Electronic Supplementary Information

Biomimetic CuO/ZTF-8 nanozyme based neoteric sensor

towards selective detection of superoxide anion

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Materials and methods

Zinc oxide powder (ZnO, 99.9%) was obtained from Sigma-Aldrich Pvt., Ltd. 5-methyl tetrazole (mtz, $C_2H_4N_4$, 98.0%), copper acetate monohydrate (Cu(CO₂CH₃)₂·H₂O, 98.0%) was purchased from Tokyo Chemical Industry Co., Ltd. Methanol (CH₃OH) was purchased from Central Drug House Pvt., Ltd. Potassium dioxide (KO₂) and dopamine hydrochloride (DA) were procured from Sigma-Aldrich. Uric acid (UA), tryptophan (L-Tryp), cholesterol (Chol), glucose (Glu), ammonia solution, N, N-Dimethyl Formamide (DMF) and sodium hydroxide pellets (NaOH) were purchased from SRL chemicals, India. Hydrogen peroxide (H₂O₂) and ascorbic acid (AA) was obtained from Merck, India. Potassium perchlorate (KClO₃) was obtained from Alfa Aesar, India and potassium nitrite (KNO₂) was bought from Avra Chemicals, India. All the chemicals and solvents were used without further purification.

Instrumentation

Powder X-ray diffraction was recorded from Bruker DS advance instrument using iron-filtered Cu K α radiation (λ =1.5406 Å). Field Emission Scanning Electron Microscopic (FE-SEM) was performed using Thermo-Fisher FEI QUANTA 250 FEG under high vacuum conditions at an operating voltage range of 5 kV – 30 kV. High resolution transmission electron microscopic studies were carried out in JEOL JEM 2100 with LaB6 as electron source. X-ray photoelectron spectroscopic studies were recorded using PHI5000 Version Probe III. Electron spin resonance studies were done using JEOL JES FA200 with X-band frequency of 8.75 - 9.65 GHz. All electrochemical studies were carried out using CHI-760E electrochemical workstation. A conventional three-electrode system with glassy carbon electrode as the working electrode, Ag/AgCl as the reference electrode and platinum coil as the counter electrode were employed to conduct all the electrochemical studies.



Fig. S1 (a) UV-vis spectrum and (b) PXRD pattern of CuONP.



Fig. S2 (a and c) HRTEM images (b) size distribution curve and (d) SAED pattern of CuONP.



Fig. S3 FESEM and elemental analysis of CuO/ZTF-8 nanocomposite.



Fig. S4 XPS (a) survey spectrum and (b) O 1s spectrum of CuO/ZTF-8 nanocomposite.



Fig. S5 (a) g-factor plot of TEMPO. (b) ESR spectrum of TEMPO in the presence and absence of O_2^{-} .



Fig. S6 Absorbance spectra of 7.1 mg of KO_2 in DMSO in the presence of 18-crown-6.



Fig. S7 (a) CVs of CuO/ZTF-8/GCE in 0.1 M PBS at different pH values (pH: 5.0, 6.0, 7.0, 8.0, and 9.0) at a scan rate of 50 mV s⁻¹. (b) Effect of pH on the cathodic peak current of CuO/ZTF-8/GCE.



Fig. S8 Electrochemical impedance spectra of bare GCE, CuO/GCE, ZTF-8/GCE and CuO/ZTF-8/GCE recorded in the presence of 2.5 mM $[Fe(CN)_6]^{3-}$ in 0.1 M KCl.



Fig. S9 (a) Effect of scan rate of CuO/ZTF-8/GCE at different scan rates from 10 mV s^{-1} to 100 mV s^{-1} . (b) Plot of peak currents vs scan rate.



Fig. S10 PXRD pattern of CuO/ZTF-8 nanocomposite before and after catalysis.

Fig. S11 Amperometric response of CuO/ZTF-8/GCE at -0.2 V for successive addition of O_2^{-1} to serum sample in 0.1 M PBS (pH = 7; N₂ saturated).