

1 Electronic Supplementary Information

2 Manipulating surface structure of quantum dots based on
3 dual response modes triggered by iron ions for visualization
4 of hydrogen sulfide with a wide detection range

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18 **Experimental section**

19 **Materials**

20 Copper(II) chloride dihydrate ($\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$, 99%), indium(III) chloride tetrahydrate
21 ($\text{InCl}_3 \cdot 4\text{H}_2\text{O}$, 99.9%), zinc acetate ($\text{Zn}(\text{Ac})_2$, 99.98%), sodium sulfide nonahydrate
22 ($\text{Na}_2\text{S} \cdot 9\text{H}_2\text{O}$, 96+%), glutathione (GSH, 97%), iron(III) chloride (FeCl_3 , 98%), 1,10-
23 phenanthroline (Phen, 99%), ferrous sulfate (FeSO_4 , 99%), L-cysteine (Cys, 99%),
24 dithiothreitol (DTT, 99%), 3-mercaptopropionic acid (3-MPA, 98%), and sodium
25 nitroprusside (SNP, 98%) were purchased from Energy Chemical. Sodium chloride
26 (NaCl , 99.5+%), sodium fluoride (NaF , 99%), potassium bromide (KBr , 99+%),
27 potassium iodide (KI , 99.5%), sodium acetate (NaAc , 99.5%), sodium carbonate
28 (Na_2CO_3 , 99.8+%), sodium bicarbonate (NaHCO_3 , 99.5%), monopotassium phosphate
29 (KH_2PO_4 , 99.5%), disodium phosphate (Na_2HPO_4 , 99+%), potassium nitrate (KNO_3 ,
30 99%), sodium sulfite (Na_2SO_3 , 98+%), sodium sulfate (Na_2SO_4 , 99+%), potassium
31 thiocyanate (KSCN , 98.5+%), sodium thiosulfate ($\text{Na}_2\text{S}_2\text{O}_3$, 98%), and sodium
32 hydroxide (NaOH , 98%) were purchased from Sinopharm Chemical Reagent Co., Ltd.
33 Trypsin (TRY, EC 3.4.4.4, 250 U mg^{-1}), Dulbecco's modified Eagle's medium
34 (DMEM), fetal calf serum (FBS) and cell counting Kit-8/CCK8 were purchased from
35 Dalian Chenyu Biotechnology. All chemical were used as received without further
36 treatment.

37 **Synthesis of $\text{CuInS}_2/\text{ZnS}$ quantum dots**

38 The synthesis of $\text{CuInS}_2/\text{ZnS}$ quantum dots (CIS/ZnS QDs) refers to An` developed
39 method with slight modification, which can be viewed in our previous research
40 report.¹⁻²

41 **Cell viability assay**

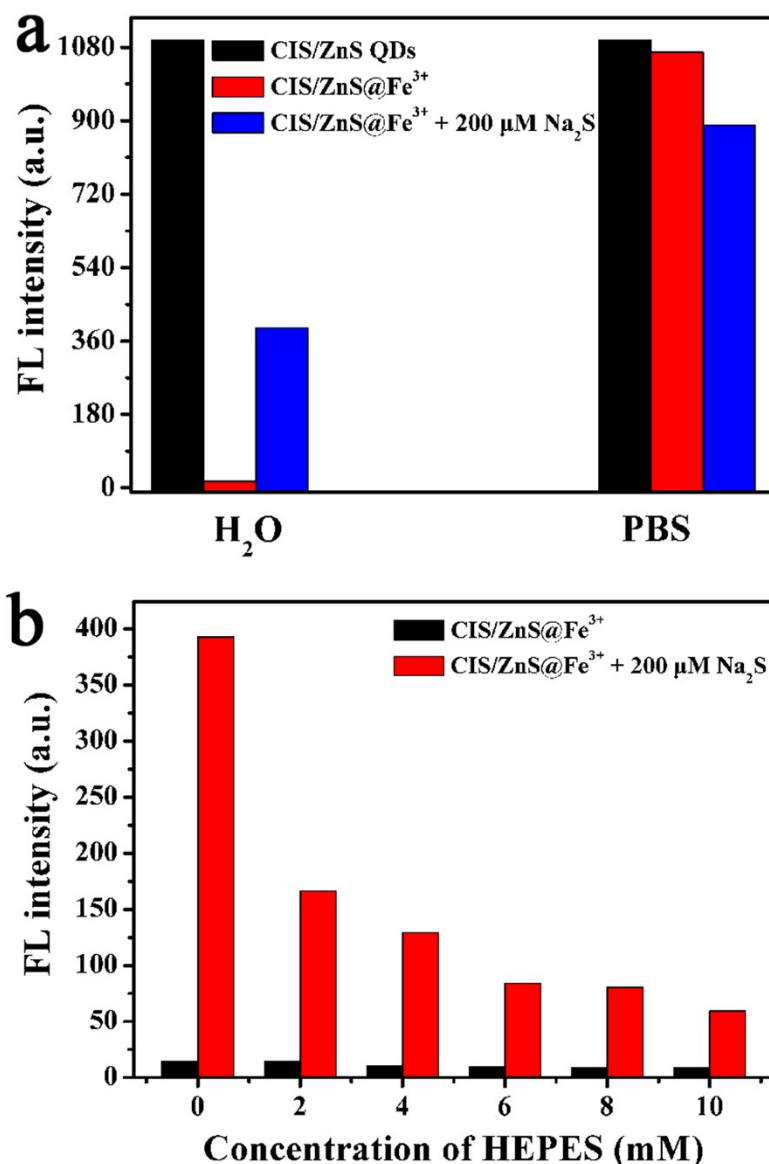
42 MCF-7 cells were provided by the Biological Laboratory of Dalian Medical University.
43 We carried out CCK-8 assay to evaluate the cytotoxicity of the CIS/ZnS QDs against
44 Hela cells. Briefly, cells were seeded at 2.5×10^4 cells/well in 96-well plates and
45 incubated at 37 °C for 24 h. Then, a fresh cultured DMEM medium (100 μ L) containing
46 CIS/ZnS QDs samples with concentrations of 0.07, 0.14, 0.28, 0.35, 0.42 μ g/mL was
47 added to plate and co-incubated with the cells for 12, 24 and 48 h, respectively. After
48 incubation, 100 μ L fresh DMEM solutions containing 10 μ L reagents was added to each
49 well and incubated at 37 °C for 1h. Finally, a microplate reader (Bio-Rad, Model, 550,
50 USA) was used to measure the absorbance at 450 nm. The free-QDs groups and culture
51 medium (without cells and CIS/ZnS QDs) was marked as control and blank
52 respectively, and incubated at the same conditions. The relative cell viability can be
53 calculated with the following equation:

54
$$\text{cell viability (\%)} = \frac{A_s - A_b}{A_c - A_b} \times 100\%$$

55 where A_s , A_c and A_b are the absorbance of the test, control and blank sample.

56 **Characterization**

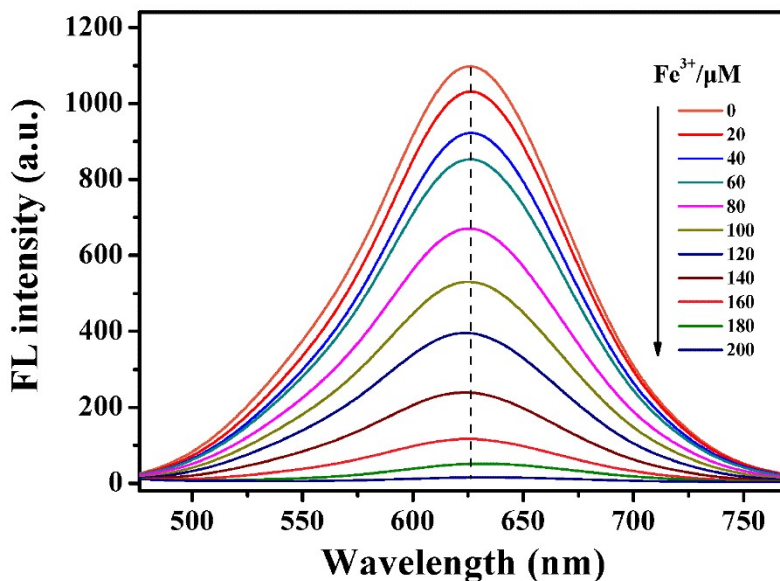
57 UV-vis absorption spectra were measured with a sp-2500 spectrophotometer (Shanghai
58 Spectrum Instruments Co. ltd., China). Photoluminescence (PL) emission spectra were
59 recorded on Hitachi F-4700 Fluorescence Spectrophotometer under the excitation
60 wavelength of 410 nm. Fluorescence images were taken with an Olympus TH4-200
61 fluorescent inverted microscope at an excitation wavelength of 530-550 nm and imaged
62 with the emission wavelength from 575 nm to near infrared region.



63

64 **Fig. S1** (a) The emission peak intensity of CIS/ZnS QDs (black column), CIS/ZnS@Fe-
 65 200 nanoprobe (red column), and CIS/ZnS@Fe-200 nanoprobe upon the addition of
 66 200 μM Na₂S (blue column) in the case of aqueous and PBS buffer (pH = 7.4, 10 mM)
 67 medium, respectively; (b) The emission peak intensity of CIS/ZnS@Fe-200 nanoprobe
 68 before (black column) and after (red column) adding 200 μM Na₂S in different
 69 concentration of HEPES buffers (pH = 7.4).

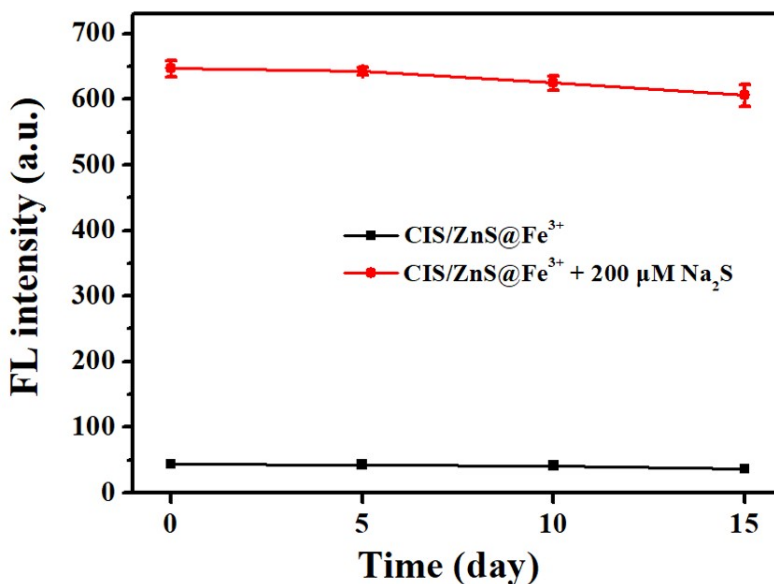
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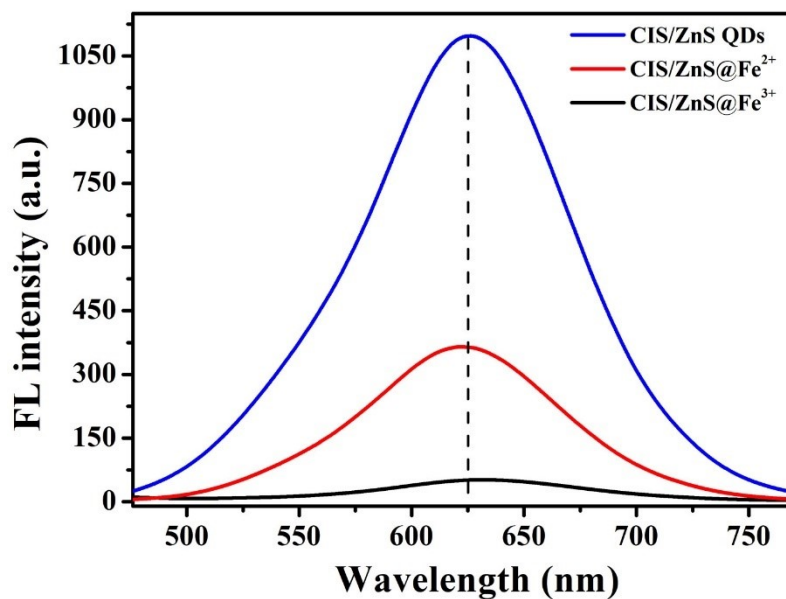
72 **Fig. S2** Fluorescent titration spectra of CIS/ZnS QDs upon gradual addition of Fe³⁺ (0-
 73 200 μM) in the aqueous medium.

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75

76 **Fig. S3** The fluorescent intensity before (black line) and after (red line) adding 200 μM
 77 Na₂S for the CIS/ZnS@Fe-180 nanoprobe, which were prepared by the CIS/ZnS QDs
 78 that storing in a refrigerator at 4 °C for 0, 5, 10 and 15-day respectively.

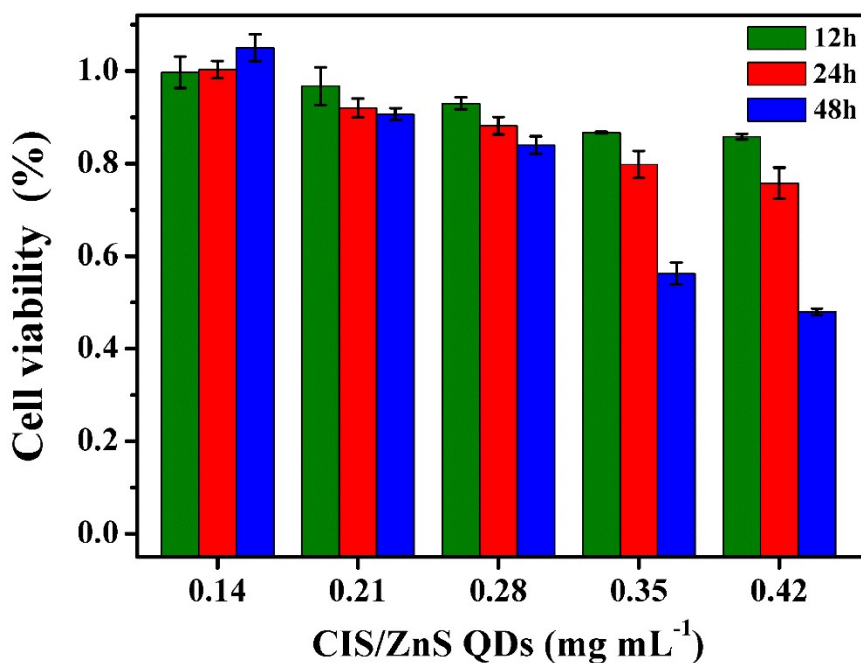


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80 **Fig. S4** The fluorescent spectra of CIS/ZnS QDs (blue line), CIS/ZnS QDs treated by
 81 180 μM Fe^{3+} (CIS/ZnS@ Fe^{3+} , black line) and 180 μM Fe^{2+} (CIS/ZnS@ Fe^{2+} , red line)

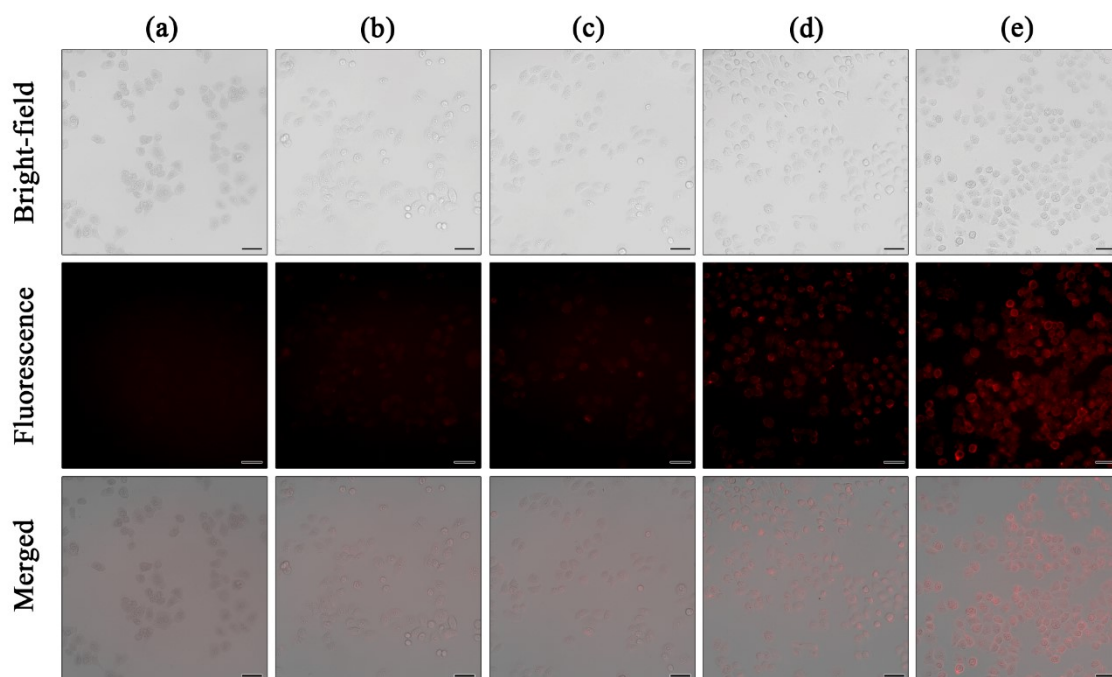
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85 **Fig. S5** The cell viability of CIS/ZnS QDs with concentration of 0.14, 0.21, 0.28, 0.35
 86 and 0.42 mg mL⁻¹ at different incubation time.

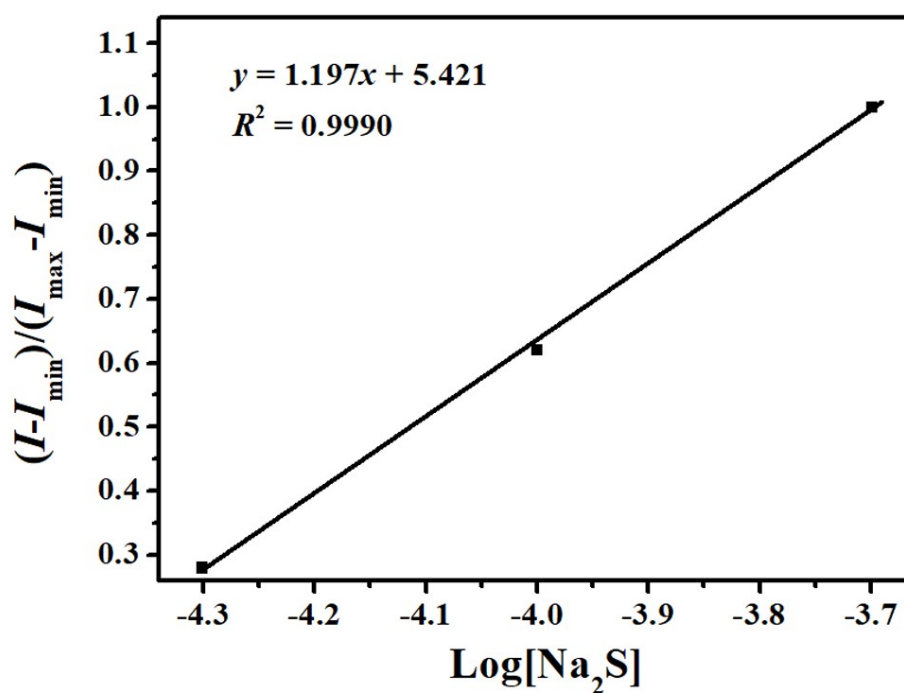


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88 **Fig. S6** Fluorescence images of MCF-7 cells. From left to right: CIS/ZnS@Fe-200

89 labeled cells incubated with exogenous 200 μM Na_2S for (a) 0 min, (b) 5 min, (c) 10min,

90 (d) 20 min and (e) 50 min. All scale bars = 100 μm .



91

92 **Fig. S7** The regression curve of normalized emission peak intensity obtained by using

93 Image J software versus the logarithm of exogenous Na₂S in MCF-7 cells.

94

95 **Table S1** The detection performance of representatively reported fluorescent turn-on
96 H₂S bio-probes

No.	Probe	LOD (μM)	Linear range (μM)	Ref.
1	CLSS-2	4.6 ± 2.0	0-250	[3]
2	Mn-doped ZnS QDs	0.2	2-100	[4]
3	TPP-H ₂ S	0.12	0.5-20	[5]
4	BH-HS	1.7	0-40	[6]
5	Ag-NPs@DNA-FAM	0.01	0.01-100	[7]
6	Flu-N ₃	0.031	0-140	[8]
7	CdTe QDs/AgNP	0.015	0.0495-5.2	[9]
8	Mito-NIR-SH	0.0893	0-30	[10]
9	Lyso-HA-HS	0.34	0-40	[11]
10	SXR	0.70	0-80	[12]
11	QCy7-HS	1.0	0-14	[13]
12	SNARF-SeSPy	0.034	0-20	[14]
13	TPA-Pz-NBD	0.70	0-125	[15]
14	NT-SH	0.080	0-50	[16]
15	CMHS	0.23	0-260	[17]
16	CPs/MOFs	7.2	0-80	[18]
17	NTR-HS	0.104	3.33-20	[19]
18	SS-N ₃	0.010	0-80	[20]
19	RDM-HS	0.31	0-80	[21]
20	ER-Nap-NBD	5.2	0-450	[22]
21	T-HS	3.6	0-100	[23]
22	WFP-PC	0.47	0-20	[24]
23	DCI-Br-NBD	0.40	0-50	[25]
24	CFB-H ₂ S	0.046	0-100	[26]
25	HN8DNP	0.31	0-50	[27]
26	DCM-H ₂ S	0.13	5-50	[28]
27	CIS/ZnS@Fe-160	0.68	0-300	This work
28	CIS/ZnS@Fe-180	0.68	0-300	This work
29	CIS/ZnS@Fe-200	0.70	0-300	This work

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