Supporting information

Water soluble synthetic dieptide-based biodegradable nanoporous materials

Samit Guha, Tushar Chakraborty, and Arindam Banerjee*



Figure S1. ¹H NMR (300 MHz, D₂O) spectrum of dipeptide 1.



Figure S2. ¹³C NMR (75 MHz, D₂O) spectrum of dipeptide 1.



Figure S3. DEPT 135 (75 MHz, D₂O) spectrum of dipeptide 1.



Figure S4. HRMS (ESI) data of dipeptide 1.



Figure S5. ¹H NMR (300 MHz, D₂O) spectrum of dipeptide **2**.



Figure S6. ¹³C NMR (75 MHz, D₂O) spectrum of dipeptide 2.



Figure S7. DEPT 135 (75 MHz, D₂O) spectrum of dipeptide 2.



Figure S8. HRMS (ESI) data of dipeptide 2.



ESI Fig. S1a EDX analysis of nanoporous material based on dipeptide **1** indicates the presence of C, N and O.



ESI Fig. S1b EDX analysis of nanoporous material based on dipeptide **2** indicates the presence of C, N and O.

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ESI Fig. S2 (a–d) Time dependent biodegradation of dipeptide **1** at 0h, 24h (1 day), 7 days and 14 days respectively after treatment with bacterial consortium quantified through ESI-MS data suggests that dipeptide **1** molecule is biodegradable. It has been observed that the intensity of the molecular ion peak for dieptide **1**, $[M+H]^+$ is gradually decreasing with respect to time. This is due to the result of the degradation of the peptide.



ESI Fig. S3 (a–d) Time dependent biodegradation of dipeptide **2** at 0h, 24h (1 day), 7 days and 14 days respectively after treatment with bacterial consortium quantified through ESI-MS data suggests that dipeptide **2** molecule is biodegradable. It has been observed that the intensity of the molecular ion peak for dieptide **2**, $[M+H]^+$ is gradually decreasing with respect to time. This is due to the result of the degradation of the peptide.



ESI Fig. S4 Bacterial colonies from the culture medium grown on M9 agar plates from 0 hr (A), 24 hr (B), 7 days (C) and 14 days (D) after growth in M9-peptide medium. The heterogeneity of the consortia is reflected in the different colony morphology.