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Supporting Information

Cooperative ion pair recognition by multitopic L-ornithine based salt receptors by Piotr Piqtek, Szymon Zdanowski, Jan Romański*

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GENERAL INFORMATION

Unless specifically indicated, all other chemicals and reagents used in this study were purchased from commercial sources and used as received. Purification of products was performed using column chromatography on silica gel (Merck Kieselgel 60, 230-400 mesh) with mixtures of chloroform/methanol. Thin-layer chromatography (TLC) was performed on silica gel plates (Merck Kieselgel 60 F254).

¹H and ¹³C NMR spectra used in the characterization of products were recorded on Bruker 300 spectrometer using a residual protonated solvent as internal standard. The following abbreviations are used to indicate the multiplicity: s - singlet; d - doublet; t - triplet; q - quartet; m - multiplet, b – broad signal.

High resolution mass spectra (HRMS) were measured on a Quattro LC Micromass unit using ESI technique.

UV-vis analyses were performed using Thermo Spectronic Unicam UV500 Spectrophotometer.



NMR SPECTRA Fig. S1 and S2: ¹H and ¹³C NMR of receptor 1.

90 80 f1 (ppm)



100 90 f1 (ppm)





"S"- solvent, "G"- grease

UV-VIS EXPERIMENTS Fig. S9: Representative Uv-Vis Titration Spectra



DILUTION AND JOB PLOTS. Fig. S10: Dilution curve of receptor **1**.













Fig. S12: Job plot (Host: Receptor 2, guest: Cl-)

	Receptor 1			Receptor 2		
	TBA ⁺	Na ⁺	K _{Na} /K _{TBA}	TBA ⁺	Na ⁺	K _{Na} /K _{TBA}
NO ₂ -	3 800	7 800	2,05	7 200	18 500	1,82
Br-	3 400	5 100	1,5	3 700	4 700	1,27
Cl-*	18 200	33 500	1,84	46 800	85 500	2,57
PhCOO-	460 000	161 000	0,35	1,19.10	526 000	0,44
Ac-	3,5.10	280 000	-	deprotonation	deprotonation	-

Table S1: Association Constants.

* Association constants for interactions of receptors **4** and **5** with chloride. Receptor **4**: K_{TBACI} = 1700 M⁻¹, K_{NaCI} = 5100 M⁻¹; Receptor **5**: K_{TBACI} = 3900 M⁻¹, K_{NaCI} = 8900 M⁻¹

Fig. S13: UV-Vis titration binding isotherms of receptor **1** with TBACl and TBACl in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S14: UV-Vis titration binding isotherms of receptor **2** with TBACl and TBACl in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S15: UV-Vis titration binding isotherms of receptor **4** with TBACl and TBACl in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S16: UV-Vis titration binding isotherms of receptor **5** with TBACl and TBACl in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S17: UV-Vis titration binding isotherms of receptor **1** with TBANO₂ and TBANO₂ in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S18: UV-Vis titration binding isotherms of receptor **2** with TBANO₂ and TBANO₂ in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S19: UV-Vis titration binding isotherms of receptor **1** with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S20: UV-Vis titration binding isotherms of receptor **2** with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S21: UV-Vis titration binding isotherms of receptor **1** with PhCOOTBA and PhCOOTBA in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S22: UV-Vis titration binding isotherms of receptor **2** with PhCOOTBA and PhCOOTBA in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



Fig. S23: UV-Vis titration binding isotherms of receptor **1** with AcTBA and AcTBA in the presence of 1 equivalent of NaPF₆. Experimental data (points), fitted curve (line).



NMR TITRATIONS

The ¹H NMR titrations were performed on a Varian UnityPlus 200 MHz spectrometer, at 298K in CD₃CN. The anion TBA and cation PF6 salts were dried under high vacuum at 30-45 °C prior to use. In each case, a 500 µL of freshly prepared 2.56 or 2.66 mM solution of receptor **1** or **2** was added to a 5mm NMR tube. Where applicable the solution also contained 1 molar equivalent of sodium hexafluorophosphate. Small aliquots of 60 mM solution of tetrabutylammonium bromide salts, containing **1** or **2** at 2.56 or 2.66 mM concentration, were added and a spectrum was acquired after each addition. Titration isotherms for NH protons were fitted to a 1:1 binding model using the HypNMR 2000 program.



Fig. S24: ¹H NMR titrations of receptors **1** and **2** with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Profiles based on the chemical shift (δ , ppm) of urea H¹ proton.





Fig. S25: ¹H NMR titrations of receptors 1 and 2 with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Profiles based on the chemical shift (δ , ppm) of urea H² proton.

Fig. S26: ¹H NMR titrations of receptors **1** and **2** with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Profiles based on the chemical shift (δ , ppm) of (thio)urea H³ proton.





Fig. S27: ¹H NMR titrations of receptors **1** and **2** with TBABr and TBABr in the presence of 1 equivalent of NaPF₆. Profiles based on the chemical shift (δ , ppm) of (thio)urea H⁴ proton.

Fig S 28: Partial ¹H NMR spectra (200 MHz, 298 K) of receptor **1** upon addition of 0, 0.6, 1.1, 1.6, 2.1, 3, 3.9, 4.7, 6.5, 8, 10.6, 13,3 of TBABr.





Fig S29: Partial ¹H NMR spectra (200 MHz, 298 K) of receptor **1** in the presence of 1eq. of NaPF₆ upon addition of 0, 0.6, 1.1, 1.6, 2.1, 3, 3.9, 4.7, 6.5, 8, 10.6, 13,3 of TBABr.

Fig S30: Partial ¹H NMR spectra (200 MHz, 298 K) of receptor **2** upon addition of 0, 0.6, 1.1, 1.6, 2.1, 3, 3.9, 4.7, 6.5, 8, 10.6, 13,3 of TBABr.





Fig. S31: Partial ¹H NMR spectra (300 MHz, 298 K) of receptor **2** in the presence of 1eq. of NaPF₆ upon addition of 0, 0.6, 1.1, 1.6, 2.1, 3, 3.9, 4.7, 6.5, 8, 10.6, 13,3 of TBABr.

^{11.8 11.6 11.4 11.2 11.0 10.8 10.6 10.4 10.2 10.0 9.8 9.6 9.4 9.2 9.0 8.8 8.6 8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6} f1 (ppm)



