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Supporting Information

Optimization of Electron Transfer Kinetics Between Photoanode and

Biocathode for Enhanced Carbon-Neutral Pollutant Removal in

Photocatalytic Fuel Cells

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Experimental section

Materials

Formate dehydrogenase from *C. boidinii* (E-FDHCB, EC 1.2.1.2) was purchased from Shanghai Chaoyan Biotechnology Co., Ltd. Cadmium acetate (C₄H₆CdO₄·2H₂O), thiourea, titanium butoxide, glutaraldehyde, 2-bromoethylamine hydrobromide, aniline, n-hexane, pentafluorobenzyl bromide (BM-PFB), bisphenol A (BPA), congo red (CR) and Aflatoxin B₂ (AFB₂) were purchased from Aladdin (Shanghai, China). Rhodamine 123 was obtained from KeyGEN Biotech (Nanjing, China). Phytic acid was obtained from Beijing Solarbio Science & Technology Co., Ltd. Tris-HCl buffer solution (0.05 M) was obtained from Beijing LABLEAD Inc. Pyruvic Acid (PA) Content Assay Kit was from Beijing Boxbio Science & Technology Co., Ltd. PBS was purchased from Sunncell. Chlortetracycline was from Chemleader Biomedical Co., Ltd. (Shanghai, China). NAD+/NADH Colorimetric Assay Kit was from Elabscience®Biotechnology Co., Ltd. All chemicals were used without further purification. Deionized water was prepared using a Milli-Q purification system with a resistivity of 18.2 MΩ.

Instrument

¹H-NMR spectra were collected from a Bruker 600 MHz spectrometer (Bruker, German). Absorbance measurement was performed through using a 2450 UV-visible spectrophotometer (Agilent, USA). Fluorescence (FL) spectra were carried out on a FluoroMax-4 spectrofluorometer with xenon discharge lamp excitation (HORIBA, USA). The morphology of CdS was characterized using scanning electron microscopy (SEM, FEI Inspect F50, America) and corresponding EDX elemental mapping images were obtained by using a JEOL model JEM 2100 (Japan). The morphology of TiO₂@CdS was characterized using high resolution transmission electron microscopy (HRTEM, Talos F200X, America) and corresponding EDX elemental mapping images were obtained by using a JEOL model JEM 2100 (Japan). X-ray diffraction (XRD) patterns of TiO₂, CdS and TiO₂@CdS were recorded by Ultima IV. X-ray photoelectron spectroscopy was carried out with Thermo ESCALAB 250XI, performed with Al K α radiation (λ = 0.8339 nm). ICP-OES was performed with Agilent 725 (USA). The Electron Paramagnetic Resonance (EPR) measurement was performed on a Bruker A300 system at room temperature. Total organic carbon (TOC) was recorded by TOC-L. The concentration of the BPA and formate were detected by a highperformance liquid chromatography (HPLC, Agilent, USA) equipped with an X Bridge column (4.6 x 250 mm, C18, 5 mm) and an UV detector. Electrochemical impedance spectroscopy (EIS, Gamry) analysis was performed in the frequency range of 105 to 0.1 Hz at a bias potential of 0 V versus Ag/AgCl. All electrochemical experiments were performed with CHI 660B.For photocurrent-time (I-T) experiments, ITO glass was cut into 4 cm × 6 cm pieces. A 0.5 mg of TiO₂, CdS and TiO₂@CdS were mixed with 100 μ L of Nafion solution (1% Nafion in CH₃CH₂OH/distilled water = 8/2) and drop-casted on the exposed area (2.65 cm × 2.65 cm) of the ITO electrode, respectively. The resulting electrode was air-dried and used as working electrode for electrochemical measurements.



Fig. S1 Schematic diagram of $TiO_2@CdS$ (a) and FDH/DA/PANi/CC (b) synthesis.

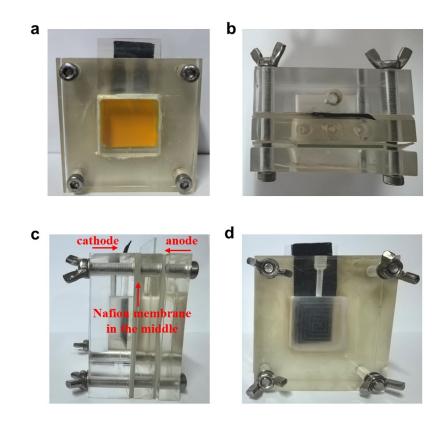


Fig. S2 Front (a), TOP (b), side (c) and back (d) view picture of PFC system.

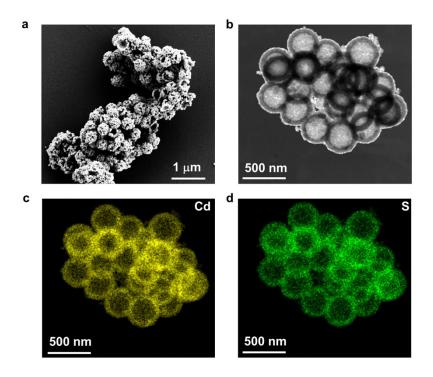


Fig. S3 SEM (a) and TEM (b) image of CdS and corresponding TEM-EDS elemental mapping images of Cd and S (c and d).

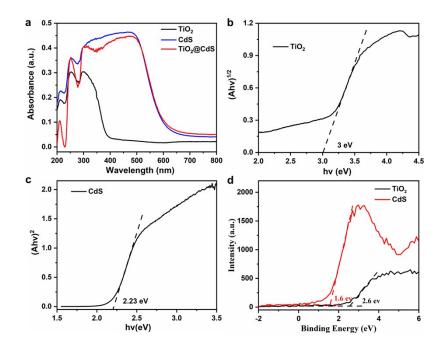


Fig. S4 (a) UV-vis diffuse reflection spectra of TiO_2 , CdS and $TiO_2@CdS$; Bandgap of TiO_2 (b) and CdS (c) estimated from the Kubelka–Munk equation according to UV-vis diffuse reflection spectra; (d) Valence band obtained from XPS.

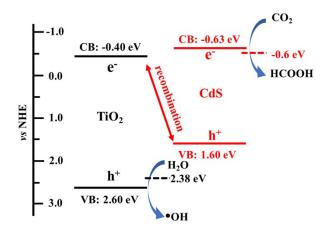


Fig. S5 The band spectrum of $TiO_2@CdS$ heterostructures for H_2O oxidated to $\cdot OH$ and CO_2 reduced to HCOOH.

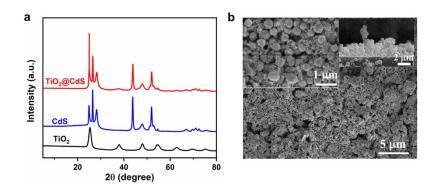


Fig. S6 (a) PXRD patterns of TiO₂, CdS and TiO₂@CdS; (b) SEM image of TiO₂@CdS/ITO (inset: the left one is a partially enlarged SEM image and the right one is the cross-sectional SEM image of TiO₂@CdS/ITO).

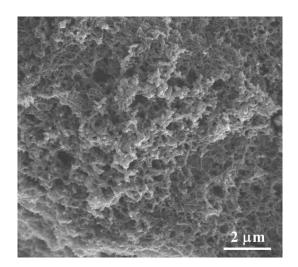


Fig. S7 SEM image of PANi/CC electrode.

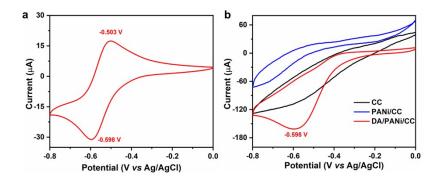


Fig. S8 (a) Cycle voltammetry curve of DA; (b) Cycle voltammetry curve of CC, PANi/CC and DA/PANi/CC.

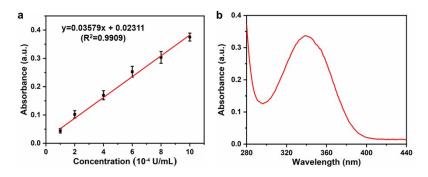


Fig. S9 (a) Standard curve of UV-vis absorbance of NADH versus concentration of FDH; (b) UV-vis absorbance of NADH of FDH/DA/PANi/CC after repeatedly washing with PBS.

After cross-linking FDH with DA/PANi/CC for 4 hours, the electrode was thoroughly washed with PBS, and the washing solutions were collected. Then, 0.1 mL of above collected solution was incubated with NAD⁺ at 37 °C for 10 min. The absorption of NADH was measured by UV-vis (Fig. S9b), which could be transformed to concertation of free NADH trough the linear equation (Fig. S9a).

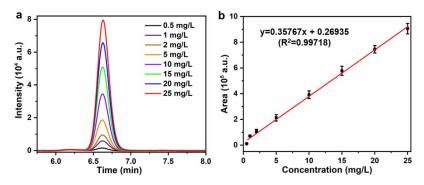


Fig. S10 (a) Curve of intensity of BPA versus its concentration; (b) Standard curve of peak area of BPA measured by HPLC versus its concentration.

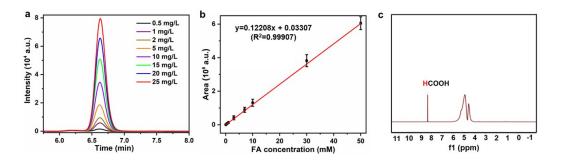


Fig. S11 (a) Curve of intensity of HCOOH versus its concentration; (b) Standard curve of peak area of HCOOH measured by HPLC versus its concentration; (c) ¹H-NMR spectrum of the solution after photocatalysis (400 μ L solution + 100 μ L D₂O).

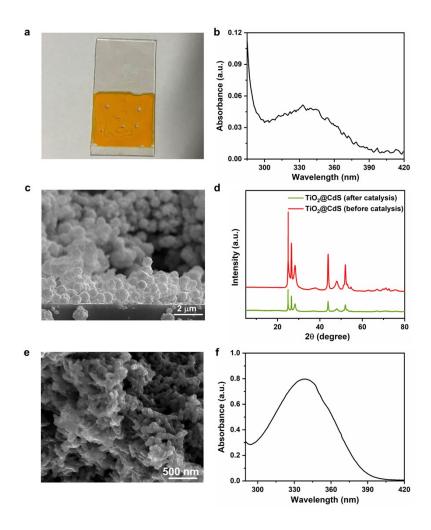


Fig. S12 (a) The image of TiO₂@CdS photoanode after photocatalytic experiment; (b) The detection of FDH of the cathode solution after photocatalytic experiment by regeneration of NADH; (c) SEM image of anode of the PFC after the 16th cycle of catalysis; (d) XRD pattern spectrum of TiO₂@CdS after the 16th cycle of catalysis; (d) SEM image of cathode of the PFC after the 16th cycle of catalysis; (f) The detection of FDH of FDH/DA/PANi/CC cathode after photocatalytic experiment by regeneration of NADH.

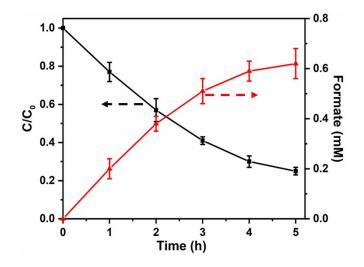


Fig. S13 The BPA degradation rate and formate production rate in the PFC system driven by the natural sunlight.

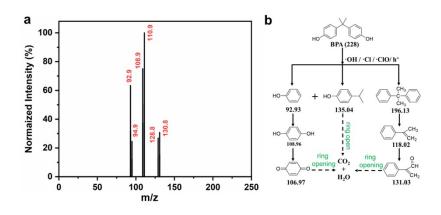


Fig. S14 (a) LC-MS spectra and proposed byproducts structure of BPA degradation after 3 h PFC reaction; (b) The proposed schematic pathway of BPA degradation via a PFC process.

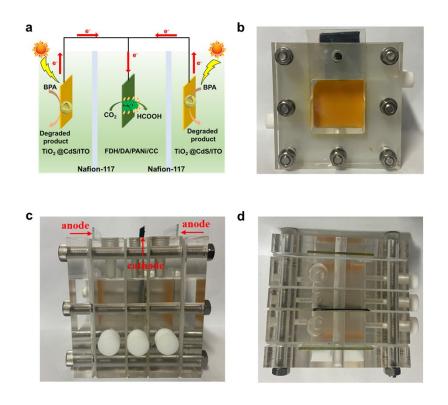


Fig. S15 (a) Schematic illustration of the 2-PFC system for BPA degradation and CO_2 reduction; Front (b), side (c) and TOP (d) view picture of 2-PFC system.

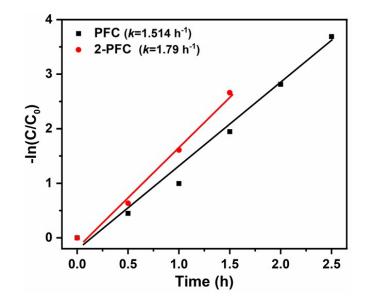


Fig. S16 Plots of -In (C/C₀) versus time for the BPA degradation by PFC and 2-PFC systems.

Anode	Cathode	Catalyst	Electron mediator	Productivity (μmol·U ⁻¹ ·h ⁻¹)	Ref
FeOOH/BiVO4/F 3D TiN-CIFDH TO		CIFDH	-	1.56	1
TiO ₂ /FTO	CFP	FDH	MV	0.0192	2
FeOOH-BiVO ₄	CIFDH-TiO ₂ -CFO	CIFDH	-	0.098	3
FTO IO- TiO ₂ dpp POs- PSII	FTO IO- TiO ₂ FDH	FDH	-	0.0925	4
TK/TiO ₂	FDH- CH₃V(CH₂)9COOH	FDH	MV derivative	0.0025	5
$Ta_3N_5 NTs$	g-C ₃ N ₄	FDH	NAD ⁺	0.1225	6
FeOOH/BiVO ₄	FDH/ITO	FDH	NAD ⁺	0.0078	7
SnTPyP/SnO ₂	RuCAT-RuC₂- <i>PolyPyr</i> -PRu/NiO	-	-	0.062 µmol∙h ⁻¹	8
CoOx/ BiVO4	NiO/PRu- PolyPyr- RuC₂RuCAT₁	-	-	0.34 μmol·h ⁻¹ (-0.7 V <i>vs</i> Ag/AgCl)	9
TiO₂@CdS/ITO	FDH/DA/PANi/C C	FDH	DA	0.74 (3.88 µmol·h⁻¹)	Our work
TiO₂@CdS/ITO (two)	FDH/DA/PANi/C C	FDH	DA	1.36 (7.13 μmol·h ⁻¹)	Our work

Table S1 Comparison of the performance of CO_2 reduction.

Anode	Cathode	Organic compound	Degrad ation rate (%)	V _{oc} (V)	J _{sc} (mA cm ⁻²)	P _{max} (μW· cm ⁻²)	Ref.
Fe@MoS ₂	Carbon fiber clothe	Berberine	92.8	0.325	0.01481	0.8	10
Carbon felt	g-FeOOH	Ametryn	98.7	0.31	-	44.6	11
3DP graphene- TiO ₂ aerogel	CNT/PVDF film	Phenol	96	-	0.63	110	12
GDH/SA- TCPP/TiO₂	CoNOC/BP	Phenol	100	0.83	0.87	296.9	13
TiO₂/ Ti	Cu ₂ O/Cu	Volatile organic compounds	22.6	0.41	0.1	20	14
WO ₃	Cu ₂ O	chlorophenol	96.8	0.65	0.34	120	15
CN-WO ₃ /W	Pt	PFOA	95	0.47	0.27	17	16
g- C ₃ N ₄ /BiOI/Ti	Cu ₂ O/Cu	RhB	95.39	0.55	0.62	103.8	17
TiO₂@CdS/IT O	FDH/DA/P ANi/CC	BPA	99.9	0.64	0.3792	94.1	Our work
TiO₂@CdS/IT O (two)	FDH/DA/P ANi/CC	BPA	99.9	0.74	1.3616	186.3	Our work

 Table S2 Performance comparison of different PFC systems.

Experimental group	TiO ₂	CdS	TiO₂@CdS
PLQY (%)	0.31	5.61	2.10

Table S3 Photoluminescence quantum yield (PLQY) of TiO₂, CdS and TiO₂@CdS.

	,	2, 2,	_	
$ au_1$ (ns)	$1/\tau_1$ (ns ⁻¹)	τ ₂ (ns)	τ_2/τ_1	τ/ns
0.0042	238	0.28	66.64	0.28
22.33	0.045	70.24	3.16	0.78
20.22	0.05	150.77	7.54	1.71
-	0.0042 22.33	τ_1 (ns) $1/\tau_1$ (ns-1)0.004223822.330.045	τ_1 (ns) $1/\tau_1$ (ns-1) τ_2 (ns)0.00422380.2822.330.04570.24	0.0042 238 0.28 66.64 22.33 0.045 70.24 3.16

Table S4 Fitting parameters of PL decay curves for TiO₂, CdS and TiO₂@CdS.

Supplemented note 1

The related equations for the generation of active species are shown as following:

$$\begin{split} h^+ + H_2O &\rightarrow \bullet OH + H^+ \\ h^+ + Cl^- &\rightarrow \bullet Cl \\ \bullet Cl + \bullet Cl &\rightarrow Cl_2 \\ Cl_2 + H_2O &\rightarrow HCIO + HCl \\ \bullet OH + HCIO &\rightarrow \bullet CIO + H_2O \\ \bullet Cl + HCIO &\rightarrow \bullet CIO + Cl^- \end{split}$$

References

- S.K. Kuk, Y. Ham, K. Gopinath, P. Boonmongkolras, Y. Lee, Y.W. Lee, S. Kondaveeti, C. Ahn, B.
 Shin, J.K. Lee, S. Jeon, C.B. Park, *Adv. Energy Mater.*, 2019, 9, 1900029.
- 2 T. Ishibashi, M. Higashi, S. Ikeda, Y. Amao, *ChemCatChem.*, 2019, **11**, 6227-6235.
- 3 S.K. Kuk, J. Jang, J. Kim, Y. Lee, Y.S. Kim, B. Koo, Y.W. Lee, J.W. Ko, B. Shin, J.K. Lee, C.B. Park, *ChemCatChem.*, 2020, **13**, 2940-2944.
- K.P. Sokol, W.E. Robinson, A.R. Oliveira, J. Warnan, M.M. Nowaczyk, A. Ruff, I.A.C. Pereira, E.
 Reisner, J. Am. Chem. Soc., 2018, 140, 116418-16422.
- Y. Amao, M. Fujimura, M. Miyazaki, A. Tadokoro, M. Nakamura, N. Shuto, *New J. Chem.*, 2018,
 42, 9269-9280.
- K.Q. Xu, A. Chatzitakis, P.H. Backe, Q.S. Ruan, J.W. Tang, F. Rise, M. Bjoras, T. Norby, Appl.
 Catal. B: Environ., 2021, 296, 120349.
- J. Kim, Y.W. Lee, E. Choi, P. Boonmongkolars, B.W. Jeon, H. Lee, S.T. Kim, S.K. Kuk, Y.H. Kim, B.
 Shin, C.B. Park, *J. Mater. Chem. A*, 2020, *8*, 88496-8502.
- F. Kuttassery, Y. Ohsaki, A. Thomas, R. Kamata, Y. Ebato, H. Kumagai, R. Nakazato, A. Sebastian,
 S. Mathew, H. Tachibana, O. Ishitani, H. Inoue, *Angew. Chem. Int. Ed.*, 2023, 62, e202308956.
- F. Kuttassery, H. Kumagai, R. Kamata, Y. Ebato, M. Higashi, H. Suzuki, R. Abe, O. Ishitani, *Chem. Sci.*, 2021, **12**, 13216-13232.
- 10 L. Xu, L.F. Liu, Appl. Catal. B: Environ., 2022, **304**, 120953.
- 11 L. Cai, H.M. Zhang, B. Dong, J. Du, Y. Tian, F. Zhang, J. Hazard. Mater., 2023, 448, 130980.
- C.J. Zhang, X.Y. Qiao, Y.H. You, Z. He, Y.F. Wang, P. Li, Y.Y. Zhang, H.L. Fu, Z.P. Yang, *Chem. Eng. J.*, 2024, **490**, 151480.
- 13 J. Ding, J.G. Zhao, H. Zhang, S.J. Dong, Biosens. Bioelectron., 2024, 266, 116714.
- 14 C.Y. Wang, Y.X. Liu, R. Chen, X. Zhu, D.D. Ye, Y. Yang, Q. Liao, *J. Hazard. Mater.*, 2023, 447, 130769.
- 15 X.F. Liu, S.J. You, N.Q. Ren, H. Zhou, J.N. Zhang, J. Hazard. Mater., 2021, 416, 11125682.
- 16 D. Zhang, W.J. Zhang, J.J. Zhang, L.M. Dong, X.P. Chen, Y.H. Guan, Z.N. Wang, Y.M. Li, *Chem. Eng. J.*, 2024, **480**, 147910.
- 17 Y.D. Zeng, Y.L. Xu, D.J. Zhong, H.Y. Yao, N.B. Zhong, J. Hazard. Mater., 2022, 425, 127967.