Electronic Supplementary Information

Enhanced Blue Phosphorescence in Platinum Acetylide Complexes via a Secondary Heavy Metal and Anion-Controlled Aggregation

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General procedures

Materials

Starting materials and reagents were purchased from commercial sources (Sigma-Aldrich, TCI Chemicals, and AmBeed) and used without further purification unless otherwise specified. All other solvents were dried by a commercial solvent purification system and stored over 3 Å molecular sieves. The precursors [Pt(COD)Cl₂] (COD = 1,5-cyclooctadiene) and AgBAr^F₄ (BAr^F₄ = tetrakis[3,5-bis(trifluoromethyl)phenyl]borate) were synthesized according to previous literature.^{1,2}

Physical Methods

¹H, ¹³C{¹H}, ¹¹B{¹H}, ¹⁹F, and ³¹P{¹H} NMR spectra were recorded at room temperature using a JEOL ECA-400, JEOL ECA-500, or ECA-600 NMR spectrometer. Infrared (IR) spectra were obtained on neat powders using a Thermo Nicolet Avatar FT-IR spectrometer with a diamond ATR. UV-vis absorption spectra were recorded in dichloromethane or acetonitrile in screwcapped 1 cm quartz cuvettes using an Agilent Cary 8454 UV-vis spectrophotometer. Steady-state photoluminescence (PL) and excitation spectra were recorded using a Horiba FluoroMax-4 spectrofluorometer. Air-free samples for PL spectra were prepared in a nitrogen-filled glovebox using dry, deoxygenated solvents and housed in 1 cm quartz cuvettes with septum-sealed screw caps. PL quantum yields in solution were measured with respect to a standard of quinine sulfate in 0.05 M sulfuric acid, which has a reported quantum yield (Φ_F) of 0.52.³ The quantum yields of complexes doped into poly(methyl methacrylate) (PMMA) thin films were recorded using a Spectralon-coated integrating sphere (150 mm diameter, Labsphere) exciting at 310 nm. Phosphorescence lifetimes were measured on a Horiba DeltaFlex Lifetime System, using 330 nm excitation. Cyclic voltammograms were recorded using a CH Instruments 602E potentiostat interfaced with a nitrogen-filled glovebox. Samples were dissolved in MeCN with 0.1 M tetrabutylammonium hexafluorophosphate as the supporting electrolyte, and recorded using a glassy carbon working electrode, platinum wire counter electrode, and silver wire pseudoreference electrode. Ferrocene was added at the end of each measurement as an internal standard, and all potentials are referenced to the ferrocenium/ferrocene redox couple.

PMMA Film Fabrication

A solution of PMMA in dichloromethane or acetonitrile was prepared at room temperature inside a nitrogen-filled glovebox. Then, the respective platinum complex was added to the solution and stirred until the solution became clear. The mass concentration was set at 100 mg/mL for 1–5 wt% PMMA films and 50 mg/mL for 10 wt% PMMA films. The solution was then drop-coated onto a quartz substrate and dried at room temperature overnight prior to use.

X-ray Crystallography Details

Single crystals were mounted on a Bruker Apex II three-circle diffractometer using Mo K α radiation ($\lambda = 0.71073$ Å). The data was collected at 123(2) K, then processed and refined within the APEXII software. Structures were solved by intrinsic phasing methods in SHELXT and refined by standard difference Fourier techniques in the program SHELX. All non-hydrogen atoms were

refined with anisotropic displacement parameters. Hydrogen atoms bonded to carbon were placed in calculated positions using the standard riding model and refined isotropically. Crystallographic details are summarized in Tables S1–S2.



Scheme S1. Synthetic routes for new platinum complexes.



Preparation of [Pt(COD)(C=C-2-py)₂]. In the glovebox, 500 mg of [Pt(COD)Cl₂] (1.3 mmol) was dissolved in 10 mL of methanol. In a separate vial, 380 mg of *t*BuOK (3.4 mmol) was dissolved in 10 mL of methanol, followed by the addition of 330 mg of 2-ethynylpyridine (3.2 mmol). Both vials were placed in the freezer at -37 °C for 10 minutes. Then, the solution of 2-enhynylpyridine and *t*BuOK was slowly added to the [Pt(COD)Cl₂] solution. The reaction mixture was stirred at room temperature for 2 h. Finally, the solid was filtered and washed with MeOH and Et₂O. Yield: 570 mg (84%). ¹H NMR (400 MHz, CHLOROFORM-*D*) δ (ppm) 8.48 (d, *J* = 4.9 Hz, 2H, Ar<u>*H*</u>), 7.56 (td, *J* = 7.7, 1.8 Hz, 2H, Ar<u>*H*</u>), 7.40 (d, *J* = 7.9 Hz, 2H, Ar<u>*H*</u>), 7.08 (ddd, *J* = 7.6, 4.9, 1.2 Hz, 2H, Ar<u>*H*</u>), 5.77 (s with ¹⁹⁵Pt satellites, ³*J*Pt-H = 44, 4H, COD), 2.56 (s, 8H, COD). These data matched what is reported in the literature.⁴



Preparation of *cis*-[Pt(CN-*t*Bu)₂(C=C-2-py)₂] (1). In the glovebox, 200 mg of [Pt(COD)(C=C-2-py)₂] (0.40 mmol) was dissolved in 20 mL of CH₂Cl₂. Then, *tert*-butyl isocyanide (79 mg, 0.95 mmol) was added to Pt(COD)(C=C-2-py)₂ solution. The mixture was stirred at room temperature for 1 h. The solvent was removed under vacuum. The product was purified via precipitation from CH₂Cl₂/Et₂O, collected by filtration, and washed with Et₂O. Yield: 178 mg (79%). ¹H NMR (400 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.39 (ddd, *J* = 4.9, 1.9, 1.0 Hz, 2H, Ar<u>H</u>), 7.57 (td, *J* = 7.7, 1.9 Hz, 2H, Ar<u>H</u>), 7.23 (dt, *J* = 7.9, 1.1 Hz, 2H, Ar<u>H</u>), 7.08 (ddd, *J* = 7.6, 4.9, 1.2 Hz, 2H, Ar<u>H</u>), 1.54 (s, 18H, $-C\underline{H_3}$). ¹³C {¹H} NMR (151 MHz, ACETONITRILE-*D*₃) δ (ppm) 149.5 (Ar), 146.1 (Ar), 135.9 (Ar), 126.6 (Ar), 122.1 (t, ¹J¹³C=¹⁴N = 22 Hz, <u>C</u>=N-*t*Bu), 121.0 (Ar), 105.9 (s, ¹⁹⁵Pt satellites, ²J_{Pt-C} = 330 Hz, Pt-C=<u>C</u>-py), 95.1 (s, ¹⁹⁵Pt satellites, ¹J_{Pt-C} = 1178 Hz, Pt-<u>C</u>=C-py), 59.2 [-<u>C</u>(CH₃)₃], 29.1 (-<u>C</u>H₃). *****T-IR (cm⁻¹): 2232 (ṽ_{CN}), 2211 (ṽ_{CN}), 2130 (ṽ_{CC}).



Preparation of *cis*-[Pt(CN-*adamantyl*)₂(C=C-2-py)₂] (2). In the glovebox, 200 mg of [Pt(COD)(C=C-2-py)₂] (0.40 mmol) was dissolved in 20 mL of CH₂Cl₂. Then, 1-adamantyl isocyanide (153 mg, 0.95 mmol) was added to the Pt(COD)(C=C-2-py)₂ solution. The mixture was stirred at room temperature for 1 h. The solvent was removed under vacuum, and the product was purified via precipitation from CH₂Cl₂/pentane, collected by filtration, and washed with pentane. Yield: 213 mg (74%). ¹H NMR (400 MHz, CHLOROFORM-*D*) δ (ppm) 8.43 (d, *J* = 3.8 Hz, 2H, Ar<u>H</u>), 7.48 (td, *J* = 7.7, 1.9 Hz, 2H, Ar<u>H</u>), 7.39 (d, *J* = 7.9 Hz, 2H, Ar<u>H</u>), 6.99 (ddd, *J* = 7.5, 4.9, 1.3 Hz, 2H, Ar<u>H</u>), 2.12 (s, 14H, -adamantly group), 1.90 (s, 4H, -adamantly group), 1.71 – 1.63 (m, 12H, -adamantly group). ¹³C{¹H} NMR (151 MHz, CHLOROFORM-*D*) δ (ppm) 149.1 (Ar), 146.5 (Ar), 135.4 (Ar), 127.3 (Ar), 123.6 (<u>C</u>=N-adamantyl), 120.6 (Ar), 105.8 (Pt-C=<u>C</u>-py), 95.1 (Pt-<u>C</u>=C-py), 58.3 (-adamantly group), 42.7 (-adamantly group), 35.3 (-adamantly group), 28.8 (-adamantly group). *****T-IR (cm⁻¹): 2228 (ṽcN), 2206 (ṽcN), 2131 (ṽcc).



Preparation of complex [1-Cu]PF₆. In the glovebox, 50 mg of complex 1 (0.088 mmol) was dissolved in 10 mL of CH₂Cl₂. A solution of [Cu(CH₃CN)₄]PF₆ (41 mg, 0.13 mmol) in 5 mL of CH₂Cl₂ was added to the solution of complex 1. The mixture was stirred overnight at RT. The solid was filtered, washed with CH₂Cl₂, and dried under vacuum. Yield: 52 mg (76%). ¹H NMR (500 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.63 (d, *J* = 5.4 Hz, 2H, Ar<u>*H*</u>), 7.76 (t, *J* = 7.8 Hz, 2H, Ar<u>*H*</u>), 7.36 – 7.32 (m, 4H, Ar<u>*H*</u>), 1.50 (s, 18H, –C<u>*H*₃). ¹⁹F NMR (470 MHz, ACETONITRILE-*D*₃) δ (ppm) –72.8 (d, *J* = 707 Hz, 6F, PF₆). ³¹P{¹H} NMR (202 MHz, ACETONITRILE-*D*₃) δ (ppm) –144.0 (sept, *J* = 707 Hz, 1P, PF₆). **#**T-IR (cm⁻¹): 2225 (\tilde{v}_{CN}), 2135 (\tilde{v}_{CC}).</u>



Preparation of complex [1-Ag]BF₄. In the glovebox, 50 mg of complex **1** (0.088 mmol) was dissolved in 10 mL of CH₂Cl₂. A solution of AgBF₄ (26 mg, 0.13 mmol) or [Ag(CH₃CN)₄]BF₄ (47 mg, 0.13 mmol) in 5 mL of CH₂Cl₂ was added to the solution of complex **1**. A solid formed after adding AgBF₄. The mixture was stirred for 1 h at RT. The solid was filtered and washed with CH₂Cl₂. The product was dried under vacuum. Yield: 55 mg (82%). ¹H NMR (400 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.52 (d, *J* = 5.3 Hz, 2H, Ar<u>*H*</u>), 7.78 (t, *J* = 7.8 Hz, 2H, Ar<u>*H*</u>), 7.37 (t, *J* = 6.6 Hz, 2H, Ar<u>*H*</u>), 7.29 (d, *J* = 7.9 Hz, 2H, Ar<u>*H*</u>), 1.57 (s, 18H, -C<u>*H*₃). ¹⁹F NMR (376 MHz, ACETONITRILE-*D*₃) δ (ppm) –151.6. ¹¹B{¹H} NMR (128 MHz, ACETONITRILE-*D*₃) δ (ppm) –2.2. **F**T-IR (cm⁻¹): 2240 ($\tilde{\nu}$ CN), 2225 ($\tilde{\nu}$ CN), 2109 ($\tilde{\nu}$ CC).</u>



Preparation of complex [1-Ag]PF₆. In the glovebox, 50 mg of complex 1 (0.088 mmol) was dissolved in 10 mL of CH₂Cl₂. A solution of AgPF₆ (33 mg, 0.13 mmol) in 5 mL of CH₂Cl₂ was added to the solution of complex 1. A solid precipitated after adding AgPF₆. The mixture was stirred for 1 h at RT. The solid was filtered and washed with CH₂Cl₂. The product was dried under

vacuum. Yield: 60 mg (83%). ¹H NMR (400 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.52 (ddd, *J* = 5.3, 1.8, 1.0 Hz, 2H, Ar<u>*H*</u>), 7.37 (ddd, *J* = 7.6, 5.3, 1.3 Hz, 2H, Ar<u>*H*</u>), 7.32 (d, *J* = 7.9 Hz, 2H, Ar<u>*H*</u>), 1.57 (s, 18H, $-C\underline{H}_3$). ¹⁹F NMR (470 MHz, ACETONITRILE-*D*₃) δ (ppm) -72.8 (d, *J* = 707 Hz, 6F, PF₆). ³¹P{¹H} NMR (202 MHz, ACETONITRILE-*D*₃) δ (ppm) -144.0 (sept, *J* = 707 Hz, 1P, PF₆). *****T-IR (cm⁻¹): 2240 (\tilde{v} CN), 2225 (\tilde{v} CN), 2109 (\tilde{v} CC).



Preparation of complex [1-Ag]SbF₆. In the glovebox, 25 mg of complex 1 (0.044 mmol) was dissolved in 5 mL of CH₂Cl₂. A solution of AgSbF₆ (23 mg, 0.066 mmol) in 5 mL of CH₂Cl₂ was added to the solution of complex 1, forming a solid. The mixture was stirred for 1 h at RT. The solid was filtered and washed with CH₂Cl₂. The product was dried under vacuum. Yield: 34 mg (86%). ¹H NMR (500 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.52 (ddd, *J* = 5.3, 1.8, 0.9 Hz, 2H, Ar<u>*H*</u>), 7.81 (td, *J* = 7.8, 1.7 Hz, 2H, Ar<u>*H*</u>), 7.38 (ddd, *J* = 7.7, 5.3, 1.3 Hz, 2H, Ar<u>*H*</u>), 7.35 (dt, *J* = 7.9, 1.1 Hz, 2H, Ar<u>*H*</u>), 1.57 (s, 18H, $-CH_3$). ¹⁹F NMR (470 MHz, ACETONITRILE-*D*₃) δ (ppm) -196.3. FT-IR (cm⁻¹): 2256 (\tilde{v}_{CN}), 2244 (\tilde{v}_{CN}), 2070 (\tilde{v}_{CC}).



Preparation of complex [1-Ag]BAr^F₄. In the glovebox, 25 mg of complex 1 (0.044 mmol) was dissolved in 5 mL of CH₂Cl₂. A solution of AgBAr^F₄ (50 mg, 0.052 mmol) in 5 mL of DCM was added to the solution of complex 1. The mixture was stirred for 1 h at RT. Then, the solvent was removed under vacuum. Et₂O was added to the crude product, and the mixture was stirred for 10 min. The white solid product was collected and washed with Et₂O. Finally, the product was dried under vacuum overnight. Yield: 51 mg (75%). ¹H NMR (400 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.52 (d, *J* = 5.2 Hz, 2H, Ar<u>H</u>), 7.67 – 7.64 (m, 12H, Ar<u>H</u>), 7.39 – 7.34 (m, 4H, Ar<u>H</u>), 1.57 (s, 18H, –C<u>H₃</u>). ¹⁹F NMR (470 MHz, ACETONITRILE-*D*₃) δ (ppm) –63.1. ¹¹B{¹H} NMR (160 MHz, ACETONITRILE-*D*₃) δ (ppm) –7.7.**F**T-IR (cm⁻¹): 2238 (\tilde{v}_{CN}), 2219 (\tilde{v}_{CN}), 2142 (\tilde{v}_{CC}).



Preparation of complex [2-Ag]BAr^F₄. In the glovebox, 25 mg of complex **2** (0.035 mmol) was dissolved in 5 mL of CH₂Cl₂. A solution of AgBAr^F₄ (41 mg, 0.042 mmol) in 5 mL of CH₂Cl₂ was added to the solution of complex **2**. The solution was stirred for 1 h at RT. Then, the solvent was removed under vacuum. The product was purified via precipitation from CH₂Cl₂/pentane. The product was collected by filtration and washed with pentane. Yield: 40 mg (67%). ¹H NMR (400 MHz, ACETONITRILE-*D*₃) δ (ppm) 8.50 (d, *J* = 5.0 Hz, 2H, Ar<u>*H*</u>), 7.77 (td, *J* = 7.8, 1.7 Hz, 2H, Ar<u>*H*</u>), 7.68 – 7.64 (m, 12H, Ar<u>*H*</u>), 7.35 (ddd, *J* = 7.7, 5.3, 1.3 Hz, 2H, Ar<u>*H*</u>), 7.29 (dt, *J* = 7.9, 1.1 Hz, 2H, Ar<u>*H*</u>), 2.15 – 2.13 (m, 18H, –adamantly group), 1.75 – 1.71 (m, 12H, –adamantly group). ¹⁹F NMR (376 MHz, ACETONITRILE-*D*₃) δ (ppm) –7.7.**F**T-IR (cm⁻¹): 2234 (\tilde{v}_{CN}), 2218 (\tilde{v}_{CN}), 2139 (\tilde{v}_{CC}).

X-ray Crystallography Summary

	1	2
CCDC	2394176	2394177
Chemical formula	C24H27N4Pt	C36H38N4Pt
Mr	566.58	721.79
Crystal system, space group	Orthorhombic, <i>P</i> ca2 ₁	Triclinic, <i>P</i> 1
Temperature (K)	123	123
<i>a</i> , <i>b</i> , <i>c</i> (Å)	26.4912(17), 11.0501(7), 22.1450(15)	11.975(3), 15.698(4), 19.951(5)
α, β, γ (°)	90, 90, 90	79.386(3), 84.090(4), 71.728(3)
$V(Å^3)$	6482.5(7)	3496.4(16)
Ζ	8	4
$\mu (mm^{-1})$	4.34	4.04
Crystal size (mm)	$0.22\times0.09\times0.04$	0.16 imes 0.08 imes 0.03
T_{\min}, T_{\max}	0.567, 0.746	0.597, 0.746
No. of measured, independent and observed $[I > 2\sigma(I)]$ reflections	37826, 14783, 12168	43231, 13257, 9388
R _{int}	0.044	0.058
$(\sin \theta / \lambda)_{max} (\text{\AA}^{-1})$	0.651	0.610
$R[F^2 > 2\sigma(F^2)], wR(F^2),$ S	0.033, 0.066, 1.02	0.036, 0.079, 0.98
No. of reflections	14783	13257
No. of parameters	536	821
No. of restraints	915	2178
$\Delta \rho_{\text{max}}, \Delta \rho_{\text{min}} (e \text{ Å}^{-3})$	1.16, -0.80	0.73, -0.95

 Table S1. Summary of crystallographic data for complexes 1 and 2.

	[1-Cu]PF ₆ ·CH ₂ Cl ₂ ·C ₄ H ₁₀ O	[1-Ag]PF ₆ ·2(CH ₂ Cl ₂)
CCDC	2394178	2394179
Chemical formula	C48H52Cu2N8Pt2·2(PF6)·CH2Cl2· C4H10O	$C_{48}H_{52}Ag_2N_8Pt_2\cdot 2(PF_6)\cdot 2(CH_2Cl_2)$
Mr	1707.22	1806.68
Crystal system, space group	Triclinic, <i>P</i> 1	Monoclinic, <i>P</i> 2 ₁ / <i>n</i>
Temperature (K)	123	123
<i>a</i> , <i>b</i> , <i>c</i> (Å)	10.1312(18), 13.882(3), 25.808(5)	11.611(5), 13.011(6), 21.235(9)
α, β, γ (°)	89.634(2), 78.885(2), 75.512(2)	90, 101.158(5), 90
$V(Å^3)$	3445.0(11)	3147(2)
Ζ	2	2
μ (mm ⁻¹)	4.85	5.34
Crystal size (mm)	0.28 imes 0.18 imes 0.05	$0.21\times0.09\times0.08$
T_{\min}, T_{\max}	0.520, 0.746	0.562, 0.746
No. of measured, independent and observed $[I > 2\sigma(I)]$ reflections	50989, 13087, 10157	15284, 5558, 4517
R _{int}	0.062	0.051
$(\sin \theta / \lambda)_{max} (\text{\AA}^{-1})$	0.610	0.595
$\frac{R[F^2 > 2\sigma(F^2)]}{S}, wR(F^2),$	0.037, 0.091, 1.06	0.048, 0.099, 1.06
No. of reflections	13087	5558
No. of parameters	753	367
No. of restraints	700	0
$\Delta ho_{ m max}, \Delta ho_{ m min} (e { m \AA}^{-3})$	1.25, -1.72	1.94, -1.94

Table S2. Summary of crystallographic data for complexes [1-Cu]PF₆ and [1-Ag]PF₆.



Fig. S1 1 H NMR spectra of complex 1 in CD₃CN with different concentrations.



Fig. S2 ¹H NMR spectra of complex [1-Ag]PF₆ in CD₃CN with different concentrations.



Fig. S3 ¹H NMR spectra of complex **1** in CD₃CN with different amounts of AgPF₆. The solutions of complex **1** (5 mg, 0.009 mmol in 1 mL of CD₃CN) and AgPF₆ (8 mg, 0.032 mmol in 1 mL of CD₃CN) were prepared. The NMR tube was filled with 1 mL of complex **1** solution. The ¹H NMR spectra were recorded in the absence of AgPF₆ and after adding different amounts of AgPF₆ stock solution. mol **1** = mol AgPF₆ when the added volume of AgPF₆ solution = 0.28 mL.



Fig. S4 ¹H NMR spectra of the mixture complex 1 and AgPF₆ in CD₃CN with different temperatures. The solutions of complex 1 (5 mg, 0.009 mmol in 1 mL of CD₃CN) and AgPF₆ (8 mg, 0.032 mmol in 1 mL of CD₃CN) were prepared. The NMR tube was filled with 1 mL of complex 1 solution and 0.1 mL of AgPF₆ solution. The ¹H NMR spectra were recorded over the temperature range of 25 °C to -35 °C.



Fig. S5. Magnification of the structure of **[1-Cu]PF**₆, determined from single-cyrstal X-ray diffraction, zooming in on the secondary interactions involving the Cu(I) ions. Carbon atoms are displayed translucently for clarity.



Fig. S6. Magnification of the structure of $[1-Ag]PF_6$, determined from single-cyrstal X-ray diffraction, zooming in on the secondary interactions involving the Ag(I) ions. Carbon atoms and a noncoordinated nitrogen atom are displayed translucently for clarity.

Photophysical Measurements

Complex	$\Phi_{ ext{PL}}$	τ/μs	$k_{\rm r} \times 10^{-4} / {\rm s}^{-1}$	$k_{\rm nr} \times 10^{-4}/{\rm s}^{-1}$
1	0.0009	0.14	0.64	714
[1-Cu]PF6 ^b	-	-	-	-
[1-Ag]BF ₄	0.003	0.51	0.59	195
[1-Ag]PF ₆	0.005	0.31	1.6	321
[1-Ag]SbF ₆	0.004	0.35	1.1	285
[1-Ag]BAr ^F 4	0.002	0.39	0.51	256
2 ^b	-	-	-	-
[2-Ag]BArF ₄	0.004	0.35	1.1	284

Table S3. Summary of photoluminescence data in solution^a

^{*a*} Recorded in CH₂Cl₂ (complexes 1, 2, [1-Ag]BArF, and [2-Ag]BArF) or CH₃CN (complexes [1-Ag]BF₄, [1-Ag]PF₆, and [1-Ag]SbF₆). ^{*b*} No photoluminescence in solution.



Fig. S7 Photoluminescence spectra of complexes [1-Ag]X in solution.



Fig. S8 UV–vis absorption (a) and photoluminescence spectra (b) of complex 1 in CH₃CN with different amounts of AgPF₆. The stock solutions of complex 1 (1.3 mg, 0.0023 mmol in 1 mL of CH₃CN) and AgPF₆ (1.2 mg, 0.0047 mmol in 1.5 mL of CH₃CN) were prepared inside the glovebox. The cuvette was filled with 3 mL of CH₃CN and 200 μ L of complex 1 stock solution. The absorption and PL spectra were recorded in the absence of AgPF₆ and after adding different amounts of AgPF₆ stock solution. mol 1 = mol AgPF₆ when the volume of AgPF₆ solution = 147 μ L.



Fig. S9 PL spectra of new platinum complexes in PMMA films with different concentrations. Samples were excited at $\lambda = 310$ nm.

Complex	Φ_{PL}			
Complex	1 wt%	2 wt%	5 wt%	10 wt%
1	0.034	0.014	0.017	0.006
[1-Ag]BF ₄	0.045	< 0.01	< 0.01	< 0.01
[1-Ag]PF ₆	0.060	0.048	< 0.01	< 0.01
[1-Ag]SbF ₆	0.071	0.016	< 0.01	< 0.01
[1-Ag]BAr ^F 4	0.16	0.14	0.05	0.03
2	0.032	0.024	0.024	0.017
[2-Ag]BAr ^F 4	0.14	0.12	0.074	0.027

Table S4. Summary of photoluminescence quantum yields in PMMA films with different concentrations.

Cyclic voltammograms



Fig. S10 Cyclic voltammogram of complex **1**, recorded in MeCN with 0.1 M (NBu₄)(PF₆) supporting electrolyte. The arrow shows the scan direction.



Fig. S11 Cyclic voltammogram of complex **2**, recorded in MeCN with 0.1 M (NBu₄)(PF₆) supporting electrolyte. The arrow shows the scan direction.



Fig. S12 Cyclic voltammogram of complex $[1-Cu]PF_6$, recorded in MeCN with 0.1 M (NBu₄)(PF₆) supporting electrolyte. The arrow shows the scan direction.



Fig. S13 Cyclic voltammogram of complex $[1-Ag]BAr^{F_4}$, recorded in MeCN with 0.1 M (NBu₄)(PF₆) supporting electrolyte. The arrow shows the scan direction.



Fig. S14 Cyclic voltammogram of complex $[2-Ag]BAr^{F_4}$, recorded in MeCN with 0.1 M (NBu₄)(PF₆) supporting electrolyte. The arrow shows the scan direction.



Fig. S15 The overlaid cyclic voltammograms of complexes 1, 2, $[1-Cu]PF_6$, $[1-Ag]BAr^{F_4}$, and $[2-Ag]BAr^{F_4}$.

Table S5. Summary of cyclic voltammetry data.

Complex	$E^{ m ox}$ (Pt complex) (V vs Fc ⁺ /Fc) ^a	$\frac{E^{\mathrm{ox}}(\mathrm{M}^{2+}\!/\mathrm{M}^{+})}{(\mathrm{V}\mathrm{vs}\mathrm{Fc}^{+}\!/\mathrm{Fc})^{b}}$	E^{red} (Pt complex) (V vs Fc ⁺ /Fc) ^a
1	$N.D.^{c}$	-	-2.31
2	N.D. ^c	-	-2.36
[1-Cu]PF ₆	+0.95	-1.02	-2.30
[1-Ag]BAr ^F 4	+1.37	-0.54	-2.30
[2-Ag]BAr ^F ₄	+1.33	-0.54	-2.34

^a Redox waves are irreversible. The half-peak potential is reported.⁷ ^b Redox waves are reversible. ^c No clear oxidation was observed within the solvent's electrochemical window.

FT-IR spectra



Fig. S16 FT-IR spectrum of complex 1, recorded as a neat powder.



Fig. S17 FT-IR spectrum of complex 2, recorded as a neat powder.



Fig. S18 FT-IR spectrum of complex [1-Cu]PF₆, recorded as a neat powder.



Fig. S19 FT-IR spectrum of complex [1-Ag]BF₄, recorded as a neat powder.



Fig. S20 FT-IR spectrum of complex [1-Ag]PF₆, recorded as a neat powder.



Fig. S21 FT-IR spectrum of complex [1-Ag]SbF₆, recorded as a neat powder.



Fig. S22 FT-IR spectrum of complex [1-Ag]BAr^F₄, recorded as a neat powder.



Fig. S23 FT-IR spectrum of complex [2-Ag]BAr^F₄, recorded as a neat powder.

NMR spectra



Fig. S24 ¹H NMR spectrum of [Pt(COD)(C=C-2-py)₂], recorded in CDCl₃ at 400 MHz.



Fig. S25 ¹H NMR spectrum of complex 1, recorded in CD₃CN at 400 MHz.



Fig. S27 ¹H NMR spectrum of complex 2, recorded in CDCl₃ at 400 MHz.



Fig. S29 ¹H NMR spectrum of complex [1-Cu]PF₆, recorded in CD₃CN at 500 MHz.



Fig. S31 ³¹P{¹H} NMR spectrum of complex [1-Cu]PF₆, recorded in CD₃CN at 202 MHz.









----2.2



Fig. S35 ¹H NMR spectrum of complex [1-Ag]PF₆, recorded in CD₃CN at 400 MHz.



~ -71.9 ~ -73.8

Fig. S37 ³¹P{¹H} NMR spectrum of complex [1-Ag]PF₆, recorded in CD₃CN at 202 MHz.



Fig. S39¹⁹F NMR spectrum of complex [1-Ag]SbF₆, recorded in CD₃CN at 470 MHz.





Fig. S41 ¹⁹F NMR spectrum of complex [1-Ag]BAr^F₄, recorded in CD₃CN at 470 MHz



--7.7

Fig. S42 ¹¹B{¹H} NMR spectrum of complex [**1-Ag**]**BAr**^F₄, recorded in CD₃CN at 160 MHz.





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