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Supplementary Information For:

An aluminium-organic framework unveiling ultra-sensitive fluorometric detection of pesticide paraoxon-methyl and pharmaceutical drug azathioprine in fruits, vegetables, and wastewater

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Materials and Characterization Methods:

The synthesis and characterisation procedures for 2-((2-hydroxy-4-methoxy benzyl) amino) terephthalic acid linker was followed by previously reported procedure.¹ All the chemicals were purchased from commercial sources and used without further purification. Attenuated Total Reflectance Infrared (ATR-IR) spectroscopy data were recorded in the region 400-4000 cm⁻¹ at room temperature with the Perkin Elmer Spectrum ATR-IR spectrometer. The following indications were used to indicate the corresponding absorption bands: very strong (vs), strong (s), medium (m), weak (w), shoulder (sh) and broad (br). Thermogravimetric (TG) experiments were carried out with a heating rate of 4 °C min⁻¹ under N₂ atmosphere using a SDT Q600 thermogravimetric analyser. Powder X-ray diffraction (PXRD) data were collected in transmission mode using Rigaku Smartlab X-ray diffractometer with Cu-Ka radiation ($\lambda = 1.54056$ Å), 40 kV of operating voltage and 125 mA of operating current. Specific surface area for N₂ sorption was calculated on a Quantachrome Autosorb iQMP gas sorption analyser at -196 °C. FE-SEM images were collected with a Zeiss (Sigma 300) scanning electron microscope. FE-TEM images were collected with JEOL transmission electron microscope having model number 2100F. Fluorescence emission studies were performed at room temperature using a HORIBA JOBIN YVON Fluoromax-4 spectrofluorometer. Pawley refinement was carried out using Materials Studio software.² Computational analysis was performed by using the B3LYP function and the Pople diffuse basis set 6-31G+(d,p) in Gaussian 09W software.³ The DICVOL program incorporated within STOE's WinXPow software package was used to determine the lattice parameters.⁴

Preparation of MOF (1') Suspension for Fluorescence Titration Experiment:

The probe 1' (4 mg) was collected in a 5 mL glass vial and 4 mL Milli-Q water was added to it to make a homogeneous suspension. Then, the suspension was sonicated for 30 min and kept it for overnight to make the suspension stable. During the fluorescence titration experiment, we used 200 μ L of the above-mentioned suspension of 1', and 3000 μ L of Milli-Q water was added to it in a quartz cuvette. All the fluorescence spectra were collected by exciting the suspension at 328 nm, within the range of 340-600 nm. Paraoxon-methyl and azathioprine solutions were prepared by dissolving in methanol and DMSO, respectively. The aforementioned solutions were recorded in the same range.

Fluorometric Detection of Azathioprine in Human Blood Serum Sample:

From the right arm vein of a healthy human, 10 mL of blood sample was collected. The sample was centrifuged at 10,000 rpm for 15 min to obtain blood plasma, from which the light-yellow blood serum was collected and stored in a Falcon tube at -20 °C. To conduct fluorescence detection experiments, varying concentrations of azathioprine were added to different aliquots of the human blood serum sample containing MOF suspension.

Fluorometric Detection of Azathioprine in Human Urine Sample:

A 10 mL urine sample was collected from a healthy individual and treated with 500 mL of HNO₃ to eliminate any interfering living organisms. The sample was then centrifuged at 8000 rpm for 10 min, and the supernatant was used for the experiments. To conduct fluorescence

experiments, various amounts of azathioprine were added to the urine samples containing MOF suspension.

Calculation of Corrected Fluorescence Emission Intensity:

The corrected fluorescence intensity was calculated using the following equation:

$$\frac{F_{corrected}}{F_{observed}} = \frac{2.3dA_{exc}}{1 - 10^{-dA_{exc}}} 10^{gA_{em}} \frac{2.3sA_{em}}{1 - 10^{-sA_{em}}}$$

where F_{observed} is the maximum fluorescence intensity; F corrected is the corrected fluorescence intensity, which is the fluorescence intensity obtained after removing the inner filter effect; A_{exc} and A_{em} represent the absorbance at the excitation wavelength of probe ($\lambda_{\text{exc}} = 328 \text{ nm}$) and maximum emission wavelength ($\lambda_{\text{em}} = 428 \text{ nm}$), respectively.; *d* is the width of the quartz cell (d = 1.0 cm); *g* is the distance between the edges of the cuvette and the excitation beam (0.4 cm in this case); *s* is the thickness of excitation light (s = 0.1 cm).



Figure S1. ATR-IR spectra of (a) 2-((2-hydroxy-4-methoxy benzyl amino) terephthalic acid linker (b) as-synthesised **1** (red) and (b) activated **1'** (black).



 Table S1. Unit cell parameters of 1 compared with those of reported Al-MIL-53.

Compound Name	[Al(OH)(L)] ·0.5H2O (1)	Al-MIL-53	
	(this work)	(reported) ⁵	
Crystal System	Orthorhombic	Orthorhombic	
$a \neq b \neq c (Å)$	$16.590(14) \neq 12.942(8) \neq$	$16.893(3) \neq 12.592(2) \neq$	
	6.588(5)	6.623(2)	
$\alpha = \beta = \gamma (^{\circ})$	90	90	
V (Å ³)	1414.6(22)	1408.8 (5)	
Radiation	Cu Kal	Cu Kal	



(red).



Figure S4. EDX image of 1.



Figure S5. Thermogravimetric analysis curves of as-synthesized 1 (black) and activated 1' (red) recorded under N_2 atmosphere in the temperature range of 30-700 °C with a heating rate of 4 °C min⁻¹.



Figure S6. PXRD patterns of (a) non-treated 1' and 1' after stirring in (b) MeOH, (c) DMSO, (d) H_2O , (e) pH = 2, and (f) pH = 12 for 24 h.



Figure S7. Excitation (black) and emission (red) spectra of 1' in water.



Figure S8. Time-dependent variation in the fluorescence spectrum of 1' after addition of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S9. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM bisphenol A solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S10. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM aqueous cyromazine solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S11. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM dodin solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S12. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM flonicamid solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S13. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM paraquat solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S14. Fluorescence quenching intensity of the aqueous solution of **1'** after gradual addition of 300 μ L of 10 mM glufosinate ammonium solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S15. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM bisphenol F solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S16. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 10 mM chlortoluron solution in presence of 300 μ L of 10 mM solution of paraoxon-methyl.



Figure S17. Change of fluorescence intensity of 1' as a function of the concentration of paraoxon-methyl in water.

	0.5H ₂ O					work
7	$[Al(OH)(C_{16}H_{13}NO_6)]$	water	38.5×10 ⁶	0.30	10	this
		buffer				
6	TPE-Peptide	phosphate	-	600.00	900	11
	ThT@ZnCPs					
5	$Ru(bpy)_3^{2+}$ and	water	-	0.93	3600	10
4	Eu-NDC MOF	ethanol	8.05×10^{3}	85.13	30	9
3	CdTe / ZnS QDs	water	-	0.65		8
	nanocomposite					
2	C based fluorescent	water	-	1250.00	-	7
1	B, N CDs	water	-	99.94	120	6
No.		Medium	(M ⁻¹)		Time (s)	
S1.	Probe	Sensing	$K_{\rm SV}$	LOD (nM)	Response	Ref.

Table S2. Comparison of the response time, detection limit, K_{sv} and sensing media used for the reported fluorometric sensors for paraoxon-methyl sensing.



Figure S18. Photoluminescence spectrum for recyclability test of probe **1'** recovered from 1st cycle for the sensing of 10 mM paraoxon-methyl in aqueous medium.



Figure S19. Photoluminescence spectrum for recyclability test of probe **1'** recovered from 2nd cycle for the sensing of 10 mM paraoxon-methyl in aqueous medium.



Figure S20. Photoluminescence spectrum for recyclability test of probe 1' recovered from 3rd cycle for the sensing of 10 mM paraoxon-methyl in aqueous medium.



Figure S21. Photoluminescence spectrum for recyclability test of probe 1' recovered from 4th cycle for the sensing of 10 mM paraoxon-methyl in aqueous medium.



Figure S22. Recyclability test of probe 1' for the sensing of 10 mM paraoxon-methyl in aqueous medium.



Figure S23. Stern-Volmer plot for the fluorescence quenching of 1' in presence of paraoxonmethyl in aqueous medium.



Figure S24. Variation of quenching efficiency after addition of paraoxon-methyl to a suspension of 1' in aqueous solution of wide range of pH.



Figure S25. Time-dependent variation in the fluorescence spectrum of 1' after addition of $300 \ \mu L$ of 5 mM solution of azathioprine.



Figure S26. Fluorescence intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous arginine solution in the presence of 300 μ L of 5 mM solution of azathioprine.



Figure S27. Fluorescence intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous histidine solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S28. Fluorescence intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous glycine solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S29. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous Mg²⁺ solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S30. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous Zn²⁺ solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S31. Fluorescence quenching intensity of the solution of 1' after gradual addition of 300 μ L of 5 mM aqueous methionine solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S32. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous glutamic acid solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S33. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous K⁺ solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S34. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous Cl⁻ solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S35. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous F⁻ solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S36. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous phenylalanine solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S37. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous proline solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S38. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous serine solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S39. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous tryptophan solution in presence of 300 μ L of 5 mM solution of azathioprine.



Figure S40. Fluorescence quenching intensity of the aqueous solution of 1' after gradual addition of 300 μ L of 5 mM aqueous Na⁺ solution in the presence of 300 μ L of 5 mM solution of azathioprine.



Figure S41. Photoluminescence spectrum for recyclability test of probe **1'** recovered from 1st cycle for the sensing of 5 mM azathioprine in aqueous medium.



Figure S42. Photoluminescence spectrum for recyclability test of probe 1' recovered from 2nd cycle for the sensing of 5 mM azathioprine in aqueous medium.



Figure S43. Photoluminescence spectrum for recyclability test of probe **1'** recovered from 3rd cycle for the sensing of 5 mM azathioprine in aqueous medium.



Figure S44. Photoluminescence spectrum for recyclability test of probe 1' recovered from 4th cycle for the sensing of 5 mM azathioprine in aqueous medium.



Figure S45. Recyclability test of probe 1' for the sensing of 5 mM azathioprine in aqueous medium.



Figure S46. Change of fluorescence intensity of 1' as a function of the concentration of azathioprine in water.



Figure S47. Stern-Volmer plot for the fluorescence quenching of 1' in the presence of azathioprine in aqueous medium.



Figure S48. ATR-IR spectra of 1' before sensing (black), after azathioprine sensing (red) and after paraoxon-methyl sensing (blue).



Figure S49. PXRD patterns of 1' before sensing (black), after azathioprine sensing (red) and after paraoxon-methyl sensing (blue).



Figure S50. Overlap plot for UV-Vis spectra of all the analytes for paraoxon-methyl sensing with the fluorescence excitation and emission spectra of 1'.



Figure S51. Overlap plot for UV-Vis spectra of all the analytes for azathioprine sensing with the fluorescence excitation and emission spectra of 1'.



Figure S52. Photoluminescence spectrum of the aqueous solution of 1' before and after addition of 300 μ L of 5 mM paraoxon-methyl solution at 388 nm excitation.



Figure S53. Photoluminescence spectra of the aqueous solution of 1' before and after the addition of 300 μ L of 5 mM azathioprine solution at 388 nm excitation.



Figure S54. Quenching efficiency of observed (black curve) and corrected (red curve) measurements for 1' after addition of paraoxon-methyl having different concentrations. Corrected quenching efficiency refers to the quenching efficiency when IFE contribution is not considered.



Figure S55. Quenching efficiency of observed (black curve) and corrected (red curve) measurements for 1' after addition of azathioprine having different concentrations. Corrected quenching efficiency refers to the quenching efficiency when IFE contribution is not considered.

Sl.	Probe	Sensing	$K_{\rm SV}({\rm M}^{-1})$	LOD (pM)	Response	Ref.
No		Medium	, , , , , , , , , , , , , , , , , , ,		Time (s)	
1	[Al(OH)(C ₁₆ H ₁₃ NO ₆)]·	water	26.9×10^{8}	4.23	5	this
	0.5H ₂ O					work
2	AgBiS ₂ /AGr	water	-	7×10^{3}	-	12
	composite by					
	electrochemical					
	methods					
3	Sm ₂ Sn ₂ O ₇ NPs by	phosphate	-	4×10^{3}	-	13
	electrochemical	buffer				
	methods					
4	Spectrophotometry	water	-	-	-	14
5	Spectrophotometry	acetonitrile	-	2.15×10^{6}	-	15
6	AgNP	water	-	90×10^{3}	-	16
7	Spectrofluorometry	methanol	_	1.5×10^{3}	_	17

Table S3. Comparison of the response time, detection limit, K_{sv} and sensing media used for the reported sensors for azathioprine sensing.

Table S4. IFE correction table for paraoxon-methyl sensing.

Paraoxon-	A _{ex}	A _{em}	Correction	Fobserved	F _{corrected}	F _{corrected} (0)/
methyl			Factor			Fcorrected
(µM)			(CF)			
0	0.1530	0.1209	1.3268	272814.2249	361962.9047	1
6.21	0.1581	0.1225	1.3359	266145.6044	355563.3512	1.0179
12.35	0.1692	0.1226	1.3523	260280.3628	351968.7863	1.0284
18.40	0.1789	0.1255	1.3701	255959.5438	350687.7385	1.0322
24.39	0.1818	0.1265	1.3757	251845.1419	346470.7553	1.0447
30.30	0.1890	0.1293	1.3901	246349.7739	342465.5594	1.0569
36.14	0.2018	0.1295	1.4095	240163.4810	338506.0283	1.0693
41.92	0.2080	0.1308	1.4206	236546.9155	336033.1385	1.0772
47.62	0.2150	0.1343	1.4359	232099.0396	333270.5729	1.0861

 Table S5. IFE correction table for paraoxon-azathioprine sensing.

Azathioprine	A _{ex}	A _{em}	Correction	Fobserved	Fcorrected	F _{corrected} (0)/
(µM)			Factor			F corrected
			(CF)			
0	0.0533	0.0469	1.1086	258924.1456	287033.5867	1
6.21	0.0955	0.0472	1.1625	245563.7790	285462.6246	1.0055
12.35	0.1379	0.0473	1.2181	230314.1176	280536.1321	1.0232
18.40	0.1766	0.0493	1.2724	219620.3617	279452.9985	1.0271
24.39	0.2209	0.0499	1.3346	205301.5780	273994.1493	1.0476
30.30	0.2601	0.0499	1.3904	193555.3321	269118.2866	1.0666
36.14	0.2976	0.0507	1.4463	182726.5099	264269.5498	1.0861
41.92	0.3261	0.0509	1.4889	172872.9461	257388.9597	1.1152

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