

Supporting Information for:

Nucleophilic activity of a linked bis{guanidine} leading to formation of a dicationic C₄N₄-heterocycle

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$[H_2C\{hpp\}_2CH_2][Cl]_2$ (**2a-H₂**)

$H_2C\{hpp\}_2$ (**1**, 1.00g, 3.44 mmol) was dissolved in CH_2Cl_2 (~ 2 mL) and left at room temperature for 3 days, during which time colourless crystals deposited from solution. Yield 0.77 g, (41 %, calculated for the bis-dichloromethane solvate). Elemental analysis calcd (%) for $C_{16}H_{28}Cl_2N_6 \cdot 2(CH_2Cl_2)$ (542.08): C 39.85, H 5.95, N 15.50; found: C 39.73, H 6.18, N 15.41. **2a-H₂**: 1H NMR (500 MHz, D_2O , 298 K): Major Isomer: δ 5.15 and 4.22 (d, $^2J_{HH} = 15.6$, exocyclic CH_2). Minor Isomer: δ 5.31 and 4.31 (d, $^2J_{HH} = 15.8$, exocyclic CH_2). The remaining methylene groups of the bicyclic framework overlap in the following regions: δ 3.43 (m, 4H, hpp- CH_2), 3.29 (m, 8H, hpp- CH_2), 3.20 (m, 4H, hpp- CH_2), 1.88 (m, 8H, hpp- CH_2). ^{13}C NMR (125 MHz, D_2O , 298 K): δ 160.8 (CN_3), 152.0 (CN_3), 74.8 (exocyclic CH_2), 68.1 (exocyclic CH_2), 47.5 (hpp- CH_2), 47.4 (hpp- CH_2), 47.2 (hpp- CH_2), 21.4 (hpp- CH_2), 21.1 (hpp- CH_2). **2a-D₂**: [2H]- NMR (76.8 MHz, D_2O , 298 K): 5.05 (br, CD_2), 4.13 (br, CD_2).

$[H_2C\{hpp\}_2CH_2][BPh_4]_2$ (**2b**)

A solution of $NaBPh_4$ (0.25 g, 0.73 mmol) in 2 mL H_2O was added *via* syringe to **2a-H₂** (0.20 g, 0.37 mmol) in 1 mL H_2O , causing the immediate precipitation of a white solid. The product was isolated by filtration and purified by slow cooling of a saturated MeCN solution, affording pure **2b** as colourless prisms. Yield 0.25 g, (72 %). Elemental analysis calcd (%) for $C_{64}H_{68}B_2N_6$ (931.56): C 81.53, H 7.21, N 8.92; found: C 81.46, H 7.25, N 8.89. 1H NMR (300 MHz, CD_3CN , 298 K): Anion resonances: δ 7.25 (br m, 16H, *o*- C_6H_5), 6.99 (t, $^3J_{HH} = 7.40$, 16H, *m*- C_6H_5), 6.83 (t, $^3J_{HH} = 7.22$, 8H, *p*- C_6H_5). Major Isomer: δ 5.06 and 4.15 (d, $^2J_{HH} = 15.7$, exocyclic CH_2). Minor Isomer: 5.17 and 4.20 (d, $^2J_{HH} = 15.9$, exocyclic CH_2). The remaining methylene groups of the bicyclic framework overlap in the following regions: δ 3.42 (m, 4H, hpp- CH_2), 3.34 (m, 8H, hpp- CH_2), 3.26 (m, 4H, hpp- CH_2), 1.93 (m, 8H, hpp- CH_2). ^{13}C NMR (75 MHz, CD_3CN , 353 K): δ 164.2 (q, $^1J_{CB} = 49.1$, *i*- C_6H_5), 161.4 (CN_3), 136.1 (C_6H_5), 125.8 (q, $J_{CB} = 2.7$, C_6H_5), 122.0 (C_6H_5), 75.4 (exocyclic CH_2), 68.7 (exocyclic CH_2), 48.1 (hpp- CH_2), 48.0 (hpp- CH_2), 47.9 (hpp- CH_2), 47.8 (hpp- CH_2), 21.9 (hpp- CH_2), 21.6 (hpp- CH_2).

General procedure for the preparation of $[H_2C\{hppR\}_2][X]_2$

A sample of **1** was dissolved in neat R–X reagent and stirred for 1-3 days. Addition of Et₂O resulted in formation of a white solid that was separated from excess RX by filtration.

General procedure for the preparation of $\{H_2C\{hppR\}_2\}[BPh_4]_2$

A solution of two equivalents of NaBPh₄ in H₂O was added to an aqueous stirred solution of $\{H_2C\{hppR\}_2\}[X]_2$. The product generally precipitated from the reaction mixture and was isolated by filtration. Purification was achieved by crystallization, as detailed below:

$[H_2C\{hppMe\}_2][I]_2$ (3**).**

Compound **3a** was purified by crystallization from MeOH at -30 °C. Yield 98 %. Elemental analysis calcd (%) for C₁₇H₃₂N₆I₂ (574.08): C 35.54, H 5.62, N 14.64; found: C 35.65, H 5.72, N 14.53. ¹H NMR (300 MHz, D₂O, 298 K): δ 4.72 (s, 2H, H₂C{hppMe}₂), 3.38 (m, 8H, hpp-CH₂), 3.20 (m, 8H, hpp-CH₂), 2.91 (s, 6H, hppMe), 1.96 (m, 8H, hpp-CH₂). ¹³C NMR (75 MHz, D₂O, 298 K): δ 158.2 (CN₃), 66.1 (H₂C{hppMe}₂), 48.9 (hpp-CH₂), 48.3 (hpp-CH₂), 48.1 (hpp-CH₂), 44.1 (hpp-CH₂), 41.6 (hppMe), 21.4 (hpp-CH₂), 20.6 (hpp-CH₂).

$[H_2C\{hppCH_2Ph\}_2][BPh_4]_2$ (4b**).**

Compound **4b** was purified by crystallization from MeOH / acetone at room temperature. Yield 65 %. Elemental analysis calcd (%) for C₇₇H₈₀B₂N₆ (1111.12): C 83.19, H 7.26, N 7.56; found: C 83.23, H 7.30, N 7.61. ¹H NMR (300 MHz, D₆-acetone, 298 K): Anion resonances: δ 7.36 (br m, 16H, *o*-C₆H₅), 6.95 (t, ³J_{HH} = 7.38, 16H, *m*-C₆H₅), 6.81 (t, ³J_{HH} = 7.19, 8H, *p*-C₆H₅). Cation resonances: δ 7.41 (m, 6H, *m*- and *p*-C₆H₅), 7.27 (d, ³J_{HH} = 7.50, 4H, *o*-C₆H₅), 5.01 (s, 2H, H₂C{hppCH₂Ph}₂), 4.44 (s, 4H, hppCH₂Ph), 3.49 (m, 12H, hpp-CH₂), 3.27 (m, 4H, hpp-CH₂), 2.14 (m, 4H, hpp-CH₂), 1.88 (m, 4H, hpp-CH₂). ¹³C NMR (75 MHz, D₆-acetone, 273K): δ 164.6 (q, ¹J_{CB} = 49.1, *i*-C₆H₅-BPh₄), 158.9 (CN₃), 136.6 (C₆H₅-BPh₄), 135.1 (CH₂Ph), 129.8 (CH₂Ph), 128.8 (CH₂Ph), 127.6 (CH₂Ph), 125.7 (q, J_{CB} = 2.8, C₆H₅-BPh₄), 122.0 (C₆H₅), 66.2 (H₂C{hppCH₂Ph}₂), 57.0 (H₂C{hppCH₂Ph}₂), 49.5 (hpp-CH₂), 48.8 (hpp-CH₂), 46.1 (hpp-CH₂), 44.1 (hpp-CH₂), 22.0 (hpp-CH₂), 20.9 (hpp-CH₂).

Reaction of $H_2C\{hpp\}_2$ with Me_2CHI

Excess Me_2CHI was added to an NMR tube containing a solution of $H_2C\{hpp\}_2$ in CD_3CN and the sample was sealed. After 15h the 1H NMR spectrum showed 100% conversion to a new guanidine containing species, with resonances at δ 6.07 (m, 1H, =CHMe), 5.29-5.13 (m, 2H, =CH₂) and 1.37 (d, J = 6.6 Hz, =CHMe) corresponding to propene.

1H NMR (300 MHz, CD_3CN , 298 K): δ 5.19 (s, 2H, $H_2C\{hpp\}\{hppH\}$), 3.63 (m, 4H, hpp-CH₂), 3.51 (m, 4H, hpp-CH₂), 3.41 (m, 8H, hpp-CH₂), 2.17 (m, 8H, hpp-CH₂).[‡] ^{13}C NMR (75 MHz, CD_3CN , 298 K): δ 152.2 (CN₃), 66.6 ($H_2C\{hpp\}\{hppH\}$), 48.2 (hpp-CH₂), 48.0 (hpp-CH₂), 47.7 (hpp-CH₂), 41.0 (hpp-CH₂), 22.5 (hpp-CH₂), 22.2 (hpp-CH₂).

([‡] resonance corresponding to the NH proton(s) not observed).

Optimized coordinates for 'chair'-conformation (red)

1	2.241790	1.229019	-2.511783
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1	1.332151	2.624797	-1.935761
1	-1.337054	2.599384	-1.965057
1	-3.847433	2.909408	-1.682427
1	3.831294	2.974299	-1.605982
6	1.993604	1.816288	-1.621033
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1	2.967622	2.955200	-0.067408
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1	1.337053	-2.599384	1.965057
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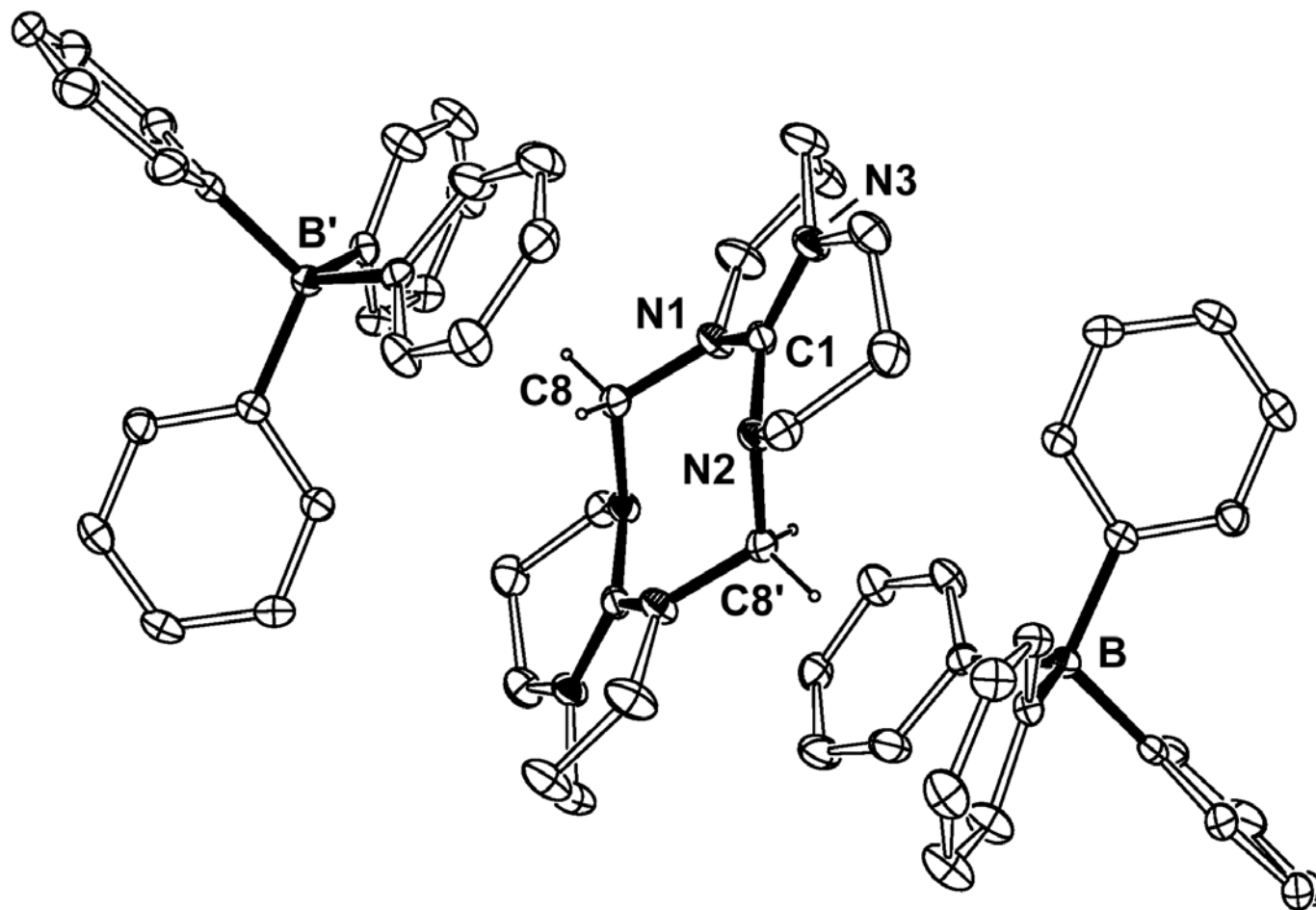
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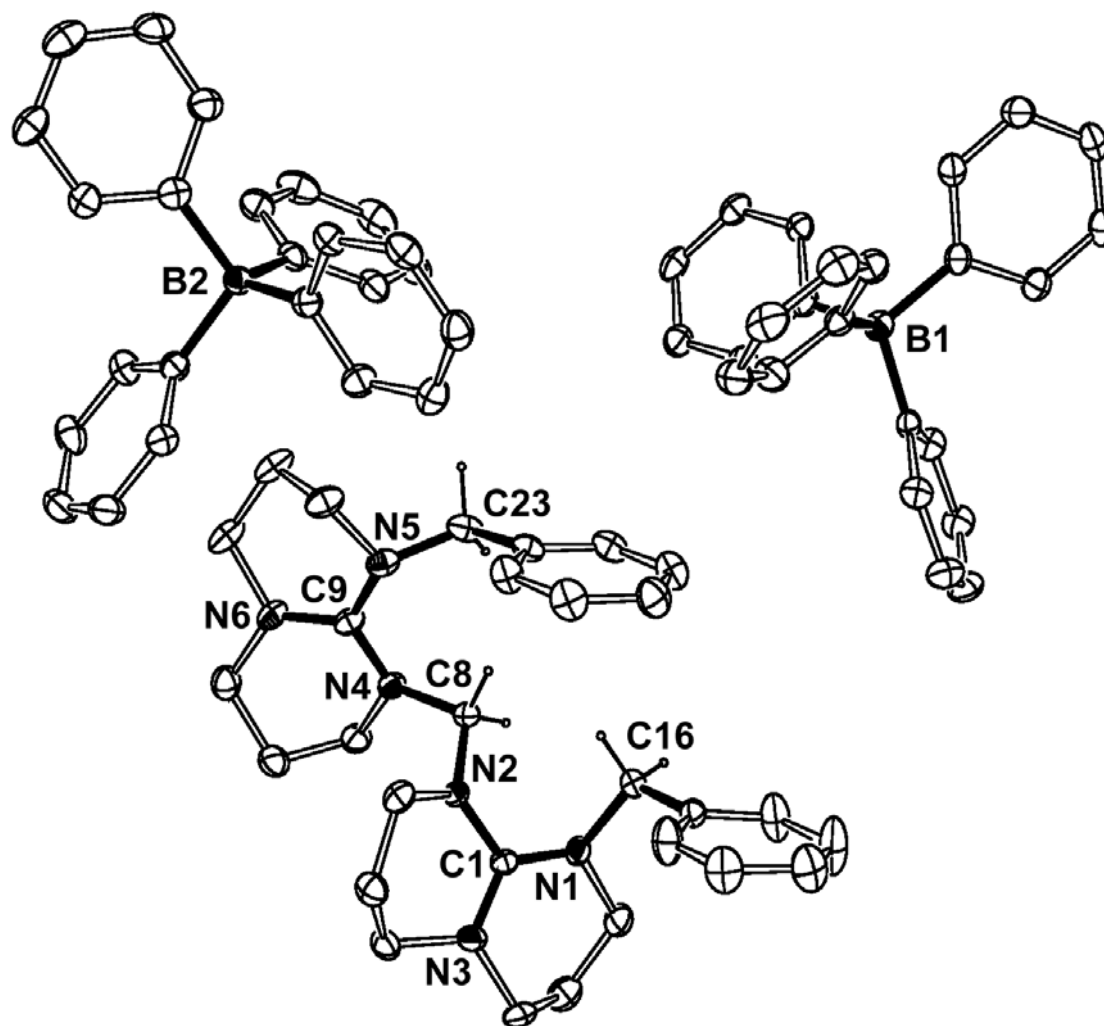
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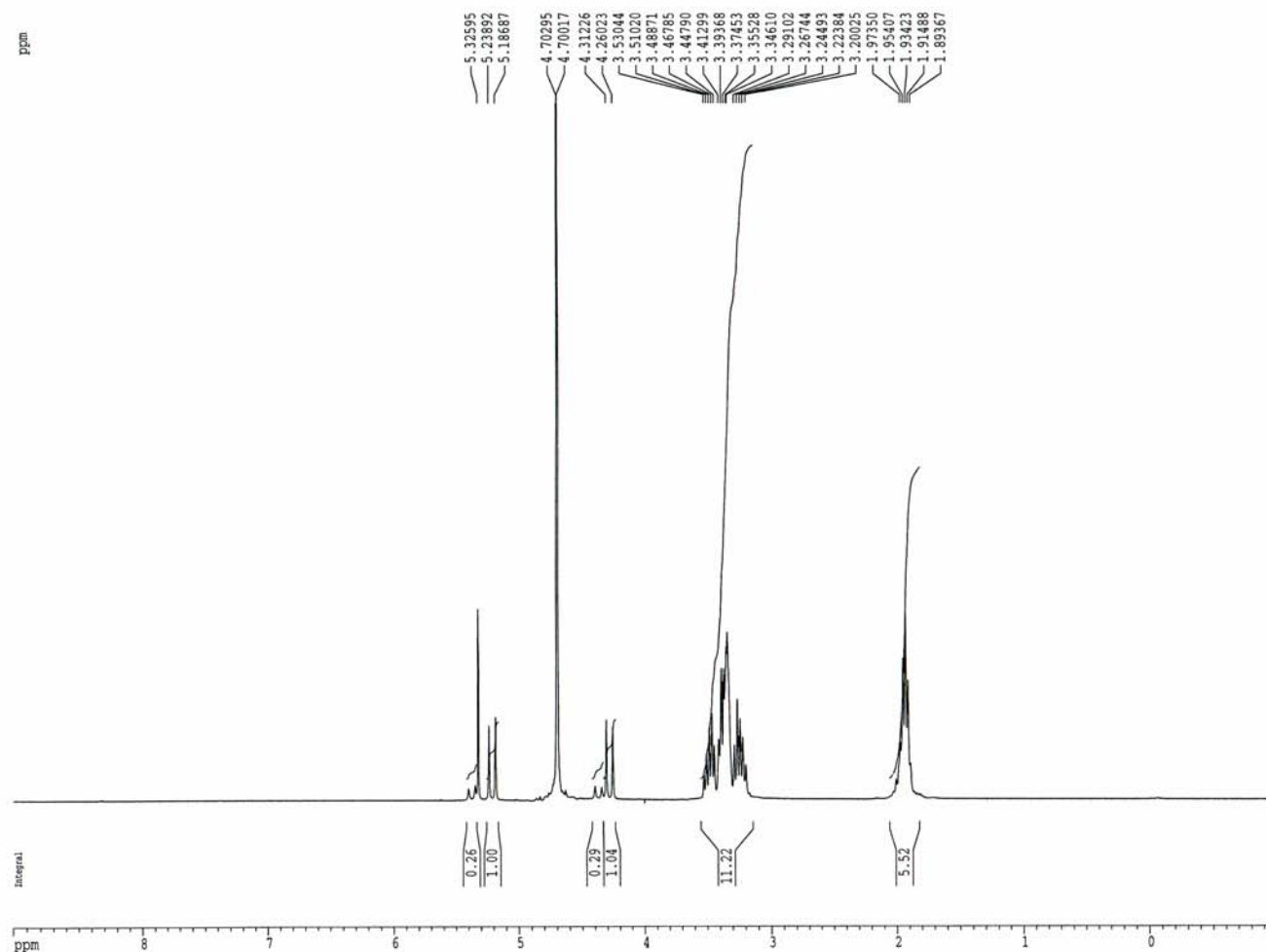
ORTEP for $[H_2C(hpp)_2CH_2][BPh_4]_2$ (2b)



ORTEP for $[H_2C\{hppCH_2Ph\}_2][BPh_4]_2$ (4b)



¹H NMR [H₂C{hpp}₂CH₂] [Cl]₂ (2a-H₂): D₂O, 298K, 300 MHz



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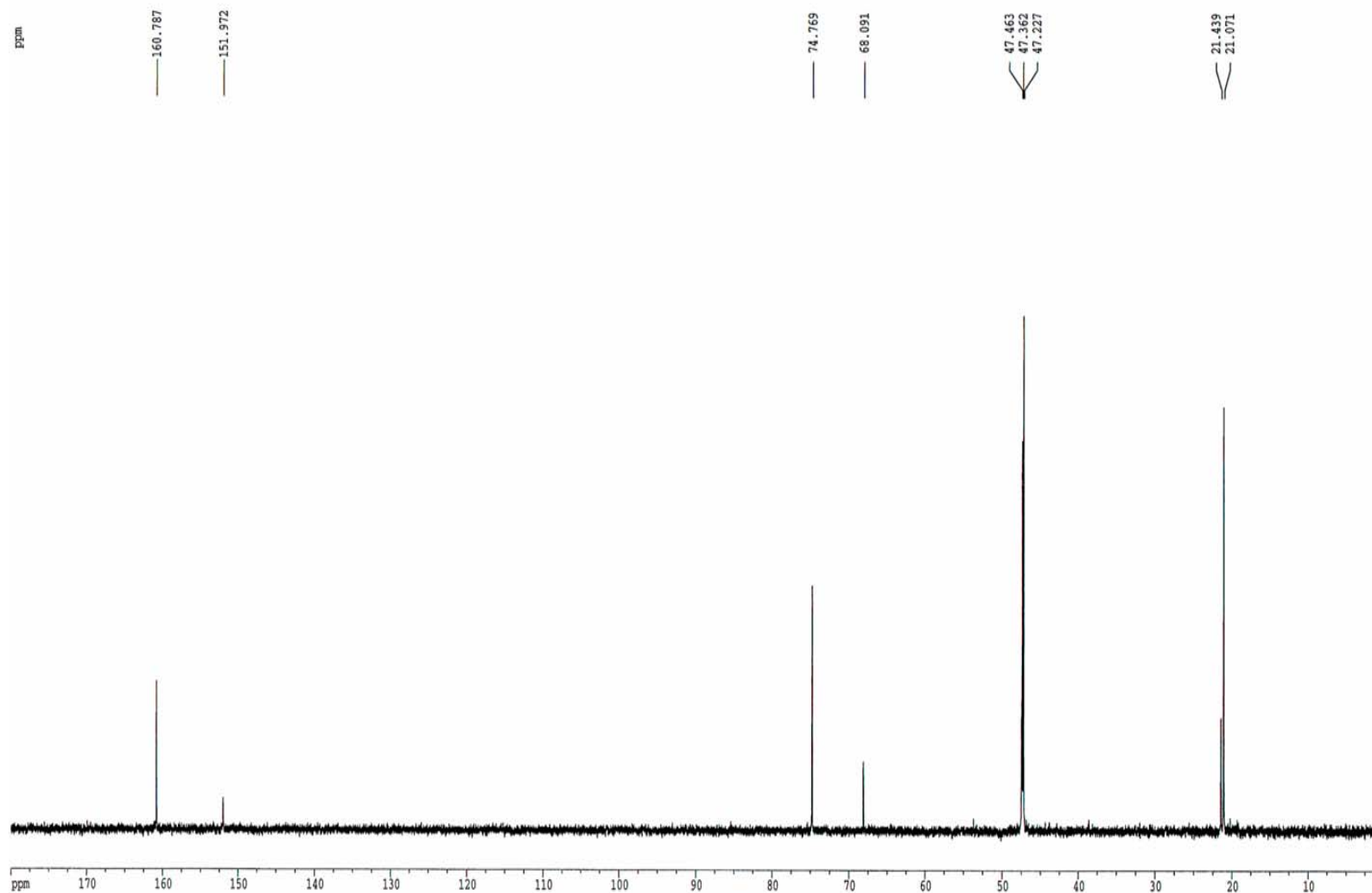
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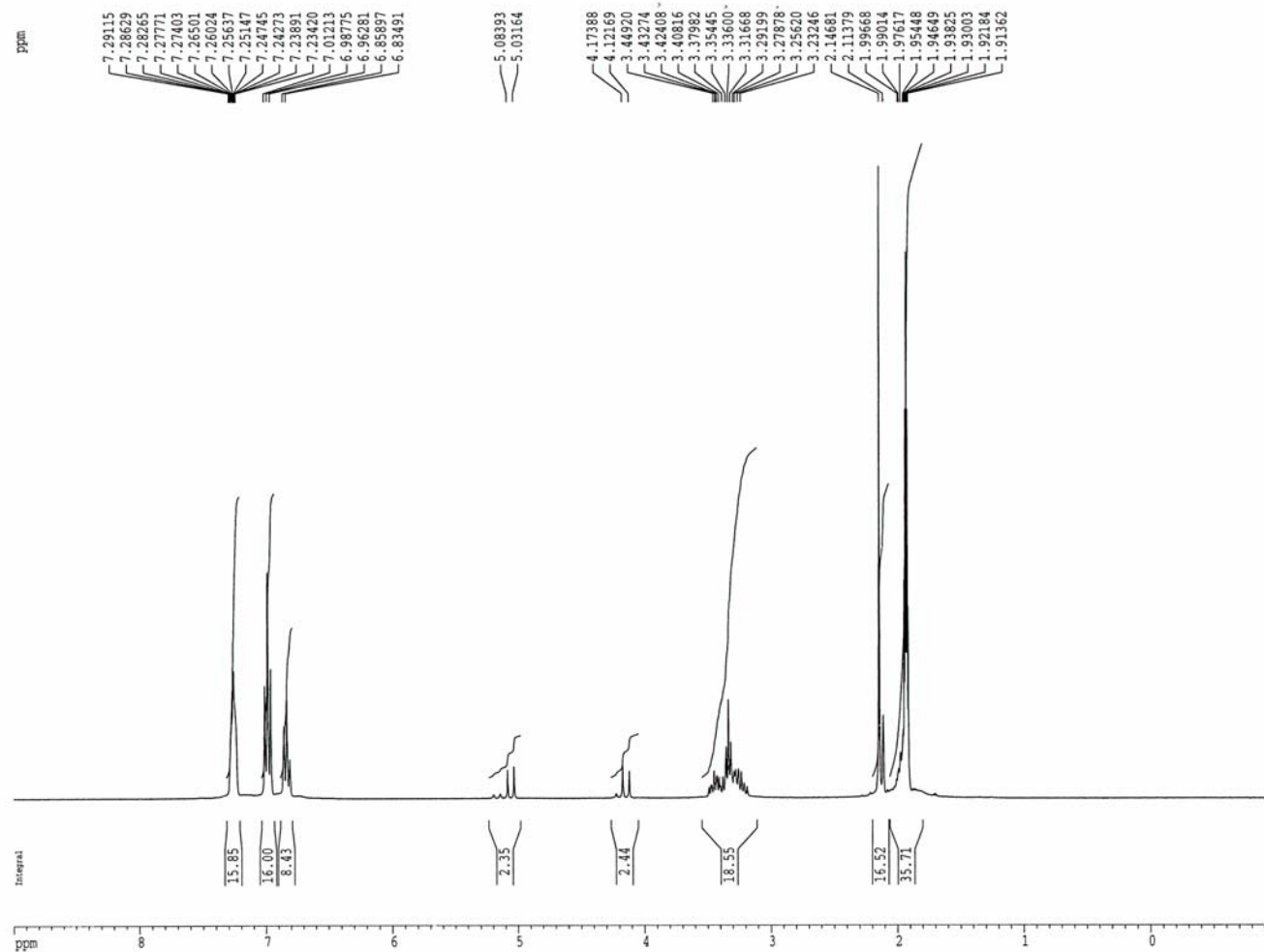
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^{13}C NMR [$\text{H}_2\text{C}\{\text{hpp}\}_2\text{CH}_2$] [Cl] $_2$ (2a-H $_2$): D_2O , 298K, 125 MHz



^1H NMR $[\text{H}_2\text{C}\{\text{hpp}\}_2\text{CH}_2]$ $[\text{BPh}_4]_2$ (2b): CD_3CN , 298K, 300 MHz



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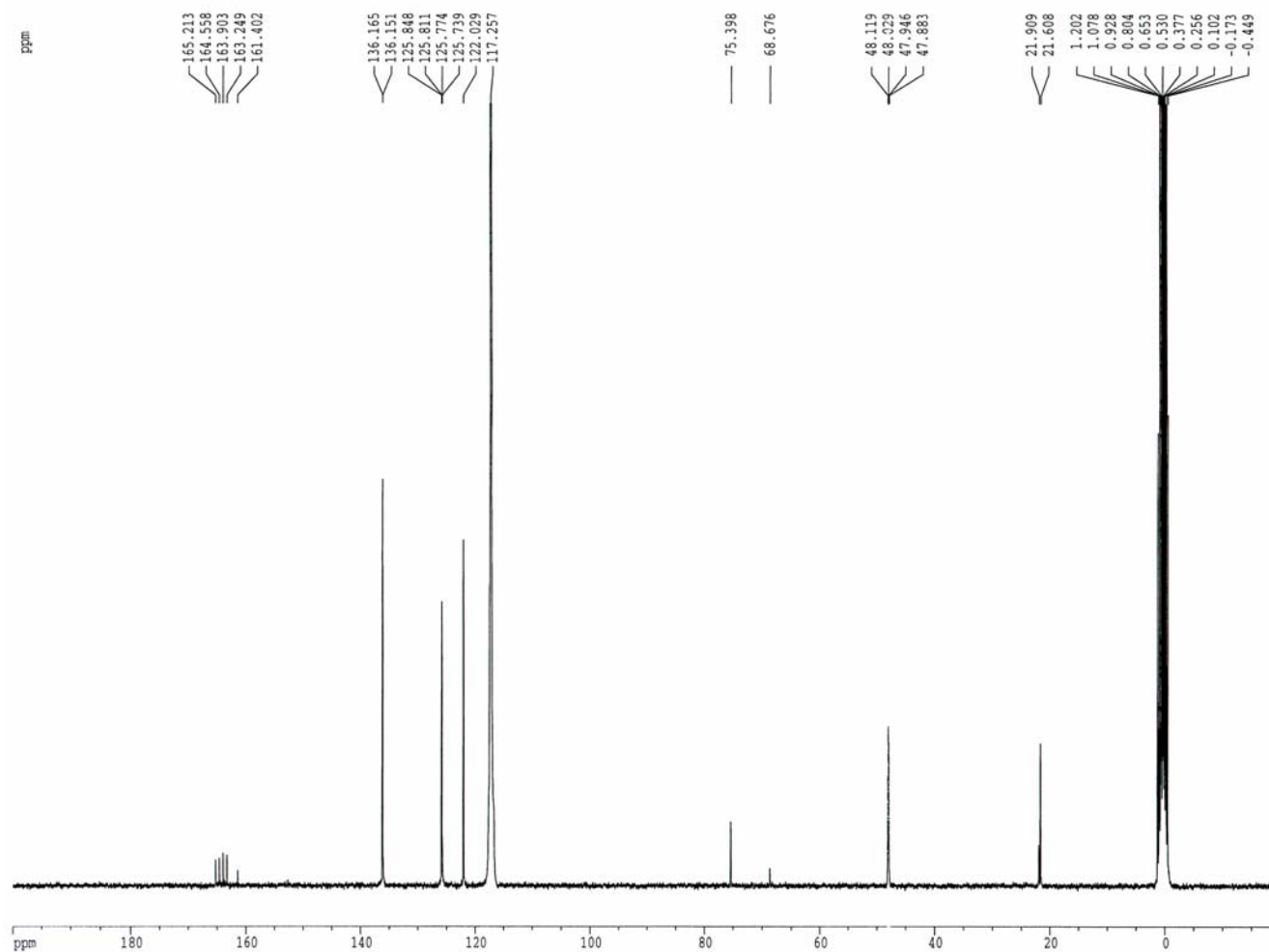
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^{13}C NMR [$\text{H}_2\text{C}\{\text{hpp}\}_2\text{CH}_2$] [BPh_4] $_2$ (2b): CD_3CN , 333K, 75 MHz



Current Data Parameters
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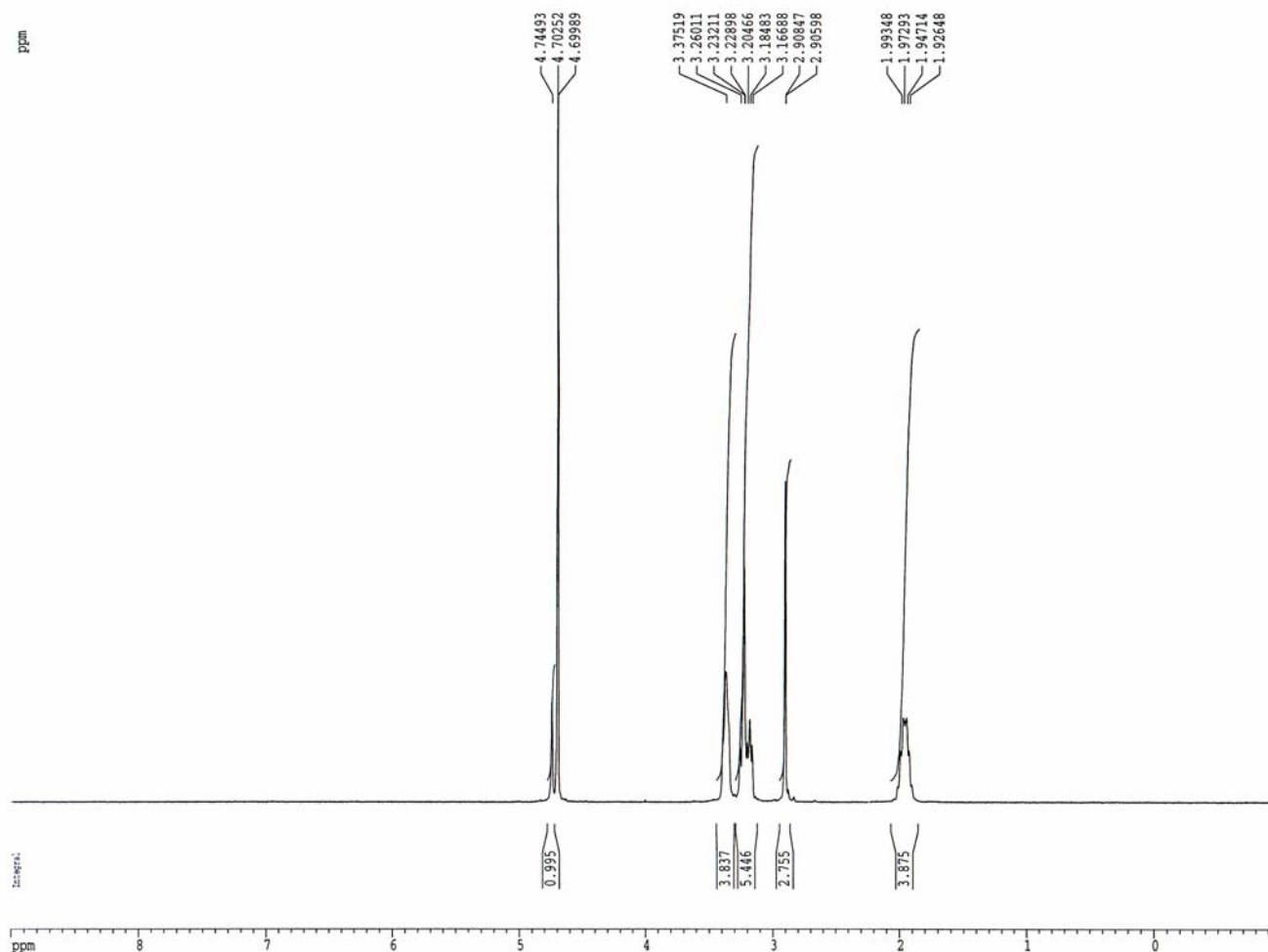
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 CX 30.00 cm
 FIP 200.000 ppm
 F1 15093.55 Hz
 F2P -20.000 ppm
 F2 -1509.35 Hz
 PPMCM 7.33333 ppm/cm
 HZCM 553.42999 Hz/cm

¹H NMR [H₂C{hppMe}₂] [I]₂ (3): D₂O, 298K, 300 MHz



Current Data Parameters
NAME MPC2004
EXPNO 1
PROCNO 1

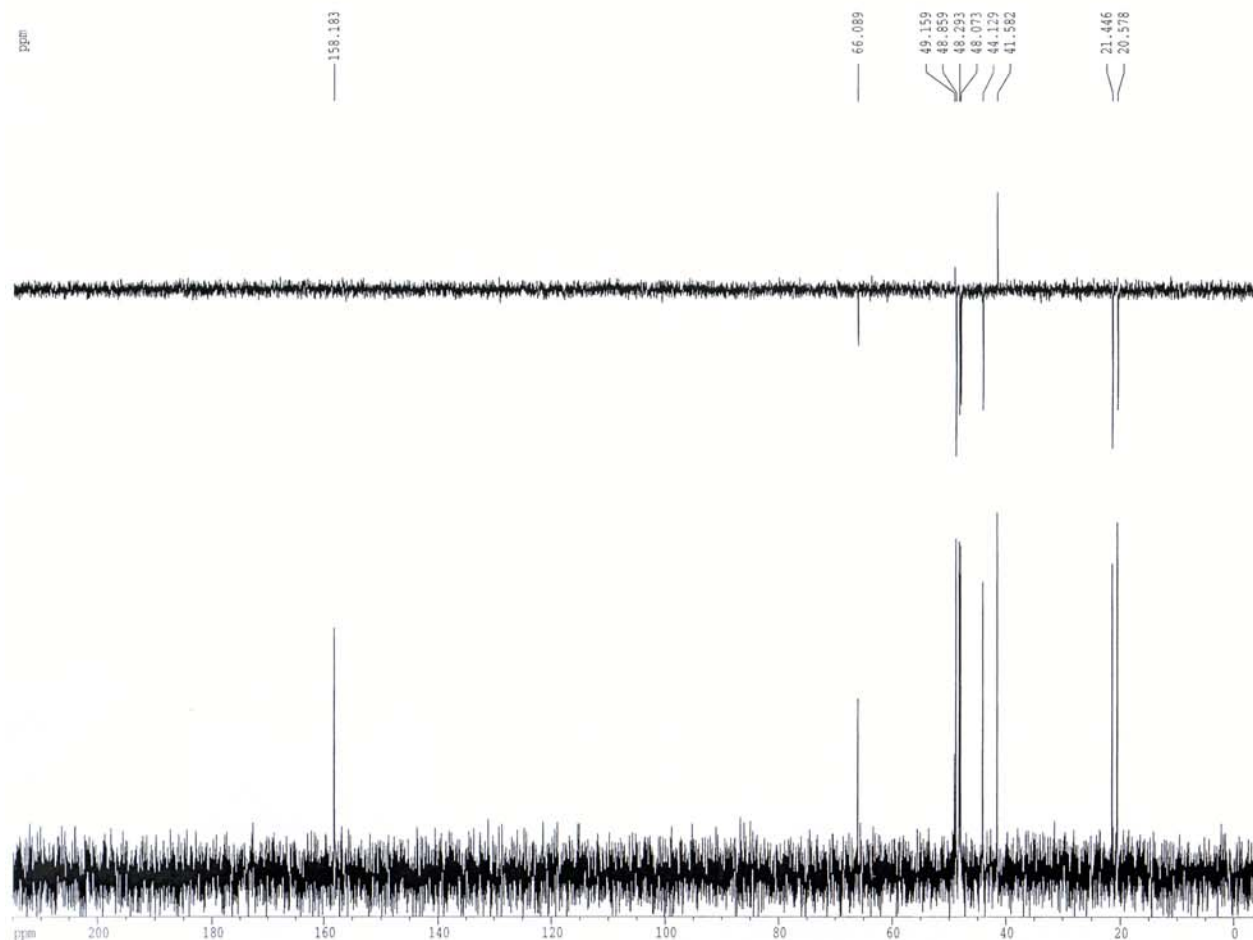
F2 - Acquisition Parameters
Date_ 20041217
Time 12.07
INSTRUM spect
PROBHD 5mm QUAD 1H/13
PULPROG zg30
TD 32768
SOLVENT C6D6
NS 8
DS 2
SWH 3612.717 Hz
FIDRES 0.110251 Hz
AQ 4.5351410 sec
RG 912.3
DW 138.400 usec
DE 5.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.50 usec
PL1 -6.00 dB
SFO1 300.1312000 MHz

F2 - Processing parameters
SI 16384
SF 300.1299971 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 30.00 cm
FLP 9.000 ppm
F1 2701.17 Hz
F2P -1.000 ppm
F2 -300.13 Hz
PPMCM 0.33333 ppm/cm
HZCM 100.04333 Hz/cm

¹³C NMR [H₂C{hppMe₂}₂] [I]₂ (3): D₂O, 298K, 75 MHz (+ DEPT 135)



```

Current Data Parameters
NAME          Martyn
EXPNO         11
PROCNO        1

F2 - Acquisition Parameters
Date_         20041217
Time          17.06
INSTRUM       spect
PROBHD        dual

PULPROG       zgpg30
TD            65536
SOLVENT       D2O
NS            256
DS            4
SWH           18832.393 Hz
FIDRES        0.287360 Hz
AQ            1.7400308 sec
RG            8192
DW            26.550 usec
DE            6.00 usec
TE            300.0 K
D1            2.00000000 sec
D11           0.03000000 sec
D12           0.00002000 sec

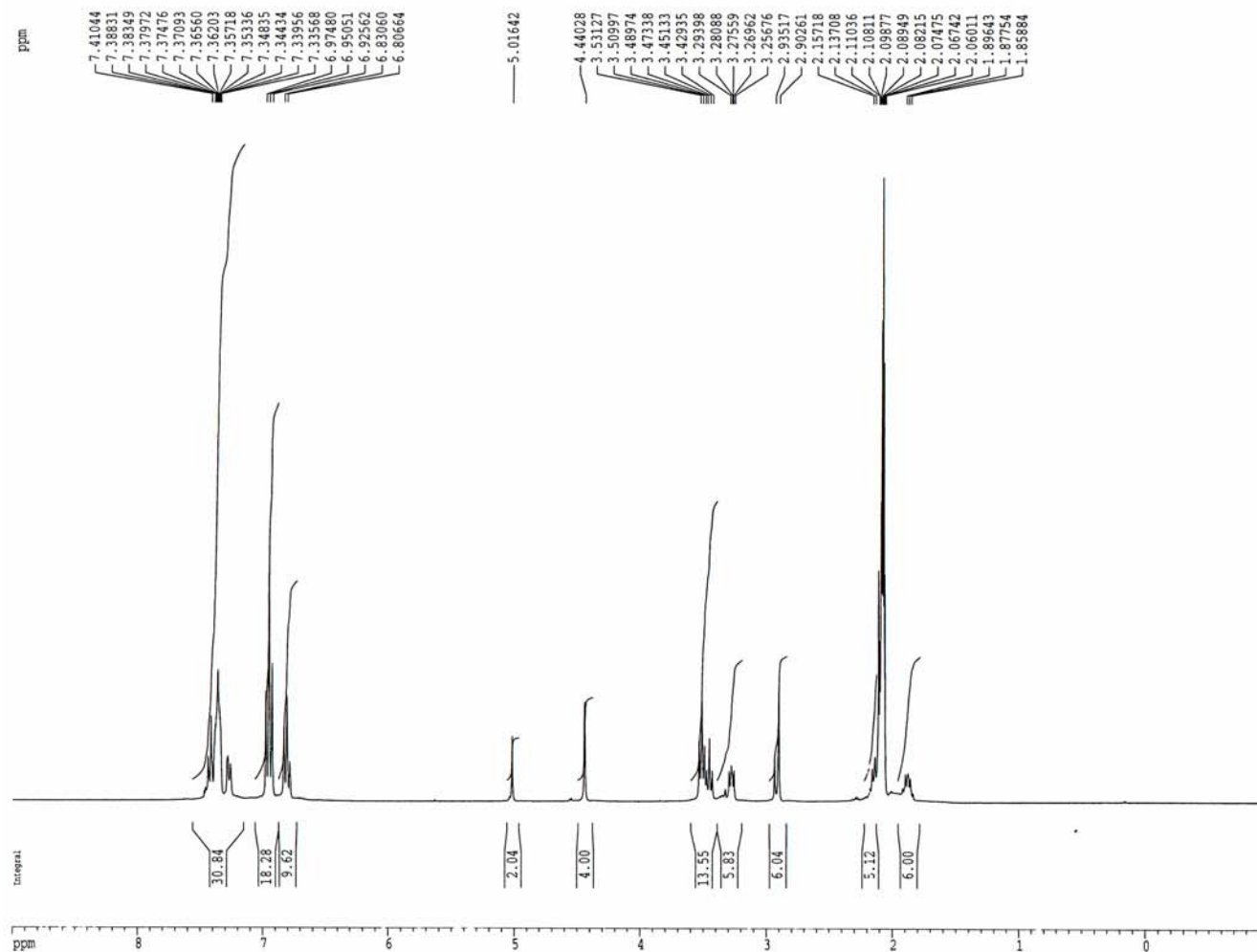
===== CHANNEL f1 =====
NUC1          13C
P1            5.50 usec
PL1           -3.00 dB
SF01          75.4760200 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         100.00 usec
PL2           -6.00 dB
PL12          16.00 dB
PL13          18.00 dB
SFO2          300.1312005 MHz

F2 - Processing parameters
SI            32768
SF            75.4677190 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40

1D NMR plot parameters
CX            30.00 cm
FIP           215.000 ppm
F1            16225.56 Hz
F2P           -5.000 ppm
F2            -377.34 Hz
PPMCM         7.33333 ppm/cm
HZCM          553.42993 Hz/cm
    
```

¹H NMR [H₂C{hppCH₂Ph}₂] [BPh₄]₂ (4b): D₆-acetone, 298K, 300 MHz



```

Current Data Parameters
NAME      MPC523e
EXPNO    1
PROCNO   1

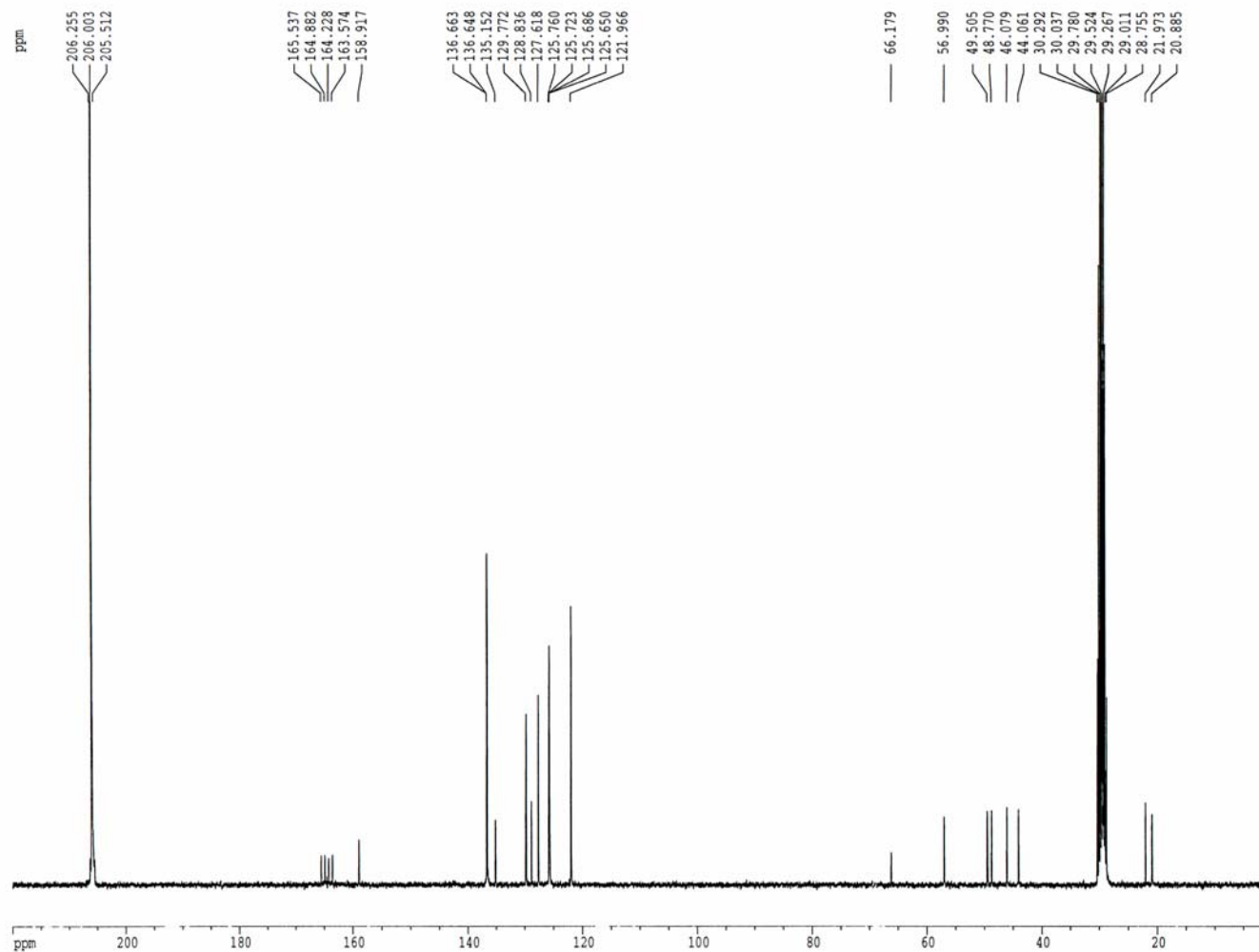
F2 - Acquisition Parameters
Date_    20060709
Time     12.29
INSTRUM  spect
PROBHD   5mm QUAD 1H/13
PULPROG  zg30
TD       32768
SOLVENT  C6D6
NS       8
DS       2
SWH      3612.717 Hz
FIDRES   0.110251 Hz
AQ       4.5351410 sec
RG       143.7
DW       138.400 usec
DE       5.50 usec
TE       300.0 K
D1       1.000000000 sec

===== CHANNEL f1 =====
NUC1     1H
P1       10.50 usec
PL1      -6.00 dB
SFO1    300.1312000 MHz

F2 - Processing parameters
SI       16384
SF       300.1299966 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       30.00 cm
F1P      9.000 ppm
F1       2701.17 Hz
F2P      -1.000 ppm
F2       -300.13 Hz
PPMCM    0.33333 ppm/cm
HZCM     100.04333 Hz/cm
    
```


^{13}C NMR [$\text{H}_2\text{C}\{\text{hppCH}_2\text{Ph}\}_2$] [BPh_4] $_2$ (4b): D_6 -acetone, 298K, 75 MHz



```

Current Data Parameters
NAME      MPC523e
EXPNO    13
PROCNO   1

F2 - Acquisition Parameters
Date_    20060709
Time     12.35
INSTRUM  spect
PROBHD   5mm QUAD 1H/13
PULPROG  zgpg30
TD       65536
SOLVENT  C6D6
NS       2509
DS       2
SWH      19267.822 Hz
FIDRES   0.294004 Hz
AQ       1.7007092 sec
RG       1625.5
DW       25.950 usec
DE       5.50 usec
TE       300.0 K
D1       1.00000000 sec
D11      0.03000000 sec
D12      0.00002000 sec

===== CHANNEL f1 =====
NUC1     13C
P1       5.50 usec
PL1      -6.00 dB
SFO1     75.4752690 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    100.00 usec
PL2      -6.00 dB
PL12     17.00 dB
PL13     19.00 dB
SFO2     300.1312005 MHz

F2 - Processing parameters
SI       32768
SF       75.4677084 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40

1D NMR plot parameters
CX       30.00 cm
FLP      220.000 ppm
F1       16602.89 Hz
F2P      0.00 ppm
F2       0.00 Hz
PFMCM    7.33333 ppm/cm
HZCM     553.42981 Hz/cm
    
```