

## Supplementary Information

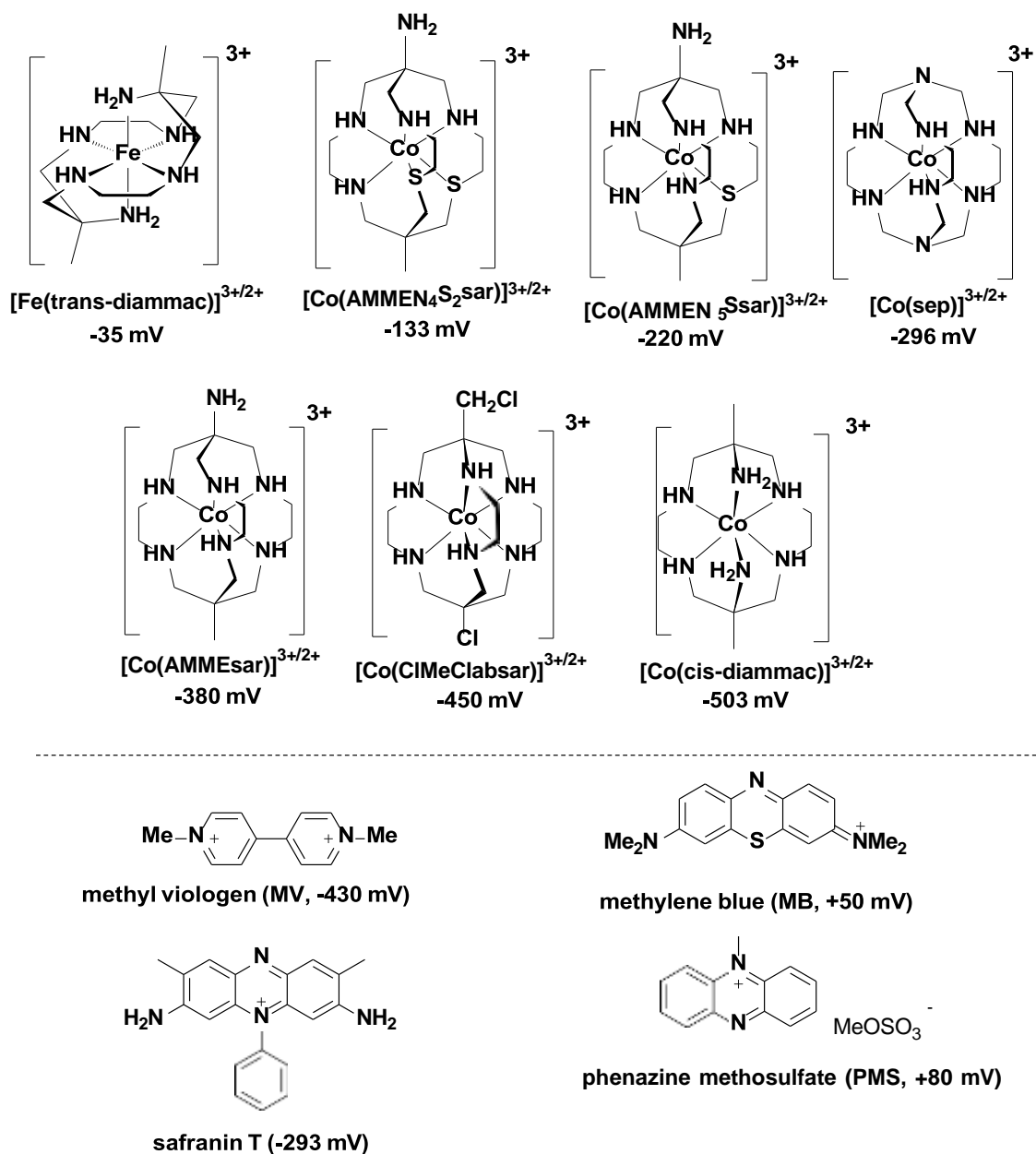
### Reversible Enzyme-Catalysed $\text{NAD}^+/\text{NADH}$ Electrochemistry

Peter D. Giang,<sup>a</sup> Dimitri Niks,<sup>b</sup> Sheron Hakopian,<sup>b</sup> Russ Hille,<sup>b</sup> Paul V. Bernhardt<sup>a\*</sup>

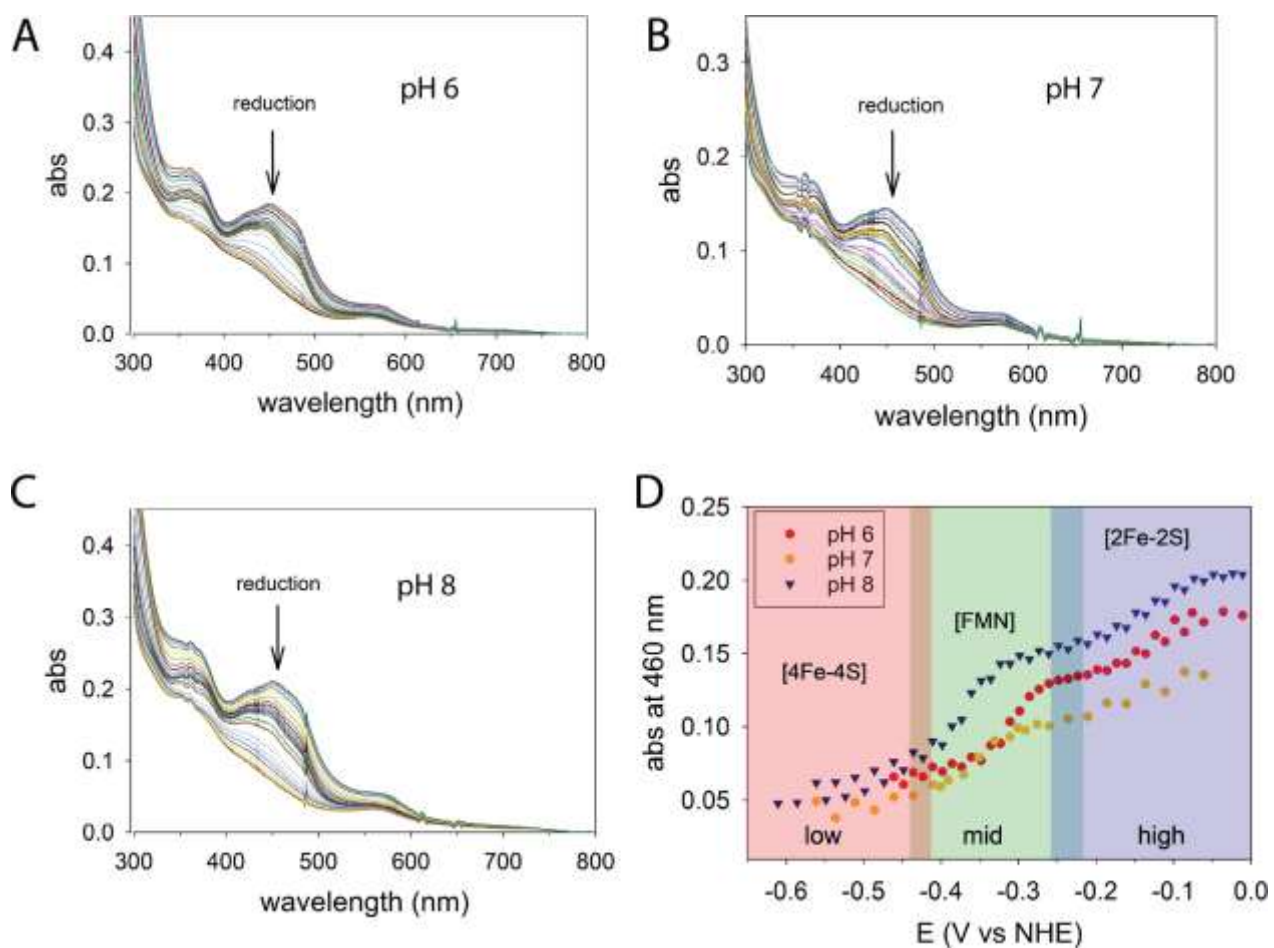
<sup>a</sup> School of Chemistry and Molecular Biosciences, University of Queensland, Brisbane 4072, Australia

<sup>b</sup> Department of Biochemistry, University of California, Riverside, USA

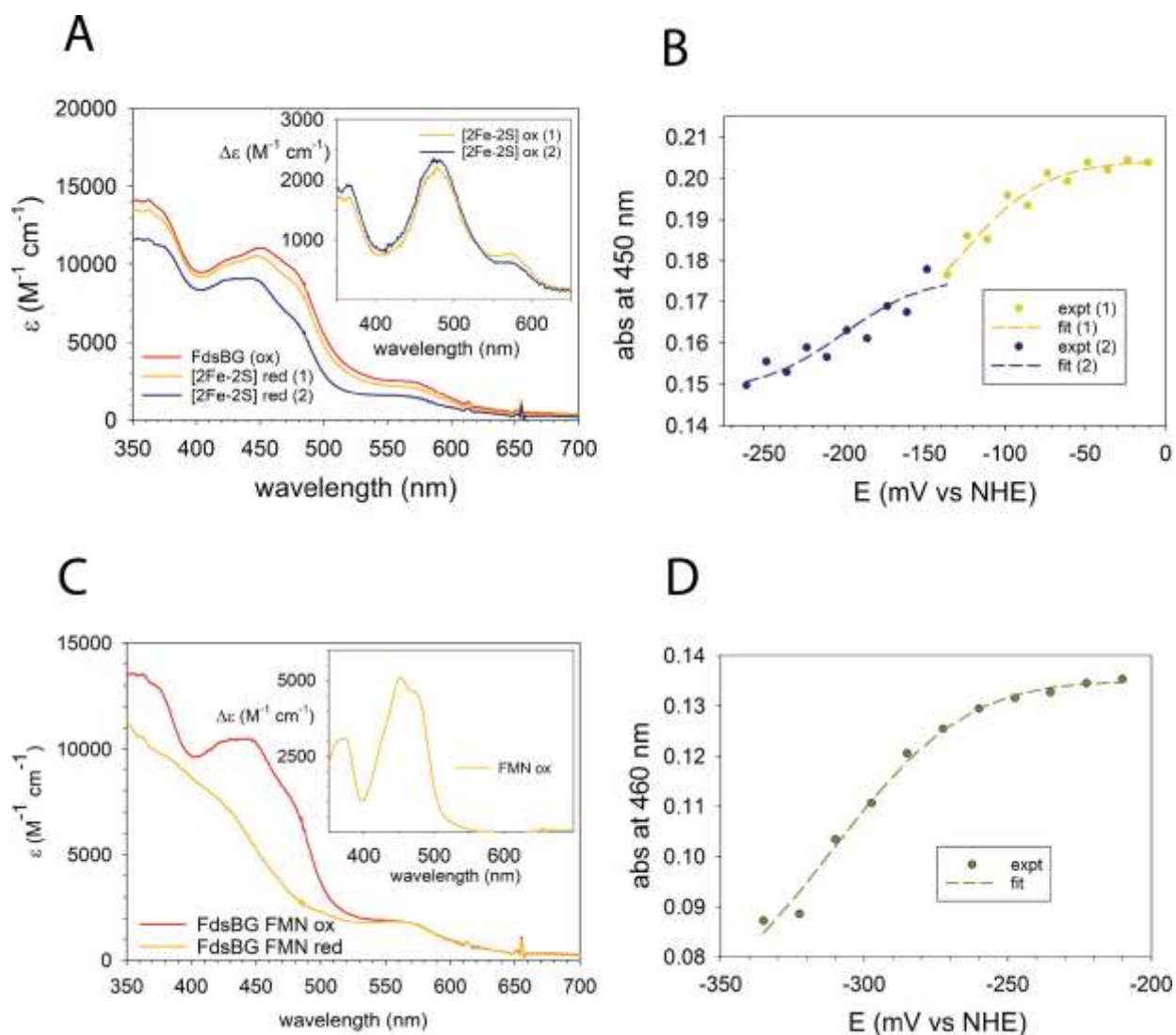
\*p.bernhardt@uq.edu.au



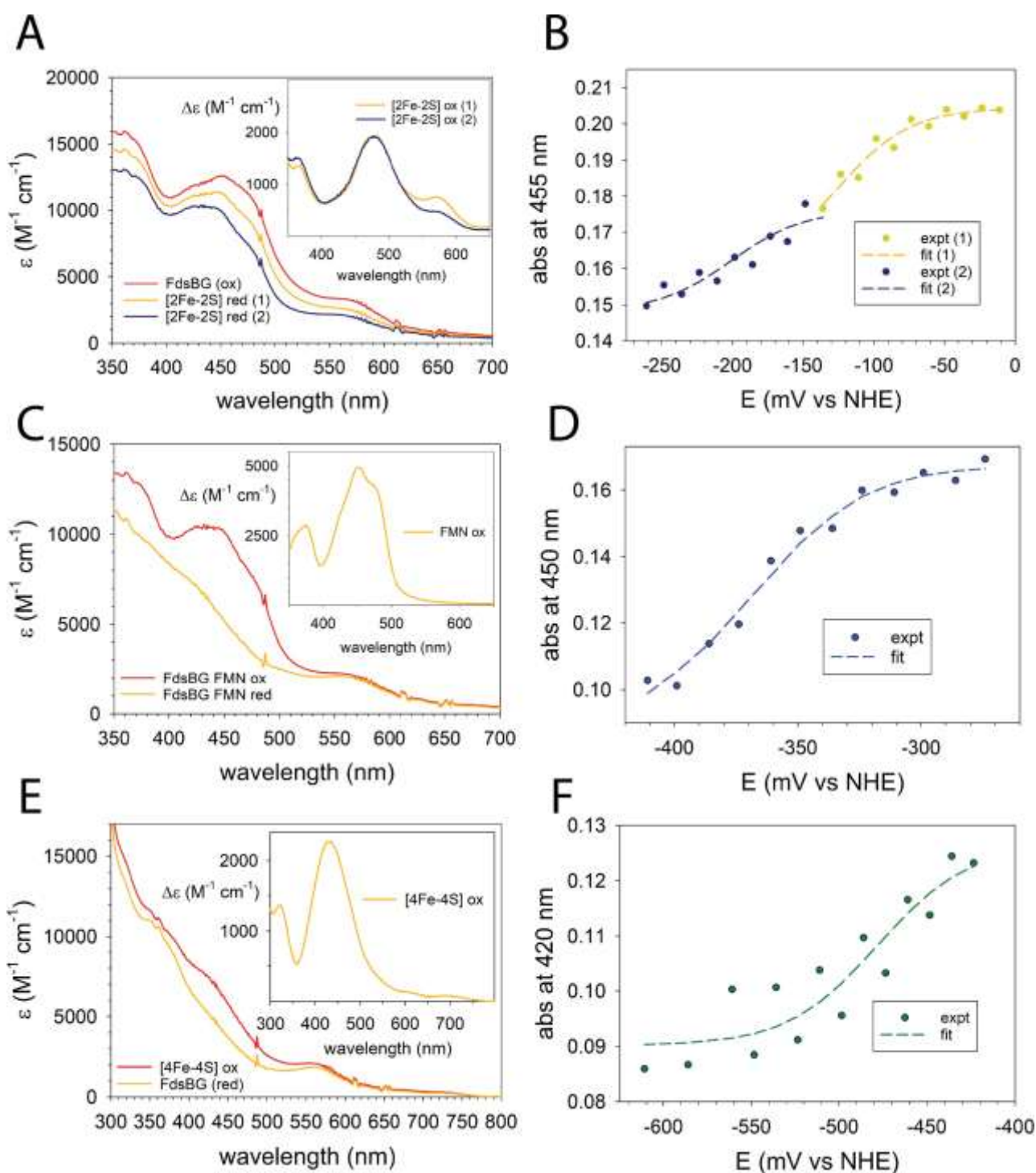
**Figure S1.** Chemical structures of all mediators used in this work and their approximate redox potentials vs NHE at pH 7.



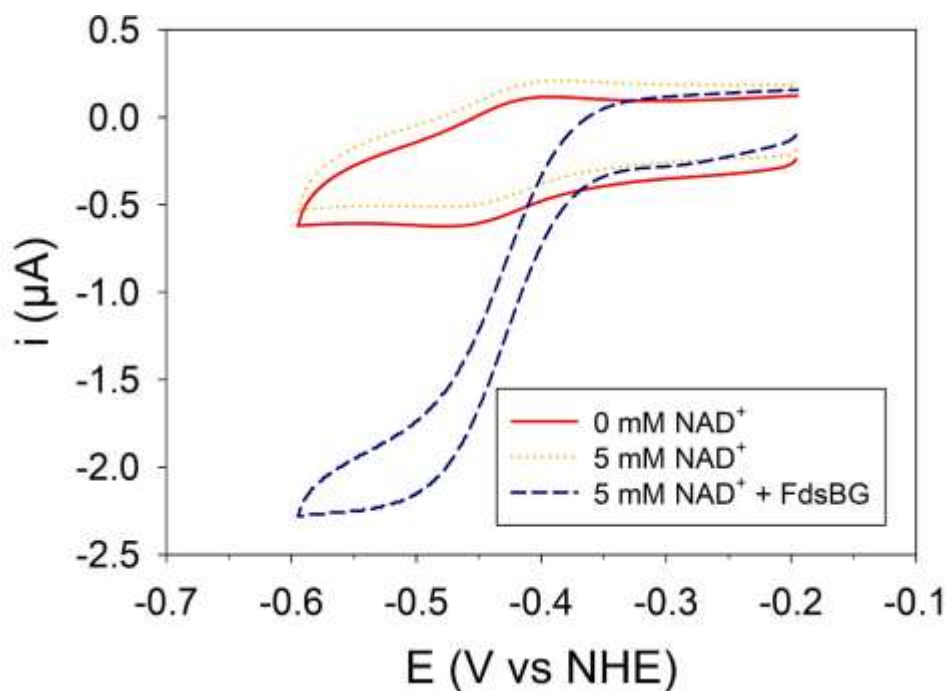
**Figure S2.** Optical spectroelectrochemistry data for FdsBG (100  $\mu\text{M}$ ) at (A) pH 6 ( $-0.45 < E < 0$  V vs NHE); (B) pH 7 ( $-0.55 < E < -0.05$  V vs NHE) and (C) pH 8 ( $-0.55 < E < 0$  V vs NHE); (D) absorbance changes at 460 nm as a function of potential and pH indicating the three data regions that were fit independently.



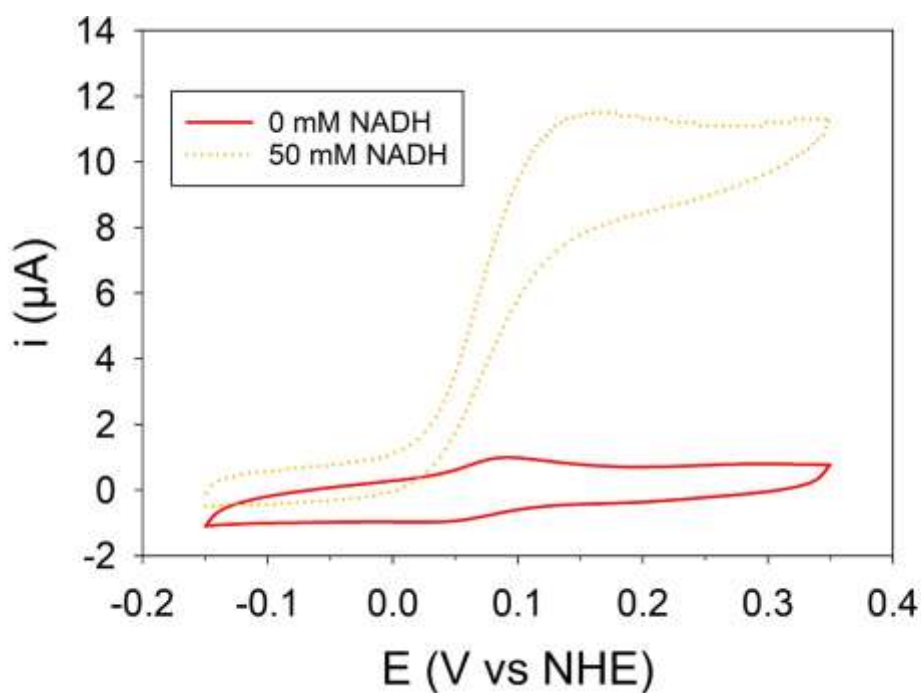
**Figure S3.** Reactlab Redox spectral analysis for FdsBG (100  $\mu$ M, pH 6) showing the (A) high potential region  $-210 < E < 0$  mV vs NHE (difference spectra shown in the inset); (B) calculated and experimental absorbances at 450 nm as a function of potential ( $E_1$   $-120$ ,  $E_2$   $-150$  mV); (C) mid potential region  $-335 < E < -210$  mV (difference spectrum shown in the inset); (D) calculated and experimental absorbances at 460 nm as a function of potential ( $E_1$   $-240$ ,  $E_2$   $-310$  mV). Difference spectra are shown in the insets of panels A and C.



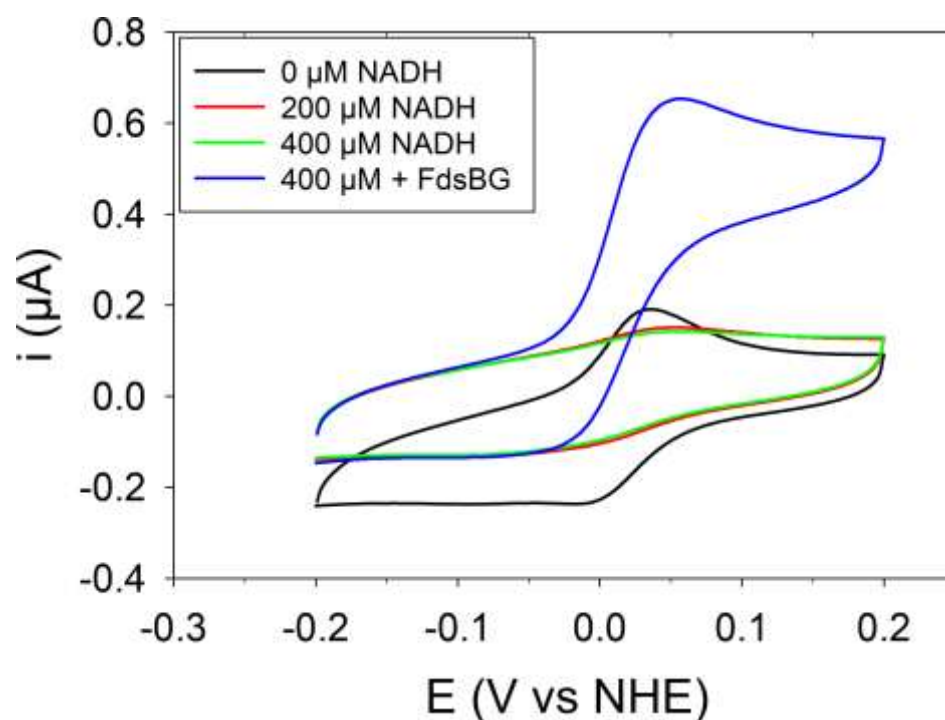
**Figure S4.** Reactlab Redox spectral analysis for FdsBG (100  $\mu$ M, pH 8) showing the (A) high potential region  $-260 < E < -10$  mV vs NHE; (B) calculated and experimental absorbances at 455 nm as a function of potential ( $E_1$  -130,  $E_2$  -200 mV); (C) mid-potential range  $-410 < E < -260$  mV; (D) calculated and experimental absorbances at 450 nm as a function of potential ( $E_1$  -320,  $E_2$  -370 mV); (E) low potential range  $-610 < E < -420$  mV; (F) calculated and experimental absorbances at 420 nm as a function of potential ( $E_1$  -480 mV). Difference spectra are shown in the insets of panels A, C and E.



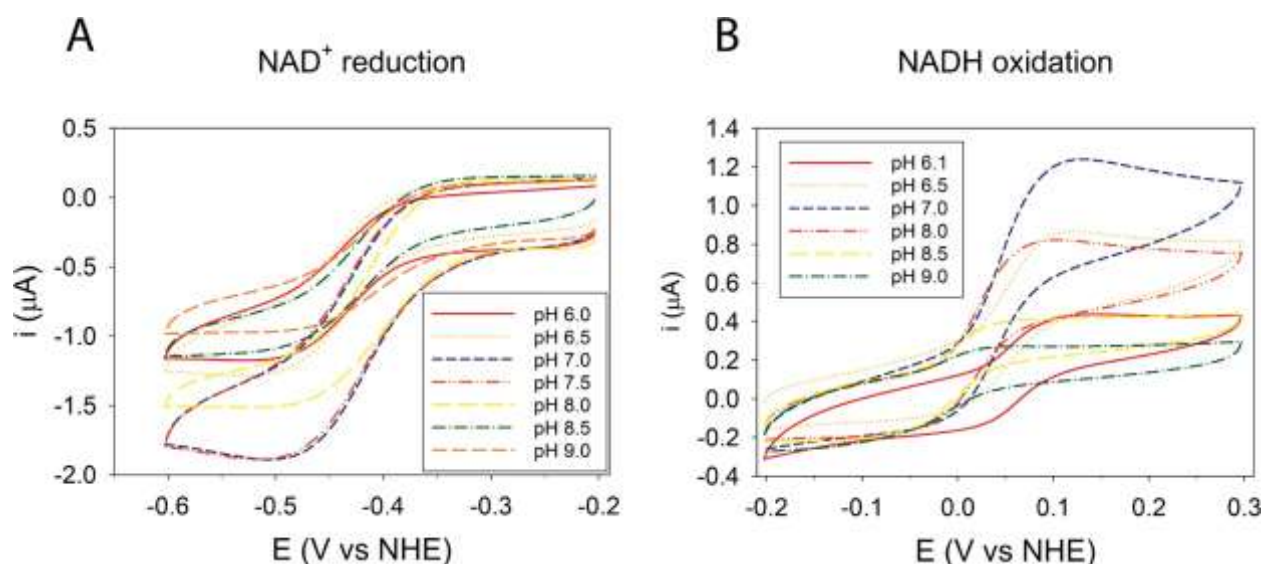
**Figure S5.** Cyclic voltammetry of methyl viologen (60  $\mu\text{M}$ ) (red curve); methyl viologen (60  $\mu\text{M}$ ) plus  $\text{NAD}^+$  (5 mM) (yellow curve) and methyl viologen (60  $\mu\text{M}$ ) plus  $\text{NAD}^+$  (5 mM) and FdsBG (0.34  $\mu\text{M}$ ) (blue curve). Scan rate 5  $\text{mV s}^{-1}$ , 50 mM phosphate buffer, pH 7.5.



**Figure S6.** Cyclic voltammetry of phenazine methosulfate (40  $\mu\text{M}$ ) (red curve) and phenazine methosulfate (40  $\mu\text{M}$ ) plus NADH (50 mM) (yellow curve). Scan rate 5  $\text{mV s}^{-1}$ , 50 mM phosphate buffer, pH 7.5.



**Figure S7.** Cyclic voltammetry of methylene blue (20  $\mu\text{M}$ ) (black curve); methylene blue (20  $\mu\text{M}$ ) plus NADH (200  $\mu\text{M}$ ) (red curve); methylene blue (20  $\mu\text{M}$ ) plus NADH (400  $\mu\text{M}$ ) (green curve); methylene blue (20  $\mu\text{M}$ ) plus NADH (400  $\mu\text{M}$ ) plus FdsBG (0.34  $\mu\text{M}$ , blue curve). Scan rate 5  $\text{mV s}^{-1}$ , 50 mM phosphate buffer, pH 7.5.



**Figure S8.** pH-dependent CVs of (A) methyl viologen (60  $\mu\text{M}$ ),  $\text{NAD}^+$  (5 mM) and FdsBG (0.34  $\mu\text{M}$ ) and (B) methylene blue (20  $\mu\text{M}$ ), NADH (500  $\mu\text{M}$ ) and FdsBG (0.34  $\mu\text{M}$ ) and as a function of pH. See experimental section for buffer compositions (each 50 mM). Scan rate 5  $\text{mV s}^{-1}$ .

**Table S1.** All Electrochemical simulation parameters

Diffusion coefficients	Electrochemical parameters	
MV <sup>2+</sup> /MV <sup>+</sup> $3.8 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$	$k_{0,MV} 1.0 \times 10^{-2} \text{ cm s}^{-1}$ ( $n = 1, \alpha = 0.5$ )	
MB <sup>+</sup> /MBH $2.1 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$	$k_{0,MB} 1.0 \times 10^{-2} \text{ cm s}^{-1}$ ( $n = 2, \alpha = 0.5$ )	
NAD <sup>+</sup> /NADH $4.0 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$	Double layer capacitance $2.0 \times 10^{-5} \text{ F}$	
FdsBG(all forms) $5.9 \times 10^{-7} \text{ cm}^2 \text{ s}^{-1}$	Electrode surface area (A) $0.056 \text{ cm}^2$	
Mediator dependent rate constants and potentials		
MV <sup>2+</sup> + e <sup>-</sup> ⇌ MV <sup>+</sup>	$E^{\circ}$ (pH 7.5) $-0.43 \text{ V vs NHE}$	
MB <sup>+</sup> + 2e <sup>-</sup> ⇌ MBH	$E^{\circ}$ (pH 7.5) $+0.06 \text{ V vs NHE}$	
MV <sup>+</sup> + FdsBG <sub>ox</sub> ⇌ MV <sup>2+</sup> + FdsBG <sub>int</sub>	$k_3 3.5 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$	$k_4 3.5 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$
MV <sup>+</sup> + FdsBG <sub>int</sub> ⇌ MV <sup>2+</sup> + FdsBG <sub>red</sub>	$k_3^a 3.5 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$	$k_4^b 3.5 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$
MB <sup>+</sup> + FdsBG <sub>red</sub> ⇌ MBH + FdsBG <sub>ox</sub>	$k_5 1.6 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$	$k_6 1.0 \times 10^3 \text{ M}^{-1} \text{ s}^{-1}$
Mediator independent rate constants		
FdsBG <sub>red</sub> + NAD <sup>+</sup> ⇌ FdsBG <sub>ox</sub> + NADH	$k_{red} 2.0 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$	$k_{ox} 2.0 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$

<sup>a</sup>  $k_3$  assumed to be equal for reduction of FdsBG<sub>ox</sub> and FdsBG<sub>int</sub>; <sup>b</sup>  $k_4$  assumed to equal for oxidation of FdsBG<sub>int</sub> and FdsBG<sub>red</sub>.