Quantitative Local State of Charge Mapping by Operando Electrochemical Fluorescence Microscopy in Porous Electrodes Anton M. Graf^{a,†}, Thomas Cochard^{a,†}, Kiana Amini^{a,b}, Michael S. Emanuel^a, Shmuel M. Rubinstein*,^c and Michael J. Aziz*,a a. Harvard John A. Paulson School of Engineering and Applied Sciences, 29 Oxford St., Cambridge, MA02138, USA. b. Present address: Department of Materials Engineering, University of British Columbia, Vancouver, BC Canada V6T 1Z4 c. Racah Institute of Physics, The Hebrew University of Jerusalem, Jerusalem, Israel. † These authors contributed equally to this work. Electronic Supporting Information (ESI) 15 ESI 1: Electrochemical setup and electrolyte preparation 17 ESI 2: *In situ* confocal microscopy ESI 3: Ex-situ photophysical characterization ESI 4: 1:1 (AQDS:H₂AQDS) binding dimer model ESI 5: Theoretical background of concentration fields in flow devices 25 ESI 6: Data calibration and SOC mapping

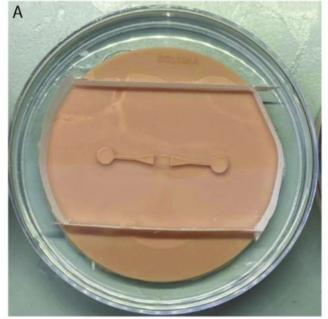
ESI 1: Electrochemical setup and electrolyte preparation

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35 Fabrication of the electrochemical cell: The electrochemical cell was constructed by assembling two acrylic plates, which sandwiched two Polydimethylsiloxane (PDMS) half-cells separated by 36 a Nafion 212 membrane, as illustrated in Figure S1(A). The PDMS components were fabricated 37 through 3D printing and molding. Initially, a mold design was created, and 3D printed using 38 Stereolithography (Form 3, Formlabs). Subsequently, PDMS was poured into each mold and 39 subjected to vacuum treatment to eliminate any air bubbles that could compromise transparency 40 and proper sealing of the components. The PDMS parts were then cured overnight at 65°C. Figure S1(B) showcases the transparency achieved with the PDMS 3D printed mold. To assemble the 42 43 PDMS half-cells, electrodes, and membrane, transparent acrylic endplates and screws were used 44 to compress the components.

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lary 2019: Accontod: 11 March 2019: Published: 15 March 2019 tion microfluidic devices by soft lithography i. y far y and low cost. In this approach PDMS (pol) imr to enerate replicas that are then sealed against g de onstrated fabrication of soft photolithograph u s alternative to the now classic SU-3 resist m ing ADEX dry film include the easily-achievable 1/2 for of the process with significant time savings due and cor. erns due to less toxic developing solution ad a te, ti process can be performed in a low-co mpr place of a convention of and the state of nomine of J.I.

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Figure S1: PDMS molding process. (A) 3D printed mold for the AQDS side with the flow channel feature in the center (orange disk) in a petri-dish where the PDMS is poured. (B) AQDS side half-cell removed from the mold after curing. The transparency of the pieces obtained is showcased by the text which is clearly readable.

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Electrolyte: For the electrolyte concentration series, a 40 mM stock solution was prepared by dissolving 2,7 anthraquinone disulfonate (AQDS) salt in 1M sulfuric acid. AQDS with a purity of 98% was procured from BocSciences. Electrolyte solutions of 40, 30, 20, and 10 mM were

54 obtained by diluting the stock solution with 1M sulfuric acid. To eliminate oxygen from the

electrolyte, each solution was purged with nitrogen (N2, ultra-high purity, Airgas) for 30 minutes

56 prior to each experiment. The electrochemical cell itself was also purged with nitrogen for 30

57 minutes, bringing the system to operating pressures to ensure the absence of leaks and remove any

58 remaining oxygen. In all experiments detailed in this report, the electrolyte was continuously

59 pumped through the cell at a constant flow rate of 2 mm/sec using a Harvard syringe pump. Further

60 information can be found in the main text.

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- 62 Electrochemical experiments:
- 63 AvCarb MGL 190 carbon paper of nominal thickness 190 μm was used as the working electrode.
- 64 The electrode was baked for 24 h at 400 °C prior to the experiment. On the counter electrode side,
- 65 a 0.5 mg/cm² 60% Platinum on Vulcan Carbon Paper Electrode from FuelCellStore was used to
- oxidize hydrogen gas. Nafion 212, used as membrane, was soaked in 1 M sulfuric acid prior to the
- 67 experiments.
- For the electrochemical experiments, a portable Gamry Instruments Reference 3000
- 69 potentiostat/galvanostat/ZRA was employed. The electrochemical reaction was induced using a
- 70 multi-step chronoamperometry protocol. Initially, the open circuit potential (OCV) was
- 71 determined for the two-electrode configuration of the hybrid cell. Subsequently, incremental
- 72 potential steps were applied and maintained at a constant value for 60 seconds each. In between
- 73 each potential step, the cell was purged at OCV conditions for 30 seconds. A sampling rate of 0.5
- 74 seconds was employed.

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76 ESI 2: *In situ* confocal microscopy

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- 78 Fluorescence microscopy was conducted using a spinning disk microscope while the
- 79 electrochemical flow cell was in operation. The setup consisted of a Leica MDi8 microscope body
- 80 and an Andor CSU-W spinning disk system. A Leica 10x/0.3NA HC PL FLUOTAR lens was
- 81 employed for imaging.
- 82 For all experiments presented in this study, the following parameters were utilized: a pinhole size
- 83 of 50 µm, an exposure time of 2 seconds, and an acquisition time of 2.5 seconds per frame.

Excitation light at a wavelength of 405 nm was provided by a laser, and emission light was filtered through a green bandpass filter (approximately 475-525 nm). The laser intensity was set to 50% of its maximum capacity. No gain or offset was applied. The laser was consistently on with the exposure time mentioned above. The intensity and exposure time are crucial for controlling image brightness. These parameters were adjusted to optimize the number of discrete intensity values across the grayscale (each image was captured at 16-bit depth, corresponding to 65,536 gray levels), ensuring optimal brightness.

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93 ESI 3: Ex-situ photophysical characterization

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The photophysical experiments involving emission and absorption scans were conducted using a BioTek Synergy HT plate reader equipped with a 405 nm excitation source and an emission scan capability. In each well of a GREINER MICROPLATE (96 WELL, PS, F-BOTTOM, μCLEAR®, BLACK, MED. BINDING), 200 μL of each sample was loaded.

99 We prepared the 40 mM AQDS solutions following the procedure outlined in ESI 1. These 100 solutions were then transferred to a glovebox and subjected to charging versus V3+/V4+ under a nitrogen (N2) atmosphere inside a bench-scale flow battery. The 40 mM solution was charged to 101 SOCs of 0%, 30%, 40%, 60%, 70%, 80%, and 90% and results are shown in the main text. For the ex-situ tests, flow battery experiments utilized cell components sourced from Fuel Cell 103 Technologies Inc. (Albuquerque, NM). Each half-cell of the battery was equipped with interdigitated flow fields made from pyro-sealed POCO graphite flow plates, with 3 layers of 105 106 carbon paper (SGL 39AA) per side, baked at 400 °C overnight. During cell assembly, the applied torque was 60 lb-in (6.78 N·m) across eight 3/8"-24 bolts, resulting in a load of approximately 107 108 800 lbs per bolt. The geometric area of each electrode was 5 cm². A Nafion 212 cation exchange 109 membrane and a 10 mil Viton gasket separated the two half-cells. The cells were operated inside a nitrogen-filled glovebox. To ensure accuracy, each solution underwent two battery cycles, 110 resulting in a consistent charge efficiency of 98% for all solutions, thereby minimizing systematic 111 112 errors. To prevent reoxidation due to the diffusion of atmospheric oxygen into the electrolytes, the 113 electrolyte solutions were transferred to a well plate inside the glovebox and sealed with a lid.

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115 ESI 4: 1:1 (AQDS:H₂AQDS) binding dimer model

117 The quinhydrone concentration at each SOC value can be calculated from a system of equations.

Using the equilibrium constant of dimer formation of $K_{dim} = 80 \, M^{-1}$ from the literature(1) and the total concentration of the reactants participating in the electrochemical reaction, which is equal to the initial concentration of AQDS in each experiment, a parametric curve for each species concentration as a function of SOC can be computed. The system of equations is shown below:

$$K_{dim} = 80 M^{-1} = \frac{[QH]}{[AQDS][H_2AQDS]}$$
 (S4.1)

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$$[AQDS]_0 = [AQDS] + [H_2AQDS] + 2[QH]$$
 (S4.2)

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$$SOC = \frac{\left[H_2AQDS\right] + \left[QH\right]}{\left[AQDS\right] + \left[H_2AQDS\right] + 2\left[QH\right]}$$
(S4.3)

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125 The system of equations is solved for different [AQDS]₀.

126 For the reactant concentrations used in this study the plots in Figure S2 were obtained.

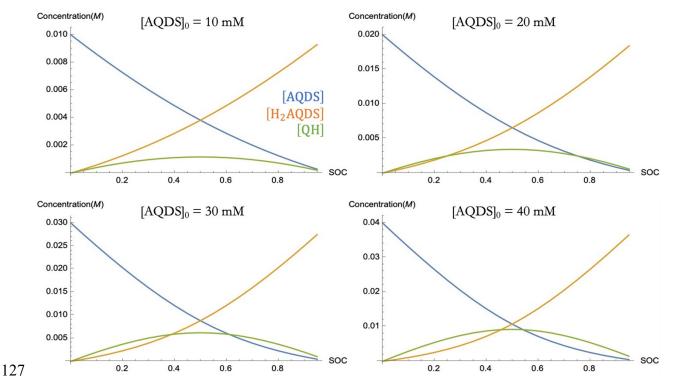


Figure S2: 1:1 (AQDS:H₂AQDS) binding dimer model. Prediction of the theoretical concentration profiles for AQDS, H₂AQDS,

and the QH as a function of SOC, calculated using the dimer model. The simulations are performed at fixed $[AQDS]_0$ of 10, 20,

130 30, and 40 mM and $K_{dim} = 80 \text{ M}^{-1}$.

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- 132 With increasing [AQDS]₀, it is evident that the proportion of dimer to the total concentration
- 133 grows in the presented cases, and the concentrations of individual molecular species exhibit
- 134 increasingly nonlinear variations with respect to the SOC. Consequently, it is essential to account
- 135 for the influence of dimer formation on fluorescence intensity, particularly at high concentrations,
- in order to accurately quantify the SOC. The plate reader data in Figure 2(C) demonstrate that the
- 137 intensity of the 40 mM solution follows the same nonlinear trend as the H₂AQDS concentration.

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142 ESI 5: Theoretical background of concentration fields in flow devices

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- 144 In electrochemical flow devices, the temporal change of the concentration fields can be modeled
- 145 by the sum of mass-transport and electrochemical source terms, namely advection A_{i} , diffusion D_{i} ,
- 146 electromigration ε_j and the (Butler-Volmer) source term S.

$$\frac{\partial C_j}{\partial t} = A_j + D_j + \varepsilon_j + S \tag{S5.1}$$

- 147 A_j describes the net flow through the electrode and is given by a function of the geometry u and
- 148 the gradient of the concentration of interest ∇C_j ,

$$A_j = -u \cdot \nabla C_j \tag{S5.2}$$

- D_j as derived by Fick's law can be calculated from the diffusion coefficient of the molecular
- 150 species d_j and the Laplacian of the concentration of interest $\nabla^2 C_j$,

$$D_j = d_j \cdot \nabla^2 C_j \tag{S5.3}$$

- 151 ε_j describes the bias of charged entities through the electric field and is given by the charge z_j ,
- 152 Faraday's constant F, the gas constant R, the temperature T, and the electric field $\nabla \phi_L$,

$$\varepsilon_{j} = \frac{z_{j} \cdot D_{j} \cdot F}{R \cdot T} \cdot \nabla \cdot (C_{j} \cdot \nabla \phi_{L})$$
(S5.4)

153 Adding all terms together we obtain the following partial differential Eq. A.1 for the temporal

154 change of the concentration as a function of current, potential and mass-transport:

$$\frac{\partial C_j}{\partial t} = -u \cdot \nabla C_j + d_j \cdot \nabla^2 C_j + \frac{z_j D_j F}{R \cdot T} \cdot \nabla \cdot (C_j \cdot \nabla \phi_L) + S_j$$
(S5.5)

- 155 The connection between the applied potential and the current is given through the electrochemical
- source term (Butler-Volmer equation) measuring the overall rate of the redox reaction of AQDS.
- 157 The source term can be computed given the overpotential (η), the geometric area of the electrode
- interface (a), the reaction rate constant (k), the charge transfer coefficients of the respective redox-
- 159 state (α_i) as well as the concentration of the reactant in each redox-state (α_i).

$$S = a \cdot k \cdot C_A^{\alpha_A} \cdot C_B^{\alpha_B} \cdot \left[\exp\left(\frac{n_e \cdot F \cdot \alpha_A}{R \cdot T} \cdot \eta\right) - \exp\left(\frac{n_e \cdot F \cdot \alpha_B}{R \cdot T} \cdot \eta\right) \right]$$
 (S5.6)

- 160 The applied overpotentials cause the current response which is monitored over the duration of the
- 161 experiment. Consequently, current and overpotential as well as fluorescence intensity and
- 162 concentration can be directly related. Both pairs of physical parameters must be connected to
- 163 finally interpret the fluorescence data and derive conclusions about current, potential and flow.

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165 ESI 6: Data calibration and SOC mapping

- 166 As part of our calibration procedure, we considered the fluorescence contributions of each
- 167 molecular species involved, i.e. AQDS, H₂AQDS and QH, which are required for proper
- 168 quantification of the local SOC. The various contributions are displayed schematically in Figure
- 169 S3.

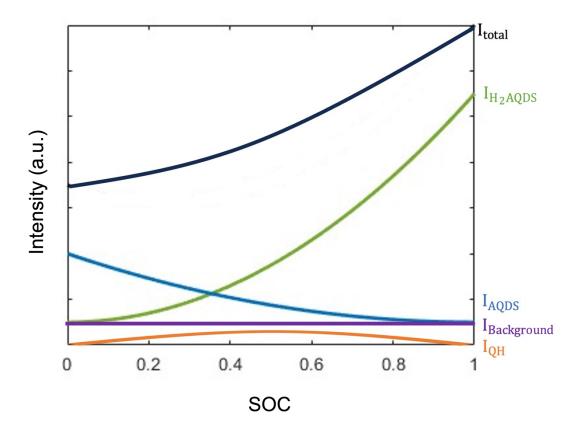


Figure S3: Schematic depiction (not to scale) of different contributions to the spatially averaged raw intensity I_{total} . For the calibration of the intensity maps to obtain SOC maps we need to distinguish between the different species contributing to I_{total} . The black solid line schematically shows I_{total} that is obtained in experiment. $I_{Background}$ is the background signal which remains constant in every image. I_{QH} corresponds to the dimer intensity which is expected to have small contribution to I_{tot} but is estimated independently. I_{AQDS} is the native fluorescence of AQDS and I_{H_2AQDS} is the fluorescence of I_{LQDS} which has the highest contribution to I_{total} .

Our experimental data for the SOC estimation comprised 29 images. A single *ex situ* image taken at a high and homogeneous (but unknown) H₂AQDS concentration; and 28 images taken for varying applied overpotentials with 4 total AQDS concentrations crossed with 7 applied voltages. As the analysis is done at the individual pixel level, it is critical for the images to be aligned on that scale based on the *ex-situ* image. This was done using the CV2 computer vision library and resulted in a mean shift of 1.8 pixels. Once all the images were aligned, we assigned pixels into three categories: excessively dim; excessively bright; and reliable pixels to be included in the analysis. We hypothesize that excessively dim pixels were due to obstructing fibers, and excessively bright pixels were caused by highly reflective heterophase impurities. We set

thresholds for dim and bright pixels at quantiles 0.55 and 0.995, respectively. These thresholds were set by a combination of a qualitative visual review of the data with a Gaussian mixture model to set the lower threshold. As the location of the fibers remains fixed in every image, a pixel I_{ri} on one of the reaction images was considered valid when the corresponding pixel B_i was valid on the *ex-situ* image, and the pixel fell within the specified quantiles of intensity for that reaction. The remaining pixels, corresponding to fibers and other heterophase impurities, were set to be NaN and were not further considered in our analysis, as demonstrated for the example in Figure S4.



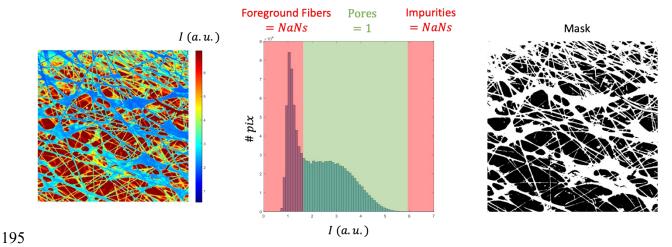


Figure S4: Selection of valid pixels for SOC maps. (Left) shows the raw intensity image in arbitrary units. (Middle) the histogram of the image is shown with schematic of the manually selected low and high intensity cutoffs. The red areas correspond to the pixels that were set to NaN values and are identified as fibers or impurities. The green area corresponds to the "good" pixels in the pores filled with electrolyte that are considered for further image processing. (Right) shows a visualization of the mask image that contains all valid pixels (in black) and NaN pixels in white.

This data filtering reduced the size of our calibration set from 29.4 million to 11.7 million pixels. The result of the image data processing was an array of ex situ intensities B_i and arrays I_r for each reaction condition.

We also had 28 utilizations u_r corresponding to each reaction and calculated by Eq. 3.2. We posit a linear relationship between the electrochemical utilization u_r and the mean state of charge S_r . When the system is not mass transport limited, the mean field theory should be a highly accurate approximation *on average* over a mesoscale patch of space, even if it does not hold uniformly over very small areas. Most of our reactions had utilizations below 50%, justifying the claim that mass transport limitations were not binding and the use of this approximation. In the simple mean field

- 211 theory without mass transport limitations, the solution is a smooth constant gradient in the state of
- 212 charge between 0 at the inlet and u at the outlet. The mean state of charge \mathfrak{T} and utilization u are
- 213 thus related by $\overline{S} = ux_m/L$ where x_m is the position of the midpoint of the image and L is the length
- 214 of the electrode. In our study, the ratio $x_m / L = 0.63$.
- There are three electroactive species in this study, namely the oxidized state AQDS; the
- 216 reduced state H₂AQDS; and the dimer QH. The concentrations of these species are subject to two
- 217 constraints,

$$[AQDS] + [H_2AQDS] + 2[QH] = [AQDS]_0$$
 (S6.1a)

$$[QH] = K [AQDS] [H_2AQDS]$$
 (S6.1b)

- 218 Eq. S6.1a is conservation of the anthraquninone species in their various forms, and Eq. S6.1b is
- 219 the chemical equilibrium governing dimer formation. It is straightforward to solve Eq. S6.1 for the
- 220 concentration of all three species using the relative concentration Y of the reduced species defined
- 221 in Eq. 3.5. The solutions are:

$$[AQDS] = \frac{1 - Y}{1 + 2RY} \cdot [AQDS]_0 \tag{S6.2a}$$

$$[H_2AQDS] = Y \cdot [AQDS]_0 \tag{S6.2b}$$

$$[QH] = \frac{RY}{1 + 2RY} [AQDS]_0$$
 (S6.2c)

- where $R = K [AQDS]_0$ is the dimensionless rate constant for the dimer formation is.
- We next made an initial estimate of the brightness coefficients α , β , γ , δ . To formulate this
- 224 estimate, we made the preliminary assumption (later relaxed in the final estimation) that the
- 225 concentrations of the electroactive species were uniform for each reaction. Using Eq. S6.2, we
- 226 constructed a numerical table mapping an input reduced fraction Y to an output state of charge.
- 227 There is one table for each initial concentration [AQDS]₀. These tables were all monotonically
- 228 increasing as we expected, and we used the data points to construct four numerical functions
- 229 mapping between SOC and Y at the various [AQDS]₀. We converted the utilizations u_r to a mean
- 230 SOC S_r for each reaction; interpolated S_r to obtain Y_r ; and then solved for the equation of each
- 231 species using Eq. S6.2. We completed this step by solving a linear least squares problem of the
- 232 form I = XC. The left-hand side was the mean image intensity for the 28 reactions. The design
- 233 matrix X has 28 rows and four columns [AQDS], [H₂AQDS], [QH], 1. The estimated coefficients
- 234 are $C = [\alpha, \beta, \gamma, \delta]$. This regression achieved an excellent fit with an R squared value of 0.9632.

- 235 The estimated coefficients included a large, positive value for β (the brightness of H_2AQDS) and
- 236 much smaller values for AQDS and QH, as expected.
- Once the brightness coefficients were available, we built a table with the predicted light
- 238 production P as a function of Y by Eq. 3.3. This led to a monotonic function that was interpolated
- 239 to produce a mapping function Y(P). We finally estimated the state of charge by applying Eq. 3.4
- 240 once we have the optical factor F_i at each pixel. We obtained the optical factor by applying the
- 241 assumption that the bright ex situ image is homogeneous, yielding the direct calculation $F_i = B_i/B$,
- 242 where B is the mean intensity on the ex-situ image. The resulting estimates of the state of charge
- 243 are visually plausible and have an excellent fit to the experimental brightness.
- 244 At this stage we made a further refinement of the model. We relinquished the assumption used
- 245 to estimate the brightness coefficients that the concentration was uniform on each reaction image.
- 246 We also insisted that the model recover the assumed mean state of charge on each reaction; the
- 247 initial estimate was close but had an RMS error of 0.031. This model was estimated by an iterative
- 248 application of three computational steps until convergence was achieved. These steps are: (1)
- 249 refine the estimate of the optical factor; (2) refine the estimate of the brightness coefficients; (3)
- 250 shift the estimated concentrations to match the mean state of charge S_r .
- The concentration shifts are performed after applying a logit transform of the estimated Y for
- 252 each reaction. At each step, logit(Y) is shifted by one step of Newton's method using a numerical
- 253 derivative. Four steps of Newton's method are sufficient to match S_r to six decimal places. The
- 254 update to the brightness coefficients is analogous to the initial estimate. This time, however,
- 255 instead of a least squares problem with 28 rows (one per reaction), there are 11.7 million rows (one
- 256 per valid pixel). The left-hand side of the least squares equation is the observed intensity, and the
- 257 design matrix X is as before, but each row is now scaled by the optical factor. We can succinctly
- 258 write the design matrix entries as
- $X_{ri} = [[AQDS], [H_2AQDS], [QH], 1] \cdot F_i$ This regression $\mathbf{I} = \mathbf{X} \mathbf{C}$ provides both an updated estimate
- 260 of the brightness coefficients and a standard error σ_I on the recovered brightness.
- The update to the optical factors F is new to this stage of the estimation. If we consider pixel
- 262 *i* in isolation, Eq. 3.4 implies that we can construct a linear regression $I_r = P_r$ F, where I_r and P_r
- are both column vectors with 28 rows, and F is the optical factor to be estimated. This regression

can be augmented to include an additional 29th row from the ex situ image with left and right sides B_i and B_i , respectively. This regression also provides a standard error σ_F on the optical factor. 265

This model was initialized with preliminary estimates for the brightness coefficients and 267 optical factors described above. It was then iterated for nine steps until convergence criteria for small changes in parameter values were met. The converged model includes estimates for the concentration of all three species, from which we can easily compute the estimated SOC. The model also estimates standard errors in I and F. We then compute a total standard error by assuming that the errors in I and F are independent. The result of this calculation is an RMS error of 0.00526 SOC units over all 28 reactions. The attribution of the error is 0.00422 due to uncertainty in I and 0.00314 due to uncertainty in F. It is important to acknowledge that even after the calibration procedure, certain residual background fibers and pore depth effects persisted, which have the potential to introduce minor systematic errors.

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277 Comparison with a constant SOC gradient assumption:

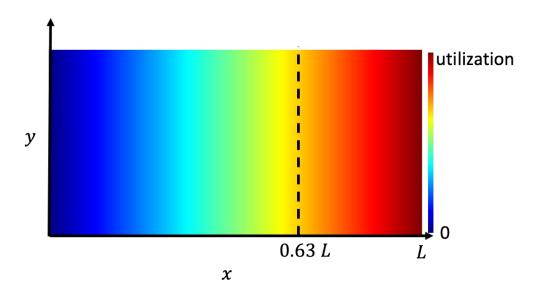
We assumed that over large scales, as the 1.2x1.2 mm imaging frame, a simplified assumption of a constant SOC gradient along x and uniform along y can be used to estimate 3. The results are then given by $\overline{S} = ux_m/L$ as demonstrated in Figure S5.

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Figure S5: Schematic of simple constant gradient assumption. The rectangle corresponds to the working electrode where the fully oxidized electrolyte enters on the left side and gets reduced in the form of a constant gradient along x towards the outlet. The color map indicates the utilization of 0% in blue and the maximum utilization, at the end of the electrode, in red. In our study, the image is centered at $x_m/L = 0.63$.

For a comparison of our results with the simple assumption of a smooth, constant gradient, we generated simulated SOC maps at two distinct utilizations (0.3 and 0.6), maintaining x_m consistent with the experimental location of the imaging center at 0.63L, as depicted in Figure S6. A mask was superimposed on the images, designating pixels in regions obscured by fibers as NaNs. Notably, the histograms indicate that the distributions largely resemble rectangular shapes, even after the exclusion of the masked pixels.

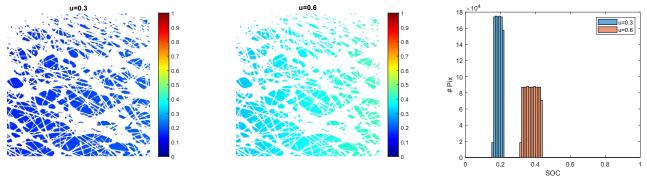


Figure S6: Spatial mapping of the simplified model featuring a constant gradient, tailored to the specific imaging frame and incorporating the position and mask derived from the actual experiment. On the left, a simulated 2D SOC corresponds to an electrolyte utilization of 0.3, while the central image represents a utilization of 0.6. The rightmost image presents the associated histograms, which largely maintain rectangular distributions even in the presence of NaN values.

301 References

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