

## **Robust PET-Based Chiral Schiff Base Molecular Probe for Recognition of Hg<sup>2+</sup> and Pb<sup>2+</sup> and Its Live Cell Imaging**

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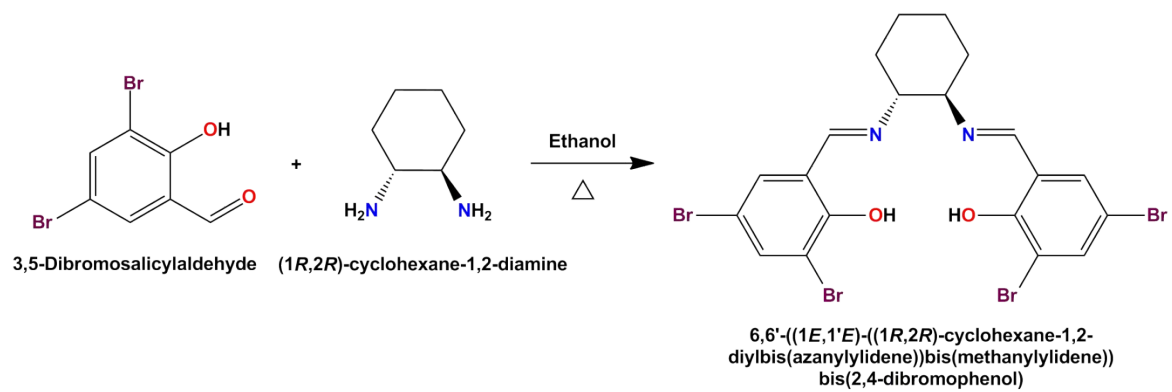
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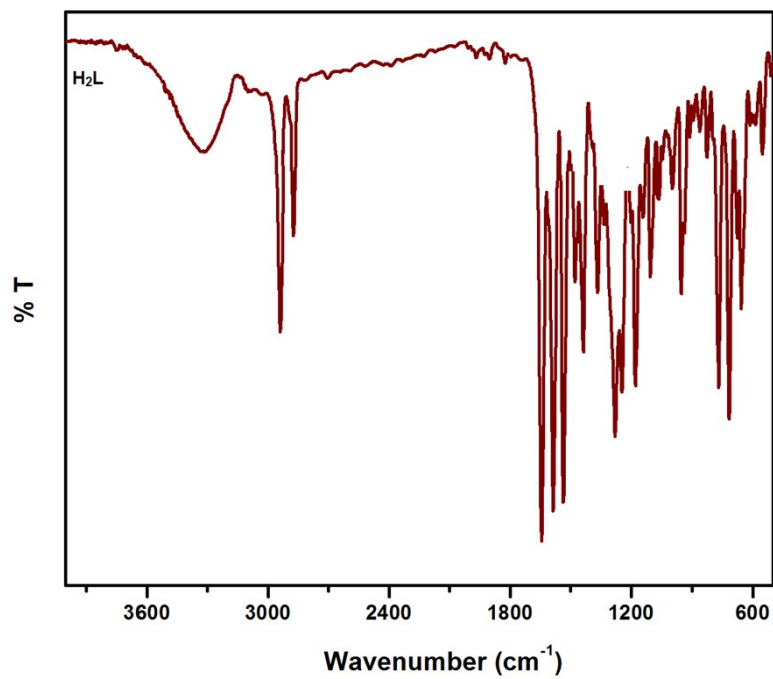
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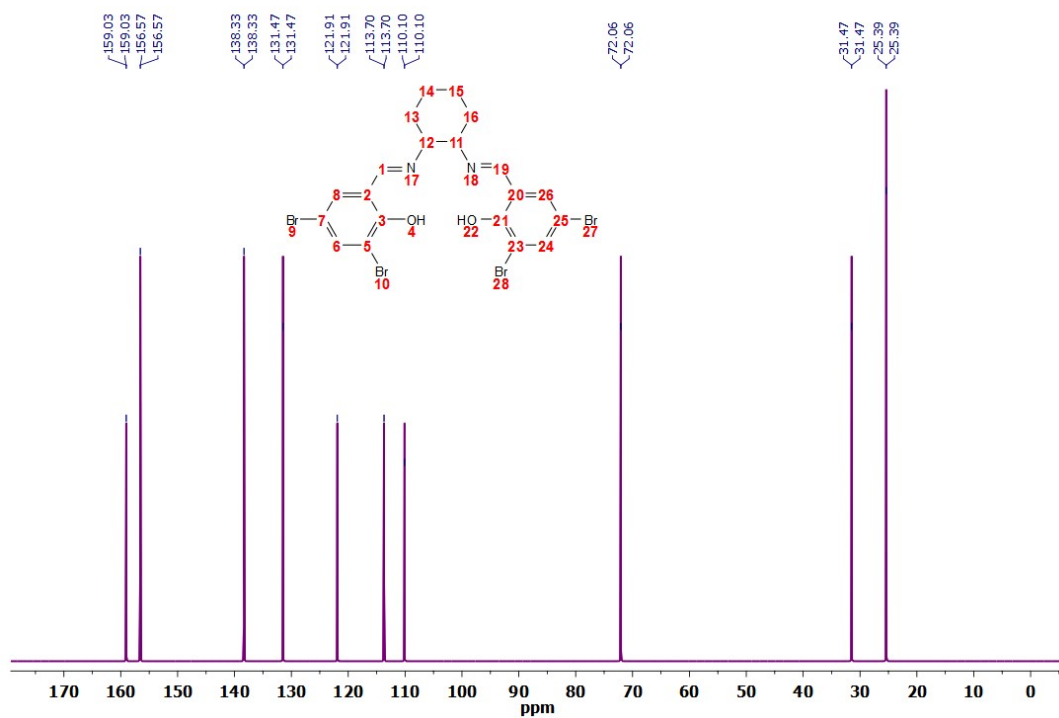
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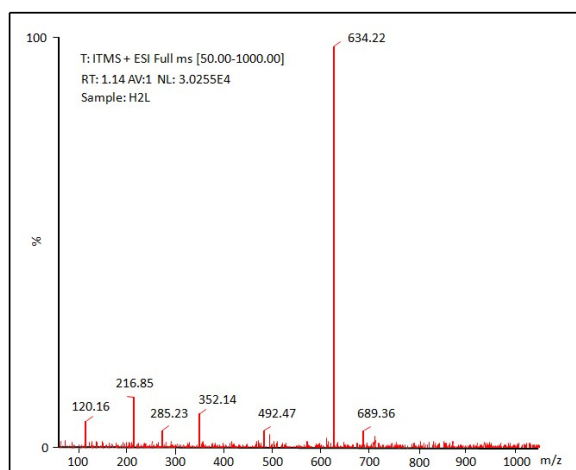
**Scheme S1.** Synthetic scheme for H<sub>2</sub>L



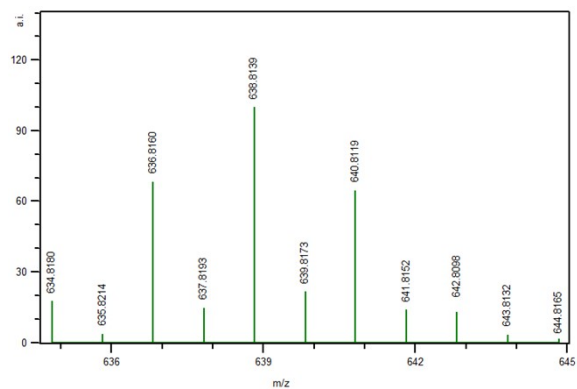
**Fig. S1.** FT-IR spectrum of H<sub>2</sub>L



**Fig. S2.**  $^{13}\text{C}$  NMR spectrum of  $\text{H}_2\text{L}$

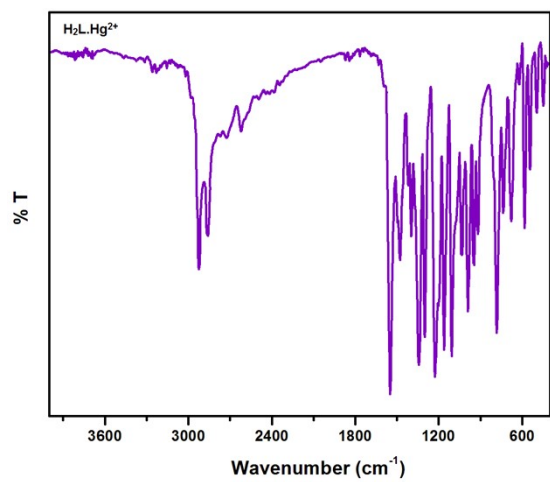


(a)

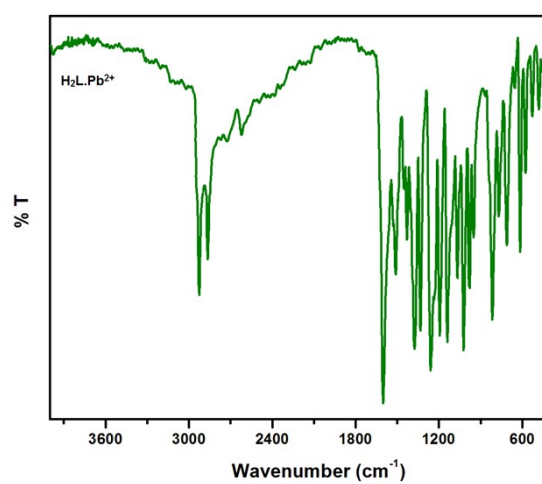


(b)

**Fig. S3.** ESI-mass spectra of H<sub>2</sub>L, (a) experimental; (b) simulated



(a)



(b)

**Fig. S4.** FT-IR spectra of (a)  $H_2L.Hg^{2+}$ ; (b)  $H_2L.Pb^{2+}$

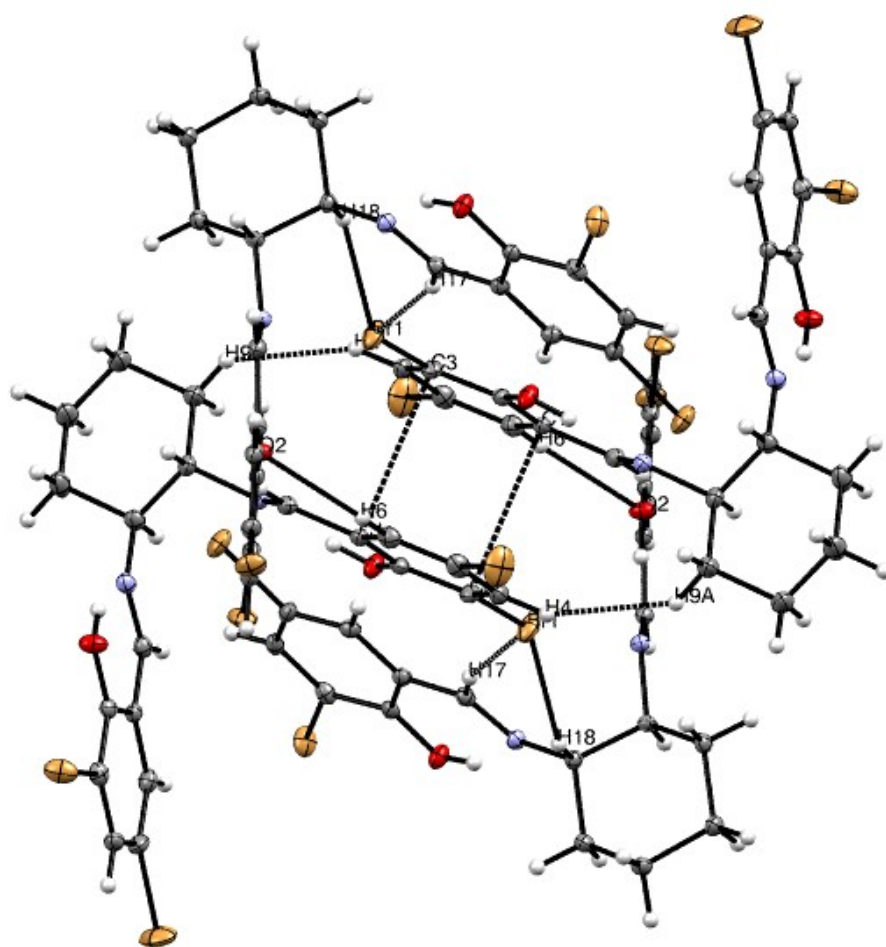


Fig. S5. 2D-coordination polymeric chain of H<sub>2</sub>L

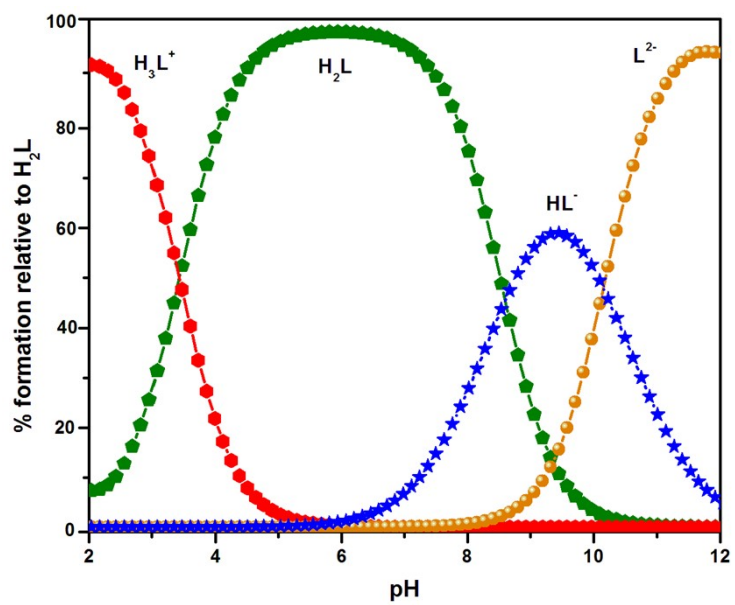
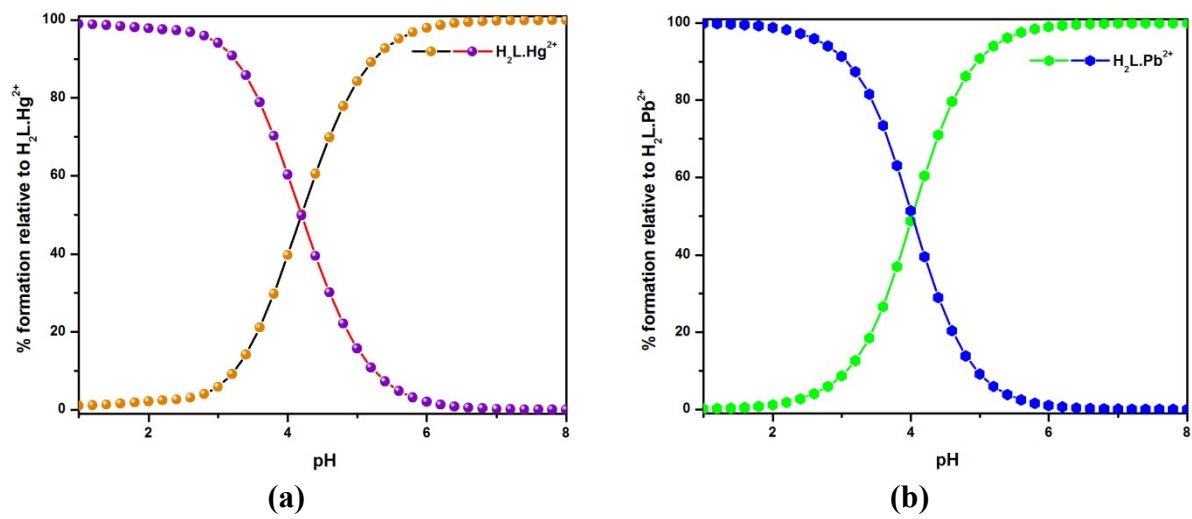
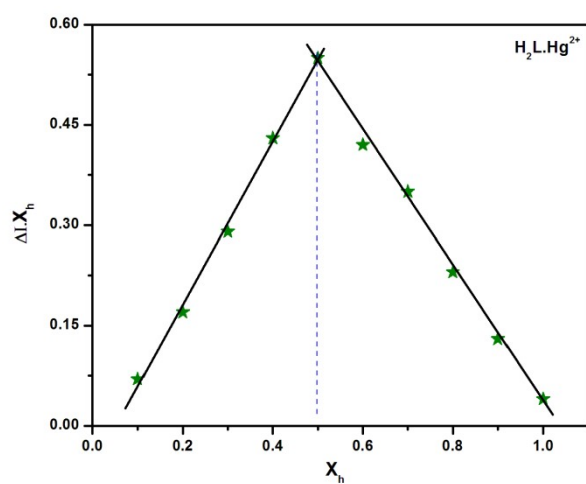


Fig. S6. Protonation equilibria diagram of  $H_2L$

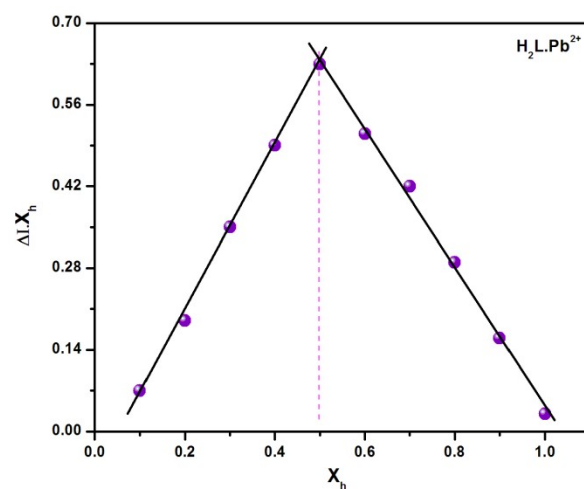




**Fig. S7.** Species distribution diagram of (a)  $H_2L.Hg^{2+}$  and (b)  $H_2L.Pb^{2+}$  equilibria in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v)

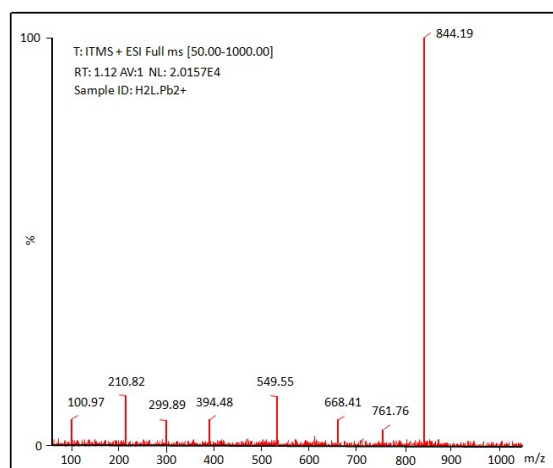
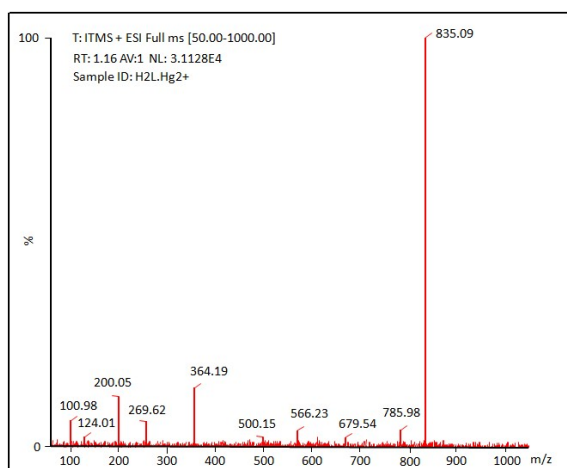


(a)



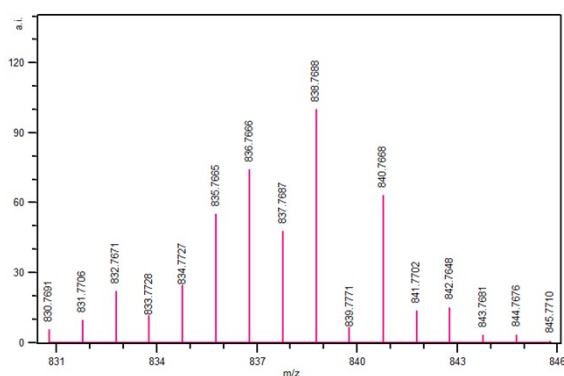
(b)

Fig. S8. Job's plot analysis  $H_2L$  with (a)  $Hg^{2+}$  and (b)  $Pb^{2+}$

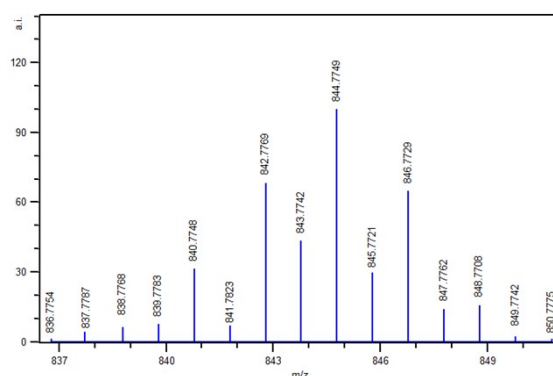


(a)

(c)

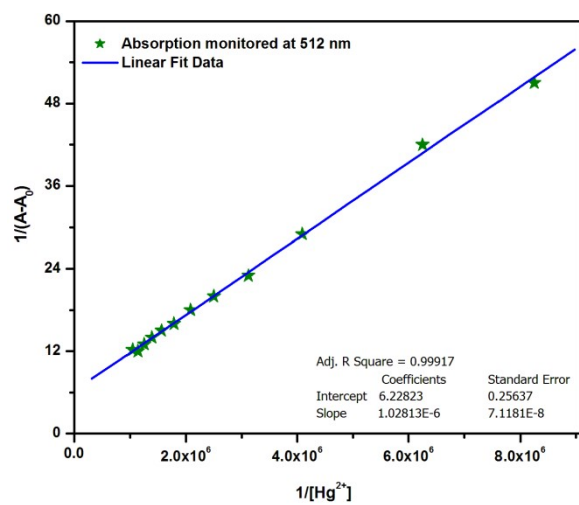


(b)

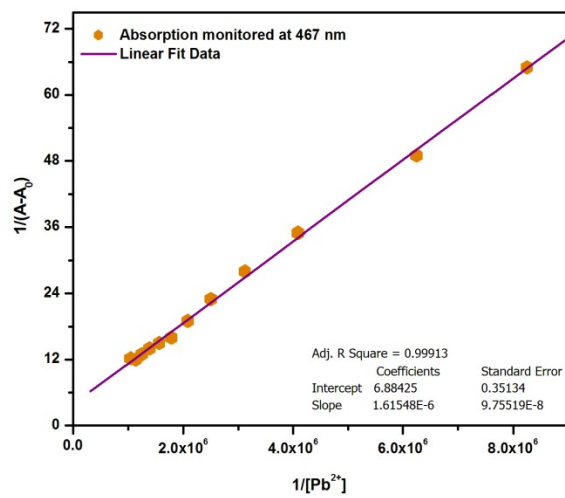


(d)

**Fig. S9.** ESI-mass spectra of (a) experimental  $\text{H}_2\text{L.Hg}^{2+}$ ; (b) simulated  $\text{H}_2\text{L.Hg}^{2+}$ ; (c) experimental  $\text{H}_2\text{L.Pb}^{2+}$  and (d) simulated  $\text{H}_2\text{L.Pb}^{2+}$

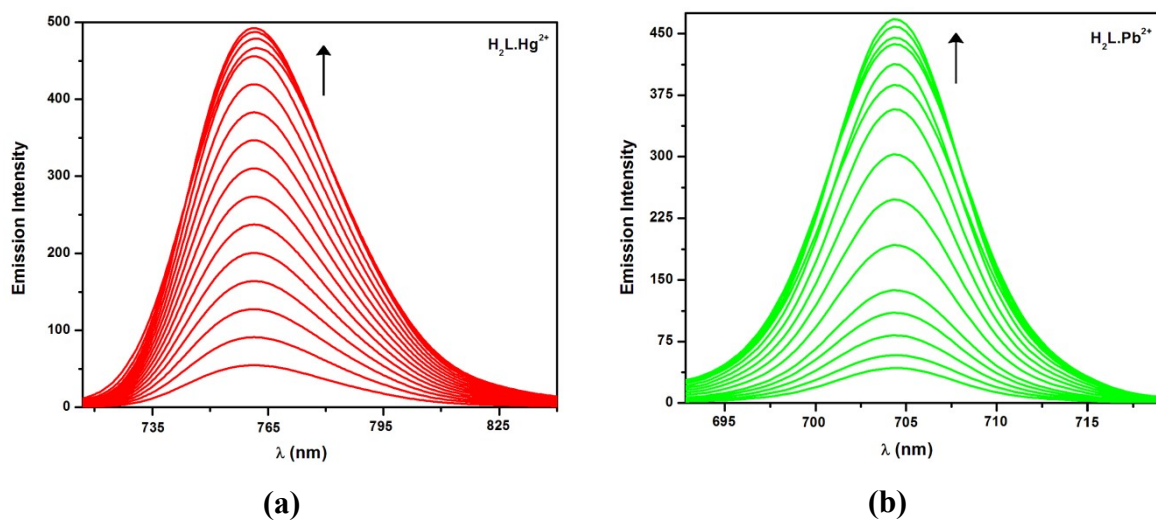


(a)

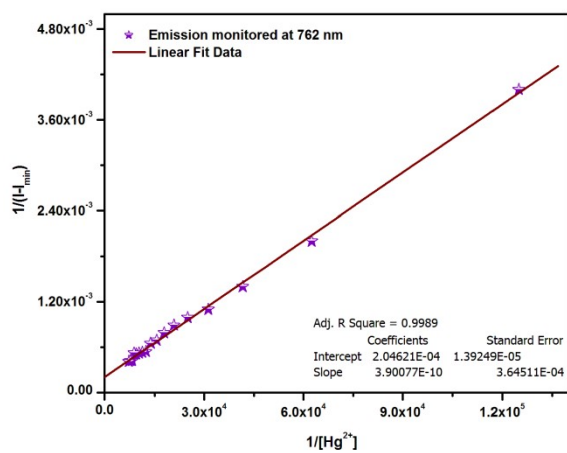


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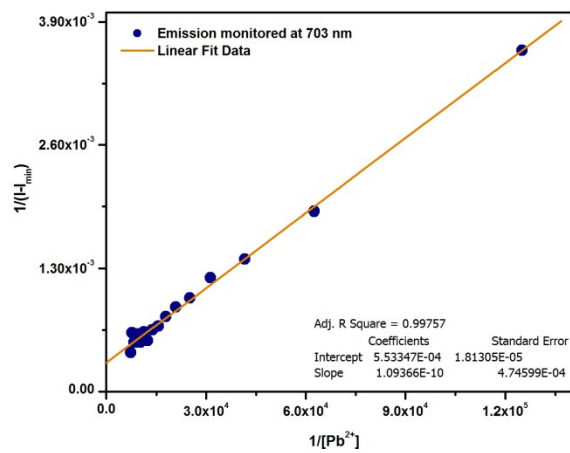
**Fig. S10.** B-H plot from UV-vis titration data of with (a)  $\text{Hg}^{2+}$  and (b)  $\text{Pb}^{2+}$  concentration



**Fig. S11.** Fluorescence spectra of H<sub>2</sub>L (10 μM) upon incremental addition of (a) Hg<sup>2+</sup> (0.0 - 5.0 equiv.) and (b) Pb<sup>2+</sup> (0.0 - 5.0 equiv.) in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v)

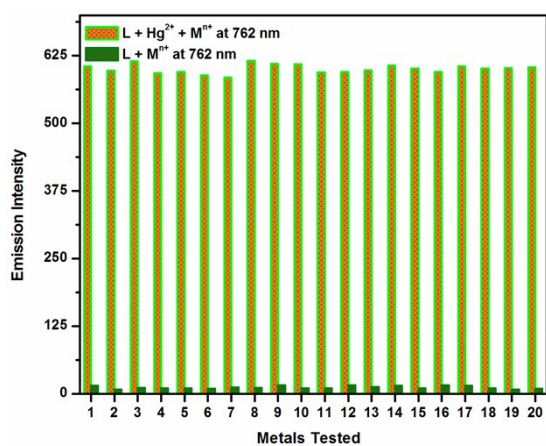


(a)

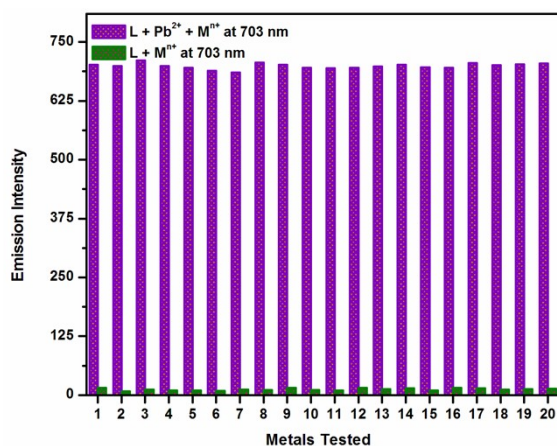


(b)

**Fig. S12.** B-H plot from fluorescence titration data of with (a)  $\text{Hg}^{2+}$  and (b)  $\text{Pb}^{2+}$  concentration

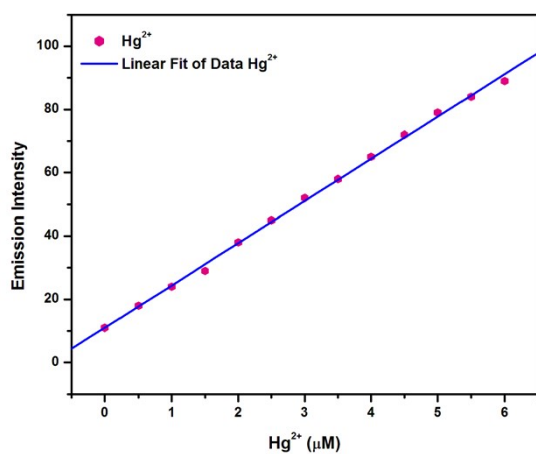


(a)

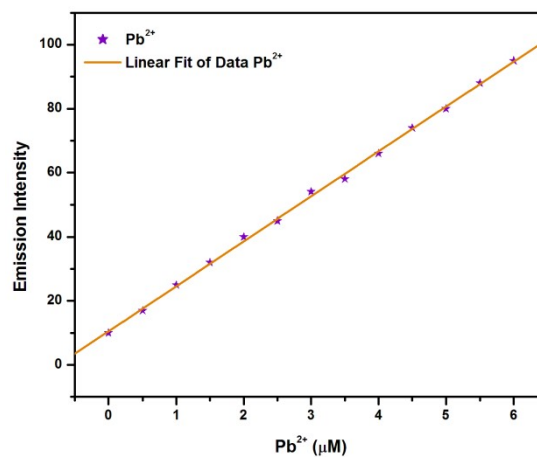


(b)

**Fig. S13.** Fluorescence intensity of H<sub>2</sub>L (10 μM) with (a) Hg<sup>2+</sup> and (b) Pb<sup>2+</sup> in the presence of other metal ions in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v) at room temperature



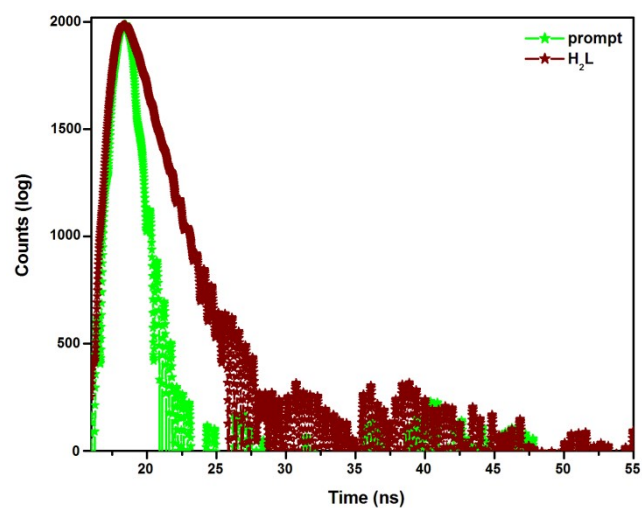
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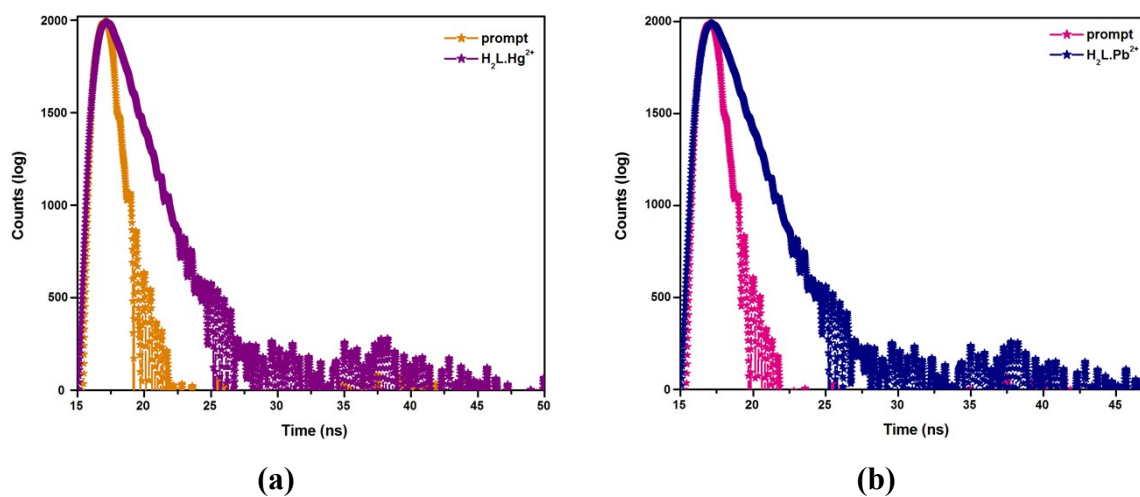
(b)

**Fig. S14.** (a) Linear dynamic plot of fluorescence intensity (at 762 nm) vs.  $[\text{Hg}^{2+}]$  for the determination of S (slope) and (b) Linear dynamic plot of fluorescence intensity (at 703 nm) vs.  $[\text{Pb}^{2+}]$  for the determination of S (slope);  $[\text{H}_2\text{L}] = 10 \mu\text{M}$

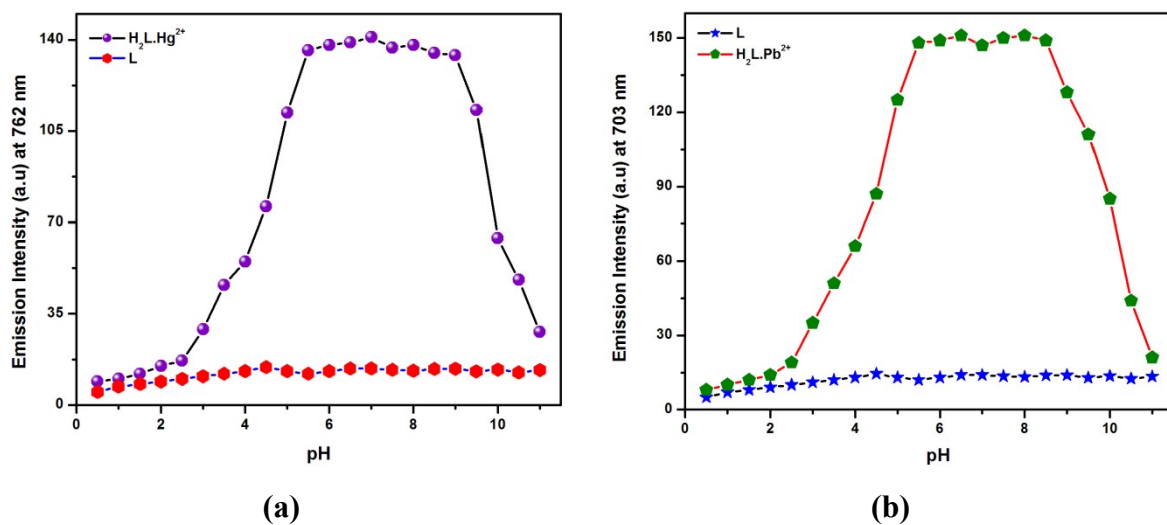




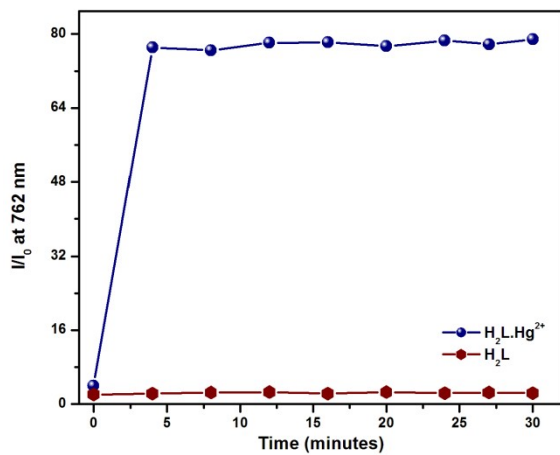
**Fig. S15.** Lifetime emission decay of H<sub>2</sub>L (10 μM) in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v)



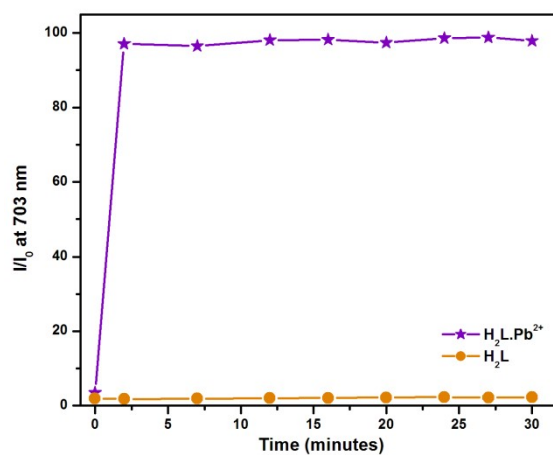
**Fig. S16.** Lifetime emission decay of (a)  $\text{H}_2\text{L.Hg}^{2+}$  and (b)  $\text{H}_2\text{L.Pb}^{2+}$  in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v)



**Fig. S17.** Fluorescence intensity of H<sub>2</sub>L in the presence of **(a)** Hg<sup>2+</sup> ( $\lambda_{em} = 600$  nm;  $\lambda_{ex} = 762$  nm); **(b)** Pb<sup>2+</sup> ( $\lambda_{em} = 575$  nm;  $\lambda_{ex} = 703$  nm) at various pH values in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v)

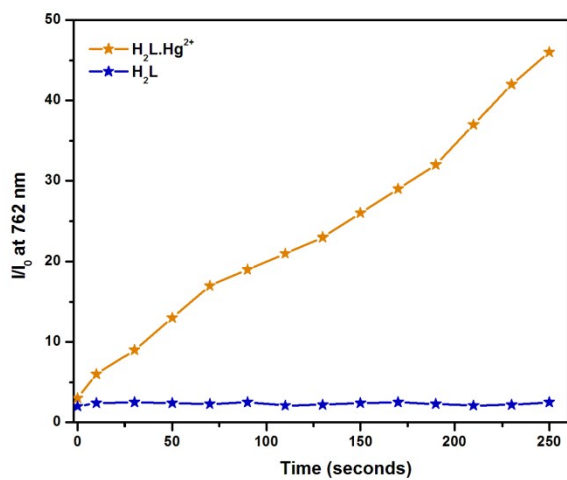


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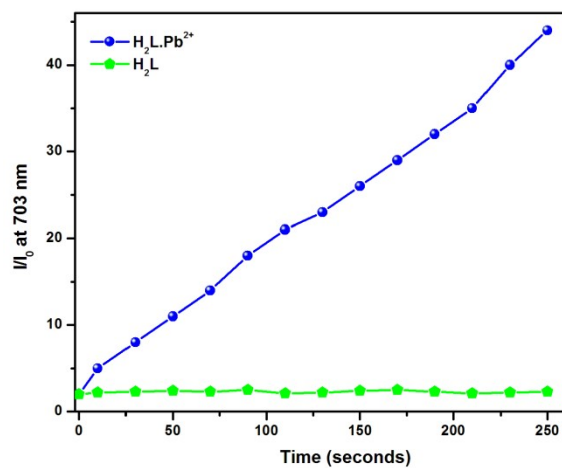


(b)

**Fig. S18.** Emission intensity of (a)  $H_2L.Hg^{2+}$ ; (b)  $H_2L.Pb^{2+}$  as a function of time (0-30 minutes)

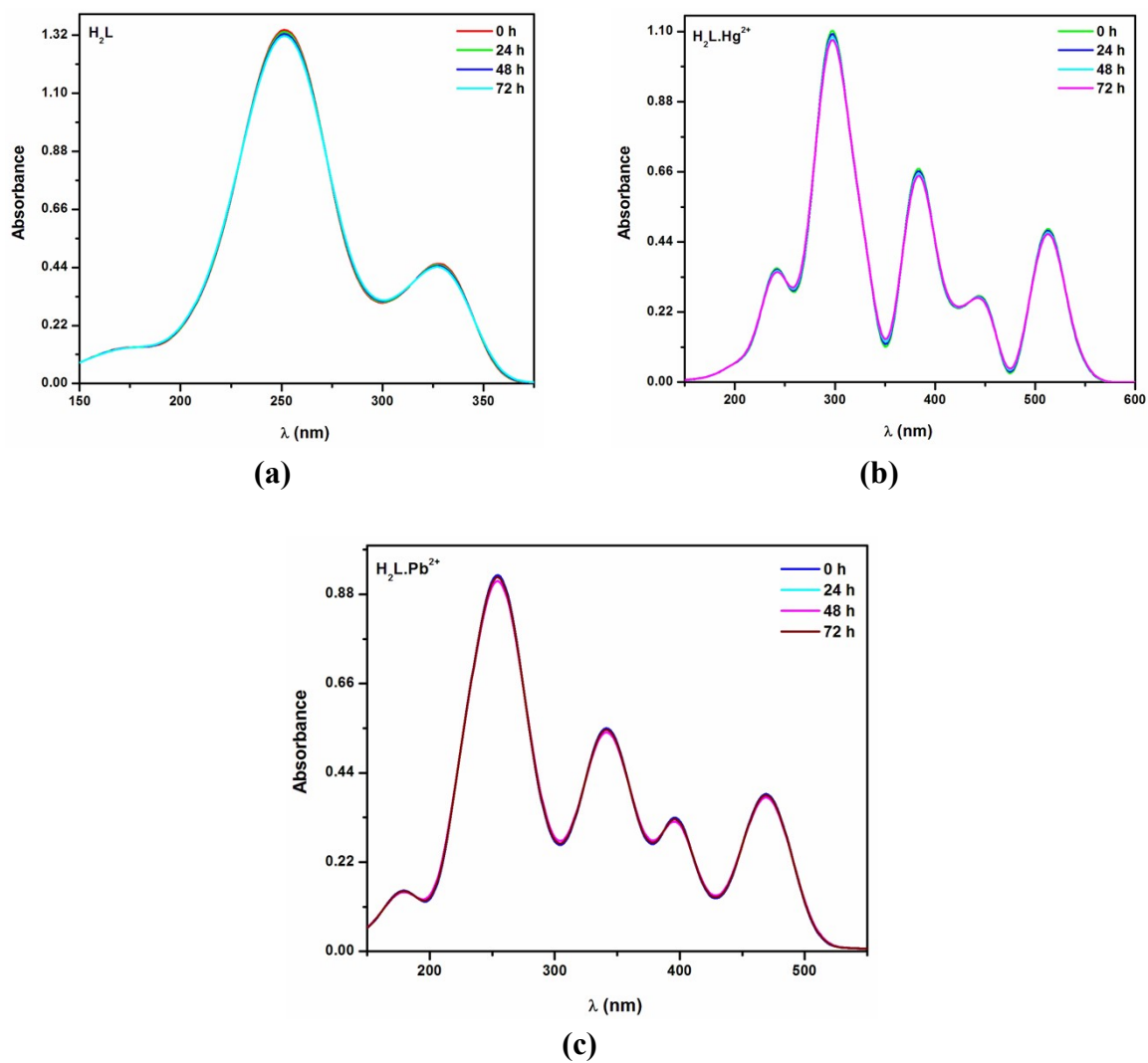


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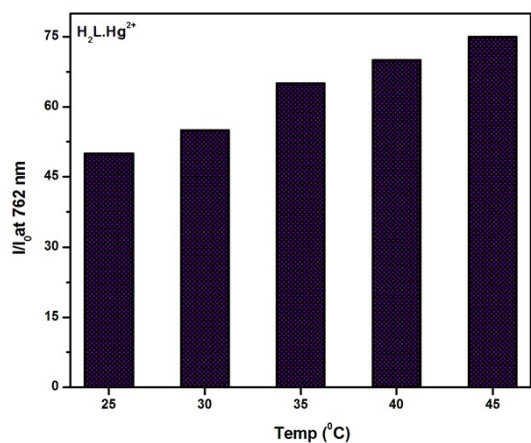


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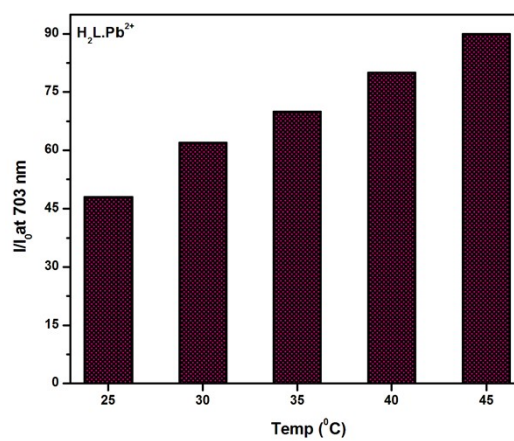
**Fig. S19.** Fluorescence intensity of (a)  $H_2L.Hg^{2+}$ ; (b)  $H_2L.Pb^{2+}$  as a function of time (seconds)



**Fig. S20.** Photostability tests of (a) H<sub>2</sub>L, (b) H<sub>2</sub>L.Hg<sup>2+</sup> and (c) H<sub>2</sub>L.Pb<sup>2+</sup> measured by absorption spectroscopy in methanol/HEPES buffer (5 mM, pH 7.2, 1:9 v/v).

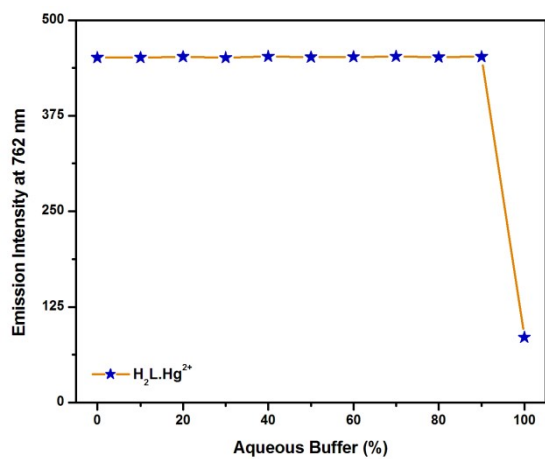


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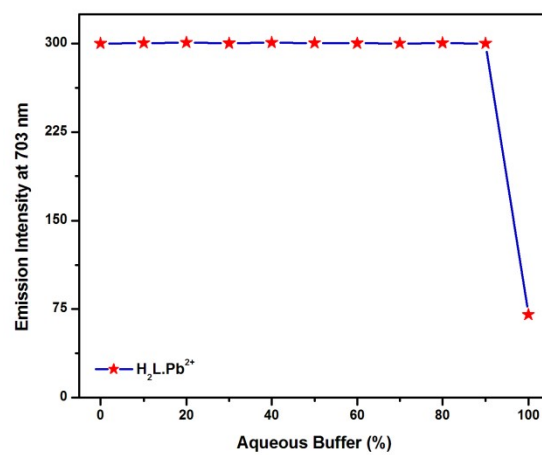


(b)

**Fig. S21.** Fluorescence spectral changes of (a)  $H_2L.Hg^{2+}$ ; (b)  $H_2L.Pb^{2+}$  as a function of temperature (25-45 °C)



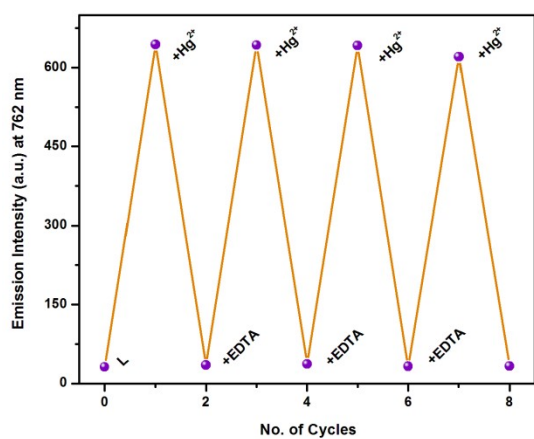
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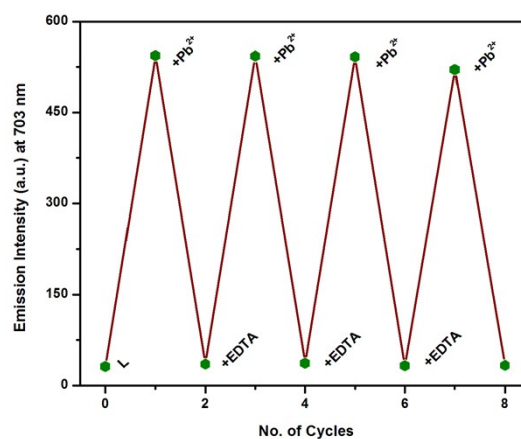
(b)

**Fig. S22.** Emission intensity of (a)  $H_2L.Hg^{2+}$ ; (b)  $H_2L.Pb^{2+}$  as a function of aqueous buffer concentration (0-99%)



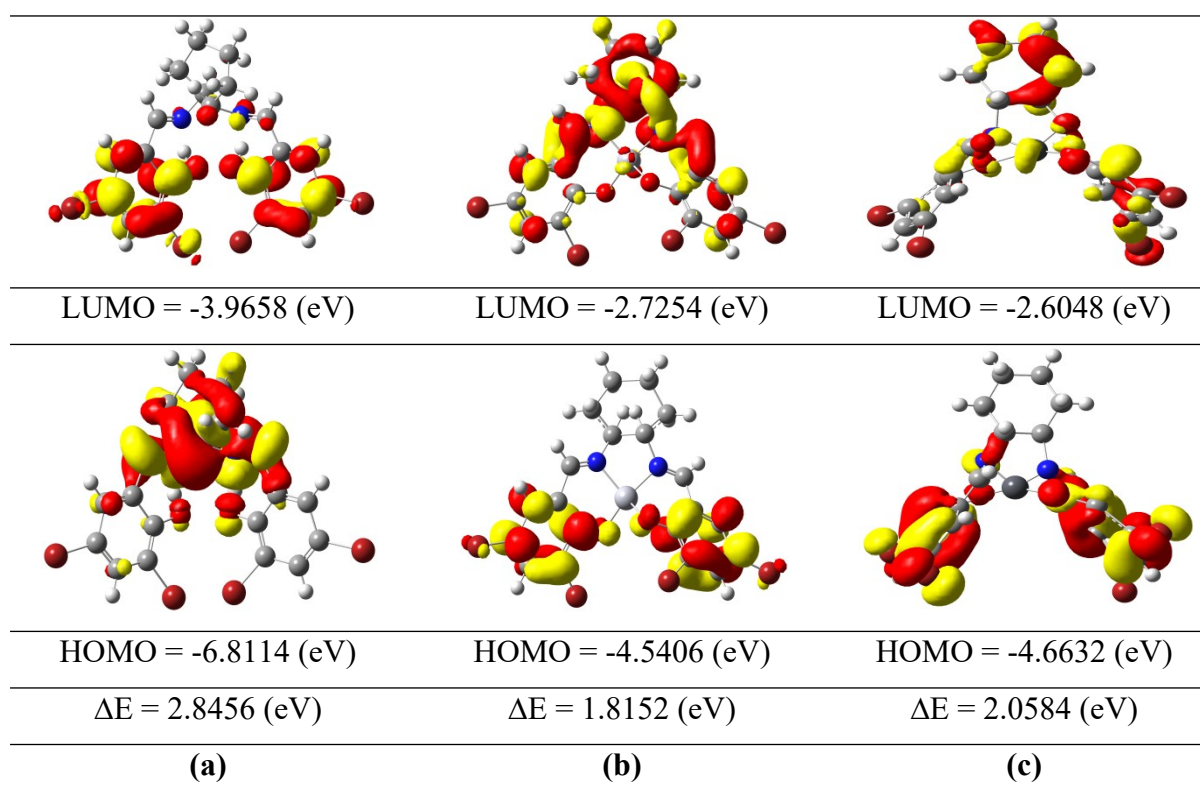


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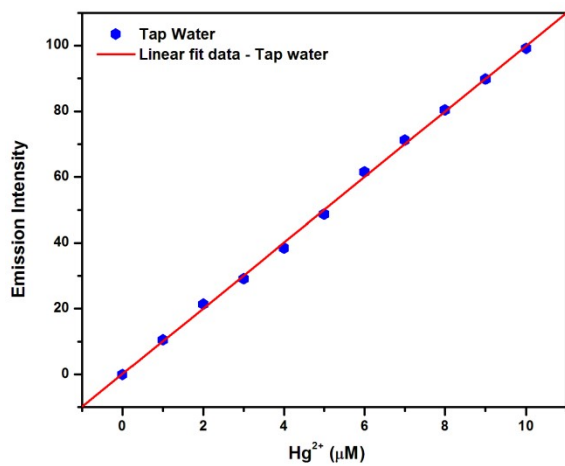


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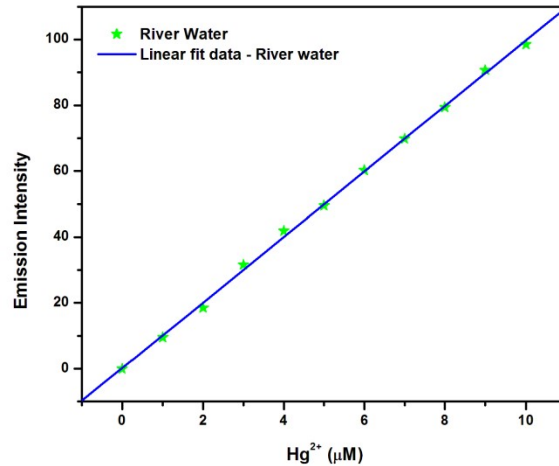
**Fig. S23.** (a) Fluorescence intensities of H<sub>2</sub>L.Hg<sup>2+</sup> (1:1) in the presence of EDTA for many cycles ( $\lambda_{\text{ex}} = 600 \text{ nm}$ ;  $\lambda_{\text{em}} = 762 \text{ nm}$ ); (b) Fluorescence intensities of H<sub>2</sub>L.Pb<sup>2+</sup> (1:1) in the presence of EDTA for many cycles ( $\lambda_{\text{ex}} = 575 \text{ nm}$ ;  $\lambda_{\text{em}} = 703 \text{ nm}$ )



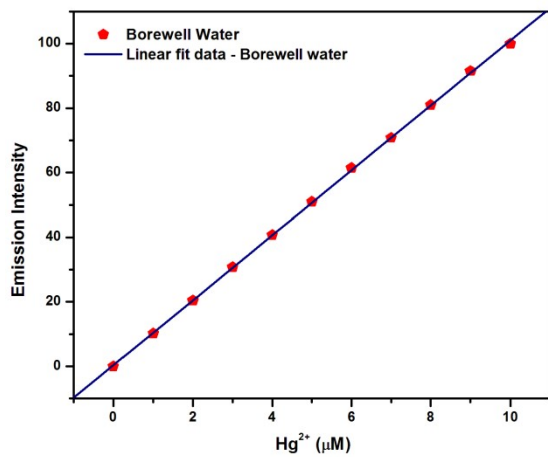
**Fig. S24.** FMO diagrams of (a)  $H_2L$ ; (b)  $H_2L.Hg^{2+}$  and (c)  $H_2L.Pb^{2+}$  with energy gap as calculated from the DFT method



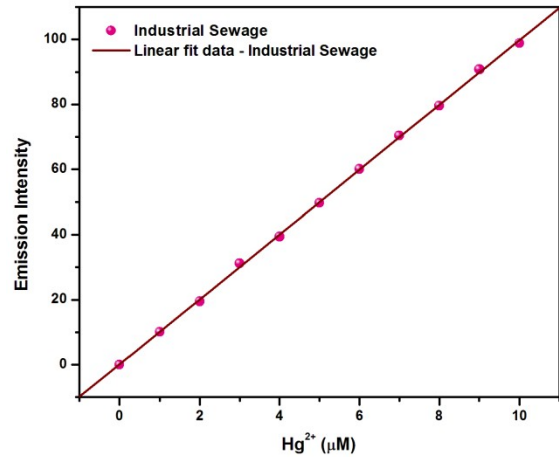
(a)



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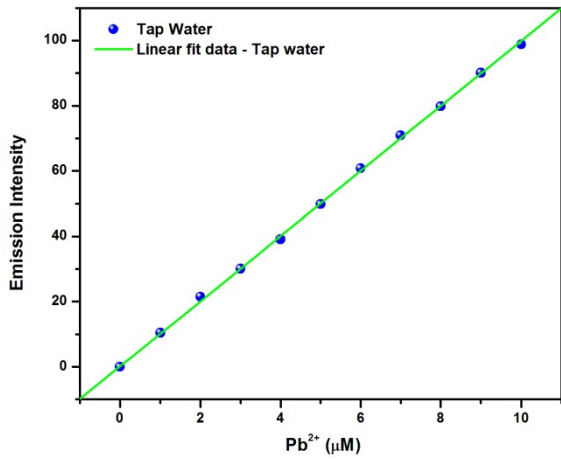


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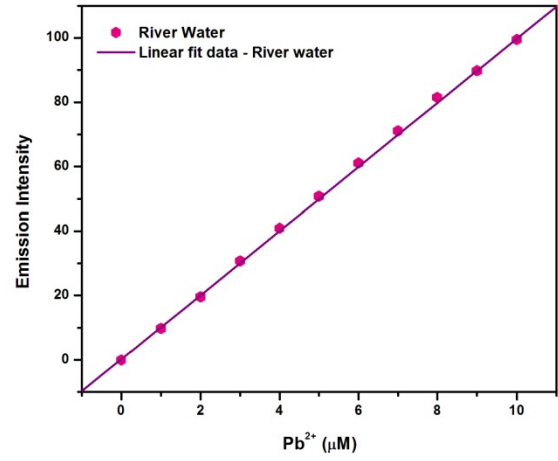


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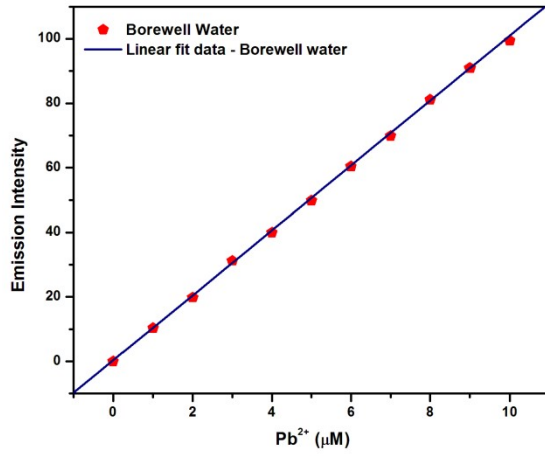
**Fig. S25.** Calibration sensitivity plot of H<sub>2</sub>L for Hg<sup>2+</sup>; (a) Tap water; (b) River water; (c) Bore well water and (d) Industrial sewage water



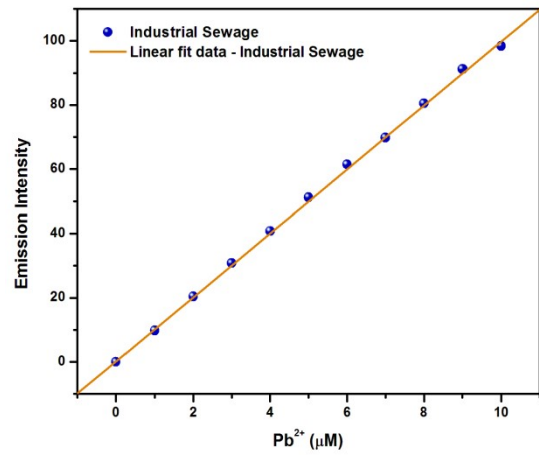
(a)



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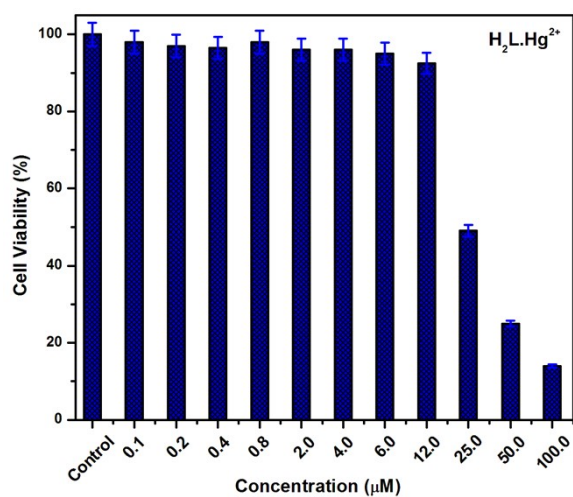


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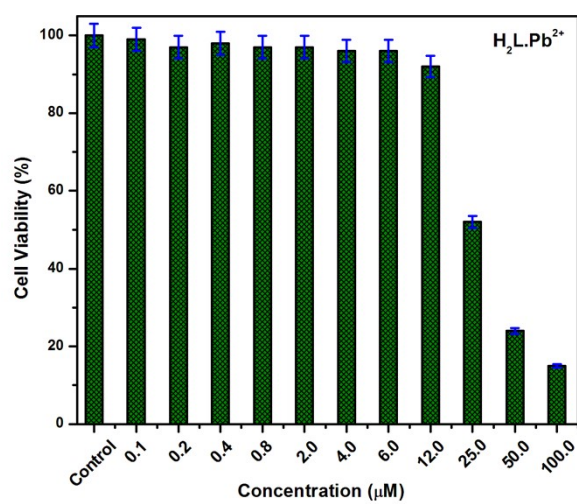


(d)

**Fig. S26.** Calibration sensitivity plot of H<sub>2</sub>L for Pb<sup>2+</sup>; (a) Tap water; (b) River water; (c) Bore well water and (d) Industrial sewage water

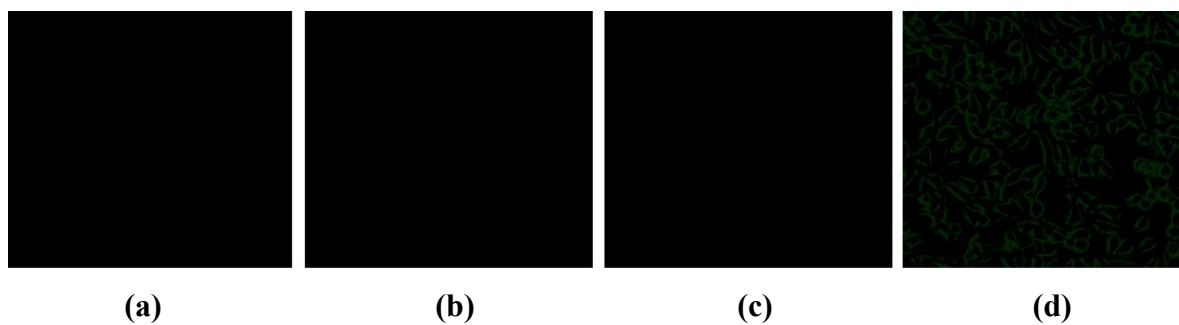


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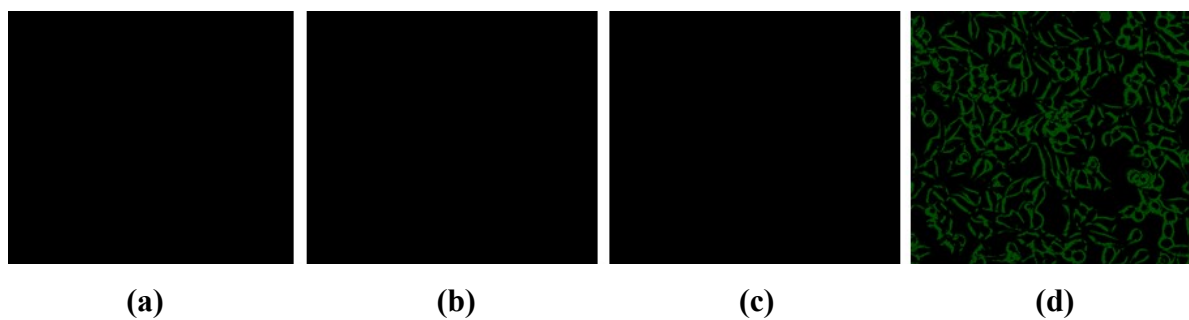


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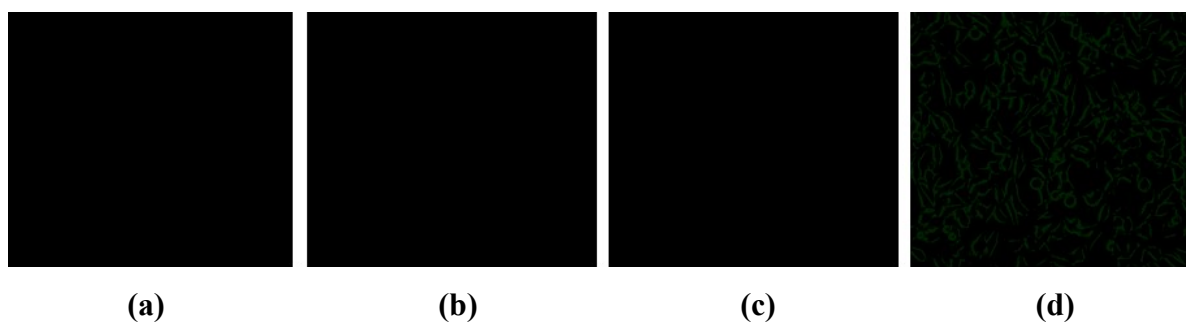
**Fig. S27.** MTT assay of (a)  $\text{H}_2\text{L.Hg}^{2+}$  and (b)  $\text{H}_2\text{L.Pb}^{2+}$  complex in the HeLa cell line



**Fig. S28.** Fluorescence bioimaging of HeLa cells; (a) control cells; (b) cells treated with 2  $\mu\text{M}$  H<sub>2</sub>L; (c) cells treated with 2  $\mu\text{M}$  Hg<sup>2+</sup>; (d) cells treated with 2  $\mu\text{M}$  Hg<sup>2+</sup> and H<sub>2</sub>L.

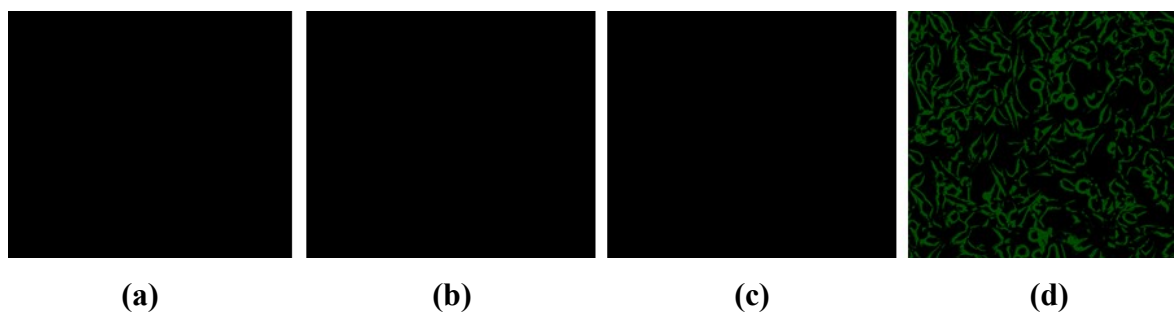


**Fig. S29.** Fluorescence bioimaging of HeLa cells; (a) control cells; (b) cells treated with 4  $\mu\text{M}$   $\text{H}_2\text{L}$ ; (c) cells treated with 4  $\mu\text{M}$   $\text{Hg}^{2+}$ ; (d) cells treated with 4  $\mu\text{M}$   $\text{Hg}^{2+}$  and  $\text{H}_2\text{L}$ .



**Fig. S30.** Fluorescence bioimaging of HeLa cells; (a) control cells; (b) cells treated with 2 μM H<sub>2</sub>L; (c) cells treated with 2 μM Pb<sup>2+</sup>; (d) cells treated with 2 μM Pb<sup>2+</sup> and H<sub>2</sub>L.



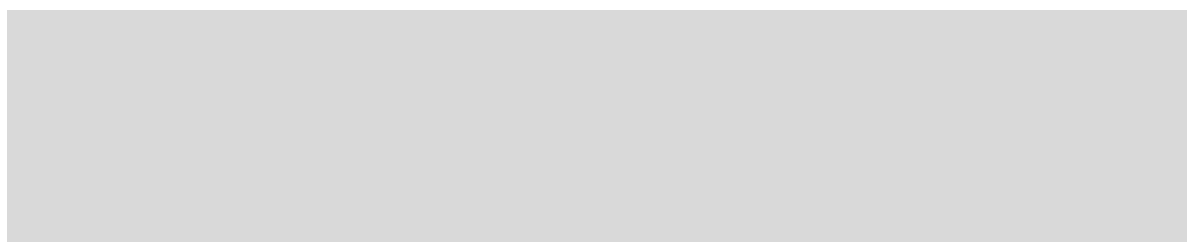


**Fig. S31.** Fluorescence bioimaging of HeLa cells; (a) control cells; (b) cells treated with 4 μM H<sub>2</sub>L; (c) cells treated with 4 μM Pb<sup>2+</sup>; (d) cells treated with 4 μM Pb<sup>2+</sup> and H<sub>2</sub>L.



(a) (b) (c) (d)

**Fig. S32.** Fluorescence images of HeLa cells with 1  $\mu\text{M}$  propidium iodide; (a) control cells; (b) cells treated with 2  $\mu\text{M}$   $\text{H}_2\text{L}$ ; (c) cells treated with 2  $\mu\text{M}$   $\text{Hg}^{2+}$ ; (d) cells treated with 2  $\mu\text{M}$   $\text{Hg}^{2+}$  and  $\text{H}_2\text{L}$ .



(a)

(b)

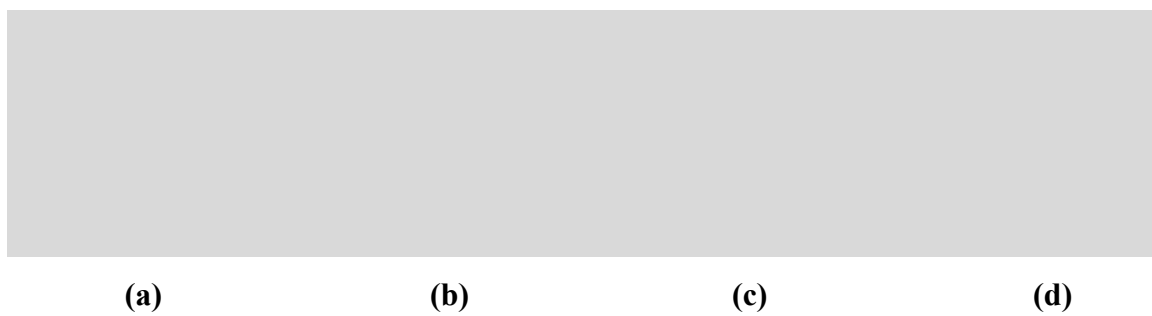
(c)

(d)

**Fig. S33.** Fluorescence images of HeLa cells with 1  $\mu\text{M}$  propidium iodide; (a) control cells; (b) cells treated with 4  $\mu\text{M}$   $\text{H}_2\text{L}$ ; (c) cells treated with 4  $\mu\text{M}$   $\text{Hg}^{2+}$ ; (d) cells treated with 4  $\mu\text{M}$   $\text{Hg}^{2+}$  and  $\text{H}_2\text{L}$ .



**Fig. S34.** Fluorescence images of HeLa cells with 1  $\mu\text{M}$  propidium iodide; (a) control cells; (b) cells treated with 2  $\mu\text{M}$   $\text{H}_2\text{L}$ ; (c) cells treated with 2  $\mu\text{M}$   $\text{Pb}^{2+}$ ; (d) cells treated with 2  $\mu\text{M}$   $\text{Pb}^{2+}$  and  $\text{H}_2\text{L}$ .



**Fig. S35.** Fluorescence images of HeLa cells with 1  $\mu\text{M}$  propidium iodide; (a) control cells; (b) cells treated with 4  $\mu\text{M}$   $\text{H}_2\text{L}$ ; (c) cells treated with 4  $\mu\text{M}$   $\text{Pb}^{2+}$ ; (d) cells treated with 4  $\mu\text{M}$   $\text{Pb}^{2+}$  and  $\text{H}_2\text{L}$ .

### Binding of H<sub>2</sub>L with Hg<sup>2+</sup>/Pb<sup>2+</sup> by UV-vis Method

The H<sub>2</sub>L.Hg<sup>2+</sup>/Pb<sup>2+</sup> binding constant was calculated using the Benesi-Hildebrand (B-H) plot.<sup>S1-S3</sup>

$$1/(A - A_0) = a/(a - b) \{1/K_a[M] + 1\} \dots\dots\dots(1)$$

where  $A_0$  is absorbance of free H<sub>2</sub>L,  $A$  is absorbance of H<sub>2</sub>L with Hg<sup>2+</sup>/Pb<sup>2+</sup> ions,  $K_a$  is the binding constant (M<sup>-1</sup>) and  $[M]$  is the concentration of Hg<sup>2+</sup>/Pb<sup>2+</sup> ions added during titration. The association constant ( $K_a$ ) could be determined from the slope of the straight line of the plot of  $1/(A - A_0)$  vs.  $[1/M^{\text{II}}]$ .

### Binding of H<sub>2</sub>L with Hg<sup>2+</sup>/Pb<sup>2+</sup> by Emission Method

In addition the binding constant value of Hg<sup>2+</sup>/Pb<sup>2+</sup> ions with H<sub>2</sub>L has been examined by emission spectroscopic method using the modified Benesi-Hildebrand equation,

$$1/(I - I_0) = 1/\{K(I_{\text{max}} - I_0)C\} + 1/(I_{\text{max}} - I_0) \dots\dots\dots(2)$$

where  $I_0$  is the emission intensity of H<sub>2</sub>L in the absence of Hg<sup>2+</sup>/Pb<sup>2+</sup> ions,  $I$  is the observed fluorescence intensity at that particular wavelength in the presence of a certain concentration of the Hg<sup>2+</sup>/Pb<sup>2+</sup> ( $C$ ),  $I_{\text{max}}$  is the maximum emission intensity value that was obtained during the titration with varying Hg<sup>2+</sup>/Pb<sup>2+</sup> concentration,  $K$  is the binding constant (M<sup>-1</sup>) and was determined from the slope of the linear plot and  $C$  is the concentration of the Hg<sup>2+</sup>/Pb<sup>2+</sup> ions added during titration.

### **Job Plot Technique**

The methanol/HEPES buffer (5 mM, pH 7.2; 1:9v/v) was used to dissolve the H<sub>2</sub>L (0.01M). 5.0 ml bottles were filled with 100, 90, 80, 70, 60, 50, 40, 30, 20, 10, and 0  $\mu$ L of the H<sub>2</sub>L solution. Each bottle received water until its total content reached 4.0 ml. Water was used to dissolve Hg<sup>2+</sup>/Pb<sup>2+</sup> ions (0.01M). Each diluted solution of H<sub>2</sub>L was mixed with 0, 10, 20, 30, 40, 50, 60, 70, 80, 90, and 100  $\mu$ L of the Hg<sup>2+</sup>/Pb<sup>2+</sup> ion solution. The total volume of each bottle was 5.0 ml. Absorption spectra were recorded at room temperature following a 5 min shake. Job's plots were drawn by plotting  $\Delta I \cdot X_h$  vs.  $X_h$ , where  $\Delta I$  = change of absorbance at 512 nm (for Hg<sup>2+</sup>) and 467 nm (for Pb<sup>2+</sup>) during titration and  $X_h$  is the mole fraction of Hg<sup>2+</sup>/Pb<sup>2+</sup> ions).

## pH Study

An automatic potentiometric titrator (HANNA-HI-902, USA) was used to perform the pH titrations at 310 K using a combination glass electrode (accuracy  $\pm 0.01$  pH unit). To calibrate the equipment, standard buffer solutions were used.<sup>S4</sup> Using  $\text{NaClO}_4$  as the supporting electrolyte, the ionic strength of each solution was brought down to 0.10 M. Based on measurements of  $[\text{H}^+]$ ,  $[\text{OH}^-]$ , and pH in many tests, the ion product of water ( $K = [\text{H}^+][\text{OH}^-]$ ) at 0.10 M  $\text{NaClO}_4$  in methanol/HEPES buffer (5 mM, pH 7.2; 1:9 v/v) combination was determined. The solution was bubbled with nitrogen gas both before to and throughout the titrations. There were several titrations performed for every system. From its solutions of concentrations ranging from  $1.0 \times 10^{-3}$  to  $3.0 \times 10^{-3}$  M, the dissociation constants ( $\text{p}K_a$ ) of  $\text{H}_2\text{L}$  were determined. The MINQUAD-75 program was used to calculate the  $\text{p}K_a$  values. Using HYSS, the concentration distribution profiles were acquired.<sup>S5</sup>



## Quantum Yield Calculation

Quantum yield ( $\Phi_x$ ) for H<sub>2</sub>L and H<sub>2</sub>L.Hg<sup>2+</sup>/Pb<sup>2+</sup> was calculated using standard solutions of fluorescein ( $\Phi_x = 0.79$ ) in methanol at an excitation wavelength 441 nm. The quantum yield was determined by the following eqn,

$$\Phi_x = \Phi_{st} \cdot (A_{st}/A_x) \cdot (F_x/F_{st}) \cdot (n_x^2/n_{st}^2) \cdot (D_x/D_{st})$$

Where,

$\Phi_x$  - Quantum yield of the sample

$\Phi_{st}$  - Quantum yield of the reference

$A_x, A_{st}$  - Absorbance of the sample and the reference

$F_x, F_{st}$  - Areas of emission for the sample and the reference

$n_x^2, n_{st}^2$  - Refractive indexes of the solvents

$D_x, D_{st}$  - Dilution factor of the sample and reference, respectively

### Calculation:

#### For Hg<sup>2+</sup>

$\Phi_{st} = 0.79$ ,  $A_{st} = 441$  nm,  $A_x = 512$  nm,  $F_x = 64025.8$ ,  $F_{st} = 109921.3$ ,  $(n) = 1.3335$ ,  $D_x = 0.002$  and  $D_{st} = 0.003$

$$\Phi_x = 0.79 \cdot (441/512) \cdot (64025.8/109921.3) \cdot (1.3335/1.3335) \cdot (0.002/0.003)$$

$$\Phi_x = 0.68045 \cdot (64025.8/109921.3) \cdot 1 \cdot 0.66667$$

$$\Phi_x = 0.45363 \cdot (64025.8/109921.3)$$

$$\Phi_x = 0.2642$$

#### For Pb<sup>II</sup>

$$\Phi_{st} = 0.79, A_{st} = 441 \text{ nm}, A_x = 467 \text{ nm}, F_x = 64098.2, F_{st} = 128456.8, (n) = 1.3335, D_x = 0.002 \text{ and } D_{st} = 0.003$$

$$\Phi_x = 0.79 \cdot (441/467) \cdot (64098.2/128456.8) \cdot (1.3335/1.3335) \cdot (0.002/0.003)$$

$$\Phi_x = 0.74602 \cdot (64098.2/128456.8) \cdot 1 \cdot 0.66667$$

$$\Phi_x = 0.49737 \cdot (64098.2/128456.8)$$

$$\Phi_x = 0.2132$$

**For H<sub>2</sub>L**

$$\Phi_{st} = 0.79, A_{st} = 441 \text{ nm}, A_x = 332 \text{ nm}, F_x = 25122.7, F_{st} = 189874.8, (n) = 1.3252, D_x = 0.002 \text{ and } D_{st} = 0.003$$

$$\Phi_x = 0.79 \cdot (441/332) \cdot (25748.9/179799.5) \cdot (1.3252/1.3252) \cdot (0.002/0.003)$$

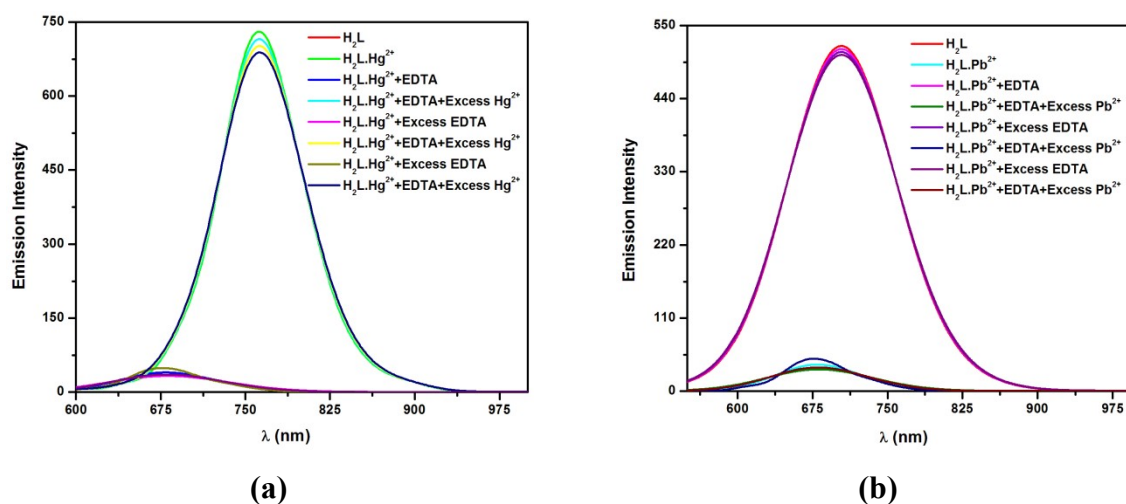
$$\Phi_x = 0.74602 \cdot (25748.9/179799.5) \cdot 1 \cdot 0.66667$$

$$\Phi_x = 0.69961 \cdot (25748.9/259799.5)$$

$$\Phi_x = 0.0693$$

## Reversibility Studies

EDTA disodium salt has demonstrated the reversible capability of the proposed sensor. Methanol (1.0 mL) was used to dissolve the H<sub>2</sub>L (10 μM), and 1.0 mL of H<sub>2</sub>L solution was combined with 4.0 ml of the Hg<sup>2+</sup>/Pb<sup>2+</sup> solution. After dissolving 0.5 mmol of EDTA in 5.0 ml of water, 2.0 mL of the EDTA solution was added to the H<sub>2</sub>L.Hg<sup>2+</sup>/Pb<sup>2+</sup> solution. Fluorescence spectra were recorded at room temperature after two minutes of mixing. The emission intensity of the H<sub>2</sub>L.Hg<sup>2+</sup>/Pb<sup>2+</sup> is quenched by the disodium salt of EDTA, a heavy metal ion chelator, suggesting that H<sub>2</sub>L reversibly coordinates to metal ions.



**Fig. S36.** Fluorescence changes of H<sub>2</sub>L (10 μM) after the addition of (a) Hg<sup>2+</sup>; (b) Pb<sup>2+</sup> and EDTA (1 equiv.) in methanol/HEPES buffer (5 mM, pH 7.2, 1:9 v/v)

**Table S1** Bond lengths [ $\text{\AA}$ ] for H<sub>2</sub>L

<b>Bond lengths [<math>\text{\AA}</math>]</b>	
Br(1)-C(3)	1.8882(18)
Br(2)-C(5)	1.8929(18)
O(1)-H(1)	0.8400
O(1)-C(2)	1.341(2)
N(1)-C(8)	1.462(2)
N(1)-C(7)	1.276(2)
C(8)-C(8)#1	1.533(3)
C(8)-C(9)	1.528(3)
C(9)-C(10)	1.523(3)
C(10)-C(10)#1	1.523(4)
Br(3)-C(13)	1.8911(17)
Br(4)-C(15)	1.8985(17)
O(2)-H(2)	0.8400
O(2)-C(12)	1.341(2)
N(2)-C(17)	1.281(2)
N(2)-C(18)	1.468(2)

**Table S2** Bond angles [°] for H<sub>2</sub>L

<b>Bond angles [°]</b>	
C(2)-O(1)-H(1)	109.5
C(7)-N(1)-C(8)	118.58(15)
N(1)-C(8)-C(8)#1	108.95(12)
N(1)-C(8)-H(8)	109.2
N(1)-C(8)-C(9)	109.62(14)
N(1)-C(7)-H(7)	119.3
N(1)-C(7)-C(1)	121.44(16)
C(4)-C(3)-Br(1)	118.56(13)
C(2)-C(3)-Br(1)	119.42(13)
O(1)-C(2)-C(3)	120.19(16)
O(1)-C(2)-C(1)	122.22(15)
C(3)-C(2)-C(1)	117.59(16)
C(6)-C(5)-Br(2)	120.15(14)
C(6)-C(5)-C(4)	121.20(17)
C(4)-C(5)-Br(2)	118.65(14)
C(12)-O(2)-H(2)	109.5
C(17)-N(2)-C(18)	116.96(14)
C(16)-C(15)-Br(4)	120.09(14)
C(14)-C(15)-Br(4)	118.72(13)
N(2)-C(17)-C(11)	122.36(16)
N(2)-C(17)-H(17)	118.8
C(12)-C(13)-Br(3)	119.19(13)
C(14)-C(13)-Br(3)	119.07(13)
N(2)-C(18)-C(18)#2	111.83(11)
N(2)-C(18)-H(18)	108.9
N(2)-C(18)-C(19)	109.54(13)
O(2)-C(12)-C(11)	121.78(15)
O(2)-C(12)-C(13)	120.25(15)

**Table S3** Torsion angles [°] for H<sub>2</sub>L

Torsion angles [°]	
Br(1)-C(3)-C(2)-O(1)	2.4(2)
Br(1)-C(3)-C(2)-C(1)	-176.88(12)
N(1)-C(8)-C(9)-C(10)	176.38(16)
N(1)-C(7)-C(1)-C(6)	175.51(16)
N(1)-C(7)-C(1)-C(2)	-2.6(3)
C(8)-N(1)-C(7)-C(1)	-178.38(15)
C(6)-C(1)-C(2)-O(1)	-179.18(16)
C(4)-C(3)-C(2)-O(1)	-179.92(16)
C(7)-N(1)-C(8)-C(8)#1	-121.56(18)
C(7)-N(1)-C(8)-C(9)	117.33(18)
C(7)-C(1)-C(2)-O(1)	-1.1(2)
C(3)-C(4)-C(5)-Br(2)	-179.19(13)
C(1)-C(6)-C(5)-Br(2)	-179.95(13)
C(5)-C(4)-C(3)-Br(1)	176.65(13)
Br(3)-C(13)-C(12)-O(2)	-3.3(2)
Br(3)-C(13)-C(12)-C(11)	175.36(12)
Br(3)-C(13)-C(14)-C(15)	-176.18(13)
Br(4)-C(15)-C(16)-C(11)	177.50(12)
Br(4)-C(15)-C(14)-C(13)	-178.55(13)
N(2)-C(18)-C(19)-C(20)	177.75(14)
C(17)-N(2)-C(18)-C(18)#2	108.82(19)
C(17)-N(2)-C(18)-C(19)	-130.47(16)
C(17)-C(11)-C(12)-O(2)	3.9(2)
C(16)-C(11)-C(17)-N(2)	178.70(16)
C(16)-C(11)-C(12)-O(2)	179.66(15)
C(18)-N(2)-C(17)-C(11)	175.40(14)
C(12)-C(11)-C(17)-N(2)	-5.5(2)
C(14)-C(13)-C(12)-O(2)	179.24(15)

Symmetry transformations used to generate equivalent atoms: #1  $-x+3/2, y, -z+3/2$ ; #2  $-x+3/2, y, -z+1/2$

**Table S4.** Hydrogen bonds for H<sub>2</sub>L (Å)

<b>Atom1</b>	<b>Atom2</b>	<b>Length</b>
H4	H9A	2.366
C3	C1	3.363
H6	O2	2.546
Br	H17	2.846
Br	H18	2.907

**Table S5.** Comparison of reported detection limit of Hg<sup>2+</sup> and Pb<sup>2+</sup> sensors with the present work.

S. No.	Compounds	Detection Limit	Ref.
1.	Cou-S	8.3 nM for Hg <sup>2+</sup> ; 10.5 nM for Pb <sup>2+</sup>	S6
2.	TBA	5.0 nM for Hg <sup>2+</sup> ; 300 pM for Pb <sup>2+</sup>	S7
3.	FNA	10.45 nM for Hg <sup>2+</sup> ; 2.65 nM for Pb <sup>2+</sup>	S8
4.	RPU	$7 \times 10^{-9}$ M for Pb <sup>2+</sup> ; $3.5 \times 10^{-8}$ M for Hg <sup>2+</sup>	S9
5.	MB	0.36 nmol/L for Hg <sup>2+</sup> ; 0.16 nmol/L for Pb <sup>2+</sup>	S10
6.	Cys-AgNPs	$45.39 \times 10^{-9}$ M for Hg <sup>2+</sup> ; $49.39 \times 10^{-9}$ M for Pb <sup>2+</sup>	S11
7.	AgNPs	$8.0 \times 10^{-7}$ M for Hg <sup>2+</sup> ; $2.0 \times 10^{-7}$ M for Pb <sup>2+</sup>	S12
8.	DNA-based sensor	10 pM for Pb <sup>2+</sup> ; 0.1 nM for Hg <sup>2+</sup>	S13
9.	PBA and HBA	0.98 pM for Pb <sup>2+</sup> ; 19 pM for Hg <sup>2+</sup>	S14
10	poly(2-VP-MBAm-AA)	10 $\mu\text{g L}^{-1}$ for Hg <sup>2+</sup> & Pb <sup>2+</sup>	S15
11.	Present Work	8.29 nM for Hg <sup>2+</sup> ; 7.65 nM for Pb <sup>2+</sup>	-



**Table S6.** Determination of Hg<sup>2+</sup> in real water samples with H<sub>2</sub>L

<b>Sample</b>	<b>pH</b>	<b>Added Hg<sup>2+</sup> (<math>\mu</math>M)</b>	<b>Found Hg<sup>2+</sup> (<math>\mu</math>M)</b>	<b>Recovery (%)</b>	<b>RSD (%)</b>
Tap water	6.94	0.1	0.097	97.0	1.40
		0.5	0.496	99.2	
		1.0	0.996	99.6	
River water	7.08	0.1	0.096	96.0	2.08
		0.5	0.497	99.4	
		1.0	0.998	99.8	
Bore well water	7.43	0.1	0.099	99.0	0.31
		0.5	0.493	98.6	
		1.0	0.992	99.2	
Industrial Sewage	7.99	0.1	0.097	97.0	1.47
		0.5	0.496	99.2	
		1.0	0.998	99.8	

RSD = Relative standard deviation

**Table S7.** Determination of Pb<sup>2+</sup> in real water samples with H<sub>2</sub>L

<b>Sample</b>	<b>pH</b>	<b>Added Pb<sup>2+</sup> (<math>\mu</math>M)</b>	<b>Found Pb<sup>2+</sup> (<math>\mu</math>M)</b>	<b>Recovery (%)</b>	<b>RSD (%)</b>
Tap water	6.94	0.1	0.099	99.0	0.38
		0.5	0.498	99.6	
		1.0	0.997	99.7	
River water	7.08	0.1	0.098	98.0	0.79
		0.5	0.496	99.2	
		1.0	0.995	99.5	
Bore well water	7.43	0.1	0.099	99.0	0.35
		0.5	0.497	99.4	
		1.0	0.997	99.7	
Industrial Sewage	7.99	0.1	0.096	96.0	2.11
		0.5	0.498	99.6	
		1.0	0.997	99.7	

RSD = Relative standard deviation

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