Catalytic regioselective synthesis of pyrazole based pyrido[2,3-*d*]pyrimidinediones and their biological evaluation

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Supporting information

1. Experimental part of biological evaluation

1.1 Instruments and Materials

All reactions were performed with commercially available reagents. They were used without further purification. The solvents used were of analytical grade. All reactions were monitored by thin-layer chromatography (TLC) on aluminum plates coated with silica gel 60 F₂₅₄, 0.25 mm thickness (Merck). Detection of the components was made by exposure to iodine vapors or UV light. Melting points were taken in melting point apparatus μ ThermoCal₁₀ (Analab Scientific Pvt. Ltd, India) and are uncorrected. The IR spectra were recorded in KBr on a Perkin-Elmer Spectrum GX FT-IR Spectrophotometer (Perkin-Elmer, USA) and only the characteristic peaks are reported in cm⁻¹. Mass spectra were recorded on Shimadzu LCMS 2010 spectrometer (Shimadzu, Tokyo, Japan) purchased under PURSE program of DST at Sardar Patel University, Vallabh Vidyanagar. The synthesized compounds were identified by 1H and 13C NMR spectra recorded in DMSO-d6 as a solvent on a Bruker Avance 400 MHz spectrometer (Bruker Scientific Corporation Ltd., Switzerland) using the residual solvent signal as an internal standard at 400 MHz and 100 MHz respectively. Chemical shifts are reported in parts per million (ppm). The elemental analysis was carried out by using Perkin-Elmer 2400 series-II elemental analyzer (Perkin-Elmer, USA) and all compounds are within $\pm 0.4\%$ of the theoretical compositions. Ampicillin, griseofulvin, isoniazid and nystatin were purchased from local market.

1.2. In vitro antimicrobial assay

The *in vitro* antimicrobial activity of all synthesized pyrazole based pyrido [2,3- *d*]pyrimidinones derivatives **4a-r** was carried out by broth micro dilution method. Mueller – Hinton broth was used as nutrient medium to grow and dilute the compound suspension for the test bacteria. Sabouraud Dextrose broth was used for fungal nutrition. 2% DMSO in water was used as the diluent to get desired concentration of compounds to test upon standard bacterial strains. Inoculum size for test strain was adjusted to 10^8 CFU mL⁻¹ by comparing the turbidity. Serial dilutions were prepared in primary and secondary screening. Each synthesized compound and the standard drugs were diluted obtaining 2000 µg/mL concentration as a stock solution. The drugs which were found to be active in primary screening (i.e. 500, 250 and 200 µg/mL concentrations) were further screened in their second set of dilution at 100, 50, 25 and 12.5 µg/mL concentration against all microorganisms. 10 micro liter suspensions were further inoculated on appropriate media and growth was noted after 1 and 2 days. The control tube containing no antibiotic was instantaneously sub cultured (before inoculation) by spreading a loopful evenly over an area of plate of medium suitable for the growth of the test organism. The tubes were then put for incubation at 37°C overnight. The highest dilution preventing appearance

of turbidity after spot subculture was considered as minimal inhibitory concentration (MIC, μ /L). All the tubes showing no visible growth (same as control tube) were subcultured and incubated overnight at 37 °C. The amount of growth from the control tube before incubation was compared. In this study Ampicillin, Norfloxacin and Chloramphenicol were used as the standard antibacterial drugs. Nystatin and Griseofulvin were used as standard antifungal drugs. The results are summarized in **Table 4**.

1.3. In vitro antituberculosis assay

The antitubercular activity of all synthesized compound against *Mycobacterium tuberculosis* H37Rv was performed by Lowensteine-Jensen method [1] with minor modification where 250 μ g/mL and 100 μ g/mL dilution of each compound was added to Lowensteine-Jensen medium and then media was uncontaminated by inspissation method. A culture of *Mycobacterium tuberculosis* H37Rv growing on Lowensteine-Jensen medium was harvested in 0.85% saline in bijou bottle. The stock solutions of the all compounds were prepared in 2% DMSO IN WATER i.e. 250 μ g/mL and 100 μ g/mL. These tubes were then incubated at 37 °C for 24 h followed by streaking of *Mycobacterium tuberculosis* H37Rv (5 _ 104 bacilli per tube). The growth of bacilli was seen after two weeks, three weeks and finally after four weeks of incubation. The tubes having the compounds were compared with control tubes where medium alone was incubated with *Mycobacterium tuberculosis* H37Rv. The concentration at which complete inhibition of colonies occurred was taken as active concentration of the tested compound. The standard strain *Mycobacterium tuberculosis* H37Rv was tested with known drug isoniazid and rifampicin for comparison purpose. The results are summarized in **Table 5**.

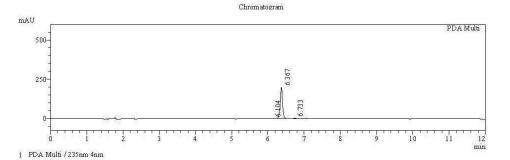
1.4. In vitro antimalarial assay

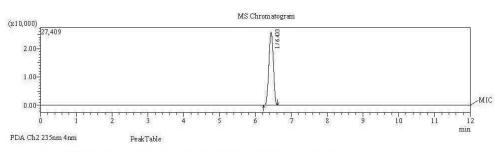
All the synthesized compounds **4a-r** were screened for their antimalarial activity against the *P. falciparum* strain. The *P. falciparum* strain was acquired from Shree R. B Shah Mahavir Superspeciality hospital, Surat, Gujarat, India, and was used in the vitro tests. The *P. falciparum* strains were cultivated by a modified method described by Trager and Jensen [2]. Compounds were dissolved in 2% DMSO in water. The final concentration of DMSO used was not toxic and did not interfere with the assay. The antiparasitic effect of the compounds was measured by growth inhibition percentage as described by Carvalho and Krettli [3]. For experimental purposes, the cultures were synchronized with 5% D-sorbitol when the parasites were in the ring stage [4]. The parasite suspension, consisting of predominately the ring stage, was adjusted to a 1-2 % parasitaemia and 2.5 % haematocrit in hypoxanthine-free RPMI-1640 culture medium with 10% human plasma and was exposed to 7 concentrations of each compound for a single cycle of parasite growth of 48 h at 37 °C. A positive control with reference to antimalarial drugs

in standard concentrations was used in each experiment. The stock solutions were additionally diluted in whole medium (RPMI 1640 plus 10% human serum) to each of the used concentrations. The concentration that inhibited 50% of parasite growth (IC₅₀ value) was determined by interpolation using Microcal Origin software. The standard drugs chloroquine and quinine were used as the reference antimalarial agents, blood smears were read blind and each duplicate experiment was repeated three times. The results are summarized in **Table 6**.

LC-MS of compound 4a





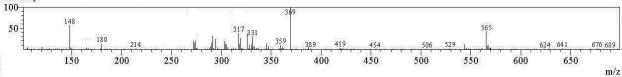


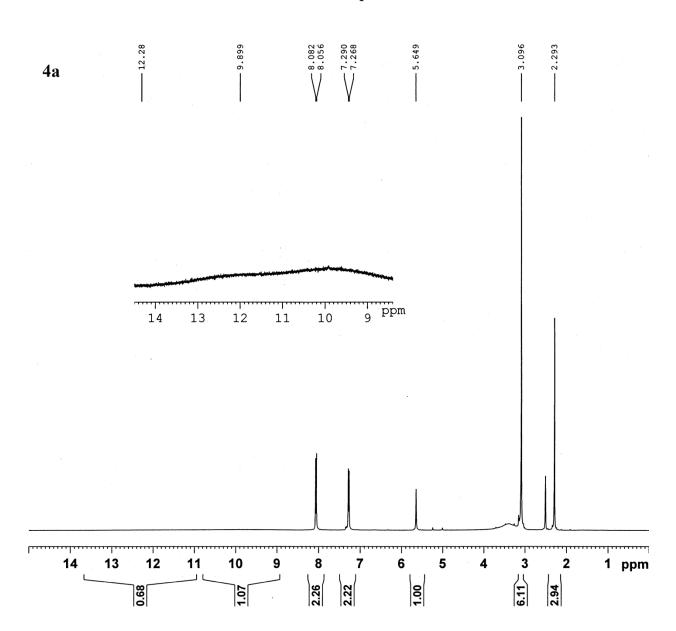
Peak#	Ret. Time	Area	Height	Area %
1	6.104	3962	826	0.442
2	6.367	879809	199559	98.143
3	6.733	12681	2990	1.415
Total		896452	203375	100.000

		MSTAB	LE MIC1		
Peak#	Ret.Time	Base Peak m/z	Area	Area%	Event#
1	6.433	369.20	224750	100.00	1-1
Total	n tha the she was a set		224750	100.00	S GUNANDSIADSIA

MS Spectrum Graph

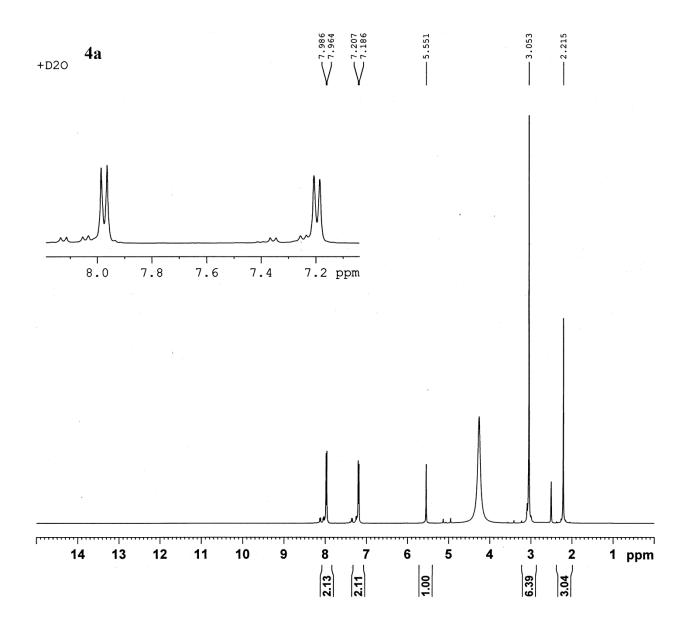
#:1 Ret.Time:Averaged 6.417-6.450(Scan#:386-388) BG Mode:Calc 6.233<->6.617(375<->398) Base Peak:369.20(41512) Polarity:Pos Segment1 - Event1 Intensity





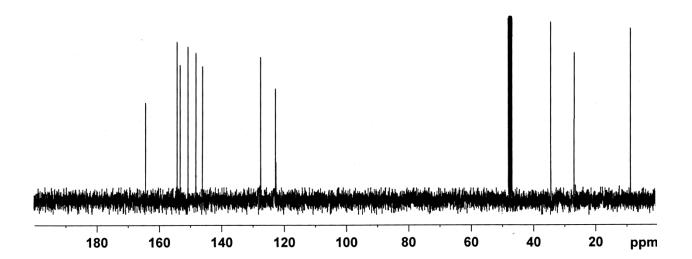
¹HNMR of compound **4a**

¹HNMR of compound **4a** (DMSO+D₂O)



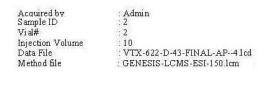
¹³CNMR of compound **4a**

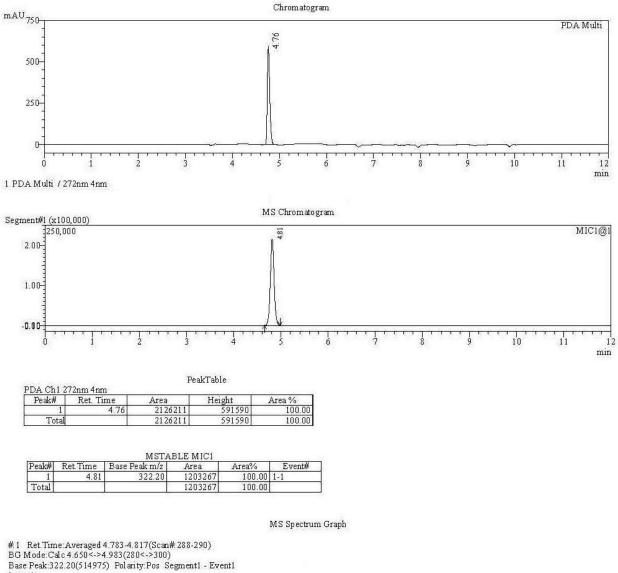


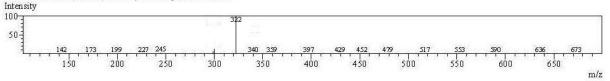


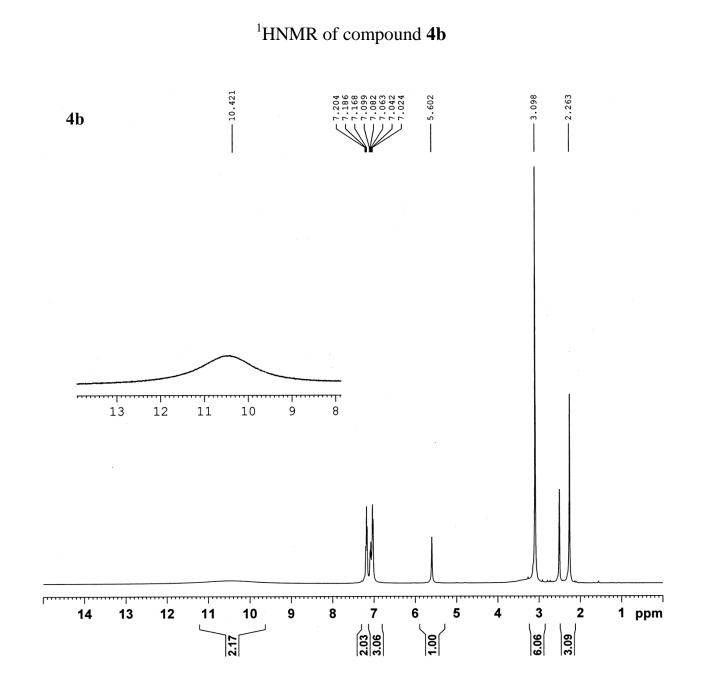
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LC-MS of compound 4b

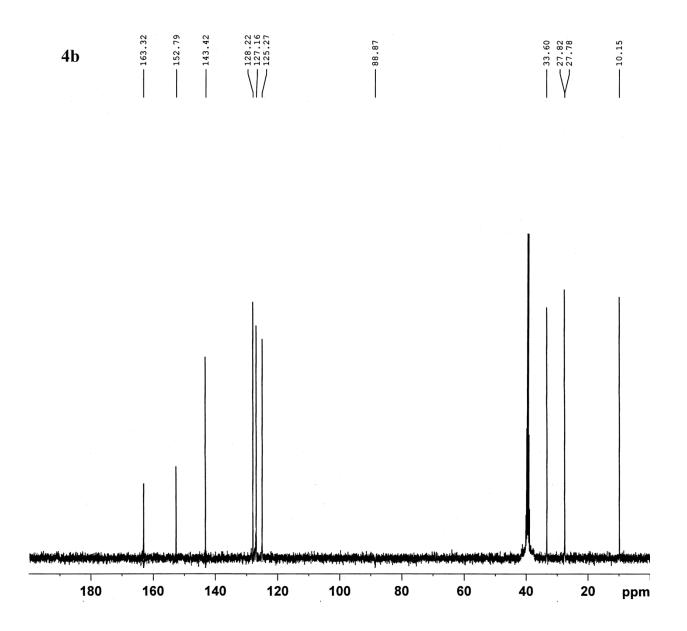




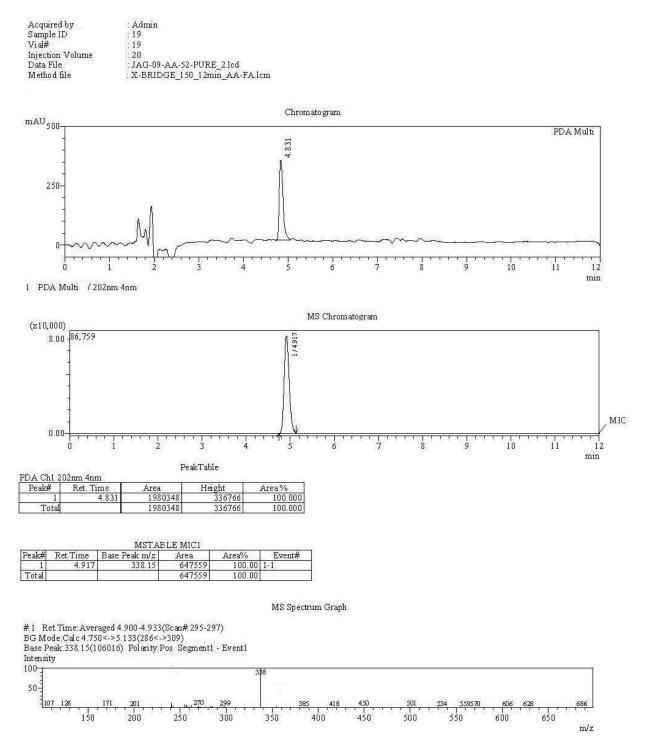


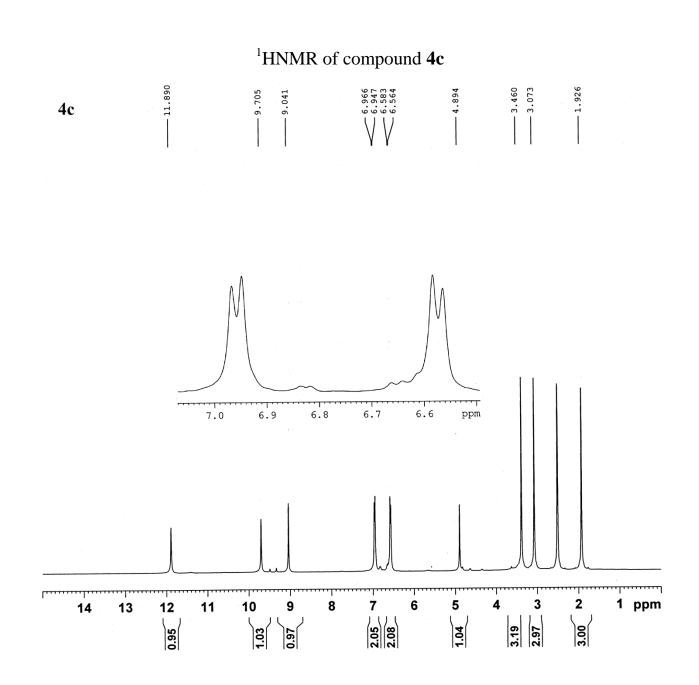


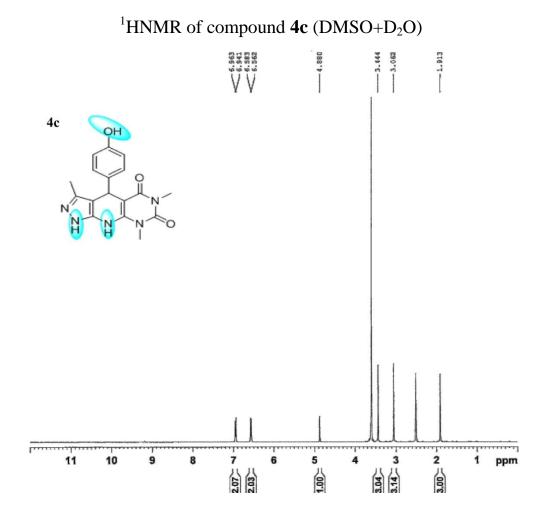
¹³CNMR of compound **4b**

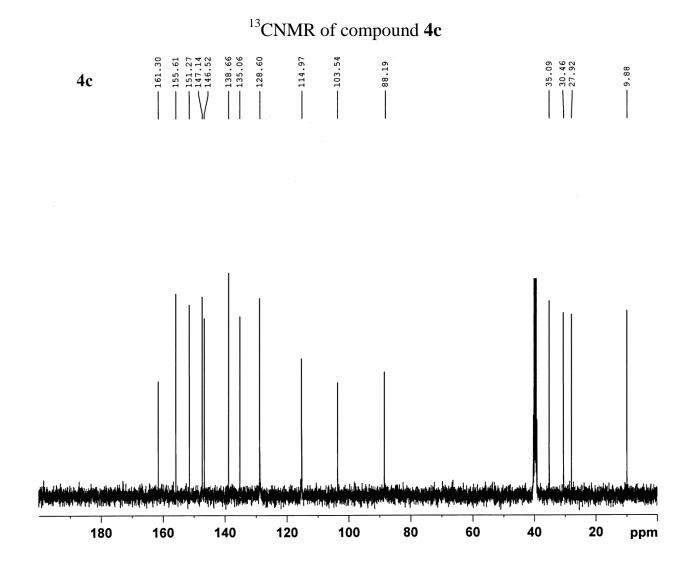


LC-MS of compound 4c

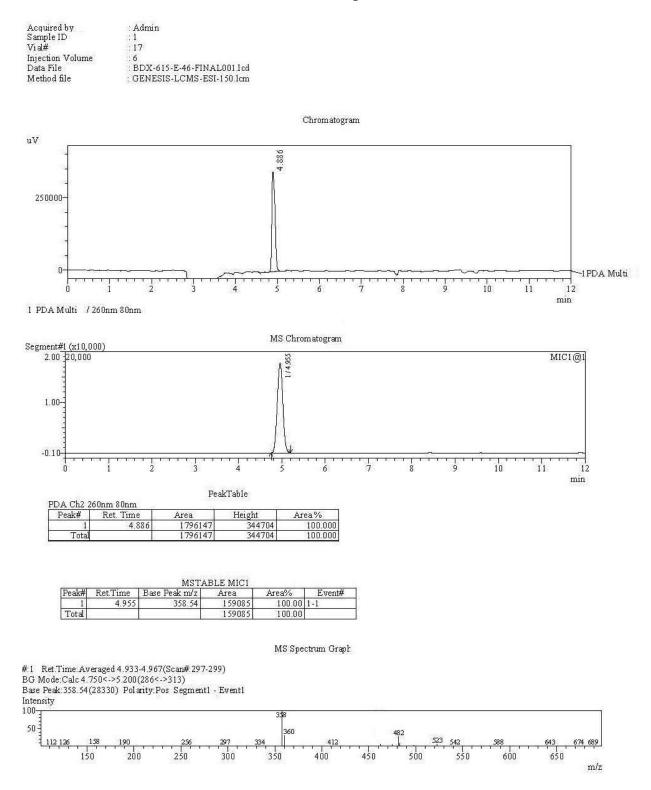


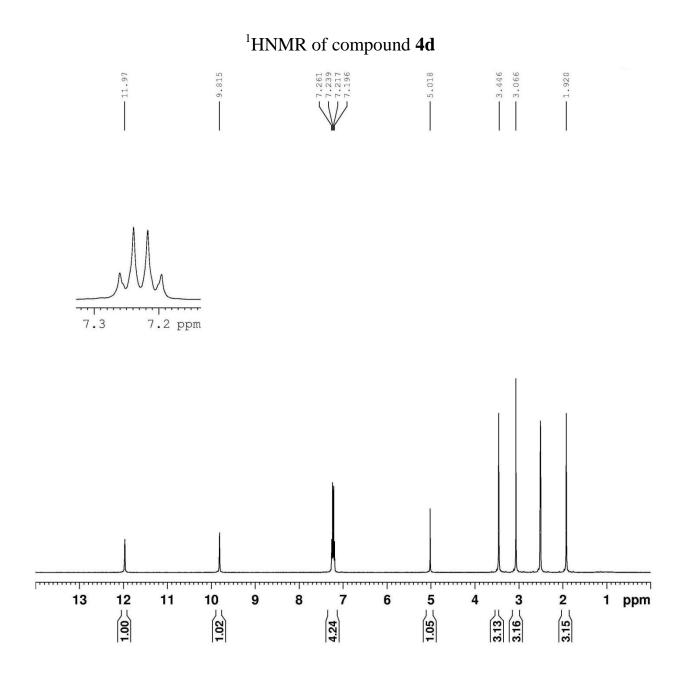




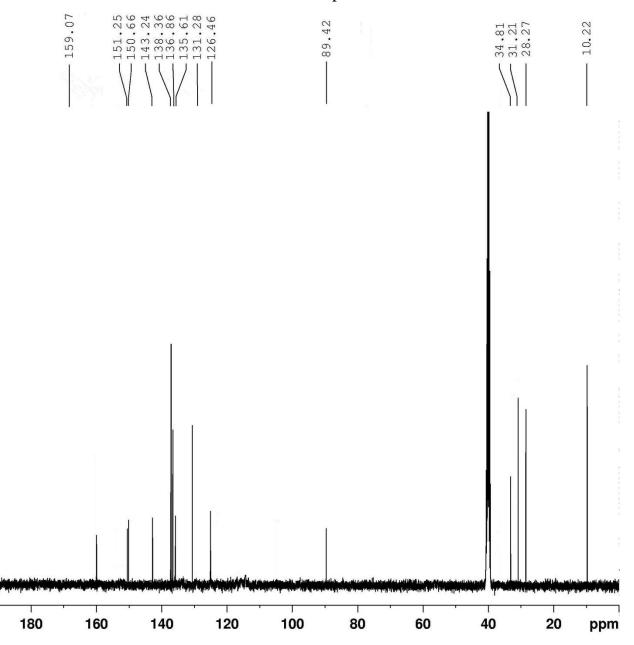


LC-MS of compound 4d

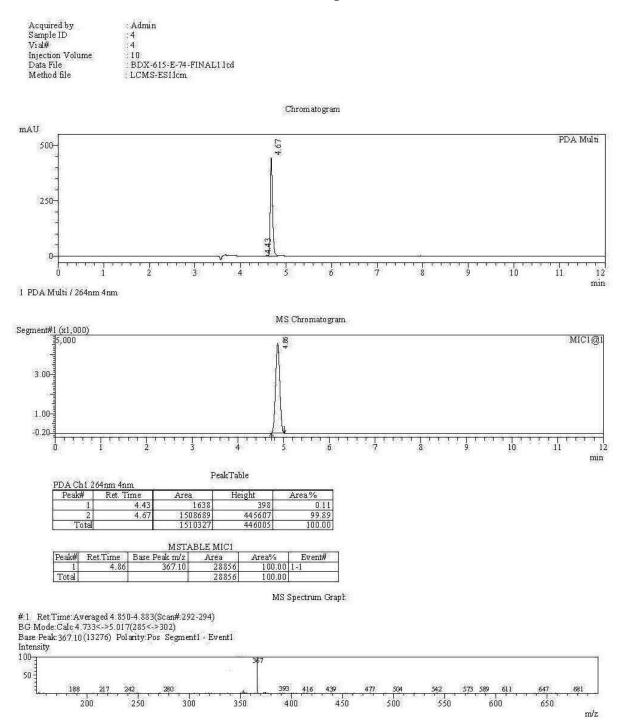


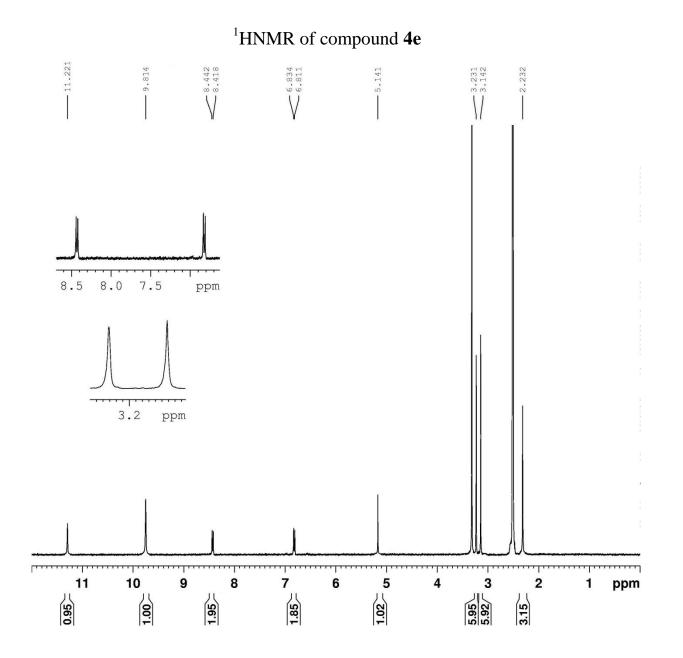


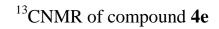
¹³CNMR of compound **4d**

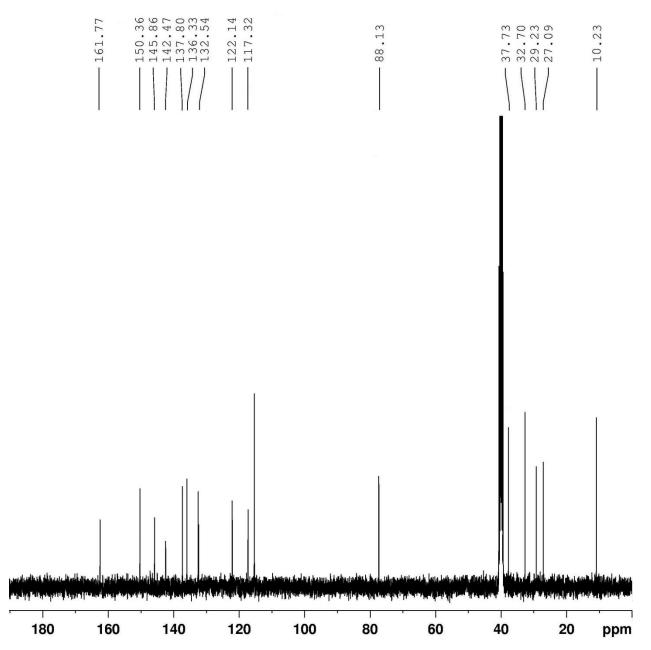


LC-MS of compound 4e

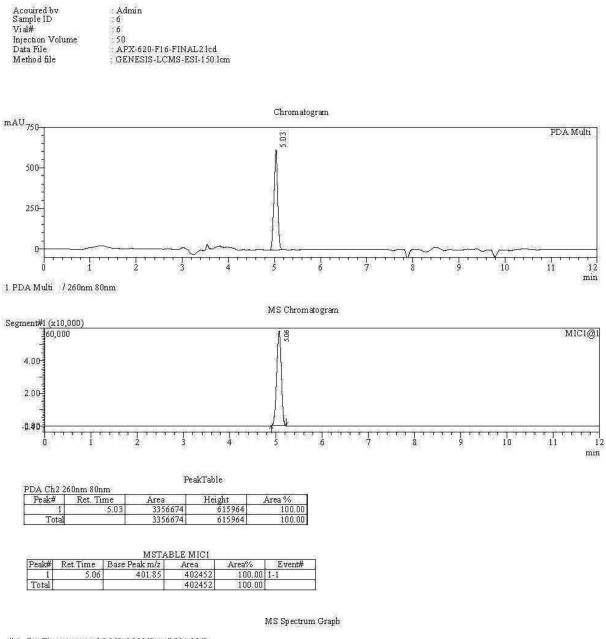




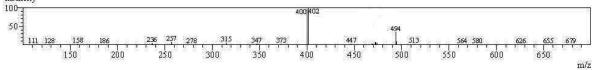


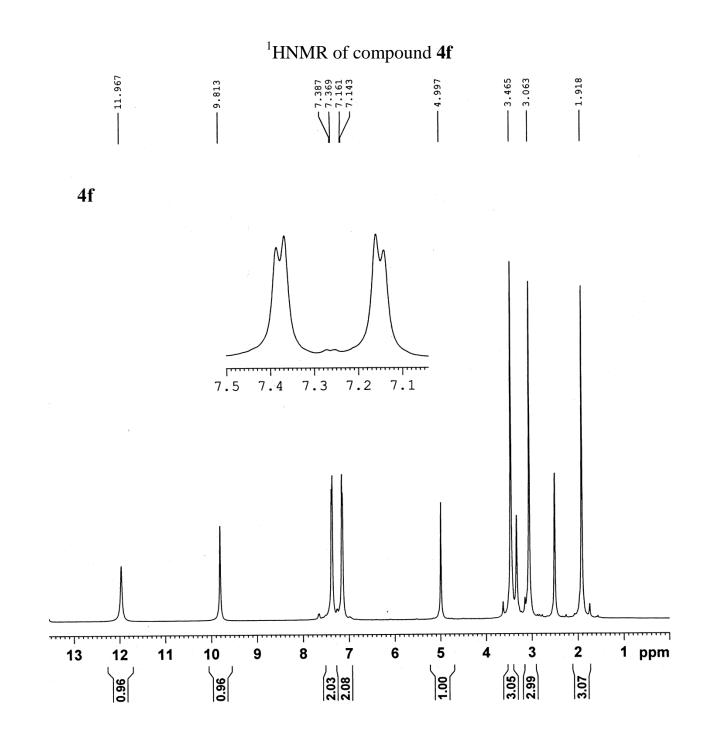


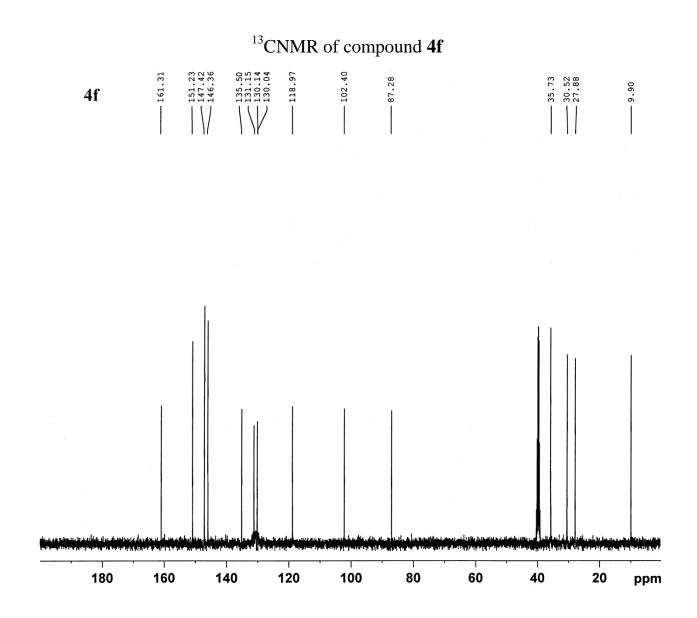
LC-MS of compound 4f



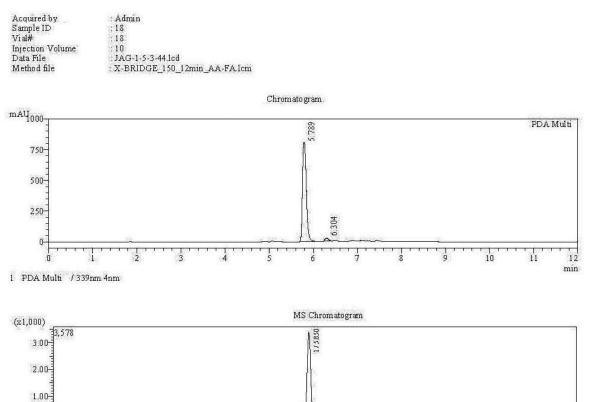
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LC-MS of compound 4g



MIC

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			0000-000000000		
117163127	Arrest States	MSTAB	LE MICI		- 1450 - 141
Peak#	Ret.Time	MSTAB Base Peak m/z	LE MIC1 Area	Area%	Event#

2

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MS Spectrum Graph

5

2.566

Area% 97.434

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100.00

3

PeakTable

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22816

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6

N

#1 Ret Time: Averaged 5.833-5.867(Scan#351-353) BG Mode: Calc 5.717<->6.000(344<->361) Base Peak 369.15(27763) Polarity:Pos Segment1 - Event1 Intensity

0.00-

PDA Chi 339nm 4nm Peak# Ret. Tim

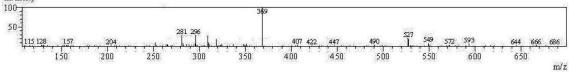
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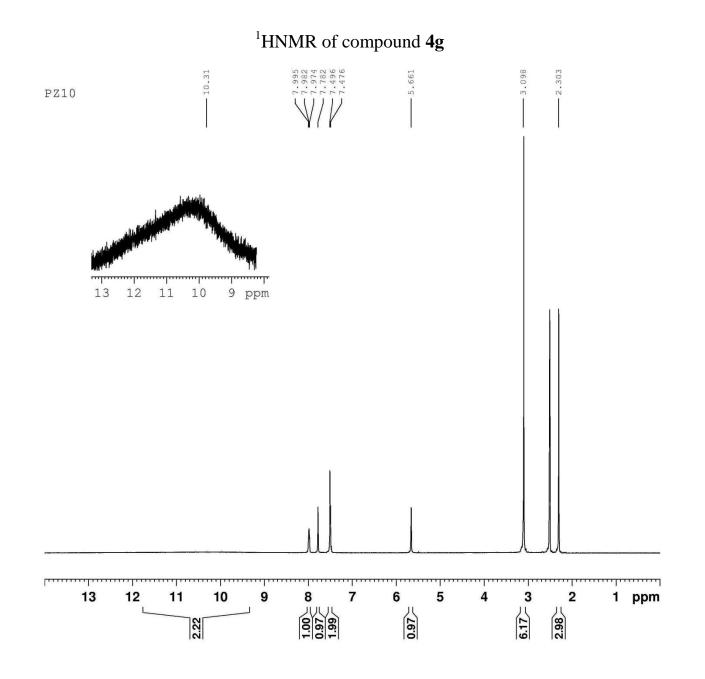
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Ret. Time

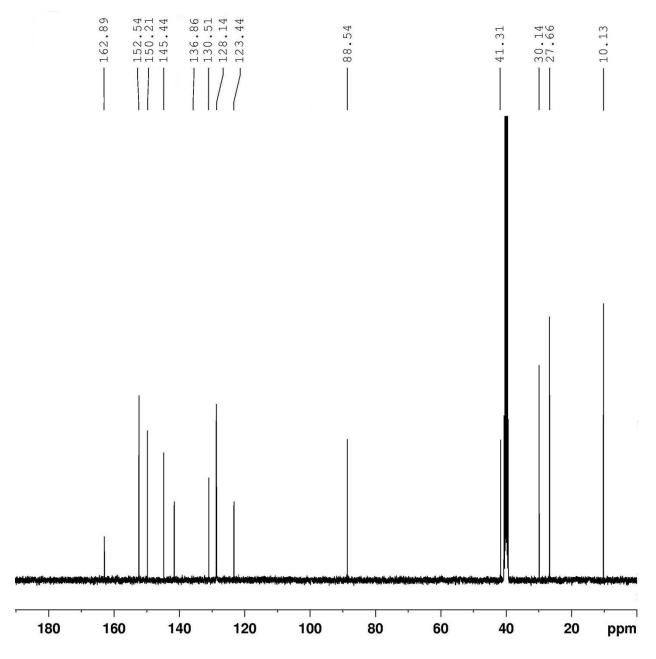
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6.304

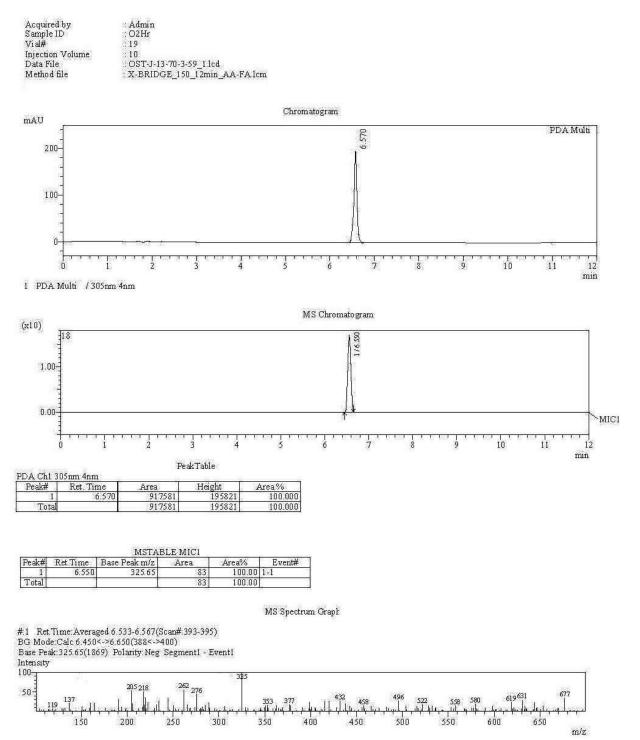


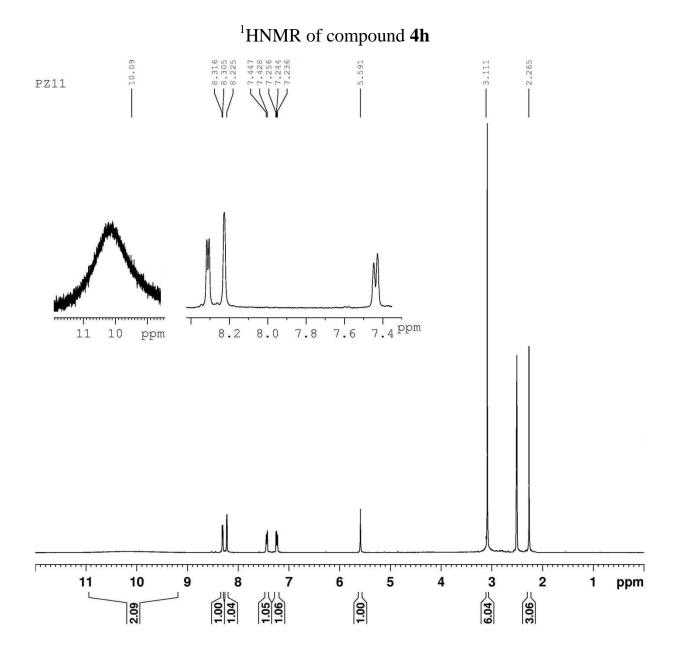


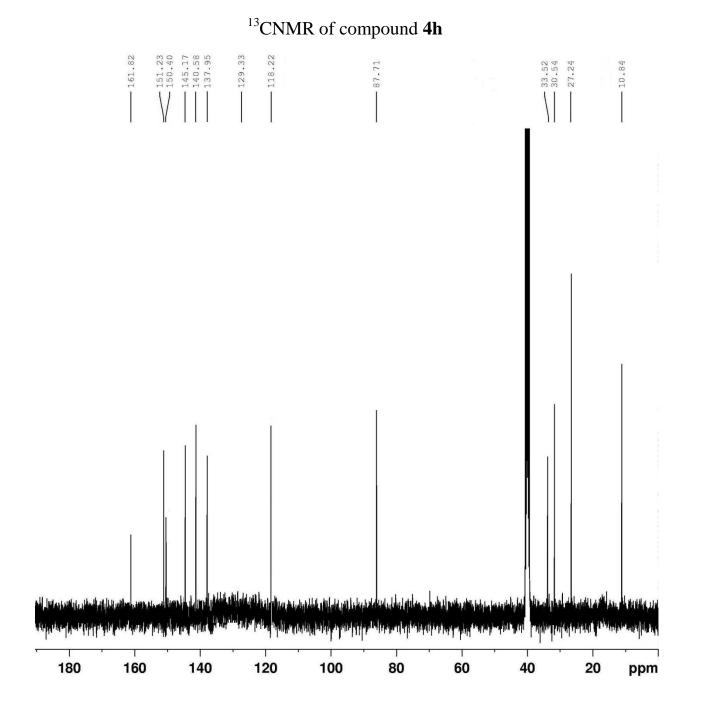
¹³CNMR of compound **4g**



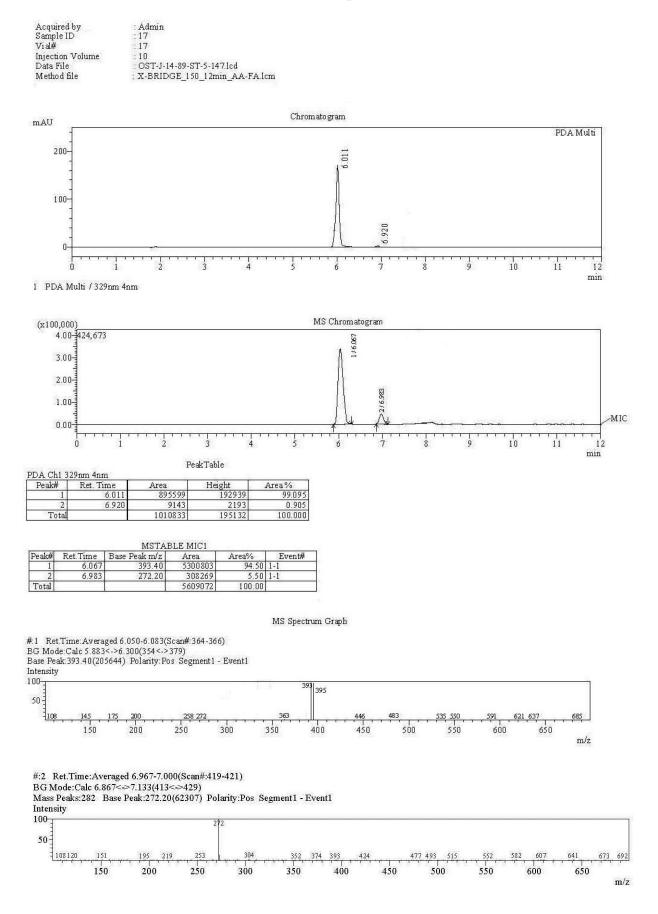
LC-MS of compound 4h







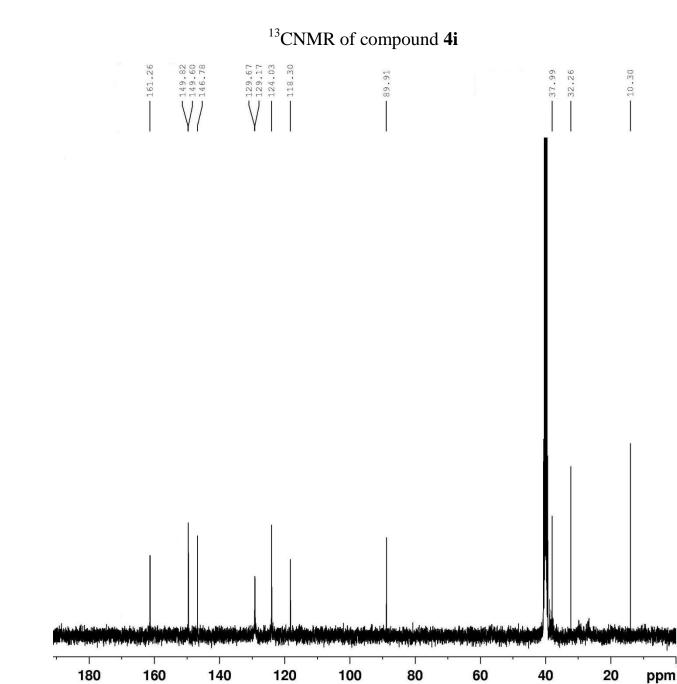
LC-MS of compound 4i



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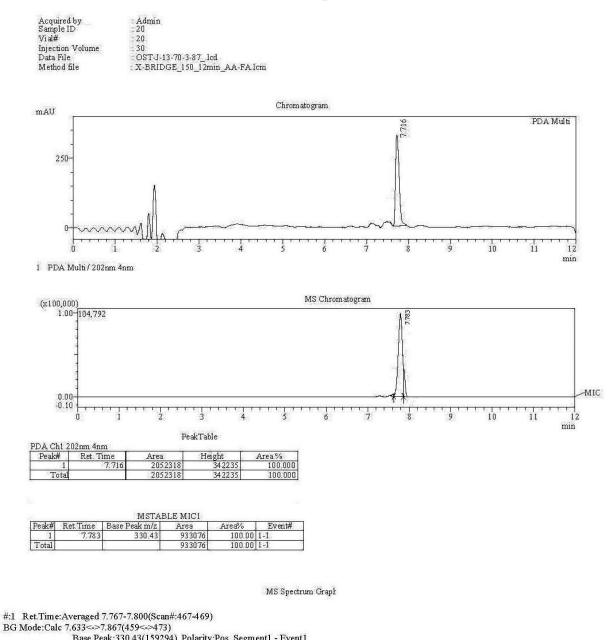
-11.524 5,420 -7.261-7.245- 3.451 ► 7.444 - 9.876 -1.907di.an.an.da.i.a.i.d 7.4 7.3 ppm πŋ 1 1 Т Т 12八001 3.33 3.20 × 6 3.35 10 7 11 9 8 6 5 4 ٦ ppm 1.02 1.04 1:03

¹HNMR of compound **4i**

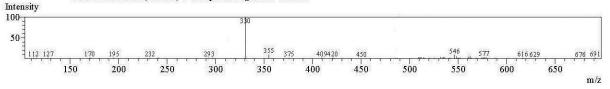


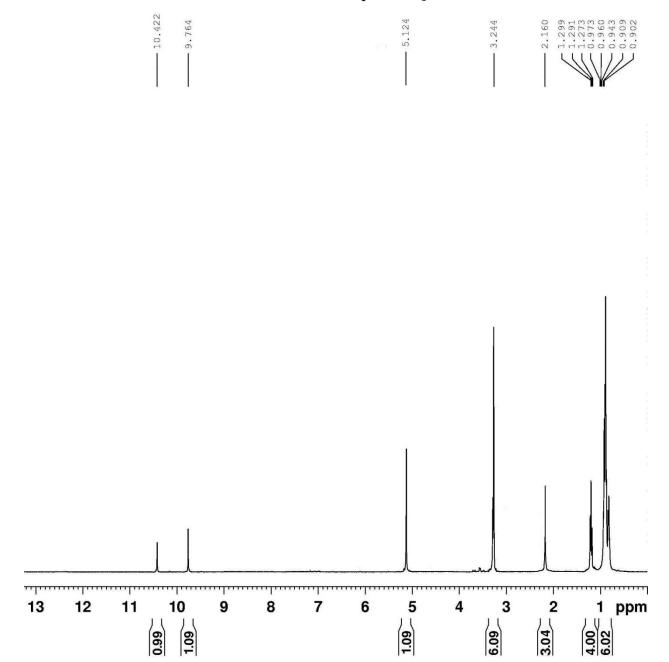
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LC-MS of compound 4j

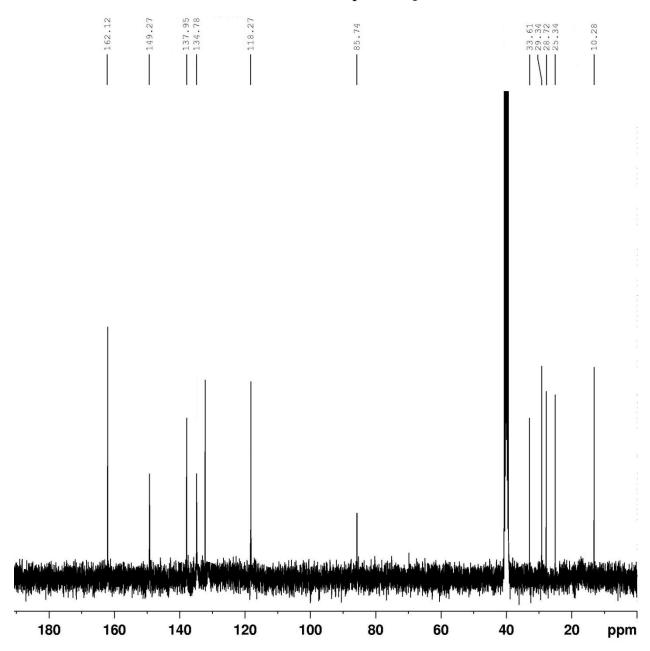


Base Peak: 330.43(159294) Polarity:Pos Segment1 - Event1



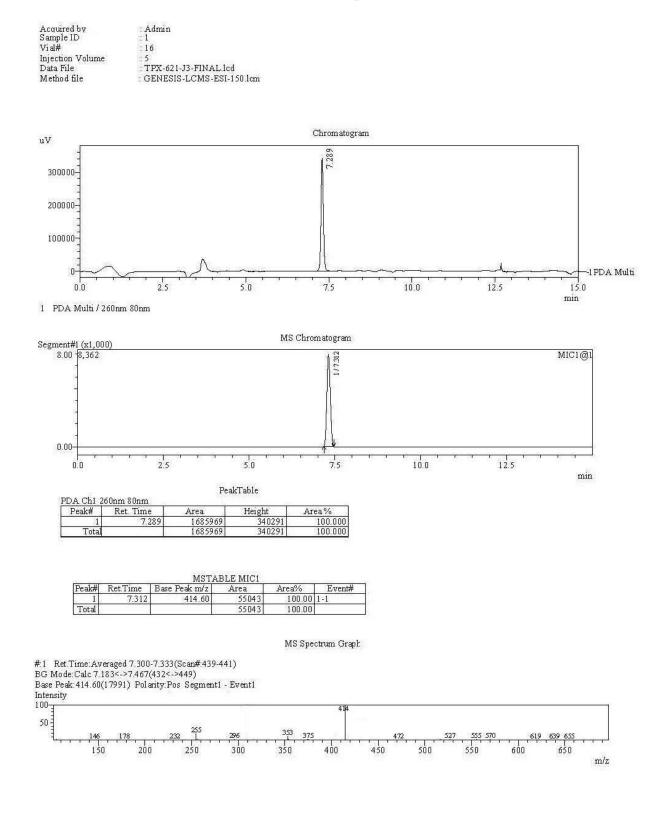


¹HNMR of compound **4**j

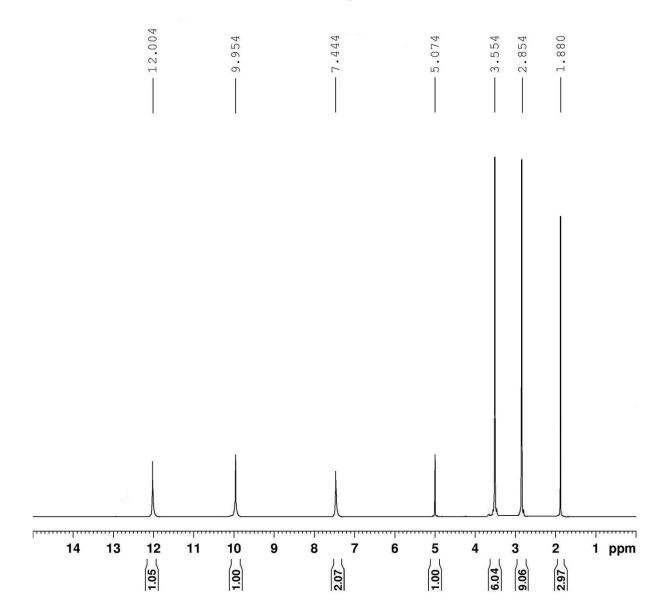


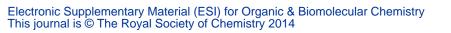
¹³CNMR of compound **4j**

LC-MS of compound 4k

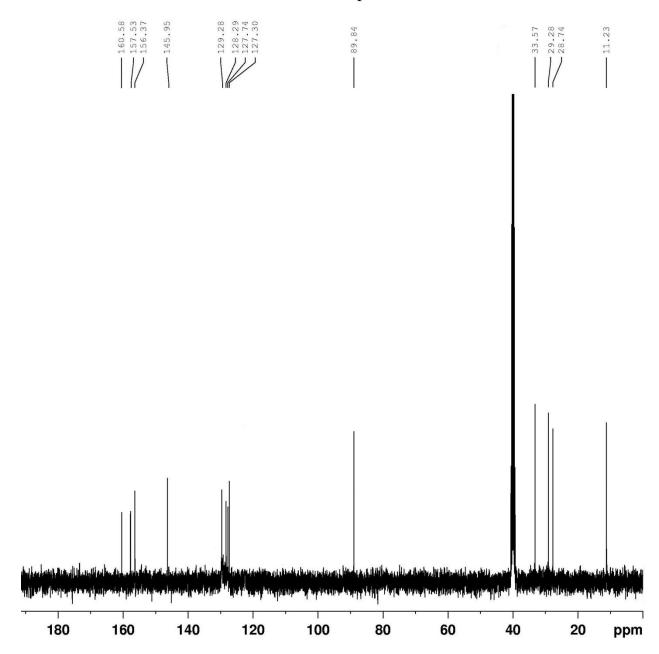


¹HNMR of compound **4**k





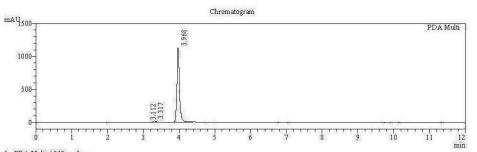
¹³CNMR of compound **4**k



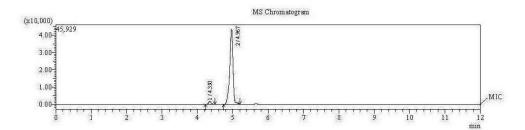
LC-MS of compound 41



: Admin :21 :21 :10 :10G-13-1-4-US.lcd : X-BRIDGE_150_12min_AA-FA.lcm



1 PDA Multi / 240nm 4nm



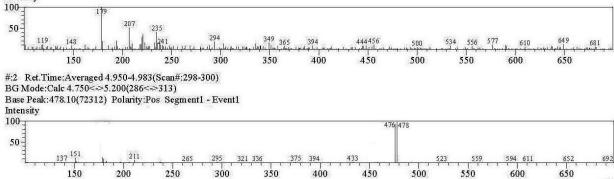
PDA Ch2 240nm 4nm

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2	3.317	91974	19111	1.560
3	3.968	5786190	1185506	98.130
Total		5896455	1208561	100.000

MSTABLE MIC1					
Peak#	Ret.Time	Base Peak m/z	Area	Area%	Event#
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Total			285831	100.00	

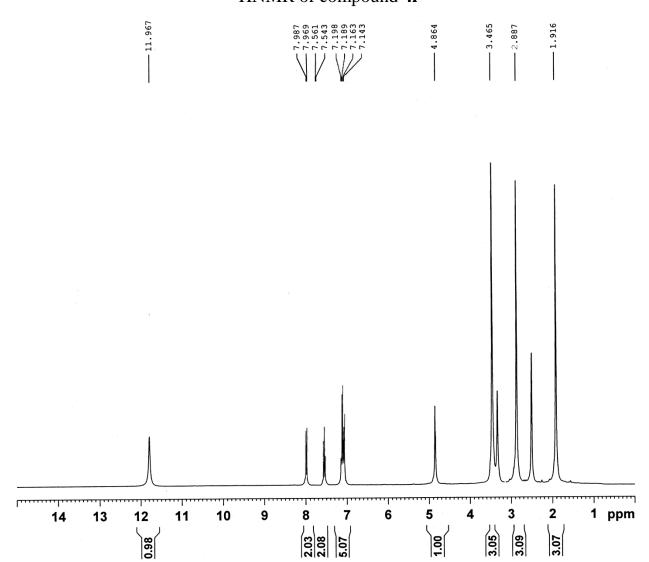
MS Spectrum Graph

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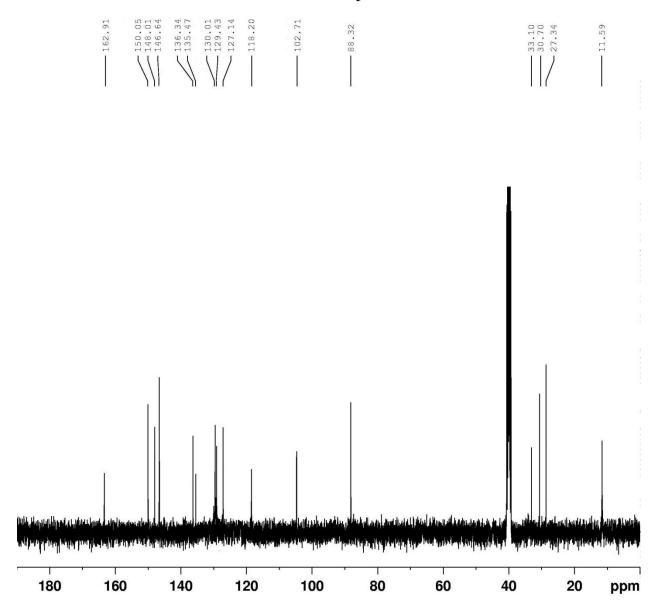
m/z

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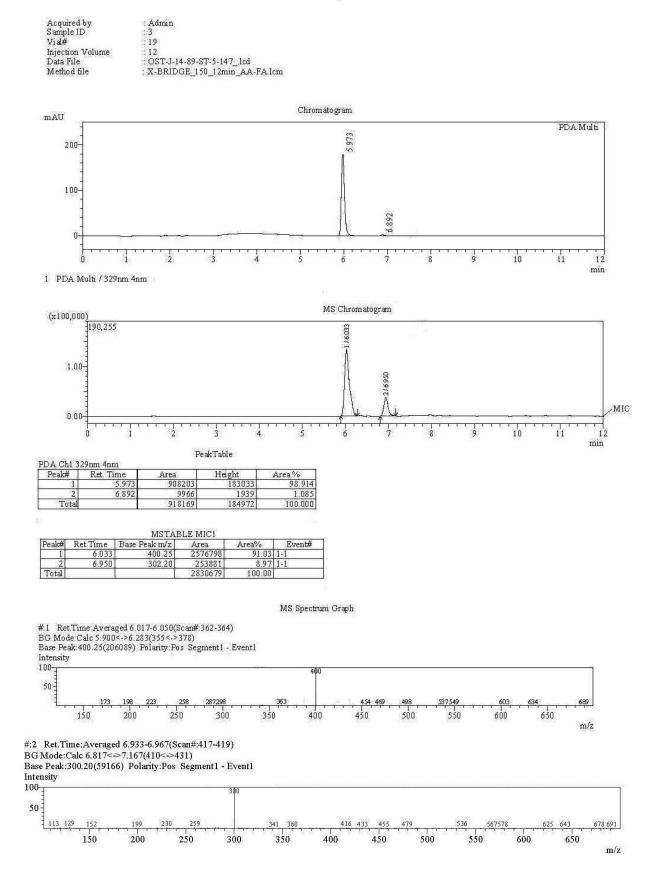


¹HNMR of compound **4**

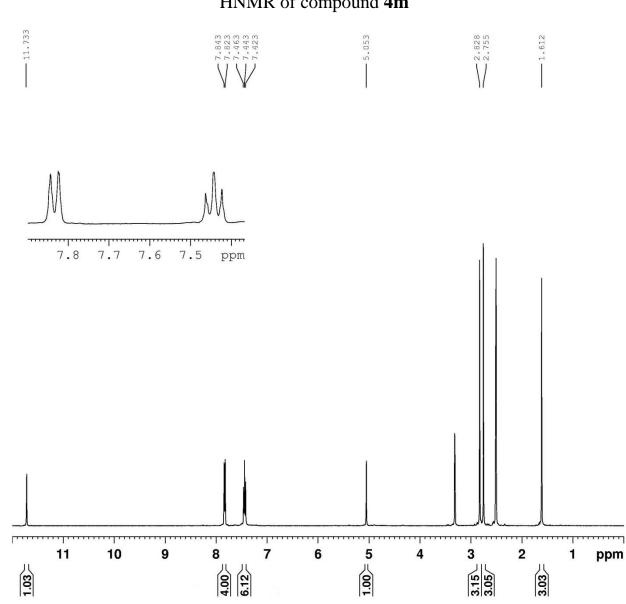
¹³CNMR of compound **4**l



LC-MS of compound 4m

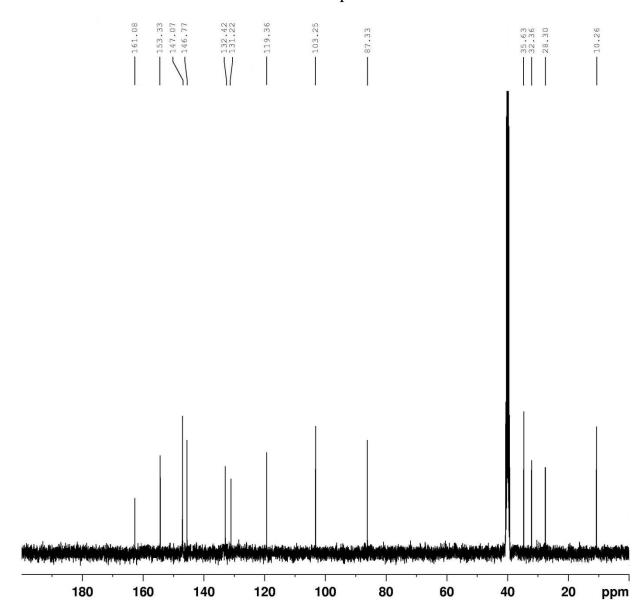


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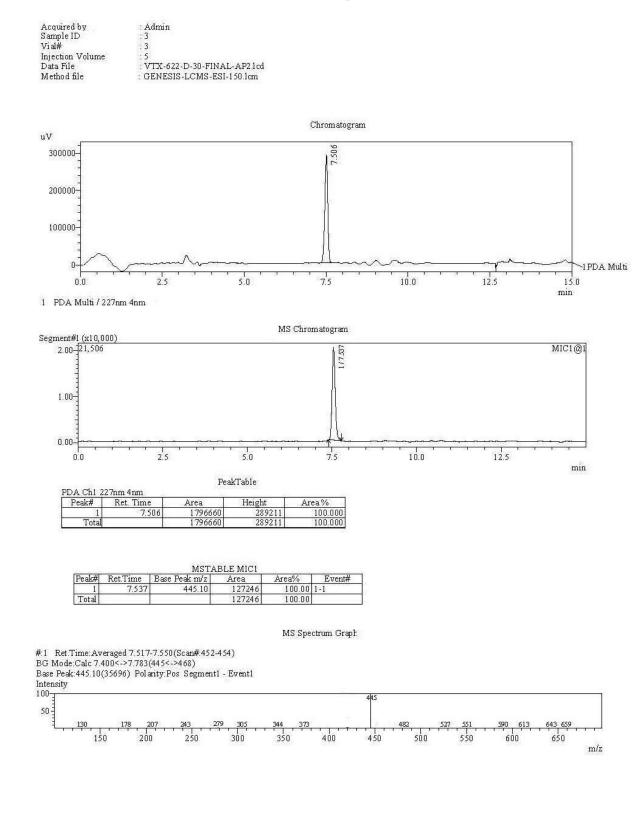
¹HNMR of compound **4m**

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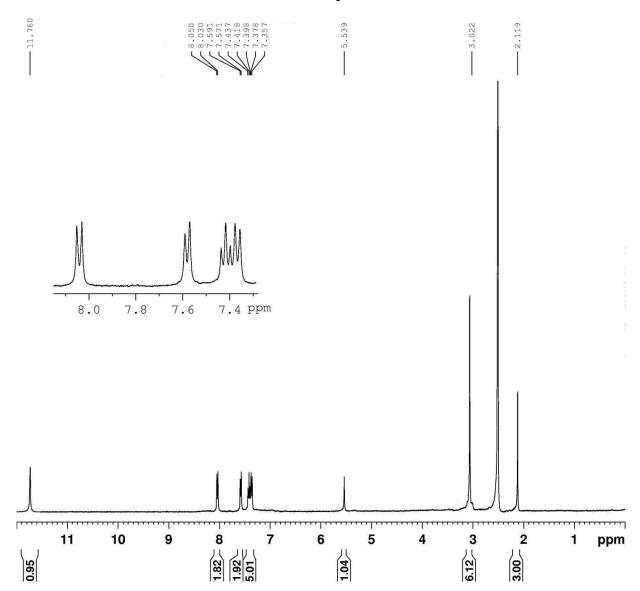
¹³CNMR of compound **4m**

LC-MS of compound 4n

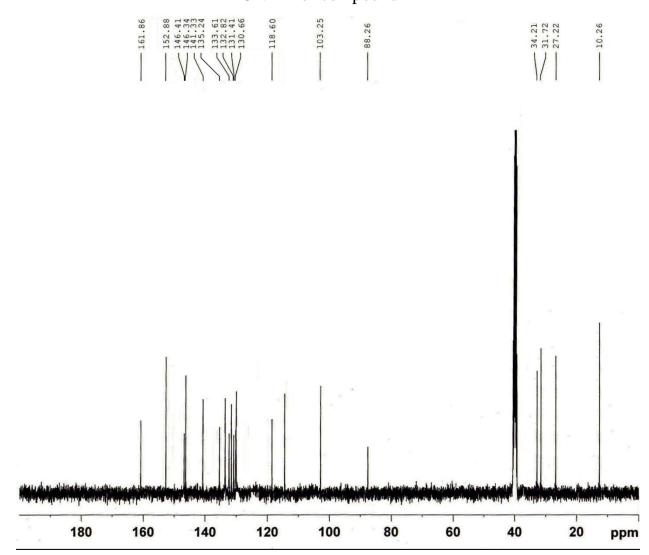


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¹HNMR of compound **4n**

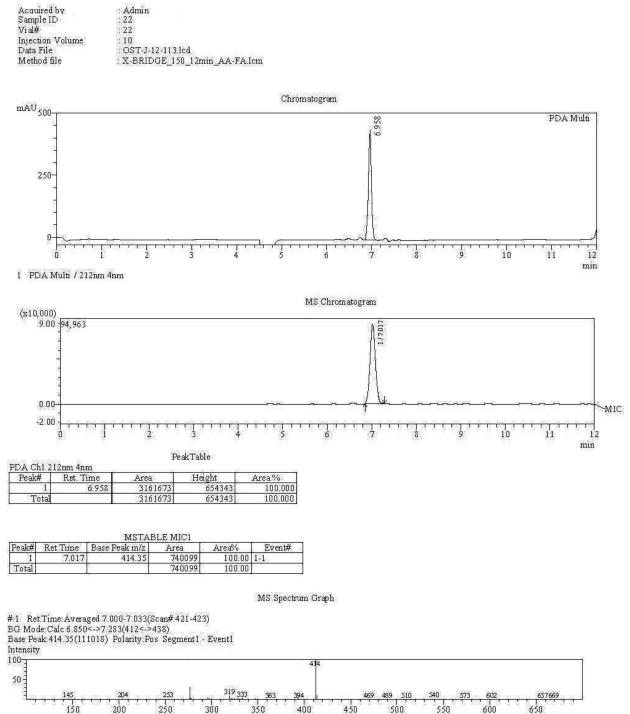


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¹³CNMR of compound **4n**

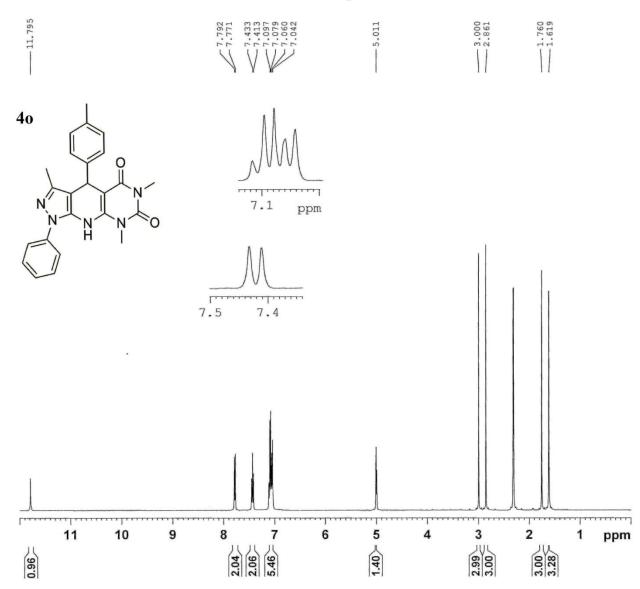
LC-MS of compound 40



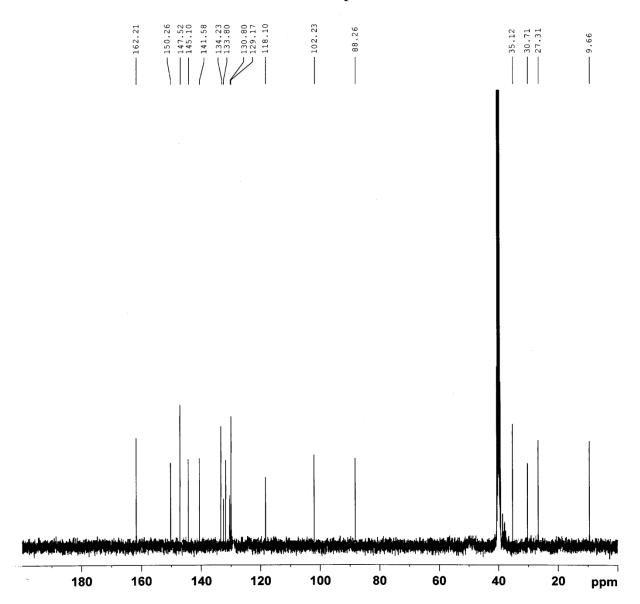
m/z

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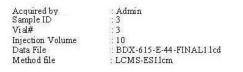
¹HNMR of compound **40**

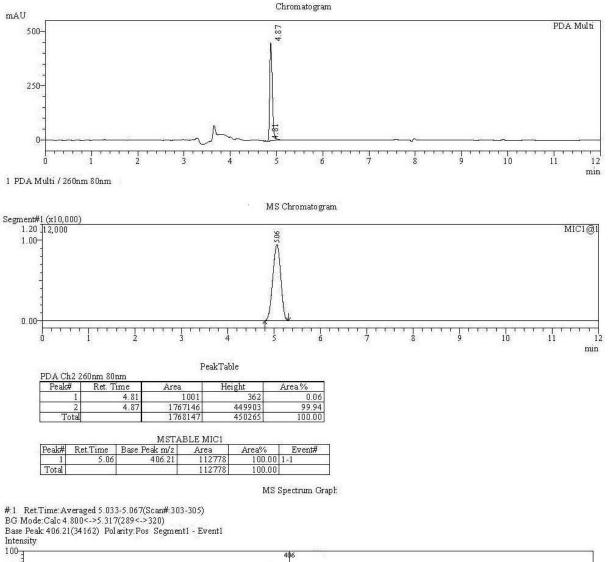


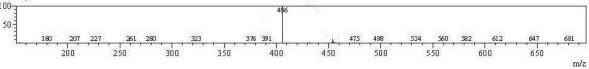
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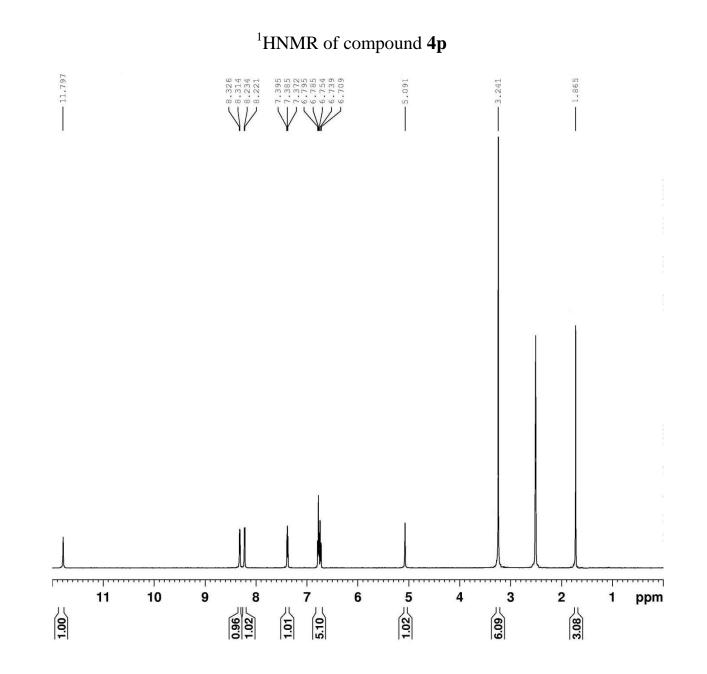


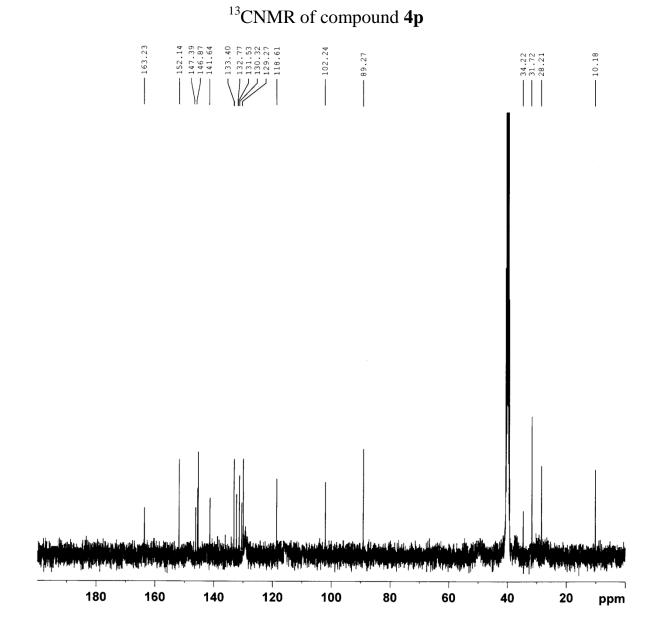
LC-MS of compound 4p



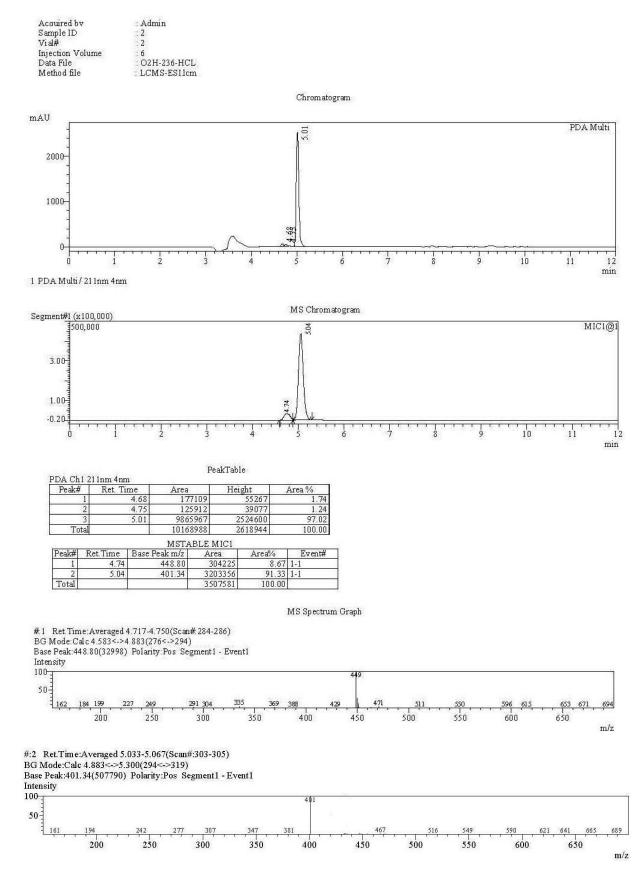


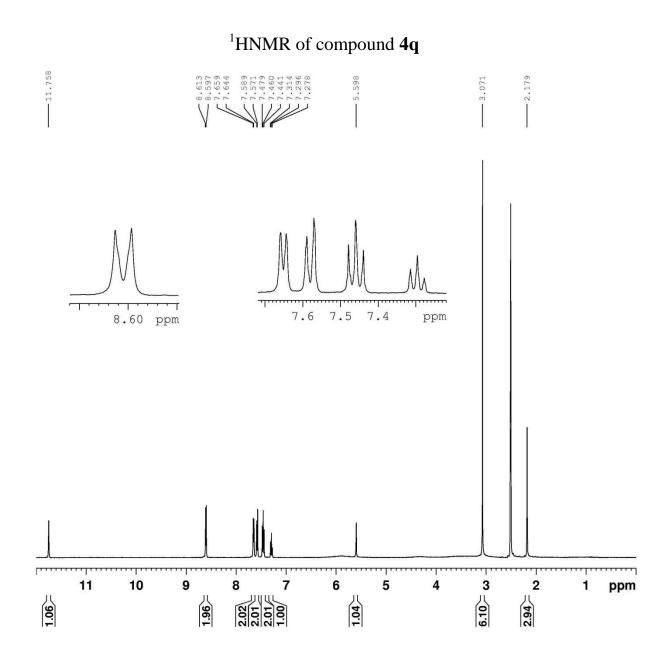




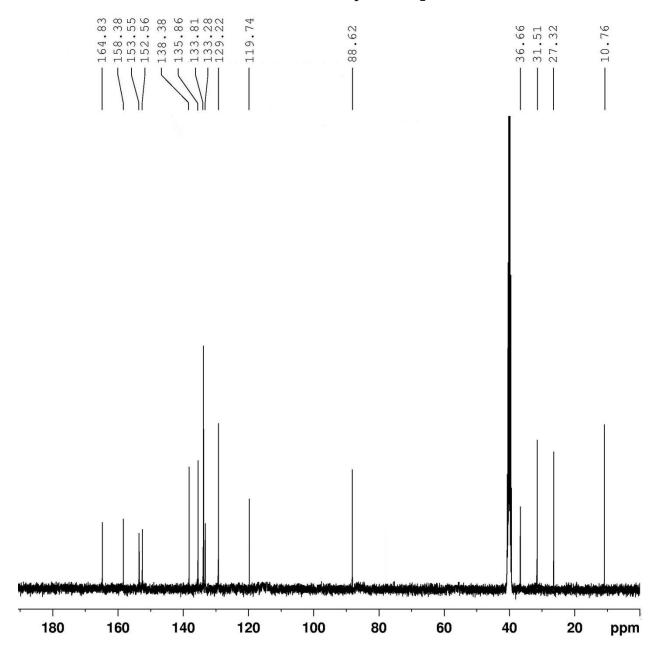


LC-MS of compound 4q





¹³CNMR of compound **4**q



LC-MS of compound 4r

 Acquired by
 : Admin

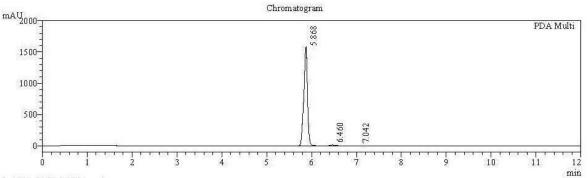
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 : O2Hr

 Vial#
 : 25

 Injection Volume
 : 10

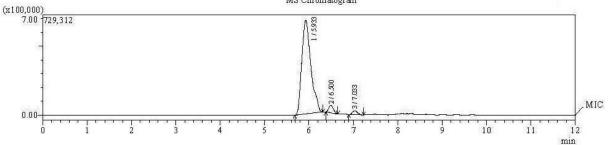
 Data File
 : OST-J-11-84-01-127 .lcd

 Method file
 : X-BRIDGE_150_12min_AA-FA.lcm



1 PDA Multi / 283nm 4nm

MS Chromatogram



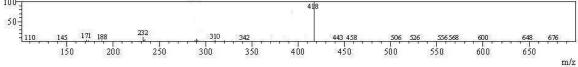
DA Ch1 28	Peaklable 33nm 4nm				
Peak#	Ret. Time	Area	Height	Area%	
1	5.868	8841872	1575526	99.049	
2	6.460	82816	11704	0.928	
3	7.042	20.64	740	0.023	
Total		8926752	1587970	100.000	

- -----

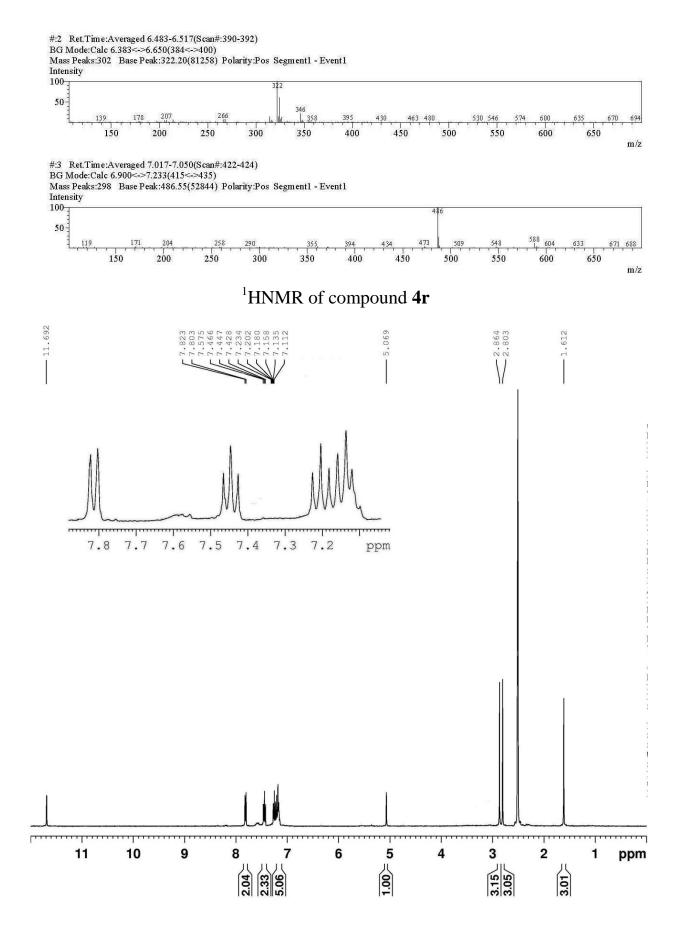
Peak#	Ret.Time	Base Peak m/z	Area	Area%	Event#
1	5.933	418.20	9641423	93.92	1-1
2	6.500	322.20	397415	3,87	1-1
3	7.033	486.55	227350	2.21	1-1
Total			10266188	100.00	<u> </u>

MS Spectrum Graph

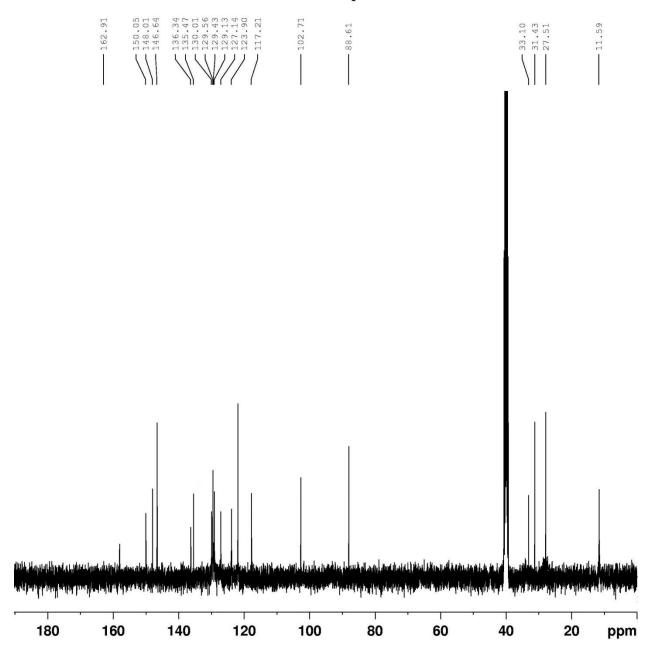
#1 Ret.Time:Averaged 5.917-5.950(Scan#356-358) BG Mode:Calc 5.683<->6.317(342<->380) Base Peak:418.20(751691) Polarity:Pos Segment1 - Event1 Intensity 100



Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry This journal is The Royal Society of Chemistry 2014



¹³CNMR of compound **4r**



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