

Co-Expression of Type 1 Fimbriae and Flagella in *Escherichia coli*: Consequences for Adhesion at Interfaces

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ELECTRONIC SUPPLEMENTARY INFORMATION

Strain construction:

Strain MG1655 Δ *fimA* was constructed by deleting *fimA* from *E. coli* K-12 MG1655 using the λ Red recombinase-based method, as described by Datsenko and Wanner.¹ The following primers and plasmids were used to generate the knockout cassette: *fimAP1* (forward), *fimAP2* (forward), *fimAP1* (reverse), *fimAP2* (reverse) and plasmid pKD13. The kanamycin resistance gene was removed using plasmid pCP20.

Plasmid construction:

Plasmid pPCC2000 (“p(blank)”) was derived from vector pPCC1322², carrying the *lac* repressor gene (*lacI*) and apramycin resistance gene (*aac*).

Plasmid pPCC2208 was constructed by linearizing plasmid pPCC1401 with XbaI restriction enzyme (New England Biolabs). Next, a gene construct containing *Ptac-fliC-flhD-flhC* was assembled by amplifying *fliC* and *flhDC* from *E. coli* MG1655 genomic DNA, and subcloning these into lab plasmid pPCC2206. *Ptac-fliC-flhD-flhC* was then PCR-amplified using Phusion Polymerase (New England Biolabs) and primers p2207P1 (forward) and p2207P1 (reverse). The PCR product was assembled into the linearized pPCC1401 using HiFi DNA Assembly (New England Biolabs).

The complete nucleotide sequences of the plasmids are provided below. The plasmids were sequenced by Plasmidsaurus.

Nucleotide sequence of plasmid pPCC2000:

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Nucleotide sequence of plasmid pPCC2208:

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CTGCTGCTTGCAAAC

Primer	Sequence (5'-3')
<i>fimAP1</i> (forward)	gtttttgaaaggaaagcagcatggattgtgtaggctggagctgc
<i>fimAP2</i> (forward)	cgactgcccattgtcgatttagaaatagtttttgaaggaaagcagcatgg
<i>fimAP1</i> (reverse)	cctgggtaggttattgatactgaaccttgaaattccgggatccgtcg
<i>fimAP2</i> (reverse)	ccgtaatgacgtccctgaacctgggtaggttattgatactgaaccttg
<i>fliC</i> (forward)	ggtgccgcccagttctccaaccgcggtcaaactggg
<i>fliC</i> (reverse)	ggttgagaactggcggcaccggaagagtcagtatag
p2207P1 (forward)	aaacaaagcgcactttgctggtctgtcccctcatctgttgacaattaatcatcggtc
p2207P1 (reverse)	tcagcagttcaacctgtgatagtaggggatcctgcgagcaggggaattgatcc

Table S1 Primers used in this study.

Strains	CFU mL ⁻¹
MG1655+p(blank)	2.0×10 ¹¹
MG1655Δ <i>fimA</i> +p(blank)	2.3×10 ¹¹
MG1655Δ <i>fimA</i> +p(<i>fim</i>)	5.9×10 ¹⁰
MG1655Δ <i>fimA</i> +p(<i>fim-flhDC</i>)	4.8×10 ¹⁰

Table S2 Number of Colony Forming Units (CFU) per mL of OD₆₀₀ = 1 for different strains induced with 10 μM concentration of IPTG.

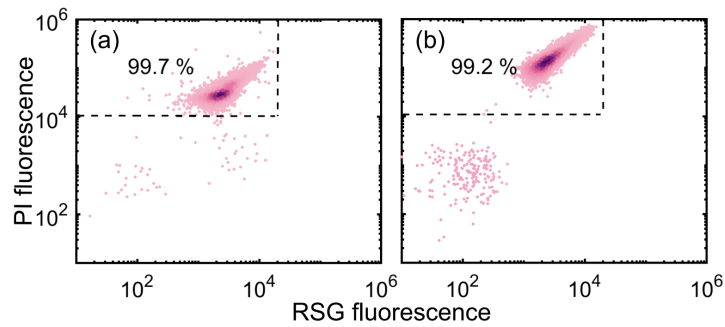


Fig. S1 (a) Cells treated with 70% ethanol for 1 h and (b) cells killed by heating at 90°C for 15 mins were stained with Propidium Iodide and Redox Sensor Green and analyzed by flow cytometry as a positive control to determine cell viability. The gating cutoffs for the PI and RSG Fluorescence were 10,000 and 20,000 respectively. The horizontal and vertical dashed lines represent the gating cutoffs for the PI and RSG Fluorescence respectively. The percentage value corresponds to the percent of dead and membrane-compromised cells in the cell population.

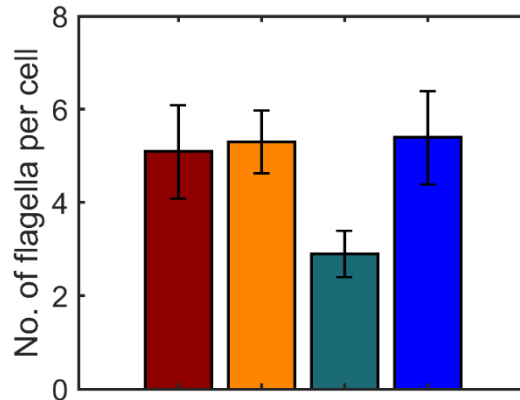


Fig. S2 Average number of flagella per cell for different strains: (■) MG1655+p(blank) + 10 μM IPTG; (■) MG1655Δ*fimA*+p(blank) + 10 μM IPTG; (■) MG1655Δ*fimA*+p(*fim*) + 10 μM IPTG; (■) MG1655Δ*fimA*+p(*fim-flhDC*) + 10 μM IPTG.

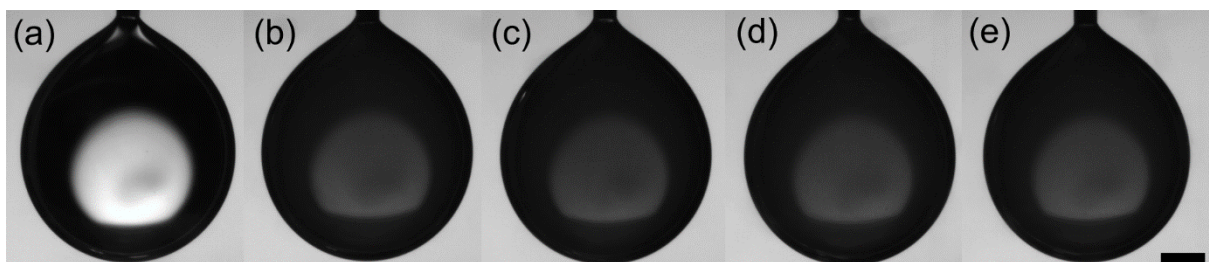


Fig. S3 Images of the pendant drop of bacterial suspension ($OD_{600} = 2$) of different strains (a) PBS with no cells; (b) MG1655+p(blank) + 10 μM IPTG; (c) MG1655Δ*fimA*+p(blank) + 10 μM IPTG; (d) MG1655Δ*fimA*+p(*fim*) + 10 μM IPTG; (e) MG1655Δ*fimA*+p(*fim-flhDC*) + 10 μM IPTG in hexadecane. Scale bar is 1 mm.

	MG1655+ p(blank)	MG1655 Δ <i>fimA</i> + p(blank)	MG1655 Δ <i>fimA</i> + p(<i>fim</i>)	MG1655 Δ <i>fimA</i> + p(<i>fim-fliC-flhDC</i>)
MG1655+ p(blank)	-	N	Y	Y
MG1655 Δ <i>fimA</i> + p(blank)	N	-	Y	Y
MG1655 Δ <i>fimA</i> + p(<i>fim</i>)	Y	Y	-	N
MG1655 Δ <i>fimA</i> + p(<i>fim-fliC-flhDC</i>)	Y	Y	N	-

Table S3 A 4×4 matrix indicating whether the difference in the interfacial tension values between different strains is significant at the $p < 0.05$ level.

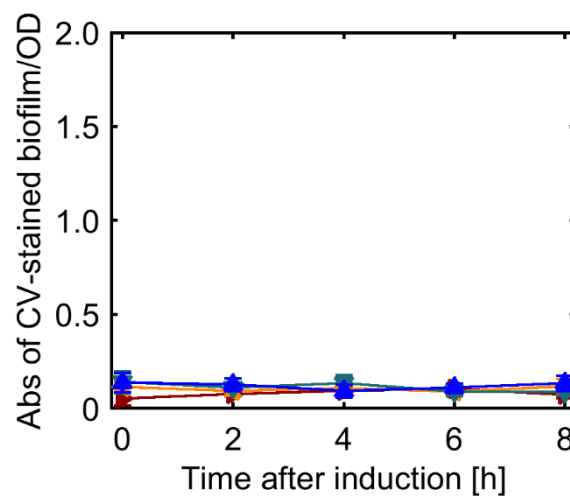


Fig. S4 MG1655 Δ *fimA*+p(*fim*) and MG1655 Δ *fimA*+p(*fim-fliC-flhDC*) strains do not form biofilm in the absence of IPTG. Symbols: (►) MG1655+p(blank) + 0 μ M IPTG; (◄) MG1655 Δ *fimA*+p(blank) + 0 μ M IPTG; (▼) MG1655 Δ *fimA*+p(*fim*) + 0 μ M IPTG; (▲) MG1655 Δ *fimA*+p(*fim-fliC-flhDC*) + 0 μ M IPTG.

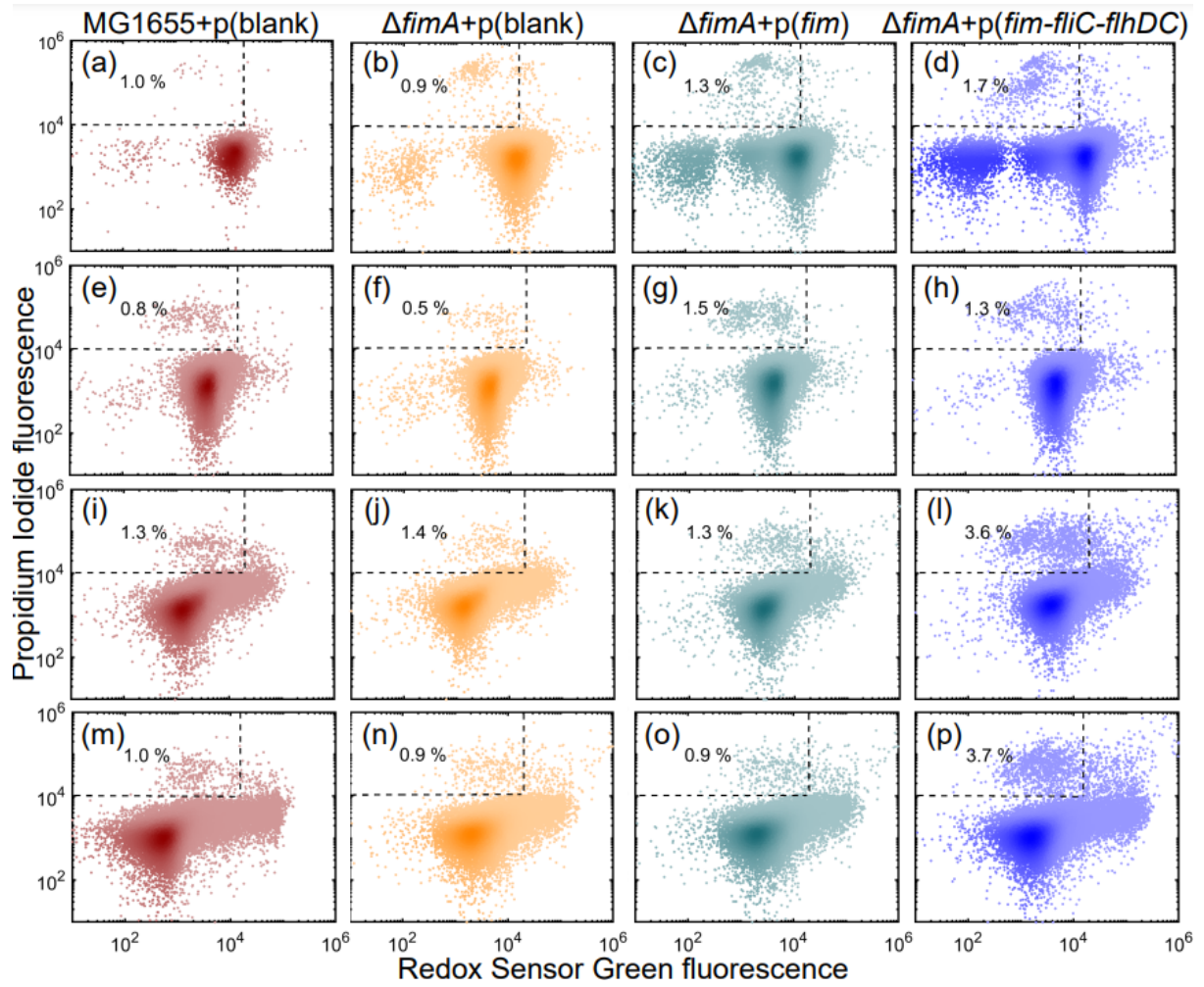


Fig. S5 PI and RSG staining of different strains (from left to right: MG1655+p(blank), MG1655 $\Delta fimA+p(\text{blank})$, MG1655 $\Delta fimA+p(\text{fim})$, and MG1655 $\Delta fimA+p(\text{fim-fliC-flhDC})$) at (a – d) 0 h, (e – h) 2 h, (i – l) 6 h, and (m – p) 8 h after induction with 10 μM IPTG.

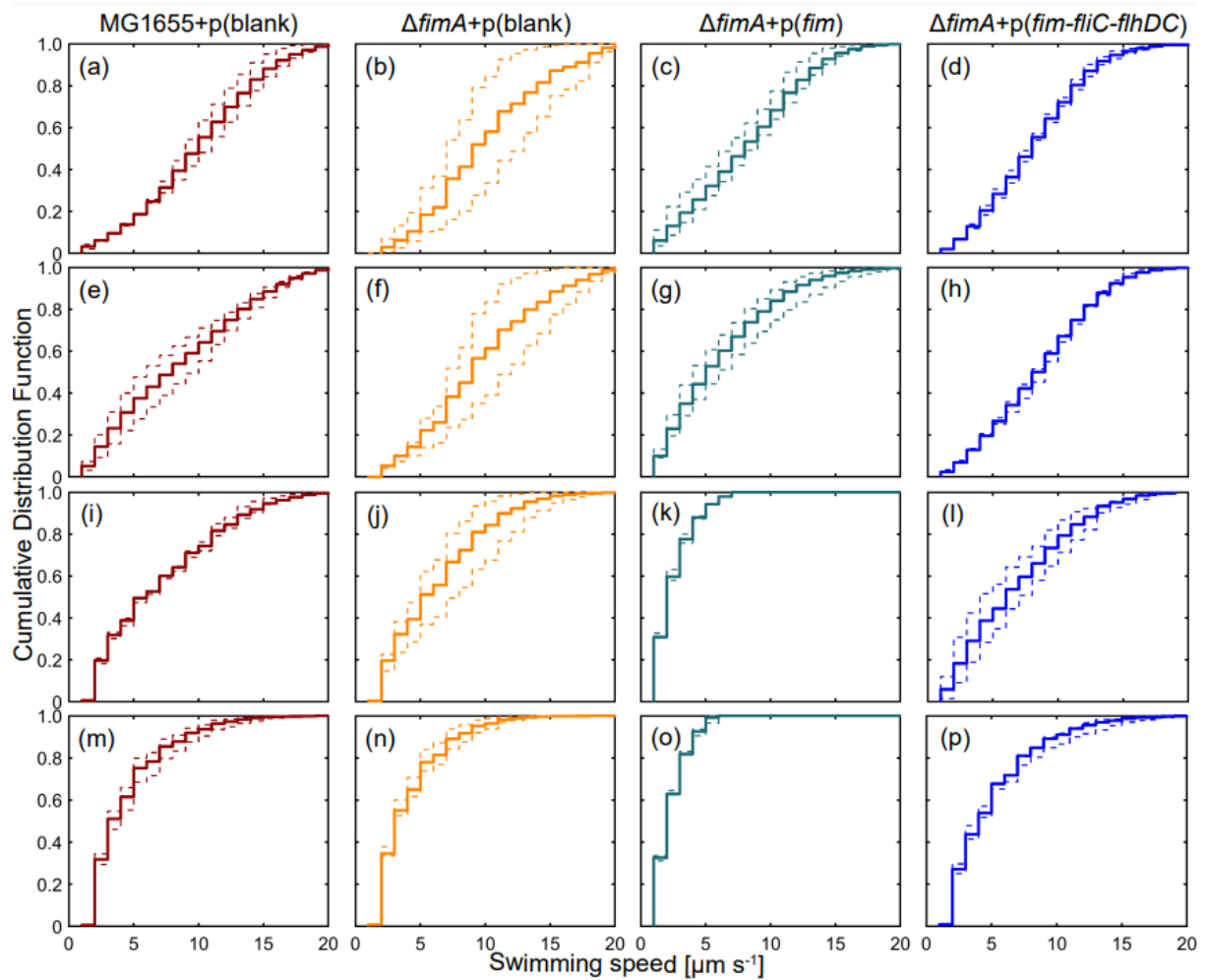


Fig. S6 The cumulative distribution function of instantaneous swimming speeds of different strains (from left to right: MG1655+p(blank), MG1655 Δ *fimA*+p(blank), MG1655 Δ *fimA*+p(*fim*), and MG1655 Δ *fimA*+p(*fim-fliC-flhDC*)) at (a – d) 0 h, (e – h) 2 h, (i – l) 6 h, and (m – p) 8 h after induction with 10 μ M IPTG.

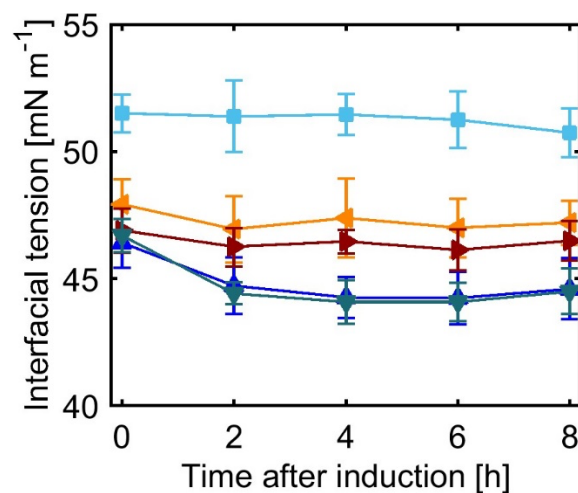


Fig. S7 Measurement of interfacial tension of the bacteria-hexadecane interface for different strains at various time points after induction with 10 μ M IPTG. Symbols: (►) MG1655+p(blank) + 10 μ M IPTG; (◄) MG1655 Δ *fimA*+p(blank) + 10 μ M IPTG; (▼) MG1655 Δ *fimA*+p(*fim*) + 10 μ M IPTG; (▲) MG1655 Δ *fimA*+p(*fim-fliC-flhDC*) + 10 μ M IPTG.

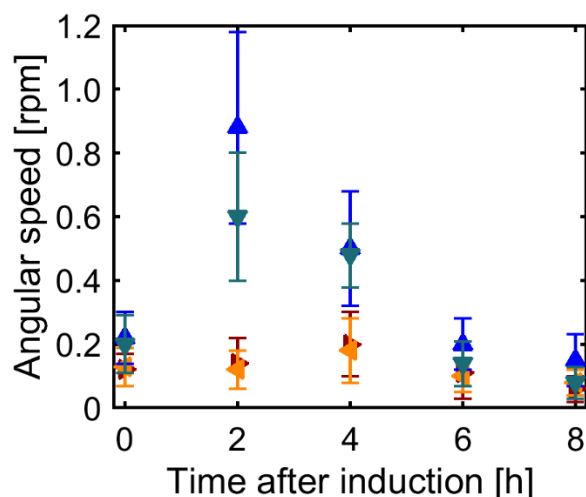


Fig. S8 Angular speed of hexadecane droplets of 25 μm diameter driven by different strains at various time points after induction with 10 μM IPTG. Symbols: (\blacktriangleright) MG1655+p(blank) + 10 μM IPTG; (\blacktriangleleft) MG1655 Δ *fimA*+p(blank) + 10 μM IPTG; (\blacktriangledown) MG1655 Δ *fimA*+p(*fim*) + 10 μM IPTG; (\blacktriangle) MG1655 Δ *fimA*+p(*fim-flhDC*) + 10 μM IPTG.

Supplementary movies:

[S1.avi](#)

[S2.avi](#)

[S3.avi](#)

[S4.avi](#)

S1.avi: Rotation behavior of hexadecane droplet driven by MG1655+p(blank) strain induced with 10 μM IPTG at cell concentration of $\text{OD}_{600} = 1$.

S2.avi: Rotation behavior of hexadecane droplet driven by MG1655 Δ *fimA*+p(blank) strain induced with 10 μM IPTG at cell concentration of $\text{OD}_{600} = 1$.

S3.avi: Rotation behavior of hexadecane droplet driven by MG1655 Δ *fimA*+p(*fim*) strain induced with 10 μM IPTG at cell concentration of $\text{OD}_{600} = 1$.

S4.avi: Rotation behavior of hexadecane droplet driven by MG1655 Δ *fimA*+p(*fim-flhDC*) strain induced with 10 μM IPTG at cell concentration of $\text{OD}_{600} = 1$.

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