

A new route towards fimbrolide analogues: importance of the exomethylene motif in LuxR dependent quorum sensing inhibition

Mohamad Sabbah,^{a,b} Maud Bernollin,^{a,b} Alain Doutheau,^{a,b} Laurent Soulere,^{a,b}
and Yves Queneau^{a,b}

^a INSA Lyon, ICBMS, Bât J. Verne, 20 av A. Einstein, 69621 Villeurbanne Cedex, France

^b CNRS, UMR 5246, ICBMS, Université Lyon 1, INSA-Lyon, CPE-Lyon, Bât CPE, 43 bd
du 11 novembre 1918, 69622 Villeurbanne Cedex, France

Supplementary Information

| | |
|---|-----------|
| I. SYNTHESIS ----- | 3 |
| I.1. General ----- | 3 |
| I.2. General procedure A for aldolisation reaction of lactone 4 ----- | 3 |
| I.2.1. 3-butyl-3-(phenylthio)dihydrofuran-2(3H)-one (5a)----- | 4 |
| I.2.2. 3-hexyl-3-(phenylthio)dihydrofuran-2(3H)-one (5b) ----- | 4 |
| I.2.3. 3-(1-hydroxybutyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5c) ----- | 5 |
| I.2.4. 3-(1-hydroxyhexyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5d)----- | 5 |
| I.2.5. 3-(1-hydroxyoctyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5e, 5e')----- | 6 |
| I.3. General procedure B for oxydation-elimination reaction of lactones 5 ----- | 6 |
| I.3.1. 3-butylfuran-2(5H)-one (6a) ¹ ----- | 6 |
| I.3.2. 3-hexylfuran-2(5H)-one (6b) ----- | 7 |
| I.3.3. 3-(1-hydroxybutyl)furan-2(5H)-one (6c)----- | 7 |
| I.3.4. 3-(1-hydroxyhexyl)furan-2(5H)-one (6d)----- | 7 |
| I.3.5. 3-(1-hydroxyoctyl)furan-2(5H)-one (6e)----- | 8 |
| I.4. General procedure C for methylenation reaction of furanone 6 ----- | 8 |
| I.4.1. 3-butyl-5-methylenefuran-2(5H)-one (8a)----- | 8 |
| I.4.2. 3-hexyl-5-methylenefuran-2(5H)-one (8b) ² ----- | 9 |
| I.4.3. 3-(1-hydroxybutyl)-5-methylenefuran-2(5H)-one (8c) ² ----- | 9 |
| I.4.4. 3-(1-hydroxyhexyl)-5-methylenefuran-2(5H)-one (8d) ³ ----- | 9 |
| I.4.5. 3-(1-hydroxyoctyl)-5-methylenefuran-2(5H)-one (8e) ² ----- | 9 |
| I.5. General procedure D for bromation reaction of methylene-furanone 8 ⁴ ----- | 10 |
| I.5.1. (Z)-5-(bromomethylene)-3-butylfuran-2(5H)-one (10a)----- | 10 |

| | | |
|--|---|-----------|
| I.5.2. | (Z)-5-(bromomethylene)-3-hexylfuran-2(5H)-one (10b) ⁵ ----- | 10 |
| I.5.3. | (Z)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)----- | 11 |
| | (E)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c) ----- | 11 |
| I.5.4. | (Z)-5-(bromomethylene)-3-(1-hydroxyhexyl)furan-2(5H)-one (10d) ----- | 11 |
| I.5.5. | (Z)-5-(bromomethylene)-3-(1-hydroxyoctyl)furan-2(5H)-one (10e)----- | 11 |
| II. BIOLOGICAL EVALUATIONS ----- | | 12 |
| II.1. | Agonistic activity ----- | 12 |
| II.2. | Antagonistic activity ----- | 12 |
| II.3. | Antibacterial activity ----- | 12 |
| III. MOLECULAR MODELLING.----- | | 14 |
| IV. ¹H AND ¹³C NMR SPECTRUM ----- | | 16 |
| IV.1.1. | 3-butyl-3-(phenylthio)dihydrofuran-2(3H)-one (5a)----- | 16 |
| IV.1.2. | 3-hexyl-3-(phenylthio)dihydrofuran-2(3H)-one (5b) ----- | 17 |
| IV.1.3. | 3-(1-hydroxybutyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5c)----- | 18 |
| IV.1.4. | 3-(1-hydroxyhexyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5d) ----- | 19 |
| IV.1.5. | 3-(1-hydroxyoctyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5e, 5e') ----- | 20 |
| IV.1.6. | 3-butylfuran-2(5H)-one (6a)----- | 22 |
| IV.1.7. | 3-hexylfuran-2(5H)-one (6b) ----- | 23 |
| IV.1.8. | 3-(1-hydroxybutyl)furan-2(5H)-one (6c)----- | 25 |
| IV.1.9. | 3-(1-hydroxyhexyl)furan-2(5H)-one (6d) ----- | 26 |
| IV.1.10. | 3-(1-hydroxyoctyl)furan-2(5H)-one (6e)----- | 28 |
| IV.1.11. | 3-butyl-5-methylenefuran-2(5H)-one (8a)----- | 29 |
| IV.1.12. | 3-hexyl-5-methylenefuran-2(5H)-one (8b)----- | 32 |
| IV.1.13. | 3-(1-hydroxybutyl)-5-methylenefuran-2(5H)-one (8c)----- | 33 |
| IV.1.14. | 3-(1-hydroxyoctyl)-5-methylenefuran-2(5H)-one (8e)----- | 36 |
| IV.1.15. | (Z)-5-(bromomethylene)-3-butylfuran-2(5H)-one (10a)----- | 37 |
| IV.1.16. | (Z)-5-(bromomethylene)-3-hexylfuran-2(5H)-one (10b) ----- | 40 |
| IV.1.17. | (Z)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)----- | 41 |
| | (E)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c) ----- | 41 |
| IV.1.18. | (Z)-5-(bromomethylene)-3-(1-hydroxyhexyl)furan-2(5H)-one (10d) ----- | 44 |
| IV.1.19. | (Z)-5-(bromomethylene)-3-(1-hydroxyoctyl)furan-2(5H)-one (10e)----- | 45 |
| V. REFERENCES ----- | | 46 |

I. SYNTHESIS

I.1. General

Solvents were distilled and dried before use: dichloromethane from calcium hydride, tetrahydrofuran from sodium benzophenone ketyl. Organic solutions were dried over anhydrous sodium sulfate. The reactions were performed under a constant flow of nitrogen. The reactions were monitored by t.l.c. on Silica Gel 60 F254 (Merck) and detection was carried out by UV light (254 nm) and/or charring with a 5 % phosphomolybdic acid solution in ethanol containing 10 % of H₂SO₄, or a 1 % potassium permanganate solution in water. Silica gel (Kieselgel 60, 70-230 mesh ASTM, Merck) was employed for column chromatography. Melting points were determined on a Kofler block apparatus.

The ¹H NMR (300 MHz or 400 MHz) and ¹³C NMR (75 MHz or 100 MHz) spectra were recorded with a Bruker ALS300, DRX300, and DRX400 spectrometers. Chemical shifts are given in ppm. Coupling constants are expressed in Hertz and splitting pattern abbreviations are: s, singulet; d, doublet; t, triplet; q, quartet; m, multiplet; M, massif, p, pseudo. Multiplicity (¹³C NMR) was determined by DEPT sequences. High resolution mass spectra were obtained by electro spray technique, positive mode with a ThermoFinnigan MAT 95 XL spectrometer. **α-phenylthio-γ-butyrolactone (4)** obtained following the procedures reported in literature.¹

I.2. General procedure A for aldolisation reaction of lactone 4

To a stirred solution of lithium diisopropylamide (LDA, 1.5 eq), prepared from diisopropylamine (2 eq.) and 1.6 M of n-BuLi in hexane (1.5 eq.), in anhydrous THF at -78 °C, was slowly added a solution of lactone **4** (1 eq) in THF. The temperature was raised to -50 °C and the solution was stirred for 1 h. Then, a solution of aldehyde (1 eq.) in THF was added dropwise. The crude mixture was left at -50 °C for 2 h, at 0 °C for 30 mn and it was poured over saturated ammonium chloride solution. The ether extracts were dried over anhydrous sodium sulfate and the solvent was removed under reduced pressure. The crude product was purified by column chromatography through silica gel.

1.2.1. 3-butyl-3-(phenylthio)dihydrofuran-2(3H)-one (5a)

Diisopropylamine (3.6 mL, 25.7 mmol) was added to anhydrous THF (20 mL) under nitrogen. After cooling the solution to 0 °C, n-butyllithium (12 mL of a 1.6 M solution in hexane, 19.2 mmol) was added and the reaction mixture was stirred for 30 min. The solution was then cooled to -78 °C and subsequently treated with the lactone **4** (2.5 g, 12.8 mmol), dissolved in dry THF (4 mL). After 30 min, a solution of iodobutane (1.5 mL, 12.8 mmol) dissolved in 1 eq. (1.4 mL) of hexamethylphosphoramide (HMPA) was added. When addition was complete, stirring was maintained at -78 °C for 4 h. The reaction was then allowed to warm to room temperature and stirred overnight. The mixture was then quenched by addition of saturated solution of ammonium chloride. The phases were separated and the aqueous layer was extracted with 3 portions (60 mL) of ether. The combined organic extract was washed with water, and saturated NaCl solution (50 mL portions of each). The organic solution was dried over anhydrous Na₂SO₄, and concentrated in vacuum to yield the desired crude products as slightly yellow oils. This crude product was purified by flash chromatography through silica gel using pentane-ether (8/2) as eluent to provide **5a** (2.6 g, 76 %) as mixture of enantiomers. Orange oil; R_f (SiO₂ ; P/Et₂O : 8/2) = 0.23. ¹H NMR (300MHz, CDCl₃) : δ 0.89 (t, J = 7.2Hz, 3H); 1.20 – 1.70 (m, 6H); 2.24 (m, 1H); 2.45 (m, 1H) ; 4.22 (m, 2H); 7.33-7.40 (m, 3H); 7.53 (m, 2H). ¹³C NMR (75MHz, CDCl₃) : δ 14.0; 22.8; 26.7; 34.3; 34.6; 54.1; 65.1; 129.1; 129.5 (2C); 130.1 (2C); 137.2; 176.2. HRMS (CI⁺): Calculated for C₁₄H₁₈O₂SNa⁺, 273.0920; founded MN⁺ = 273.0919.

1.2.2. 3-hexyl-3-(phenylthio)dihydrofuran-2(3H)-one (5b)

Diisopropylamine (3.6 mL, 25.7 mmol) was added to anhydrous THF (20 mL) under nitrogen. After cooling the solution to 0 °C, n-butyllithium (12 mL of a 1.6 M solution in hexane, 19.2 mmol) was added and the reaction mixture was stirred for 30 min. The solution was then cooled to -78 °C and subsequently treated with the lactone **4** (2.5 g, 12.8 mmol), dissolved in dry THF (4 mL). After 30 min, a solution of iodohexane (1.9 mL, 12.8 mmol) dissolved in 1 eq. (1.4 mL) of hexamethylphosphoramide (HMPA) was added. When addition was complete, stirring was maintained at -78 °C for 4 h. The reaction was then allowed to warm to room temperature and stirred overnight. The mixture was then quenched by addition of saturated solution of ammonium chloride. The phases were separated and the aqueous layer was extracted with 3 portions (60 mL) of ether. The combined organic extract was washed with water, and saturated NaCl solution (50 mL portions of each). The organic solution was dried

over anhydrous Na_2SO_4 , and concentrated in vacuum to yield the desired crude products as slightly yellow oils. This crude product was purified by flash chromatography through silica gel using pentane-ether (80:20) as eluent to provide **5b** (2.7 g, 64 %) as mixture of enantiomers. Orange oil; Rf (SiO_2 ; P/Et₂O : 80/20) = 0.29 and 0.47. ¹H NMR (300MHz, CDCl₃) : δ 0.81 (t, *J* = 7.2Hz, 3H); 1.22 (m, 8H); 1.50-1.78 (m, 2H); 2.19 (m, 1H); 2.35 (m, 1H); 4.17 (m, 2H); 7.30-7.36 (m, 3H); 7.40-7.50 (m, 2H). ¹³C NMR (75MHz, CDCl₃) : δ 14.1; 22.6; 24.5; 29.3; 31.7; 34.3; 34.8; 54.1; 65.1; 129.1 (2C); 129.4; 130.1; 137.2 (2C); 176.2. HRMS (CI⁺): Calculated for C₁₆H₂₂O₂SNa⁺, 301.1233; founded MNa⁺ = 301.1232.

I.2.3. 3-(1-hydroxybutyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5c)

According to general procedure A, freshly distilled butanal (926 μL, 10.3 mmol), LDA (1.5 eq), and lactone **4** (2 g, 10.3 mmol) gave the products **5c** (2.2 g, 80 %) as mixture of 3 diastereoisomers (by TLC) after purification by flash chromatography on silica gel. Eluent: dichloromethane-ether (97/3). Characterization of one diastereoisomer was done: yellow solid; mp = 97- 98 °C; Rf (SiO_2 ; CH₂Cl₂/Et₂O: 97/3) = 0.31. ¹H NMR (300MHz, CDCl₃) : δ 0.89 (t, *J* = 7.2Hz, 3H); 1.24 (m, 2H); 1.55 (m, 2H); 1.88 (ddd, *J* = 7.5; 6.6; 1.7Hz, 1H); 2.7 (m, 1H); 3.23 (s, 1H, OH); 3.78 (dd, *J* = 9; 2.8 Hz, 1H); 4.2 (m, 2H); 7.1-7.34 (m, 3H); 7.44 (m, 2H). ¹³C NMR (75MHz, CDCl₃) : δ 14.0; 19.4; 29.0; 32.3; 58.1; 65.7; 70.5; 128.4; 129.1 (2C); 130.3 (2C); 137.2; 175.8. HRMS (CI⁺): Calculated for C₁₄H₁₈O₃SNa⁺, 289.0869; founded MNa⁺: 289.0870.

I.2.4. 3-(1-hydroxyhexyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5d)

According to general procedure A, hexanal (614 μl, 5.12 mmol), LDA and lactone **4** (1 g, 5.12 mmol) gave the products **5d** (836 mg, 55 %) were obtained as mixture of diastereoisomers as yellow oil after purification by flash chromatography on silica gel. Eluent: pentane-ether (6/4). Characterization of one diastereoisomer was done: yellow oil; Rf (SiO_2 ; P/Et₂O: 60/40) = 0.29. ¹H NMR (300MHz, CDCl₃) : δ 0.87 (t, *J* = 7.0Hz, 3H); 1.29 (m, 6H); 1.57-1.64 (m, 2H); 2.14 (ddd, *J* = 7.5; 6.6; 1.7Hz, 1H); 2.55 (m, 1H); 3.19 [s(OH) , 1H]; 3.78 (d, *J* = 10.0Hz, 1H); 4.13 (m, 1H); 4.25 (p td, *J* = 8.9; 1.5Hz, 1H); 7.33-7.44 (m, 3H); 7.58 (m, 2H). ¹³C NMR (75MHz, CDCl₃) : δ 14.3; 22.8 (2C); 26.4; 31.4; 32; 59.5; 65.8; 73.0; 129.0; 129.4 (2C); 130.6 (2C); 137.6; 175.7. HRMS (ESI⁺): Calculated for C₁₆H₂₂O₃S Na⁺, 317.1187; founded MNa⁺, 317.1186.

I.2.5. 3-(1-hydroxyoctyl)-3-(phenylthio)dihydrofuran-2(3H)-one (**5e**, **5e'**)

According to general procedure A, octanal (1.6 mL, 10.24 mmol), LDA and lactone **4** (2 g, 10.24 mmol), gave the diastereoisomers **5e**, **5e'** [2.29 g (3/7), 69 %] in elution order after purification by repeated column chromatography through silica gel: Eluent 1 pentane-ether (6/4), Eluent 2 pure dichloromethane. **5e**: white solid; mp: 42-44°C; Rf (SiO₂; CH₂Cl₂) = 0.52. ¹H NMR of **5e** (300MHz, CDCl₃): δ 0.87 (t, *J* = 7.0Hz, 3H); 1.26-1.37 (m, 10H); 1.64 (m, 2H); 2.14 (ddd, *J* = 14.1; 6.6; 1.7Hz, 1H); 2.55 (m, 1H); 3.18 (s, 1H, OH); 3.78 (d, *J* = 10.0Hz, 1H); 4.13 (m, 1H); 4.27 (p td, *J* = 9; 1.7Hz, 1H); 7.35-7.44 (m, 3H); 7.58 (m, 2H). ¹³C NMR of **5e** (75MHz, CDCl₃): δ 14.2; 22.7 (2C); 26.7; 29.3; 29.6; 31.3; 31.9; 59.3; 65.7; 72.8; 128.6; 129.3 (2C); 130.5 (2C); 137.5; 175.4. **5e'**: white solid; mp: 54-56°C; Rf (SiO₂; CH₂Cl₂) = 0.44. ¹H NMR of **5e'** (300MHz, CDCl₃): δ 0.90 (t, *J* = 7.0Hz, 3H); 1.32 (m, 10H); 1.57-1.65 (m, 1H); 2.00 (m, 1H); 2.15 (m, 1H); 2.74-2.85 (m, 1H); 3.61 (s, 1H, OH); 3.83 (m, 1H); 4.29 (m, 2H); 7.34-7.38 (m, 3H); 7.53 (m, 2H). ¹³C NMR of **5e'** (75MHz, CDCl₃): δ 14.2; 22.8; 26.4; 29.0; 29.4; 29.6; 30.2; 32.0; 58.2; 65.7; 70.9; 129.2 (3C); 130.3 (2C); 137.3; 175.8. HRMS (ESI+): Calculated for C₁₈H₂₆O₃S Na⁺, 345.1500; founded MNa⁺, 345.1500.

I.3. General procedure B for oxydation-elimination reaction of lactones **5**

To a stirred solution of lactones **5** in dichloromethane at 0 °C, was slowly added a solution of *m*-chloroperbenzoic acid (1.5 eq) in dichloromethane during 5 min. The solution was kept for 20 mn at 0 °C. The precipitate was filtered off and dichloromethane was added. The resulting organic phase was washed successively with aqueous 10 % sodium bisulfite, saturated NaHCO₃ solution, water, dried over anhydrous sodium sulphate and the solvent removed under reduced pressure to afford crude sulfoxides. This material was dissolved in toluene and was heated at reflux during 90 min. The solvent was removed under reduced pressure and the crude product was purified by column chromatography through silica gel.

I.3.1. 3-butylfuran-2(5H)-one (**6a**)¹

According to general procedure B, lactones **5a** (1.2g, 4.7 mmol) and *m*-CPBA (1g, 6.3 mmol) gave furanone **6a** (530 mg, 81 %) after purification by flash chromatography on silica gel. Eluent: pentane-ether (60/40). Aspect: yellow oil; Rf (SiO₂; P/Et₂O: 60/40) = 0.15. ¹H NMR

(300MHz, CDCl₃): δ 0.93 (t, J = 7.3 Hz, 3H); 1.30-1.80 (m, 4H); 2.29 (m, 2H); 4.83 (dd, J = 1.8; 2 Hz); 7.09 (ps, 1H). ¹³C NMR (75MHz, CDCl₃): δ 13.8; 22.3; 25.0; 29.5; 70.2; 134.4; 144.2; 174.6. HRMS (IC⁺): Calculated for C₈H₁₂O₂ Na⁺ = 163.0730; founded MNa⁺ = 163.0730.

I.3.2. 3-hexylfuran-2(5H)-one (6b)

According to general procedure B, lactone **5d** (900 mg, 3.2 mmol) and *m*-CPBA (615 mg, 3.6 mmol) gave furanone **6d** (468 mg, 86 %) after purification by flash chromatography on silica gel. Eluent: pentane-ether (50/50). Aspect: yellow oil; Rf (SiO₂; P/Et₂O: 50/50) = 0.56. ¹H NMR (300MHz, CDCl₃): δ 0.87 (t, J = 7.5 Hz, 3H); 1.29 (m, 6H); 1.54 (m, 2H); 2.27 (m, 2H); 4.75 (d, J = 2 Hz, 2H); 7.09 (ps, 1H). ¹³C NMR (75MHz, CDCl₃): δ 14.1; 22.6; 25.4; 27.4; 28.9; 31.6; 70.2; 134.6; 144.1; 174.6. HRMS (IC⁺): Calculated for C₁₀H₁₆O₂ Na⁺ = 191.1043; founded MNa⁺ = 191.1043.

I.3.3. 3-(1-hydroxybutyl)furan-2(5H)-one (6c)

According to general procedure B, lactones **5c** (3g, 4,2 mmol) and *m*-CPBA (943 mg, 5.46 mmol) gave furanone **6c** (1.35 g, 78 %) after purification by flash chromatography on silica gel. Eluent: dichloromethane-ether (85/15). Aspect: yellow oil; Rf (SiO₂; CH₂Cl₂/Et₂O: 85/15) = 0.15. ¹H NMR (300MHz, CDCl₃): δ 0.96 (t, J = 7.3 Hz, 3H); 1.30-1.80 (m, 4H); 2.50 (s, 1H, OH); 4.53 (m, 1H); 4.83 (t, J = 1.7Hz, 2H); 7.30 (ps, 1H). ¹³C NMR (75MHz, CDCl₃): δ 13.8; 18.5; 37.5; 66.7; 70.6; 136.7; 145.4; 173.5. HRMS (IC⁺): Calculated for C₈H₁₂O₃ Na⁺ = 179.0679; founded MNa⁺ = 179.0680.

I.3.4. 3-(1-hydroxyhexyl)furan-2(5H)-one (6d)

According to general procedure B, lactones **5d** (283 mg, 96 mmol) and *m*-CPBA (166 mg, 96 mmol) gave furanone **6d** (148 mg, 84 %) after purification by flash chromatography on silica. Eluent: dichloromethane-ether (85/15). Aspect: uncolored oil; Rf (SiO₂; CH₂Cl₂/Et₂O: 85/15) = 0.28. ¹H NMR (300MHz, CDCl₃) : δ 0.89 (pt, 3H); 1.31-1.48 (m, 6H); 1.70-1.78 (m, 2H); 2.50 (s(OH), 1H, OH); 4.21 (m, 1H); 4.84 (t, J = 1.7Hz, 2H); 7.30 (td, J = 3.2Hz; 1.7Hz, 1H). ¹³C NMR (75MHz, CDCl₃) : δ 14.1; 22.6; 25.0; 31.6; 35.5; 67.2; 70.6; 136.7; 144.9; 173.4. HRMS (IC⁺): Calculated for C₁₀H₁₆O₃ H⁺, 185.1178; founded MH⁺ = 185.1180.

I.3.5. 3-(1-hydroxyoctyl)furan-2(5H)-one (6e)

According to general procedure B, lactones **5e**, **5e'** (1.1 g, 3.4 mmol) and *m*-CPBA (1 g, 5.1 mmol) gave furanone **6e** (516 mg, 71 %) after purification by flash chromatography on silica gel using dichloromethane-ether (85/15). Uncolored oil R_f (SiO₂; CH₂Cl₂-Et₂O: 85/15)= 0.33. ¹H NMR (300MHz, CDCl₃): δ 0.84 (t, *J* = 6.6Hz, 3H); 1.24-1.41 (m, 10H); 1.66-1.74 (m, 2H); 2.84 (s(OH), 1H); 4.47 (m, 1H); 4.80 (t, *J* = 1.5Hz, 2H); 7.31 (td, *J* = 3.0; 1.5Hz, 1H). ¹³C NMR (75MHz, CDCl₃): δ 14.3; 22.9; 25.5; 29.5; 29.6; 32.0; 35.7; 67.3; 70.8; 136.9; 145.2; 173.6. HRMS (ESI+): Calculated for C₁₂H₂₀O₃Na⁺ = 235.1310; founded MNa⁺ = 235.1311.

I.4. General procedure C for methylenation reaction of furanone 6

Diisopropylamine (2 eq.) was added to anhydrous THF under nitrogen. After cooling the solution to 0 °C, *n*-butyllithium (1.5 eq. of a 1.6 M solution in hexane) was added and the reaction mixture was stirred for 15 min. The solution was then cooled to -78 °C and subsequently treated with furanones **6** (1eq.), dissolved in dry THF. Stirring was continued for one hour followed by addition of Eshenmoser's salt (7 eq.). The reaction mixture was stirred for one hour then allowed to warm to room temperature and stirred for 2h. The reaction mixture was poured into saturated ammonium chloride solution and extracted with ethyl acetate. The organic phase extracted was washed with water and brine and dried over anhydrous sodium sulphate, filtered and the solvent removed under reduced pressure. The crude product was purified by column chromatography through silica gel. Elimination of the amine occurs spontaneously on the silica gel.

I.4.1. 3-butyl-5-methylenefuran-2(5H)-one (8a)

According to general procedure C, furanone **6a** (1.2 g, 8.5 mmol) and Eshenmoser's salt (6.46 g, 34.9 mmol) gave methylene-furanone **8a** (435 mg, 34 %) after purification by flash chromatography on silica gel. Eluent: dichloromethane-ether (90:10). Aspect: yellow oil; R_f (SiO₂; CH₂Cl₂/Et₂O: 90/10) = 0.63. ¹H NMR (300MHz, CDCl₃): δ 0.93 (t, *J* = 7.8 Hz, 3H); 1.31-1.40 (m, 2H); 1.42-1.70 (m, 2H); 2.37 (m, 2H); 4.76 (d, *J* = 2 Hz, 1H); 5.10 (d, *J* = 2 Hz, 1H); 7.02 (s, 1H). ¹³C NMR (75MHz, CDCl₃): δ 13.7; 22.3; 24.9; 29.5; 95.5; 136.4; 136.5; 154.0; 170.6. HRMS (IC+): Calculated for C₉H₁₂O₂ H⁺ = 153.0916; founded MH⁺ = 153.0916.

I.4.2. 3-hexyl-5-methylenefuran-2(5H)-one (8b)²

According to general procedure C, furanone **6b** (1.1 g, 6.5 mmol) and Eshenmoser's salt (8.4 g, 45.7 mmol) gave methylene-furanone **8b** (680 mg, 58 %) after purification by flash chromatography on silica gel. Eluent: dichloromethane-pentane (90:10). Aspect: yellow oil; Rf (SiO₂; CH₂Cl₂/P: 90/10) = 0.8. ¹H NMR (300MHz, CDCl₃): δ 0.89 (t, *J* = 7.8 Hz, 3H); 1.20-1.45 (m, 6H); 1.50-1.60 (m, 2H); 2.37 (t, *J* = 9 Hz, 2H); 4.76 (d, *J* = 2 Hz, 1H); 5.10 (d, *J* = 2 Hz, 1H); 7.01 (s, 1H). HRMS (IC⁺): Calculated for C₁₁H₁₆O₂ Na⁺ = 203.1043; founded MNa⁺ = 203.1041.

I.4.3. 3-(1-hydroxybutyl)-5-methylenefuran-2(5H)-one (8c)²

According to general procedure C, furanone **6c** (300 mg, 1.92 mmol) and Eshenmoser's salt (2.48 g, 13.44 mmol) gave methylene-furanone **8c** (174 mg, 54 %) after purification by flash chromatography on silica gel. Eluent: dichloromethane-ether (90:10). Aspect: yellow oil; Rf (SiO₂; CH₂Cl₂/Et₂O: 90/10) = 0.38. ¹H NMR (300MHz, CDCl₃): δ 0.96 (t, *J* = 7.3 Hz, 3H); 1.38-1.80 (m, 4H); 2.33 (s, 1H, OH); 4.60 (m, 1H); 4.89 (d, *J* = 2.5Hz, 1H); 5.20 (d, *J* = 2.5Hz, 1H); 7.20 (d, *J* = 1.2Hz, 1H). ¹³C NMR (75MHz, CDCl₃): δ 14.7; 18.4; 37.7; 66.4; 97.6; 136.7; 138.6; 153.6; 169.4. HRMS (IC⁺): Calculated for C₉H₁₂O₃ H⁺ = 169.0865; founded MH⁺ = 169.0864.

I.4.4. 3-(1-hydroxyhexyl)-5-methylenefuran-2(5H)-one (8d)³

According to general procedure C, furanone **6d** (400 mg, 2.17 mmol) and Eshenmoser's salt (2.81 mg, 15.2 mmol) gave methylene-furanone **8d** (268 mg, 63 %) after purification by flash chromatography on silica gel. Eluent: pentane- ethyl acetate (60:40). Aspect: yellow oil; Rf (SiO₂; P/AcOEt: 60/40) = 0.51. ¹H NMR (300MHz, CDCl₃): δ 0.89 (pt, 3H); 1.30-1.42 (m, 6H); 1.69-1.77 (m, 2H); 2.30 (s, 1H, OH); 4.59 (m, 1H); 4.89 (d, *J* = 2.5Hz, 1H); 5.20 (d, *J* = 2.6Hz, 1H); 7.20 (m, 1H). ¹³C NMR (75MHz, CDCl₃): δ 14.3; 22.7; 25.2; 31.8; 36.0; 67.3; 97.9; 136.6; 138.6; 154.0; 169.5.

I.4.5. 3-(1-hydroxyoctyl)-5-methylenefuran-2(5H)-one (8e)²

According to general procedure C, furanone **6e** (475 mg, 2.56 mmol) and Eshenmoser's salt (2.9 g, 15.7 mmol) gave methylene-furanone **8e** (208 mg, 42 %) after purification by flash chromatography on silica gel. Eluent: pentane- ethyl acetate (60:40). Aspect: yellow oil; Rf (SiO₂; P/AcOEt: 60/40) = 0.74. ¹H NMR (300MHz, CDCl₃): δ 0.87 (pt, 3H); 1.27-1.42 (m, 10H); 1.71-1.80 (m, 2H); 2.28 (s, 1H, OH); 4.56-4.62 (m, 1H); 4.89 (d, *J* = 2.6Hz, 1H); 5.20

(dd, $J = 2.5$; 0.4Hz, 1H); 7.20 (m, 1H). ^{13}C NMR (75MHz, CDCl_3): δ 14.2; 22.7; 25.3; 29.3; 29.4; 31.9; 35.8; 67.1; 97.7; 136.4; 138.4; 153.8; 169.3. HRMS (IC+): Calculated for $\text{C}_{13}\text{H}_{20}\text{O}_3$ $\text{H}^+ = 225.1491$; founded $\text{MH}^+ = 225.1492$.

I.5. General procedure D for bromation reaction of methylene-furanone **8**⁴

A solution of Br_2 (1 eq.) in CCl_4 was added to the solution of methylene-furanone **8** (1 eq.) in CCl_4 at 0 °C. The mixture was stirred at room temperature for 30 mn and then the solvent was removed in vacuum. The obtained residue was dissolved in CHCl_3 and Et_3N (1 eq.), one drop of DBU was added at 0 °C. The mixture was further stirred at room temperature for 1 to 3 h. The solvent was removed in vacuum and the obtained crude product was purified by column chromatography through silica gel.

I.5.1. (Z)-5-(bromomethylene)-3-butylfuran-2(5H)-one (**10a**)

According to general procedure D, furanone **8a** (60 mg, 0.39mmol) and Br_2 (63 mg, 0.39 mmol) gave bromofuranone **10a** (51 mg, 56 %) after purification by flash chromatography on silica gel. Eluent: pentane-ether (80:20). Aspect: yellow oil; R_f (SiO_2 ; P/ether : 80/20) = 0.27. ^1H NMR (300MHz, CDCl_3): δ 0.93 (t, $J = 7.3\text{Hz}$, 3H); 1.30-1.40 (m, 2H); 1.53-1.80 (m, 2H); 2.34 (m, 2H); 5.93 (s, 1H); 7.01 (d, $J = 1.3\text{ Hz}$, 1H). ^{13}C NMR (75MHz, CDCl_3): δ 13.8; 22.3; 25.2; 29.5; 89.6; 135.0; 136.0; 151.5; 169.3. HRMS (IC+): Calculated for $\text{C}_9\text{H}_{11}\text{BrO}_2$ $\text{H}^+ = 231.0021$; founded $\text{MH}^+ = 231.0020$.

I.5.2. (Z)-5-(bromomethylene)-3-hexylfuran-2(5H)-one (**10b**)⁵

According to general procedure D, furanone **8b** (75 mg, 0.41 mmol) and Br_2 (66 mg, 0.41 mmol) gave bromofuranone **10a** (40 mg, 37 %) after purification by flash chromatography on silica gel. Eluent: pentane-ether (80:20). Aspect: yellow oil; R_f (SiO_2 ; P/ether : 80/20) = 0.49. ^1H NMR (300MHz, CDCl_3): δ 0.89 (t, $J = 7.3\text{Hz}$, 3H); 1.30-1.40 (m, 6H); 1.53-1.80 (m, 2H); 2.34 (m, 2H); 5.93 (s, 1H); 7.01 (d, $J = 1.3\text{ Hz}$, 1H). ^{13}C NMR (75MHz, CDCl_3): δ 14.2; 22.6; 25.5; 27.4; 28.9; 31.5; 89.6; 135.0; 136.2; 151.5; 169.3. HRMS (IC+): Calculated for $\text{C}_{11}\text{H}_{15}\text{BrO}_2$ $\text{Na}^+ = 281.0148$; founded $\text{MNa}^+ = 281.0148$.

I.5.3. (Z)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)

(E)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)

According to general procedure D, furanone **8c** (65 mg, 0.38 mmol) and Br₂ (62 mg, 0.39 mmol) gave bromofuranone **10c-Z** and **10c-E** (45 mg and 6 mg, 48 %) after purification and separation by flash chromatography on silica gel. Eluent: dichloromethane-ether (95:5). Aspect **10c-Z**: yellow oil; R_f (SiO₂; CH₂Cl₂/ether : 95/5) = 0.42. ¹H NMR **10c-Z** (300MHz, CDCl₃) : δ 0.96 (t, *J* = 7.3Hz, 3H); 1.30-1.55 (m, 2H); 1.58-1.80 (m, 2H); 2.45 (s, 1H, OH); 4.57 (m, 1H); 6.06 (s, 1H); 7.22 (d, *J* = 1.3 Hz, 1H). ¹³C NMR **10c-Z** (75MHz, CDCl₃): δ 13.9; 18.5; 37.7; 66.9; 91.8; 135.1; 137.8; 151.3; 167.9. Aspect **10c-E**: yellow oil ; R_f (SiO₂; CH₂Cl₂/ether : 95/5) = 0.6 ¹H NMR **10c-E** (300MHz, CDCl₃) : δ 0.97 (t, *J* = 7.3 Hz, 3H); 1.4-1.55 (m, 2H); 1.58-1.80 (m, 2H); 2.25 (1H, OH); 4.61 (m, 1H); 6.46 (s, 1H); 7.56 (s, 1H). HRMS (IC+): Calculated for C₉H₁₁BrO₃ H⁺ = 246.9970; founded MH⁺ = 246.9972.

I.5.4. (Z)-5-(bromomethylene)-3-(1-hydroxyhexyl)furan-2(5H)-one (10d)

According to general procedure D, furanone **8d** (130 mg, 0.66 mmol) and Br₂ (149 mg, 0.66 mmol) gave bromofuranone **10d** (56 mg, 31 %) after purification by flash chromatography on silica gel. Eluent: pentane-ethyl acetate (80:20). Aspect: yellow oil; R_f (SiO₂; P/AcOEt : 80/20)= 0.30. ¹H NMR (300MHz, CDCl₃): δ 0.87 (t, *J* = 7.3 Hz, 3H); 1.25-1.41 (m, 6H); 1.67-1.80 (m, 2H); 2.29 (s(OH), 1H); 4.56 (m, 1H); 6.06 (s, 1H); 7.21 (pd, 1H). ¹³C NMR (75MHz, CDCl₃): δ 14.1; 22.7; 24.9; 31.6; 35.7; 67.2; 91.8; 135.1; 137.8; 151.3; 167.9. HRMS (IC+): Calculated for C₁₁H₁₅BrO₃ H⁺ = 275.0283; founded MH⁺ = 275.0283.

I.5.5. (Z)-5-(bromomethylene)-3-(1-hydroxyoctyl)furan-2(5H)-one (10e)

According to general procedure D, furanone **8e** (86 mg, 0.38 mmol) and Br₂ (61 mg, 0.38 mmol) gave bromofuranone **10e** (57 mg, 49 %) after purification by flash chromatography on silica gel. Eluent: pentane-ethyl acetate (80:20). Aspect: yellow oil; R_f (SiO₂; P/AcOEt : 80/20) = 0.32. ¹H NMR (300MHz, CDCl₃): δ 0.87 (t, *J* = 7.3Hz, 3H); 1.24-1.44 (m, 10H); 1.66-1.80 (m, 2H); 2.42 (s (OH), 1H); 4.55 (m, 1H); 6.06 (s, 1H); 7.22 (pd, 1H). ¹³C NMR (75MHz, CDCl₃) : δ 14.2; 22.7; 25.3; 29.3; 29.4; 31.9; 35.7; 67.2; 91.8; 135.1; 137.8; 151.3; 167.9. HRMS (ESI+): Calculated for C₁₃H₁₉BrO₃ Na⁺ =325.0415; founded MNa⁺: 325.0415.

II. Biological evaluations

The recombinant *Escherichia coli* strain NM522 containing the sensor plasmid pSB401 was used. In pSB401, the *LuxR* and the *LuxI* promoter from *V. fischeri* have been coupled to the entire *Lux* structural operon (*LuxCDABE*) from *Photobacterium luminescens*. Bacterial cultures were grown in exponential phase in Luria broth, in the presence of tetracycline ($20 \mu\text{g mL}^{-1}$), at $30 \text{ }^\circ\text{C}$.

II.1. Agonistic activity

The inducing activity of the various acyl-HSL analogues was monitored using the *E. coli* biosensor strain. Acyl-HSL activity was measured in a microtitre plate format, with bioluminescence quantified using a Luminoskan luminometer. Concentrations of analogues, ranging from 0.1 to 200 μM , were made up to 0.1 mL volumes with growth medium. The amount of light produced by the bacteria was detected after 4–5 h. The amount of light measured was expressed in relative light units (RLU).

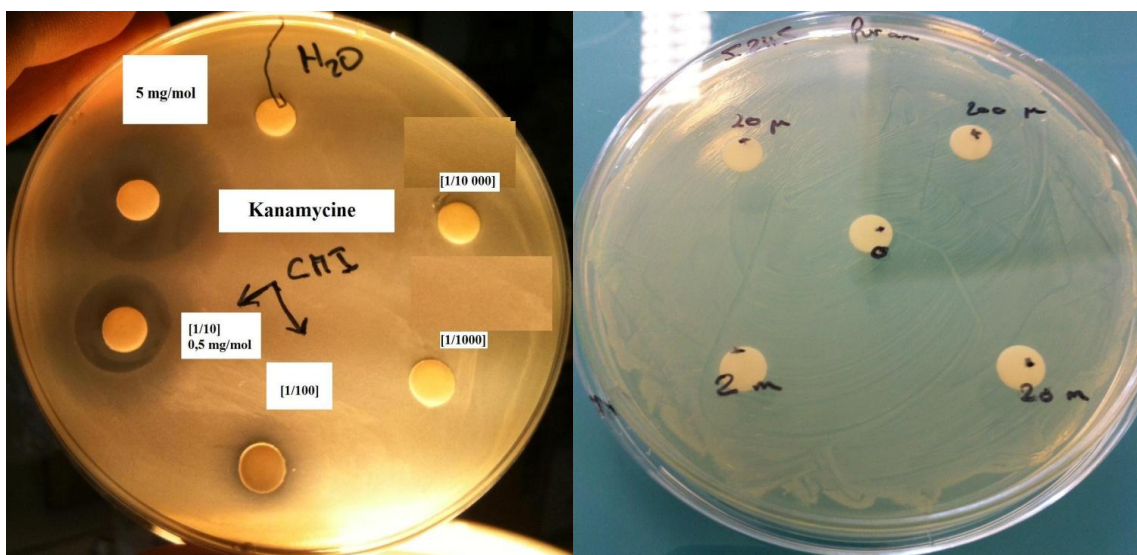
II.2. Antagonistic activity

The influence of acyl-HSL analogues on the induction of bioluminescence by 3-oxo-C₆-HSL was determined at concentrations ranging from 1 to 200 μM as described above, except that 3-oxo-C₆-HSL was included at a final concentration of 200 nM together with the analogue. 3-Oxo-C₆-HSL at 200 nM induced a 124 ± 8 RLU after 4-5 h, when the ratio of induced to background light was at its maximum.

II.3. Antibacterial activity

Furanone **10a**, the strongest inhibitor of this series of compounds, was tested using the disk diffusion assay, a common assay for antibiotic susceptibility standardized according to the Clinical and Laboratory Standards Institute guidelines.⁶ A control disk diffusion assay was conducted using the relevant kanamycin antibiotic. The compound **10a** was tested at concentrations 20 μM , 200 μM , 2 mM and 20 mM. After overnight incubation at $37 \text{ }^\circ\text{C}$, the inhibition zones of each compound were measured. No inhibition zone appears for the

furanone **10a** in contrast with the witness compound, where these zones are very clear for this antibiotic.



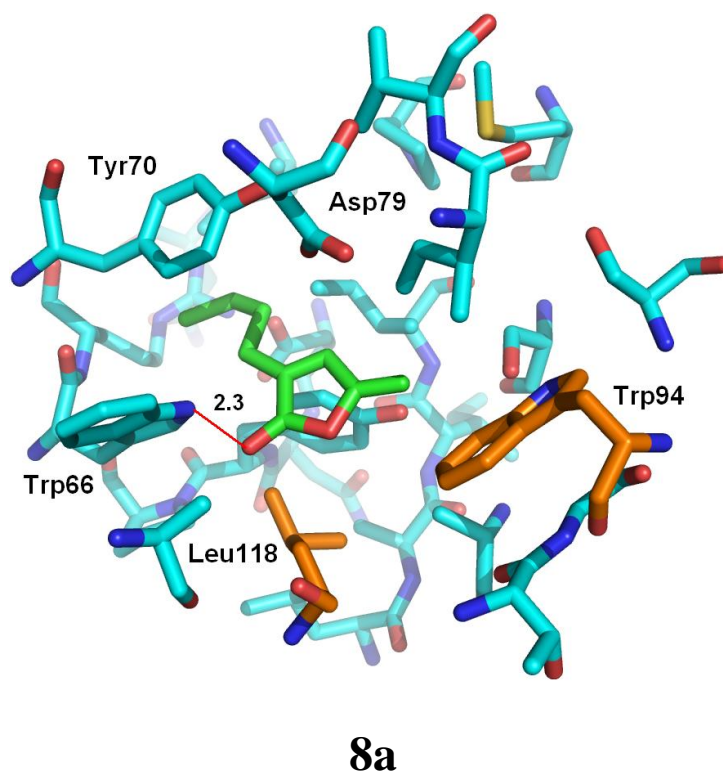
Kanamycin antibiotic

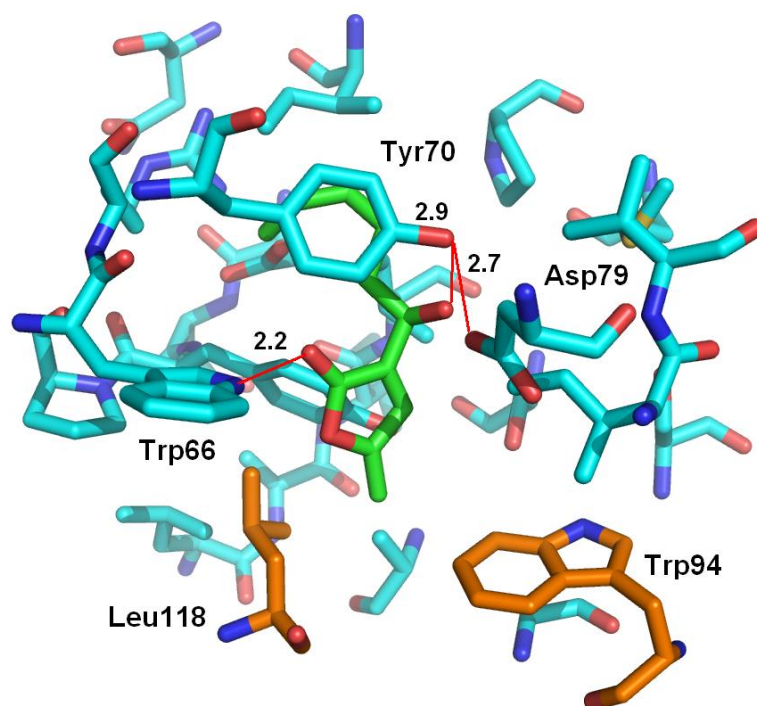
Furanone 10a

III. Molecular modelling.

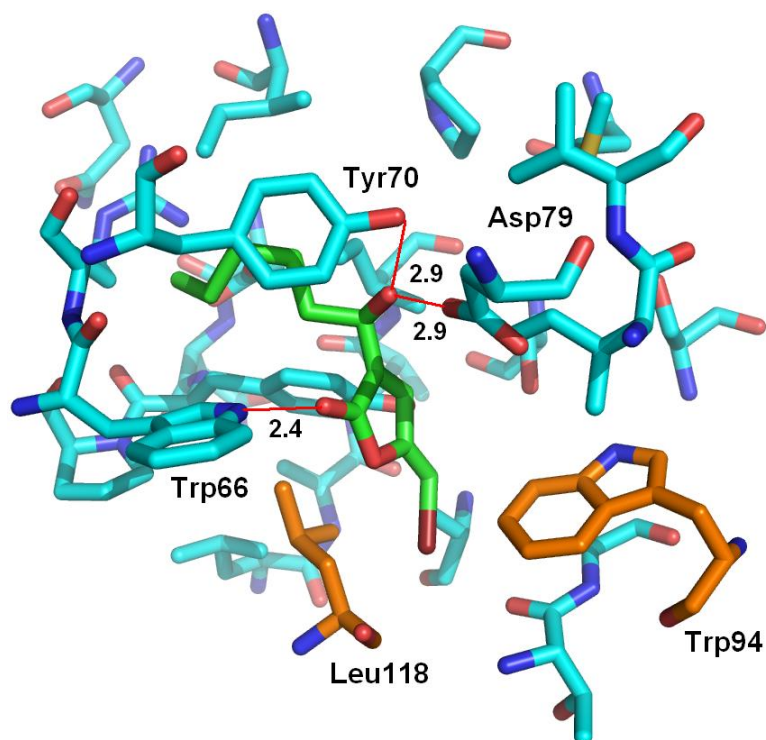
All calculations were performed with ArgusLab⁷ as software on a Dell OPTIPLEX GX620 PC. Docking experiments were performed with the docking module of ArgusLab. The protein model of LuxR⁸ was created using SWISS-MODEL⁹ with ClustalW.¹⁰

Docking studies were performed using the ligand binding site of LuxR created as previously described,⁸ subsequently to the docking of 3-oxo-C₆-HSL.¹¹ Docking experiments were performed with the following parameters: Docking box centred on 3-oxo-C₆-HSL: X = Y = Z = 15 Å, ligand option: flexible; calculation type: Dock; Docking engine: GADock (Genetic Algorithm).¹² Genetic algorithm dock engine settings: default advanced parameters; hydrogen bonds were assigned within a distance of 3 Å.





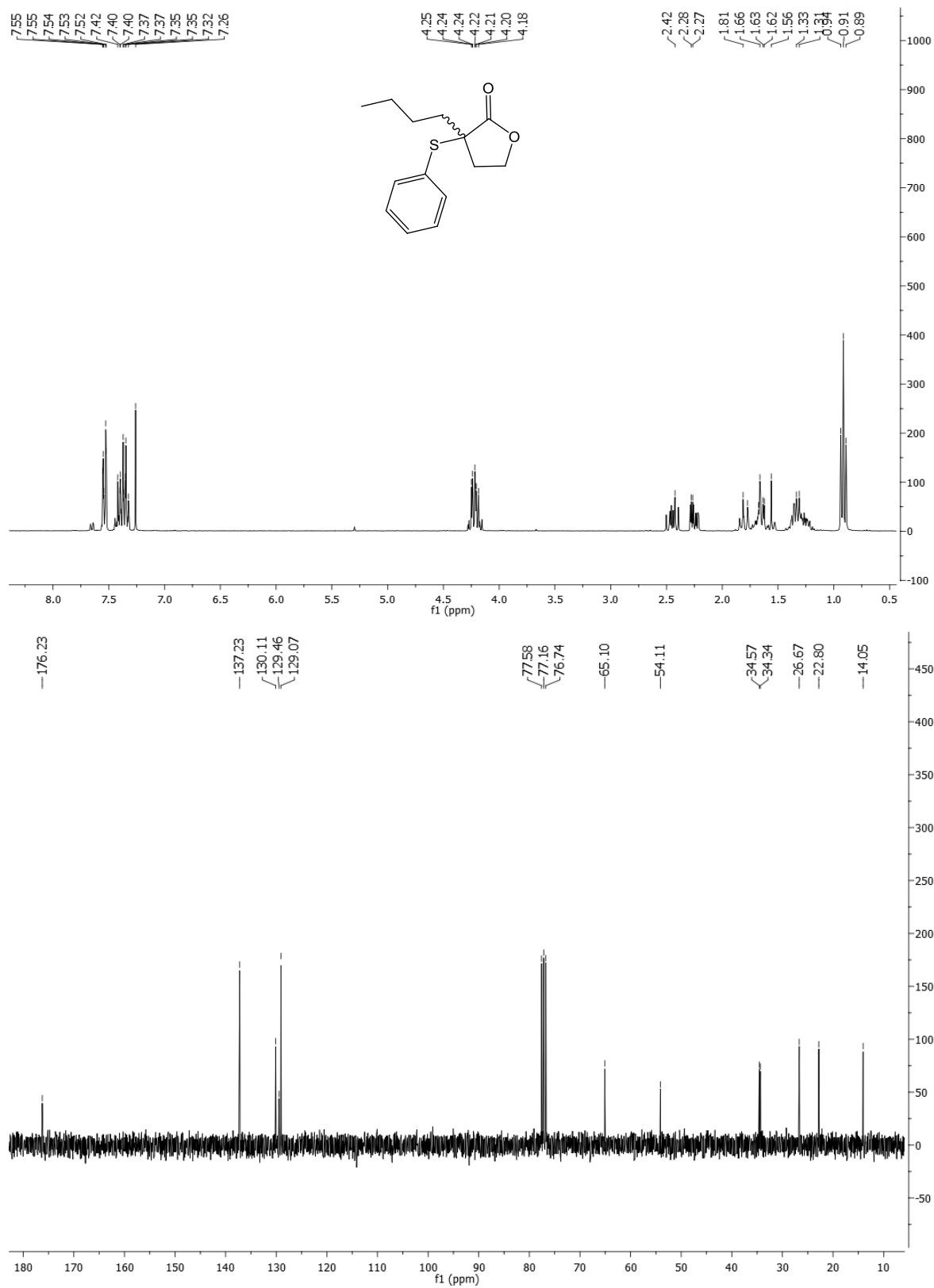
8d



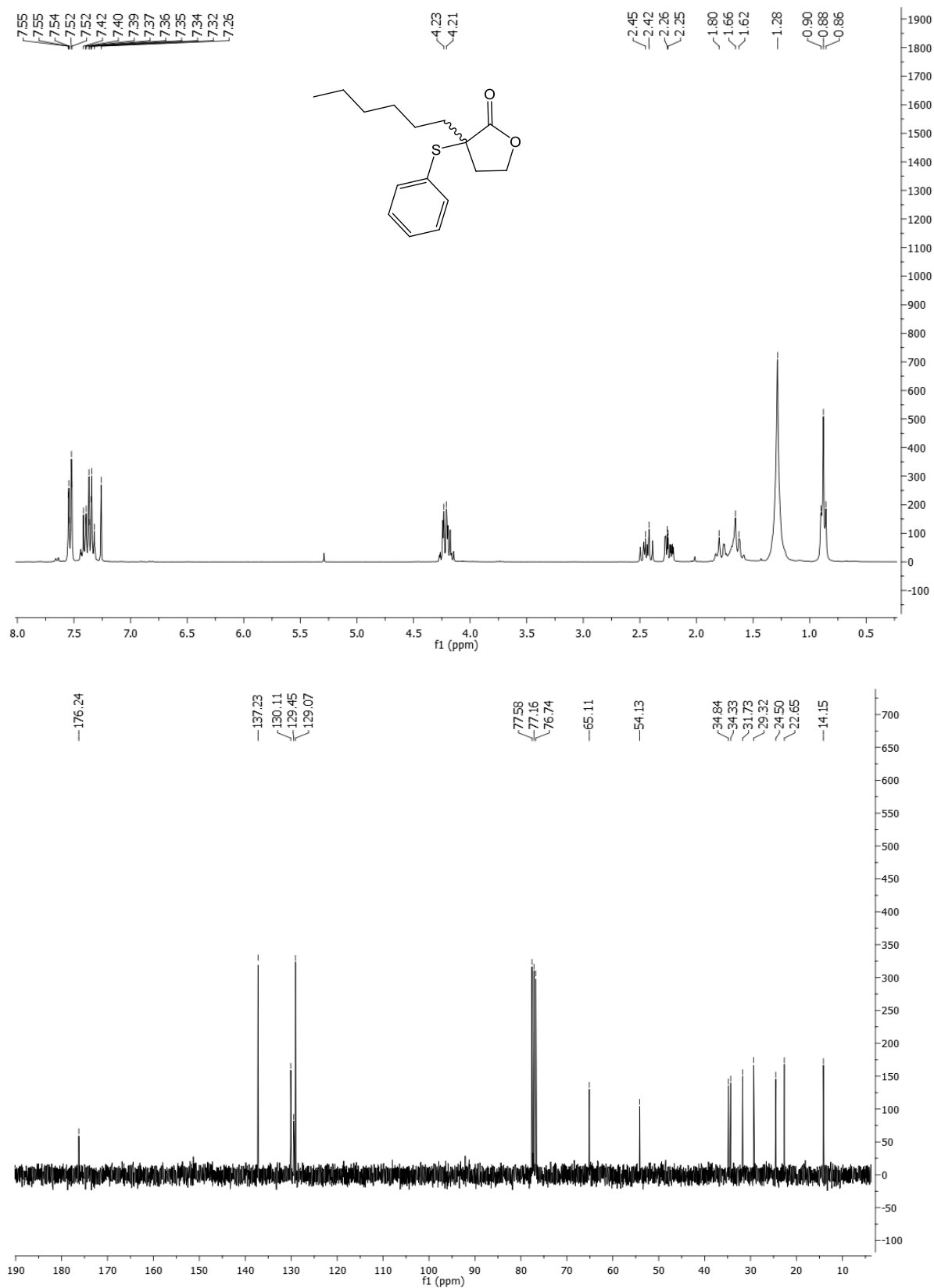
10d

IV. ¹H and ¹³C NMR spectrum

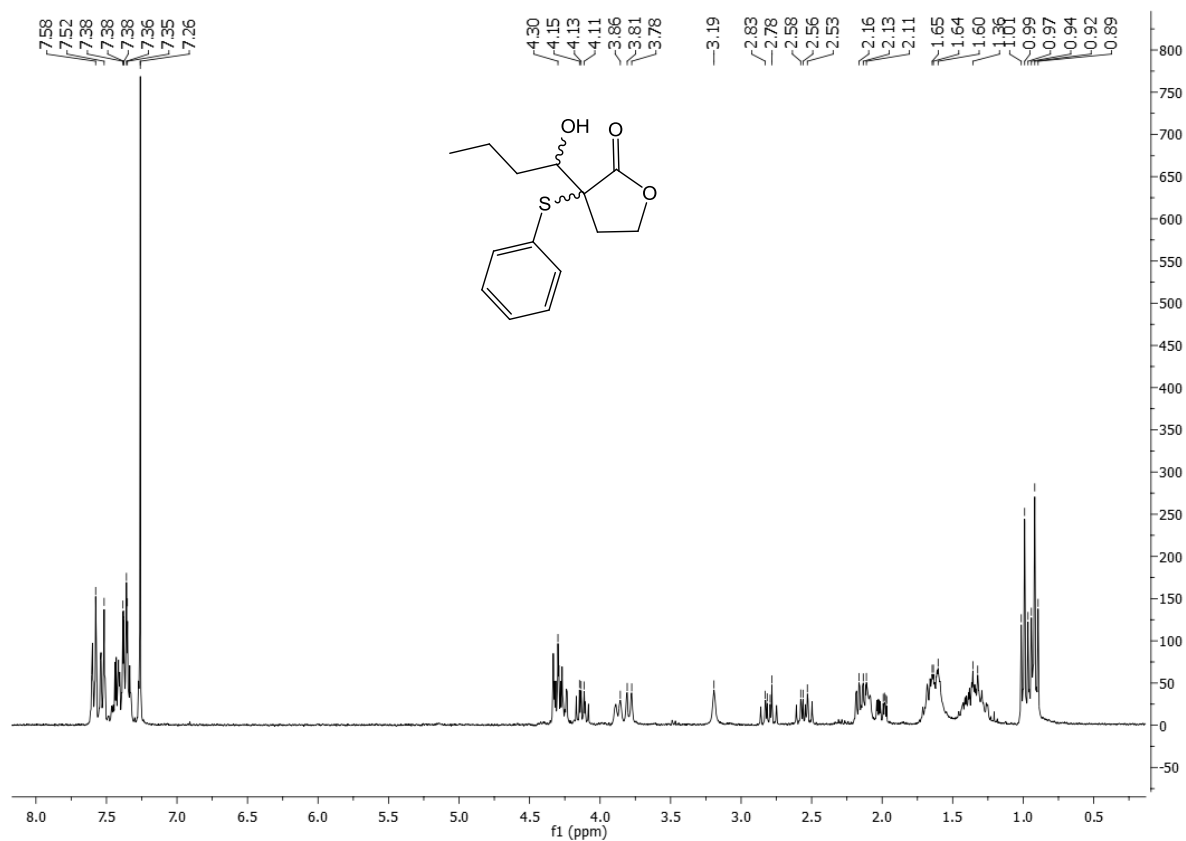
IV.1.1. 3-butyl-3-(phenylthio)dihydrofuran-2(3H)-one (5a)

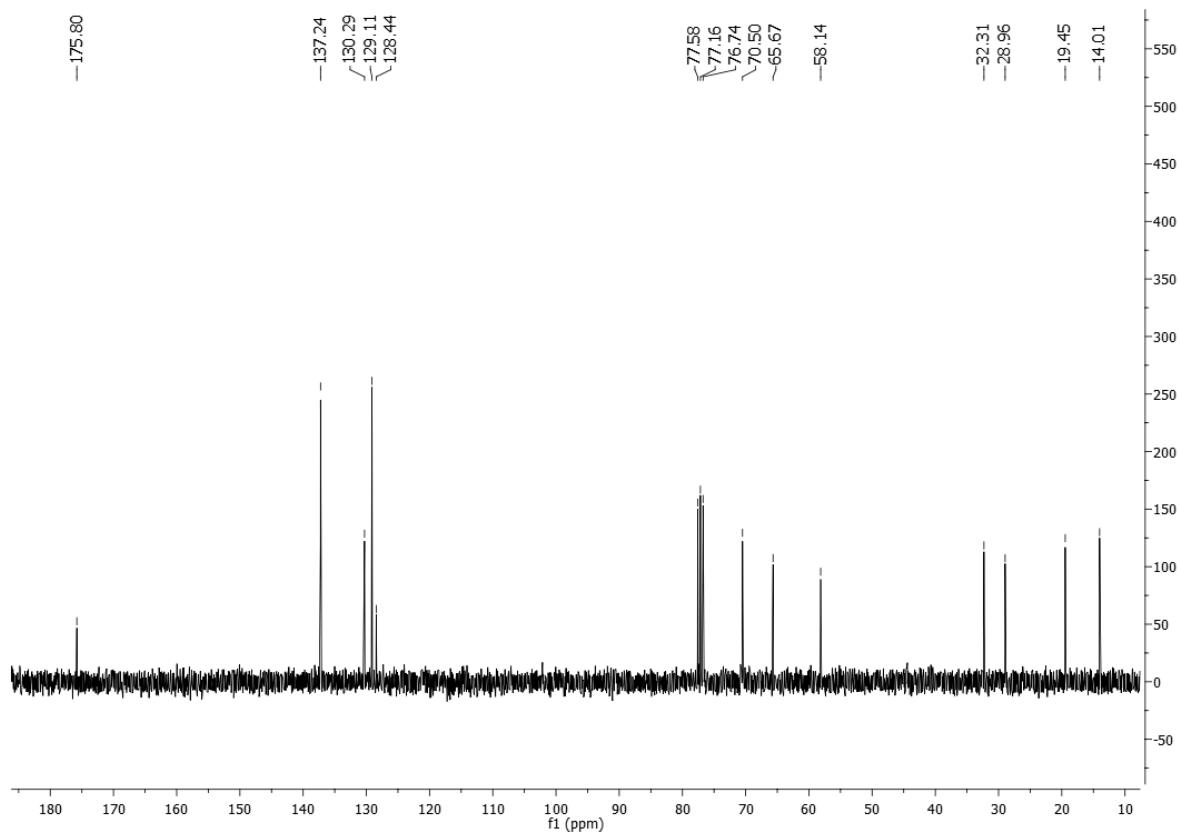


IV.1.2. 3-hexyl-3-(phenylthio)dihydrofuran-2(3H)-one (5b)

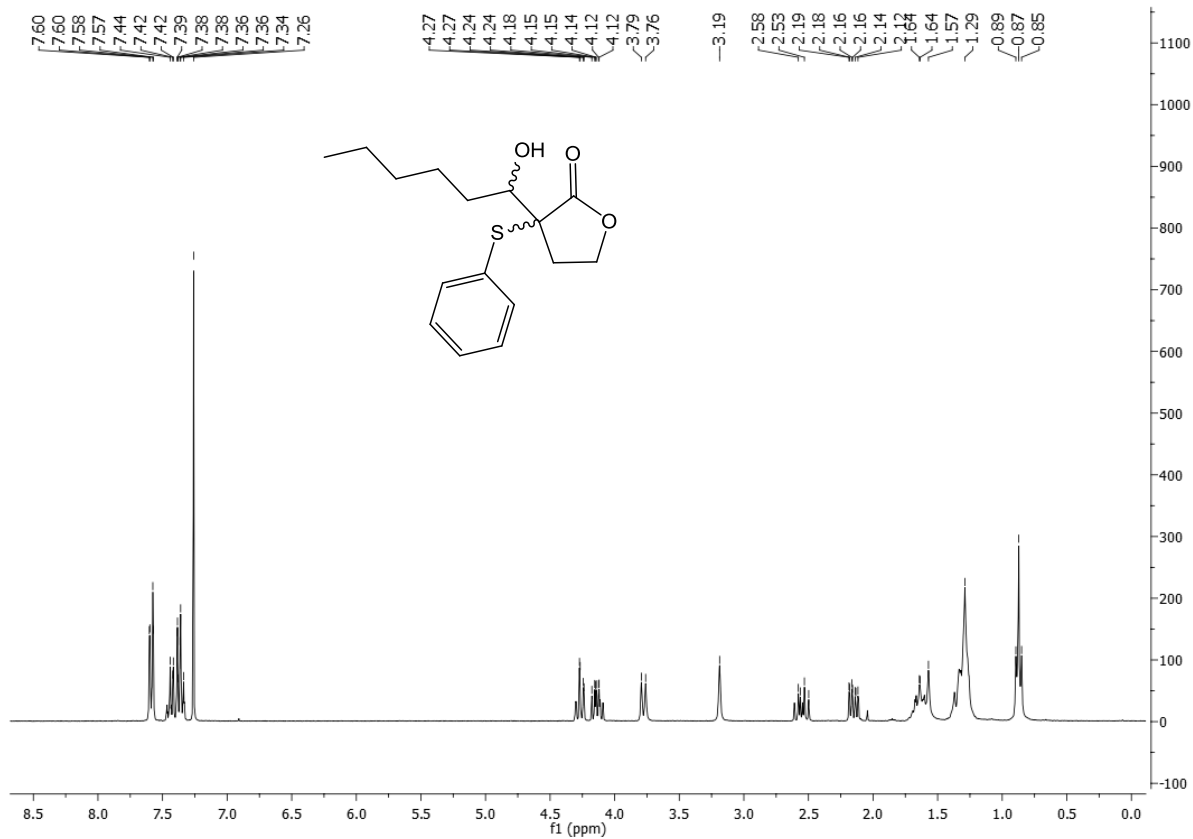


IV.1.3. 3-(1-hydroxybutyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5c)



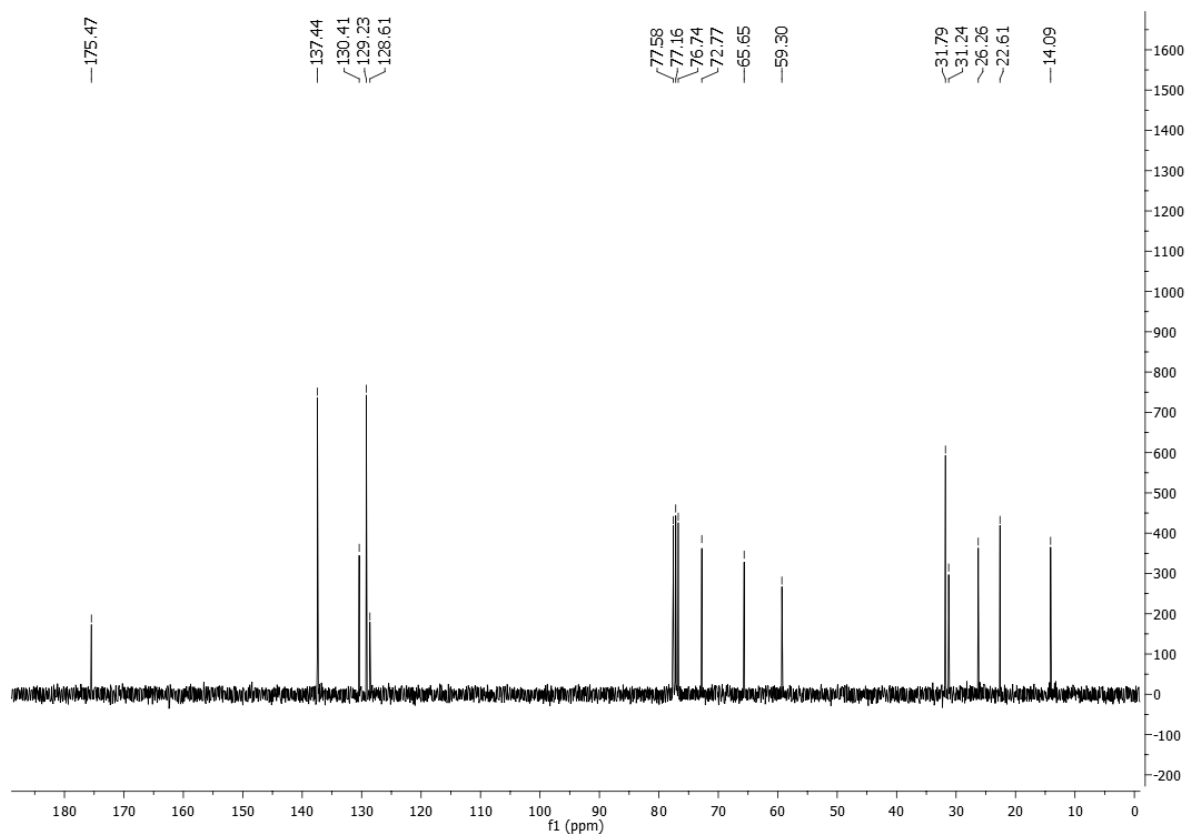


IV.1.4. 3-(1-hydroxyhexyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5d)

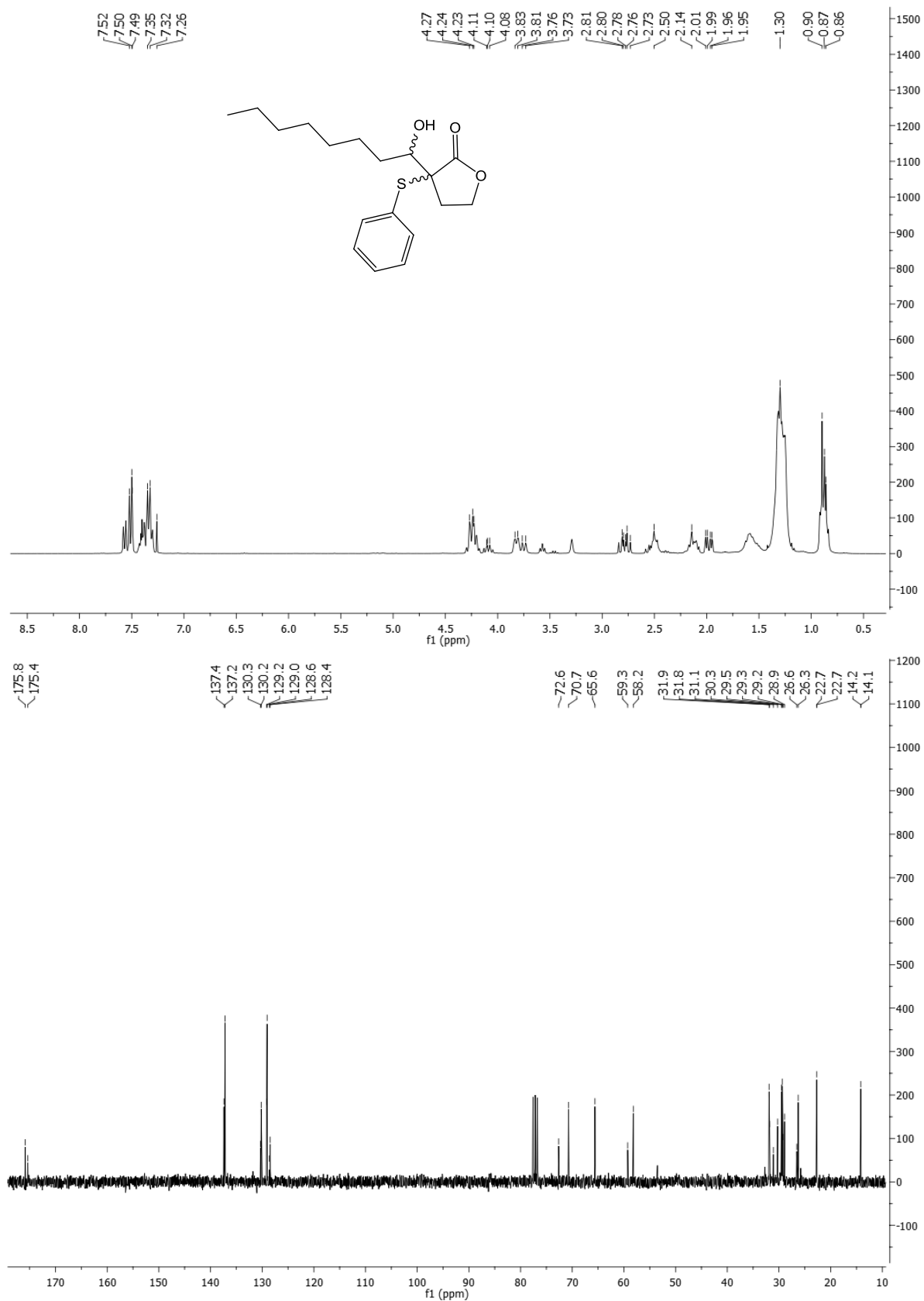


Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

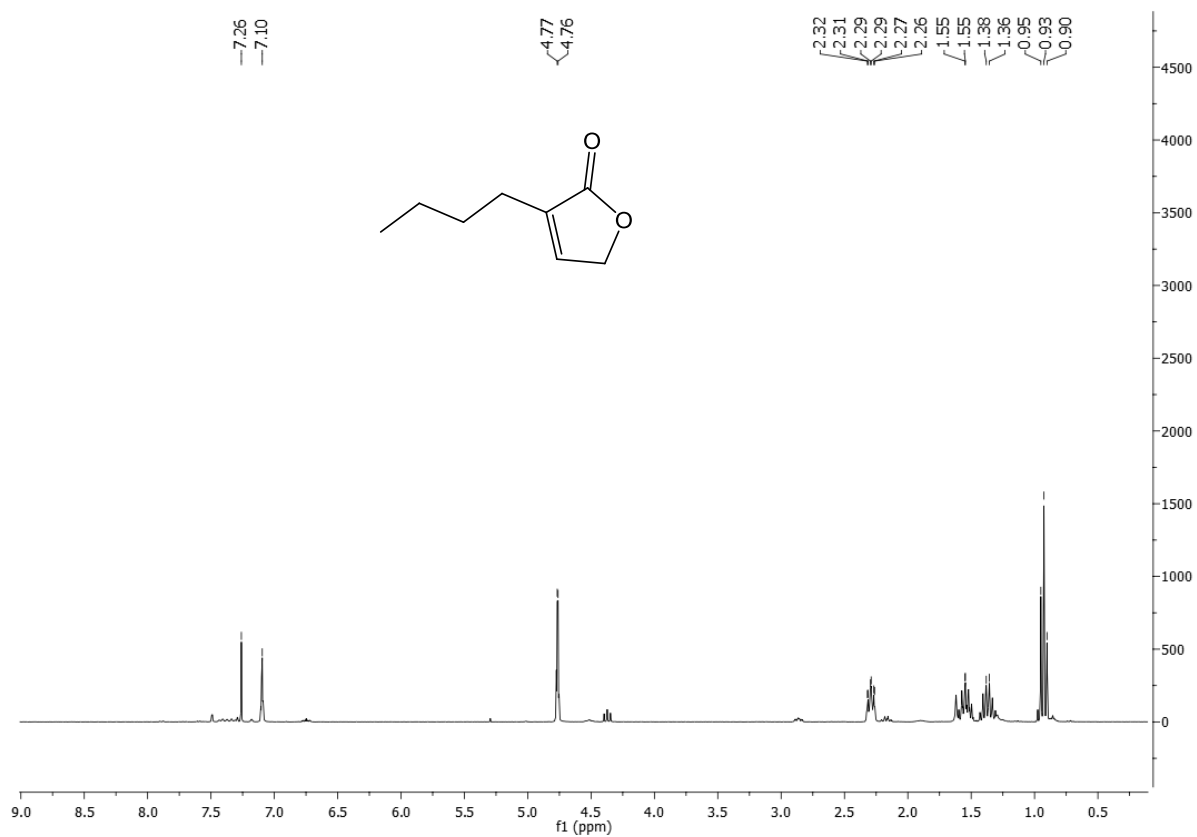
^1H and ^{13}C NMR spectrum
Sabbah et al.



IV.1.5. 3-(1-hydroxyoctyl)-3-(phenylthio)dihydrofuran-2(3H)-one (5e, 5e')

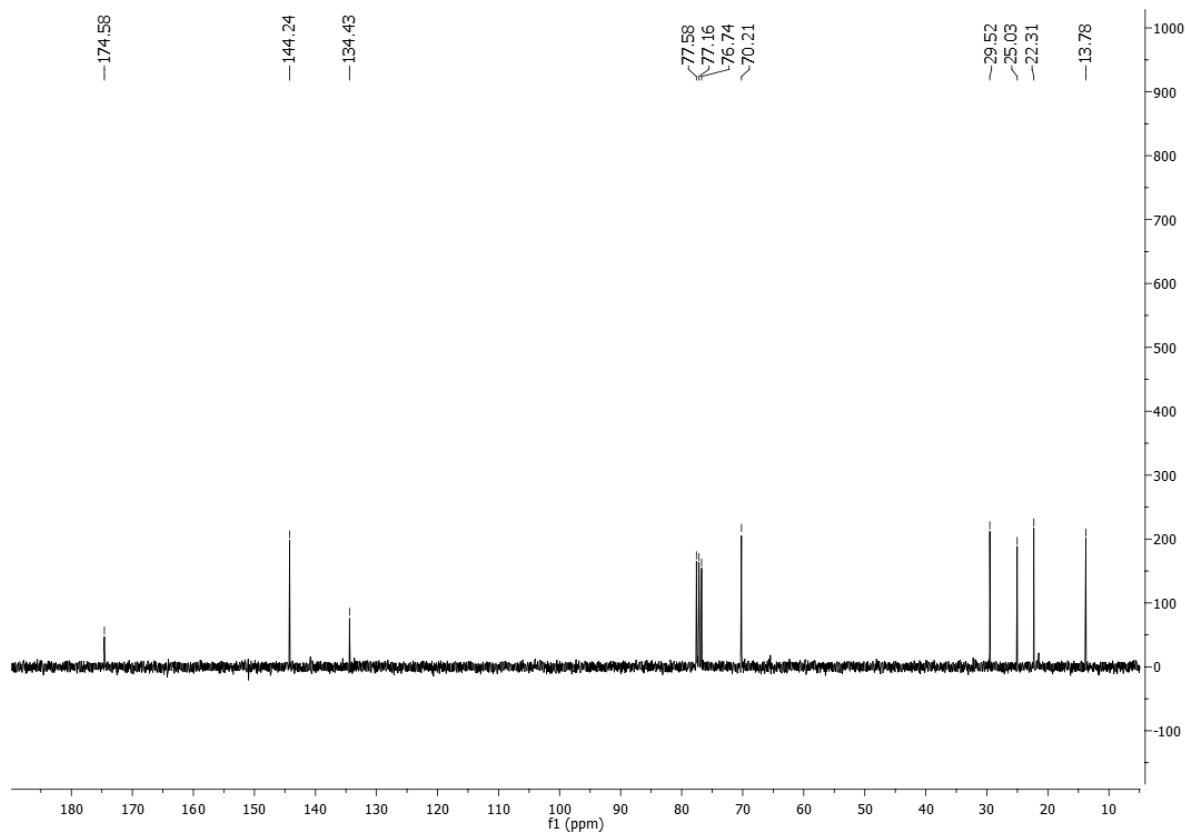


IV.1.6. 3-butylfuran-2(5H)-one (6a)



Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

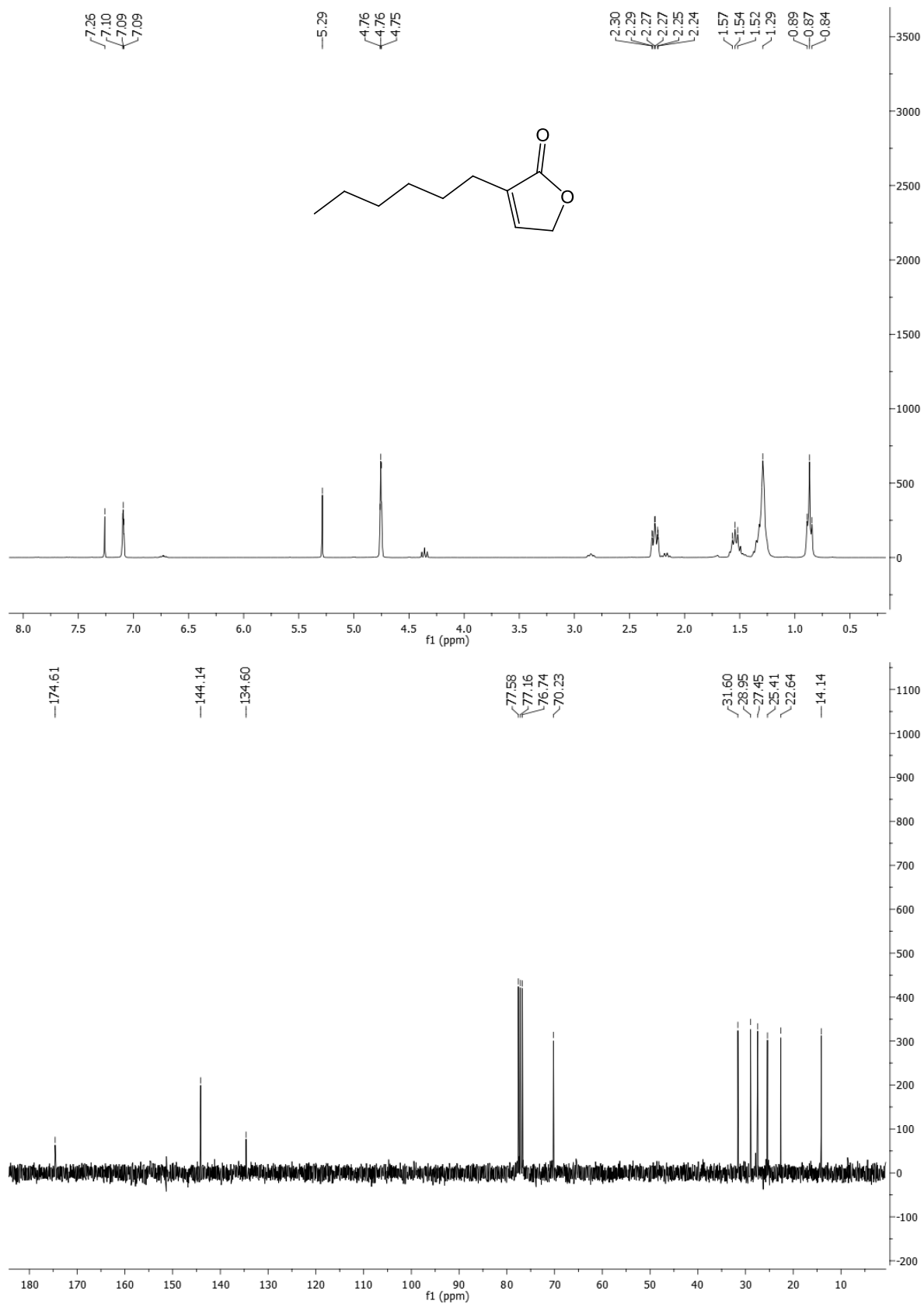
^1H and ^{13}C NMR spectrum
Sabbah et al.



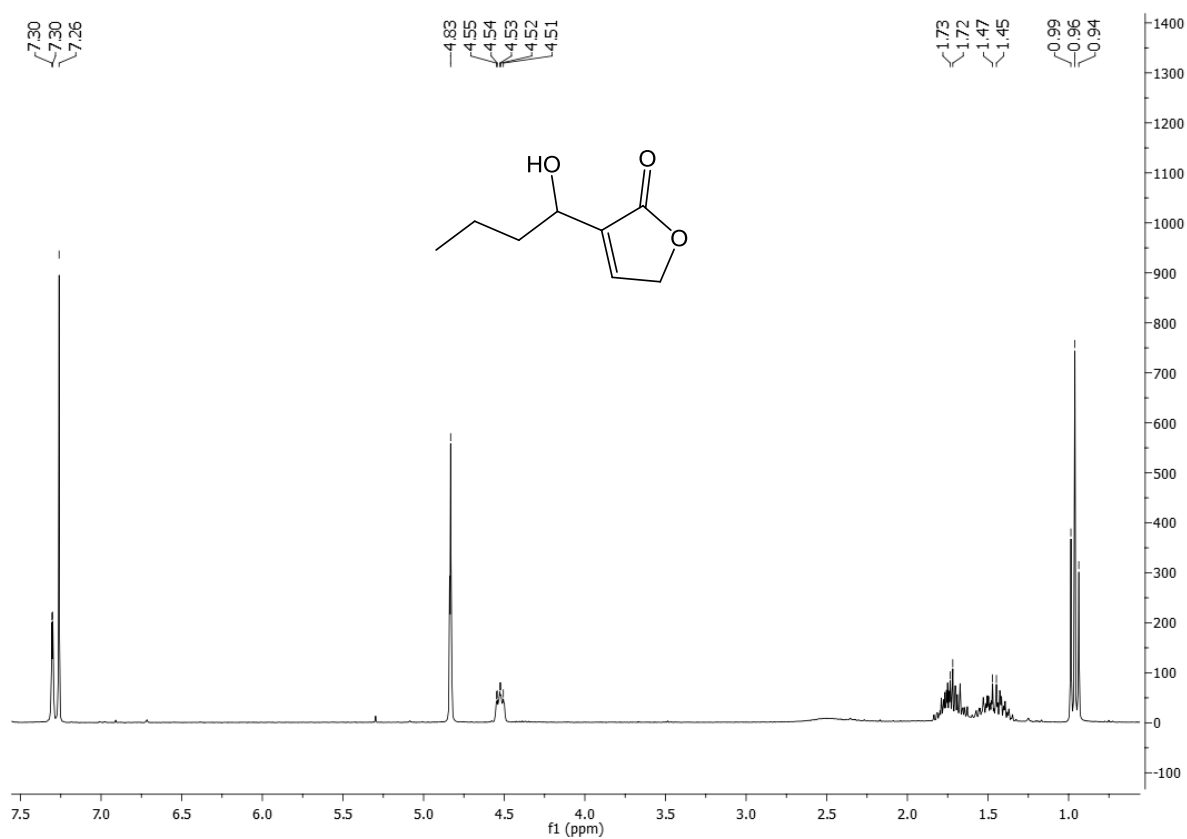
IV.1.7. 3-hexylfuran-2(5H)-one (6b)

Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

^1H and ^{13}C NMR spectrum
Sabbah et al.

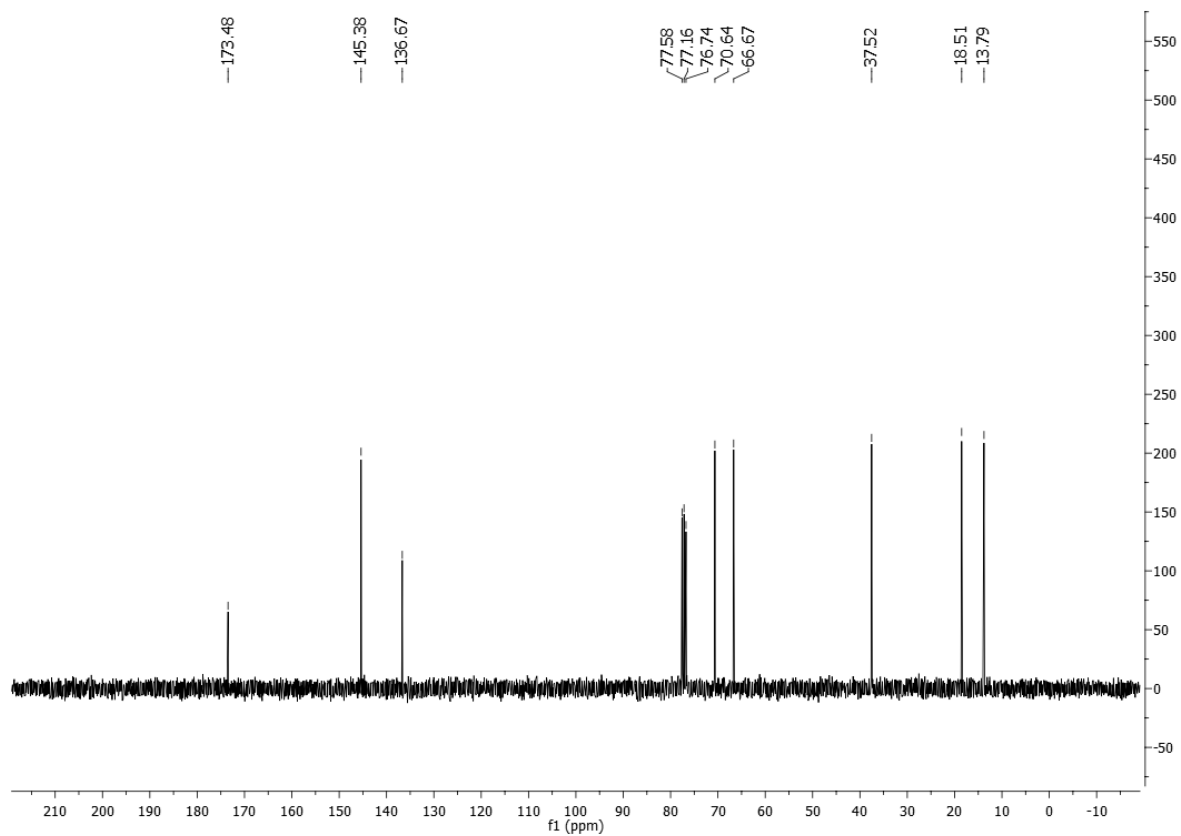


IV.1.8. 3-(1-hydroxybutyl)furan-2(5H)-one (6c)

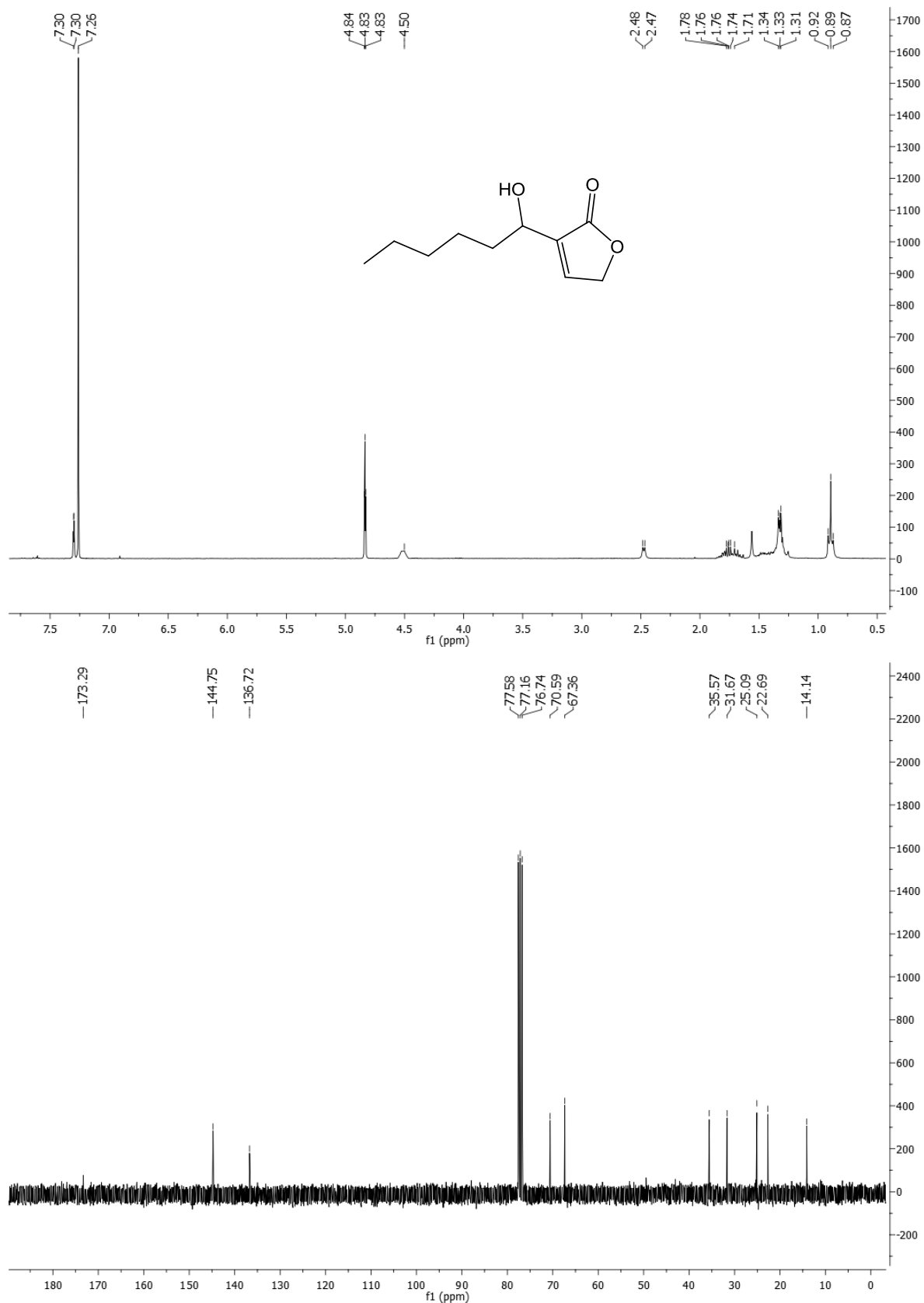


Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

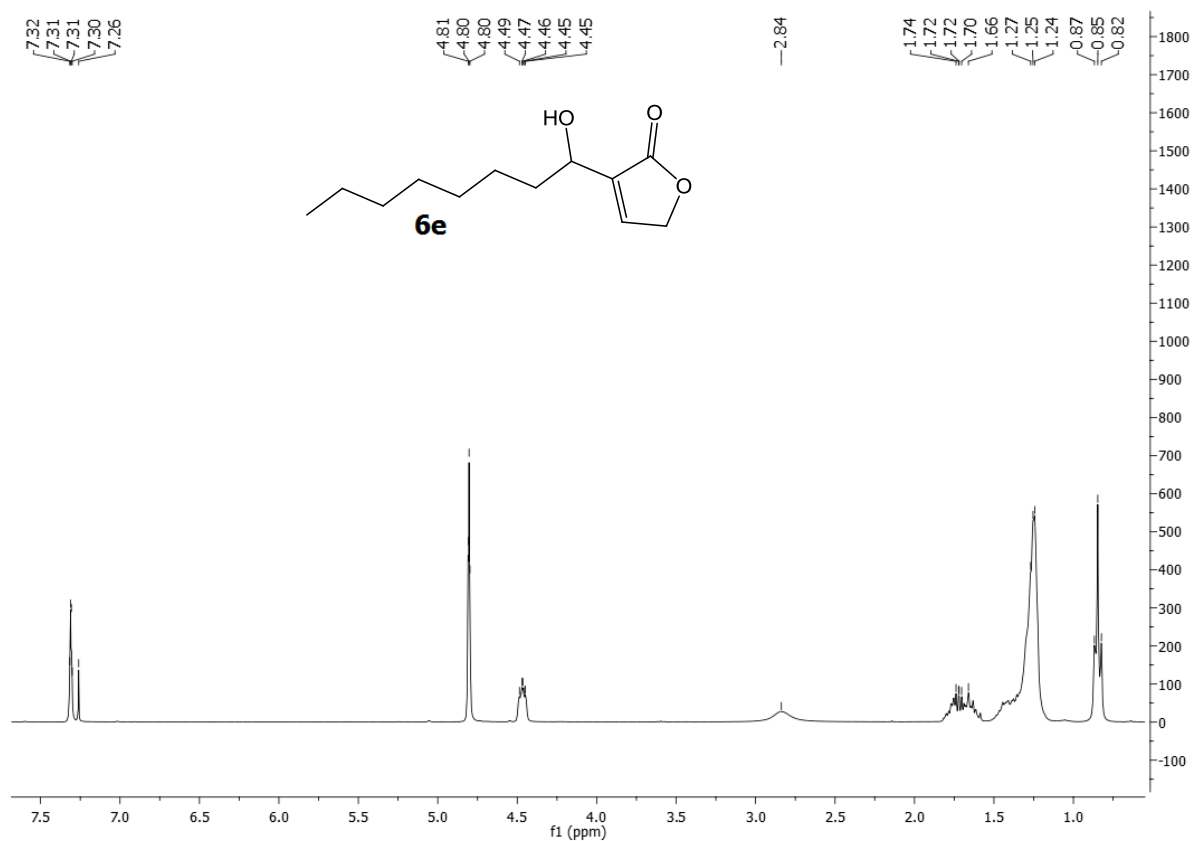
^1H and ^{13}C NMR spectrum
Sabbah et al.



IV.1.9. 3-(1-hydroxyhexyl)furan-2(5H)-one (6d)

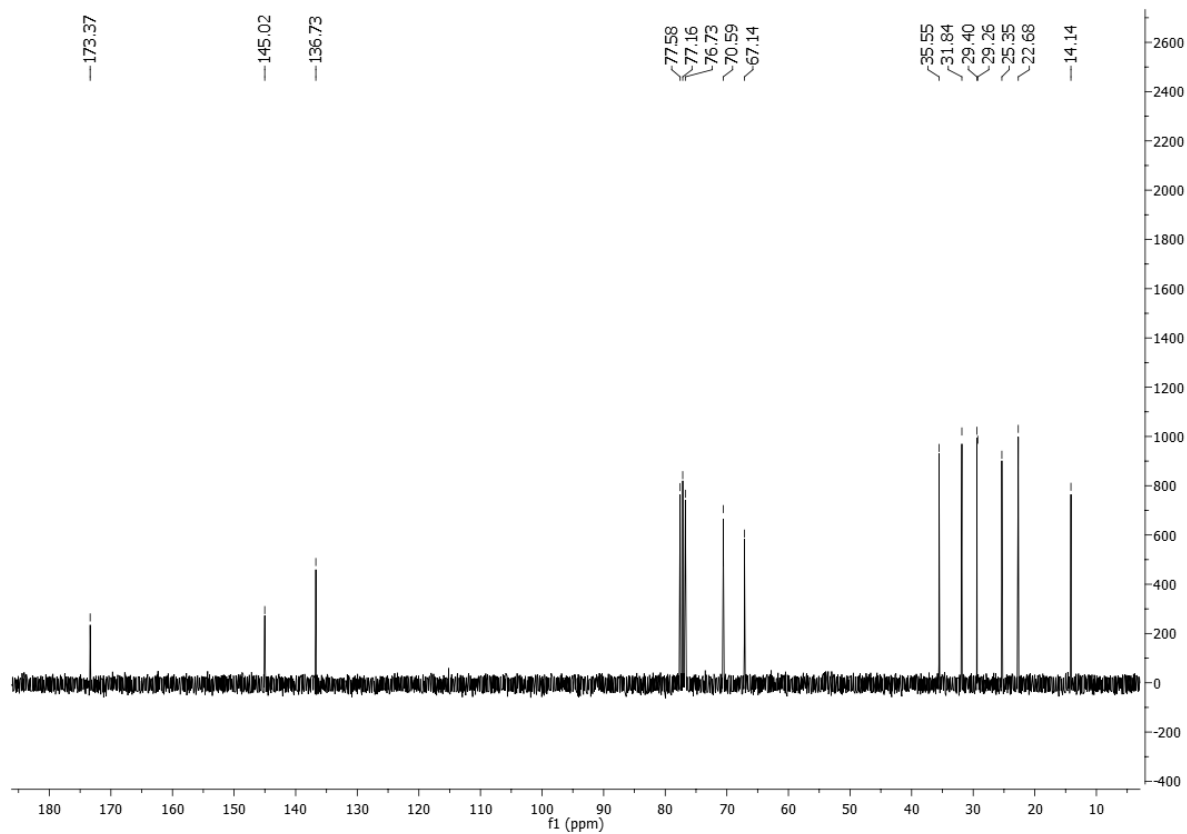


IV.1.10. 3-(1-hydroxyoctyl)furan-2(5H)-one (6e)



Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

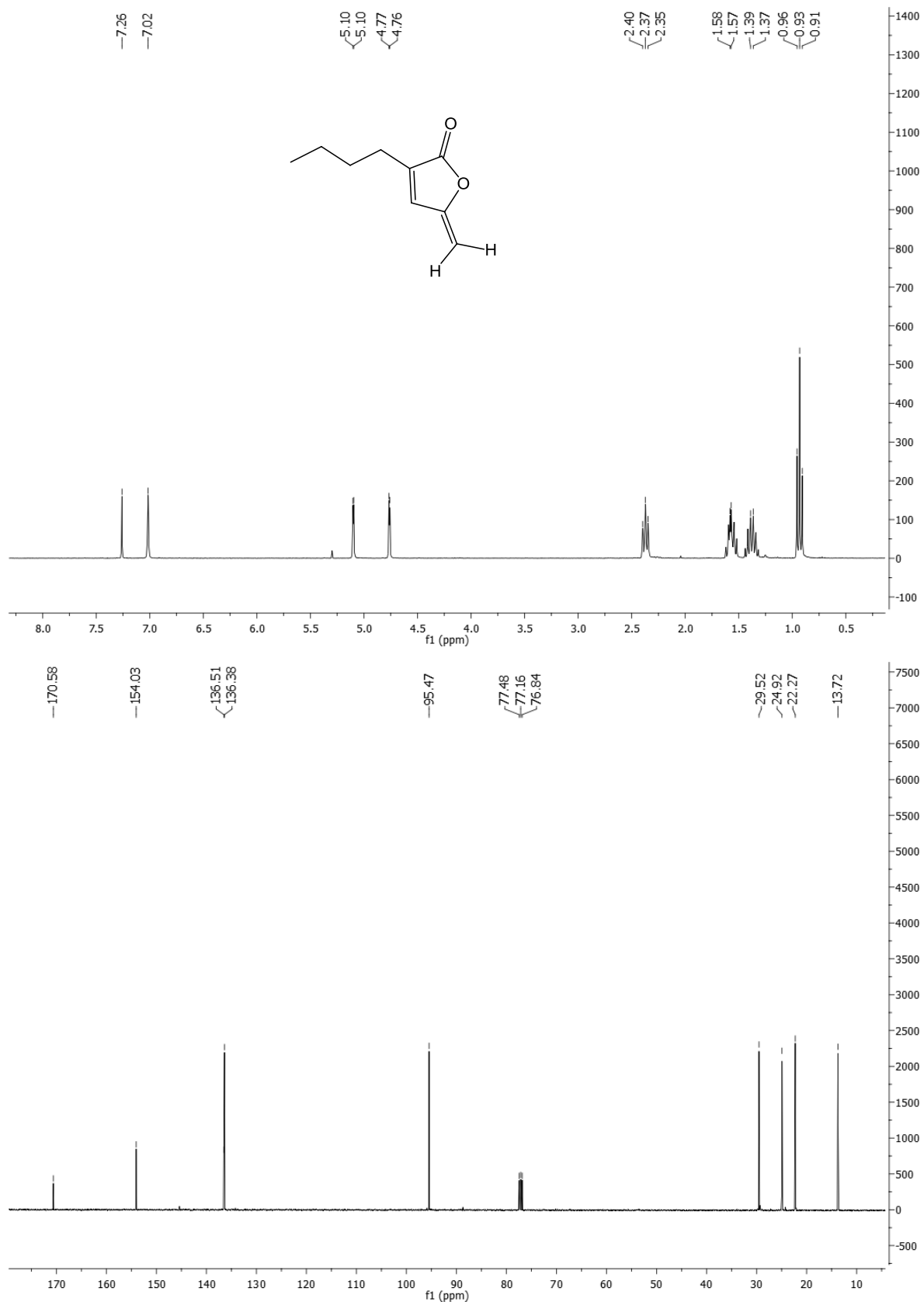
^1H and ^{13}C NMR spectrum
Sabbah et al.

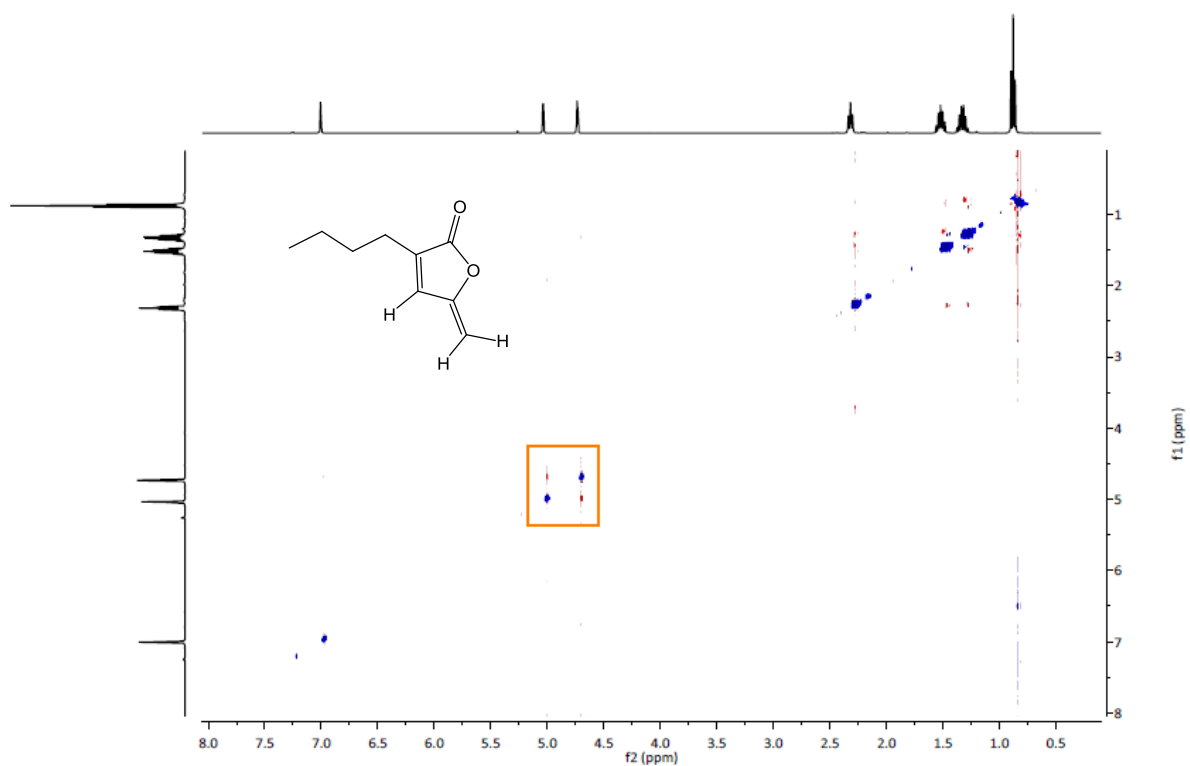


IV.1.11. 3-butyl-5-methylenefuran-2(5H)-one (8a)

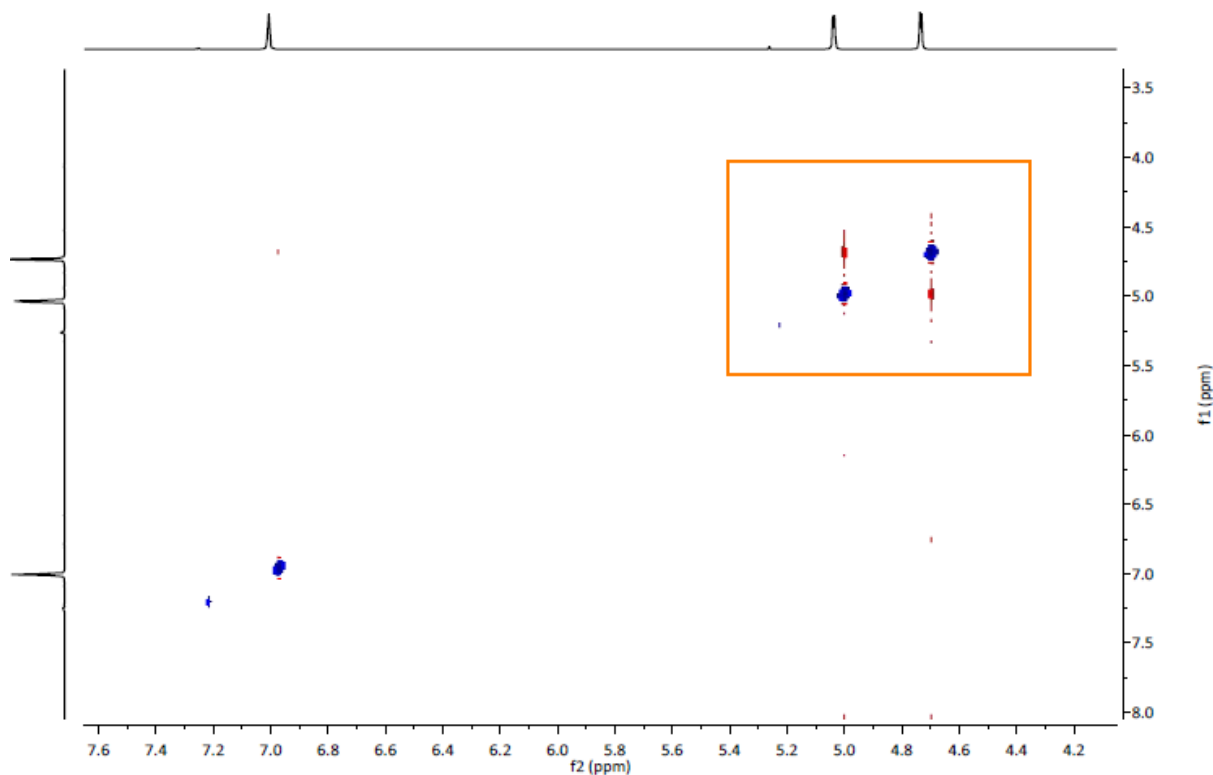
Supplementary Material (ESI) for Chemical Communications
This journal is (c) The Royal Society of Chemistry 2012

¹H and ¹³C NMR spectrum
Sabbah et al.



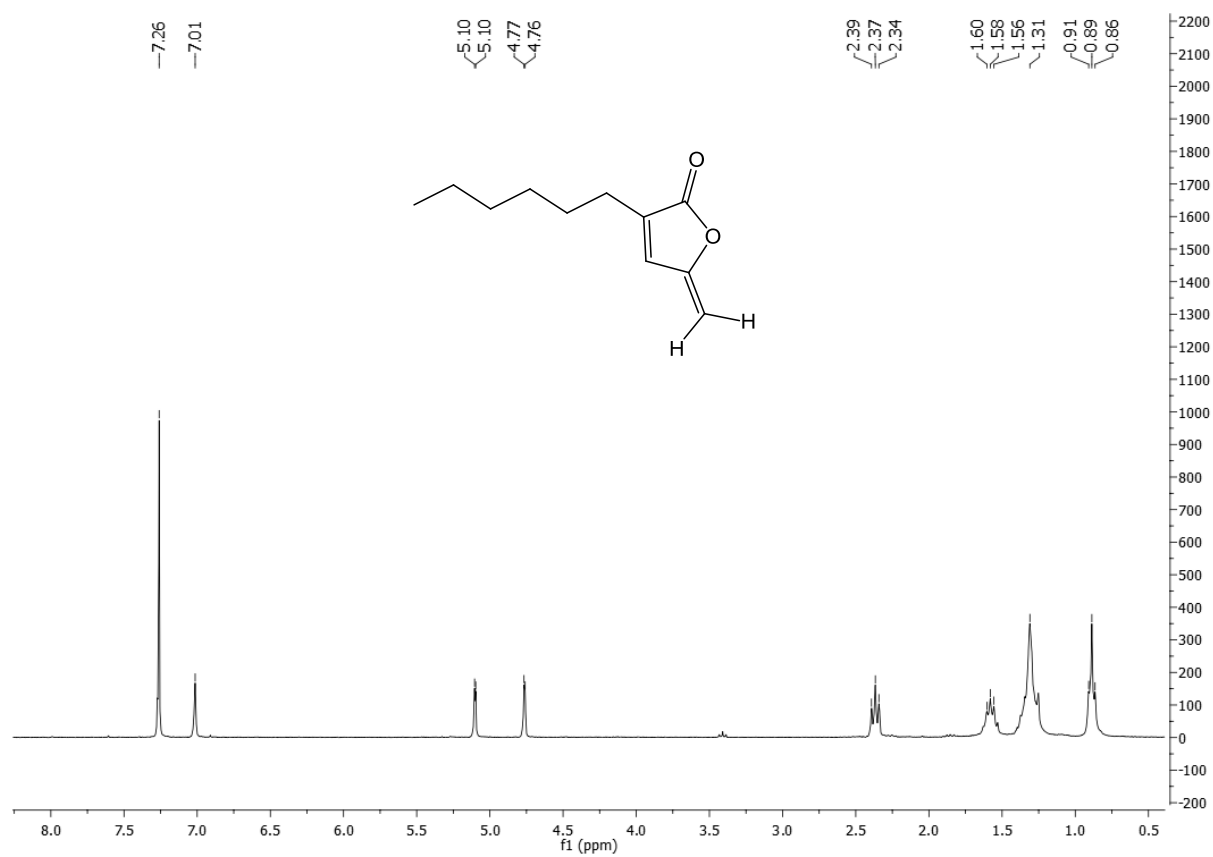


NOESY

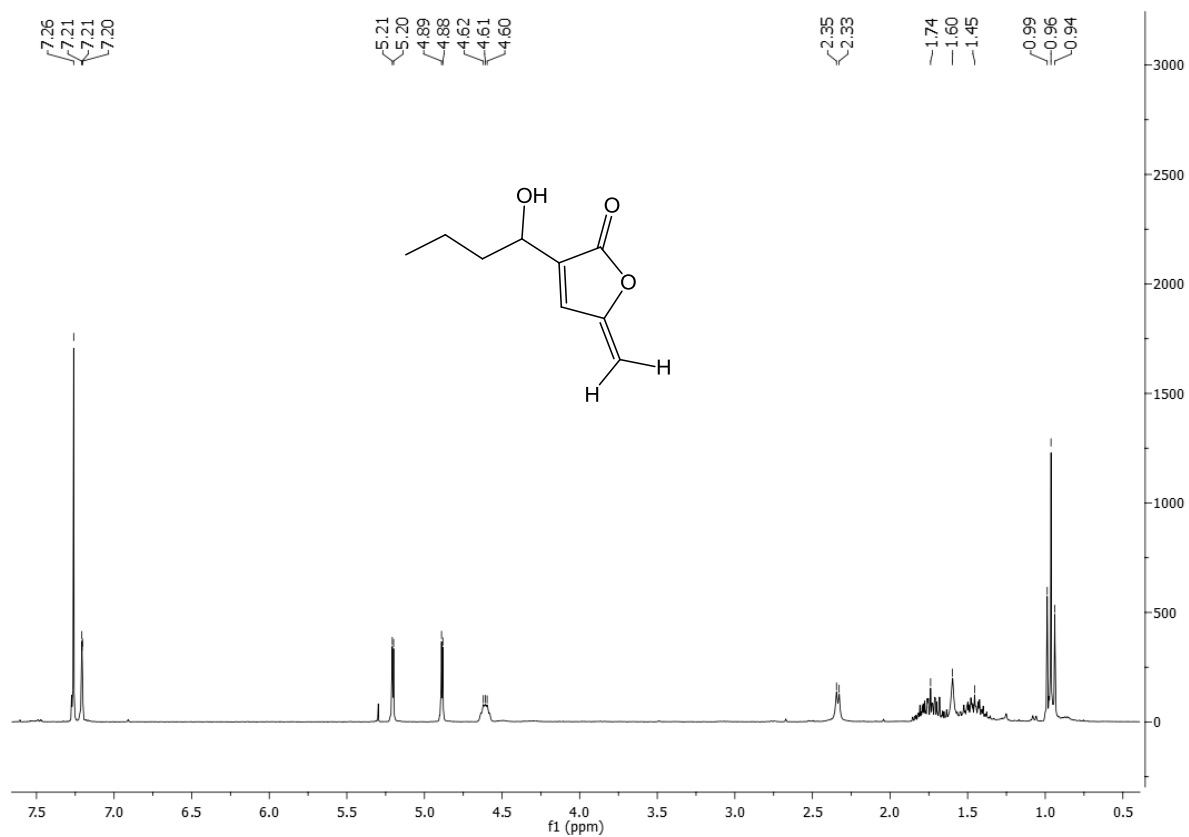


NOESY (Zoom)

IV.1.12. 3-hexyl-5-methylenefuran-2(5H)-one (8b)



IV.1.13. 3-(1-hydroxybutyl)-5-methylenefuran-2(5H)-one (8c)

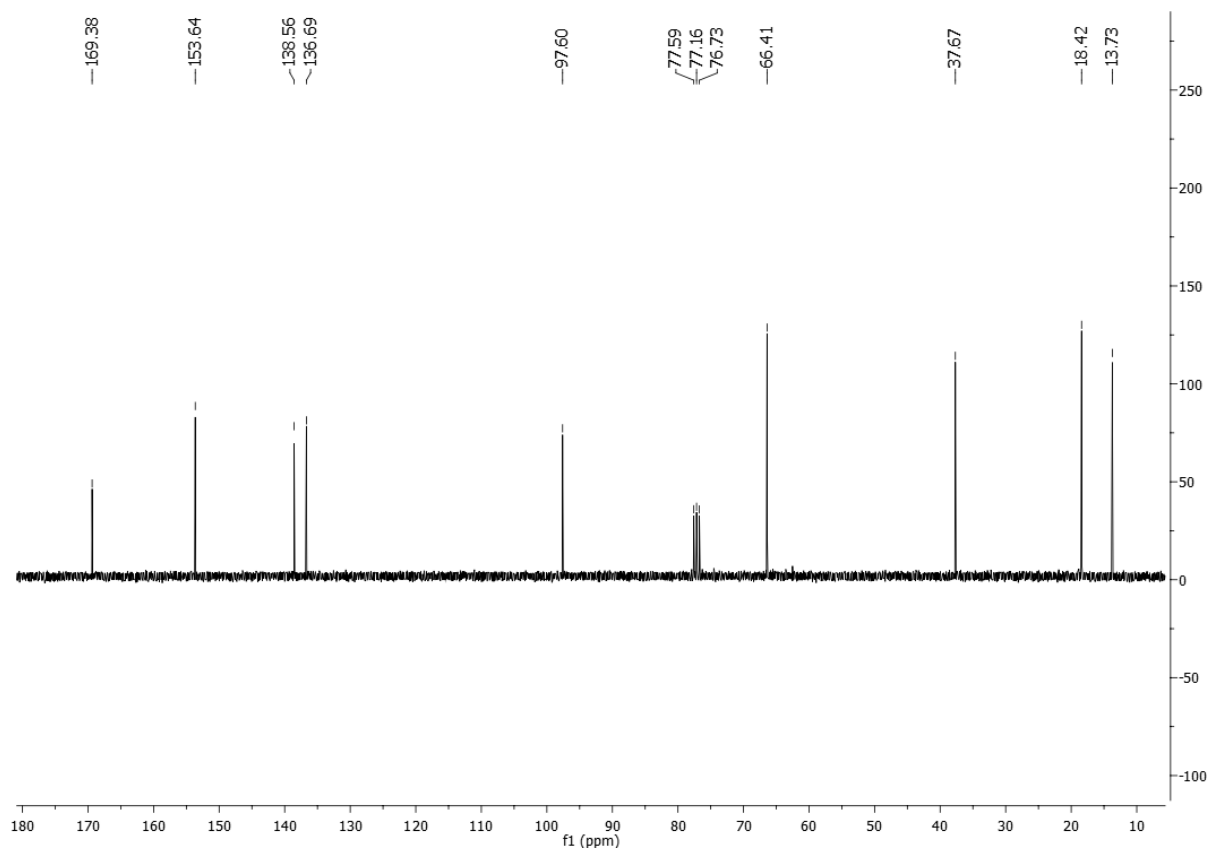


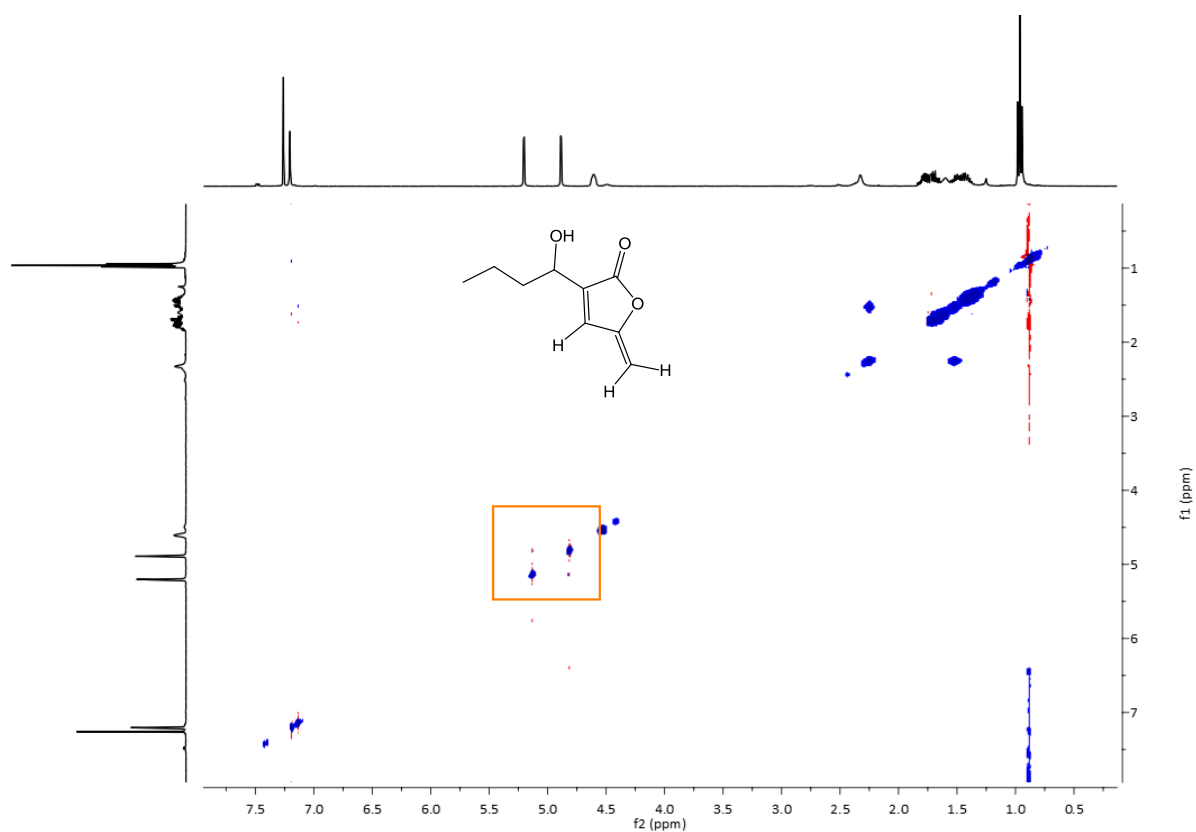
Supplementary Material (ESI) for Chemical Communications

This journal is (c) The Royal Society of Chemistry 2012

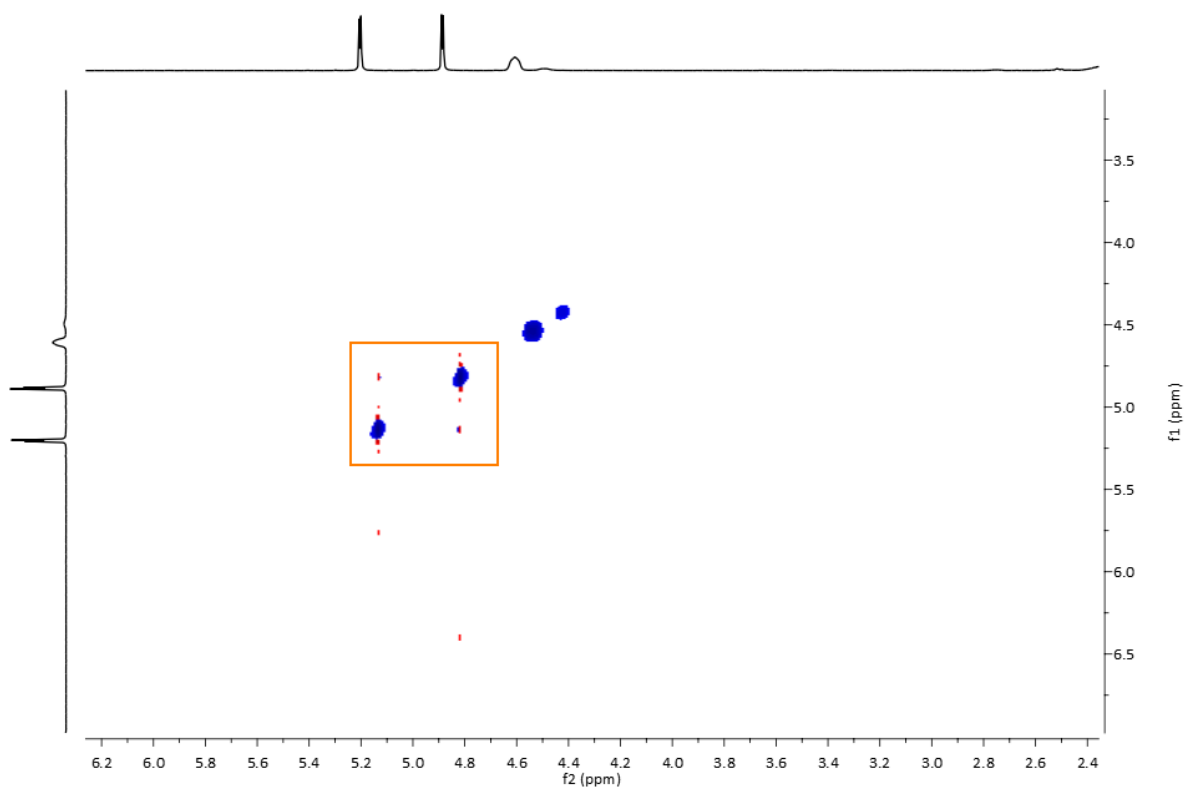
^1H and ^{13}C NMR spectrum

Sabbah et al.



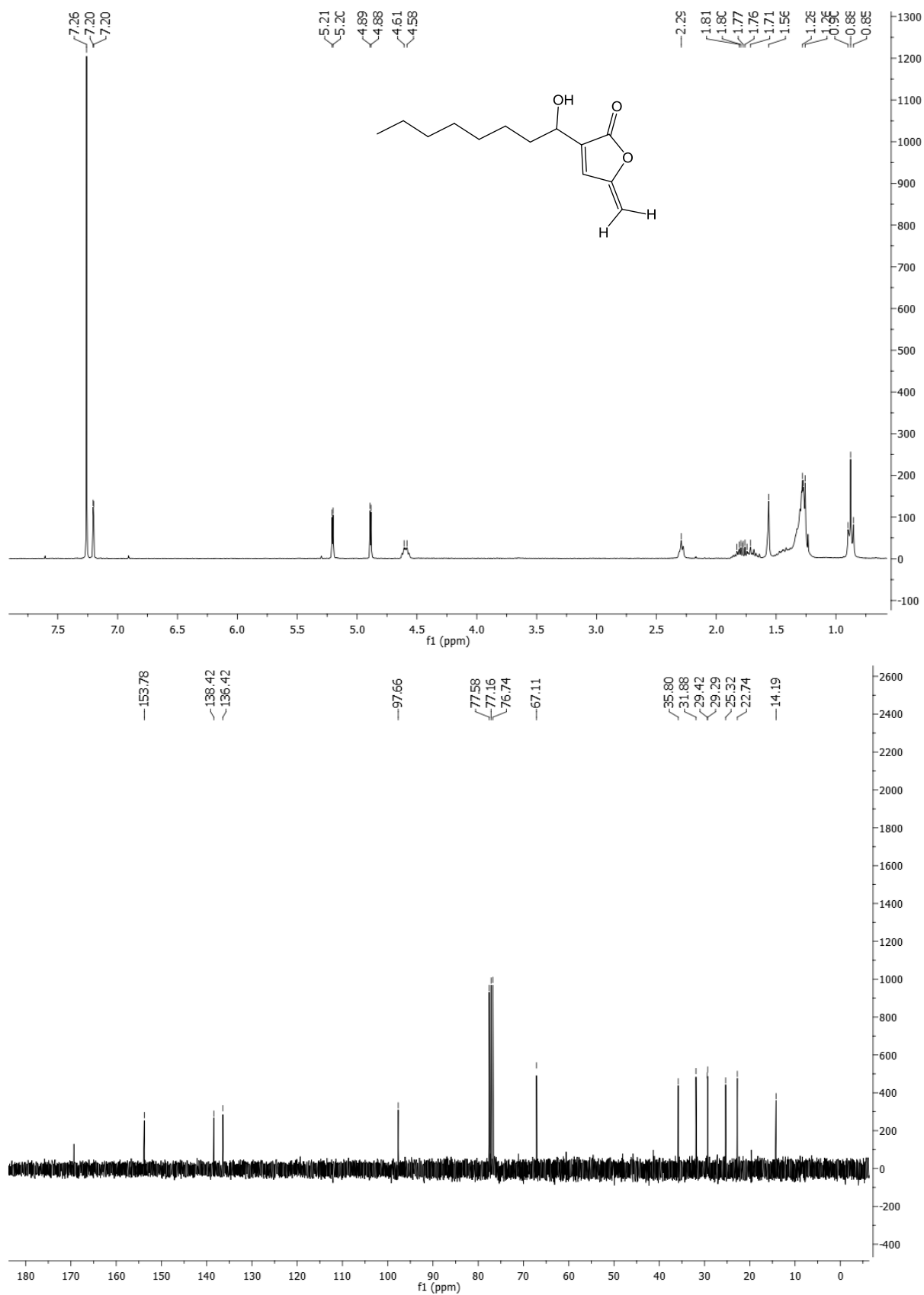


NOESY

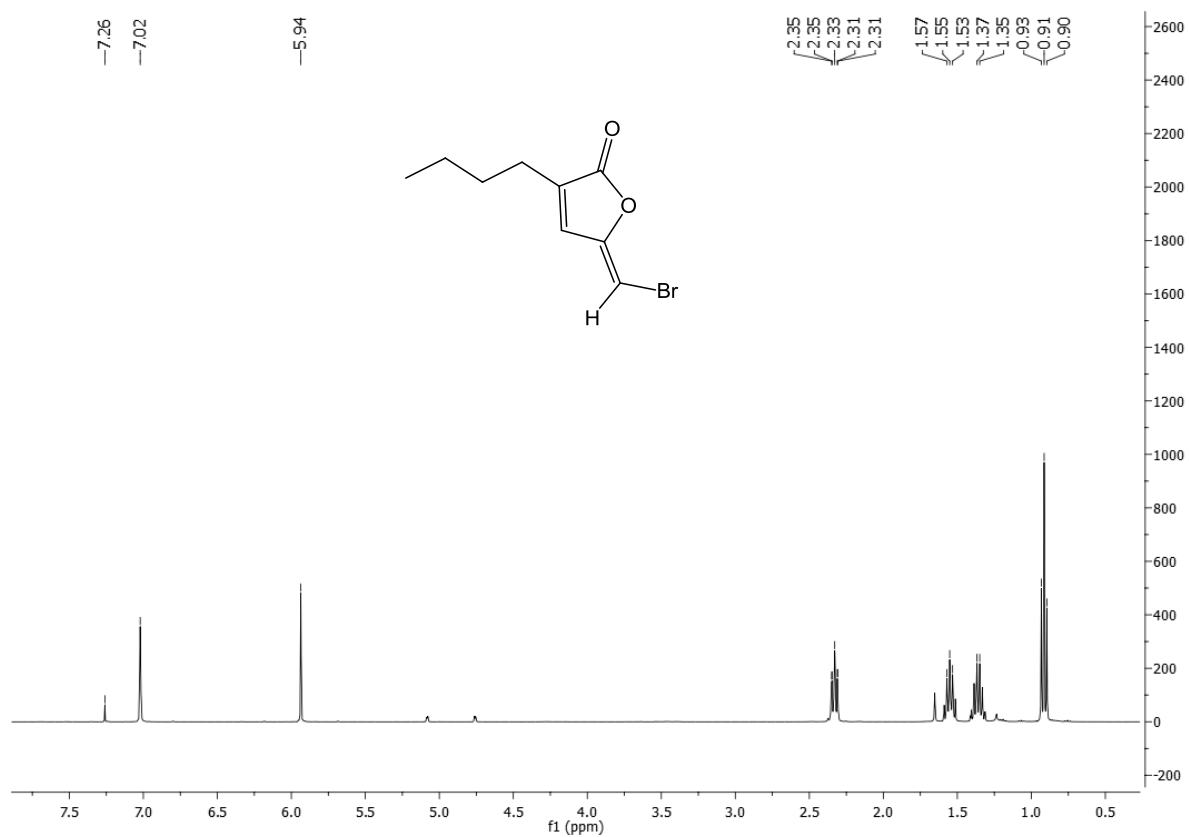


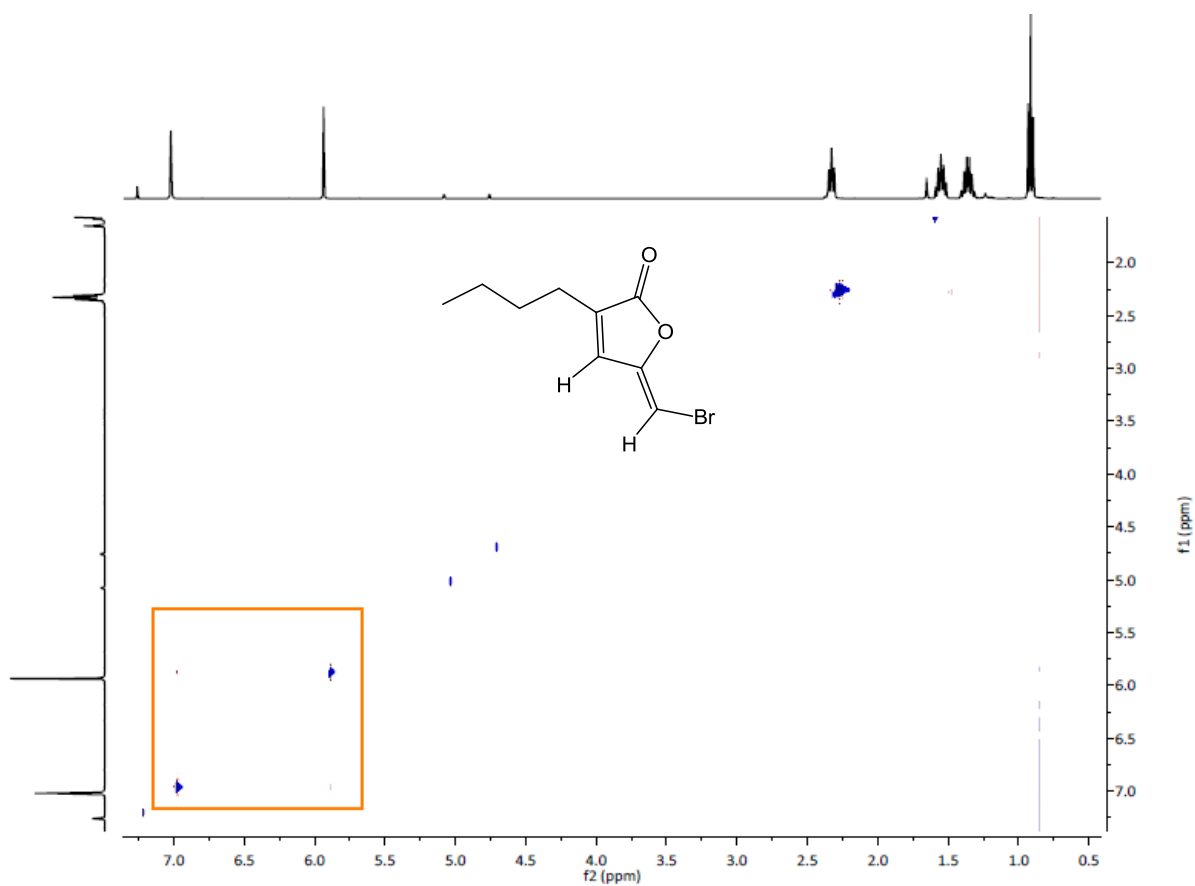
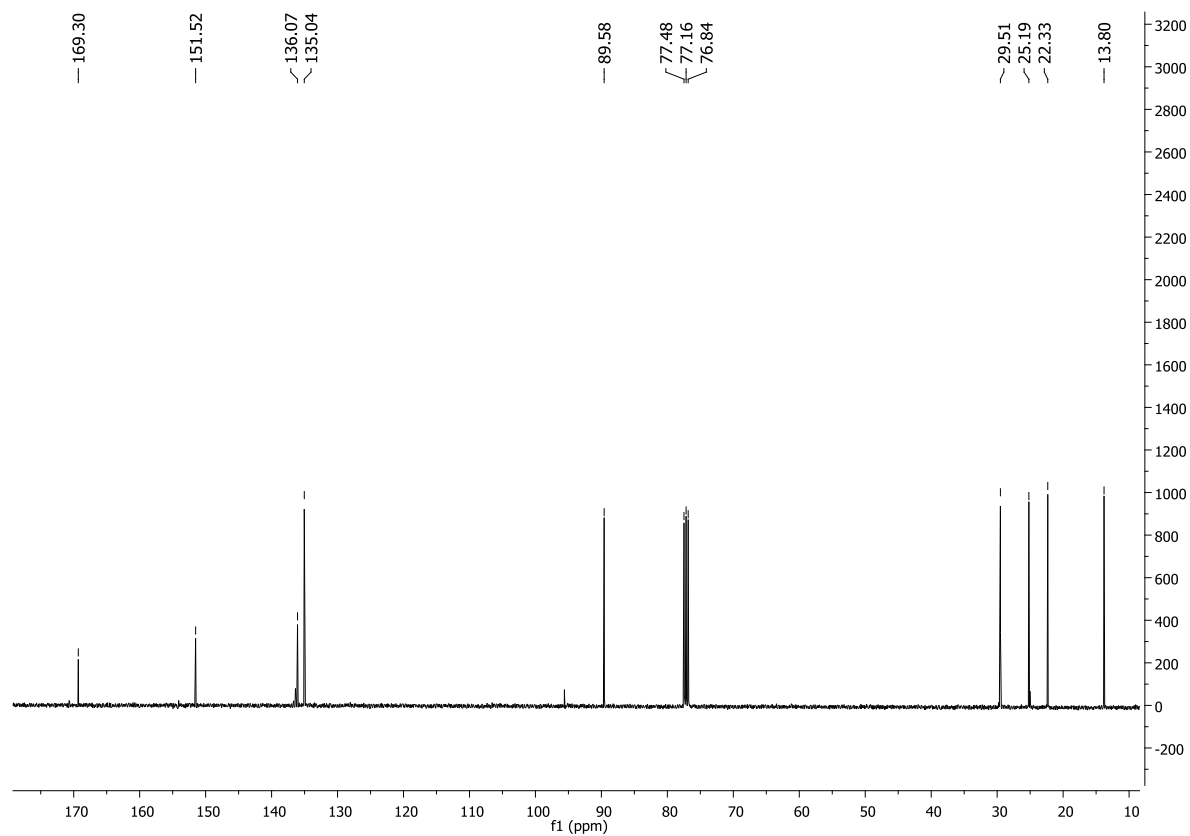
NOESY (Zoom)

IV.1.14. 3-(1-hydroxyoctyl)-5-methylenefuran-2(5H)-one (8e)

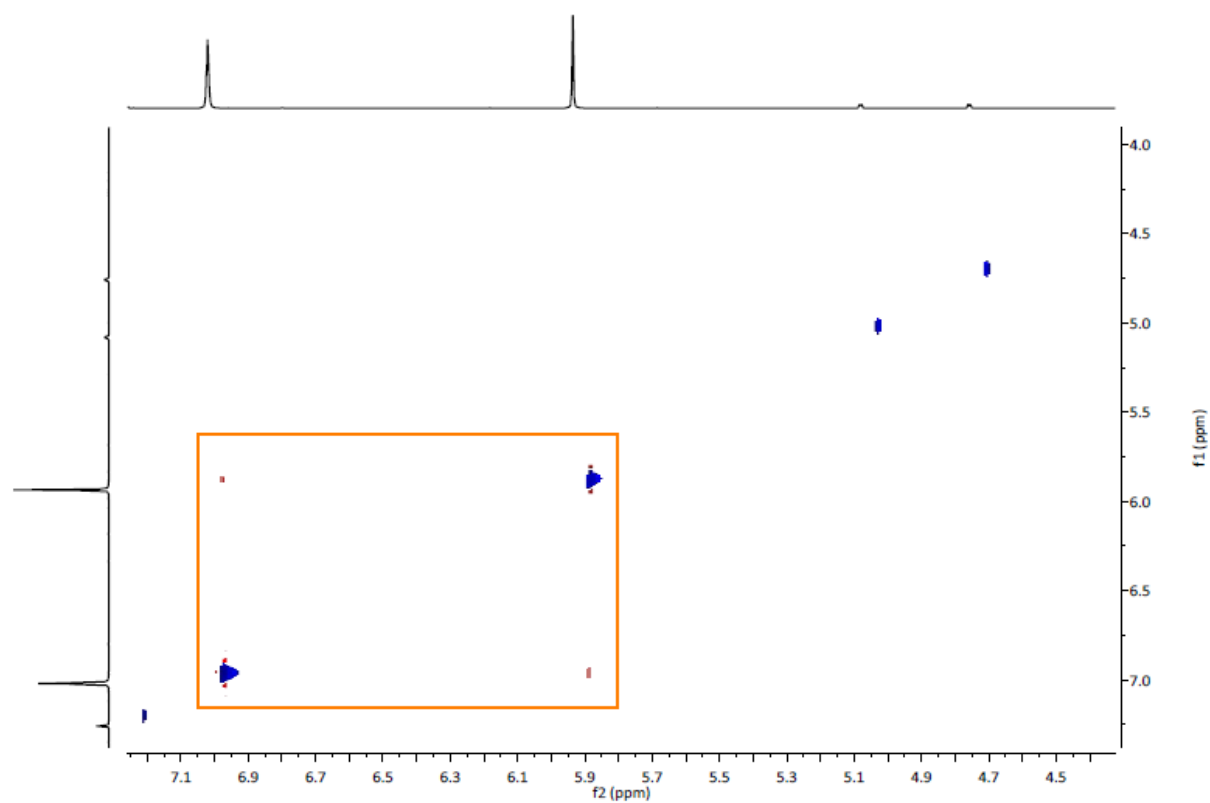


IV.1.15. (Z)-5-(bromomethylene)-3-butylfuran-2(5H)-one (10a)



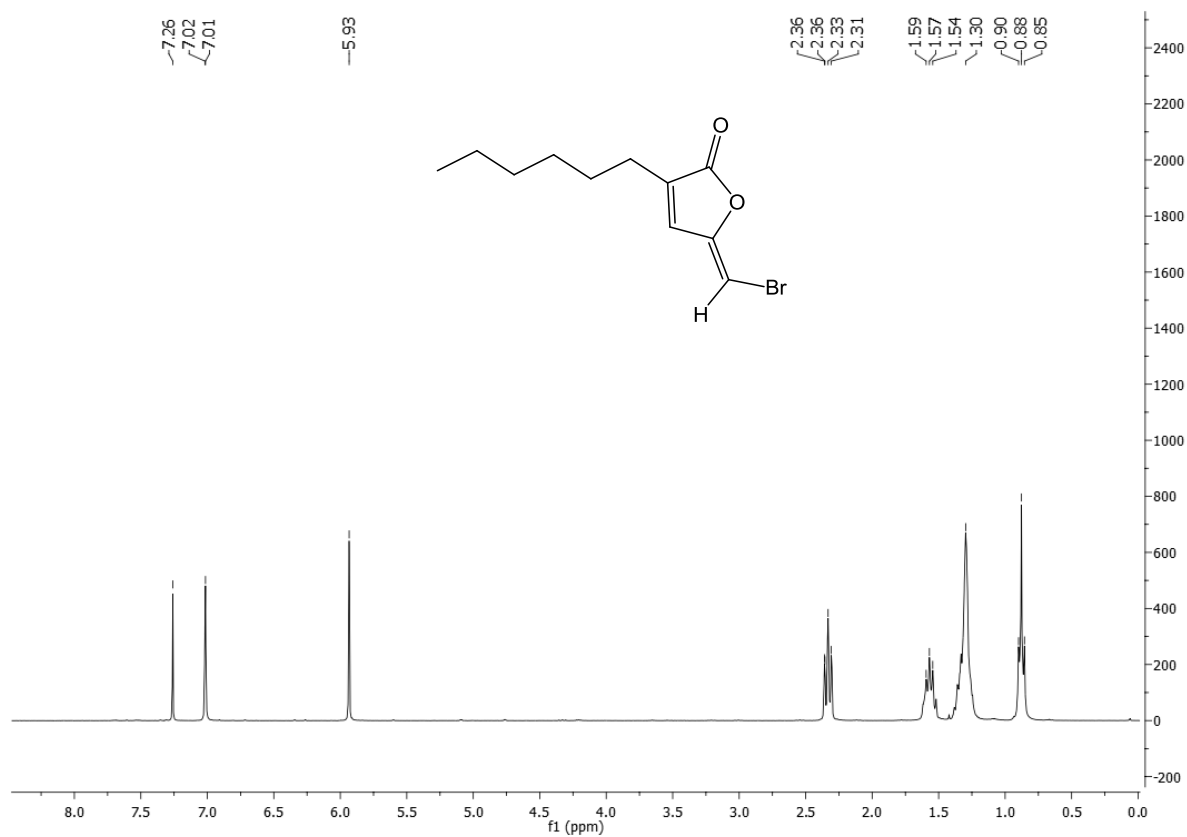


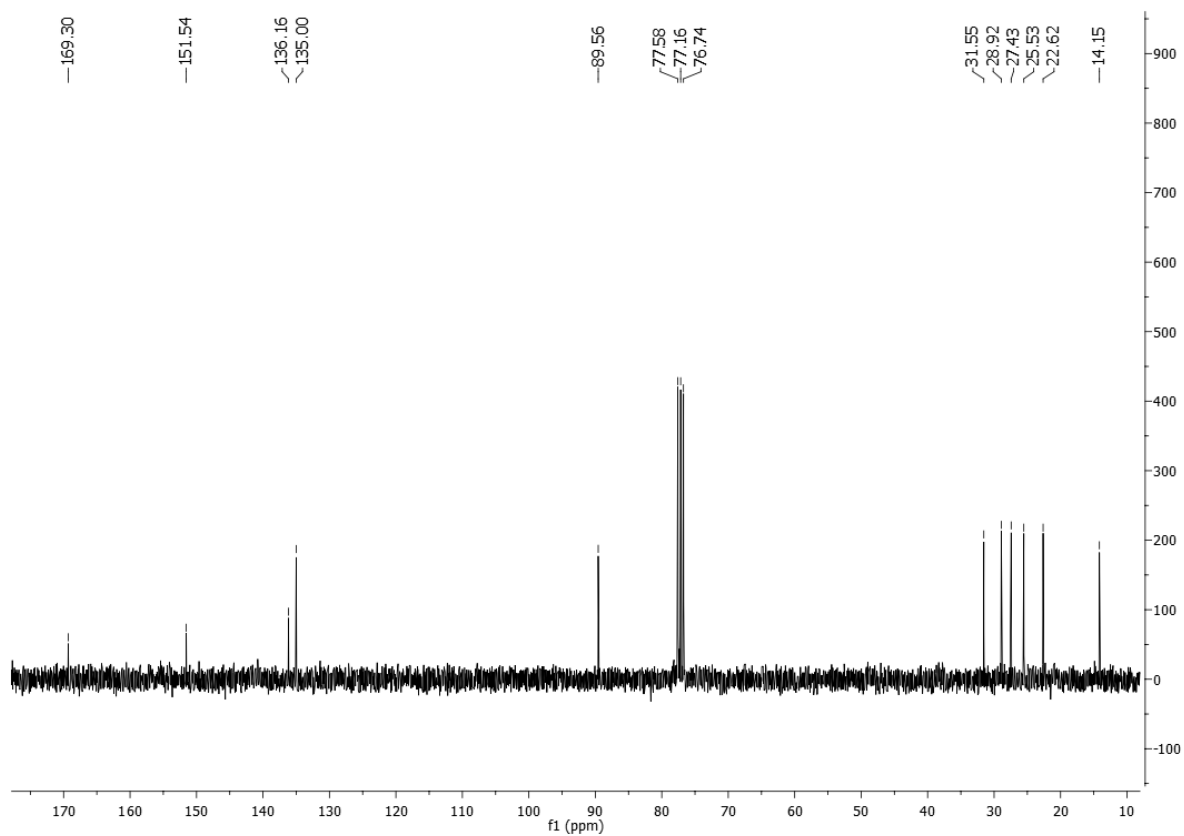
NOESY



NOESY (Zoom)

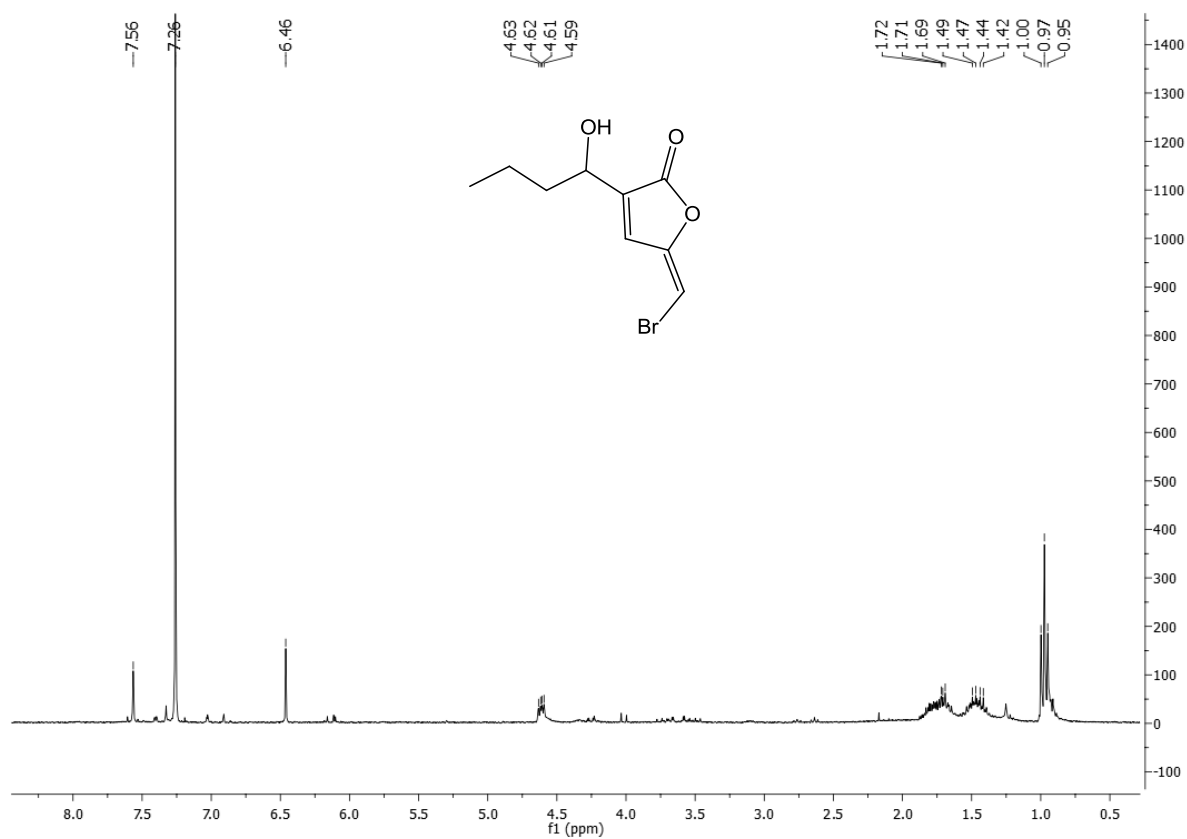
IV.1.16. (Z)-5-(bromomethylene)-3-hexylfuran-2(5H)-one (10b)

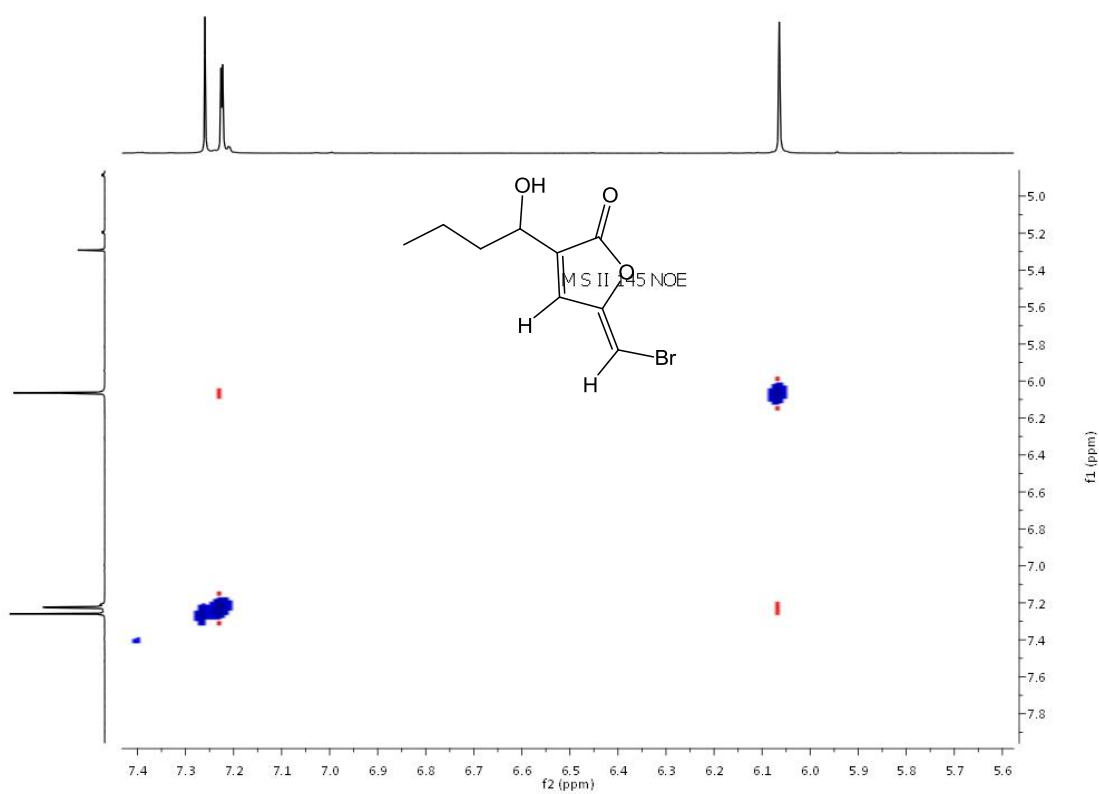
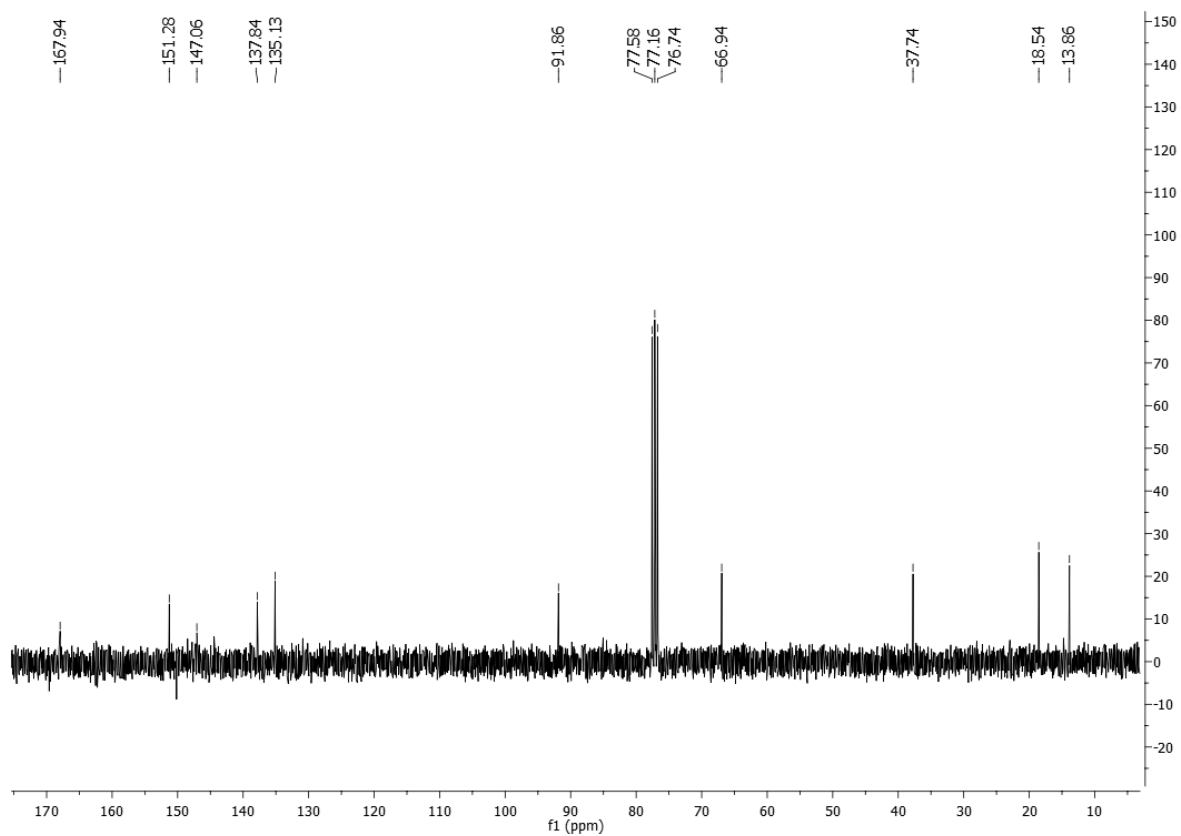




IV.1.17. (Z)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)

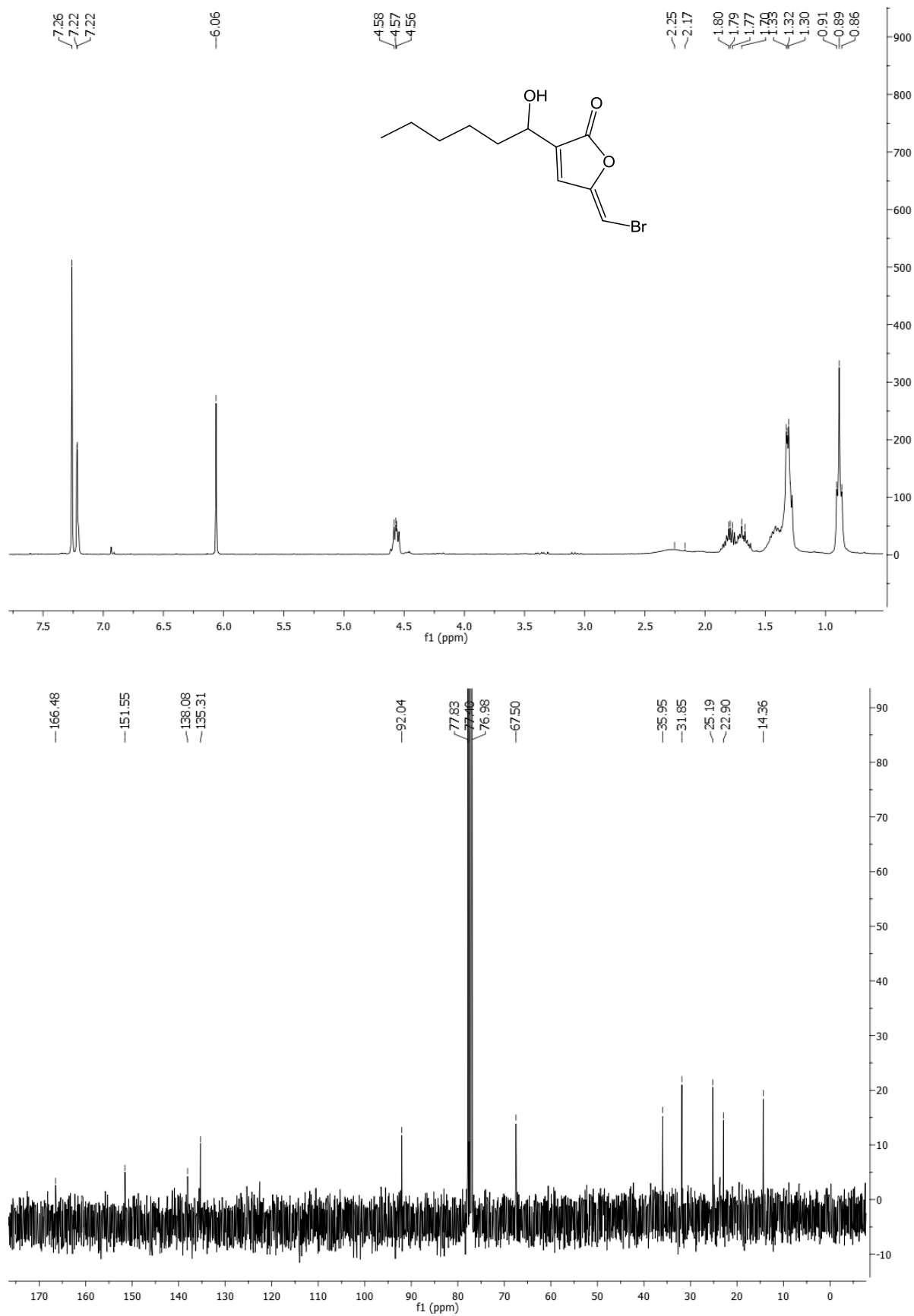
(E)-5-(bromomethylene)-3-(1-hydroxybutyl)furan-2(5H)-one (10c)



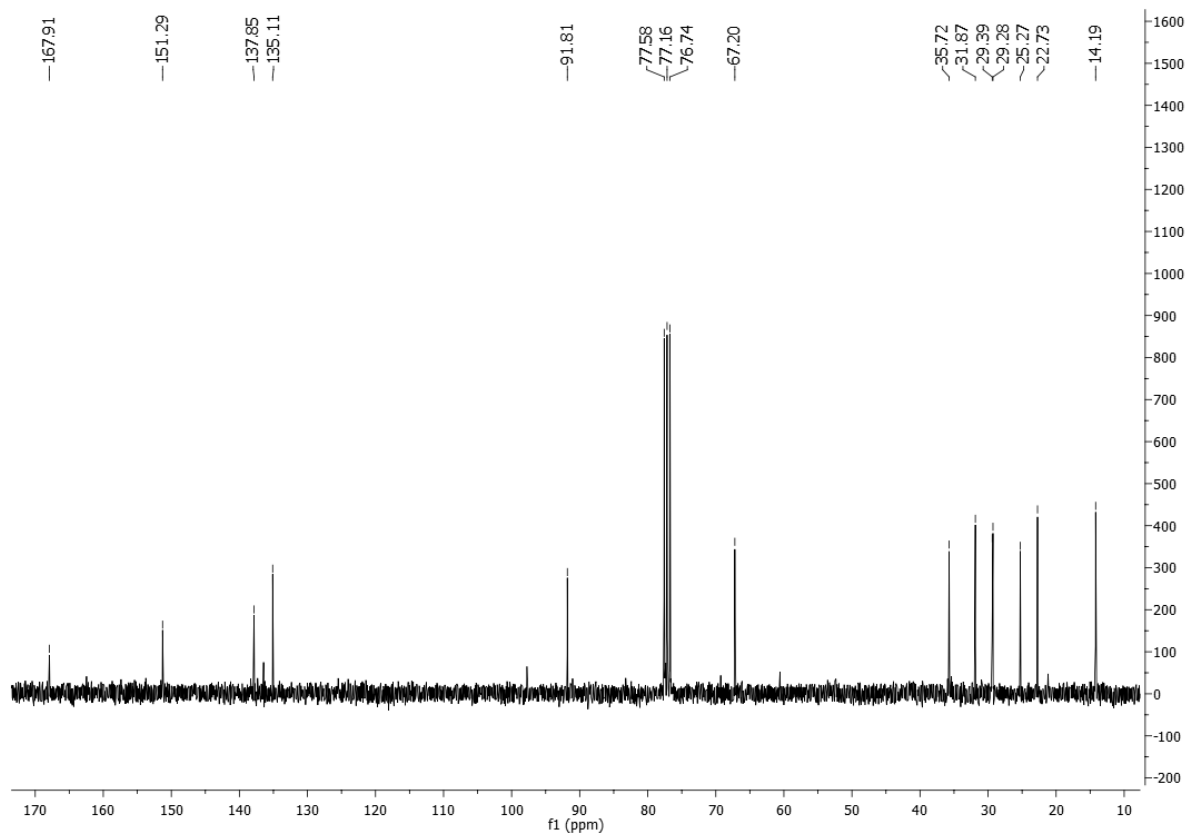
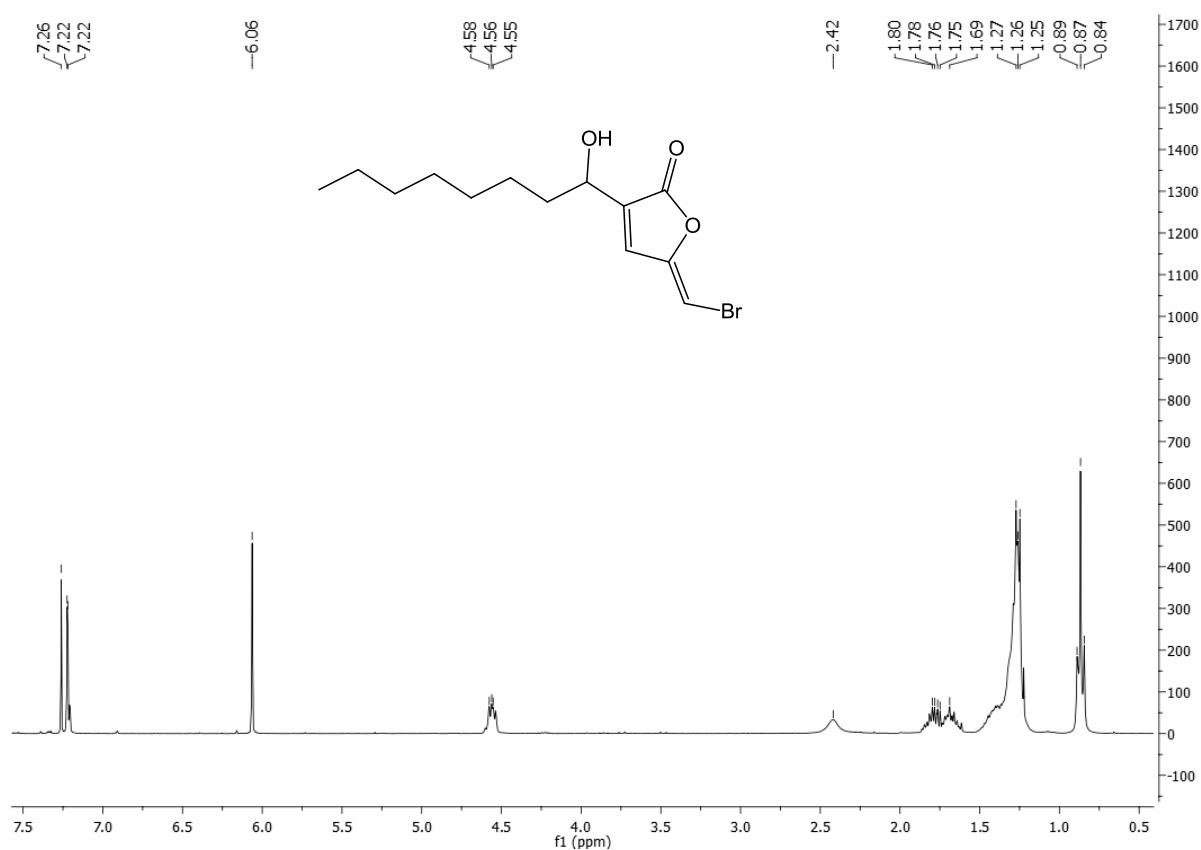


NOESY (Zoom)

IV.1.18. (Z)-5-(bromomethylene)-3-(1-hydroxyhexyl)furan-2(5H)-one (10d)



IV.1.19. (Z)-5-(bromomethylene)-3-(1-hydroxyoctyl)furan-2(5H)-one (10e)



V. REFERENCES

1. A. Calderón, P. de March, M. el Arrad and J. Font, *Tetrahedron*, 1994, **50**, 4201-4214.
2. K. Tanaka, M. Terauchi and A. Kaji, *chem. Lett.*, 1982, 351-352.
3. Y. Sekiyama, H. Araya, K. Hasumi, A. Endo and Y. Fujimoto, *Nat. Prod. Lett.*, 1997, **11**, 61 - 66.
4. K. P. Haval and N. P. Argade, *Synthesis*, 2007, **2007**, 2198,2202.
5. H. P. Steenackers, J. Levin, J. C. Janssens, A. D. Weerd, J. Balzarini, J. Vanderleyden, D. E. De Vos and S. C. De Keersmaecker, *Bioorg. Med. Chem.*, 2010, **18**, 5224-5233.
6. G. Kronvall and S. Ringertz, *APMIS*, 1991, **99**, 295-306.
7. M. A. Thompson, *ArgusLaB 4.0.1*, Seattle, WA planetaria Software LLC, 2004.
8. L. Soulère, M. Frezza, Y. Queneau and A. Doutheau, *J. Mol. Graph. Model.*, 2007, **26**, 581-590.
9. K. Arnold, L. Bordoli, J. Kopp and T. Schwede, *Bioinformatics*, 2006, **22**, 195-201.
10. J. D. Thompson, D. G. Higgins and T. J. Gibson, *Nucleic Acids Research*, 1994, **22**, 4673-4680.
11. J. Estephane, J. Dauvergne, L. Soulere, S. Reverchon, Y. Queneau and A. Doutheau, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 4321-4324.
12. G. M. Morris, D. S. Goodsell, R. S. Halliday, R. Huey, W. E. Hart, R. K. Belew and A. J. Olson, *J. Comp. Chem.*, 1998, **19**, 1639.