Supplementary Information for

Development of a Xanthene-Based NIR Fluorescent Probe for Accurate and Sensitive Detection of γ-Glutamyl Transpeptidase in Cancer Diagnosis and Treatment

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Synthesis of probe XM-Glu



Scheme S1 Synthetic route of probe XM-Glu.



Scheme S2 Proposed reaction mechanism of XM-Glu reacted with GGT.

Synthesis of 2-bromocyclohex-1-ene-1-carbaldehyde (1)¹

DMF (7.8 mL, 100 mmol) was mixed with 20 mL anhydrous chloroform. The reaction mixture was cooled to 0°C under N₂ condition. Then PBr₃ (3.8 mL, 40 mmol) was added dropwise to the solution, and it was stirred for 30 minutes. Cyclohexanone (2.06 mL, 20 mmol) was then added, and was reacted for another 16 hours at 25°C. After the reaction, the mixture was poured into ice H₂O and adjusted to pH 7 with solid NaHCO₃. The solution was extracted with DCM and H₂O. The organic layer was then dried over MgSO₄ and the solvent was removed to yield orange oil. The crude product was directly used in next step without further purification.¹H NMR (400 MHz, CDCl₃) δ 9.90 (s, 1H), 2.65 (m, 2H), 2.17 (m, 2H), 1.67 (m, 2H), 1.59 (m, 2H). ¹³C NMR(100MHz, CDCl₃) δ 194.0, 144.0, 135.4, 38.5, 24.6, 23.9, 20.7.

Synthesis of 6-methoxy-2,3-dihydro-1*H*-xanthene-4-carbaldehyde $(2)^{1}$

Compound **1** (1 g, 5.32 mmol), 2-hydroxy-4-methoxybenzaldehyde (402 mg, 2.64 mmol) and cesium carbonate (5 g, 15.35 mmol) were dissolved in 10 mL anhydrous DMF. The reaction mixture was stirred for 16 hours at room temperature. After the reaction, the mixture was filtered and the filtrate was extracted with DCM and H₂O for three times. The organic layer was then dried over MgSO₄ and the solvent was removed. The crude product was purified with column chromatography (Hexane/EtOAc = 4:1) to obtain compound **2** as a yellow solid (210 mg, 32%). ¹H NMR (400 MHz, CDCl₃) δ 10.32 (s, 1H), 7.08 (d, J = 9.2 Hz, 1H), 6.67-6.63 (m, 3H), 3.84 (s, 3H), 2.57 (t, J = 5.96 Hz, 2H), 2.45 (t, J = 6.08 Hz, 2H), 1.72 (tt, J₁= 6.16 Hz, J₂ = 5.96 Hz, 2H). ¹³C

NMR (100 MHz) δ 187.6, 161.4, 160.8, 153.4, 127.4, 126.8, 126.6, 114.6, 112.6, 110.9, 100.5, 55.6, 29.9, 21.5, 20.4. LRMS m/z (FD) calcd. for C₁₅H₁₄O₃: 242.1; found for: 242.1.

Synthesis of 2-((6-methoxy-2,3-dihydro-1*H*-xanthen-4yl)methylene)malononitrile (3)²

Compound **2** (100 mg, 0.41 mmol) and malononitrile (41 mg, 0.62 mmol) were dissolved in 10 mL acetic anhydride. And the reaction mixture was refluxed under N₂ condition for 16 hours. After the reaction, 20 mL methanol was added and the reaction was refluxed for another 2 hr. The reaction mixture was concentrated, which was then purified by column chromatography with Hexane/EtOAc 3:1 (v/v) as the eluent to obtain compound **3** as a deep red solid (80 mg, 67%).¹H NMR (400 MHz, CDCl₃) δ 8.06 (s, 1H), 7.18 (d, J = 8.3 Hz, 1H), 6.90 (s, 1H), 6.80 – 6.75 (m, 2H), 3.89 (s, 3H), 2.87 (t, J = 6.0 Hz, 2H), 2.61 (t, J = 5.76 Hz, 2H), 1.83 (tt, J₁= 6.0 Hz, J₂ = 5.76 Hz, 2H). ¹³C (100 MHz) δ 162.3, 158.6, 154.0, 150.3, 130.8, 127.9, 126.1, 117.6, 115.7, 114.8, 112.9, 109.8, 100.1, 70.2, 55.8, 29.1, 24.7, 20.5. MS m/z (FD) calcd. for C₁₈H₁₄N₂O₂: 290.1; found for: 290.1.

Synthesis of 2-((6-hydroxy-2,3-dihydro-1*H*-xanthen-4-

yl)methylene)malononitrile (XM-OH)²

Compound **3** (252 mg, 0.87 mmol) was dissolved in 30 mL ultra-dry DCM, and BBr₃ (1.7 mL, 17.37 mmol) was added at 0°C under N₂ condition. The reaction mixture was stirred at 0°C for 1 hour and refluxed for another 16 hours. After the reaction, the reaction mixture was neutralized by saturated NaHCO₃ solution at 0°C, which was extracted with DCM and H₂O. The organic layer was then dried over MgSO₄ and the solvent was evaporated. The crude product was purified by column chromatography with Hexane/EtOAc 2:1 (v/v) as the eluent to obtain **XM-OH** as a deep red solid (144 mg, 60%).¹H NMR (400 MHz, DMSO-d₆) δ 10.6 (s, 1H), 8.15 (s, 1H), 7.37 (d, J = 8.44

Hz, 1H), 7.34 (s, 1H), 6.91 (d, J = 2.04 Hz, 1H), 6.75 (dd, $J_1 = 8.44$ Hz, $J_2 = 2.04$ Hz, 1H), 2.73 (t, J = 6.0 Hz, 2H), 2.60 (t, J = 5.76 Hz, 2H), 1.74 (tt, $J_1 = 6.0$ Hz, $J_2 = 5.76$ Hz, 2H). ¹³C NMR (150 MHz, DMSO-d₆) 116.89, 114.25, 114.03, 109.33, 102.67, 66.65, 28.46, 24.77, 20.51 LRMS m/z (FD) calcd. for $C_{17}H_{12}N_2O_2$: 276.1; found for: 276.1.

Synthesis of tert-butyl N²-(tert-butoxycarbonyl)-N⁵-(4-(hydroxymethyl)phenyl)glutaminate (4)³

N-(tert-butoxycarbonyl)glutamic acid tert-butyl ester (Boc-Glu-OtBu) (455 mg, 1.5 mmol), hexafluorophosphate benzotriazole tetramethyl uranium (HBTU) (569 mg, 1.5 mmol) and N,N-diisopropylethylamine (DIPEA) (412 µL, 3 mmol) were added to anhydrous THF (8 mL) and stirred for 30 minutes. Then the reaction mixture was treated with p-aminobenzeyl alcohol (222 mg, 1.8 mmol) and stirred at 0°C for 10 minutes and then at room temperature for 3 hours. After the reaction, the solvent was removed under reduced pressure. The reaction mixture was extracted with EtOAc and brines. The organic layer was then dried over MgSO₄ and the solvent was evaporated. The crude product was purified by column chromatography with Hexane/EtOAc 1:1 (v/v) as the eluent to obtain compound 4 as a light yellow solid (560 mg, 91%).¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ (Figure S22) δ 8.84 (s, 1H), 7.60 (d, J = 8.12 Hz, 2H), 7.32 (d, J = 8.12 Hz, 2H), 5.36 (d, J = 7.60 Hz 1H), 4.64 (d, J = 4.00 Hz 2H), 4.22 (m, 1H), 2.43 (m, 2H), 2.26 (m, 2H), 1.46 (s, 18H) ¹³C NMR (100 MHz, CDCl₃) (Figure **S23**) δ 171.24, 170.58, 156.56, 137.88, 136.50, 127.75, 119.83, 82.75, 80.51, 65.01, 5 3.20, 34.14, 30.68, 28.30, 27.96 LRMS m/z (FD) (Figure S24) calcd. for C₂₁H₃₂N₂O₆: 408.2; found for: 408.2.

Synthesis of *tert*-butyl N⁵-(4-(bromomethyl)phenyl)-N²-(tertbutoxycarbonyl)glutaminate (5)³

Compound 4 (244 mg, 0.6 mmol) was dissolved in anhydrous THF (10 mL), and

PBr₃ (57 μ L, 0.6 mmol) was added at 0°C under N₂ condition. The reaction mixture was stirred at 0°C for 1 hours. Then, the reaction mixture was added to a saturated NaHCO₃ solution (20 mL) at 0°C, which was then extracted with EtOAc and H₂O. The organic layer was then dried over MgSO₄ and the solvent was evaporated. The crude product was used in next step without further purification. (172 mg, 61%).

Preparation of stock solution

Solutions of cysteine, homocysteine, glutathione, hydrogen peroxide, and HOCl were prepared in distilled water at a concentration of 1 mM. Enzymes including acetylcholinesterase, alkaline phosphatase, b-galactosidase, g-glutamyl transpeptidase, and tyrosinase were produced at 1U/mL in 10 mM PBS buffer. Singlet oxygen was created by combining 1 mM hydrogen peroxide with sodium hypochlorite, while O2-was created by dissolving 1 mM potassium superoxide in distilled water. Solutions with pH ranging from 4.0 to 12.0 were prepared in a 10 mM PBS buffer by adding various concentrations of 1M HCl or 1M NaOH. For photophysical studies, a solution of **XM-Glu** (20 μ M) was prepared by diluting the stock solution in a mixture of H₂O and DMSO (volume ratio of 6:4) with 6 mM PBS buffer (pH 7.4).

Cytotoxicity test

The cytotoxicity of the probe **XM-Glu** probe was evaluated using MTT tests. HepG2 and HEK293 cells were planted on 96 well plates with 200 L Dulbecco modified Eagle medium (DMEM) and incubated for 24 hours at 37°C under 5% CO₂. Various concentrations of XM-Glu (0, 5, 10, 15, and 25 μ M) were incubated with the cells for an additional 24 hours. The cells were treated with 1 mg/mL of 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) for 4 hours, and then the purple crystals were dissolved in DMSO. The absorbance (570 nm) was measured by microplate readers. The following equation shows the calculation for cell viability:



Figure S1 (a) Absorbance changes at 610 nm and (b) fluorescence changes at 648 nm of XM-Glu (20 μ M) in response to various concentration of GGT (0-100 mU/mL) in DMSO-H₂O (v/v = 4/6, 6 mM PBS buffer, pH 7.4) solution for 4 h. λ_{ex} = 595 nm.



Figure S2 The fluorescence intensity at 648 nm of the probe **XM-Glu** (20 μ M) in response to GGT (0-20 mU/mL) in DMSO-H₂O (v/v = 4/6, 6 mM PBS, pH 7.4) solution for 4 h. (λ_{ex} = 595 nm) The detection limit was calculated to be 0.067 mU/mL.



Figure S3 (a) The fluorescence intensity at 648 nm of fluorophore **XM-OH** (0 ~ 12 μ M) in DMSO- H₂O (v/v = 4/6, 6 mM PBS, pH 7.4) solution. (b) Lineweaver-Burk plot of probe **XM-Glu** in response to GGT (100 mU/mL). λ_{ex} = 595 nm



Figure S4 The fluorescence changes of **XM-Glu** (20 μ M), and **XM-Glu** (20 μ M) with GGT (100 mU/mL) in different pH value of DMSO-H₂O (v/v = 4/6, 6 mM PBS) solution for 4 h. λ_{ex} = 595 nm.



Figure S5 HPLC chromatograms of fluorophore XM-OH (100 μ M), probe XM-Glu (100 μ M), and XM-Glu (100 μ M) + GGT (400 mU/mL). The absorption peak was measured at 520 nm. (HPLC conditions: 0 min: 80% H₂O + 20% MeOH, 5 ~ 15 min: 4.5 % H₂O + 95.5% MeOH, 20 min: 100% MeOH)



Figure S6 ESI-MS spectra of XM-Glu reacted with GGT.



Figure S7 Cell viability of HepG2 and HEK293 cells treated with XM-Glu (0, 5, 10, 15, 20, 25 μ M) at 37°C for 24 h. The results are the mean and standard deviation of three independent experiments.



Figure S8 Fluorescence images of HeLa cells. (a ~ d) Control group. (e ~ h) HeLa cells were incubated with **XM-Glu** (10 μ M) at 37°C for 2 h (Blue fluorescence: $\lambda_{ex} = 405$ nm. $\lambda_{em} = 435 \sim 485$ nm. Red fluorescence: $\lambda_{ex} = 561$ nm. $\lambda_{em} = 600 \sim 700$ nm).



Figure S9 Fluorescence images of HEK293 cells. Control group (a–d). Experimental group treated with **XM-Glu** (10 μ M) at 37°C for 2 hours (e–h). DAPI fluorescence: Excitation wavelength = 405 nm, Emission wavelength = 435–485 nm. Red fluorescence: Excitation wavelength = 561 nm, Emission wavelength = 600–700 nm.



Figure S10 Fluorescence intensity at tumor site pre-injection, 5 min after injection, 1 h after injection, and 2 h after injection.





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Figure S11 (a) Fluorescence imaging of tissue sections from tumors and organs of mouse incubated with XM-Glu (50 μ M) for 2 h. Fluorescence: $\lambda_{ex} = 545$ nm. $\lambda_{em} = 590 \sim 650$ nm. (b) Relative intensity plots of the images.



Figure S12 ¹H NMR (400 MHz) spectrum of Compound 1 in CDCl₃.



Figure S13 ¹³C NMR (100 MHz) spectrum of Compound 1 in CDCl₃.



Figure S15¹³C NMR (100 MHz) spectrum of Compound 2 in CDCl₃.



Figure S16 FD mass spectrum of compound 2.



Figure S17 ¹H NMR (400 MHz) spectrum of Compound 3 in CDCl₃.



Figure S18¹³C NMR (100 MHz) spectrum of Compound 3 in CDCl₃.



Figure S19 FD mass spectrum of compound 3



CARBON_01 XM-OH-3 V159.8467 -159.8467 -154.2220 -150.1393 ~133.3049 -129.1481 ~125.0081 ~118.2869 ~118.2869 ~114.2473 ~114.0330 ~109.3282 -102.6659 ~28.4588 -24.7689 -20.5132 -66.6548 -38 -36 -34 -32 HO -30 114.0300 W ∥N 10 -28 -26 N -24 -20 -18 NHA'Y 'Y HAV -16 114 f1 (ppm) 113 -14 -12 170 160 150 140 130 70 60 50 120 110 100 90 fl (ppm) 80 40 30 20 10 0

Figure S21 ¹³C NMR (150 MHz) spectrum of XM-OH in DMSO-d₆.



Figure S22 FD mass spectrum of XM-OH.



Figure S23 ¹H NMR (400 MHz) spectrum of Compound 4 in CDCl₃.



Figure S24 ¹³C NMR (100 MHz) spectrum of Compound 4 in CDCl₃.



Figure S25 FD mass spectrum of compound 4.



Figure S26 ¹H NMR (600 MHz) spectrum of Compound 6 in DMSO-d₆.



Figure S27¹³C NMR (150 MHz) spectrum of Compound 6 in DMSO-d₆.



Figure S28 FD mass spectrum of compound 6.



Figure S29 HR-FD mass spectrum of compound 6.



Figure S30 ¹H NMR (400 MHz) spectrum of XM-Glu in DMSO-d₆.



Figure S31 ¹³C NMR (150 MHz) spectrum of XM-Glu in DMSO-d₆.



Figure S32 ESI mass spectrum of XM-Glu.



Figure S33 HR-ESI mass spectrum of XM-Glu.

Table S1 Published fluorescent probe	s for the detection of GGT.
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	Probe structure	$\lambda_{ex}/\lambda_{em}$	Detection limit (mU / mL)	K _m (μM) V _{max} (μM s ⁻¹) k _{cat} (s ⁻¹)	Ref.
$HOOC \underbrace{\begin{array}{c} 0 \\ NH_2 \end{array}}_{(R=n\cdot C_{12}H_{11})} \underbrace{\begin{array}{c} 0 \\ N \\ 0 \\ R \end{array}}_{(R=n\cdot C_{12}H_{11})} \underbrace{\begin{array}{c} 0 \\ N \\ 0 \\ R \end{array}}_{(R=n\cdot C_{12}H_{11})} \underbrace{\begin{array}{c} 0 \\ N \\ 0 \\ R \end{array}}_{(R=n\cdot C_{12}H_{11})} \underbrace{\begin{array}{c} 0 \\ N \\ N \\ R \end{array}}_{(R=n\cdot C_{12}H_{11})} \underbrace{\begin{array}{c} 0 \\ N \\$	451 nm /	1.47	N.D.	4	
	610 nm				

	390 nm / 640 nm	2.27	$K_m = 19.3$ $k_{cat} = 0.118$	5
C C NH NH NH NH NH NH NH NH NH NH	680 nm / 727 nm	0.4	$K_{m} = 4.264$ $V_{max} = 0.04$	6
$ \begin{pmatrix} 0 & 0 & 0 & 0 \\ HO & 1 & 0 & 0 & 0 \\ \vdots & \vdots & \vdots & \vdots & \vdots \\ HO & 1 & 0 & 0 & 0 & 0 \\ \vdots & \vdots & \vdots & \vdots & \vdots \\ \vdots & \vdots & \vdots & \vdots & \vdots$	687 nm / 714 nm	N.D.	N.D.	7
$\begin{array}{c} \overline{o_3}S \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$	710 nm / 770 nm	N.D.	K _m =16	8
O O NH NH2 OH	355 nm / 500 nm	0.3	K _m =9.8	9
	365 nm / 650 nm	2.9	K _m =7.68	10
	408 nm / 550 nm	0.76	N.D.	11
$\begin{array}{c} HOOC\\ CN \\ F \\ $	405 nm / 490 nm	0.117	N.D.	12
HO J HN CO CI O'O	λ_{em} =540 nm (Chemilumin escence)	0.016	N.D.	13

	680 nm / 720 nm	0.0036	$K_m = 1.26$ $k_{cat} = 0.004$	3
with the stand of	600 nm / 670 nm	0.0785	K _m =6.62 V _{max} =0.103	14
	λ _{em} = 600 nm (Bioluminesc ence)	0.192	$K_m = 12.57$ $V_{max} = 0.057$ $k_{cat} = 0.917$	15
HO J HN G O J HN G S SCOOH	λ_{em} = 600 nm (Bioluminesc ence)	0.442	$K_m = 18.42$ $V_{max} = 0.031$ $k_{cat} = 0.5$	15
	585 nm / 615 nm	0.0056	K _m =7.64	16
H ₂ N COOH	578 nm / 601 nm	N.D.	$K_{m} = 18.7$ $V_{max} = 0.06$	17
	530 nm / 565 nm	N.D.	N.D.	18
	540 nm / 640 nm	0.03	$K_{\rm m} = 9.87$ $V_{\rm max} = 0.021$	19
$(R = (CH_2)_{0}CO_2H)$	460 nm / 557 nm	0.15	N.D.	20
	354 nm / 473 nm	0.21	$K_{m} = 17.64$ $V_{max} = 0.024$	21

H ₂ N O COOH NH ₂	496 nm / 525 nm	N.D.	$K_m = 145$ $V_{max} = 0.051$ $k_{cat} = 0.078$	22
	555 nm / 582 nm	N.D.	$K_m = 35.4$ $V_{max} = 0.01$ $k_{cat} = 77.7$	23
N SI N H2 NH2	637 nm / 662 nm	N.D.	N.D.	24
	360 nm / 472 nm	0.59	K _m =15.17 V _{max} =0.018	25
NC NC NC NC NC NC NC NC NC NC NC NC NC N	510 nm / 613 nm	0.0379	K _m =11.48	26
NC CN NC CN NC O NC O	490 nm / 635 nm	0.057	K _m =10.27	27
CI HN O COOH COOH R (R = c-RGD)	660 nm / 712 nm	0.0029	$K_m = 1.85$ $V_{max} = 0.000109$ $k_{cat} = 0.005$	28
COOH NH N≓ HOOC ''NH ₂	680 nm / 708 nm	0.5	K _m =7.01	29
	595 nm / 645 nm	0.067	$K_m = 21.46$ $V_{max} = 0.0023$ $k_{cat} = 0.0094$	This work

*N.D. Not determined.

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