Electronic Supplementary Information

Synthesis of apiose-containing oligosaccharide fragments of the plant cell wall: fragments of rhamnogalacturonan-II side chains A and B, and apiogalacturonan

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¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) data for protected saccharide derivatives and oligosaccharides $1-4$ (in D ₂ O)
1-O-Acetyl-3-C-acetoxymethyl-2,3-O-isopropylidene-β-D-erythrofuranose (8)
p-Tolyl 3- <i>C</i> -acetoxymethyl-2,3- <i>O</i> -isopropylidene-1-thio- α -D-erythrofuranoside (9a)
p-Tolyl 3- <i>C</i> -acetoxymethyl-2,3- <i>O</i> -isopropylidene-1-thio-β-D-erythrofuranoside (9b)
Methyl (methyl 3,4- O -isopropylidene- α -D-galactopyranosid)uronate (12)
Methyl (3- <i>C</i> -acetoxymethyl-2,3- <i>O</i> -isopropylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 3,4- <i>O</i> -isopropylidene- α -D-galactopyranosid)uronate (13)
Methyl (2,3- <i>O</i> -isopropylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 3,4- <i>O</i> -isopropylidene- α -D-galactopyranosid)uronate (14)
Methyl (4- <i>O</i> -acetyl-2,3- <i>O</i> -carbonyl- β -L-rhamnopyranosyloxymethyl)-(1 \rightarrow 3)- <i>C</i> -(2,3- <i>O</i> -isopropylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 3,4- <i>O</i> -isopropylidene- α -D-galactopyranosid) uronate (16)
Methyl (β -L-rhamnopyranosyloxymethyl)-(1 \rightarrow 3)- <i>C</i> -(2,3- <i>O</i> -isopropylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 3,4- <i>O</i> -isopropylidene- α -D-galactopyranosid)uronate (17) S12
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Methyl (methyl 3,4-O-(1-ethoxyethylidene)-α-D-galactopyranosid)uronate (24) S15
Methyl (methyl 2-O-benzoyl-3,4-O-(1-ethoxyethylidene)-α-D-galactopyranosid)uronate (25) S16
Methyl (methyl 4-O-acetyl-2-O-benzoyl-α-D-galactopyranosid)uronate (26)
Methyl (metyl 4-O-acetyl-α-D-galactopyranosid)uronate (27)
Methyl (3- <i>C</i> -acetoxymethyl-2,3,di- <i>O</i> -acetyl- β -D-erythrofuranosyl)-(1 \rightarrow 3)-(methyl 4- <i>O</i> -acetyl-2-S19)

O -benzoyl- α -D-galactopyranosid)uronate (28)
$ \begin{array}{l} Methyl \ (3-C-acetoxymethyl-2,3,di-{\it O}-acetyl-\beta-D-erythrofuranosyl)-(1\rightarrow 2)-[(3-C-acetoxymethyl-2,3-di-{\it O}-acetyl-\beta-D-erythrofuranosyl)-(1\rightarrow 3)]-(methyl \ 4-{\it O}-acetyl-\alpha-D-galactopyranosid)uronate \\ ({\bf 29}) \qquad \qquad$
L-Arabinose di(<i>p</i> -tolyl)dithioacetal (30)
2,3:4,5-Di- <i>O</i> -isopropylidene-L-arabinose di(<i>p</i> -tolyl) dithioacetal (31)
2,3- <i>O</i> -Isopropylidene-L-arabinose di(<i>p</i> -tolyl) dithioacetal (32)
2,3-O-Isopropylidene-L- <i>erythro</i> -tetrodialdose di(<i>p</i> -tolyl) dithioacetal (33)
3-C-Hydroxymethyl-2,3-O-isopropylidene- D-glycero-tetrose di(p-tolyl) dithioacetal (34)
<i>p</i> -Tolyl 3-C-hydroxymethyl-2,3- <i>O</i> -isopropylidene-1-thio- α , β -D-erythrofuranoside (35a,b) S26
<i>p</i> -Tolyl 3- <i>C</i> -chloroacetoxymethyl-2,3- <i>O</i> -isopropylidene-1-thio- α -D-erythrofuranoside (36a) S27
$p-Tolyl \ 3-C-chloroacetoxymethyl-2, 3-O-isopropylidene-1-thio-\alpha-D-erythrofuranoside \ \textbf{(36b)} \ \dots \ \textbf{S28}$
<i>p</i> -Tolyl 3- <i>C</i> -chloroacetoxymethyl-1-thio- α , β -D-erythrofuranoside (37)
<i>p</i> -Tolyl 3- <i>C</i> -chloroacetoxymethyl-2,3- <i>O</i> -(S)-benzylidene-1-thio- α -D-erythrofuranoside (38a) S30
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Methyl (2,3-O-(<i>S</i>)-benzylidene-3- <i>C</i> -chloroacetoxymethyl- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 2- <i>O</i> -acetyl-3,4- <i>O</i> -isopropylidene- β -D-galactopyranosid)uronate (39)
Methyl (2,3- O -(S-benzylidene-3- C -hydroxymethyl- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 2- O -acetyl-3,4- O -isopropylidene- α -D-galactopyranosid)uronate) (40)
Methyl <i>C</i> -(4- <i>O</i> -acetyl-2,3- <i>O</i> -carbonyl- β -L-rhamnopyranosyloxymethyl)-(1 \rightarrow 3')-(2,3- <i>O</i> -(<i>S</i>)-benzylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl 2-O-acetyl-3,4-O-isopropylidene- β -D-galactopyranosid)uronate (41)
Methyl <i>C</i> -(4-O-acetyl-2,3- <i>O</i> -carbonyl- β -L-rhamnopyranosyloxymethyl)-(1 \rightarrow 3')-(β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl (2- <i>O</i> -acetyl-3,4- <i>O</i> -isopropylidene- β -D-galactopyranosid) uronate)
Methyl <i>C</i> -(β -L-rhamnopyranosyloxymethyl)-($1\rightarrow 3$)- β -D-erythrofuranosyl-($1\rightarrow 2$)-(methyl α -D-galactopyranosid)uronate (42)
Methyl <i>C</i> -(β -L-rhamnopyranosyloxymethyl)-($1\rightarrow 3$)- β -D-erythrofuranosyl-($1\rightarrow 2$)- α -D-galactopyranosiduronic acid (1)
Methyl (3- <i>C</i> -hydroxymethyl- β -D-erythrofuranosyl)-(1 \rightarrow 2)- α -D-galactopyranosiduronic acid (2) S38
Methyl (3- <i>C</i> -hydroxymethyl- β -D-erythrofuranosyl)-(1 \rightarrow 3)- α -D-galactopyranosiduronic acid (3)
Methyl (3- <i>C</i> -hydroxymethyl- β -D-erythrofuranosyl)-(1 \rightarrow 2)-[3- <i>C</i> -hydroxymethyl- β -D-erythrofuranosyl-(1 \rightarrow 3)]-(α -D-galactopyranosiduronic acid) (4)

Crystal structure analysis of Methyl *C*-(4-*O*-acetyl-2,3-*O*-carbonyl- β -L-rhamnopyranosyloxymethyl)-(1 \rightarrow 3')-(2,3-*O*-(*R*)-benzylidene- β -D-erythrofuranosyl)-(1 \rightarrow 2)-(methyl (2-*O*-acetyl-3,4-*O*-isopropylidene- α -D-galactopyranosid)uronate) (41)

Crystal data: $C_{32}H_{40}O_{17}$, CH_2Cl_2 , M = 781.6. Orthorhombic, space group $P2_12_12_1$ (no. 19), a = 11.2799(6), b = 15.1788(9), c = 21.4815(13) Å, V = 3678.0(4) Å^3. Z = 4, Dc = 1.411 g cm⁻³, F(000) = 1640, T = 140(1) K, μ (Mo-K α) = 2.5 cm⁻¹, λ (Mo-K α) = 0.71069 Å.

Crystals are beautiful, colourless prisms. One, *ca* 0.52 x 0.25 x 0.24 mm, was mounted in oil on a glass fibre and fixed in the cold nitrogen stream on an Oxford Diffraction Xcalibur-3 CCD diffractometer equipped with Mo-K α radiation and graphite monochromator. Intensity data were measured by thin-slice ω - and φ -scans. Total no. of reflections recorded, to $\theta_{max} = 27.5^{\circ}$, was 49119 of which 8410 were unique (Rint = 0.045); 7002 were 'observed' with I > 2 σ_{I} .

Data were processed using the CrysAlis-CCD and -RED (1) programs. The structure was determined by the direct methods routines in the SHELXS program (2A) and refined by full-matrix least-squares methods, on F^2 's, in SHELXL (2B). The non-hydrogen atoms were refined with anisotropic thermal parameters. Hydrogen atoms were included in idealised positions and their Uiso values were set to ride on the Ueq values of the parent carbon atoms. At the conclusion of the refinement, $wR_2 = 0.093$ and $R_1 = 0.049$ (2B) for all 8410 reflections weighted $w = [\sigma^2(F_o^2) + (0.0556P)^2]^{-1}$ with $P = (F_o^2 + 2F_c^2)/3$; for the 'observed' data only, $R_1 = 0.037$.

In the final difference map, the highest peak (*ca* $0.7 \text{ e}\text{\AA}^{-3}$) was close to Cl(2).

Scattering factors for neutral atoms were taken from reference (3). Computer programs used in this analysis have been noted above, and were run through WinGX (4) on a Dell Precision 370 PC at the University of East Anglia.

References

- 1. Programs CrysAlis-CCD and -RED, Oxford Diffraction Ltd., Abingdon, UK (2005).
- 2. G. M. Sheldrick, SHELX-97 Programs for crystal structure determination (SHELXS) and refinement (SHELXL), *Acta Crystallogr.*, (2008), A64, 112-122.
- 3. 'International Tables for X-ray Crystallography', Kluwer Academic Publishers, Dordrecht (1992). Vol. C, pp. 500, 219 and 193.
- 4. L. J. Farrugia, J. Appl. Cryst., (1999) 32, 837-838.



Fig. 1 X-Ray crystal structure of disaccharide derivative 41.

$^{1}\mathrm{H}$ NMR (400 MHz) and $^{13}\mathrm{C}$ NMR (100 MHz) data for protected saccharide derivatives and oligosaccharides 1–4





Compound 9a





Compound 12

CO2Me Me





0 ppm



Compound 14



CDCl₃









Compound 22a,b



4.8 4.6 4.4 4.2 7.4 7.2 7.0 5.8 5.6 5.4 5.2 5.0 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 6.8 6.6 6.4 6.2 6.0









































Compound 37a,b













170

160





20

10

ppm





Compound 42



 D_2O





Compound 1

HC но ю нó

D₂O (1D NOESY presaturation)





HO CO2H но Ю ò

D₂O (1D NOESY presaturation)





D₂O (1D NOESY presaturation)

