Immobilization of laccase on functional groups and degree of conjugation modified COFs for simultaneous Photochemical-Enzymatic removal of 2,4-dichlorophenol and uranium U(VI)

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1. Chemicals

1,3,5-triformylphloroglucinol (Tp, 97%) was obtained from Jilin Yanshen Technology (China). Laccase (from *Trametes versicolor*, 0.5 U·mg⁻¹, EC 1.10.3.2), 2,4dichlorophenol (2,4-DCP), glutaraldehyde (GA, 25 % in v/v, aqueous solution), UO₂(NO₃)₂·6H₂O (99.5%), mesitylene (98%), acetic acid (99.5%), 1,4-dioxane (99.5%), 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid (ABTS, 99%), 1,4diaminobenzene (Pa-1, 99%), benzidine (BD, 97%), 4,4"-Diamino-p-terphenyl (TPA, 98%), 2,5-diaminohydroquinone dihydrochloride (Pa-(OH)₂·2HCl, 97%), 2-nitro-1,4phenelynediamine (Pa-NO₂, 98%), 3,3'-dinitrobenzidine (BD(-NO₂)₂, 95%), 3,3'dihydroxybenzidine (BD(-OH)₂, 99%) and other reagents were purchased from Macklin Biochemistry Co., Ltd (Shanghai, China).

2. Characterization

XRD spectra were obtained on a Bruker D8 diffractometer. FT-IR spectra were collected on a ThermoFisher Nicolet NEXUS 670 spectrophotometer. The morphologies of COFs-TpPa-X were photographed on SEM (JEOL-7800F) and HRTEM-Mapping (HAADF-STEM, JEOL-2100F). Thermogravimetric analysis (TGA) was conducted on a SDT650-TA Instruments by heating from 20 to 800 °C with 10 °C/min in N₂ atmosphere. The BET specific surface areas were obtained from Micromeritics ASAP 2020 apparatus. XPS analysis and work function calculation (PS spectra, He lamp 21.21 eV) were performed measured on using a Thermo Fisher ESCALAB 250Xi. The UV-visible diffused reflectance spectra (UV-vis DRS) were tested on a Shimadzu UV-2600 spectrophotometer with BaSO₄ as the background.

Confocal microscope images were captured by laser confocal scanning microscope (OLYMPUS FV1200). The electron spin resonance (ESR) spectra were detected by a Bruker MS 5000 instrument under visible light irradiation (300 W, lamp Au-Light). Free radicals $\cdot O_2^-$ and 1O_2 were detected by ESR using 5, 5-dimethyl-1-pyrroline-N-oxide (DMPO) and 2, 2, 6, 6tetramethyl-4-piperidinol (TEMP) as trapping agents, respectively. Transient photocurrent responses (*i-t*) and electrochemical impedance spectra (EIS) were carried out on a Shanghai Chenhua CHI 660E workstation with a standard three-electrode system.

3. Figures and tables



Fig. S1 The XRD pattern of Tp-TPA



Fig. S2 The $N_{\rm 2}$ adsorption-desorption isotherms of COFs and laccase immobilized COFs.



Fig. S3 The TGA curves of (a) Tp-TPA and (b) Lac@Tp-TPA.



Fig. S4 (a) the XPS survey spectrum of Tp-TPA and Lac@Tp-TPA, the O 1s spectra of (b) Tp-TPA and (c) Lac@Tp-TPA, the N 1s spectra of (d) Tp-TPA and (e) Lac@Tp-TPA, the C 1s spectra of (f) Tp-TPA and (g) Lac@Tp-TPA.



Fig. S5 Solid-state ¹³C CP-MAS NMR spectrum of Tp-TPA.



Fig. S6 (a) the performance of U(VI) photoreduction by Tp-TPA, (b) the generation of H_2O_2 during photo-immobilization of U(VI) in Tp-TPA system under visible light (C_0 =30 mg/L, T=298 K, pH=5.0, m/V=0.1667 g/L, 3 mL CH₃OH).



Fig. S7 The EPR spectra TEMPO-*e*⁻ of Tp-TPA and Lac@COFs under illumination for different times.



Fig. S8 (a) The XPS survey spectrum of Tp-TPA after photoreduction and the high-resolution (b) U 4f, (c) O 1s, (d) N 1s and (e) C 1s spectra of Tp-TPA after photoreduction.



Fig. S9 The high-resolution (a) N 1s and (b) C 1s spectra of Lac@Tp-TPA after photoreduction, and the high-resolution (c) N 1s and (d) C 1s spectra of Lac@TpBD- $(OH)_2$ after photoreduction.



Fig. S10 The XRD pattern of Tp-TPA after U(VI) photoreduction under illumination.



Fig. S11 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S12 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S13 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S14 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S15 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S16 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S17 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S18 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S19 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S20 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S21 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S22 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S23 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S24 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S25 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S26 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S27 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S28 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S29 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S30 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S31 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S32 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S33 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.



Fig. S34 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.







Fig. S36 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.







Fig. S37 The intermediates generated during 2,4-DCP degradation on Lac@Tp-TPA.