

## Supporting Information

### Acid-Responsive Singlet Oxygen Nanodepot

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## Experimental Section

**Materials.** *N,N*-diethylaminoethyl methacrylate (DEAEMA) purchased from Aldrich was removed inhibitor by distillation under vacuum and stored at -20 °C prior to use. Dulbecco's modified Eagle's medium (DMEM, GIBCO) was used as received. Methanol, dioxane, tetrahydrofuran (THF), dimethylformamide (DMF), dimethyl sulfoxide (DMSO), and all other reagents were purchased from Adamas-beta and used as received. Water was deionized with a Milli-Q SP reagent water system (Millipore) to a specific resistivity of 18.4 MΩ cm. RAFT chain transfer agent PEG-CEP<sup>1</sup> was synthesized according to literature procedures.

### Sample Preparation.

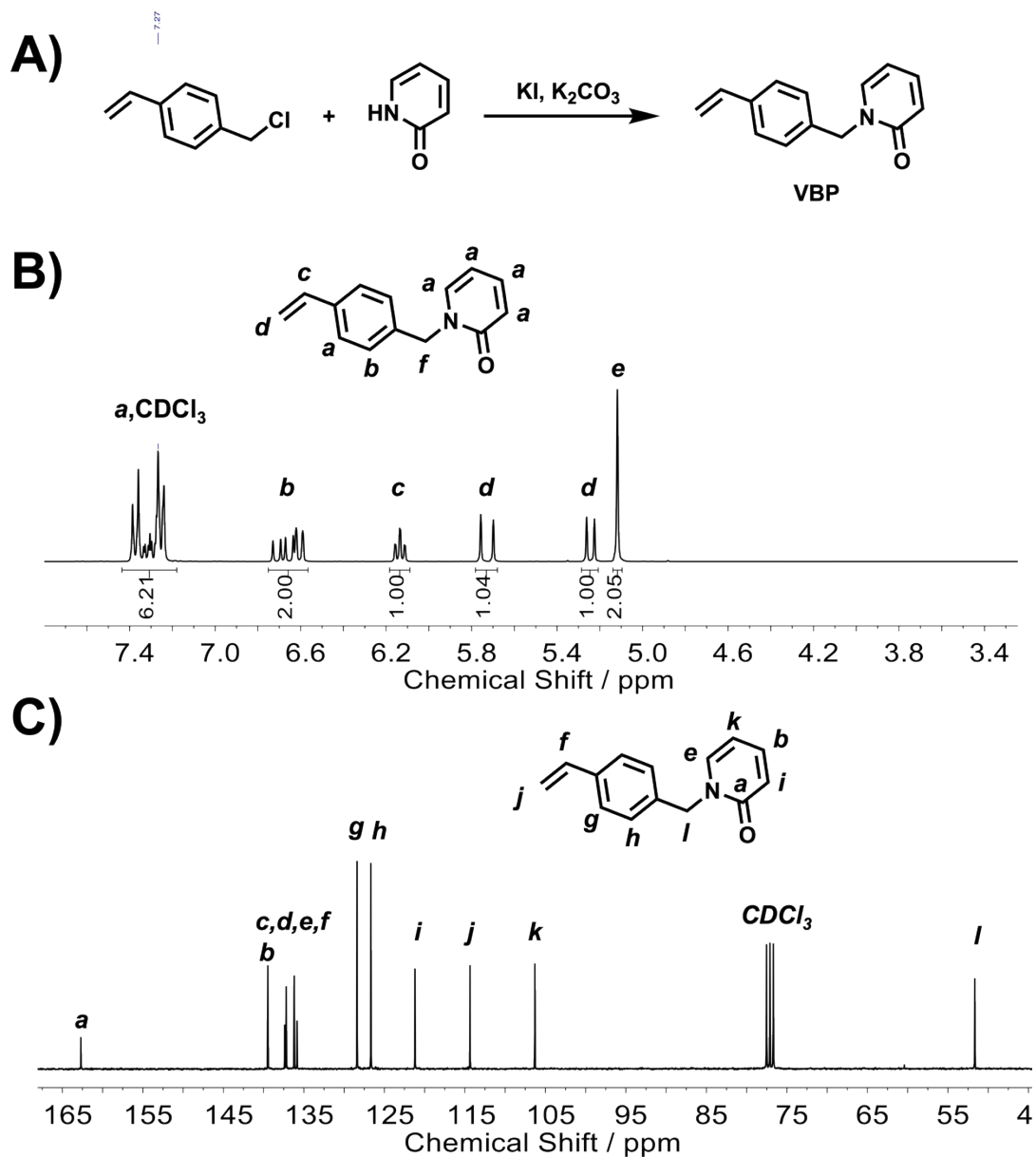
*Synthesis of VBP.* Pyridin-2(1H)-one (10 mmol), 4-chloromethylstyrene (10 mmol), K<sub>2</sub>CO<sub>3</sub> (20 mmol), KI (10 mmol) and acetone (5 mL) were charged into a reaction flask, the mixture was refluxed for 12 h. After evaporating all the solvents, the residues were dissolved in CH<sub>2</sub>Cl<sub>2</sub>, successively washed with saturated aqueous NaCl and water, dried over anhydrous MgSO<sub>4</sub>, filtered, and then concentrated on a rotary evaporator. The crude product was subjected to further purification by silica gel column chromatography using ethyl acetate/petroleum ether as the eluent, affording 1-(4-vinylbenzyl)pyridin-2(1H)-one (VBP) as a white solid (67% yield). <sup>1</sup>H NMR (Figure S1, CDCl<sub>3</sub>, δ, ppm, TMS): 7.4 (6H, aromatic protons), 6.6 (1H, -CH=CH<sub>2</sub>), 5.7 (1H, -CH=CHH), 5.2 (1H, -CH=CHH), 5.1 (2H, -CH<sub>2</sub>-).

*Synthesis of PI-P4.* A reaction flask was charged with PEO-CEP (1 mmol), DEAEMA (0-30 mmol), VBP (45-60 mmol), AIBN (0.2 mmol) and dioxane (25 mL). The reaction system was deoxygenated and backfilled with nitrogen through three freeze-vacuum-thaw cycles. Polymerization was performed at 70 °C for 8 h. Monomers and other impurities were removed by precipitation three times in excess cold ether. The final polymer was dried in a vacuum oven overnight at room temperature to give a yellow sticky solid.

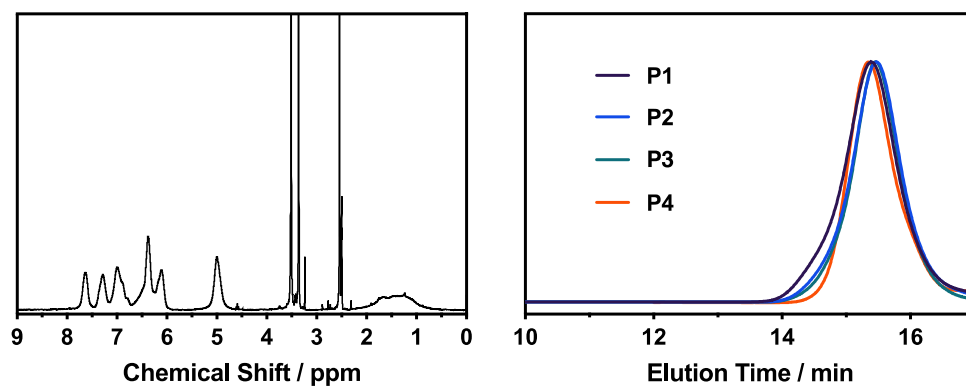
*RAFT End Group Removing of PI-P4 Block Copolymers.* A glass ampoule was charged with PI-P4 (100 mg), excess AIBN (5 mg), and DMSO (2 mL). The reaction system

was deoxygenated and backfilled with nitrogen through three freeze-vacuum-thaw cycles. After stirring at 70 °C for 2 h, the mixture was precipitated into an excess of diethyl ether for three times. The final product was dried in a vacuum oven overnight at room temperature to yield a white solid.

**Characterization.** Nuclear magnetic resonance (NMR) spectra were performed on a Bruker AV300 NMR 300 MHz spectrometer (samples dissolved in CDCl<sub>3</sub> or D<sub>2</sub>O). The molecular weight and molecular weight distribution of the block polymers and their precursors were determined by size exclusion chromatography (SEC) with a Waters 1515 pump and a Waters 2414 differential refractive index detector set at 30 °C. The SEC separation column used two Styragel columns (HR2 and HR4) at 45 °C and THF as eluent (flow rate 1.0 mL/min). Calibration was performed using a series of low polydispersity polystyrene standards. A Hitachi H-800 electron microscope was used to acquire transmission electron microscopy (TEM) images. The sample for TEM observations was prepared by placing 10 μL of dispersion of nanoparticle (0.5 g/L) on copper grids coated with thin films of Formvar and carbon successively. Confocal lasers scanning microscopy (CLSM) images were acquired using a Leica TCS SP5 microscope. Dynamic Laser Scattering (DLS) was measured using an ALV5000.



**Figure S1.** Synthetic schemes (A),  $^1\text{H}$  NMR (B), and  $^{13}\text{C}$  NMR (C) recorded for 1-(4-vinylbenzyl)pyridin-2(1H)-one (VBP).

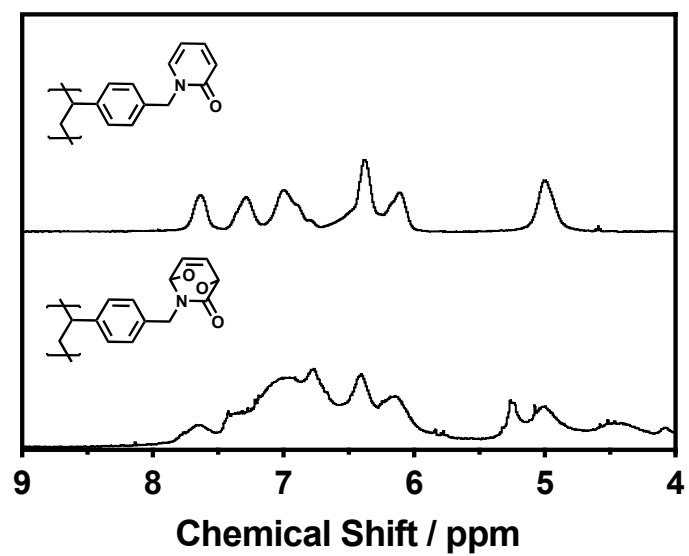


**Figure S2.**  $^1\text{H}$  NMR spectra recorded (left) and THF GPC traces (right) for **P1-P4**.

**Table S1. Molecular parameters of polymers.**

<b>Entry</b>	<b><math>M_{n, \text{NMR}}</math> (kDa)<sup>a</sup></b>	<b><math>M_{n, \text{GPC}}</math> (kDa)<sup>b</sup></b>	<b><math>M_w/M_n</math><sup>b</sup></b>
<b>P1</b>	15.0	16.4	1.20
<b>P2</b>	13.3	14.7	1.21
<b>P3</b>	13.5	14.9	1.23
<b>P4</b>	13.8	15.1	1.24

<sup>a</sup> Calculated from <sup>1</sup>H NMR results. <sup>b</sup> Determined by GPC using THF as the eluent (1.0 mL/min).



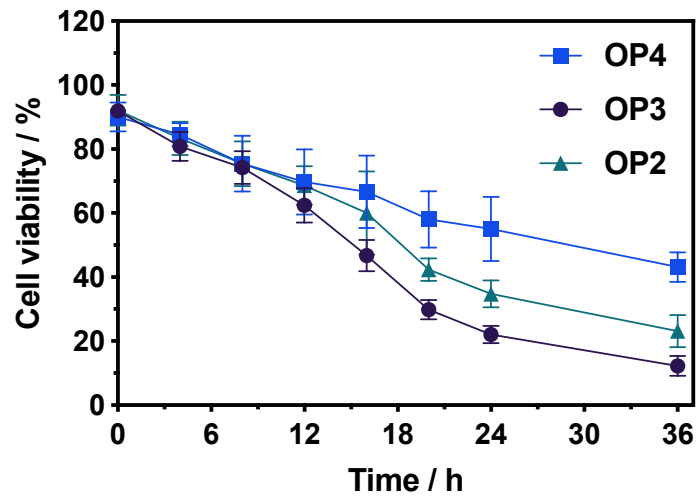
**Figure S3.** <sup>1</sup>H NMR spectra recorded in for **P1** before and after <sup>1</sup>O<sub>2</sub> loading.

**Table S2. The content of  $^1\text{O}_2$  loaded on OP1-OP4.**

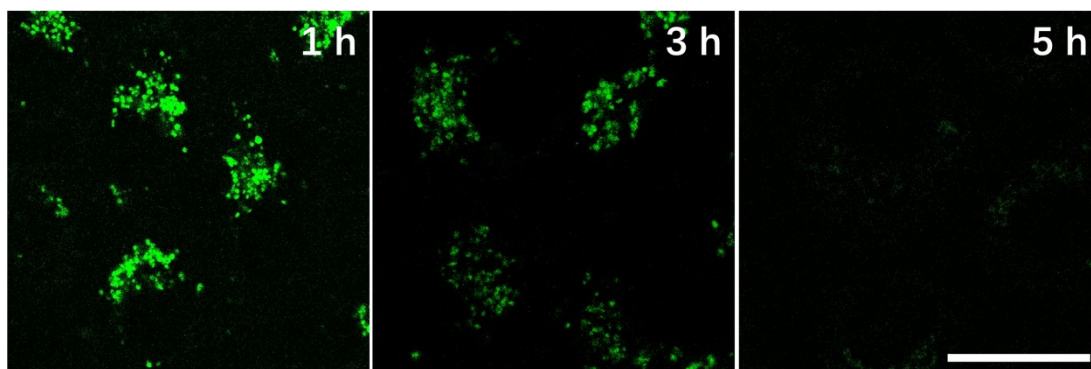
<b>Entry</b>	<b>Content of <math>^1\text{O}_2</math> loaded <sup>a</sup></b>
<b>OP1</b>	51%
<b>OP2</b>	53%
<b>OP3</b>	62%
<b>OP4</b>	56%

<sup>a</sup> Calculated from  $^1\text{H}$  NMR results.

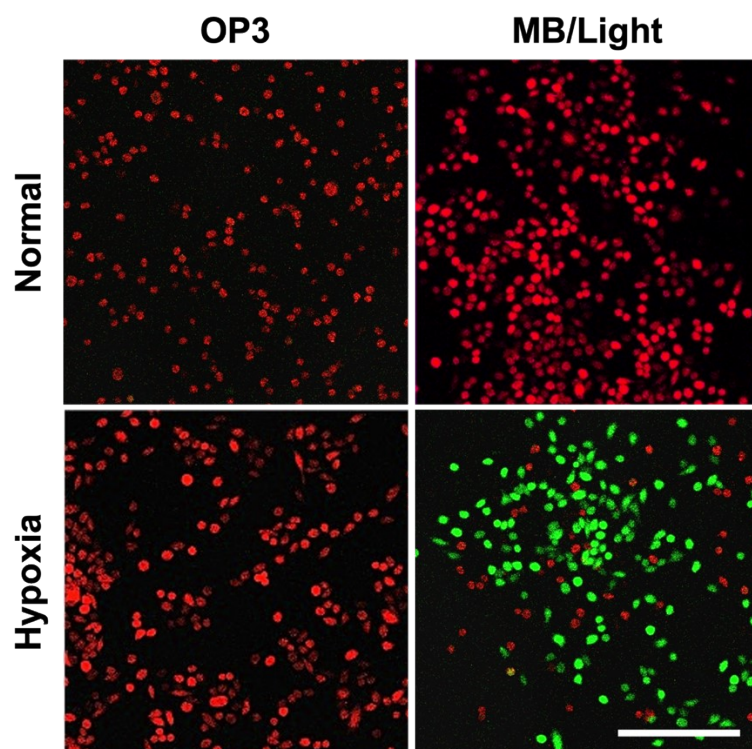




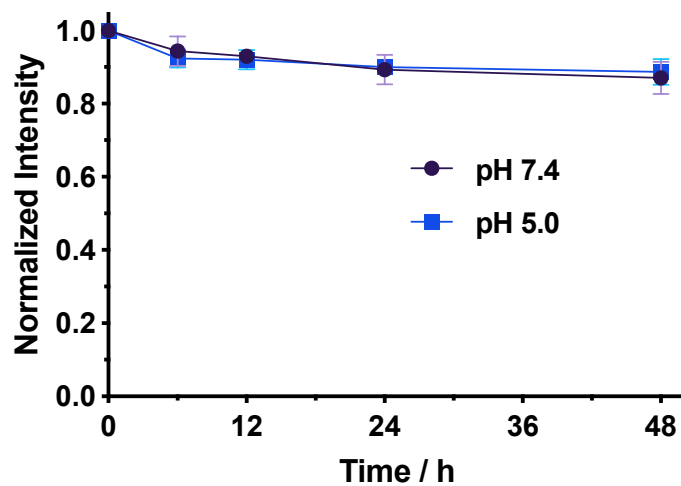
**Figure S4.** Time-dependent cytotoxicity of OP2-OP4 (0.1 g/L) to HepG2 cells by MTT assay.



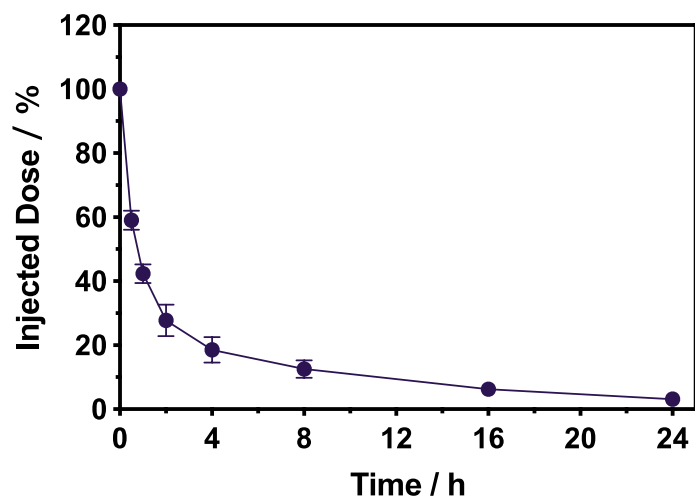
**Figure S5.** CLSM images of Lysotracker Green incubated with HepG2 cells after treating with **OP3** for varying times. Scale bar: 25  $\mu\text{m}$ .



**Figure S6.** Live (green channel) / dead (red channel) assay with HepG2 cells incubated of **OP3** aSOND (0.1 g/L) and MB (10  $\mu$ M) based PDT under normal and hypoxia condition. Green and red channel emissions were collected at  $530 \pm 20$  nm and  $610 \pm 20$  nm, respectively. Scale bar: 100  $\mu$ m.



**Figure S7.** Fluorescence emission intensity for DOX ( $2 \mu\text{M}$ ,  $\lambda_{\text{ex}} = 540 \text{ nm}$ ) incubating with **OP3** ( $0.1 \text{ g/L}$ ) at pH 7.4 and 5.0.



**Figure S8.** Time-dependent blood level upon tail vein injection of **OP5**, calculated as percentage of injected dose remaining in the blood.

- 1 Xu, X.; Flores, J. D.; McCormick, C. L. *Macromolecules*, 2011, **44**, 1327-1334.