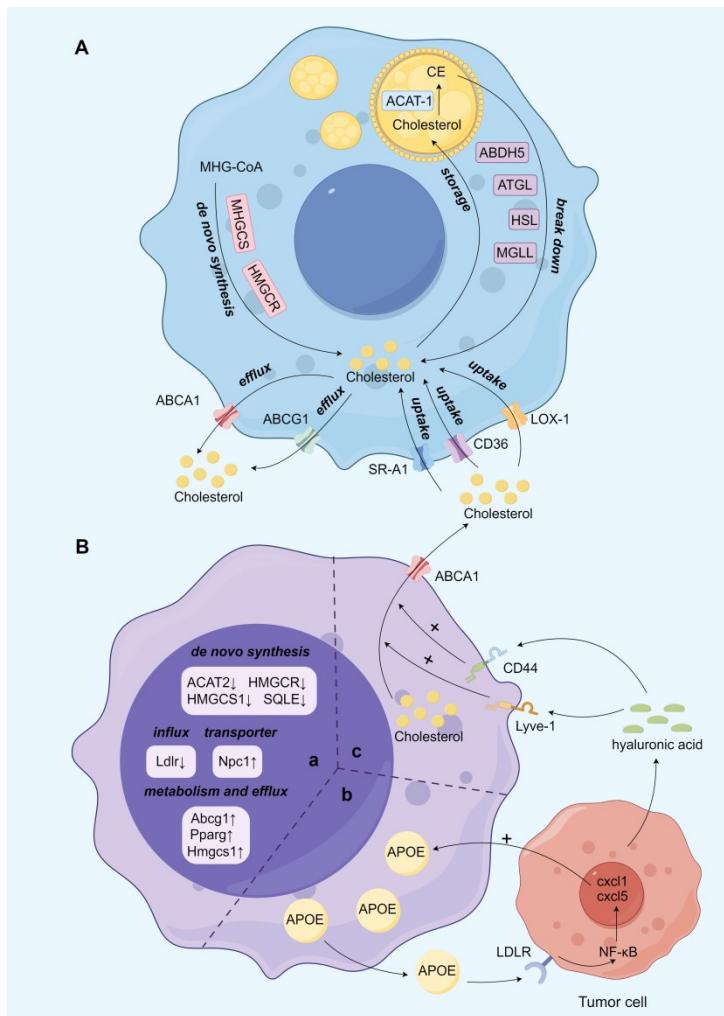
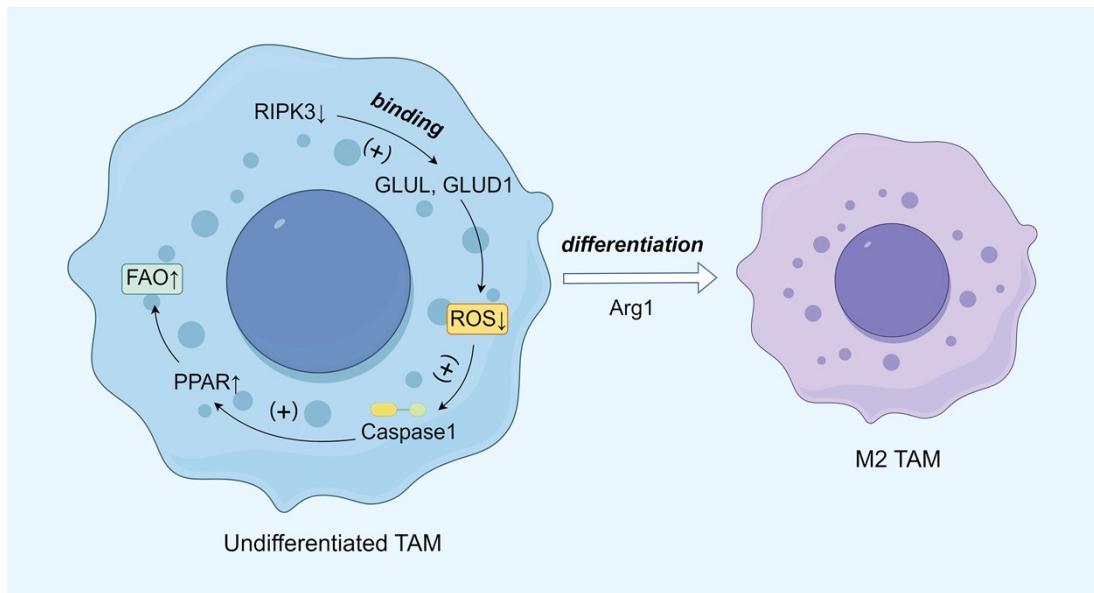


Supplemental Figures:



**Figure S1.** Cholesterol metabolism in Macrophages versus TAMs. **A:** Normal macrophages perform cholesterol metabolism through uptake, de novo synthesis, storage and efflux. **B:** **(a)** TAMs showed reduced expression of genes involved in cholesterol de novo synthesis and cholesterol influx, whereas the expression of genes involved in cholesterol metabolism, efflux, and cholesterol transport was upregulated. **(b)** TAMs interact with tumor cells to elevate ApoE expression, leading to increased cholesterol efflux. **(c)** The tumor cell product hyaluronic acid acts on TAMs to facilitate cholesterol efflux.



**Figure S2.** Macrophages mediated by RIPK3 deficiency are susceptible to differentiation towards the M2 phenotype. RIPK3 deficiency reduces ROS production, which subsequently leads to a reduction in Caspase-1-mediated cleavage of PPAR, and elevated levels of PPAR regulate the gene expression of molecules encoding molecules used for FAO, such as inducing the expression of Arg1, which predisposes macrophages to differentiate toward the M2 phenotype.

**Supplemental Tables:**

**Table 1.** The senescent cells produced SASPs.

SASP	Senescent cells	Role in	
		tumor	Ref.
		immunity	
IL-6	Osteoblast		1
CCL5	Melanocyte		2
CXCL14	Hepatocyte	Promoting tumor	3
VEGF	Pancreatic ductal cell	immunity	4
IL-6, IFN-γ	Hepatic stellate cell		5-6
IL-1β	Hepatic stellate cell		7
CCL2	Hepatocyte		8
MMPs	Fibroblast		9
CXCL1	Prostate epithelial cell	Suppressing tumor	10
CXCL2	Thyroid follicular cell	immunity	11
CXCL12			
IL-6	Melanocyte		12
IL-1β	Bronchial cell		13

**Table S2.** Drugs in clinical trials targeting abnormal cholesterol metabolism in tumor-associated macrophages.

Active Ingredient	Tumor type	Target Gene	Interventions	Locations	Clinical Status	Ref.
Atorvastatin	Prostate cancer	HMG-CoA	Atorvastatin	Finland	Phase 1	<sup>14</sup>
Statin	Solid tumors	HMG-CoA	Statin	Egypt	Phase 1/2	<sup>15</sup>
Oncolytic Virus	Triple-negative breast cancer	ApoA1	Oncolytic virus (ADV-ApoA1) injection	China	Not applicable	<sup>16</sup>
Fenofibrate	Multiple myeloma	PPAR $\alpha$	Fenofibrate	United States	Phase 2	—
GSK5733584	Advanced solid tumor	B7-H4	Intravenous injection	Japan	Phase 1	—
Cholesterol-fus1	Lung cancer	fus1	DOTAP : Chol-fus1	United States	Phase 1	<sup>14</sup>
TAS-117 and TAS-120	Advanced solid tumors	Akt and FGFR	TAS-117 combined with TAS-120	Japan	Phase 1/2	—

**Table S3.** Nanomaterials in clinical trials targeting abnormal cholesterol metabolism.

Vehicle Type	Active Ingredient	Specific Materials	Type of disease	Tar get	Intervention s	Locati ons	Clinica l	Ref.
Status								
Liposome e	Methotrexat	LDE	COVID-19	DHF	Methotrexate -LDE	Brazil	Phase 1/2	17
Liposome	Paclitaxel	LDE	Stable coronary disease	tubulin	LDE- Paclitaxel	Brazil	Phase 2/3	18
Liposome e	Methotrexat	LDE	Stable coronary disease	DHF	Methotrexate -LDE	Brazil	Phase 2/3	19
Liposome vit	Phospholipo	Phospholipids	atherosclerosis	HDL	Phospholipo	Russia	Phase 2/3	—
			nanoemulsion		vitamin	United Federation		
Complex	IL-12 DNA	PEG-PEI-	Ovarian	EGF	PEG-PEI-	United States	Phase 1	—
	Plasmid	cholesterol	epithelial, fallopian tube and primary	R	cholesterol lipopolymer-encased IL-12 DNA			
	Vector	ol Lipopol						
	GEN-1 and Pegylated	ymer						
	Liposomal		peritoneal		Plasmid			
	Doxorubicin		cancer		Vector GEN-1			
	Hydrochlori							

de

---

**Table S4.** Key studies on cholesterol metabolism abnormalities in TAMs and tumor immune suppression

<b>Key Finding</b>	<b>Mechanism</b>	<b>Impact on Tumor Immunity</b>	<b>Ref.</b>
Tumor cells promote cholesterol efflux from TAMs, depleting membrane cholesterol.	Depleted cholesterol activates PI3K-AKT-mTORC2, enhancing IL-4-induced M2 polarization.	Promotes immunosuppressive TME; reduces CD8 <sup>+</sup> T cell activity.	20
NPC1 upregulation in TAMs reduces intracellular cholesterol levels.	Enhanced cholesterol export via NPC1 disrupts lipid raft signaling.	Impairs antigen presentation; dampens anti-tumor immunity.	21
CH25H/25HC axis in TAMs depletes arginine via Arg1 upregulation.	25HC inhibits T cell proliferation by reducing arginine availability.	Suppresses cytotoxic T cell responses; facilitates immune escape.	22

Low cholesterol in TME induces CD8+ T cell dysfunction and autophagy.	Cholesterol deficiency downregulates hypoxia-induced anti-tumor pathways.	Reduces T cell cytotoxicity and survival.	23
Impaired cholesterol efflux in prostate cancer TAMs reduces androgen synthesis in tumor cells.	Low cholesterol limits androgen receptor activation in cancer cells.	Slows tumor progression but sustains immunosuppressive TME.	24

---

## References

- 1 H. Ying, Z. Q. Li, M. P. Li and W. C. Liu, *FRONT ENDOCRINOL*, 2023, **14**, 1217669.
- 2 A. F. Rezk, D. M. Kemp, M. El-Domyati, W. H. El-Din, J. B. Lee, J. Uitto, O. Igoucheva and V. Alexeev, *J. INVEST. DERMATOL.*, 2017, **137**, 1126.
- 3 D. Umbaugh, N. Nguyen, G. S. Guerrero, C. Villanueva, B. Hagenbuch, A. Ramachandran and H. Jaeschke, *The Journal of Pharmacology and Experimental Therapeutics*, 2023, **385**, 226.
- 4 C. A. Wang, I. H. Chang, P. C. Hou, Y. J. Tai, W. N. Li, P. L. Hsu, S. R. Wu, W. T. Chiu, C. F. Li, Y. S. Shan and S. J. Tsai, *J. EXTRACELL VESICLES*, 2020, **9**, 1746529.
- 5 S. Iwahasi, F. Rui, Y. Morine, S. Yamada, Y. U. Saito, T. Ikemoto, S. Imura and M. Shimada, *ANTICANCER RES.*, 2020, **40**, 743.
- 6 L. Basset, S. Chevalier, Y. Danger, M. I. Arshad, C. Piquet-Pellorce, H. Gascan and M. Samson, *J. MOL. MED.*, 2015, **93**, 1355.
- 7 S. Robert, T. Gicquel, A. Bodin, A. Fautrel, E. Barreto, T. Victoni, V. Lagente and E. Boichot, *INT. IMMUNOPHARMACOL.*, 2019, **72**, 12.

- 8 T. Deng, J. Zhao, Y. Tong, Z. Chen, B. He, J. Li, B. Chen, R. Li, L. Deng, H. Yu, B. Zhang, T. Zhang, Z. Shi, B. Gao, J. Jiang, Y. Shan, Z. Yu, Y. Jin, Y. Wang, J. Xia and G. Chen, *ONCOGENE*, 2024, **43**, 944.
- 9 Y. Kurihara, M. Hatori, Y. Ando, D. Ito, T. Toyoshima, M. Tanaka and S. Shintani, *CLIN. EXP. METASTAS.*, 2009, **26**, 425.
- 10 S. L. Ciummo, L. D'Antonio, C. Sorrentino, C. Fieni, P. Lanuti, G. Stassi, M. Todaro and E. Di Carlo, *FRONT CELL DEV BIOL*, 2021, **9**, 689286.
- 11 X. Zhu, Q. Bai, Y. Lu, Y. Lu, L. Zhu, X. Zhou and L. Wu, *INT. J. ONCOL.*, 2016, **48**, 2321.
- 12 M. J. Alasady, A. R. Terry, A. D. Pierce, M. C. Cavalier, C. S. Blaha, K. A. Adipietro, P. T. Wilder, D. J. Weber and N. Hay, *PLOS ONE*, 2021, **16**, e0256238.
- 13 K. You, P. Parikh, K. Khandalavala, S. A. Wicher, L. Manlove, B. Yang, A. Roesler, B. B. Roos, J. J. Teske, R. J. Britt, C. M. Pabelick and Y. S. Prakash, *AM. J. PHYSIOL.-LUNG C.*, 2019, **317**, L525.
- 14 C. Lu, D. J. Stewart, J. J. Lee, L. Ji, R. Ramesh, G. Jayachandran, M. I. Nunez, I. I. Wistuba, J. J. Erasmus, M. E. Hicks, E. A. Grimm, J. M. Reuben, V. Baladandayuthapani, N. S. Templeton, J. D. McMannis and J. A. Roth, *PLOS ONE*, 2012, **7**, e34833.
- 15 B. A. Radman, A. Alhameed, G. Shu, G. Yin and M. N. Wang, *J. MATER CHEM B*, 2024, **12**, 5299.
- 16 J. Dong, L. Kong, S. Wang, M. Xia, Y. Zhang, J. Wu, F. Yang, S. Zuo and J. Wei, *J. EXP. CLIN. CANC. RES.*, 2024, **43**, 102.
- 17 N. Chen, M. Zhou, X. Dong, J. Qu, F. Gong, Y. Han, Y. Qiu, J. Wang, Y. Liu, Y. Wei, J. Xia, T. Yu, X. Zhang and L. Zhang, *LANCET*, 2020, **395**, 507.
- 18 R. C. Maranhao, C. G. Vital, T. M. Tavoni and S. R. Graziani, *EXPERT OPIN DRUG DEL*, 2017, **14**, 1217.
- 19 D. H. Solomon, E. W. Karlson, E. B. Rimm, C. C. Cannuscio, L. A. Mandl, J. E. Manson, M. J. Stampfer and G. C. Curhan, *CIRCULATION*, 2003, **107**, 1303.
- 20 P. Goossens, J. Rodriguez-Vita, A. Etzerodt, M. Masse, O. Rastoin, V. Gouirand, T. Ulas, O. Papantonopoulou, M. Van Eck, N. Auphan-Anezin, M. Bebien, C. Verthuy, P. V. M. Thien, M. Turner, M. Dalod, J. L. Schultze and T. Lawrence, *CELL METAB*, 2019, **29**, 1376.
- 21 N. Shao, H. Qiu, J. Liu, D. Xiao, J. Zhao, C. Chen, J. Wan, M. Guo, G. Liang, X. Zhao and L. Xu, *J. ADV RES*, 2025, **68**, 99.
- 22 J. Xiao, S. Wang, L. Chen, X. Ding, Y. Dang, M. Han, Y. Zheng, H. Shen, S. Wu, M. Wang, D. Yang, N. Li, C. Dong, M. Hu, C. Su, W. Li, L. Hui, Y. Ye, H. Tang, B. Wei and H. Wang, *IMMUNITY*, 2024, **57**, 1087.
- 23 H. Xu, S. Zhou, Q. Tang, H. Xia and F. Bi, *BBA-REV. CANCER*, 2020, **1874**, 188394.
- 24 I. Liikanen, C. Lauhan, S. Quon, K. Omilusik, A. T. Phan, L. B. Bartroli, A. Ferry, J. Goulding, J. Chen, J. P. Scott-Browne, J. T. Yustein, N. E. Scharping, D. A. Witherden and A. W. Goldrath, *J. CLIN. INVEST.*, 2021, **131**.