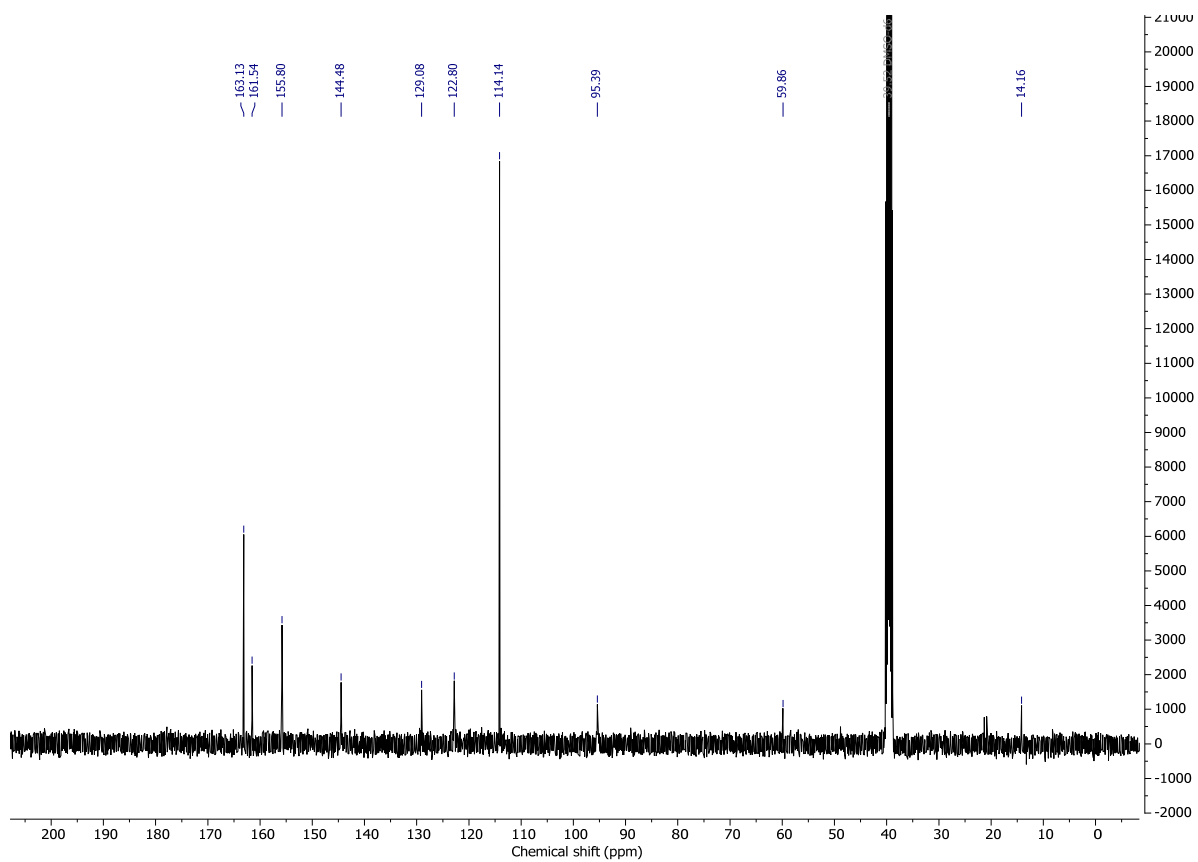
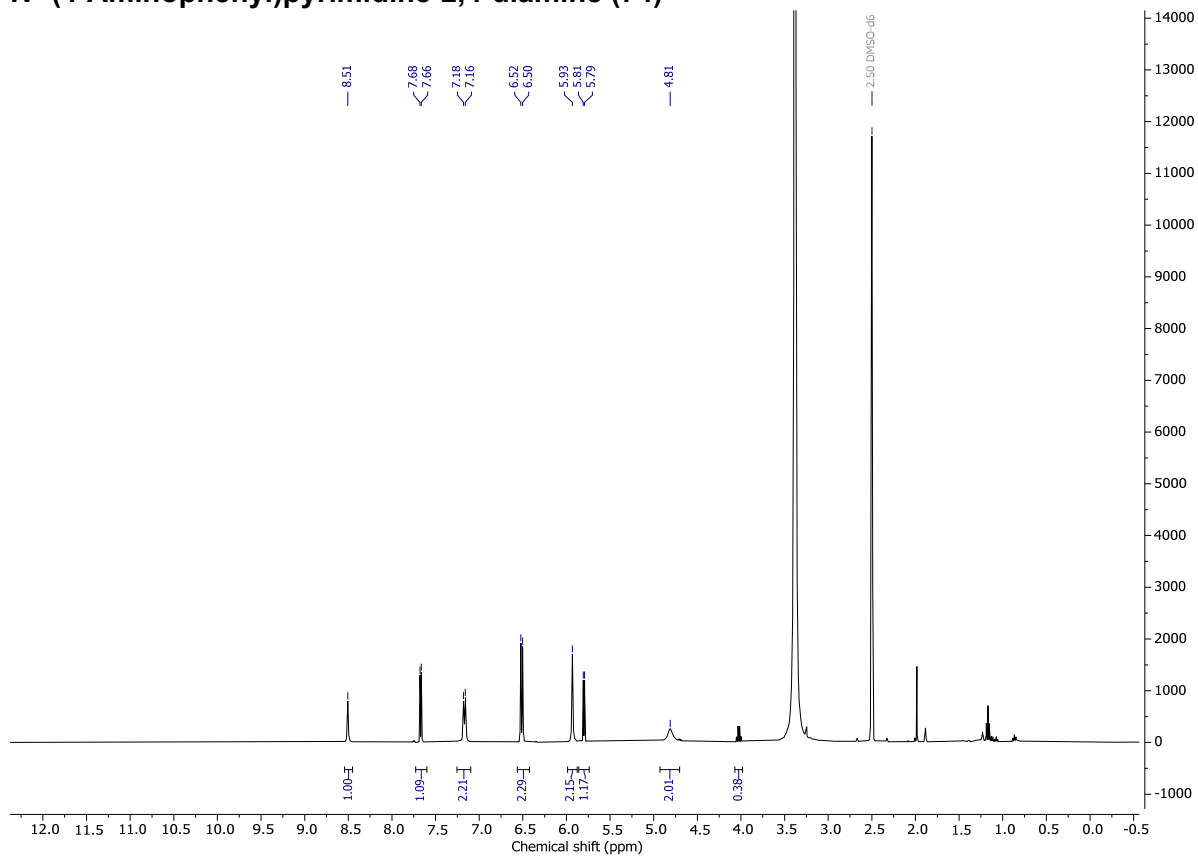




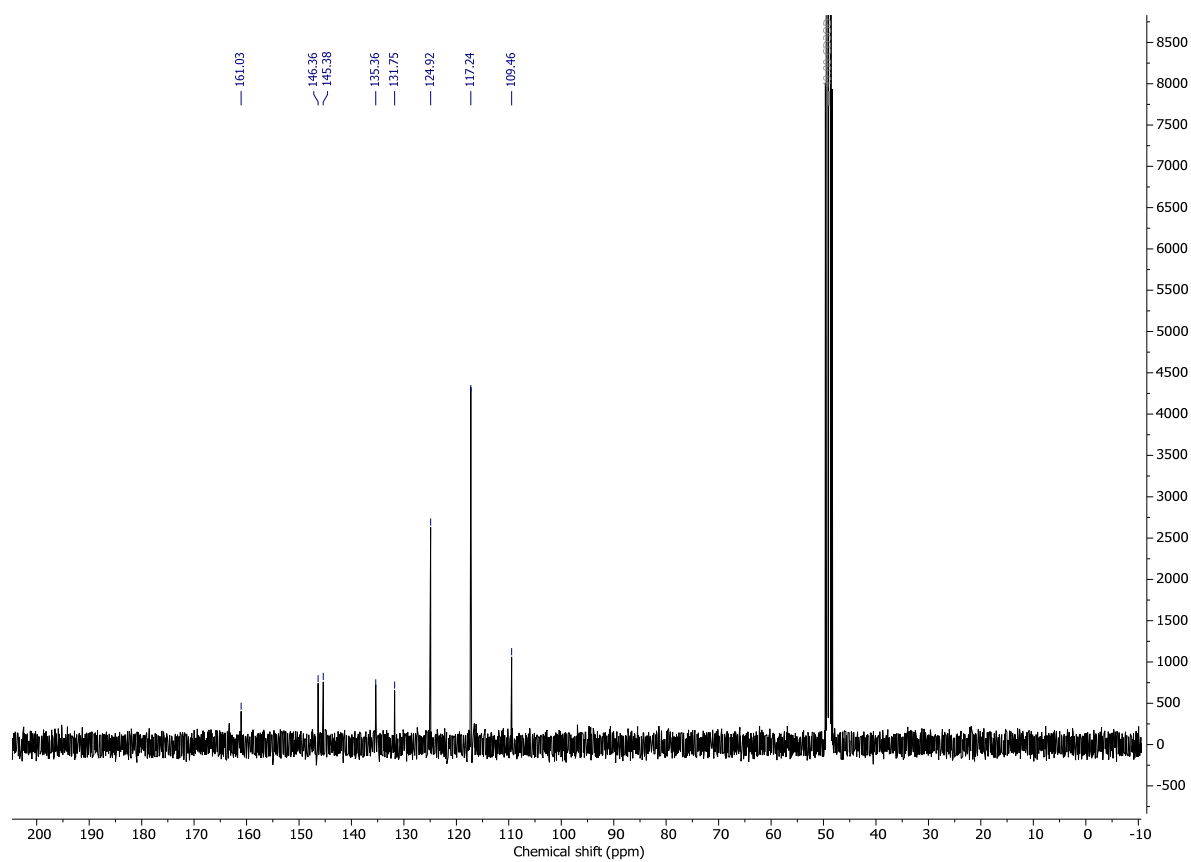
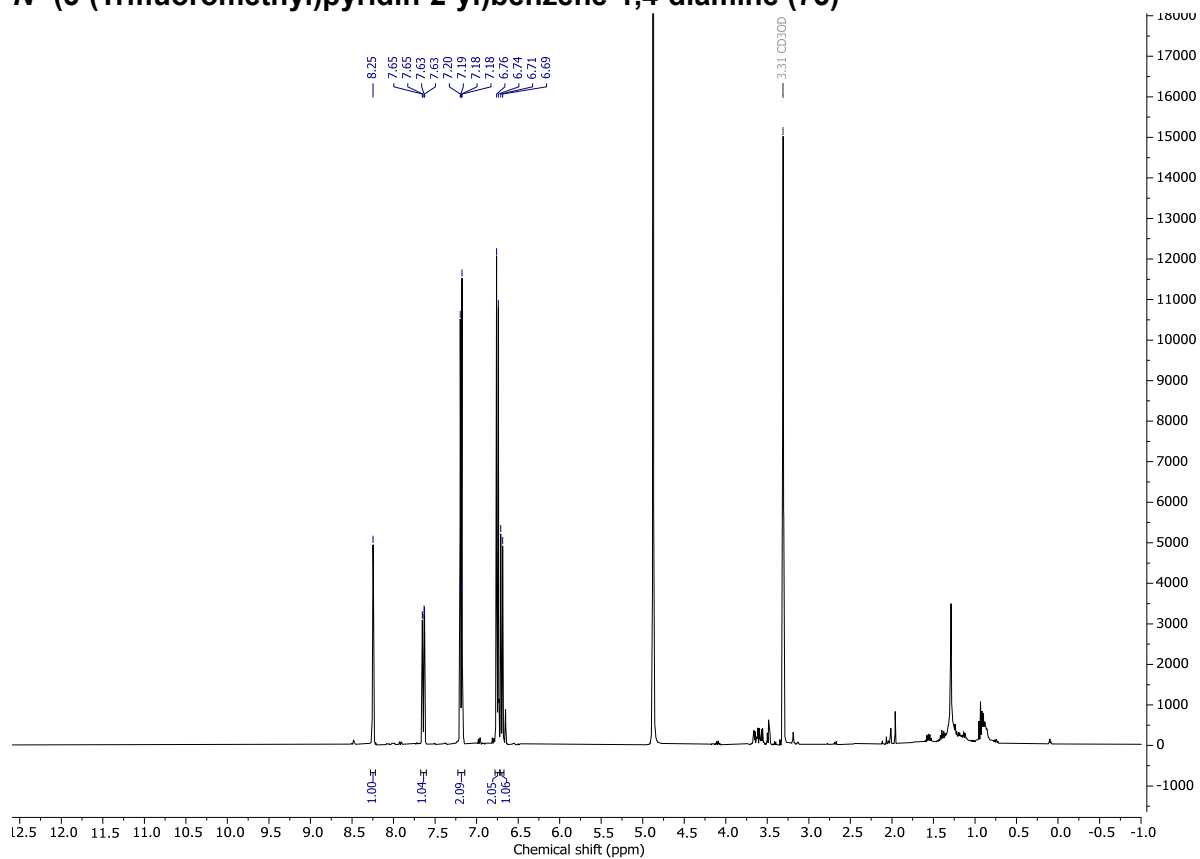
***N*⁴-(4-Aminophenyl)-6-methylpyrimidine-2,4-diamine (73)**



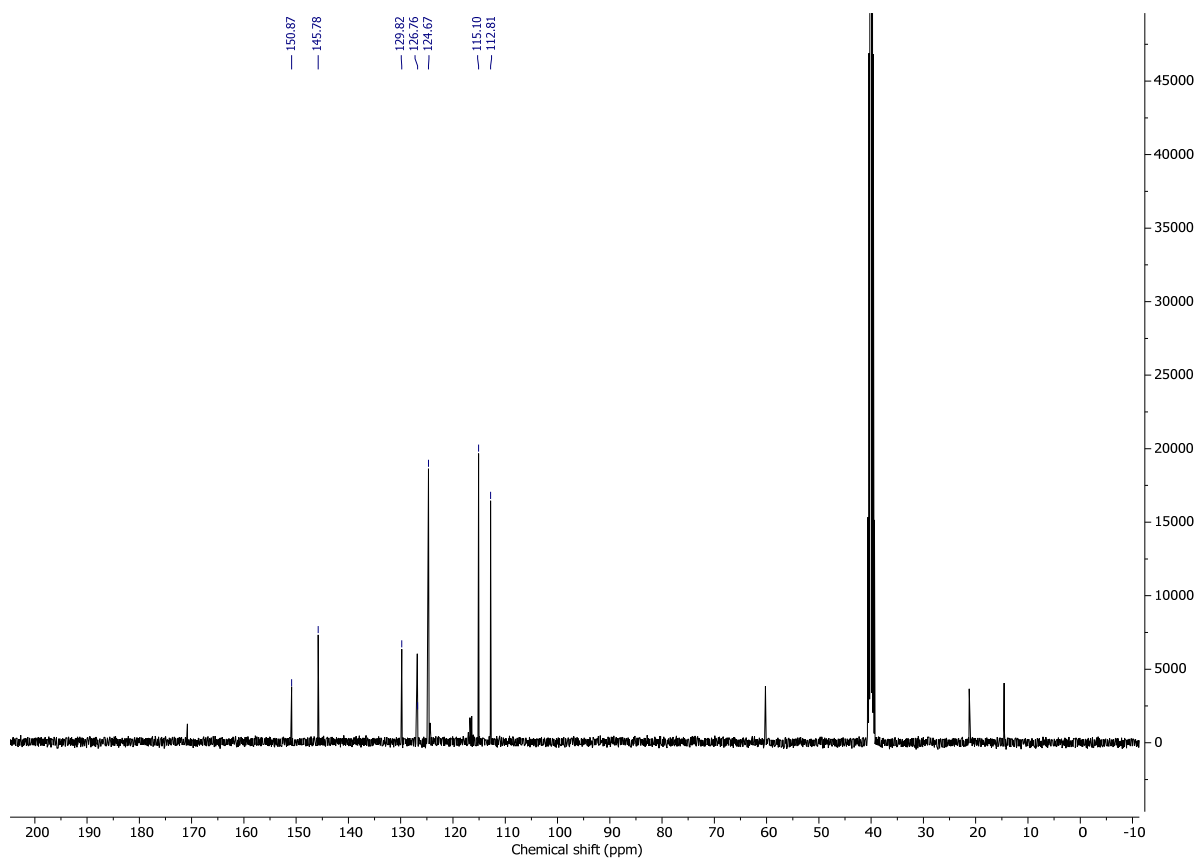
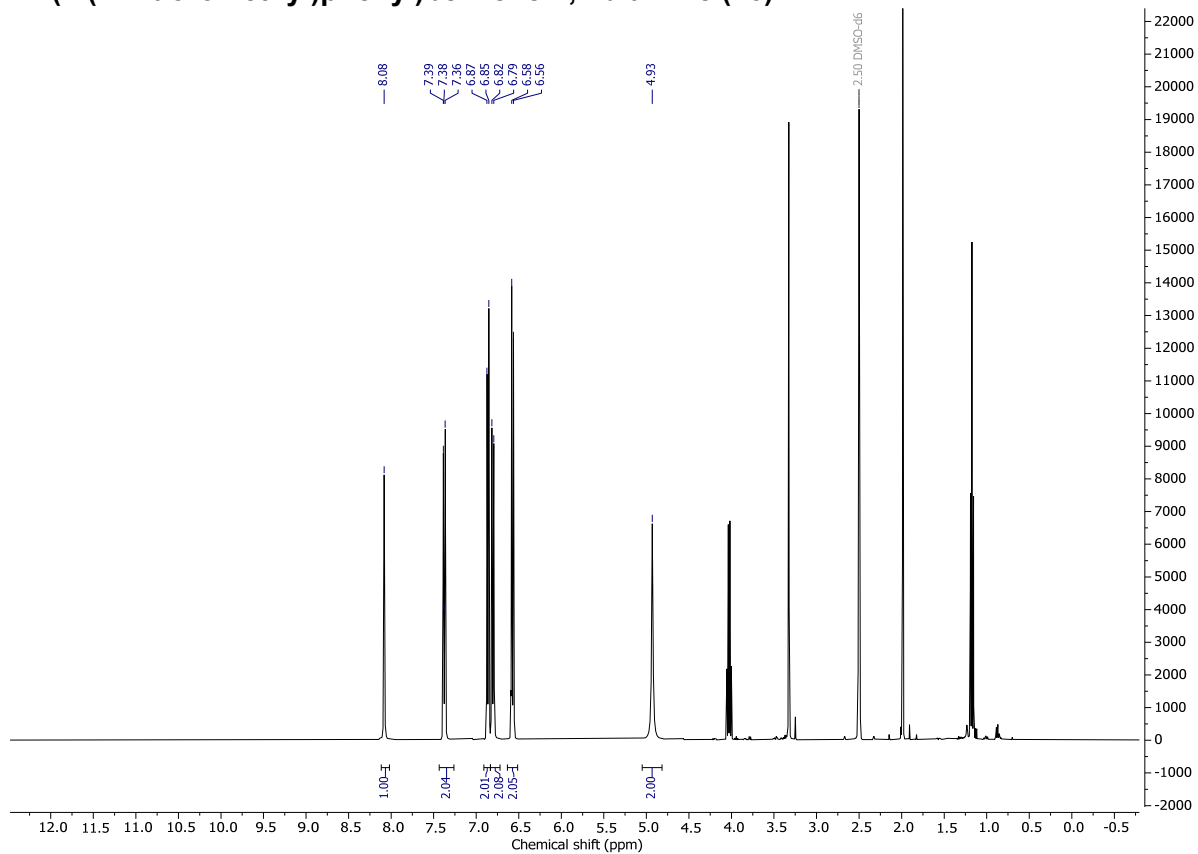
***N*⁴-(4-Aminophenyl)pyrimidine-2,4-diamine (74)**



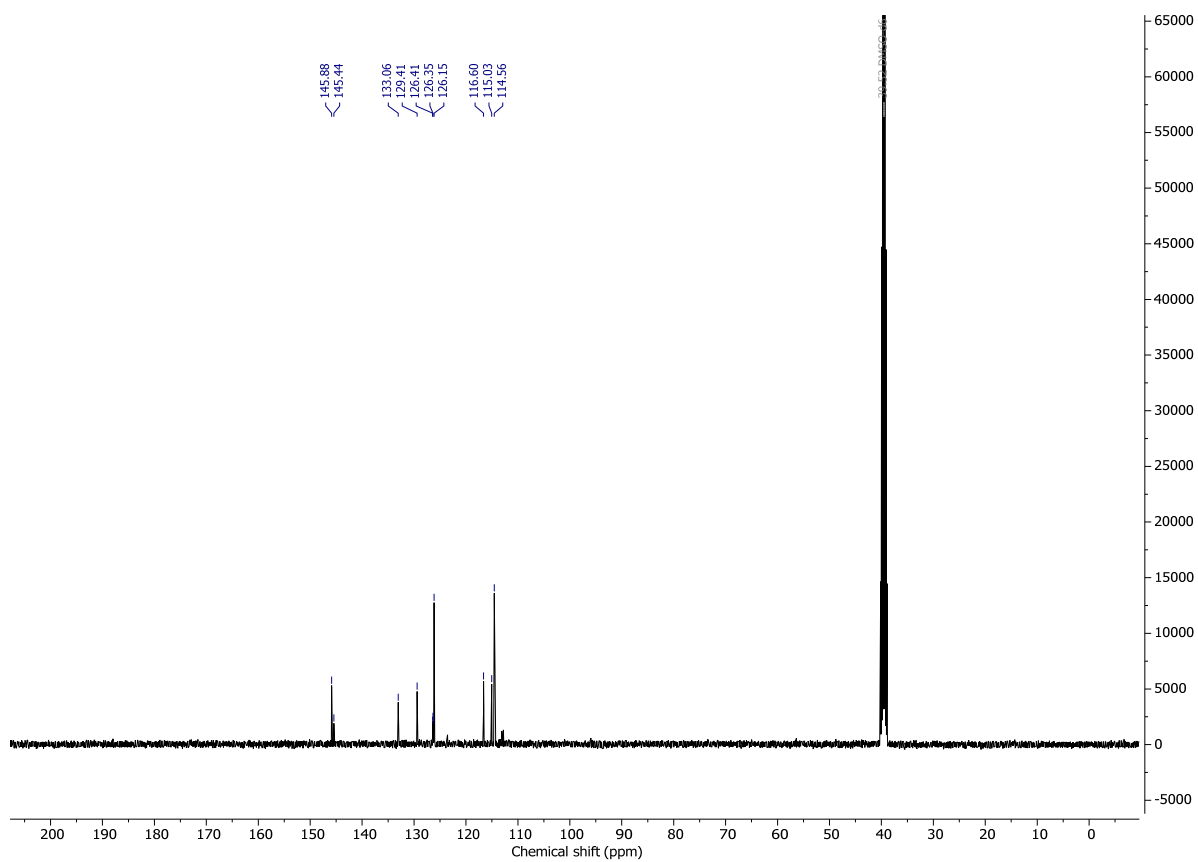
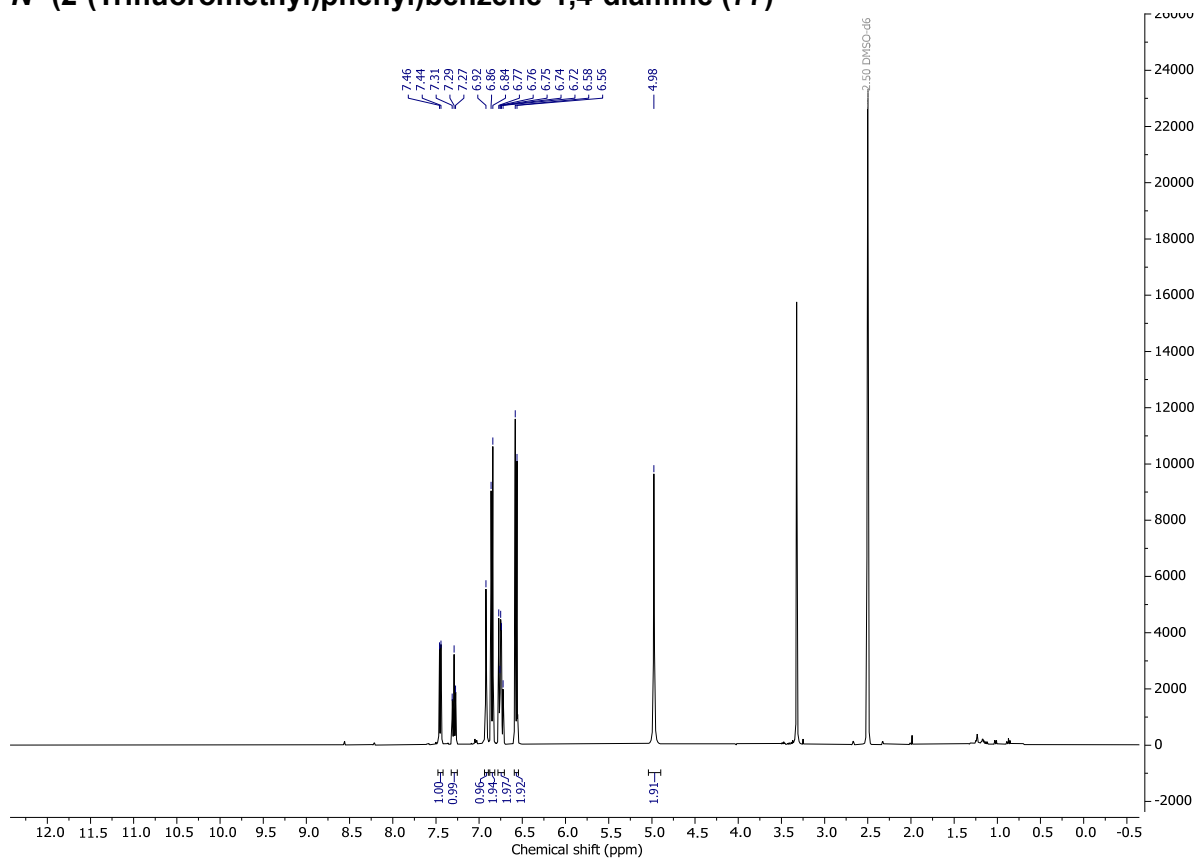
***N*'-(5-(Trifluoromethyl)pyridin-2-yl)benzene-1,4-diamine (75)**



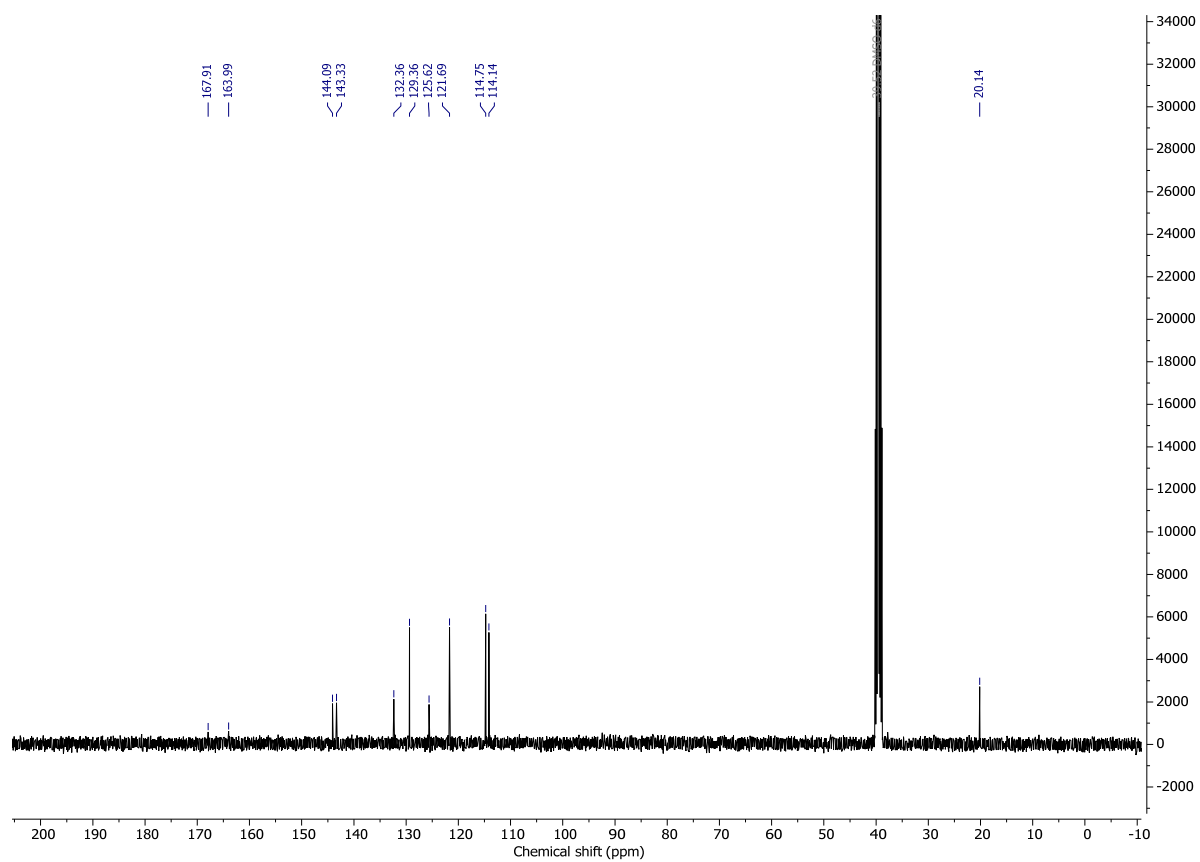
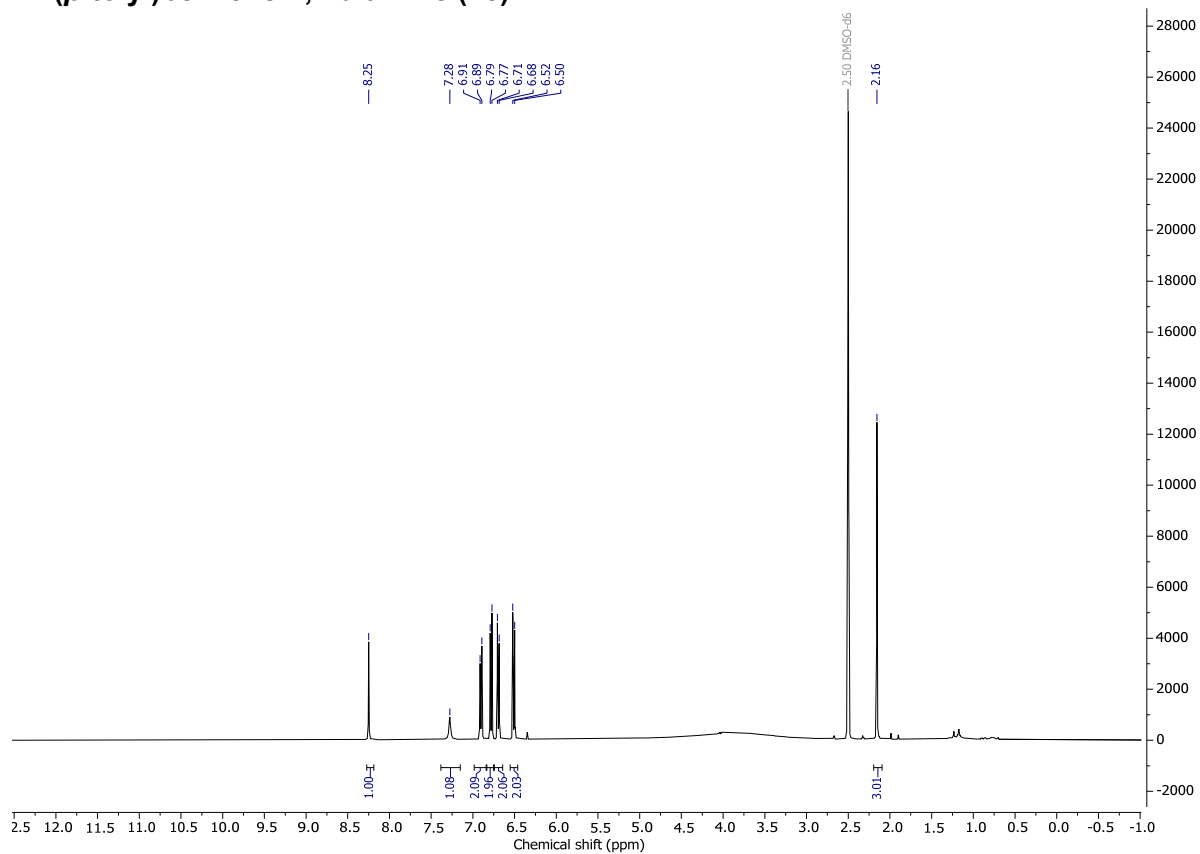
*N*¹-(4-(Trifluoromethyl)phenyl)benzene-1,4-diamine (76)



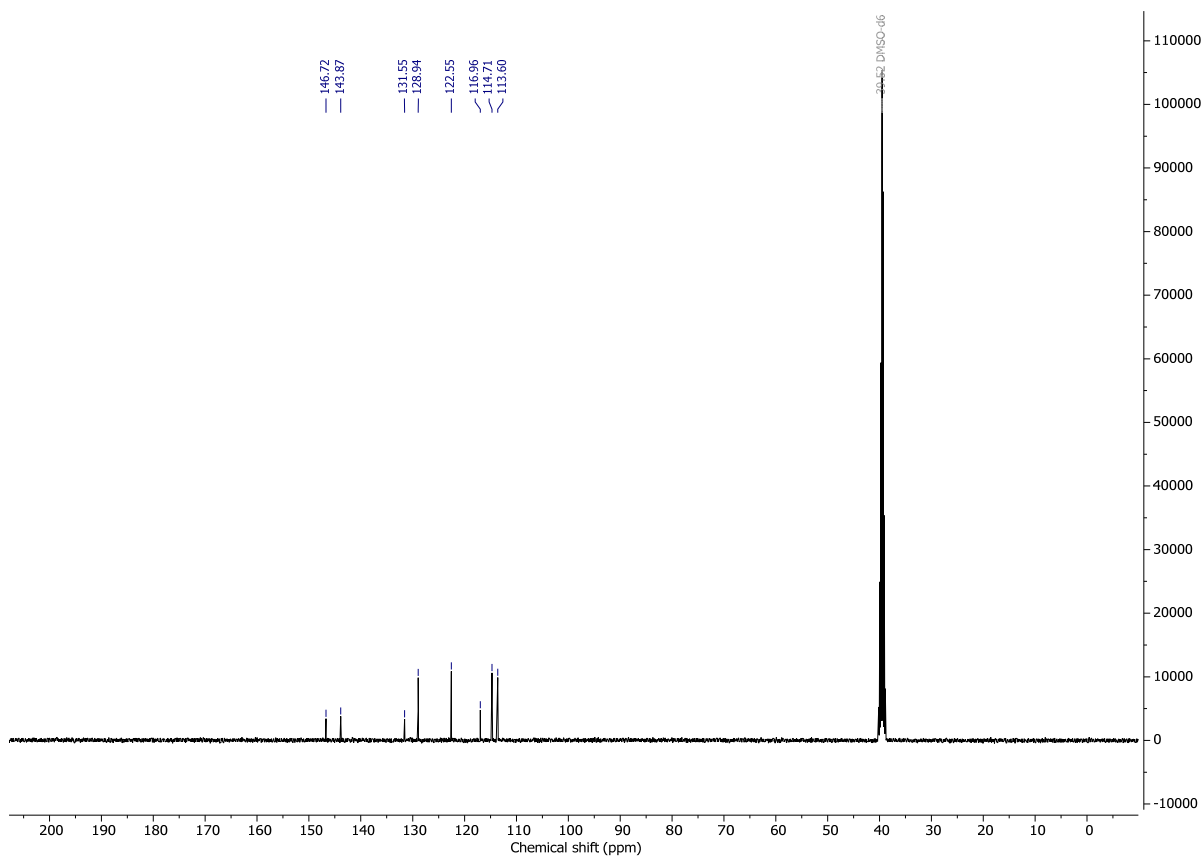
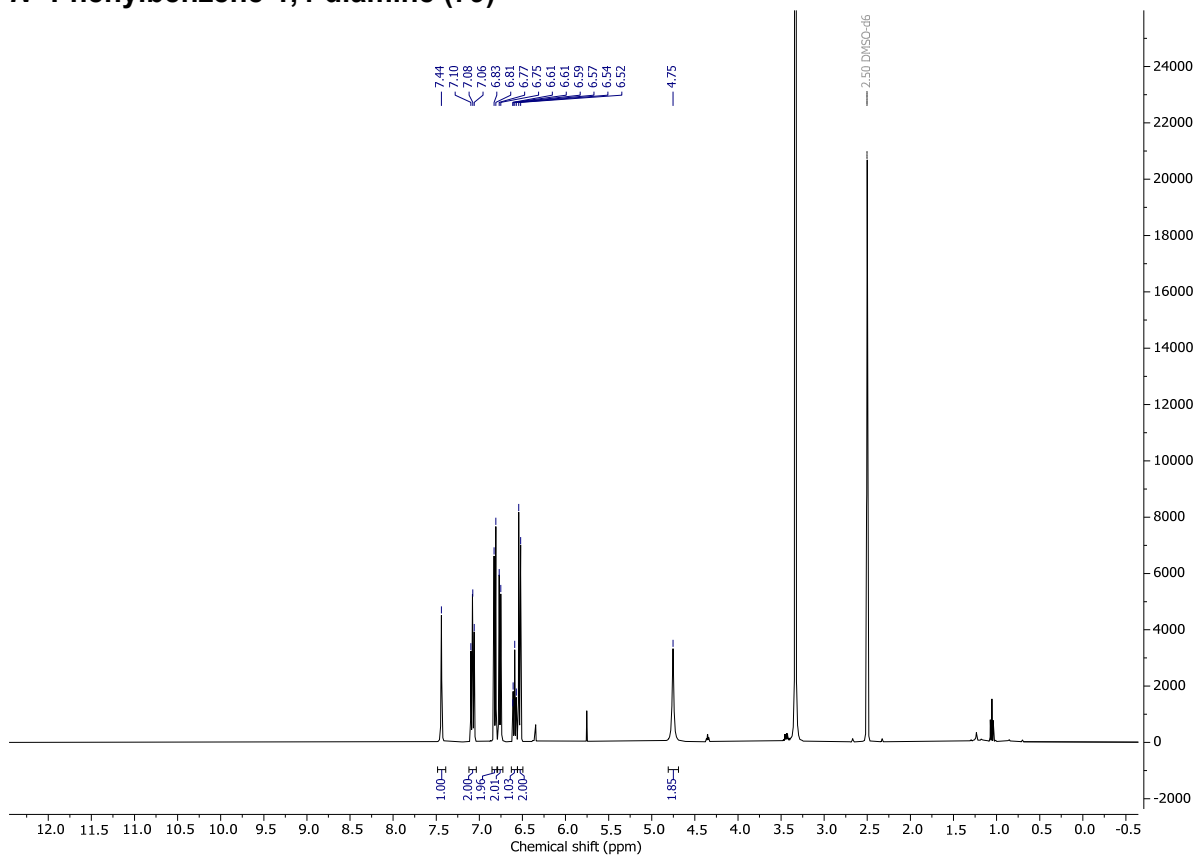
*N*¹-(2-(Trifluoromethyl)phenyl)benzene-1,4-diamine (77)



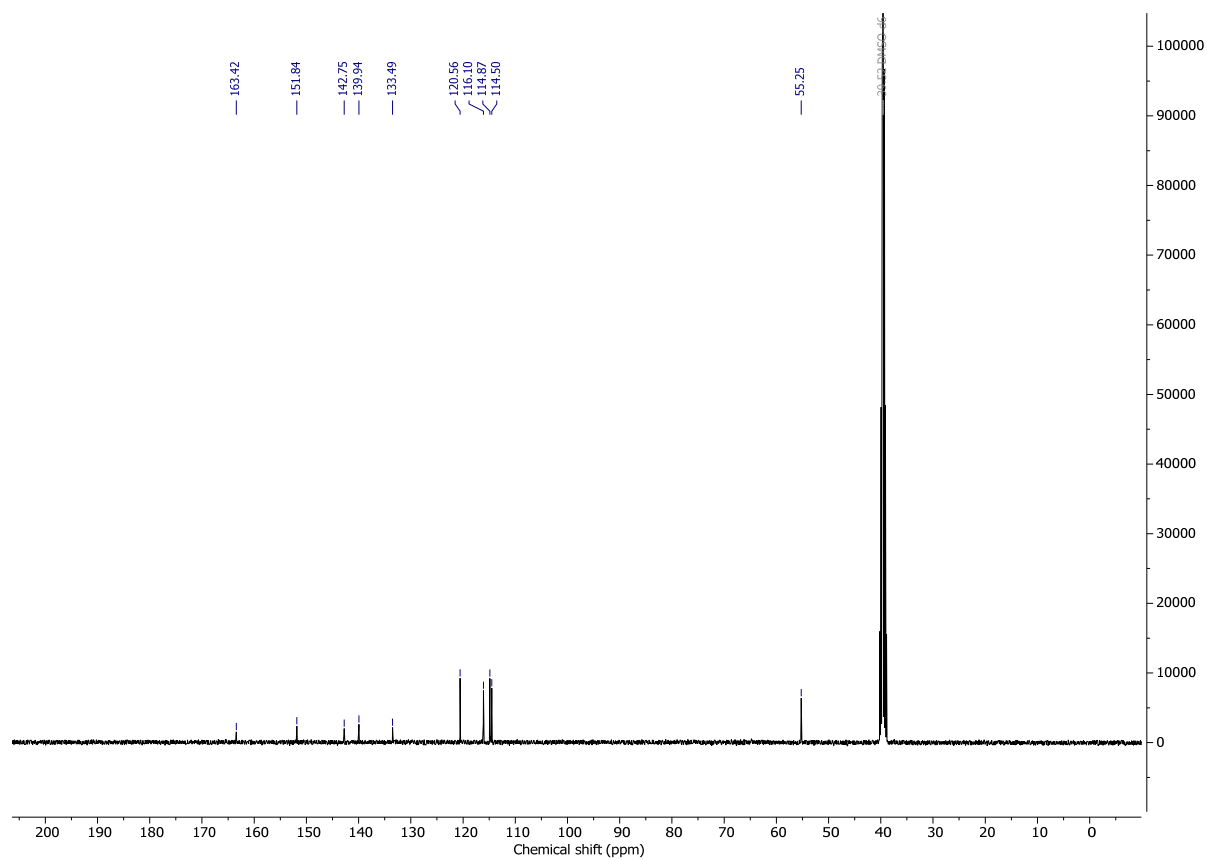
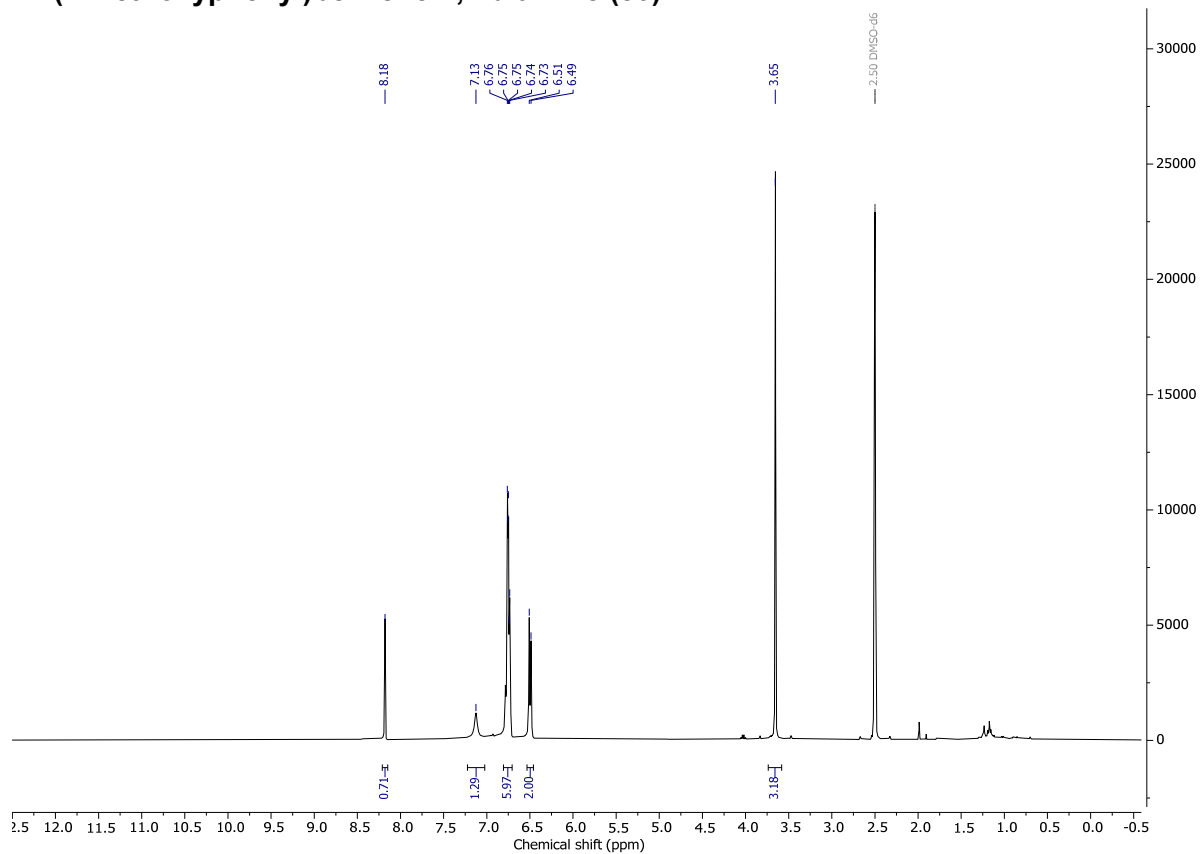
*N*¹-(*p*-tolyl)benzene-1,4-diamine (78)



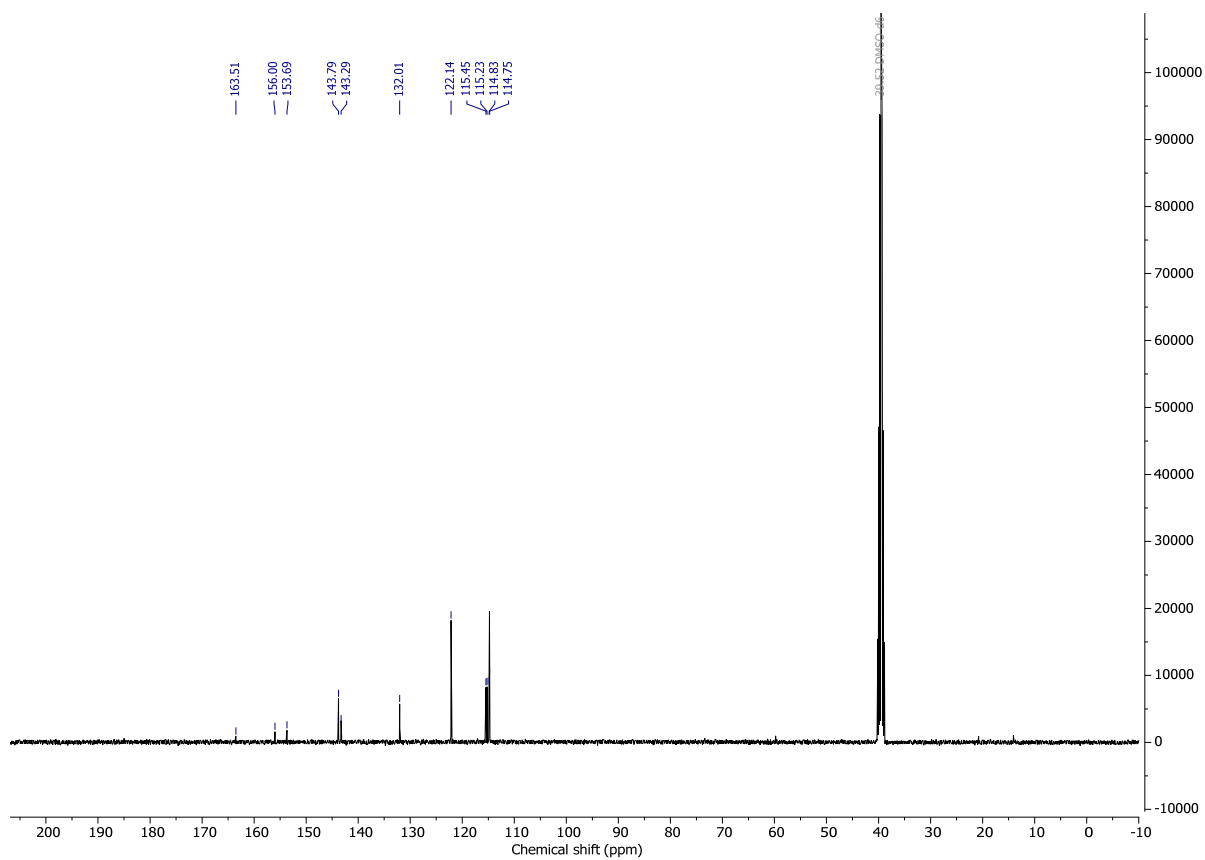
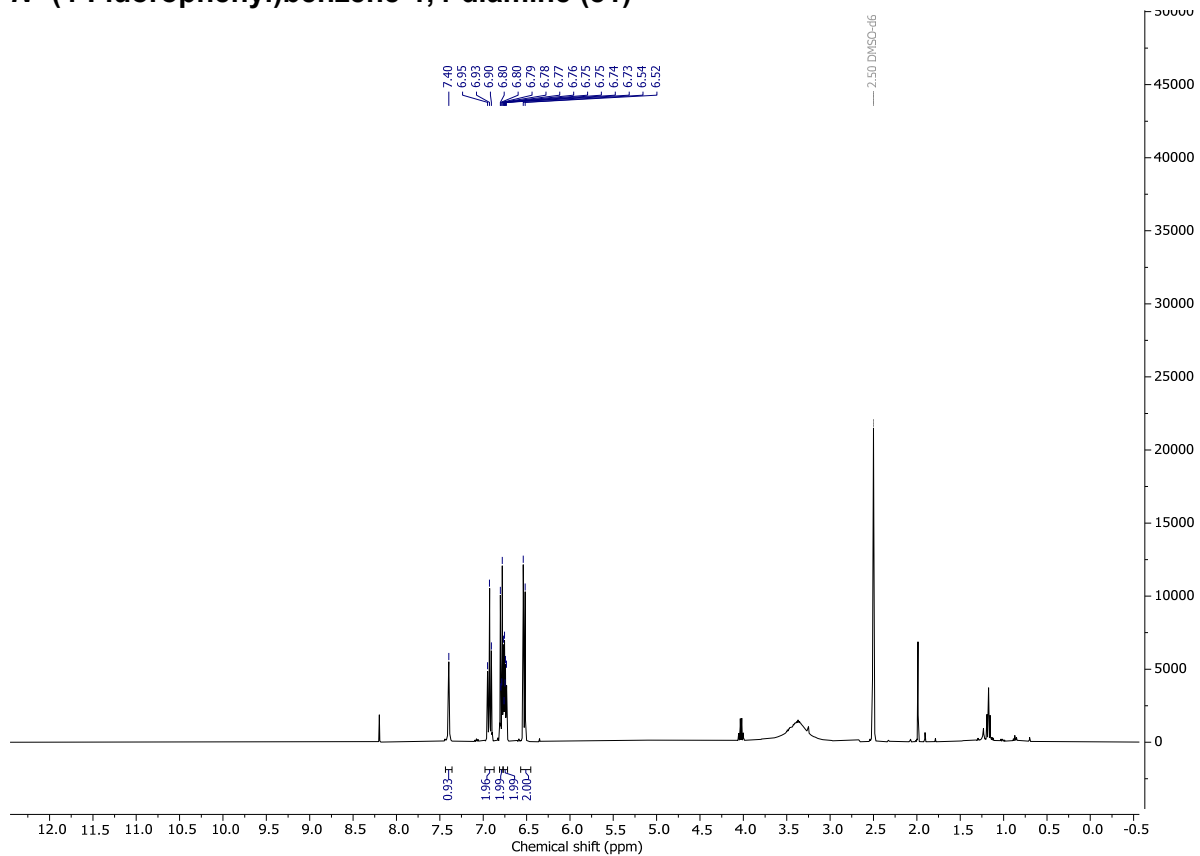
N¹-Phenylbenzene-1,4-diamine (79)



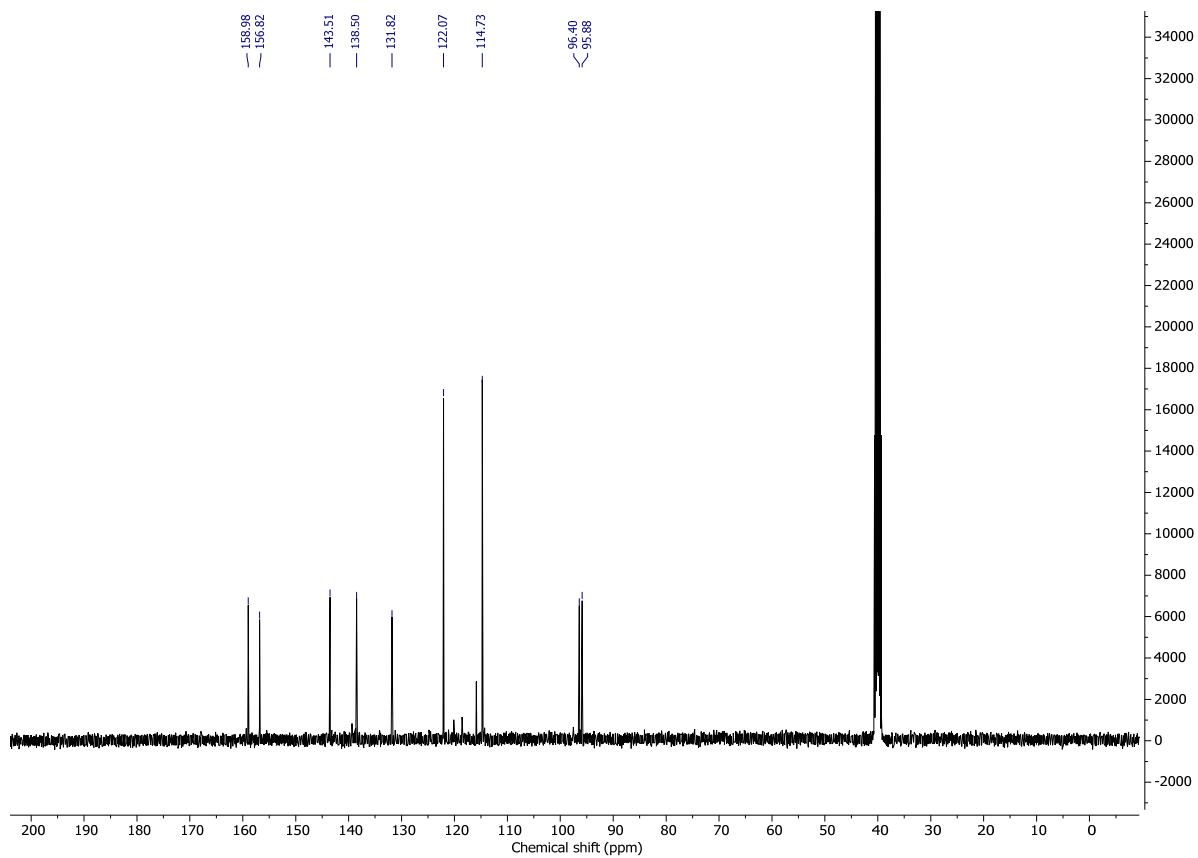
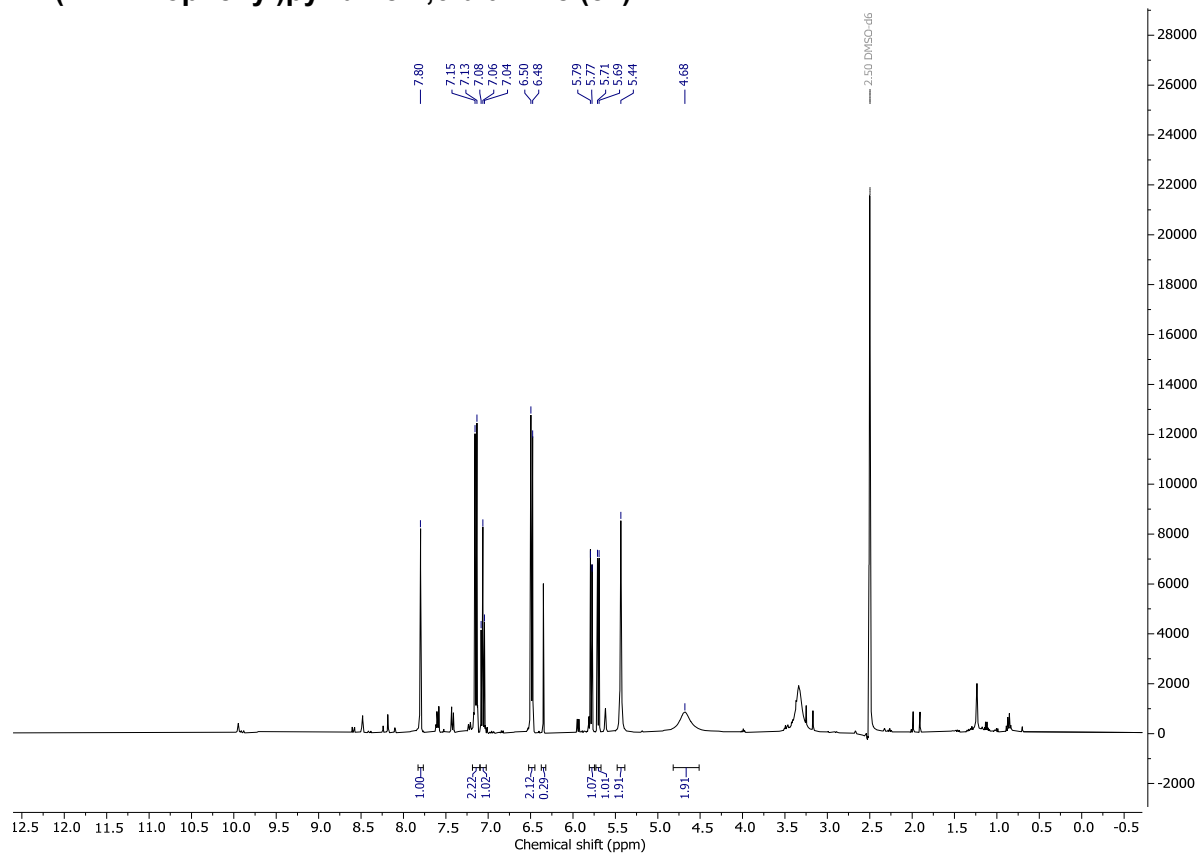
*N*¹-(4-Methoxyphenyl)benzene-1,4-diamine (80)



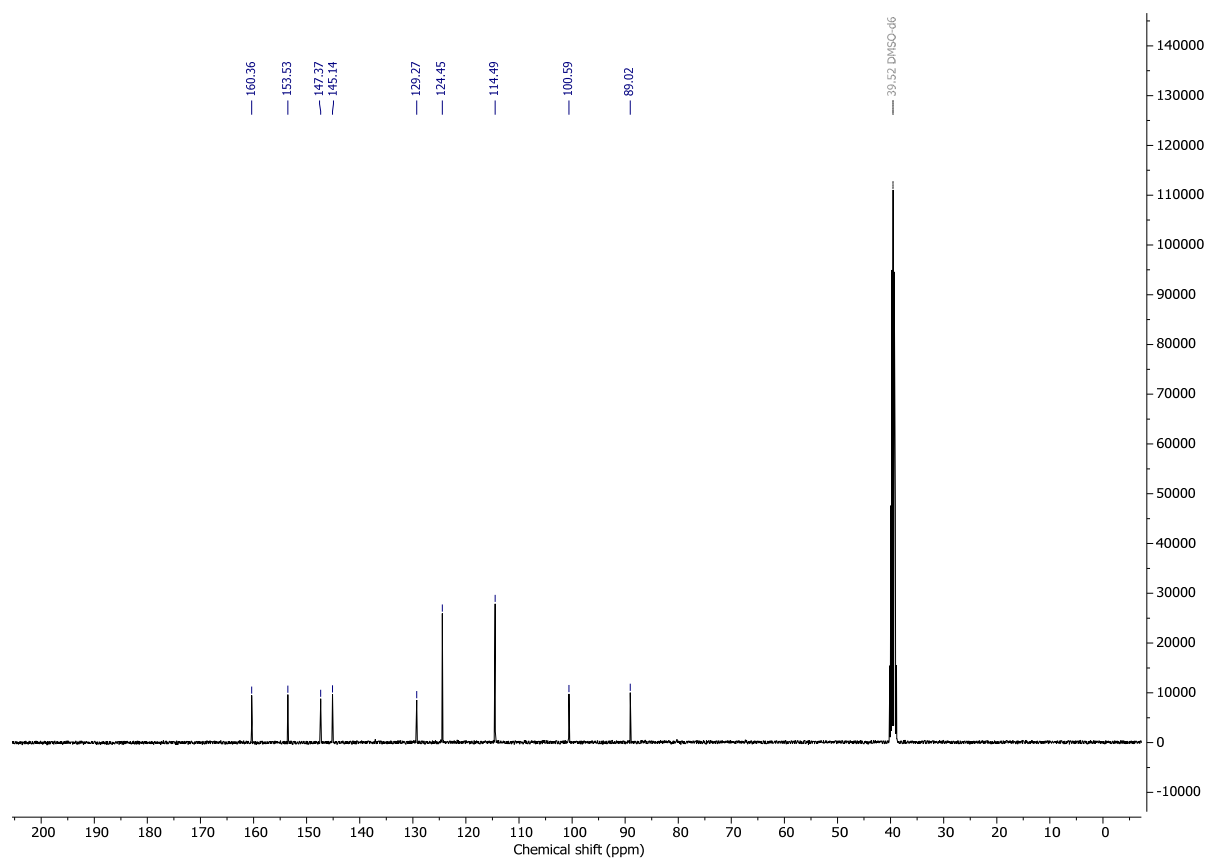
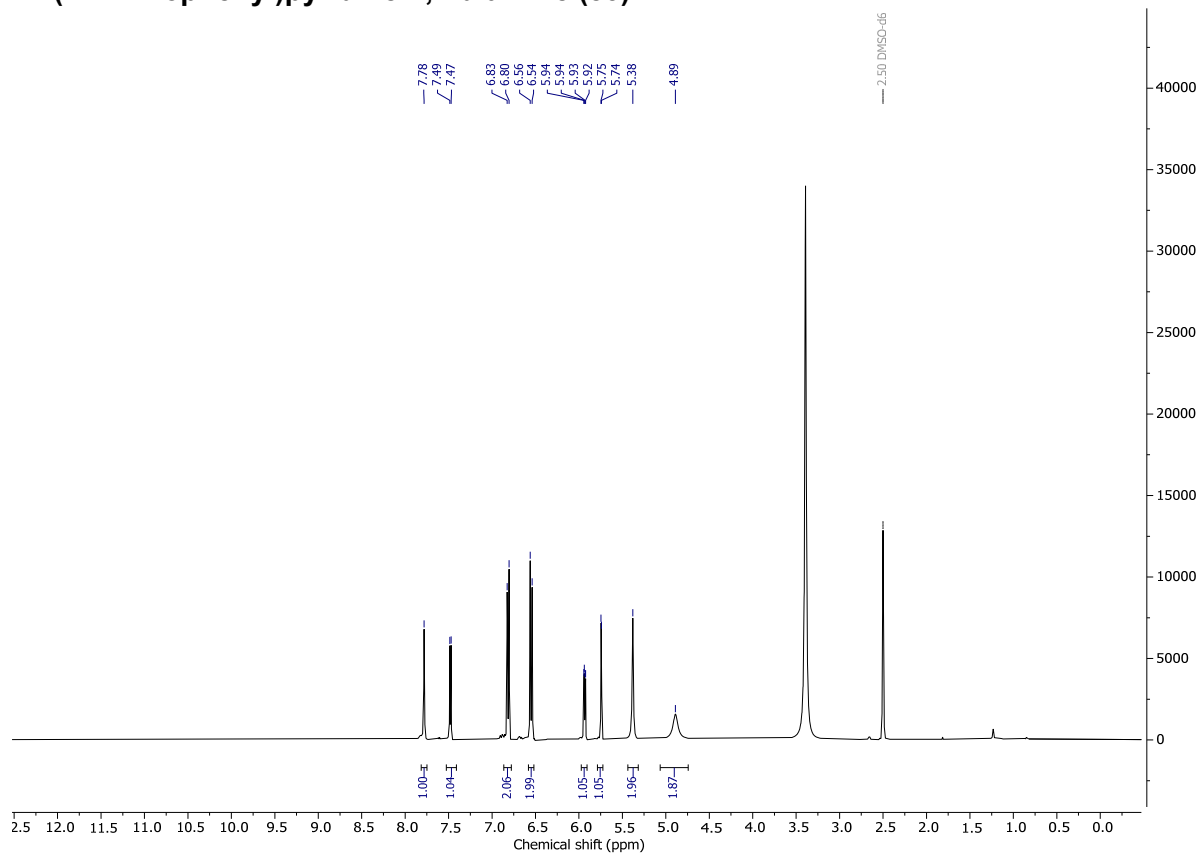
*N*¹-(4-Fluorophenyl)benzene-1,4-diamine (81)



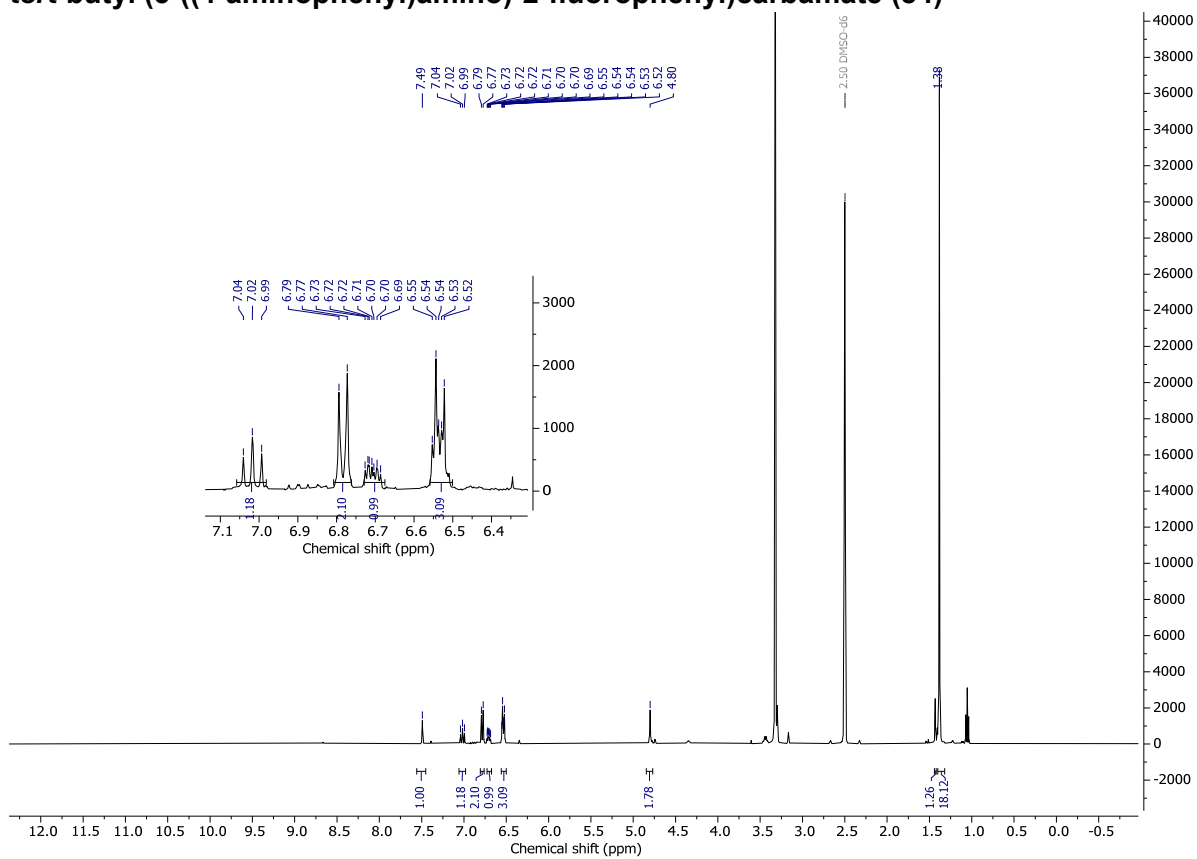
*N*²-(4-Aminophenyl)pyridine-2,6-diamine (82)



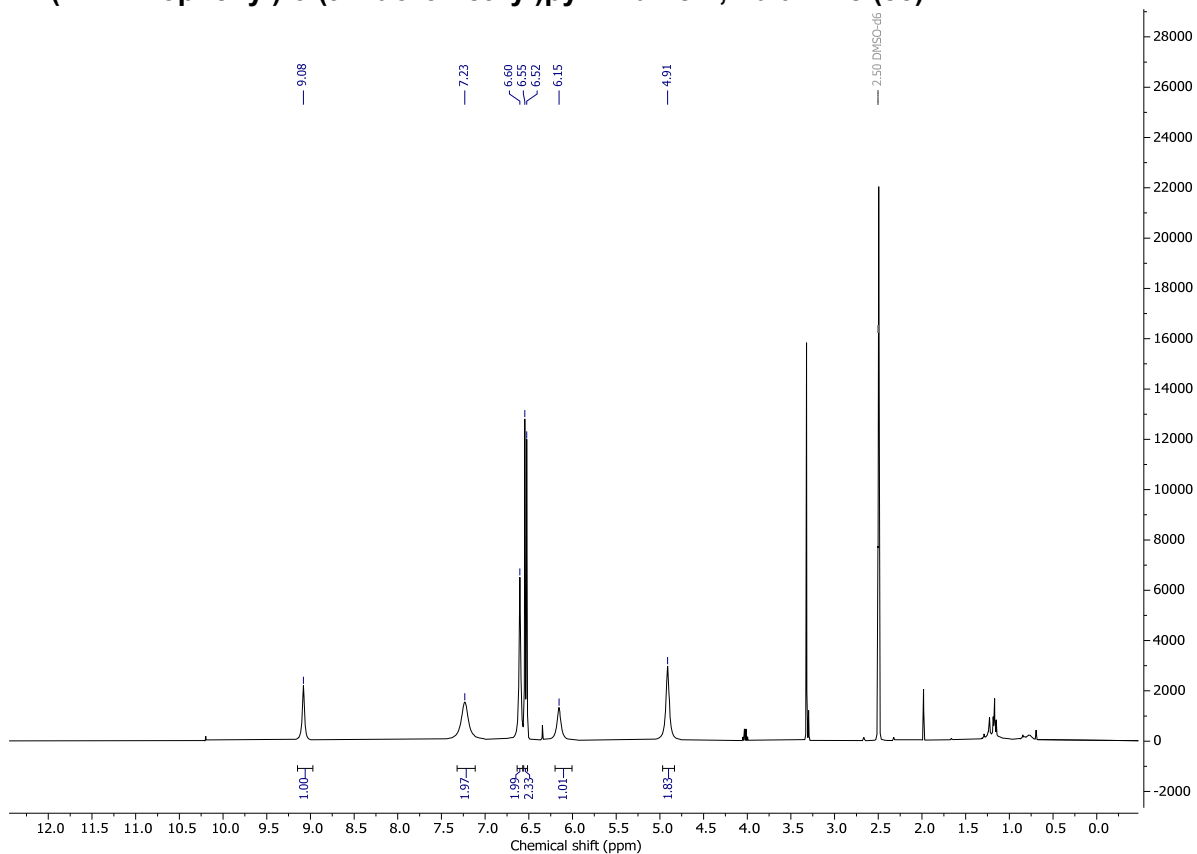
N⁴-(4-Aminophenyl)pyridine-2,4-diamine (83)



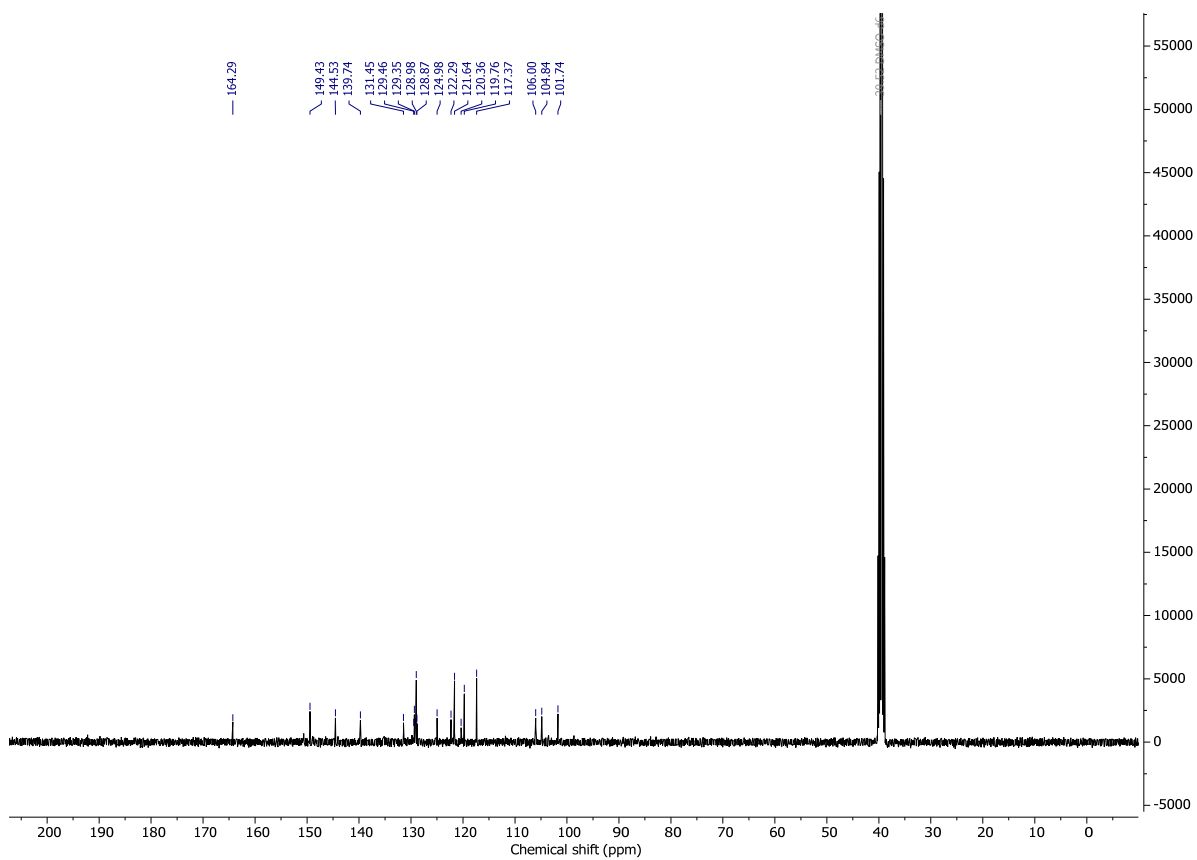
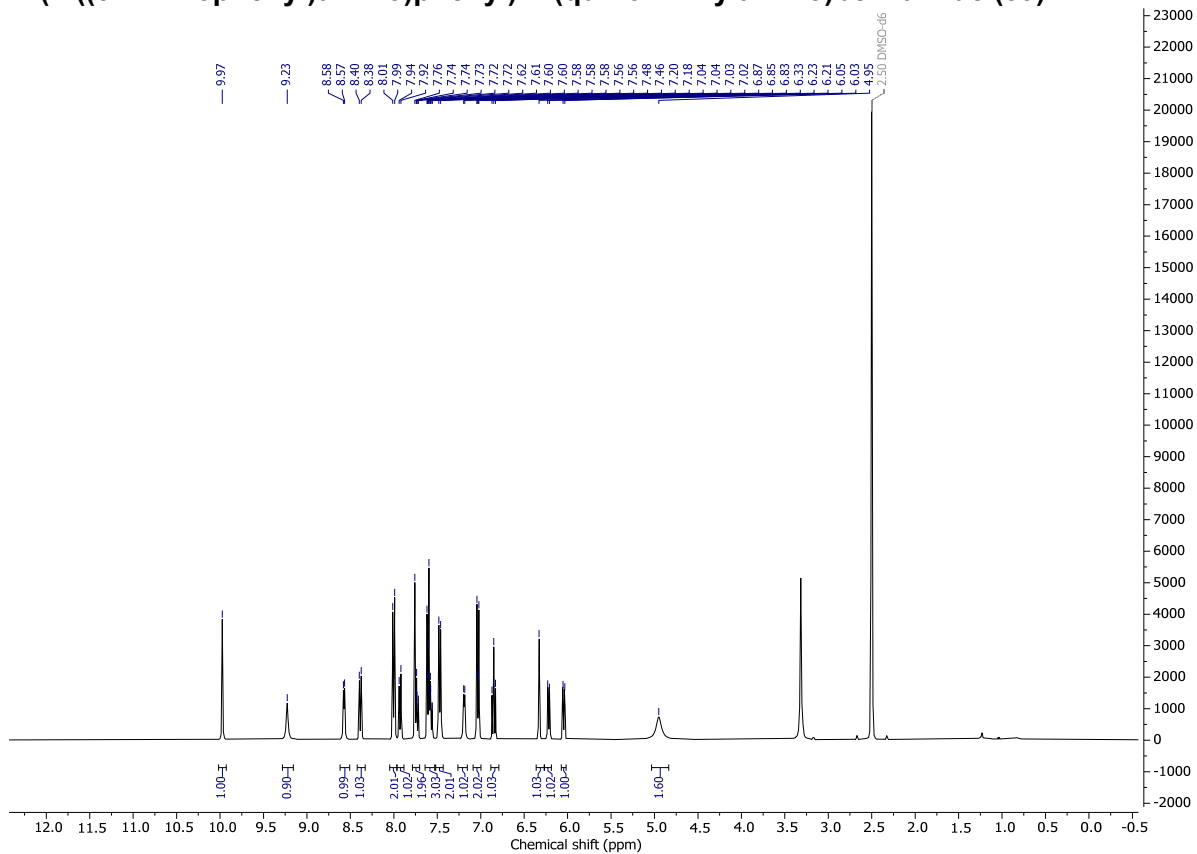
tert-Butyl (5-((4-aminophenyl)amino)-2-fluorophenyl)(tert-butoxycarbonyl)carbamate & tert-butyl (5-((4-aminophenyl)amino)-2-fluorophenyl)carbamate (84)



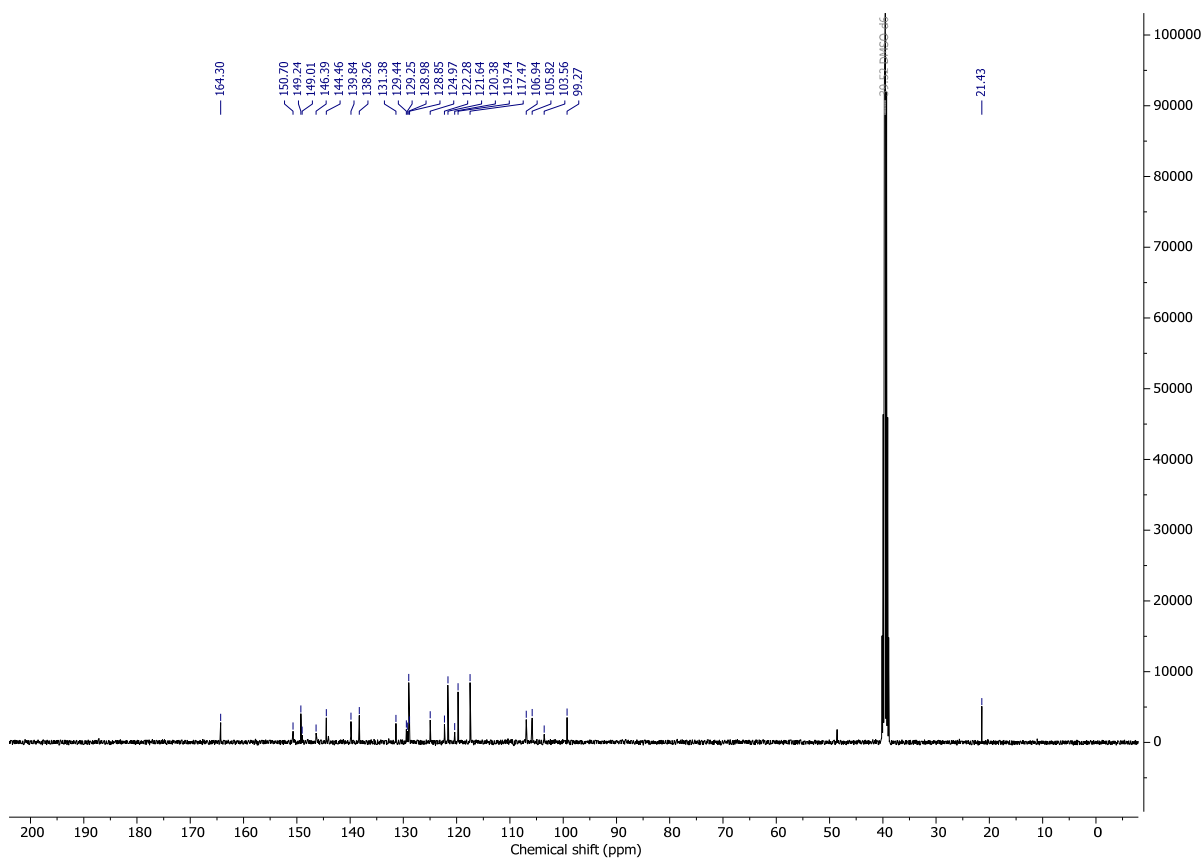
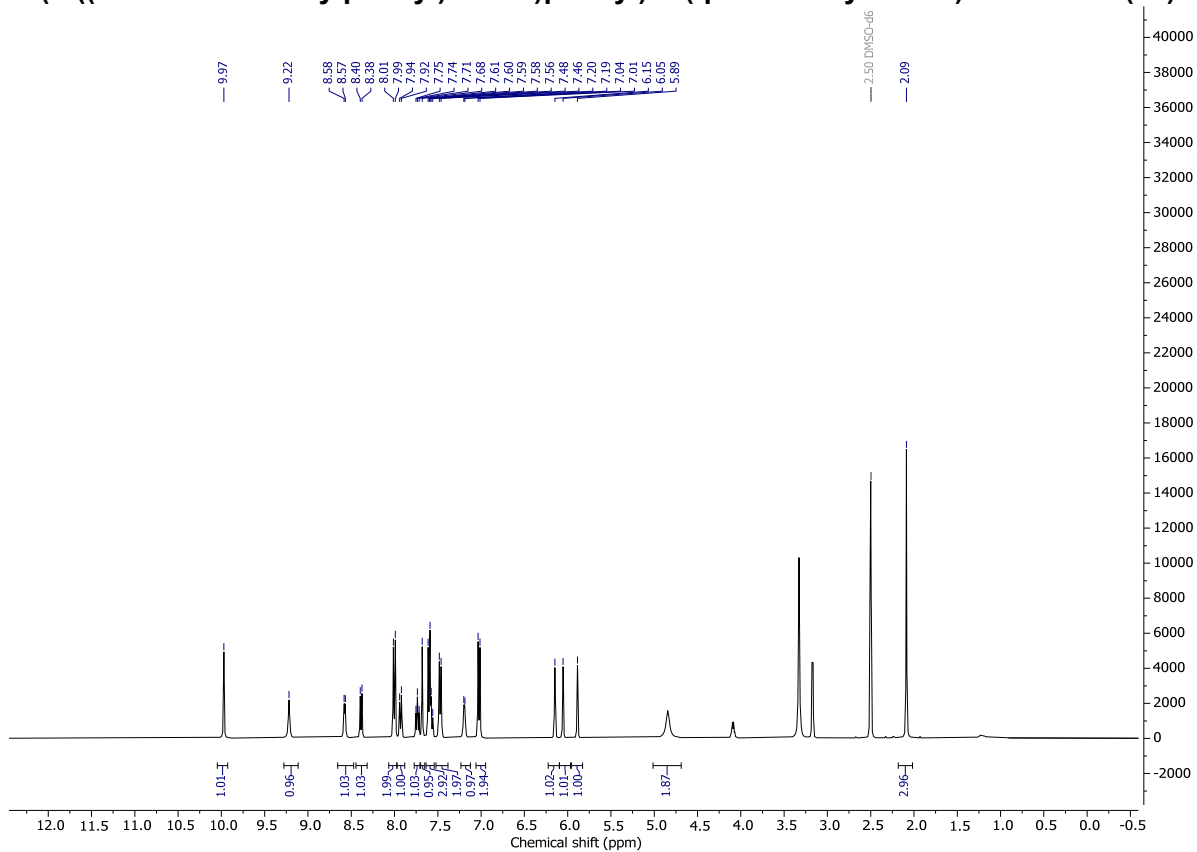
N⁴-(4-Aminophenyl)-6-(trifluoromethyl)pyrimidine-2,4-diamine (85)



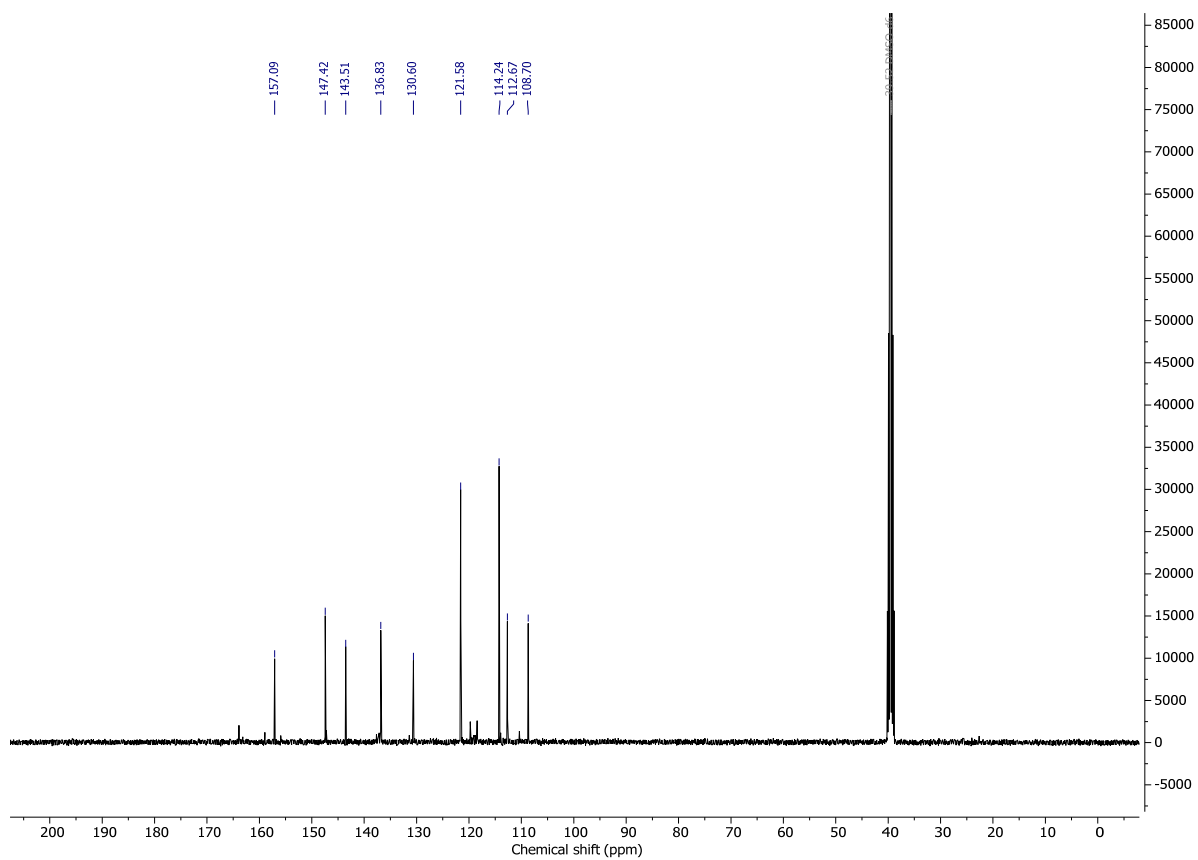
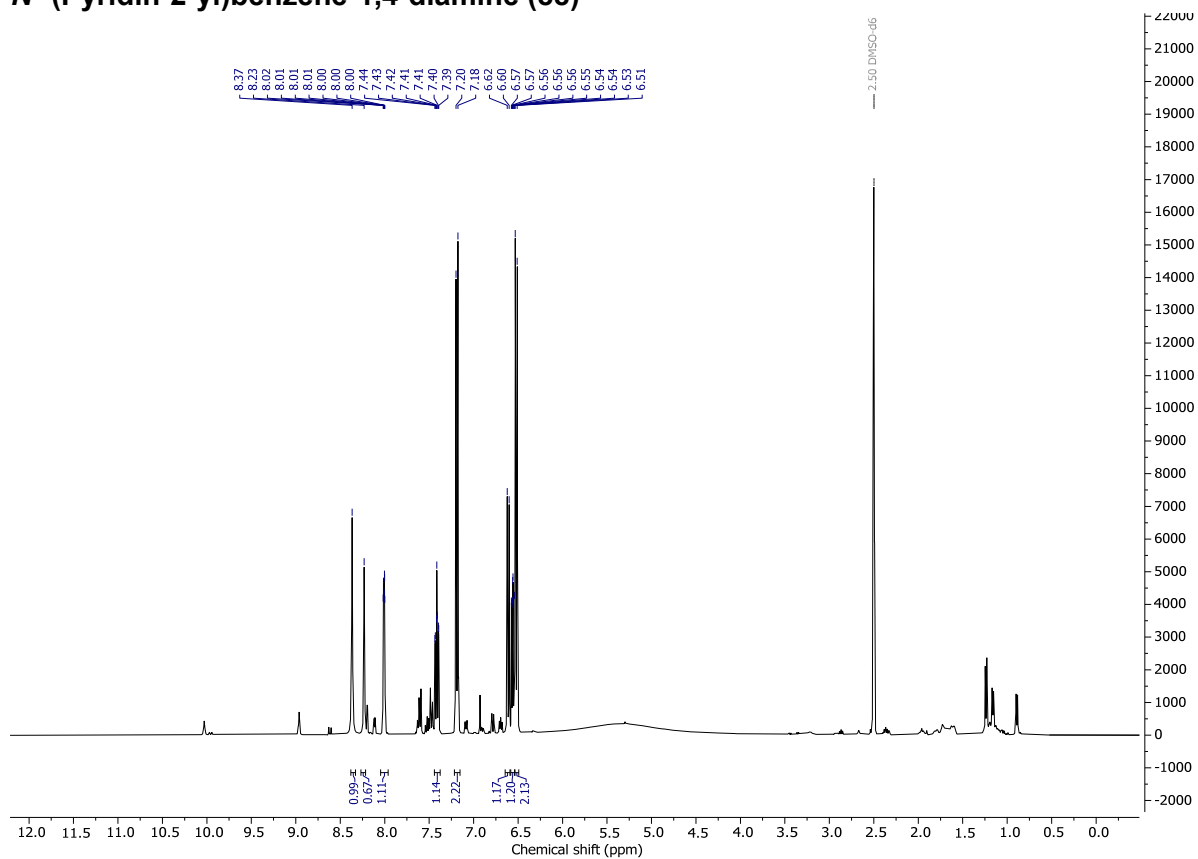
N-(4-((3-Aminophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (39)



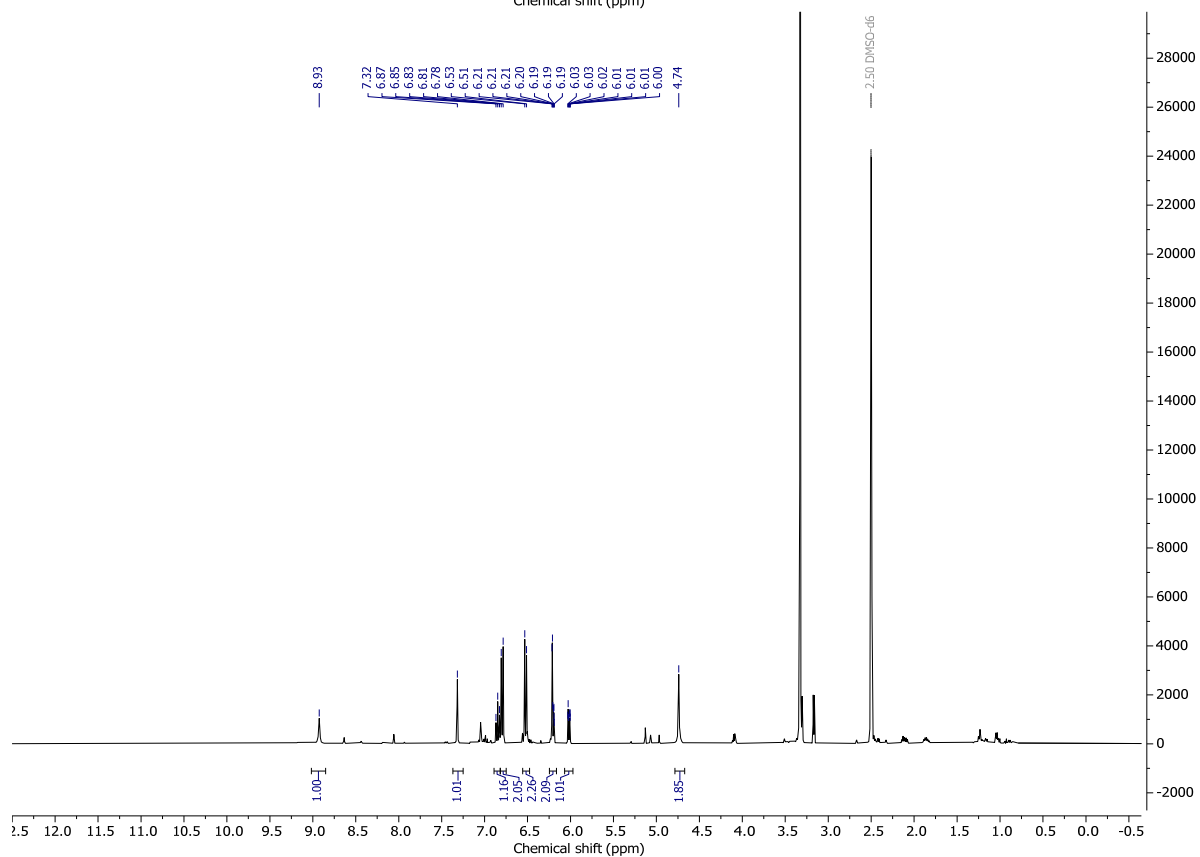
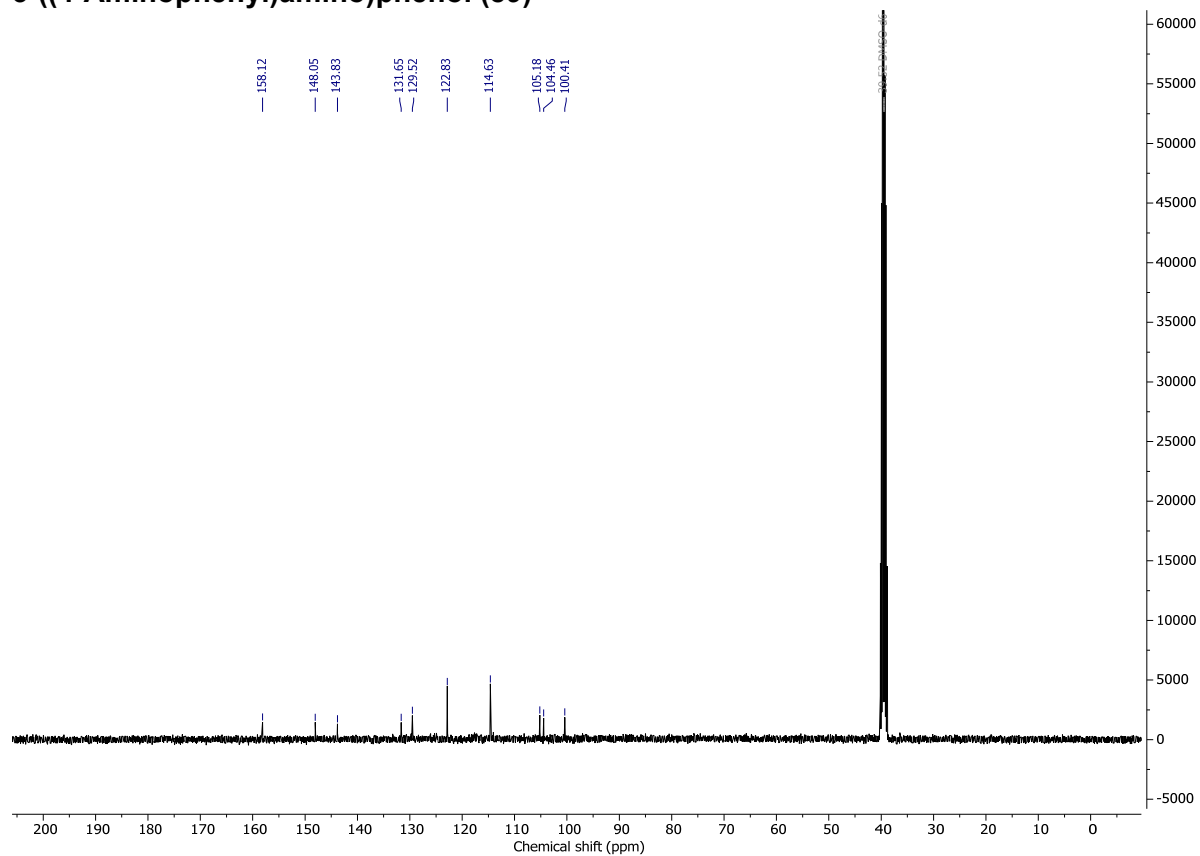
N-(4-((3-Amino-5-methylphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (43)



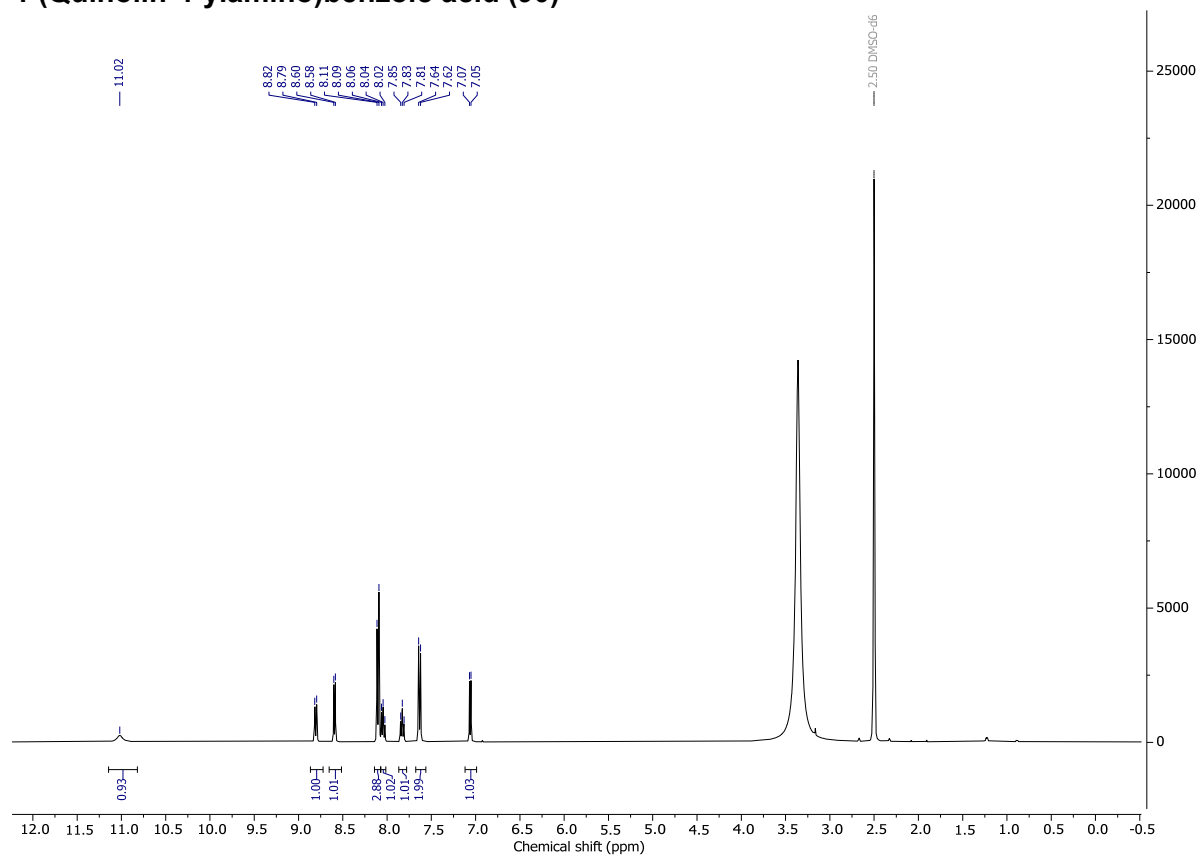
*N*¹-(Pyridin-2-yl)benzene-1,4-diamine (88)



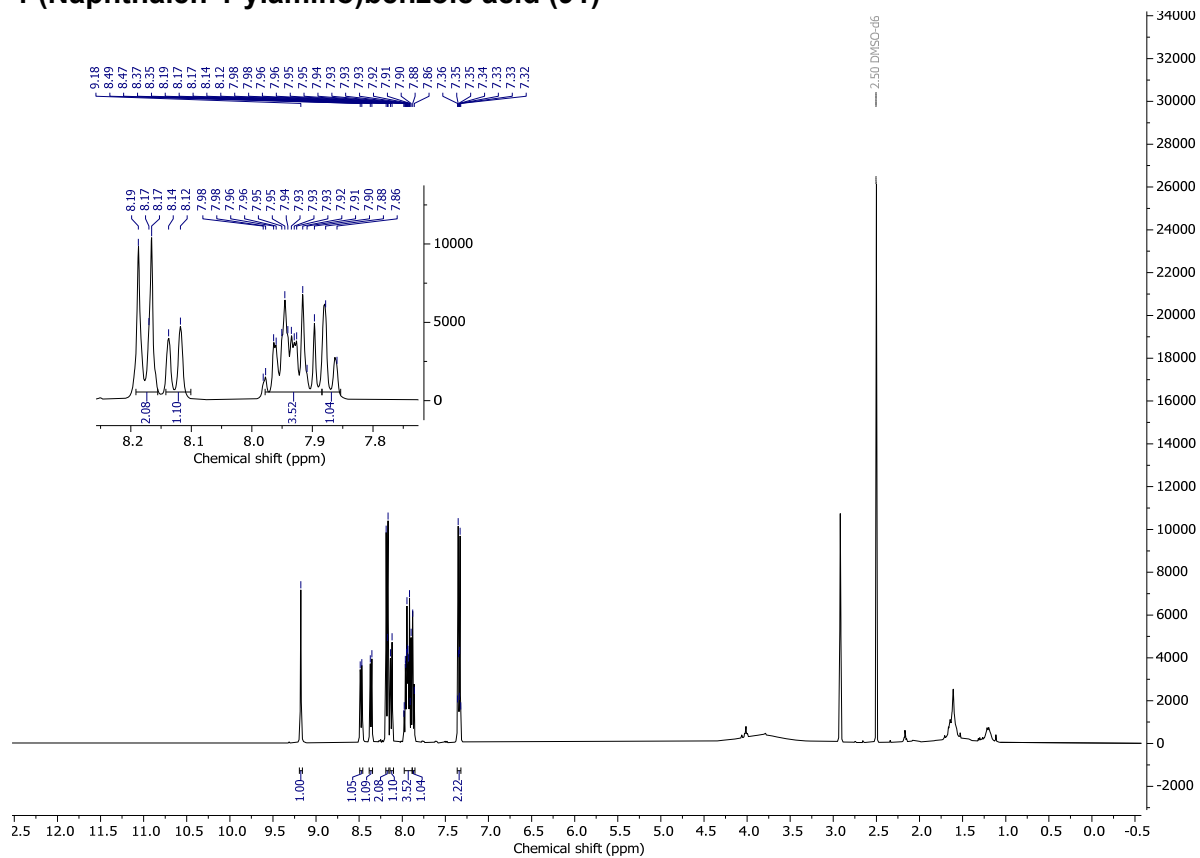
3-((4-Aminophenyl)amino)phenol (89)



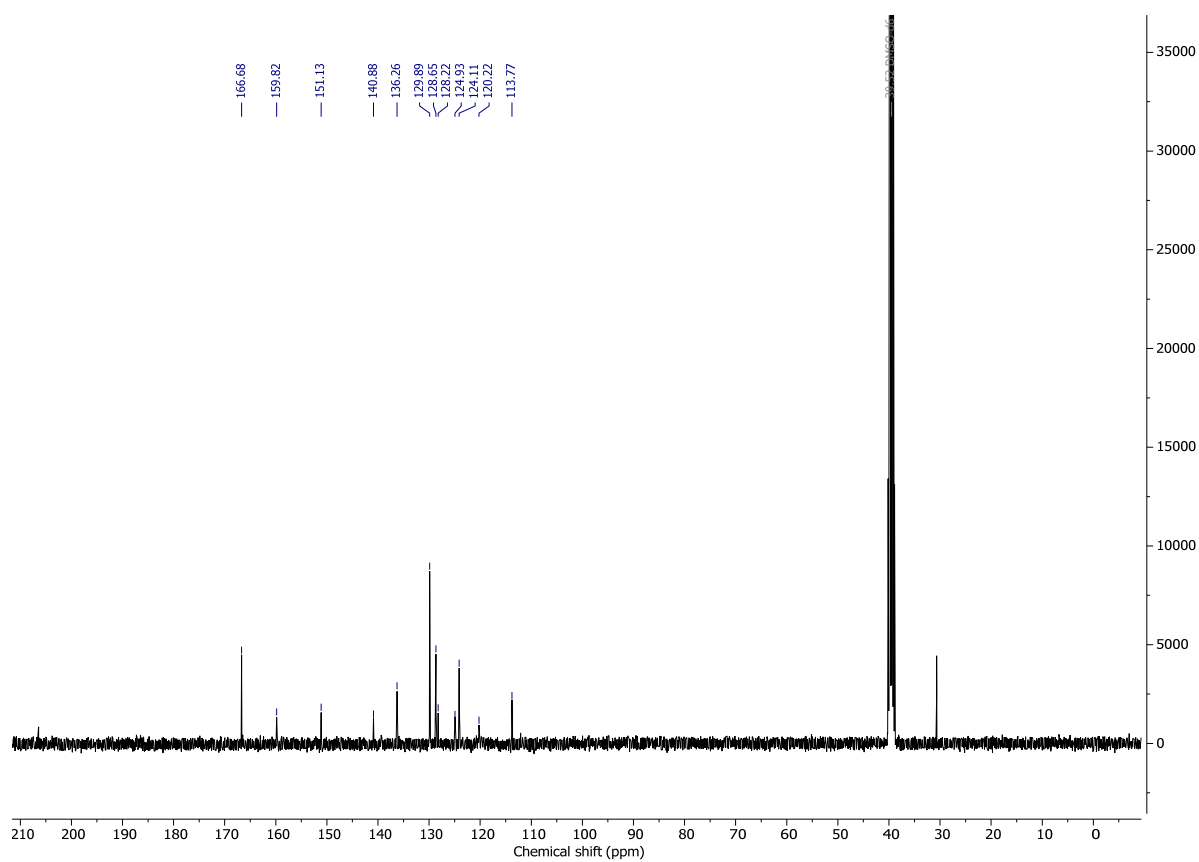
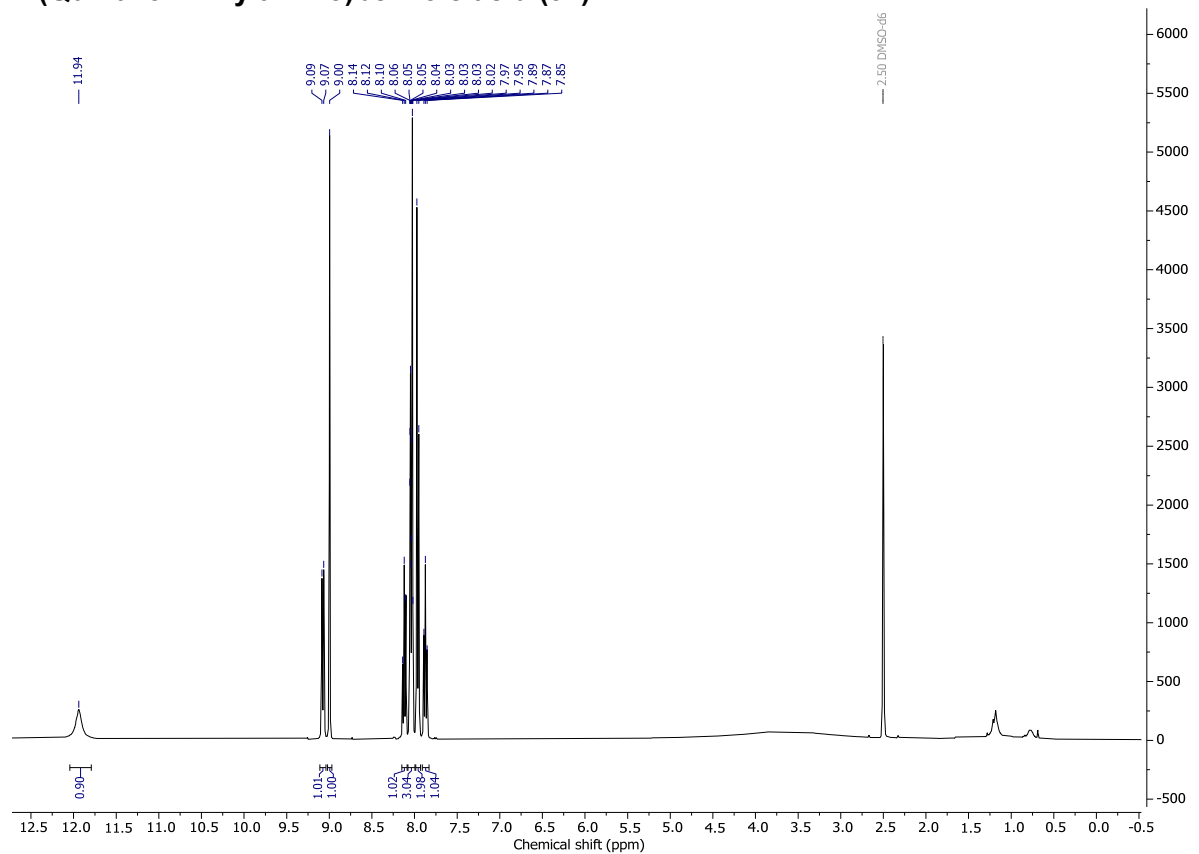
4-(Quinolin-4-ylamino)benzoic acid (90)



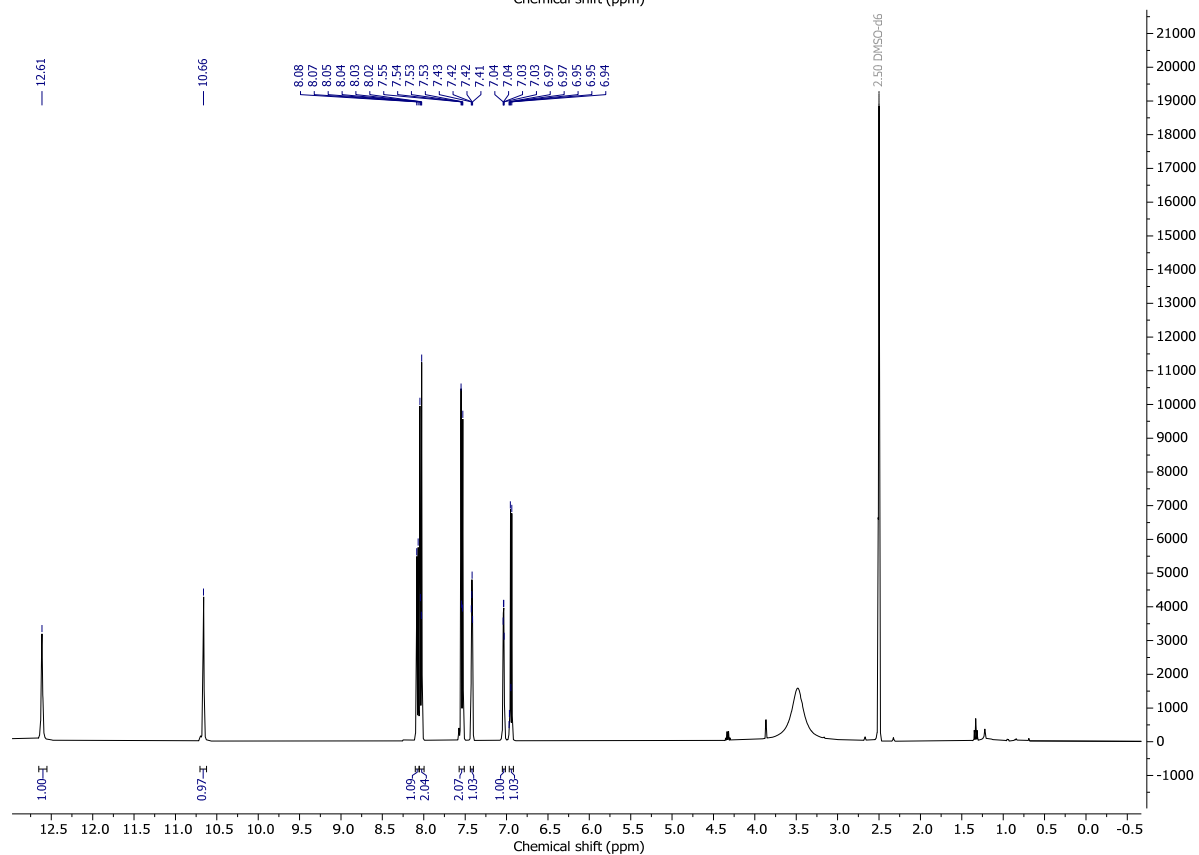
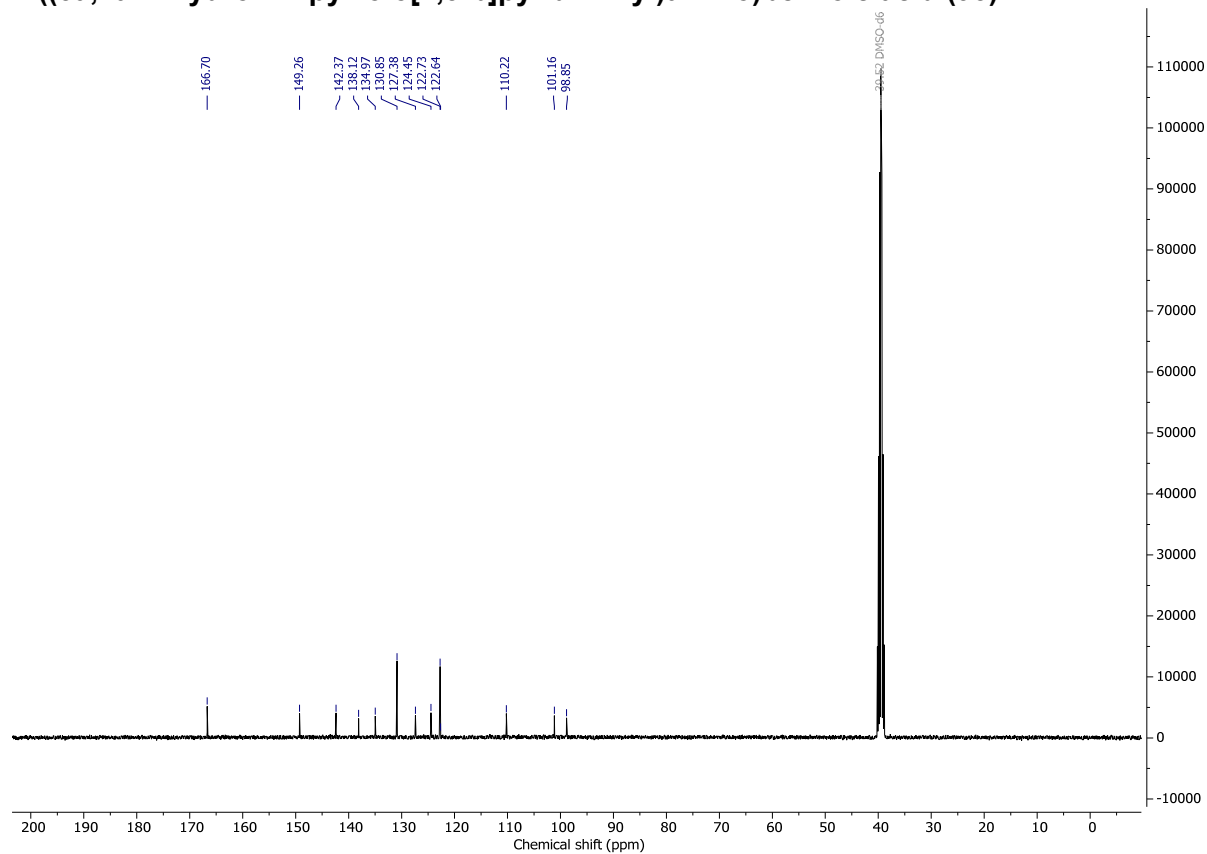
4-(Naphthalen-1-ylamino)benzoic acid (91)



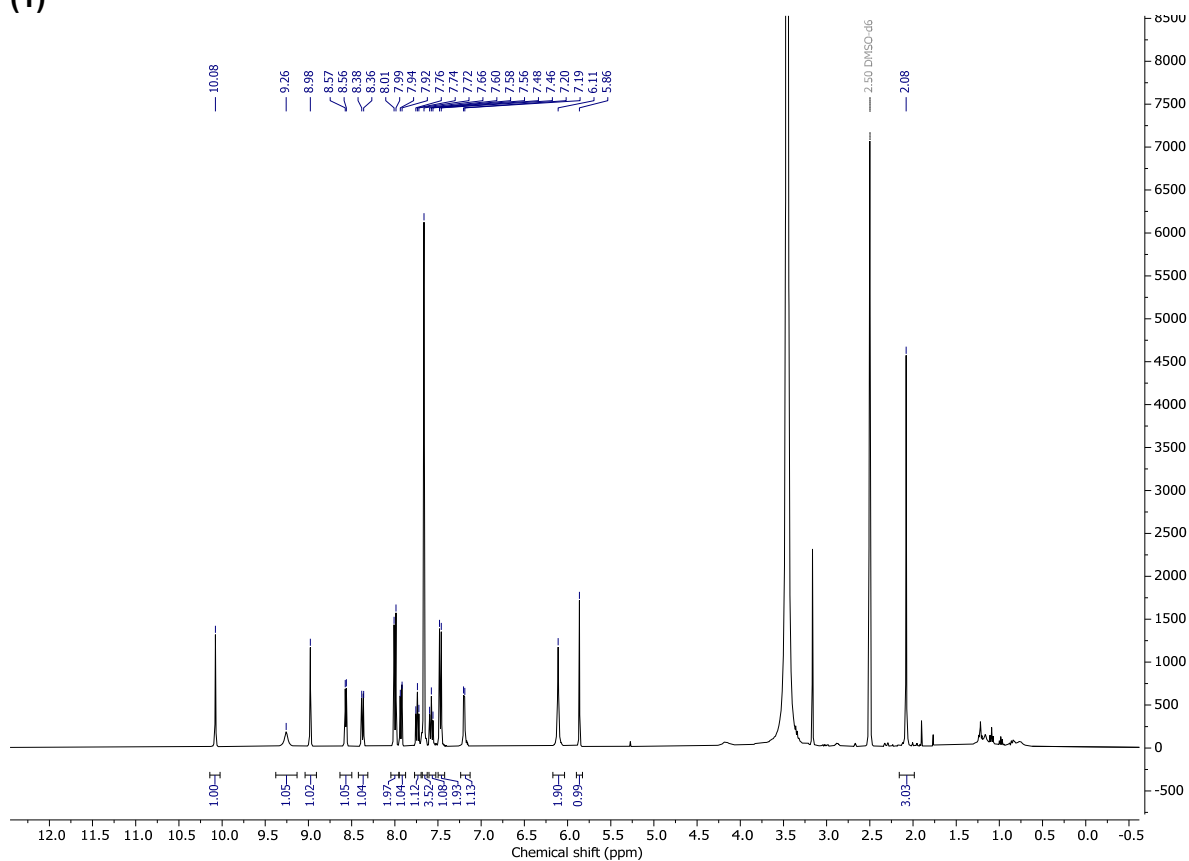
4-(Quinazolin-4-ylamino)benzoic acid (92)



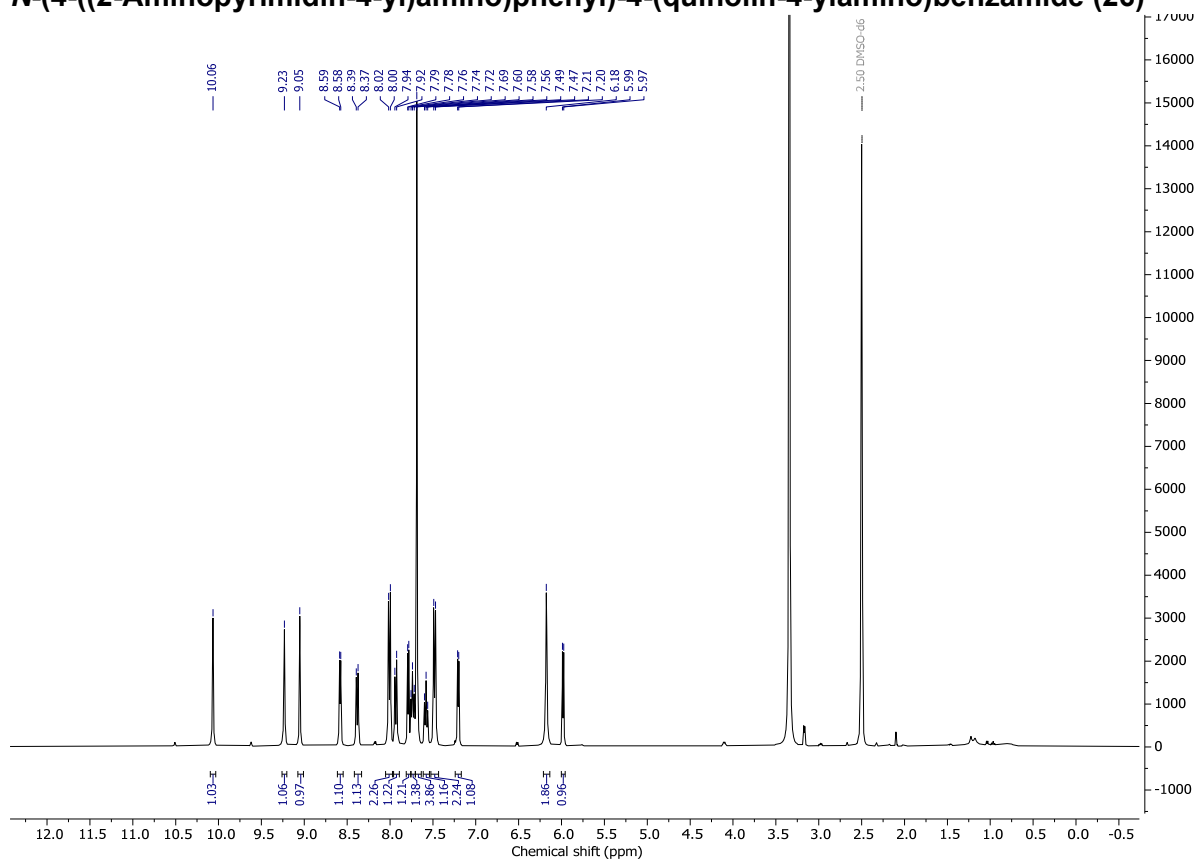
4-((3a,7a-Dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzoic acid (93)

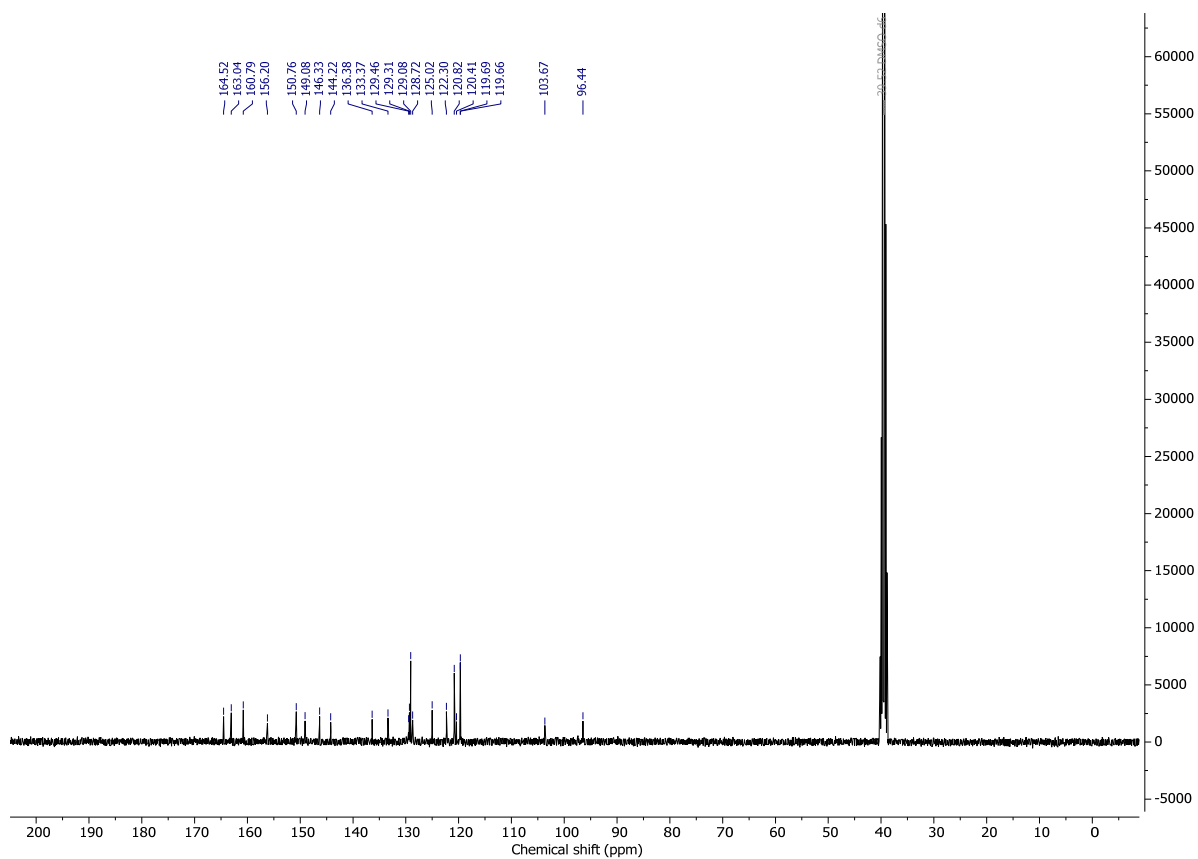


***N*-4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (1)**

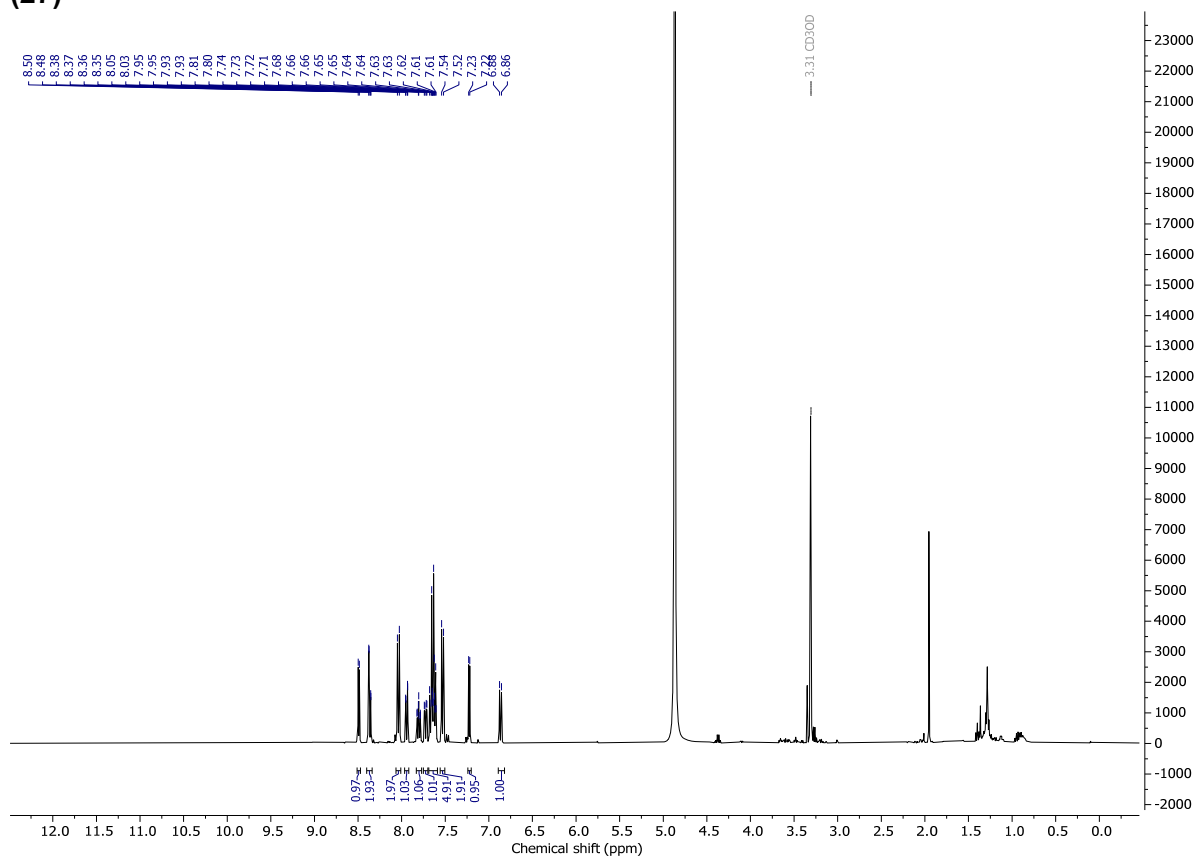


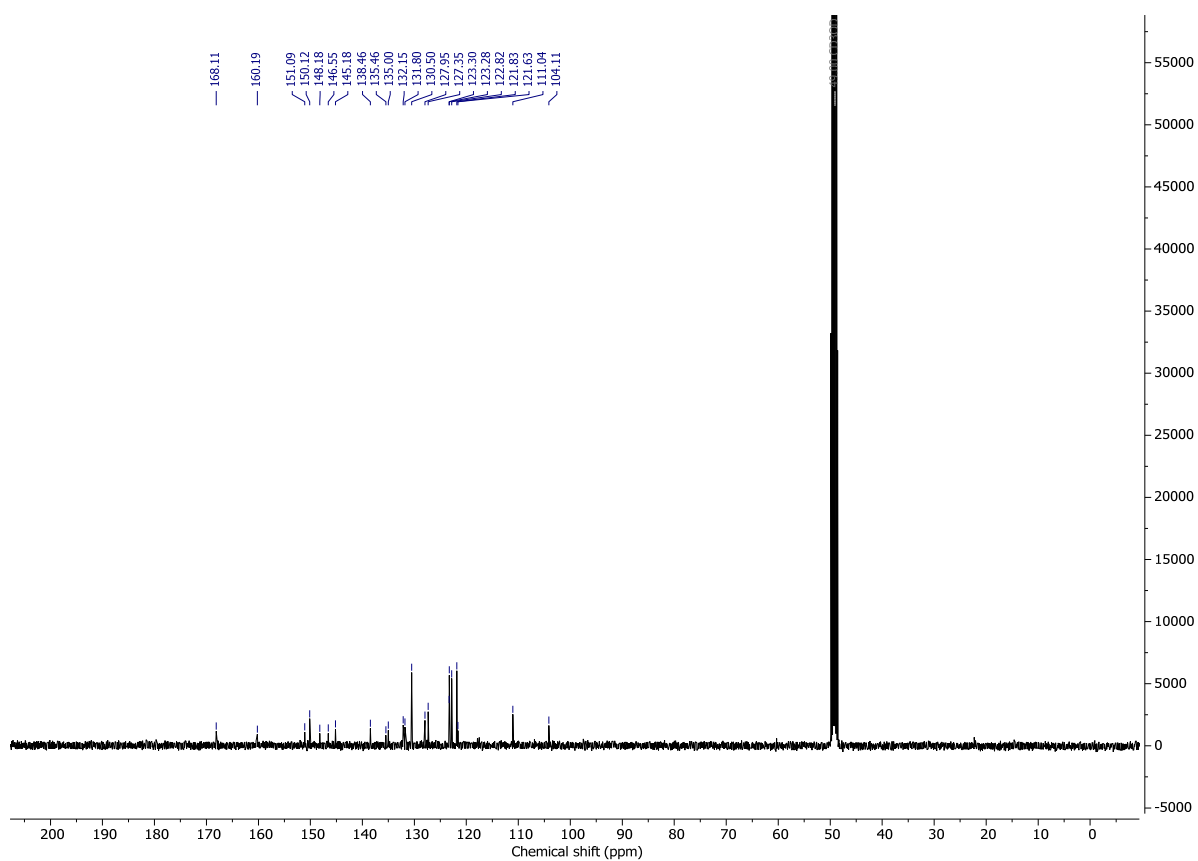
***N*-4-((2-Aminopyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (26)**



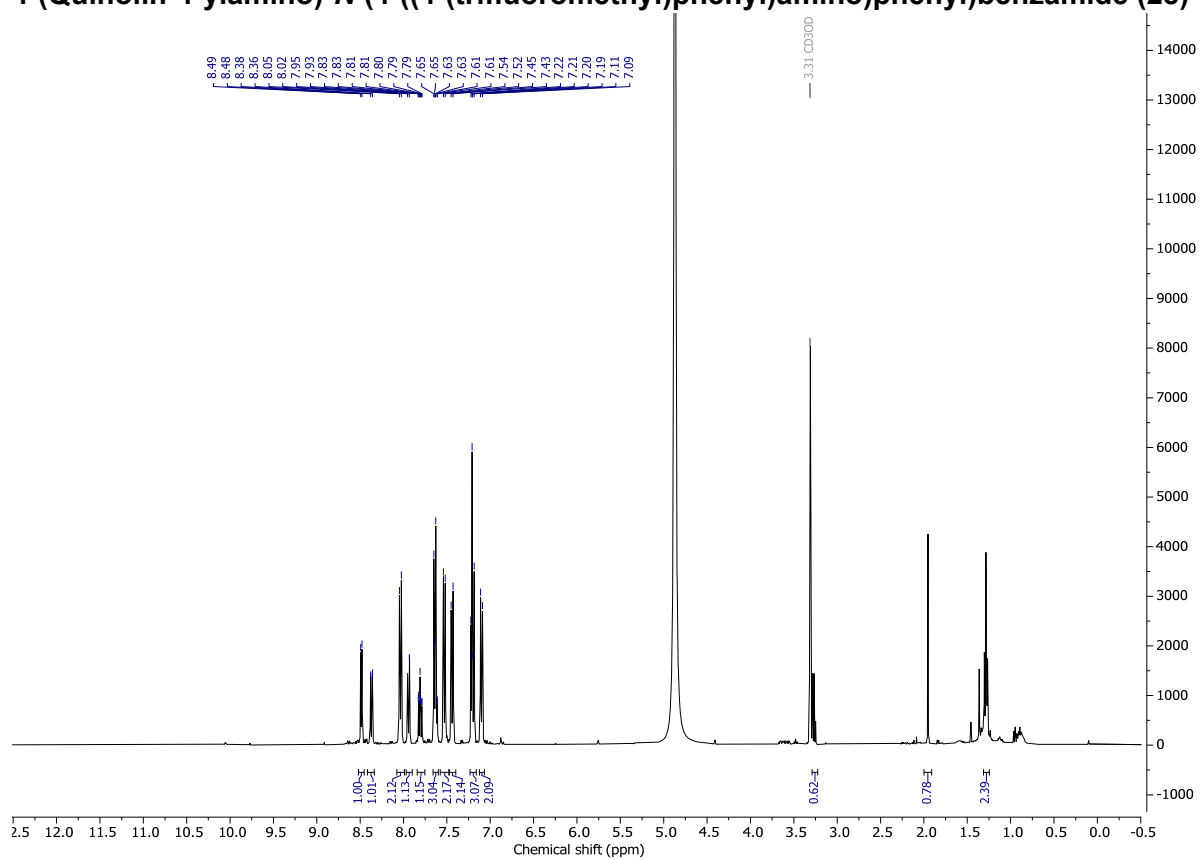


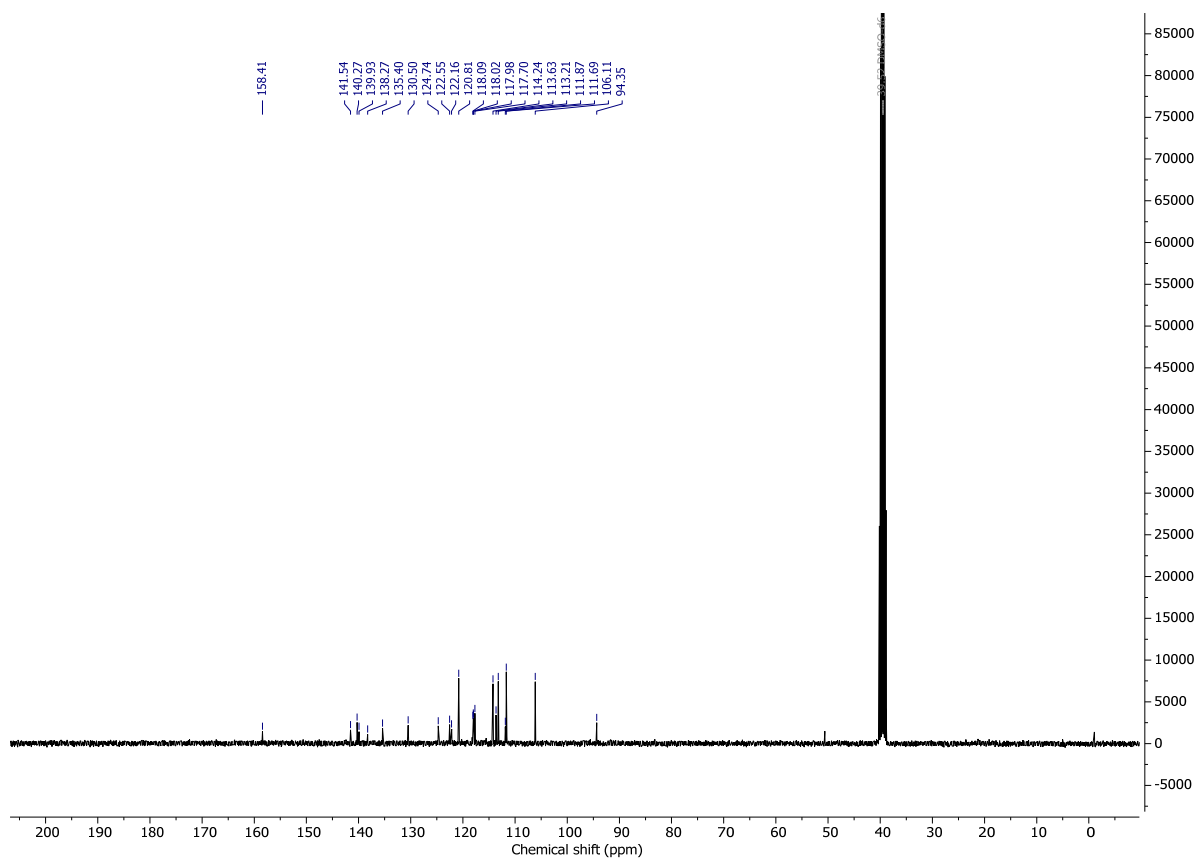
4-(Quinolin-4-ylamino)-N-(4-((5-(trifluoromethyl)pyridin-2-yl)amino)phenyl)benzamide (27)



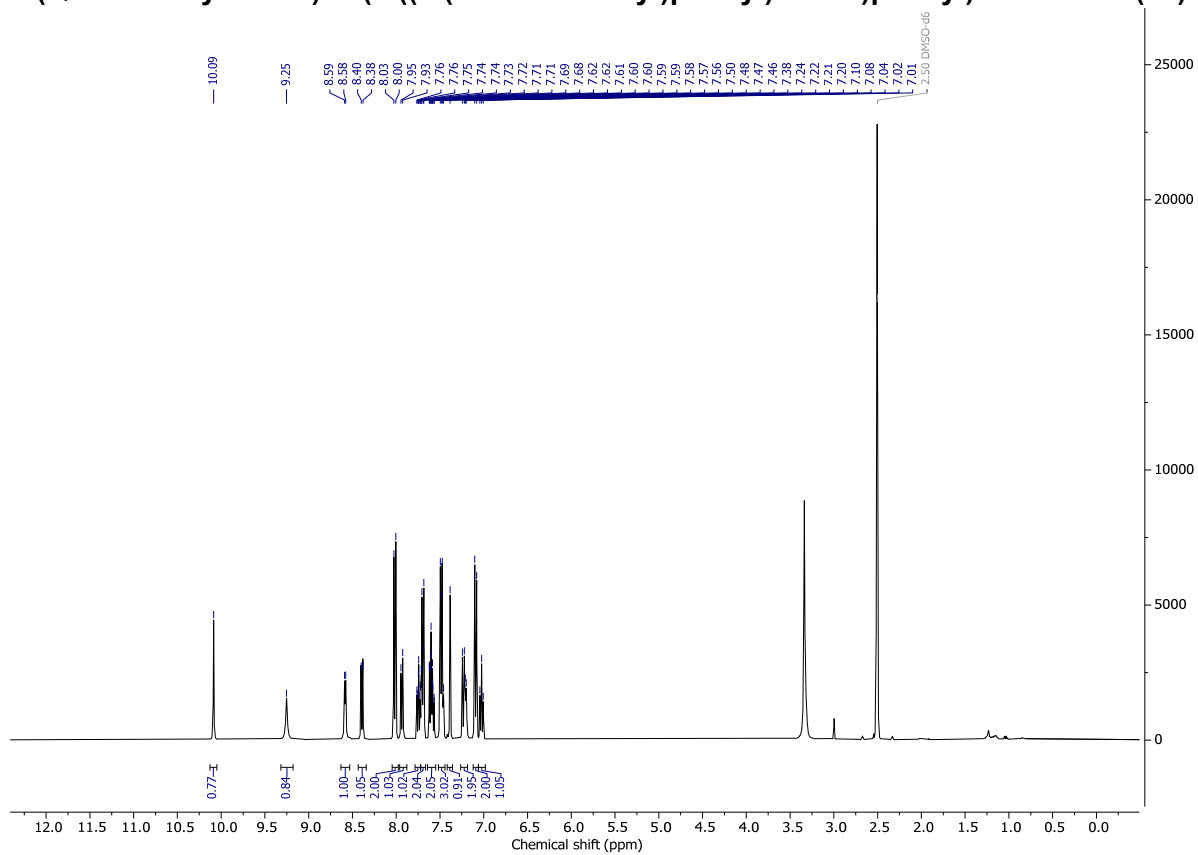


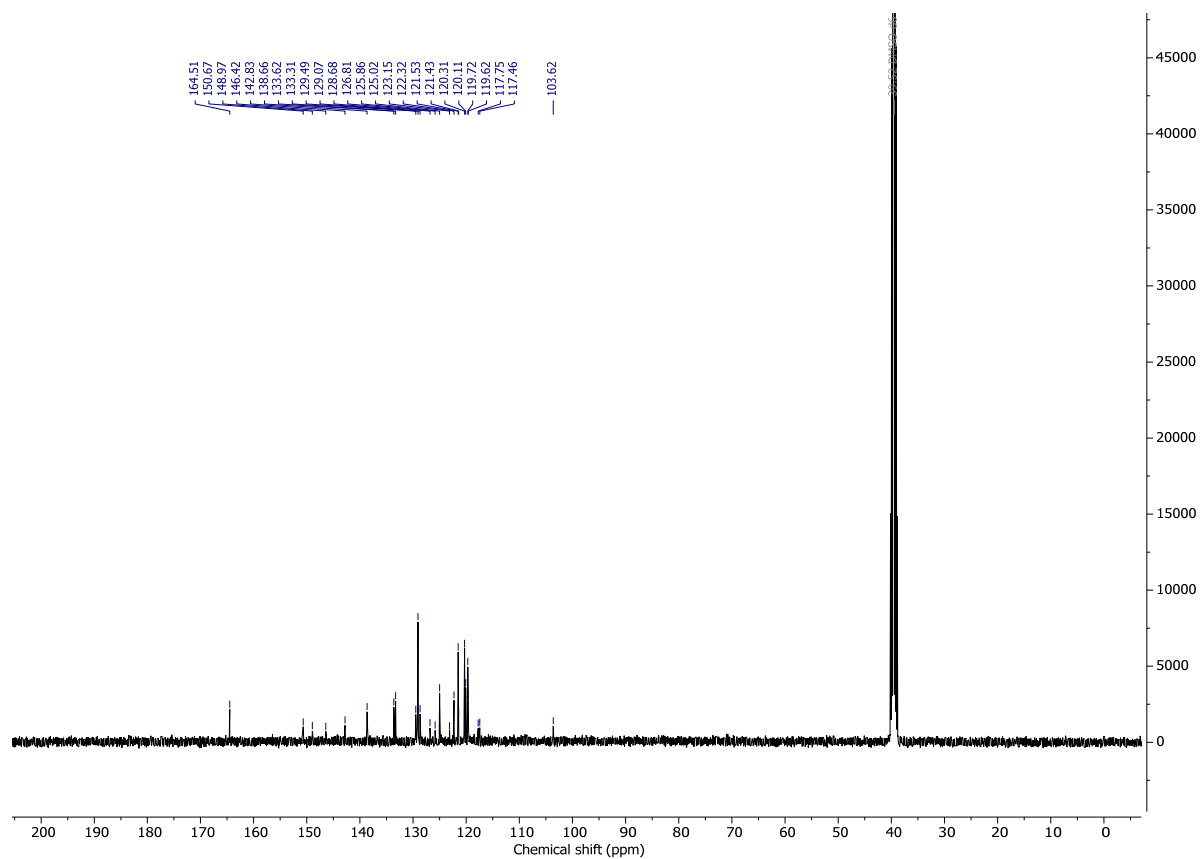
4-(Quinolin-4-ylamino)-N-(4-((4-(trifluoromethyl)phenyl)amino)phenyl)benzamide (28)



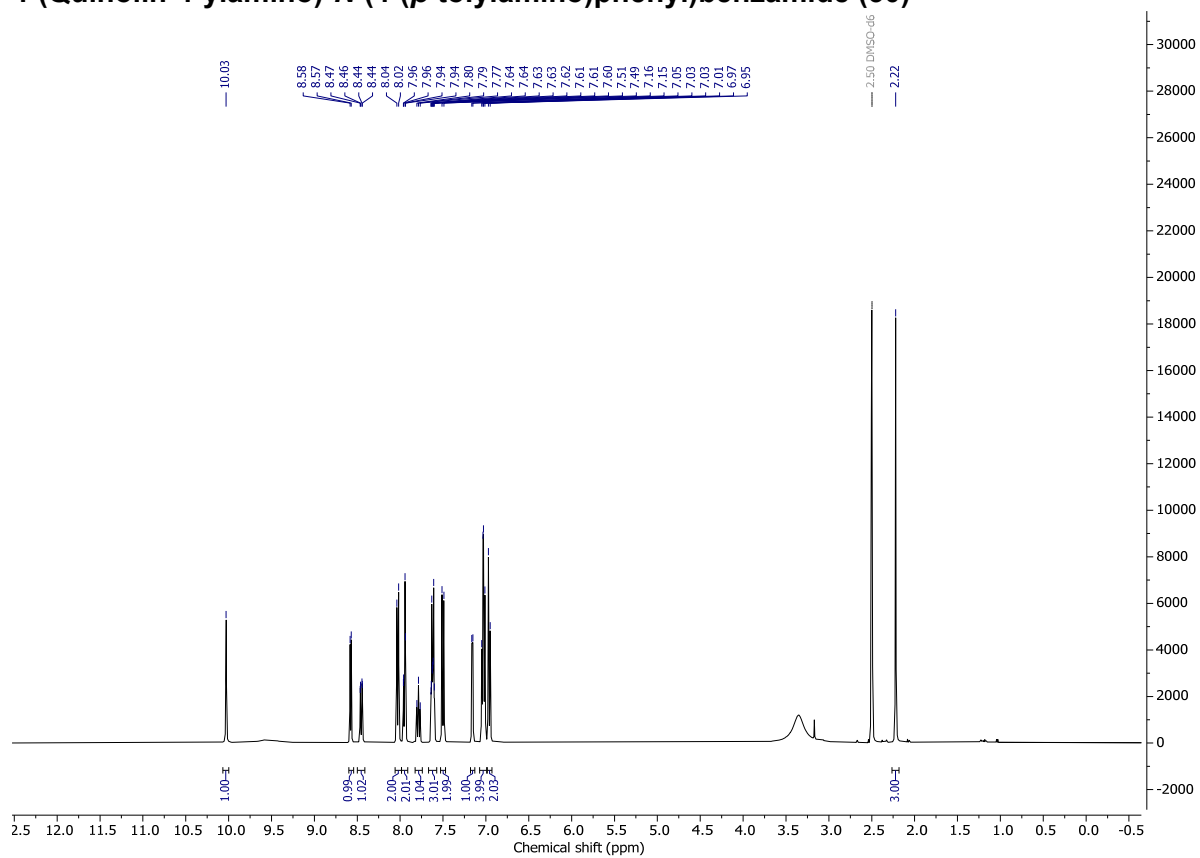


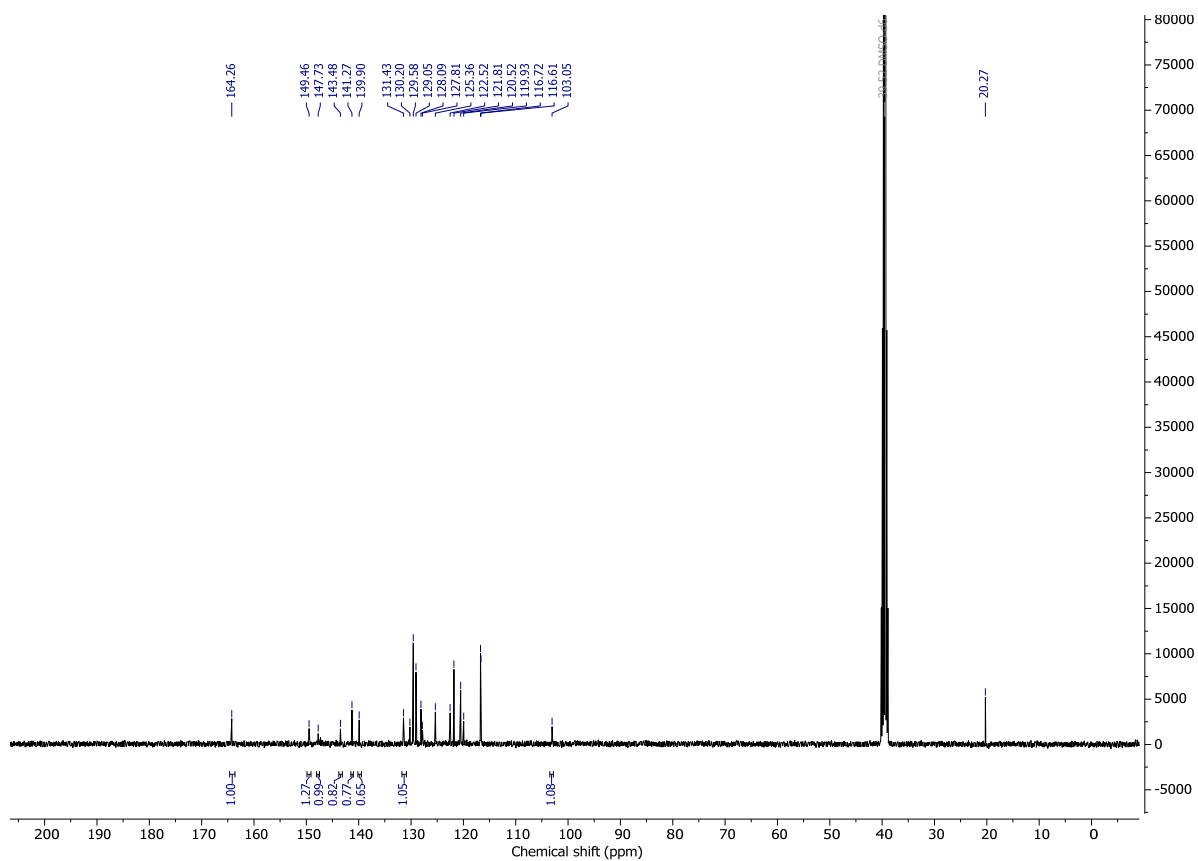
4-(Quinolin-4-ylamino)-N-(4-((2-(trifluoromethyl)phenyl)amino)phenyl)benzamide (29)



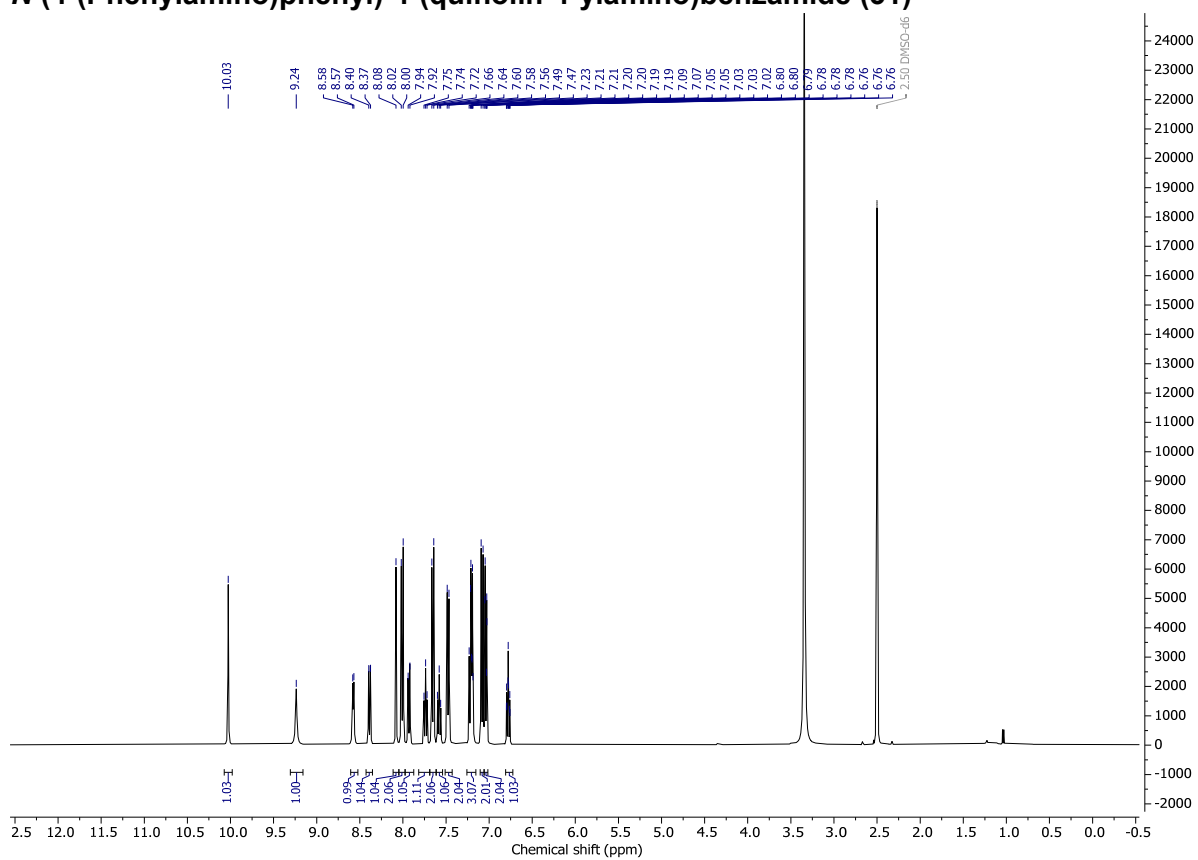


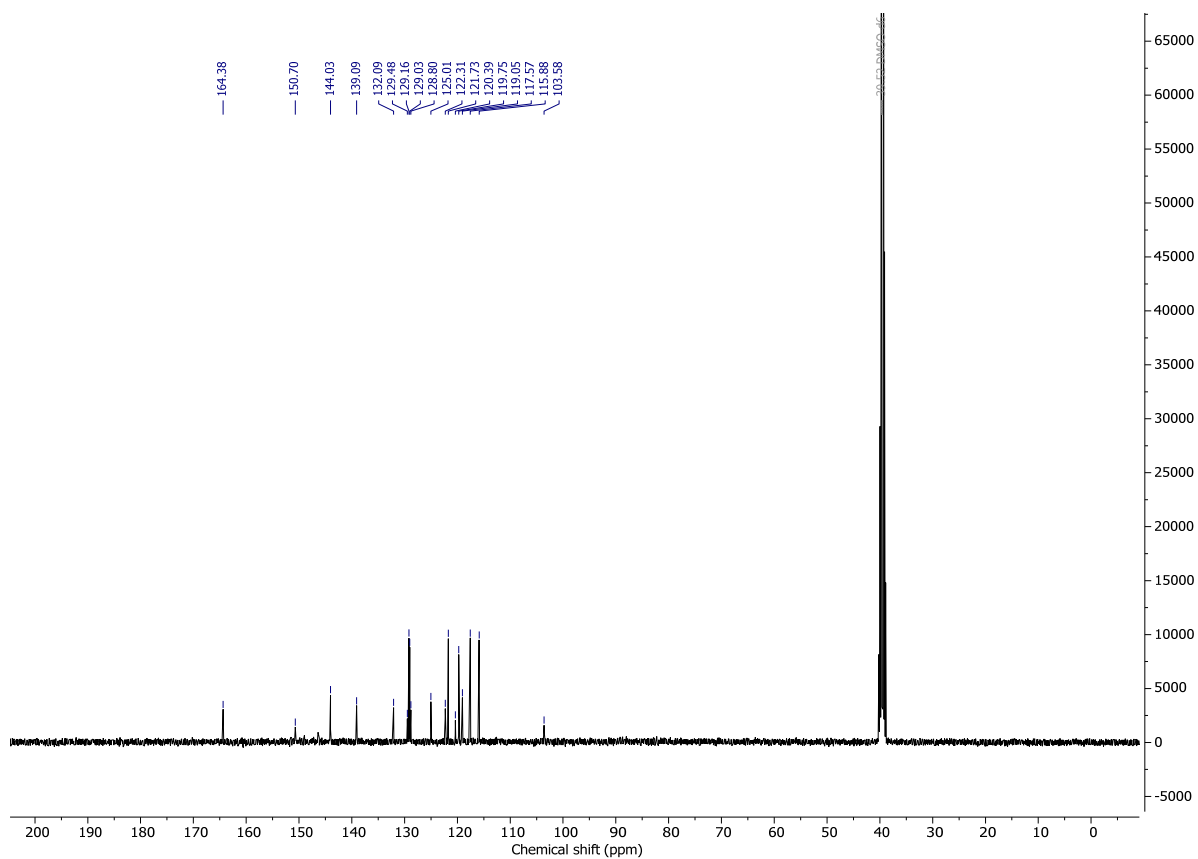
4-(Quinolin-4-ylamino)-N-(4-(p-tolylamino)phenyl)benzamide (30)



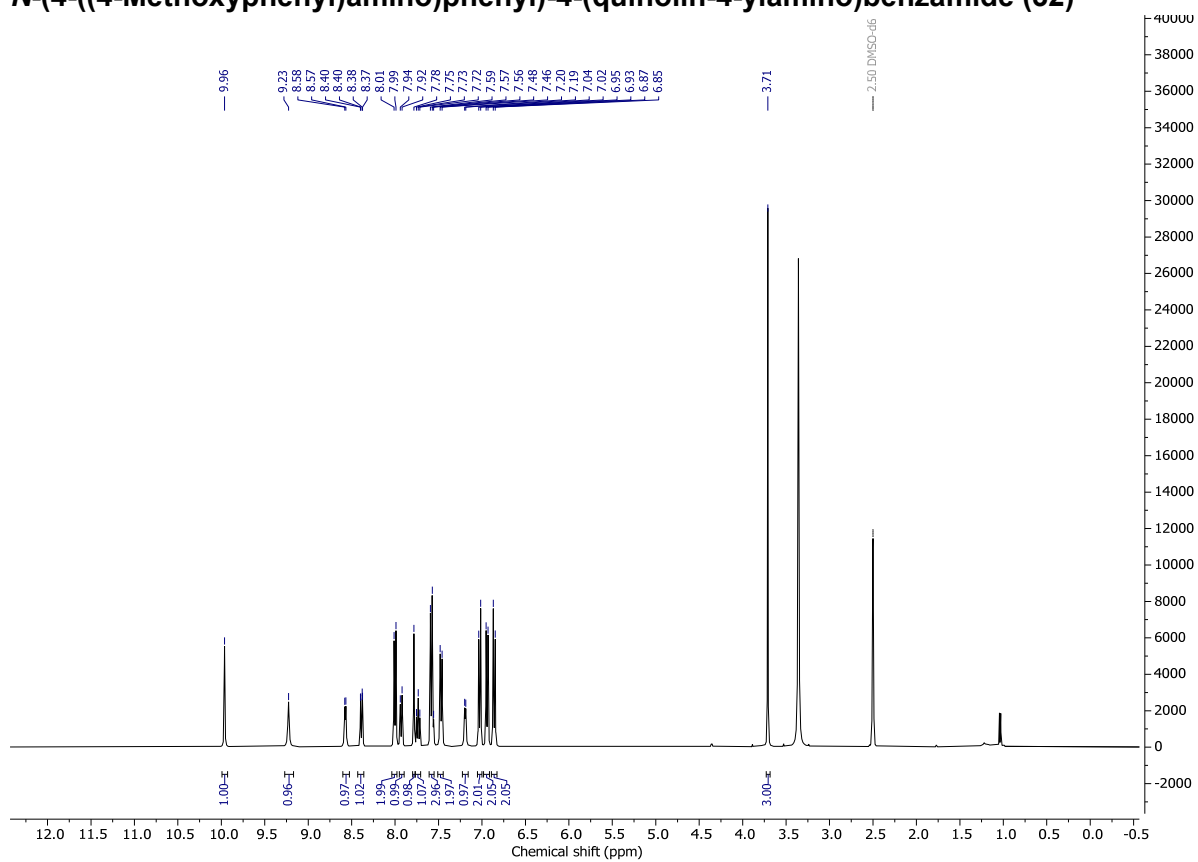


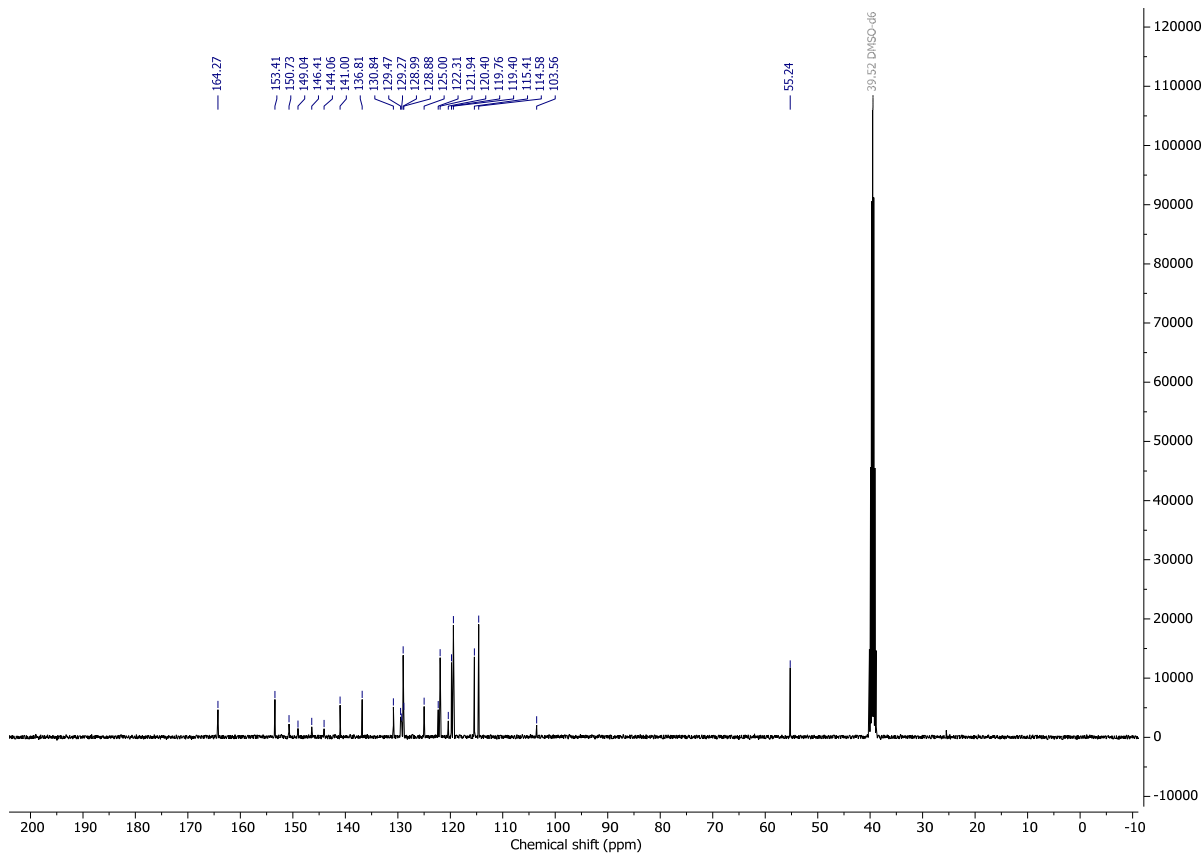
***N*-(4-(Phenylamino)phenyl)-4-(quinolin-4-ylamino)benzamide (31)**



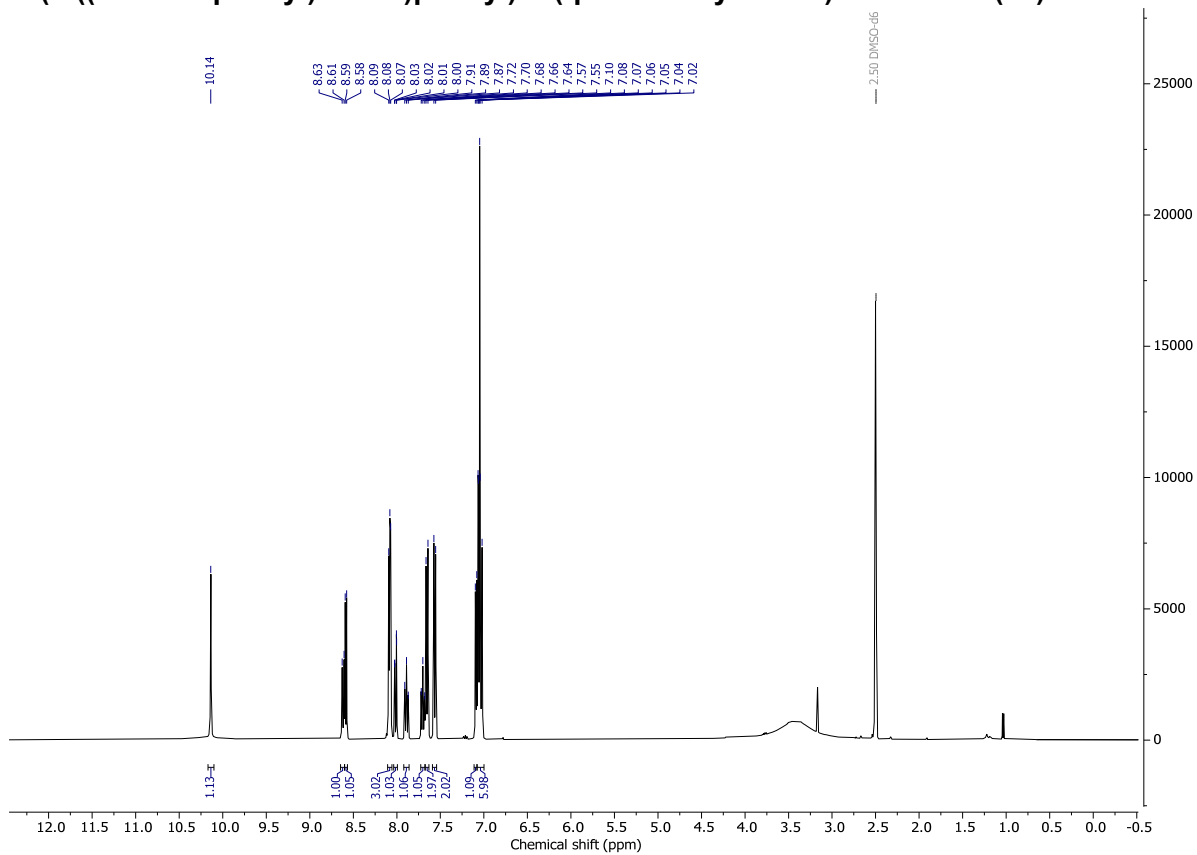


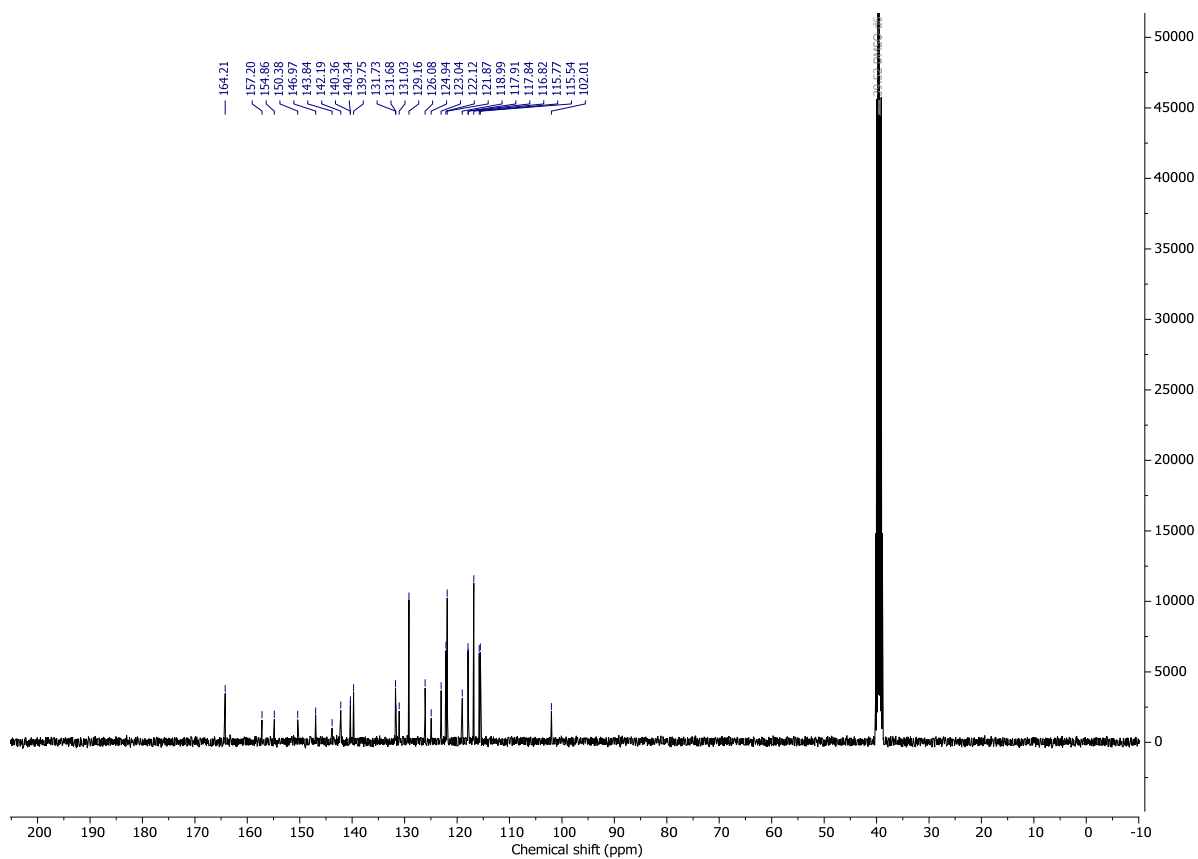
***N*-(4-((4-Methoxyphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (32)**



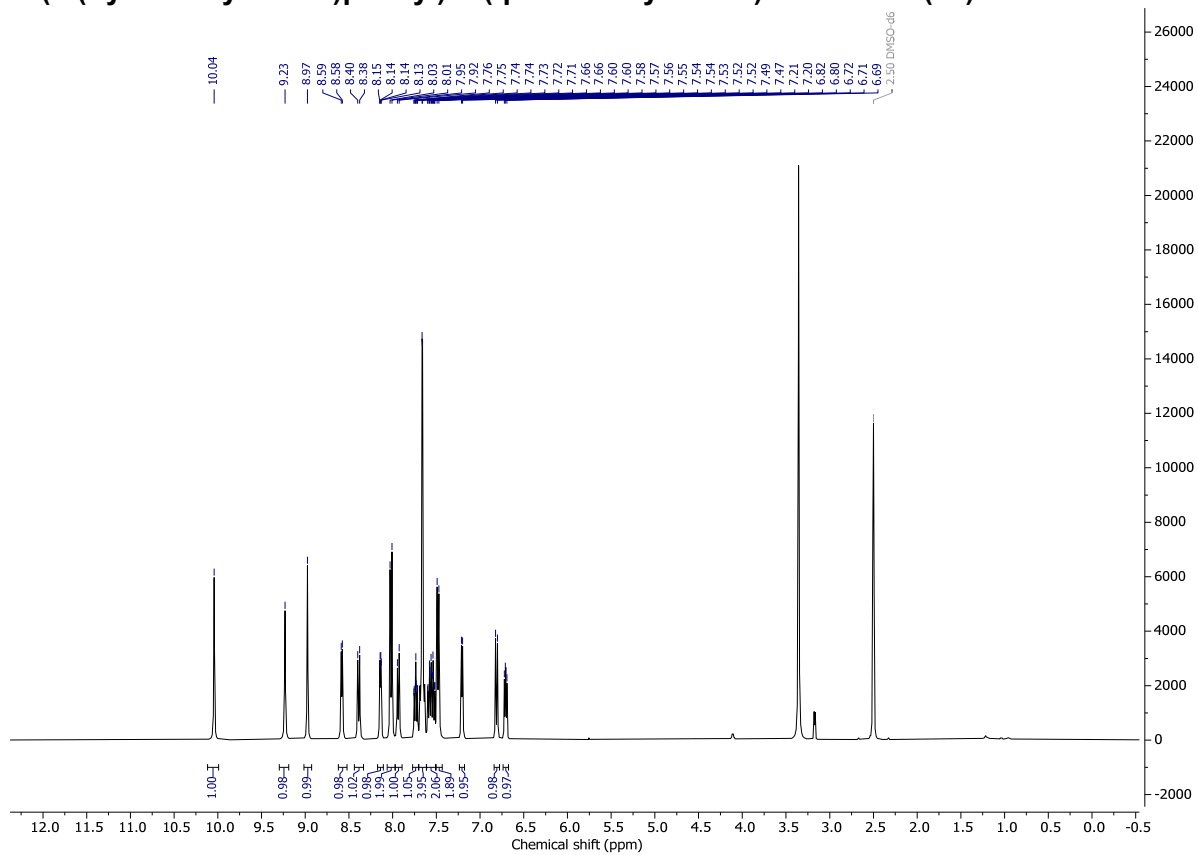


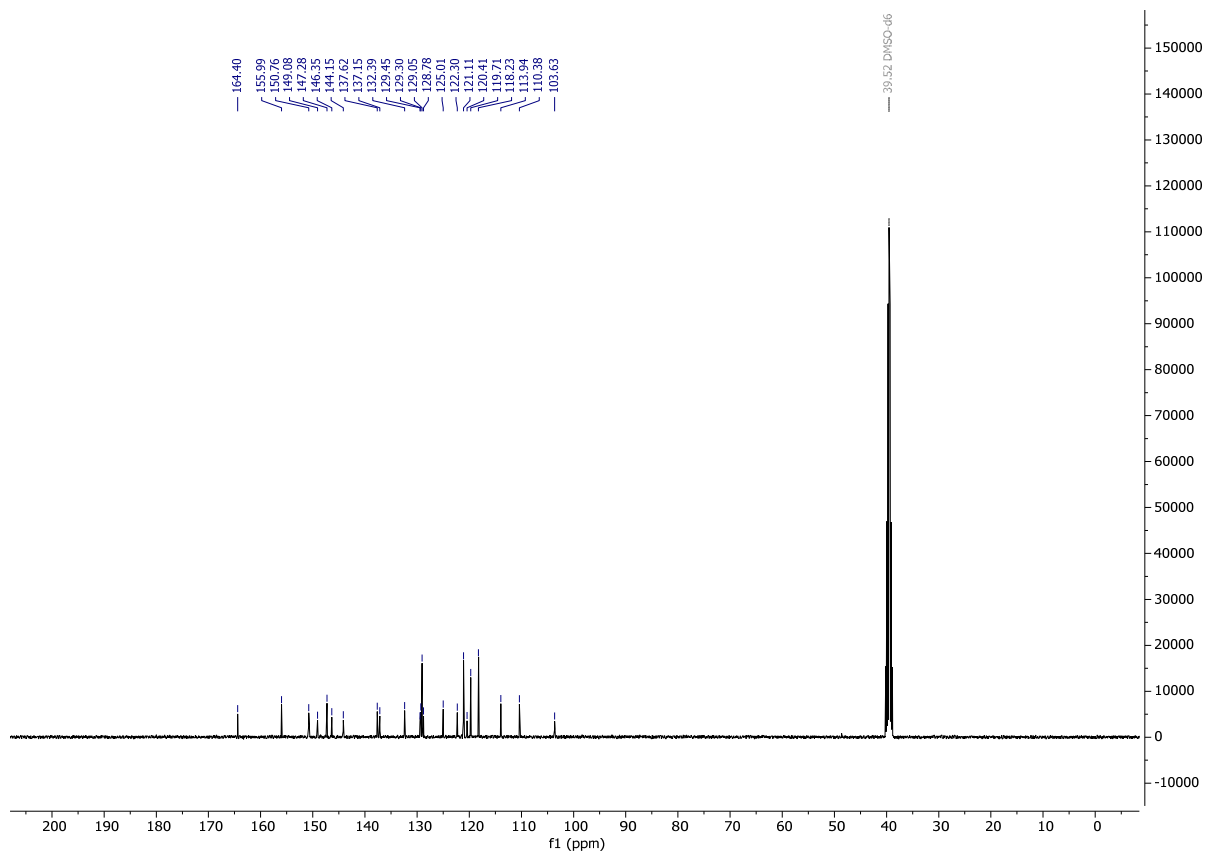
N-(4-((4-Fluorophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (33)



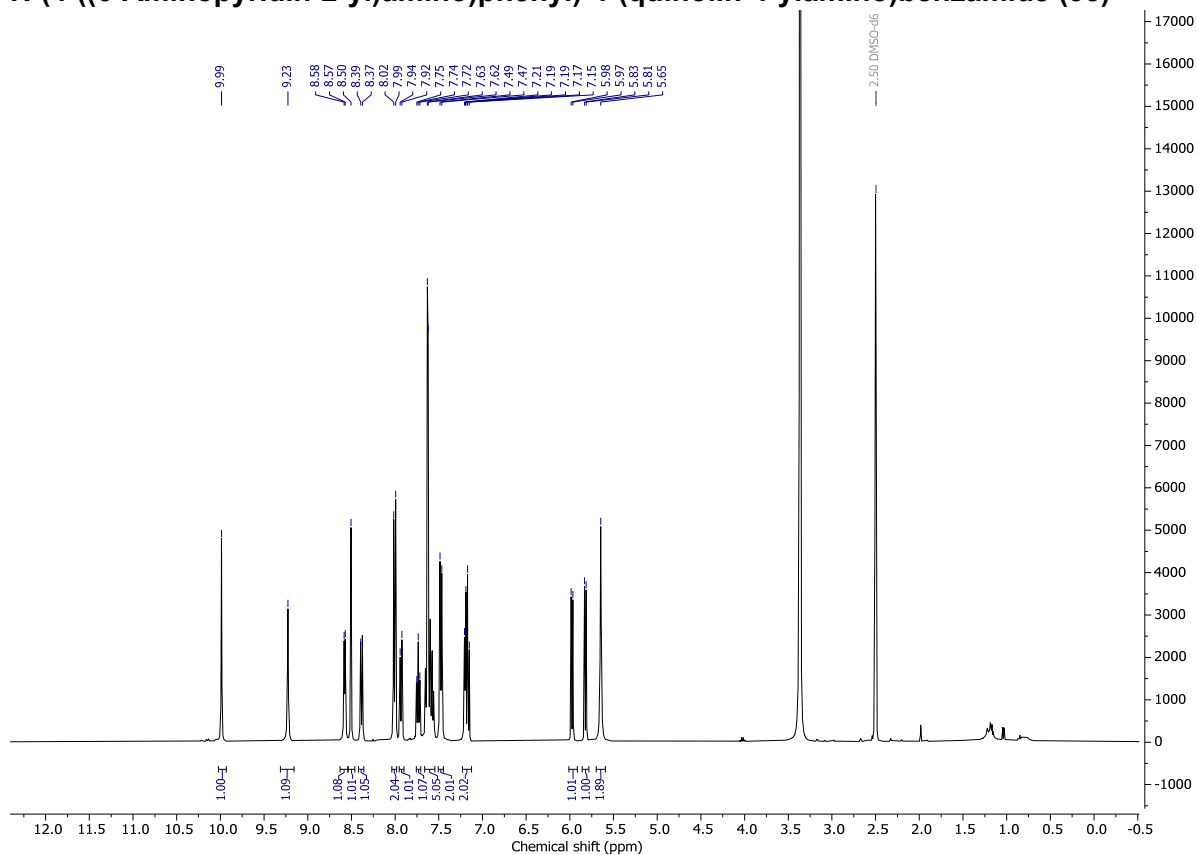


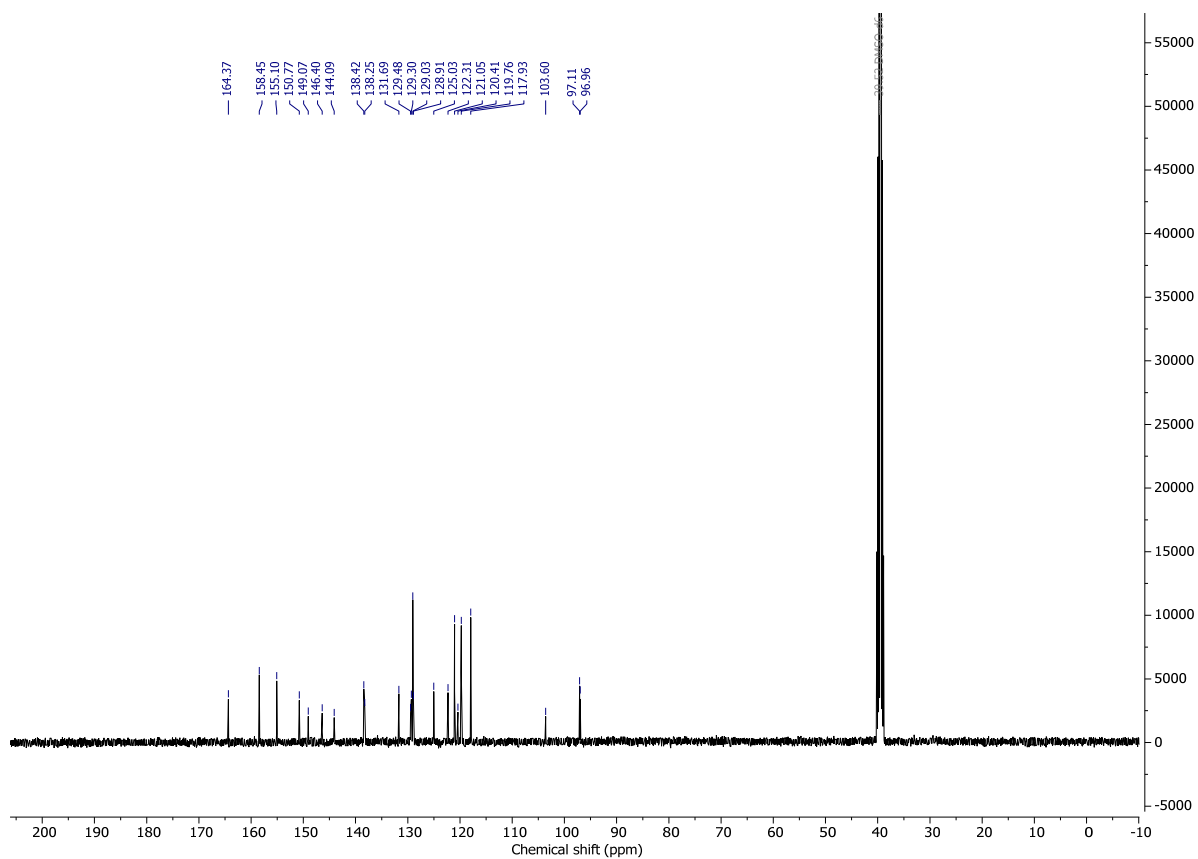
***N*-(4-(Pyridin-2-ylamino)phenyl)-4-(quinolin-4-ylamino)benzamide (34)**



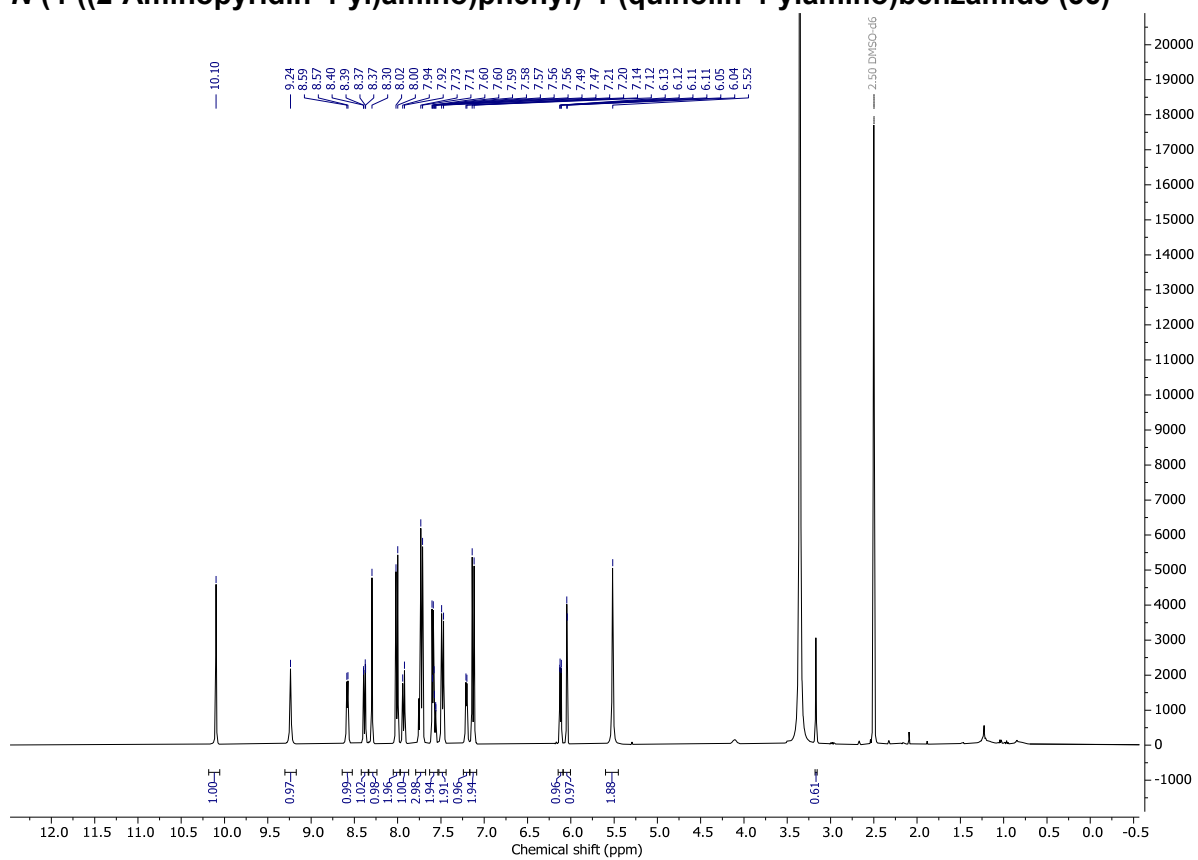


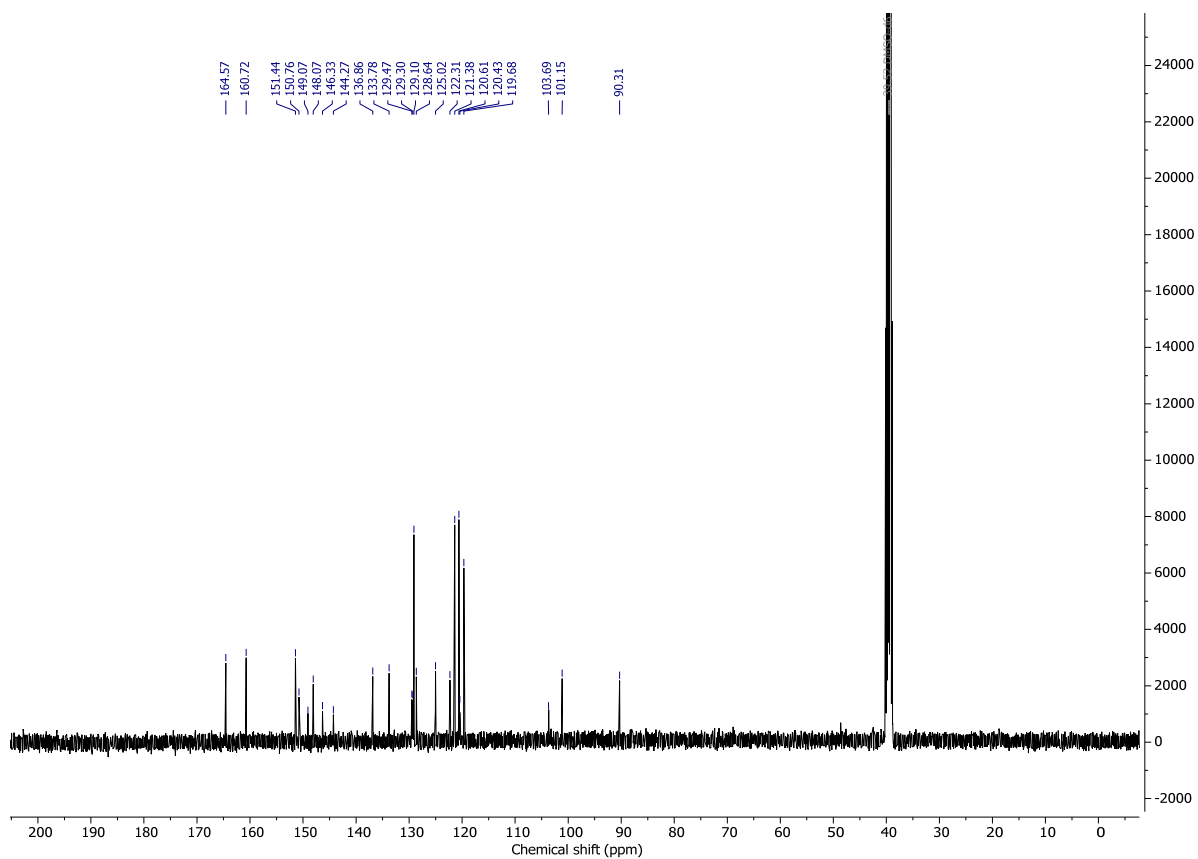
***N*-(4-((6-Aminopyridin-2-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (35)**



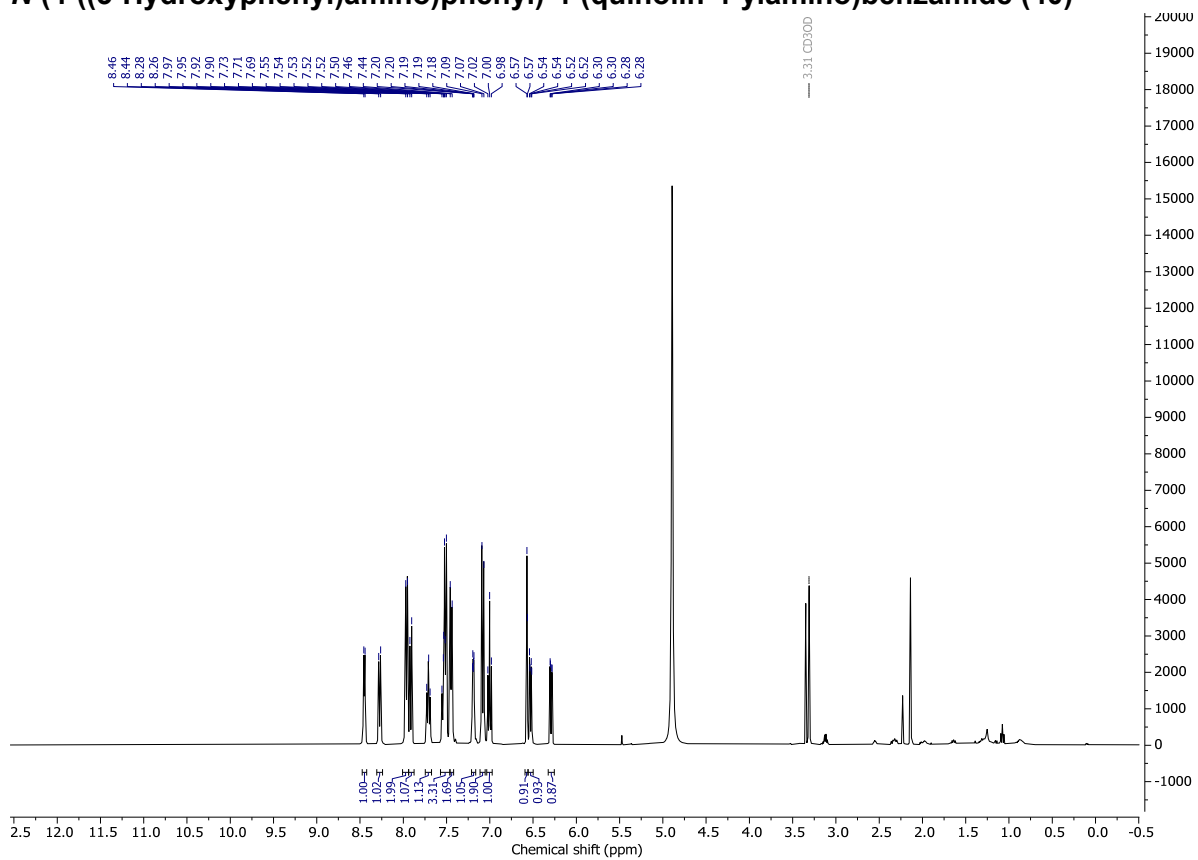


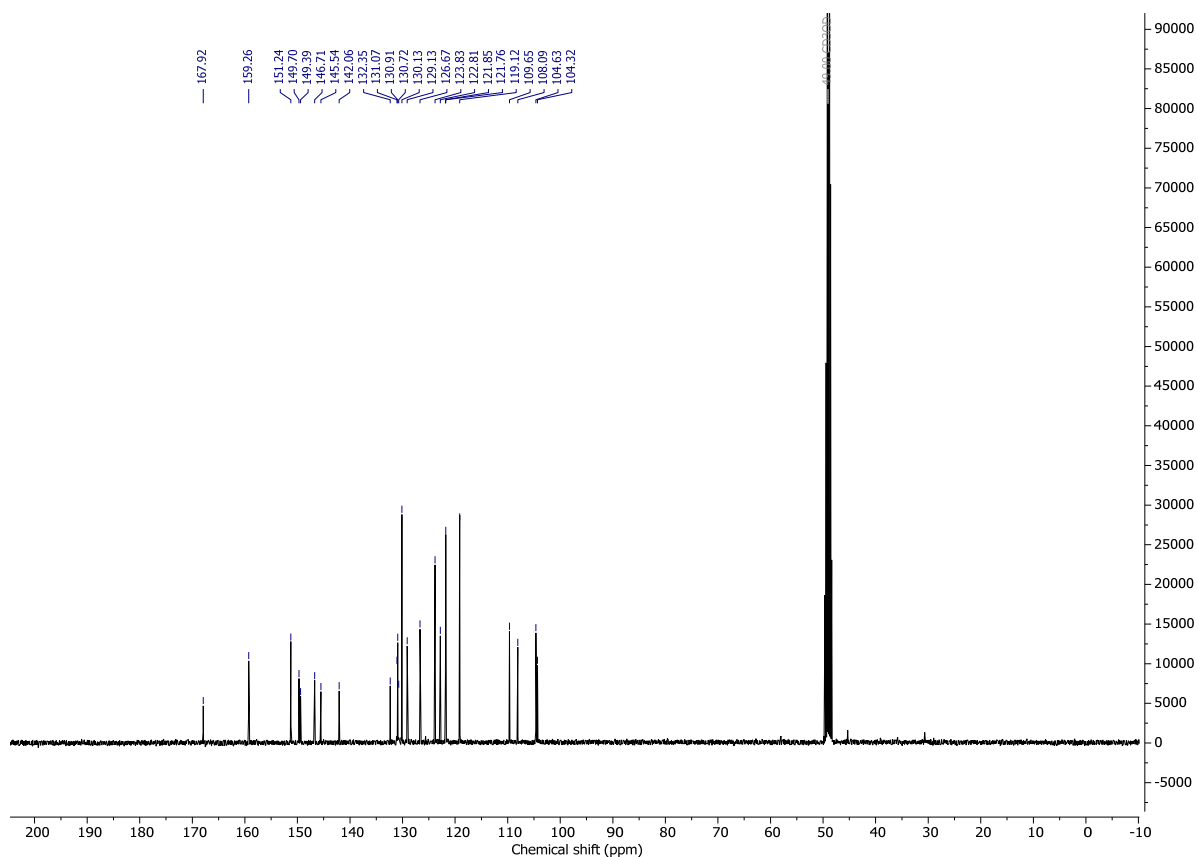
***N*-(4-((2-Aminopyridin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (36)**



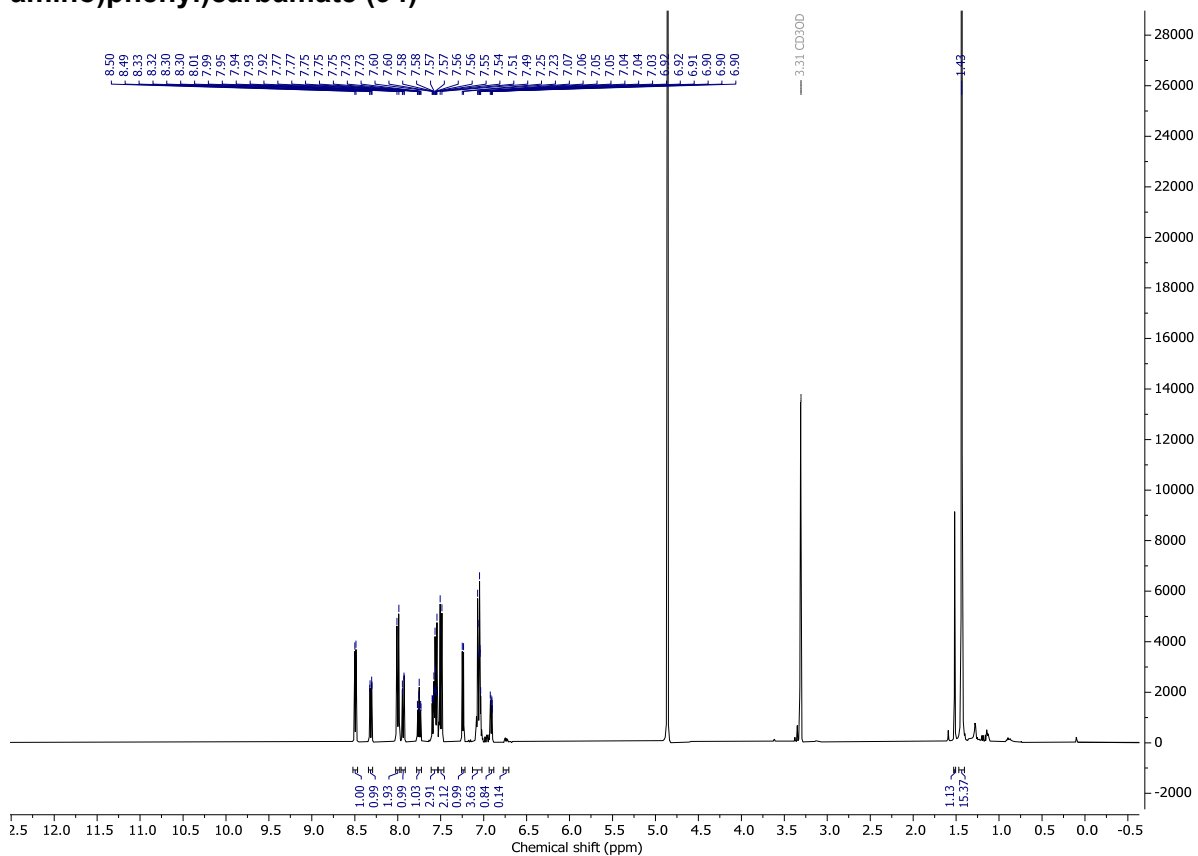


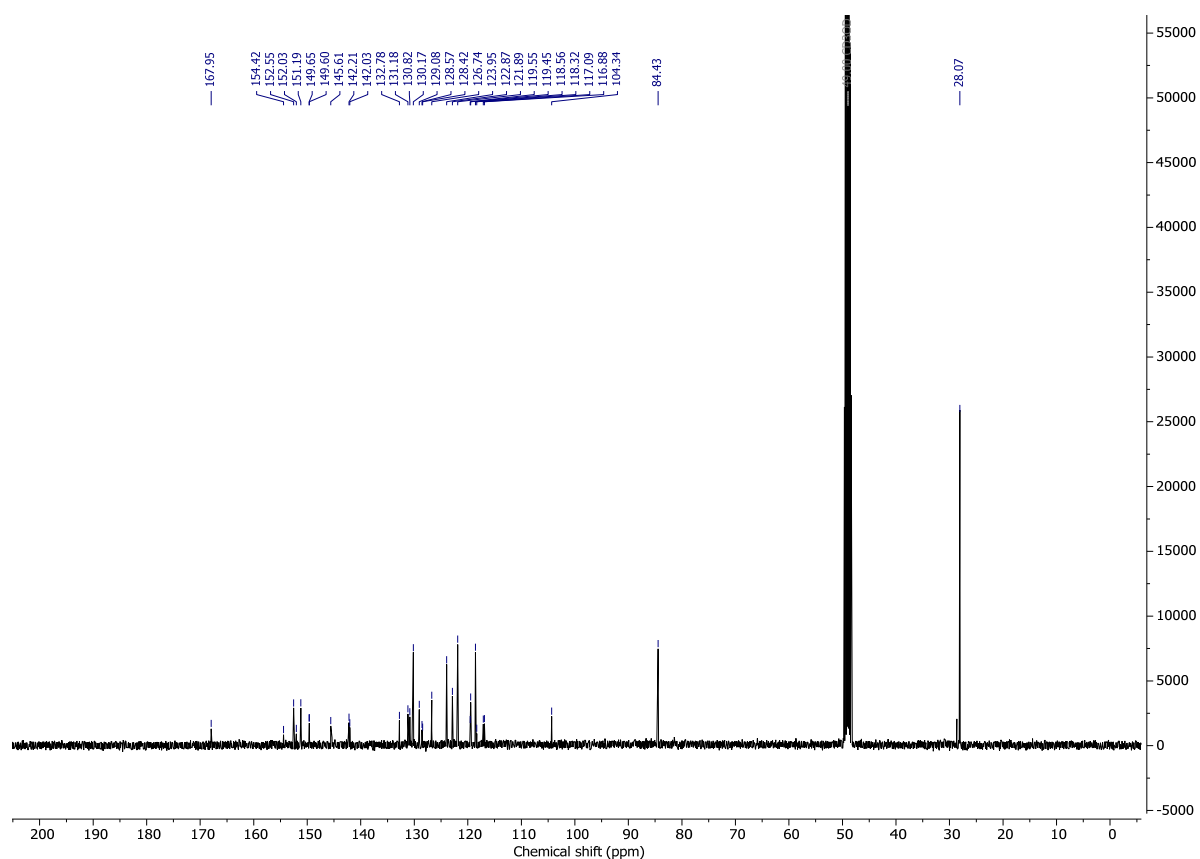
***N*-(4-((3-Hydroxyphenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (40)**



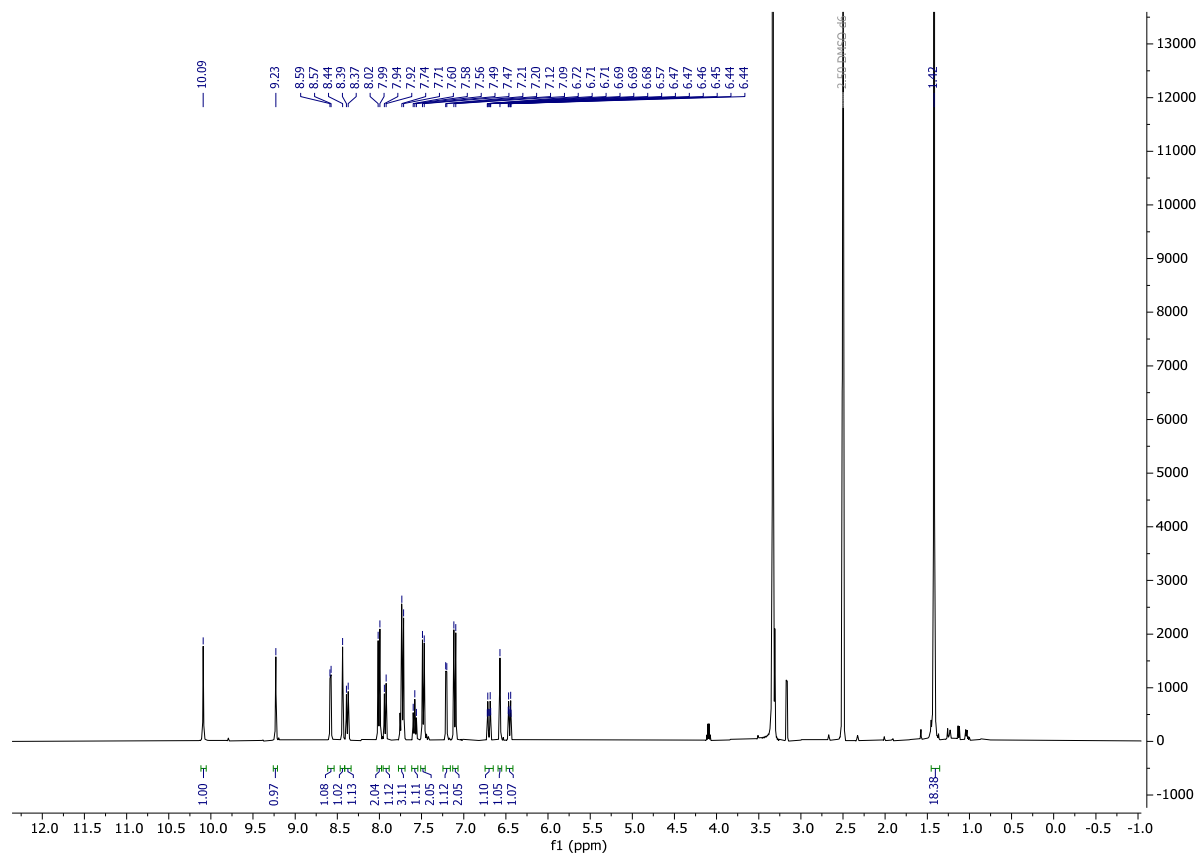


***tert*-Butyl (*tert*-butoxycarbonyl)(2-fluoro-5-((4-(4-(quinolin-4-ylamino)benzamido)phenyl)amino)phenyl)carbamate (94)**

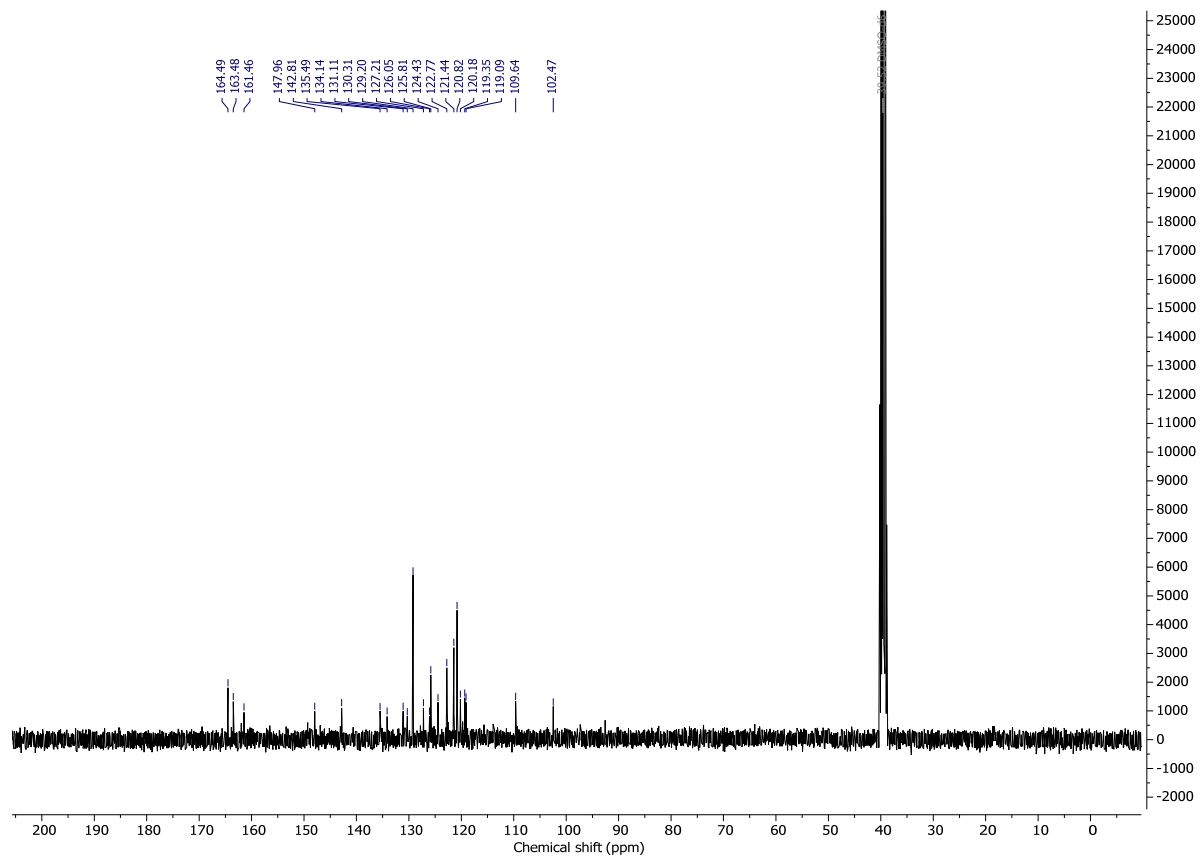
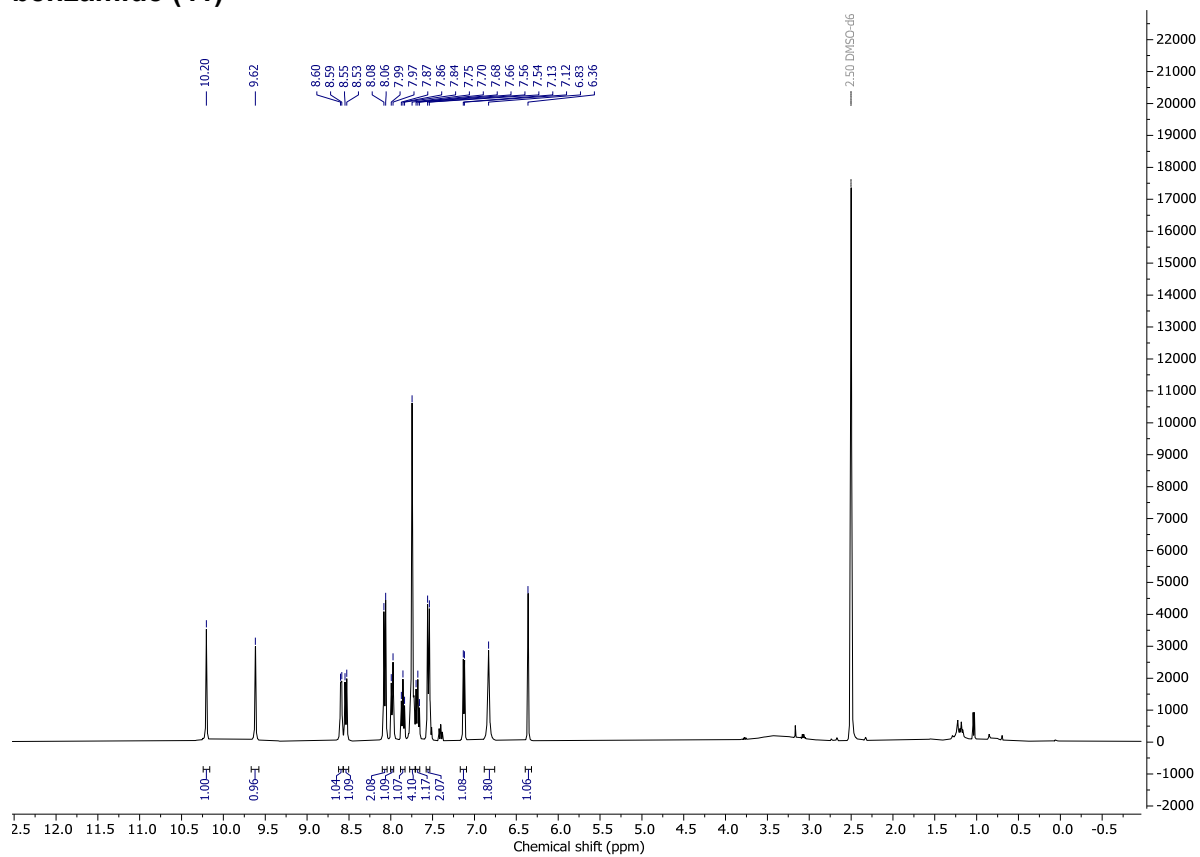


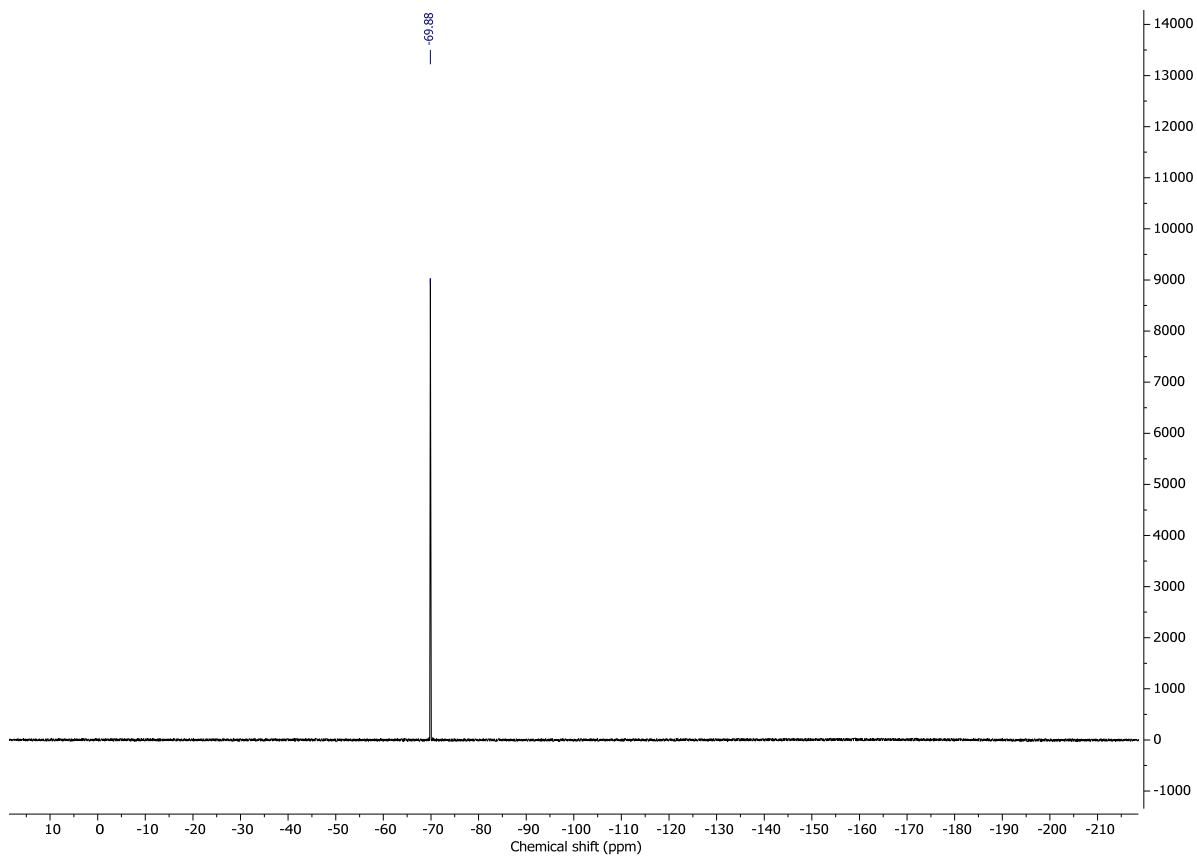


***tert*-Butyl (*tert*-butoxycarbonyl)(3-fluoro-5-((4-(4-(quinolin-4-ylamino)benzamido)phenyl)amino)phenyl)carbamate (95)**

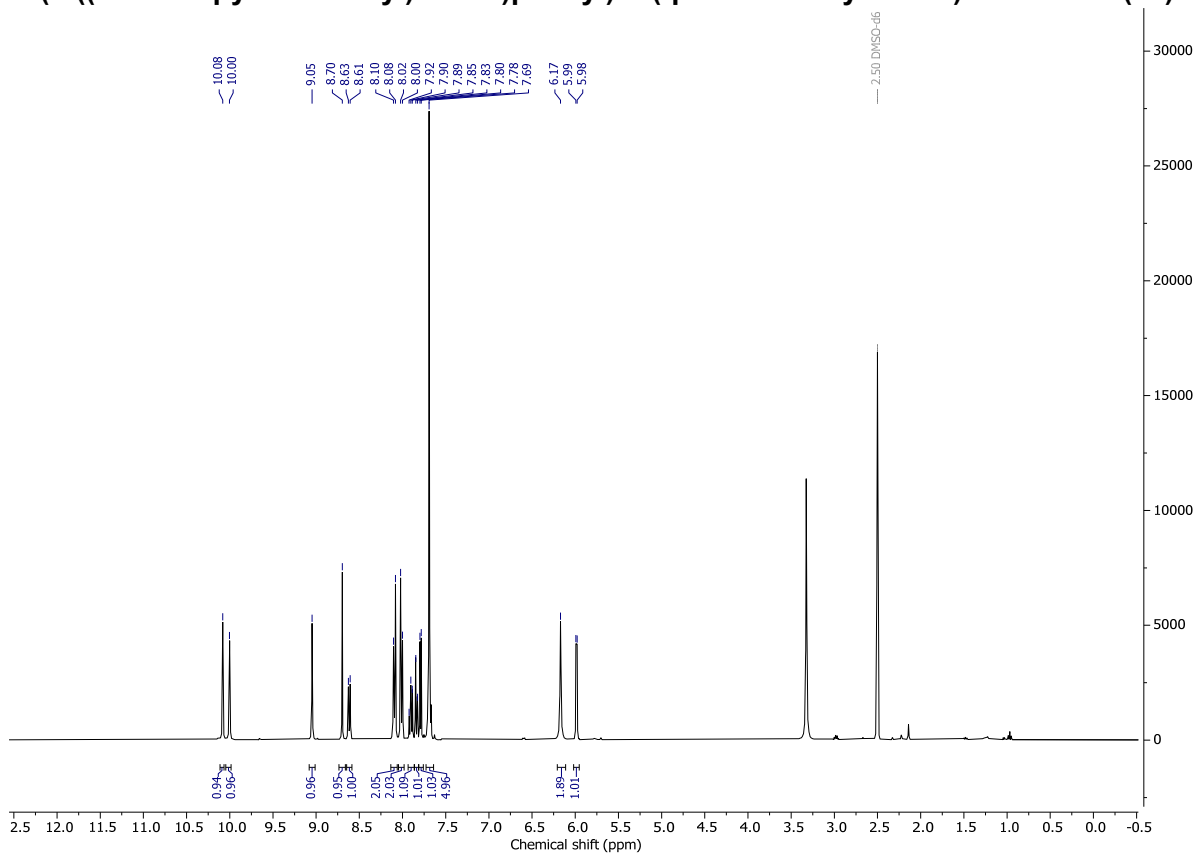


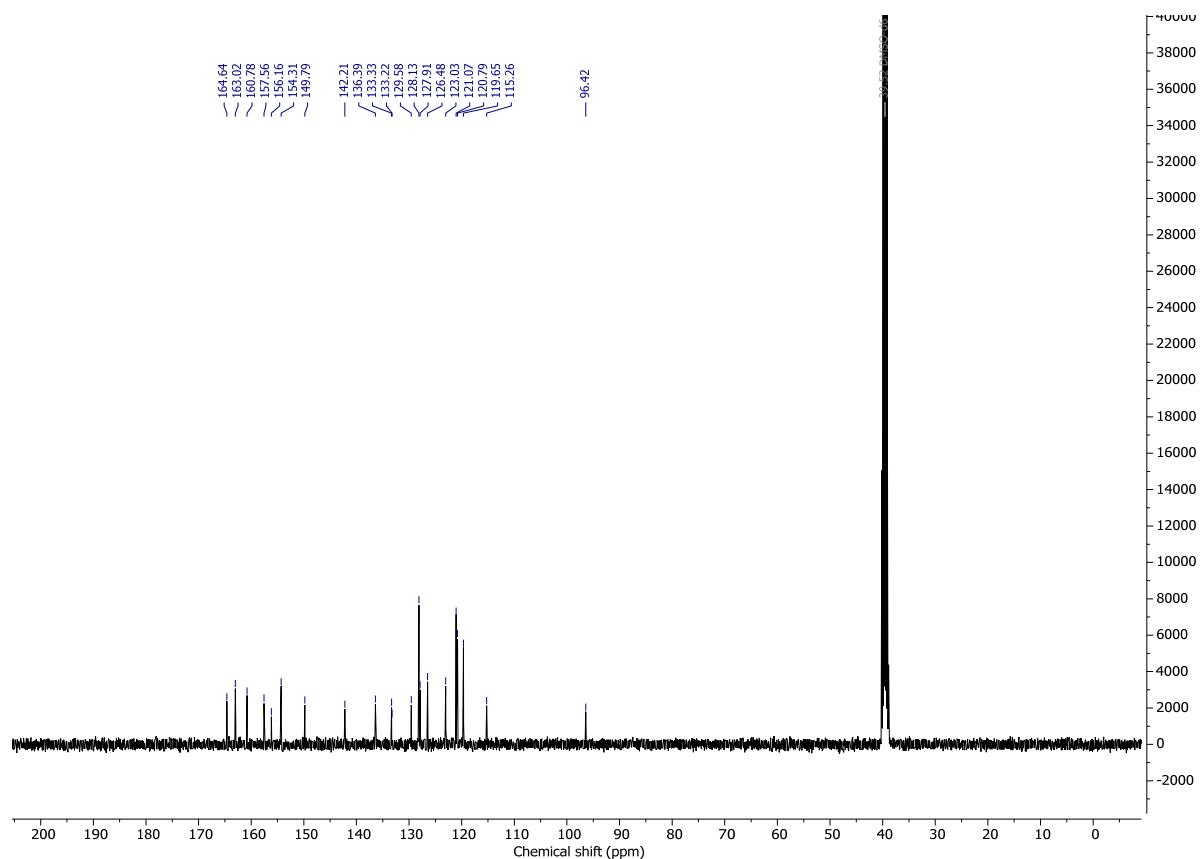
***N*-[4-((2-Amino-6-(trifluoromethyl)pyrimidin-4-yl)amino)phenyl]-4-(quinolin-4-ylamino)benzamide (41)**



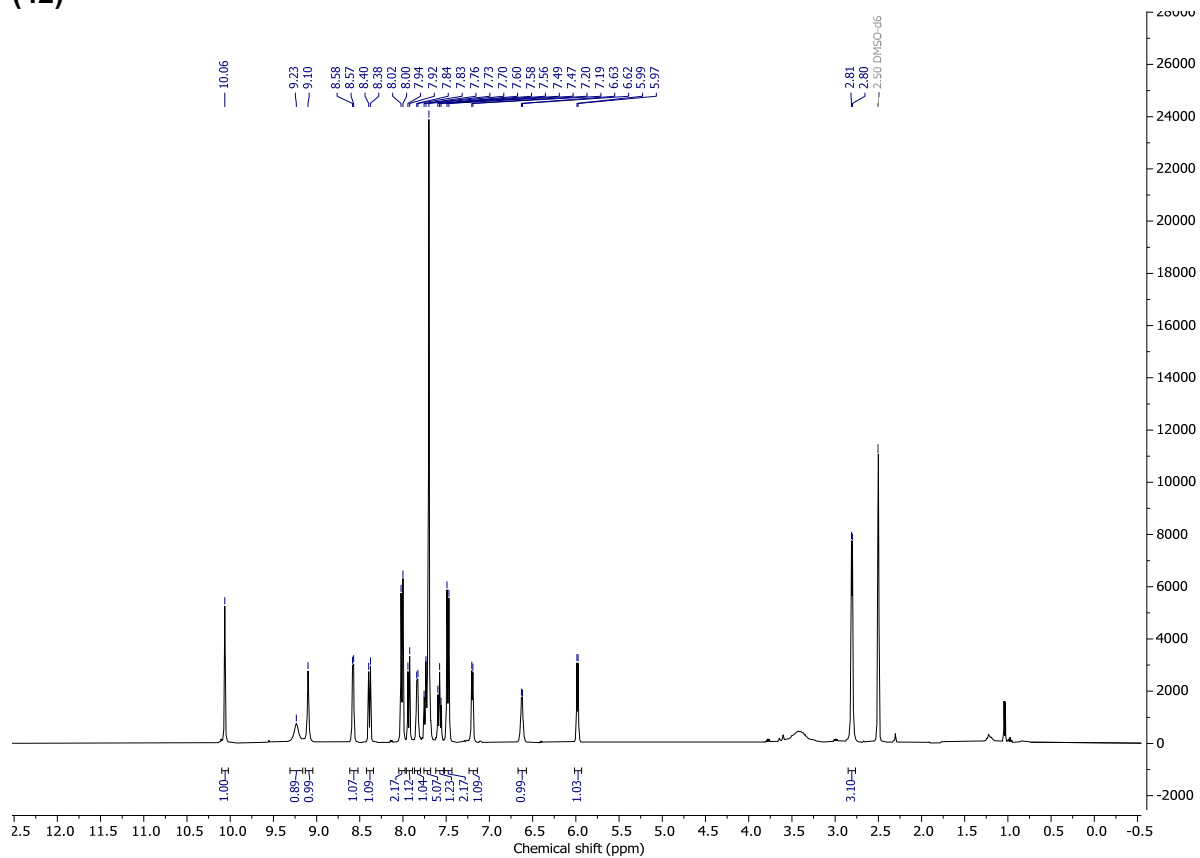


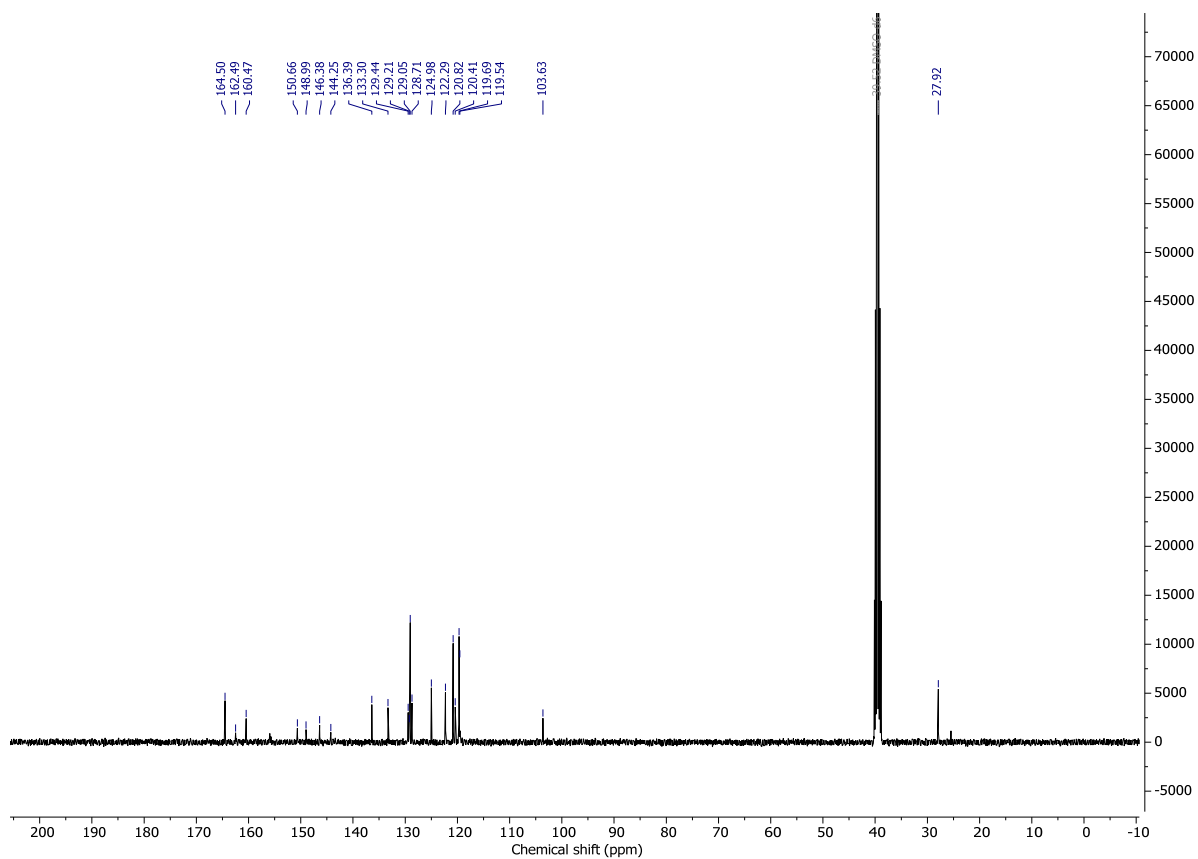
***N*-(4-((2-Aminopyrimidin-4-yl)amino)phenyl)-4-(quinazolin-4-ylamino)benzamide (47)**



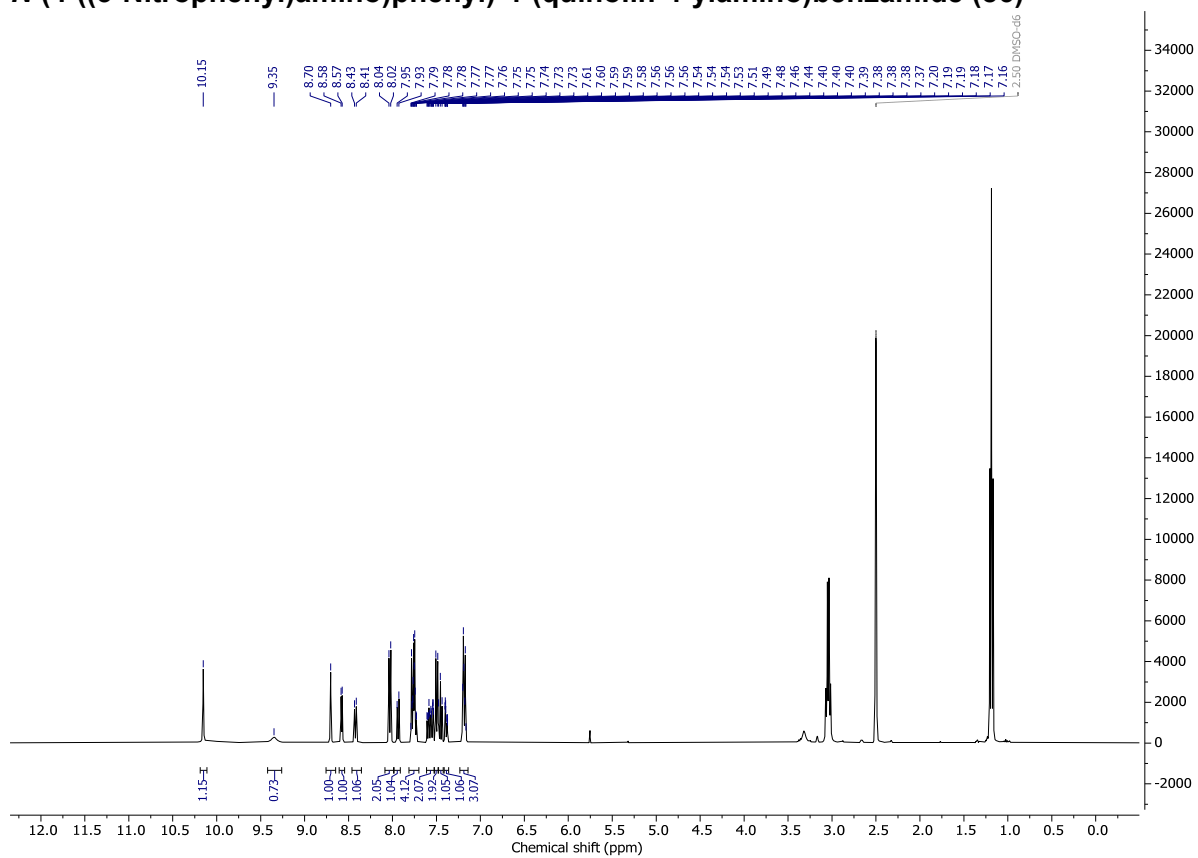


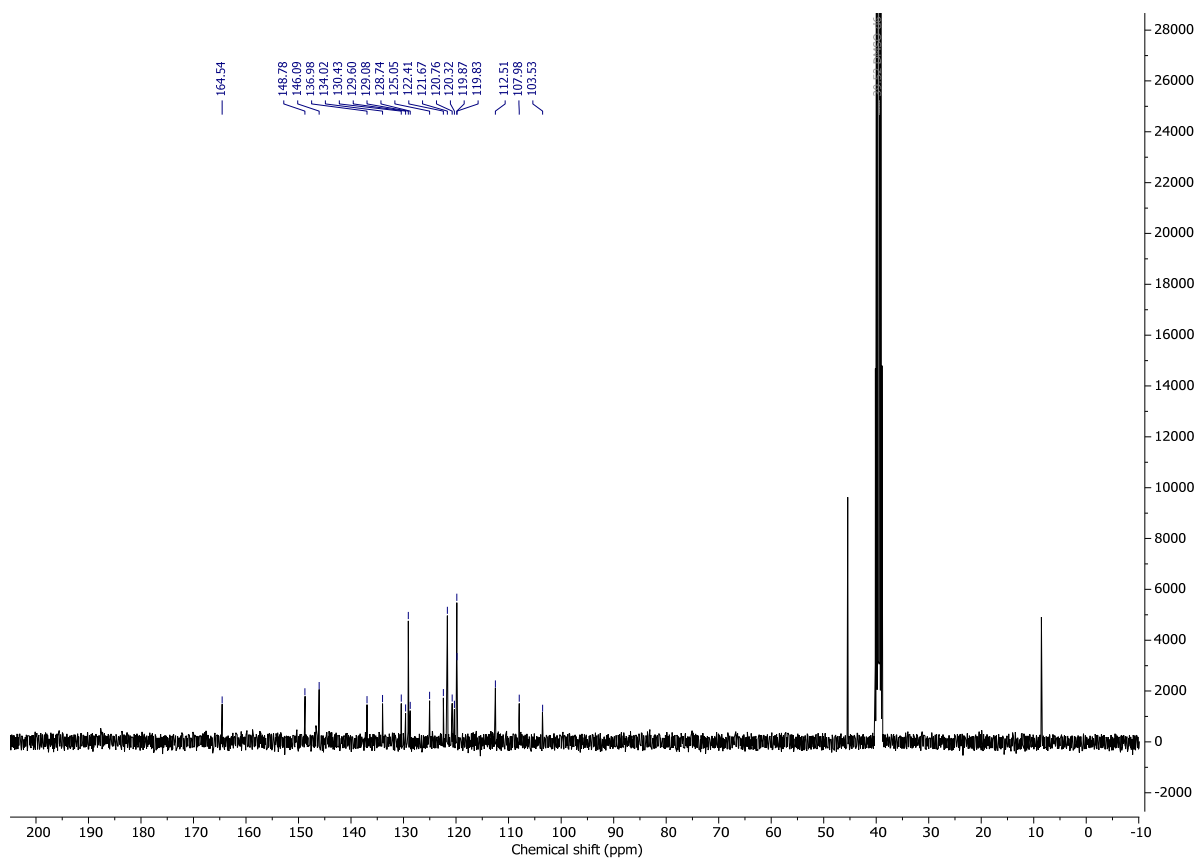
***N*-(4-((2-(Methylamino)pyrimidin-4-yl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (42)**



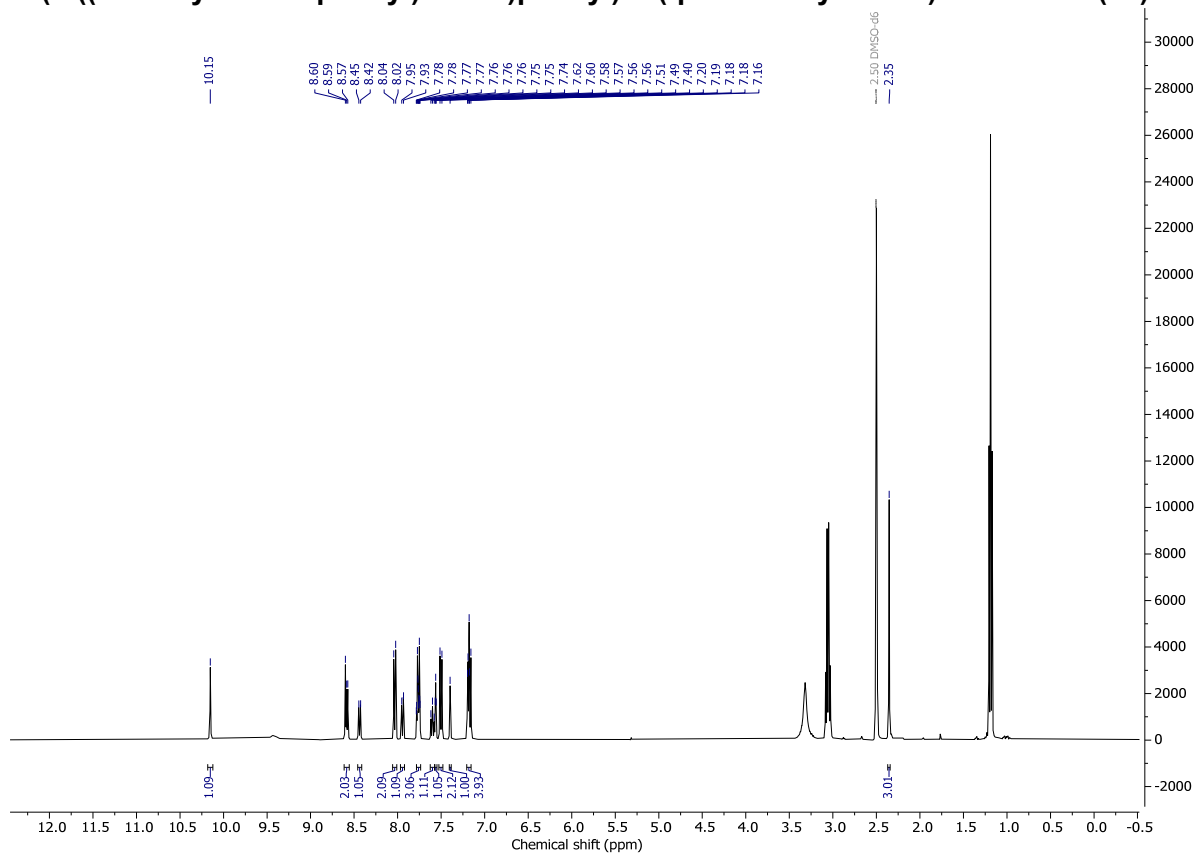


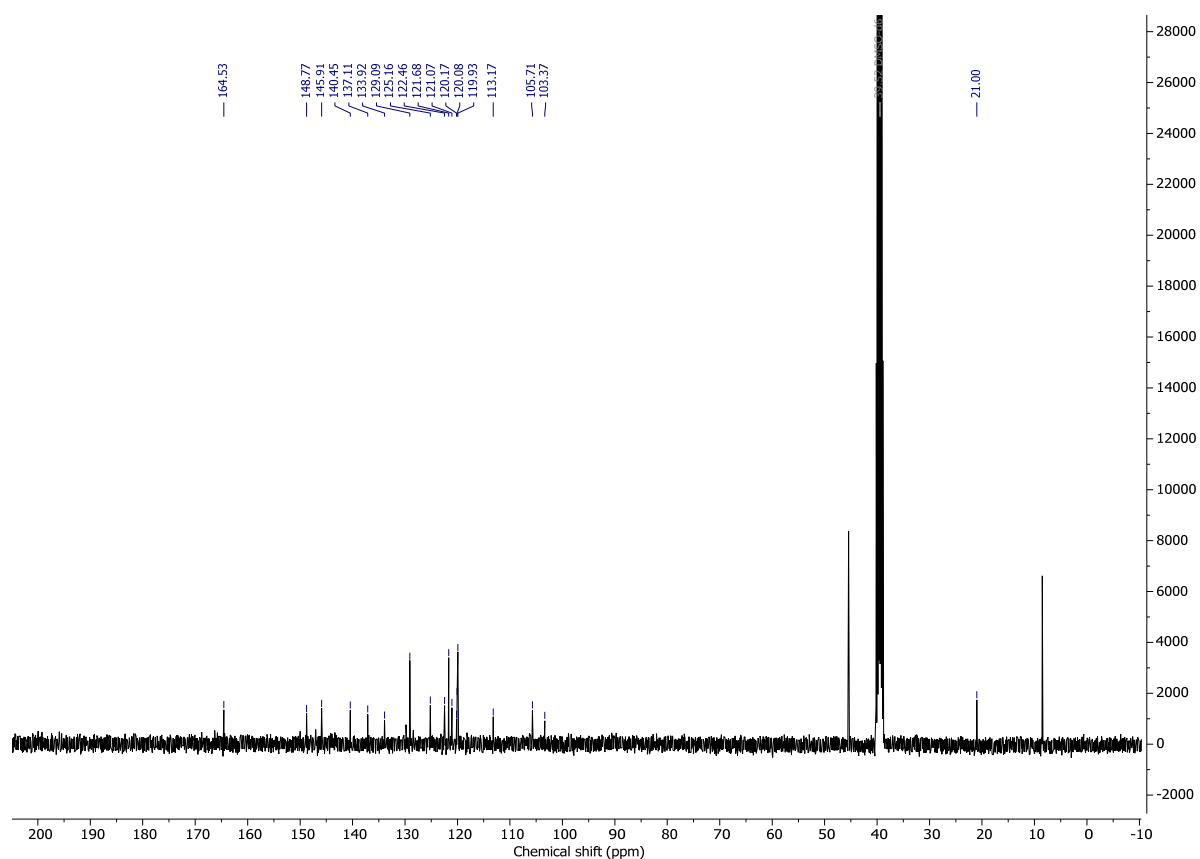
***N*-(4-((3-Nitrophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (86)**



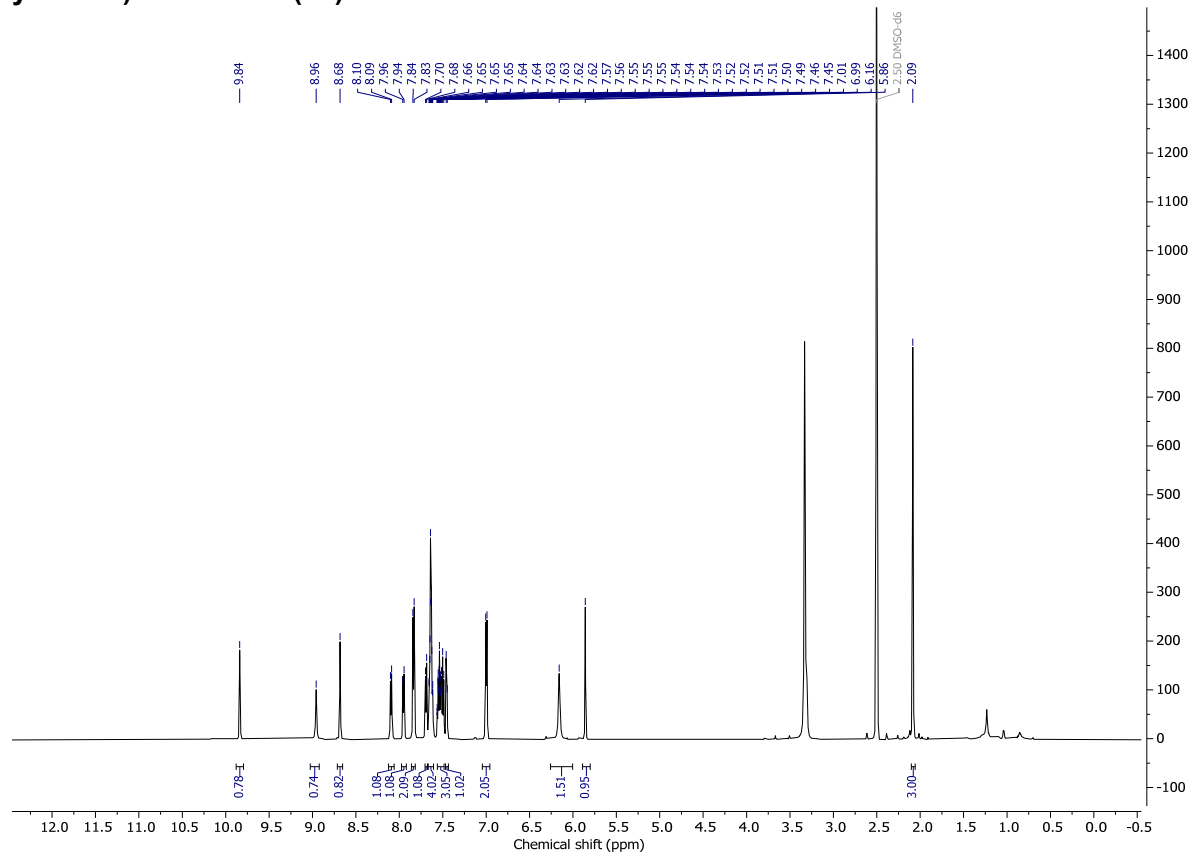


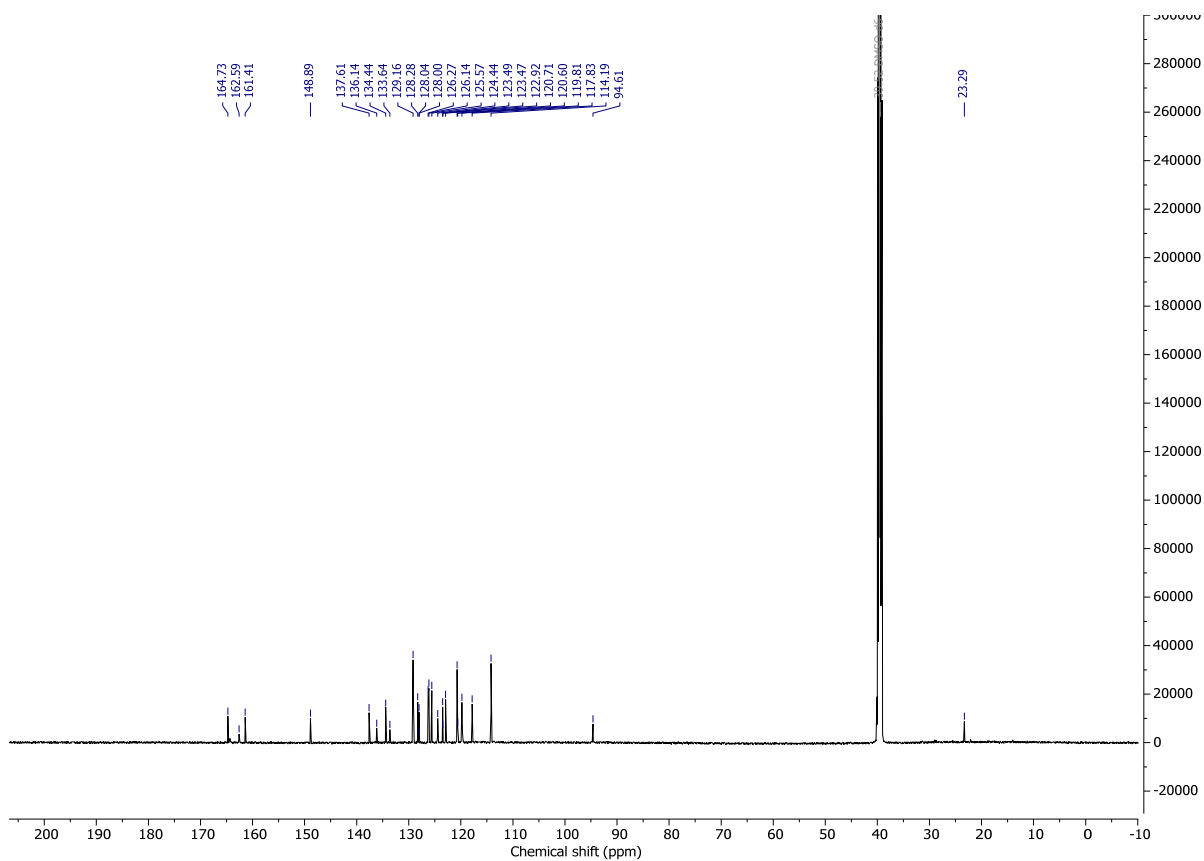
***N*-(4-((3-Methyl-5-nitrophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (87)**



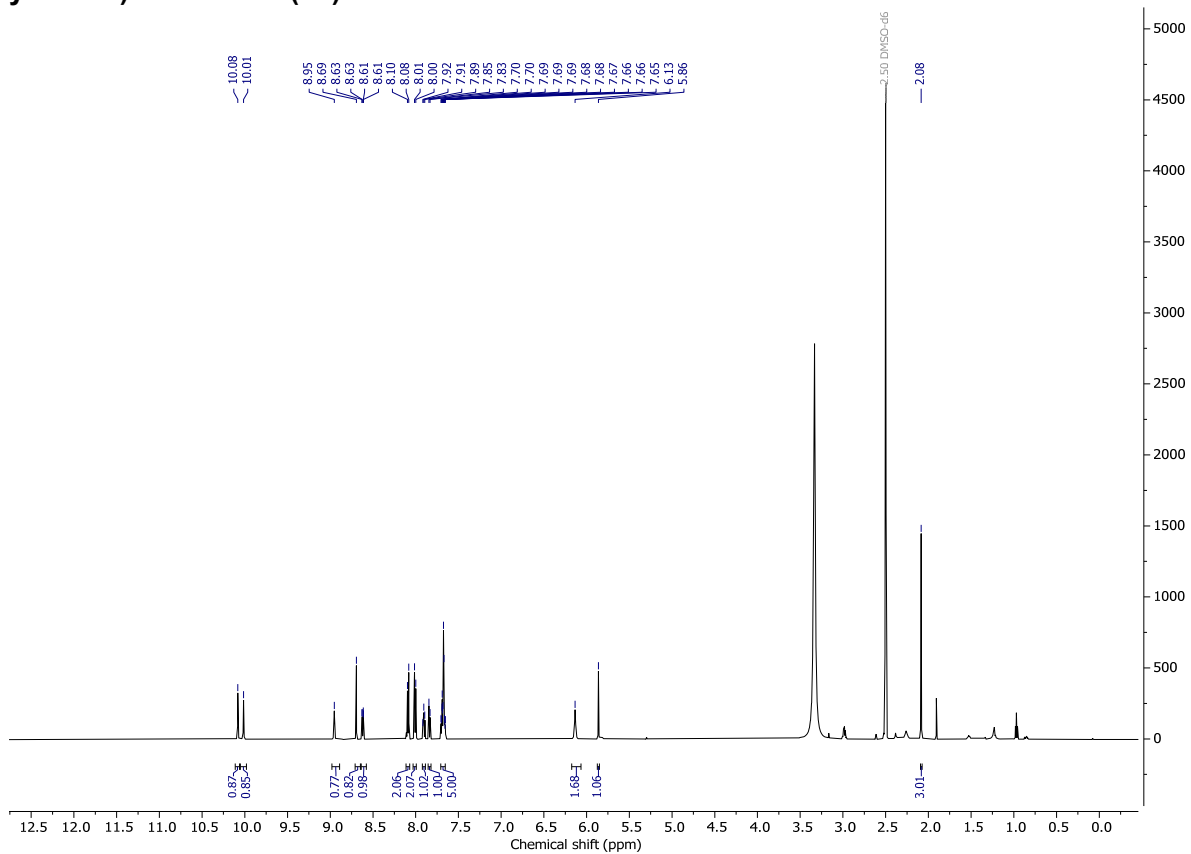


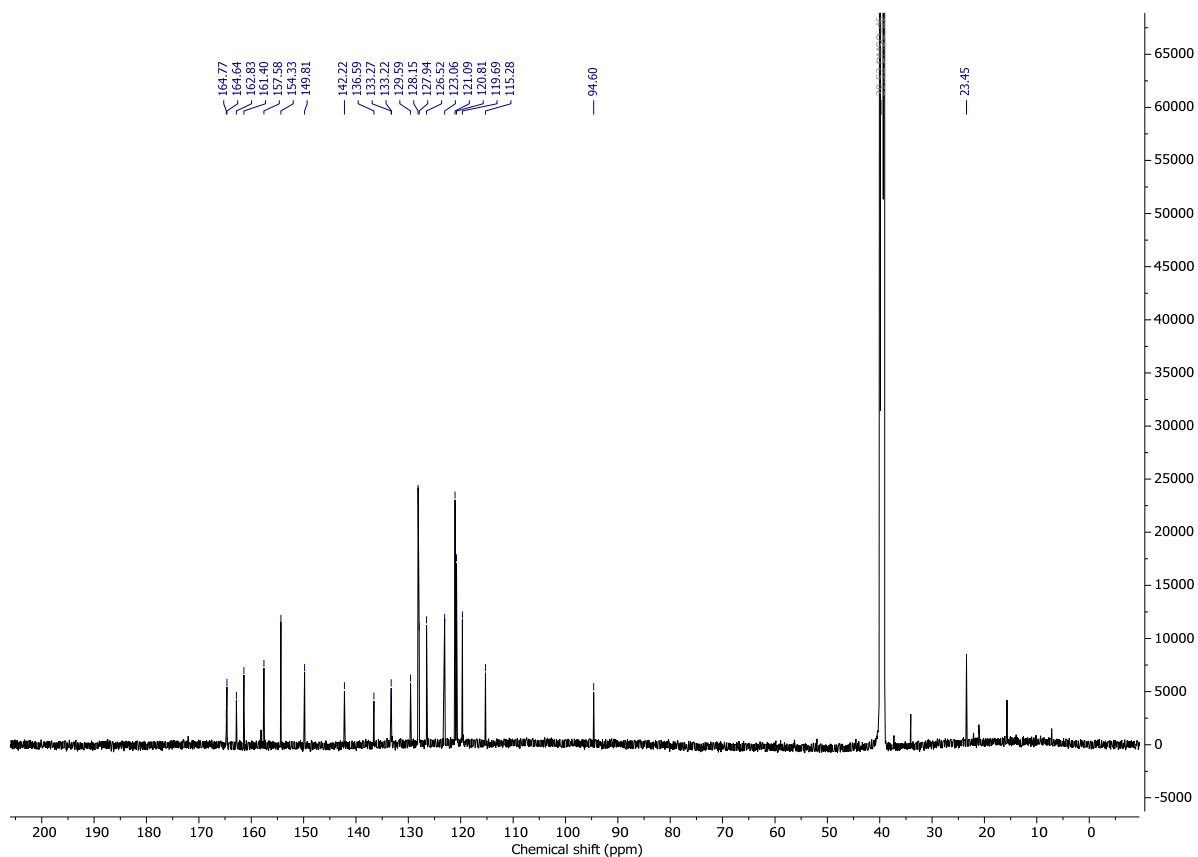
***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(naphthalen-1-ylamino)benzamide (44)**



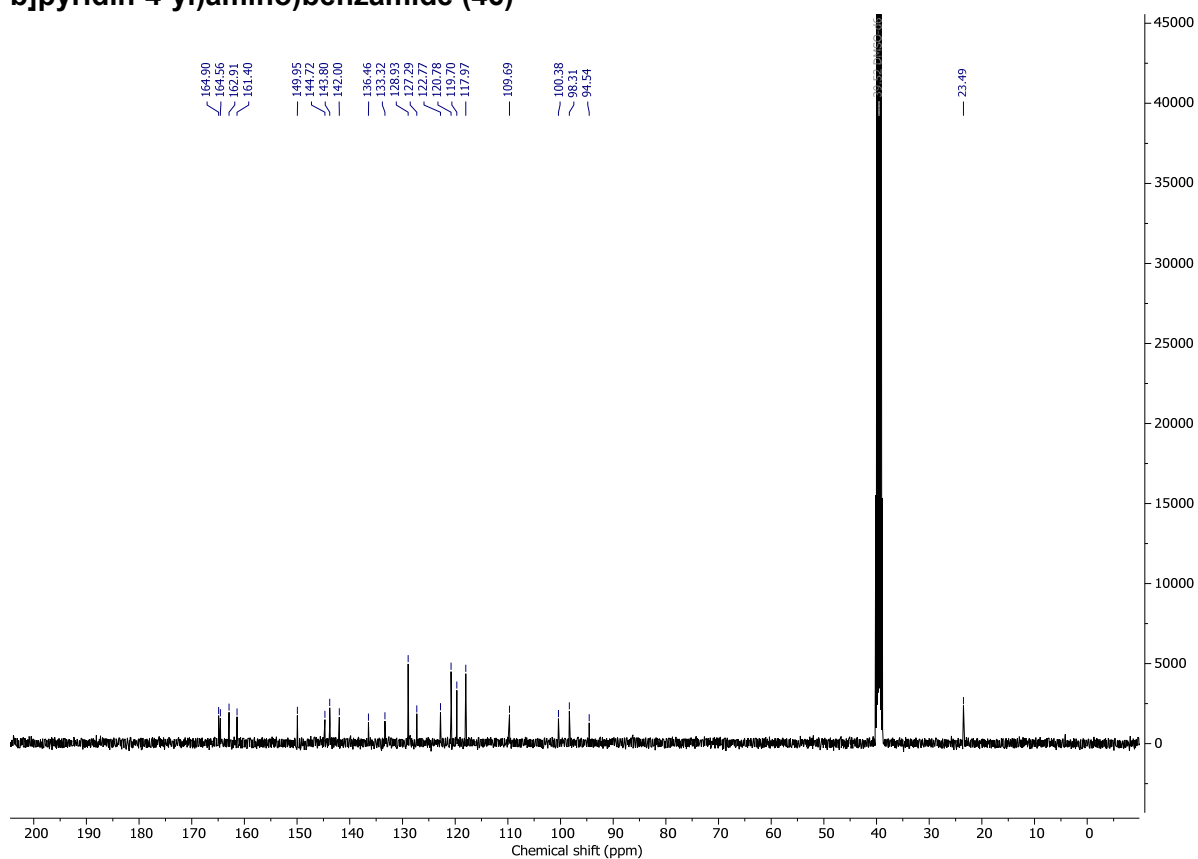


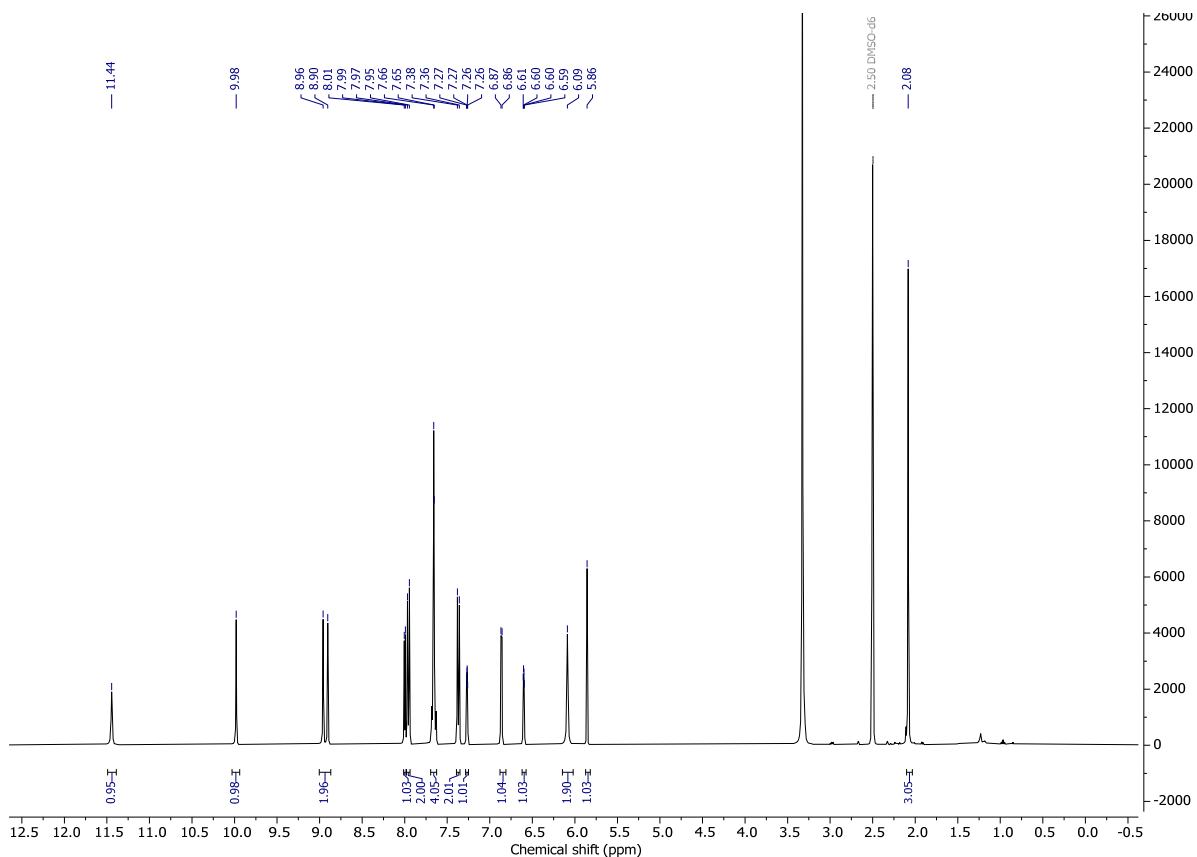
***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-(quinazolin-4-ylamino)benzamide (45)**



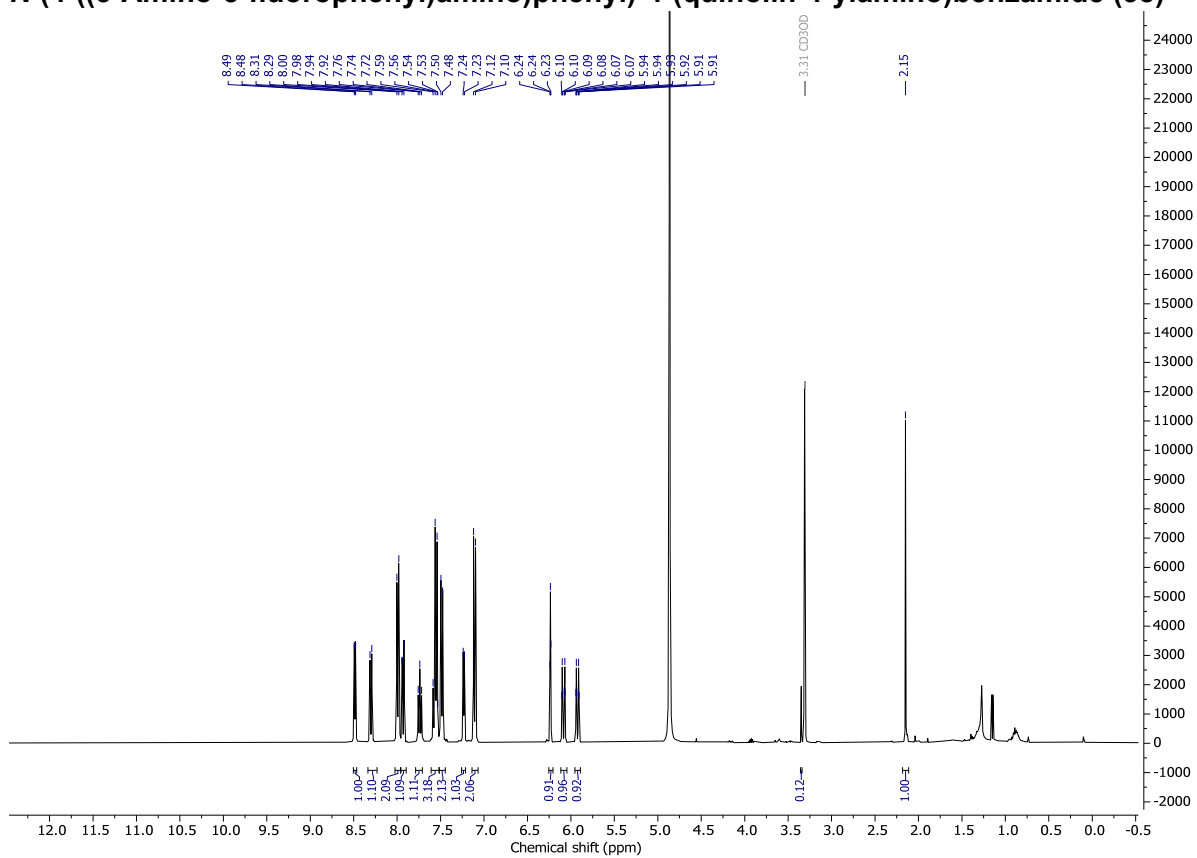


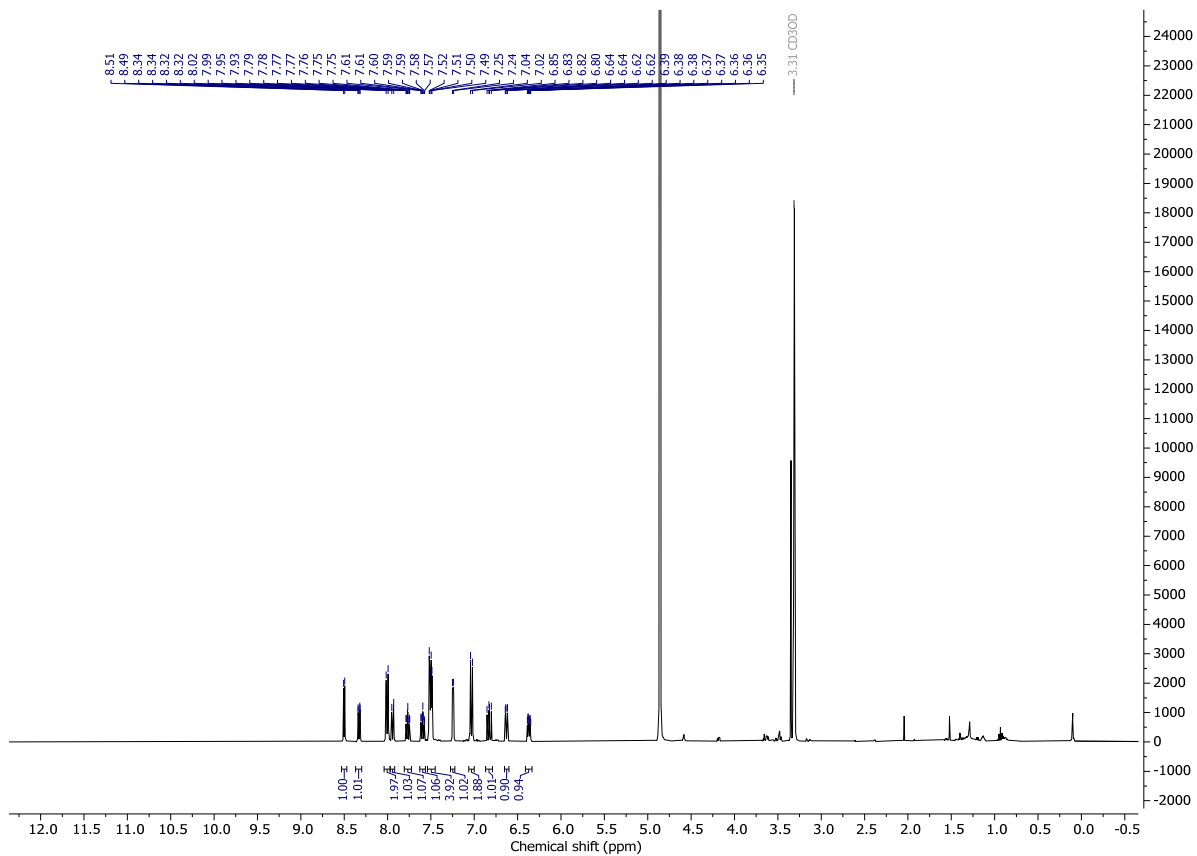
***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-((3a,7a-dihydro-1H-pyrrolo[2,3-b]pyridin-4-yl)amino)benzamide (46)**



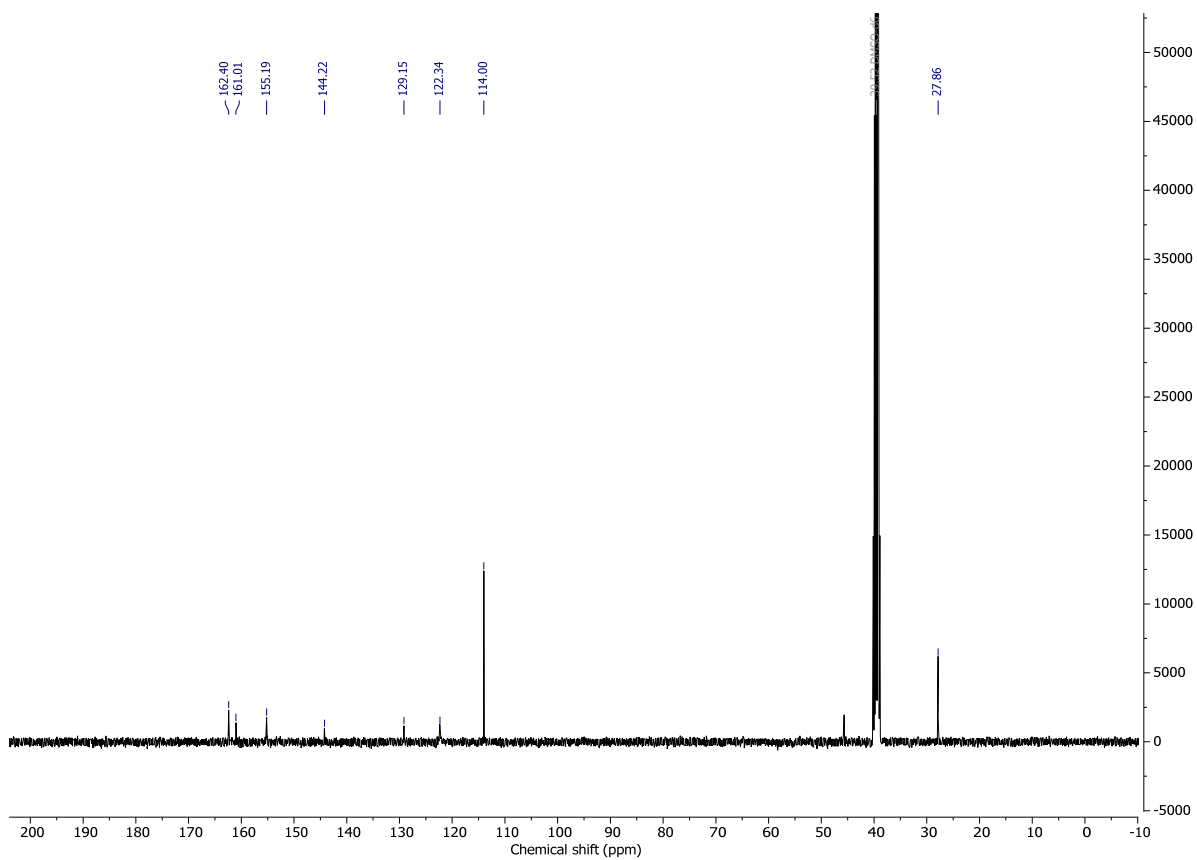


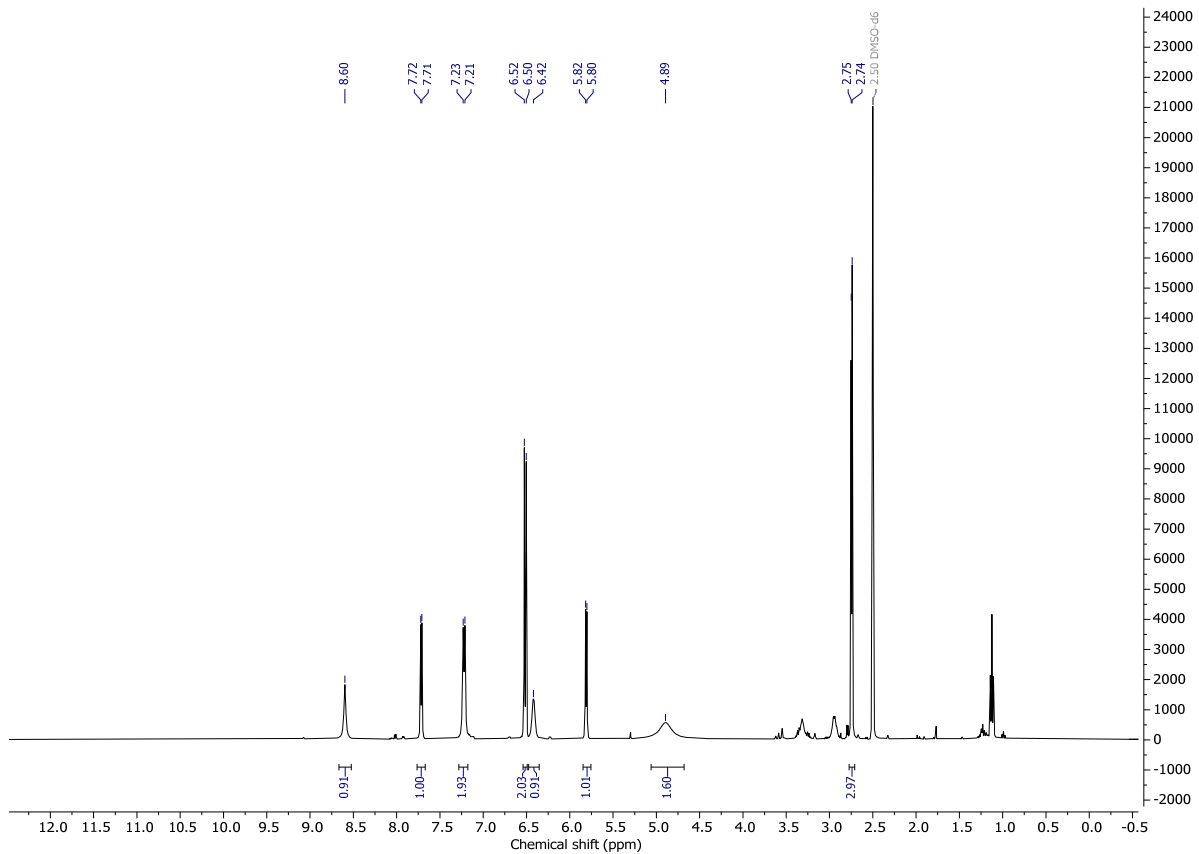
***N*-(4-((3-Amino-5-fluorophenyl)amino)phenyl)-4-(quinolin-4-ylamino)benzamide (38)**



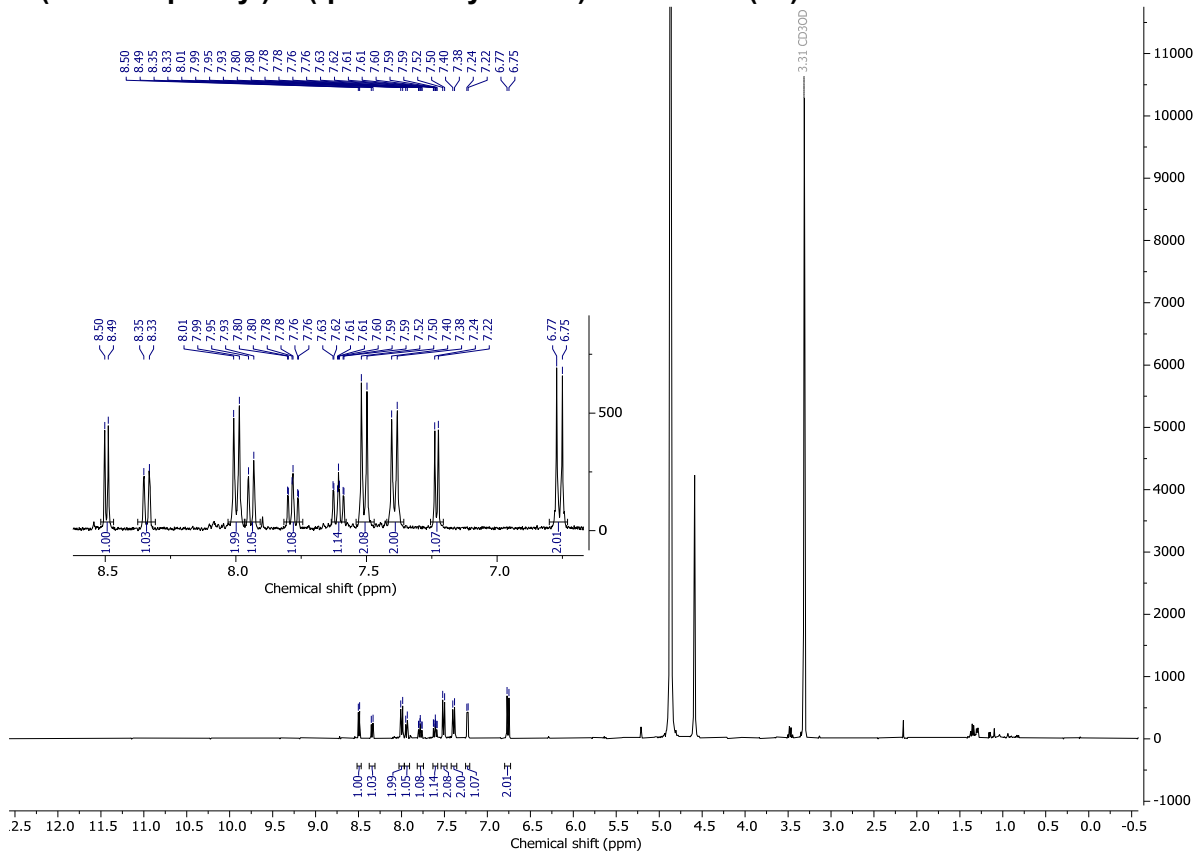


***N*⁴-(4-Aminophenyl)-*N*²-methylpyrimidine-2,4-diamine (96)**

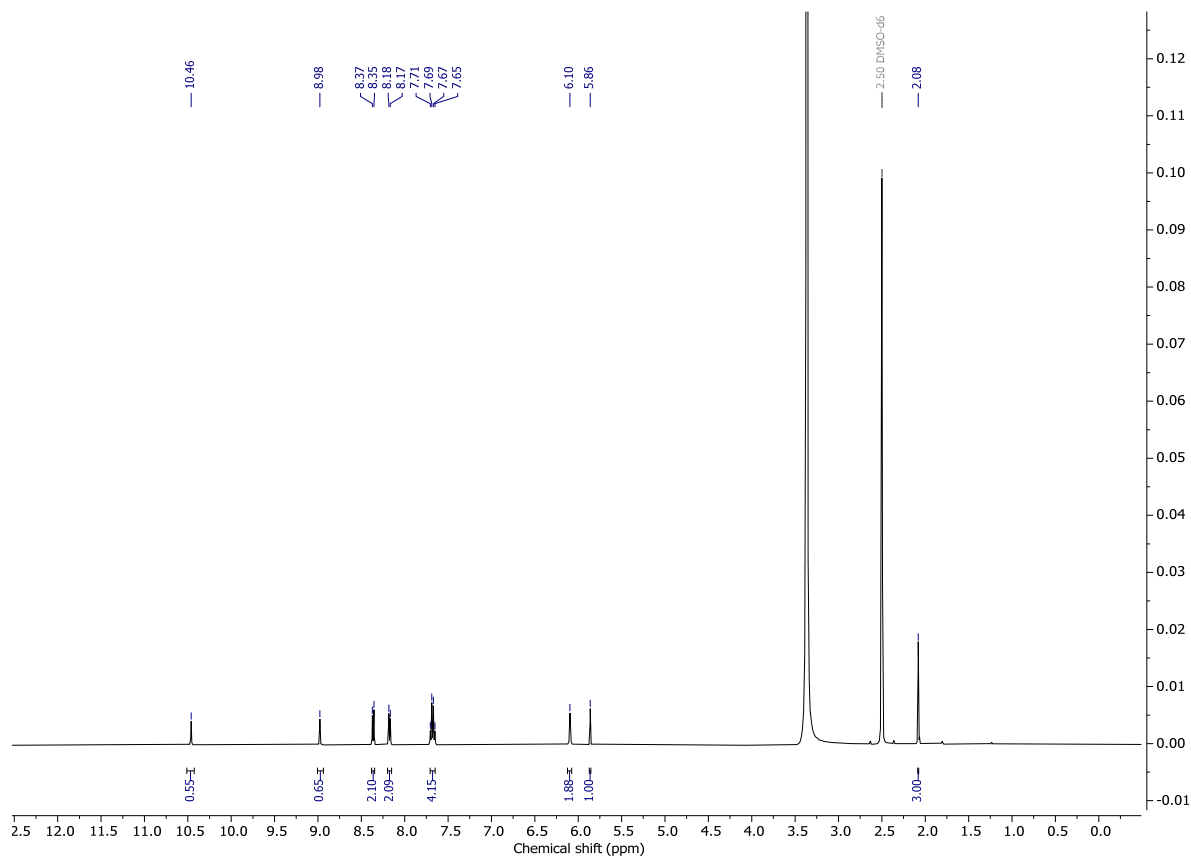




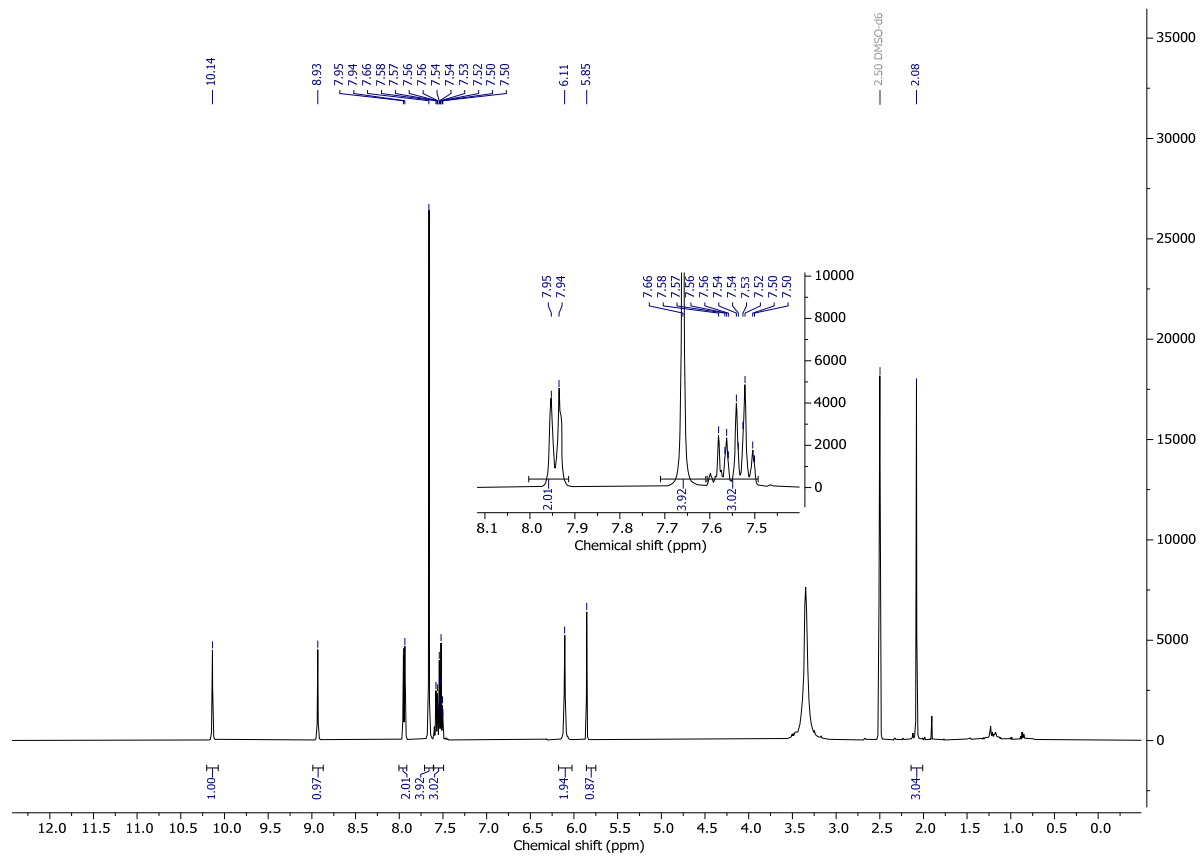
N-(4-Aminophenyl)-4-(quinolin-4-ylamino)benzamide (22)

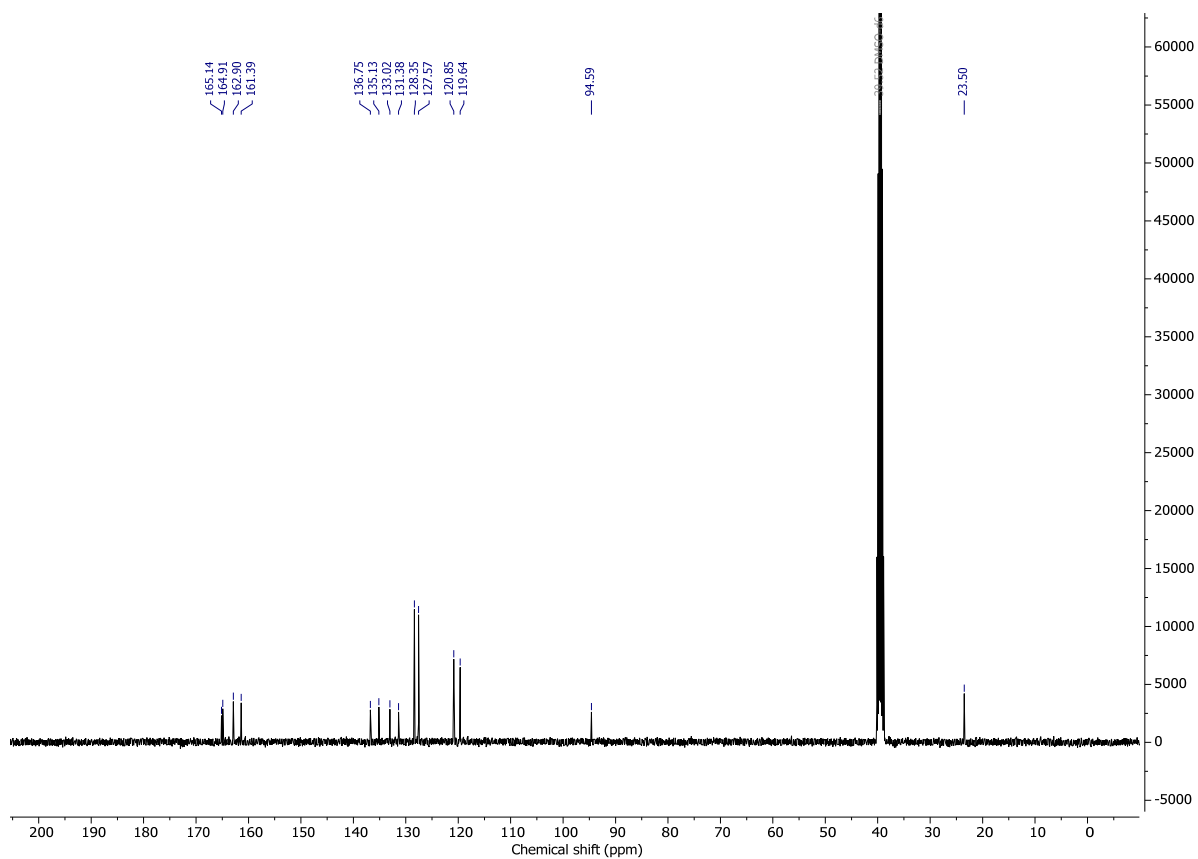


***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)-4-nitrobenzamide (24)**



***N*-(4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)benzamide (23)**





4-((4-((2-Amino-6-methylpyrimidin-4-yl)amino)phenyl)carbamoyl)benzenaminium (25)

