

Electronic Supplementary Information for

Cystine-derived Bis-naphthalimides as Stimuli-Responsive Fluorescent Gelators

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Details of synthesis, characterization of the ligands **L1** and **L2** (Figures S1-S4 to S6-S9, copies of the ^1H & ^{13}C NMR spectra), ES-MS (Figure S5, S10), along with the relevant UV-vis, fluorescence, FT-IR and ^1H NMR spectra (Figures S11-S24).

1. General experimental techniques.

All chemicals were commercially available from Sigma-Aldrich or Spectrochem (India) and used as received. Solvents for spectroscopic experiments were distilled under nitrogen atmosphere before use. All ^1H and ^{13}C NMR were measured on a 300 MHz Bruker spectrometer, and reported in δ/ppm . The electronic absorption spectra were recorded on a Shimadzu UV-VIS spectrophotometer.

2. Copies of $^1\text{H}/^{13}\text{C}$ NMR spectra for ND1, ND2, L1 and L2.

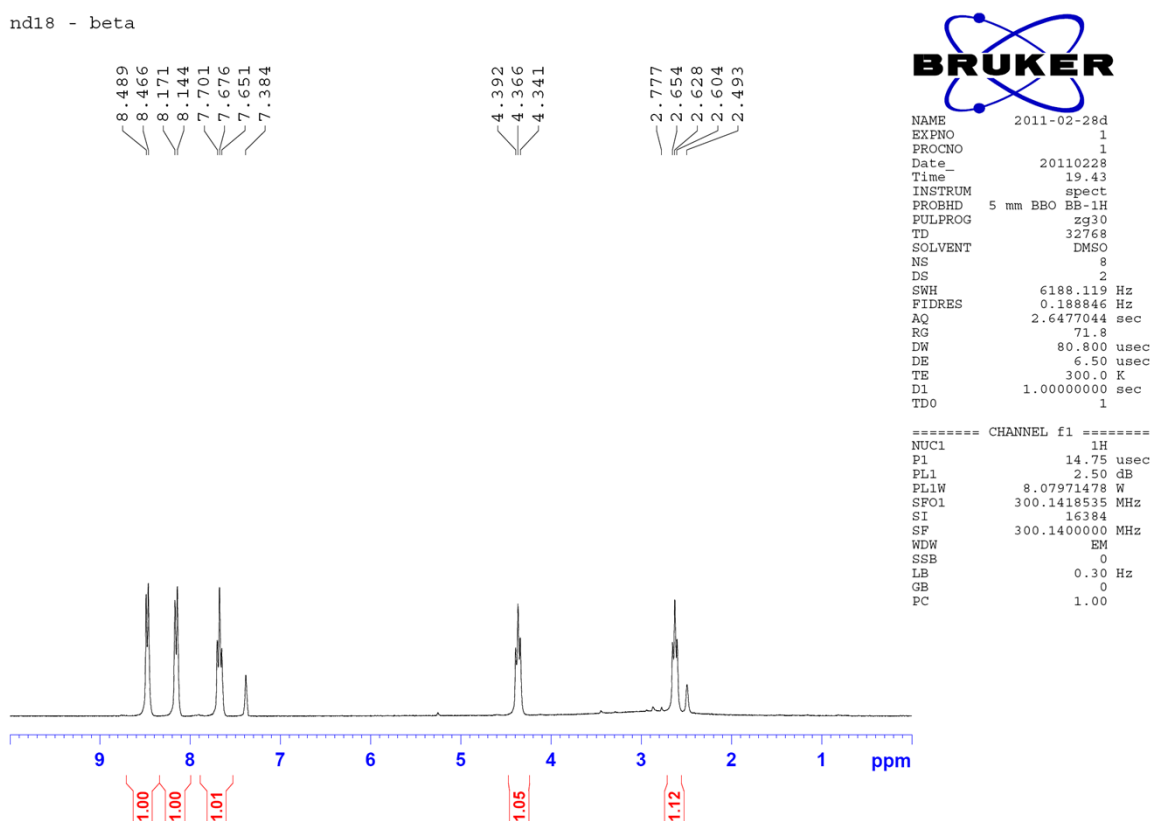


Figure S1: ^1H spectra of **ND1** in $\text{DMSO-d}_6/\text{CDCl}_3$

nd18-beta 13C

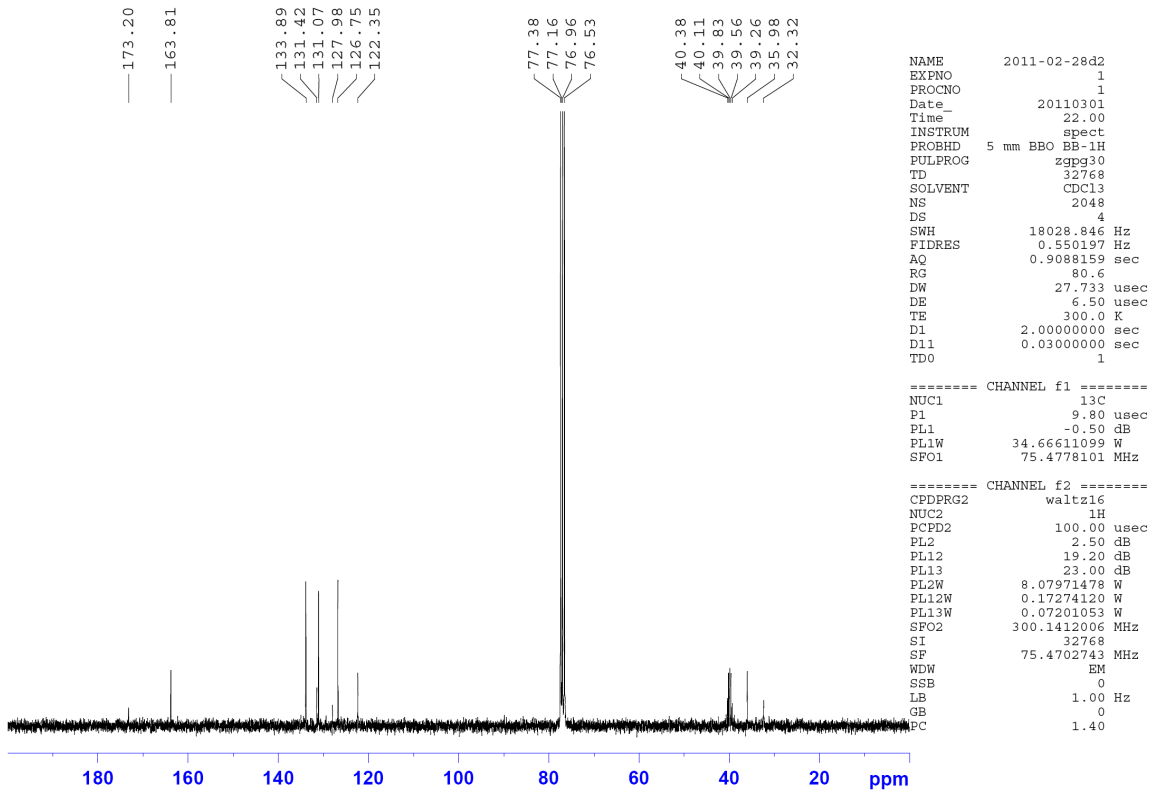


Figure S2: ^{13}C NMR spectra of ND1 in DMSO- d_6 /CDCl $_3$

nd18B - CysOMe (# re)

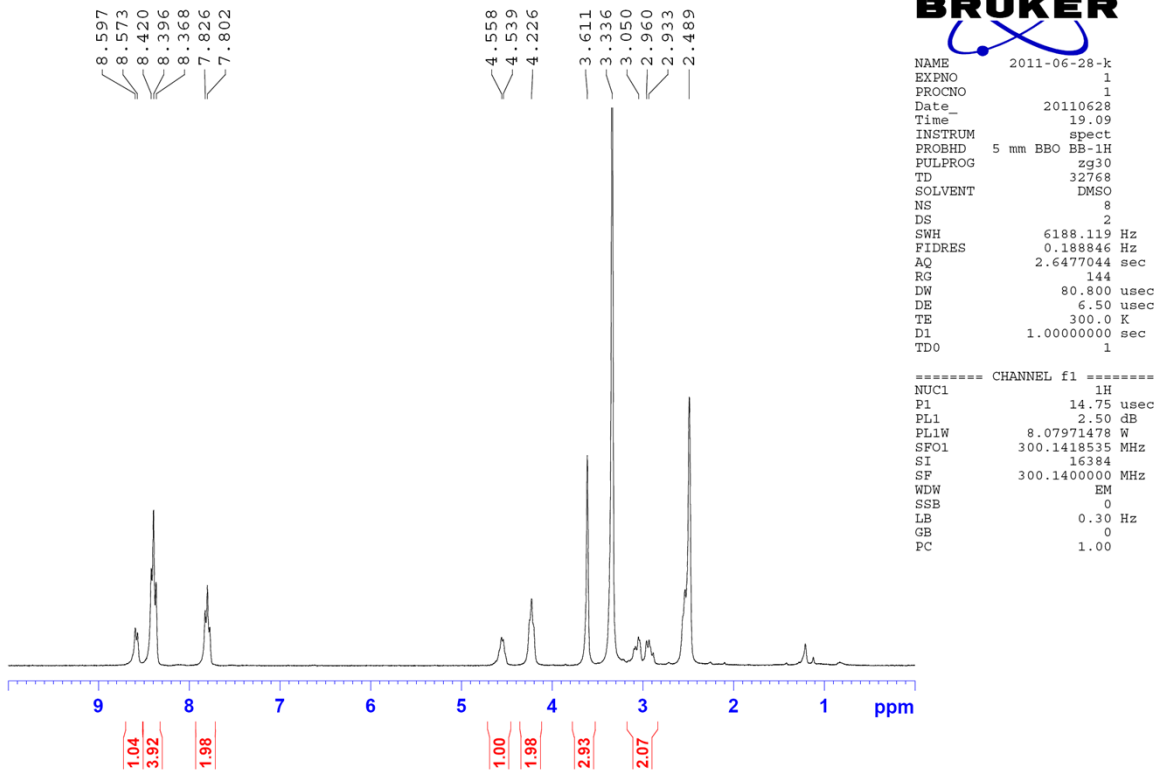


Figure S3: ^1H NMR spectra of L1 in DMSO- d_6

nd18B - CysOMe (13C)

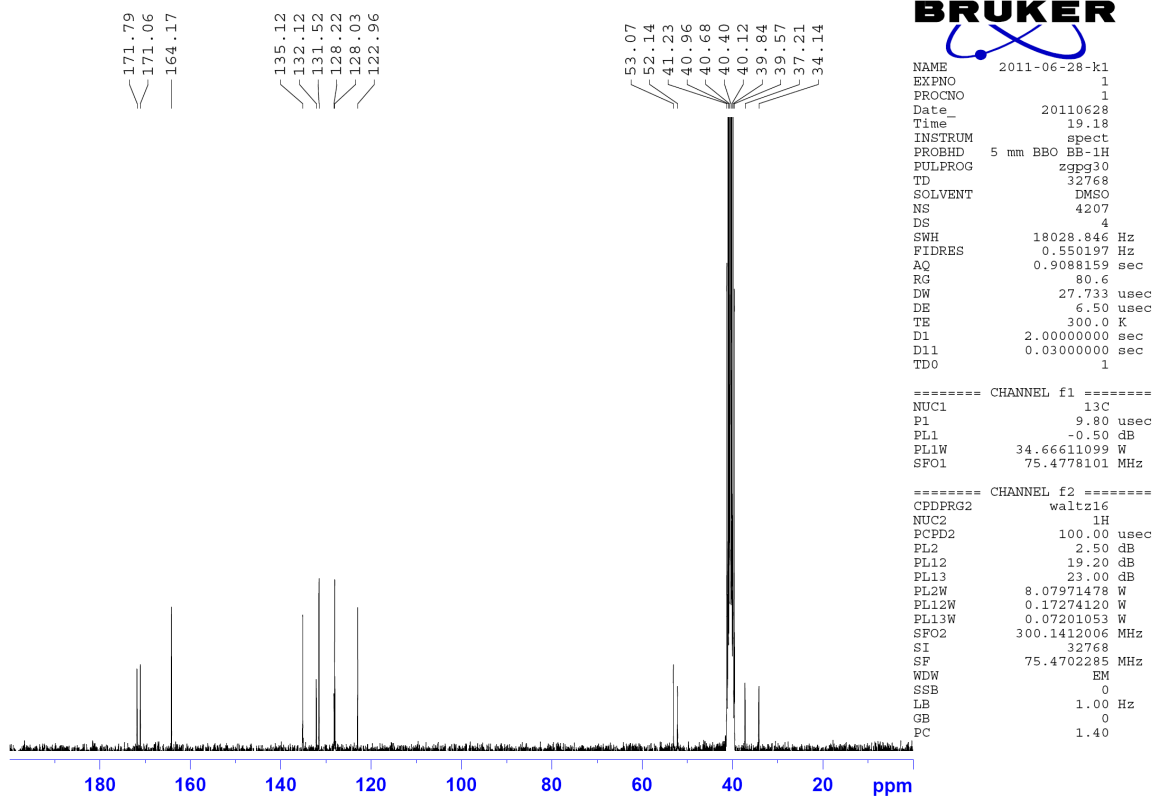


Figure S4: ^{13}C NMR spectra of **L1** in DMSO-d_6

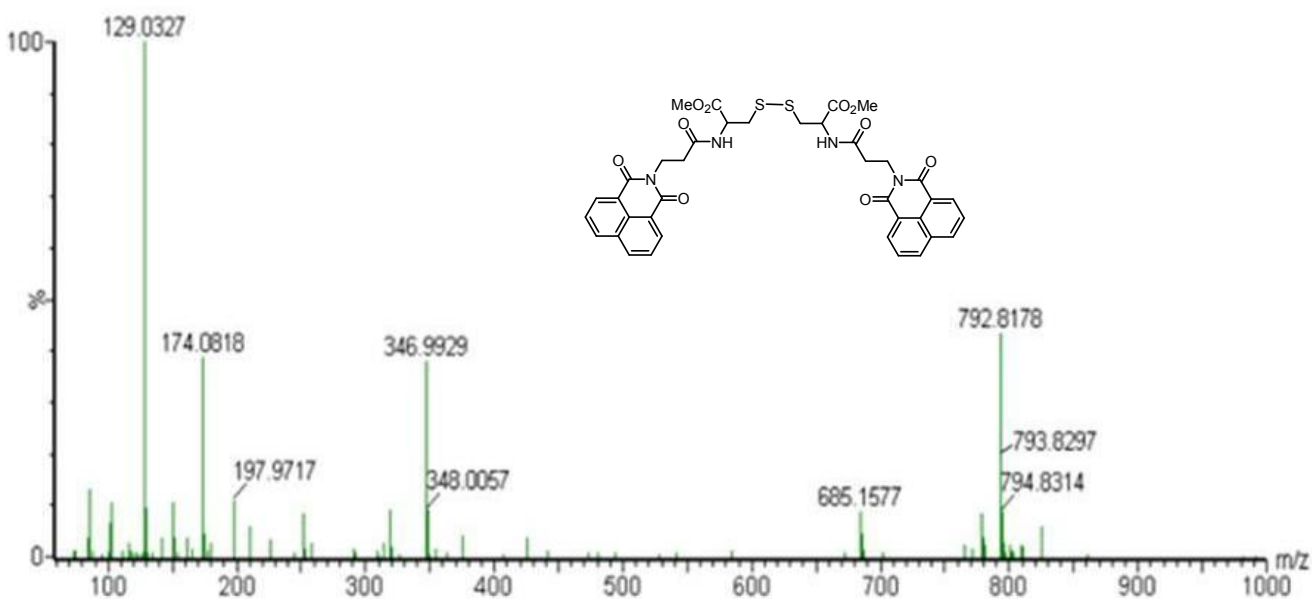


Figure S5: ES-MS of **L1** in MeCN ($m/z = 793.81$, calc. for $\text{M}+\text{Na}^+$)

nd18 - gaba 1H

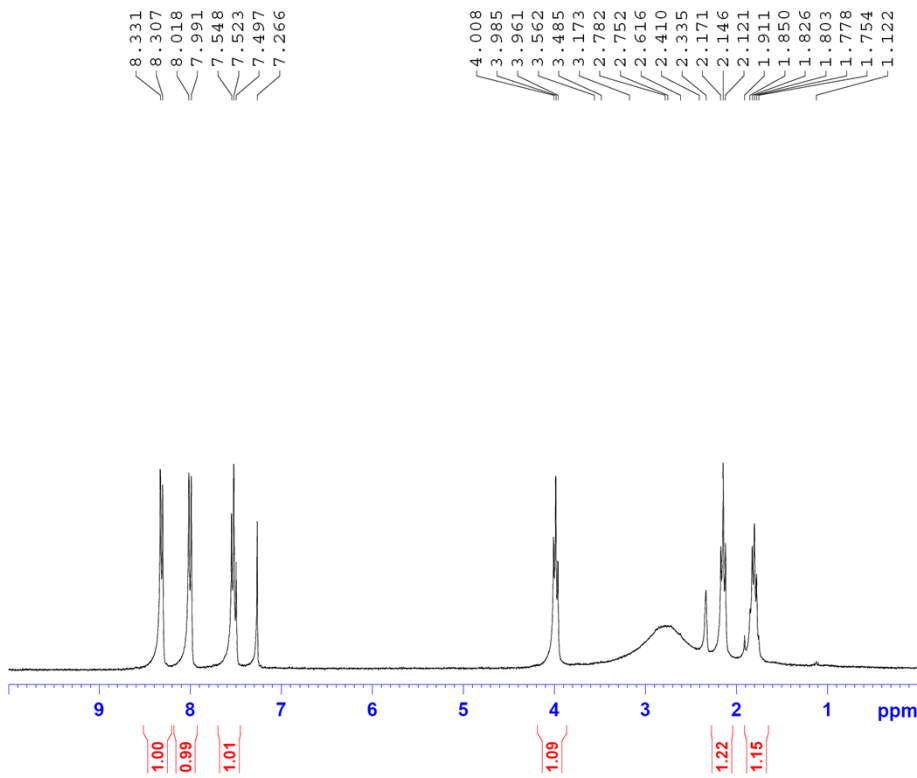


Figure S6: ¹H NMR spectra of ND2 in DMSO-d₆/CDCl₃

nd18 - gaba 13C

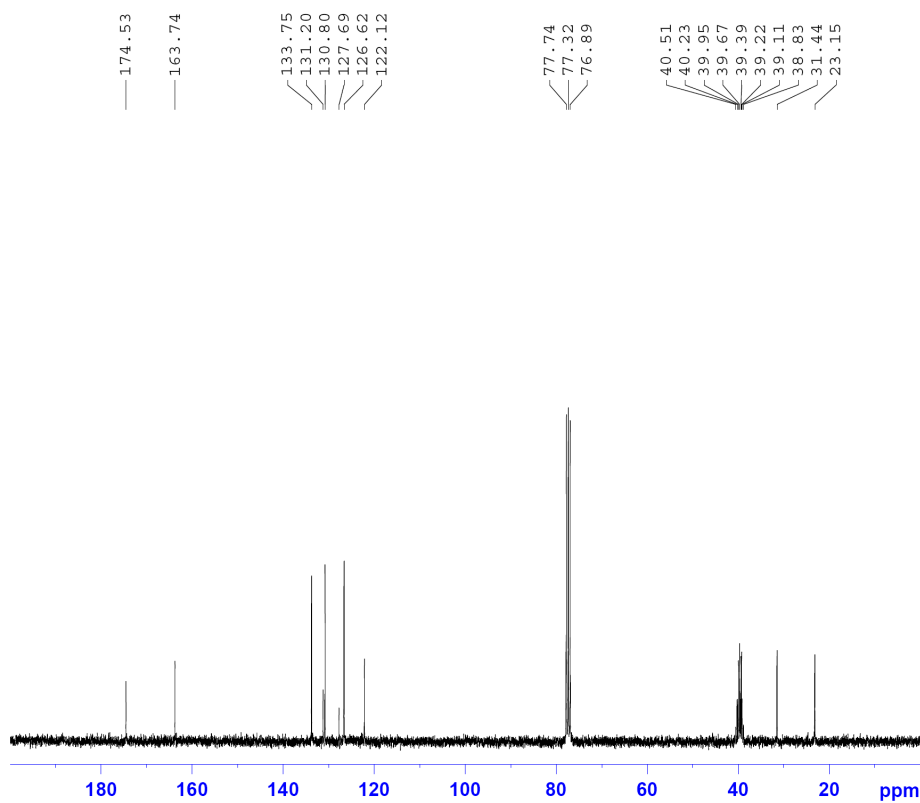
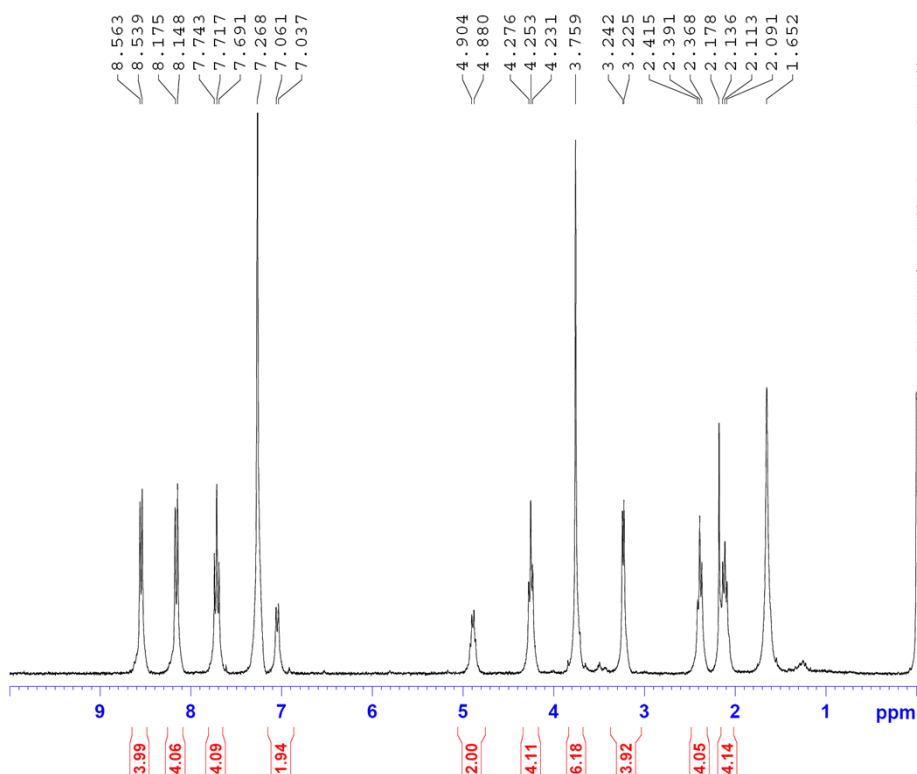


Figure S7: ¹³C NMR spectra of ND2 in DMSO-d₆/CDCl₃

nd18G - CysOMe (# 1)



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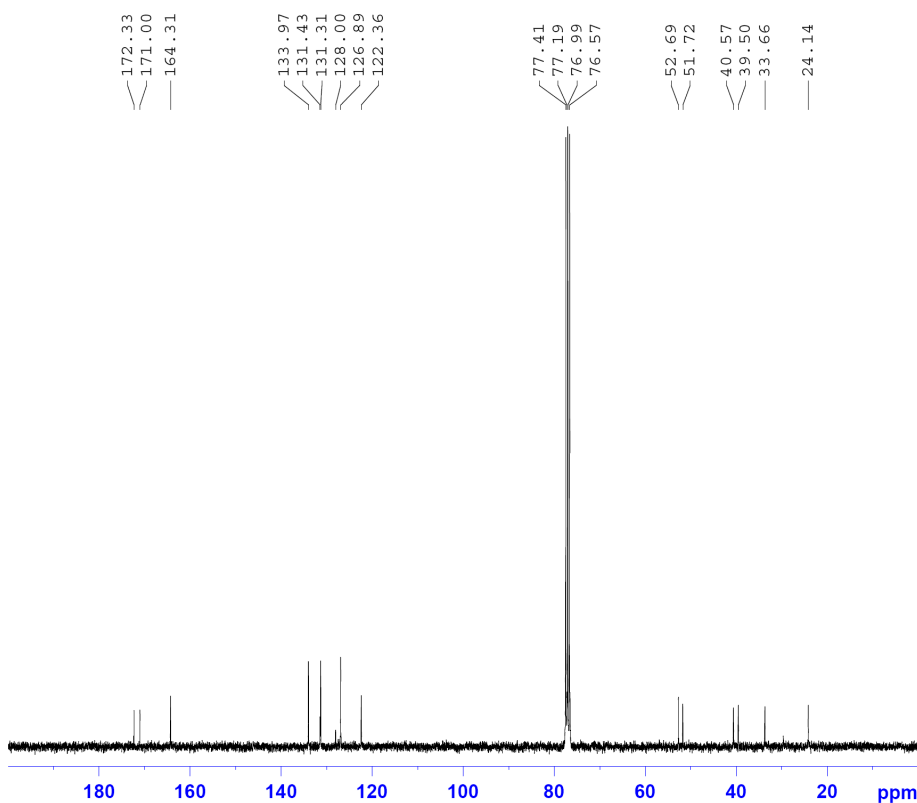
NAME      2011-06-24c
EXPNO     1
PROCNO    1
Date_     20110624
Time      18.32
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PROBHD    5 mm BBO BB-1H
PULPROG   zg30
TD        32768
SOLVENT   CDCl3
NS        8
DS        2
SWH       6188.119 Hz
FIDRES    0.188846 Hz
AQ        2.6477044 sec
RG        80.6
DW        80.800 usec
DE        6.50 usec
TE        300.0 K
D1        1.00000000 sec
TD0       1
  
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===== CHANNEL f1 =====
NUC1      1H
P1        14.75 usec
PL1       2.50 dB
PL1W      8.07971478 W
SFO1      300.1418535 MHz
SI        16384
SF        300.1400000 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
  
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Figure S8: ^1H NMR spectra of L2 in CDCl_3

nd18G - CysOMe (# 13C)



```

NAME      2011-06-27-e2
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PROCNO    1
Date_     20110627
Time      17.16
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PULPROG   zgpg30
TD        32768
SOLVENT   CDCl3
NS        2002
DS        4
SWH       18028.846 Hz
FIDRES    0.550197 Hz
AQ        0.9088159 sec
RG        80.6
DW        27.733 usec
DE        6.50 usec
TE        300.0 K
D1        2.00000000 sec
D11       0.03000000 sec
TD0       1
  
```

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===== CHANNEL f1 =====
NUC1      13C
P1        9.80 usec
PL1       -0.50 dB
PL1W      34.66611099 W
SFO1      75.4778101 MHz

===== CHANNEL f2 =====
CDPRG2    waltz16
NUC2      1H
PCPD2     100.00 usec
PL2       2.50 dB
PL12      19.20 dB
PL13      23.00 dB
PL2W      8.07971478 W
PL12W     0.17274120 W
PL13W     0.07201053 W
SFO2      300.1432006 MHz
SI        32768
SF        75.4702658 MHz
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LB        1.00 Hz
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PC        1.40
  
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Figure S9: ^{13}C NMR spectra of L2 in CDCl_3

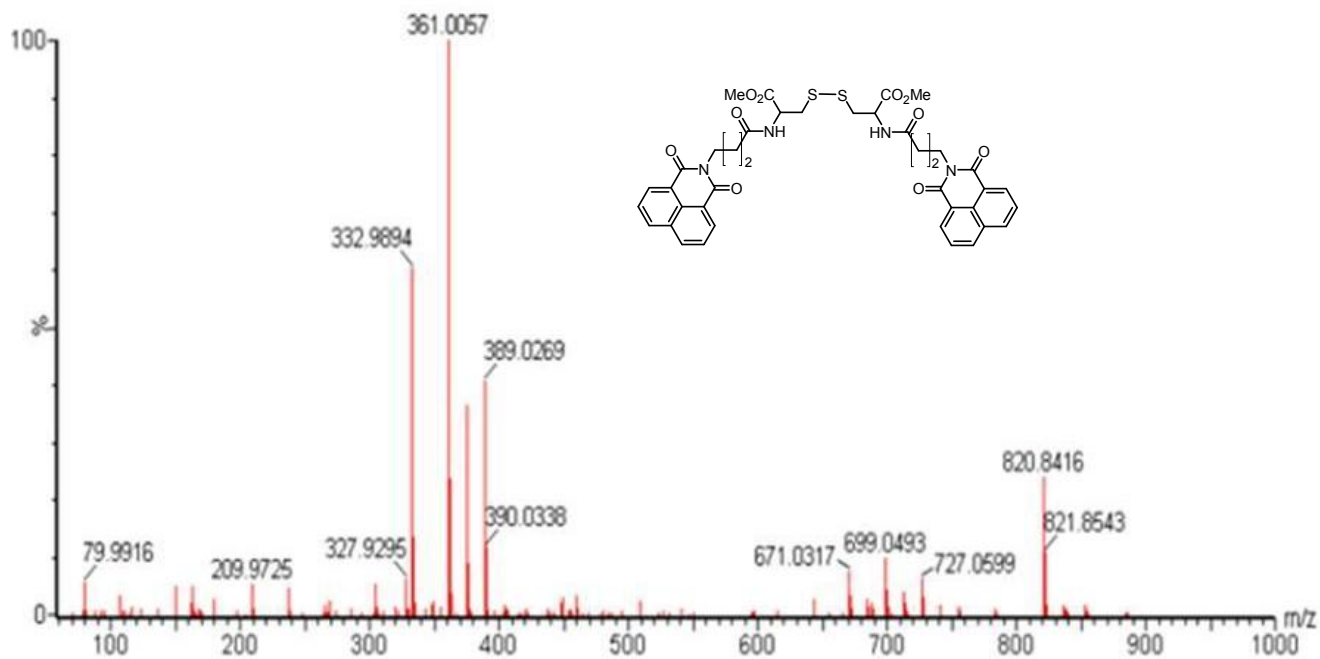


Figure S10: ES-MS of L2 ($m/z = 821.85$, calc. for $M+Na^+$)

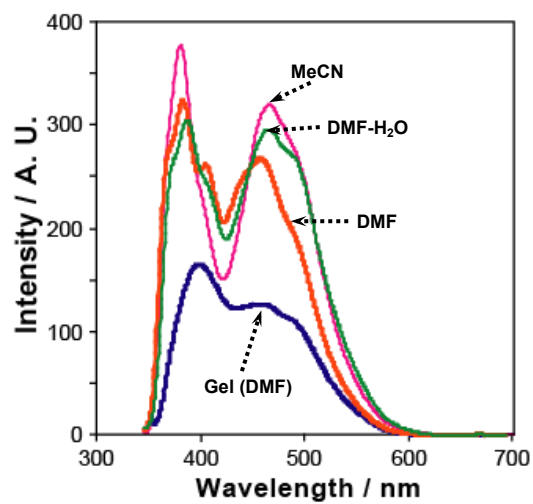


Figure S11: Solvent-dependent fluorescence emission spectra of L2 (λ_{ex} 340nm).

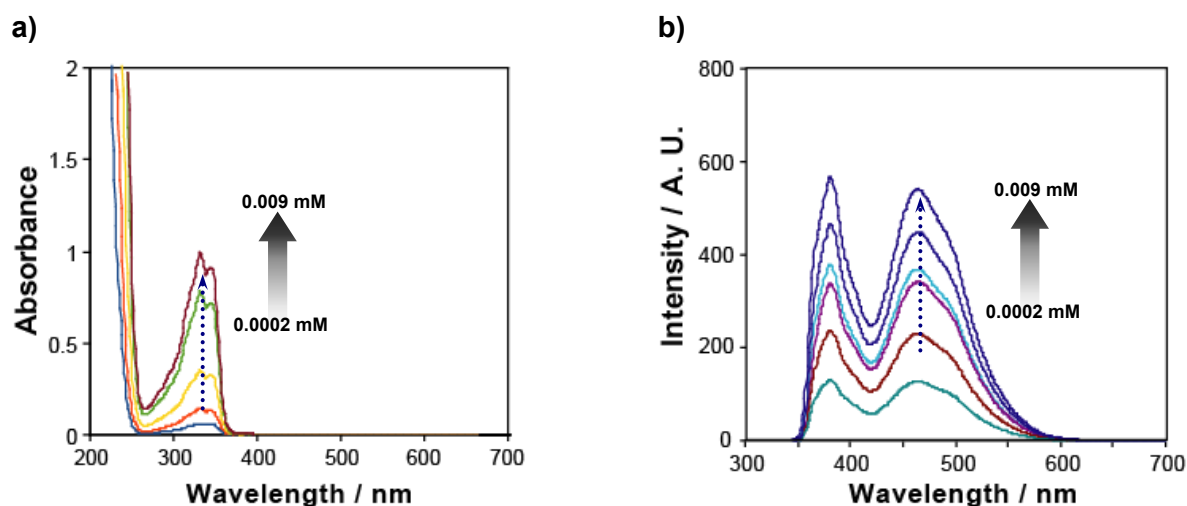


Figure S12: Concentration-dependent spectra of **L2** (in MeCN): (a) UV-visible, and (b) fluorescence emission (λ_{ex} 340nm); Note the monomer/excimer emission for **L2**.

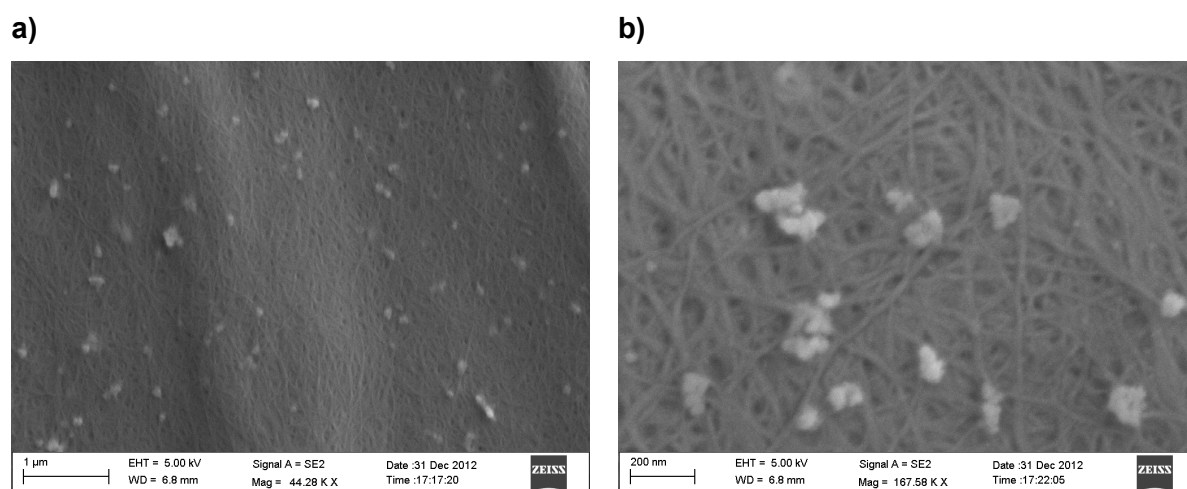


Figure S13. (a), (b) SEM images of the dried organogel obtained from **L1** in DMF (5mg/mL; the particles were initially formed due to spontaneous aggregation of the **L1** gelator (as tiny nucleation sites) while the formation of fibres was relatively slow (ageing 2-3 days).

Such features were not observed for **L2** in DMF, except at high concentration (>12mg/mL). It appears that the gelation of acetonitrile and DMF by **L1** and **L2** occurs due to hydrogen bonding between the amide NH groups and the solvent (immobilisation of the solvent molecules), with concomitant hydrophobic packing (stacking interactions) of the bis-naphthalimide motifs, which result in the formation of networks of self-assembled fibres.

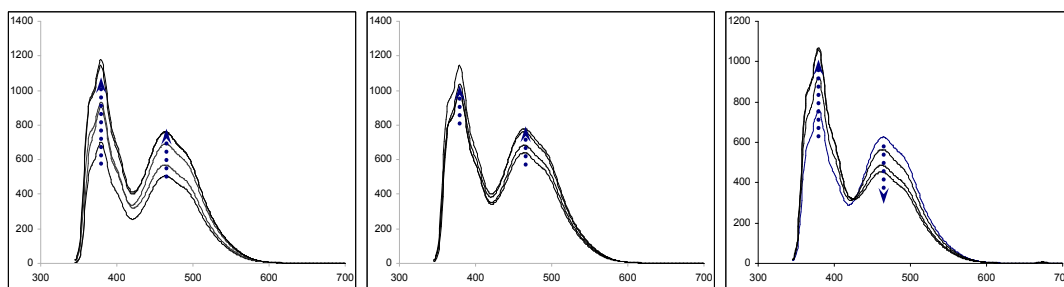


Figure S14. Fluorescence titration of **L1** with chloride, bromide and acetate anions in acetonitrile (10% water).

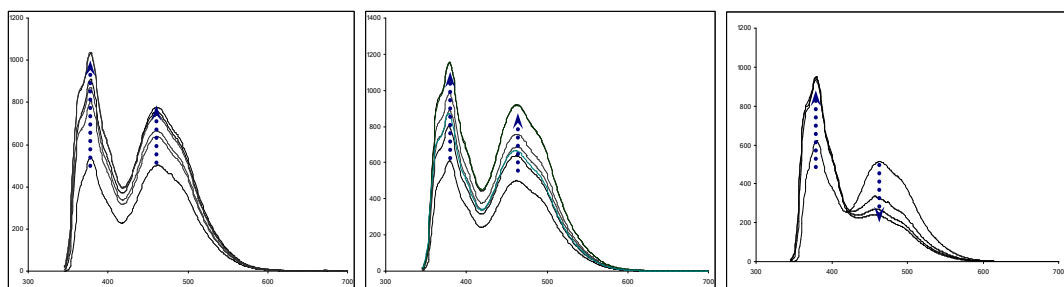


Figure S15. Fluorescence titration of **L2** with chloride, bromide and acetate anions in acetonitrile (10% water).

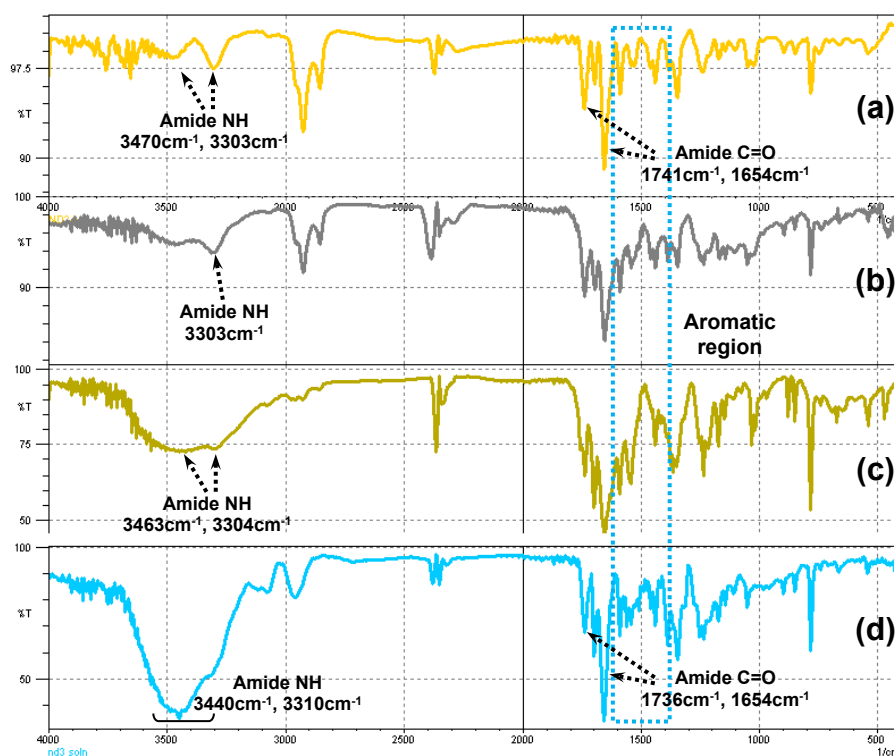


Figure S16. Comparative study of FT-IR spectra of **L2** under various conditions: (a) 0.1mg/mL in chloroform; (b) 1.0mg/mL in chloroform; (c) dried **L2**/acetonitrile gel, after removal of solvent; (d) **L2**/acetonitrile gel, when suspended in chloroform; the absorptions at 3303cm^{-1} in dilute solutions and in the aromatic region were distinct, of which the latter was apparently affected by changes in gelator concentration.

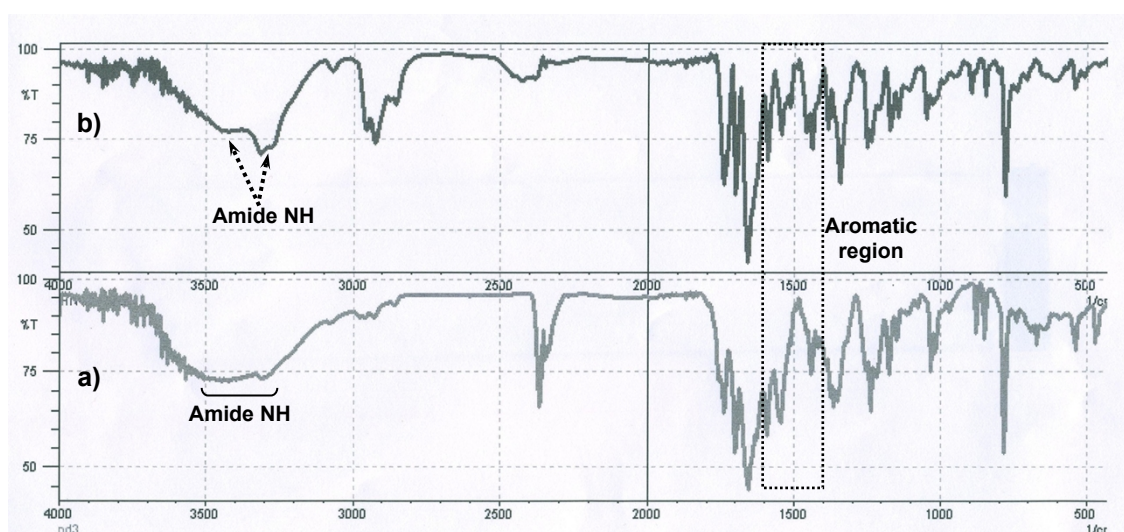


Figure S17. FTIR spectra of (a) **L2**/acetonitrile gel, and (b) upon addition of BF_4^- anions (**L2**/ NaBF_4). Changes in the amide NH absorption and the aromatic region are noteworthy.

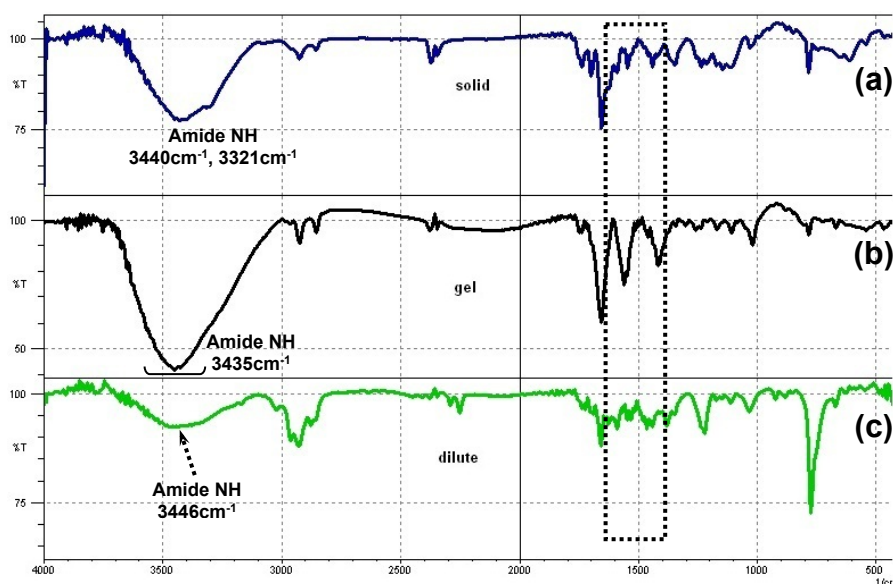


Figure S18. Comparative study of FT-IR spectra of **L1** under various conditions: (a) **L1**/DMF gel after removal of solvent; (b) **L1**/DMF, when suspended in chloroform ; (c) **L1** when suspended in acetonitrile; the absorptions at 3446 cm^{-1} in dilute solutions was affected by changes in gelator concentration.

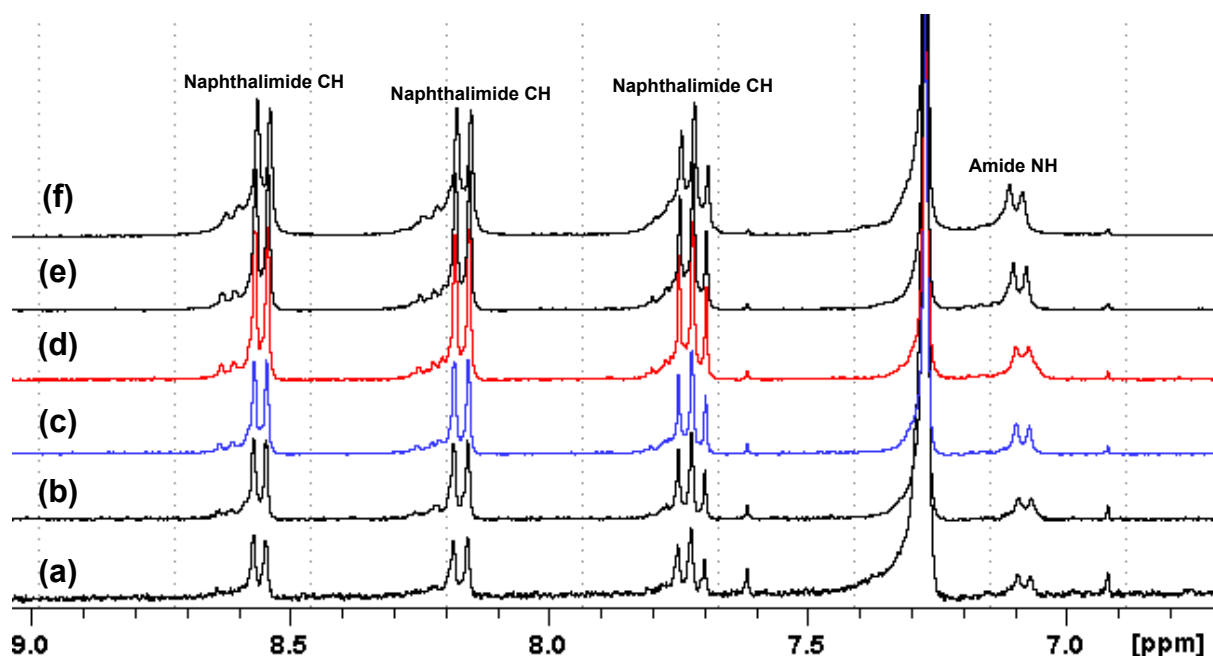


Figure S19. Concentration dependent ^1H NMR spectra of **L2** in CDCl_3 : (a) 0.3 mg/mL ; (b) 1.0 mg/mL ; (c) 2.0 mg/mL ; (d) 4.0 mg/mL ; (e) 6.0 mg/mL , with formation of partial gel; and (f) after the gelation process is complete. The amide NH resonance at 7.078 ppm in dilute solutions, i.e. $[\text{L2}] < 2\text{ mg/mL}$ remains relatively unaffected by changes in gelator concentration. Beyond the critical gelation concentration of 6 mg/mL , the amide NH resonances shifted downfield, to 7.082 ppm , such that $\Delta\delta = 0.018\text{ ppm}$, indicating that H-bonding interactions were not dominant. However, the onset of gelation did cause the naphthalimide CH resonances to shift upfield by $\sim 0.01\text{ ppm}$; for instance, from 7.723 (i.e. spectra 'a') to 7.714 ppm (i.e. spectra 'f'), and from 8.17 ppm to 8.16 ppm .

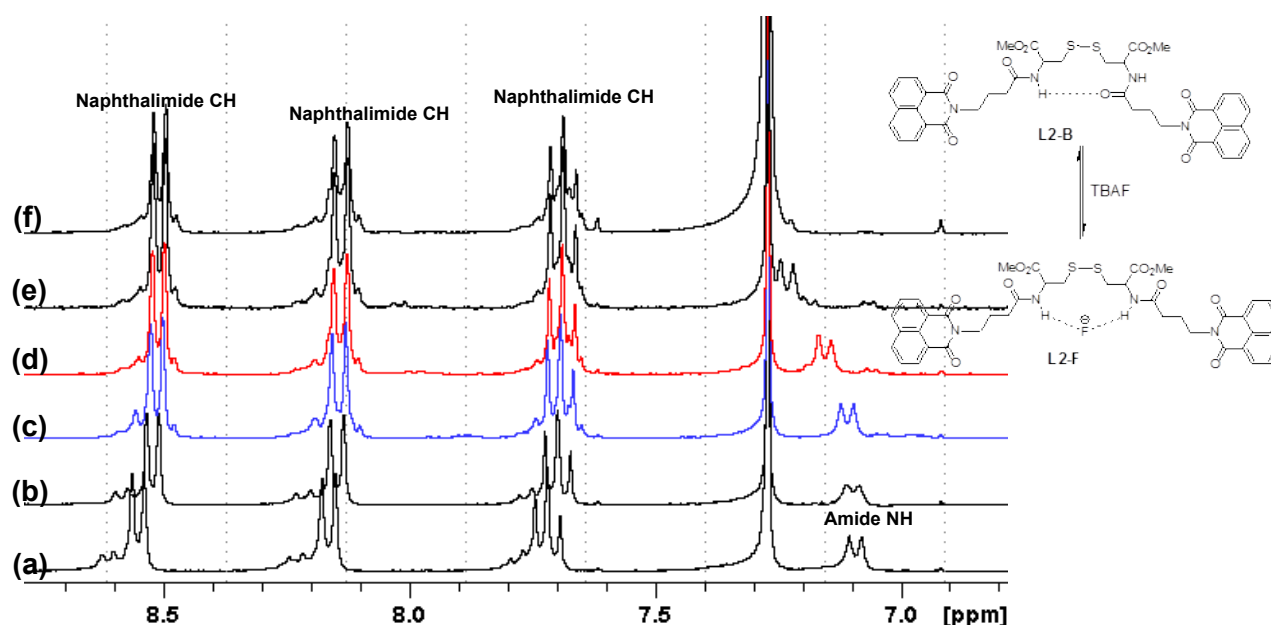


Figure S20. Partial ^1H NMR spectra showing the effect of TBAF on **L2** (3.0 mg in 0.5mL chloroform-d), during the gel-to-sol transformation; (a) Partially formed **L2**/chloroform gel; (b) **L2**/chloroform gel; (c)-(f), after TBAF addition (0.5, 1.0, 2.0 and 3.0 equiv); (Inset: Plausible hydrogen bonding interactions of **L2** with fluoride anions, i.e. TBAF, in chloroform-d)

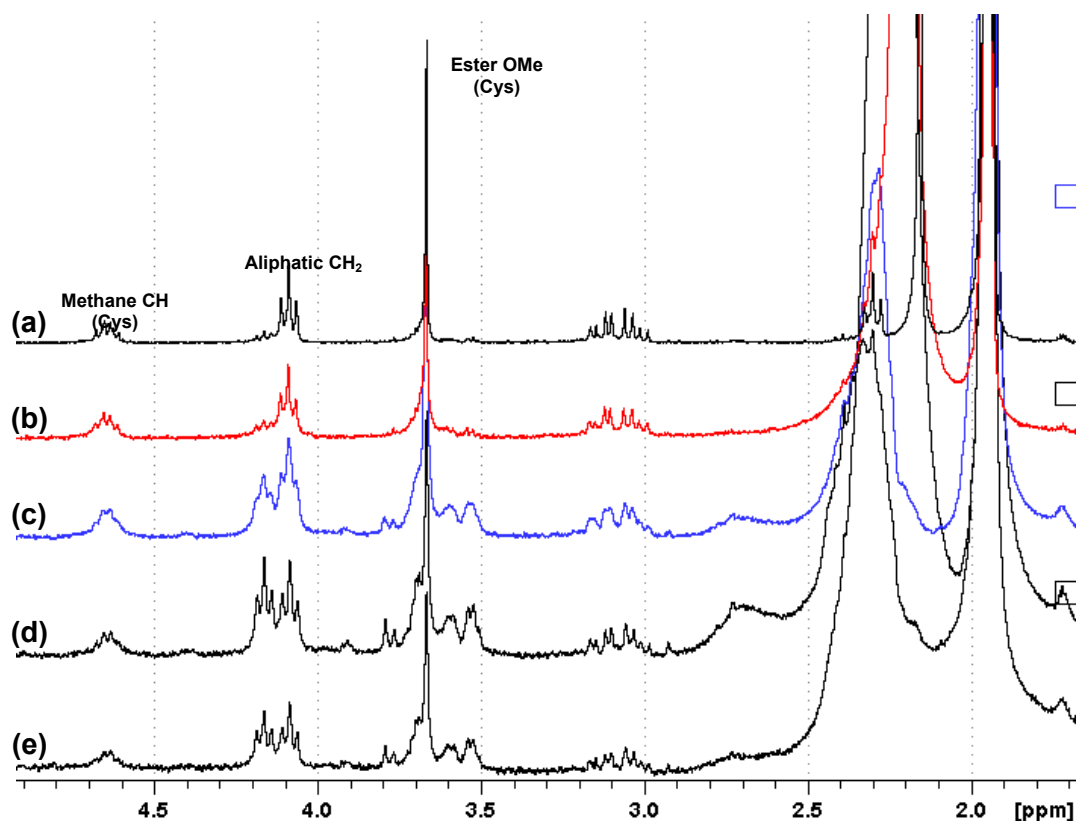


Figure S21. Partial ^1H NMR spectra of **L2** showing the temperature-dependent variations of the naphthalimide CH and amide NH resonances in acetonitrile- d_3 : (a) 50°C; (b) 45°C; (c) 35°C; (d) 25°C; (e) 25°C after 12h. Splitting/broadening of the resonance due to the ester group (Cys) also indicated multiple interactions of this residue during the gel forming process.

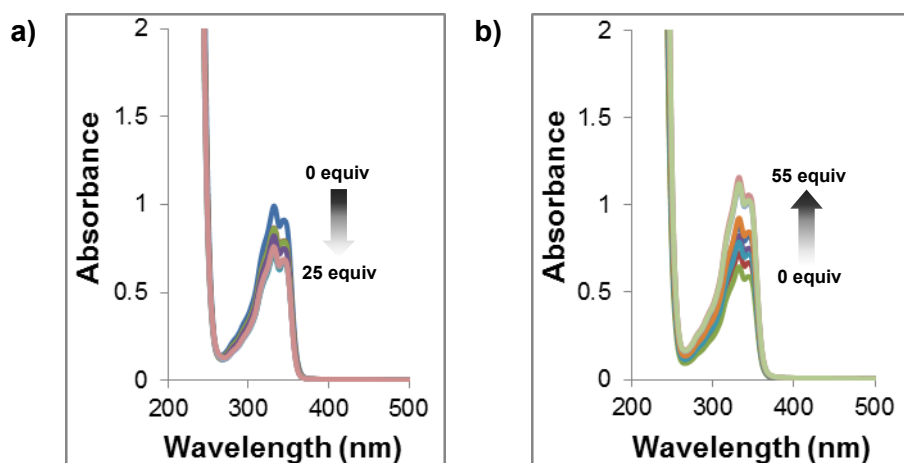


Figure S22. UV-vis spectra of **L2** in acetonitrile: (a) following the addition of fluoride anions (as TBAF, upto – equiv.); (b) and the addition of NaBF₄ (upto – equiv.); [**L2**] = 0.001mM

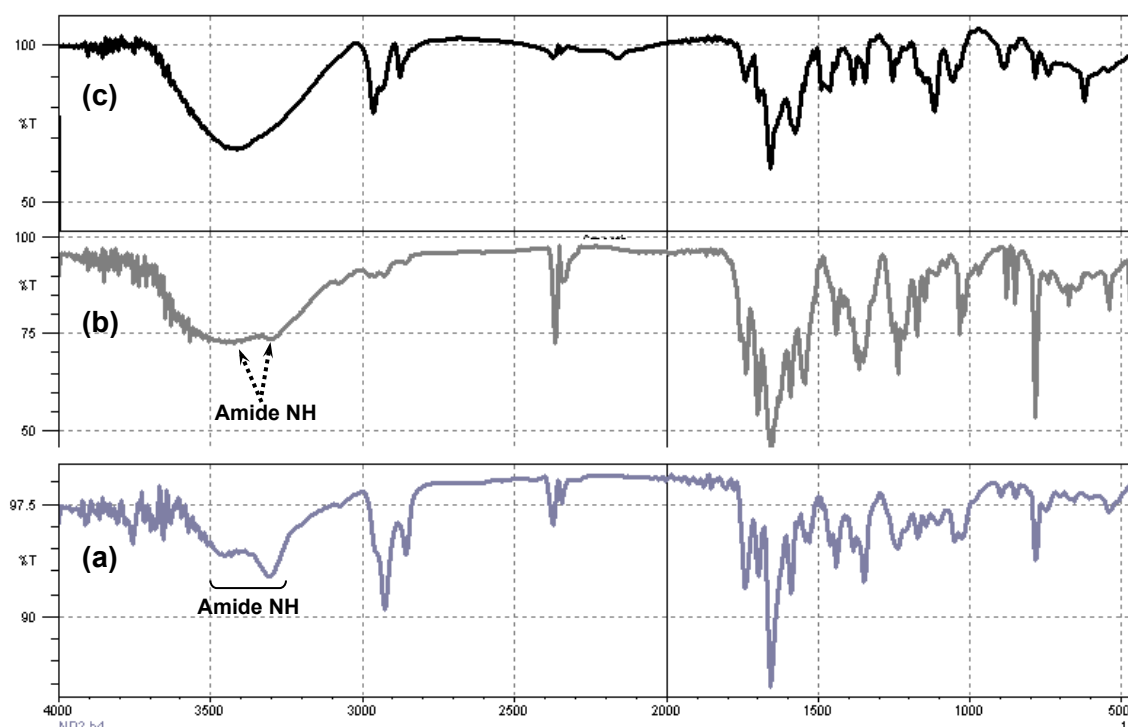


Figure S23. FTIR spectra of (a) **L2** (1.0mg/mL in chloroform); (b) **L2**/acetonitrile gel, after removal of solvent, and (c) upon addition of fluoride anions (as TBAF, ~ 20uL of 1mM stock); Changes in the amide NH absorption at 3304cm⁻¹ and the aromatic region are noteworthy.

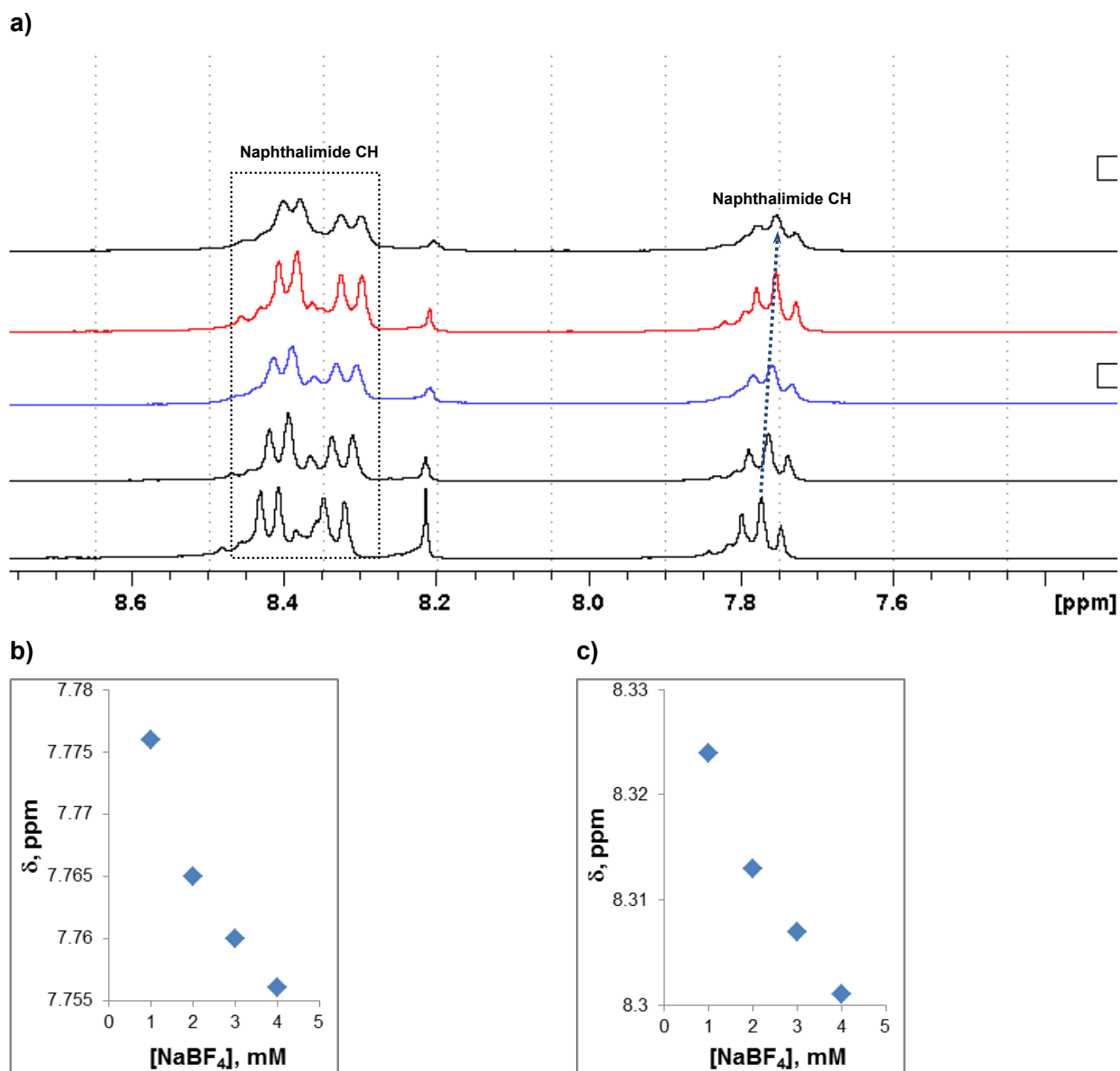


Figure S24. (a) Partial ^1H NMR spectra of **L2** (3mg in DMSO-d_6) following addition of BF_4^- anions (as NaBF_4). (b), (c) The chemical-shift changes for the naphthalimide CH resonances were noteworthy, which were indicative of anion- π interactions between the BF_4^- anions and the naphthalimide motif.