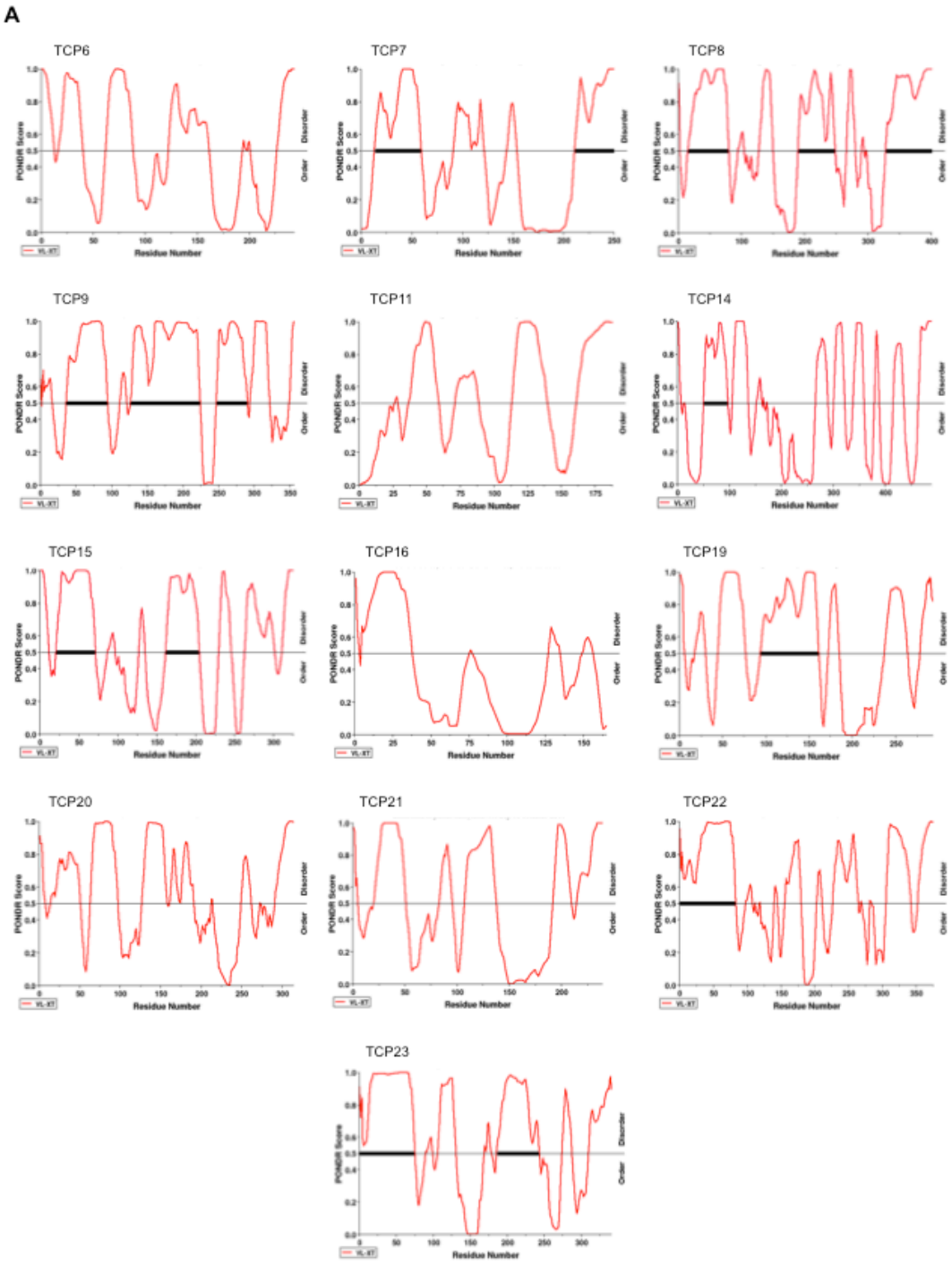

Figure S1

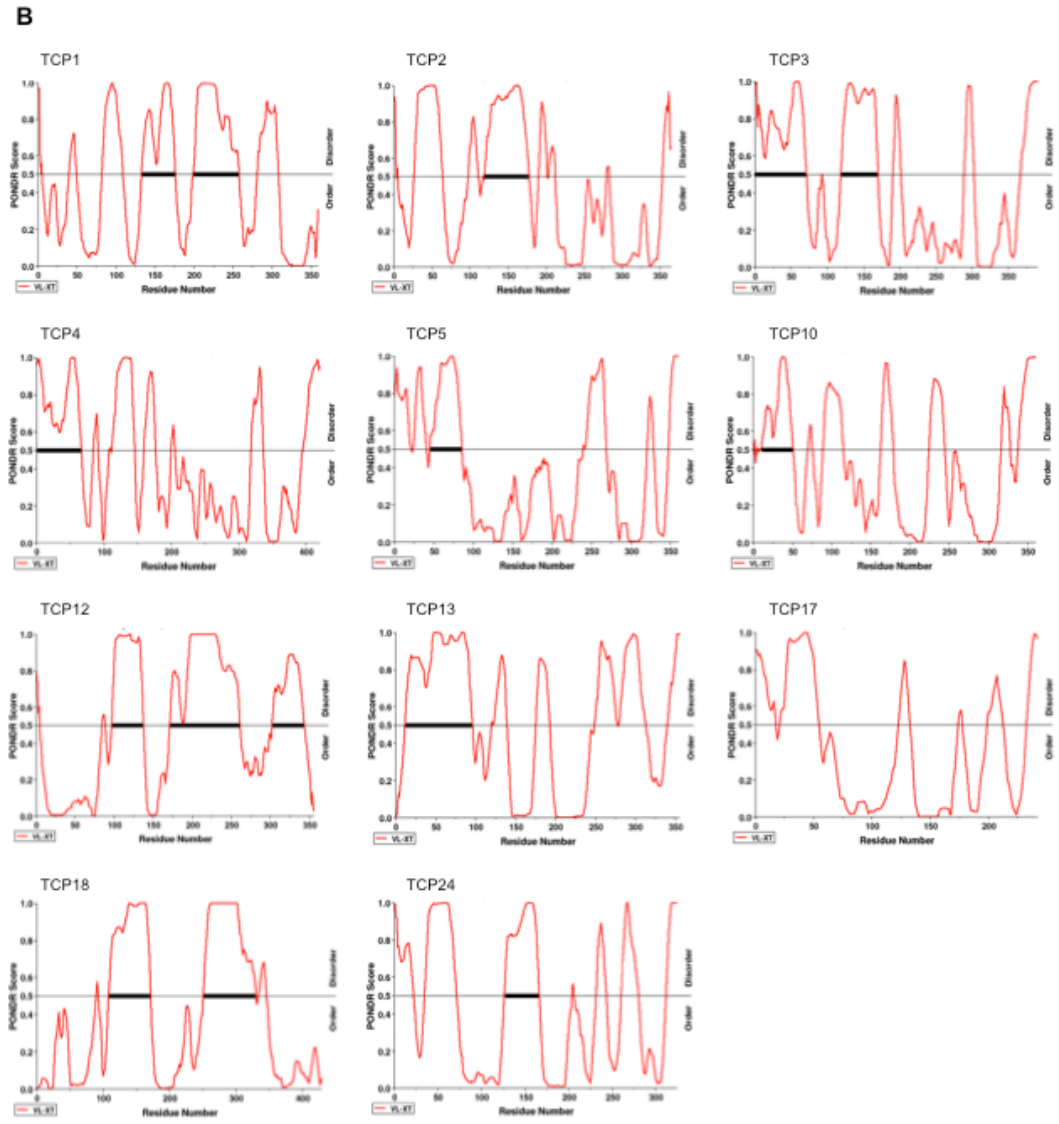

Figure S1. PONDR prediction of disorder in the TCP family. Disorder-prediction scores were plotted against each amino acid residue for class I TCPs **(A)** and class II TCPs **(B)**. The residues that show values above 0.5 are considered as disordered. Thicker line on the threshold line indicates a region of 40 or more consecutive disordered residues.

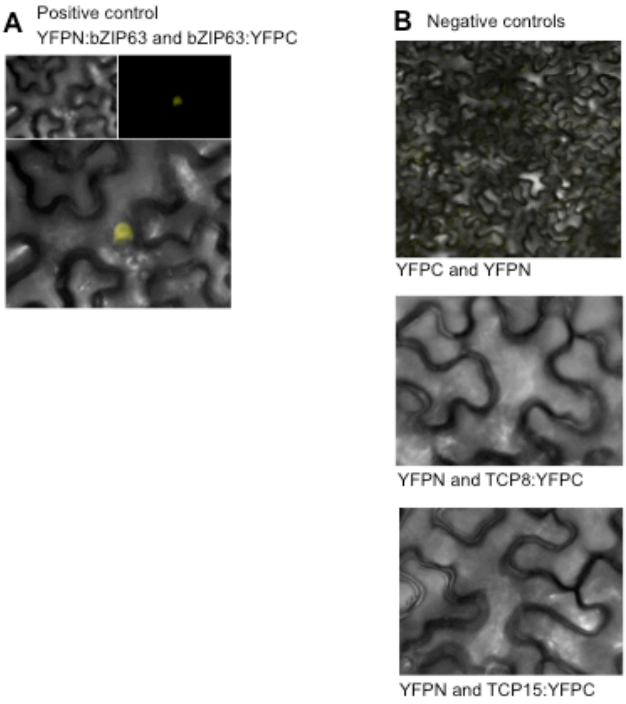

Figure S2. Control for BiFC experiments. Positive **(A)** and negative **(B)** BiFC controls, merge of bright-field and YFP fluorescence.

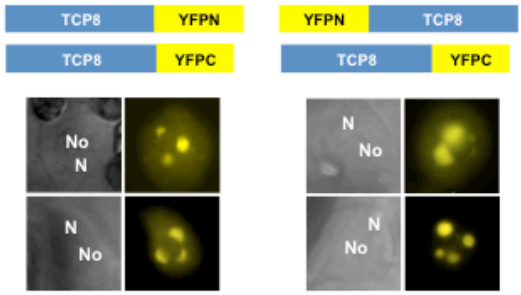

Figure S3. Additional BiFC views depending on the TCP8 position versus YFP.

N. benthamiana leaves were infiltrated with a mix of *Agrobacterium tumefaciens* transformed with BiFC plasmids carrying the full-length cDNA of *TCP8* in frame with YFPN or YFPC (as indicated in the figure). Brightfield (left panels), YFP fluorescence (right panels); N, nucleus; No, nucleolus. Views are representative of 3 biologically independent experiments.

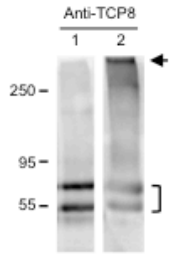

Figure S4. The C-terminal part of TCP8 is involved in its aggregation capacity.

Proteins from *N. benthamiana* leaves infiltrated with TCP8 fused with YFPN or YFPC at its C- (lane 1) or N- (lane 2) terminal part, were extracted under denaturating conditions, loaded on SDS-PAGE and blotted. Membranes were probed with anti-TCP8 antibodies. Molecular weight markers (kDa) are indicated on the left.

Primers used for two-hybrid constructions

F *TCP8* EcoRI	5'-TCGAATTCGATCTCTCCGAC-3'
R *TCP8* BamHI	5'-TAGGATCCTCACTCAGAGCT-3'

Primers used for BiFC constructions

F1 *TCP8* BamHI	5'-ATGGATCCATGGATCTCTC-3'
R1 *TCP8* XhoI	5'-ATCTCGAGCTCAGAGCTAT-3'
F2 *TCP8* BamHI	5'-ATGGATCCGATCTCTCCGA-3'
R2 *TCP8* XhoI	5'-ATCTCGAGTCACTCAGAGC-3'
F1 *TCP15* StuI	5'-ATAGGCCTATGGATCCGGAT-3'
R1 *TCP15* XhoI	5'-TCTCGAGGGAATGATGACTG-3'
F3 *TCP8*	5'-GAACTCGGCTTATTAAAGAGGCGGAAACGCAAACGC-3'
R3 *TCP8*	5'-GCGTTTGCGTTTCCGCCTCTTTAATAAGCCGAGTTC-3'

Table S1. Pairs of primers used for the cloning of *TCP8* and *TCP15* in two-hybrid and BiFC plasmids.

F1 and R1 primers are used to clone *TCP* cDNA in pSPYCE(MR) or pSPYNE(R)173.

F2 and R2 primers are used to clone *TCP* cDNA in pSPYNE173 or pSPYCE(M).

F3 and R3 primers are used to generate the truncated version of *TCP8*.