

Supporting Information

Palladium/Norbornenes-Catalyzed C–H Bond Activation and Annulation to Construct Polycyclic Aromatic Hydrocarbons-based Fluorescent Materials

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1. General Information

Unless otherwise noted, commercial available reagents were purchased from commercial suppliers (such as Energy Chemical, Bide Pharmatech Ltd. and Adamas) and used as received. Solvents were generally dried over 4Å molecular sieves. Purification of products was performed by flash column chromatography using silica gel. ^1H and ^{13}C NMR spectra were recorded on a Agilent 400MR DD2 spectrometer (400 MHz and 101 MHz, respectively). Chemical shifts are reported parts per million (ppm) referenced to CDCl_3 (δ 7.26 ppm), tetramethylsilane (TMS, δ 0.00 ppm) for ^1H NMR; CDCl_3 (δ 77.16 ppm) for ^{13}C NMR. The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublet, td = triplet of doublet and m = multiplet. To distinguish, some ^{13}C NMR chemical shifts retain two decimal places. High-resolution mass spectra (HRMS) were obtained on an Agilent 6546 Q-TOF LC/MS system with an ESI source at the School of Pharmaceutical Sciences of Chongqing University. UV-vis spectra were recorded on Agilent Cary60 spectrometer. Fluorescence spectra were collected on HITACHI F-700003040428 Fluorescence Spectrometer or Agilent Technologies Cary Eclipse Fluorescence Spectrometer. The absolute quantum yields were taken using Edinburgh Instruments FLS1000 fluorescence spectrometer with a calibrated integrating sphere system. The excited-state lifetimes were performed using Edinburgh Instruments FLS1000.

2. DFT Calculation

All calculations were performed using the Gaussian 16, Revision A.03.^[S1] Geometries were optimized in toluene with the SMD solvation model by the B3-LYP-D3 functional and a basis set of 6-31G(d). Vibrational frequencies were calculated at the same level and check the optimized structure. Solvation energies were computed through a self-consistent reaction field using the SMD model. Calculations were performed at the M06/SDD-6-31+G(d,p)-SMD(toluene) level of

theory.

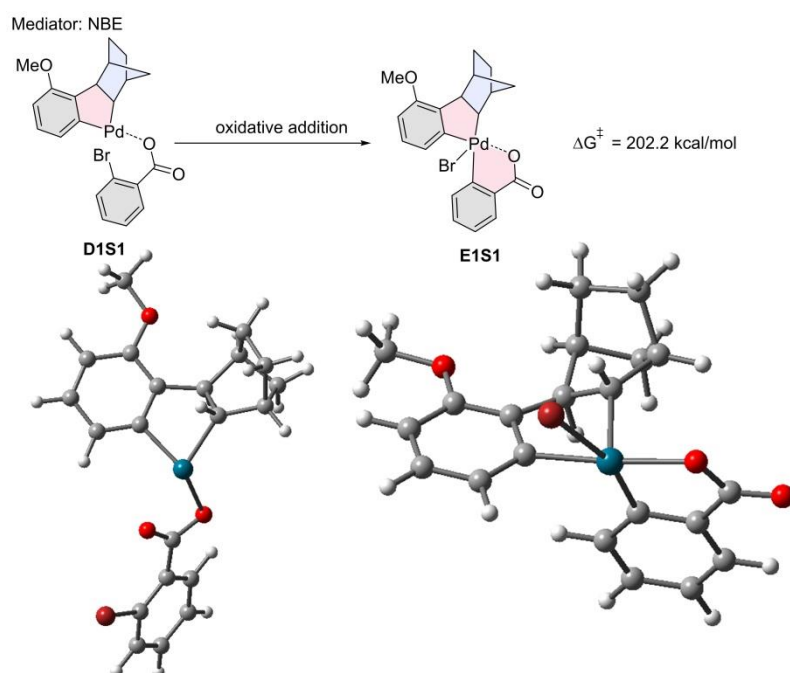


Fig. S1. DFT calculates the free-energy barrier of the reaction of the rate-determining step using norbornene (NBE) as a mediator.

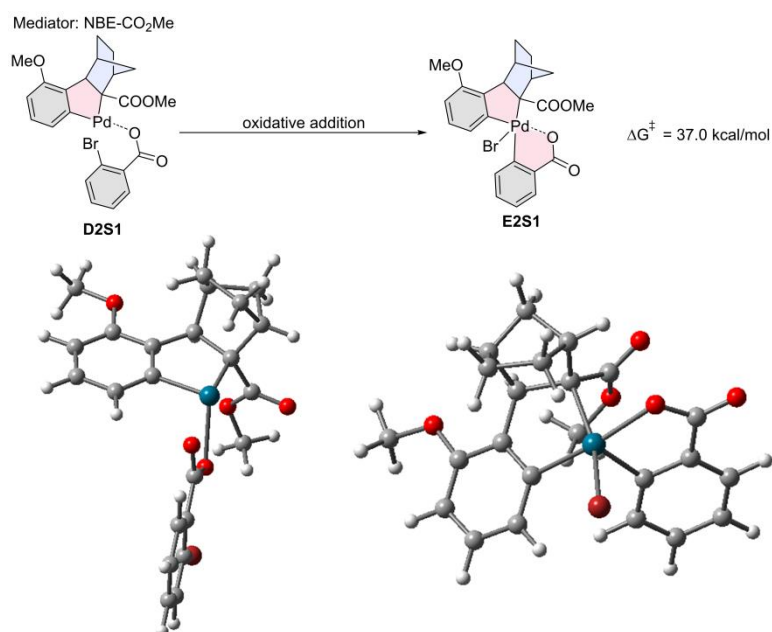


Fig. S2. DFT calculates the free-energy barrier of the reaction of the rate-determining step using methyl bicyclo[2.2.1]hept-2-ene-2-carboxylate (NBE-CO₂Me) as a mediator.

Cartesian coordinates of the structures

D1S1

C	-2.00716	-1.35636	-0.32571
C	-2.62379	3.31508	1.17352
C	-1.44274	3.57769	0.45362
C	-0.78516	2.55658	-0.25536
C	-1.3515	1.27305	-0.23643
C	-2.52727	0.97071	0.46851
C	-3.16388	2.01622	1.18084
C	-4.26955	-0.09583	-1.79398
C	-4.36042	-0.71481	-0.37102
C	-3.0324	-0.44225	0.42772
C	-3.22945	-1.01968	-2.52092
C	-2.83758	-2.05852	-1.42299
C	-4.20128	-2.2309	-0.67786
Pd	-0.46398	-0.24275	-1.22047
C	5.80371	0.58778	0.25377
C	6.3287	1.33408	-0.81154
C	5.49278	1.7191	-1.87301
C	4.1441	1.35166	-1.85307
C	3.58989	0.59818	-0.78825
C	4.44786	0.22077	0.26925
Br	3.88601	-0.8049	1.81801
C	2.13653	0.27648	-0.89713
O	1.5118	-0.42133	0.00047
O	1.46732	0.71723	-1.92283
O	-4.32008	1.658	1.86568
C	-5.02381	2.64445	2.66166
C	-1.14855	-2.26722	0.47886
O	-0.59014	-3.28981	0.03156
O	-0.98005	-1.85739	1.77622
C	-0.00882	-2.58372	2.59904
H	-3.10802	4.12514	1.72141
H	-1.03041	4.59153	0.44866
H	0.13457	2.76111	-0.80917
H	-3.95512	0.96144	-1.7699
H	-5.25475	-0.13325	-2.29298
H	-5.25731	-0.41761	0.19181
H	-3.18025	-0.80996	1.45888
H	-3.67358	-1.53589	-3.39151
H	-2.35469	-0.45733	-2.90028
H	-2.36452	-2.97384	-1.80549
H	-4.99541	-2.63423	-1.3326
H	-4.12848	-2.86048	0.22702
H	6.44959	0.28661	1.08199
H	7.38694	1.61061	-0.80635
H	5.88987	2.30095	-2.70906
H	3.47111	1.63703	-2.66385

H	-5.87751	2.11178	3.10652
H	-4.37785	3.04563	3.4655
H	-5.39403	3.47696	2.03365
H	-0.07017	-2.11984	3.59367
H	-0.26983	-3.65424	2.64583
H	1.00048	-2.46321	2.17172
E1S1			
C	0.60361	1.62205	-0.00889
C	3.48631	-2.36522	0.80719
C	2.38377	-3.16964	0.46346
C	1.17873	-2.59879	0.02566
C	1.0719	-1.19731	-0.07192
C	2.17037	-0.37298	0.25384
C	3.37544	-0.97004	0.70856
C	2.38703	1.13015	-2.39376
C	2.79602	1.81789	-1.06553
C	2.06851	1.12735	0.1535
C	0.88558	1.53913	-2.54396
C	0.60569	2.45328	-1.29897
C	2.00396	3.15085	-1.14219
Pd	-0.59934	-0.13198	-0.63962
C	-1.60308	-1.48091	1.92641
C	-2.57248	-1.60902	2.94384
C	-3.72629	-0.80482	2.94145
C	-3.93008	0.13051	1.9148
C	-2.97841	0.27221	0.89065
C	-1.81784	-0.51882	0.93768
Br	-1.4884	-2.0978	-2.03715
C	-3.17227	1.21657	-0.26042
O	-2.1743	1.18151	-1.14107
O	-4.18487	1.93994	-0.37743
O	4.38637	-0.08247	1.04782
C	5.65905	-0.58078	1.53792
C	-0.12069	2.27689	1.15194
O	-0.82756	3.27517	0.95114
O	-0.00058	1.85672	2.44496
C	0.81624	0.78412	3.00694
H	4.41278	-2.83284	1.14485
H	2.46791	-4.25817	0.54055
H	0.33888	-3.24361	-0.23699
H	2.99597	1.51954	-3.22872
H	2.53184	0.03692	-2.36651
H	3.8807	1.89145	-0.90506
H	2.5145	1.56997	1.06391
H	0.2063	0.66473	-2.63031
H	0.69442	2.12375	-3.46051
H	-0.27463	3.09478	-1.39812
H	2.25141	3.76804	-2.02447
H	2.08253	3.77537	-0.23482

H	-0.71996	-2.12124	1.93069
H	-2.41787	-2.35063	3.73362
H	-4.47239	-0.91882	3.73301
H	-4.82445	0.7597	1.87173
H	6.26806	0.31355	1.74015
H	5.53091	-1.15817	2.47345
H	6.16455	-1.20602	0.77785
H	0.35185	0.55982	3.97921
H	0.81245	-0.11562	2.3773
H	1.8489	1.13891	3.16932

D2S2

C	2.48081	-1.28104	0.65082
C	2.142	3.34026	-0.2073
C	0.94322	3.27496	0.49893
C	0.38519	2.04918	0.86589
C	1.08443	0.91196	0.50708
C	2.26263	0.94621	-0.22676
C	2.80714	2.17248	-0.58641
C	5.13023	-0.79989	0.10779
C	4.11563	-0.91813	-1.06012
C	2.72982	-0.42708	-0.598
C	4.75448	-1.99407	1.05818
C	3.41186	-2.50478	0.45114
C	3.78835	-2.44234	-1.05736
Pd	0.47828	-0.95681	1.01287
C	-5.36954	0.04562	-1.22582
C	-5.38847	-0.58213	-2.46163
C	-4.25903	-1.25886	-2.90984
C	-3.13227	-1.29576	-2.10991
C	-3.10488	-0.6712	-0.85892
C	-4.23824	0.00658	-0.42251
Br	-4.36262	0.93044	1.24632
C	-1.83234	-0.79909	-0.10721
O	-0.83895	-1.40762	-0.69211
O	-1.66822	-0.35285	1.07866
O	3.98831	2.12572	-1.30167
C	4.59442	3.38028	-1.69928
H	2.80343	-0.73151	1.54812
H	2.54416	4.30833	-0.47033
H	0.43302	4.19363	0.76139
H	-0.56393	1.97988	1.37985
H	5.03555	0.17277	0.59551
H	6.15519	-0.9028	-0.25684
H	4.45105	-0.48471	-1.99898
H	2.02567	-0.76711	-1.37403
H	5.51064	-2.78367	1.00429
H	4.64559	-1.68334	2.10008
H	3.07436	-3.46627	0.83579
H	2.96172	-2.69515	-1.72566

H	4.65552	-3.06859	-1.28172
H	-6.2388	0.57867	-0.868
H	-6.28247	-0.53917	-3.07052
H	-4.25994	-1.75025	-3.87373
H	-2.23337	-1.80702	-2.42236
H	5.49738	3.10349	-2.24138
H	3.93369	3.95685	-2.35554
H	4.86043	3.99253	-0.83079
E2S2			
C	0.06398	0.40018	1.20071
C	-3.74184	0.00752	-1.68353
C	-3.14362	-1.12643	-2.21022
C	-1.85103	-1.48933	-1.83168
C	-1.14377	-0.68022	-0.91686
C	-1.76417	0.47488	-0.39466
C	-3.04449	0.82885	-0.77884
C	-1.58029	1.87853	2.76817
C	-1.05833	2.46479	1.43001
C	-0.82109	1.32984	0.39648
C	-0.30759	1.21225	3.40637
C	0.74143	1.29098	2.24932
C	0.44619	2.73358	1.75276
Pd	0.78811	-0.99752	-0.14916
C	1.35702	1.21011	-2.17676
C	2.25308	2.1125	-2.75451
C	3.56002	2.22059	-2.28181
C	3.98704	1.41407	-1.23414
C	3.10443	0.50994	-0.66463
C	1.79958	0.42194	-1.12559
Br	-0.93973	-2.93402	0.62152
C	3.5287	-0.39743	0.44622
O	2.55794	-1.25744	0.80134
O	4.64384	-0.3582	0.9534
O	-3.54174	1.99852	-0.25507
C	-4.87827	2.42454	-0.63205
H	-0.48836	-0.40932	1.68043
H	-4.74873	0.26186	-1.98295
H	-3.69209	-1.74117	-2.91259
H	-1.39402	-2.3778	-2.24597
H	-2.38159	1.15733	2.59044
H	-1.96779	2.671	3.4125
H	-1.64015	3.29099	1.03111
H	-0.18857	1.80855	-0.36042
H	0.05337	1.79763	4.25649
H	-0.48563	0.18667	3.73691
H	1.75861	1.03355	2.53179
H	1.02514	3.01807	0.87124
H	0.5903	3.46844	2.54774
H	0.34225	1.13239	-2.54714

H	1.92453	2.73197	-3.58032
H	4.24018	2.92947	-2.73657
H	4.9945	1.45324	-0.83633
H	-5.03838	3.35965	-0.09848
H	-4.9554	2.59799	-1.71019
H	-5.63277	1.69316	-0.32564

The time-dependent density functional theory (TDDFT) calculations of the excitation energies were calculated at the optimized geometries of the ground states. Calculations were performed at the B3LYP-6-31+G(d) (**6u**) and MPWLPW91-6-31+G(d) (**6w**) level of theory.

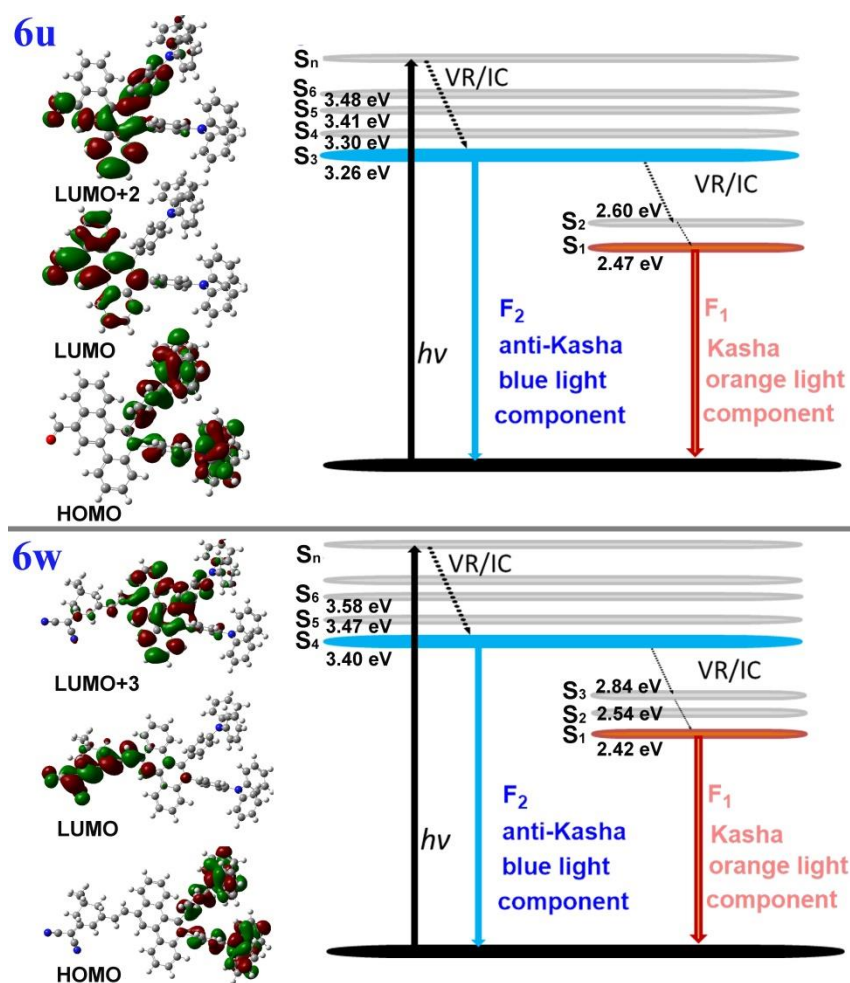


Fig. S3. Molecular orbitals of the S_0 , S_1 , S_3 , or S_4 states of **6u** and **6w**; Jablonski diagram illustrating the anti-Kasha dual-emission mechanism of **6u** and **6w**.

Cartesian coordinates of the structures

6u

C	-6.74854	-0.45937	0.38766
C	-6.06282	0.74942	-0.00307
C	-6.09785	-1.66765	0.29141
C	-4.72465	-1.78524	-0.0533
C	-3.95104	-0.60275	-0.23434
C	-4.66898	0.67204	-0.35517
C	-2.50155	-0.72984	-0.28322
C	-4.11452	-3.09744	-0.16862
C	-4.87573	-4.29631	-0.13281
C	-4.27338	-5.53575	-0.25993
C	-2.87512	-5.63039	-0.42192
C	-2.10752	-4.4812	-0.4634
C	-2.70239	-3.19379	-0.35011
C	-1.90697	-1.98796	-0.34839
C	-1.59638	0.45695	-0.11741
C	-0.41248	-2.13113	-0.35468
C	-4.07113	1.84667	-0.89147
C	-4.76469	3.03921	-1.00534
C	-6.10599	3.12397	-0.58704
C	-6.74103	1.9932	-0.10847
C	-1.57773	1.16175	1.10019
C	-0.70918	2.2337	1.30415
C	0.18438	2.63185	0.29221
C	0.17122	1.93359	-0.93
C	-0.70677	0.8674	-1.12682
C	0.28551	-2.42114	-1.54085
C	1.67444	-2.55842	-1.54864
C	2.41345	-2.39877	-0.36175
C	1.7207	-2.11991	0.83133
C	0.33091	-1.99833	0.83123
N	1.08884	3.71395	0.50534
C	1.35171	4.64878	-0.54328
C	2.66226	5.10345	-0.78253
C	2.91249	6.02465	-1.80268
C	1.8692	6.49454	-2.60992
C	0.56572	6.0379	-2.37877
C	0.30327	5.12975	-1.35014
C	1.75306	3.84969	1.76527
C	2.27722	2.71959	2.42028
C	2.91889	2.85701	3.65378
C	3.06335	4.11846	4.24444
C	2.55005	5.24497	3.58988
C	1.89219	5.11545	2.36412
N	3.83531	-2.51319	-0.37077
C	4.50961	-3.17672	0.70101
C	5.69958	-2.64577	1.2333
C	6.35957	-3.30311	2.27458

C	5.83932	-4.48645	2.81305
C	4.65128	-5.01223	2.29052
C	3.99338	-4.37113	1.23793
C	4.58914	-1.94198	-1.444
C	5.67474	-2.64136	-2.00323
C	6.41452	-2.07563	-3.04473
C	6.07646	-0.81638	-3.55556
C	4.9916	-0.12227	-3.00571
C	4.25615	-0.67258	-1.95291
C	-8.14139	-0.4775	0.87018
O	-8.77053	-1.52159	1.1327
H	-6.67609	-2.54595	0.54836
H	-5.95193	-4.25237	-0.02595
H	-4.87992	-6.43528	-0.23738
H	-2.40339	-6.60323	-0.51489
H	-1.03428	-4.55382	-0.58313
H	-3.05527	1.81147	-1.24815
H	-4.27188	3.90711	-1.43147
H	-6.64741	4.06103	-0.66699
H	-7.78837	2.05811	0.15863
H	-2.26106	0.87344	1.89276
H	-0.71682	2.76496	2.24895
H	0.85308	2.22633	-1.72029
H	-0.69342	0.33886	-2.07372
H	-0.26647	-2.54626	-2.46775
H	2.19204	-2.78553	-2.47359
H	2.27506	-1.99736	1.75478
H	-0.18351	-1.7747	1.75997
H	3.47544	4.73291	-0.16885
H	3.92873	6.3658	-1.97428
H	2.06833	7.20523	-3.40513
H	-0.25384	6.40009	-2.99178
H	-0.70983	4.79142	-1.16424
H	2.17981	1.74277	1.95981
H	3.31887	1.97659	4.14711
H	3.56775	4.22191	5.19943
H	2.64927	6.22783	4.04002
H	1.48406	5.98814	1.86691
H	6.09938	-1.72278	0.82901
H	7.27642	-2.88089	2.67416
H	6.35127	-4.99068	3.62591
H	4.24044	-5.93278	2.69342
H	3.08194	-4.79034	0.82697
H	5.93147	-3.62224	-1.61942
H	7.2491	-2.62736	-3.46613
H	6.64921	-0.38292	-4.36871
H	4.72373	0.85824	-3.38701
H	3.42564	-0.1253	-1.52103
H	-8.63759	0.49209	1.02219

6w

C	-11.32954	-1.65787	-0.19484
C	-9.02175	-0.83752	-0.10565
C	-10.4112	-0.79448	0.30746
C	-10.77766	0.22029	1.36132
C	-9.98681	1.53624	1.21941
C	-8.47945	1.20879	1.21012
C	-8.10616	0.06015	0.30123
C	-6.71677	-0.1045	-0.13436
C	-5.68308	0.6538	0.23614
C	2.47195	-2.03257	0.80061
C	3.82193	-2.33078	0.89196
C	4.55434	-2.64839	-0.24852
C	3.9013	-2.66531	-1.47802
C	2.55545	-2.34851	-1.56235
C	1.91514	1.0101	-1.27217
C	2.88645	1.9653	-1.02236
C	2.91794	2.63966	0.19537
C	1.95319	2.33577	1.1511
C	0.99244	1.37117	0.89706
C	-3.95334	2.80042	-0.72385
C	-3.17899	3.82985	-1.17855
C	-1.8347	3.58787	-1.48482
C	-1.29294	2.34588	-1.29231
C	1.8195	-2.02013	-0.42763
C	0.95382	0.69264	-0.31749
C	0.35899	-1.6842	-0.52342
C	-0.57947	-2.77976	-0.54842
C	-0.14575	-4.12753	-0.59583
C	-1.04018	-5.1599	-0.59176
C	-2.41655	-4.89011	-0.53968
C	-2.85902	-3.59855	-0.47976
C	-1.95522	-2.50773	-0.47262
C	-0.07115	-0.38527	-0.53301
C	-2.04822	1.265	-0.7745
C	-1.49726	-0.0817	-0.58539
C	-2.40186	-1.13175	-0.42901
C	-3.77841	-0.84363	-0.16936
C	-3.4208	1.50601	-0.53004
C	-4.28083	0.4112	-0.15648
C	-10.30282	2.44042	2.42148
C	-10.39023	2.26625	-0.07579
C	-10.98352	-2.65168	-1.16343
N	-10.68715	-3.44359	-1.9402
C	-12.69679	-1.63551	0.22104
N	-13.79236	-1.60664	0.56404
N	5.94024	-2.94567	-0.16277
C	6.84977	-2.361	-1.08698
C	7.83728	-3.13647	-1.68995

C	8.72919	-2.56295	-2.58291
C	8.63908	-1.21503	-2.89979
C	7.65063	-0.4424	-2.30701
C	6.76601	-1.00623	-1.40061
C	6.4233	-3.80248	0.86431
C	7.54138	-3.446	1.61465
C	8.01375	-4.28811	2.60954
C	7.36979	-5.48671	2.88158
C	6.25095	-5.84033	2.14086
C	5.78411	-5.01047	1.13333
N	3.91824	3.61284	0.46028
C	4.62244	3.58859	1.69538
C	4.85204	4.76859	2.39928
C	5.54737	4.74407	3.59825
C	6.00809	3.54392	4.12053
C	5.77255	2.36621	3.42573
C	5.09224	2.38553	2.21793
C	4.23342	4.59189	-0.52135
C	3.21822	5.29055	-1.17056
C	3.52612	6.23734	-2.13522
C	4.84827	6.51299	-2.45365
C	5.86175	5.82504	-1.80171
C	5.55967	4.865	-0.84825
H	-8.72435	-1.6296	-0.76429
H	-10.55589	-0.22291	2.3307
H	-11.83962	0.42402	1.3474
H	-8.15465	0.96466	2.21997
H	-7.93191	2.09791	0.91975
H	-6.54418	-0.92156	-0.81135
H	-5.8449	1.49559	0.88307
H	1.92737	-1.78654	1.69155
H	4.30895	-2.31742	1.84678
H	4.44892	-2.91912	-2.36387
H	2.07164	-2.36357	-2.52092
H	1.91329	0.50178	-2.21613
H	3.62268	2.18729	-1.7693
H	1.96097	2.84989	2.09165
H	0.25817	1.15198	1.64851
H	-4.99658	2.96868	-0.54857
H	-3.60285	4.80375	-1.32954
H	-1.22643	4.37346	-1.88963
H	-0.27755	2.19759	-1.56976
H	0.90195	-4.33724	-0.63635
H	-0.69337	-6.17417	-0.63327
H	-3.12235	-5.69797	-0.55292
H	-3.91392	-3.42544	-0.46382
H	-4.4242	-1.64714	0.10978
H	-10.05759	1.95208	3.35858
H	-9.73605	3.36371	2.36613
H	-11.35572	2.69883	2.44328

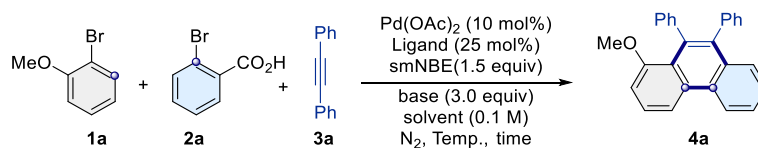
H	-9.86896	3.21419	-0.15323
H	-10.15971	1.68761	-0.96151
H	-11.45483	2.47257	-0.07943
H	7.90331	-4.18069	-1.45671
H	9.48584	-3.17286	-3.03806
H	9.32633	-0.77492	-3.59596
H	7.57284	0.6024	-2.53962
H	6.01007	-0.40231	-0.93873
H	8.03596	-2.51613	1.41485
H	8.87786	-4.00114	3.17758
H	7.73346	-6.13364	3.65609
H	5.74691	-6.76712	2.33758
H	4.92715	-5.29339	0.55471
H	4.48784	5.69685	2.00587
H	5.71805	5.66171	4.1279
H	6.54043	3.52675	5.05164
H	6.12729	1.43157	3.81601
H	4.92127	1.47301	1.6817
H	2.19606	5.08995	-0.91665
H	2.73328	6.7669	-2.62794
H	5.08445	7.25083	-3.19555
H	6.88831	6.02636	-2.0412
H	6.3456	4.32889	-0.35412

3. Experimental Section

3.1 Optimization of the reaction conditions

The oven-dried Schlenk tube (10 mL) equipped with a stirring bar was charged with substrates **1a** (0.1 mmol, 1.0 equiv), **2a** (0.2 mmol, 2.0 equiv), **3a** (0.15 mmol, 1.5 equiv), Pd(OAc)₂ (10 mol%), ligand (25 mol%) and base (2.5-4.5 equiv) in the air. Followed by the mediator smNBE (1.5 equiv) and the anhydrous solvent (0.1 M) was added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N₂ for three times. Finally, the reaction mixture was stirred at 130 °C for 24-72 h and then cooled to room temperature. Upon completion of the reaction, the mixture was diluted with 5 mL EtOAc and filtered through a short pad of Celite and the Celite pad were washed with an additional EtOAc (10 mL × 5). The filtrate was concentrated *in vacuo* to give the crude product. The crude ¹H NMR spectrum was taken using CH₂Br₂ (0.1 mmol) as internal standard.

Table S1. Screening of the reaction conditions.



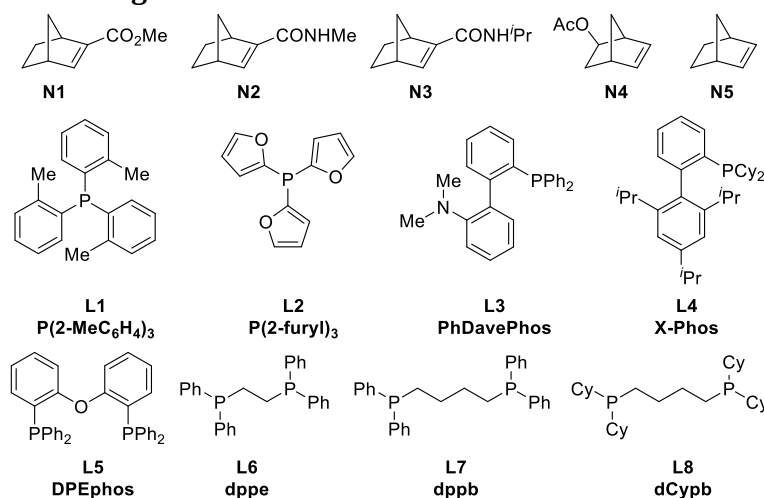
NO.	Ligand (25 mol%)	smNBE (1.5 eq.)	Base (3.0 eq.)	Solvent (0.1M)	Temp.	time	Yield ^a (%)
1	L1	N5	K_2CO_3	DMF	130 °C	24 h	n.d.
2	L1	N1	K_2CO_3	toluene	130 °C	24 h	33
3	L2	N1	K_2CO_3	toluene	130 °C	24 h	7
4	L3	N1	K_2CO_3	toluene	130 °C	24 h	trace
5	L4	N1	K_2CO_3	toluene	130 °C	24 h	4
^c 6	L5	N1	K_2CO_3	toluene	130 °C	24 h	11
^c 7	L6	N1	K_2CO_3	toluene	130 °C	24 h	4
^c 8	L7	N1	K_2CO_3	toluene	130 °C	24 h	2
^c 9	L8	N1	K_2CO_3	toluene	130 °C	24 h	2
10	L1	N1	Na_2CO_3	toluene	130 °C	24 h	13
11	L1	N1	Cs_2CO_3	toluene	130 °C	24 h	8
12	L1	N1	$^t\text{BuONa}$	toluene	130 °C	24 h	n.d.
13	L1	N1	KOAc	toluene	130 °C	24 h	trace
14	L1	N1	CsOAc	toluene	130 °C	24 h	5
15	L1	N1	K_3PO_4	toluene	130 °C	24 h	36
16	L1	N2	K_3PO_4	toluene	130 °C	24 h	21
17	L1	N3	K_3PO_4	toluene	130 °C	24 h	35
18	L1	N4	K_3PO_4	toluene	130 °C	24 h	trace
19	L1	N5	K_3PO_4	toluene	130 °C	24 h	n.d.
20	L1	-	K_3PO_4	toluene	130 °C	24 h	n.d.
21	L1 (20 mol%)	N1	K_3PO_4	toluene	130 °C	24 h	30
22	L1 (15 mol%)	N1	K_3PO_4	toluene	130 °C	24 h	29
23	L1	N1	K_3PO_4	xylene	130 °C	24 h	34
24	L1	N1	K_3PO_4	DMF	130 °C	24 h	n.d.
25	L1	N1	K_3PO_4 (2.5 eq.)	toluene	130 °C	24 h	16
26	L1	N1	K_3PO_4 (3.5 eq.)	toluene	130 °C	24 h	50
27	L1	N1	K_3PO_4 (4.0 eq.)	toluene	130 °C	24 h	40
28	L1	N1	K_2CO_3 (3.5 eq.)	toluene	130 °C	24 h	38
29	L1	N1	K_2CO_3 (4.0 eq.)	toluene	130 °C	24 h	42
30	L1	N1	K_2CO_3 (4.5 eq.)	toluene	130 °C	24 h	51
31	L1	N1	K_2CO_3 (5.0 eq.)	toluene	130 °C	24 h	29
32	L1	N1	K_2CO_3 (4.5 eq.)	toluene	90 °C	24 h	5

33	L1	N1	K ₂ CO ₃ (4.5 eq.)	toluene	110 °C	24 h	13
34	L1	N1	K ₂ CO ₃ (4.5 eq.)	toluene	120 °C	24 h	26
35	L1	N1	K ₂ CO ₃ (4.5 eq.)	toluene	140 °C	24 h	40
36	L1	N1	K ₂ CO ₃ (4.5 eq.)	dioxane	130 °C	24 h	7
37	L1	N1	K ₂ CO ₃ (4.5 eq.)	<i>t</i> -Amyl OH	130 °C	24 h	4
38	L1	N1	K ₂ CO ₃ (4.5 eq.)	toluene	130 °C	48 h	63
39	L1	N1	K ₂ CO ₃ (4.5 eq.)	toluene	130 °C	72 h	71 ^b

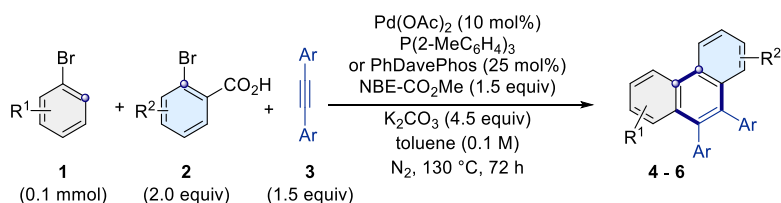
^aYield was determined by ¹H NMR with CH₂Br₂ (0.1mmol) as internal standard.

^bIsolated yield. ^c12 mol% ligand was used. n.d. = not detected.

NBE mediators and ligands:



3.2 General procedures for synthesis of products



General procedure A:

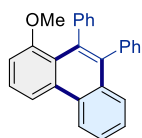
The oven-dried Schlenk tube (10 mL) equipped with a stirring bar was charged with substrates **1** (0.1 mmol, 1.0 equiv), **2** (0.2 mmol, 2.0 equiv), **3** (0.15 mmol, 1.5 equiv), Pd(OAc)₂ (10 mol%), P(2-MeC₆H₄)₃ (25 mol%) and K₂CO₃ (4.5 equiv) in the air. Followed by the mediator NBE-CO₂Me (1.5 equiv) and the anhydrous toluene (1.0 mL) were added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N₂ for three times. Finally, the reaction mixture was stirred at 130 °C for 72 h and then cooled to room temperature. Upon completion of the

reaction, the mixture was diluted with 5 mL EtOAc and filtered through a short pad of Celite and the Celite pad were washed with an additional EtOAc (10 mL $\times$ 5). The filtrate was concentrated *in vacuo* to give the crude product. The crude ^1H NMR spectrum was taken using CH_2Br_2 (0.1 mmol) as internal standard. Finally, the crude product was purified via silica gel column chromatography using PE/EtOAc as the eluent.

General procedure B:

The oven-dried Schlenk tube (10 mL) equipped with a stirring bar was charged with substrates **1** (0.1 mmol, 1.0 equiv), **2** (0.2 mmol, 2.0 equiv), **3** (0.15 mmol, 1.5 equiv), $\text{Pd}(\text{OAc})_2$ (10 mol%), PhDavePhos (25 mol%) and K_2CO_3 (4.5 equiv) in the air. Followed by the mediator NBE- CO_2Me (1.5 equiv) and the anhydrous toluene (1.0 mL) were added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N_2 for three times. Finally, the reaction mixture was stirred at 130 $^\circ\text{C}$ for 72 h and then cooled to room temperature. Upon completion of the reaction, the mixture was diluted with 5 mL EtOAc and filtered through a short pad of Celite and the Celite pad were washed with an additional EtOAc (10 mL $\times$ 5). The filtrate was concentrated *in vacuo* to give the crude product. The crude ^1H NMR spectrum was taken using CH_2Br_2 (0.1 mmol) as internal standard. Finally, the crude product was purified via silica gel column chromatography using PE/EtOAc as the eluent.

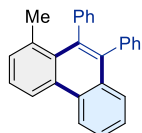
3.3 Characterization of products



1-methoxy-9,10-diphenylphenanthrene (4a)

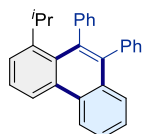
The general procedure A was followed. Yield: 71%, 25.6 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.76 (d, J = 8.4 Hz, 1H), 8.45 (d, J = 8.4 Hz, 1H), 7.66 – 7.55 (m, 2H), 7.48 – 7.39 (m, 2H), 7.26 – 7.11 (m, 3H), 7.12 – 6.99 (m, 7H), 6.96 (d, J = 8.0 Hz, 1H), 3.33 (s, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 157.9, 144.2, 139.9,

138.2, 135.2, 132.3, 132.1, 131.3, 129.7, 129.6, 127.9, 127.4, 127.0, 126.8, 126.4, 126.3, 126.2, 124.8, 123.1, 122.6, 115.6, 109.3, 56.0. HRMS (m/z, ESI): calcd for $C_{27}H_{21}O^+ (M+H)^+$ 361.1587, found 361.1551.



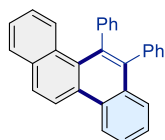
1-methyl-9,10-diphenylphenanthrene (4b)

The general procedure A was followed. Yield: 75%, 26.0 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.79 (d, J = 8.8 Hz, 1H), 8.75 (d, J = 8.8 Hz, 1H), 7.61 (t, J = 6.8 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.46 – 7.37 (m, 2H), 7.33 (d, J = 6.8 Hz, 1H), 7.22 – 7.13 (m, 3H), 7.12 – 6.99 (m, 7H), 1.91 (s, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 143.2, 140.1, 139.0, 137.0, 136.96, 131.6, 131.59, 131.5, 131.3, 130.6, 130.5, 127.8, 127.5, 127.1, 126.6, 126.5, 126.3, 126.2, 123.0, 121.4, 25.5. HRMS (m/z, ESI): calcd for $C_{27}H_{21}^+ (M+H)^+$ 345.1565, found 345.1574.



1-isopropyl-9,10-diphenylphenanthrene (4c)

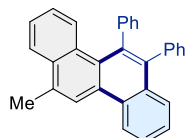
The general procedure A was followed. Yield: 79%, 29.4 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.77 (d, J = 8.4 Hz, 1H), 8.71 (d, J = 7.6 Hz, 1H), 7.69 – 7.56 (m, 3H), 7.45 – 7.36 (m, 2H), 7.24 – 7.12 (m, 3H), 7.12 – 6.98 (m, 7H), 2.88 (p, J = 6.8 Hz, 1H), 0.89 (d, J = 6.8 Hz, 6H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 148.7, 143.6, 140.3, 138.9, 136.1, 131.7, 131.5, 131.3, 130.7, 130.6, 129.8, 127.8, 127.5, 127.3, 126.6, 126.5, 126.4, 126.3, 126.2, 126.0, 123.0, 120.7, 29.4, 24.8. HRMS (m/z, ESI): calcd for $C_{29}H_{25}^+ (M+H)^+$ 373.1878, found 373.1888.



5,6-diphenylchrysene (4d)

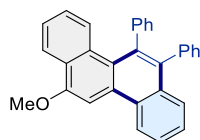
The general procedure A was followed. Yield: 95%, 36.2 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.86 (d, J = 8.4 Hz, 1H), 8.82 (d, J = 9.2 Hz, 1H), 8.01 (d, J = 8.8 Hz,

1H), 7.89 (d, $J = 7.6$ Hz, 1H), 7.68 (t, $J = 6.8$ Hz, 1H), 7.61 – 7.52 (m, 2H), 7.48 (t, $J = 7.2$ Hz, 1H), 7.39 (t, $J = 7.2$ Hz, 1H), 7.28 – 7.19 (m, 3H), 7.19 – 7.12 (m, 3H), 7.12 – 7.05 (m, 4H), 7.01 (t, $J = 7.6$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform- d) δ 143.7, 140.2, 139.5, 136.9, 133.9, 131.9, 131.4, 131.2, 130.2, 129.7, 128.8, 128.4, 128.2, 127.92, 127.9, 127.6, 126.7, 126.6, 126.4, 126.35, 125.7, 124.8, 123.3, 121.2. HRMS (m/z, ESI): calcd for $\text{C}_{30}\text{H}_{21}^+$ ($\text{M}+\text{H}$) $^+$ 381.1565, found 381.1578.



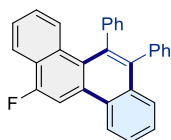
12-methyl-5,6-diphenylchrysene (4e)

The general procedure A was followed. Yield: 99%, 39.1 mg. ^1H NMR (400 MHz, Chloroform- d) δ 8.85 (d, $J = 8.4$ Hz, 1H), 8.66 (s, 1H), 8.06 (d, $J = 8.4$ Hz, 1H), 7.73 – 7.62 (m, 2H), 7.55 (d, $J = 8.0$ Hz, 1H), 7.45 (dt, $J = 12.5, 7.4$ Hz, 2H), 7.22 (m, 3H), 7.16 – 7.11 (m, 3H), 7.10 – 6.97 (m, 5H), 2.89 (s, 3H). ^{13}C NMR (101 MHz, Chloroform- d) δ 143.7, 140.2, 138.6, 136.6, 133.7, 133.3, 131.9, 131.4, 131.37, 131.2, 129.7, 129.4, 129.3, 128.1, 127.8, 127.6, 127.0, 126.5, 126.4, 126.3, 126.2, 125.6, 124.3, 124.1, 123.2, 121.7, 20.8. HRMS (m/z, ESI): calcd for $\text{C}_{31}\text{H}_{23}^+$ ($\text{M}+\text{H}$) $^+$ 395.1722, found 395.1717.



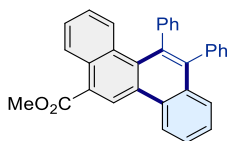
12-methoxy-5,6-diphenylchrysene (4f)

The general procedure A was followed. Yield: 93%, 38.1 mg. ^1H NMR (400 MHz, Chloroform- d) δ 8.75 (d, $J = 8.4$ Hz, 1H), 8.39 (d, $J = 8.0$ Hz, 1H), 8.04 (s, 1H), 7.64 (t, $J = 7.6$ Hz, 1H), 7.56 (t, $J = 9.6$ Hz, 2H), 7.46 (t, $J = 7.6$ Hz, 1H), 7.40 (t, $J = 7.6$ Hz, 1H), 7.28 – 7.17 (m, 3H), 7.17 – 6.96 (m, 8H), 4.24 (s, 3H). ^{13}C NMR (101 MHz, Chloroform- d) δ 154.5, 143.7, 140.3, 137.1, 136.7, 132.4, 132.1, 131.6, 131.2, 130.5, 129.6, 128.8, 128.1, 127.9, 127.5, 127.3, 126.6, 126.3, 126.2, 126.1, 125.4, 125.3, 123.2, 123.18, 121.9, 97.9, 55.7. HRMS (m/z, ESI): calcd for $\text{C}_{31}\text{H}_{23}\text{O}^+$ ($\text{M}+\text{H}$) $^+$ 411.1671, found 411.1692.



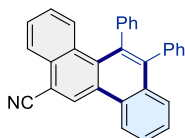
12-fluoro-5,6-diphenylchrysene (4g)

The general procedure A was followed. Yield: 91%, 36.5 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.68 (d, J = 8.4 Hz, 1H), 8.45 (d, J = 12.8 Hz, 1H), 8.21 (d, J = 8.0 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.60 – 7.52 (m, 2H), 7.52 – 7.43 (m, 2H), 7.26 – 7.19 (m, 3H), 7.18 – 7.12 (m, 3H), 7.10 – 7.02 (m, 5H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 158.1 (d, $J_{\text{C-F}}$ = 250.8 Hz), 143.3, 139.9, 138.8 (d, $J_{\text{C-F}}$ = 2.1 Hz), 136.6, 132.9 (d, $J_{\text{C-F}}$ = 4.9 Hz), 131.9, 131.3, 131.1, 130.0 (d, $J_{\text{C-F}}$ = 8.8 Hz), 129.6 (d, $J_{\text{C-F}}$ = 4.4 Hz), 128.8 (d, $J_{\text{C-F}}$ = 2.7 Hz), 128.3, 127.9, 127.6, 127.0, 126.6, 126.5, 126.4, 126.0 (d, $J_{\text{C-F}}$ = 1.6 Hz), 125.9, 125.1 (d, $J_{\text{C-F}}$ = 2.3 Hz), 124.9 (d, $J_{\text{C-F}}$ = 16.8 Hz), 123.3, 120.6 (d, $J_{\text{C-F}}$ = 6.9 Hz), 103.9 (d, $J_{\text{C-F}}$ = 21.8 Hz). HRMS (m/z , ESI): calcd for $\text{C}_{30}\text{H}_{20}\text{F}^+$ ($\text{M}+\text{H}$) $^+$ 399.1471, found 399.1473.



methyl 11,12-diphenylchrysene-6-carboxylate (4h)

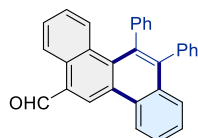
The general procedure A was followed. Yield: 83%, 36.2 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.48 (s, 1H), 8.86 (t, J = 10.0 Hz, 2H), 7.79 – 7.67 (m, 2H), 7.59 (d, J = 8.4 Hz, 1H), 7.54 – 7.42 (m, 2H), 7.28 – 7.18 (m, 3H), 7.17 – 7.05 (m, 5H), 7.05 – 6.95 (m, 3H), 4.12 (s, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 168.5, 143.1, 141.9, 139.7, 136.4, 132.0, 131.8, 131.3, 131.1, 130.8, 130.75, 130.3, 129.3, 128.2, 128.0, 127.9, 127.7, 127.2, 127.1, 126.8, 126.7, 126.6, 126.4, 126.1, 125.8, 124.8, 123.2, 52.6. HRMS (m/z , ESI): calcd for $\text{C}_{32}\text{H}_{23}\text{O}_2^+$ ($\text{M}+\text{H}$) $^+$ 439.1693, found 439.1661.



11,12-diphenylchrysene-6-carbonitrile (4i)

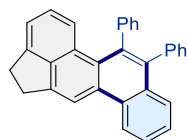
The general procedure A was followed. Yield: 61%, 25.0 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.30 (s, 1H), 8.79 (d, J = 8.4 Hz, 1H), 8.33 (d, J = 8.4 Hz, 1H), 7.77

(t, $J = 6.8$ Hz, 1H), 7.68 (d, $J = 8.8$ Hz, 1H), 7.64 – 7.51 (m, 3H), 7.32 – 7.21 (m, 3H), 7.21 – 7.14 (m, 3H), 7.15 – 7.11 (m, 1H), 7.11 – 7.06 (m, 2H), 7.06 – 6.99 (m, 2H). ^{13}C NMR (101 MHz, Chloroform- d) δ 143.0, 142.6, 139.3, 136.5, 132.0, 131.1, 131.0, 130.9, 130.7, 129.6, 129.5, 129.3, 128.4, 128.2, 127.9, 127.8, 127.76, 127.6, 127.4, 126.8, 126.79, 126.2, 125.6, 122.9, 118.7, 110.0. HRMS (m/z , ESI): calcd for $\text{C}_{31}\text{H}_{20}\text{N}^+$ ($\text{M}+\text{H}$) $^+$ 406.1517, found 406.1558.



11,12-diphenylchrysene-6-carbaldehyde (4j)

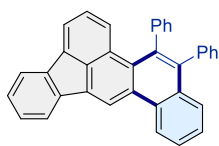
The general procedure A was followed. Yield: 54%, 22.2 mg. ^1H NMR (400 MHz, Chloroform- d) δ 10.58 (s, 1H), 9.32 (d, $J = 8.4$ Hz, 1H), 9.25 (s, 1H), 8.89 (d, $J = 8.4$ Hz, 1H), 7.77 (t, $J = 7.6$ Hz, 1H), 7.72 (d, $J = 8.8$ Hz, 1H), 7.61 (d, $J = 8.4$ Hz, 1H), 7.54 (q, $J = 7.6$ Hz, 2H), 7.31 – 7.21 (m, 3H), 7.19 – 7.11 (m, 3H), 7.11 – 7.04 (m, 3H), 7.04 – 6.96 (m, 2H). ^{13}C NMR (101 MHz, Chloroform- d) δ 193.6, 143.3, 142.8, 139.4, 136.5, 134.3, 132.0, 131.9, 131.8, 131.1, 130.9, 130.3, 130.2, 130.0, 129.2, 128.3, 128.2, 128.0, 127.7, 127.64, 127.6, 127.3, 126.8, 126.6, 125.4, 124.8, 122.9. HRMS (m/z , ESI): calcd for $\text{C}_{31}\text{H}_{21}\text{O}^+$ ($\text{M}+\text{H}$) $^+$ 409.1514, found 409.1559.



11,12-diphenyl-4,5-dihydrocyclopenta[hi]chrysene (4k)

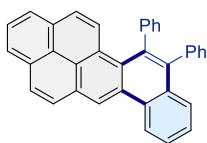
The general procedure A was followed. Yield: 86%, 35.0 mg. ^1H NMR (400 MHz, Chloroform- d) δ 8.89 (d, $J = 8.4$ Hz, 1H), 8.70 (s, 1H), 7.65 (t, $J = 7.6$ Hz, 1H), 7.53 (d, $J = 8.4$ Hz, 1H), 7.47 (t, $J = 7.6$ Hz, 1H), 7.27 (d, $J = 6.8$ Hz, 1H), 7.26 – 7.15 (m, 6H), 7.15 – 7.07 (m, 4H), 7.03 (t, $J = 8.0$ Hz, 1H), 6.80 (d, $J = 8.8$ Hz, 1H), 3.54 (dd, $J = 9.6, 5.6$ Hz, 2H), 3.45 (dd, $J = 8.8, 4.4$ Hz, 2H). ^{13}C NMR (101 MHz, Chloroform- d) δ 145.6, 145.5, 143.4, 140.2, 139.2, 137.9, 137.5, 132.0, 131.7, 131.5, 131.1, 130.4, 130.0, 128.2, 127.8, 127.5, 127.0, 126.6, 126.4, 126.3, 126.2, 125.5, 124.5, 123.6, 120.4, 114.5, 30.6, 29.9. HRMS (m/z , ESI): calcd for $\text{C}_{32}\text{H}_{23}^+$ ($\text{M}+\text{H}$) $^+$

407.1722, found 407.1722.



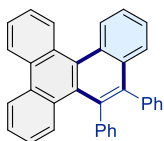
4,5-diphenylindeno[1,2,3-*hi*]chrysene (4l)

The general procedure A was followed. Yield: 98%, 45.1 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.36 (s, 1H), 9.02 (d, J = 8.4 Hz, 1H), 8.11 (d, J = 6.4 Hz, 1H), 7.87 (d, J = 6.8 Hz, 1H), 7.79 (d, J = 6.8 Hz, 1H), 7.73 (t, J = 7.2 Hz, 1H), 7.63 – 7.48 (m, 2H), 7.47 – 7.36 (m, 2H), 7.32 – 7.02 (m, 11H), 6.92 (d, J = 8.4 Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 143.0, 140.7, 139.9, 139.8, 138.5, 137.7, 136.9, 135.8, 133.0, 132.0, 131.23, 131.2, 131.19, 131.1, 129.0, 128.5, 128.4, 128.2, 128.0, 127.8, 127.6, 127.5, 127.1, 126.9, 126.74, 126.7, 126.5, 123.5, 121.7, 121.3, 119.4, 116.0. HRMS (m/z , ESI): calcd for $\text{C}_{36}\text{H}_{23}^+$ ($\text{M}+\text{H}$) $^+$ 455.1722, found 455.1714.



11,12-diphenylbenzo[*pqr*]picene (4m)

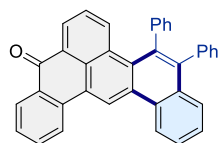
The general procedure A was followed. Yield: 75%, 34.1 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.48 (s, 1H), 9.10 (d, J = 8.4 Hz, 1H), 8.16 (d, J = 9.2 Hz, 1H), 8.08 (d, J = 7.6 Hz, 1H), 8.01 (t, J = 6.4 Hz, 2H), 7.90 (dd, J = 14.4, 7.2 Hz, 2H), 7.75 (t, J = 7.6 Hz, 1H), 7.65 – 7.51 (m, 3H), 7.30 – 7.21 (m, 3H), 7.19 – 7.09 (m, 5H), 7.09 – 7.02 (m, 2H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 143.9, 140.2, 139.1, 136.7, 131.8, 131.6, 131.4, 131.3, 130.5, 130.2, 130.1, 129.2, 128.7, 128.3, 128.2, 128.1, 128.0, 127.64, 127.6, 127.0, 126.8, 126.4, 126.3, 126.2, 126.18, 125.2, 125.1, 125.0, 124.9, 124.8, 123.5, 119.9. HRMS (m/z , ESI): calcd for $\text{C}_{36}\text{H}_{23}^+$ ($\text{M}+\text{H}$) $^+$ 455.1722, found 455.1718.



5,6-diphenylbenzo[*g*]chrysene (4n)

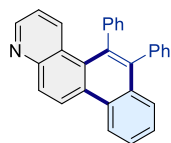
The general procedure A was followed. Yield: 61%, 26.1 mg. ^1H NMR (400 MHz,

Chloroform-*d*) δ 8.80 (t, J = 8.8 Hz, 2H), 8.65 (d, J = 7.6 Hz, 1H), 8.55 (d, J = 8.4 Hz, 1H), 7.77 (d, J = 8.4 Hz, 1H), 7.66 (p, J = 6.4 Hz, 2H), 7.62 – 7.50 (m, 2H), 7.46 (t, J = 7.6 Hz, 1H), 7.42 (t, J = 7.2 Hz, 1H), 7.37 – 7.21 (m, 3H), 7.17 (d, J = 7.2 Hz, 2H), 7.12 – 7.01 (m, 6H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 142.8, 139.5, 138.4, 135.9, 132.4, 132.0, 131.7, 131.5, 131.2, 130.8, 130.7, 129.94, 129.9, 129.6, 129.4, 128.9, 128.6, 128.5, 128.4, 127.9, 127.7, 127.0, 126.9, 126.6, 126.3, 126.2, 125.7, 125.6, 123.7, 123.2. HRMS (m/z , ESI): calcd for $\text{C}_{34}\text{H}_{23}^+$ ($\text{M}+\text{H}$) $^+$ 431.1722, found 431.1724.



5,6-diphenyl-10H-dibenzo[*gh,m*]tetraphen-10-one (4o)

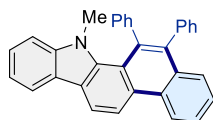
The general procedure A was followed. Yield: 84%, 40.4 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.75 (s, 1H), 8.95 (d, J = 8.4 Hz, 1H), 8.60 (dd, J = 7.2, 4.8 Hz, 2H), 8.52 (d, J = 7.6 Hz, 1H), 8.03 (d, J = 8.8 Hz, 1H), 7.82 (t, J = 7.2 Hz, 1H), 7.77 (t, J = 7.6 Hz, 1H), 7.66 – 7.51 (m, 3H), 7.33 – 7.19 (m, 4H), 7.19 – 7.14 (m, 3H), 7.10 (d, J = 7.6 Hz, 2H), 7.07 – 6.99 (m, 2H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 184.2, 143.0, 141.5, 139.6, 136.5, 136.4, 135.2, 133.5, 132.2, 131.2, 131.16, 131.1, 131.0, 130.2, 129.4, 129.3, 129.0, 128.6, 128.54, 128.5, 128.4, 128.3, 128.2, 127.8, 127.3, 127.28, 126.72, 126.7, 125.3, 125.1, 123.3, 122.9, 120.5. HRMS (m/z , ESI): calcd for $\text{C}_{37}\text{H}_{23}\text{O}^+$ ($\text{M}+\text{H}$) $^+$ 483.1743, found 483.1706.



5,6-diphenyl-10H-dibenzo[*gh,m*]tetraphen-10-one (4o)

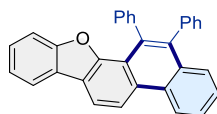
The general procedure A was followed. Yield: 44%, 17.0 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.08 (d, J = 9.2 Hz, 1H), 8.89 (d, J = 8.8 Hz, 1H), 8.77 (s, 1H), 8.28 (d, J = 9.6 Hz, 1H), 7.79 (d, J = 8.8 Hz, 1H), 7.73 (t, J = 7.2 Hz, 1H), 7.58 (d, J = 8.4 Hz, 1H), 7.56 – 7.49 (m, 1H), 7.30 – 7.16 (m, 6H), 7.15 – 7.03 (m, 4H), 6.92 (dd, J = 9.2, 4.0 Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 149.2, 149.0, 142.9, 140.2,

139.7, 136.6, 136.1, 132.1, 131.2, 131.0, 130.0, 129.5, 129.4, 128.6, 128.0, 127.7, 127.6, 127.1, 127.0, 126.8, 126.6, 125.1, 123.3, 119.3. HRMS (m/z, ESI): calcd for $C_{29}H_{20}N^+$ (M+H) $^+$ 382.1590, found 382.1567.



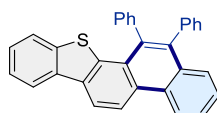
13-methyl-1,2-diphenyl-13H-naphtho[2,1-a]carbazole (4q)

The general procedure A was followed. Yield: 55%, 23.8 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.84 (d, J = 8.4 Hz, 1H), 8.66 (d, J = 8.8 Hz, 1H), 8.37 (d, J = 8.4 Hz, 1H), 8.18 (d, J = 8.0 Hz, 1H), 7.73 – 7.61 (m, 2H), 7.44 (dt, J = 16.8, 7.2 Hz, 2H), 7.38 – 7.27 (m, 4H), 7.21 – 7.05 (m, 4H), 7.05 – 6.91 (m, 4H), 2.89 (s, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 143.5, 141.4, 140.4, 139.6, 137.5, 133.9, 132.8, 131.7, 131.6, 131.4, 130.8, 127.92, 127.87, 127.0, 126.7, 126.6, 126.3, 126.2, 125.6, 123.7, 123.6, 123.0, 120.0, 119.8, 119.6, 118.3, 115.6, 110.3, 36.6. HRMS (m/z, ESI): calcd for $C_{33}H_{24}N^+$ (M+H) $^+$ 434.1830, found 434.1866.



1,2-diphenylphenanthro[1,2-b]benzofuran (4r)

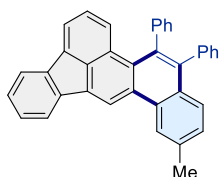
The general procedure B was followed. Yield: 50%, 21.1 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.89 (d, J = 8.4 Hz, 1H), 8.83 (d, J = 8.8 Hz, 1H), 8.22 (d, J = 8.8 Hz, 1H), 8.03 – 7.95 (m, 1H), 7.70 (t, J = 7.2 Hz, 1H), 7.63 (q, J = 8.4 Hz, 1H), 7.57 (d, J = 8.4 Hz, 1H), 7.50 (t, J = 7.2 Hz, 1H), 7.35 – 7.14 (m, 11H), 7.07 – 7.01 (m, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 155.7, 153.3, 141.5, 139.4, 138.8, 134.2, 132.4, 132.3, 132.0, 131.3, 130.9, 130.2, 129.9, 128.3, 127.6, 127.1, 126.9, 126.8, 126.6, 126.5, 126.3, 124.0, 123.2, 122.7, 122.3, 120.2, 119.2, 119.1, 118.2, 111.7. HRMS (m/z, ESI): calcd for $C_{32}H_{21}O^+$ (M+H) $^+$ 421.1587, found 421.1635.



1,2-diphenylbenzo[b]phenanthro[2,1-d]thiophene (4s)

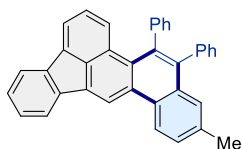
The general procedure A was followed. Yield: 67%, 29.5 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 8.95 (d, J = 8.8 Hz, 1H), 8.90 (d, J = 8.4 Hz, 1H), 8.42 (d, J = 8.8 Hz,

1H), 8.21 (d, $J = 7.6$ Hz, 1H), 7.77 – 7.61 (m, 2H), 7.57 – 7.47 (m, 2H), 7.42 (t, $J = 7.6$ Hz, 1H), 7.40 – 7.31 (m, 2H), 7.31 – 7.16 (m, 7H), 7.13 (d, $J = 6.8$ Hz, 2H). ^{13}C NMR (101 MHz, Chloroform- d) δ 140.9, 139.7, 139.3, 139.2, 138.6, 136.0, 135.2, 134.6, 132.8, 131.7, 131.1, 130.6, 129.6, 128.14, 128.1, 128.07, 127.6, 127.0, 126.9, 126.8, 126.6, 126.3, 124.1, 123.3, 122.1, 121.3, 120.4, 120.2. HRMS (m/z , ESI): calcd for $\text{C}_{32}\text{H}_{21}\text{S}^+$ ($\text{M}+\text{H}$) $^+$ 437.1286, found 437.1288.



8-methyl-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5a)

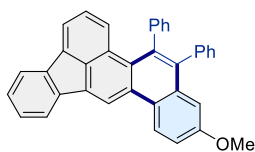
The general procedure A was followed. Yield: 97%, 45.5 mg. ^1H NMR (400 MHz, Chloroform- d) δ 9.33 (s, 1H), 8.77 (s, 1H), 8.13 (d, $J = 6.8$ Hz, 1H), 7.86 (d, $J = 6.4$ Hz, 1H), 7.77 (d, $J = 6.4$ Hz, 1H), 7.49 – 7.37 (m, 3H), 7.34 (d, $J = 8.4$ Hz, 1H), 7.28 – 7.04 (m, 11H), 6.92 (d, $J = 8.8$ Hz, 1H), 2.69 (s, 3H). ^{13}C NMR (101 MHz, Chloroform- d) δ 143.1, 140.7, 140.0, 139.8, 138.6, 136.9, 136.8, 136.4, 135.6, 133.0, 131.4, 131.24, 131.2, 130.8, 130.1, 129.0, 128.7, 128.6, 128.3, 128.1, 128.0, 127.8, 127.6, 127.4, 127.0, 126.8, 126.4, 123.0, 121.7, 121.3, 119.3, 116.1, 22.4. HRMS (m/z , ESI): calcd for $\text{C}_{37}\text{H}_{25}^+$ ($\text{M}+\text{H}$) $^+$ 469.1878, found 469.1871.



7-methyl-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5b)

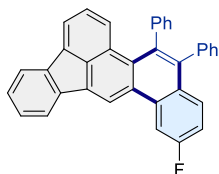
The general procedure A was followed. Yield: 94%, 44.2 mg. ^1H NMR (400 MHz, Chloroform- d) δ 9.35 (s, 1H), 8.91 (d, $J = 8.8$ Hz, 1H), 8.12 (d, $J = 7.6$ Hz, 1H), 7.88 (d, $J = 6.4$ Hz, 1H), 7.80 (d, $J = 6.8$ Hz, 1H), 7.56 (d, $J = 8.8$ Hz, 1H), 7.48 – 7.38 (m, 2H), 7.32 (s, 1H), 7.28 – 7.19 (m, 6H), 7.19 – 7.13 (m, 3H), 7.10 (d, $J = 7.6$ Hz, 2H), 6.91 (d, $J = 8.4$ Hz, 1H), 2.44 (s, 3H). ^{13}C NMR (101 MHz, Chloroform- d) δ 143.2, 140.8, 139.9, 139.6, 138.6, 137.8, 136.9, 136.5, 135.8, 132.9, 132.1, 131.3, 131.2, 129.3, 129.1, 128.8, 128.3, 128.1, 128.0, 127.8, 127.6, 127.5, 127.3, 127.1, 126.8, 126.4, 123.5, 121.7, 121.3, 119.2, 116.1, 21.9. HRMS (m/z , ESI): calcd for $\text{C}_{37}\text{H}_{25}^+$

(M+H)⁺ 469.1878, found 469.1882.



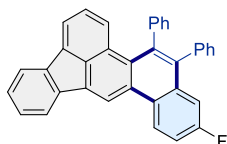
7-methoxy-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5c)

The general procedure A was followed. Yield: 98%, 47.5 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.26 (s, 1H), 8.90 (d, *J* = 9.2 Hz, 1H), 8.09 (dd, *J* = 6.0, 2.4 Hz, 1H), 7.91 – 7.83 (m, 1H), 7.77 (d, *J* = 6.8 Hz, 1H), 7.46 – 7.39 (m, 2H), 7.36 (dd, *J* = 9.6, 2.8 Hz, 1H), 7.25 – 7.20 (m, 5H), 7.20 – 7.12 (m, 4H), 7.11 – 7.06 (m, 2H), 6.91 – 6.83 (m, 2H), 3.69 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 158.2, 143.1, 140.8, 139.9, 139.3, 138.6, 138.3, 136.9, 135.9, 133.5, 132.6, 131.4, 131.2, 131.1, 129.1, 128.3, 128.1, 127.7, 127.6, 127.4, 127.1, 127.06, 126.8, 126.5, 126.1, 125.3, 121.7, 121.2, 119.0, 117.7, 115.9, 107.7, 55.2. HRMS (*m/z*, ESI): calcd for C₃₇H₂₅O⁺ (M+H)⁺ 485.1827, found 485.1837.



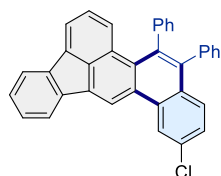
8-fluoro-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5d)

The general procedure A was followed. Yield: 71%, 34.0 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.07 (s, 1H), 8.53 (d, *J* = 11.6 Hz, 1H), 8.06 (d, *J* = 6.0 Hz, 1H), 7.84 (d, *J* = 6.8 Hz, 1H), 7.76 (d, *J* = 6.8 Hz, 1H), 7.57 – 7.47 (m, 1H), 7.42 (q, *J* = 8.0, 4.0 Hz, 2H), 7.29 – 7.16 (m, 7H), 7.16 – 7.00 (m, 5H), 6.88 (d, *J* = 8.8 Hz, 1H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 161.6 (d, *J*_{C-F} = 246.1 Hz), 142.8, 140.7, 139.6, 138.3, 136.9, 136.0, 133.2, 132.7 (d, *J*_{C-F} = 8.3 Hz), 131.1, 130.6, 130.52, 130.5, 129.2, 128.9, 128.4, 128.3, 127.8, 127.7, 127.6, 127.2, 126.9, 126.6, 121.8, 121.3, 119.6, 116.0 (d, *J*_{C-F} = 23.8 Hz), 115.8, 108.1 (d, *J*_{C-F} = 22.5 Hz). HRMS (*m/z*, ESI): calcd for C₃₆H₂₂F⁺ (M+H)⁺ 473.1627, found 473.1615.



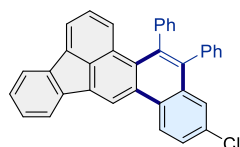
7-fluoro-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5e)

The general procedure A was followed. Yield: 95%, 44.9 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.13 (s, 1H), 8.86 (dd, $J = 9.2, 5.6$ Hz, 1H), 8.07 – 7.97 (m, 1H), 7.82 (m, 1H), 7.73 (d, $J = 6.8$ Hz, 1H), 7.45 – 7.34 (m, 3H), 7.25 – 7.15 (m, 6H), 7.15 – 7.05 (m, 4H), 7.04 – 6.98 (m, 2H), 6.83 (d, $J = 8.4$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 161.3 (d, $J_{\text{C-F}} = 246.3$ Hz), 142.7, 140.7, 139.3, 139.2 (d, $J_{\text{C-F}} = 4.3$ Hz), 138.8, 138.4, 136.9, 136.2, 133.5 (d, $J_{\text{C-F}} = 8.7$ Hz), 132.8, 131.13, 131.1, 131.0, 128.9, 128.4, 128.3, 127.94, 127.9, 127.8, 127.7, 127.5, 127.2, 127.0, 126.7, 126.0 (d, $J_{\text{C-F}} = 8.6$ Hz), 121.7, 121.3, 119.4, 116.0 (d, $J_{\text{C-F}} = 24.1$ Hz), 115.7, 111.8 (d, $J_{\text{C-F}} = 22.0$ Hz). HRMS (m/z , ESI): calcd for $\text{C}_{36}\text{H}_{22}\text{F}^+$ ($\text{M}+\text{H}$) $^+$ 473.1627, found 473.1623.



8-chloro-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5f)

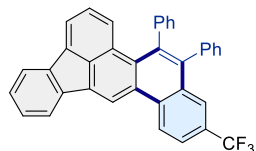
The general procedure A was followed. Yield: 96%, 47.1 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.06 (s, 1H), 8.87 (s, 1H), 8.06 (d, $J = 6.4$ Hz, 1H), 7.82 (d, $J = 6.4$ Hz, 1H), 7.72 (d, $J = 6.8$ Hz, 1H), 7.49 – 7.35 (m, 4H), 7.29 – 7.16 (m, 6H), 7.15 – 6.96 (m, 5H), 6.84 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 142.7, 140.6, 139.44, 139.36, 138.3, 137.9, 136.9, 136.1, 133.1, 132.9, 132.2, 131.1, 131.0, 130.3, 130.2, 129.7, 129.1, 128.7, 128.4, 128.3, 127.8, 127.7, 127.6, 127.23, 127.2, 127.0, 126.7, 123.0, 121.9, 121.3, 119.6, 115.6. HRMS (m/z , ESI): calcd for $\text{C}_{36}\text{H}_{22}\text{Cl}^+$ ($\text{M}+\text{H}$) $^+$ 489.1332, found 489.1330.



7-chloro-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5g)

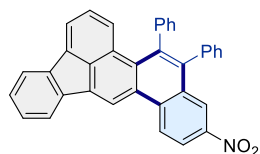
The general procedure A was followed. Yield: 98%, 48.0 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.29 (s, 1H), 8.95 (d, $J = 9.2$ Hz, 1H), 8.17 – 8.09 (m, 1H), 7.92 – 7.85 (m, 1H), 7.82 (d, $J = 7.2$ Hz, 1H), 7.67 (dd, $J = 8.8, 2.4$ Hz, 1H), 7.53 (s, 1H), 7.49 – 7.40 (m, 2H), 7.30 – 7.21 (m, 6H), 7.19 – 7.13 (m, 3H), 7.13 – 7.06 (m, 2H),

6.91 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform- d) δ 148.2, 147.4, 142.7, 140.8, 139.1, 139.0, 138.4, 137.0, 136.4, 133.1, 132.8, 131.2, 131.1, 131.0, 129.6, 128.9, 128.6, 128.4, 127.9, 127.8, 127.6, 127.4, 127.3, 127.1, 126.9, 126.8, 125.3, 121.8, 121.4, 119.6, 115.7. HRMS (m/z , ESI): calcd for $\text{C}_{36}\text{H}_{22}\text{Cl}^+$ ($\text{M}+\text{H}$) $^+$ 489.1332, found 489.1319.



4,5-diphenyl-7-(trifluoromethyl)indeno[1,2,3-*hi*]chrysene (5h)

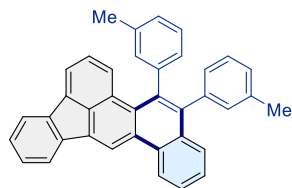
The general procedure A was followed (note: 20 mol% of $\text{Pd}(\text{OAc})_2$ and 40 mol% of $\text{P}(2\text{-MeC}_6\text{H}_4)_3$ were used). Yield: 55%, 28.7 mg. ^1H NMR (400 MHz, Chloroform- d) δ 9.30 (s, 1H), 9.08 (d, $J = 8.8$ Hz, 1H), 8.18 – 8.08 (m, 1H), 7.94 – 7.86 (m, 2H), 7.84 (s, 1H), 7.81 (d, $J = 7.2$ Hz, 1H), 7.50 – 7.40 (m, 2H), 7.31 – 7.20 (m, 6H), 7.20 – 7.11 (m, 3H), 7.11 – 7.03 (m, 2H), 6.91 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform- d) δ 142.5, 140.7, 140.1, 139.1, 138.7, 138.3, 137.0, 136.5, 133.3, 132.9, 131.3, 131.1, 131.0, 130.8, 129.8, 128.8, 128.54, 128.48, 128.36 (q, $J_{\text{C-F}} = 31.9$ Hz), 127.93, 127.9, 127.7, 127.5, 127.2, 127.0, 125.4 (q, $J_{\text{C-F}} = 4.8$ Hz), 124.5 (q, $J_{\text{C-F}} = 262.2$ Hz), 124.6, 122.3 (q, $J_{\text{C-F}} = 4.8$ Hz), 121.9, 121.4, 119.9, 115.8. HRMS (m/z , ESI): calcd for $\text{C}_{37}\text{H}_{22}\text{F}_3^+$ ($\text{M}+\text{H}$) $^+$ 523.1668, found 523.1793.



7-nitro-4,5-diphenylindeno[1,2,3-*hi*]chrysene (5i)

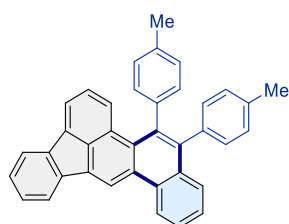
The general procedure A was followed. Yield: 50%, 24.9 mg. ^1H NMR (400 MHz, Chloroform- d) δ 9.89 (s, 1H), 9.26 (s, 1H), 8.23 (dd, $J = 9.2, 2.0$ Hz, 1H), 8.17 (d, $J = 6.8$ Hz, 1H), 7.85 (d, $J = 6.4$ Hz, 1H), 7.81 (d, $J = 6.8$ Hz, 1H), 7.67 (d, $J = 9.2$ Hz, 1H), 7.46 (p, $J = 7.2$ Hz, 2H), 7.32 – 7.23 (m, 6H), 7.20 – 7.11 (m, 3H), 7.11 – 7.05 (m, 2H), 6.88 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform- d) δ 145.9, 142.2, 141.7, 140.7, 139.3, 138.7, 138.0, 137.4, 137.2, 135.1, 134.2, 133.3, 132.0, 131.1, 130.7, 130.5, 129.6, 128.8, 128.7, 128.5, 128.0, 127.94, 127.9, 127.7, 127.4, 127.1,

122.2, 121.4, 120.2, 120.1, 120.0, 115.3. HRMS (m/z, ESI): calcd for $C_{36}H_{22}NO_2^+$ (M+H)⁺ 500.1572, found 500.1548.



4,5-di-*m*-tolyldeno[1,2,3-*hi*]chrysene (6a)

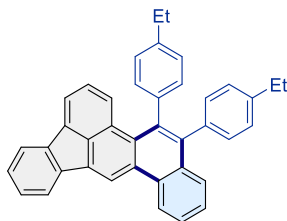
The general procedure A was followed (note: the products were isolated as an inseparable mixture of diastereoisomers, *d.r.* = 1:1). Yield: 90%, 43.5 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.37 (s, 1H), 9.01 (d, *J* = 8.8 Hz, 1H), 8.16 – 8.07 (m, 1H), 7.91 – 7.84 (m, 1H), 7.81 (d, *J* = 6.8 Hz, 1H), 7.72 (t, *J* = 7.2 Hz, 1H), 7.60 (d, *J* = 8.0 Hz, 1H), 7.51 (t, *J* = 7.2 Hz, 1H), 7.46 – 7.36 (m, 2H), 7.21 – 7.08 (m, 3H), 7.07 – 6.84 (m, 7H), 2.26 (s, 1.50H), 2.25 (s, 1.44H), 2.23 (s, 1.47H), 2.21 (s, 1.53H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 142.8, 140.7, 140.1, 140.06, 139.7, 138.6, 137.8, 137.76, 137.7, 137.0, 136.9, 136.8, 135.7, 133.0, 132.1, 132.0, 131.9, 131.8, 131.2, 131.1, 129.1, 128.7, 128.6, 128.3, 128.24, 128.2, 128.16, 128.1, 128.05, 127.5, 127.4, 127.39, 127.3, 127.1, 127.08, 127.06, 126.6, 123.4, 121.7, 121.3, 119.4, 116.1, 21.5. HRMS (m/z, ESI): calcd for $C_{38}H_{27}^+$ (M+H)⁺ 483.2035, found 483.2025.



4,5-di-*p*-tolyldeno[1,2,3-*hi*]chrysene (6b)

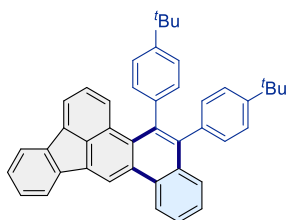
The general procedure A was followed. Yield: 97%, 47.2 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.33 (s, 1H), 8.98 (d, *J* = 8.4 Hz, 1H), 8.08 (d, *J* = 6.4 Hz, 1H), 7.85 (d, *J* = 6.4 Hz, 1H), 7.78 (d, *J* = 6.8 Hz, 1H), 7.69 (t, *J* = 7.6 Hz, 1H), 7.55 (d, *J* = 8.4 Hz, 1H), 7.48 (t, *J* = 6.8 Hz, 1H), 7.44 – 7.35 (m, 2H), 7.16 (t, *J* = 7.6 Hz, 1H), 7.10 – 6.91 (m, 9H), 2.34 (s, 3H), 2.32 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 140.7, 140.1, 140.05, 138.6, 137.8, 136.9, 136.8, 136.2, 135.8, 135.7, 133.0, 132.3, 131.2, 131.17, 131.0, 130.9, 129.1, 128.9, 128.4, 128.1, 128.0, 127.4, 127.0, 126.6, 123.4,

121.7, 121.2, 119.3, 116.0, 21.5, 21.4. HRMS (m/z, ESI): calcd for $C_{38}H_{27}^+$ (M+H)⁺ 483.2035, found 483.2026.



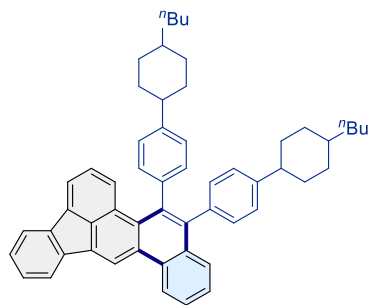
4,5-bis(4-ethylphenyl)indeno[1,2,3-*hi*]chrysene (6c)

The general procedure A was followed. Yield: 94%, 48.2 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.34 (s, 1H), 8.98 (d, *J* = 8.8 Hz, 1H), 8.13 – 8.05 (m, 1H), 7.85 (dd, *J* = 6.0, 2.0 Hz, 1H), 7.77 (d, *J* = 7.2 Hz, 1H), 7.69 (t, *J* = 7.2 Hz, 1H), 7.61 (d, *J* = 8.0 Hz, 1H), 7.49 (t, *J* = 7.2 Hz, 1H), 7.44 – 7.35 (m, 2H), 7.14 (dd, *J* = 8.4, 7.2 Hz, 1H), 7.08 – 6.94 (m, 9H), 2.61 (p, *J* = 7.4 Hz, 4H), 1.21 (t, *J* = 7.6 Hz, 6H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 142.8, 142.2, 140.7, 140.3, 140.2, 138.6, 137.9, 137.1, 136.8, 135.7, 133.0, 132.2, 131.2, 131.14, 131.1, 131.0, 129.1, 128.9, 128.14, 128.1, 128.0, 127.8, 127.4, 127.0, 126.6, 123.4, 121.7, 121.2, 119.3, 116.1, 28.8, 28.7, 16.1, 15.7. HRMS (m/z, ESI): calcd for $C_{40}H_{31}^+$ (M+H)⁺ 511.2348, found 511.2340.



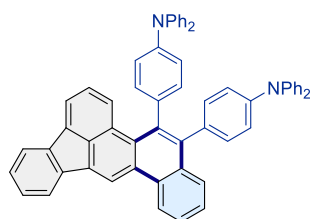
4,5-bis(4-(*tert*-butyl)phenyl)indeno[1,2,3-*hi*]chrysene (6d)

The general procedure A was followed. Yield: 89%, 50.8 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.38 (s, 1H), 9.02 (d, *J* = 8.4 Hz, 1H), 8.12 (d, *J* = 6.4 Hz, 1H), 7.87 (d, *J* = 6.8 Hz, 1H), 7.80 (d, *J* = 6.0 Hz, 1H), 7.75 – 7.67 (m, 2H), 7.51 (t, *J* = 7.6 Hz, 1H), 7.41 (p, *J* = 7.2 Hz, 2H), 7.21 – 7.08 (m, 6H), 7.01 (d, *J* = 7.6 Hz, 2H), 6.97 (d, *J* = 7.6 Hz, 2H), 1.29 (s, 9H), 1.28 (s, 9H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 149.6, 149.0, 140.8, 140.4, 140.1, 138.6, 138.1, 136.8, 136.7, 135.7, 133.0, 132.1, 131.2, 131.1, 130.9, 130.7, 129.1, 128.8, 128.2, 128.1, 128.07, 127.4, 127.0, 126.6, 124.9, 124.2, 123.4, 121.7, 121.2, 119.3, 116.1, 34.6, 34.5, 31.53, 31.49. HRMS (m/z, ESI): calcd for $C_{44}H_{39}^+$ (M+H)⁺ 567.2974, found 567.2960.



4,5-bis(4-(4-butylcyclohexyl)phenyl)indeno[1,2,3-*hi*]chrysene (6e)

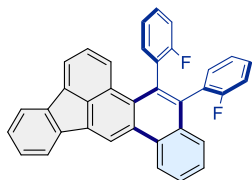
The general procedure A was followed. Yield: 69%, 50.3 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.39 (s, 1H), 9.03 (d, J = 8.4 Hz, 1H), 8.13 (d, J = 6.4 Hz, 1H), 7.88 (d, J = 6.8 Hz, 1H), 7.80 (d, J = 6.8 Hz, 1H), 7.72 (t, J = 7.6 Hz, 1H), 7.66 (d, J = 8.4 Hz, 1H), 7.51 (t, J = 7.6 Hz, 1H), 7.46 – 7.37 (m, 2H), 7.14 (t, J = 8.0 Hz, 1H), 7.06 – 6.91 (m, 9H), 2.41 (q, J = 11.6 Hz, 2H), 1.86 (d, J = 11.2 Hz, 8H), 1.47 – 1.35 (m, 4H), 1.35 – 1.19 (m, 14H), 1.06 (t, J = 11.6 Hz, 4H), 0.92 (t, J = 6.8 Hz, 6H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 146.4, 145.8, 140.8, 140.4, 140.36, 138.6, 138.1, 137.1, 136.8, 135.7, 133.0, 132.2, 131.2, 131.1, 131.05, 130.9, 129.2, 128.9, 128.2, 128.1, 128.09, 127.4, 127.0, 126.6, 126.56, 125.8, 123.4, 121.7, 121.2, 119.3, 116.1, 44.5, 44.46, 37.5, 37.3, 34.7, 34.5, 33.8, 29.4, 23.2, 14.3. HRMS (m/z , ESI): calcd for $\text{C}_{56}\text{H}_{59}^+$ ($\text{M}+\text{H}$) $^+$ 731.4539, found 731.4511.



4,4'-(indeno[1,2,3-*hi*]chrysene-4,5-diyl)bis(*N,N*-diphenylaniline) (6f)

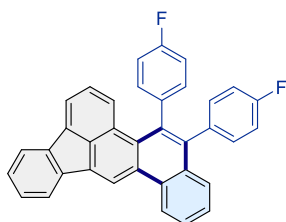
The general procedure A was followed. Yield: 73%, 57.5 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.39 (s, 1H), 9.04 (d, J = 8.8 Hz, 1H), 8.21 – 8.12 (m, 1H), 7.94 – 7.91 (m, 1H), 7.89 (d, J = 6.4 Hz, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.77 (t, J = 7.6 Hz, 1H), 7.60 (t, J = 7.6 Hz, 1H), 7.48 – 7.41 (m, 2H), 7.40 – 7.32 (m, 2H), 7.31 – 7.23 (m, 8H), 7.15 (t, J = 8.4 Hz, 8H), 7.07 – 6.99 (m, 12H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 148.04, 148.0, 146.5, 146.3, 140.8, 140.0, 138.6, 137.85, 137.8, 137.0, 135.9, 134.1, 133.1, 132.22, 132.2, 132.1, 131.3, 131.29, 129.5, 129.0, 128.8,

128.2, 128.1, 127.6, 126.9, 126.8, 126.77, 124.4, 124.3, 124.2, 123.6, 123.1, 122.9, 122.8, 121.7, 121.3, 119.4, 116.1. HRMS (m/z, ESI): calcd for $C_{60}H_{41}N_2^+$ (M+H)⁺ 789.3191, found 789.3170.



4,5-bis(2-fluorophenyl)indeno[1,2,3-*hi*]chrysene (6g)

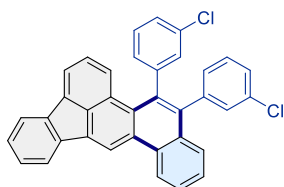
The general procedure A was followed. Yield: 45%, 22.2 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.32 (s, 1H), 8.98 (d, *J* = 8.4 Hz, 1H), 8.05 (d, *J* = 6.4 Hz, 1H), 7.84 – 7.79 (m, 1H), 7.77 (d, *J* = 6.8 Hz, 1H), 7.73 – 7.65 (m, 1H), 7.53 – 7.41 (m, 2H), 7.40 – 7.31 (m, 2H), 7.31 – 7.13 (m, 5H), 7.06 – 6.96 (m, 3H), 6.95 – 6.86 (m, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.9 (d, *J*_{C-F} = 245.5 Hz), 159.8 (d, *J*_{C-F} = 245.4 Hz), 140.7, 138.5, 137.2, 136.3, 134.9, 132.9, 132.2, 131.9, 131.6, 131.5, 131.0, 130.81, 129.8 (d, *J*_{C-F} = 34.7 Hz), 129.7 (d, *J*_{C-F} = 34.8 Hz), 128.5 (d, *J*_{C-F} = 56.6 Hz), 128.3, 127.9, 127.6, 127.4 (d, *J*_{C-F} = 17.9 Hz), 127.24, 127.22, 127.1, 126.2, 124.6 (d, *J*_{C-F} = 3.4 Hz), 123.9 (d, *J*_{C-F} = 3.7 Hz), 123.8, 121.8, 121.3, 119.6, 116.1, 115.6 (d, *J*_{C-F} = 22.0 Hz), 115.1 (d, *J*_{C-F} = 22.1 Hz). HRMS (m/z, ESI): calcd for $C_{36}H_{21}F_2^+$ (M+H)⁺ 491.1533, found 491.1545.



4,5-bis(4-fluorophenyl)indeno[1,2,3-*hi*]chrysene (6h)

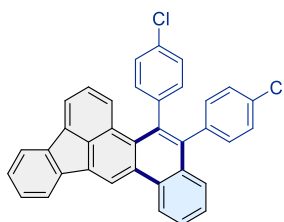
The general procedure A was followed. Yield: 90%, 44.5 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.36 (s, 1H), 9.02 (d, *J* = 8.4 Hz, 1H), 8.12 (d, *J* = 6.0 Hz, 1H), 7.89 (d, *J* = 6.8 Hz, 1H), 7.83 (d, *J* = 7.2 Hz, 1H), 7.76 (t, *J* = 6.4 Hz, 1H), 7.59 – 7.50 (m, 2H), 7.48 – 7.39 (m, 2H), 7.22 (t, *J* = 7.6 Hz, 1H), 7.13 – 7.01 (m, 4H), 7.00 – 6.88 (m, 5H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 161.9 (d, *J*_{C-F} = 247.8 Hz), 161.6 (d, *J*_{C-F} = 246.9 Hz), 140.7, 139.2, 138.9 (d, *J*_{C-F} = 3.7 Hz), 138.5, 137.1, 137.0, 136.2, 135.6

(d, $J_{C-F} = 3.4$ Hz), 133.0, 132.7, 132.64, 132.56, 132.0, 131.5, 131.3, 128.6 (d, $J_{C-F} = 39.9$ Hz), 128.1 (d, $J_{C-F} = 54.3$ Hz), 127.6, 127.5, 127.4, 127.0, 126.9, 123.6, 121.8, 121.4, 119.5, 116.0, 115.6 (d, $J_{C-F} = 21.1$ Hz), 114.9 (d, $J_{C-F} = 21.4$ Hz). HRMS (m/z , ESI): calcd for $C_{36}H_{21}F_2^+$ ($M+H$) $^+$ 491.1533, found 491.1520.



4,5-bis(3-chlorophenyl)indeno[1,2,3-*hi*]chrysene (6i)

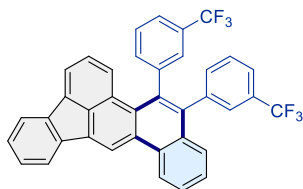
The general procedure A was followed (note: the products were isolated as an inseparable mixture of diastereoisomers). Yield: 97%, 51.0 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 9.25 (s, 1H), 8.93 (d, $J = 8.4$ Hz, 1H), 8.09 – 7.97 (m, 1H), 7.87 – 7.78 (m, 1H), 7.75 (d, $J = 6.8$ Hz, 1H), 7.68 (t, $J = 6.8$ Hz, 1H), 7.51 – 7.42 (m, 2H), 7.40 – 7.31 (m, 2H), 7.19 – 7.01 (m, 7H), 7.00 – 6.84 (m, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 144.5, 141.3, 140.7, 138.5, 138.4, 137.1, 136.3, 136.2, 134.5, 134.4, 133.9, 133.7, 133.0, 131.6, 131.4, 131.34, 131.26, 131.2, 131.1, 129.8, 129.7, 129.5, 129.4, 129.3, 129.25, 129.1, 129.0, 128.6, 128.4, 127.8, 127.6, 127.5, 127.4, 127.35, 127.2, 127.1, 127.0, 123.6, 121.8, 121.4, 119.6, 115.9. HRMS (m/z , ESI): calcd for $C_{36}H_{21}Cl_2^+$ ($M+H$) $^+$ 523.0942, found 523.0927.



4,5-bis(4-chlorophenyl)indeno[1,2,3-*hi*]chrysene (6j)

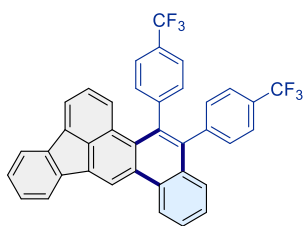
The general procedure A was followed. Yield: 99%, 52.0 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 9.25 (s, 1H), 8.94 (d, $J = 8.4$ Hz, 1H), 8.05 (dd, $J = 4.8, 2.0$ Hz, 1H), 7.90 – 7.81 (m, 1H), 7.78 (d, $J = 6.8$ Hz, 1H), 7.72 (t, $J = 7.2$ Hz, 1H), 7.51 (t, $J = 8.0$ Hz, 1H), 7.48 – 7.36 (m, 3H), 7.29 – 7.13 (m, 5H), 7.05 – 6.92 (m, 4H), 6.89 (d, $J = 8.8$ Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 141.3, 140.6, 138.6, 138.4, 138.0, 137.1, 136.4, 136.2, 133.1, 132.9, 132.7, 132.4, 132.36, 131.6, 131.5, 131.3, 128.8,

128.6, 128.4, 128.1, 128.0, 127.7, 127.6, 127.4, 127.0, 126.9, 123.6, 121.8, 121.3, 119.5, 115.8. HRMS (m/z, ESI): calcd for $C_{36}H_{21}Cl_2^+$ (M+H)⁺ 523.0942, found 523.0931.



4,5-bis(3-(trifluoromethyl)phenyl)indeno[1,2,3-*hi*]chrysene (6k)

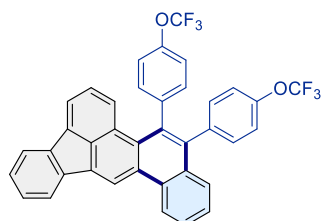
The general procedure A was followed (note: the products were isolated as an inseparable mixture of diastereoisomers, *d.r.* = 1:1). Yield: 91%, 53.9 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.21 (d, *J* = 3.2 Hz, 1H), 8.92 (d, *J* = 8.4 Hz, 1H), 8.05 – 7.95 (m, 1H), 7.84 – 7.75 (m, 1H), 7.75 – 7.65 (m, 2H), 7.48 (t, *J* = 8.0 Hz, 1H), 7.45 – 7.19 (m, 9H), 7.18 – 7.01 (m, 3H), 6.81 (d, *J* = 4.0 Hz, 0.52H), 6.78 (d, *J* = 4.4 Hz, 0.49H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 143.5, 143.46, 140.6, 140.3, 140.2, 138.4, 138.3, 137.1, 136.3, 134.6, 134.5, 134.2, 132.9, 131.6, 131.56, 131.33, 131.3, 131.2, 131.17, 128.9, 128.4, 128.31, 128.28, 128.1 (q, *J*_{C-F} = 3.6 Hz), 127.9 (q, *J*_{C-F} = 3.6 Hz), 127.6, 127.5, 127.44, 127.4, 127.2, 127.1, 127.06, 127.0, 125.5, 125.4, 124.1 (q, *J*_{C-F} = 273.4 Hz), 124.0 (q, *J*_{C-F} = 273.7 Hz), 123.8, 123.7, 123.63, 123.6, 121.8, 121.3, 119.5, 115.7. HRMS (m/z, ESI): calcd for $C_{38}H_{21}F_6^+$ (M+H)⁺ 591.1469, found 591.1501.



4,5-bis(4-(trifluoromethyl)phenyl)indeno[1,2,3-*hi*]chrysene (6l)

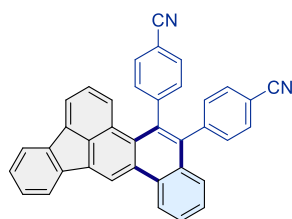
The general procedure A was followed. Yield: 93%, 55.0 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.29 (s, 1H), 8.99 (d, *J* = 8.4 Hz, 1H), 8.12 – 8.04 (m, 1H), 7.92 – 7.83 (m, 1H), 7.83 – 7.72 (m, 2H), 7.55 (t, *J* = 8.0 Hz, 1H), 7.49 (t, *J* = 7.6 Hz, 4H), 7.45 – 7.37 (m, 3H), 7.22 – 7.09 (m, 5H), 6.79 (d, *J* = 8.8 Hz, 1H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 146.5, 143.3, 140.7, 138.3, 137.2, 136.5, 136.1, 133.0, 131.7,

131.5, 131.46, 131.4, 131.2, 129.5 (q, $J_{C-F} = 32.7$ Hz), 129.2 (q, $J_{C-F} = 32.6$ Hz), 128.5, 128.4, 127.7, 127.6, 127.58, 127.5, 127.3, 127.2, 127.1, 125.44 (q, $J_{C-F} = 3.6$ Hz), 124.82 (q, $J_{C-F} = 3.6$ Hz), 124.23 (q, $J_{C-F} = 273.3$ Hz), 124.2 (q, $J_{C-F} = 273.2$ Hz), 123.7, 121.9, 121.4, 119.7, 115.8. HRMS (m/z, ESI): calcd for $C_{38}H_{21}F_6^+$ (M+H)⁺ 591.1469, found 591.1454.



4,5-bis(4-(trifluoromethoxy)phenyl)indeno[1,2,3-*hi*]chrysene (6m)

The general procedure A was followed. Yield: 81%, 50.4 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.34 (s, 1H), 9.03 (d, $J = 8.4$ Hz, 1H), 8.18 – 8.05 (m, 1H), 7.94 – 7.86 (m, 1H), 7.82 (d, $J = 6.8$ Hz, 1H), 7.77 (t, $J = 7.2$ Hz, 1H), 7.61 – 7.51 (m, 2H), 7.48 – 7.39 (m, 2H), 7.19 (t, $J = 8.0$ Hz, 1H), 7.15 – 7.03 (m, 8H), 6.93 (d, $J = 8.8$ Hz, 1H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 148.3, 148.1, 141.7, 140.7, 138.7, 138.4, 138.2, 137.2, 136.5, 136.4, 133.0, 132.5, 132.4, 131.6, 131.5, 131.4, 128.6, 128.5, 128.0, 127.74, 127.7, 127.4, 127.3, 127.2, 127.1, 123.7, 121.8, 121.4, 121.2, 120.6 (q, $J_{C-F} = 258.1$ Hz), 120.4, 119.6, 115.9. HRMS (m/z, ESI): calcd for $C_{38}H_{21}F_6O_2^+$ (M+H)⁺ 623.1367, found 623.1337.

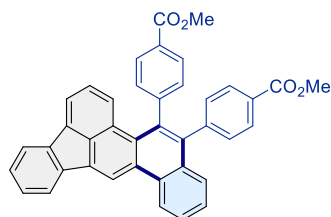


4,4'-(indeno[1,2,3-*hi*]chrysene-4,5-diyl)dibenzonitrile (6n)

The general procedure A was followed. Yield: 97%, 49.0 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.26 (s, 1H), 9.00 (d, $J = 8.4$ Hz, 1H), 8.13 – 8.04 (m, 1H), 7.94 – 7.85 (m, 1H), 7.84 – 7.76 (m, 2H), 7.60 – 7.48 (m, 5H), 7.48 – 7.41 (m, 2H), 7.34 (d, $J = 8.4$ Hz, 1H), 7.21 – 7.07 (m, 5H), 6.71 (d, $J = 8.4$ Hz, 1H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 147.5, 144.3, 140.6, 138.2, 137.6, 137.4, 136.8, 135.5, 132.9, 132.3, 132.26, 131.9, 131.85, 131.8, 131.4, 130.8, 128.7, 128.1, 127.9, 127.7, 127.6, 127.4,

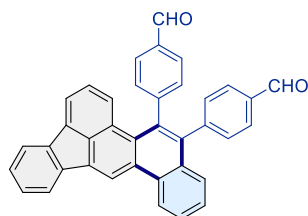
127.3, 127.0, 126.8, 123.8, 121.9, 121.5, 119.8, 118.7, 118.6, 115.7, 111.4, 111.2.

HRMS (m/z, ESI): calcd for $C_{38}H_{21}N_2^+$ (M+H) $^+$ 505.1699, found 505.1673.



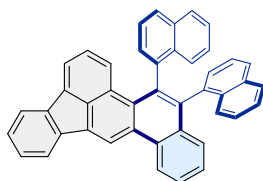
dimethyl 4,4'-(indeno[1,2,3-*hi*]chrysene-4,5-diyl)dibenzoate (6o)

The general procedure A was followed. Yield: 70%, 40.0 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 9.43 (s, 1H), 9.09 (d, J = 8.4 Hz, 1H), 8.21 – 8.14 (m, 1H), 8.00 – 7.90 (m, 5H), 7.86 (d, J = 6.8 Hz, 1H), 7.80 (t, J = 7.2 Hz, 1H), 7.55 (t, J = 7.6 Hz, 1H), 7.52 – 7.42 (m, 3H), 7.30 (d, J = 8.0 Hz, 2H), 7.22 (d, J = 8.2 Hz, 2H), 7.18 (d, J = 8.4 Hz, 1H), 6.93 (d, J = 8.4 Hz, 1H), 3.95 (s, 3H), 3.93 (s, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 167.1, 147.8, 144.6, 140.8, 137.2, 136.5, 131.7, 131.4, 131.3, 131.28, 129.8, 129.2, 129.1, 128.9, 128.7, 128.6, 128.5, 127.8, 127.7, 127.6, 127.4, 127.3, 127.1, 123.7, 121.9, 121.4, 119.7, 118.6, 118.0, 115.9, 52.3, 52.3. HRMS (m/z, ESI): calcd for $C_{40}H_{27}O_4^+$ (M+H) $^+$ 571.1831, found 571.1847.



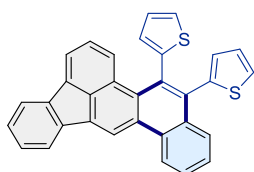
4,4'-(indeno[1,2,3-*hi*]chrysene-4,5-diyl)dibenzaldehyde (6p)

The general procedure A was followed. Yield: 89%, 45.6 mg. 1H NMR (400 MHz, Chloroform-*d*) δ 9.99 (s, 1H), 9.97 (s, 1H), 9.31 (s, 1H), 9.01 (d, J = 8.4 Hz, 1H), 8.13 – 8.07 (m, 1H), 7.90 – 7.85 (m, 1H), 7.82 – 7.77 (m, 2H), 7.74 (dd, J = 8.0, 5.2 Hz, 4H), 7.54 (t, J = 8.0 Hz, 1H), 7.47 – 7.39 (m, 3H), 7.28 (d, J = 8.0 Hz, 2H), 7.23 (d, J = 8.0 Hz, 2H), 7.13 (dd, J = 8.4, 7.2 Hz, 1H), 6.83 (d, J = 8.4 Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 192.04, 191.99, 149.3, 146.1, 140.6, 138.3, 138.2, 137.2, 136.6, 136.1, 135.0, 134.9, 133.0, 131.9, 131.87, 131.7, 131.4, 131.0, 129.8, 129.2, 128.6, 128.4, 127.8, 127.5, 127.4, 127.2, 127.1, 123.8, 121.9, 121.4, 119.7, 115.8. HRMS (m/z, ESI): calcd for $C_{38}H_{23}O_2^+$ (M+H) $^+$ 511.1620, found 511.1632.



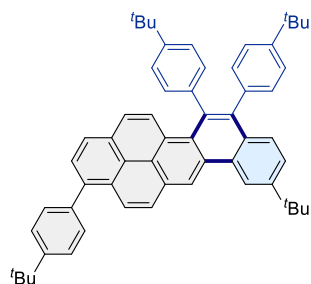
4,5-di(naphthalen-1-yl)indeno[1,2,3-*hi*]chrysene (6q)

The general procedure A was followed. Yield: 69%, 38.5 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.57 (s, 1H), 9.19 (d, J = 8.8 Hz, 1H), 8.20 (d, J = 7.2 Hz, 1H), 7.87 (d, J = 6.8 Hz, 1H), 7.83 – 7.70 (m, 4H), 7.62 (d, J = 8.4 Hz, 1H), 7.60 – 7.53 (m, 2H), 7.51 (d, J = 9.2 Hz, 1H), 7.49 – 7.38 (m, 5H), 7.37 – 7.26 (m, 3H), 7.15 (d, J = 7.2 Hz, 1H), 7.01 – 6.85 (m, 4H), 6.74 (d, J = 8.8 Hz, 1H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 140.8, 140.5, 139.1, 138.6, 137.3, 136.9, 136.8, 136.2, 133.4, 133.2, 133.1, 132.9, 132.5, 131.6, 130.1, 129.6, 129.2, 128.4, 128.3, 128.2, 128.1, 127.7, 127.66, 127.6, 127.3, 127.1, 127.05, 126.95, 126.7, 126.5, 126.1, 126.0, 125.8, 125.6, 125.4, 125.3, 124.8, 124.5, 123.7, 121.8, 121.3, 119.5, 116.3, 116.2. HRMS (m/z , ESI): calcd for $\text{C}_{44}\text{H}_{27}^+$ ($\text{M}+\text{H}$) $^+$ 555.2035, found 555.2013.



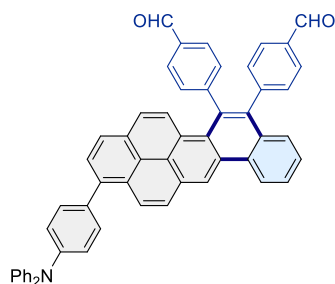
2,2'-(indeno[1,2,3-*hi*]chrysene-4,5-diyl)dithiophene (6r)

The general procedure A was followed. Yield: 85%, 39.8 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.33 (s, 1H), 9.00 (d, J = 8.4 Hz, 1H), 8.17 – 8.07 (m, 1H), 7.93 – 7.87 (m, 1H), 7.86 (d, J = 6.8 Hz, 1H), 7.83 – 7.72 (m, 2H), 7.58 (t, J = 8.0 Hz, 1H), 7.47 – 7.41 (m, 2H), 7.38 (dd, J = 10.4, 5.2 Hz, 2H), 7.31 (t, J = 8.4 Hz, 1H), 7.11 (d, J = 8.8 Hz, 1H), 7.04 (q, J = 4.0 Hz, 2H), 6.94 (dd, J = 7.2, 3.2 Hz, 2H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 143.9, 140.9, 139.9, 138.4, 137.0, 136.7, 134.7, 133.0, 132.7, 132.3, 131.9, 131.6, 129.7, 129.23, 129.2, 128.8, 128.5, 128.1, 127.64, 127.6, 127.4, 127.3, 127.2, 127.1, 126.9, 126.4, 126.3, 123.5, 121.8, 121.3, 119.7, 115.7. HRMS (m/z , ESI): calcd for $\text{C}_{32}\text{H}_{19}\text{S}_2^+$ ($\text{M}+\text{H}$) $^+$ 467.0850, found 467.0842.



8-(*tert*-butyl)-3,11,12-tris(4-(*tert*-butyl)phenyl)benzo[*pqr*]picene (6s)

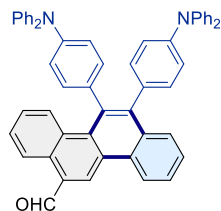
The general procedure A was followed. Yield: 46%, 34.6 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.56 (s, 1H), 9.16 (s, 1H), 8.29 – 8.21 (m, 2H), 8.17 (d, J = 9.6 Hz, 1H), 8.10 (d, J = 8.0 Hz, 1H), 7.95 (d, J = 8.0 Hz, 1H), 7.77 (d, J = 8.8 Hz, 1H), 7.70 (d, J = 8.4 Hz, 1H), 7.67 – 7.56 (m, 5H), 7.31 – 7.21 (m, 2H), 7.16 (d, J = 8.0 Hz, 2H), 7.05 (d, J = 8.0 Hz, 2H), 6.98 (d, J = 8.0 Hz, 2H), 1.61 (s, 9H), 1.49 (s, 9H), 1.35 (s, 9H), 1.33 (s, 9H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 150.2, 149.4, 148.94, 148.9, 141.2, 139.4, 138.6, 137.5, 137.3, 136.5, 131.0, 130.9, 130.5, 129.95, 129.9, 129.8, 129.5, 129.3, 128.9, 128.3, 128.2, 128.0, 127.9, 126.7, 126.0, 125.5, 125.4, 125.3, 125.2, 124.7, 124.2, 119.5, 119.2, 35.5, 34.8, 34.6, 34.5, 31.8, 31.7, 31.6, 31.5. HRMS (m/z , ESI): calcd for $\text{C}_{58}\text{H}_{59}^+$ ($\text{M}+\text{H}$) $^+$ 755.4539, found 755.4521.



4,4'-(3-(4-(diphenylamino)phenyl)benzo[*pqr*]picene-11,12-diyl)dibenzaldehyde (6t)

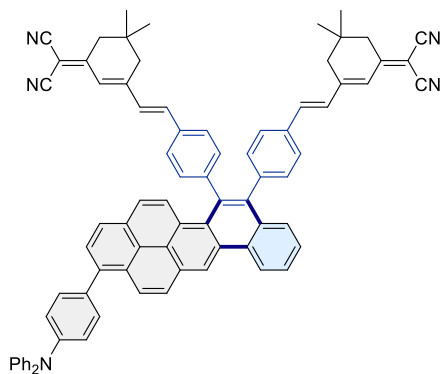
The general procedure A was followed. Yield: 72%, 54.1 mg. ^1H NMR (400 MHz, Chloroform-*d*) δ 9.93 (s, 1H), 9.89 (s, 1H), 9.40 (s, 1H), 9.06 (d, J = 8.4 Hz, 1H), 8.23 (d, J = 9.2 Hz, 1H), 8.10 (d, J = 9.6 Hz, 1H), 8.00 (d, J = 8.0 Hz, 1H), 7.89 (d, J = 8.0 Hz, 1H), 7.77 – 7.65 (m, 4H), 7.59 (d, J = 7.6 Hz, 2H), 7.51 (t, J = 9.6 Hz, 2H), 7.41 (d, J = 8.4 Hz, 3H), 7.25 (t, J = 8.0 Hz, 4H), 7.22 – 7.11 (m, 10H), 7.00 (t, J = 7.2 Hz, 2H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 192.1, 192.0, 150.3, 147.9, 147.4, 146.7, 138.0, 137.6, 135.4, 135.0, 134.9, 134.5, 132.1, 132.0, 131.5, 130.8, 130.6, 130.4,

129.7, 129.6, 129.5, 129.3, 128.72, 128.69, 128.6, 128.0, 127.6, 127.55, 127.5, 127.1, 126.7, 125.6, 125.3, 125.1, 125.0, 124.8, 123.8, 123.3, 119.5. HRMS (*m/z*, ESI): calcd for $C_{56}H_{36}NO_2^+$ (*M*+*H*)⁺ 754.2668, found 754.2647.



11,12-bis(4-(diphenylamino)phenyl)chrysene-6-carbaldehyde (6u)

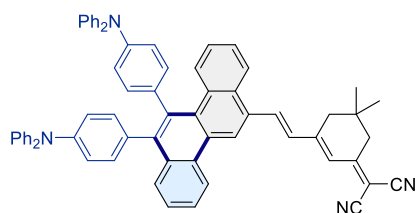
The general procedure A was followed. Yield: 64%, 47.9 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 10.47 (s, 1H), 9.24 (d, *J* = 8.0 Hz, 1H), 9.12 (s, 1H), 8.76 (d, *J* = 8.4 Hz, 1H), 7.92 (d, *J* = 8.8 Hz, 1H), 7.74 (d, *J* = 8.0 Hz, 1H), 7.66 (t, *J* = 7.2 Hz, 1H), 7.50 (q, *J* = 7.6 Hz, 2H), 7.22 – 7.10 (m, 9H), 7.08 – 6.98 (m, 8H), 6.92 (t, *J* = 8.0 Hz, 6H), 6.88 – 6.80 (m, 4H), 6.75 (d, *J* = 8.0 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 193.4, 147.9, 147.8, 146.5, 146.3, 143.4, 137.5, 136.7, 134.3, 133.6, 132.1, 132.0, 131.9, 131.8, 130.32, 130.3, 130.1, 129.5, 129.4, 129.3, 128.2, 127.9, 127.6, 127.3, 125.2, 124.9, 124.5, 124.2, 124.0, 123.0, 122.94, 122.87. HRMS (*m/z*, ESI): calcd for $C_{55}H_{39}N_2O^+$ (*M*+*H*)⁺ 743.2984, found 743.2968.



2,2'-(((1*E*,1'*E*)-((3-(4-(diphenylamino)phenyl)benzo[*qqr*]picene-11,12-diyl)bis(4,1-phenylene))bis(ethene-2,1-diyl))bis(5,5-dimethylcyclohex-2-en-3-yl-1-ylidene))di malononitrile (6v)

The general procedure A was followed (note: 0.05 mmol scale reaction was carried out). Yield: 42%, 22.9 mg. ¹H NMR (400 MHz, Chloroform-*d*) δ 9.38 (s, 1H), 9.04 (d, *J* = 8.4 Hz, 1H), 8.22 (d, *J* = 9.2 Hz, 1H), 8.08 (d, *J* = 9.6 Hz, 1H), 7.94 (d, *J* = 8.0 Hz, 1H), 7.87 (d, *J* = 8.0 Hz, 1H), 7.81 (d, *J* = 9.6 Hz, 1H), 7.76 – 7.68 (m, 1H), 7.56 –

7.48 (m, 3H), 7.40 (d, $J = 8.4$ Hz, 2H), 7.34 (d, $J = 8.0$ Hz, 2H), 7.29 – 7.20 (m, 5H), 7.21 – 7.11 (m, 7H), 7.10 (d, $J = 8.0$ Hz, 2H), 7.05 (d, $J = 8.0$ Hz, 2H), 6.99 (t, $J = 7.2$ Hz, 2H), 6.86 (d, $J = 14.8$ Hz, 2H), 6.82 (d, $J = 13.6$ Hz, 2H), 6.61 (s, 1H), 6.56 (s, 1H), 2.37 (s, 2H), 2.29 (s, 2H), 2.21 (s, 2H), 2.16 (s, 2H), 0.91 (s, 6H), 0.86 (s, 6H). ^{13}C NMR (101 MHz, Chloroform- d) δ 169.2, 169.1, 153.7, 153.6, 147.8, 147.3, 145.7, 141.9, 138.3, 137.8, 136.7, 135.9, 134.9, 134.1, 133.8, 132.1, 132.0, 131.5, 131.4, 130.3, 129.6, 129.55, 129.5, 129.45, 129.2, 129.1, 128.9, 128.7, 128.5, 128.2, 127.6, 127.32, 127.3, 127.1, 126.5, 125.6, 125.5, 125.4, 125.1, 125.0, 124.8, 123.7, 123.6, 123.5, 123.48, 123.3, 123.2, 119.6, 113.51, 113.49, 112.8, 112.7, 78.8, 78.7, 42.9, 42.8, 39.2, 39.1, 32.0, 31.9, 28.1, 28.0. HRMS (m/z , ESI): calcd for $\text{C}_{80}\text{H}_{60}\text{N}_5^+$ ($\text{M}+\text{H}$) $^+$ 1090.4770, found 1090.4719.



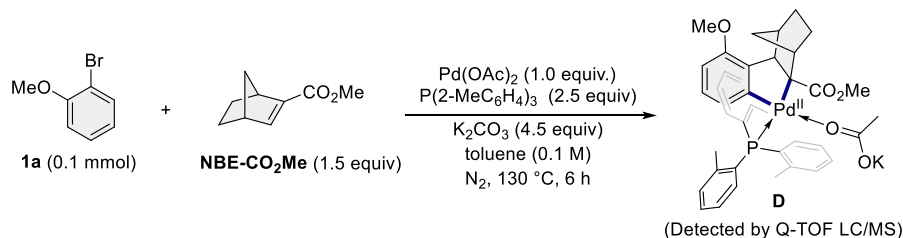
(*E*)-2-(3-(2-(11,12-bis(4-(diphenylamino)phenyl)chrysen-6-yl)vinyl)-5,5-dimethylcyclohex-2-en-1-ylidene)malononitrile (6w)

The general procedure A was followed (note: 0.05 mmol scale reaction was carried out). Yield: 40%, 18.0 mg. ^1H NMR (400 MHz, Chloroform- d) δ 8.98 (s, 1H), 8.84 (d, $J = 8.4$ Hz, 1H), 8.11 (d, $J = 8.0$ Hz, 1H), 7.96 (d, $J = 8.8$ Hz, 1H), 7.88 (d, $J = 15.6$ Hz, 1H), 7.74 (d, $J = 8.4$ Hz, 1H), 7.70 (t, $J = 7.2$ Hz, 1H), 7.52 (t, $J = 7.6$ Hz, 1H), 7.46 (t, $J = 7.2$ Hz, 1H), 7.24 – 7.13 (m, 10H), 7.04 (t, $J = 8.4$ Hz, 8H), 6.97 – 6.91 (m, 7H), 6.90 – 6.85 (m, 4H), 6.82 (d, $J = 8.4$ Hz, 2H), 2.56 (s, 2H), 2.55 (s, 2H), 1.08 (s, 6H). ^{13}C NMR (101 MHz, Chloroform- d) δ 169.4, 153.9, 148.0, 147.9, 146.4, 146.1, 140.8, 138.1, 136.6, 134.6, 134.0, 132.3, 132.2, 132.0, 131.7, 131.5, 130.1, 129.9, 129.45, 129.43, 129.2, 129.0, 128.1, 127.2, 126.2, 124.7, 124.4, 124.3, 124.2, 124.1, 123.4, 123.2, 123.1, 122.9, 122.8, 120.8, 113.6, 112.9, 79.2, 43.2, 39.5, 32.3, 28.3. HRMS (m/z , ESI): calcd for $\text{C}_{67}\text{H}_{51}\text{N}_4^+$ ($\text{M}+\text{H}$) $^+$ 911.4035, found 911.4030.

3.4 Mechanistic Studies

Control experiments:

(a) Reaction of **1a** with NBE-CO₂Me in the condition of quantitative Pd(OAc)₂:



The oven-dried Schlenk tube (10 mL) equipped with a stirring bar was charged with substrates **1a** (0.1 mmol, 1.0 equiv), Pd(OAc)₂ (1.0 equiv), P(2-MeC₆H₄)₃ (2.5 equiv) and K₂CO₃ (4.5 equiv) in the air. Followed by the mediator NBE-CO₂Me (1.5 equiv) and the anhydrous toluene (1.0 mL) were added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N₂ for three times. Finally, the reaction mixture was stirred at 130 °C for 6 h and then cooled to room temperature. Upon completion of the reaction, the mixture was analyzed by the Agilent 6546 Q-TOF LC/MS system detection and intermediate **D** were detected.

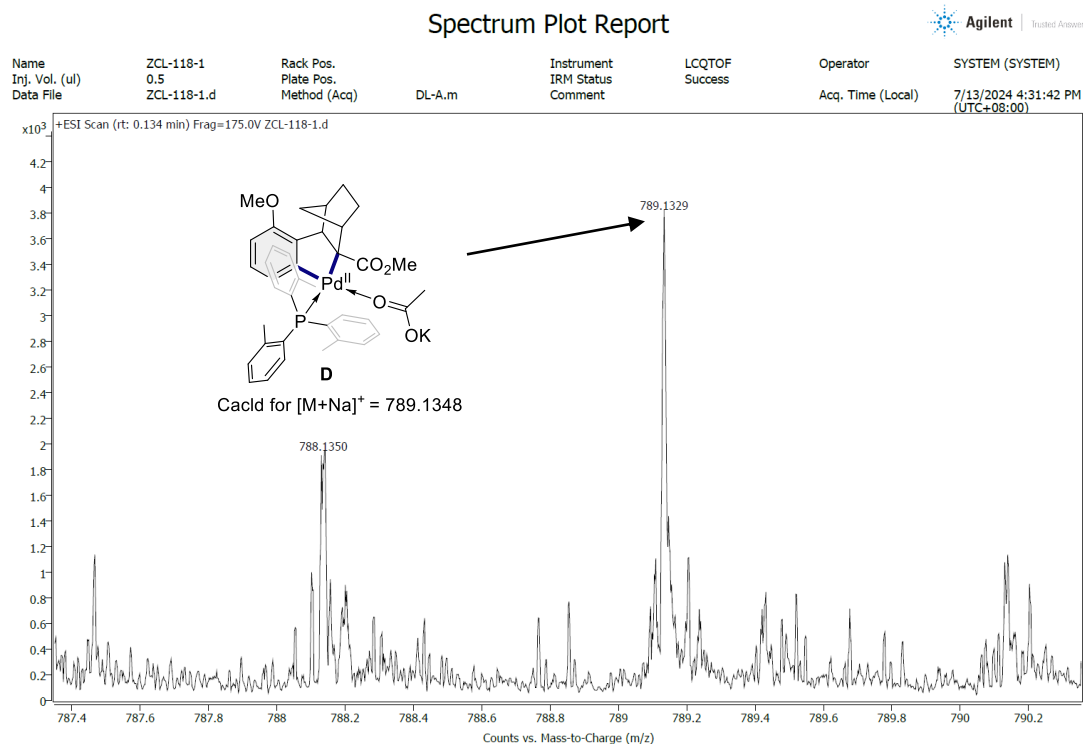
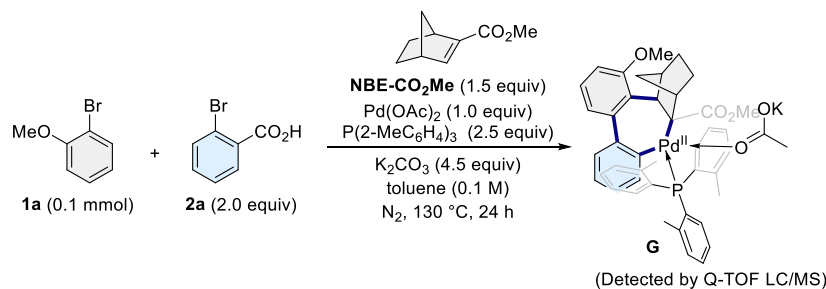


Fig. S4. MS spectrum of intermediate **D**.

(b) Reaction of **1a** with **NBE-CO₂Me** and **2a** in the condition of quantitative Pd(OAc)₂:



The oven-dried Schlenk tube (10 mL) equipped with a stirring bar was charged with substrates **1a** (0.1 mmol, 1.0 equiv), **2a** (2.0 equiv), Pd(OAc)₂ (1.0 equiv), P(2-MeC₆H₄)₃ (2.5 equiv) and K₂CO₃ (4.5 equiv) in the air. Followed by the mediator NBE-CO₂Me (1.5 equiv) and the anhydrous toluene (1.0 mL) were added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N₂ for three times. Finally, the reaction mixture was stirred at 130 °C for 24 h and then cooled to room temperature. Upon completion of the reaction, the mixture was analyzed by the Agilent 6546 Q-TOF LC/MS system detection and intermediate **G** were detected.

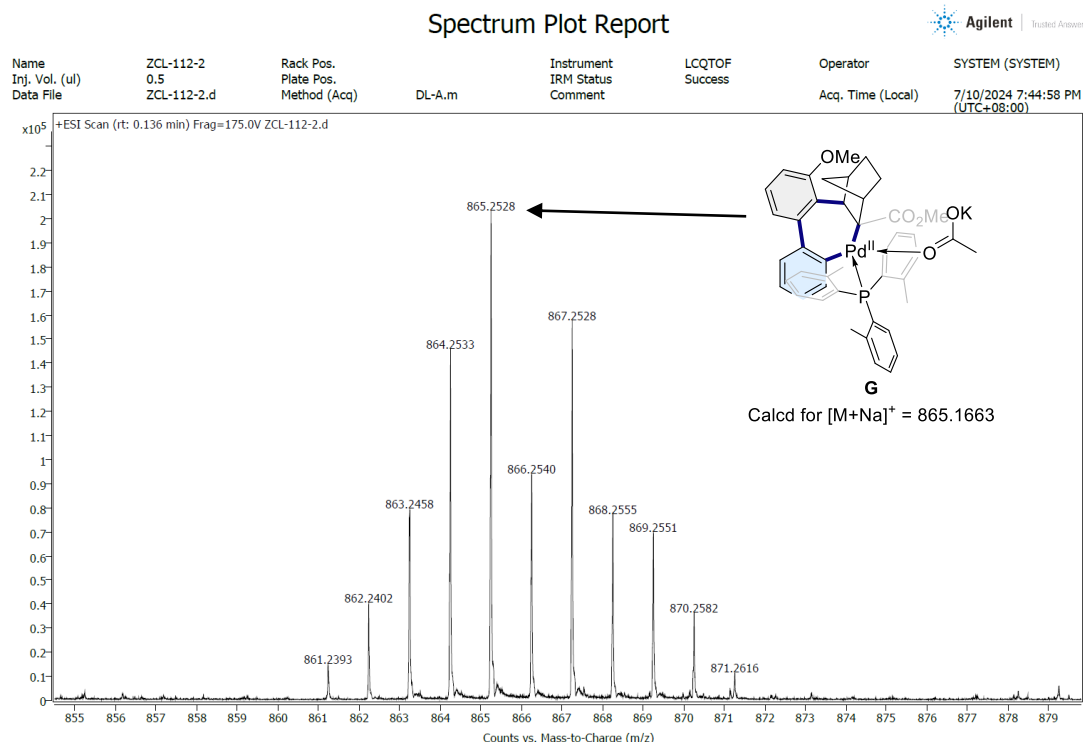


Fig. S5. MS spectrum of intermediate **G**.

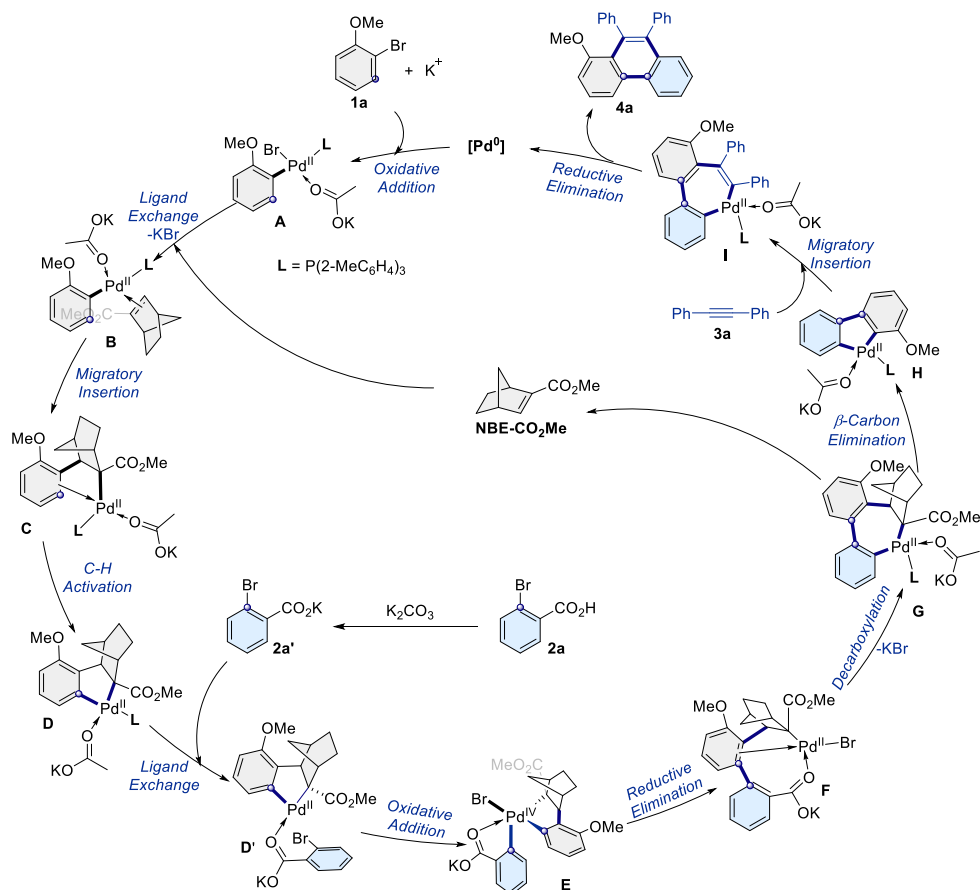
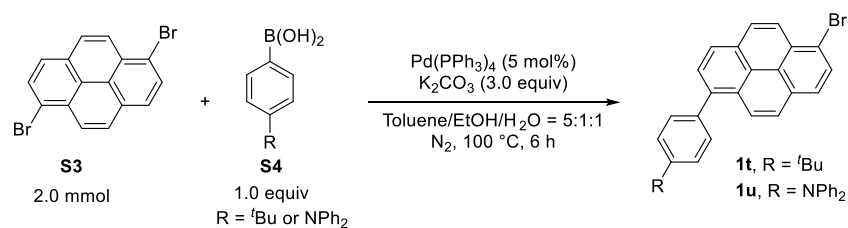


Fig. S6. Proposed reaction mechanism.

3.5 General procedures for the synthesis of substrates

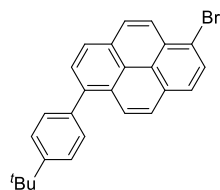
(a) The synthesis of substrates 1t and 1u:



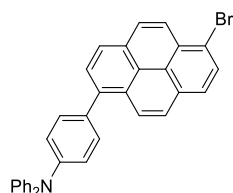
The oven-dried Schlenk tube (100 mL) equipped with a stirring bar was charged with 1,6-dibromopyrene (**S3**, 2.0 mmol, 1.0 equiv), 4-tert-butylphenylboronic acid or 4-(diphenylamino)phenyl boronic acid (**S4** 1.0 equiv), Pd(PPh₃)₄ (5 mol%), and K₂CO₃ (4.5 equiv) in the air. Followed by the toluene/EtOH/H₂O (5/1/1, 10 mL) were added via syringe under nitrogen flow. Then the tube was evacuated and back-filled with N₂ for three times. Finally, the reaction mixture was stirred at 100 °C for 6 h and then cooled to room temperature. Upon completion of the reaction, the mixture was

quenched by 2N HCl aqueous solution and ethyl acetate (EA). The organic phases were separated and the aqueous phase was extracted with EA three times. The combined organic phases were washed with brine, dried with anhydrous Na₂SO₄ and concentrated *in vacuo*. The resulting residue was purified via silica gel column chromatography using PE/EA (200/1 – 20/1) as the eluent.

Characterization of substrates 1t and 1u:



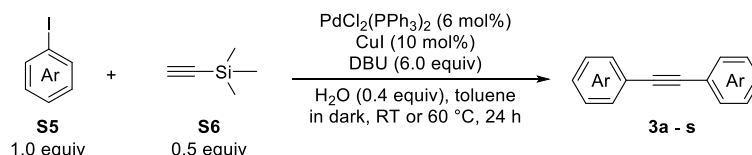
1-bromo-6-(4-(*tert*-butyl)phenyl)pyrene (1t): ¹H NMR (400 MHz, Chloroform-*d*) δ 8.45 (d, *J* = 9.2 Hz, 1H), 8.30 – 8.21 (m, 3H), 8.19 (d, *J* = 9.2 Hz, 1H), 8.02 (d, *J* = 8.0 Hz, 1H), 8.00 – 7.92 (m, 2H), 7.64 – 7.52 (m, 4H), 1.46 (s, 9H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 150.5, 138.7, 138.0, 130.6, 130.4, 130.35, 130.3, 130.1, 129.2, 128.7, 128.5, 127.1, 126.3, 126.0, 125.9, 125.5, 125.4, 125.3, 124.5, 120.0, 34.8, 31.6. HRMS (*m/z*, ESI): calcd for C₂₆H₂₄Br⁺ (*M*+H)⁺ 415.0983, found 415.0795.



4-(6-bromopyren-1-yl)-*N,N*-diphenylaniline (1u): ¹H NMR (400 MHz, Chloroform-*d*) δ 8.44 (d, *J* = 9.2 Hz, 1H), 8.31 (d, *J* = 9.2 Hz, 1H), 8.24 (dd, *J* = 7.6, 6.0 Hz, 2H), 8.18 (d, *J* = 9.2 Hz, 1H), 8.03 (d, *J* = 8.0 Hz, 1H), 7.99 (d, *J* = 8.8 Hz, 2H), 7.50 (d, *J* = 8.4 Hz, 2H), 7.34 (t, *J* = 8.0 Hz, 4H), 7.30 – 7.18 (m, 6H), 7.09 (t, *J* = 7.2 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 147.8, 147.4, 138.4, 134.7, 131.5, 130.6, 130.31, 130.29, 130.1, 129.5, 129.2, 128.6, 128.4, 127.1, 126.3, 125.9, 125.4, 125.36, 124.8, 124.6, 123.3, 120.0. HRMS (*m/z*, ESI): calcd for C₃₄H₂₅BrN⁺ (*M*+H)⁺ 526.1092, found 526.0902.

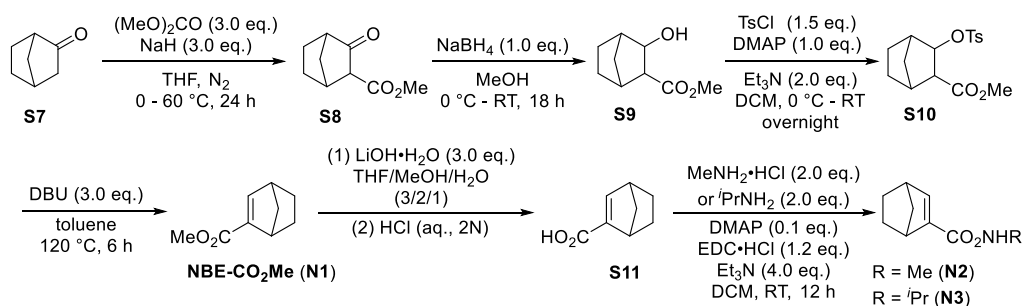
(b) The synthesis of symmetrical 1,2-diarylalkynes:

All the symmetrical 1,2-diarylalkynes were prepared according to the reported literatures^[S2].



(c) The synthesis of norbornene mediators:

The structurally modified norbornenes were synthesized following the procedure as described in literatures^[S3, S4].



4. Photophysical properties and applications of products.

4.1. The absorption and fluorescence properties of products

Fluorescence spectra were collected on HITACHI F-700003040428 Fluorescence Spectrometer. UV-vis spectra were recorded on Agilent Cary60 spectrometer. Samples for the experiments were prepared in a 4.0 mL glass cuvette with a septum screw cap. In a typical experiment, a 5.0×10^{-5} M and 1.0×10^{-6} M solution of product in DCM for the collection of emission spectrum and UV-vis spectrum were prepared respectively.

Table S2. Absorption maxima and emission maxima of products.

Compounds	Absorb.(λ_{abs} , nm) ^a	Emission (λ_{em} , nm) ^b	CIE ₁₉₃₁
4a	258, 303, 345, 362	370, 388	(0.16, 0.02)
4b	264, 298, 310	366, 381	(0.16, 0.03)
4c	265, 299, 310	369, 386	(0.16, 0.04)
4d	277, 319, 333	382, 399	(0.16, 0.02)
4e	277, 322, 335	386, 404	(0.16, 0.02)
4f	279, 327, 341, 378	393, 409	(0.16, 0.02)
4g	273, 317, 332, 373	384, 402	(0.16, 0.02)
4h	281, 345	415	(0.16, 0.03)
4i	282, 341, 356	409	(0.16, 0.02)
4j	255, 288, 360	435	(0.15, 0.06)
4k	279, 326, 341, 376	385, 403	(0.16, 0.01)
4l	267, 282, 292, 320, 371, 392	445	(0.15, 0.08)
4m	251, 290, 306, 319, 377, 398	428, 452	(0.15, 0.04)
4n	298, 333	404	(0.16, 0.02)
4o	252, 275, 327, 433	496	(0.15, 0.49)
4p	272, 332, 370	388, 401	(0.16, 0.02)
4q	310, 376, 395	428	(0.16, 0.03)
4r	290, 352, 370	379, 395, 415	(0.17, 0.01)
4s	265, 296, 359, 377	383, 403, 425	(0.17, 0.01)
5a	268, 284, 320, 375, 395	446	(0.15, 0.07)
5b	261, 284, 296, 320, 372, 391	445	(0.15, 0.08)
5c	264, 302, 322, 370, 388	446	(0.15, 0.08)
5d	283, 319, 373, 395	444	(0.15, 0.08)
5e	262, 281, 292, 320, 367, 385	450	(0.15, 0.13)
5f	271, 283, 322, 373, 395	445	(0.15, 0.10)
5g	285, 296, 321, 369, 388	451	(0.15, 0.13)
5h	286, 320, 370, 392	454	(0.15, 0.14)
5i	290, 356, 388	551	(0.37, 0.58)
6a	264, 282, 318, 372, 390	444	(0.15, 0.08)
6b	262, 284, 318, 372, 392	443	(0.15, 0.07)
6c	264, 282, 318, 372, 392	440	(0.15, 0.07)
6d	266, 282, 320, 373, 393	442	(0.15, 0.07)

6e	265, 282, 319, 373, 394	442	(0.15, 0.07)
6f	293, 376	527	(0.26, 0.60)
6g	264, 288, 318, 368, 386	449	(0.15, 0.13)
6h	264, 282, 318, 370, 390	446	(0.15, 0.10)
6i	264, 288, 318, 368, 388	449	(0.15, 0.12)
6j	264, 286, 318, 370, 388	446	(0.15, 0.10)
6k	288, 318, 377, 388	451	(0.15, 0.13)
6l	264, 288, 318, 368, 388	451	(0.15, 0.13)
6m	266, 288, 318, 370, 388	447	(0.15, 0.11)
6n	288, 320, 368, 388	456	(0.15, 0.15)
6o	260, 288, 320, 368, 388	448	(0.15, 0.12)
6p	260, 288, 318, 370, 388	449	(0.15, 0.12)
6q	288, 316, 370, 390	446	(0.15, 0.10)
6r	264, 282, 320, 372, 388	454	(0.15, 0.13)
6s	256, 292, 326, 392, 412	448, 463	(0.14, 0.09)
6t	288, 416	523	(0.24, 0.61)
6u ^c	290, 352	412, 436, 561	(0.31, 0.34)
6v	290, 414	535	(0.32, 0.62)
6w ^d	278, 306, 450	394, 418, 571	(0.33, 0.33)
6w NPs ^e	281, 305, 459	426, 465, 610	(0.33, 0.28)

^a Absorption maximum in CH₂Cl₂ (1×10⁻⁶ M). ^b Emission maximum in CH₂Cl₂ (5.0×10⁻⁵ M). ^c CH₂Cl₂ used as solvent (2.5×10⁻⁷ M). ^d Toluene used as solvent (2.4×10⁻⁶ M). ^e H₂O used as solvent (2.0×10⁻⁵ M).

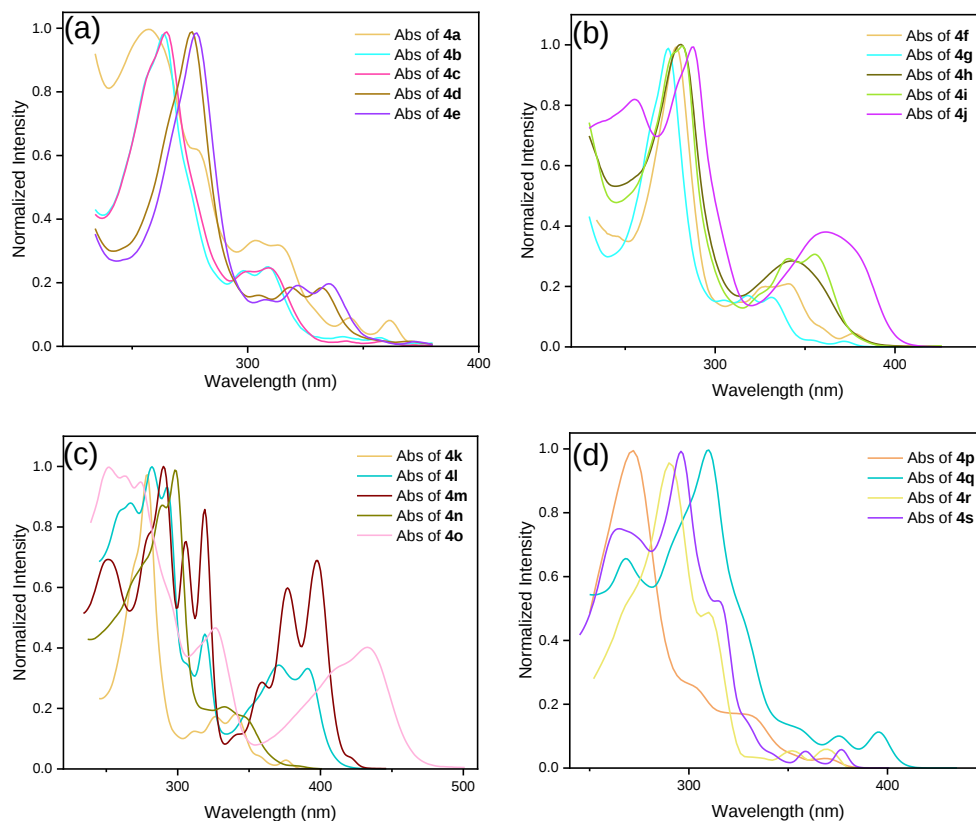


Fig. S7. Normalized UV-Vis absorption spectra of **4a-4s** in DCM (1×10^{-6} M).

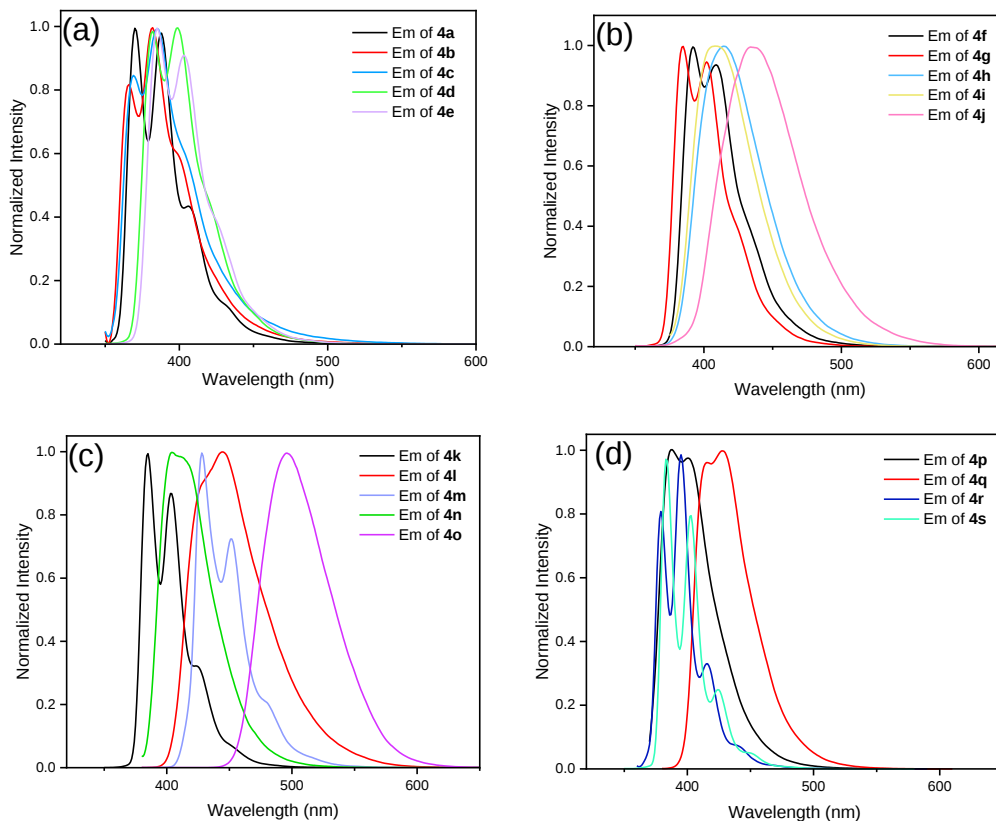


Fig. S8. Normalized fluorescence emission spectra of **4a-4s** in DCM (5.0×10^{-5} M).

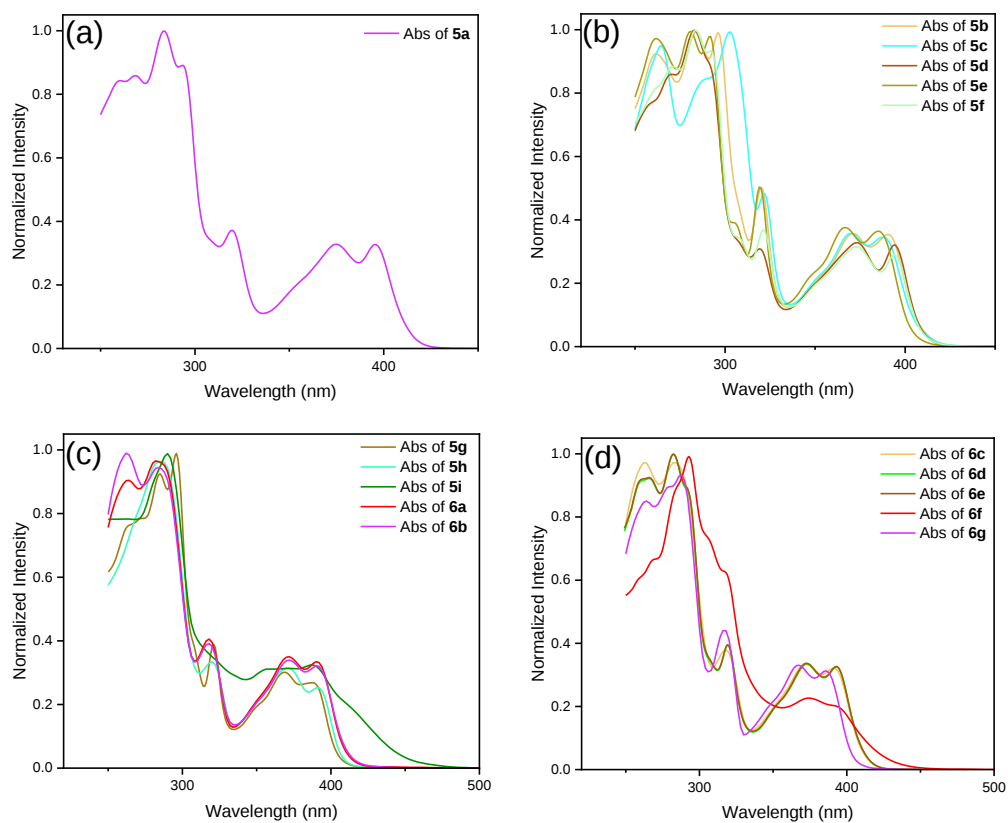


Fig. S9. Normalized UV-Vis absorption spectra of **5a-6g** in DCM (1×10^{-6} M).

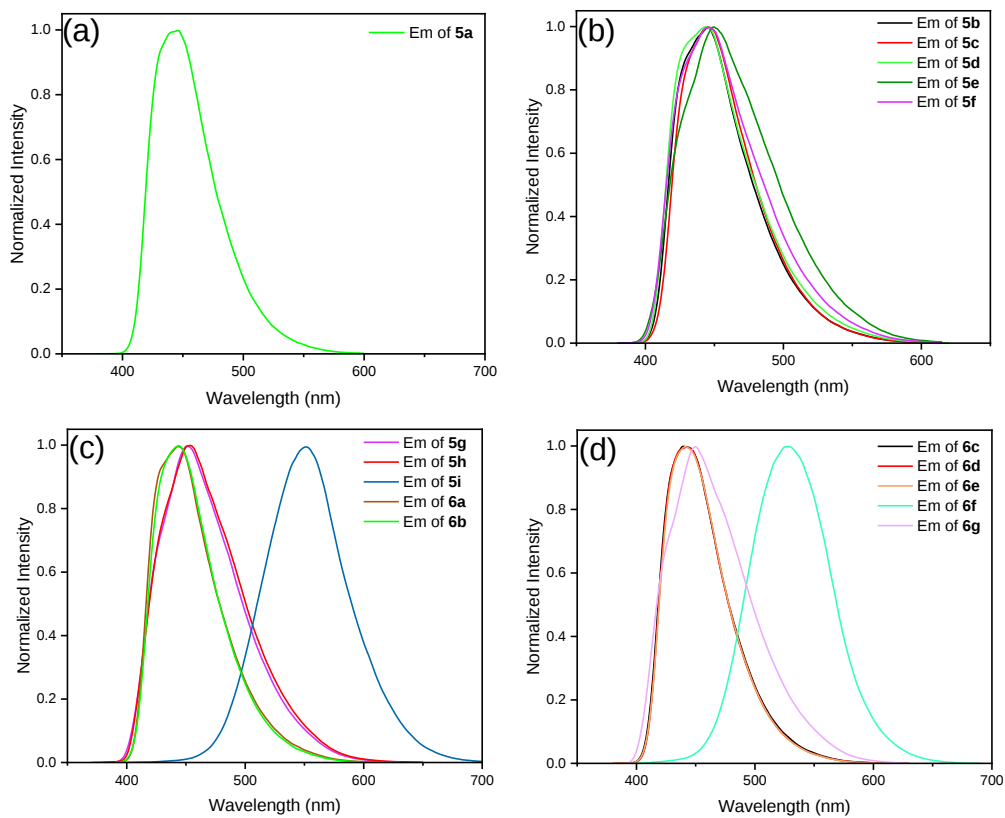


Fig. S10. Normalized fluorescence emission spectra of **5a-6g** in DCM (5.0×10^{-5} M).

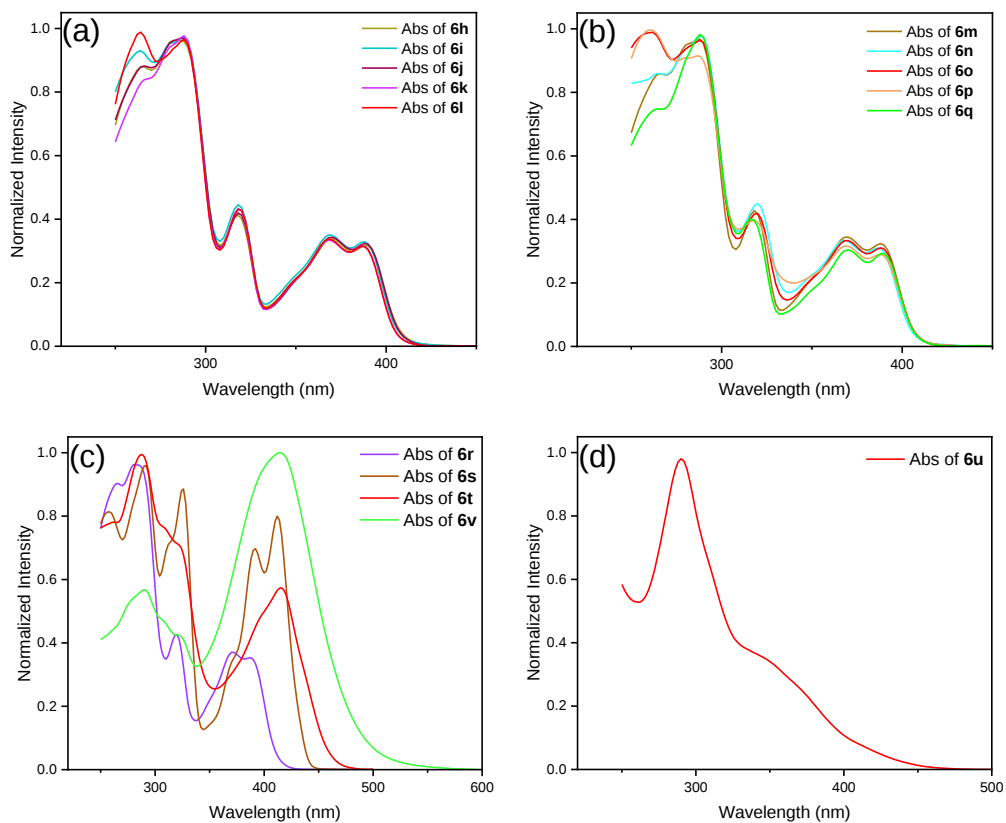


Fig. S11. Normalized UV-Vis absorption spectra of **6h-6v** in DCM (1×10^{-6} M).

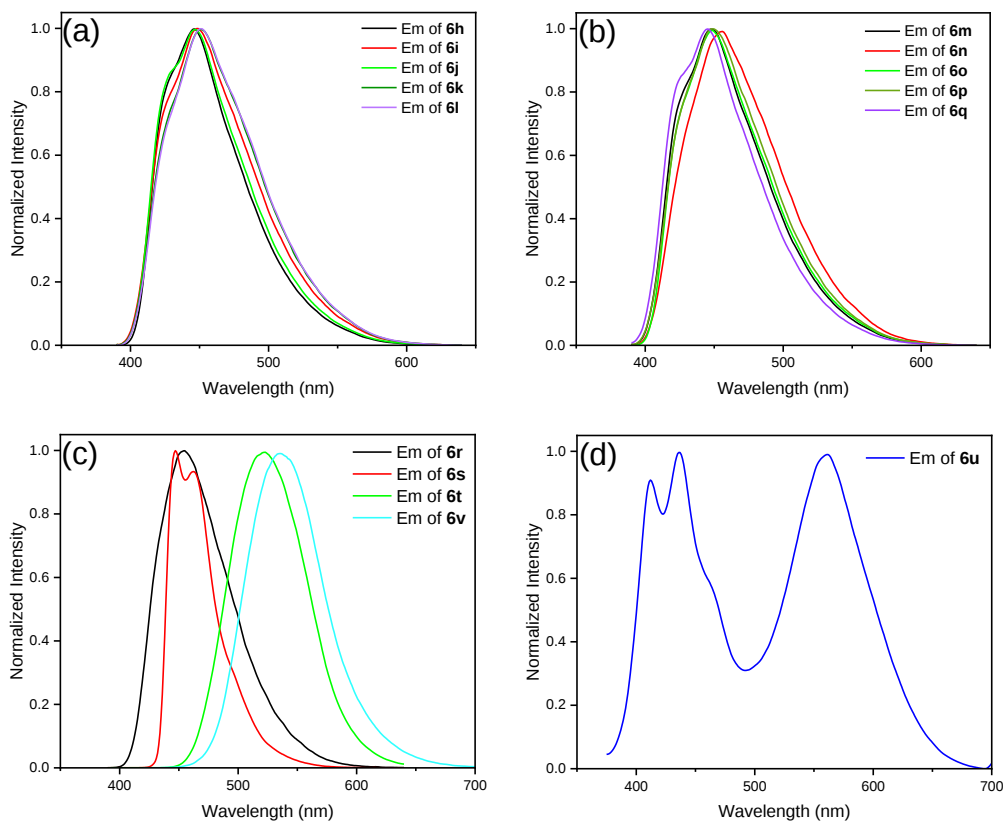


Fig. S12. (a - c) Normalized fluorescence emission spectra of **6h-6t** and **6v** in DCM

(5.0×10^{-5} M). (d) Normalized fluorescence emission spectra of **6u** in DCM (2.5×10^{-7} M).

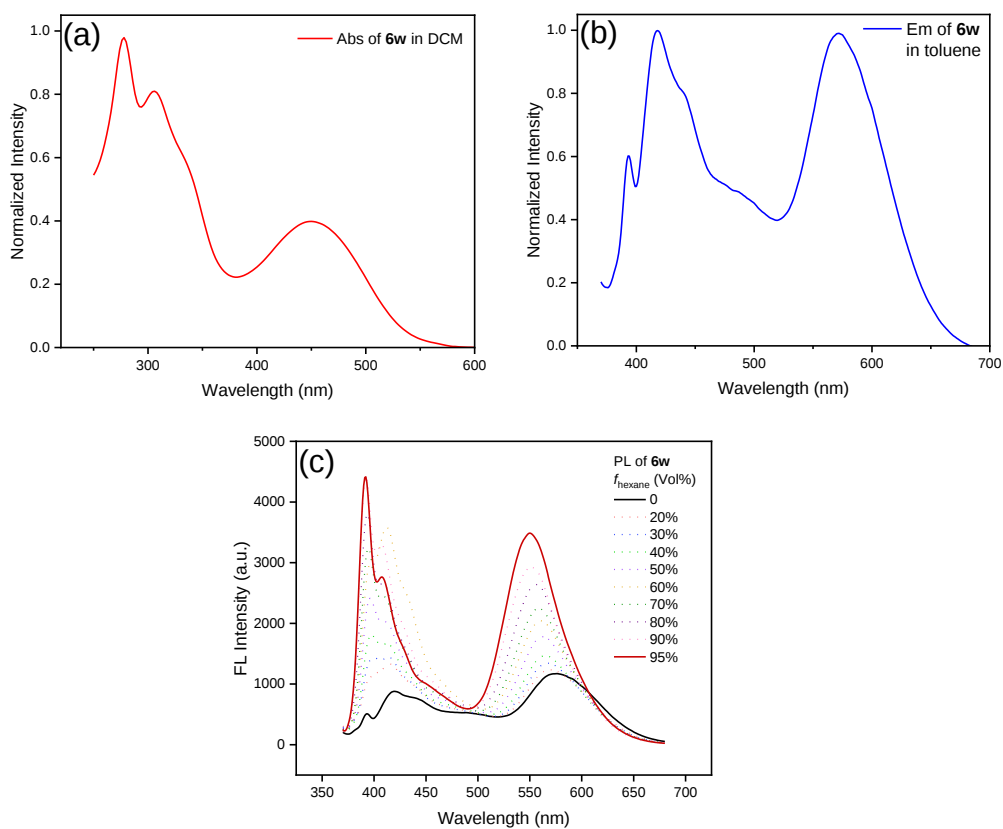


Fig. S13. (a) Normalized UV-Vis absorption spectra of **6w** in DCM (1×10^{-6} M); (b) Normalized fluorescence emission spectra of **6w** in toluene (2.4×10^{-6} M); (c) Fluorescence spectra of **6w** in toluene–hexane mixtures (2.4×10^{-6} M) with different fractions (f_{hexane}).

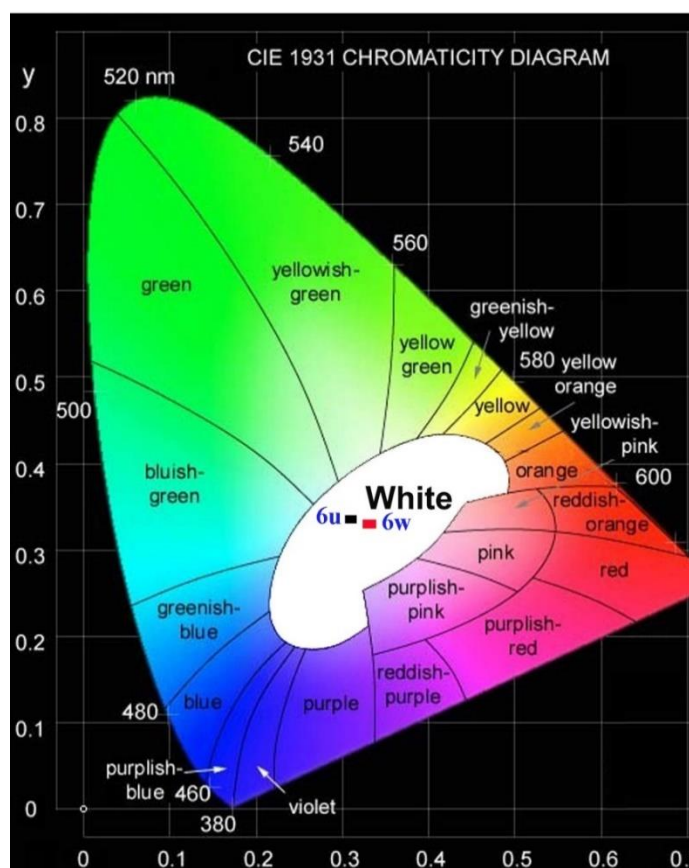


Fig. S14. The white-light emissions of **6u** and **6w**. CIE₁₉₃₁ chromaticity diagrams **6u** (0.31, 0.34) and **6w** (0.33, 0.33) in CH₂Cl₂ (Concentration: **6u**: 2.5×10^{-7} M; **6w**: 2.4×10^{-6} M).

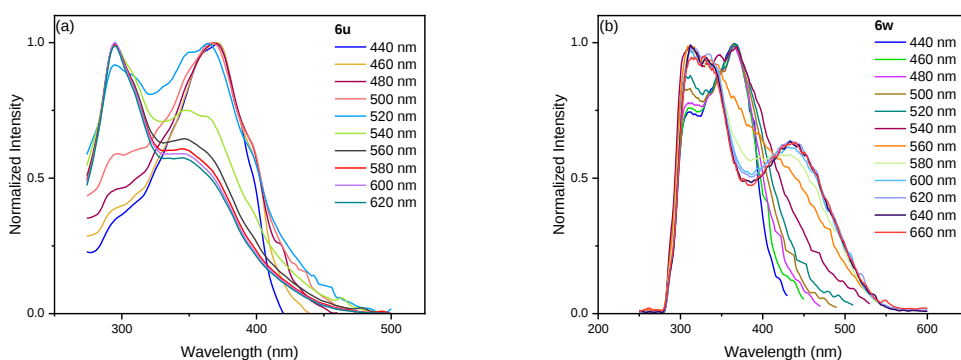


Fig. S15. (a) Emission-wavelength-dependent excitation spectra of **6u** (1.0×10^{-8} M in DCM). (b) Emission-wavelength-dependent excitation spectra of **6w** (1.3×10^{-7} M in toluene). Excitation spectra were collected by Edinburgh Instruments FLS1000.

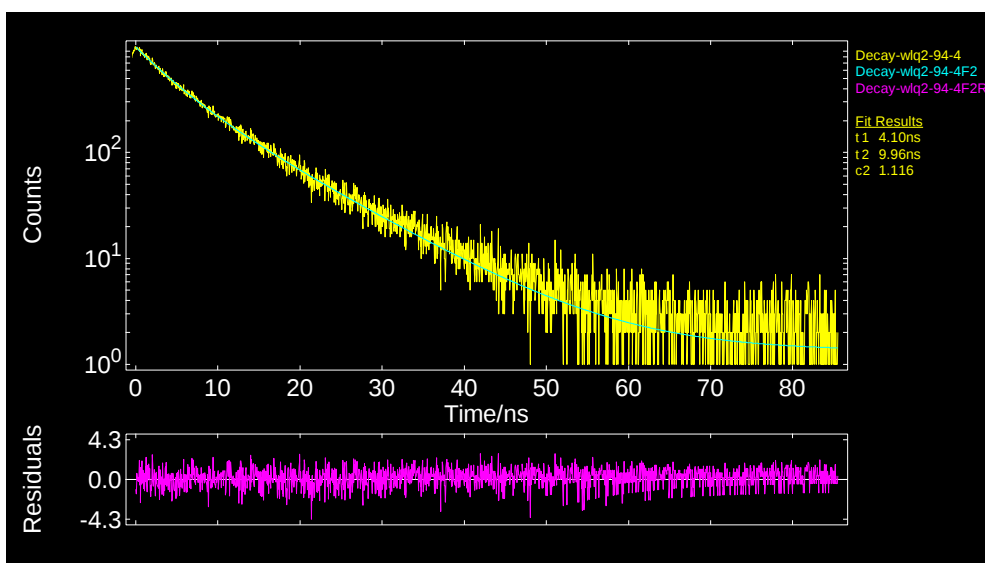


Fig. S16. Fluorescence lifetime decay curves of **6u** in TBCPF film (3 wt%).

Fix	Value / ns	Std. Dev / ns	Fix	Value	Std. Dev	Rel %
τ_1	4.1024	0.1554	B_1	524.3182	21.0846	31.39
τ_2	9.9586	0.1609	B_2	472.1648	23.1554	68.61
τ_3			B_3			
τ_4			B_4			

$\langle \tau \rangle_{amp}$	6.8773	0.1451
$\langle \tau \rangle_{int}$	8.1205	0.1533

A

$\chi^2 : 1.1161$

Fig. S17. The excited state lifetimes of **6u** in TBCPF film.

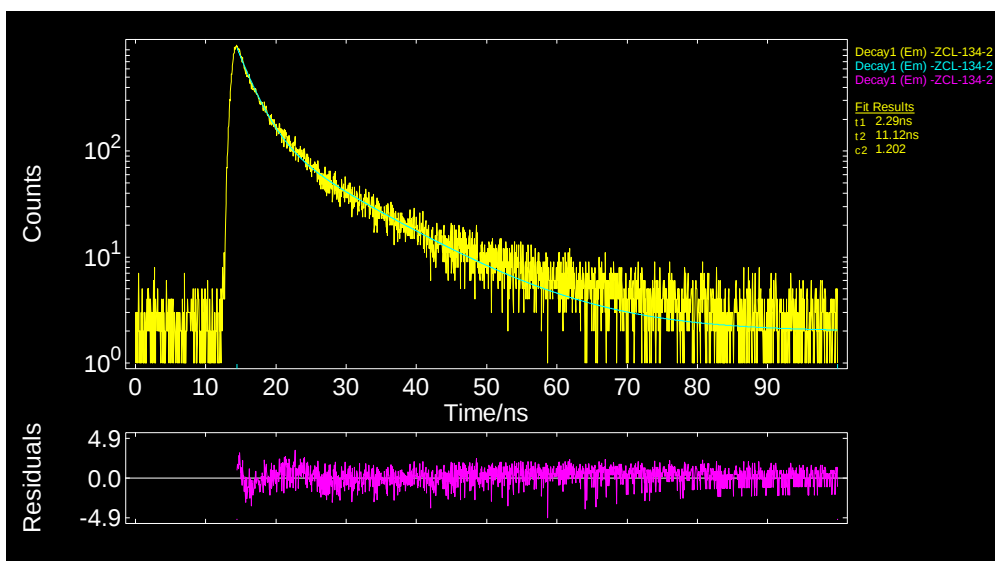


Fig. S18. Fluorescence lifetime decay curves of **6w** in TBCPF film (10 wt%).

Fix	Value / ns	Std. Dev / ns	Fix	Value	Std. Dev	Rel %
τ_1 ☐	2.2908	0.0338	B_1 ☐	799.6371	7.7316	51.32
τ_2 ☐	11.1197	0.1709	B_2 ☐	156.2660	4.3568	48.68
τ_3 ☐			B_3 ☐			
τ_4 ☐			B_4 ☐			

$\langle \tau \rangle_{amp}$	3.7341	0.0534
$\langle \tau \rangle_{int}$	6.5888	0.1348

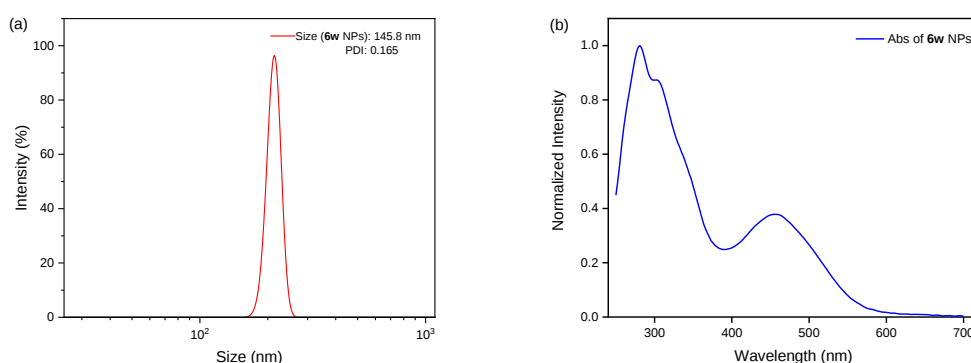
A ☐ 1.9782

$\chi^2 : 1.2024$

Fig. S19. The excited state lifetimes of **6w** in TBCPF film.

4.2. Preparation and characterization of **6w** NPs.

The compound **6w** (2.0 mg) was dissolved in THF (2 mL) followed by ultrasound for 10 min. Poloxamer 188 (60 mg) was added to the solution of **6w** in THF and the mixture was treated by ultrasound for 10 min. The resulted mixture was stirred three hours at RT. Then the solvent was removed by reduced pressure. The residue was dissolved in deionized water (2 mL) and stirred overnight at RT to drive the formation of NPs. To remove the residual THF, the nanoparticle suspensions were dialyzed by three times. The concentrations of nanoparticles were calibrated via the UV-vis spectra. Finally, the NPs solution was collected and stored at 4 °C.



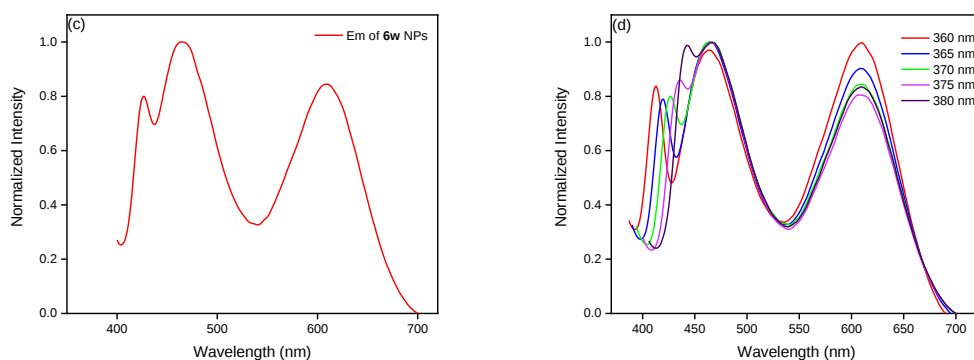


Fig. S20. (a) Hydrodynamic size distribution of **6w** NPs in aqueous solution. (b) Normalized UV-Vis absorption spectra emission spectra of **6w** NPs. in H₂O (1.0×10^{-6} M). (b) Normalized fluorescence emission spectra of **6w** NPs. in H₂O (2.0×10^{-5} M). (d) Excitation-wavelength-dependent fluorescence spectra of **6w** NPs in H₂O (2.0×10^{-5} M).

4.3. Cell imaging experiments

Cellular Experiments: The Hela cells were cultured in 10% FBS and 1% penicillin-streptomycin-containing DMEM medium at 37 °C with 5% CO₂.

Methyl thiazolyl tetrazolium (MTT) assay: To assess the safe usability of **6w** nanoparticles for biomedical applications, the cytotoxicity experiments in Hela cells were carried out using the standard methyl thiazolyl tetrazolium (MTT) assay.

Cell viability assay: The Hela cells were seeded into 96-well plates and incubated with a standard medium overnight. The cell suspensions were treated with different concentrations of **6w** NPs in a deionized water solution (0, 1, 5, 10, 20, 40 μM) and incubated for another 24 h. Live/dead cell staining was carried out after 20 min white light irradiation and without light irradiation, respectively.

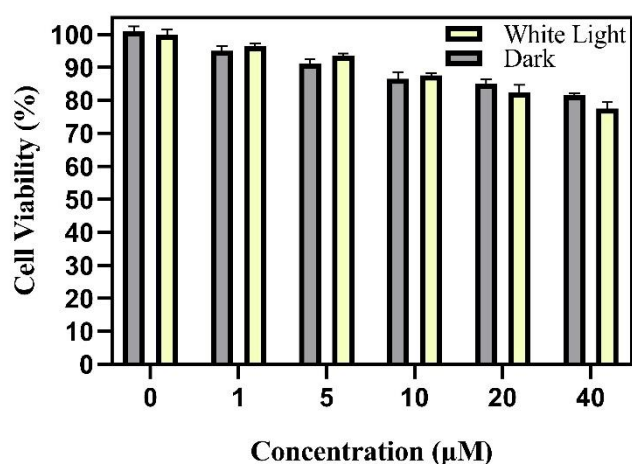


Fig. S21. In vitro cytotoxicity of the **6w** NPs. Cell viabilities of HeLa cells incubated with **6w** NPs for 24 h.

Confocal imaging experiments: The confocal imaging experiments were performed on Leica TCS SP8. The nanoparticles of product **6w** (**6w** NPs) was used as fluorescent probe.

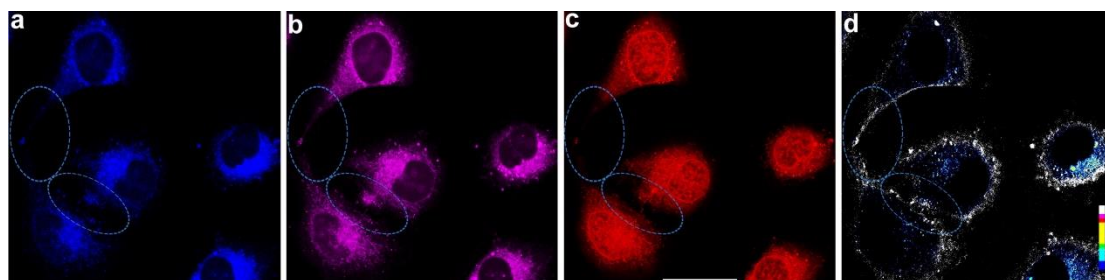


Fig. S22. Fluorescence microscopy images of HeLa cells incubated with **6w** NPs (10 μM) for 2 h at 37 °C. (a) Fluorescence microscope image from channel 1 at 425–500 nm (Excitation 405 nm). (b) Fluorescence microscope image from channel 1 at 500–700 nm (Excitation 405 nm). (c) Fluorescence microscope image from channel 2 at 500–700 nm (Excitation 488 nm). (d) The emission intensity ratio of (a) and (c) of HeLa cells (Image generation by Image J software). Elliptic: mark out the intercellular structure and cellular local imaging information. The scale bar is 25.0 μm.

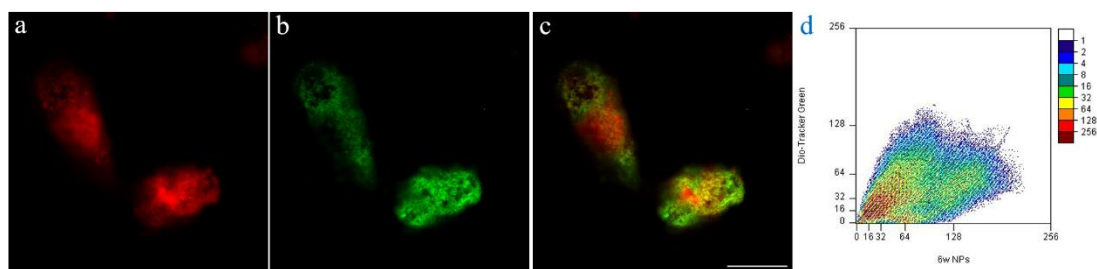


Fig. S23. The co-staining experiments. (a) Fluorescent image of HeLa cells cultured with **6w** NPs (10 μ M) (λ_{ex} = 552 nm, λ_{em} = 575–750 nm). (b) Fluorescent image of HeLa cells with Dio-Tracker Green (λ_{ex} = 488 nm, λ_{em} = 508–540 nm). (c) Merged image of (a) and (b). (d) The Pearson correlation coefficient r = 0.57; The scale bar is 25.0 μ m.

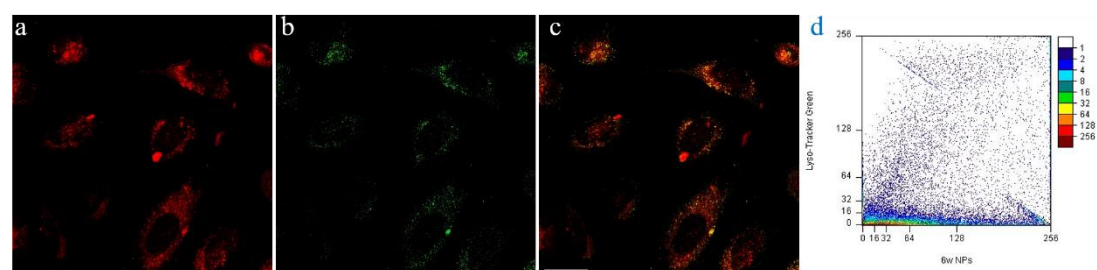


Fig. S24. The co-staining experiments. (a) Fluorescent image of HeLa cells cultured with **6w** NPs (10 μ M) (λ_{ex} = 552 nm, λ_{em} = 575–750 nm). (b) Fluorescent image of HeLa cells with Lyso-Tracker Green (λ_{ex} = 488 nm, λ_{em} = 508–540 nm). (c) Merged image of (a) and (b). (d) The Pearson correlation coefficient r = 0.44; The scale bar is 25.0 μ m.

5. References

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K., Montgomery, J. A., Peralta, Jr., J. E., Ogliaro, F., Bearpark, M. J., Heyd, J. J., Brothers, E. N., Kudin, K. N., Staroverov, V. N., Keith, T. A., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A. P., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Millam, J. M., Klene, M., Adamo, C., Cammi, R., Ochterski, J. W., Martin, R. L., Morokuma, K., Farkas, O., Foresman, J. B. & Fox, D. J., Gaussian 16, Revision A.03, Gaussian, Inc., Wallingford CT, **2016**.

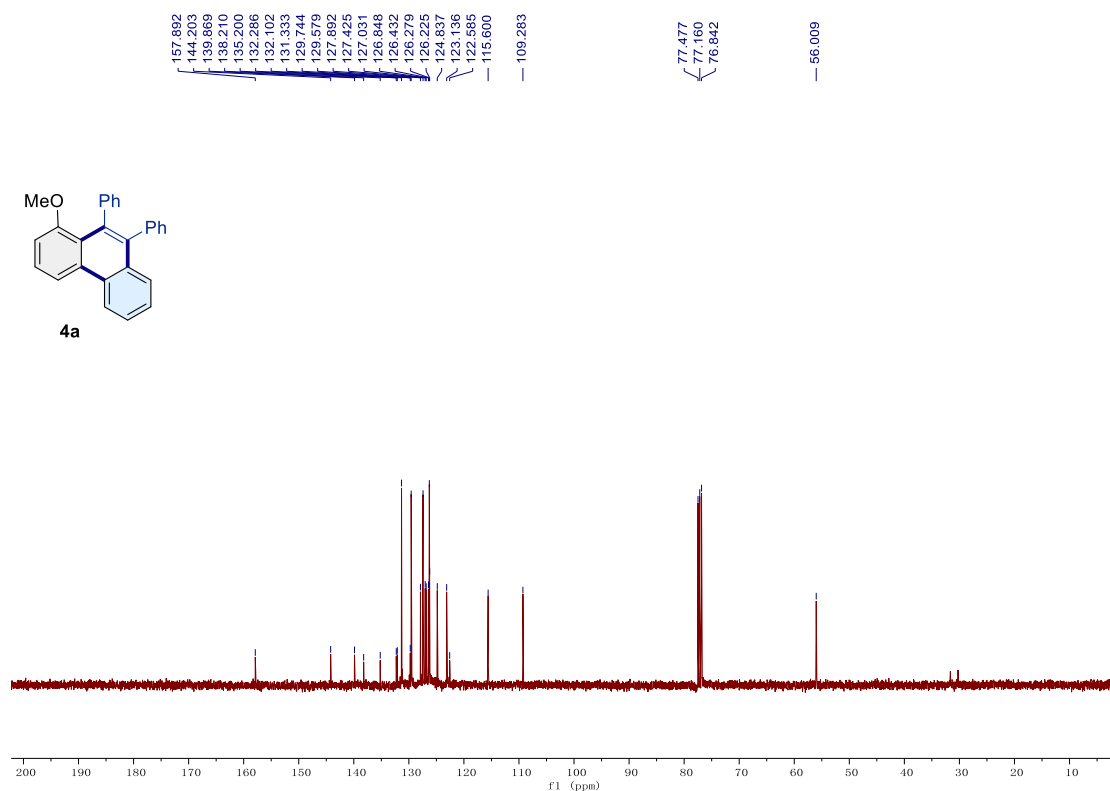
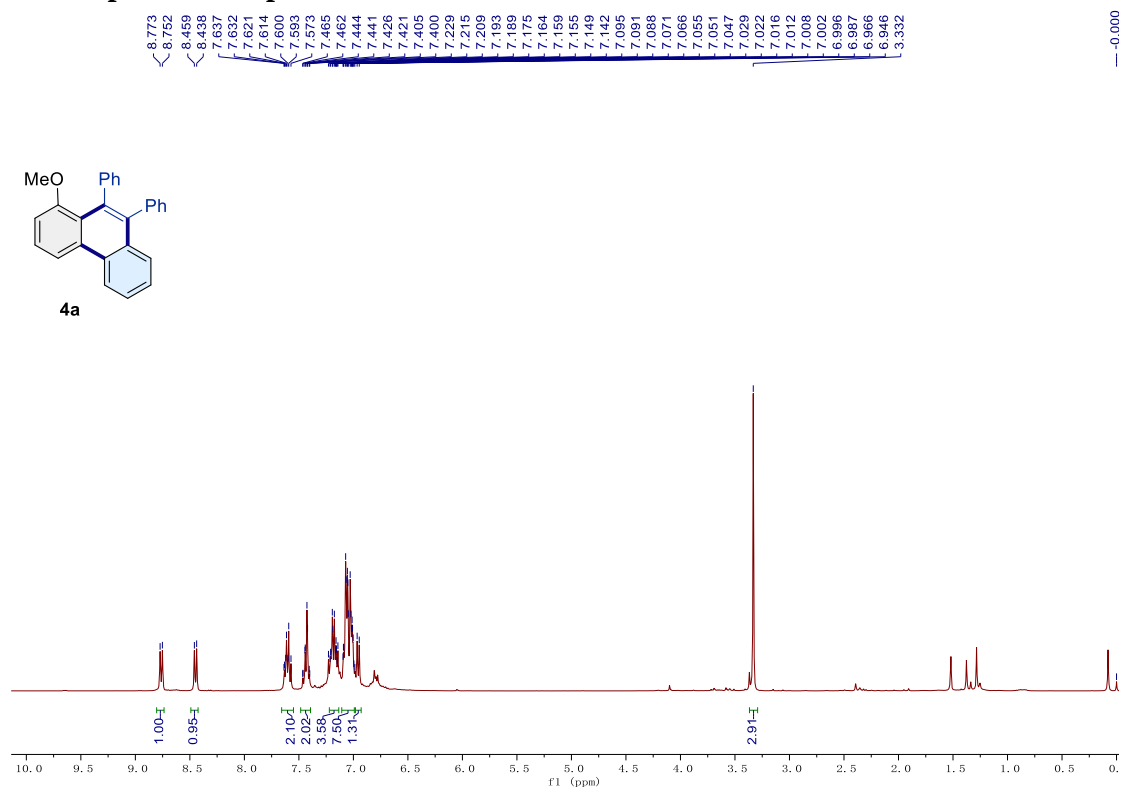
[S2] J. Bai, N. Xu, H. Wang and X. Luan. *Org. Lett.* **2022**, *24*, 5099–5104.

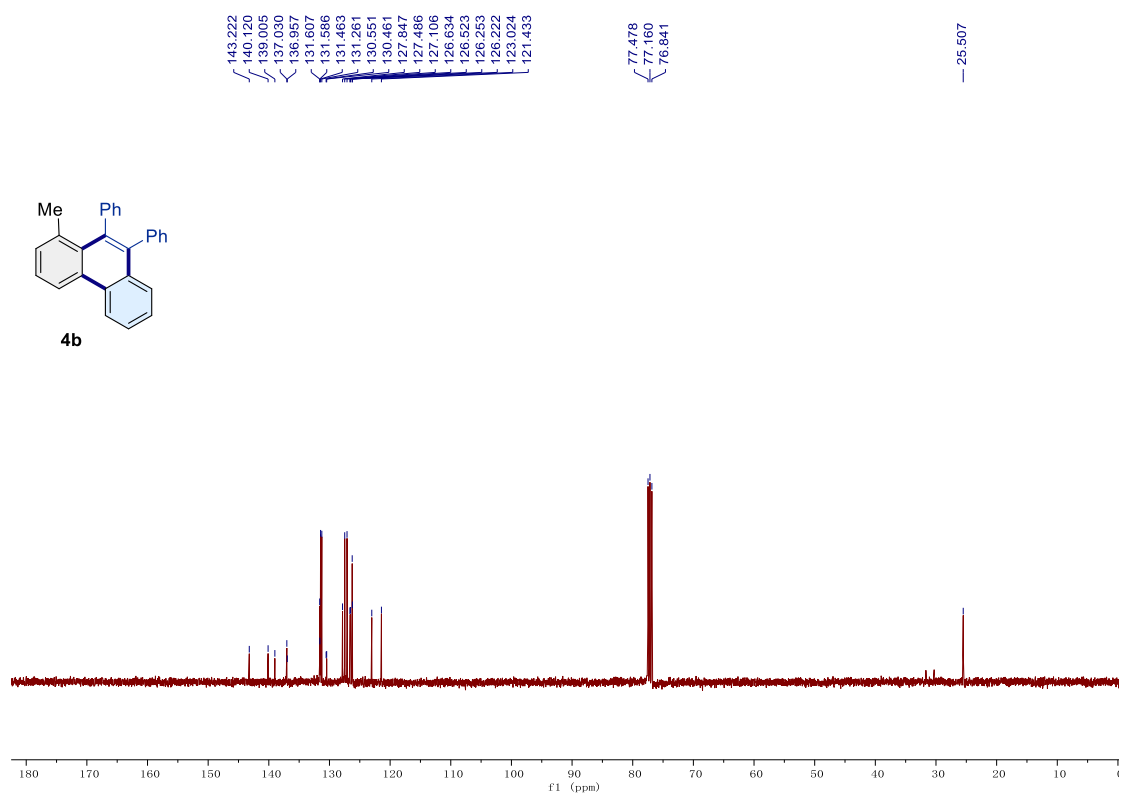
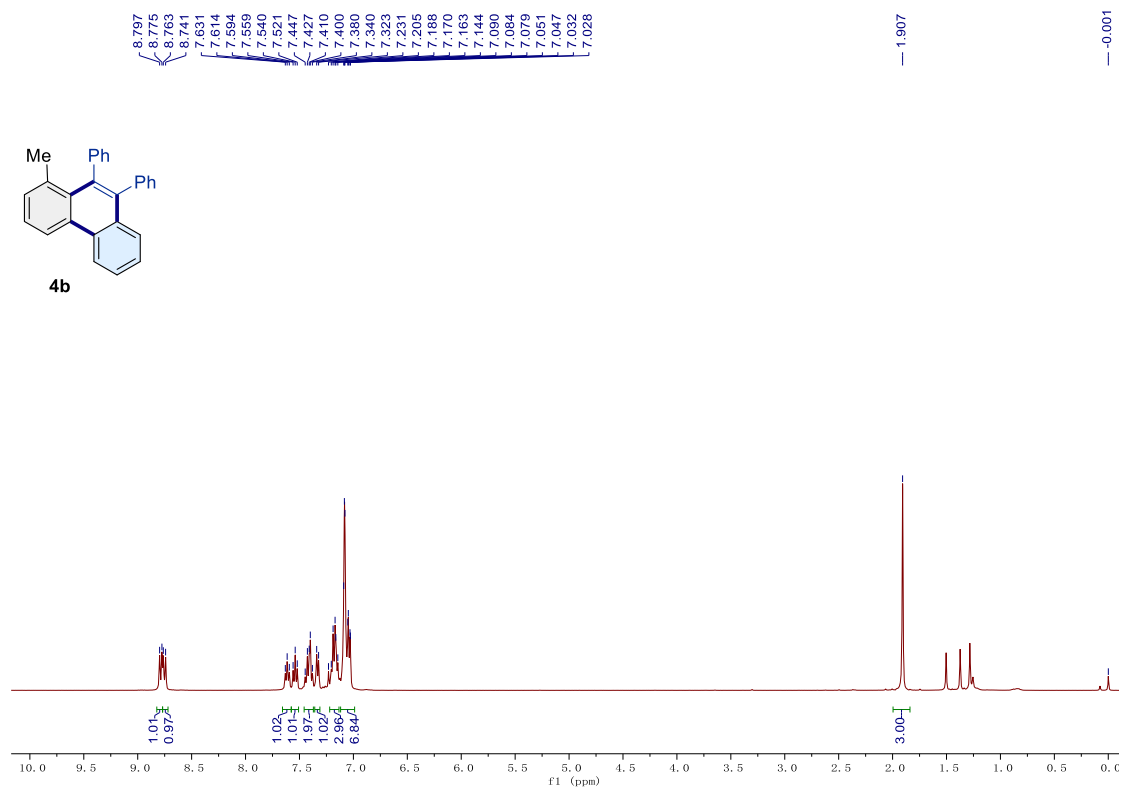
[S3] J. Wang, Z. Dong, C. Yang and G. Dong. *Nat. Chem.* **2019**, *11*, 1106–1112.

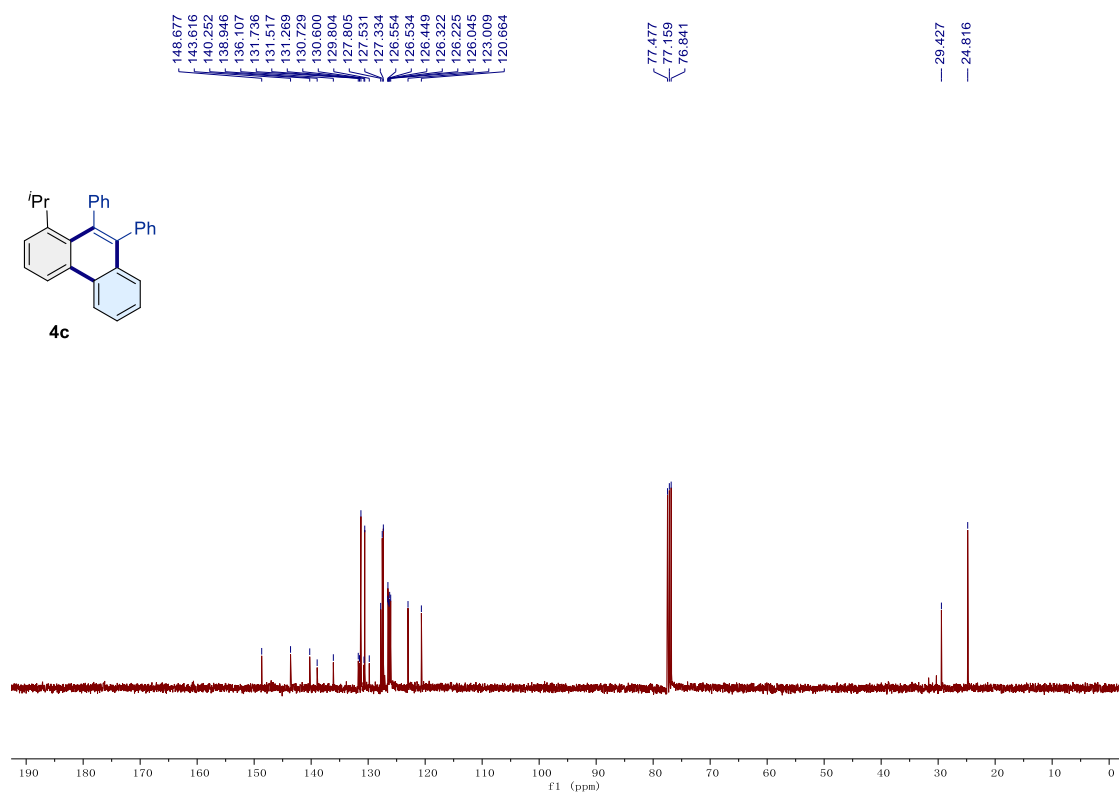
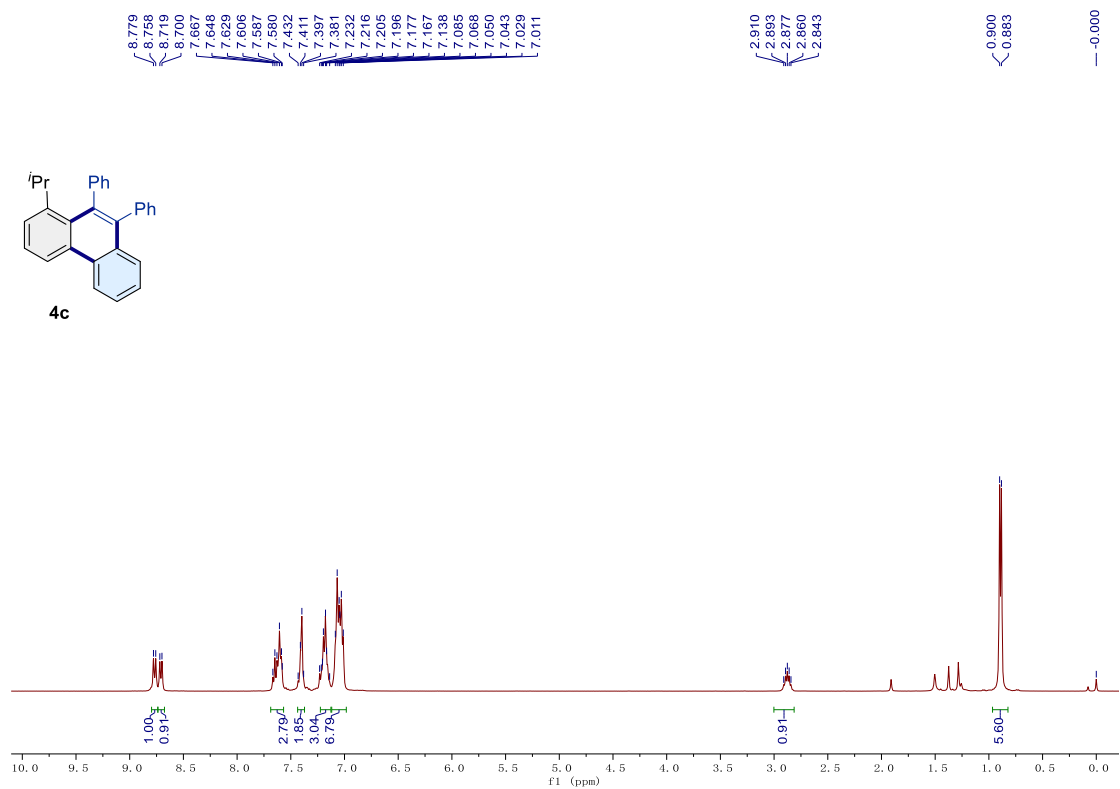
[S4] V. Sukowski, M. Borselen, S. Mathew and M. Á. Fernández-Ibáñez. *Angew. Chem. Int. Ed.* **2022**, *61*, e202201750.

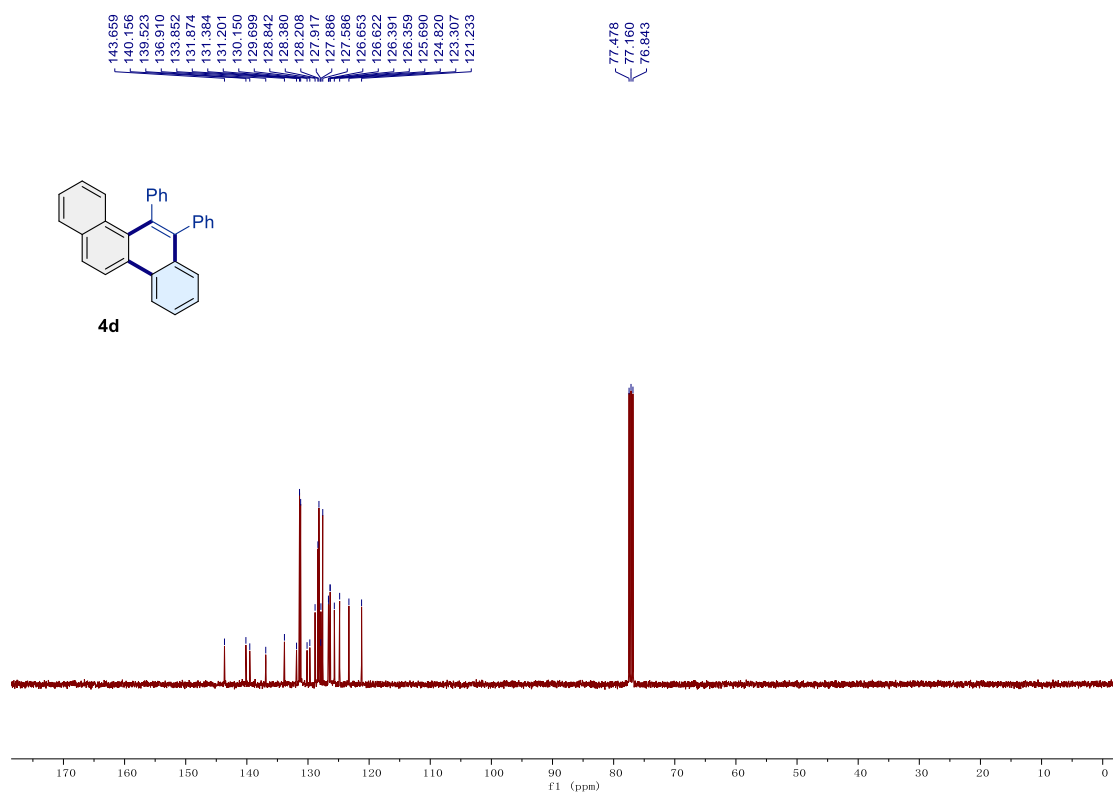
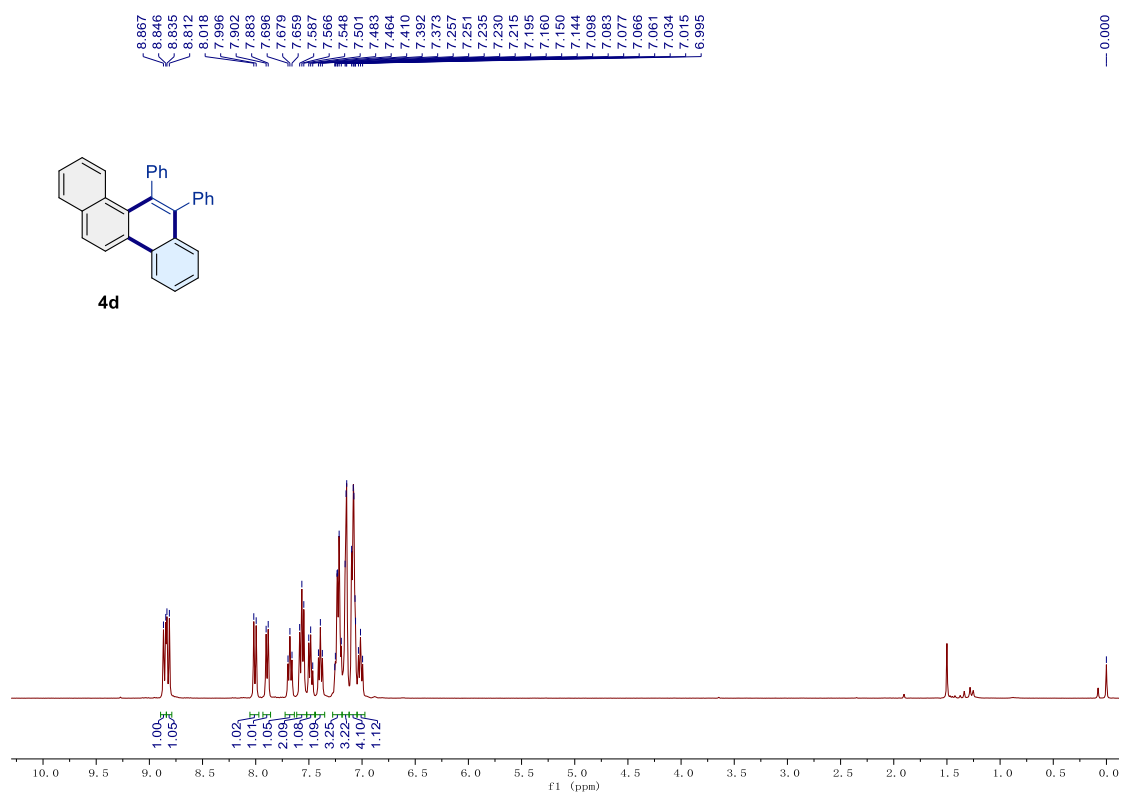
6. NMR spectrum of compounds

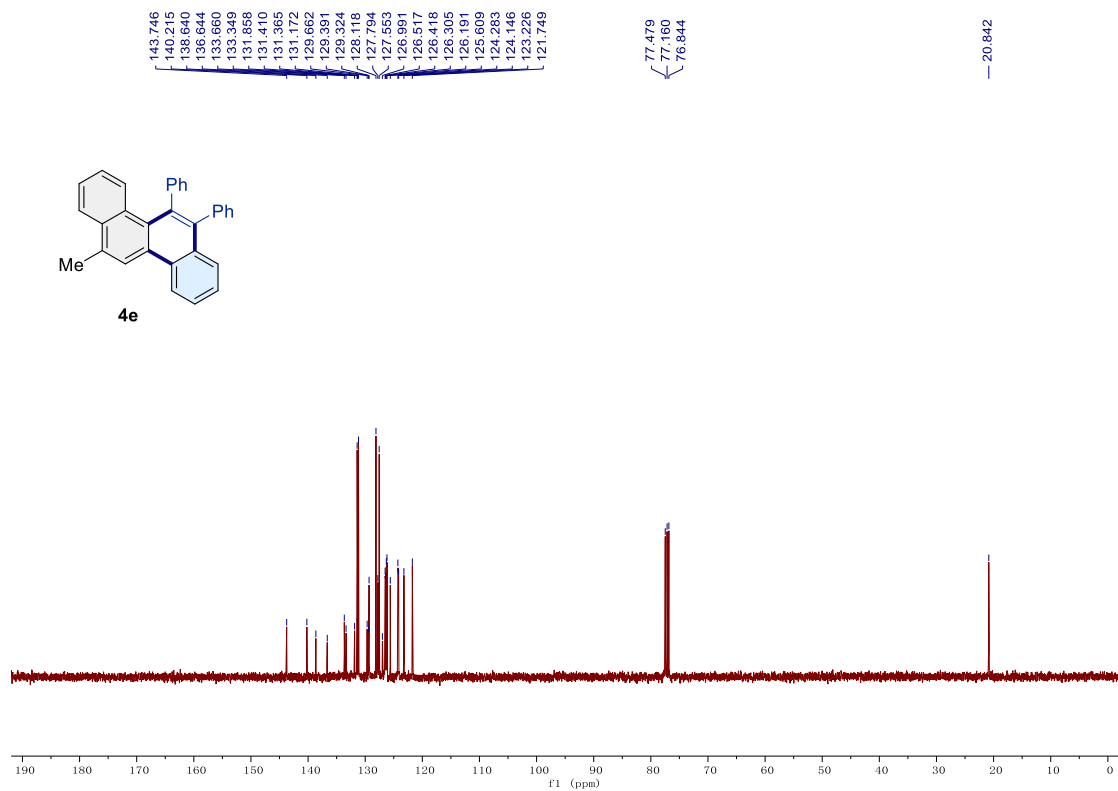
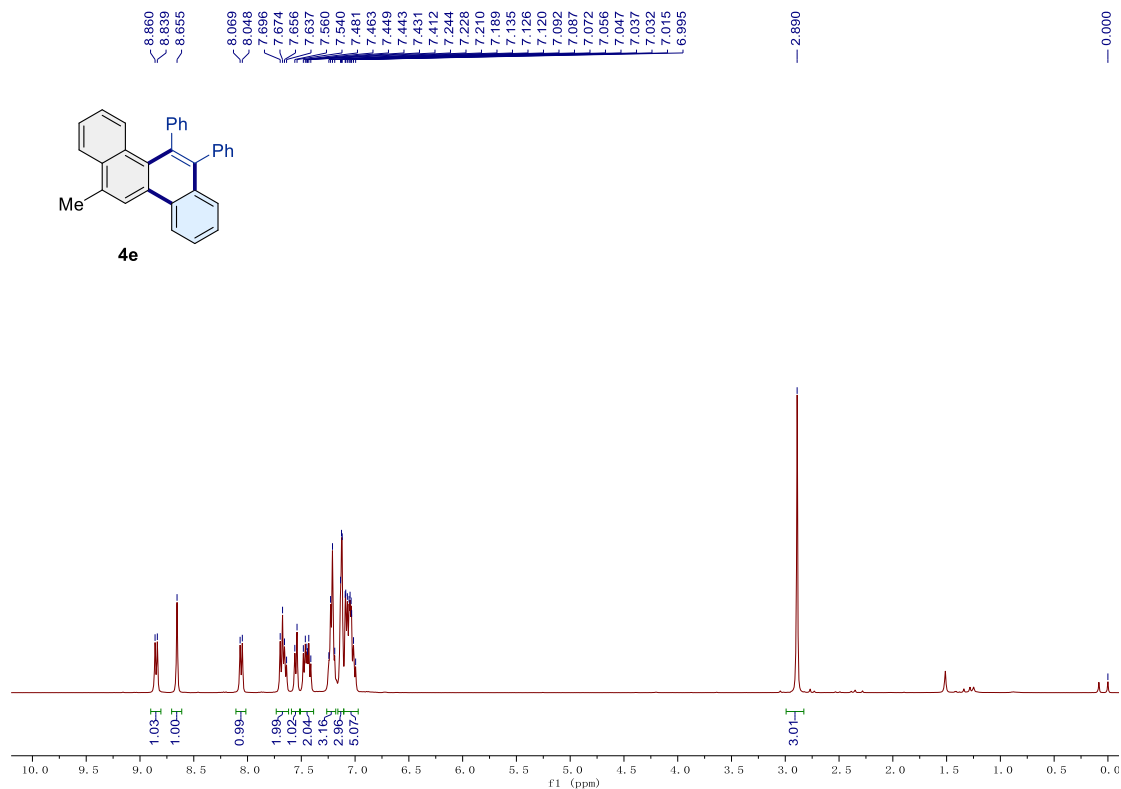
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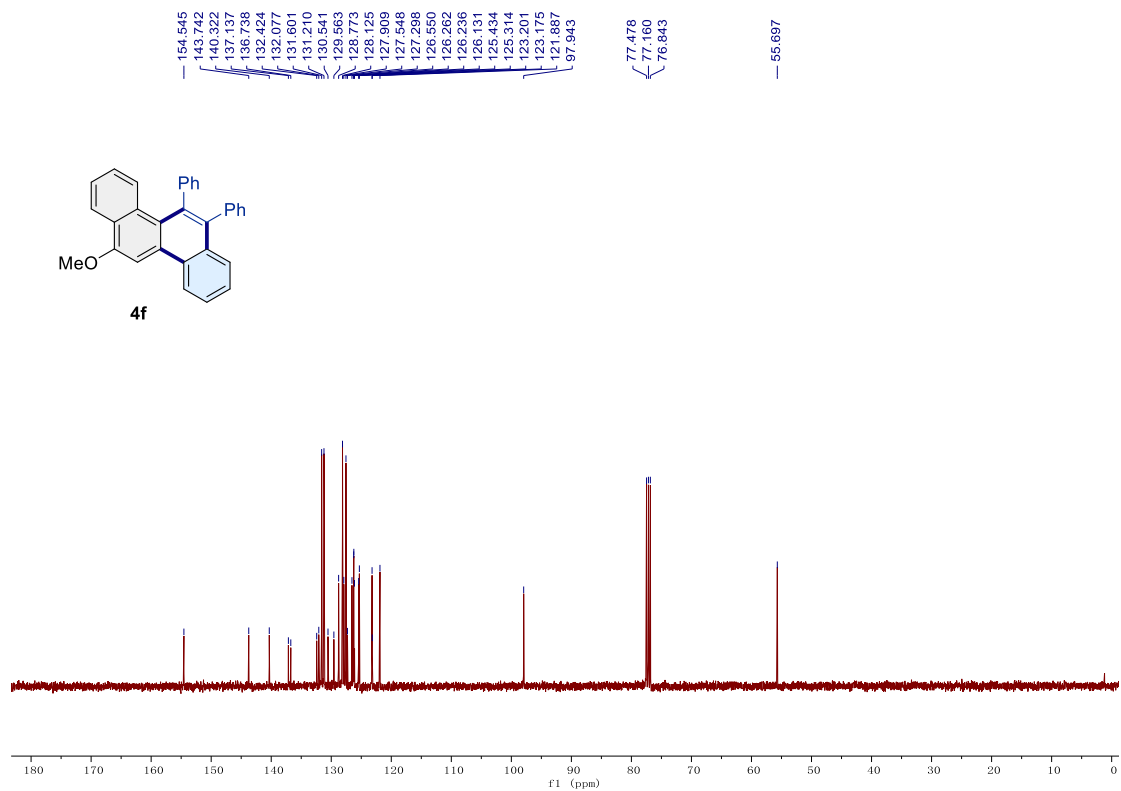
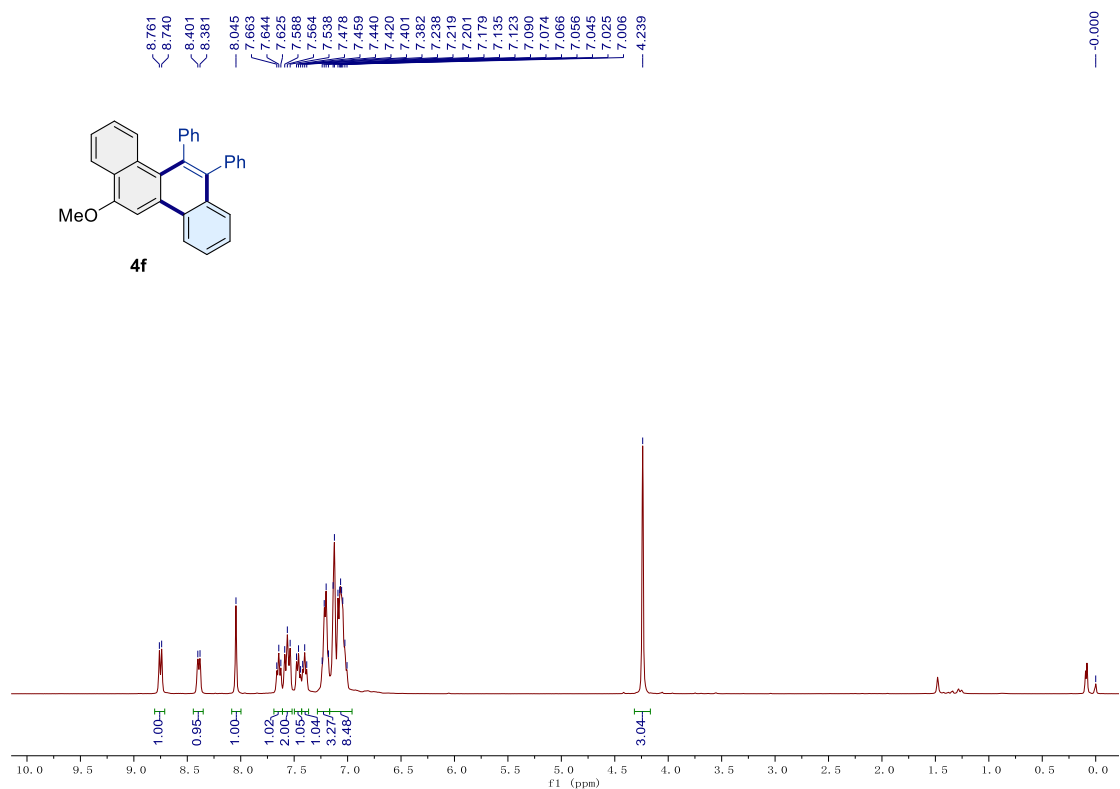


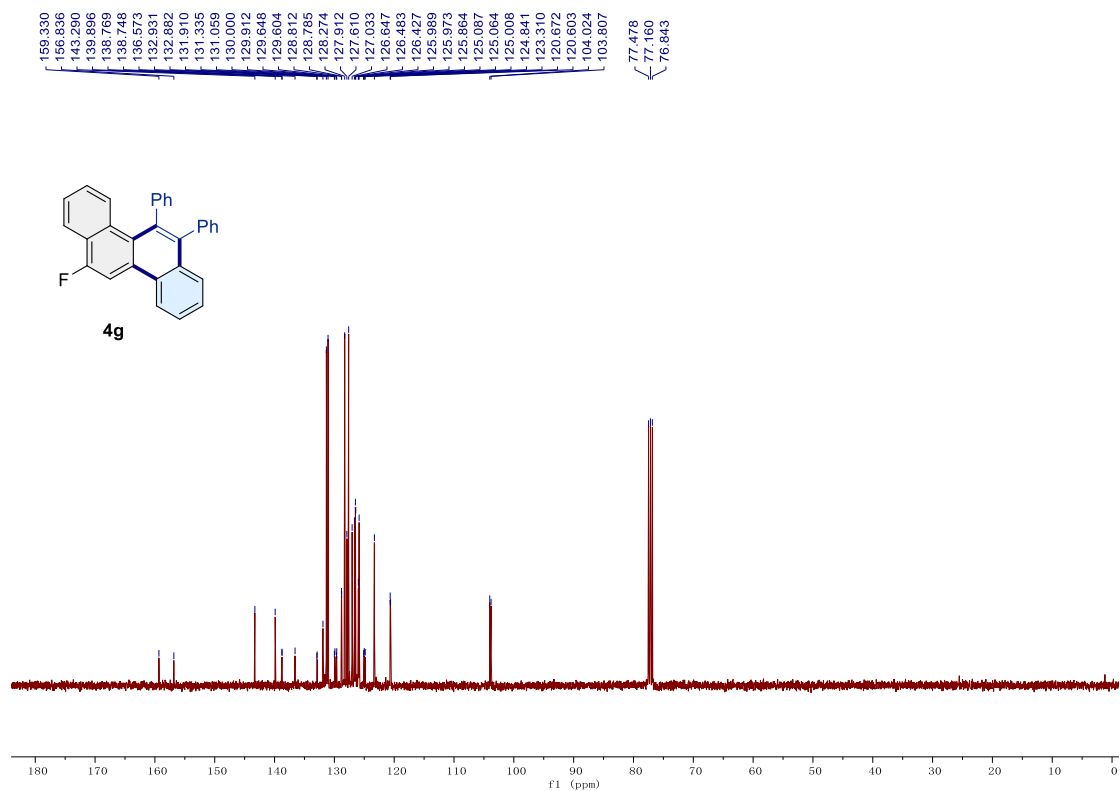
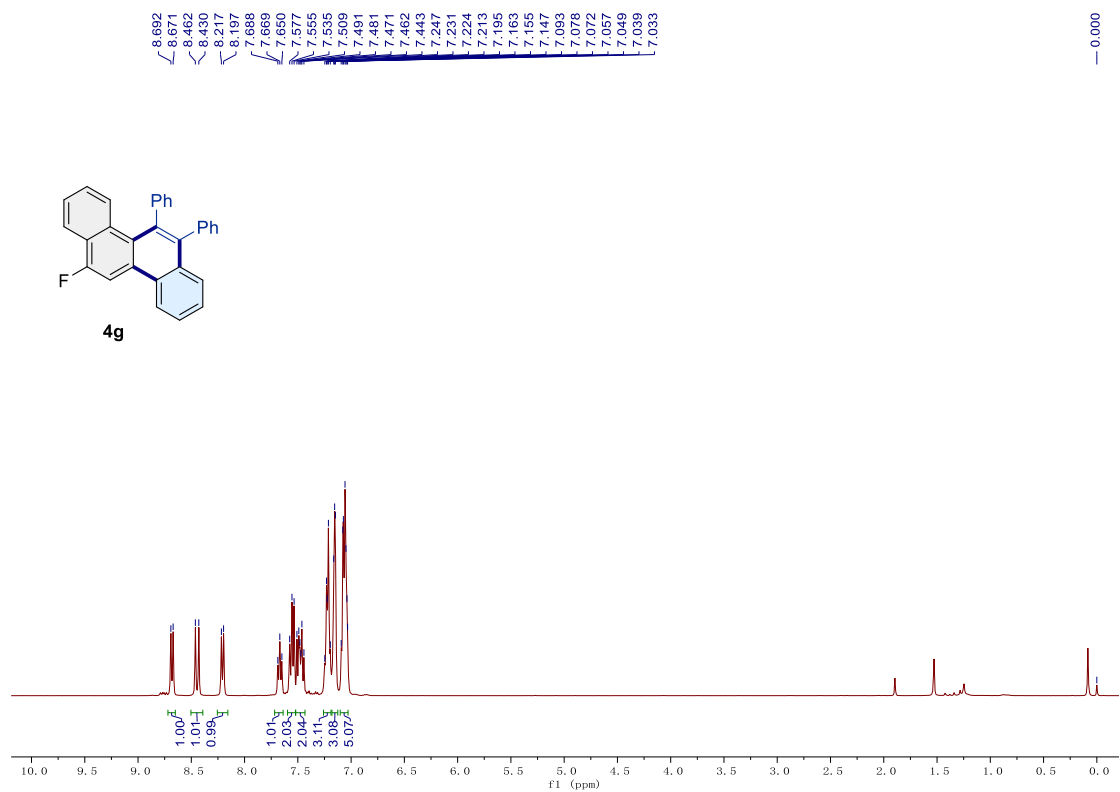


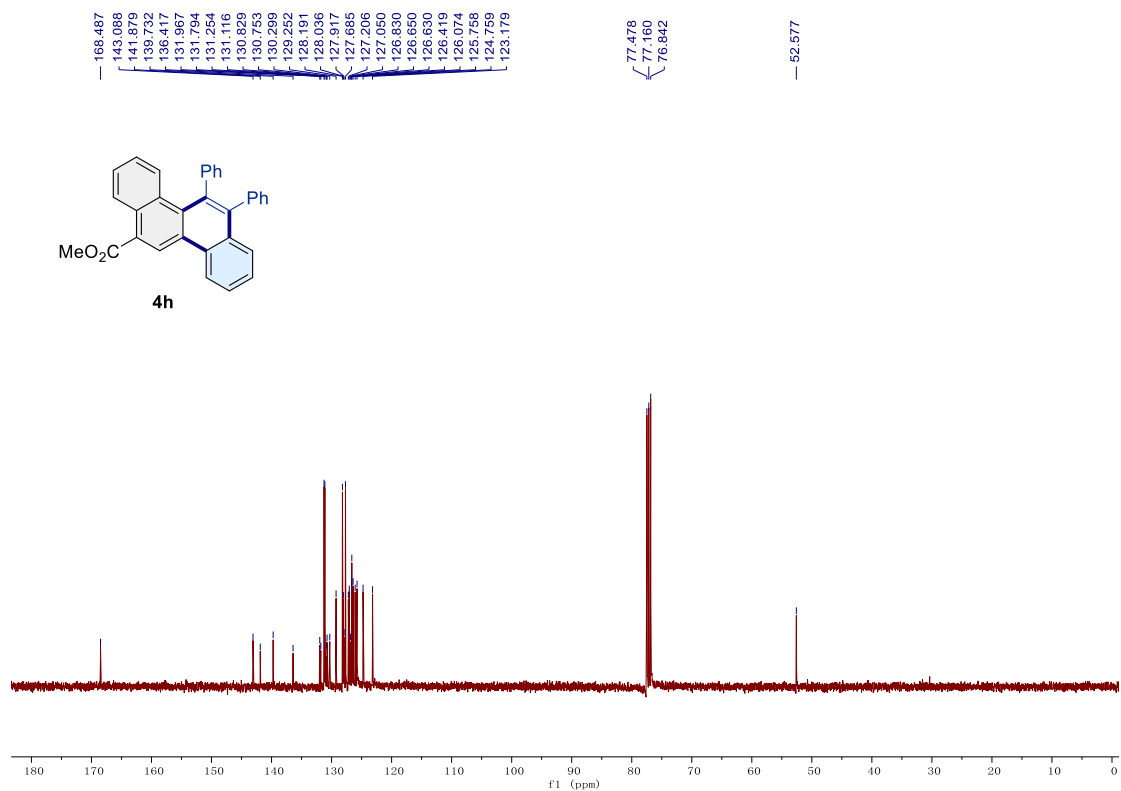
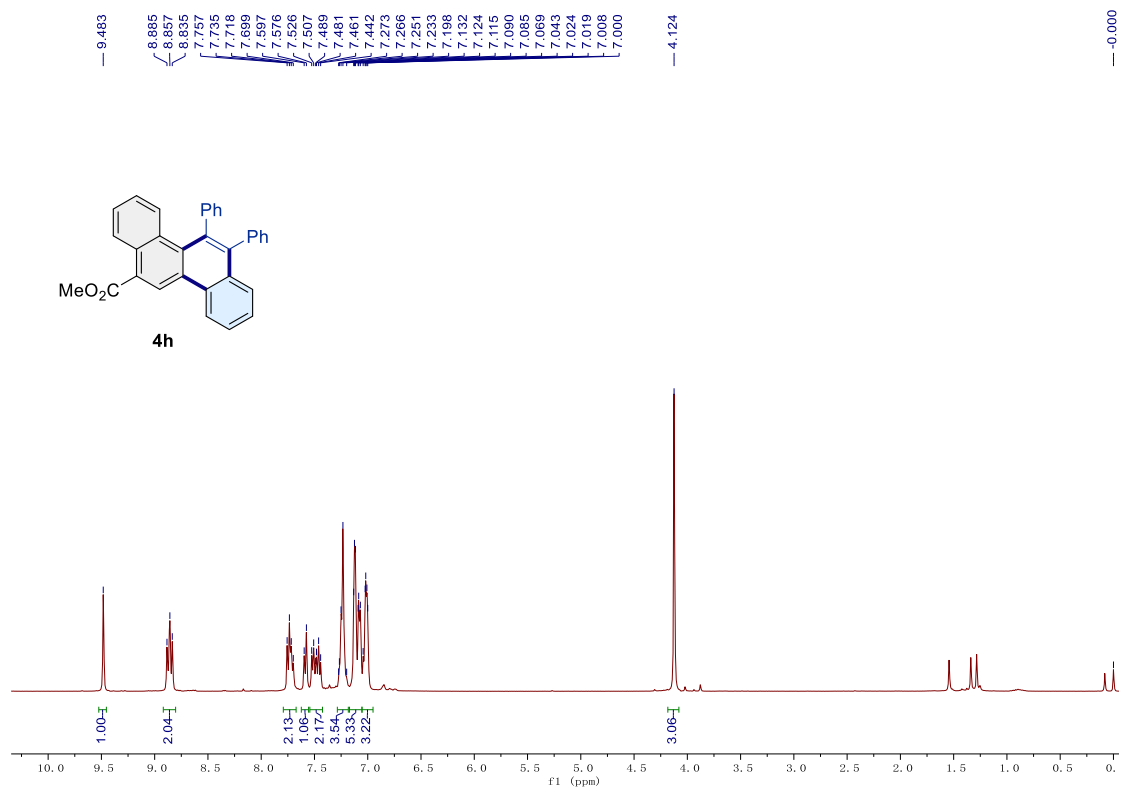


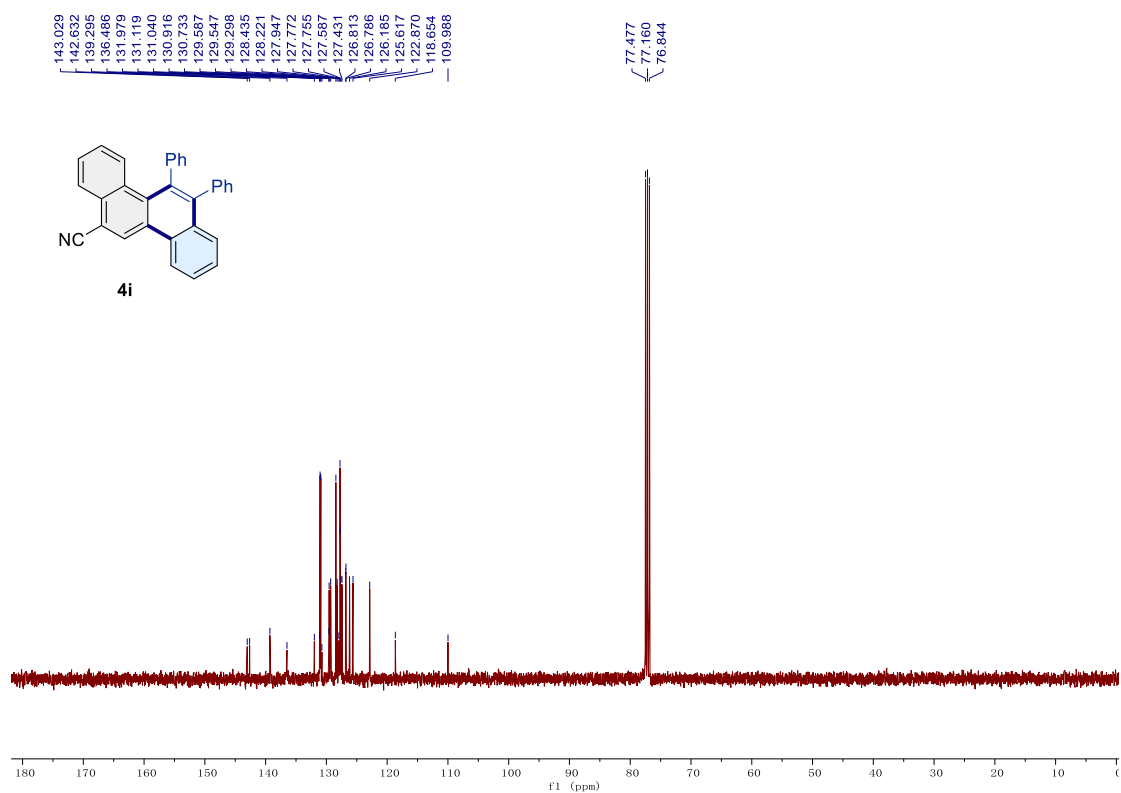
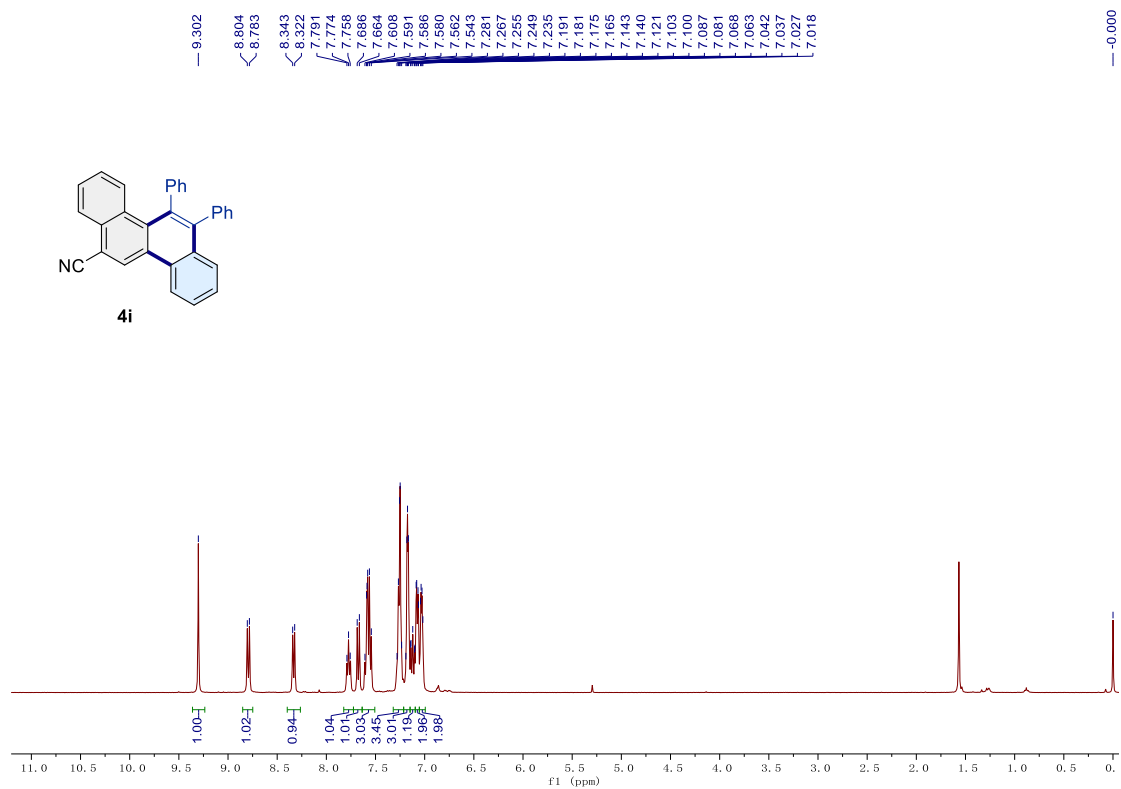


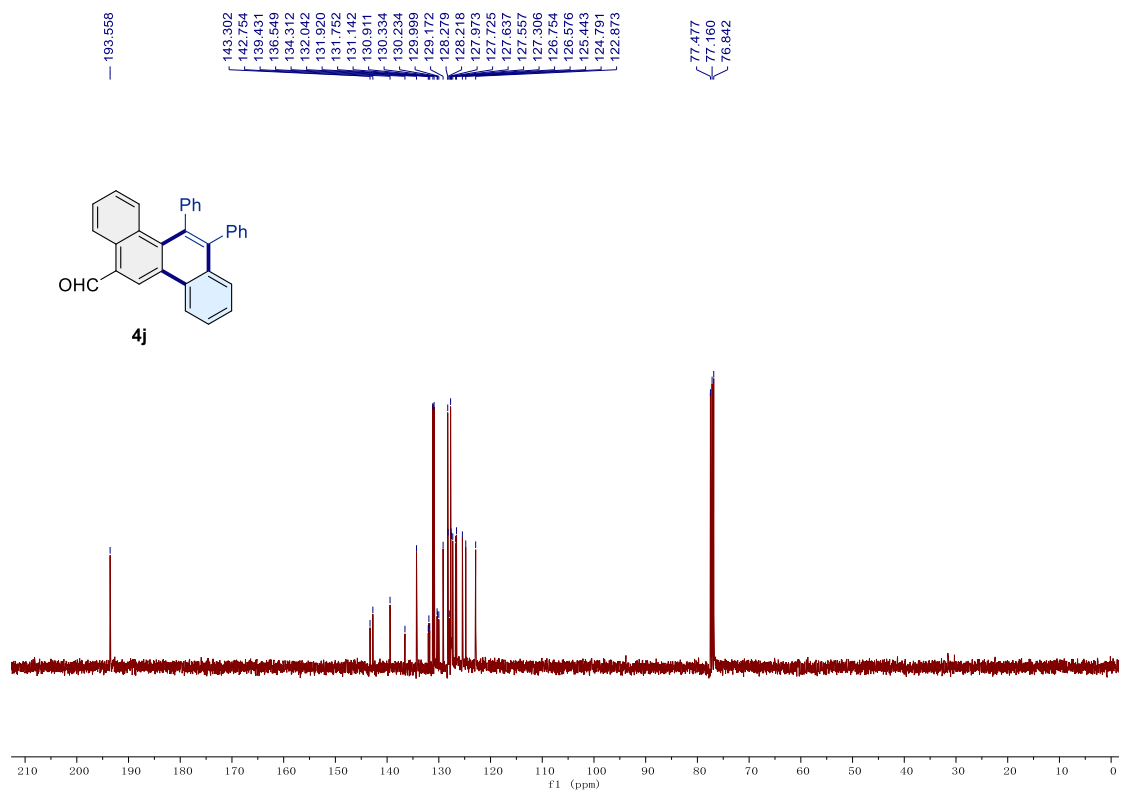
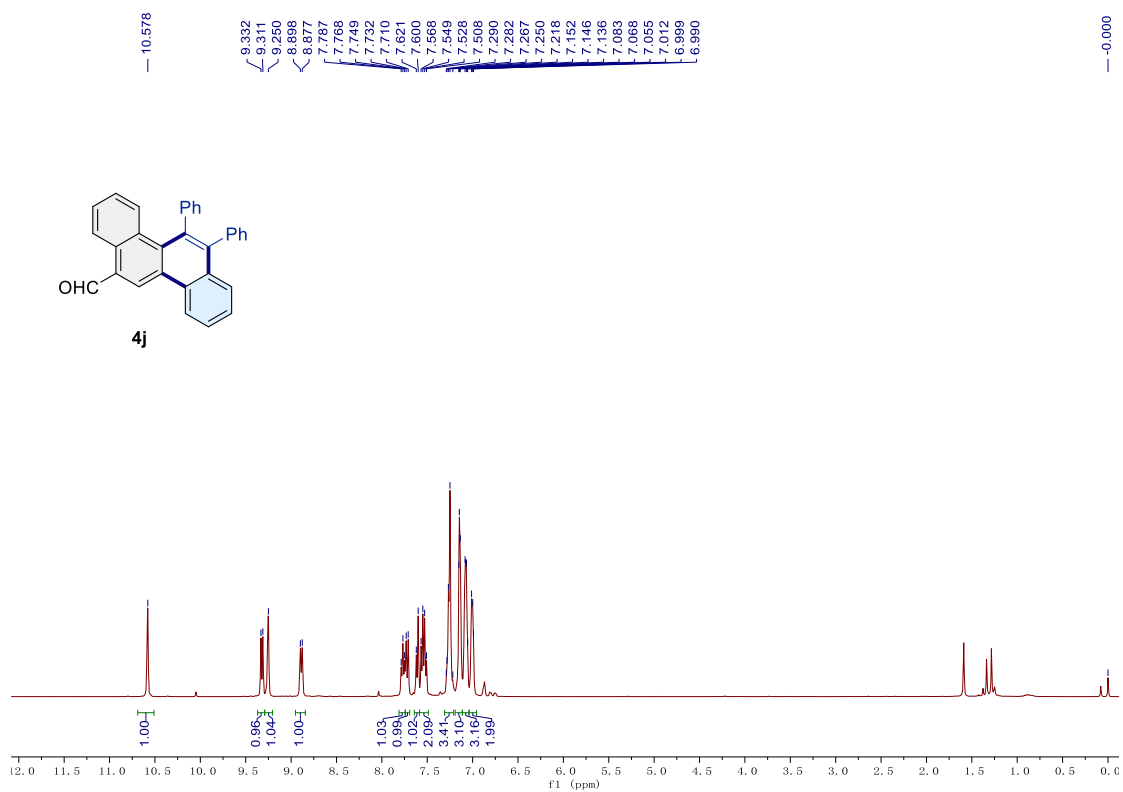


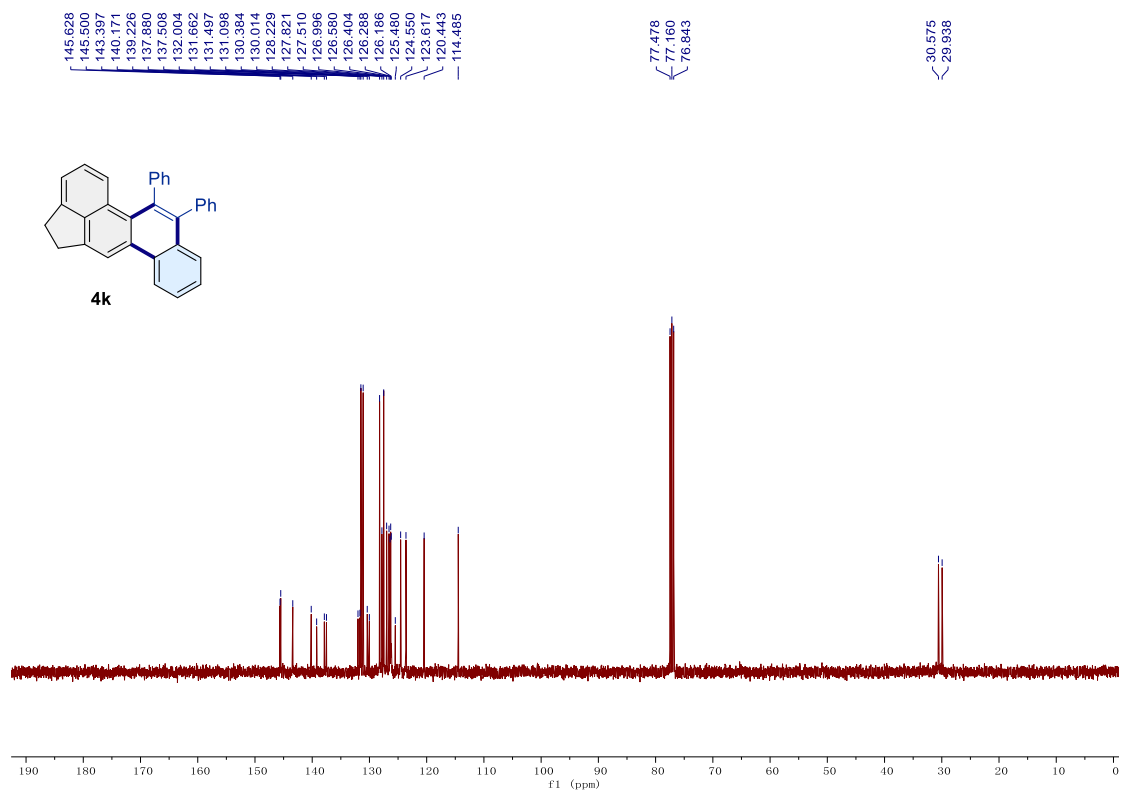
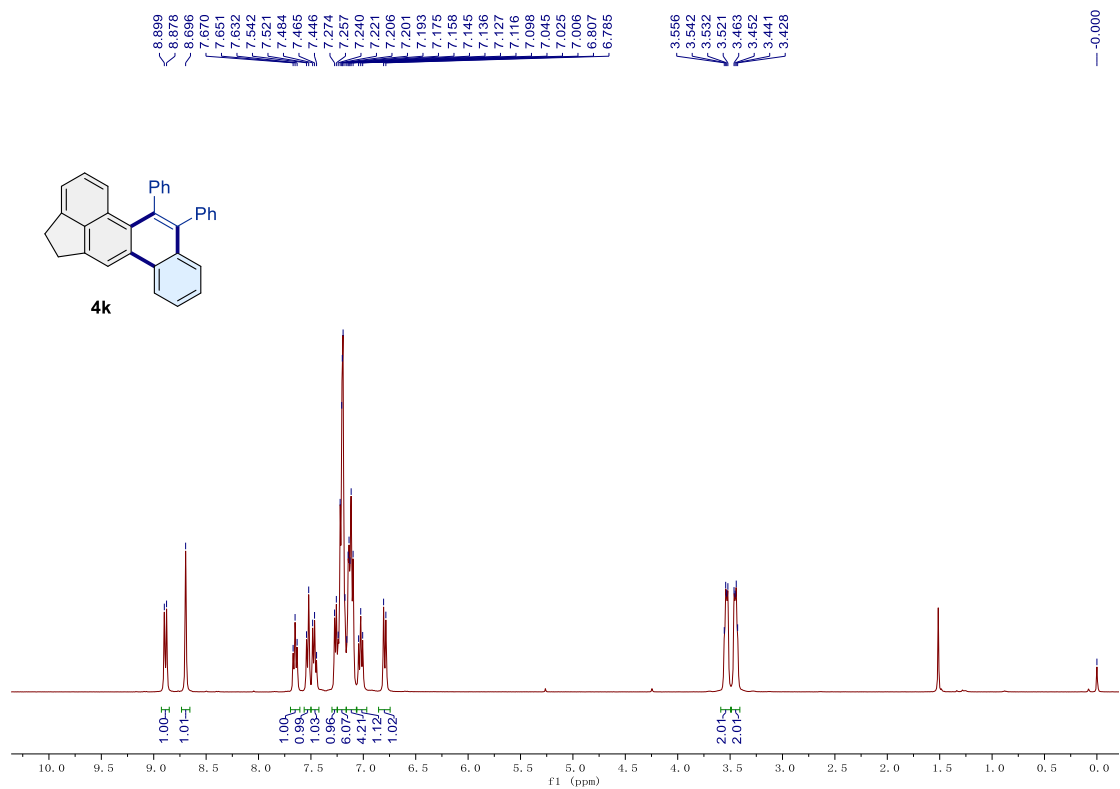


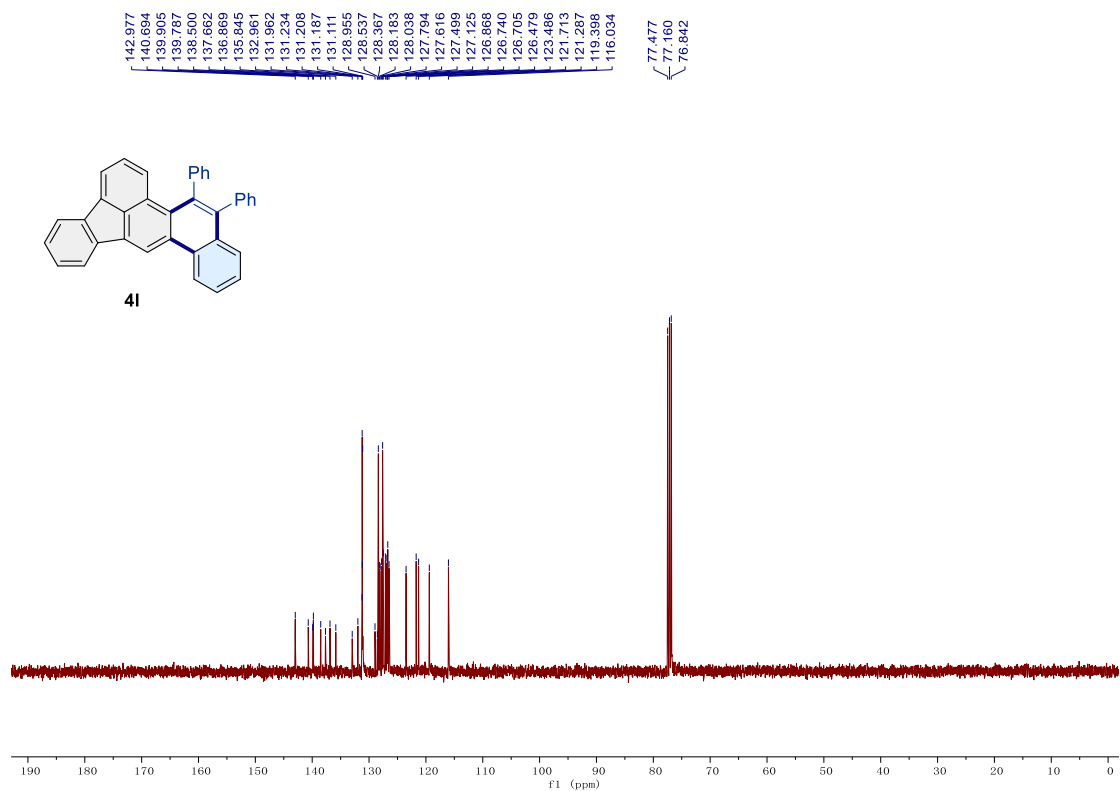
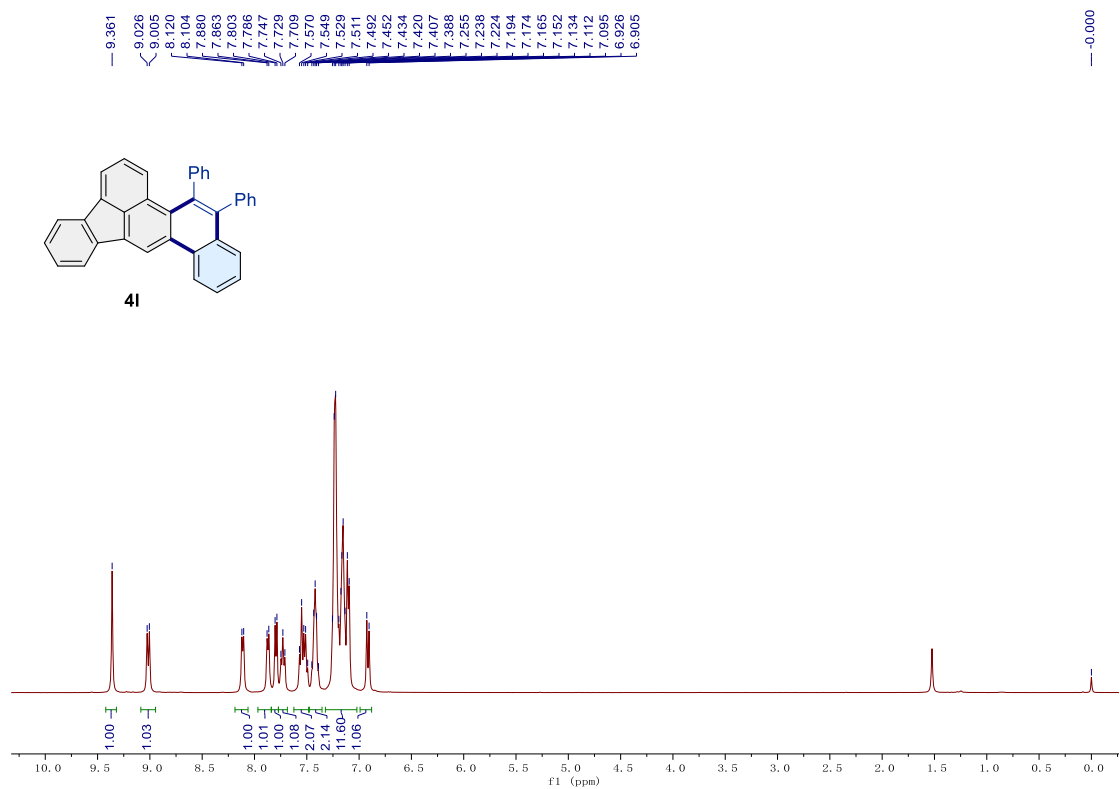


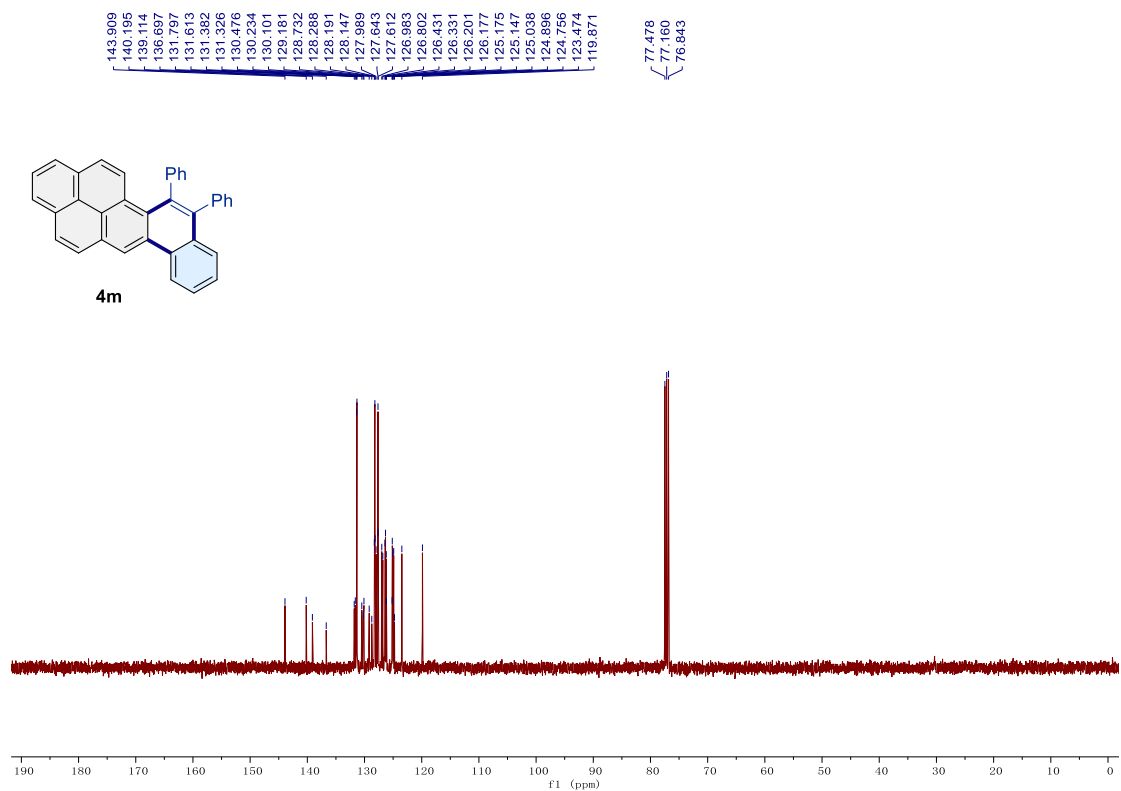
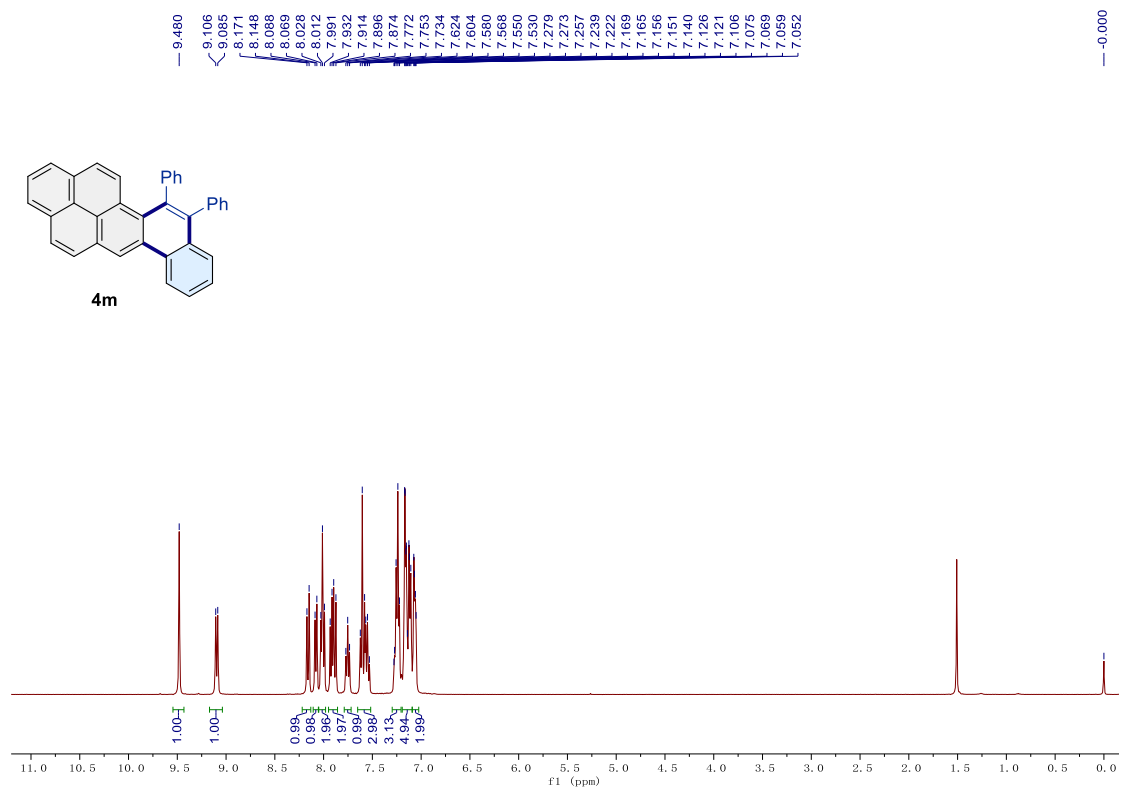


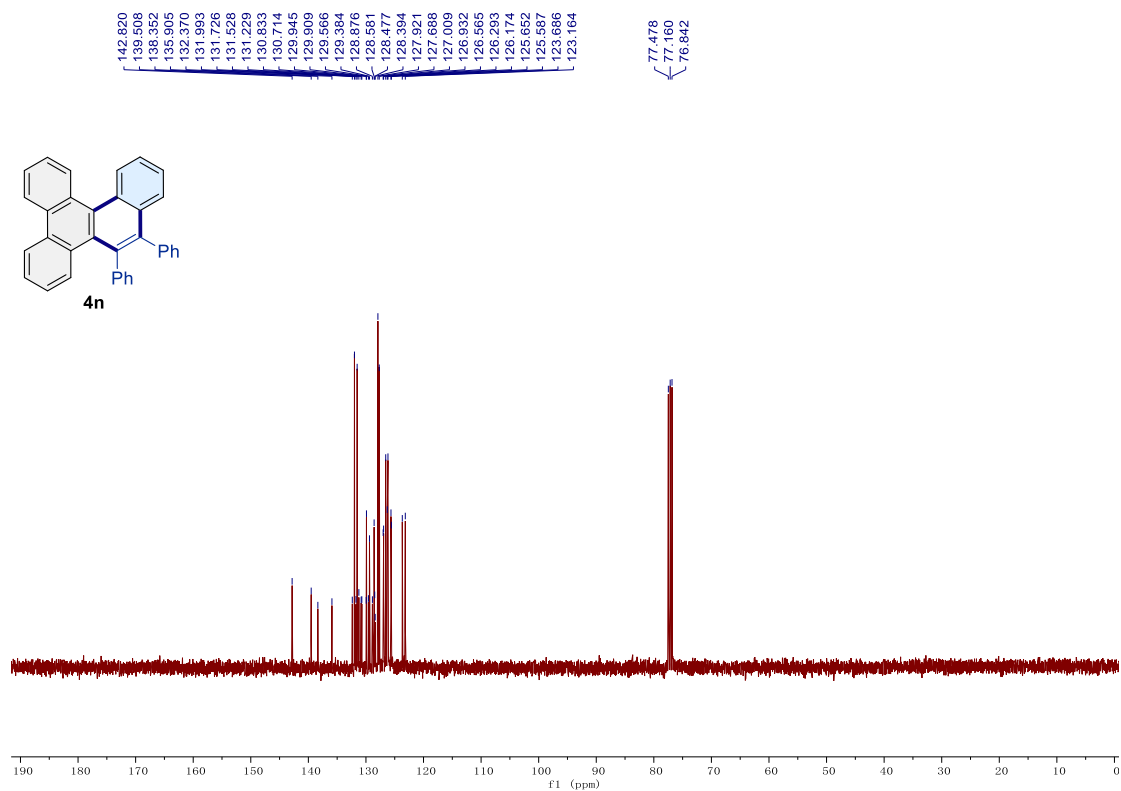
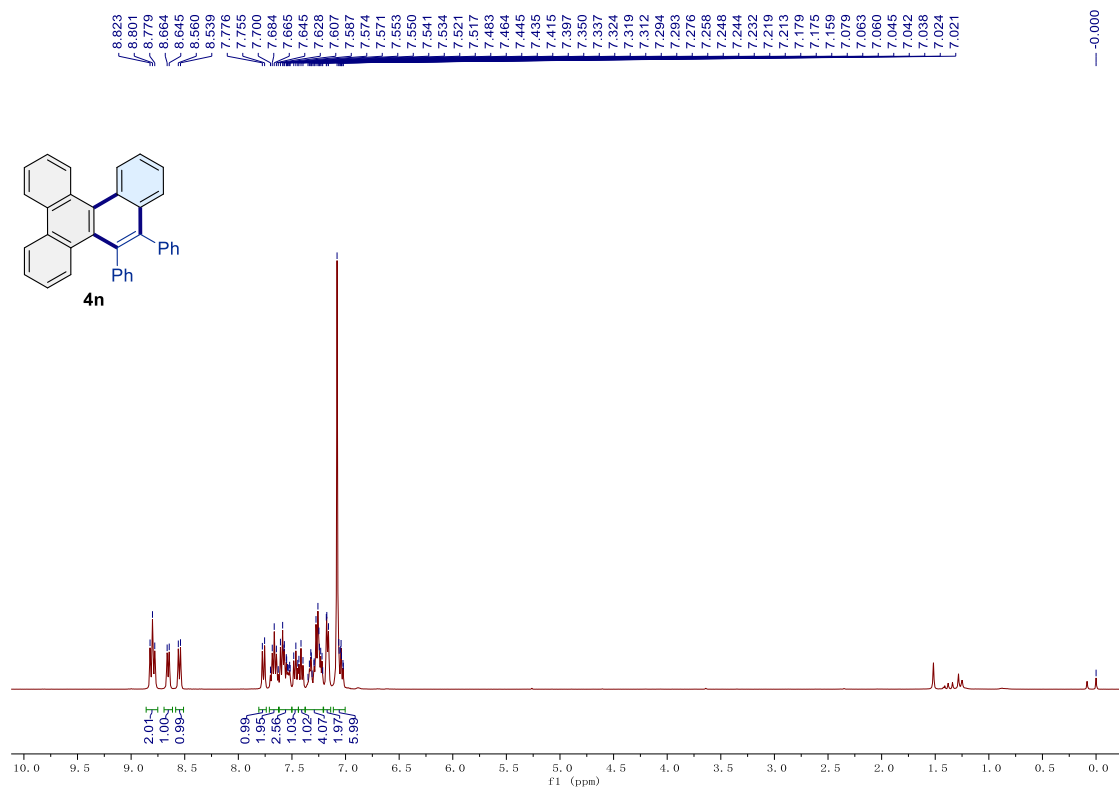


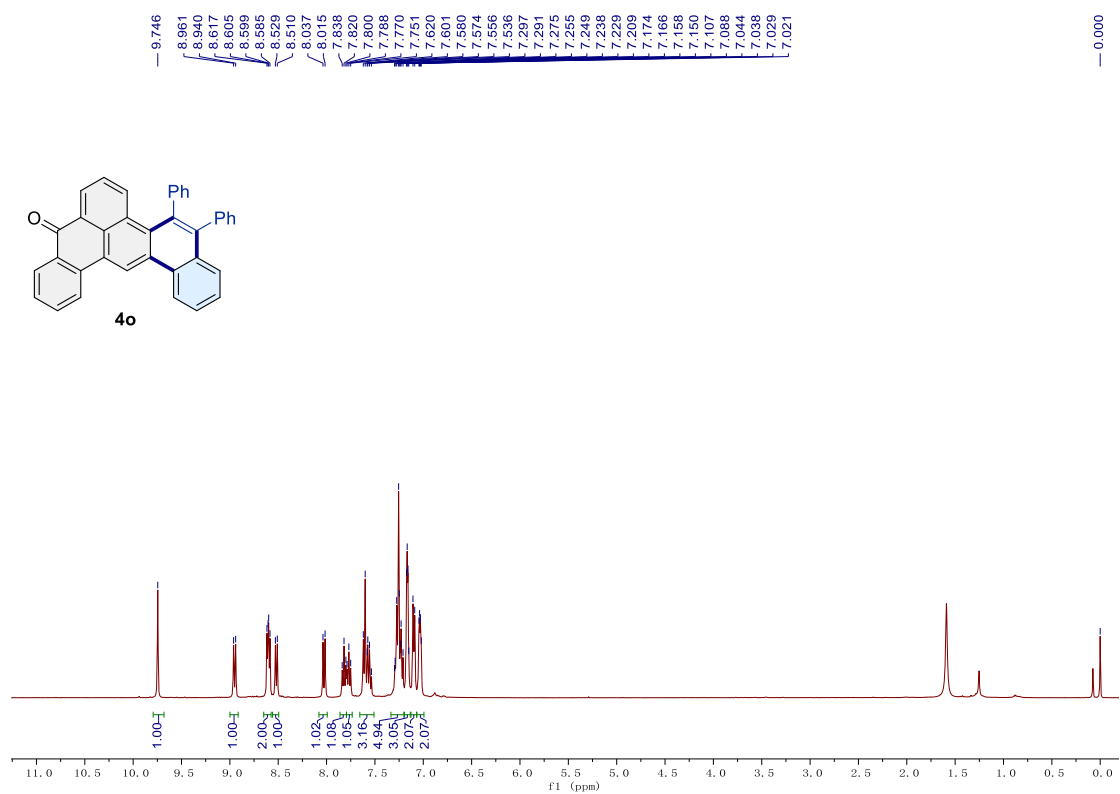


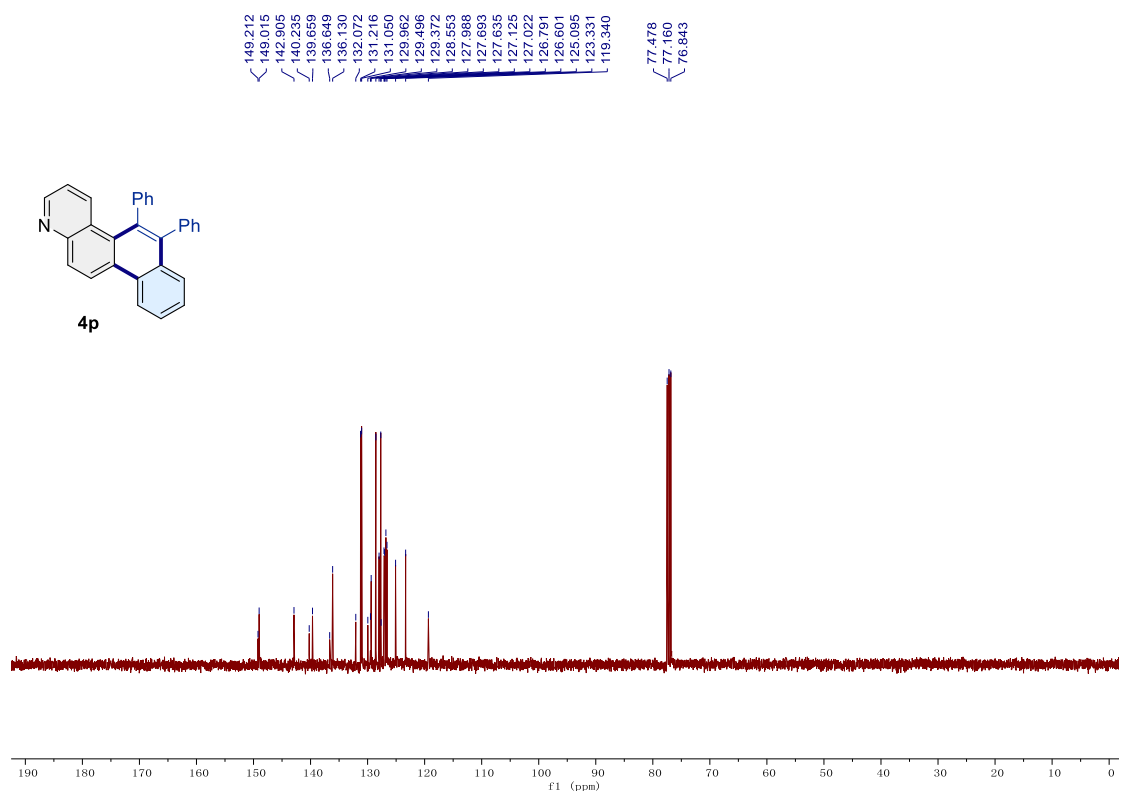
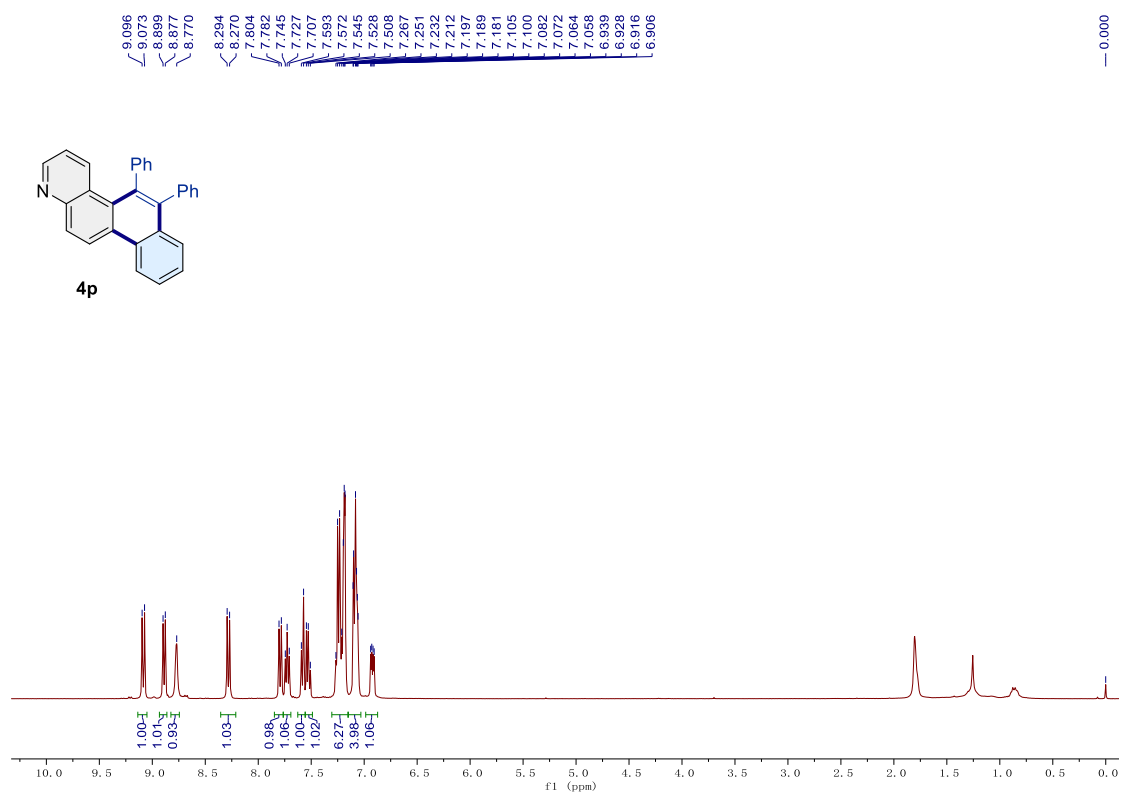


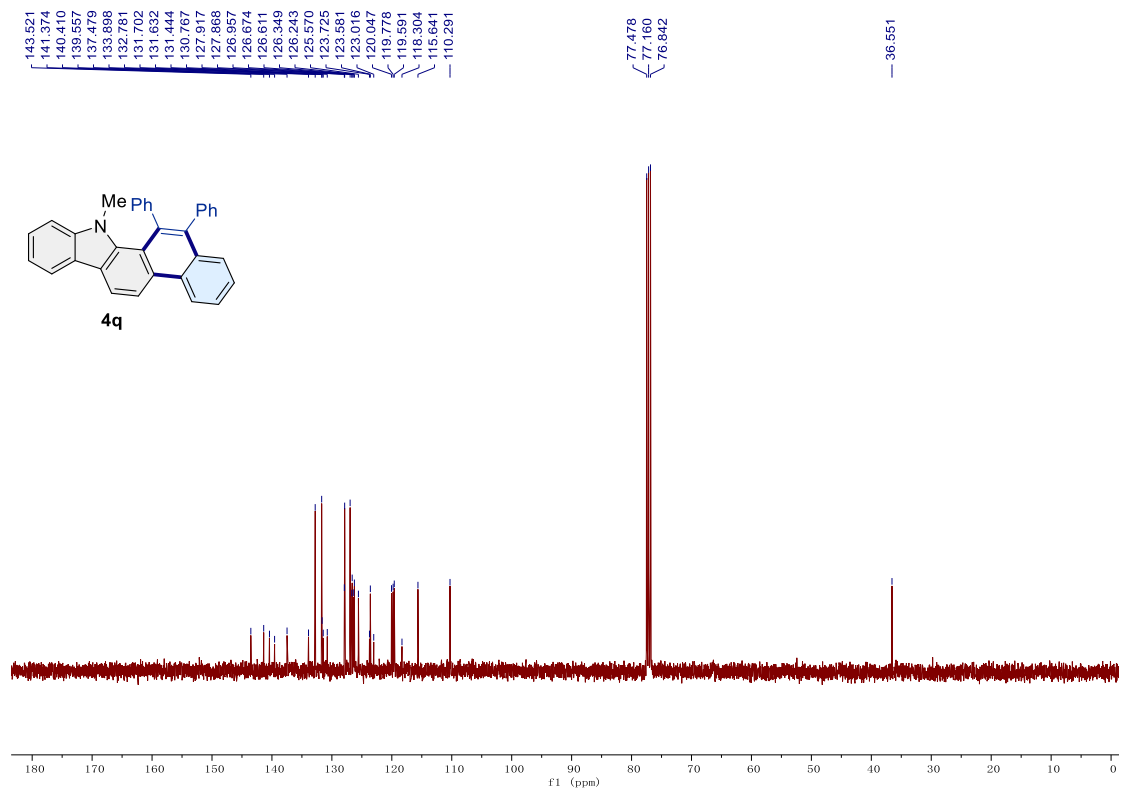
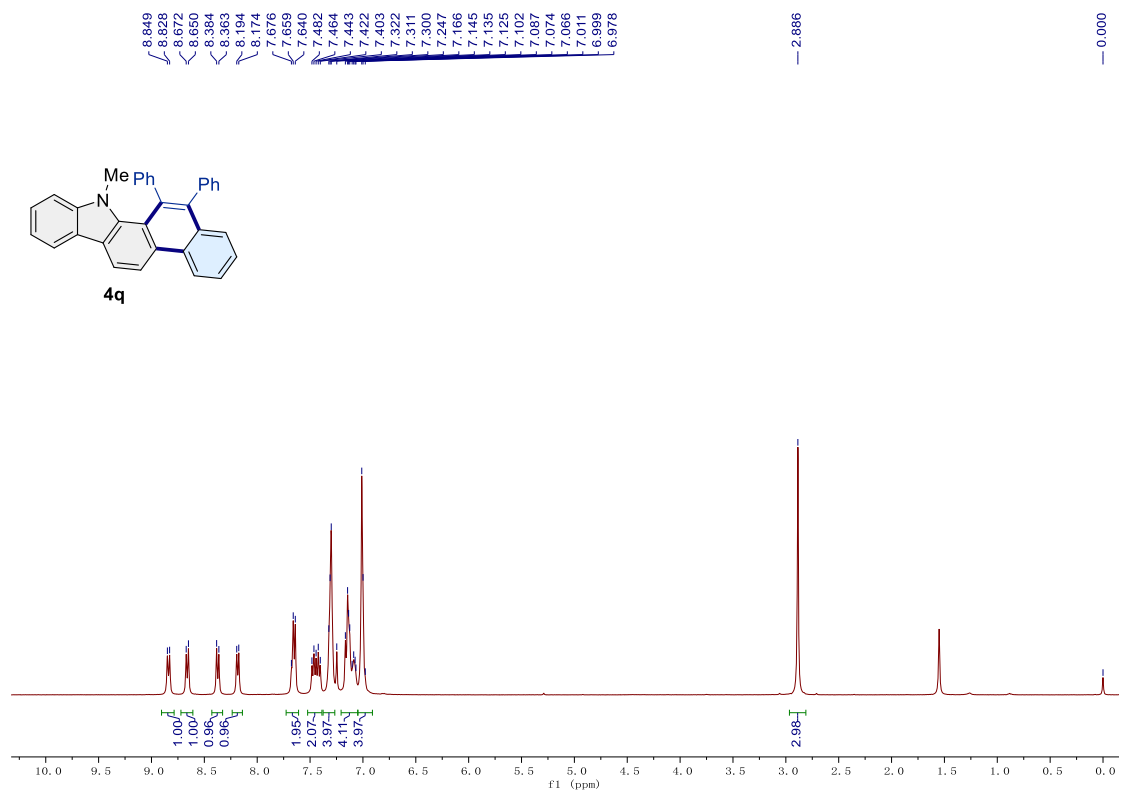


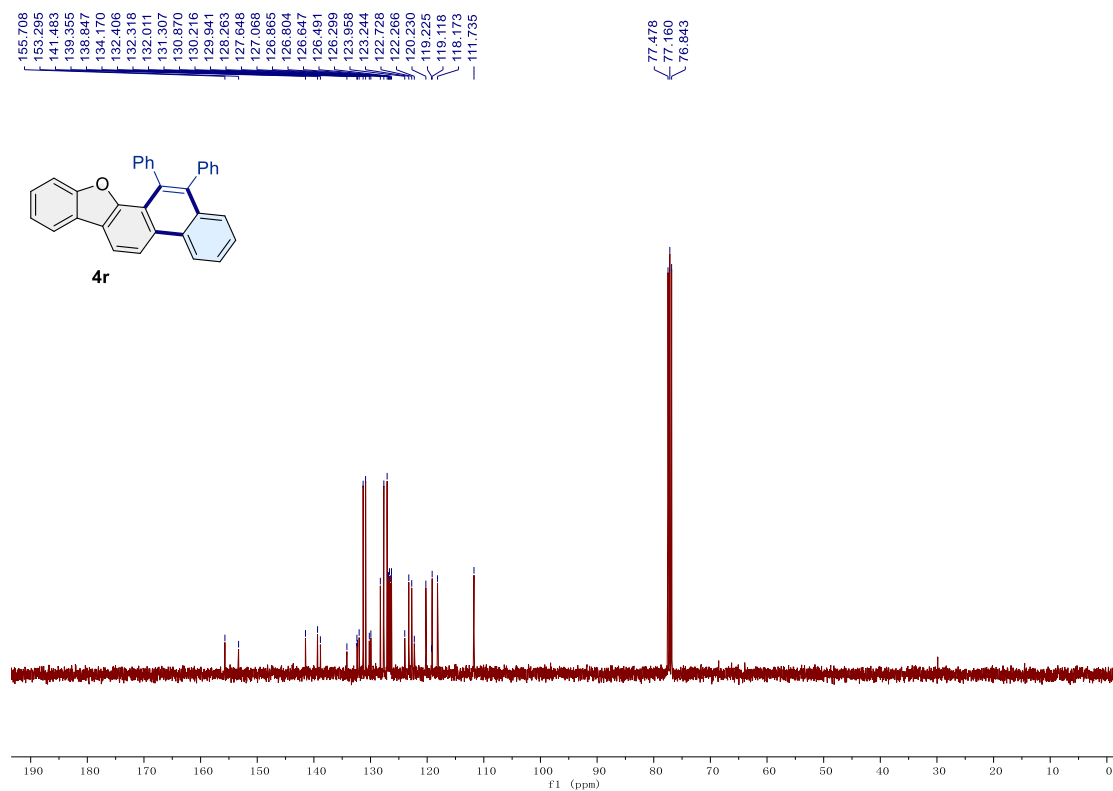
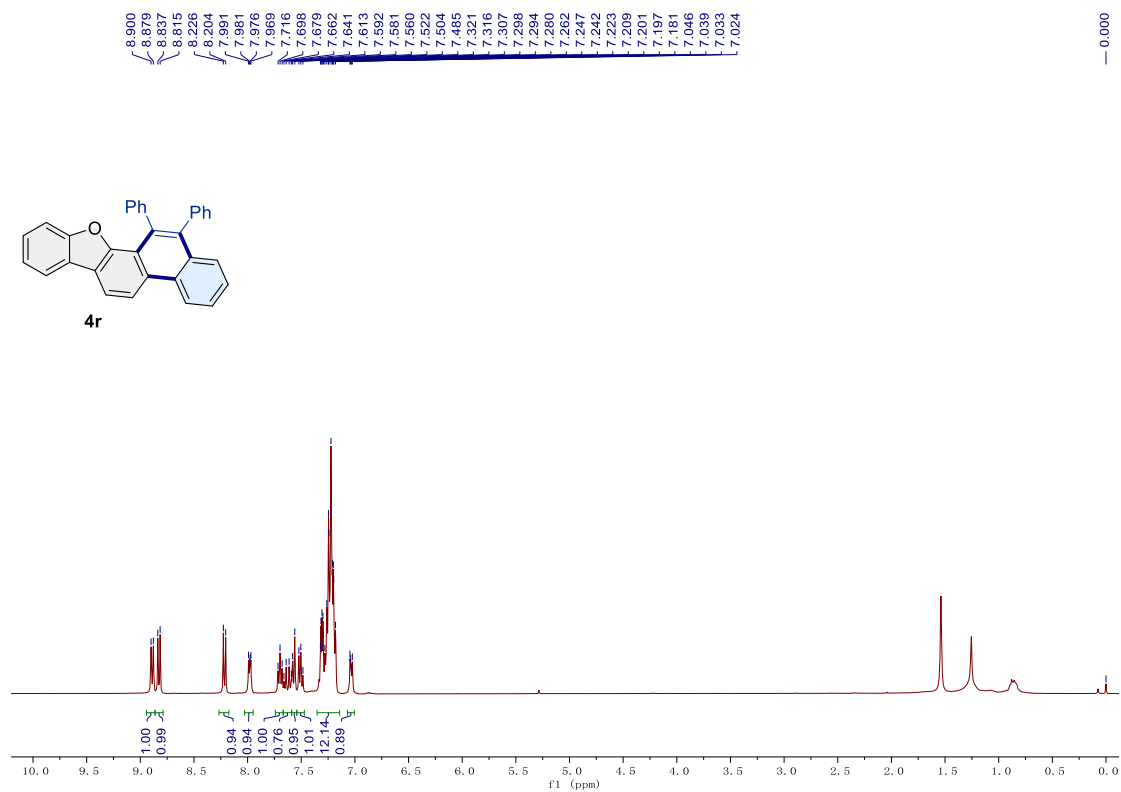


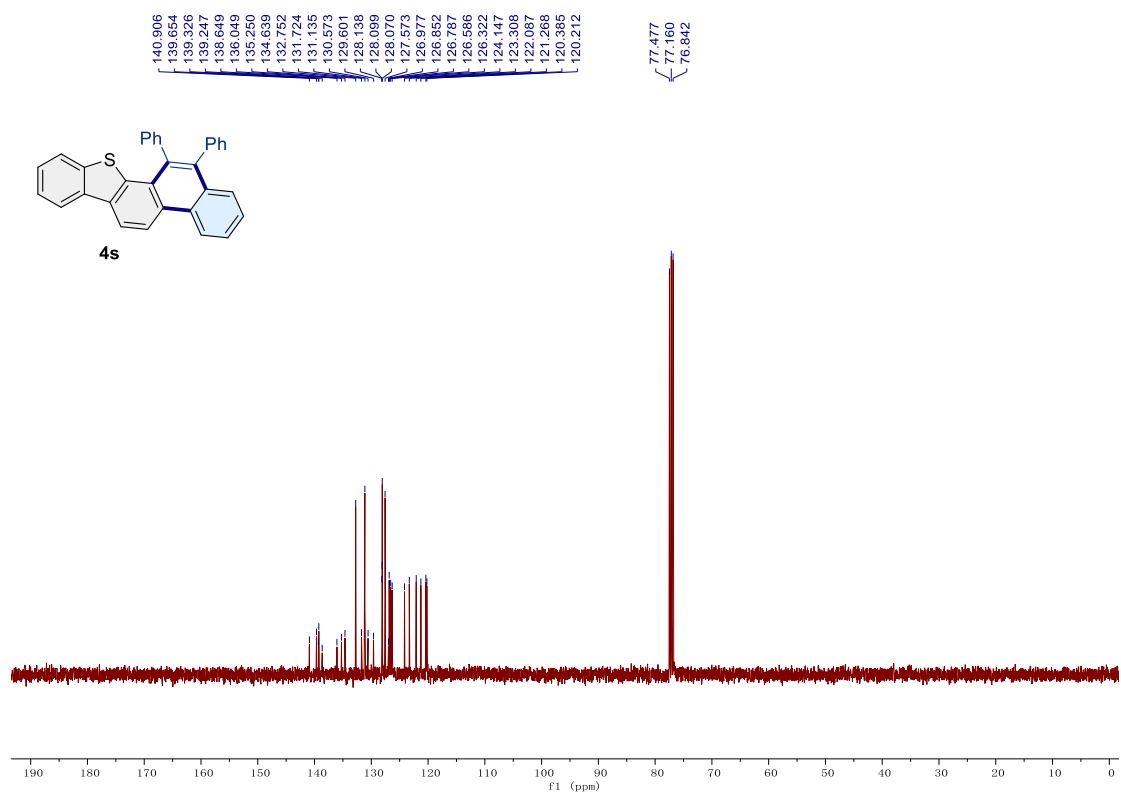
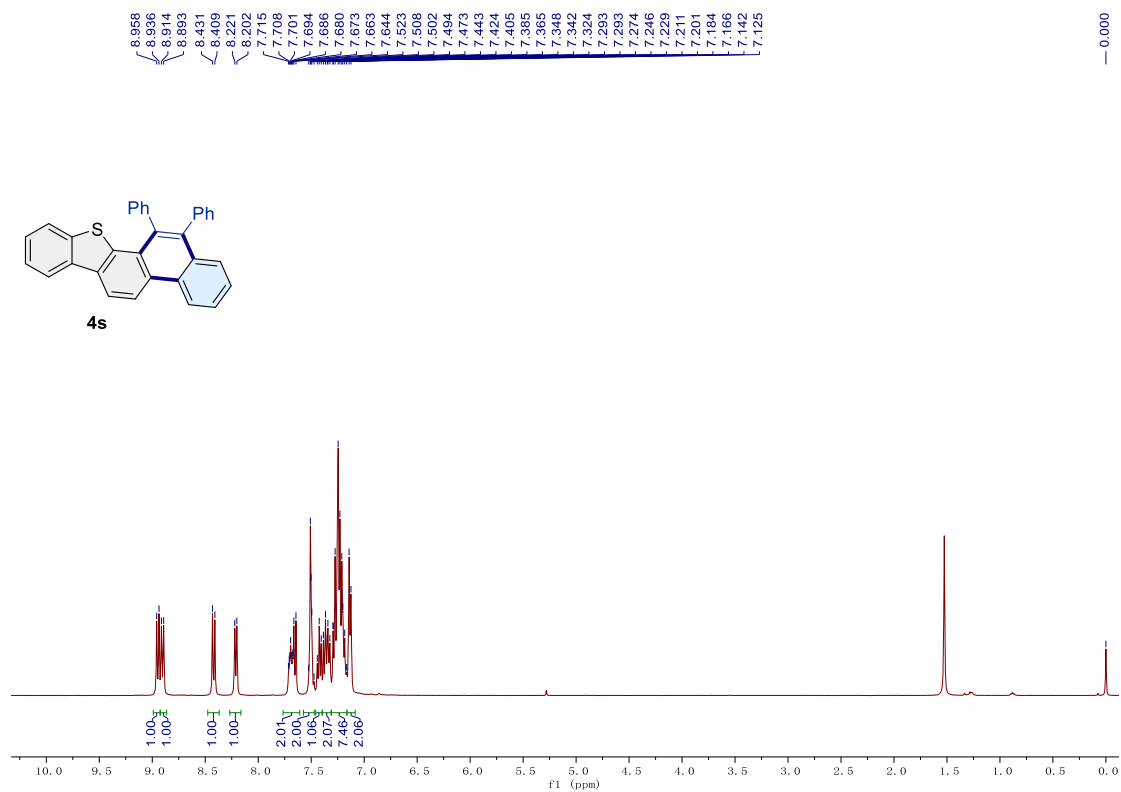


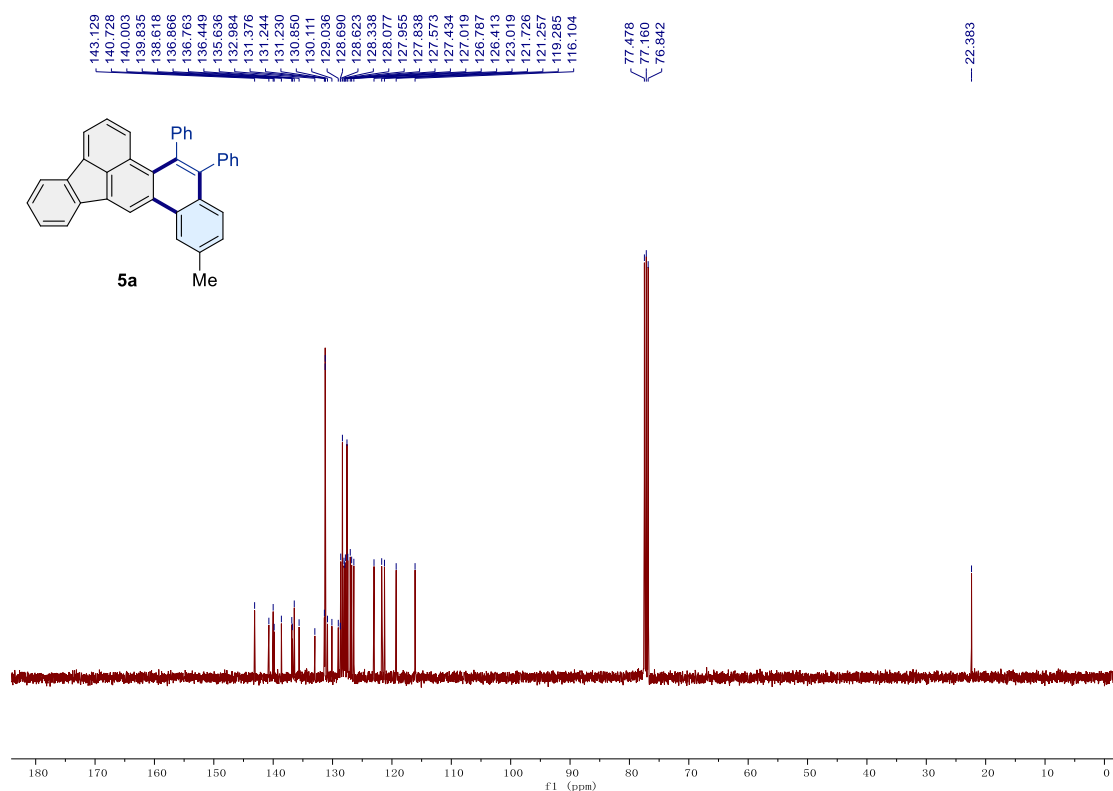
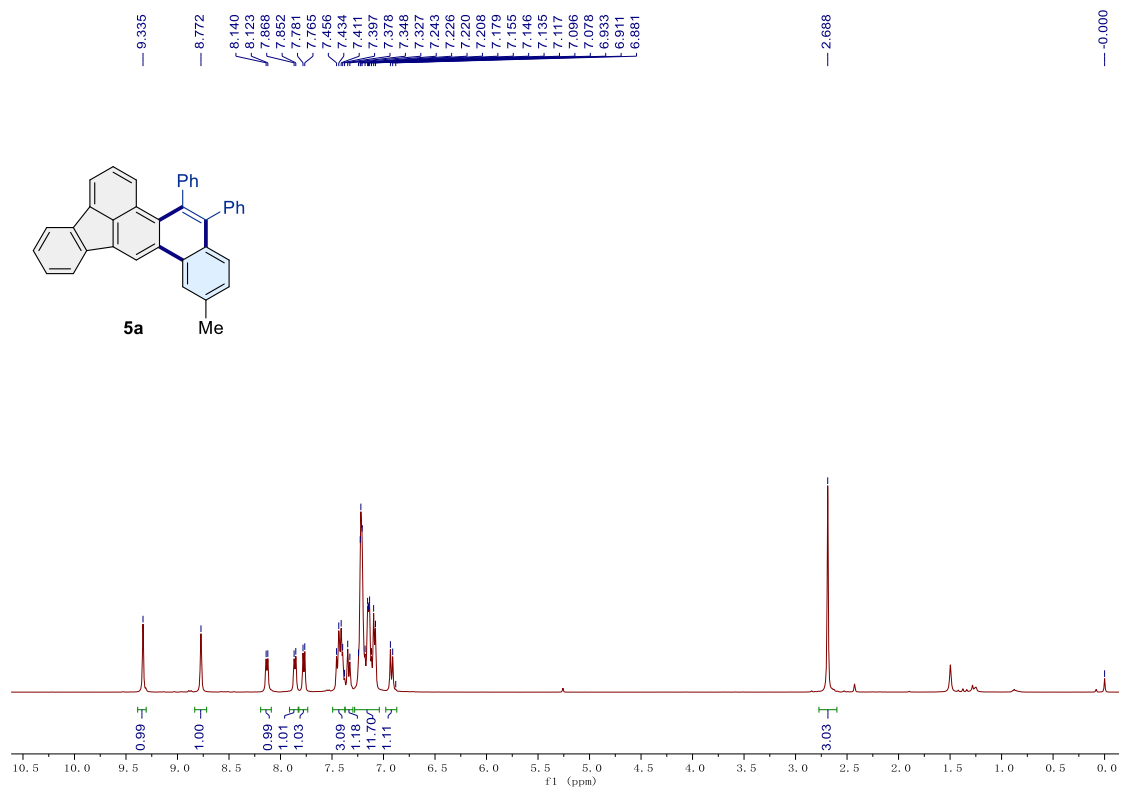


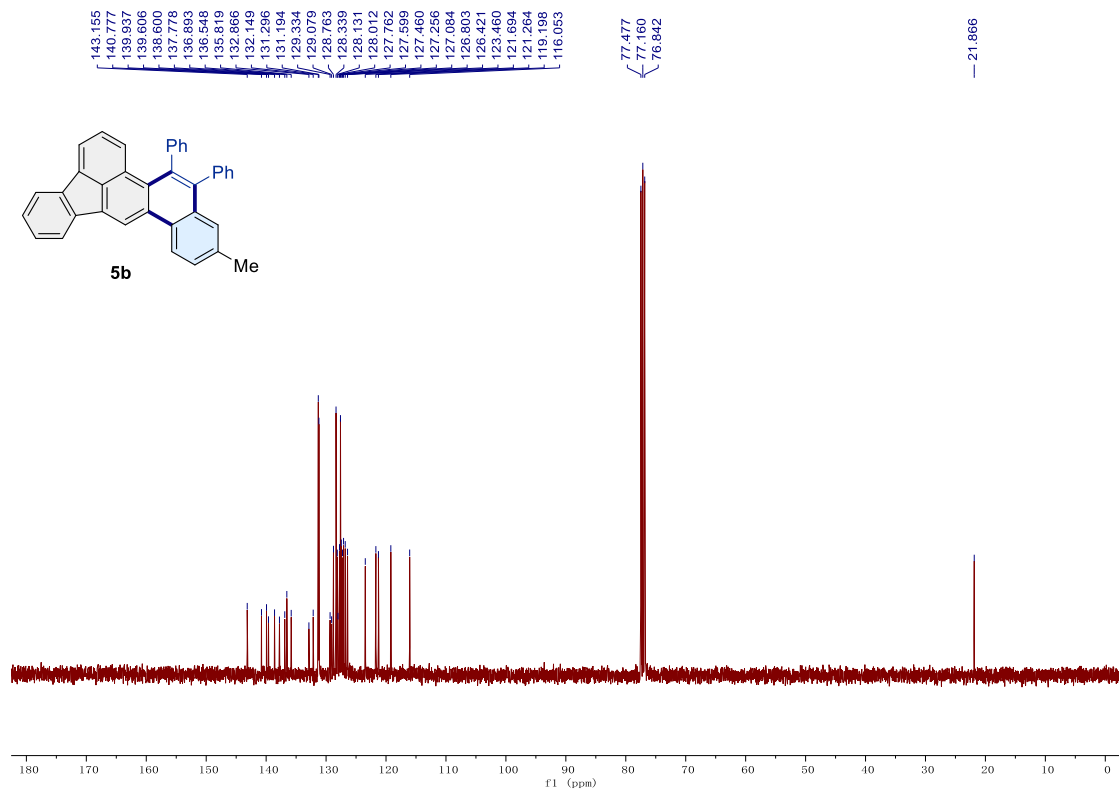
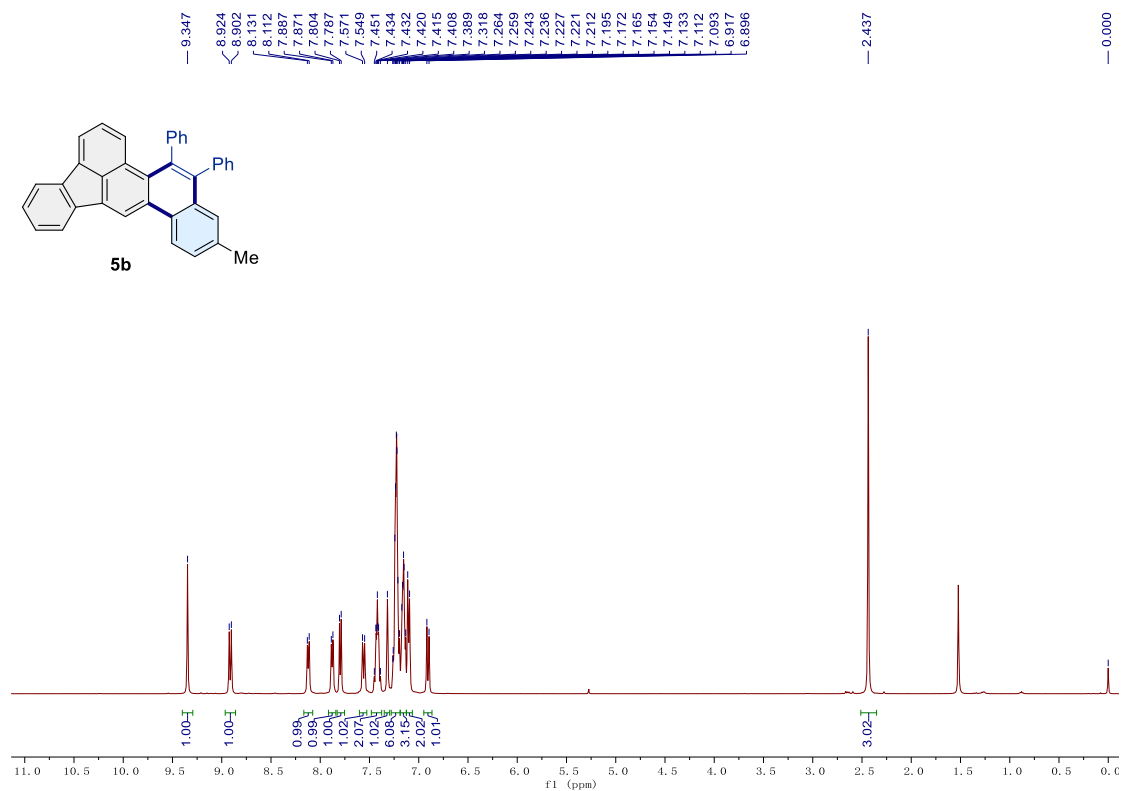


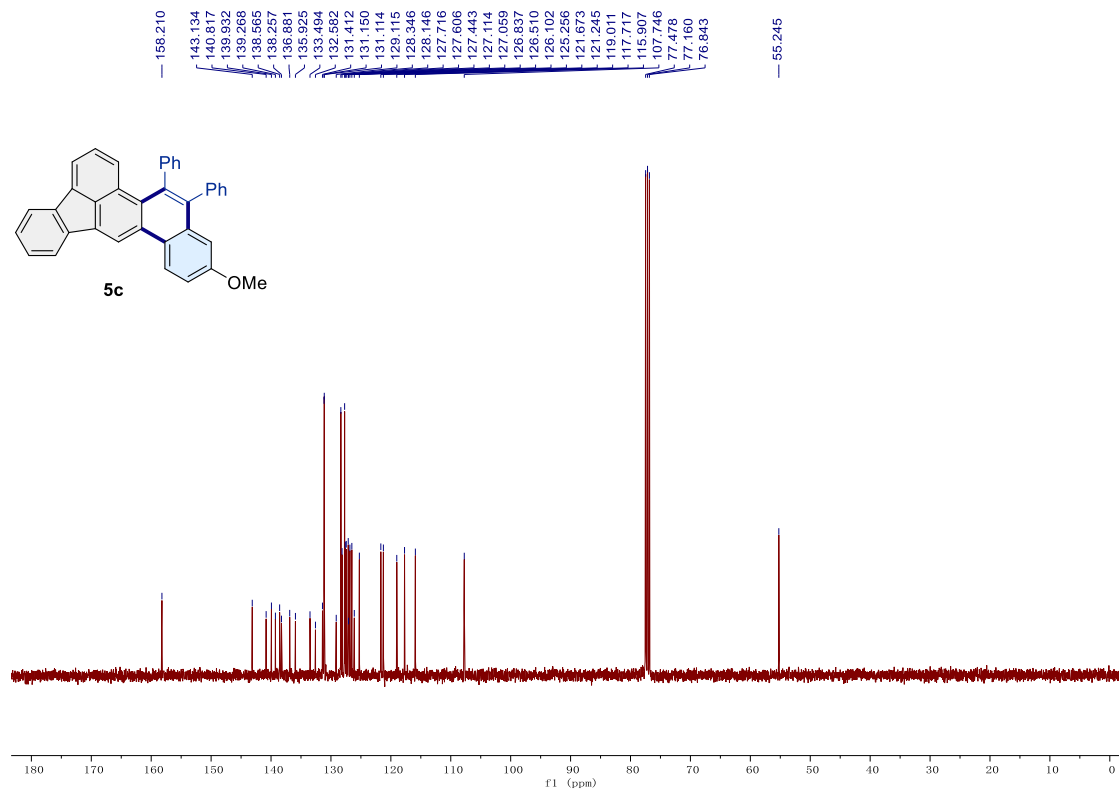
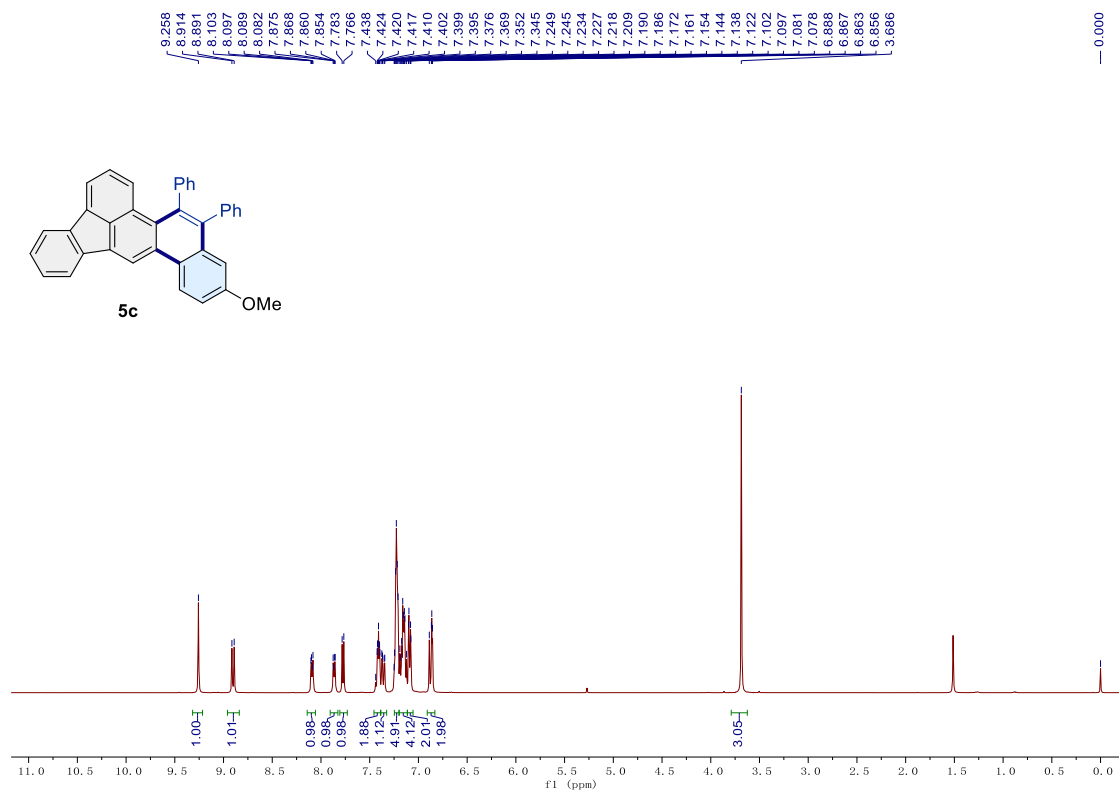


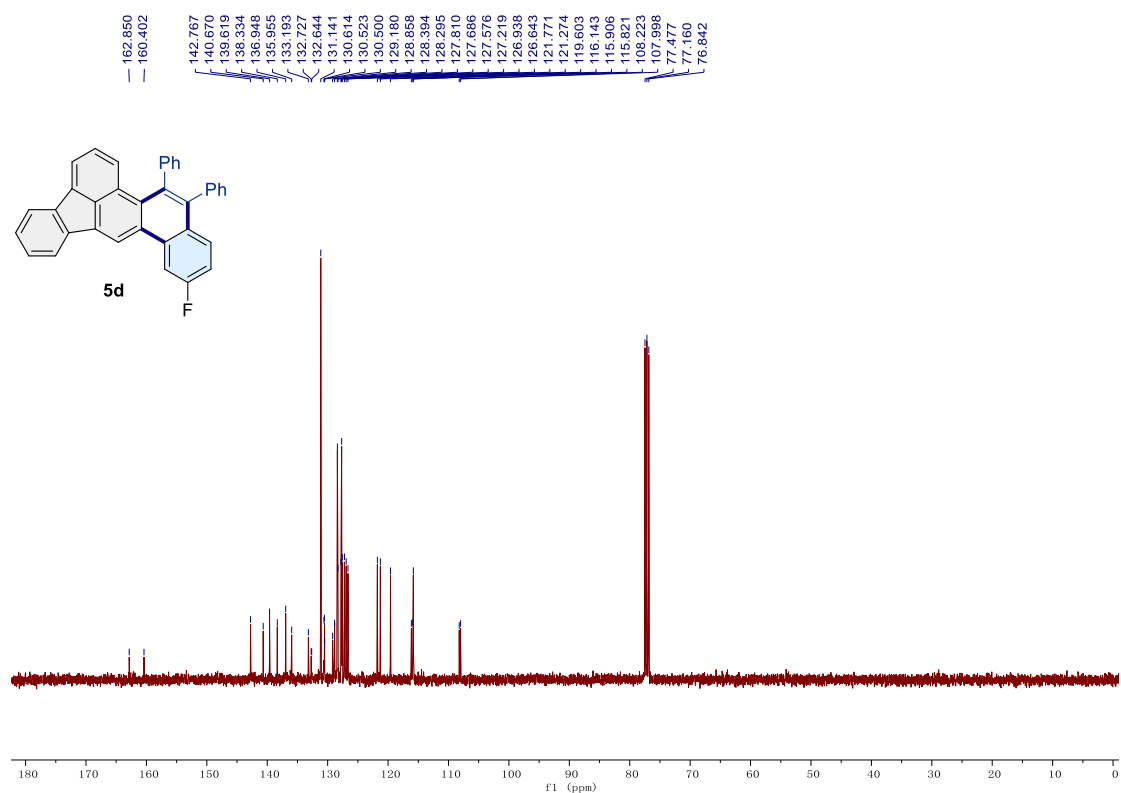
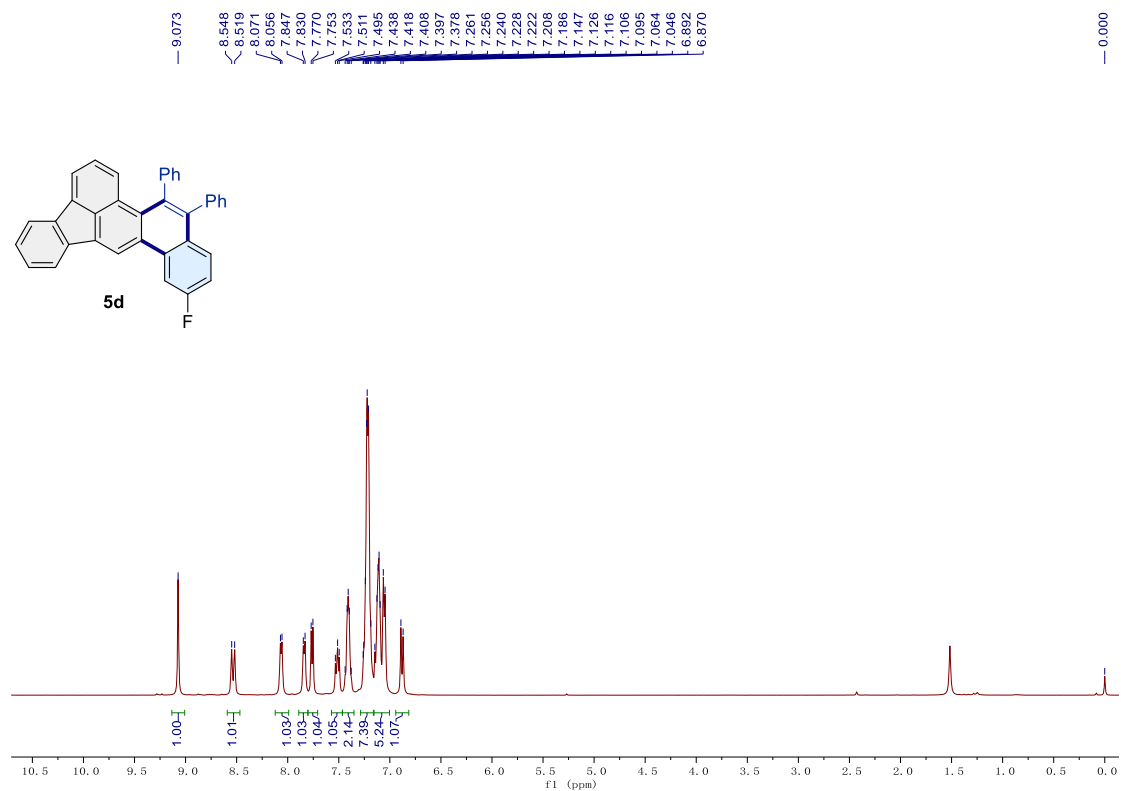


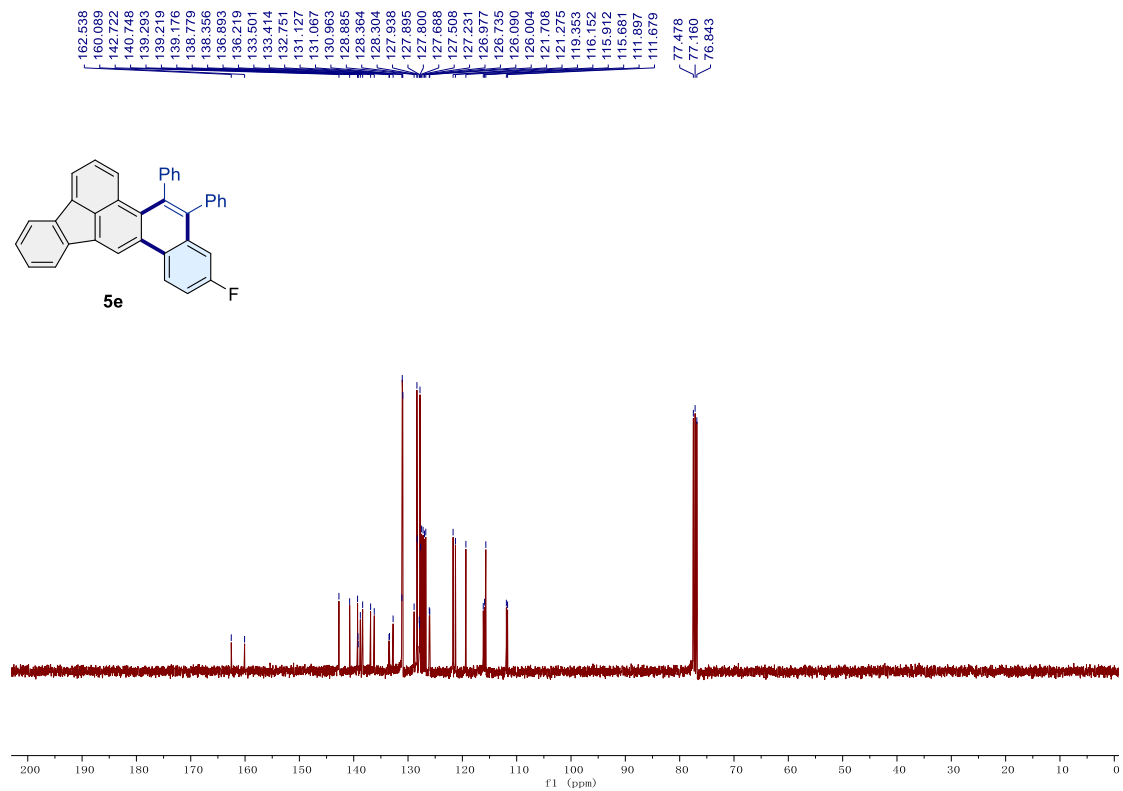
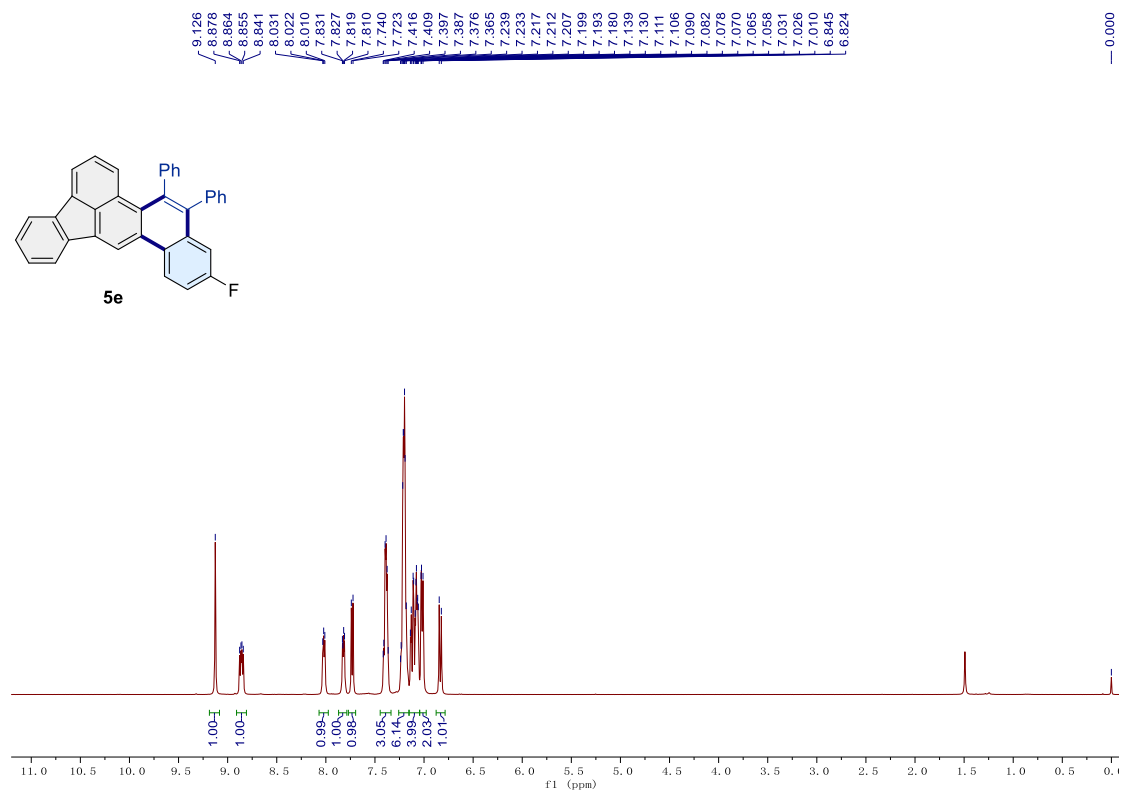


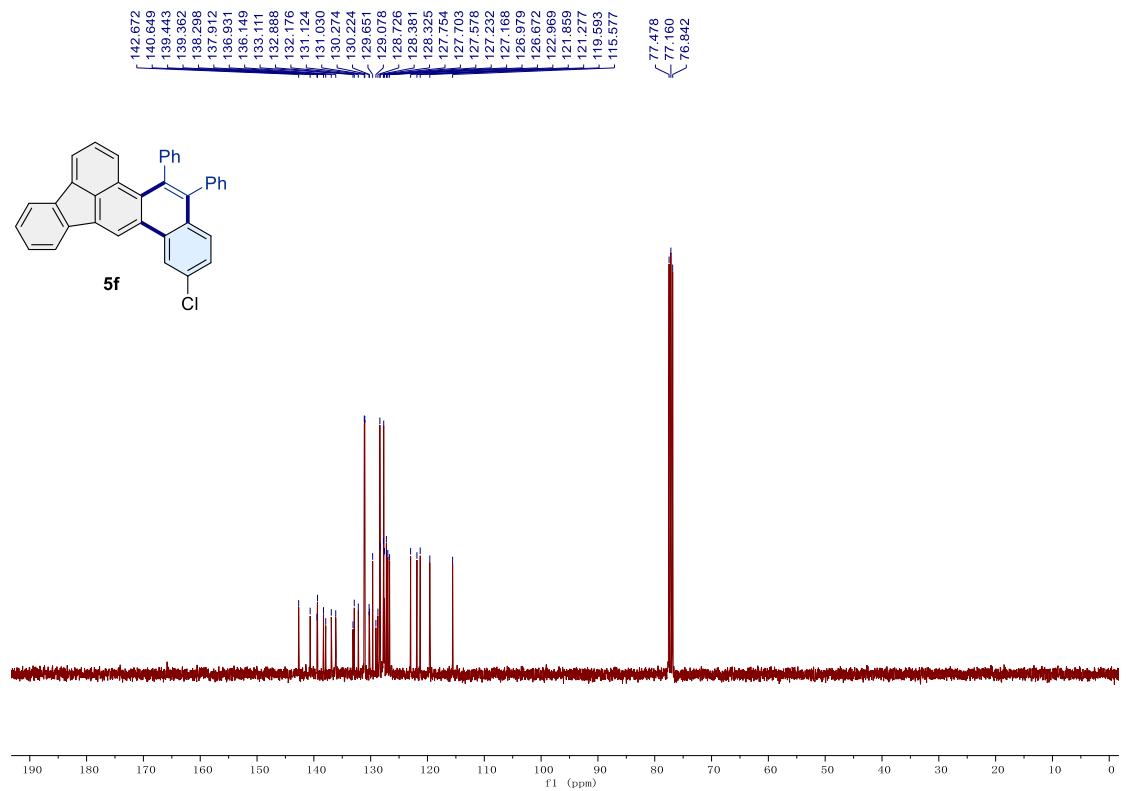
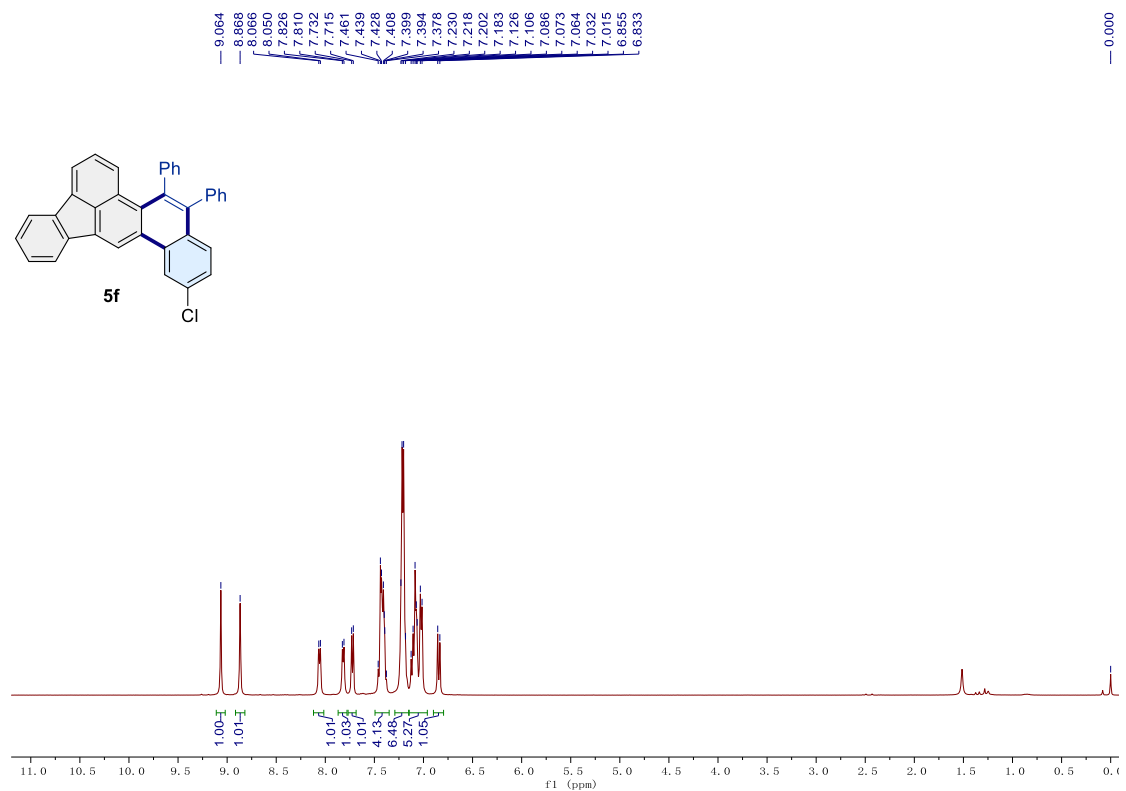


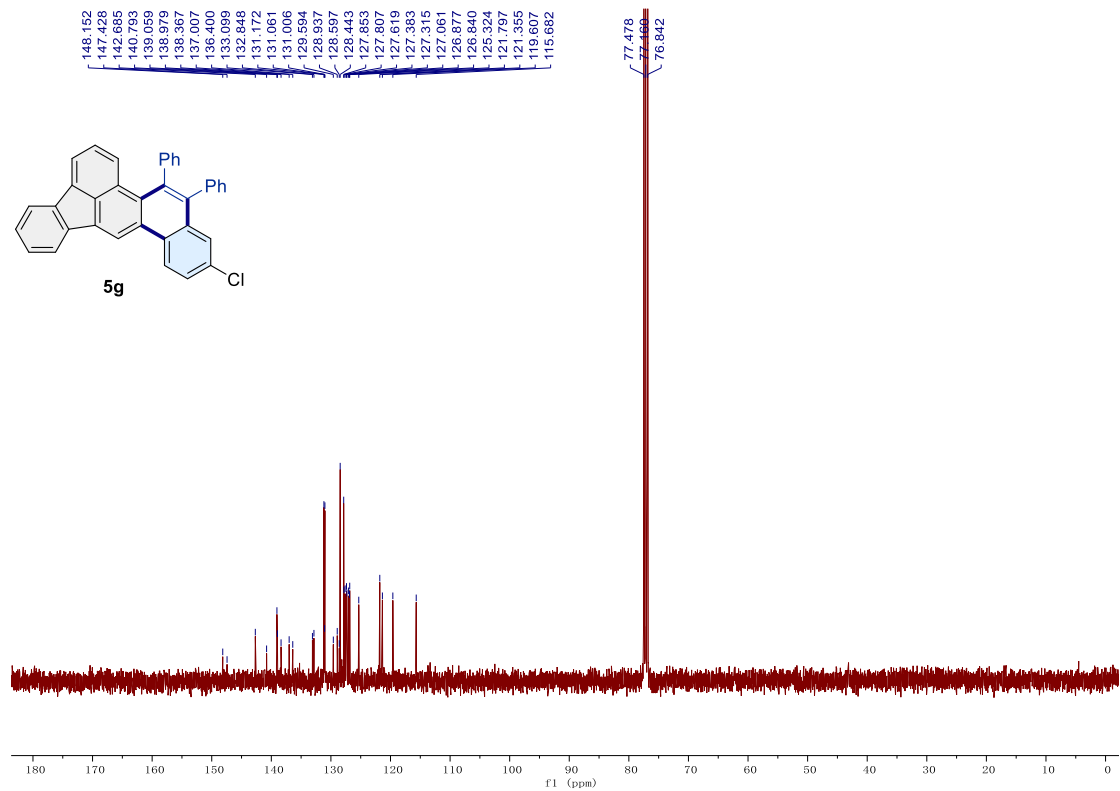
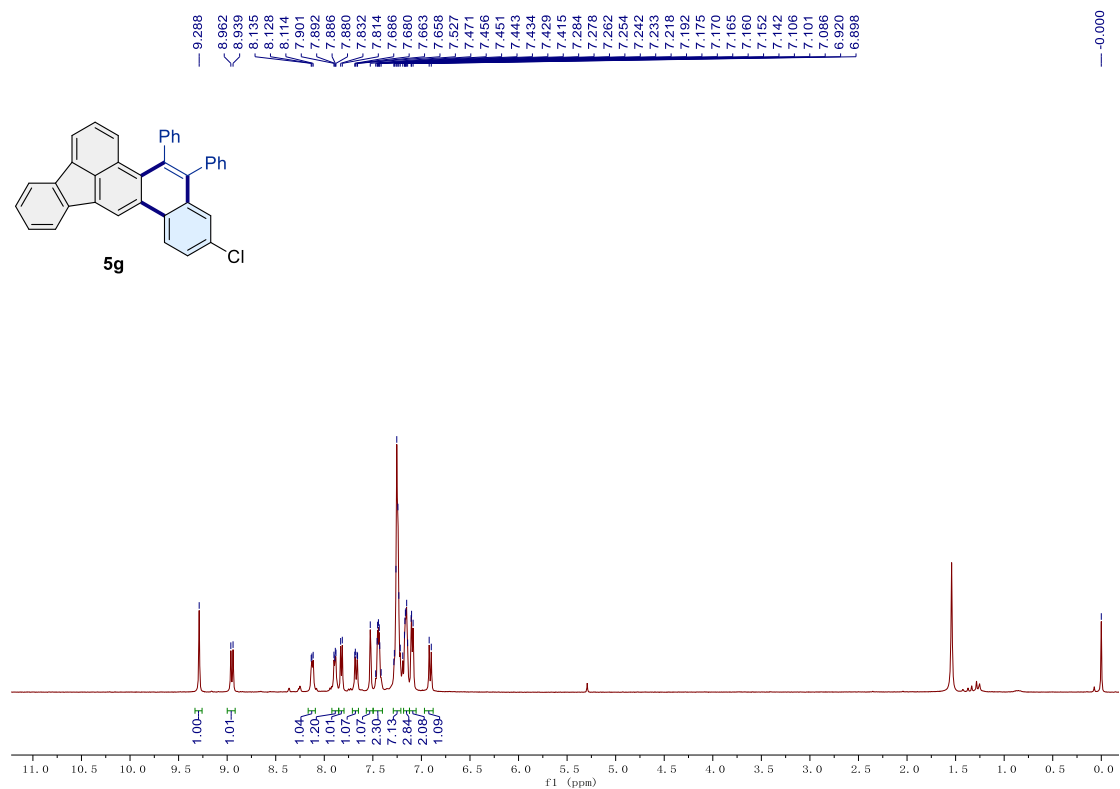


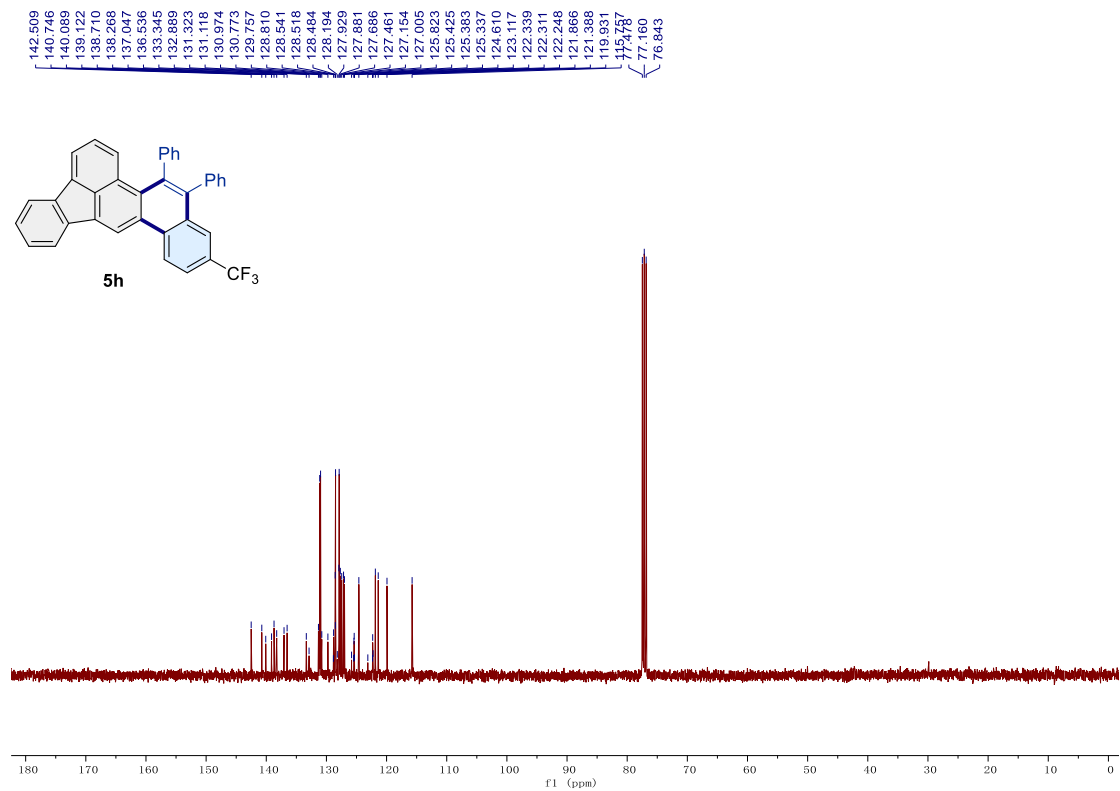
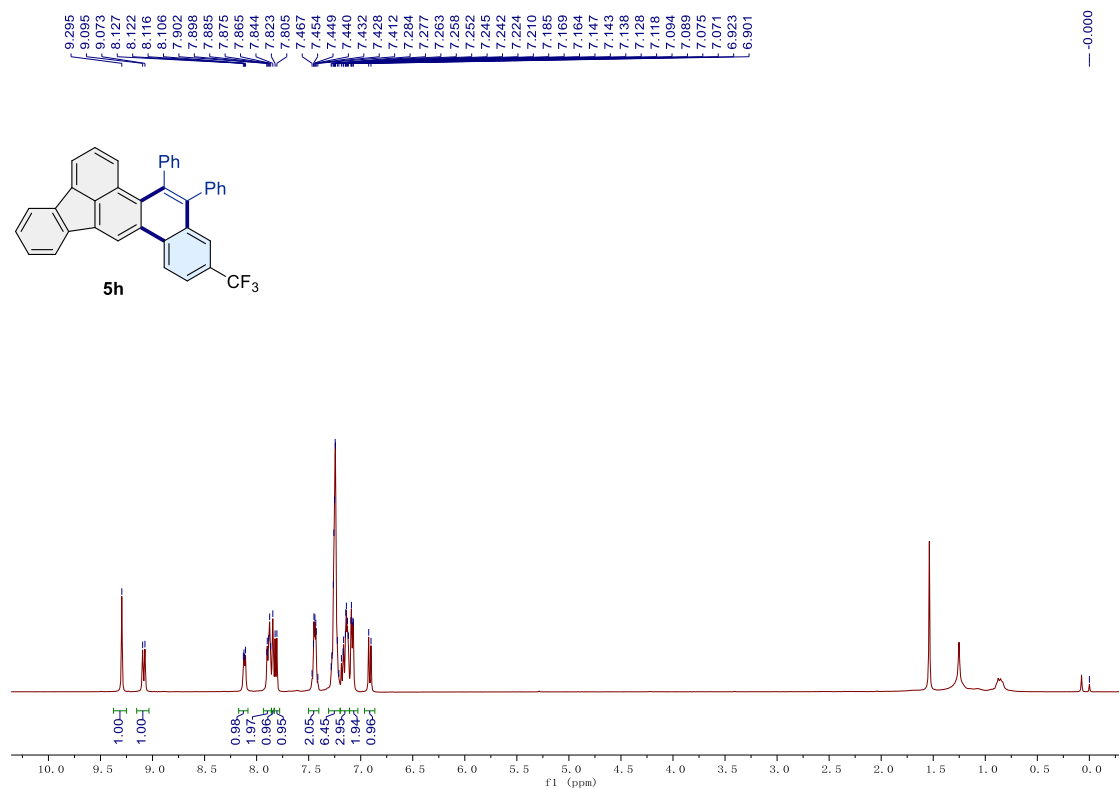


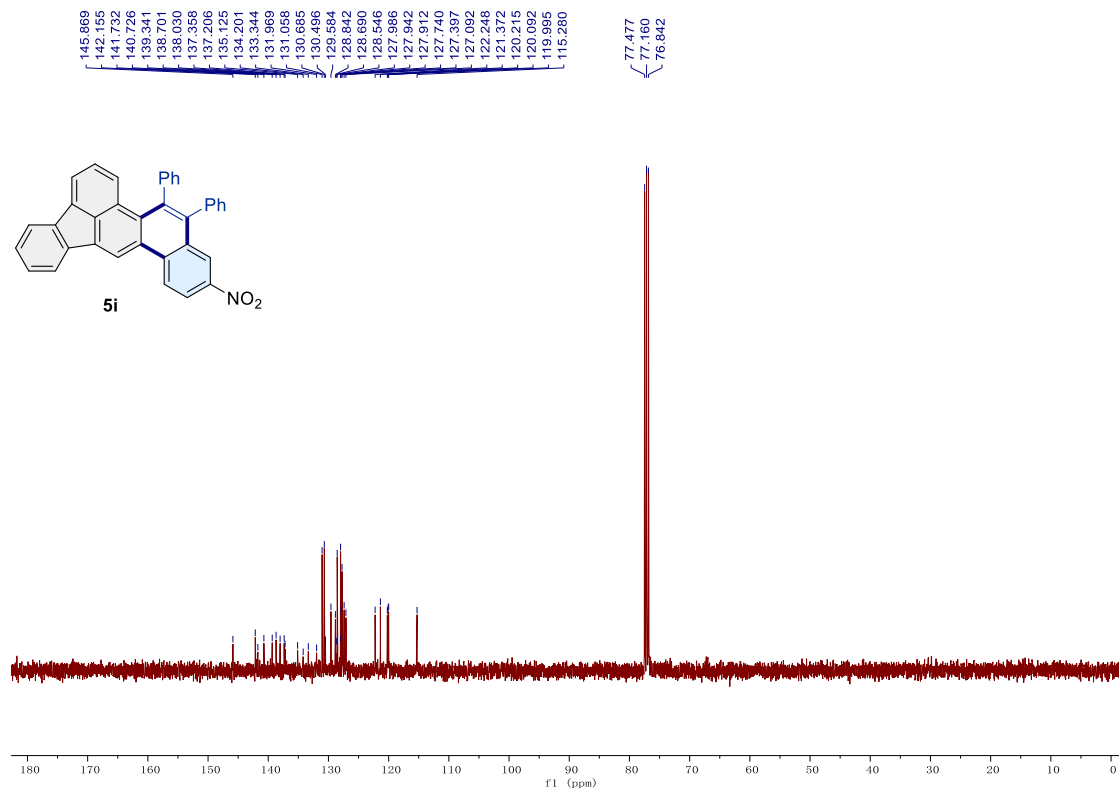
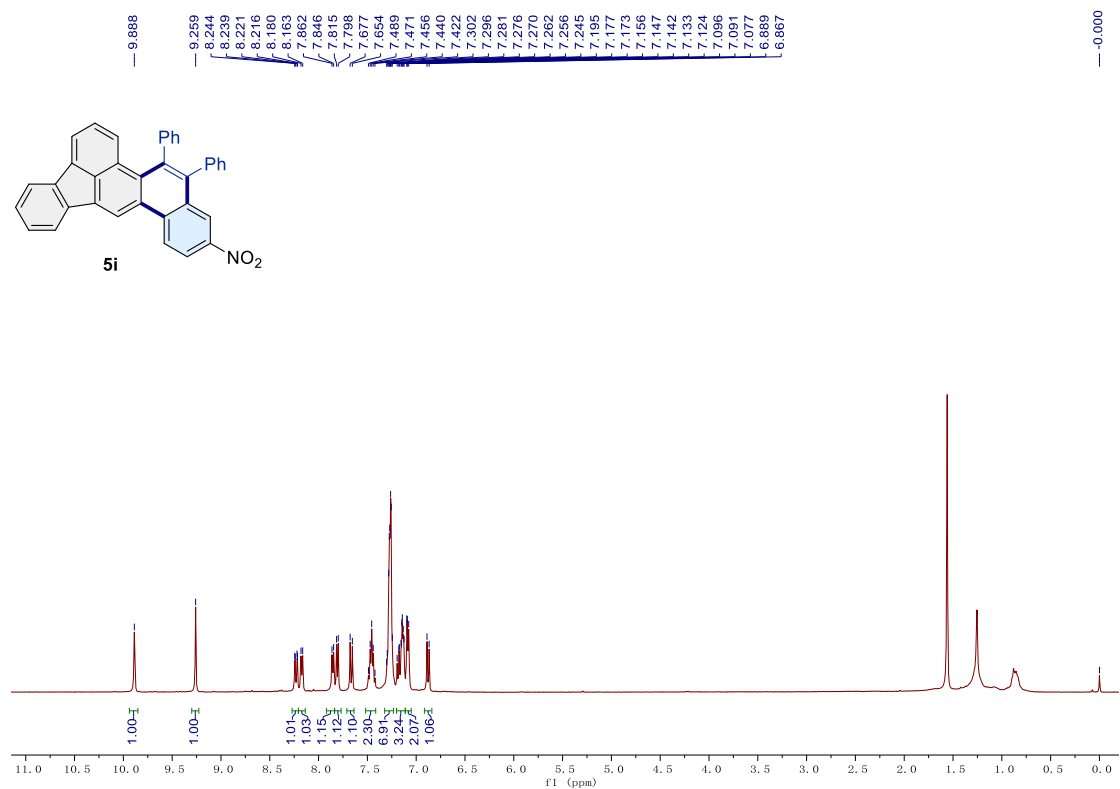


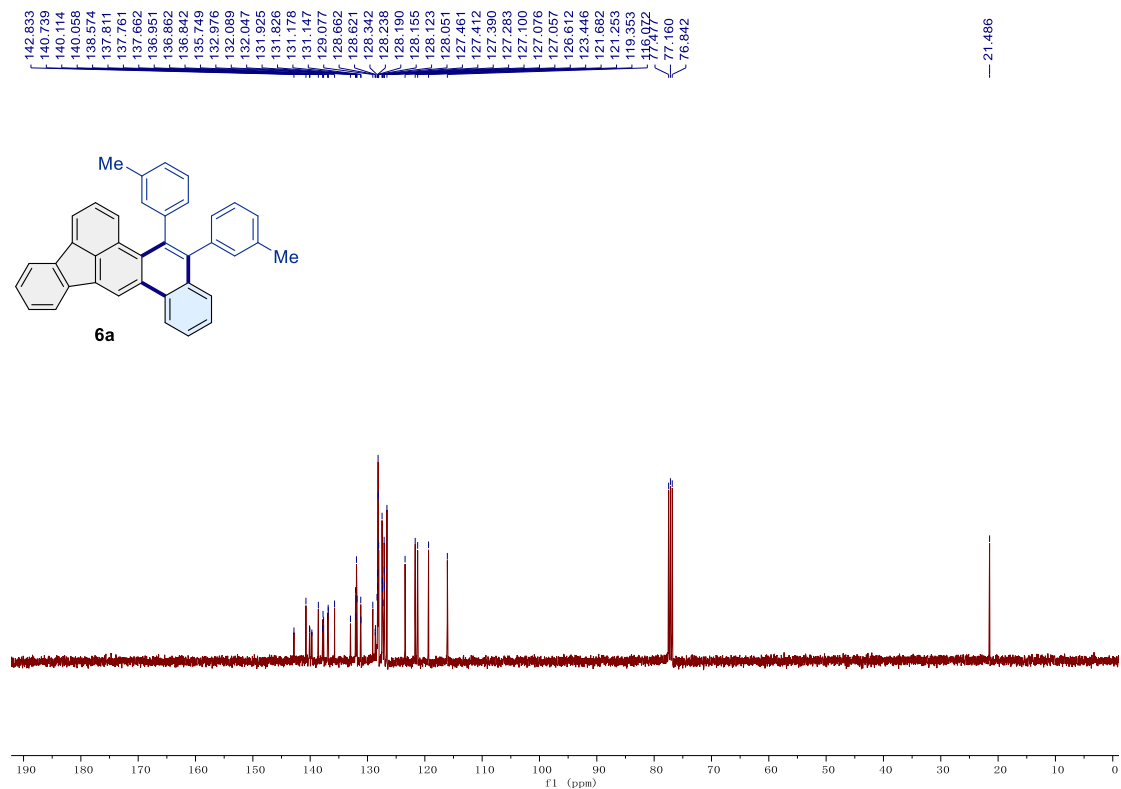
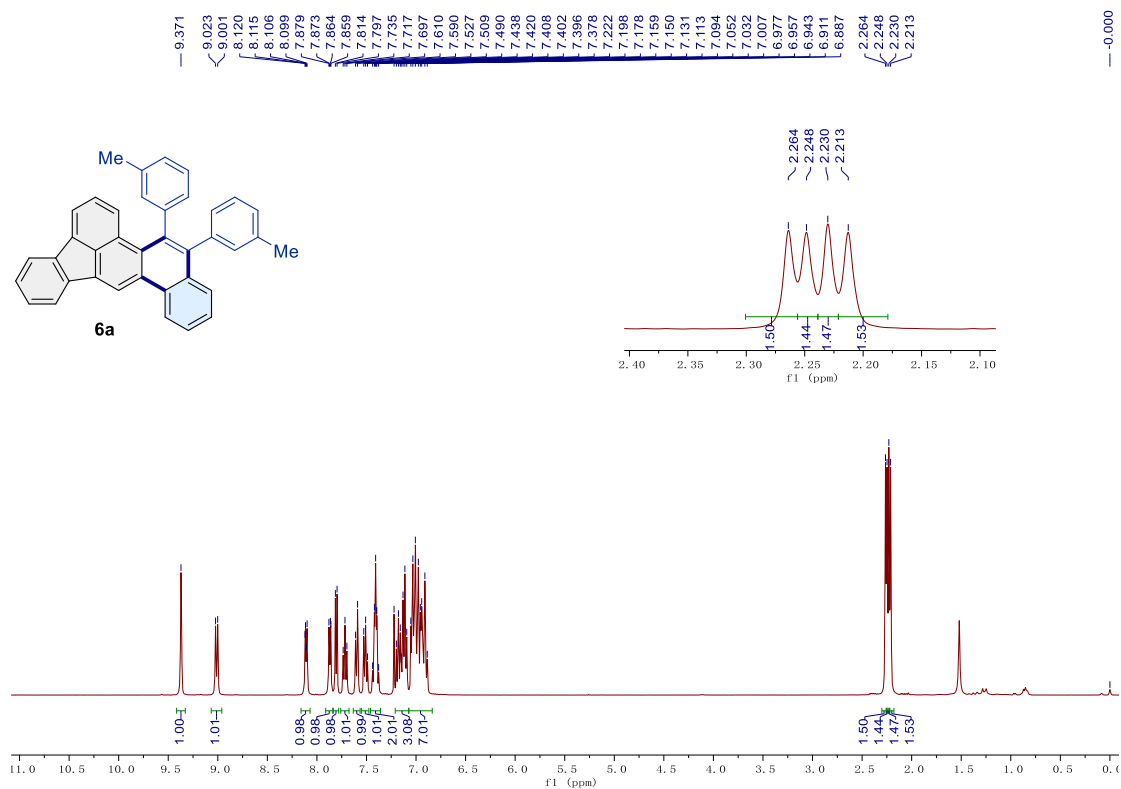


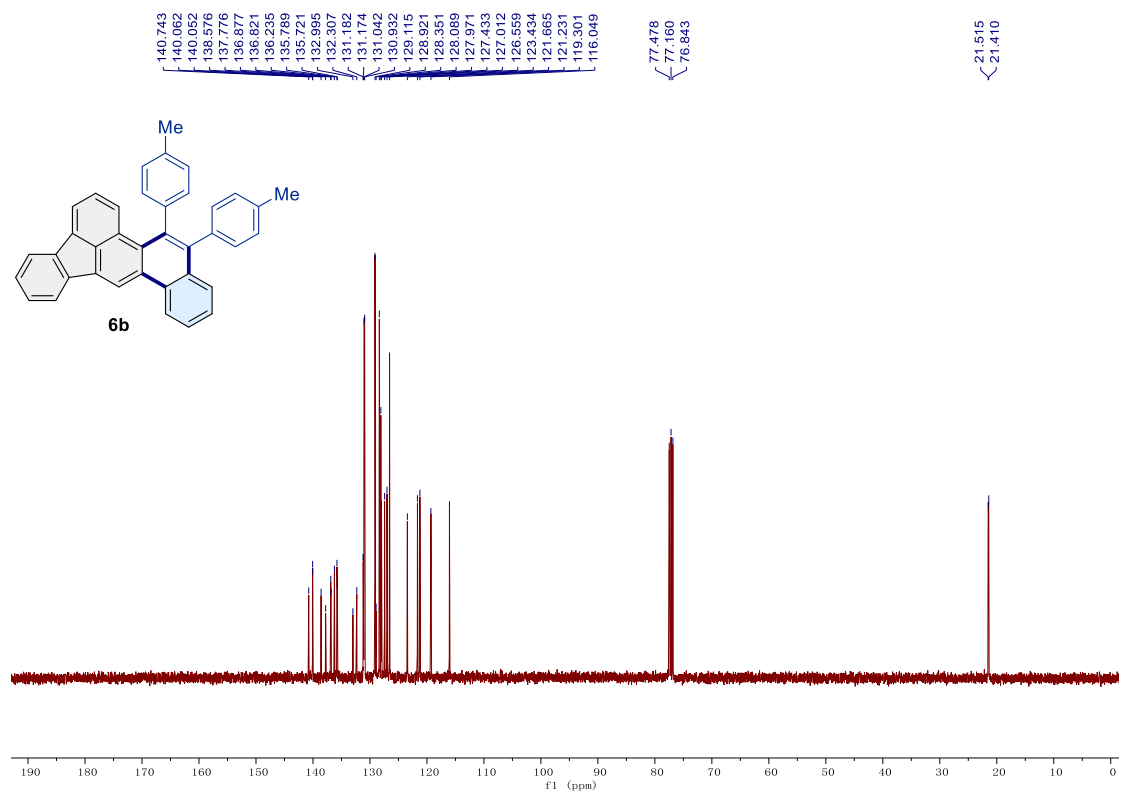
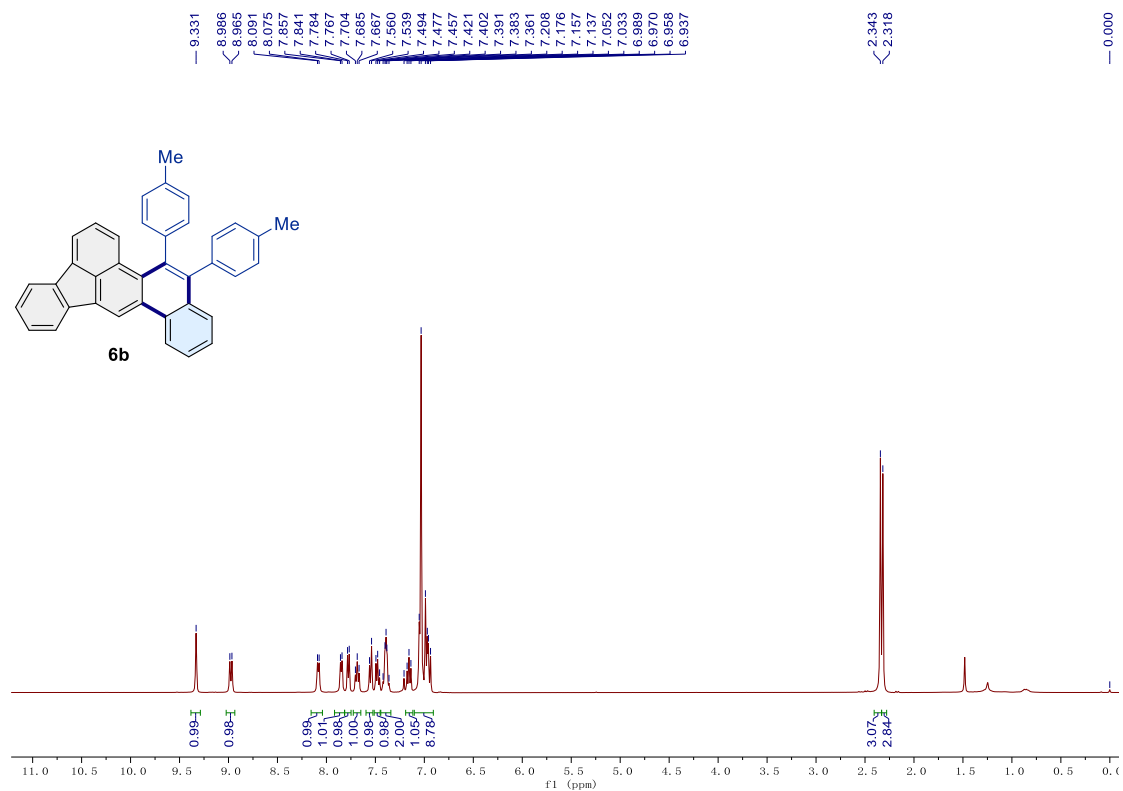


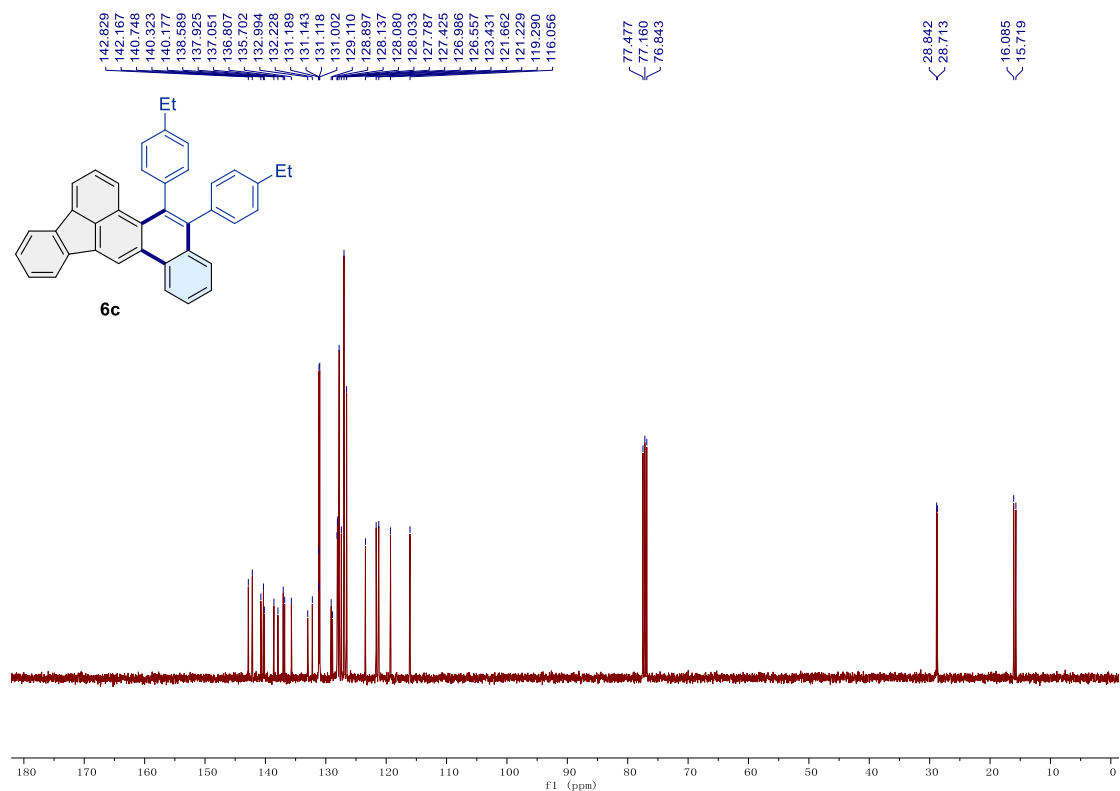
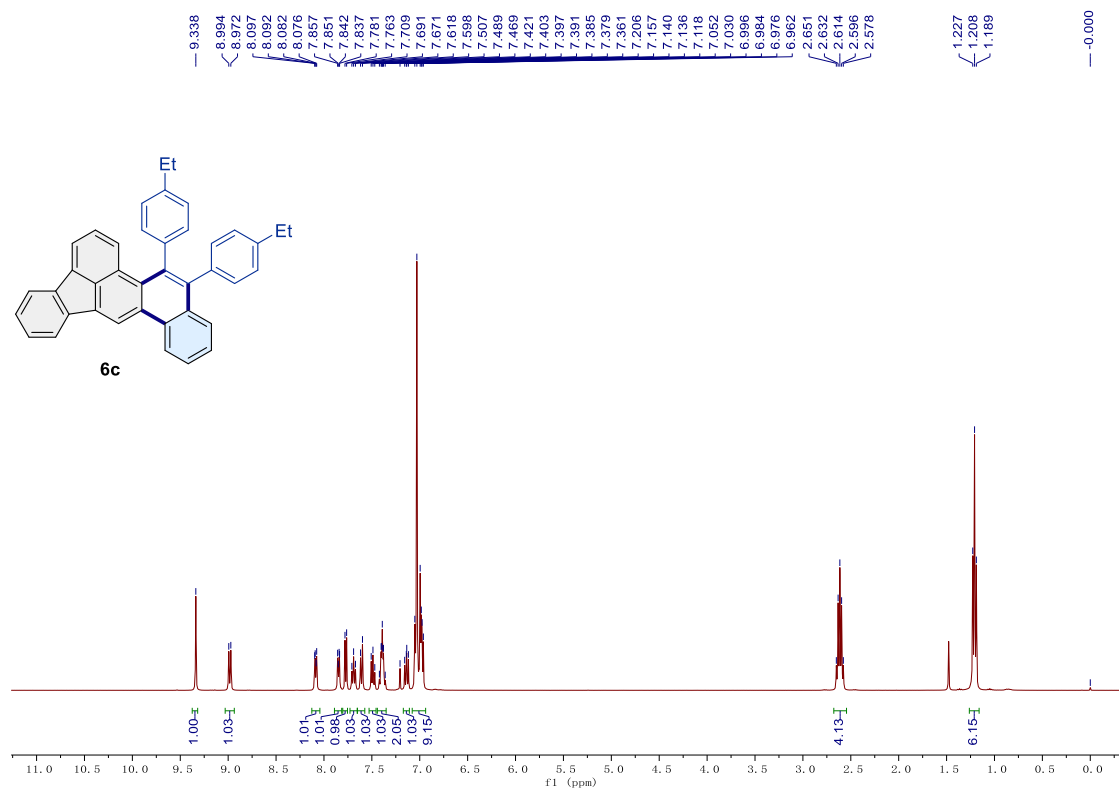


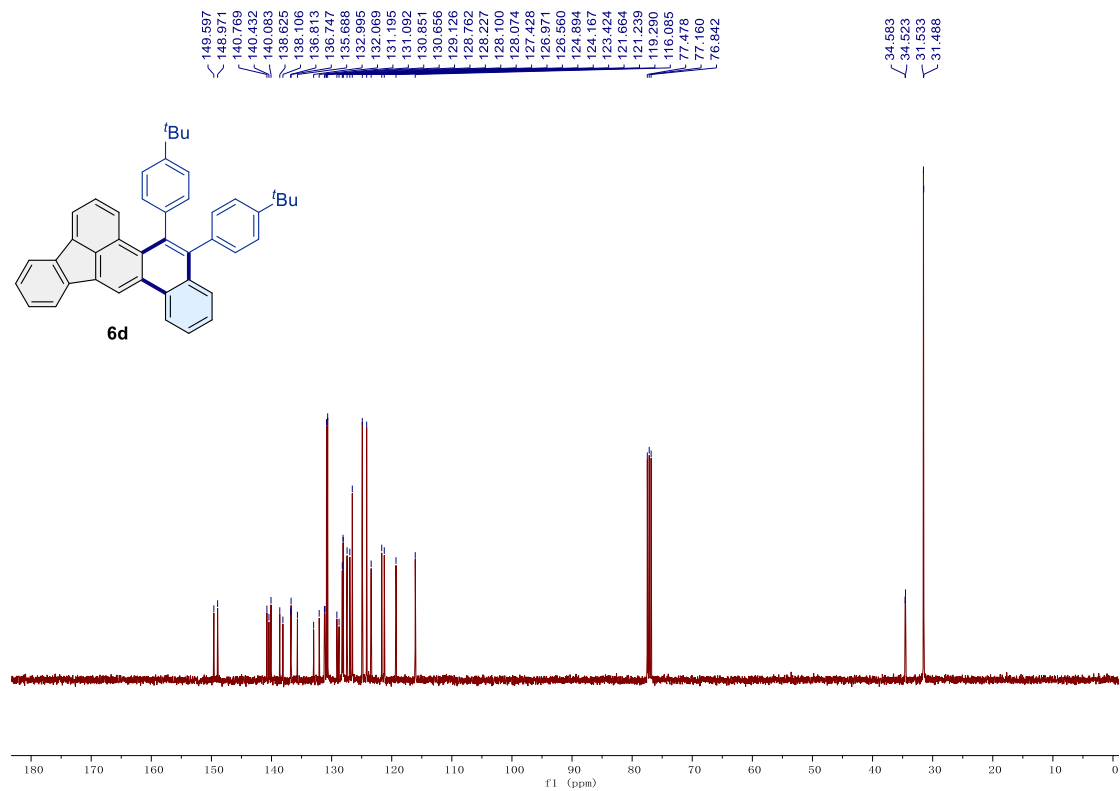
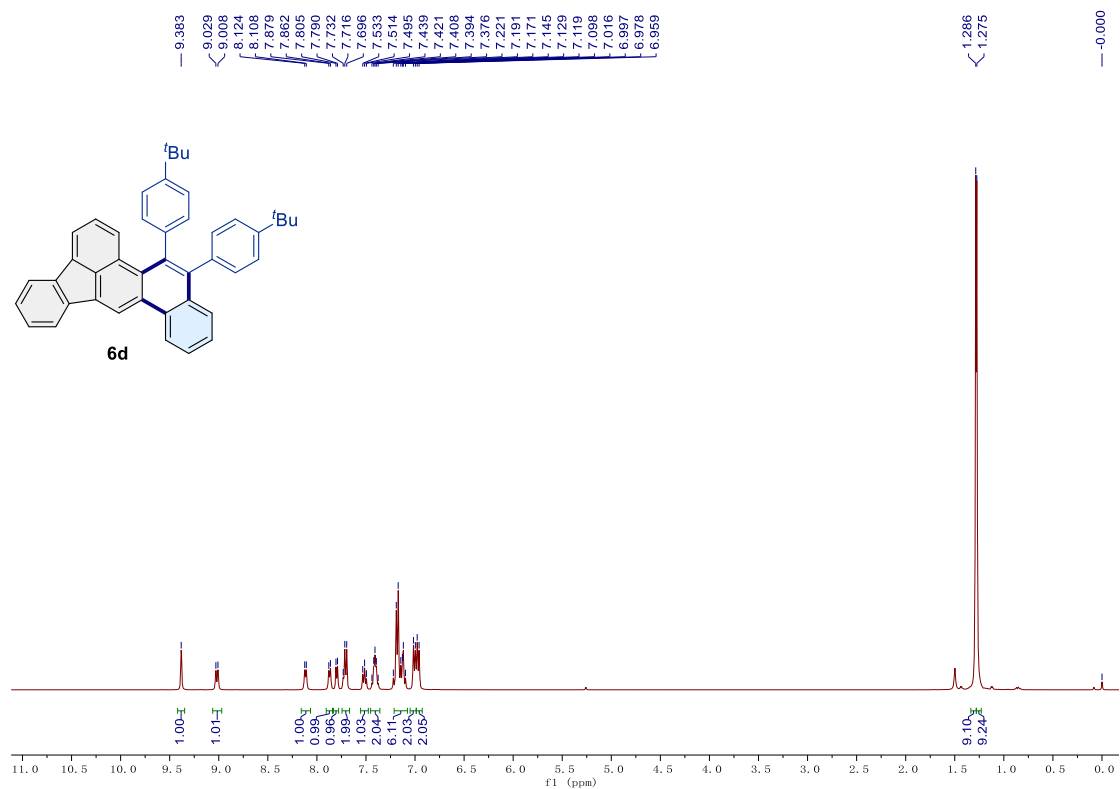


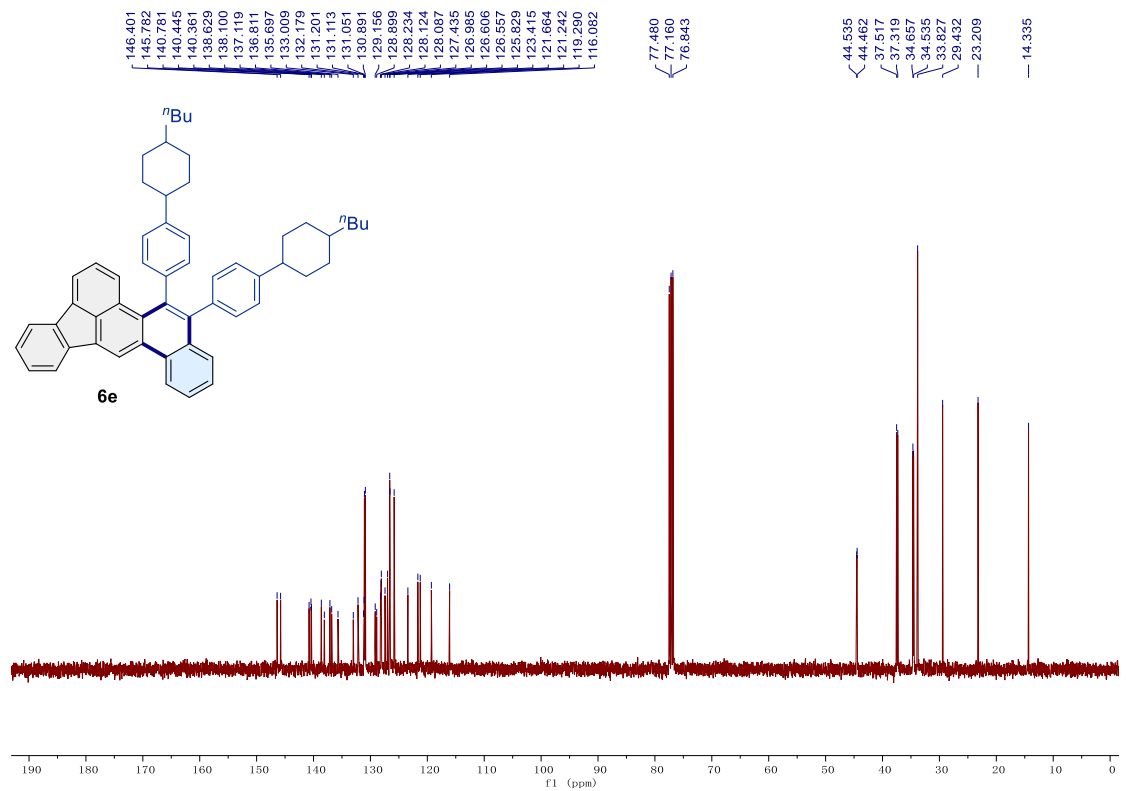
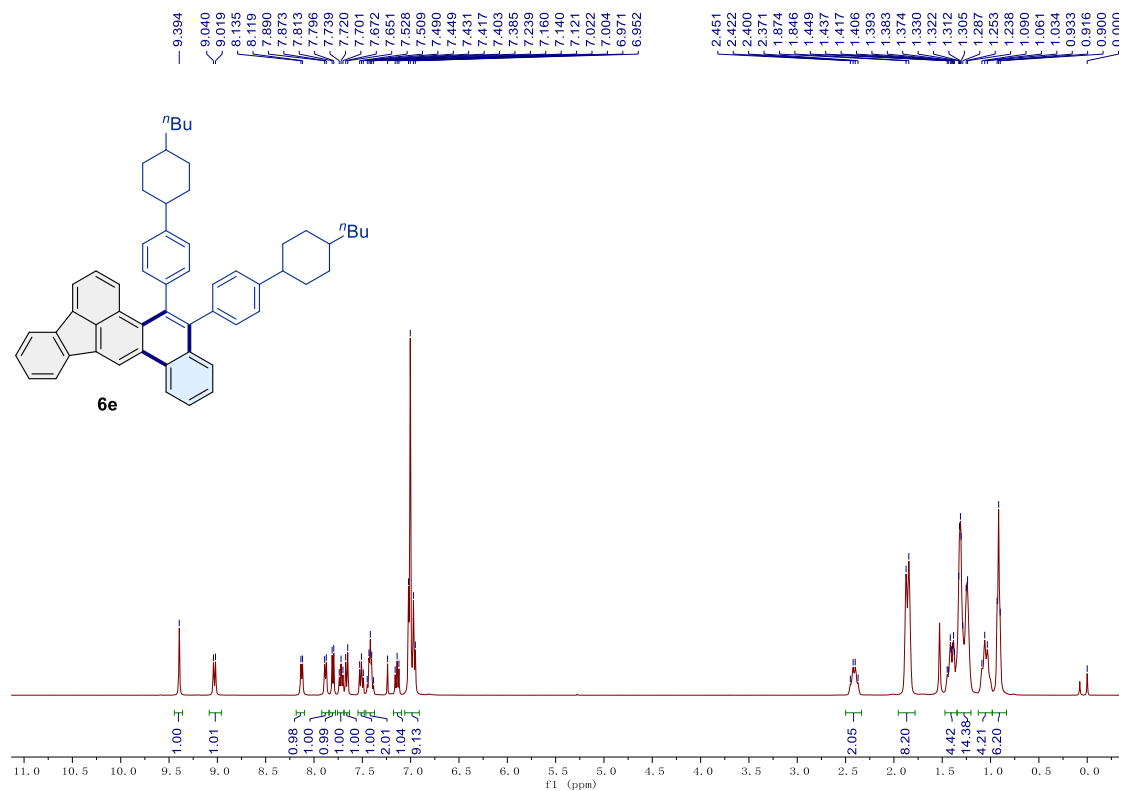


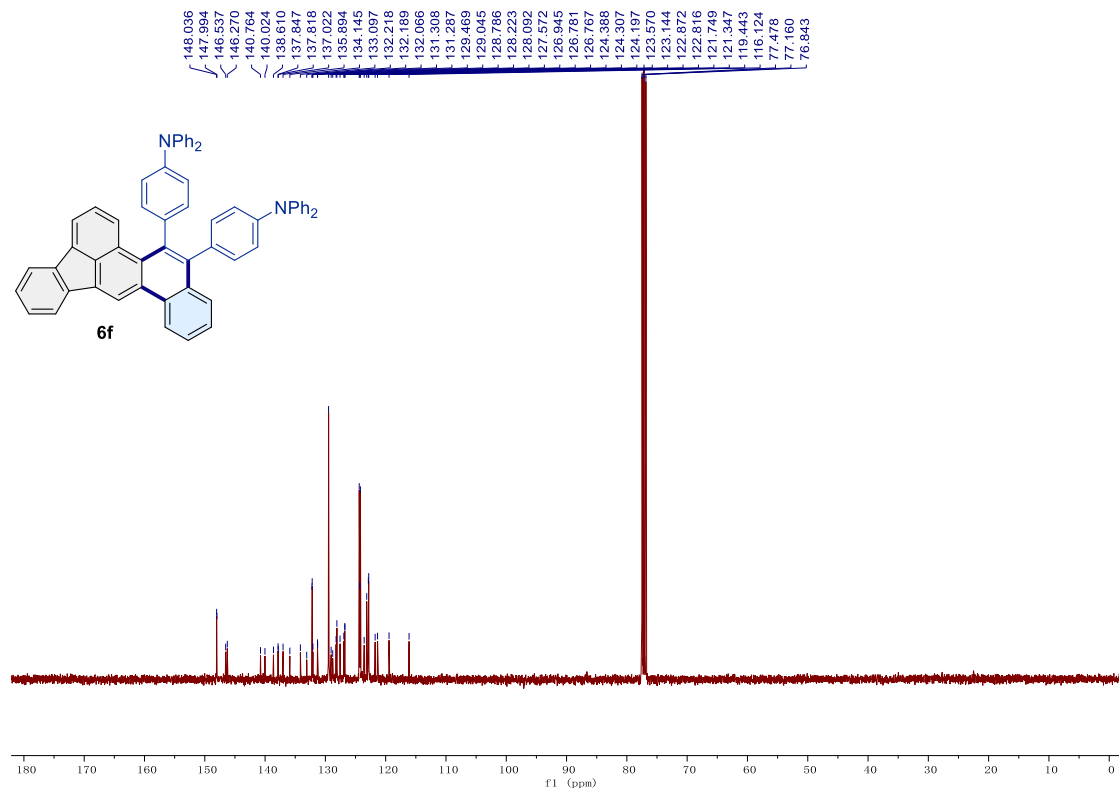
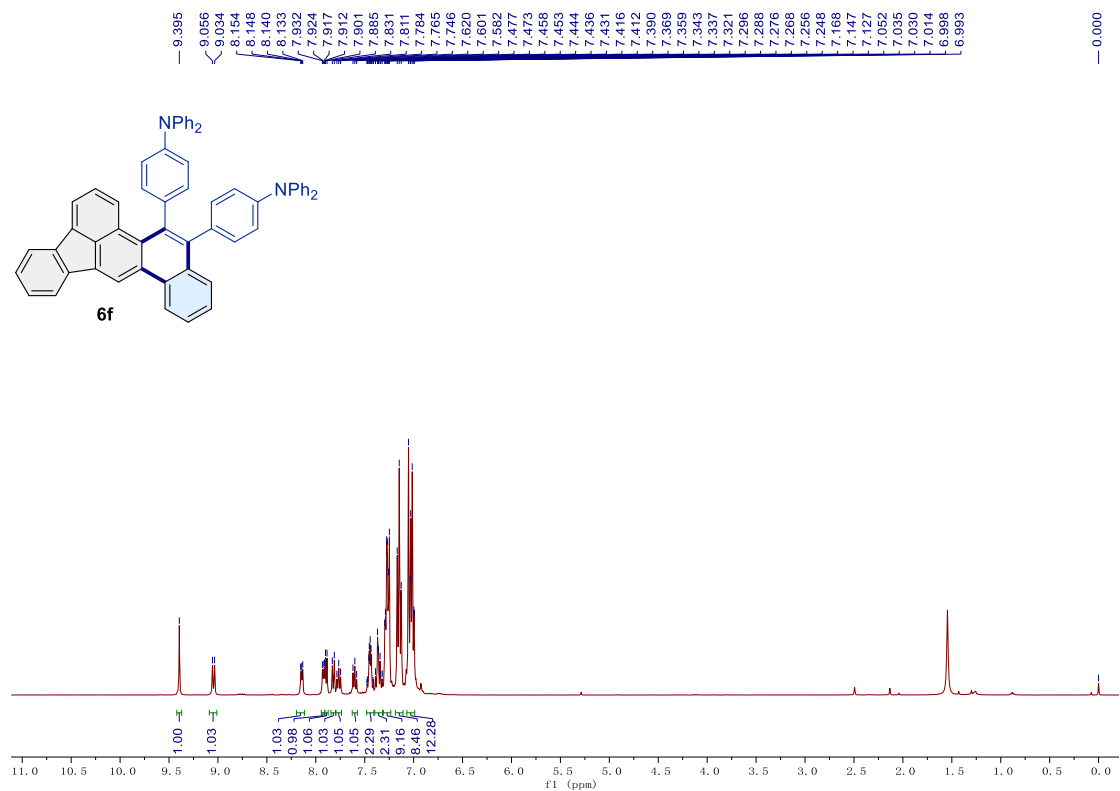


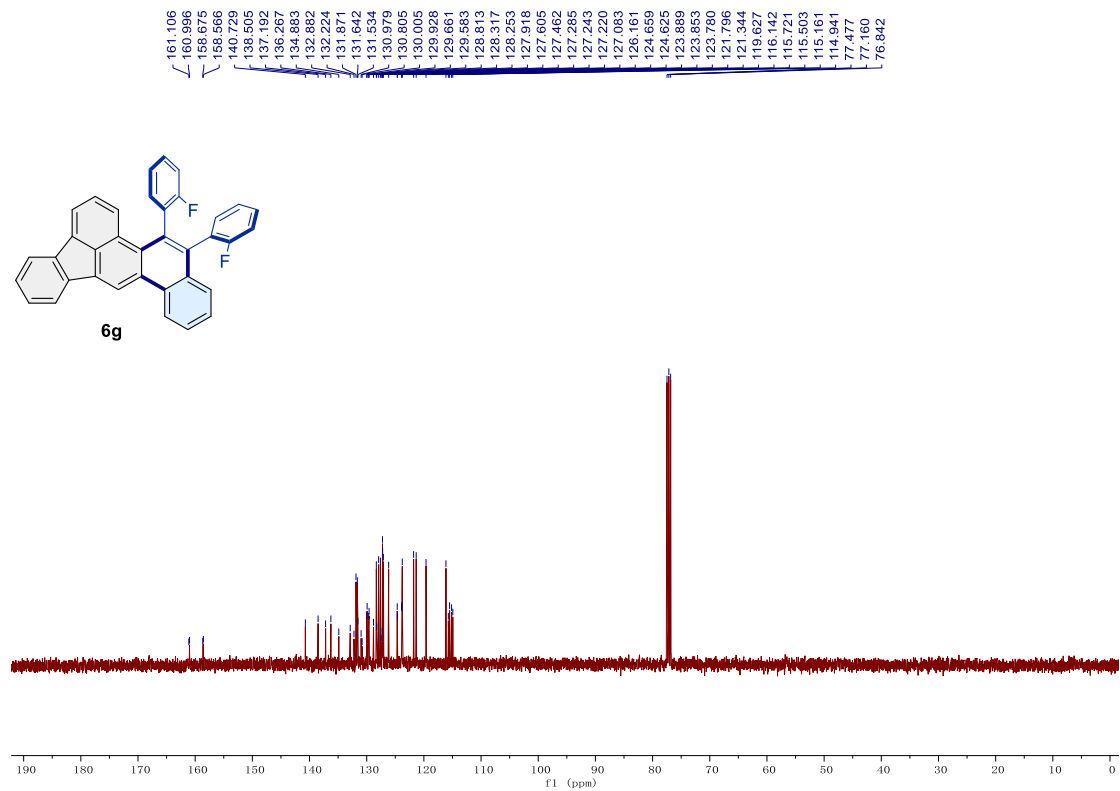
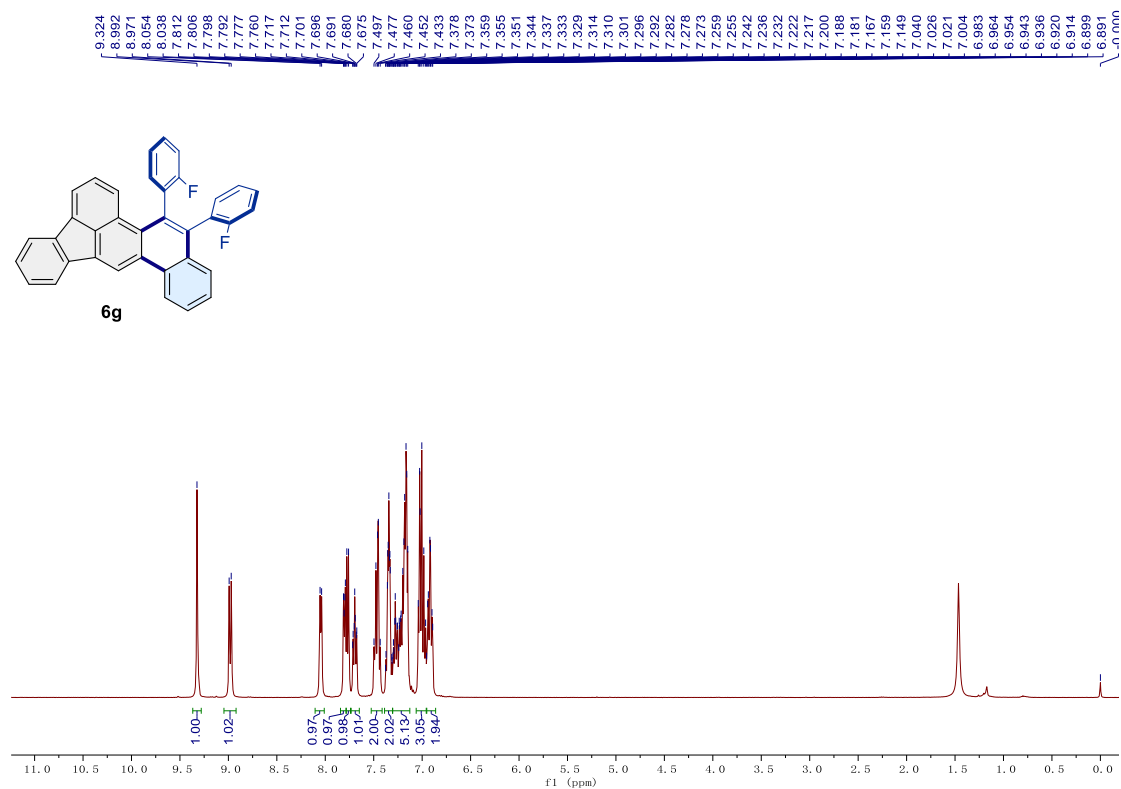


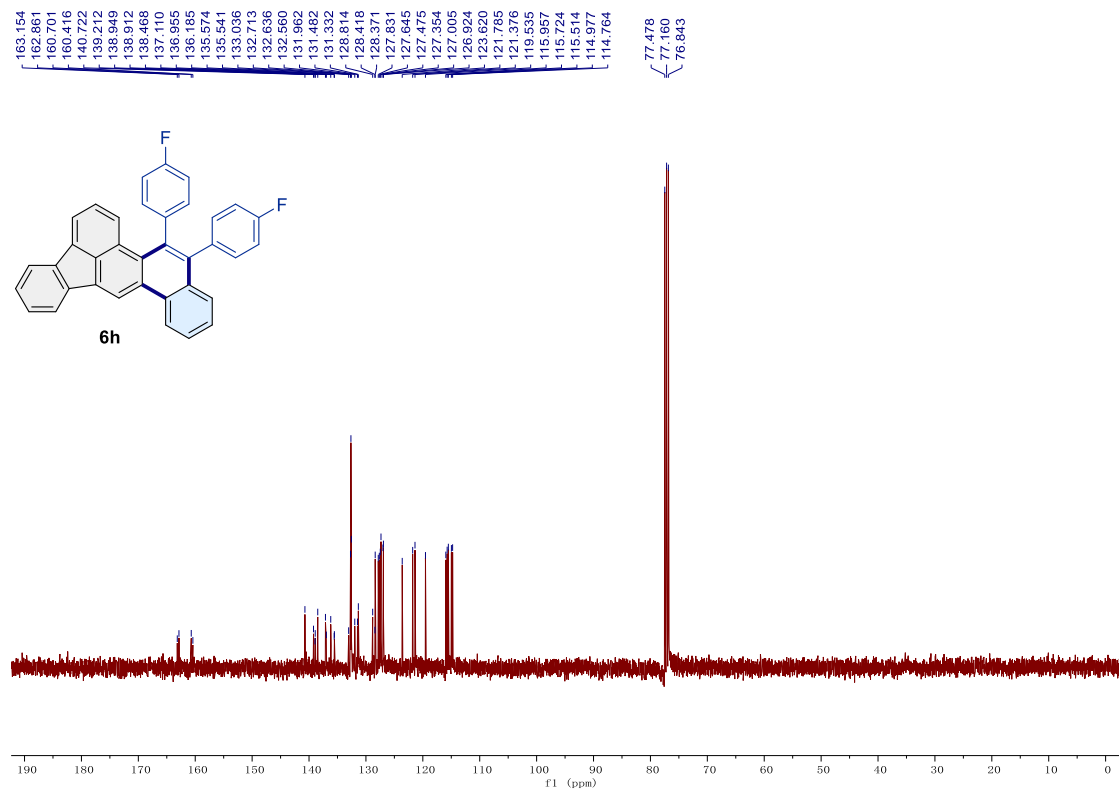
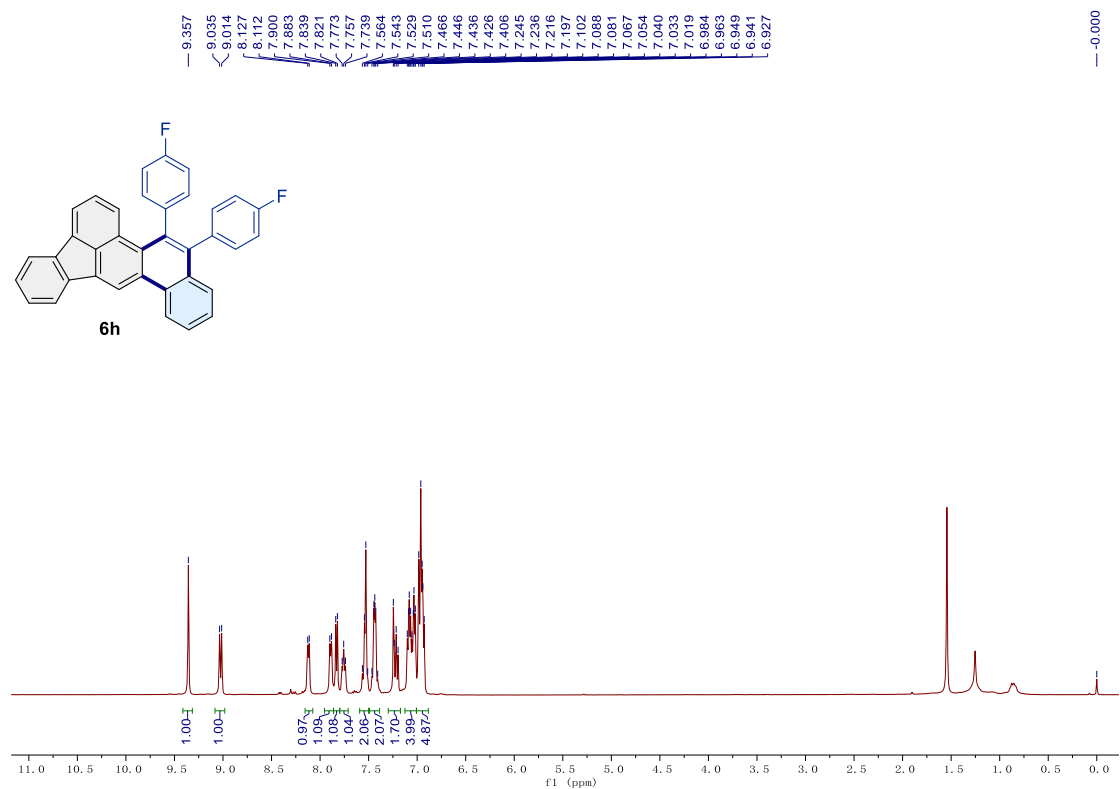


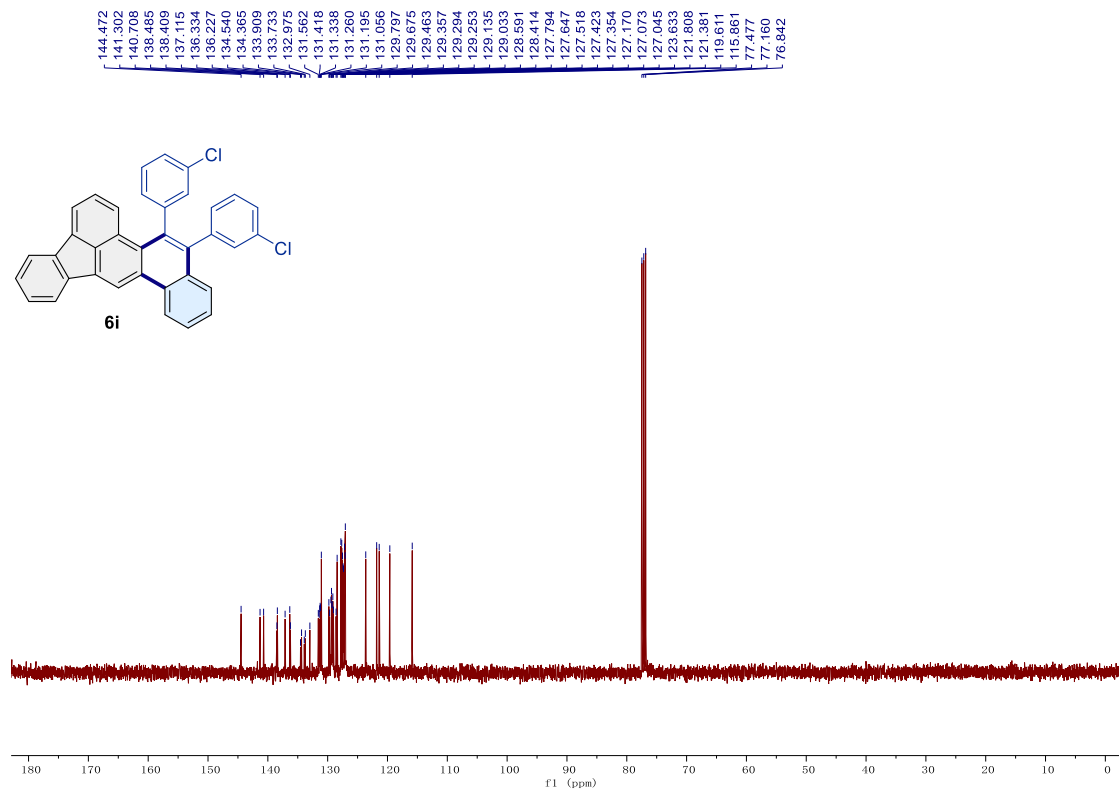
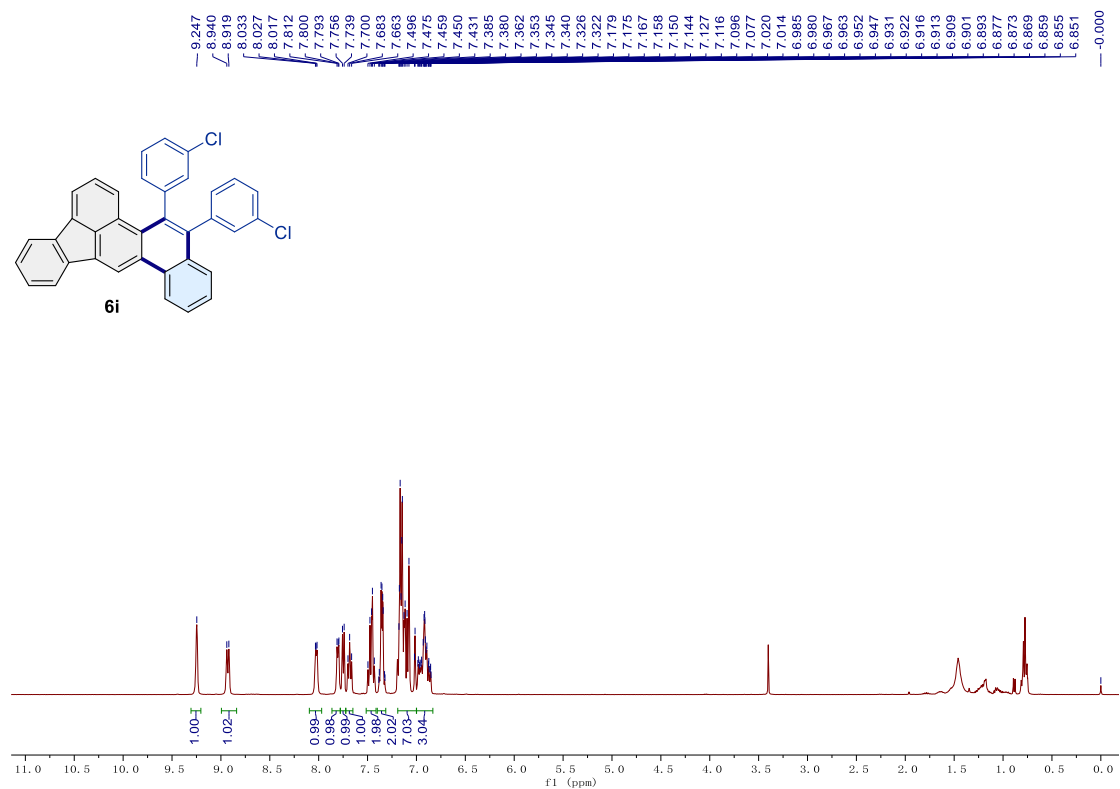


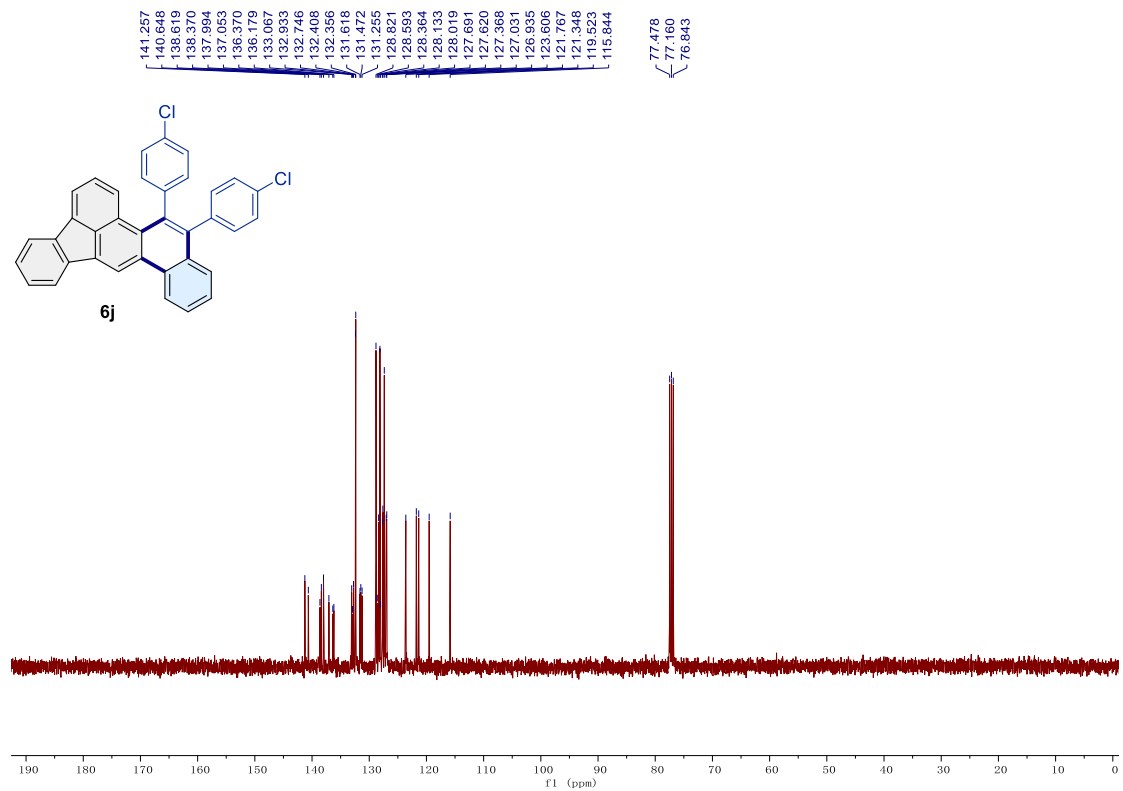
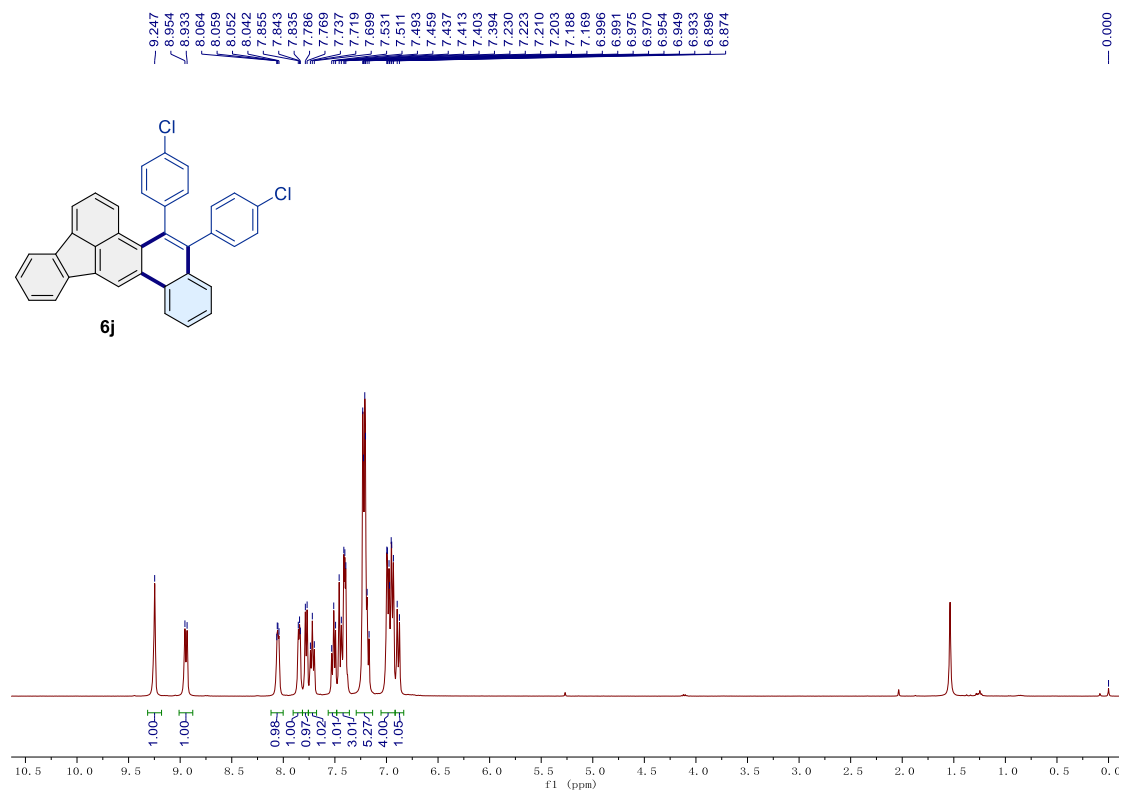


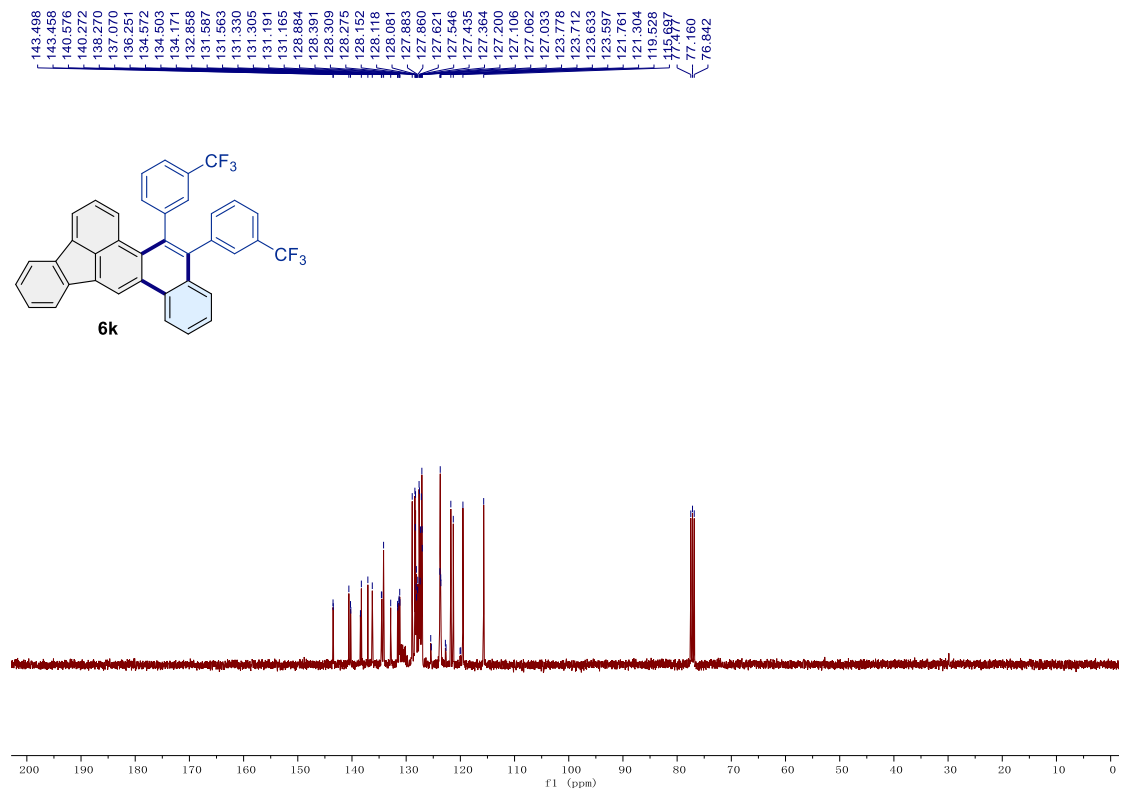
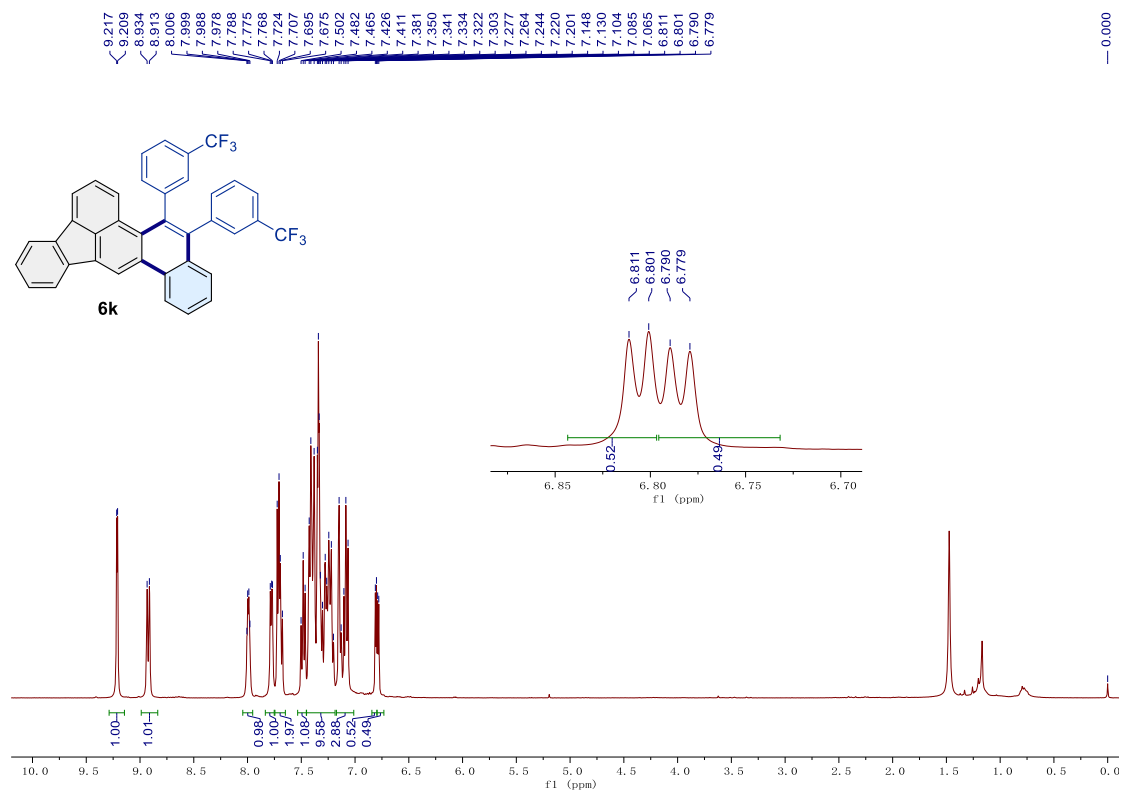


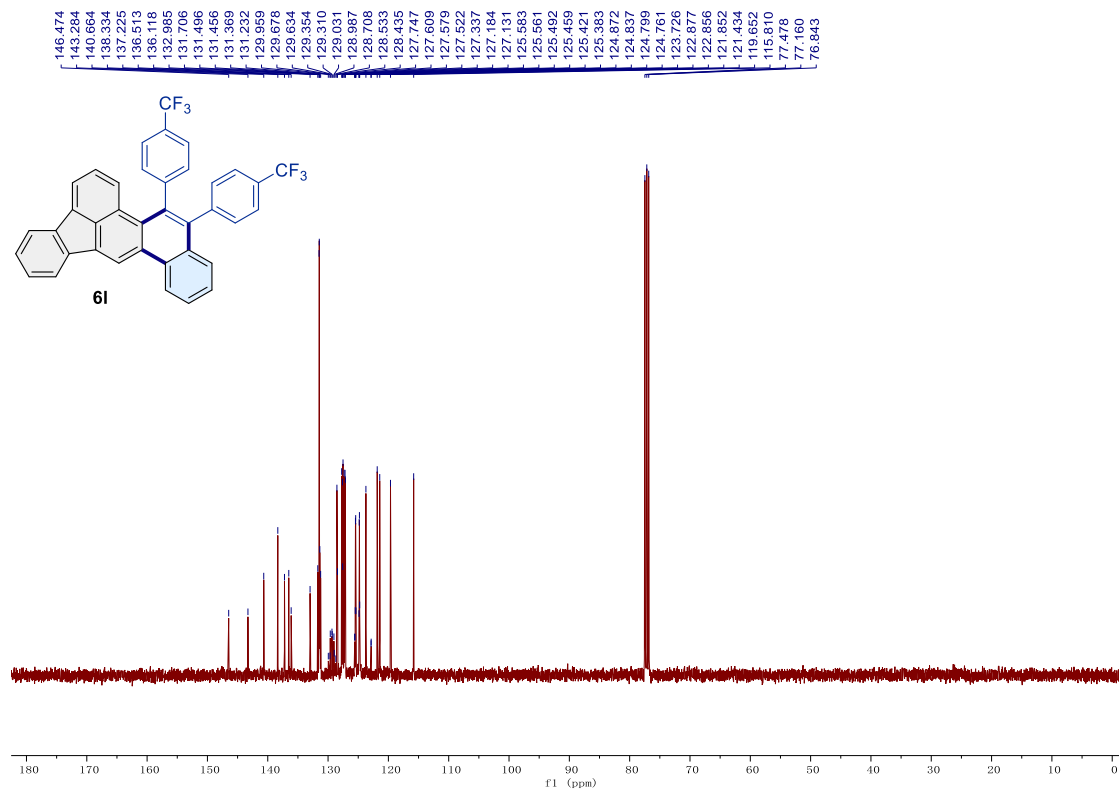
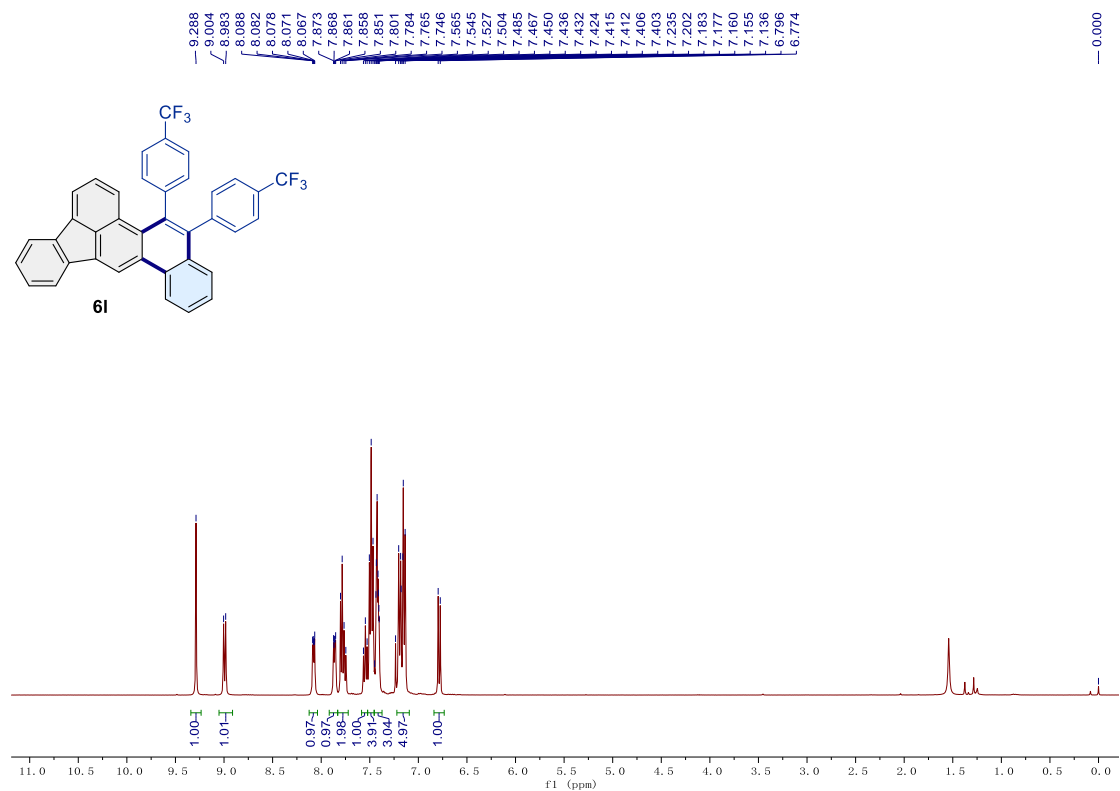


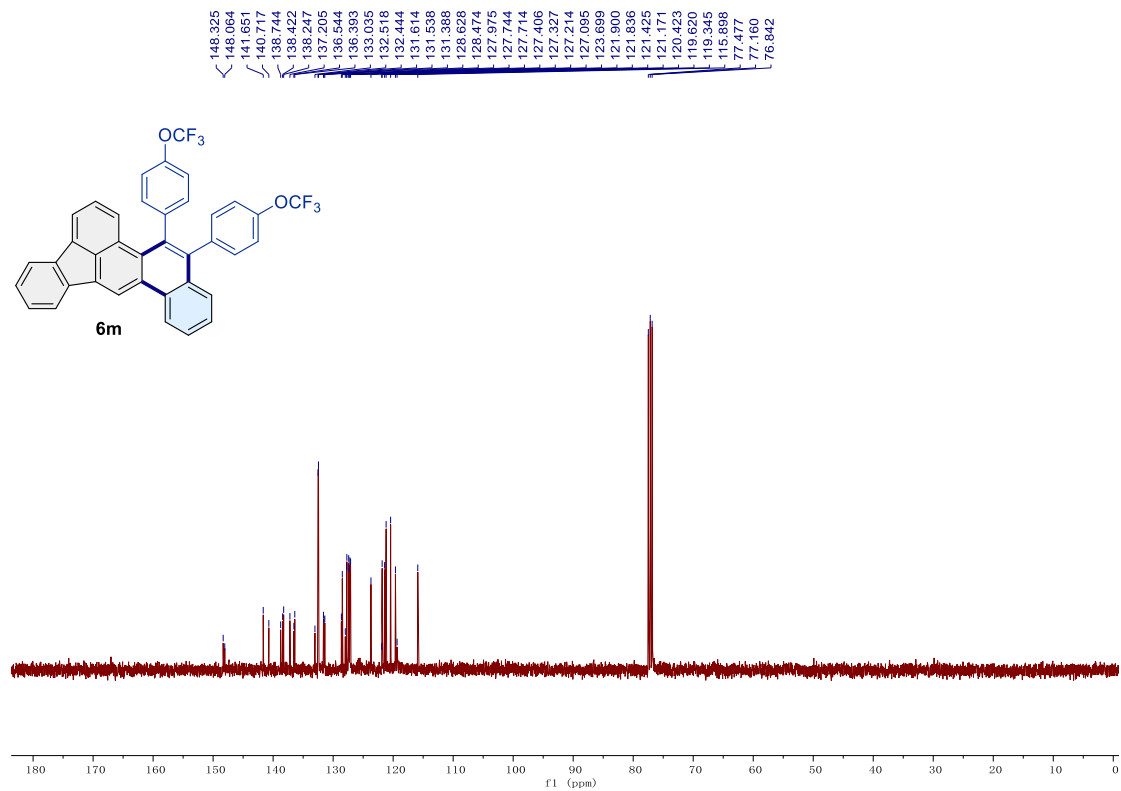
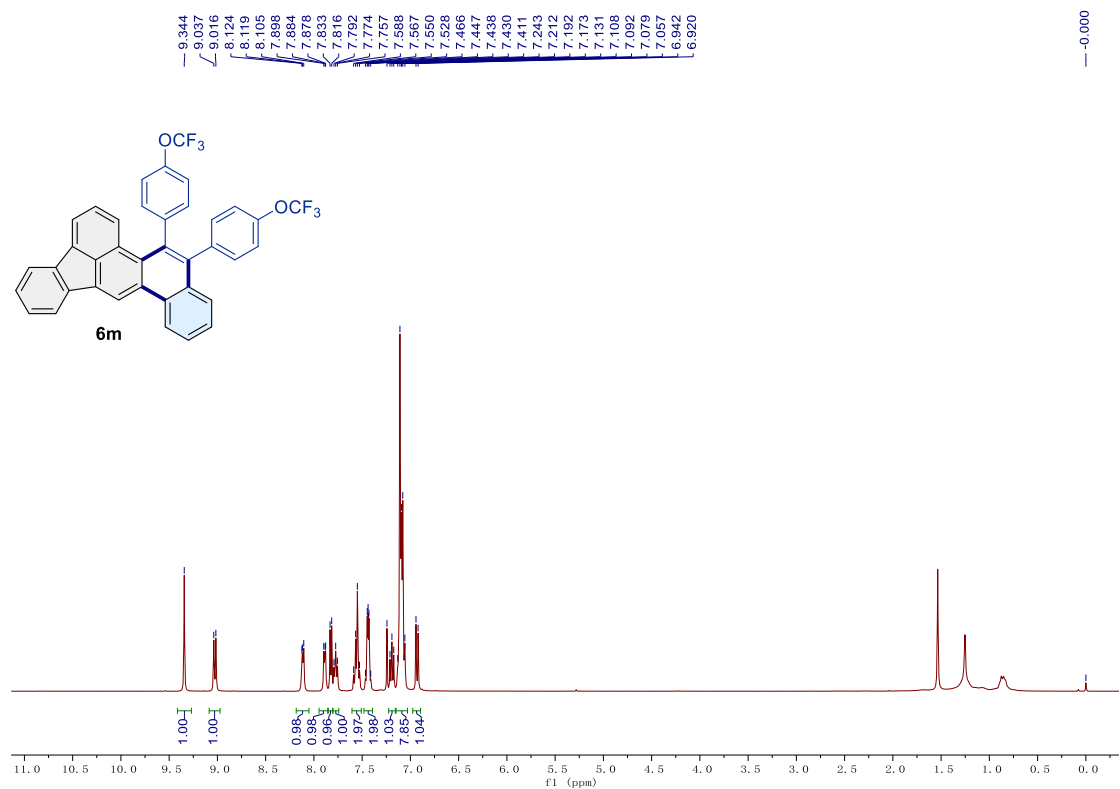


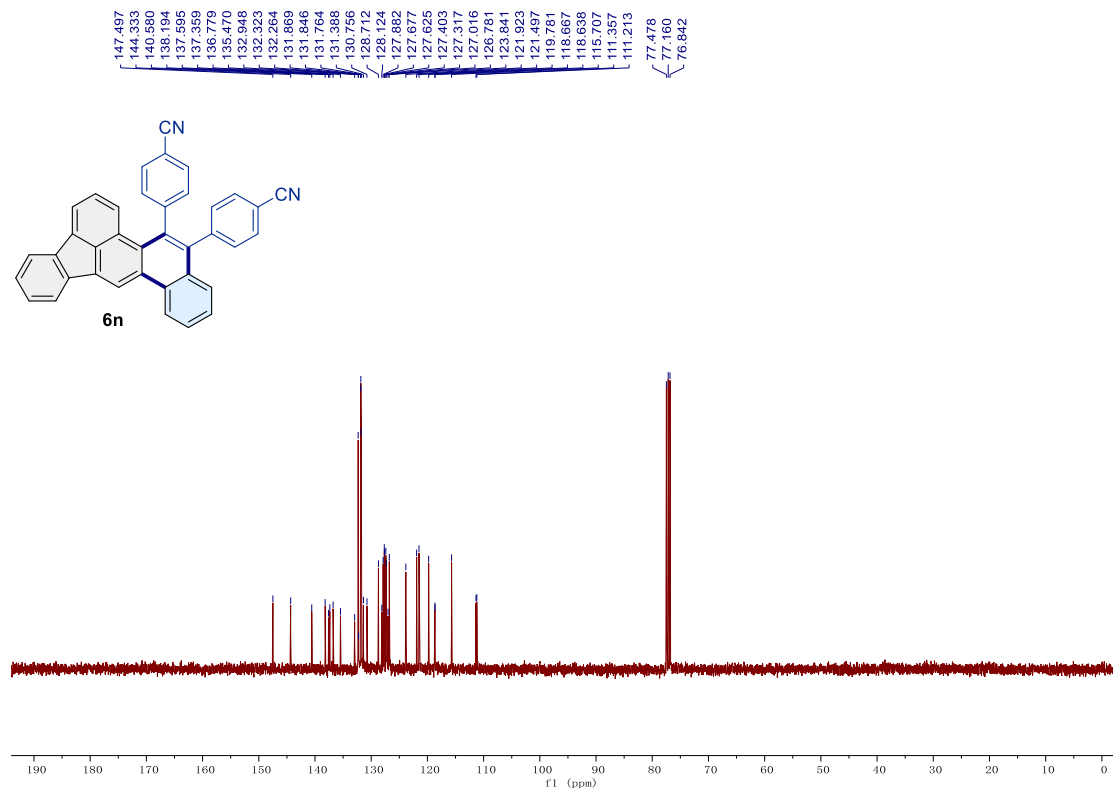
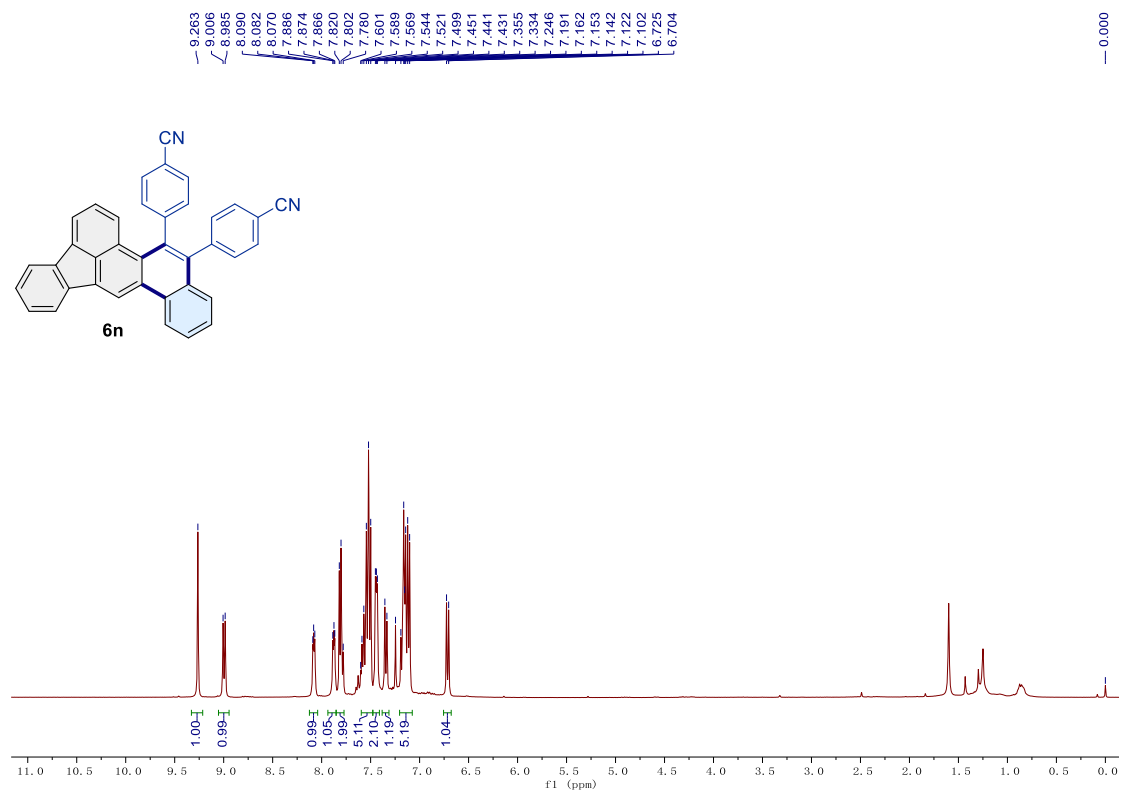


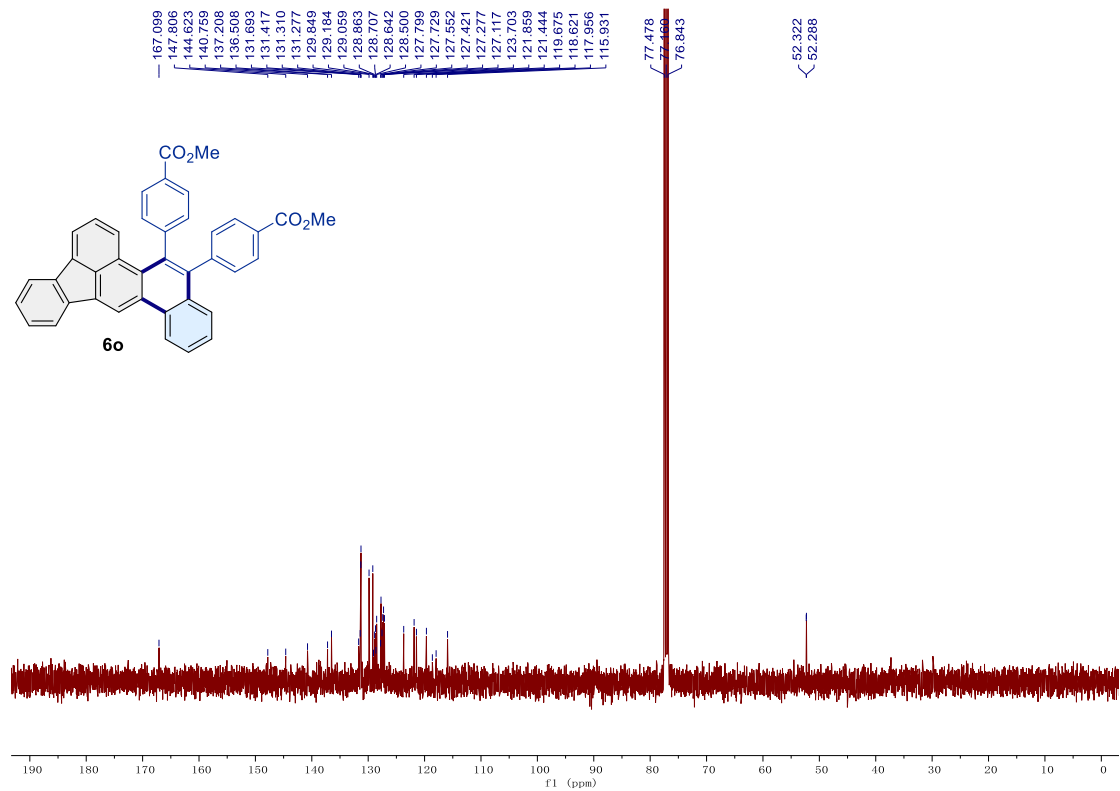
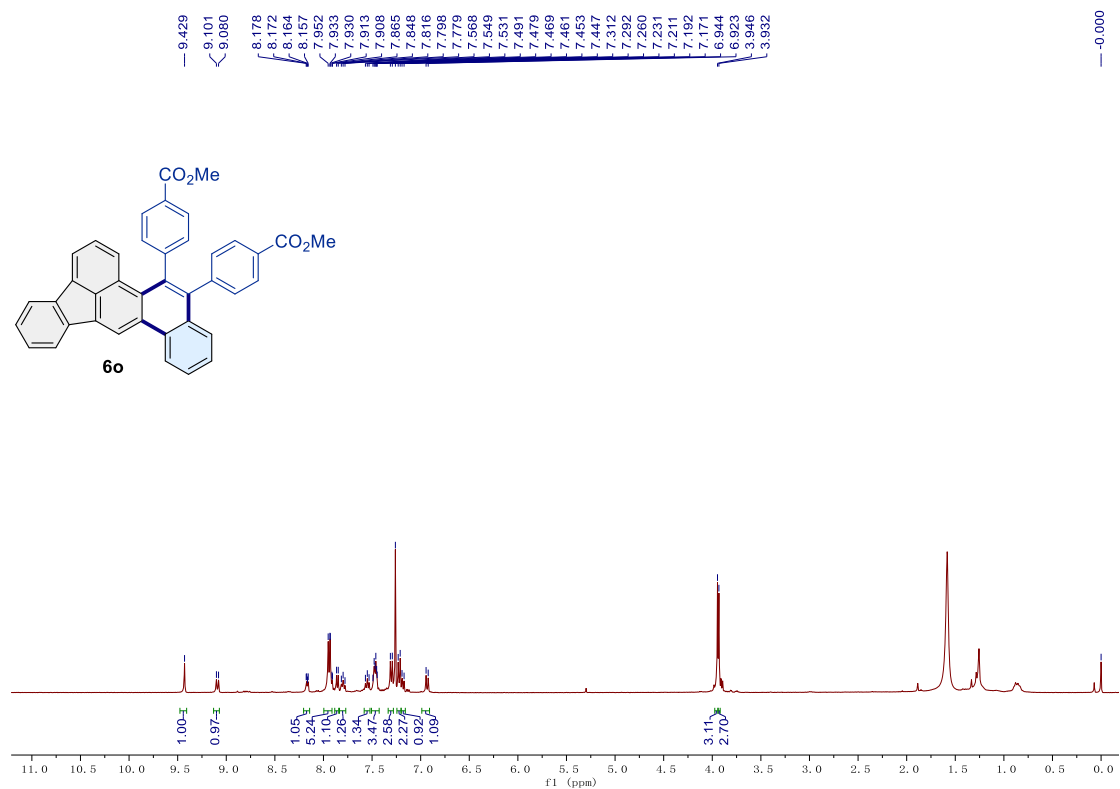


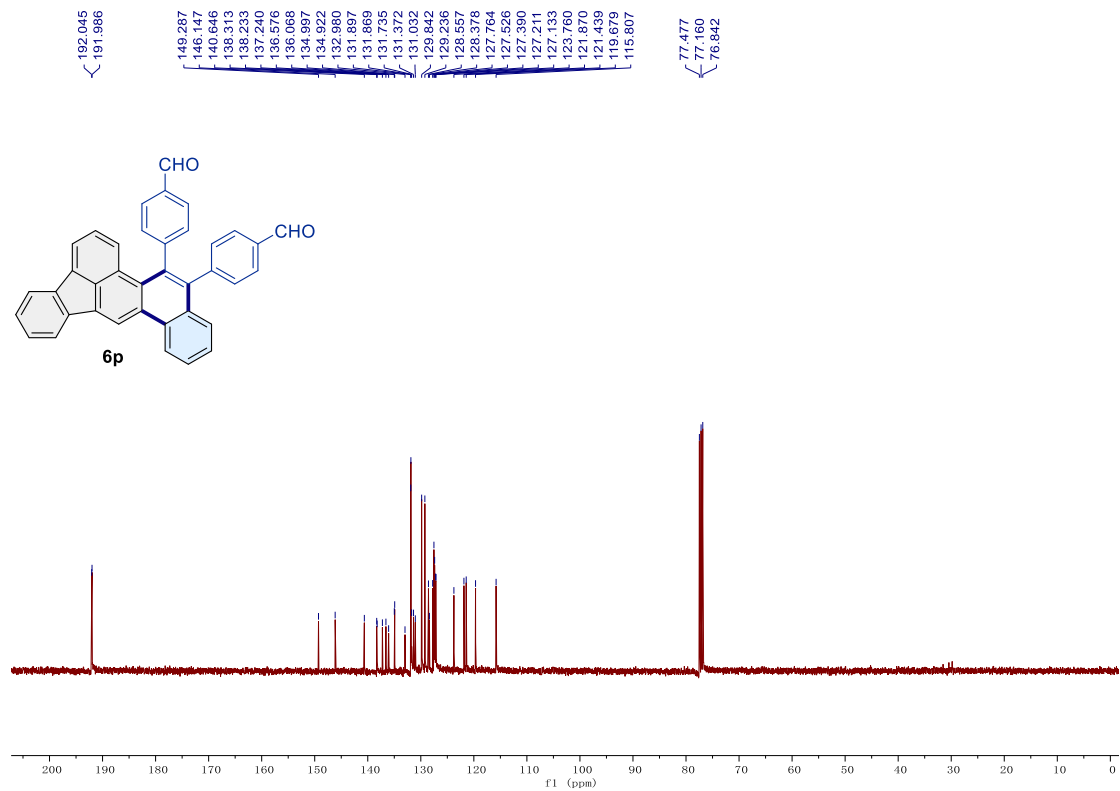
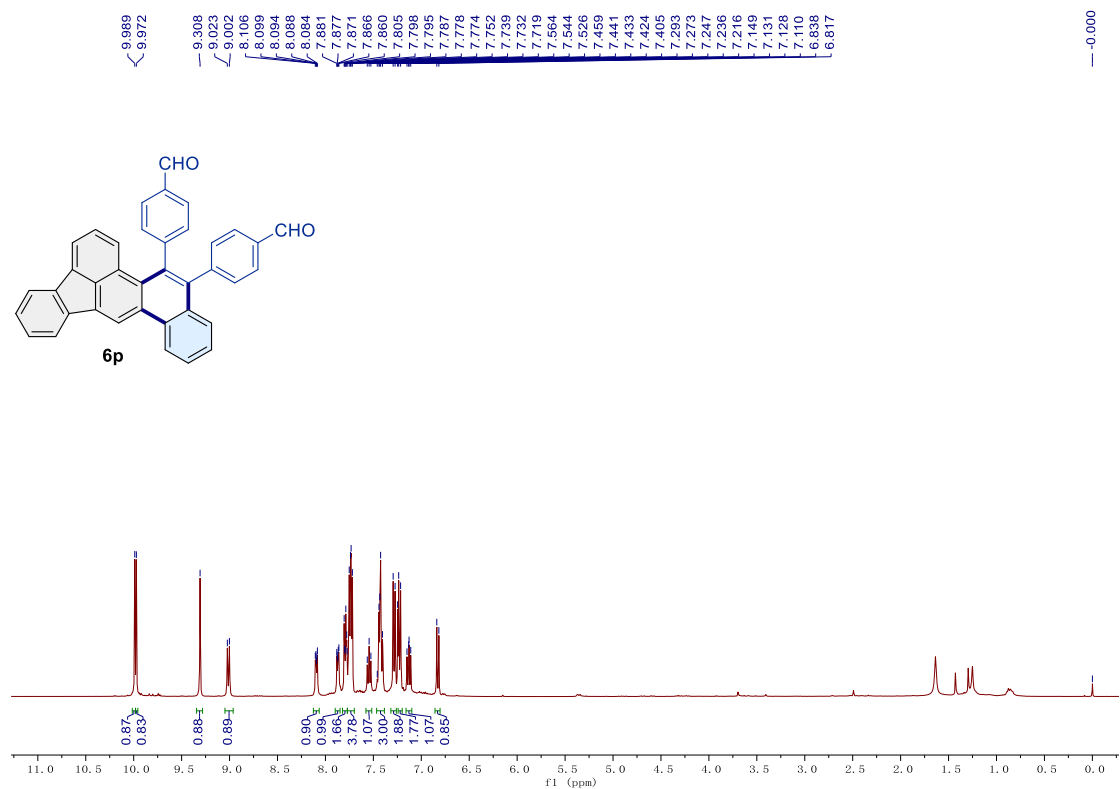


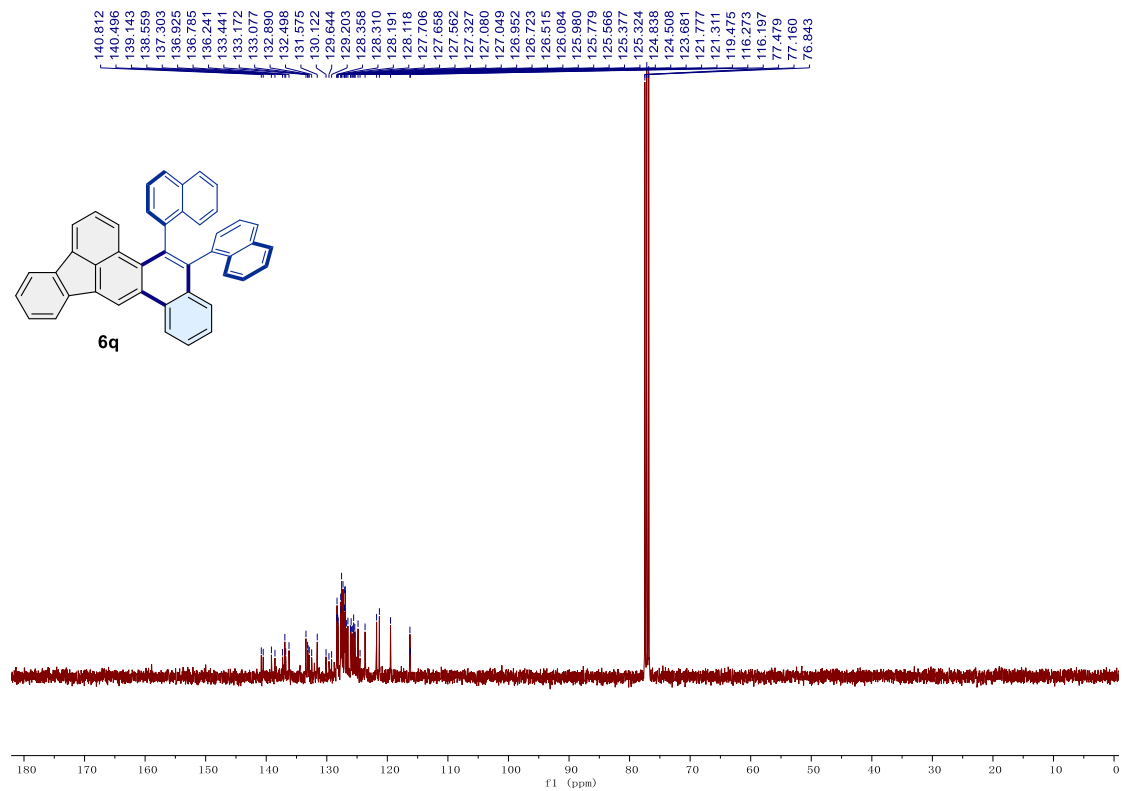
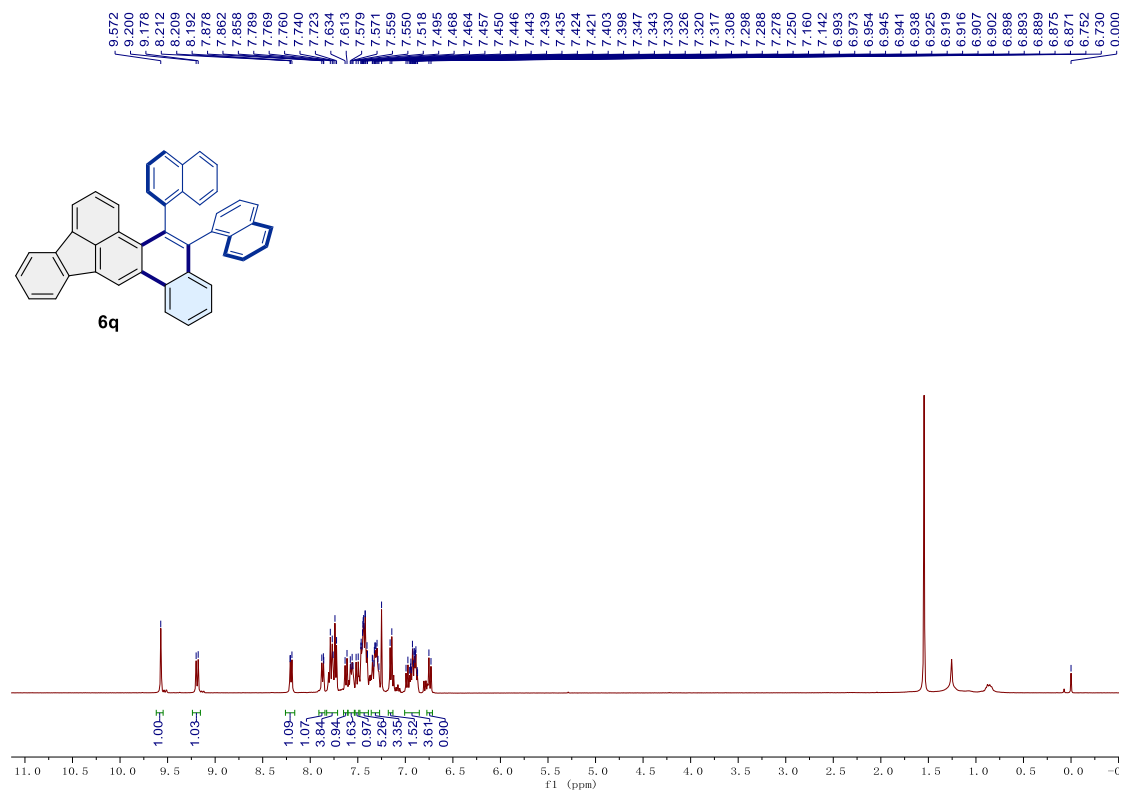


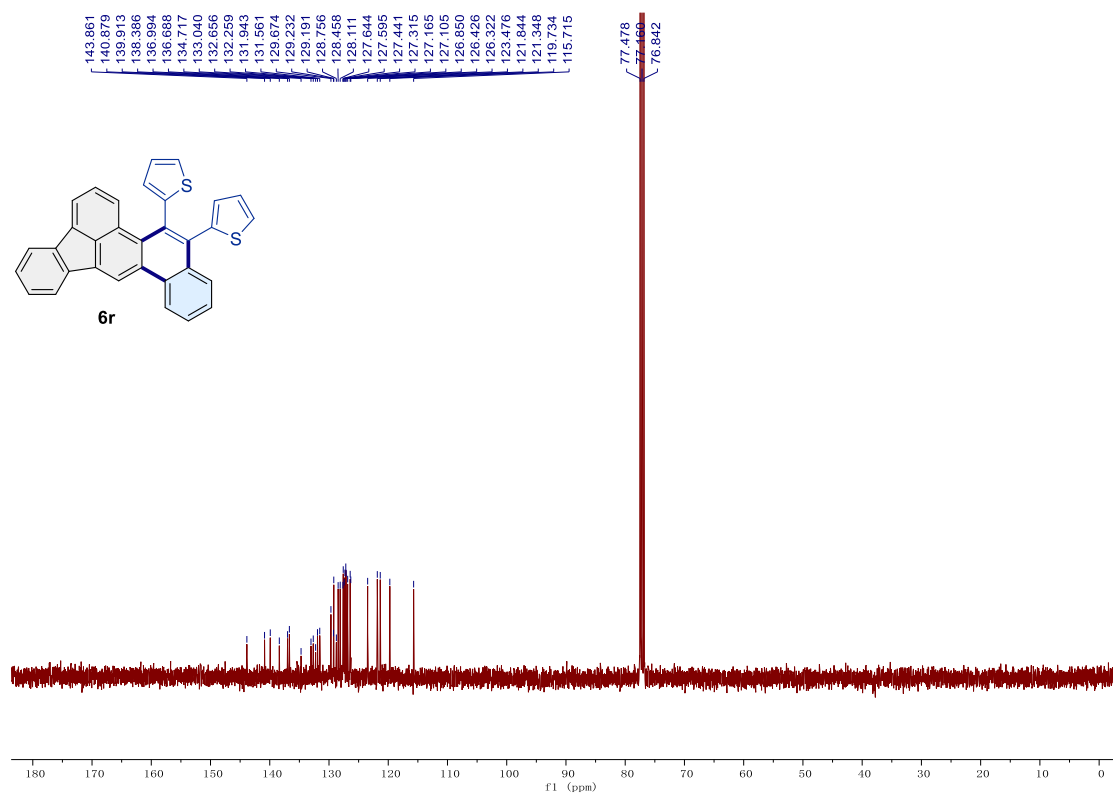
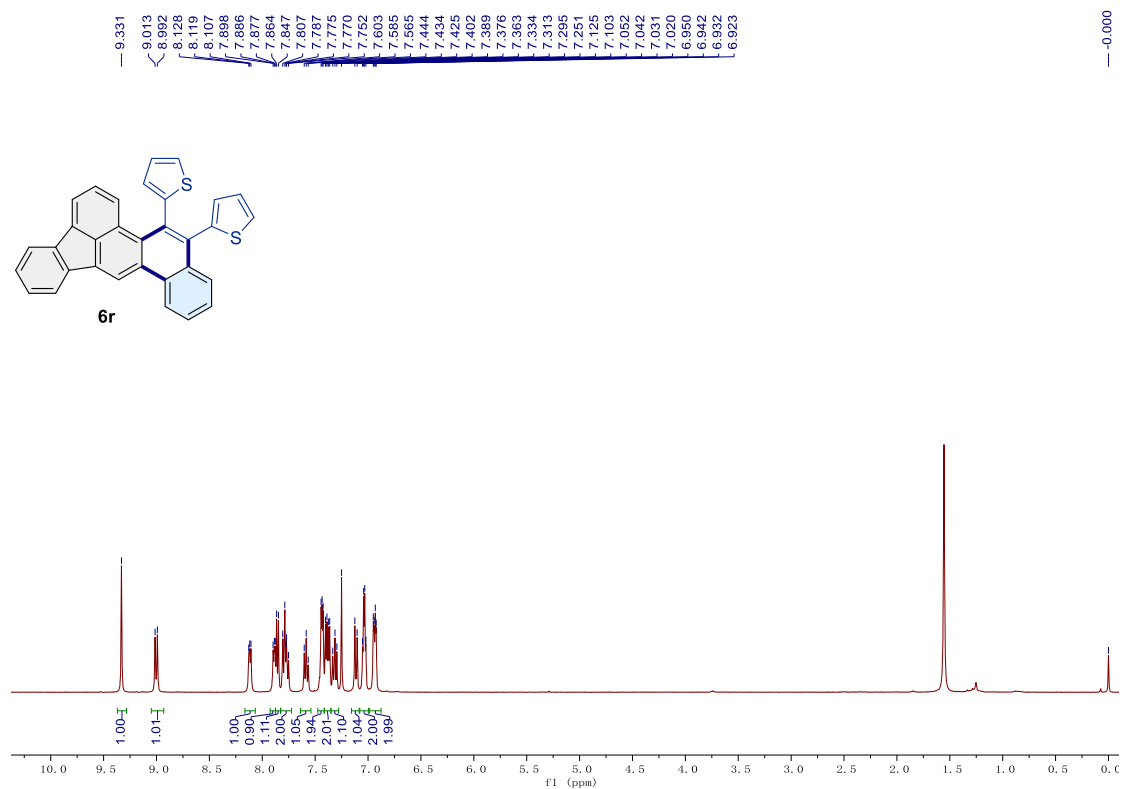


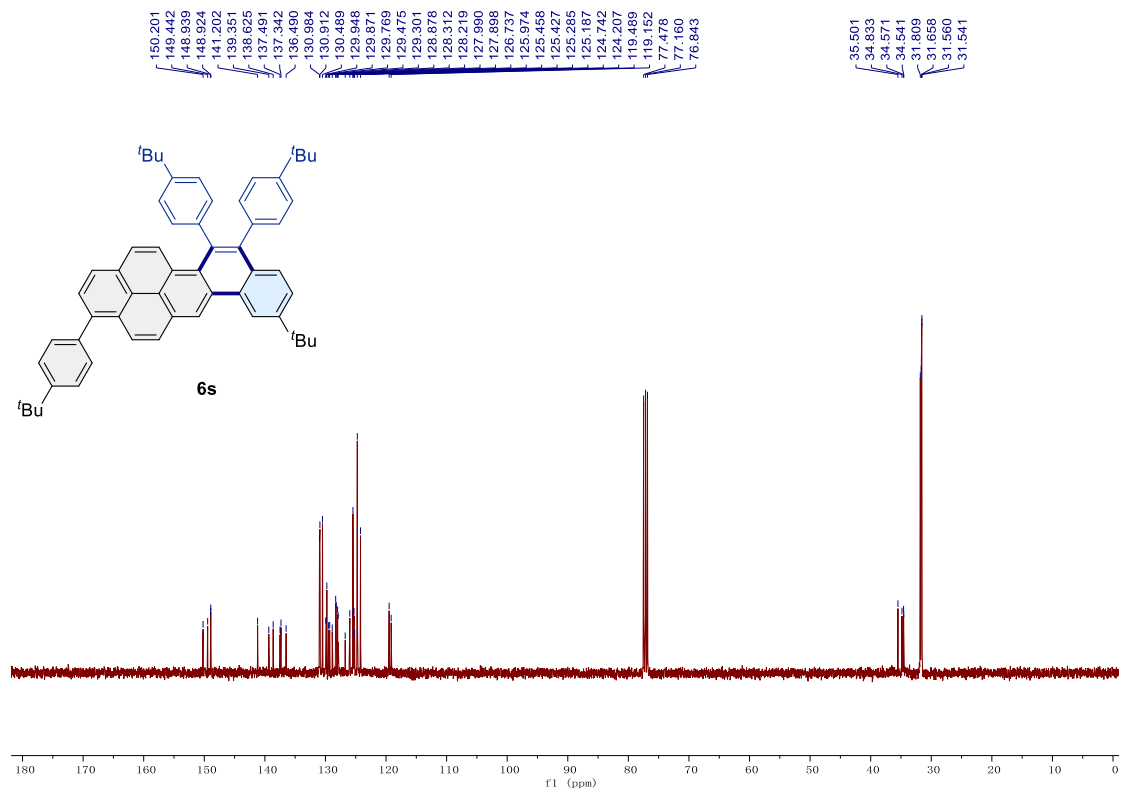
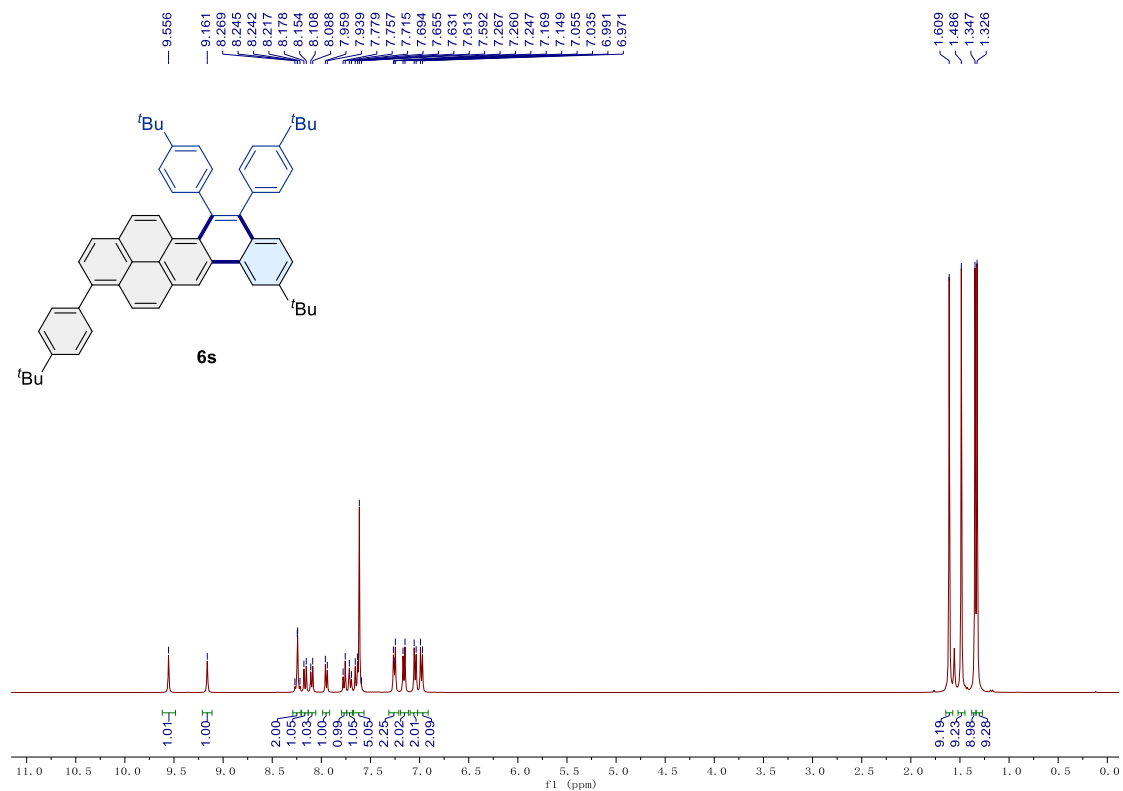


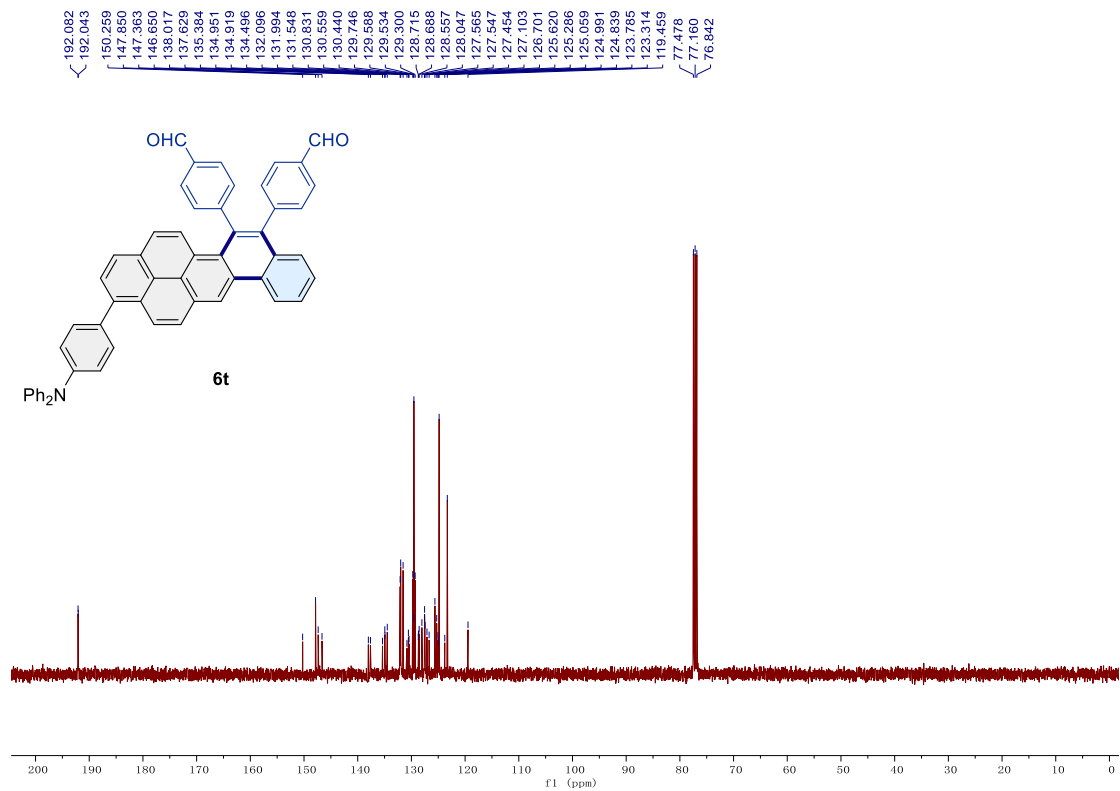
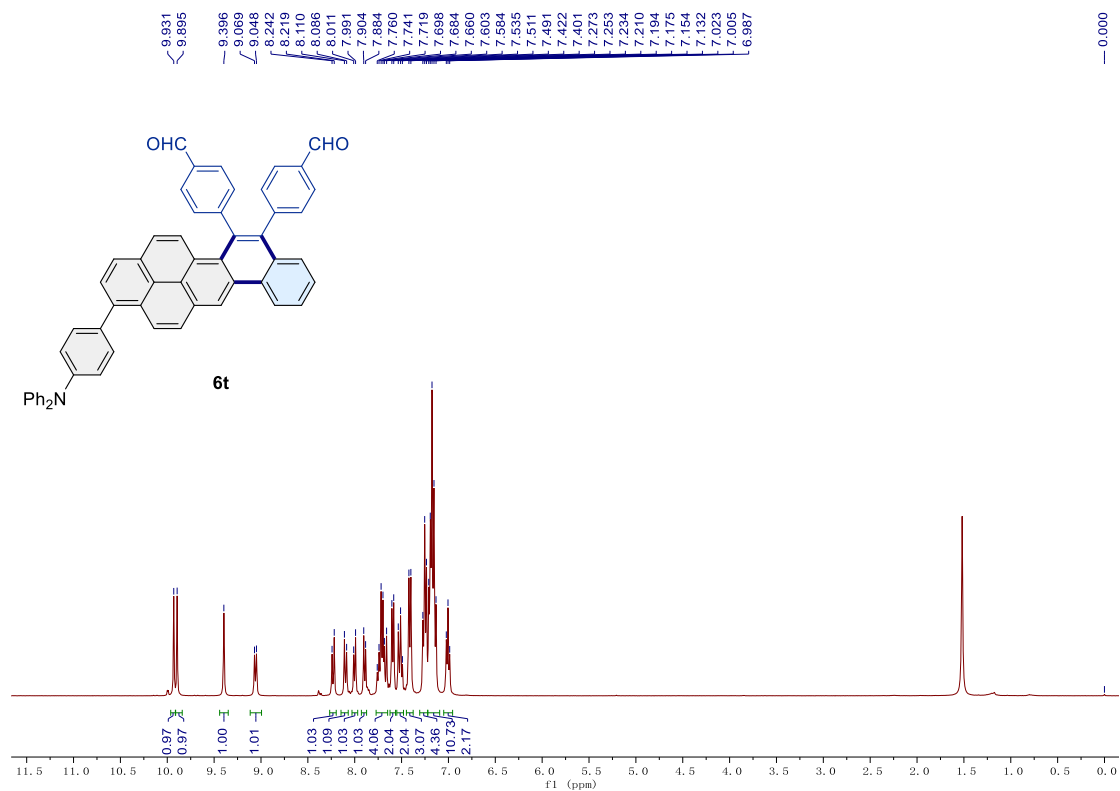


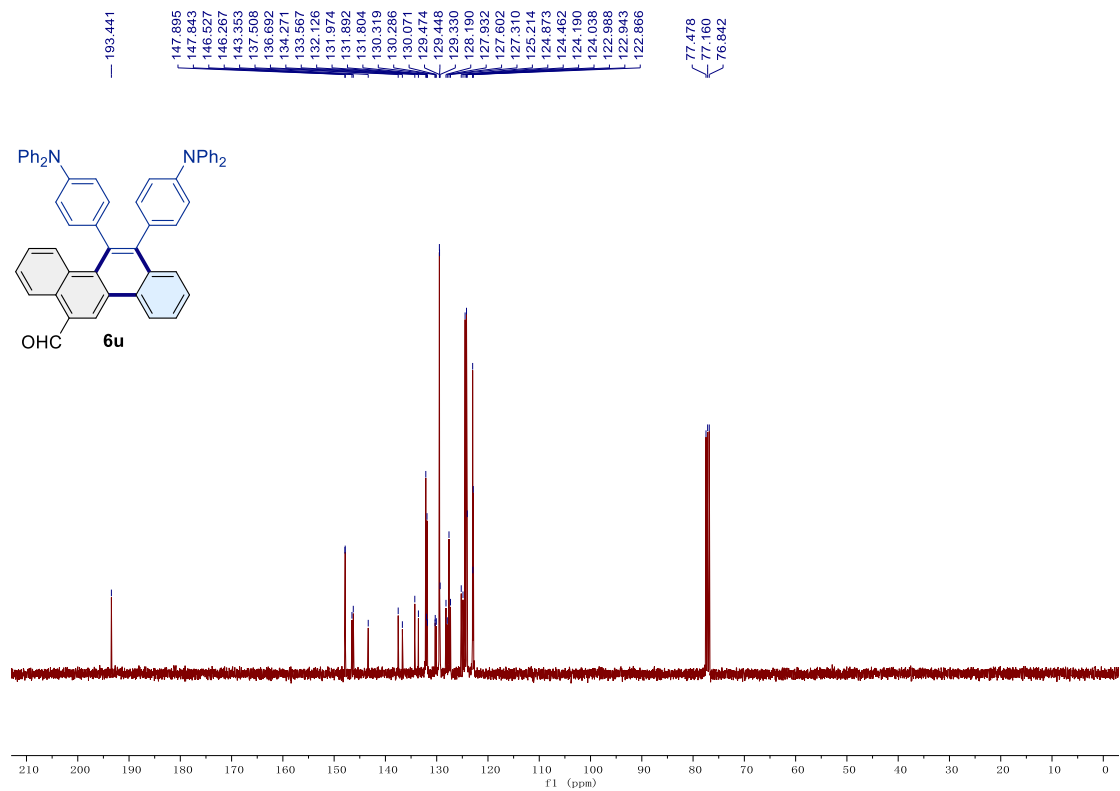
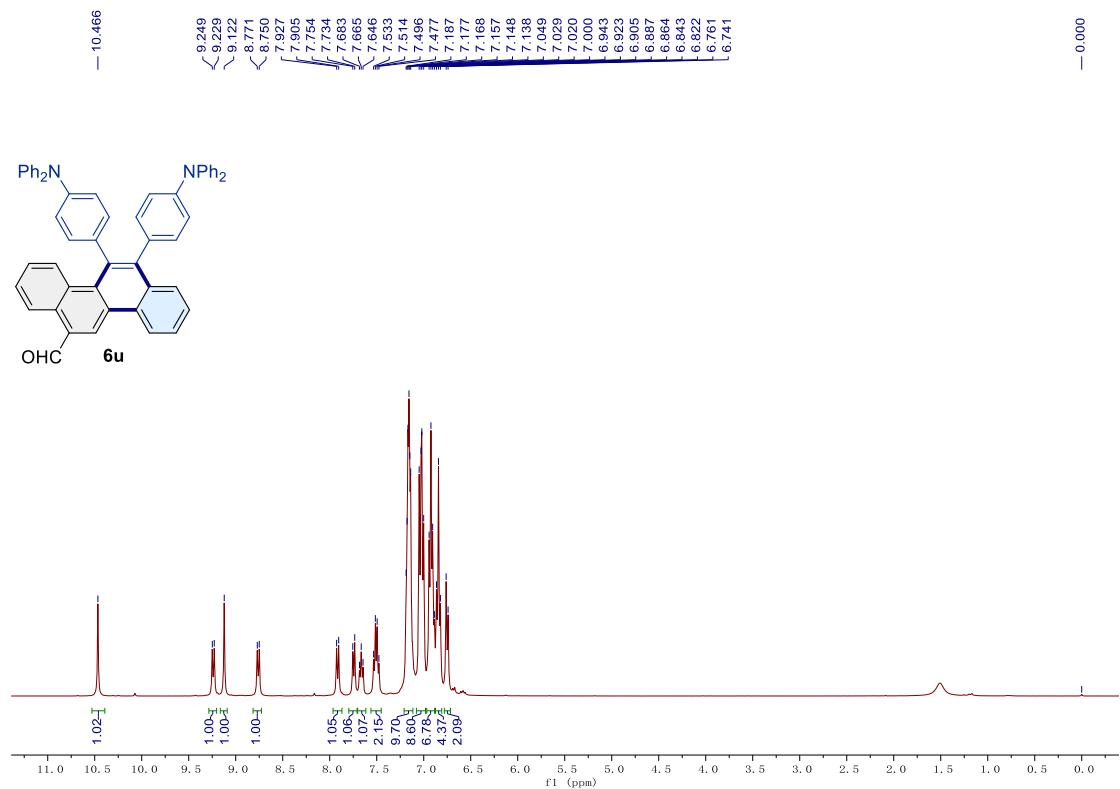


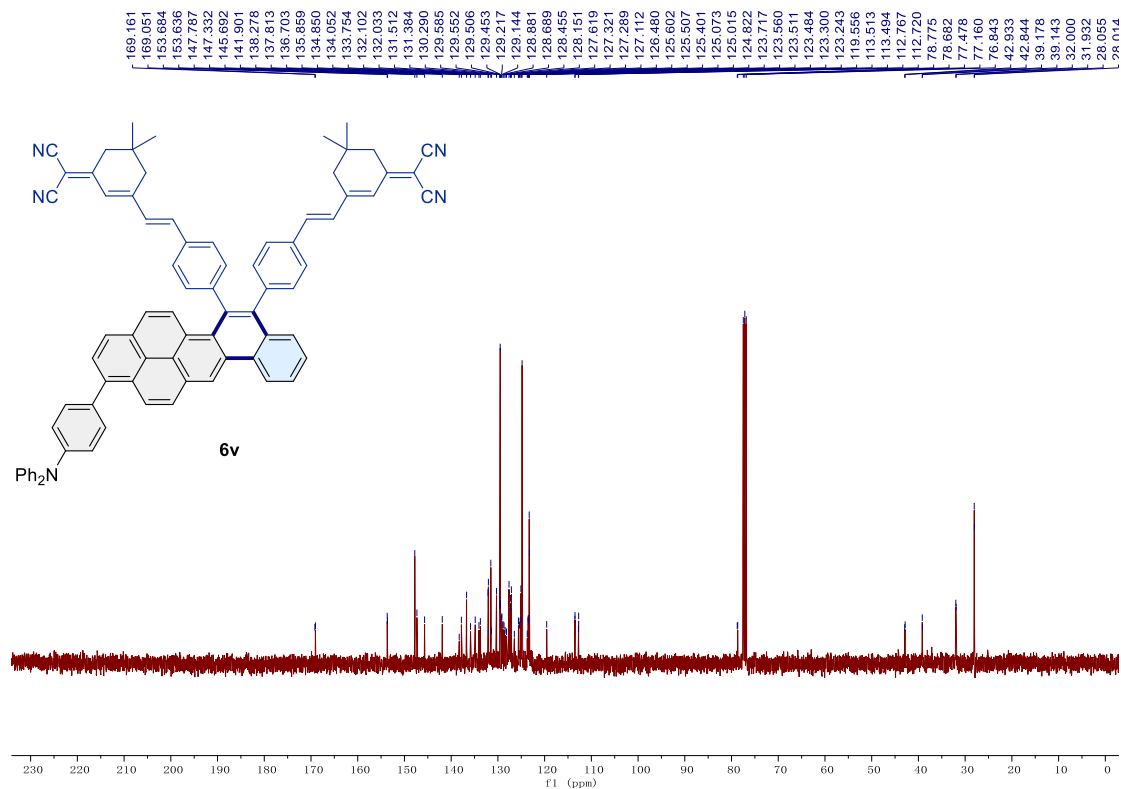
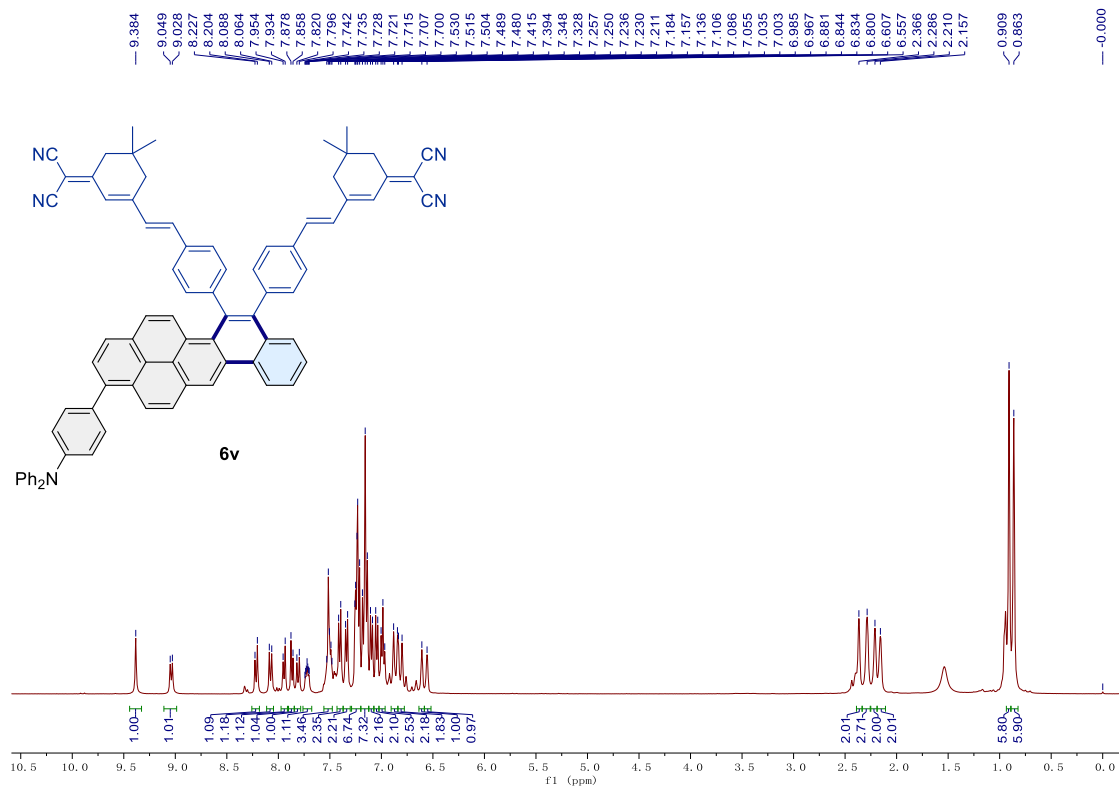


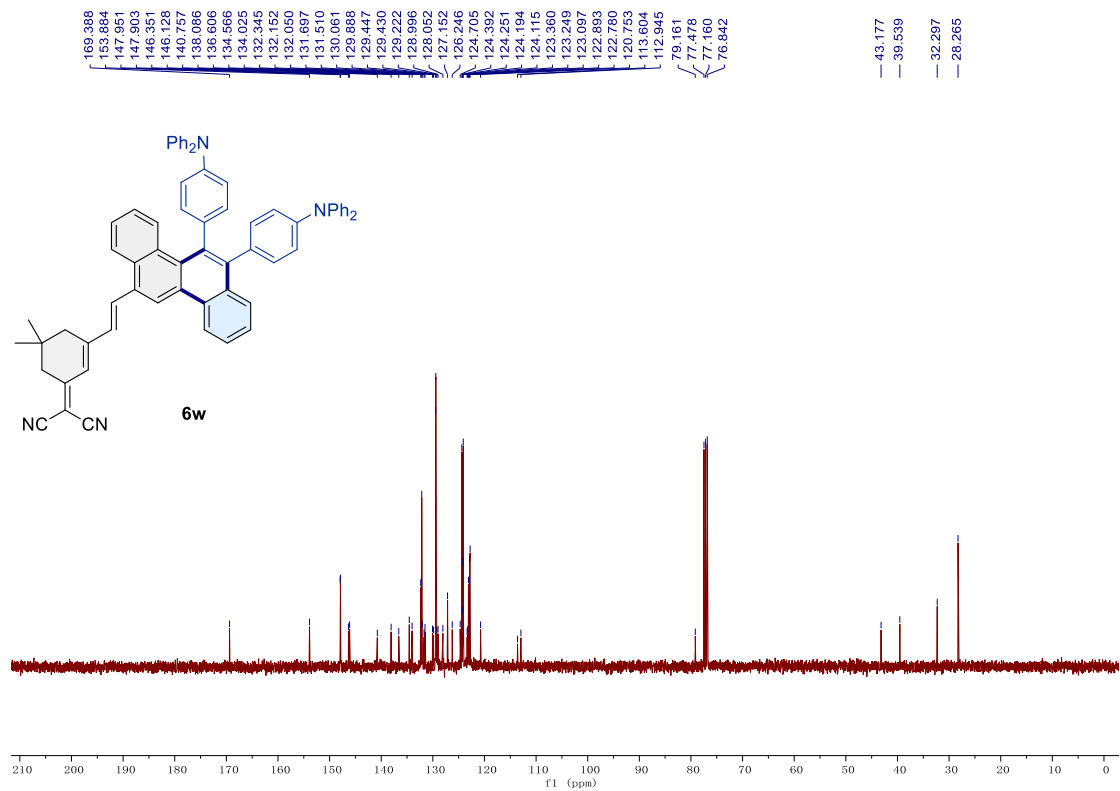
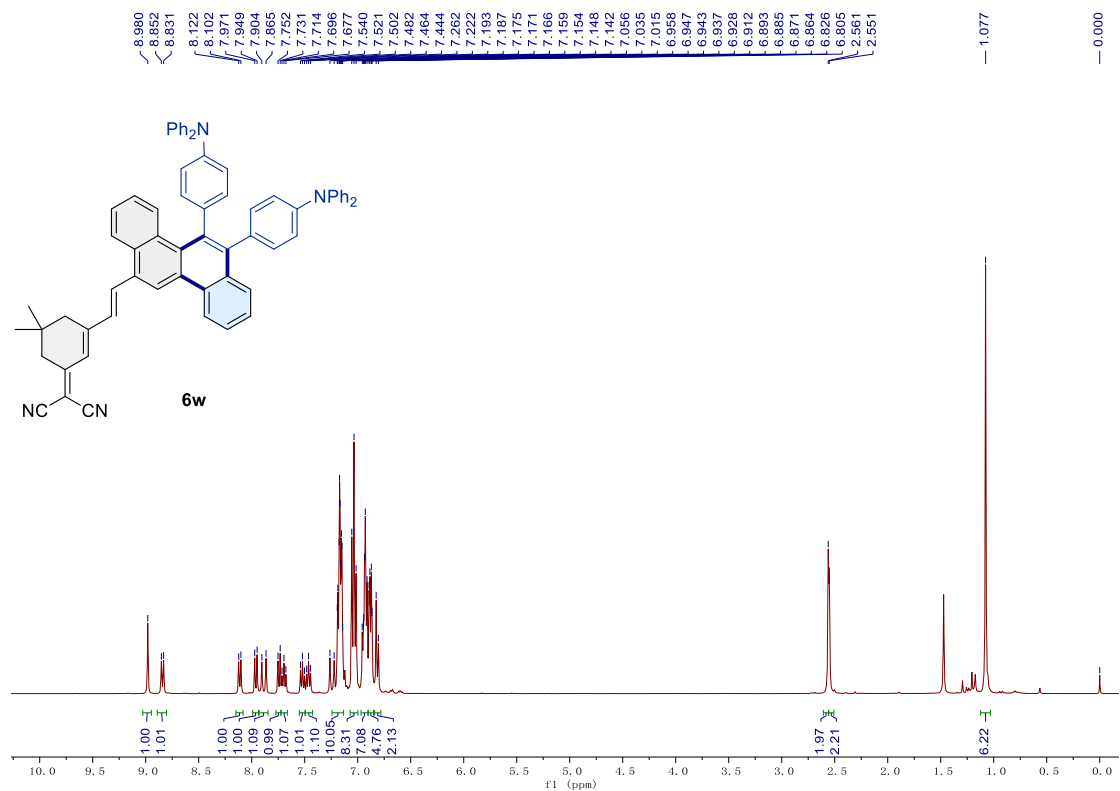












NMR spectrum of substrates 1t – u:

