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Gaining insights into nutrient and metal element distributions in radix

***Angelicae sinensis* by mass spectrometry**

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Table S1: Regression equations, determination coefficients (R^2), and linear ranges in $\mu\text{g}\cdot\text{mL}^{-1}$ of 24 standard chemicals for quantitative analysis.

No	Compound	Regression equation	Determination coefficient	Linear range/ $\mu\text{g}\cdot\text{mL}^{-1}$
1	ferulic acid	$y = 24835x - 2879.1$	0.9991	0–10.00
2	isoferulic acid	$y = 34979x - 4469.8$	0.9992	0.05–10.00
3	vanillic acid	$y = 20205x + 63599$	0.9942	0–50.00
4	caffeic acid	$y = 3824x + 2525.4$	0.9912	0.005–0.20
5	thymine	$y = 16997x + 228.94$	0.9995	0–1.00
6	guanine	$y = 16912x + 92.645$	0.9989	0–10.00
7	adenine	$y = 40789x + 554.02$	0.9999	0–2.00
8	cytidine	$y = 81618x + 1310.3$	0.9997	0–0.20
9	γ -aminobutyric acid	$y = 7926x + 778.9$	0.9995	0.08–8.00
10	<i>L</i> -hydroxyproline	$y = 64.461x + 3803.3$	0.9967	0–800.00
11	<i>L</i> -glutamine	$y = 600000x + 200000$	0.9961	0–3.00
12	<i>L</i> -asparagine	$y = 25429x + 5844.7$	0.9960	0–5.00
13	<i>Z</i> -ligustilide	$y = 657.48x + 1668.9$	0.9989	0–10.00
14	senkyunolide I	$y = 81742x + 27863$	0.9990	0–5.00
15	glutamic acid	$y = 41405x + 1927.1$	0.9985	0–3.00
16	<i>L</i> -arginine	$y = 166380x + 282.62$	0.9998	0–8.00
17	<i>L</i> -phenylalanine	$y = 154005x + 660.58$	0.9986	0–1.00
18	<i>L</i> -proline	$y = 2490.6x + 59.458$	0.9984	0–8.00
19	<i>L</i> -valine	$y = 125545x + 852.05$	0.9990	0–8.00
20	<i>L</i> -alanine	$y = 22407x + 2126.1$	0.9999	0–5.00
21	<i>L</i> -leucine	$y = 12884x + 330.02$	0.9986	0.008–5.00
22	<i>L</i> -tyrosine	$y = 124919x + 1189.2$	0.9994	0.005–0.50
23	<i>L</i> -tryptophan	$y = 107038x + 9287.3$	0.9973	0–5.00
24	<i>L</i> -ornithine	$y = 75319x + 12207$	0.9979	0–50.00

Table S2: Precision, repeatability and stability of the UPLC-MS/MS analysis of 24 standard chemicals and five individual aqueous extracts of one exemplary radix *Angelicae sinensis* powder sample, with respect to the relative standard deviation (RSD) of the corresponding integrated peak areas in percentage (%).

Compound	Intraday precision / %	Three-day stability / %	Sample repeatability / %
ferulic acid	1.68	2.16	1.81
isoferulic acid	2.09	2.17	2.41
vanillic acid	2.41	2.21	2.47
caffeic acid	1.97	2.43	0.55
thymine	1.91	1.89	2.06
guanine	1.17	1.26	1.30
adenine	2.26	1.69	1.80
cytidine	2.41	2.33	2.20
γ -aminobutyric acid	1.75	0.99	0.78
<i>L</i> -hydroxyproline	2.24	2.32	2.43
<i>L</i> -glutamine	2.38	2.19	1.01
<i>L</i> -asparagine	2.08	1.06	1.07
<i>Z</i> -ligustilide	2.25	1.67	1.85
senkyunolide I	2.06	1.17	1.31
glutamic acid	2.49	1.34	0.86
<i>L</i> -arginine	1.50	1.73	1.87
<i>L</i> -phenylalanine	1.94	1.09	1.00
<i>L</i> -proline	1.87	1.72	1.25
<i>L</i> -valine	2.25	1.46	1.00
<i>L</i> -alanine	1.75	1.96	2.11
<i>L</i> -leucine	2.15	1.91	1.61
<i>L</i> -tyrosine	2.38	1.57	1.73
<i>L</i> -tryptophan	1.25	1.22	1.34
<i>L</i> -ornithine	1.60	1.59	1.29

To inspect the stability, reproducibility and repeatability of the UPLC-MS/MS analysis, the mixture solution of 24 standard chemicals in 2 μ L was successively tested six times on one day using a single aliquot (*i.e.*, $n=6$). At the same time, over a three-day period fresh aliquots of the mixture solution were removed from cold storage, and continuously analyzed six times every day (*i.e.*, $n=18$). On the other hand, five powder aliquots were sampled from one exemplary RAS material (*i.e.*, sample #1 in sample set1) by weighing 5.0 g for each, and the aqueous extraction was followed. All of the mixture standard solution and the sample extracted solutions were sequentially loaded to the UPLC-MS/MS for measurement. The area under the chromatographic peak acquired at quantitative production ion was integrated for each compound after necessary baseline correction, and the relative standard deviation between the peak areas was accordingly calculated.

Table S3: Recovery of 24 standard chemicals concerning their concentrations in aqueous extract of powder sample, spiking solution, really measured value, and relative standard deviation (RSD) in percentage (%).

$$\text{Recovery} = (c_{\text{measured}} - c_{\text{sample}}) / c_{\text{spiking}} \times 100\%.$$

Compound	Sample concentration (c_{sample})/mg•g ⁻¹	Spiking concentration (c_{spiking})/mg•g ⁻¹	Measured concentration (c_{measured})/mg•g ⁻¹	Recovery / %	Averaged recovery / %	RSD / %
ferulic acid	1.410	0.70	2.144	104.86	103.75	1.78
	1.403	1.40	2.870	104.79		
	1.402	2.10	3.536	101.62		
isoferulic acid	0.241	0.12	0.357	96.67	97.91	1.10
	0.236	0.20	0.433	98.50		
	0.235	0.28	0.511	98.57		
vanillic acid	4.810	2.50	7.275	98.60	98.56	2.08
	4.802	3.50	8.179	96.86		
	4.815	4.50	9.342	100.60		
caffeic acid	0.326	0.15	0.477	100.67	99.06	2.03
	0.329	0.25	0.571	96.80		
	0.327	0.35	0.676	99.41		
thymine	0.038	0.02	0.058	100.00	98.06	1.77
	0.039	0.03	0.068	96.67		
	0.038	0.04	0.077	97.50		
guanine	0.387	0.20	0.589	101.00	98.61	2.10
	0.388	0.40	0.778	97.50		
	0.388	0.60	0.972	97.33		
adenine	0.038	0.02	0.057	95.00	96.39	1.32
	0.039	0.03	0.068	96.67		
	0.039	0.04	0.078	97.50		
cytidine	0.008	0.004	0.012	102.50	100.28	2.09
	0.008	0.006	0.014	98.33		
	0.008	0.008	0.016	100.00		
γ -aminobutyric acid	1.096	0.50	1.641	109.00	107.82	1.95
	1.093	1.00	2.147	105.40		
	1.098	1.50	2.734	109.07		
<i>L</i> -hydroxyproline	4.847	2.50	7.267	96.80	98.98	1.91
	4.931	4.50	9.433	100.04		
	4.764	6.50	11.271	100.11		
<i>L</i> -glutamine	1.805	0.75	2.548	99.07	101.22	1.88
	1.803	1.50	3.332	101.93		
	1.802	2.25	4.112	102.67		
<i>L</i> -asparagine	0.140	0.07	0.211	101.43	100.23	2.06
	0.142	0.14	0.279	97.86		
	0.141	0.21	0.354	101.43		

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	2.637	1.30	3.979	103.23		
Z-ligustilide	2.630	2.60	5.355	104.81	102.75	2.27
	2.633	3.90	6.541	100.21		
	0.565	0.30	0.853	96.00		
senkyunolide I	0.561	0.60	1.151	98.33	97.67	1.49
	0.567	0.90	1.455	98.67		
	0.122	0.06	0.183	101.67		
glutamic acid	0.121	0.12	0.244	102.50	100.84	2.19
	0.122	0.18	0.299	98.33		
	1.132	0.50	1.646	102.80		
L-arginine	1.138	1.00	2.145	100.70	103.19	2.62
	1.113	1.50	2.704	106.07		
	0.164	0.08	0.248	105.00		
L-phenylalanine	0.164	0.16	0.328	102.50	103.75	1.20
	0.163	0.24	0.412	103.75		
	0.576	0.30	0.873	99.00		
L-proline	0.572	0.60	1.185	102.17	101.09	1.79
	0.577	0.90	1.496	102.11		
	0.191	0.10	0.292	101.00		
L-valine	0.192	0.20	0.391	99.50	100.94	1.40
	0.192	0.30	0.499	102.33		
	2.070	1.00	3.102	103.20		
L-alanine	2.090	2.00	4.175	104.25	102.51	2.12
	2.060	3.00	5.062	100.07		
	0.198	0.10	0.299	101.00		
L-leucine	0.197	0.20	0.403	103.00	101.89	1.00
	0.200	0.30	0.505	101.67		
	0.064	0.03	0.095	103.33		
L-tyrosine	0.063	0.06	0.124	101.67	102.78	0.94
	0.062	0.09	0.155	103.33		
	0.568	0.25	0.815	98.80		
L-tryptophan	0.566	0.50	1.078	102.40	99.91	2.16
	0.569	0.75	1.308	98.53		
	1.534	0.75	2.304	102.67		
L-ornithine	1.525	1.50	3.064	102.60	101.31	2.26
	1.533	2.25	3.753	98.67		

To examine the recovery rate, a total of nine powder aliquots in three groups were first made from one single powder sample by weighing *ca.* 5.0 g for each. Each group was composed of three aliquots. Then, three different levels of each of 24 standard chemical solutions were individually spiked to the aliquots. Their aqueous solutions were prepared and tested for UPLC-MS/MS. The area under the chromatographic peak acquired at quantitative production ion was integrated after necessary baseline correction, and both the recoveries and relative standard deviations were accordingly calculated for each of 24 standard chemicals.

Table S4: ICP-MS instrumental operating parameters.

Parameter	Value	Parameter	Value
Radio-frequency power/W	1100	Intercept the cone aperture/mm	0.7
Atomizing gas flow rate/(L•min ⁻¹)	0.9	Sample depth/mm	6.6
Flow rate of plasma/(L•min ⁻¹)	11	Pulsed voltage/V	950
Auxiliary gas flow/(L•min ⁻¹)	1	Vacuum degree of analysis room/Pa	0.00067
Fog room	double access	Identify the valve	15
Atomizing chamber temperature/°C	2	Compensation standard for single bar	-17
Sampling cone aperture/mm	1.1	Compensation standard for four-stage rods	0

Table S5: AAS instrumental operating parameters.

Element	Lamp current / mA	λ / nm	Pass band / nm	Flame type	Acetylene flow / L•min ⁻¹	Oxygen flow / L•min ⁻¹
Zn	6	213.9	0.2	air-acetylene	1.2	6.5
Fe	8	248.3	0.2	air-acetylene	0.9	6.5
Mn	4	279.5	0.2	air-acetylene	1.0	6.5
Mg	4	285.2	0.5	air-acetylene	1.1	6.5
Ca	5	422.7	0.5	air-acetylene	1.4	6.5
Na	6	589.0	0.2	air-acetylene	1.1	6.5
K	5	766.5	0.5	air-acetylene	1.2	6.5

Table S6: 24 target compounds in aqueous extracts of radix *Angelicae sinensis* materials and corresponding fragment ions (M/Z), retention time (Rt) in the UPLC-MS/MS analysis in positive ion mode.

No	Compound	Ion mode	Molecular mass	Fragment ions (M/Z)	Rt/min	Formula
1	ferulic acid	[M+H] ⁺	194	88.9/177.1	8.49	C ₁₀ H ₁₀ O ₄
2	isoferulic acid	[M+H] ⁺	194	89.1/177.2	9.08	C ₁₀ H ₁₀ O ₄
3	vanillic acid	[M+H] ⁺	168	93.2/151.3	5.71	C ₈ H ₈ O ₄
4	caffeic acid	[M+H] ⁺	180	89.2/162.9	8.16	C ₉ H ₈ O ₄
5	thymine	[M+H] ⁺	126	54.1/110.2	1.33	C ₅ H ₆ N ₂ O ₂
6	guanine	[M+H] ⁺	151	110.3/135.0	0.81	C ₅ H ₅ N ₅ O
7	adenine	[M+H] ⁺	135	67.1/119.1	0.80	C ₅ H ₅ N ₅
8	cytidine	[M+H] ⁺	243	95.2/112.2	0.79	C ₉ H ₁₃ N ₃ O ₅
9	γ -aminobutyric acid	[M+H] ⁺	103	69.0/87.3	0.68	C ₄ H ₉ NO ₂
10	<i>L</i> -hydroxyproline	[M+H] ⁺	131	68.1/86.0	0.70	C ₅ H ₉ NO ₃
11	<i>L</i> -glutamine	[M+H] ⁺	146	65.1/93.1	20.5	C ₅ H ₁₀ N ₂ O ₃
12	<i>L</i> -asparagine	[M+H] ⁺	132	87.0	0.61	C ₄ H ₈ N ₂ O ₃
13	<i>Z</i> -ligustilide	[M+H] ⁺	190	115.1/173.2	18.25	C ₁₂ H ₁₄ O ₂
14	senkyunolide I	[M+H] ⁺	224.2	207.0	10.00	C ₁₂ H ₁₆ O ₄
15	glutamic acid	[M+H] ⁺	147	102.2/130.4	0.69	C ₅ H ₉ NO ₄
16	<i>L</i> -arginine	[M+H] ⁺	174	70.1/130.2	0.64	C ₆ H ₁₄ N ₄ O ₂
17	<i>L</i> -phenylalanine	[M+H] ⁺	165	120.1/103.3	2.06	C ₉ H ₁₁ NO ₂
18	<i>L</i> -proline	[M+H] ⁺	115	70.1/130.2	0.71	C ₅ H ₉ NO ₂
19	<i>L</i> -valine	[M+H] ⁺	117	55.2/72.1	0.81	C ₅ H ₁₁ NO ₂
20	<i>L</i> -alanine	[M+H] ⁺	89	44.1/72.1	0.68	C ₃ H ₇ NO ₂
21	<i>L</i> -leucine	[M+H] ⁺	131	86.4	1.20	C ₆ H ₁₃ NO ₂
22	<i>L</i> -tyrosine	[M+H] ⁺	181	136.1/165.2	1.07	C ₉ H ₁₁ NO ₃
23	<i>L</i> -tryptophan	[M+H] ⁺	204	146.1/188.2	3.78	C ₁₁ H ₁₂ N ₂ O ₂
24	<i>L</i> -ornithine	[M+H] ⁺	132	116.3	0.68	C ₅ H ₁₂ N ₂ O ₂

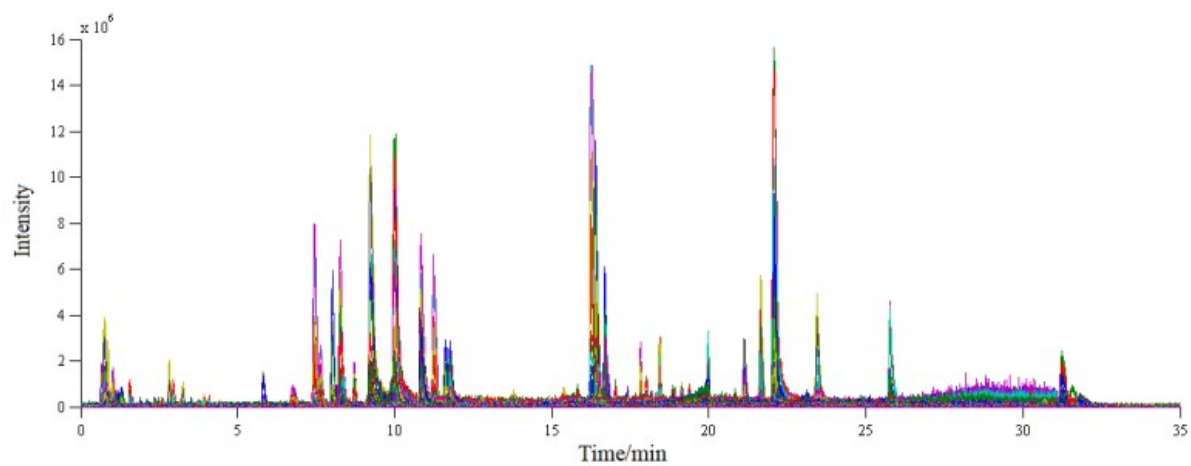


Figure S1: Original UPLC-MS/MS chromatograms acquired in the positive ion mode from exemplary aqueous extract solution of one radix *Angelicae sinensis* powder in sample set1.

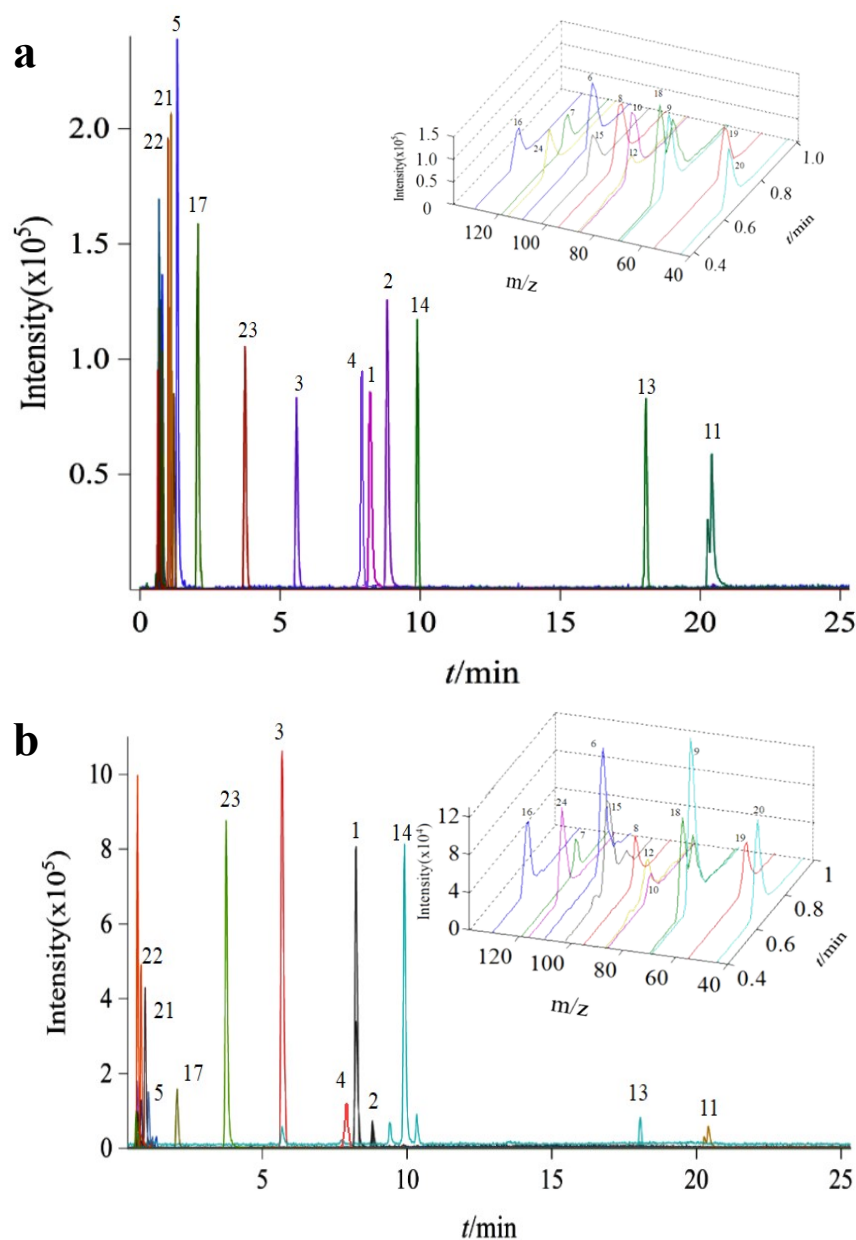


Figure S2: Chromatograms acquired at the individual quantitative production ions in the positive ion mode from (a) the solutions of 24 standard chemicals and (b) the exemplary aqueous extract solution of one radix *Angelicae sinensis* powder in sample set1.

1–ferulic acid, 2–isoferulic acid, 3–vanillic acid, 4–caffeic acid, 5–thymine, 6–guanine, 7–adenine, 8–cytidine, 9– γ -aminobutyric acid, 10–L-hydroxyproline, 11–L-glutamine, 12–L-asparagine, 13–Z-ligustilide, 14–senkyunolide I, 15–glutamic acid, 16–L-arginine, 17–L-phenylalanine, 18–L-proline, 19–L-valine, 20–L-alanine, 21–L-leucine, 22–L-tyrosine, 23–L-tryptophan, 24–L-ornithine.

Table S7: Determination (in mg•g⁻¹) of 24 target nutrient compounds in aqueous extracts of 287 radix *Angelicae sinensis* materials in the sample set1 originated from three provincial origins of Gansu (A), Qinghai (B) and Yunnan (C), together with the corresponding values of mean and standard deviation (STD).

Class	No.	Compound	Content / mg•g ⁻¹	mean / mg•g ⁻¹			STD mg•g ⁻¹
				A	B	C	
Organic acids	1	ferulic acid	0.104–7.314	0.359	0.541	1.474	1.123
	2	isoferulic acid	0.111–0.791	0.156	0.183	0.275	0.136
	3	vanillic acid	0.004–29.080	3.798	3.818	5.408	3.370
	4	caffeic acid	0.010–7.476	0.569	0.335	0.667	0.769
	5	<i>L</i> -arginine	0.000–2.543	0.678	0.218	0.683	0.643
	6	<i>L</i> -phenylalanine	0.000–0.461	0.065	0.054	0.055	0.064
	7	<i>L</i> -proline	0.001–4.679	0.781	0.409	0.911	0.881
	8	<i>L</i> -valine	0.000–1.059	0.059	0.109	0.045	0.107
	9	<i>L</i> -alanine	0.001–2.303	0.539	0.469	0.602	0.498
Amino acids	10	<i>L</i> -leucine	0.000–3.897	0.095	0.119	0.200	0.327
	11	<i>L</i> -tyrosine	0.000–0.282	0.055	0.046	0.048	0.051
	12	<i>L</i>-tryptophan	0.210–1.967	0.599	0.868	1.145	0.369
	13	<i>L</i> -ornithine	0.003–24.842	4.376	1.495	3.937	5.037
	14	<i>L</i> -glutamine	0.142–2.250	1.000	0.821	0.786	0.561
	15	<i>L</i> -hydroxyproline	0.021–349.856	29.447	15.239	16.313	32.000
	16	<i>L</i> -asparagine	0.001–1.793	0.375	0.173	0.237	0.375
	17	glutamic acid	0.000–1.538	0.181	0.099	0.189	0.226
	18	γ -aminobutyric acid	0.002–6.966	0.994	0.630	1.173	1.190
Nucleosides	19	thymine	0.001–0.127	0.017	0.015	0.011	0.020
	20	guanine	0.005–6.097	0.661	0.265	0.726	0.748
	21	adenine	0.000–1.387	0.068	0.029	0.089	0.117
	22	cytidine	0.001–7.536	0.281	0.204	0.204	0.542
Phthalides	23	<i>Z</i>-ligustilide	0.016–3.502	1.312	1.663	1.575	0.597
	24	senkyunolide I	0.001–2.597	0.587	0.662	0.875	0.444

Table S8: Result of Kruskal-Wallis H test and post-hoc multiple comparison of the content of 24 nutrients in aqueous extracts of 241 normal radix *Angelicae sinensis* batches in the sample set1 originated from three provincial origins of Gansu (A), Qinghai (B) and Yunnan (C).

Compound	H value	P value	Adjusted P value (Bonferroni)		
			A vs. B	A vs. C	B vs. C
ferulic acid	20.036	<0.001***	0.015*	<0.001***	0.589
isoferulic acid	28.995	<0.001***	0.002**	0.001**	<0.001***
vanillic acid	22.079	<0.001***	0.238	<0.001***	0.027**
guanine	38.982	<0.001***	<0.001***	0.427	<0.001***
adenine	13.065	0.001***	0.008**	0.844	0.002**
cytidine	7.978	0.019*	0.015*	1.000	0.180
γ -aminobutyric acid	11.039	0.004**	0.037*	0.516	0.004**
<i>L</i> -hydroxyproline	8.026	0.018*	0.016*	0.629	0.698
<i>L</i> -asparagine	6.778	0.034*	0.030*	1.000	0.239
<i>Z</i> -ligustilide	9.984	0.007**	0.032*	0.034*	1.000
senkyunolide I	18.958	<0.001***	0.071	<0.001***	0.216
glutamic acid	18.911	<0.001***	<0.001***	0.261	0.123
<i>L</i> -arginine	32.436	<0.001***	<0.001***	1.000	<0.001***
<i>L</i> -phenylalanine	10.883	0.004**	0.003**	0.792	0.257
<i>L</i> -proline	35.525	<0.001***	<0.001***	1.000	<0.001***
<i>L</i> -alanine	27.840	<0.001***	<0.001***	1.000	<0.001***
<i>L</i> -tyrosine	7.041	0.030*	0.034*	0.407	1.000
<i>L</i>-tryptophan	83.197	<0.001***	<0.001***	<0.001***	0.028
<i>L</i> -ornithine	25.361	<0.001***	<0.001***	1.000	0.001**
caffeic acid	4.949	0.084	–	–	–
thymine	0.863	0.649	–	–	–
<i>L</i> -glutamine	4.261	0.119	–	–	–
<i>L</i> -valine	2.362	0.307	–	–	–
<i>L</i> -leucine	2.398	0.301	–	–	–

Note: * $P < 0.05$ for statistically significant differences in the nutrient content among the batches across three origins, ** $P < 0.01$ for highly significant differences, and *** $P < 0.001$ for extremely significant differences.

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The distribution of the IBR values of the content responses of 24 nutrients of 241 radix *Angelicae sinensis* materials in the sample set1 was visualized in histogram for each class of 132 A-batches, 59 B-batches and 50 C-batches, respectively (Figure S3). It was pronounced that the IBR values were not normally distributed. Therefore, excluding 46 abnormal samples, Kruskal-Wallis *H* test and post-hoc multiple comparison were carried out on the IBR values. The comparison of the IBR distributions unraveled that: (i) there presented significant differences in content response between the A-batches, B-batches and C-batches; (ii) the C-batches had larger IBRs in terms of its median value of 9.86, than those of both A-batches (with IBR median = 5.52) and B-batches (with IBR median =2.59). The 25th–75th percentile ranges of IBR values were 2.41–9.99 for A-batches, 0.91–7.26 for B-batches, 5.48–16.76 for C-batches, respectively.

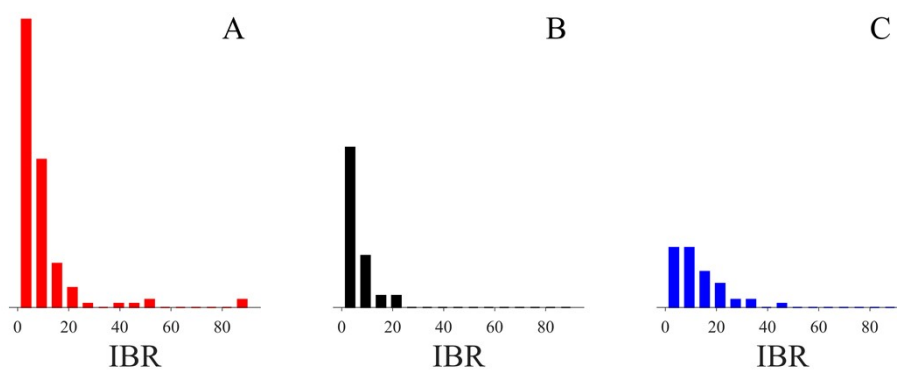


Figure S3: Distributions of the IBR values calculated from the content responses of 24 nutrients of 241 radix *Angelicae sinensis* materials in sample set1 originated from three geographic origins of Gansu (A), Qinghai (B) and Yunnan (C).

Table S9: Calibrations of 15 metal elements resulted from ICP-MS and AAS measurements ($n=3$)

Element	Regression equation	Determination coefficient (R^2)	Linear range/ $\mu\text{g}\cdot\text{L}^{-1}$	Limit of detection (LOD) $\mu\text{g}\cdot\text{L}^{-1}$, ($n=15$)
ICP-MS Method				
Pb	$y=9641.8x$	1.0000	0–200	0.0213
Cd	$y=994.1x$	0.9998	0–200	0.0196
As	$y=770.2x$	0.9996	0–200	0.0168
Hg	$y=1123.6x$	0.9998	0–200	0.0215
Sb	$y=3144.2x$	1.0000	0–200	0.0189
Ni	$y=1734.1x$	1.0000	0–200	0.0176
Cu	$y=3887.3x$	1.0000	0–200	0.0241
Cr	$y=6494.8x$	0.9988	0–200	0.0192
AAS Method				
Zn	$y=0.0857x+0.2060$	0.9978	500–2500	10
Fe	$y=0.0426x+0.0109$	0.9986	1000–5000	18
Mn	$y=0.0955x+0.0130$	0.9982	200–2500	11
Mg	$y=0.2659x+0.1035$	0.9992	500–2500	3
Ca	$y=0.0283x+0.0020$	0.9996	1000–5000	11
Na	$y=0.1970x+0.1962$	0.9994	500–2500	12
K	$y=0.1156x+0.0769$	0.9984	500–2500	15

The ICP-MS and AAS data were collected in triplicate experimental runs ($n=3$), and the mean values were subsequently calculated for each of both standard and digested solutions, and used for the quantitative calibration and prediction. Meanwhile, the limits of detection (LODs) were estimated as triple standard deviation of the signal acquired from 15 independently prepared reagent blanks ($n=15$). The linear ranges were also individually computed, and all seemed satisfactory. In order to validate this testing method, a reference material of CRM GSB-26-celery (GBW 09607) was used, as received from the Institute of Geophysical and Geochemical Exploration (IGGE), Chinese Academy of Geological Sciences, China. As a result, the ultimate determinations of the celery by such an ICP-MS and AAS method were in good agreement with the certified reference values (data not shown), which indicated that the precision of the present element quantitative analysis was acceptable.

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Table S10: Kruskal-Wallis H test result for the contents of 14 metals in the RAS materials of the sample set2 originated from ten planting origins of Gansu.

Element	P value	Element	P value
Mg	0.057	Cu	0.891
Ca	0.299	Cr	0.121
K	<u>0.026</u>	Pb	0.027
Na	<u>0.021</u>	Cd	0.835
Fe	0.308	As	0.039
Zn	0.561	Sb	0.799
Mn	0.519	Ni	0.043