

## Supporting information

for

# Bioinspired programmable coacervate droplets and self-assembled fibers through pH regulation of monomers

Satyajit Patra, Sushmitha Chandrabhas, and Subi J. George\*

New Chemistry Unit and School of Advanced Materials (SAMat), Jawaharlal Nehru Centre for Advanced Scientific Research (JNCASR), Bangalore 560064, India

Corresponding authors

\*Subi J. George

New Chemistry Unit and School of Advanced Materials (SAMat), Jawaharlal Nehru Centre for Advanced Scientific Research (JNCASR), Jakkur, Bangalore, India-560064.

[george@jncasr.ac.in](mailto:george@jncasr.ac.in)

## Table of Contents

1. General Methods and Materials
2. Synthetic Schemes and Procedures
3. Experimental Procedures
4. Supporting Figures
5. References

### 1. General Methods and Materials

**Materials:** All chemicals were purchased from commercial sources and were used as such without any further purification. Spectroscopic grade solvents were used for all spectroscopic measurements.

**Optical Measurements:** Electronic absorption spectra were recorded on JASCO V-750 UV-Visible Spectrophotometer equipped with a peltier. Fluorescence measurements were carried out on a JASCO FP-8500 spectrofluorometer equipped with a peltier. UV-Vis and emission spectra were recorded in 10 mm path-length cuvettes.

**Confocal Laser Scanning Microscopy (CLSM):** Confocal microscopy imaging was done at room temperature using Leica TCS SP8 laser scanning confocal microscope with a laser excitation of  $\lambda_{\text{exc}} = 405, 488$  nm. Imaging was performed using Leica oil-immersion objectives: HC PL APO CS2 63x/100x with numerical aperture (NA) 1.40. Fluorescence light was specially filtered with emission filters (TD 488/561/633) and imaged with HVD detector. Confocal images were processed using LAS X (Leica) and ImageJ software.

**Dynamic Light Scattering (DLS):** DLS measurements were carried out using a Zetasizer ULTRA Malvern employing a 633 nm laser at a back scattering angle of  $173^\circ$ .

### 2. Synthetic Schemes and Procedures:

NDBA molecule was synthesized according to the previously reported procedure. <sup>[s1]</sup>

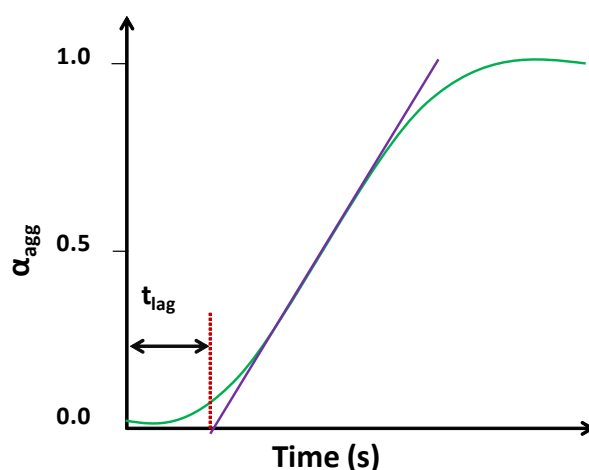
### 3. Experimental Procedure

**Protocol I:** Sample preparation protocol for self-assembly study: A stock solution of NDBA ( $c = 5 \times 10^{-3}$  M) was prepared in DMSO. An appropriate volume was injected into a mixture of DMSO and aqueous buffer to adjust the final concentration and solvent composition to the required percentages (2 % DMSO in water (v/v) for self-assembled solutions).

**Protocol II:** Preparation of sample for transient LLPS:  $5 \times 10^{-5}$  M NDBA solution at pH 9 was kept for 60 mins to get grown coacervate droplets. On to it 15 mM citric acid, 50 mM Urea, and  $25 \text{ U mL}^{-1}$  urease were added all together.

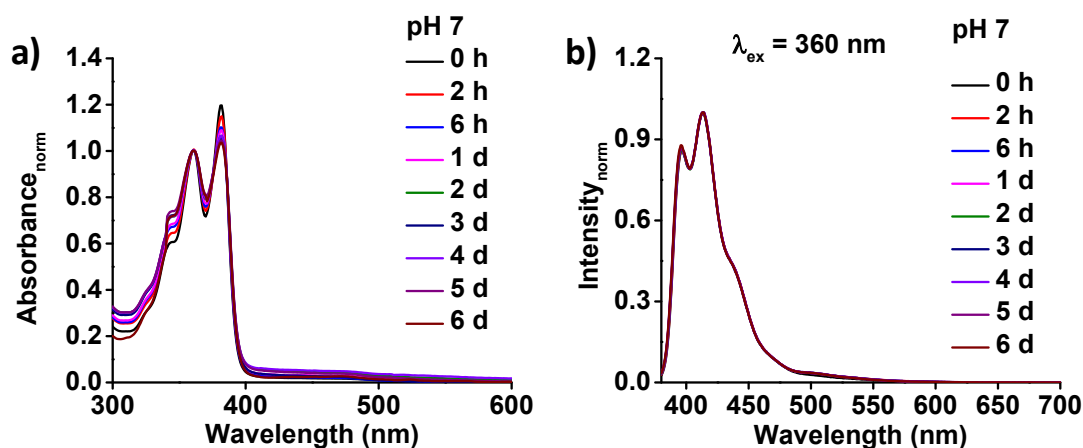
**Analysis of confocal microscopy images:** Length analysis of the confocal microscopy images was done using ImageJ software package developed by US. National Institute of Health. [s2] A frequency statistics was done on the obtained diameter.

#### Lag phase calculation:

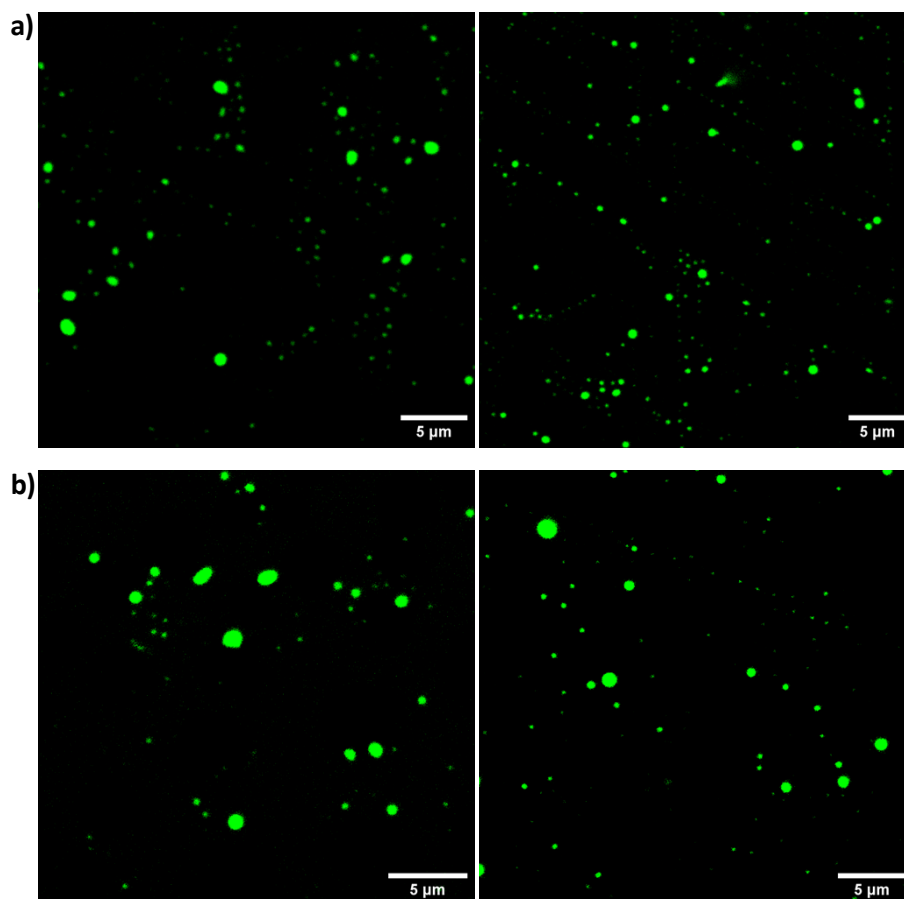


From the above-mentioned representative temporal change in the extent of aggregation ( $\alpha_{agg}$ ), the data tag (lag time, after which elongation starts) was calculated.

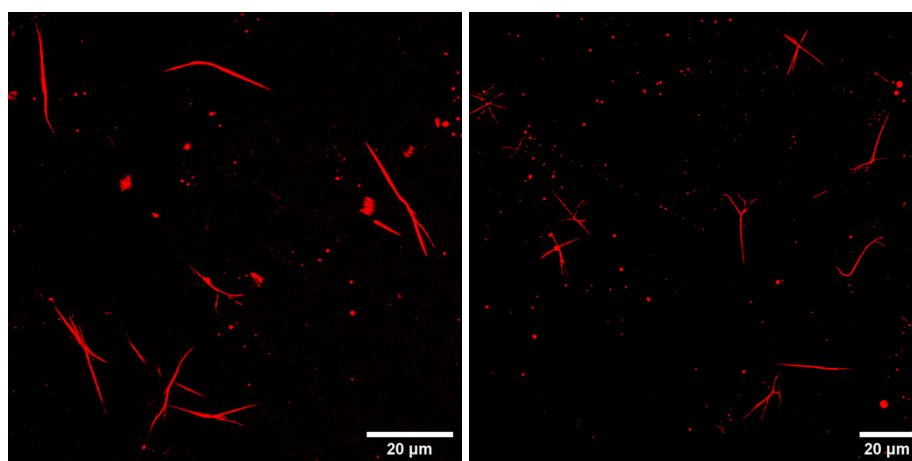
#### 4. Supporting Figures



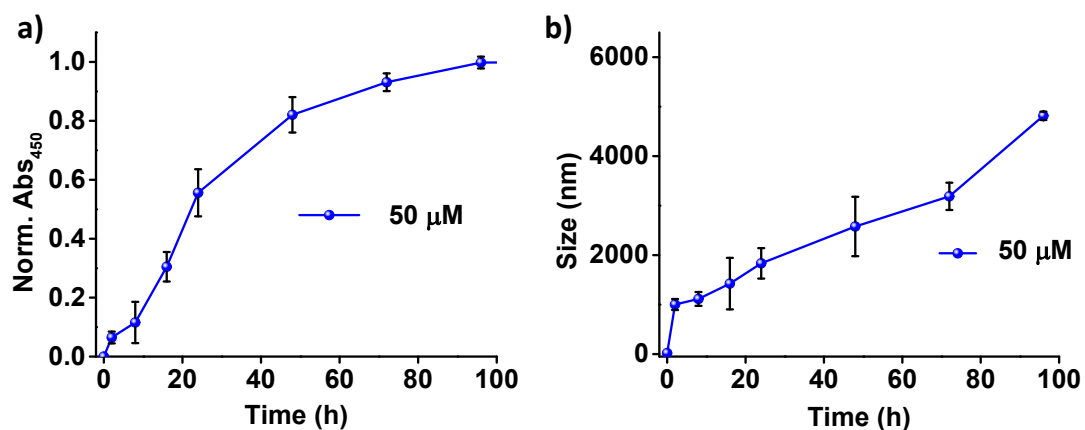
**Figure S1.** Time-dependent normalized a) absorption, and b) emission spectra ( $\lambda_{ex} = 360 \text{ nm}$ ) of NDBA monomers at pH 7.  $[\text{NDBA}] = 5 \times 10^{-5} \text{ M}$ ,  $\text{H}_2\text{O}/\text{DMSO}$ , 98/2 (v/v).



**Figure S2.** CLSM images of pH-triggered coacervate droplets with different urease units. a) 25 U<sub>mL</sub><sup>-1</sup>, and b) 50 U<sub>mL</sub><sup>-1</sup> urease. [NDBA] =  $5 \times 10^{-5}$  M, H<sub>2</sub>O/DMSO, 98/2 (v/v), 50 mM Urea.



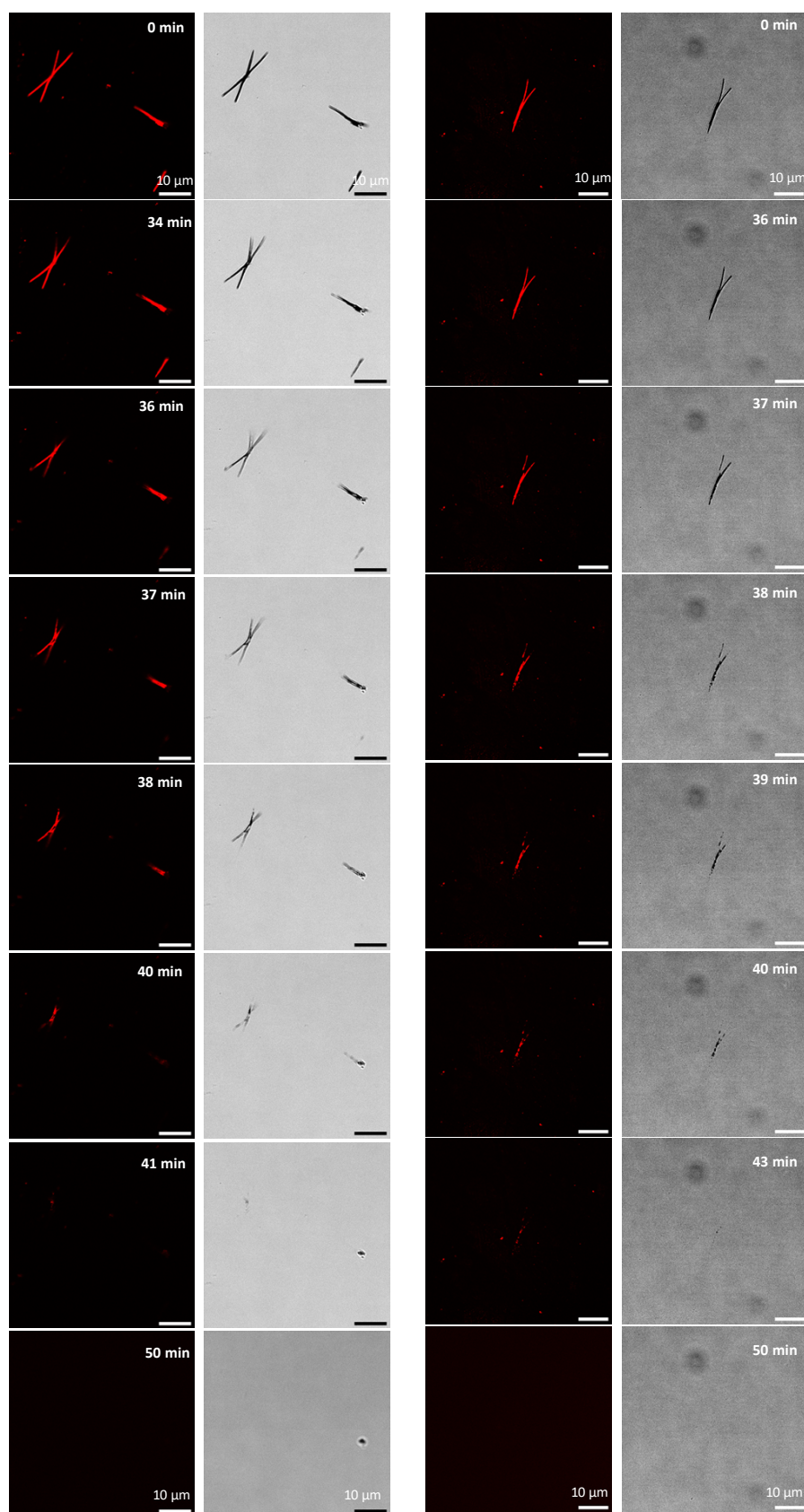
**Figure S3.** CLSM images after 96 h of the temporally grown thermodynamically stable fiber state with 1 μM Nile red. [NDBA] =  $5 \times 10^{-5}$  M, H<sub>2</sub>O/DMSO, 98/2 (v/v).



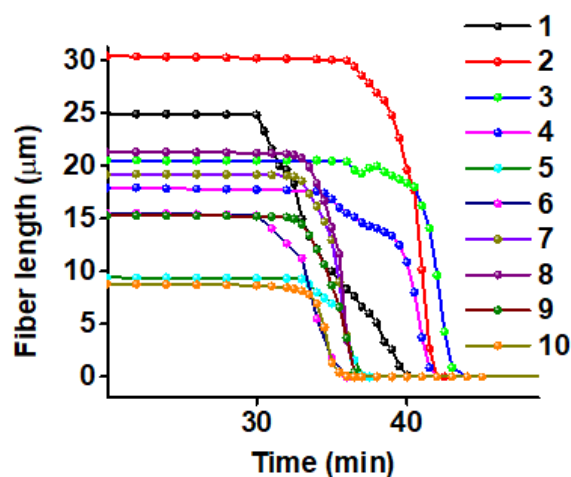
**Figure S4.** Supramolecular polymerization process was monitored by a) absorption kinetics (at 450 nm) and change in size in b) dynamic light scattering.

**Figure S5.** Time-dependent a) absorption, and b) emission spectra during the transient dissolution of coacervate droplets. 15 mM citric acid, 50 mM Urea, and 25 U mL<sup>-1</sup> urease, [NDBA] =  $5 \times 10^{-5}$  M, H<sub>2</sub>O/DMSO, 98/2 (v/v).

**Figure S6.** Disassembly study of kinetically grown fiber with 100 mM citric acid. Temporal change in a) pH profile, and b) absorption spectra after the addition of 100 mM citric acid to a kinetically grown fiber solution. [NDBA] =  $5 \times 10^{-5}$  M, H<sub>2</sub>O/DMSO, 98/2 (v/v).



**Figure S7.** Time dependent CLSM (left) and corresponding bright-field (right) images of *in-situ* visualization of fiber disassembly upon the addition of 100 mM citric acid for two different sets of experiments. Scale bar 10  $\mu\text{m}$ .  $[\text{NDBA}] = 5 \times 10^{-5} \text{ M}$ ,  $\text{H}_2\text{O}/\text{DMSO}$ , 98/2 (v/v), 1  $\mu\text{M}$  Nile Red.



**Figure S8.** The Disassembly profile was calculated from time-dependent confocal images by tracing 10 individual fibers.

Movie S1 and S2 denotes the disassembly of fibers in Fig 3 main text.

## 5. References:

1. Patra, S.; Chandrabhas, S.; Dhiman, S.; George, S. J. Controlled Supramolecular Polymerization via Bioinspired, Liquid–Liquid Phase Separation of Monomers. *J. Am. Chem. Soc.* **2024**, *146*, 12577–12586.
2. Schneider, C. A.; Rasband, W. S.; Eliceiri, K. W. NIH Image to ImageJ: 25 years of image analysis. *Nat. Methods* **2012**, *9*, 671-675. DOI: 10.1038/nmeth.2089.