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Supplementary Information

2 Liquid Metal-based Sticky Conductor for Wearable and Real-Time Electromyogram
3 Monitoring with Machine Learning Classification

4

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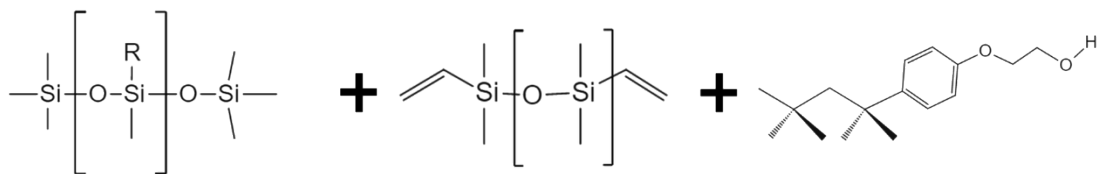
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23 **Key words:** Liquid Metal, Electromyogram, Soft electronics, Wearable Devices,
24 Signal Processing

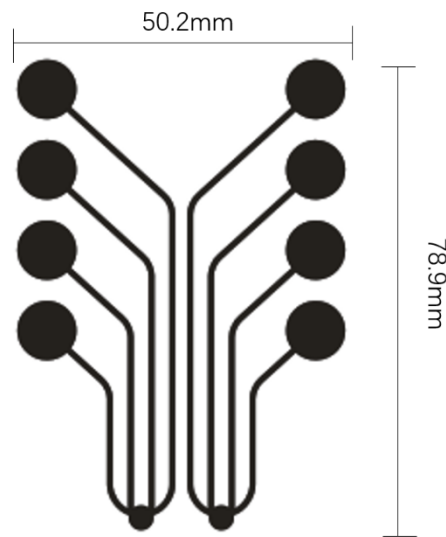


25 PDMS base

PDMS crosslinker

Triton-X

26 **Figure S1.** The molecular structure we have used for the modification of PDMS.



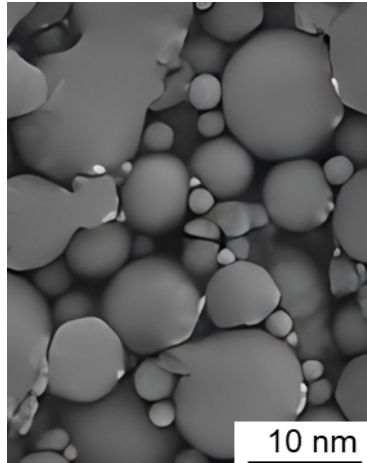
27
28 **Figure S2.** The design of the EMG electrode (50.2 mm wide, 78.9 mm long)
29



30

31 **Figure S3.** Photo of the fabricated LM ink.

32

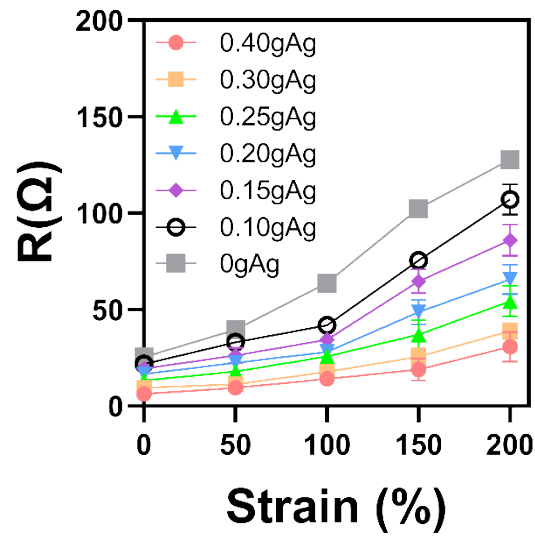


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34 **Figure S4.** SEM images of the LM ink without AgNWs.

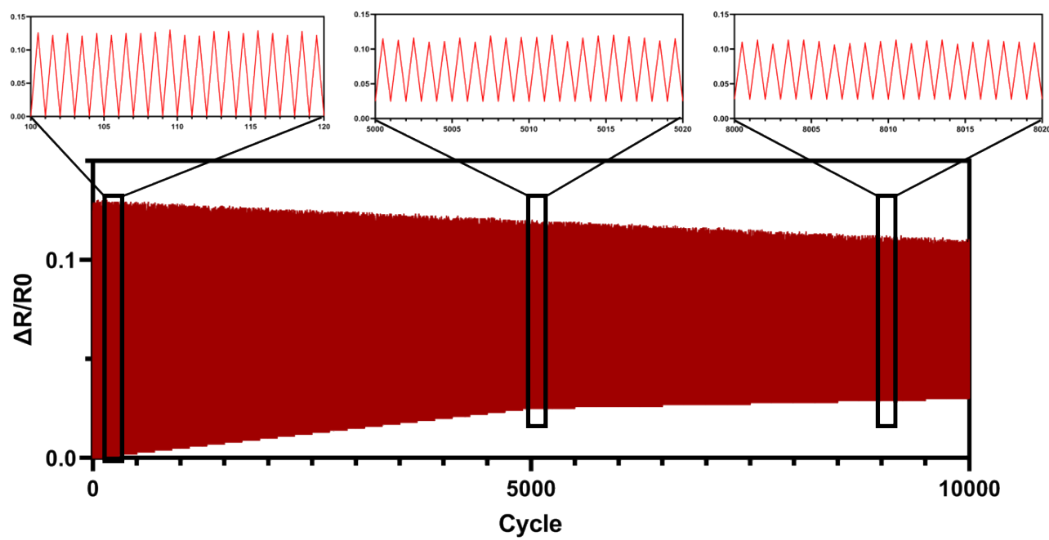
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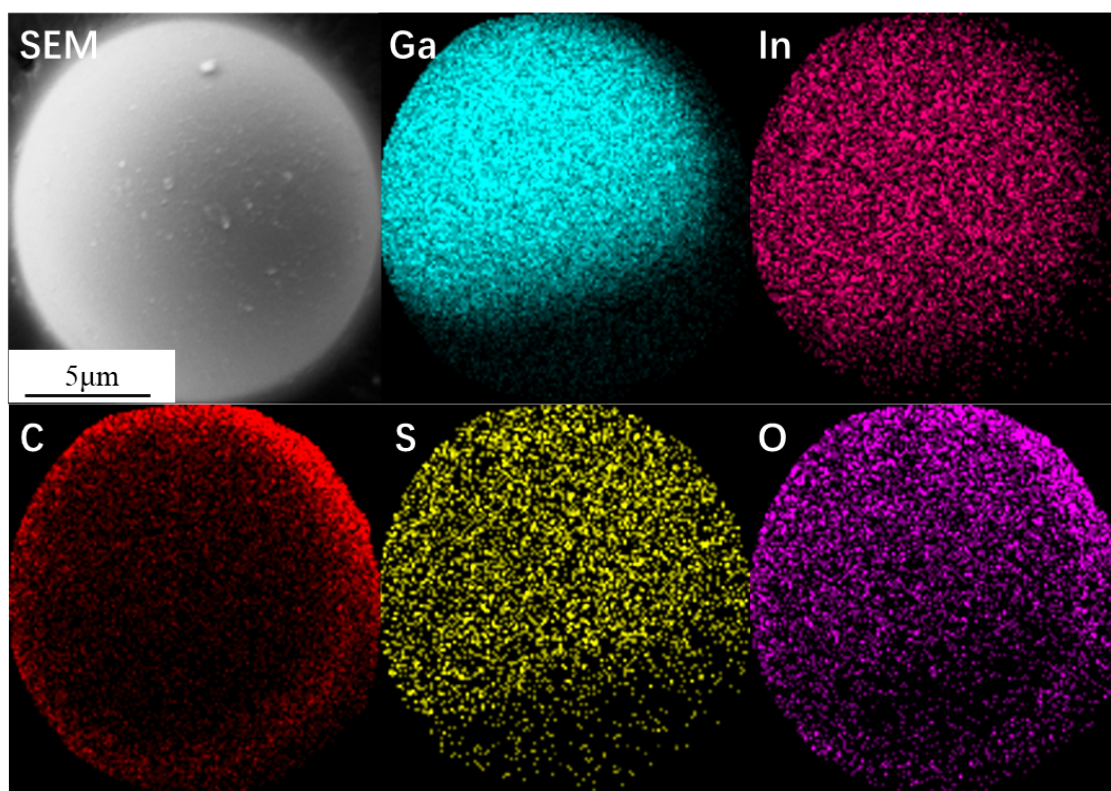
38 **Figure S5.** Resistance versus strain (%) for different concentrations of Ag (0.40g, 0.30g, 0.25g,
 39 0.20g, 0.15g, 0.10g, 0g). Error bars indicate the standard deviation of the measured resistance
 40 change values, showing the variability and reliability of the measurements over multiple
 41 experiments. Each data point represents the average of multiple independent experiments. Different
 42 color markers correspond to different silver concentrations, indicating the repeatability and
 43 reproducibility of the results under the same experimental conditions.



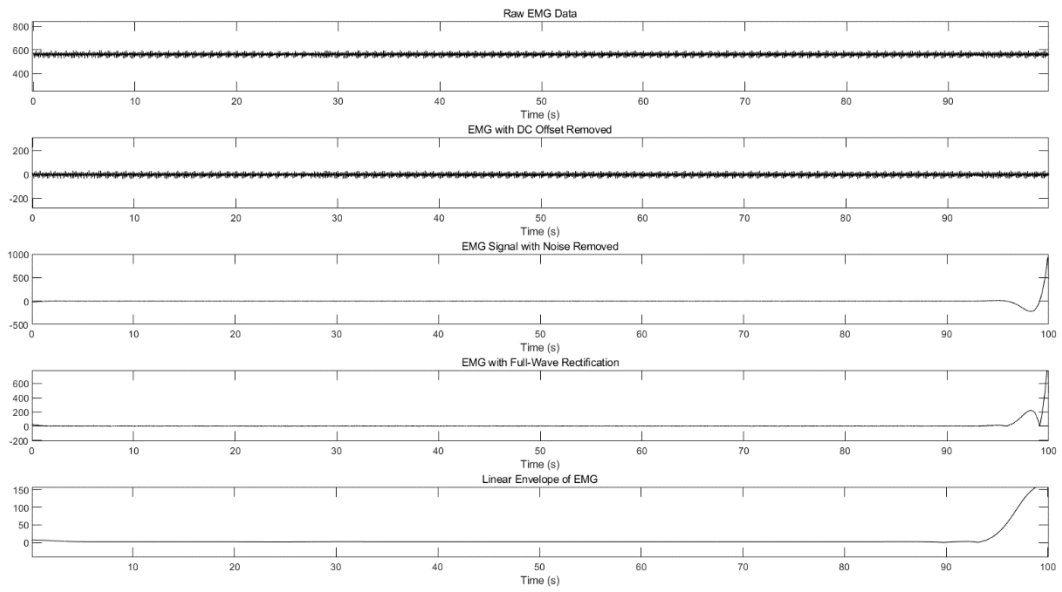
44

45 **Figure S6.** Stability data for resistance changes of the LM-PDMS electrode over
46 10,000 cycles

47

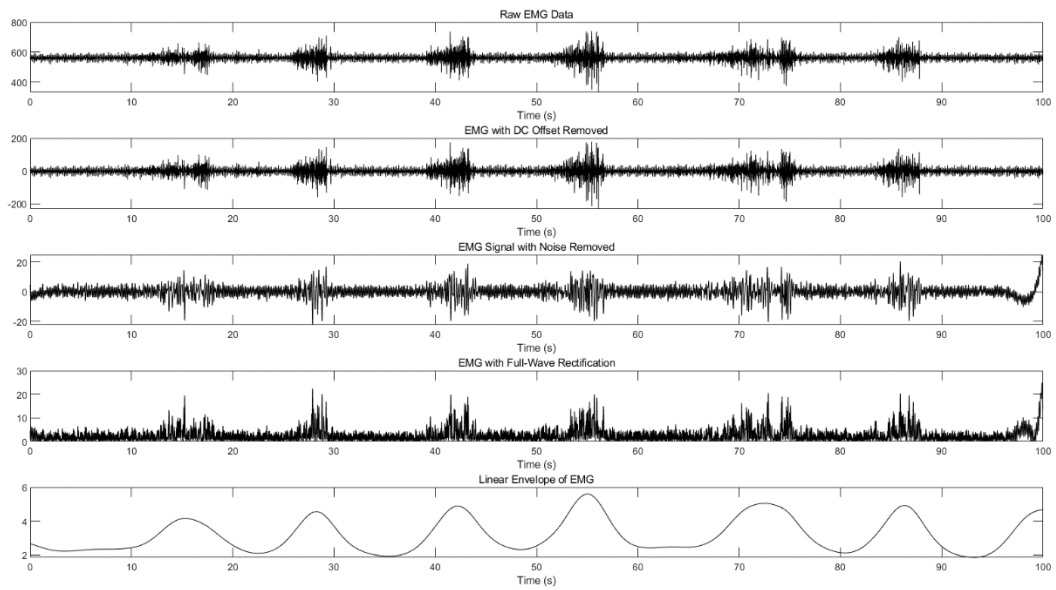


48
49 **Figure S7.** EDS images of LM-PDMS particles.
50



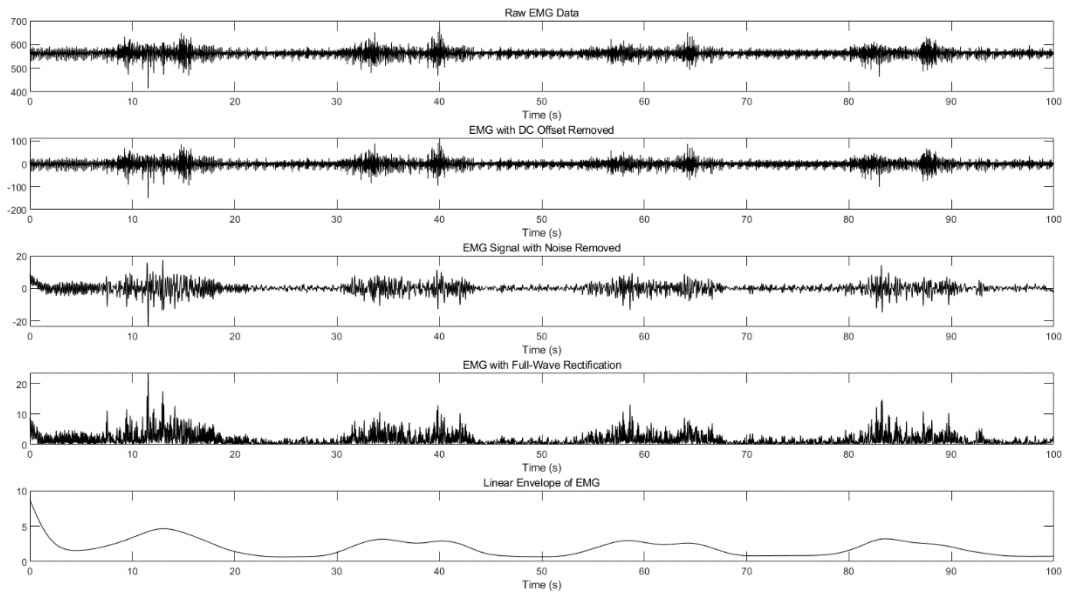
51

52 **Figure S8.** The raw EMG data in the resting state and its corresponding curve after multi-step
 53 processing.



54

55 **Figure S9.** The raw EMG data from a clenched fist and the resultant curve following multi-step
 56 processing.



57

58 **Figure S10.** The raw EMG data from the forearm during wrist rotation and the plotted curve after
 59 multi-step processing.

60

Ref.	Stable Resistance Strain Range (%)
Our study	50
[1]	36
[2]	40
[3]	26
[4]	60
[5]	100
[6]	30
[7]	4
[8]	60

61 **Table S1.** Comparison of stable resistance strain ranges in different studies

62

63 1. Lee, H.; Lee, S.; Kim, J.; Jung, H.; Yoon, K. J.; Gandla, S.; Park, H.; Kim, S., *npj Flexible*
64 *Electronics* **2023**, *7* (1), 20. DOI 10.1038/s41528-023-00246-3.

65 2. Zhao, Y.; Chen, C.; Lu, B.; Zhu, X.; Gu, G., *Advanced Functional Materials* **2024**, *34* (14), 2312480.
66 DOI <https://doi.org/10.1002/adfm.202312480>.

67 3. Gong, Q.; Jiang, X.; Liu, Y.; Yu, M.; Hu, Y., *Advanced Electronic Materials* **2023**, *9* (9), 2200916.
68 DOI <https://doi.org/10.1002/aelm.202200916>.

69 4. Zhao, Q.; Gribkova, E.; Shen, Y.; Cui, J.; Naughton, N.; Liu, L.; Seo, J.; Tong, B.; Gazzola, M.;
70 Gillette, R.; Zhao, H., *Science Advances* *10* (18), eadn7202. DOI 10.1126/sciadv.adn7202.

71 5. Wang, W.; Wang, S.; Rastak, R.; Ochiai, Y.; Niu, S.; Jiang, Y.; Arunachala, P. K.; Zheng, Y.; Xu,
72 J.; Matsuhisa, N.; Yan, X.; Kwon, S.-K.; Miyakawa, M.; Zhang, Z.; Ning, R.; Foudeh, A. M.; Yun, Y.;
73 Linder, C.; Tok, J. B. H.; Bao, Z., *Nature Electronics* **2021**, *4* (2), 143-150. DOI 10.1038/s41928-020-
74 00525-1.

75 6. Zhang, L.; Kumar, K. S.; He, H.; Cai, C. J.; He, X.; Gao, H.; Yue, S.; Li, C.; Seet, R. C.-S.; Ren, H.;
76 Ouyang, J., *Nature Communications* **2020**, *11* (1), 4683. DOI 10.1038/s41467-020-18503-8.

77 7. Kim, T.; Park, J.; Sohn, J.; Cho, D.; Jeon, S., *ACS Nano* **2016**, *10* (4), 4770-4778. DOI
78 10.1021/acsnano.6b01355.

79 8. Jang, K.-I.; Han, S. Y.; Xu, S.; Mathewson, K. E.; Zhang, Y.; Jeong, J.-W.; Kim, G.-T.; Webb, R.
80 C.; Lee, J. W.; Dawidezyk, T. J.; Kim, R. H.; Song, Y. M.; Yeo, W.-H.; Kim, S.; Cheng, H.; Rhee, S. I.;
81 Chung, J.; Kim, B.; Chung, H. U.; Lee, D.; Yang, Y.; Cho, M.; Gaspar, J. G.; Carbonari, R.; Fabiani, M.;

82 Gratton, G.; Huang, Y.; Rogers, J. A., *Nature Communications* **2014**, 5 (1), 4779. DOI
83 10.1038/ncomms5779

```

84 close all;
85 clear all;
86 clc
87 % Set sampling frequency and read EMG data
88 fs = 10000; % Sampling frequency (Hz)
89 EMG_DATA = xlsread('data.xlsx'); % Read the data from an Excel file
90 % Extract individual channels from the data
91 EMG_DATA_1 = EMG_DATA(:,1);
92 EMG_DATA_2 = EMG_DATA(:,2);
93 EMG_DATA_3 = EMG_DATA(:,3);
94 % Process each channel with the EMG processing function, including wavelet analysis
95 [emg_data_1, t, wResults1, waveletPower1] = processemg(EMG_DATA_1, fs, 6, 2);
96 [emg_data_2, t, wResults2, waveletPower2] = processemg(EMG_DATA_2, fs, 6, 2);
97 [emg_data_3, t, wResults3, waveletPower3] = processemg(EMG_DATA_3, fs, 6, 2);
98 % Normalize the processed EMG data based on the max contraction value
99 m_EMG_DATA_1 = max(emg_data_1);
100 perc_EMG_DATA_1 = (emg_data_1 / m_EMG_DATA_1) * 100;
101 m_EMG_DATA_2 = max(emg_data_2);
102 perc_EMG_DATA_2 = (emg_data_2 / m_EMG_DATA_2) * 100;
103 m_EMG_DATA_3 = max(emg_data_3);
104 perc_EMG_DATA_3 = (emg_data_3 / m_EMG_DATA_3) * 100;
105 % Create a new figure for EMG percentage of MVC plot
106 figure;
107 subplot(3,1,1);
108 plot(t, perc_EMG_DATA_1, 'black', 'LineWidth', 1.5);
109 xlabel('Time(s)');
110 ylabel('EMG (%MVC)');
111 legend('ARM Muscle in silent')
112 subplot(3,1,2);
113 plot(t, perc_EMG_DATA_2, 'black', 'LineWidth', 1.5);
114 xlabel('Time(s)');
115 ylabel('EMG (%MVC)');
116 legend('ARM Muscle with wrist twisting')
117 subplot(3,1,3);
118 plot(t, perc_EMG_DATA_3, 'black', 'LineWidth', 1.5);
119 xlabel('Time(s)');
120 ylabel('EMG (%MVC)');
121 title('EMG of ARM during Squat (%MVC)');
122 legend('ARM Muscle with fist up')

```

123 **Code S1.** The annotated MATLAB main function code for processing the forearm EMG signals.

```

124 function [output, t, waveletResult, waveletPower] = processemg(EMGRAW, fs, LOWPASSRATE,
125 NUMPASSES)
126     % Ensure NUMPASSES is reasonable, often 2 or 4.
127     % Handle NaNs or Infinities here if they exist.
128     EMGRAW(~isfinite(EMGRAW)) = 0;
129     % Read data and create time vector
130     t = linspace(0, (length(EMGRAW)-1)/fs, length(EMGRAW))*100;
131     % Plot raw EMG data
132     figure();
133     subplot(5,1,1);
134     plot(t,EMGRAW,'black');
135     xlabel('Time (s)');
136     title('Raw EMG Data');
137     % Remove DC offset
138     EMGDC = EMGRAW - mean(EMGRAW);
139     subplot(5,1,2);
140     plot(t,EMGDC,'black');
141     xlabel('Time (s)');
142     title('EMG with DC Offset Removed');
143     % Filter the noise with bandpass filter
144     Wn1 = 20/(fs/2);
145     Wn2 = 600/(fs/2);
146     [b,a] = butter(NUMPASSES,[Wn1 Wn2],'bandpass');
147     EMGFILT = filtfilt(b,a,EMGDC);
148     subplot(5,1,3);
149     plot(t,EMGFILT,'black');
150     xlabel('Time (s)');
151     title('EMG Signal with Noise Removed');
152     % Full-wave rectification
153     EMGFWR = abs(EMGFILT);
154     subplot(5,1,4);
155     plot(t,EMGFWR,'black');
156     xlabel('Time (s)');
157     title('EMG with Full-Wave Rectification');
158     % Wavelet decomposition
159     waveletName = 'db9';
160     level = wmaxlev(length(EMGFWR), waveletName);
161     [C,L] = wavedec(EMGFWR, level, waveletName);
162     % Store wavelet result
163     waveletResult.cfs = C;

```



```

164 waveletResult.lvl = L;
165 waveletResult.detailCfs = cell(1, level);
166 waveletResult.approxCfs = appcoef(C, L, waveletName);
167 for i = 1:level
168     waveletResult.detailCfs{i} = detcoef(C, L, i);
169 end
170 % Create wavelet power spectrum
171 waveletPower = cell(1, level);
172 for i = 1:level
173     waveletPower{i} = abs(waveletResult.detailCfs{i}).^2;
174 end
175 % Linear Envelope
176 Wn = LOWPASSRATE/(fs/4);
177 [b,a] = butter(NUMPASSES, Wn, 'low');
178 EMGLE = abs(filtfilt(b, a, EMGFWR));
179 % Optionally remove the end spike
180 subplot(5,1,5);
181 plot(t,EMGLE,'black');
182 xlabel('Time (s)');
183 title('Linear Envelope of EMG');
184 output = EMGLE; % Return the linear envelope result
185 end
186
187 Code S2. The MATLAB function code designed for sEMG signal processing, including removing
188 DC offset, denoising through filtering, and extracting the linear envelope.
189

```

```

190 import pandas as pd
191 import numpy as np
192 import scipy.fft as sp_fft
193 from scipy.signal import butter, filtfilt
194 from sklearn.model_selection import train_test_split, GridSearchCV
195 from sklearn.neighbors import KNeighborsClassifier
196 from sklearn.metrics import classification_report, accuracy_score
197 from sklearn.svm import SVC
198 from sklearn.preprocessing import StandardScaler
199 from sklearn.ensemble import RandomForestClassifier
200 from imblearn.over_sampling import SMOTE # Import SMOTE for handling class imbalance
201 # Load the data
202 file_path = 'D:/matlab/Fist up.xlsx'
203 data = pd.read_excel(file_path)
204 # Preprocessing function
205 def preprocess_signals(data, lowcut=20, highcut=500, fs=1000, order=5):
206     data_centered = data - data.mean()
207     nyq = 0.5 * fs
208     low = max(lowcut / nyq, 0.01)
209     high = min(highcut / nyq, 0.99)
210     b, a = butter(order, [low, high], btype='band')
211     filtered_data = filtfilt(b, a, data_centered)
212     return filtered_data
213 # Apply preprocessing
214 preprocessed_data = data.apply(preprocess_signals, axis=0)
215 def enhanced_features(data):
216     data_array = data.values
217     fft_values = sp_fft.fft(data_array)
218     psd = np.abs(fft_values) ** 2
219     normalized_psd = psd / np.sum(psd)
220     mean_val = np.mean(data_array)
221     std_val = np.std(data_array)
222     # Flatten the feature array properly
223     return np.concatenate([normalized_psd, [mean_val, std_val]])
224 # Apply enhanced feature extraction and collect features in a proper format
225 feature_list = []
226 for column in preprocessed_data.columns:
227     features = enhanced_features(preprocessed_data[column])
228     feature_list.append(features)

```

```

229 # Convert list of arrays into a 2D array
230 feature_matrix = np.array(feature_list)
231 # Prepare labels
232 labels = ['fist'] * 10 + ['wrist'] * 10
233 # Handling class imbalance
234 smote = SMOTE()
235 X_res, y_res = smote.fit_resample(feature_matrix, labels)
236 # Train-test split
237 X_train, X_test, y_train, y_test = train_test_split(X_res, y_res, test_size=0.3, random_state=42)
238 # Classifier with hyperparameter tuning
239 model = RandomForestClassifier()
240 param_grid = {'n_estimators': [10, 50, 100], 'max_depth': [None, 10, 20, 30]}
241 grid_search = GridSearchCV(model, param_grid, cv=5)
242 grid_search.fit(X_train, y_train)
243 # Predict and evaluate
244 y_pred = grid_search.predict(X_test)
245 accuracy = accuracy_score(y_test, y_pred)
246 report = classification_report(y_test, y_pred)
247 print("Best parameters:", grid_search.best_params_)
248 print("Accuracy:", accuracy)
249 print("Classification Report:\n", report)
250
251 Code S3. The Python code designed for EMG signal processing and classification using
252 random forest.
253
254

```