# **Supporting Information for Publication**

# Synthesis of 2,3,5,6-Tetrasubstituted Tetrahydropyrans via (3,5)-Oxonium-Ene Reaction

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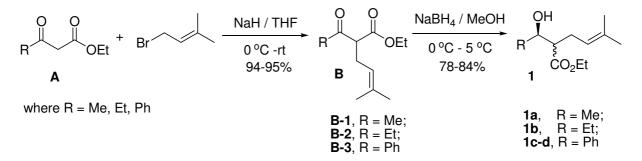
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**General Information:** All reagents are commercially obtained. BF<sub>3</sub> Et<sub>2</sub>O was distilled over CaH<sub>2</sub> prior to use. <sup>1</sup>H NMR spectra were recorded in CDCl<sub>3</sub> on 400 MHz NMR spectrometer using TMS as internal standard. The <sup>13</sup>C and <sup>19</sup>F NMR spectra were recorded at 100 MHz and 376 MHz, respectively. For <sup>13</sup>C and <sup>19</sup>F NMR CDCl<sub>3</sub> and C<sub>6</sub>F<sub>6</sub> were used as internal standard. IR spectra were recorded on FT-IR spectrometer. Melting points were measured in open capillary tubes and are uncorrected.

#### Preparation of ethyl 2-(1-hydroxyalkyl/hydroxy(phenyl)methyl)-5-methylhex-4-enoate

1: The ethyl 2-(1-hydroxyalkyl/hydroxy(phenyl)methyl)-5-methylhex-4-enoate 1 was synthesized starting from  $\beta$ -keto ester **A** and 1-bromo-3-methylbut-2-ene as shown in Scheme 1.<sup>1</sup> Thus the reaction of  $\beta$ -keto ester **A** with 1-bromo-3-methylbut-2-ene in presence



Scheme 1: Synthesis of ethyl 2-(1-hydroxyalkyl/hydroxy(phenyl)methyl)-5-methylhex-4enoate

of sodium hydride in THF afforded  $\alpha$ -substituted  $\beta$ -keto ester **B**, which after reduction with sodium borohydride in methanol gives alcohol **1** in 78-84% yields.

The methyl and ethyl substituted  $\beta$ -keto esters **B-1** and **B-2** afforded two inseparable diastereomers<sup>2</sup> **1a,b** whereas phenyl substituted  $\beta$ -ketoester **B-3** gave two separable *anti*- and *syn*-diastereomers **1c** and **1d**, respectively. The structures of all compounds are determined

from IR, <sup>1</sup>H, <sup>13</sup>C NMR and mass spectroscopy. The stereochemistry of compounds **1c** and **1d** are determined from coupling constants values.<sup>2b,3</sup>

References:

- M. Peña-López, M. M. Martinez, L. A. Sarandeses and J. S. Sestelo, J. Org. Chem. 2010, 75, 5337-5339.
- (a) V. A. Kramer and H. Pfander, *Helv. Chim. Acta* 1982, **65**, 293-301; (b) P. Galatsis,
   S. D. Millan, P. Nechala and G. Ferguson, *J. Org. Chem.* 1994, **59**, 6643-6651.
- E. Marcantoni, S. Alessaandrini, M. Malavolta, G. Bartoli, M. C. Bellucci and L. Sambri, J. Org. Chem. 1999, 64, 1986-1992.

General procedure for the synthesis of *a*-alkyl- $\beta$ -keto esters (B-1-3): To a suspension of sodium hydride (15.69 mmol, 1 equiv.) in THF (15 mL) at 0 °C was added  $\beta$ -keto ester (15.69 mmol, 1 equiv.) dropwise via syringe. After 20 min, a solution of 3,3-dimethylallyl bromide (17.26 mmol, 1.1 equiv.) in THF (5 mL) was added and the mixture was left at room temperature overnight. The solvent was removed under reduced pressure, and the residue was dissolved in Et<sub>2</sub>O (10 mL) and washed with brine (25 ml). The organic layer was dried and filtered, and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography over silica gel to afford the title compounds as colourless oil.

Synthesis of Ethyl 2-acetyl-5-methylhex-4-enoate (B-1): To a suspension of sodium hydride (0.378 g, 15.69 mmol) in THF (15 mL) at 0  $^{\circ}$ C was added ethyl acetoacetate (2 mL, 15.69 mmol) dropwise via syringe. After 20 min, a solution of 3,3-dimethylallyl bromide (2 mL, 17.26 mmol) in THF (5 mL) was added and the mixture was left at room temperature overnight. The solvent was removed under reduced pressure, and the residue was dissolved in Et<sub>2</sub>O (10 mL) and washed with brine (25 mL). The organic layer was dried and filtered, and

chromatography over silica gel (10% EtOAc/hexane) to afford ethyl 2-acetyl-5-methylhex-4enoate **B-1** (2.97 g, 95%) as a colourless oil; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.27 (t, *J* = 7.2 Hz, 3 H), 1.63 (s, 3 H), 1.68 (s, 3 H), 2.22 (s, 3 H), 2.54 (t, *J* = 7.2 Hz, 2 H), 3.43 (t, *J* = 7.6 Hz, 1 H), 4.19 (q, *J* = 7.2 Hz, 2 H), 5.03 (dt, *J* = 7.6 and 1.6 Hz, 1 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  14.1, 17.8, 25.8, 27.0, 29.1, 59.8, 61.3, 119.9, 134.7, 169.7, 203.2; IR (KBr, Neat): 2981, 2930, 1739, 1718, 1205, 1150 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>11</sub>H<sub>18</sub>O<sub>3</sub> (M+H)<sup>+</sup> requires 199.1334; found 199.1337. APCI-MS: m/z (relative intensity): 199.2 ((M+H)<sup>+</sup>, 43%), 181.1 (100), 169.1 (55), 143.1 (17), 124.1 (43), 107.1 (73).

Ethyl 5-methyl-2-propionylhex-4-enoate (B-2): Colourless oil (3.13 g, 94%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.06 (t, J = 7.2 Hz, 3 H), 1.26 (t, J = 7.2, 3 H), 1.62 (s, 3 H), 1.67 (s, 3 H), 2.47-2.60 (m, 4 H), 3.45 (t, J = 7.2 Hz, 1 H), 4.17 (q, J = 7.2 Hz, 2 H), 5.02 (dt, J = 7.6 and 1.6 Hz, 1 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  7.8, 14.2, 17.9, 25.9, 27.2, 35.6, 58.9, 61.4, 120.1, 134.8, 169.9, 206.0; IR (KBr, Neat): 2980, 2937, 1741, 1716, 1198, 1156 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>12</sub>H<sub>20</sub>O<sub>3</sub> (M<sup>+</sup>) requires 212.1412; found 212.1408. APCI-MS: m/z (relative intensity): 212.2 ((M+H)<sup>+</sup>, 100%), 194.7 (65), 168.7 (27), 123.8 (22).

Ethyl 2-benzoyl-5-methylhex-4-enoate (B-3): Colourless oil (3.88 g, 95%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.67 (t, J = 7.2 Hz, 3 H), 1.62 (s, 3 H), 1.65 (s, 3 H), 2.61-2.78 (m, 2 H), 4.14 (q, J = 7.2 Hz, 2 H), 4.30 (t, J = 7.2 Hz, 1 H), 5.11 (dt, J = 7.2 and 1.6 Hz, 1 H), 7.44-7.50 (m, 2 H), 7.55-7.61 (m, 1 H), 7.97-8.00 (m, 2 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  14.1, 17.9, 25.8, 27.8, 54.6, 61.4, 120.3, 128.7, 128.8, 133.5, 134.7, 136.4, 169.9, 195.2; IR (KBr, Neat): 2979, 2929, 1737, 1688, 1448, 1379, 1241, 1153 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>16</sub>H<sub>20</sub>O<sub>3</sub> (M+H)<sup>+</sup> requires 261.1490; found 261.1494. APCI-MS: m/z (relative intensity): 261.2 ((M+H)<sup>+</sup>, 57%), 215.1 (21), 193.1 (23), 169.1 (12), 124.1 (100).

General procedure for the synthesis of Ethyl 2-(1hydroxyalkyl/hydroxy(phenyl)methyl)-5-methylhex-4-enoate (1a-d): To a solution of αalkyl-β-keto esters (14.98 mmol, 1 equiv.) in dry MeOH (15 mL) at 0 °C, was added sodium borohydride (39.25 mmol, 2.62 equiv.) in small portions. The reaction mixture was stirred in between 0 °C to 5 °C for 1.5 h. The progress of the reaction was monitored by TLC with ethyl acetate and hexane as eluents. After completion of the reaction, the product was extracted with ethyl acetate (30 mL) and then washed with water (15 mL) and brine (15 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to leave the crude product, which was purified by column chromatography over silica gel to give the title compounds.

Synthesis of Ethyl 2-(1-hydroxyethyl)-5-methylhex-4-enoate (1a): To a solution of ethyl 2-acetyl-5-methylhex-4-enoate (2.97 g, 14.98 mmol) in dry MeOH (15mL) at 0 °C, was added sodium borohydride (1.485 g, 39.25 mmol) in small portions. The reaction mixture was stirred in between 0 °C to 5 °C for 1.5 h. The progress of the reaction was monitored by TLC with ethyl acetate and hexane (3:22) as eluents. After completion of the reaction, the product was extracted with ethyl acetate (30 mL) and then washed with water (15 mL) and brine (15 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to leave the crude products, which were purified by column chromatography over silica gel to give an inseparable mixture of two diastereomers **1a** (2.40 g, 80% overall yield) as a colourless oil; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.19-1.29 (m, 6 H), 1.62 (s, 3 H), 1.69 (s, 3 H), 2.29-2.47 (m, 3 H), 2.51 (brs 0.5 H), 2.69 (brs, 0.5 H), 3.86-3.96 (m, 0.5 H), 3.98-4.06 (m, 0.5 H), 4.08-4.24 (m, 2 H), 5.04-5.14 (m, 1 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  14.1(2C), 17.5(2C), 20.5, 21.1, 25.6(2C), 26.7, 27.7, 53.0, 53.1, 60.2, 60.3, 67.8, 67.9, 120.5, 121.1, 133.3, 133.6, 174.7, 175.0; IR (KBr, Neat): 3441, 2972, 2929, 1732, 1640, 1182, 1155 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>11</sub>H<sub>20</sub>O<sub>3</sub> (M+H)<sup>+</sup> requires 201.1490; found 201.1498. APCI-MS: m/z (relative intensity):

201.2 ((M+H)<sup>+</sup>, 30%), 137.1 (4), 125.1 (11), 124.1 (100), 123.1 (14), 82.0 (24).

Ethyl 2-(1-hydroxypropyl)-5-methylhex-4-enoate (1b): Colourless oil (2.50 g, 78%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  0.98 (t, *J* = 7.6 Hz, 3 H), 1.25 (t, *J* = 7.2 Hz, 1.5 H), 1.26 (t, *J* = 7.2 Hz, 1.5 H), 1.45-1.54 (m, 2H), 1.61 (s, 1.5 H), 1.62 (s, 1.5 H), 1.69 (s, 3 H), 2.28-2.50 (m, 3 H), 2.67 (brs, 1 H), 3.56-3.64 (m, 0.5 H), 3.70-3.78 (m, 0.5 H), 4.08-4.22 (m, 2 H), 5.04-5.16 (m, 1 H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  10.2, 10.3, 14.3(2C), 17.8(2C), 25.8, 26.1, 27.4(2C), 28.3, 28.6, 50.7, 51.2, 60.5(2C), 73.3, 73.4, 120.7, 121.3, 133.6, 134.1, 175.3, 175.5; IR (KBr, Neat): 3456, 2968, 2932, 1731, 1642, 1183, 1159 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>12</sub>H<sub>22</sub>O<sub>3</sub> (M+H)<sup>+</sup> requires 215.1647; found 215.1653. APCI-MS: m/z (relative intensity): 215.2 ((M+H)<sup>+</sup>, 100%), 197.2 (6), 169.1 (8), 151.1 (7), 124.1 (35), 123.1 (14), 82.0 (7).

Synthesis of *anti/syn*-Ethyl 2-(hydroxy(phenyl)methyl)-5-methylhex-4-enoate (1c & 1d):

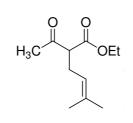
To a solution of ethyl 2-benzoyl-5-methylhex-4-enoate (3.90 g, 14.98 mmol) in dry MeOH (15 mL) at 0 °C, was added sodium borohydride (1.485 g, 39.25 mmol) in small portions. The reaction mixture was stirred in between 0 °C to 5 °C for 1.5 h. The progress of the reaction was monitored by TLC with ethyl acetate and hexane (7:43) as eluents. After completion of the reaction, the product was extracted with ethyl acetate (30 mL) and then washed with water (15 mL) and brine (15 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to leave the crude products, which were separated by column chromatography over silica gel to give **1c** (1.73 g, 44%) and **1d** (1.58 g, 40%) as a colourless oil.

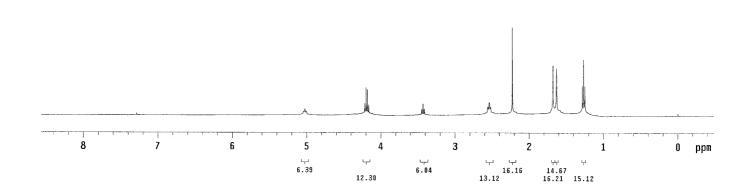
*anti*-Ethyl 2-(hydroxy(phenyl)methyl)-5-methylhex-4-enoate (1c): Colourless oil (1.73 g, 44%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 1.18 (t, *J* = 7.2 Hz, 3 H), 1.50 (s, 3 H), 1.65 (s, 3 H), 2.05 (ddd, *J* = 13.2, 7.6 and 5.6 Hz, 1 H), 2.25 (ddd, *J* = 14.8, 8.0 and 6.8 Hz, 1 H), 2.75 (ddd, *J* = 12.8, 8.8, 5.2 Hz, 1 H), 3.16 (d, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 12.8, 8.8, 5.2 Hz, 1 H), 3.16 (d, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 12.8, 8.8, 5.2 Hz, 1 H), 3.16 (d, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 12.8, 8.8, 5.2 Hz, 1 H), 3.16 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, *J* = 5.2 Hz, 1H), 4.11 (q, *J* = 7.2 Hz, 2 H), 4.80 (dd, J) = 5.2 Hz, 1H), 4.11 (q, J =

*J* = 6.4 and 4.4 Hz, 1H), 5.03 (dt, *J* = 6.8 and 1.2 Hz, 1H), 7.26-7.37 (m, 5H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  14.3, 17.8, 25.9, 28.4, 53.3, 60.7, 74.8, 120.2, 126.6, 128.0, 128.6, 134.3, 142.2, 175.2; IR (KBr, Neat): 3461, 2978, 2929, 1729, 1452, 1377, 1180, 1037, 766, 702 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>16</sub>H<sub>22</sub>O<sub>3</sub> (M<sup>+</sup>) requires 262.1569; found 262.1573. APCI-MS: m/z (relative intensity): 262.2 (M<sup>+</sup>, 3%), 244.6 (15), 177.7 (15), 176.7 (100), 123.8 (22). *syn*-Ethyl 2-(hydroxy(phenyl)methyl)-5-methylhex-4-enoate (1d): Colourless oil (1.58 g, 40%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  1.11 (t, *J* = 7.2 Hz, 3 H), 1.54 (s, 3 H), 1.64 (s, 3 H), 2.29 (ddd, *J* = 14.4, 8.8 and 5.8 Hz, 1 H), 2.46 (ddd, *J* = 14.4, 8.8 and 5.6 Hz, 1 H), 2.72 (ddd, *J* = 10.4, 5.6 and 4.4 Hz, 1 H), 3.03 (brs, 1H), 4.02 (q, *J* = 7.2 Hz, 2 H), 4.95 (d, *J* = 5.6 Hz, 1H), 5.04 (t, *J* = 6.8 Hz, 1H), 7.24-7.39 (m, 5H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  14.2, 17.8, 25.9, 26.1, 53.3, 60.7, 74.2, 121.1, 126.4, 127.8, 128.4, 134.0, 141.7, 175.0; IR (KBr, Neat): 3460, 2979, 2930, 1728, 1453, 1375, 1180, 1026, 767, 701 cm<sup>-1</sup>. HRMS (APCI) cald. for C<sub>16</sub>H<sub>22</sub>O<sub>3</sub> (M+H)<sup>+</sup> requires 263.1647; found 263.1652. APCI-MS: m/z (relative intensity): 263.2 ((M+H)<sup>+</sup>, 19%), 192.1 (5), 178.1 (15), 177.1 (100), 171.1 (21), 131.1 (13), 124.1 (75).

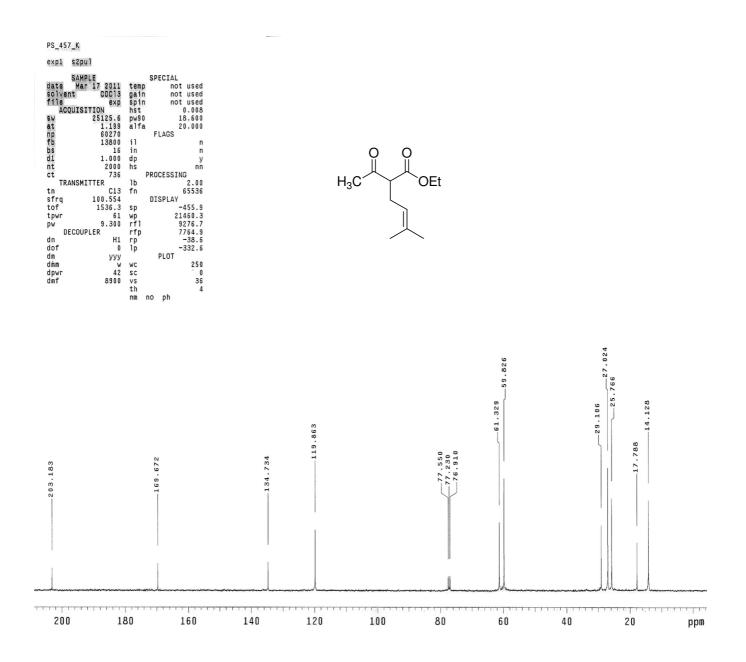
<sup>1</sup>H NMR spectra of B-1

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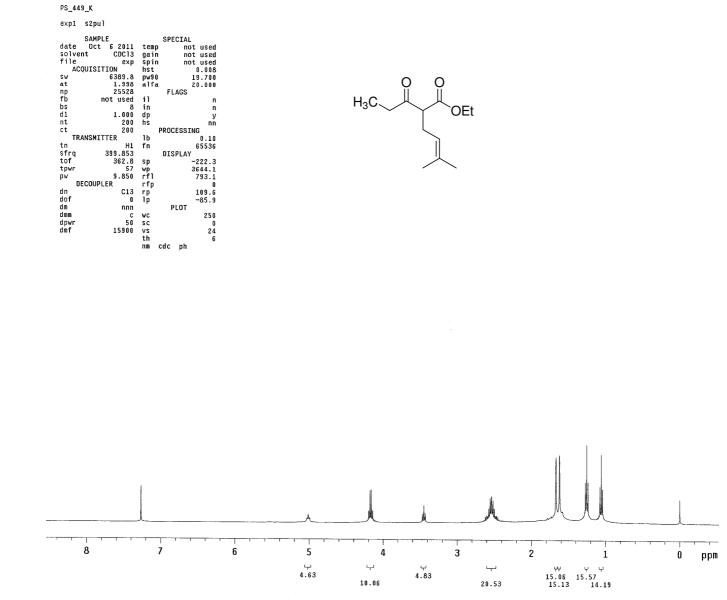




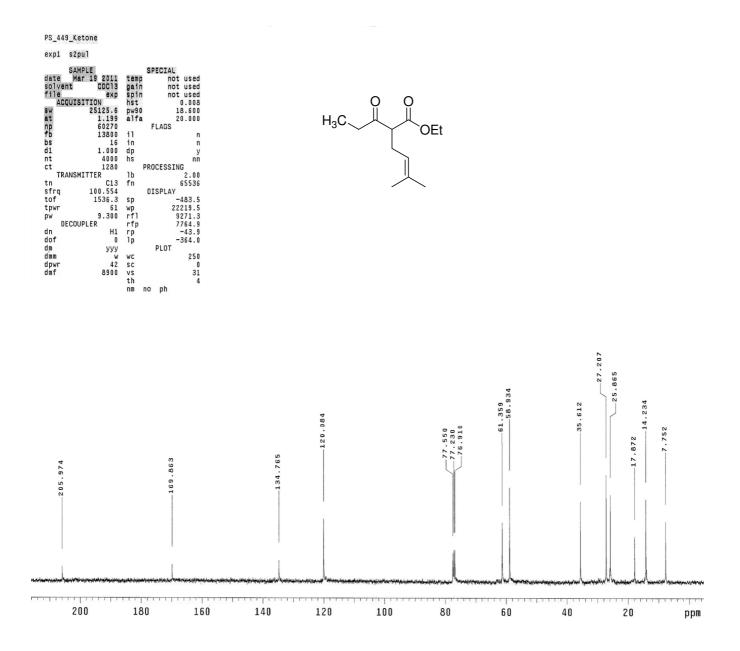
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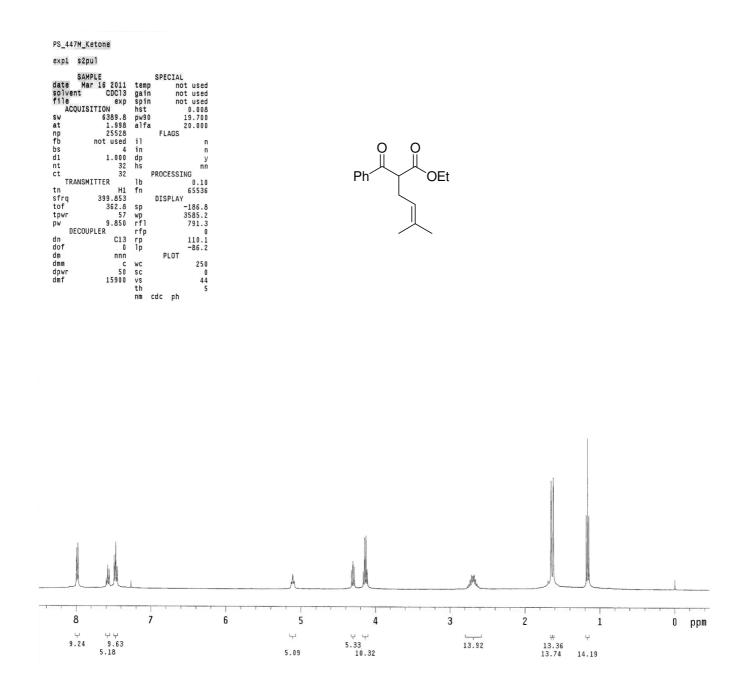
<sup>1</sup>H NMR spectra of B-2



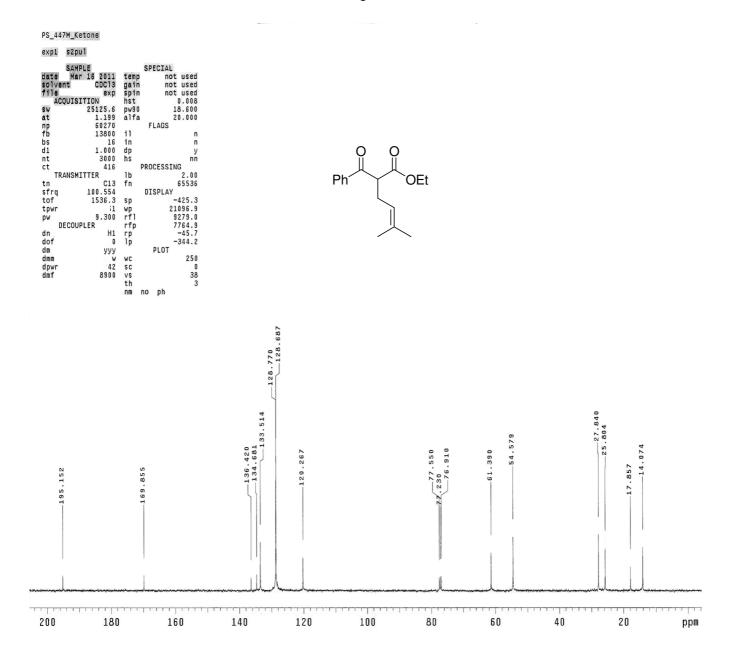
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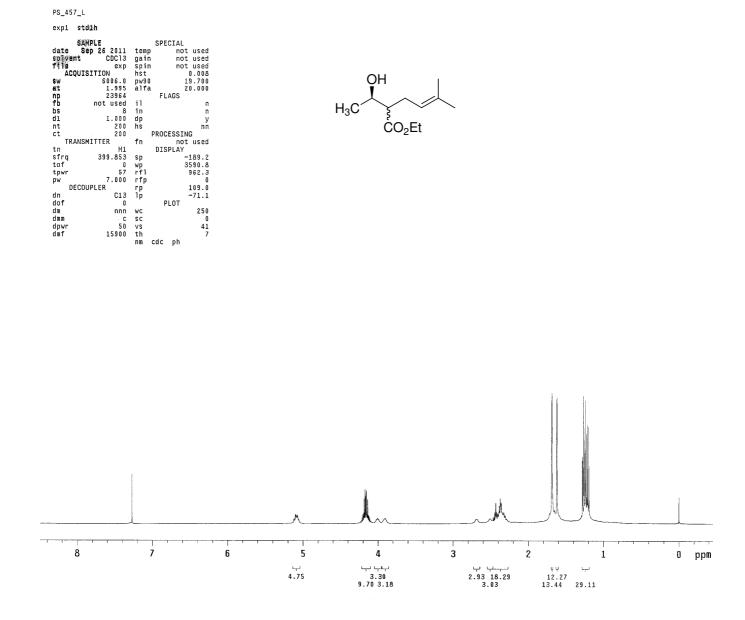
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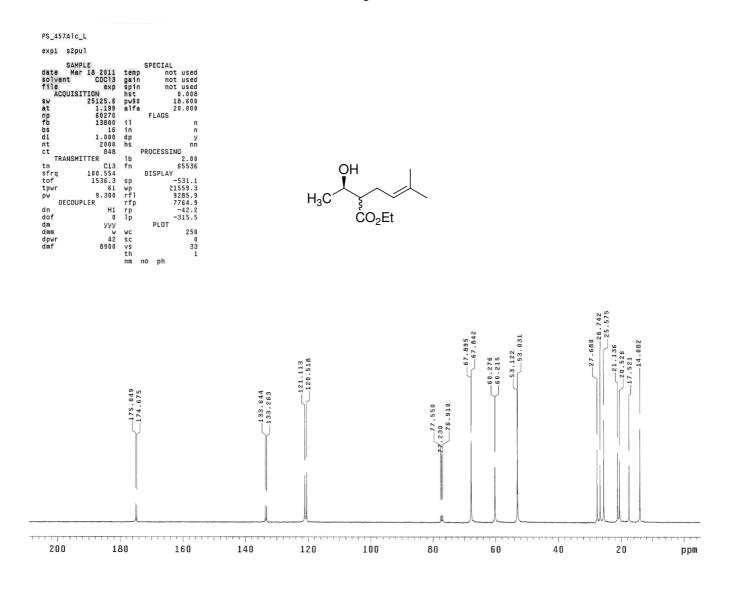
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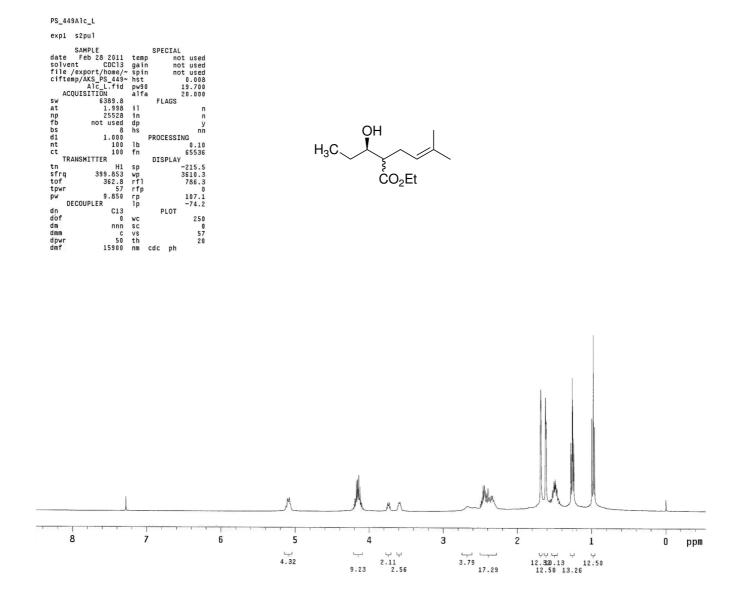
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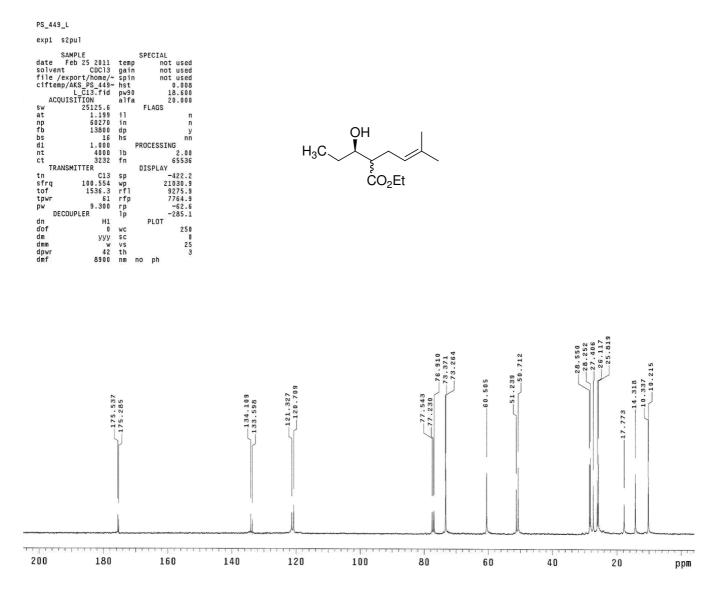
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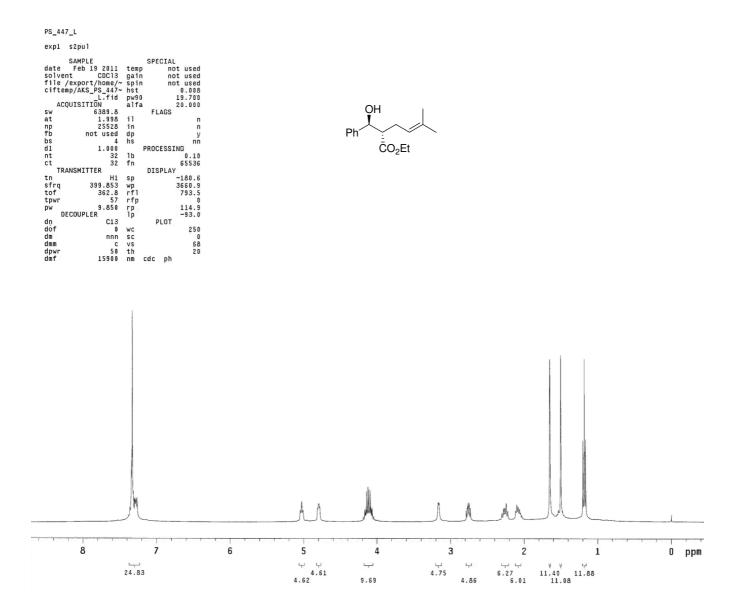
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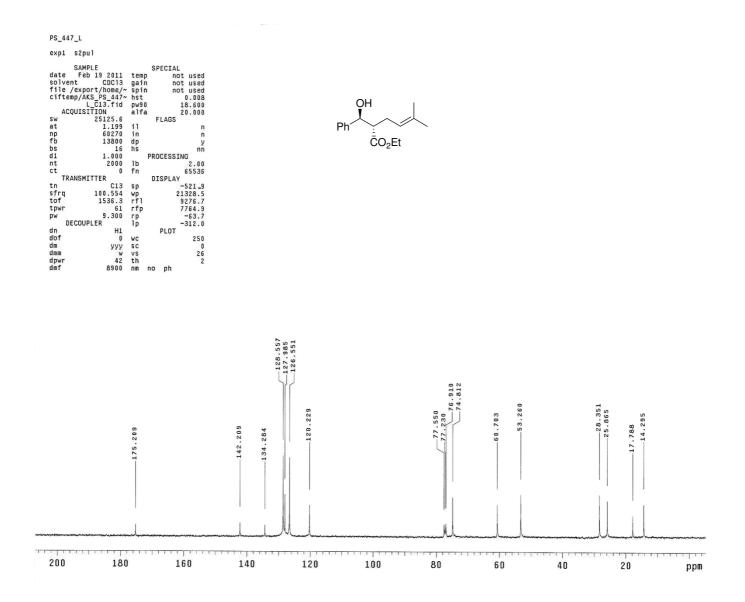
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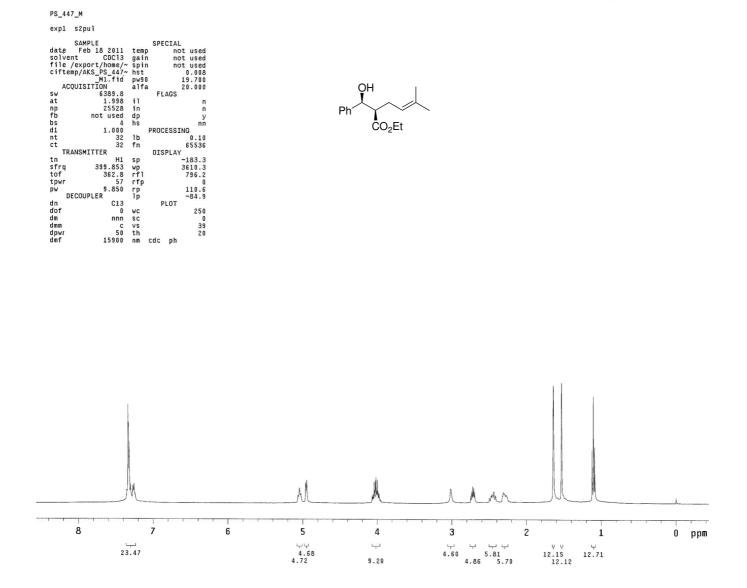




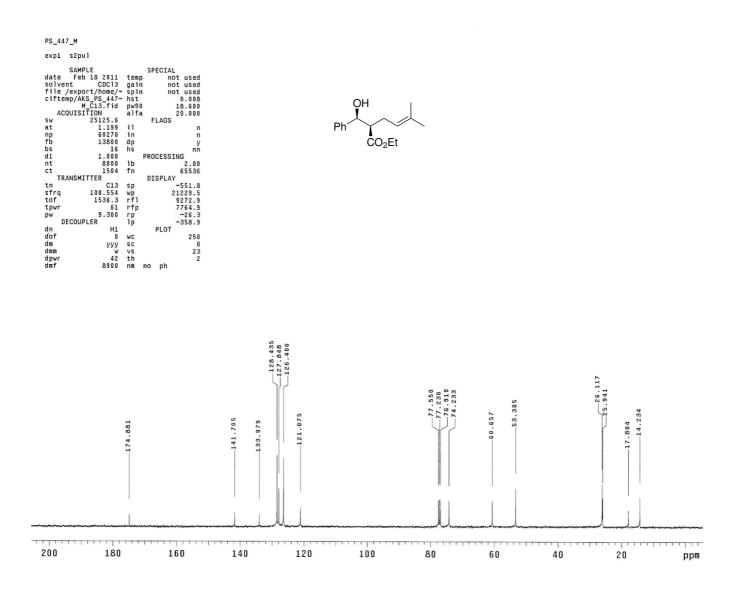
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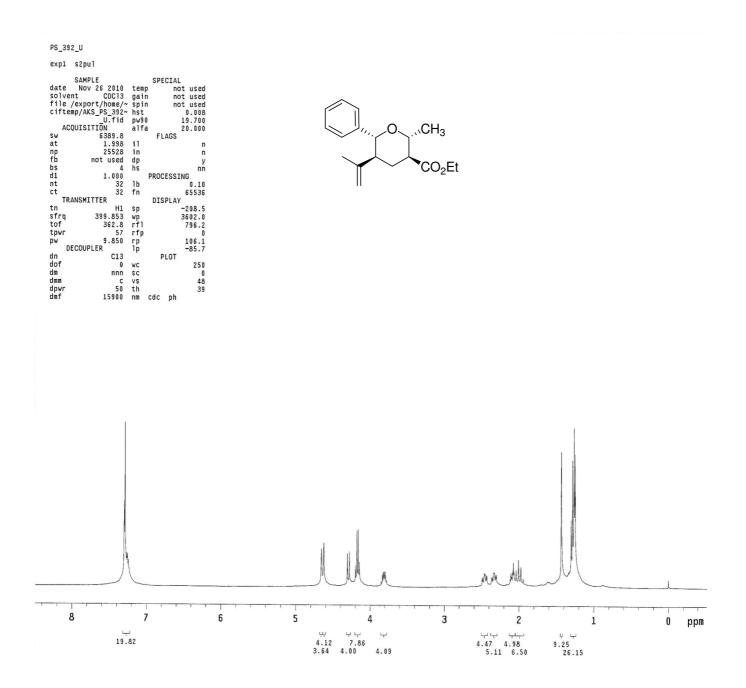
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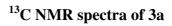


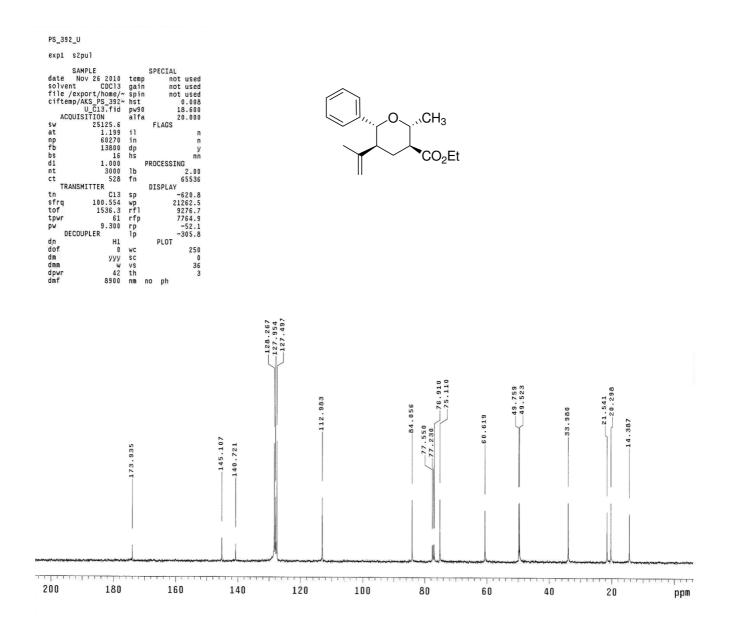
# <sup>13</sup>C NMR spectra of 1d



<sup>1</sup>H NMR spectra of 3a

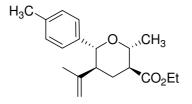


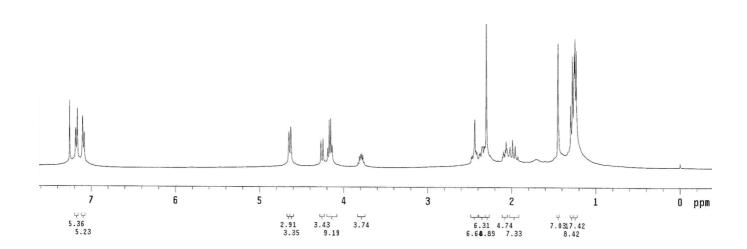




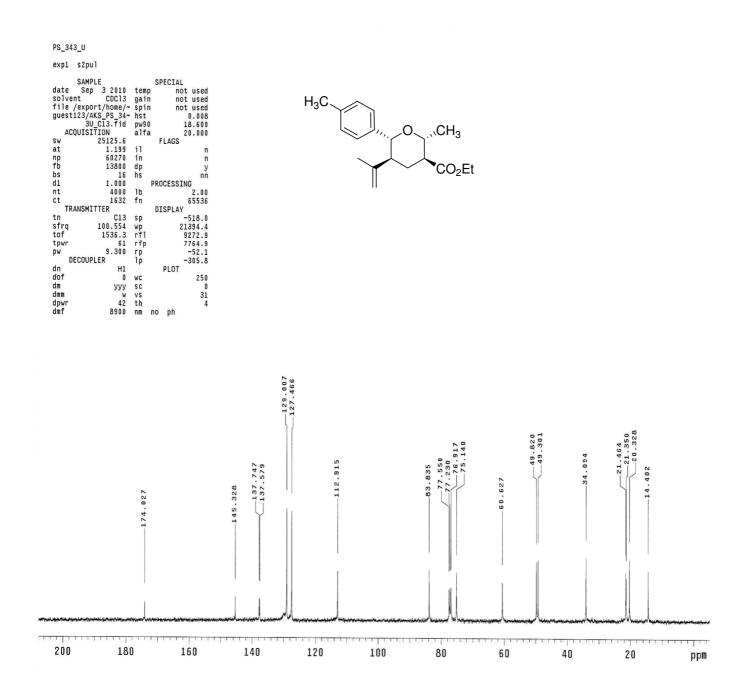
<sup>1</sup>H NMR spectra of 3b

PS\_343\_MU exp1 s2pul expl szpu: SAMPLE date Sep 2 2010 temp solvent CDCl3 gain file /export/home/~ spin guest123/AKS\_PS\_34~ hst 3\_U.Fid pw90 ACQUISITION alfa sw 6389.8 at 1.998 il np 25528 in fb not used dp bs 4 hs dl 1.000 F nt 32 lb ct 32 fn TRANSMITTER SPECIAL not used not used 0.008 19.700 20.000 FLAGS FLAGS n n nn PROCESSING 0.10 65536 TRANSMITTER DISPLAY TRANSMITTER H1 399.853 362.8 Vr 57 9.850 DECOUPLER tn sfrq tof tpwr pw dn dof dm dmm dpwr dmf -157.4 3207.5 795.4 0 sp wp rfl rfp rp lp 0 118.1 -67.4 C13 0 nnn 50 15900 PLOT wc sc vs th nm 250 0 43 39 cdc ph



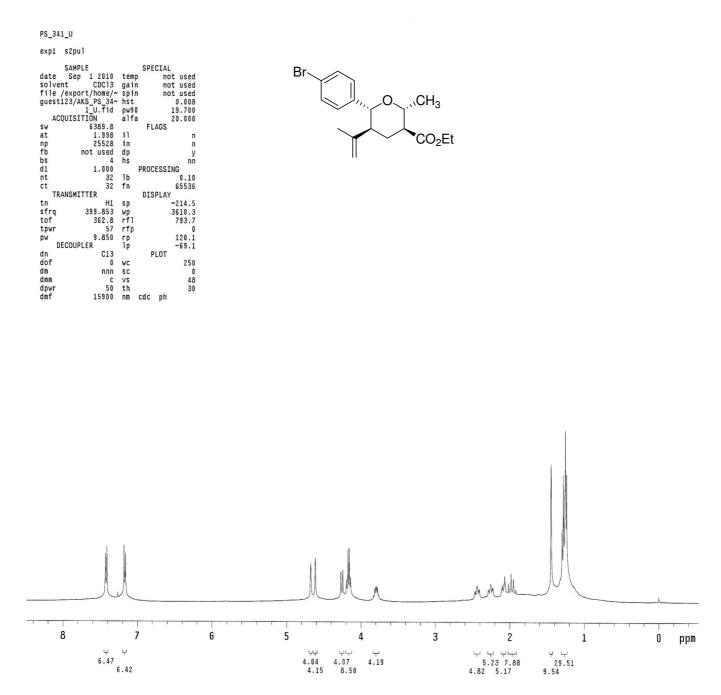


# <sup>13</sup>C NMR spectra of 3b

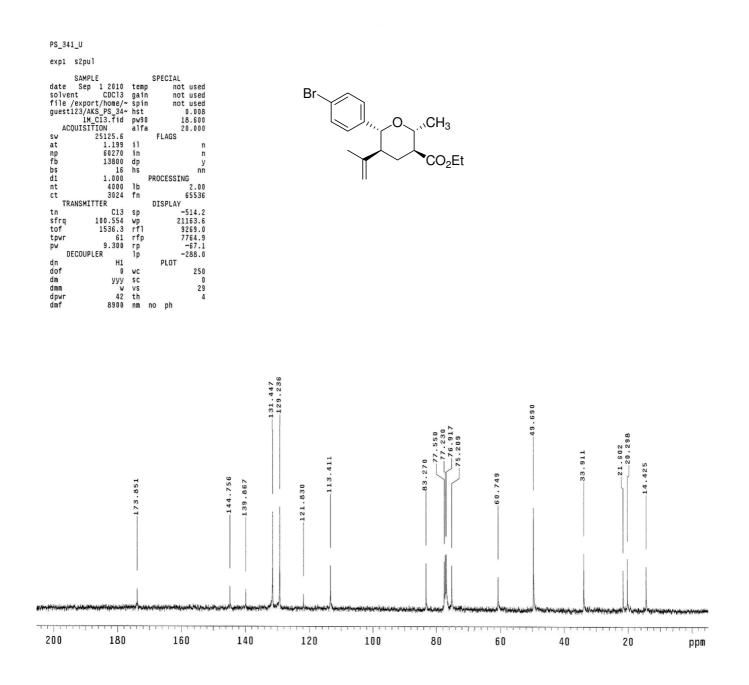


S-25

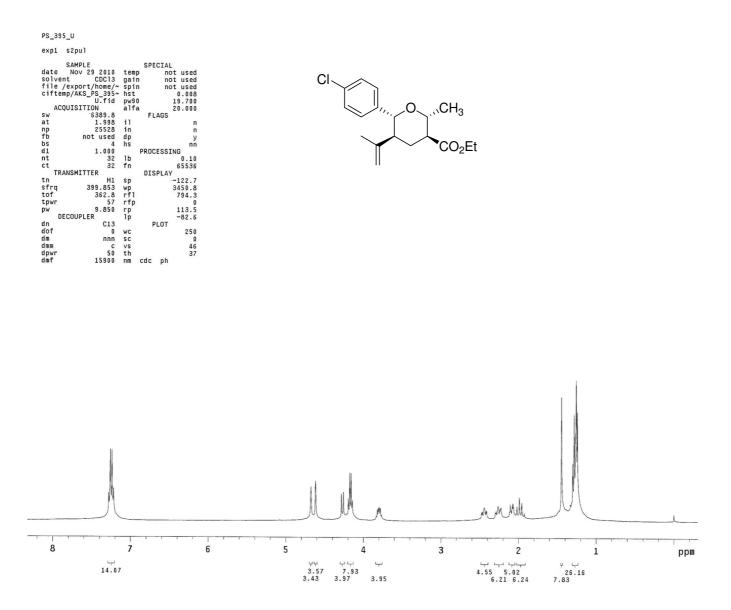
<sup>1</sup>H NMR spectra of 3c



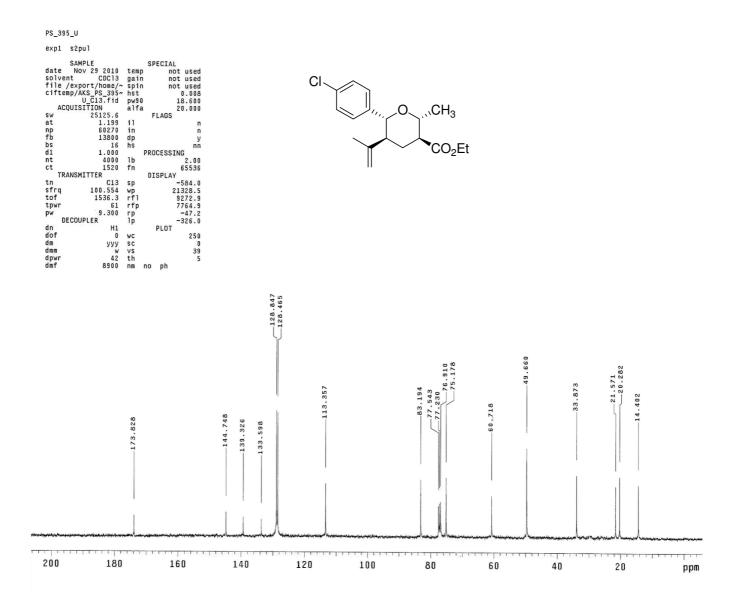
<sup>13</sup>C NMR spectra of 3c



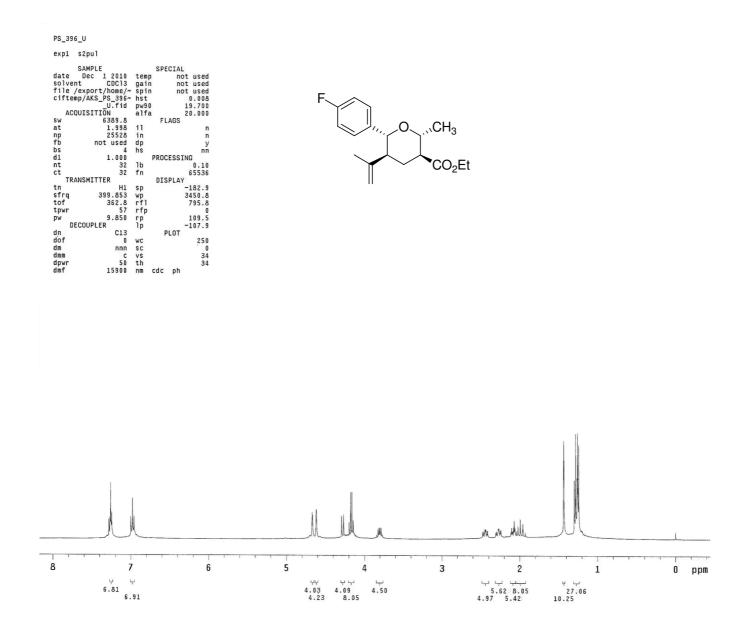
<sup>1</sup>H NMR spectra of 3d



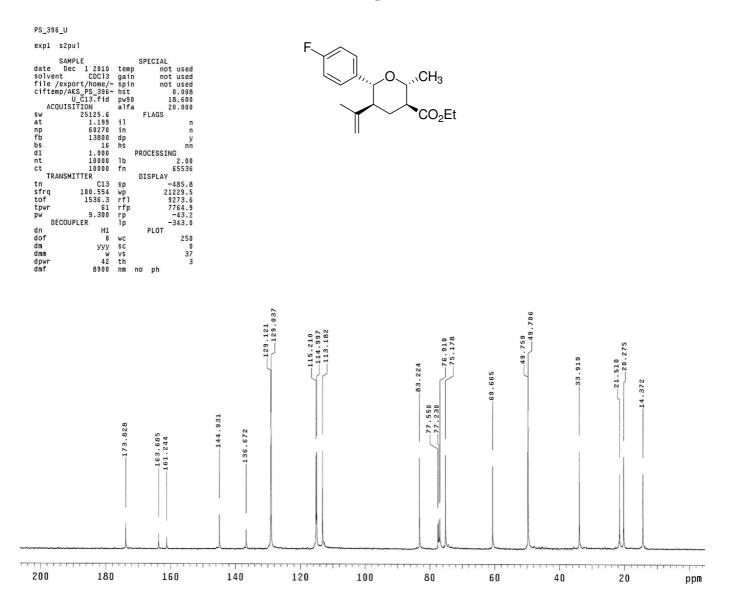
<sup>13</sup>C NMR spectra of 3d



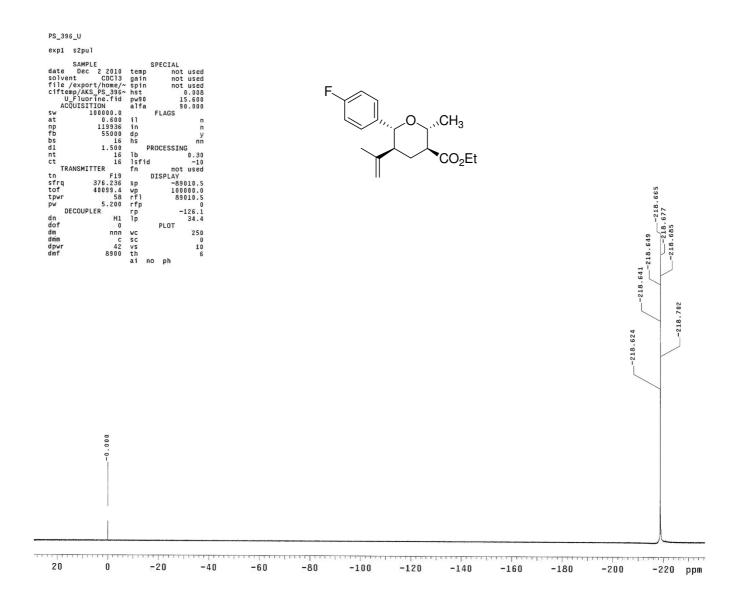
### <sup>1</sup>H NMR spectra of 3e



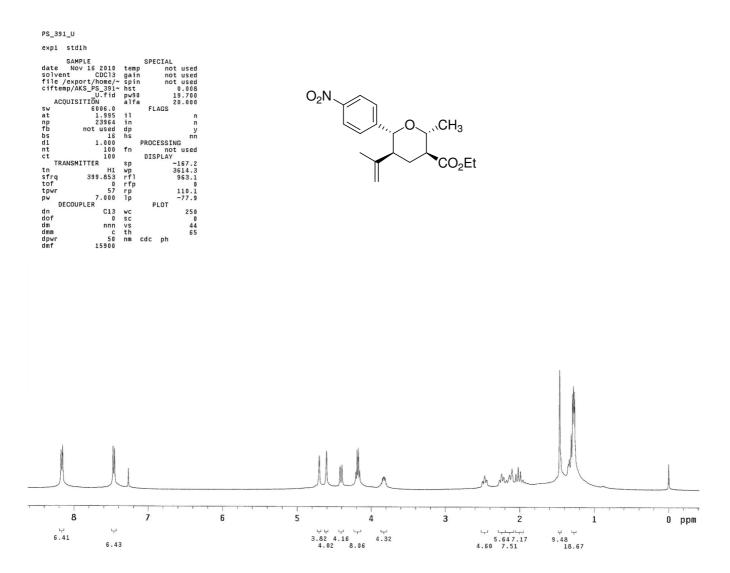
<sup>13</sup>C NMR spectra of 3e

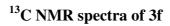


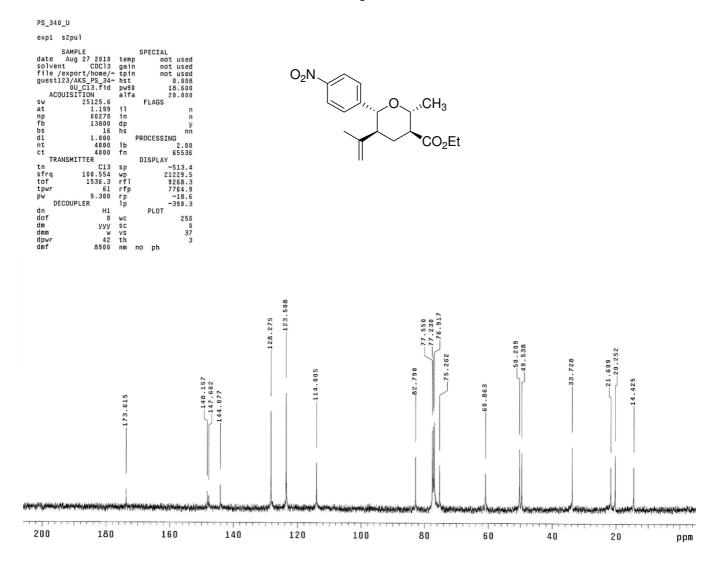
<sup>19</sup>F NMR spectra of 3e



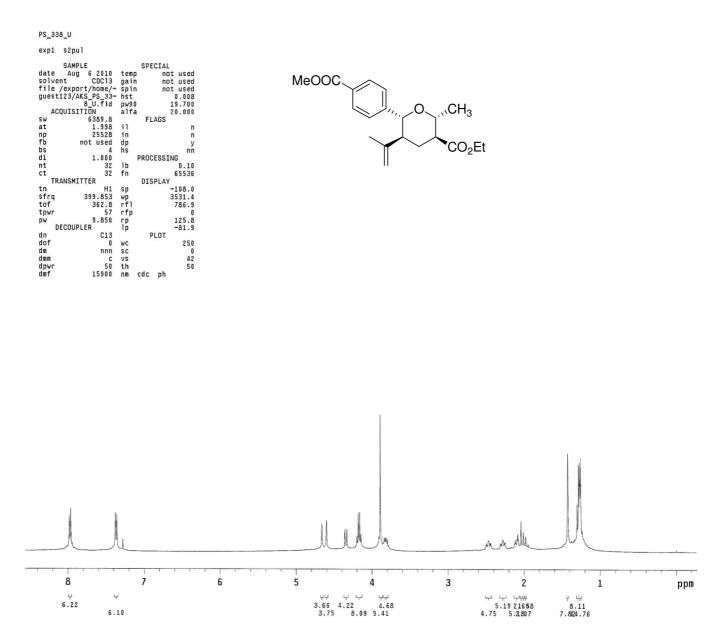
<sup>1</sup>H NMR spectra of 3f

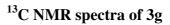


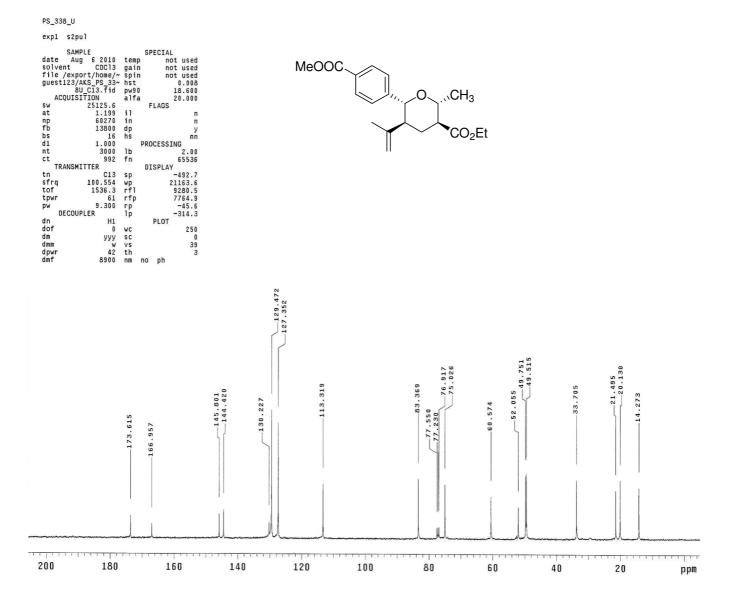




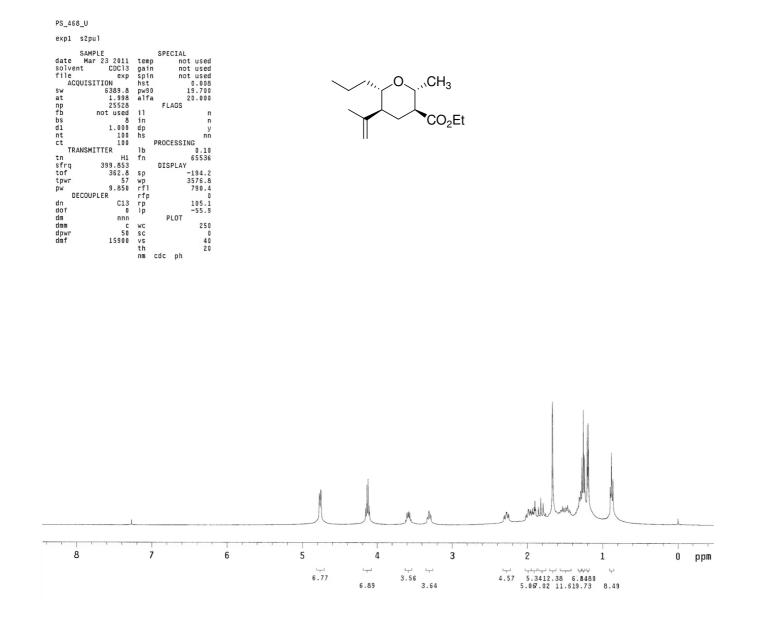
## <sup>1</sup>H NMR spectra of 3g



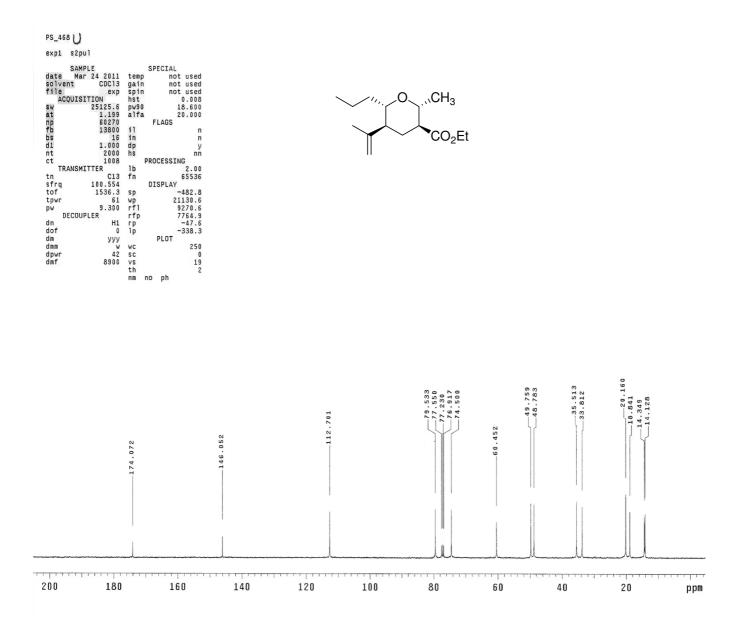




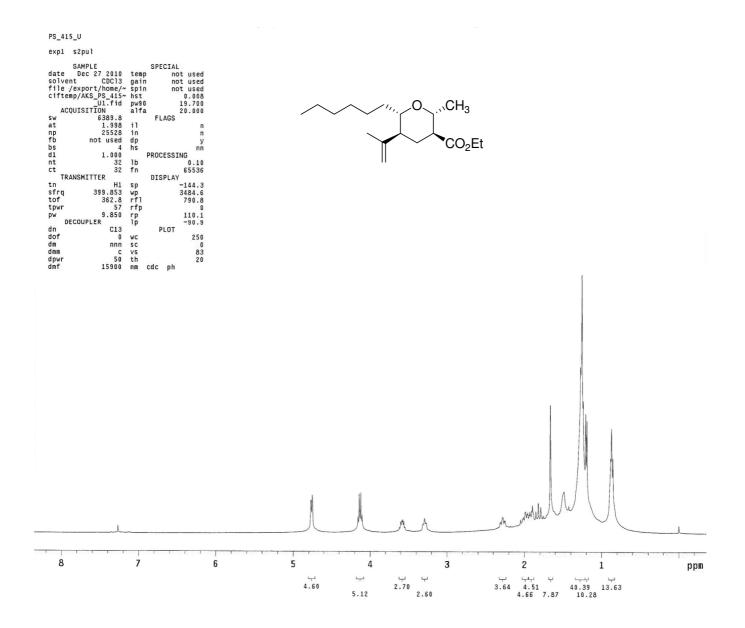




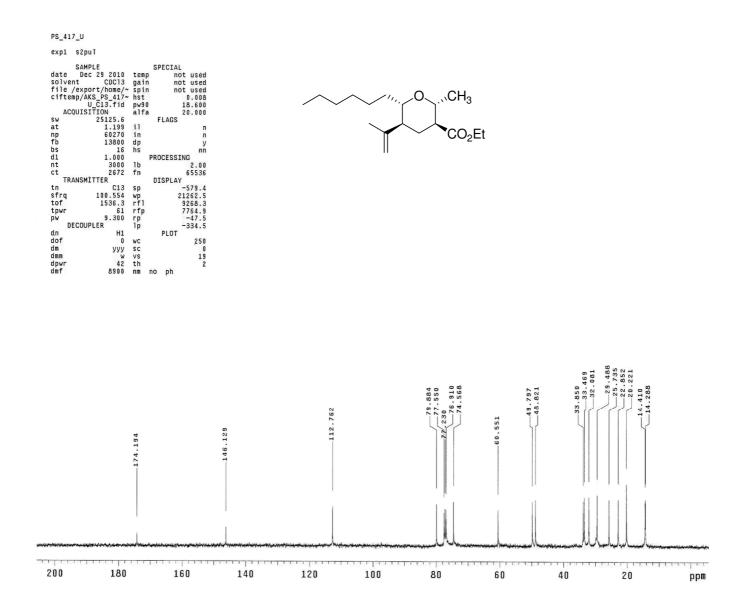
<sup>13</sup>C NMR spectra of 3h



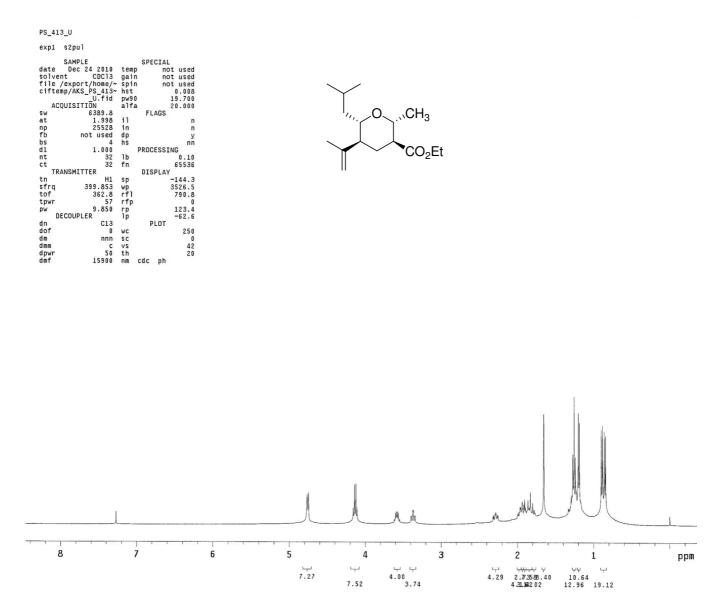
<sup>1</sup>H NMR spectra of 3i

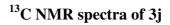


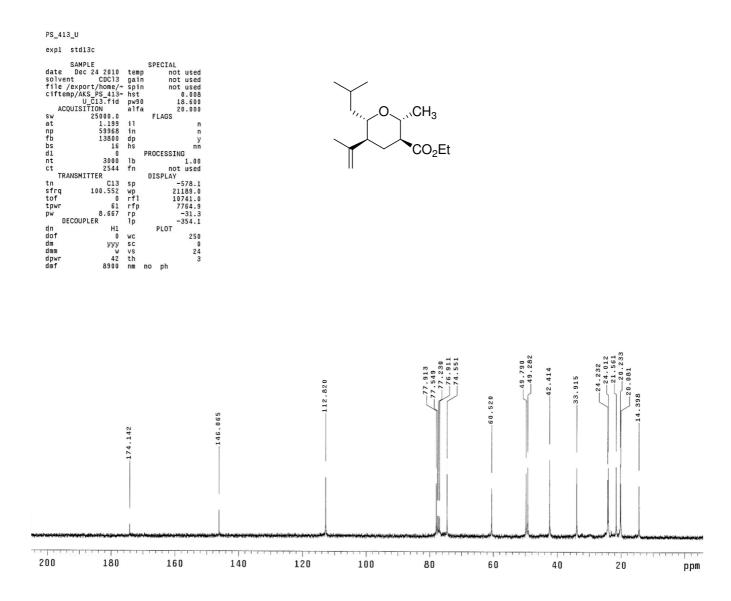
<sup>13</sup>C NMR spectra of 3i



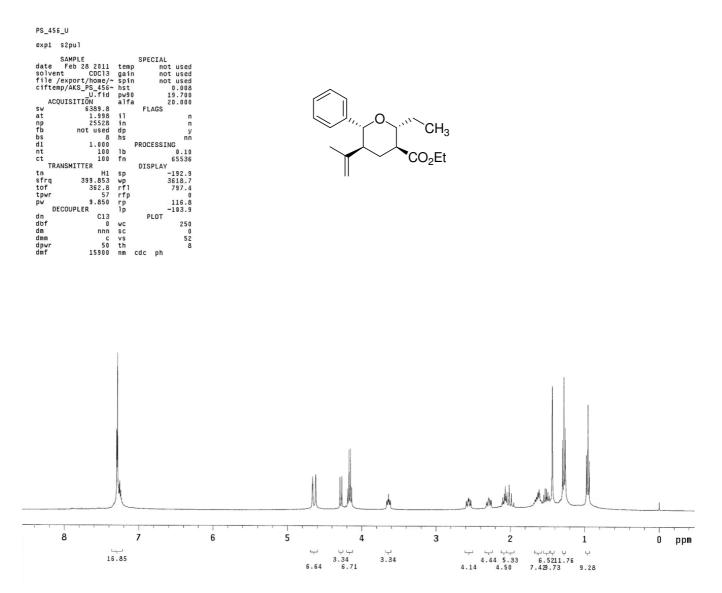
<sup>1</sup>H NMR spectra of 3j

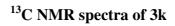


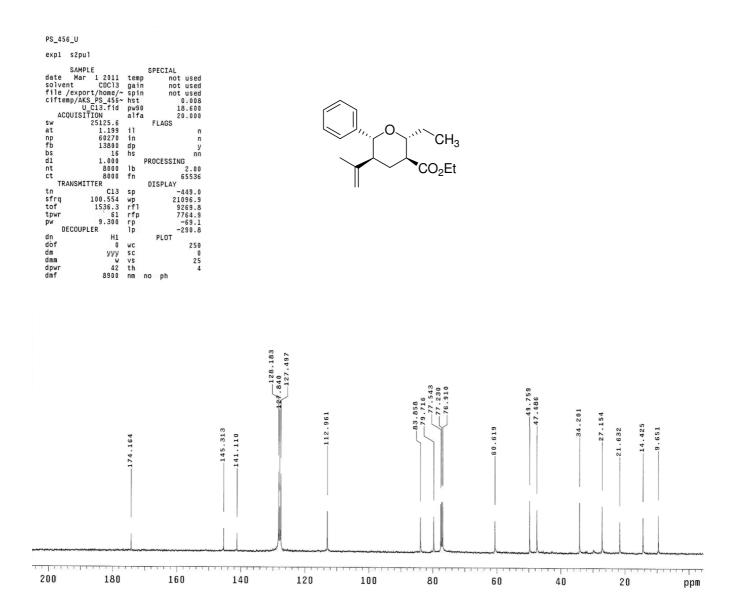


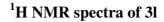


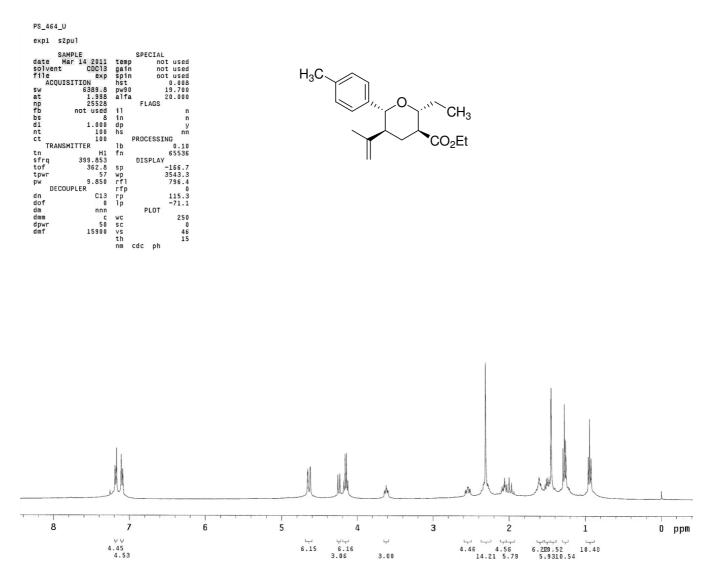
<sup>1</sup>H NMR spectra of 3k

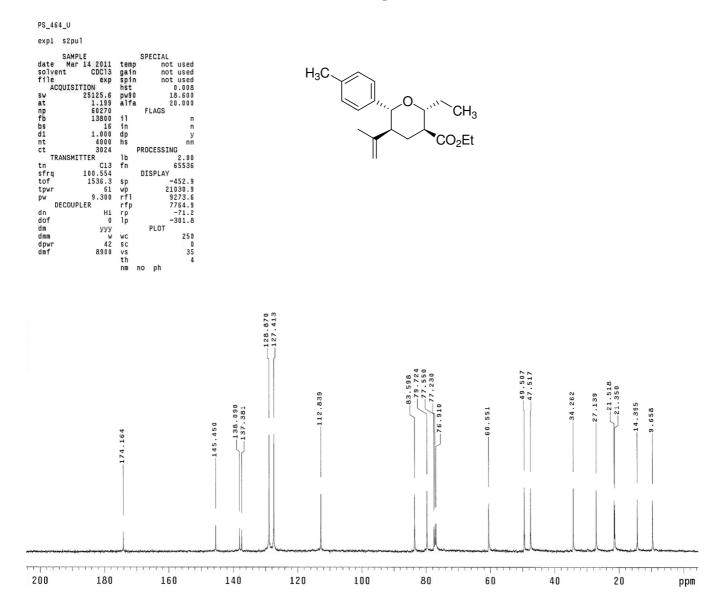








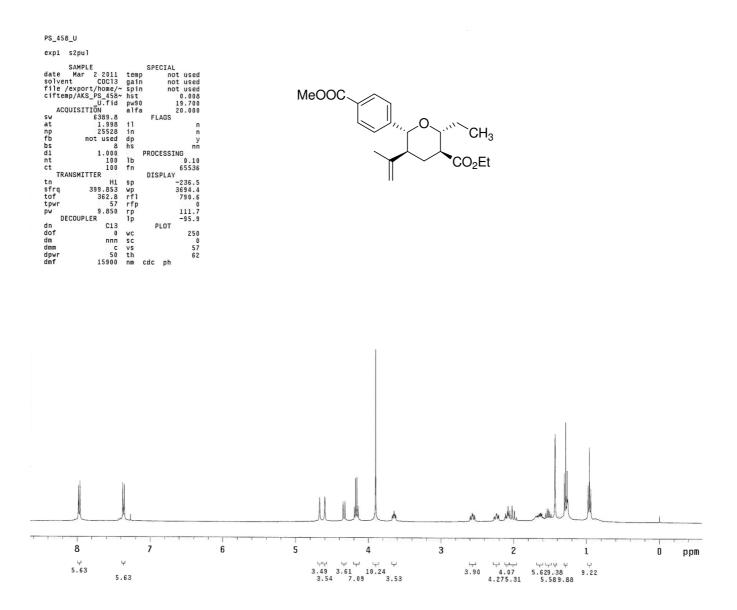


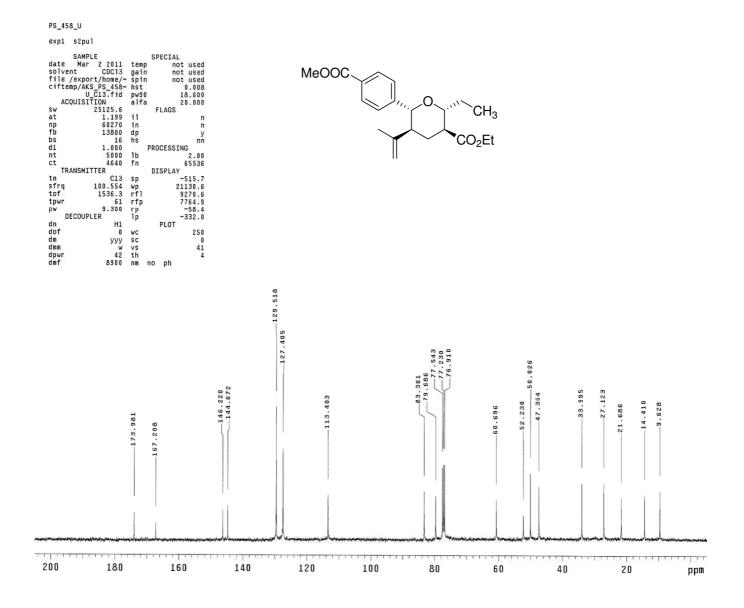


# <sup>13</sup>C NMR spectra of 3l

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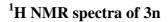
# <sup>1</sup>H NMR spectra of 3m

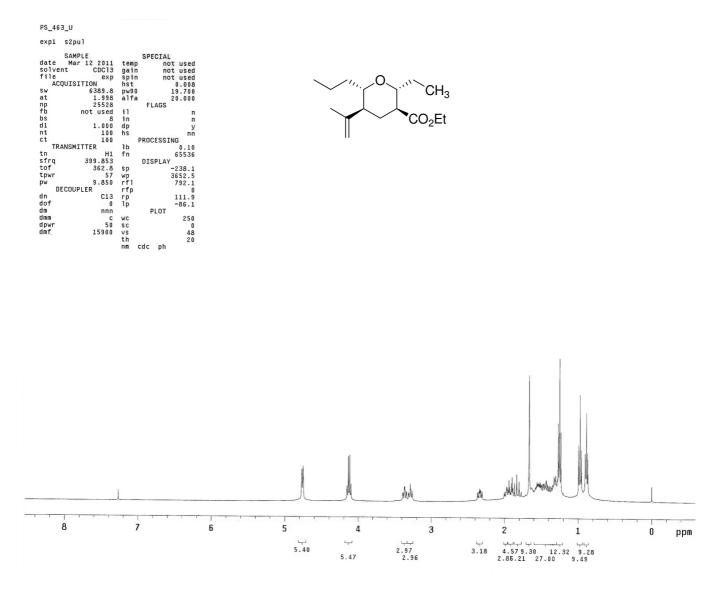


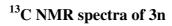


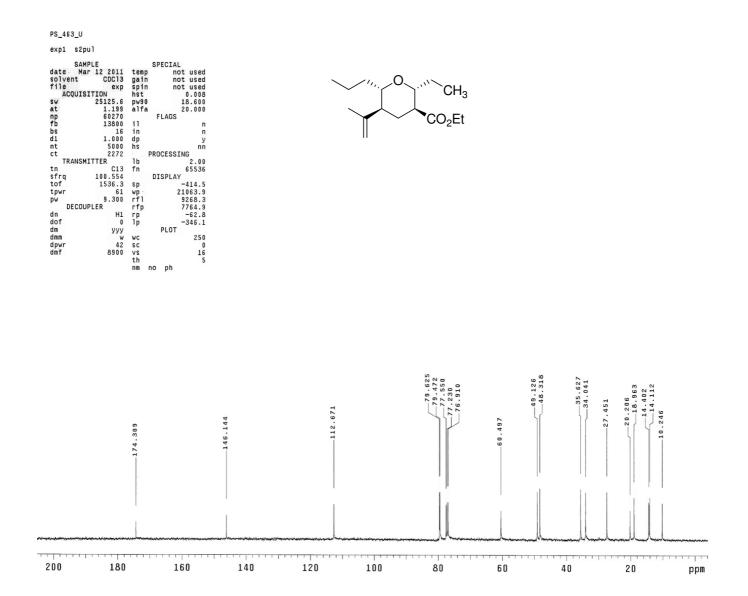
### <sup>13</sup>C NMR spectra of 3m

**S-48** 

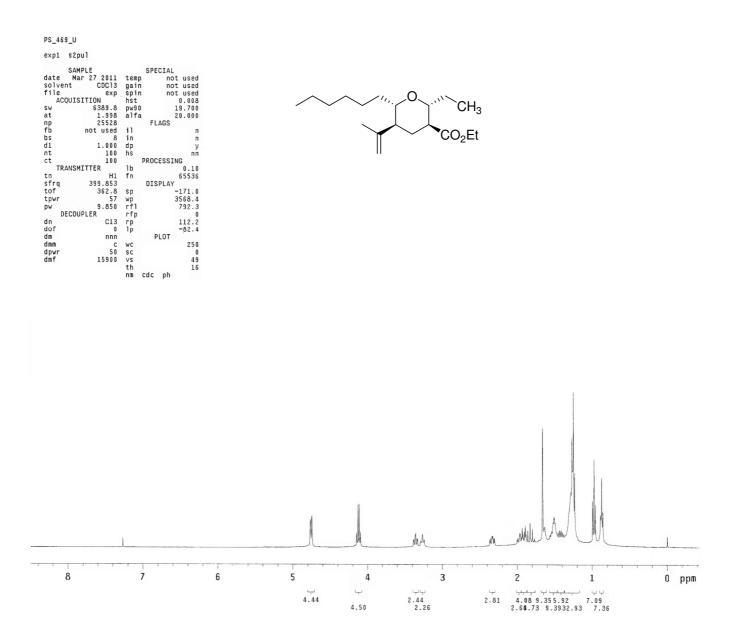




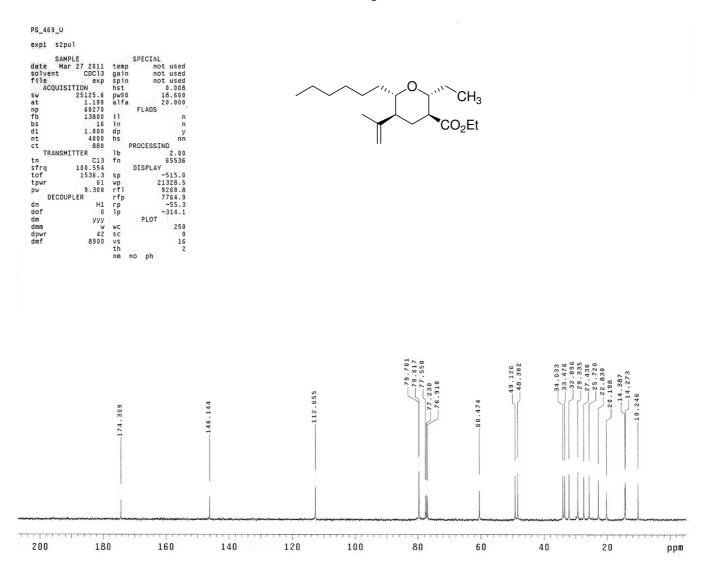




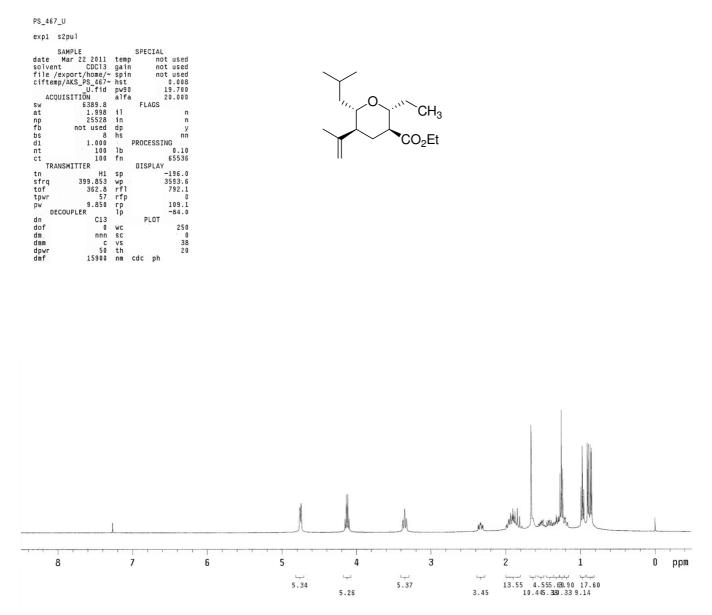
# <sup>1</sup>H NMR spectra of 30



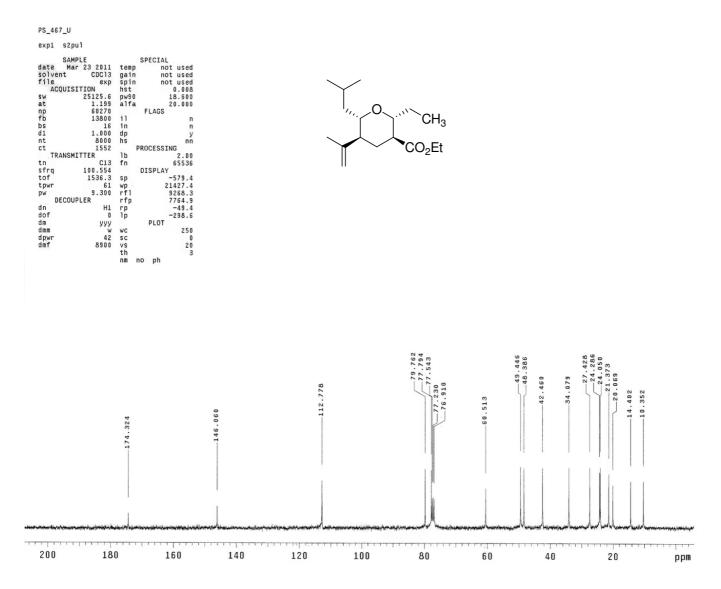
<sup>13</sup>C NMR spectra of 30



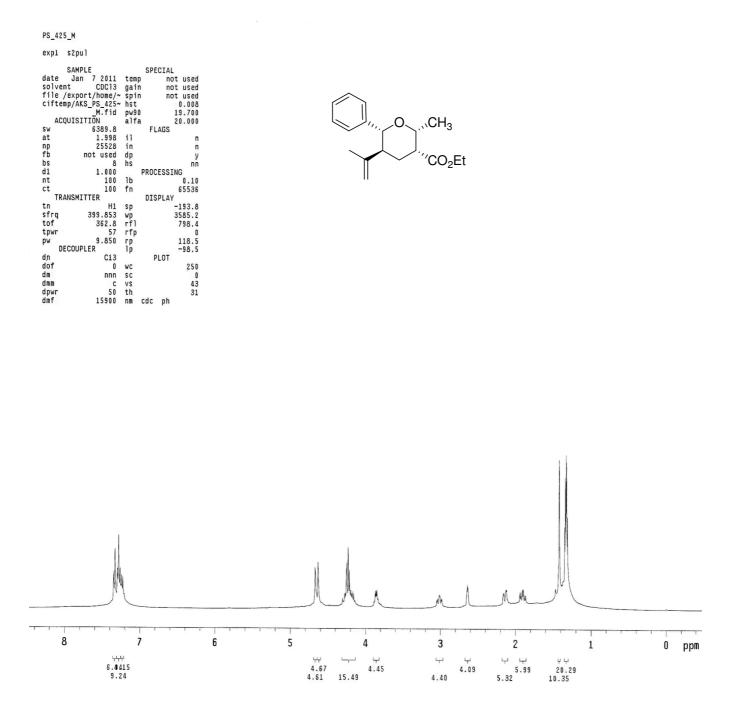
<sup>1</sup>H NMR spectra of 3p



<sup>13</sup>C NMR spectra of 3p

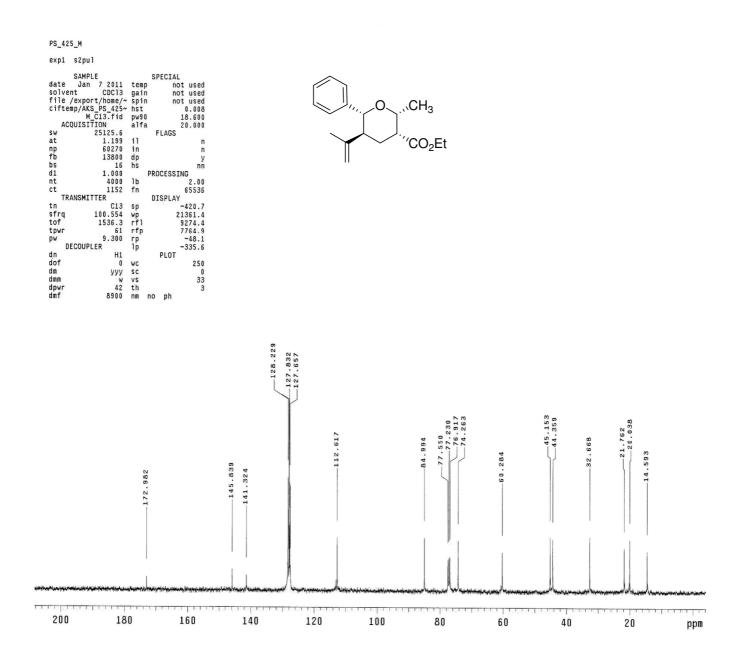


# <sup>1</sup>H NMR spectra of 4a

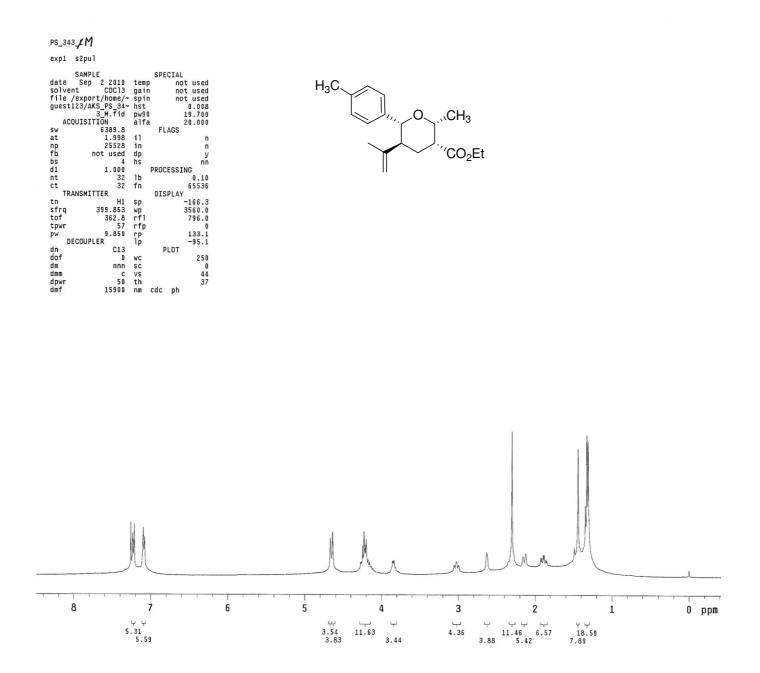


S-55

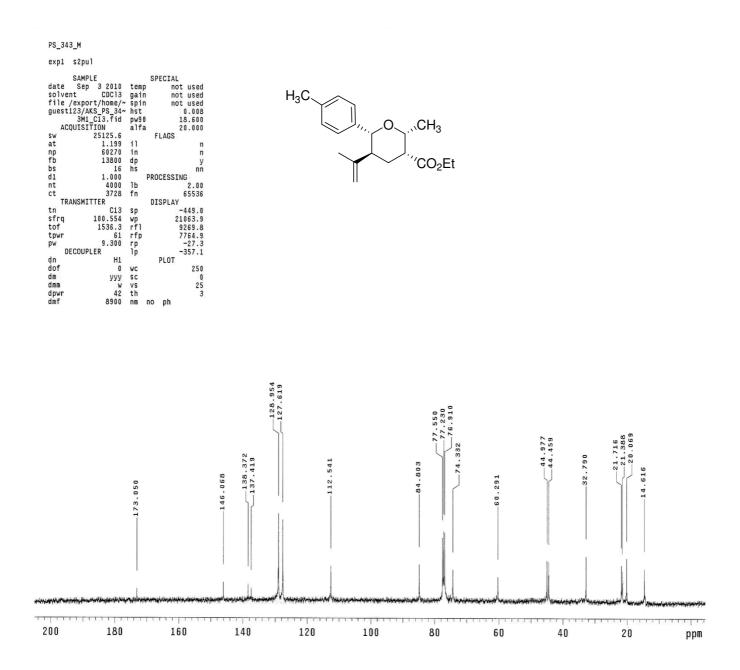
# <sup>13</sup>C NMR spectra of 4a



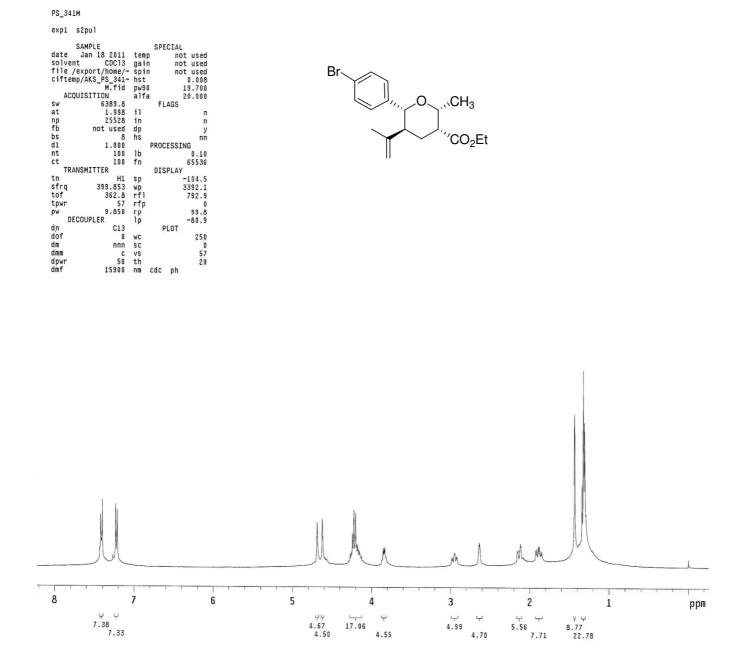
#### <sup>1</sup>H NMR spectra of 4b



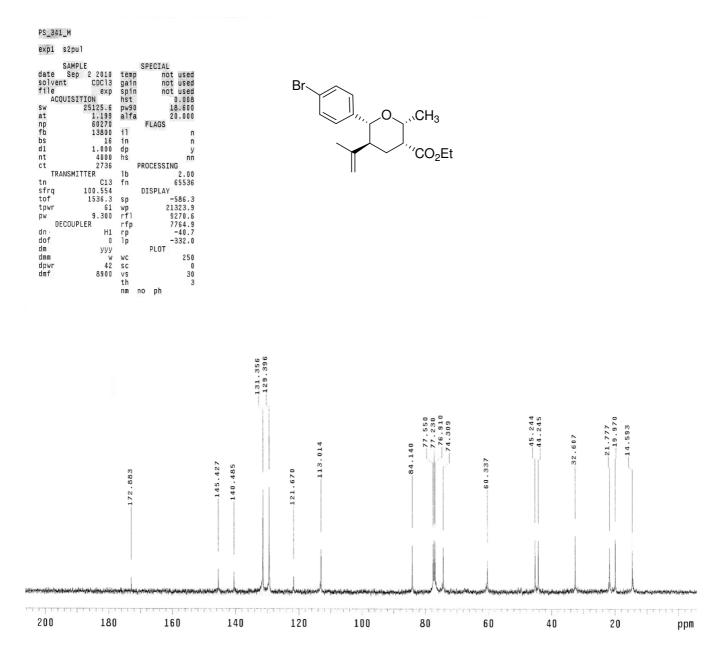
# <sup>13</sup>C NMR spectra of 4b



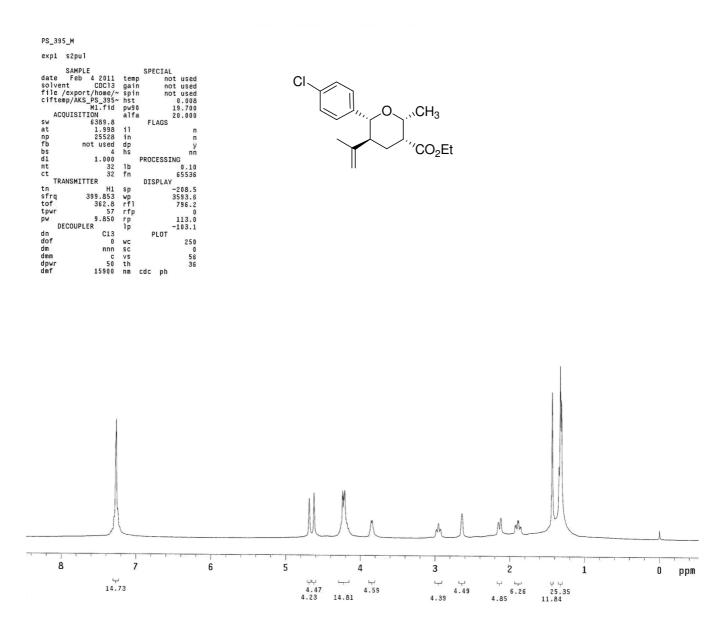
#### <sup>1</sup>H NMR spectra of 4c



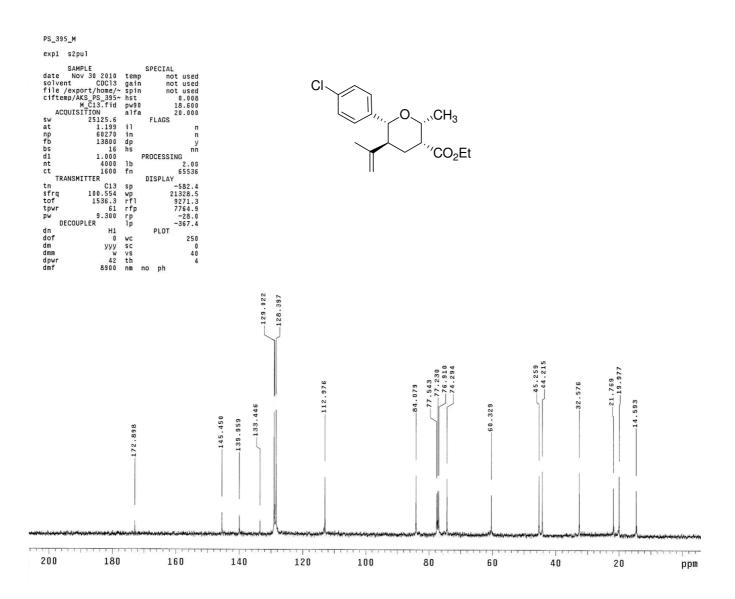
<sup>13</sup>C NMR spectra of 4c



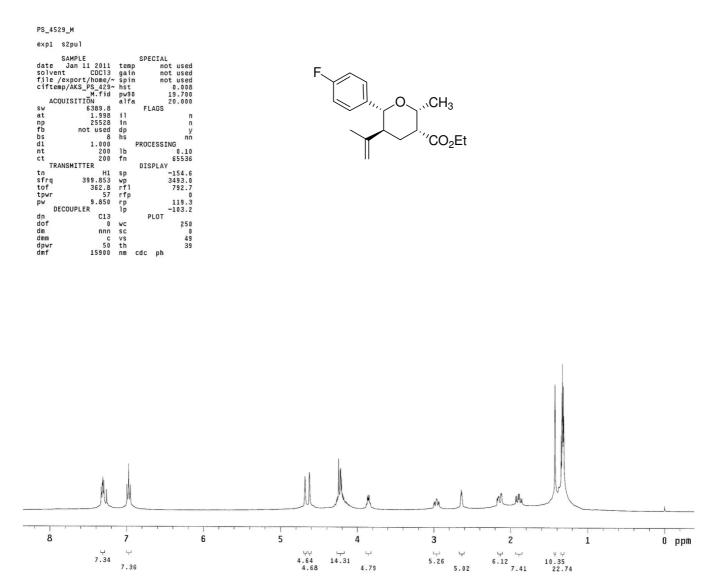
#### <sup>1</sup>H NMR spectra of 4d



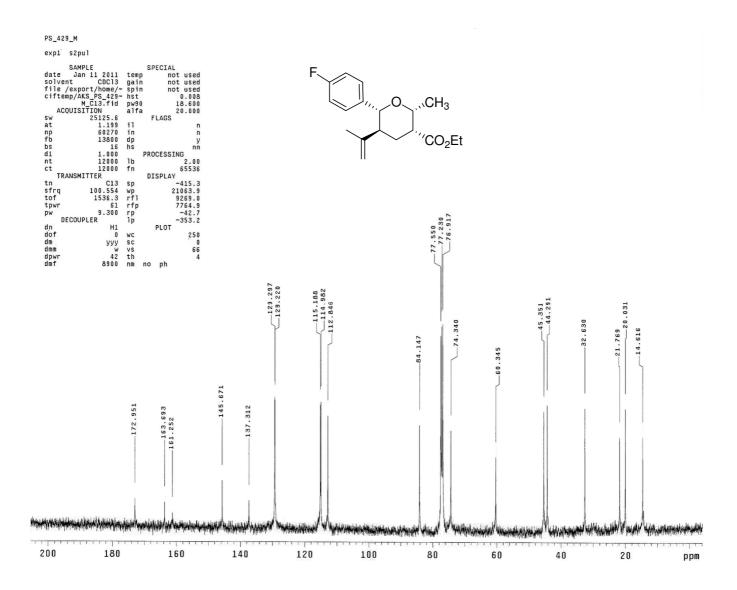
<sup>13</sup>C NMR spectra of 4d



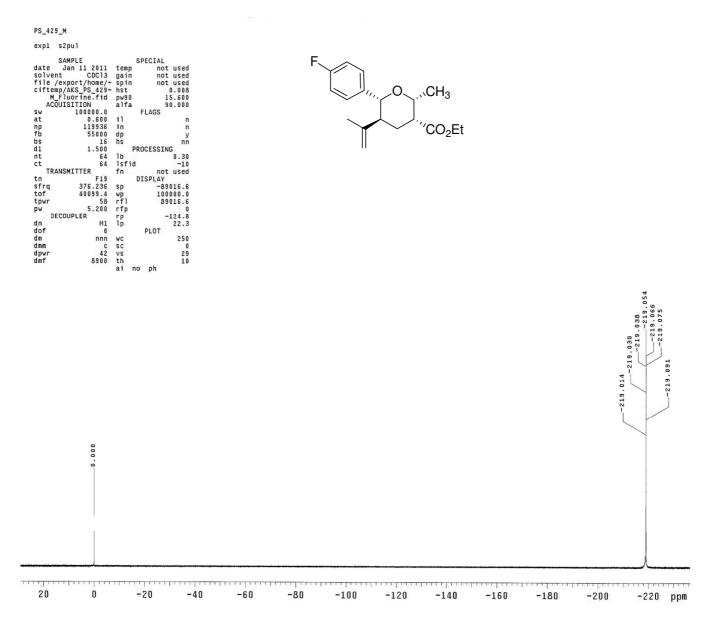
<sup>1</sup>H NMR spectra of 4e



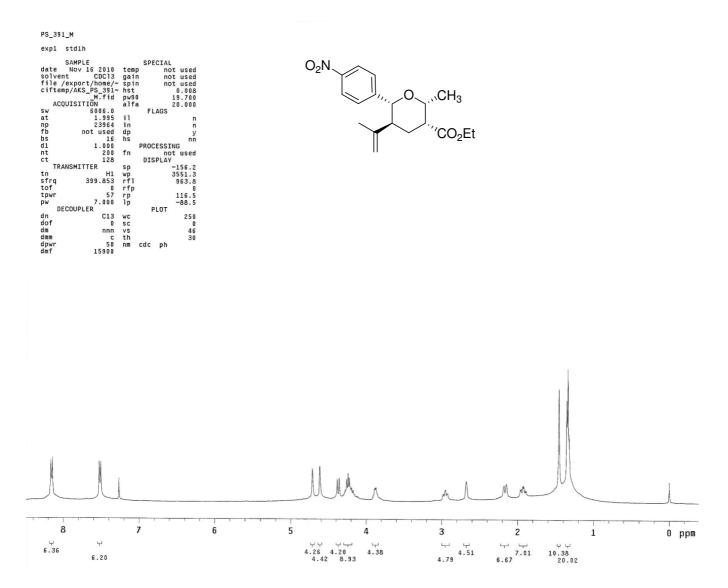
# <sup>13</sup>C NMR spectra of 4e

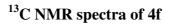


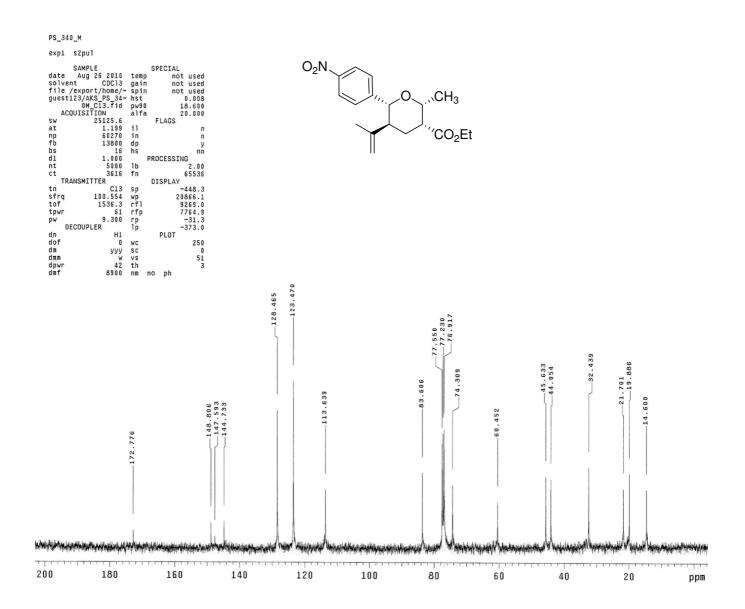
<sup>19</sup>F NMR spectra of 4e



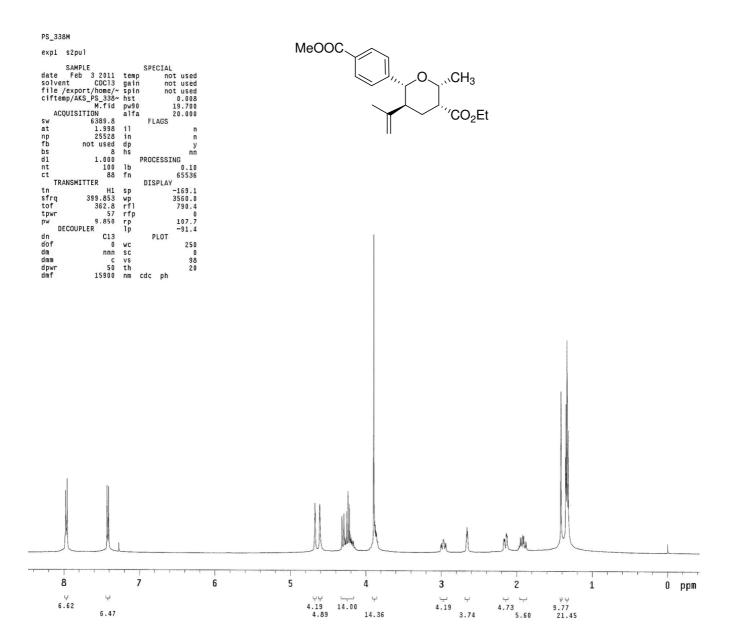
<sup>1</sup>H NMR spectra of 4f



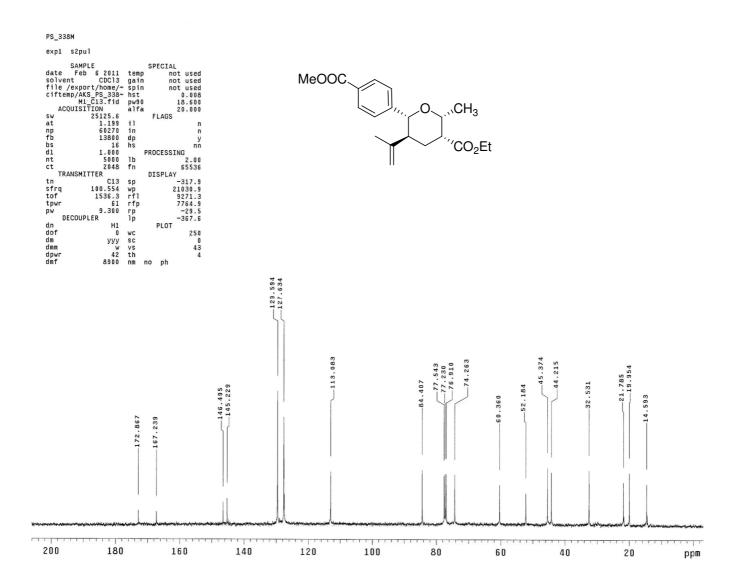




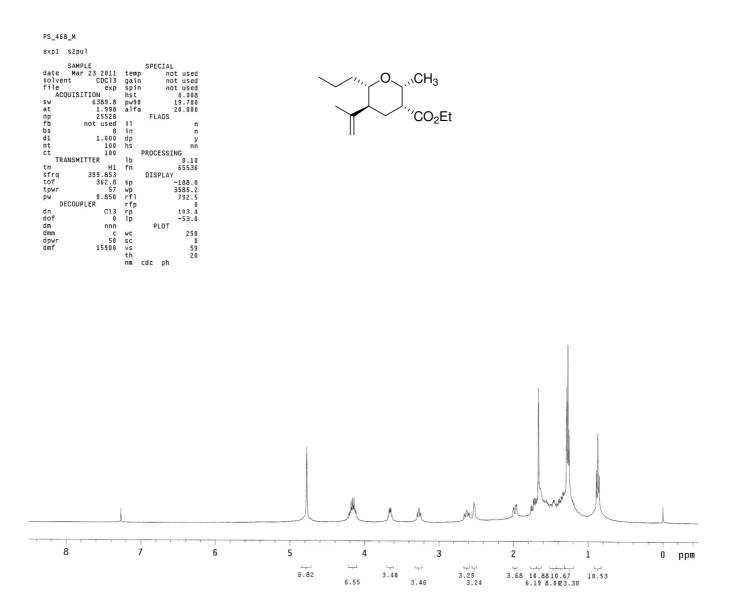
#### <sup>1</sup>H NMR spectra of 4g

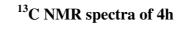


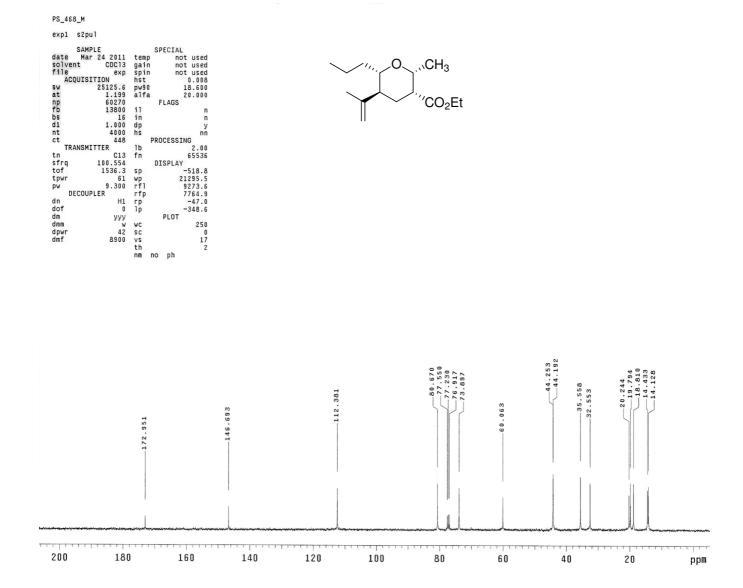
<sup>13</sup>C NMR spectra of 4g



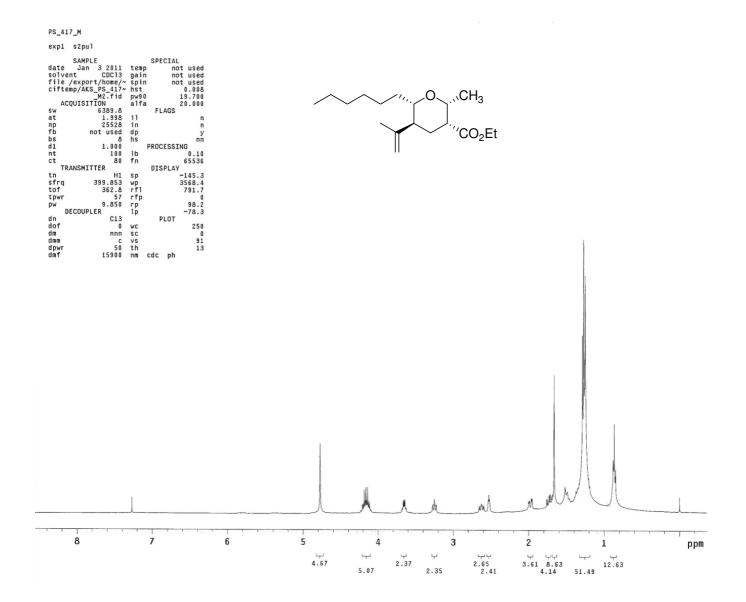


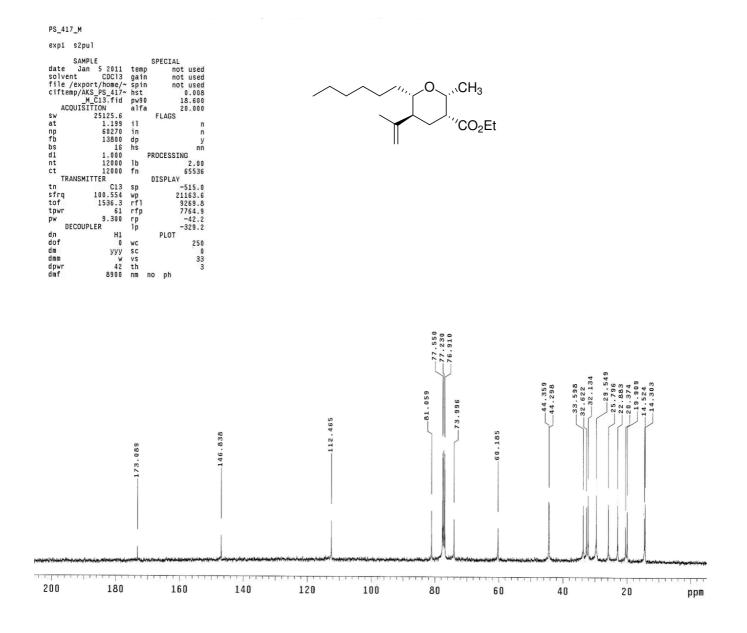






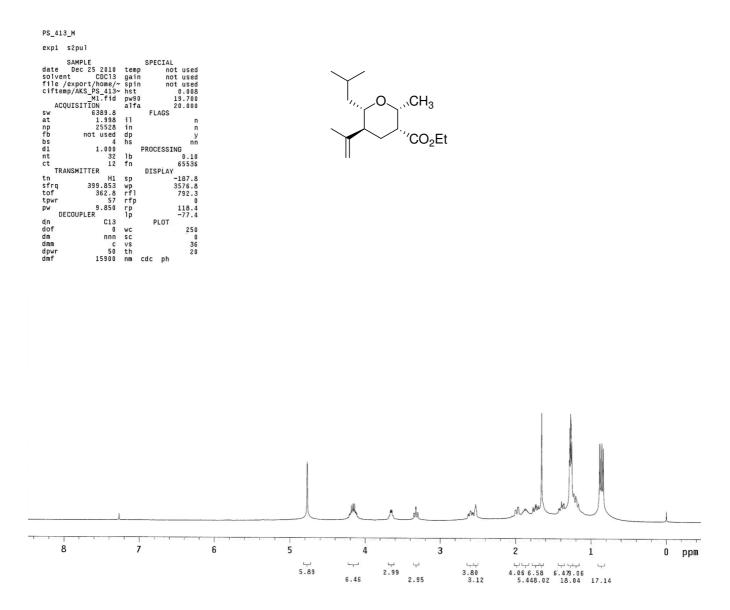
<sup>1</sup>H NMR spectra of 4i

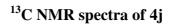


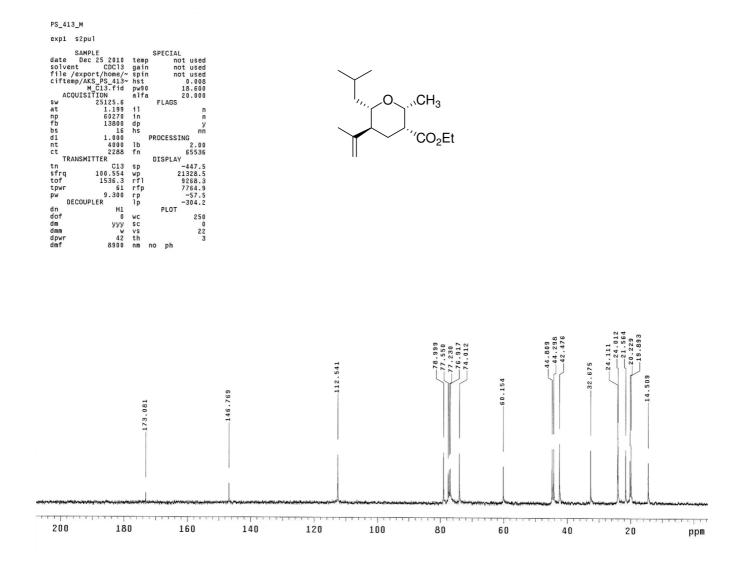


# <sup>13</sup>C NMR spectra of 4i



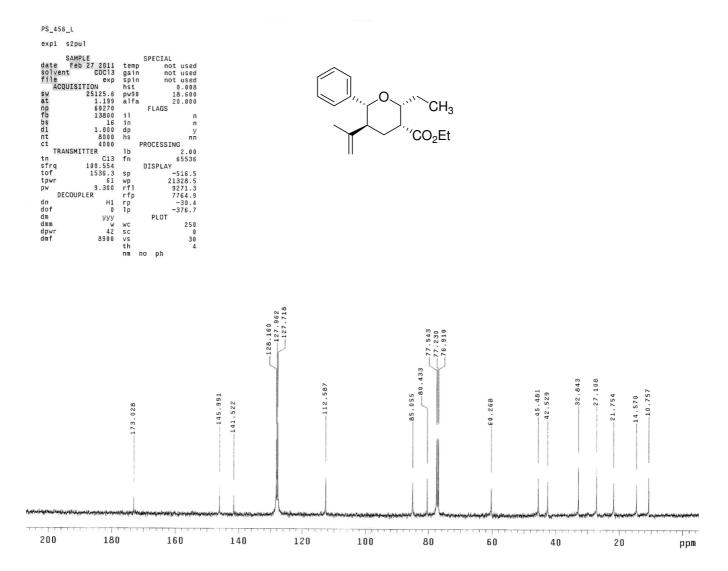




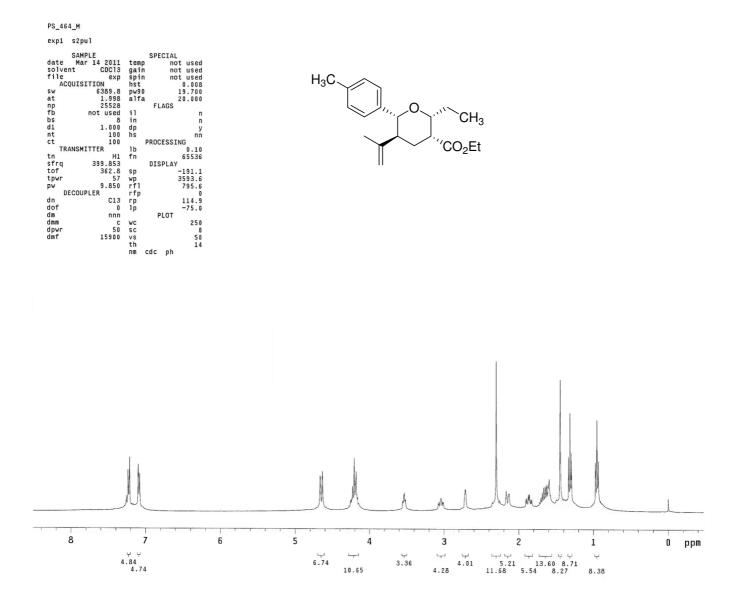




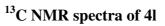


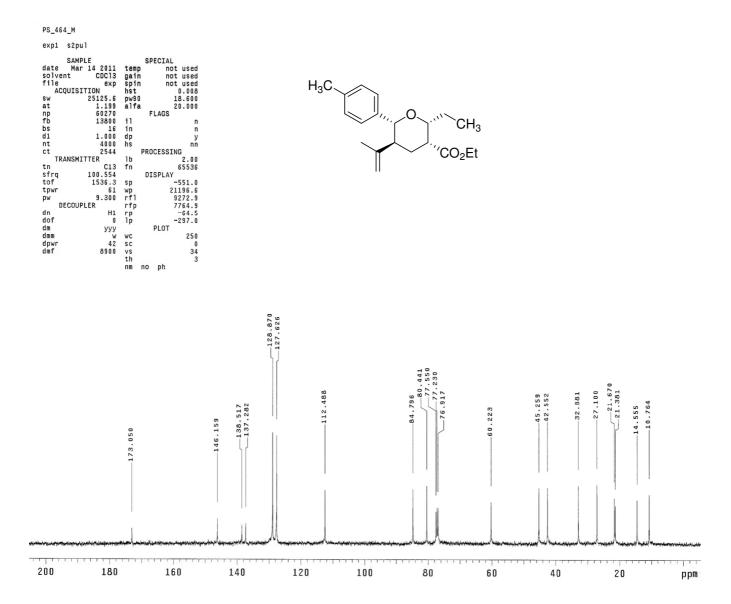


<sup>13</sup>C NMR spectra of 4k

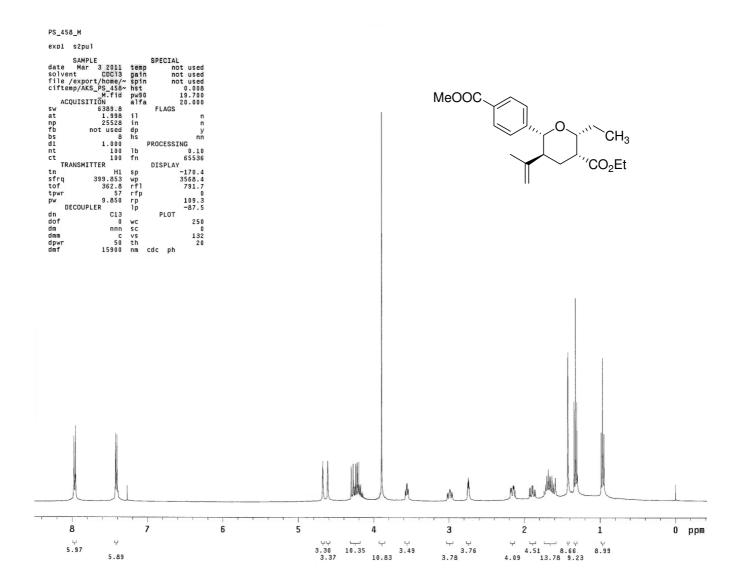


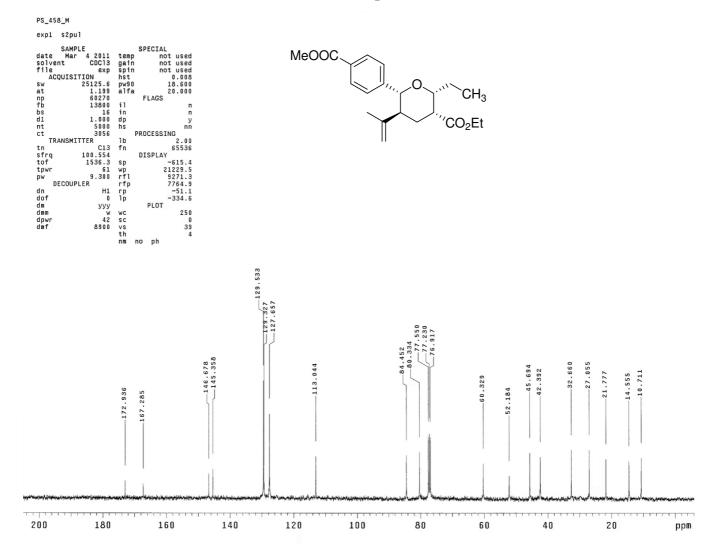
### <sup>1</sup>H NMR spectra of 4l



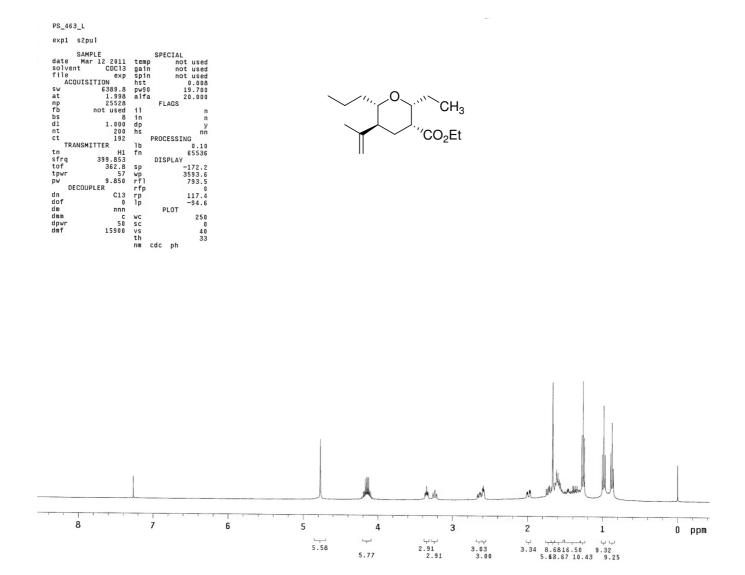


### <sup>1</sup>H NMR spectra of 4m



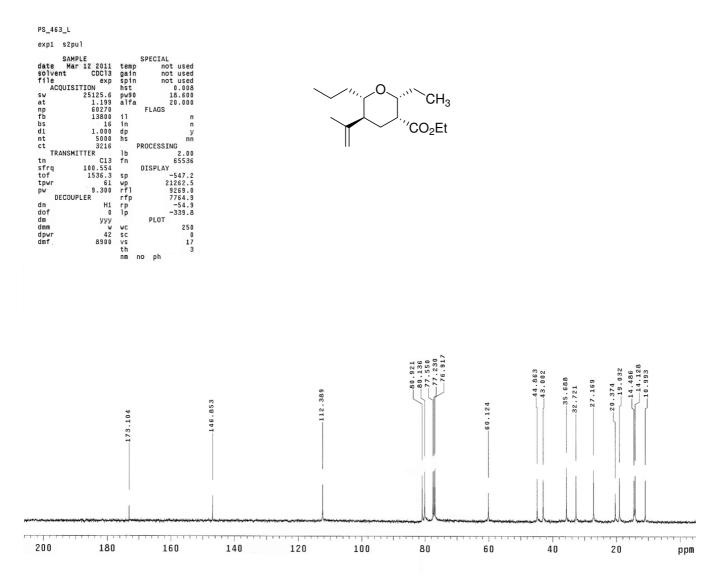


# <sup>13</sup>C NMR spectra of 4m

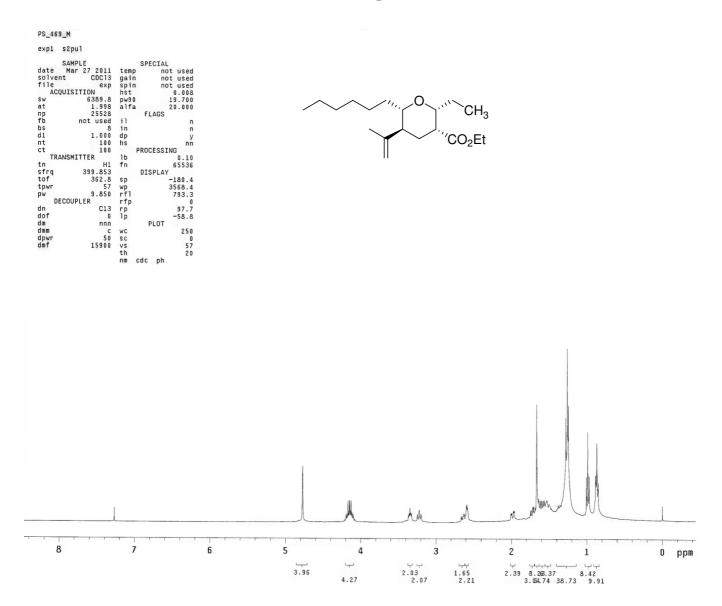


### <sup>1</sup>H NMR spectra of 4n

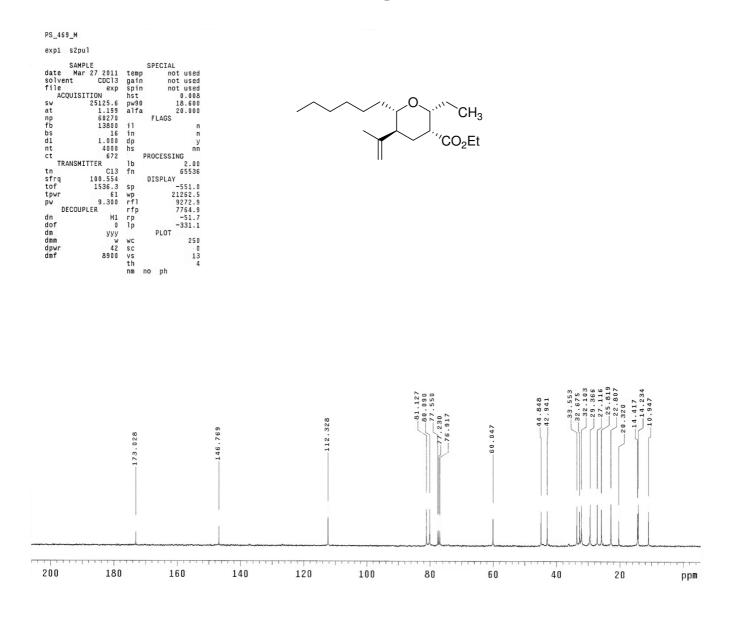
<sup>13</sup>C NMR spectra of 4n



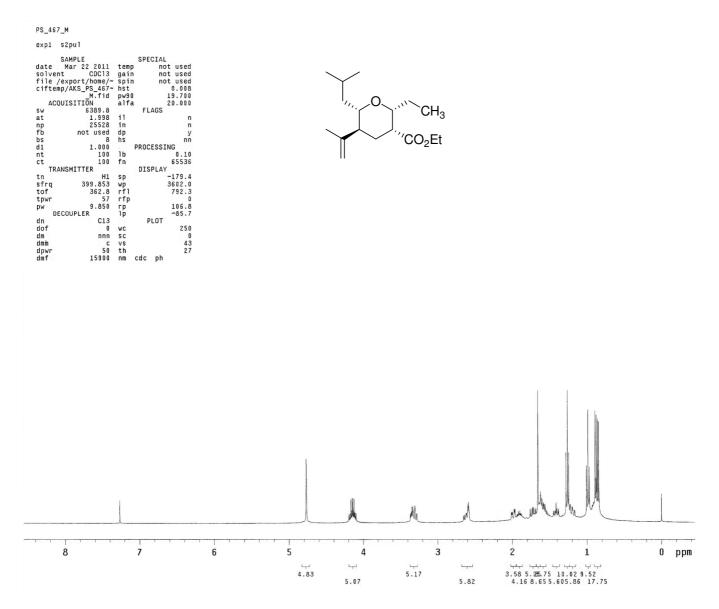
<sup>1</sup>H NMR spectra of 40



<sup>13</sup>C NMR spectra of 40



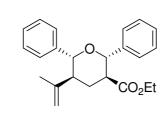
### <sup>1</sup>H NMR spectra of 4p



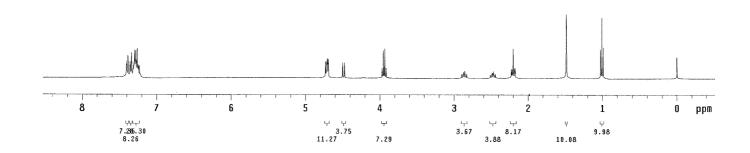
<sup>13</sup>C NMR spectra of 4p

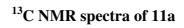


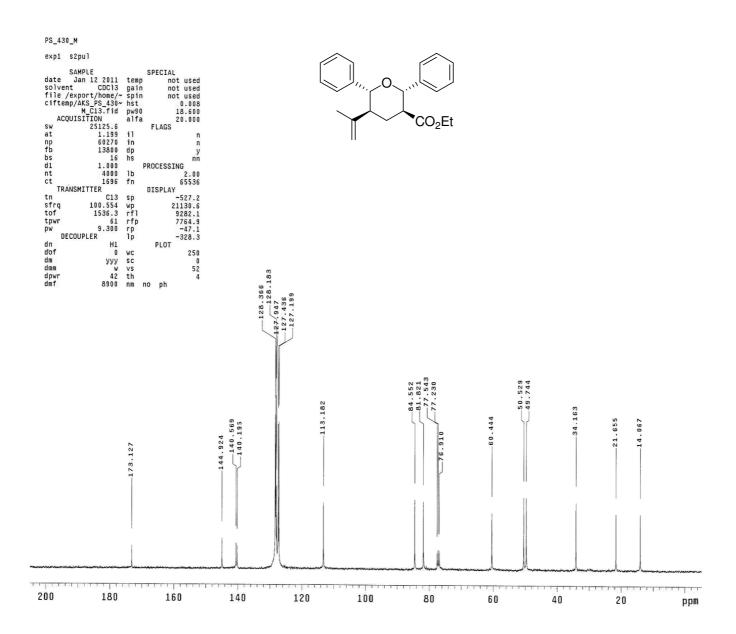
# <sup>1</sup>H NMR spectra of 11a



exp1	\$2pu1		
	SAMPLE		SPECIAL
date	Sep 7 2011	temp	not used
solver		gain	not used
file	/export/home/~	spin	not used
	np/AKS_PS_430~	hst	0.008
	M Pdt.fid	pw90	19.700
AC	QUISITION	alfa	20.000
SW	6389.8		FLAGS
at	1.998	11	n
np	25528	in	n
fb	not used	dp	У
bs	4	hs	nň
d1	1.000		PROCESSING
nt	200	fn	not used
ct	164		DISPLAY
TR/	ANSMITTER	sp	-207.1
tn	. H1	wp	3618.7
sfrq	399.853	rf1	794.5
tof	362.8	rfp	0
tpwr	57	rp	135.4
pw	9.850	1p	-84.7
DI	ECOUPLER	•	PLOT
dn	C13	WC	250
dof	0	SC	0
dm	nnn	VS	61
dmm	с	th	8
dpwr	50	nm	ph
dmf	15900		



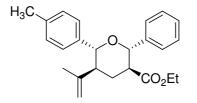


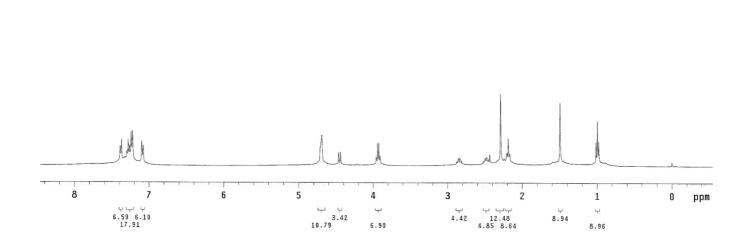


**S-89** 

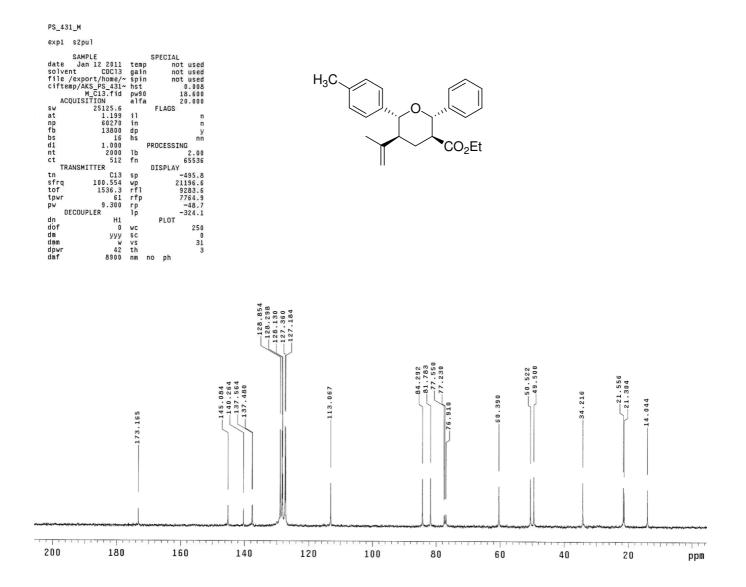
# <sup>1</sup>H NMR spectra of 11b

PS_431_M					
exp1	s2pul				
	SAMPLE		SPE	CIAL	
date	Jan 12 2011	temp	)	no	t used
solve	nt CDC13	gair		no	t used
file	/export/home/~	spir	1	no	
	mp/AKS PS 431~	hst	-		0.008
	M.fid	pw90	E Contraction of the second seco		19.700
AC	OUISITION	alfa			20.000
SW	6389.8			AGS	
at	1,998	i1			n
np	25528	in			n
fb	not used	dp			y
bs	8	hs			nn
d1	1.000		PROC	ESST	
nt	100	lb		2001	0.10
ct	100	fn			65536
	ANSMITTER		DIS	PLAY	05550
tn	H1	sp	013		-219.4
sfrq	399.853	wp			3602.0
tof	362.8	rf1			
	57				798.6
tpwr		rfp			0
pw	9.850 ECOUPLER	rp			109.2
		1 p			-82.3
dņ	C13		P	LOT	
dof	0	WC			250
dm	nnn	SC			0
dmm	С	VS			23
dpwr	50	th			24
dmf	15900	nm	cdc	ph	

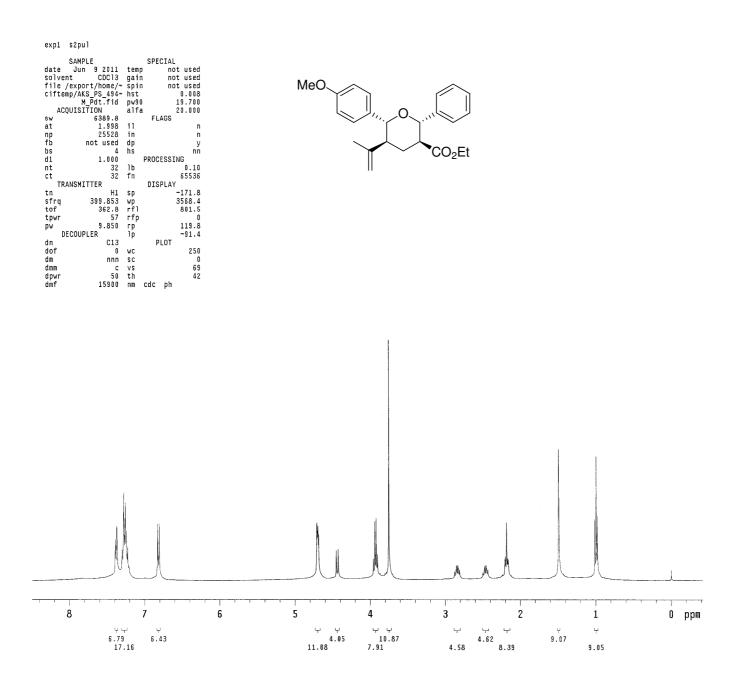




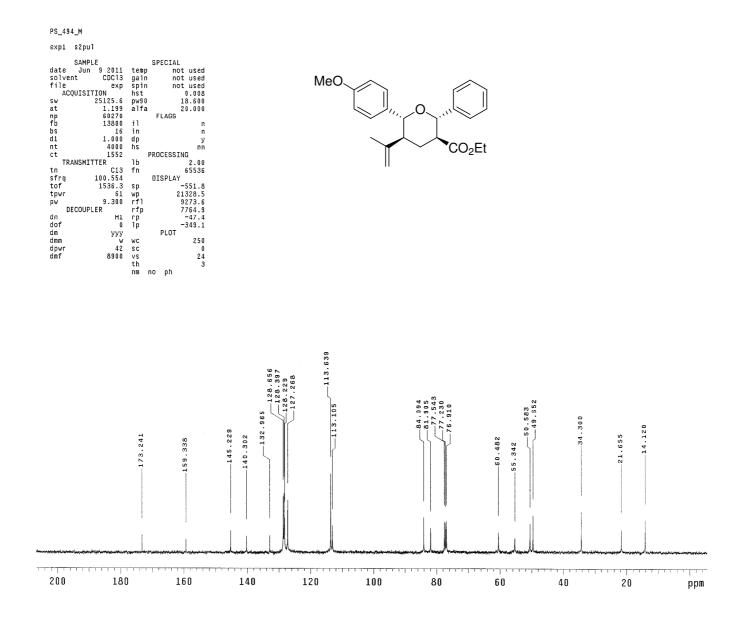
## <sup>13</sup>C NMR spectra of 11b



### <sup>1</sup>H NMR spectrta of 11c



## <sup>13</sup>C NMR spectra of 11c



SPECIAL not used not used 0.008 19.700 20.000 FLAGS

n n y nn

0.10 65536

Y -224.5 3652.5 795.2 0 119.0 -88.5

FLAGS

PROCESSING

DISPLAY

PLOT

cdc ph

sp vp rfl rfp rp lp

wc sc vs th nm

PS\_435\_M

exp1 s2pul

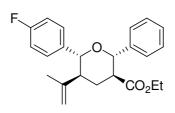
exp1 s2pul SAMPLE S date Sep.12 2011 temp solvent CDC13 gain file /export/home/~ spin ciftemp/AKS\_PS\_435~ hst ML\_Pdt.fid pw30 ACQUISITION aira sw 6389.8 at 1.998 il np 25528 in fb not used dp bs 8 hs d1 1.000 nt 300 lb ct 256 fn TANSMITTER HI Sp

H H 399.853 of 362.8 pwr 57 V 9.850 DECOUPLER

C13 0 7nn 50 15900

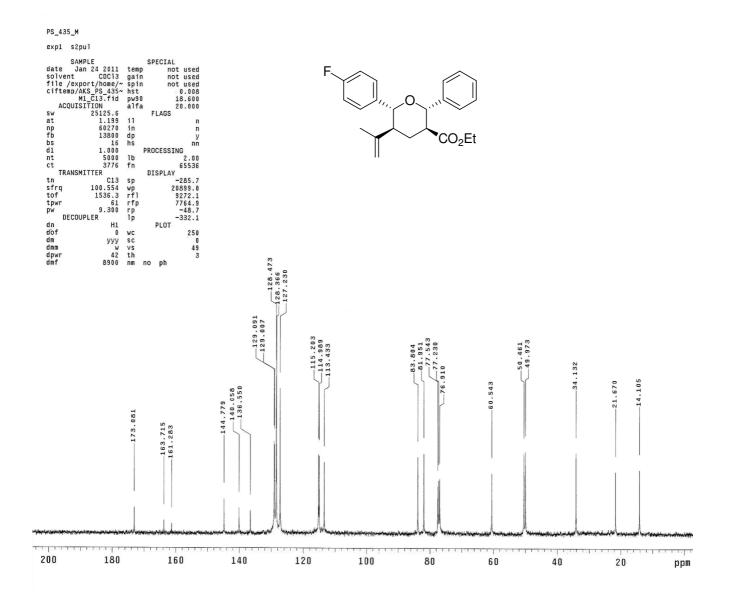
tn sfrq tof tpwr pw dn dof dm dmm dpwr dmf

### <sup>1</sup>H NMR spectra of 11d

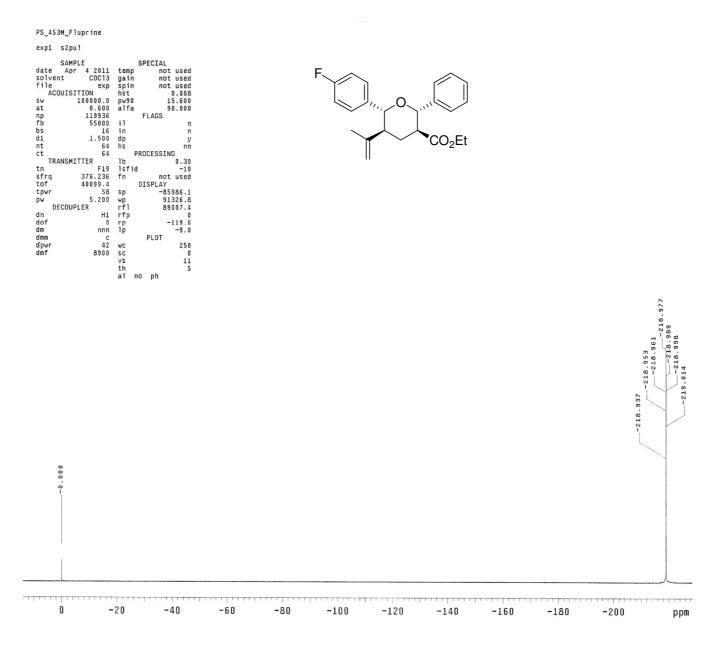


MAL M	M_uk			
8 7 6 پېپ ب 6.831.30 12.63.6.89	5 4 ب ب ب 11.09 7.36 3.63	3 2 4.80 4.41 9.15	1 y y 10.03 S.90	0 ppm

# <sup>13</sup>C NMR spectra of 11d



# <sup>19</sup>F NMR spectra of 11d



SPECIAL not used not used 0.008 19.700 20.000 FLAGS

n n y nn

0.10 65536

Y -146.6 3635.7 793.1 0 109.0 -81.3

FLAGS

PROCESSING

DISPLAY

PLOT

cdc ph

temp gain spin hst pw90 alfa

il in dp hs

1b fn

sp vp rfl rfp lp

wc sc vs th nm

PS\_450\_M exp1 s2pul

sw at np fb d1 nt ct

TF tn sfrq tof tpwr pw dn dof dm dmm dpwr dmf

SAMPLE date Feb 20 2011 solvent CDC13 file /export/home/-ciftemp/AKS\_PS\_450-fid

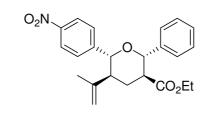
ftemp/AKS\_PS\_450-M.fid ACQUISITION 6389.8 1.998 25528 not used 8 1.000 2000 TRANSMITTER

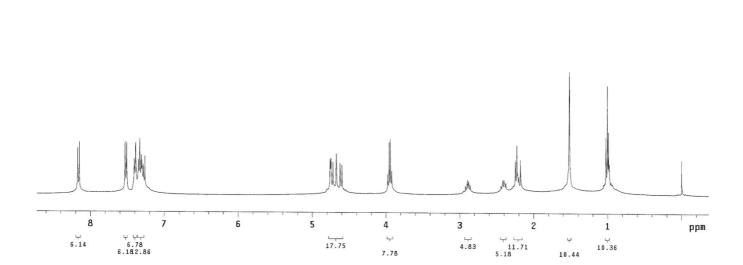
H Frq 399.853 f 362.8 Wr 57 9.850 DECOUPLER

R C13 0 nnn c 50 15900

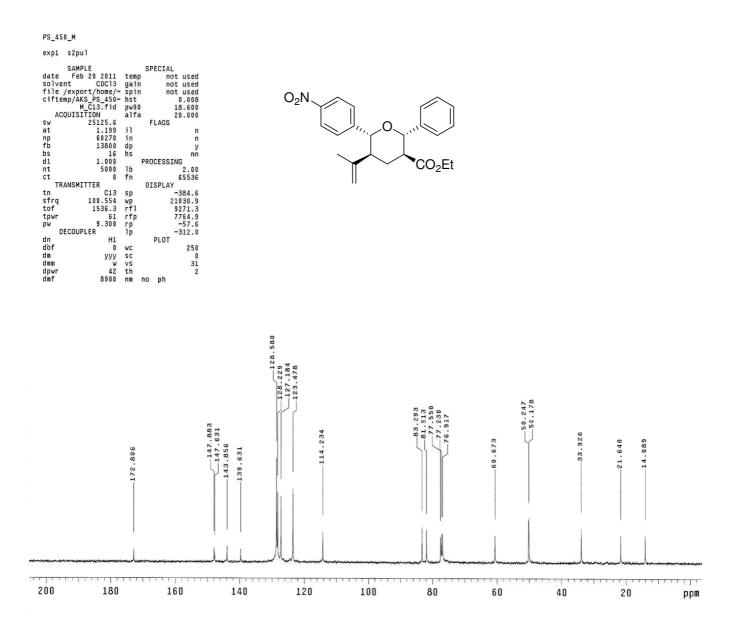
TRANSMITTER

### <sup>1</sup>H NMR spectra of 11e



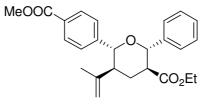


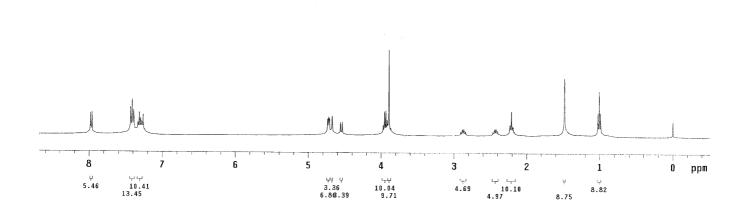
# <sup>13</sup>C NMR spectra of 11e



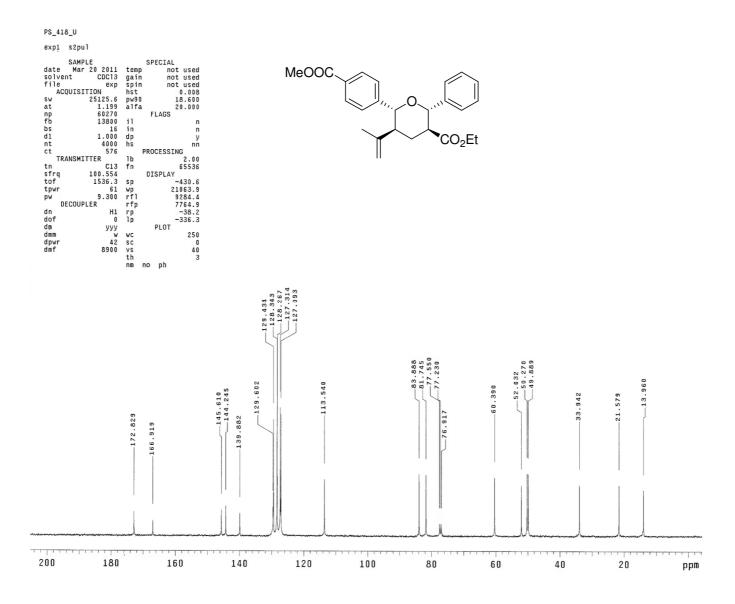
## <sup>1</sup>H NMR spectra of 11f

PS_418	3_U	
exp1	s2pu1	
	SAMPLE	SPECIAL
date	Sep 8 2011	temp not used
solver	nt CDC13	gain not used
file	exp	spin not used
ACC	DUISITION	hst 0.008
sw	6389.8	pw90 19,700
at	1.998	alfa 20.000
np	25528	FLAGS
fb	not used	il n
bs		in n
d1	1.000	dp y
nt	200	hs nn
čt	144	PROCESSING
TR	NSMITTER	1b 0.10
th	HI	fn 65536
sfrq	399.853	DISPLAY
tof	362.8	sp -206.9
tpwr	57	wp 3686.0
pw	9.850	rfl 794.7
	COUPLER	rfp 0
dn	C13	rp 124.3
dof	0	lp +82.2
dm	กกก	PLOT
dmm	c	wc 250
dpwr	5.0	sc 0
dmf	15900	vs 28
em1	13300	th 20
		nna cdc ph









S-100

SPECIAL not used not used 0.008 19.700 20.000

n n y nn

0.10 65536

-188.6 3568.4 793.1 103.0 -90.3

FLAGS

PROCESSING

DISPLAY

PLOT

cdc ph

temp gain spin hst pw90 alfa

il in dp hs

1b fn

sp wp rfl rfp rp lp

wc sc vs th nm

C13 0 nnn

C 50 15900

PS\_424\_M exp1 s2pul

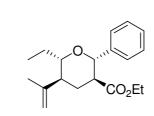
sw at np fb bs d1 nt ct tf tf tof tpwr pw dn dof dmm dpwr dmf

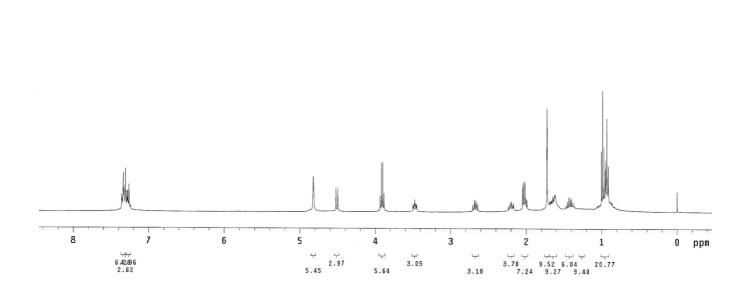
SAMPLE date Jan 5 2011 solvent CDC13 file /export/home/~ ciftemp/AKS\_PS\_424~ M.fid ACQUISITION sw 6389.8 at 1.998 np 25528 fb not used bs 8 dl 1.000 nt 100 ct 100 TRANSMITTER

TRANSMITTER

H Frq 399,853 57 362.8 1Wr 57 DECOUPLER

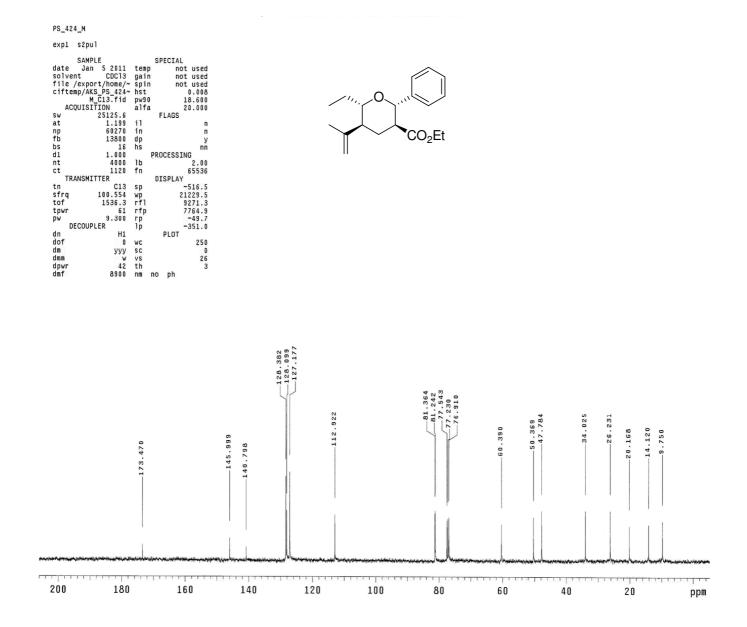
<sup>1</sup>H NMR spectra of 11g



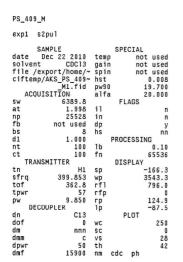


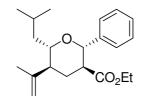
#### S-101

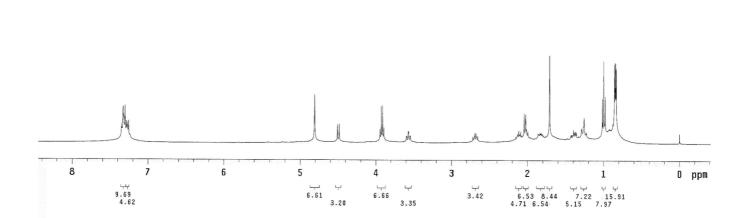
### <sup>13</sup>C NMR spectra of 11g



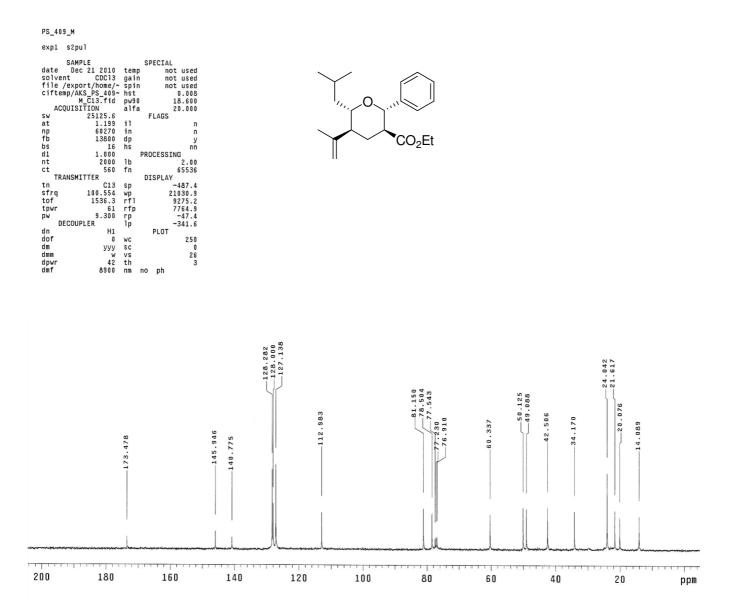
## <sup>1</sup>H NMR spectra of 11h





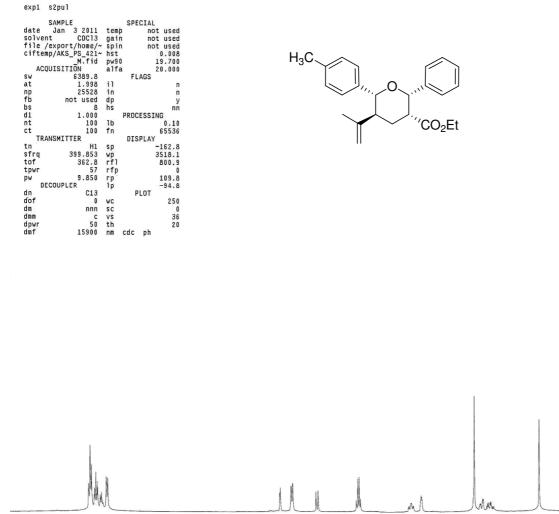


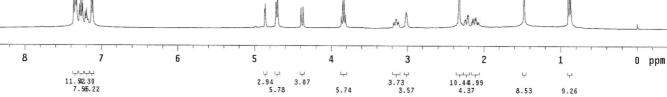
<sup>13</sup>C NMR spectra of 11h



PS\_421\_M

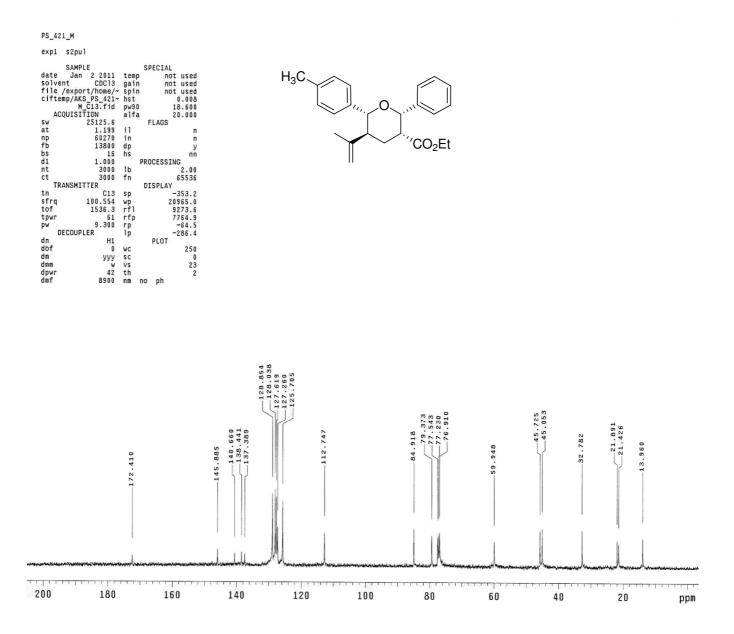




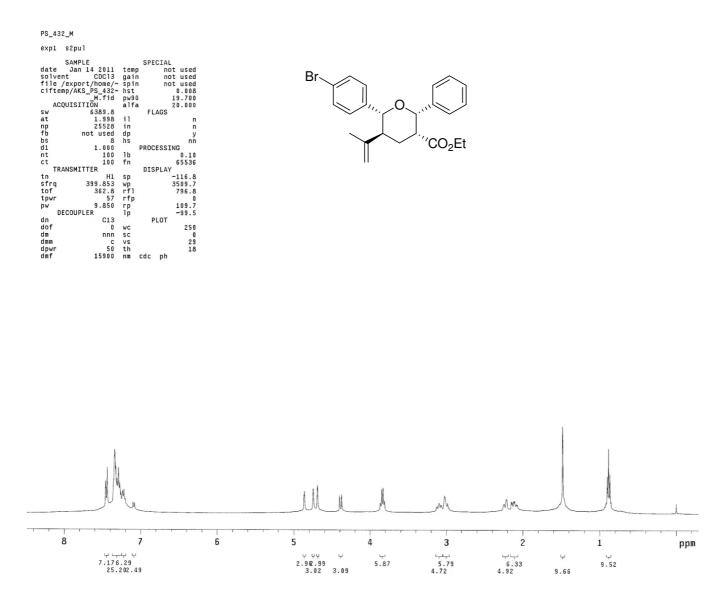


#### S-105

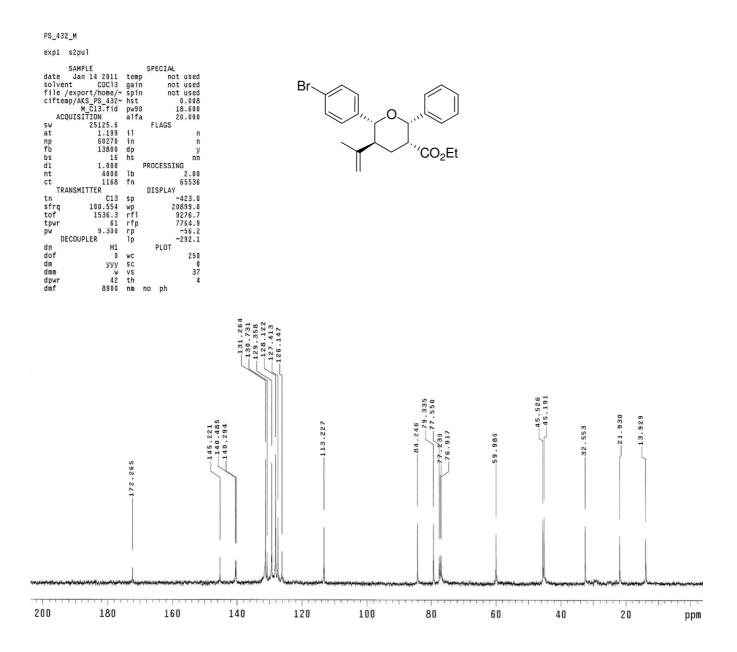
# <sup>13</sup>C NMR spectra of 12a



### <sup>1</sup>H NMR spectra of 12b

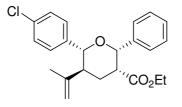


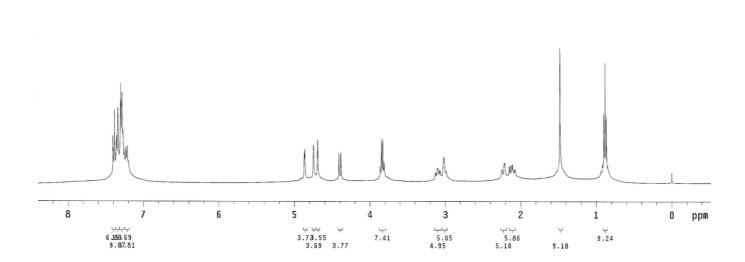
### <sup>13</sup>C NMR spectra of 12b



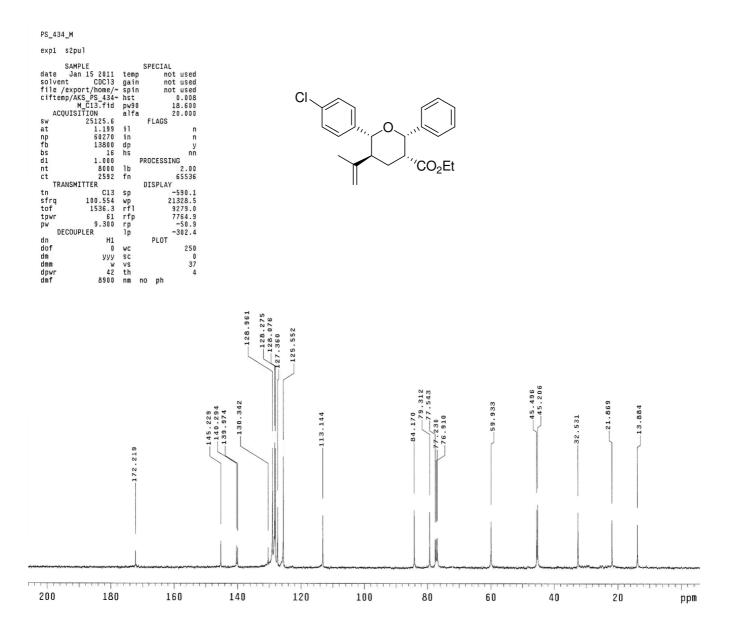
## <sup>1</sup>H NMR spectra of 12c

PS_43	4_M		
exp1	s2pu1		
	SAMPLE		SPECIAL
date	Jan 15 2011	temp	not used
solve	nt CDC13	gain	not used
	/export/home/~	spin	
cifter	mp/AKS_PS_434~	hst	0.008
	_M.fid	pw90	19.700
AC	QUISITION	alfa	
SW	6389.8		FLAGS
at	1.998	<b>i</b> 1	n
np	25528	in	n
fb	not used	dp	У
bs	8	hs	nn
d1	1.000		PROCESSING
nt	100	1b	0.10
ct	100	fn	65536
	ANSMITTER		DISPLAY
tn	H1	sp	-209.6
sfrq	399.853	wp	3568.4
tof	362.8	rfl	797.4
tpwr	57	rfp	0
pw	9.850	rp	115.2
	ECOUPLER	1p	-109.8
dn	C13		PLOT
dof	0	WC	250
dm	nnn	SC	0
dmm	С	VS	42
dpwr	50	th	20
dmf	15900	nm	cdc ph

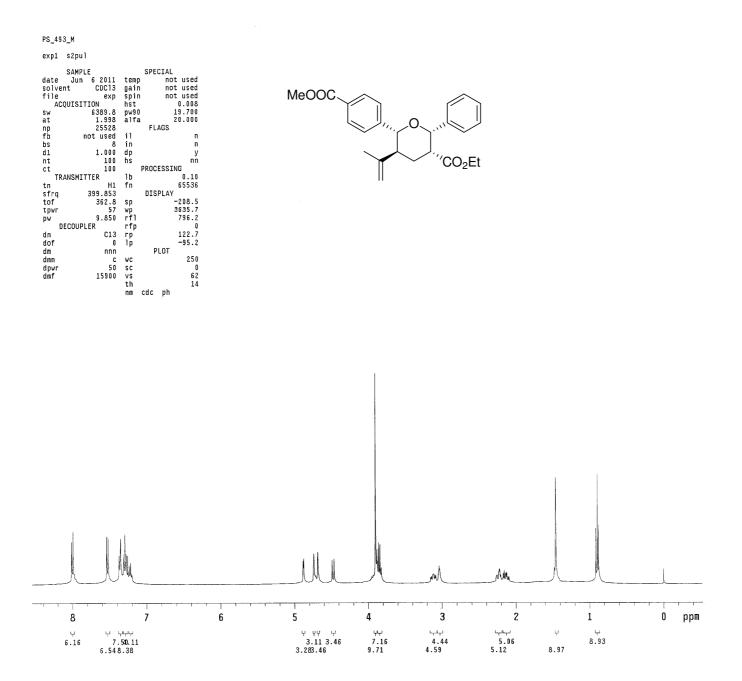




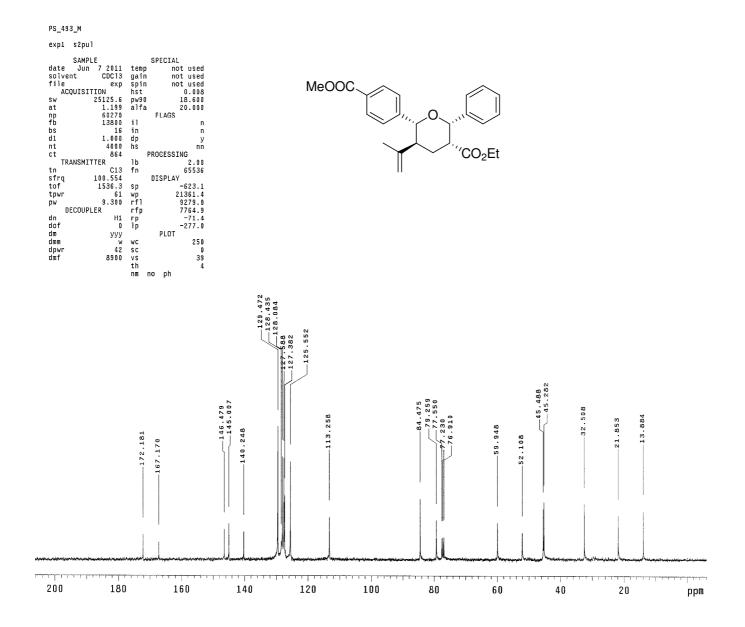
<sup>13</sup>C NMR spectra of 12c



### <sup>1</sup>H NMR spectra of 12d



<sup>13</sup>C NMR spectra of 12d



SPECIAL not used not used 0.008 19.700 20.000 FLAGS

n n

nn

0.10

-175.1 3568.4 796.4 0

0 119.8 -83.9

FLAGS

PROCESSING

DISPLAY

PLOT

cdc ph

temp gain spin hst pw90 alfa

il in dp hs

1b fn

sp wp rfl rfp Ip lp

R 1p C13 0 wc nnn sc c vs 50 th 15900 nm

PS\_411\_M exp1 s2pul

sw at np fb bs d1 nt ct tf tf tof tpwr pw dn dof dmm dpwr dmf

 SAMPLE

 Gate
 Dec 22 2010

 solvent
 CDC13

 file /export/home/~
 ciftemp/AKS\_PS\_411~

 M.fid
 ACQUISITION

 sw
 688.8

 at
 1.998

 np
 25528

 fb
 not used

 bs
 8

 di
 1.000

 nt
 100

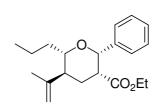
 Ct
 100

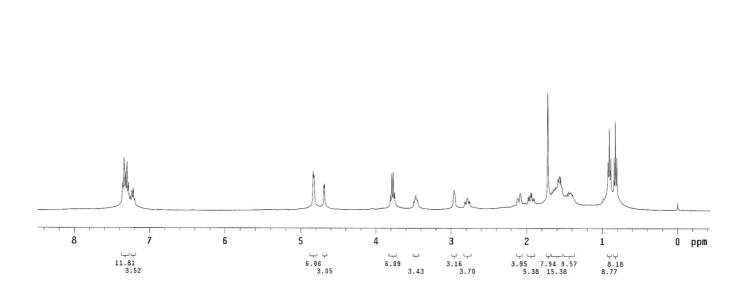
 TRANSMITTER
 110

TRANSMITTER

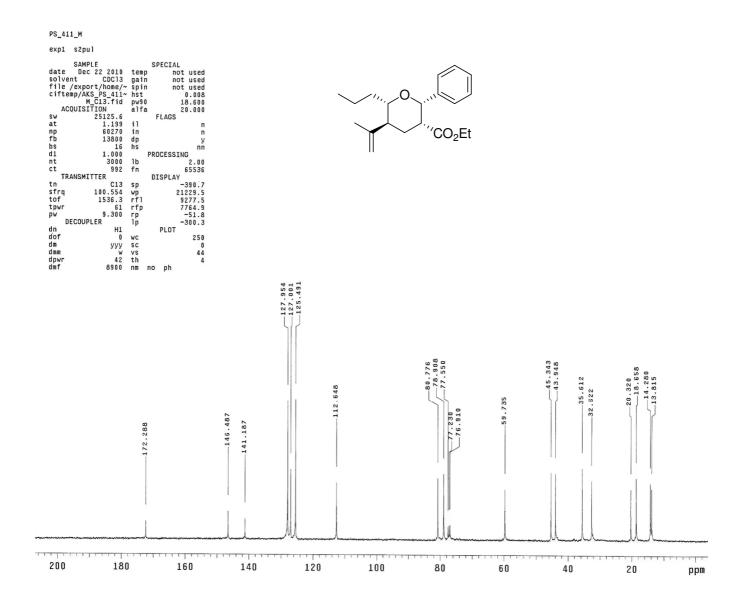
rq 399.853 f 362.8 wr 57 DECOUPLER

# <sup>1</sup>H NMR spectra of 12e

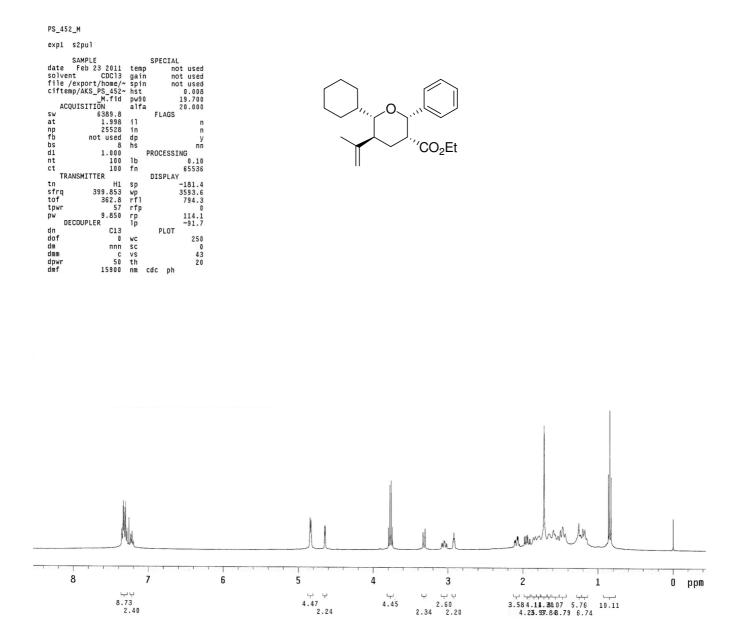




<sup>13</sup>C NMR spectra of 12e



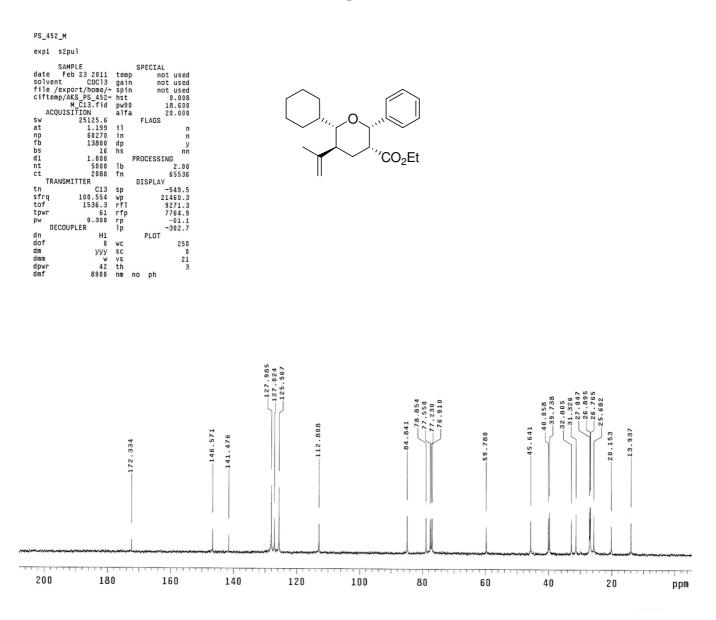
<sup>1</sup>H NMR spectra of 12f



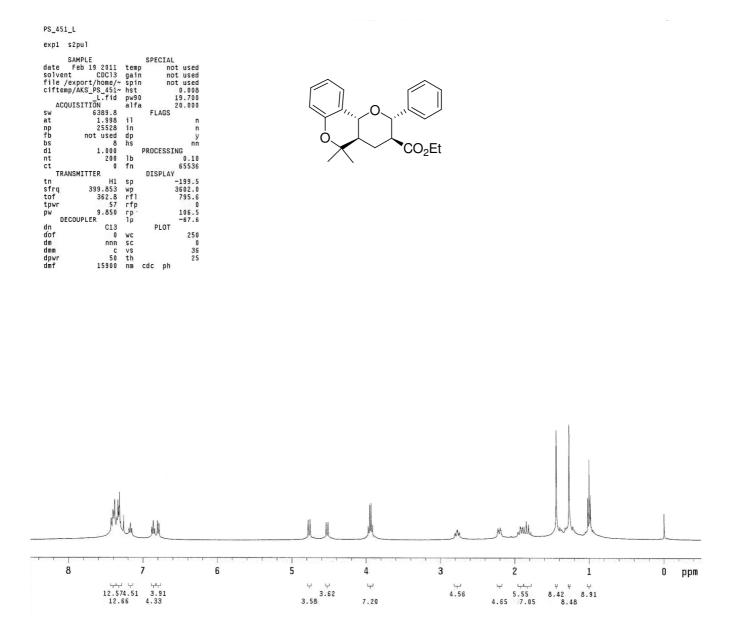
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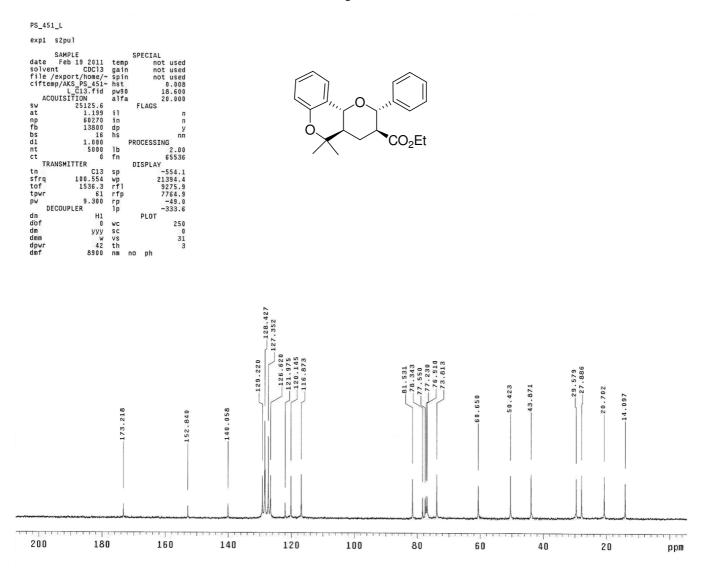
<sup>13</sup>C NMR spectra of 12f



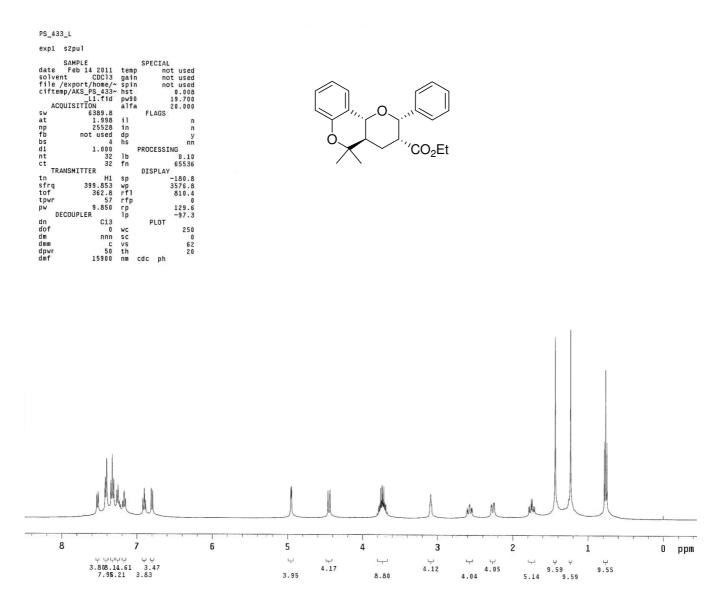
### <sup>1</sup>H NMR spectra of 13



<sup>13</sup>C NMR spectra of 13

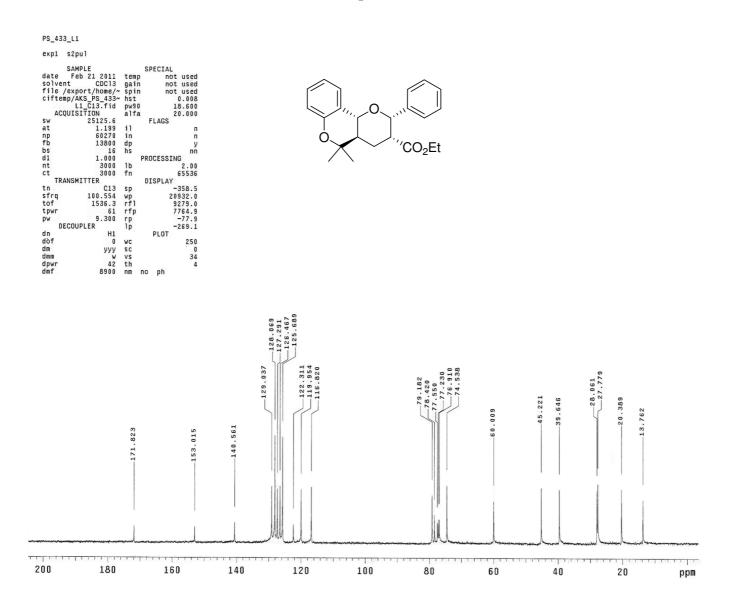


<sup>1</sup>H NMR spectra of 14



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<sup>13</sup>C NMR spectra of 14



	4g-CCDC 819363
Formula	$C_{20} H_{26}O_5$
Formula weight	346.41
T/K	296(2)
Crystal system	Orthorhombic
Space group	P n a 21
a/Å	19.8370(10)
b/Å	15.2026(8)
c/Å	6.3744(4)
α/°	90.00
βľ°	90.00
$\gamma/^{\circ}$	90.00
ŃÅ <sup>3</sup>	1922.35(18)
Z	4
Abs. Coeff./mm <sup>-1</sup>	0.085
Abs. Correction	multi-scan
GOF on $F^2$	0.943
Final <i>R</i> indices $[I > 2\sigma(I)]$	R1 = 0.0677
	wR2 = 0.1644
R indices [all data]	R1 = 0.0856
	wR2 = 0.1735

## The crystal parameters of compound 4g

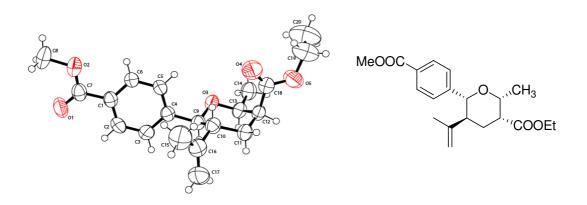


Figure 1: ORTEP diagram of 4g

	<b>11f</b> -CCDC 819365
Formula	$C_{25} H_{28} O_5$
Formula weight	408.47
T/K	296(2)
Crystal system	Monoclinic
Space group	P 21/c
aĺÅ	12.0752(3)
b/Å	15.3970(4)
c/Å	12.4647(3)
α/°	90.00
β/°	104.3910(10)
y/°	90.00
V/Å <sup>3</sup>	2244.74(10)
Ζ	4
Abs. Coeff./mm <sup>-1</sup>	0.083
Abs. Correction	multi-scan
GOF on $F^2$	1.070
Final <i>R</i> indices $[I > 2\sigma(I)]$	<i>R1</i> =0.0615
	wR2 = 0.1806
R indices [all data]	R1 = 0.0759
	wR2 = 0.2203

### The crystal parameters of compound 11f

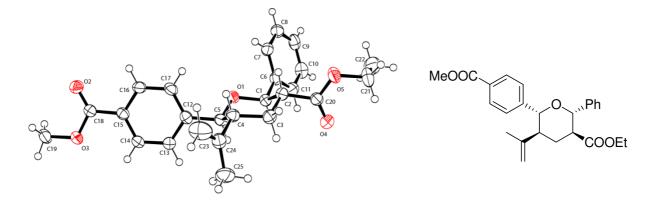
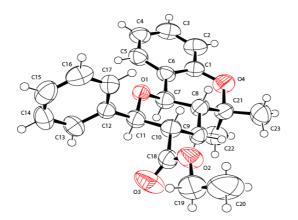


Figure 2: ORTEP diagram of 11f

	<b>13-</b> CCDC 819364
Formula	$C_{23}H_{26}O_4$
Formula weight	366.44
T/K	296(2)
Crystal system	Monoclinic
Space group	P 21/n
a/Å	8.9913(7)
b/Å	19.6654(14)
c/Å	11.4452(9)
α/°	90.00
β/°	103.239(4)
γ/°	90.00
V/Å <sup>3</sup>	1969.9(3)
Z	4
Abs. Coeff./mm <sup>-1</sup>	0.084
Abs. Correction	Multi-scan
GOF on $F^2$	1.064
Final <i>R</i> indices $[I > 2\sigma(I)]$	R1 = 0.0667
	wR2 = 0.1815
R indices [all data]	R1 = 0.0923
	wR2 = 0.1973

#### The crystal parameters of compound 13



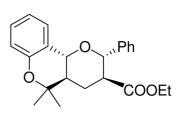


Figure 3: ORTEP diagram of 13

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	14-CCDC 819366
Formula	$C_{23}H_{26}O_4$
Formula weight	366.44
<i>T</i> /K	296(2)
Crystal system	Monoclinic
Space group	P 21/c
a/Å	12.5282(8)
b/Å	8.7884(6)
c/Å	18.2013(12)
α/°	90.00
βľ°	94.582(2)
γ/°	90.00
$V/Å^3$	1997.6(2)
Ζ	4
Abs. Coeff./mm <sup>-1</sup>	0.082
Abs. Correction	Multi-scan
GOF on $F^2$	1.373
Final <i>R</i> indices $[I > 2\sigma(I)]$	R1 = 0.0613
	wR2 = 0.1695
R indices [all data]	R1 = 0.0967
	wR2 = 0.2108

#### The crystal parameters of compound 14

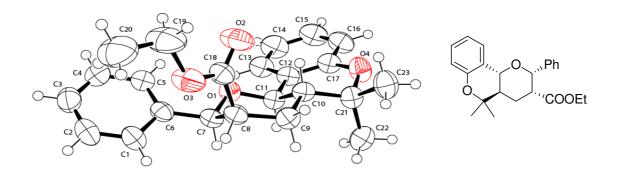


Figure 4: ORTEP diagram of 14