

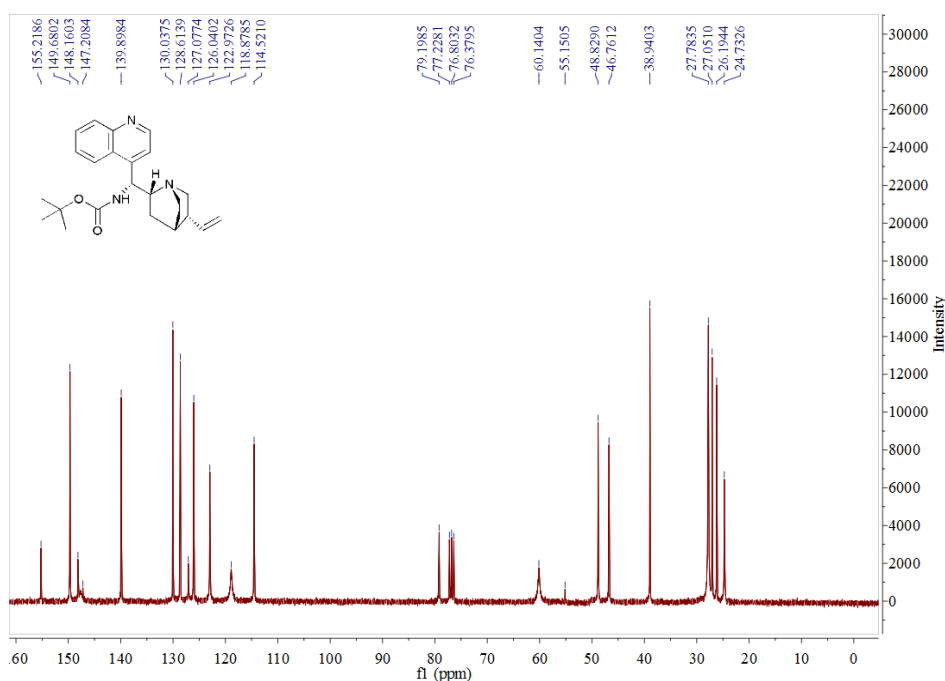
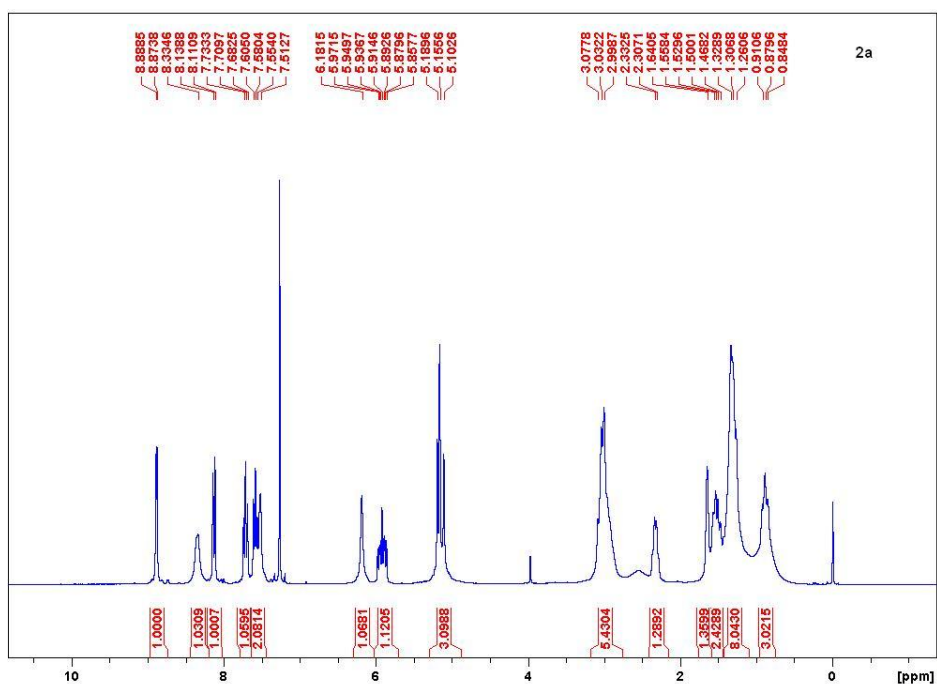
9-amino-(9-deoxy)cinchona alkaloids-derived novel chiral phase-transfer catalysts

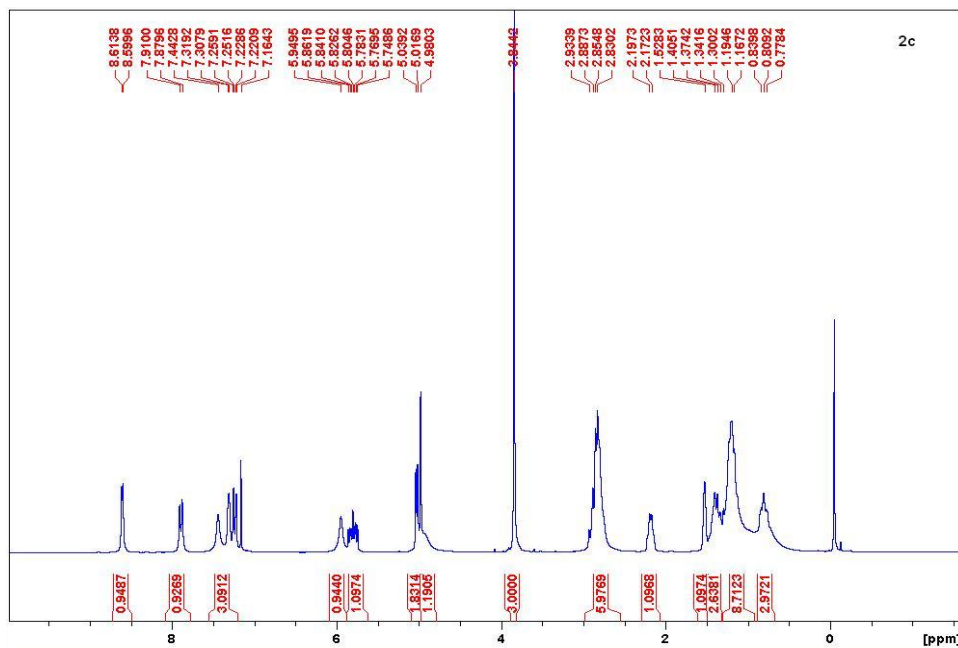
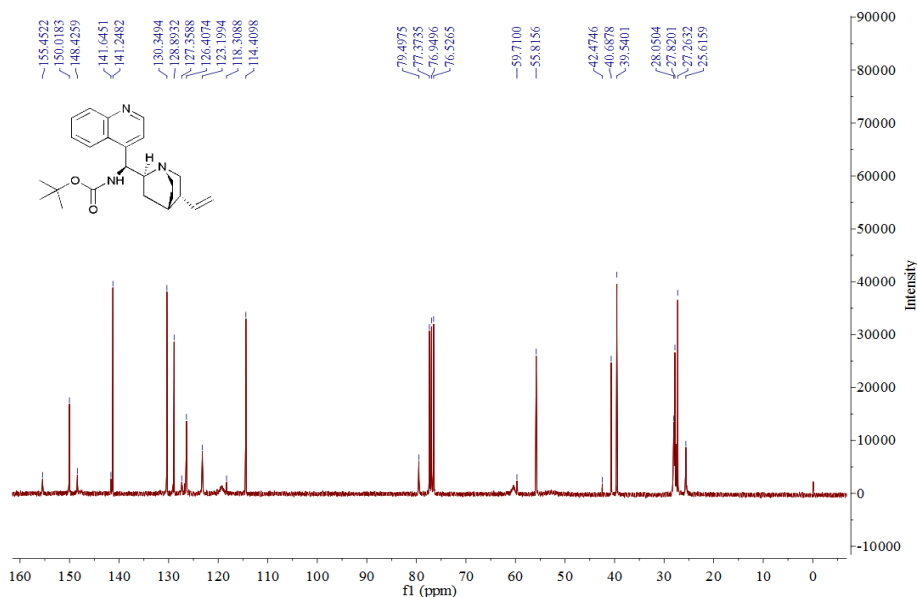
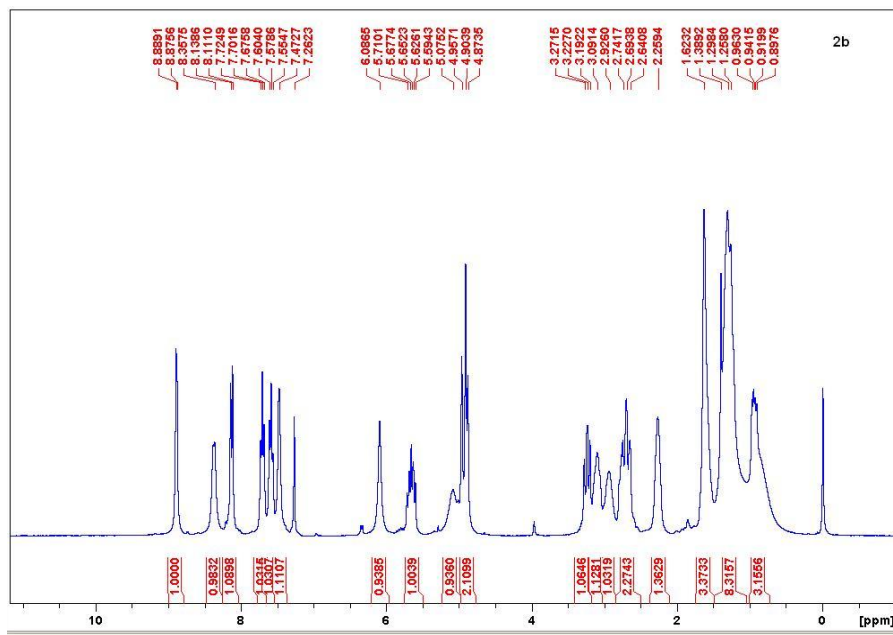
Wenwen Peng,^a Jun Cao,^a Jingwei Wan,^a Xuebing Ma^{*a} and Bing Xie,^{*b}

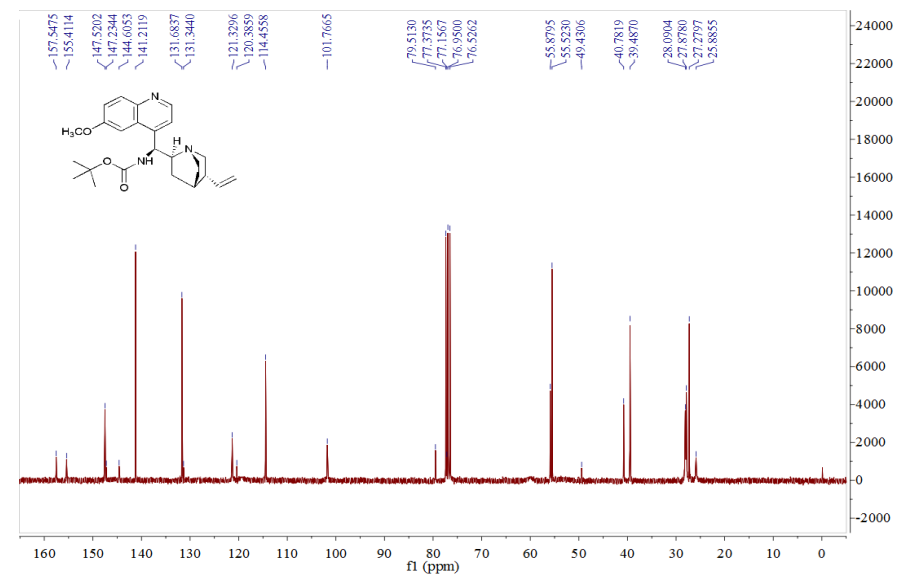
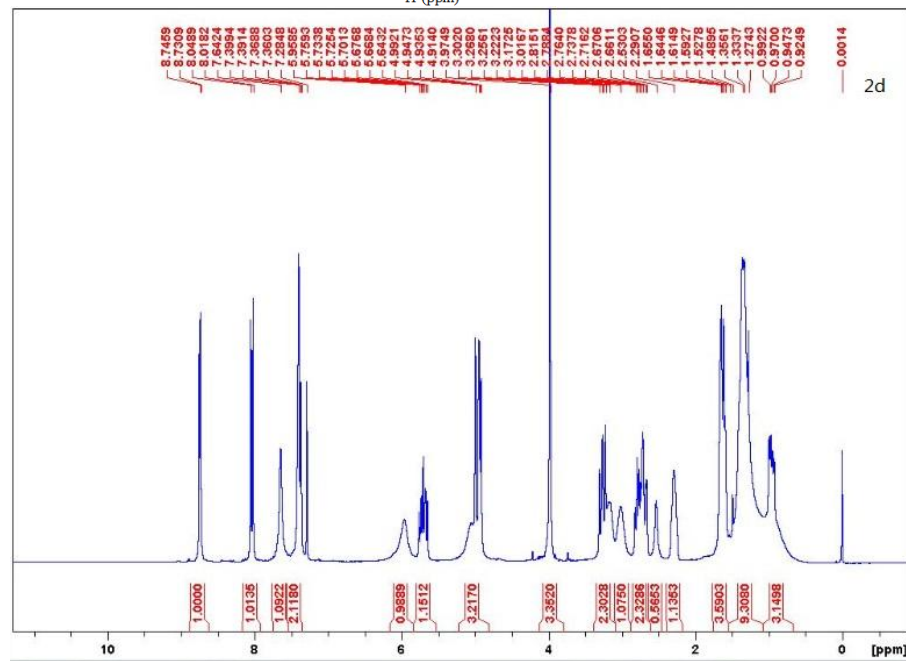
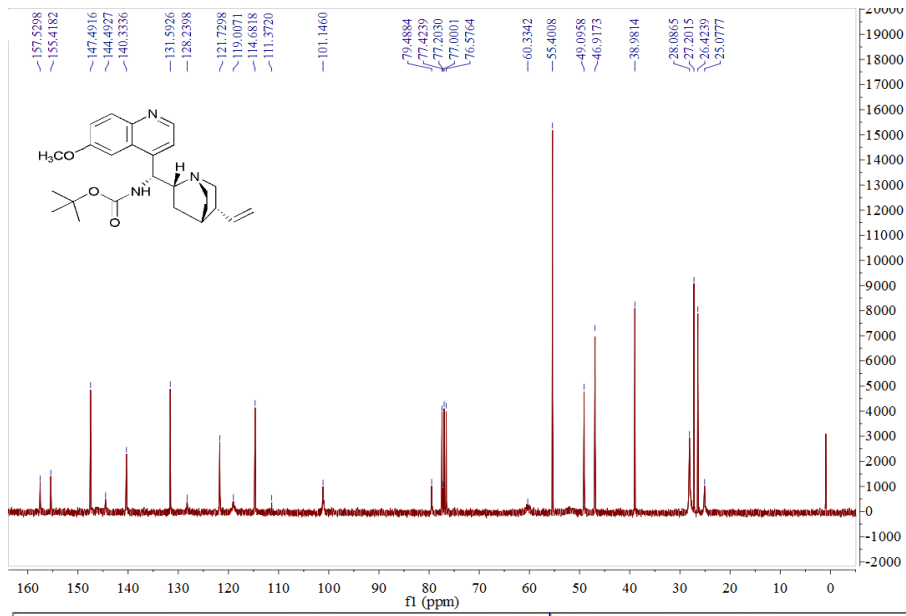
College of Chemistry and Chemical Engineering, Southwest University,
Chongqing, 400715, P. R. China

1. NMR spectra

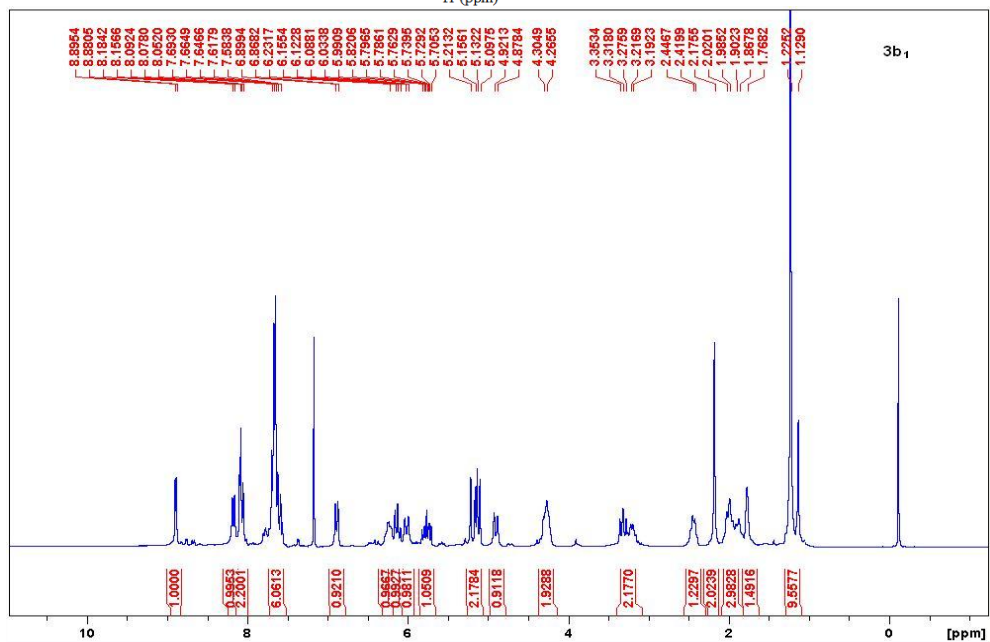
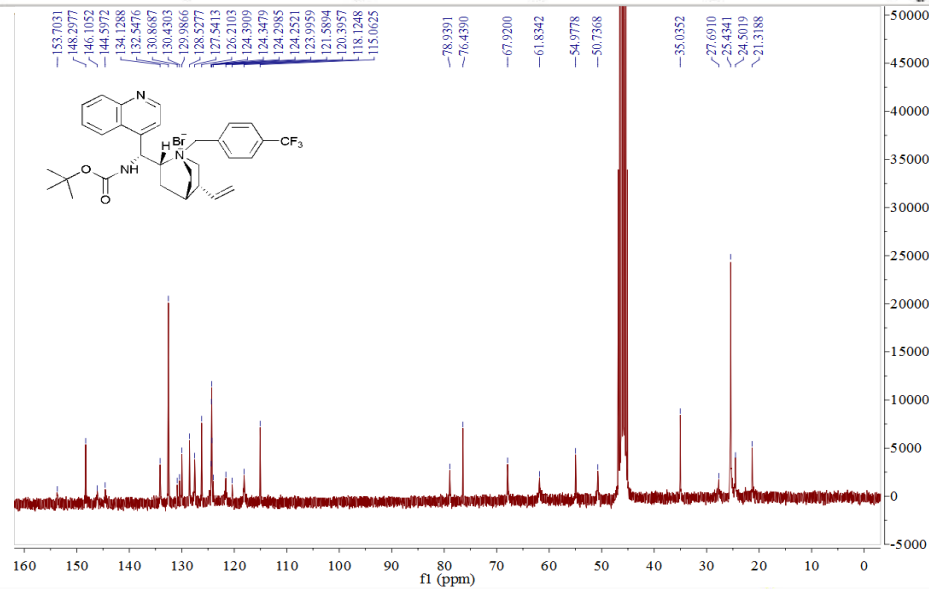
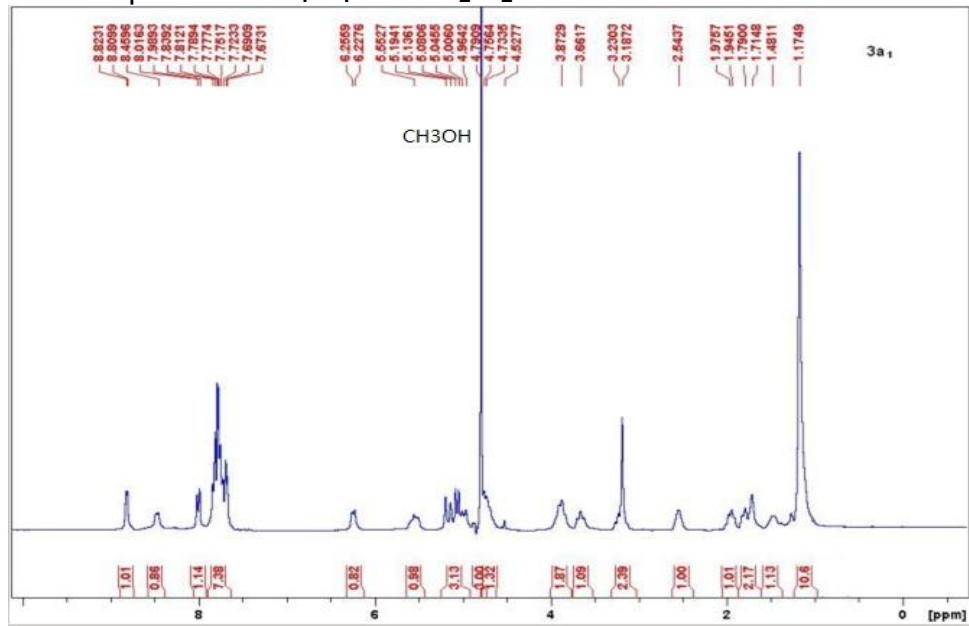
1.1 ¹H and ¹³C NMR spectra of **2a-d**

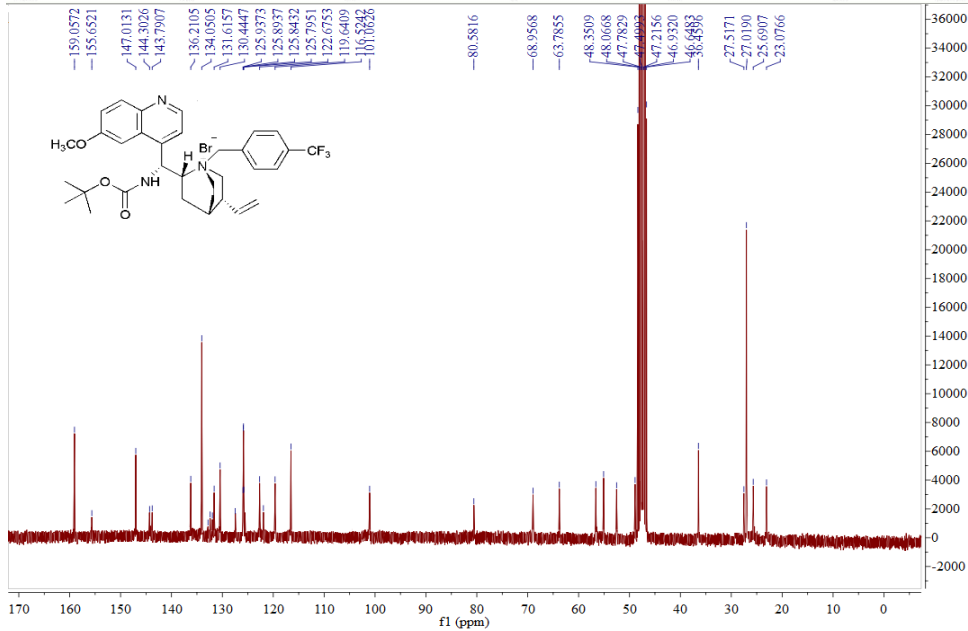
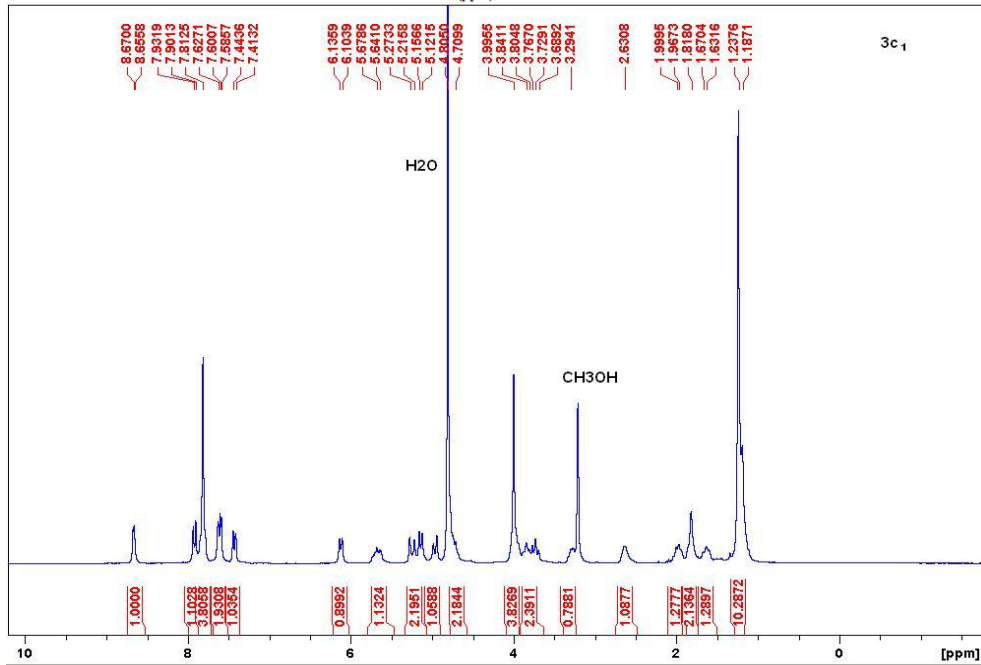
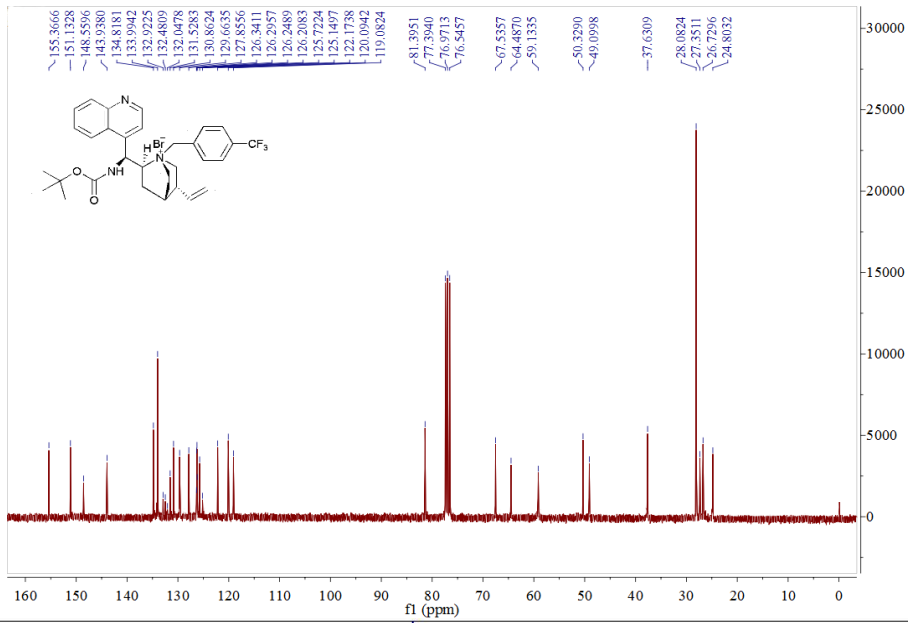


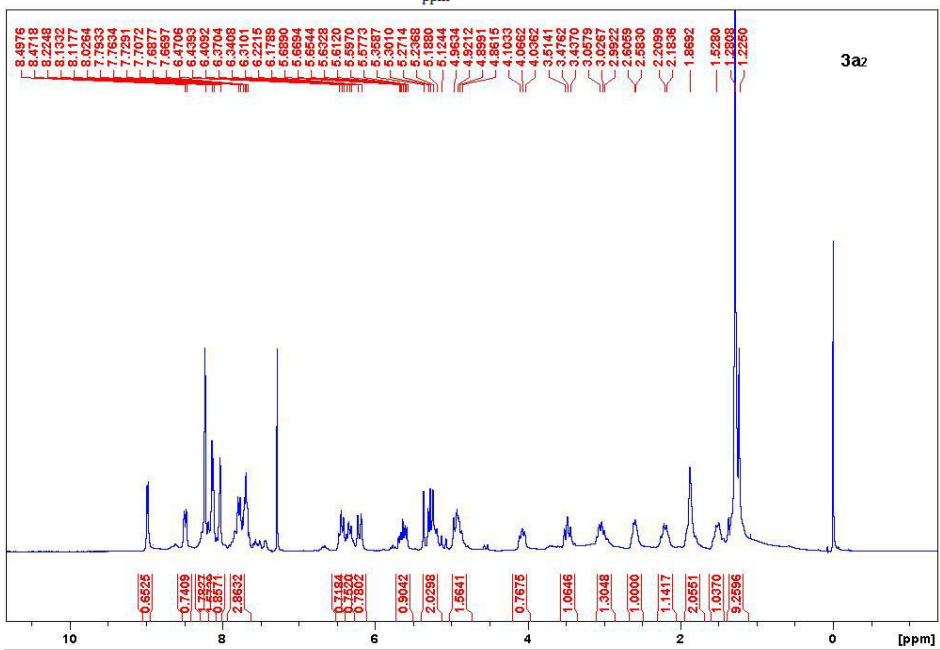
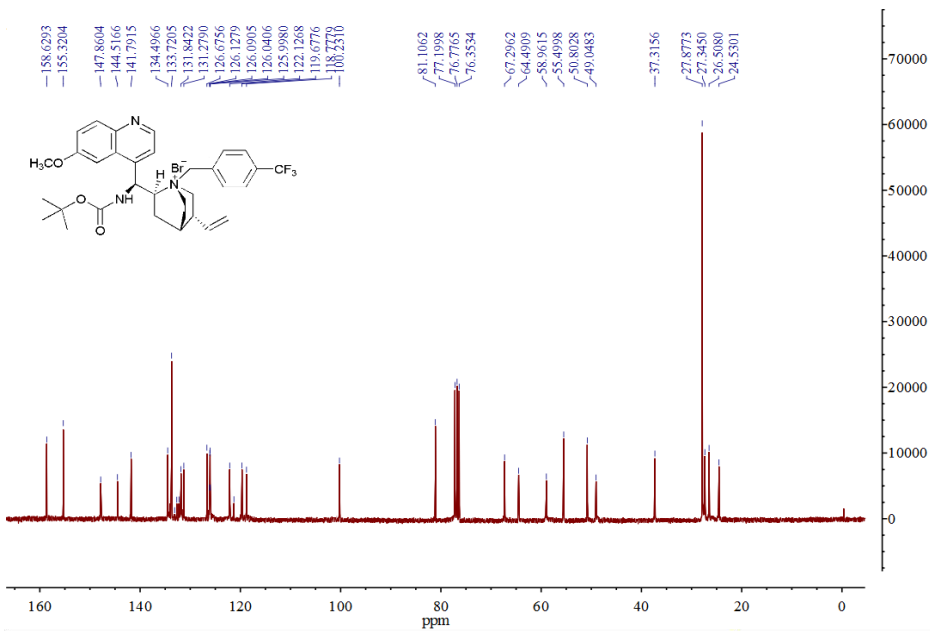
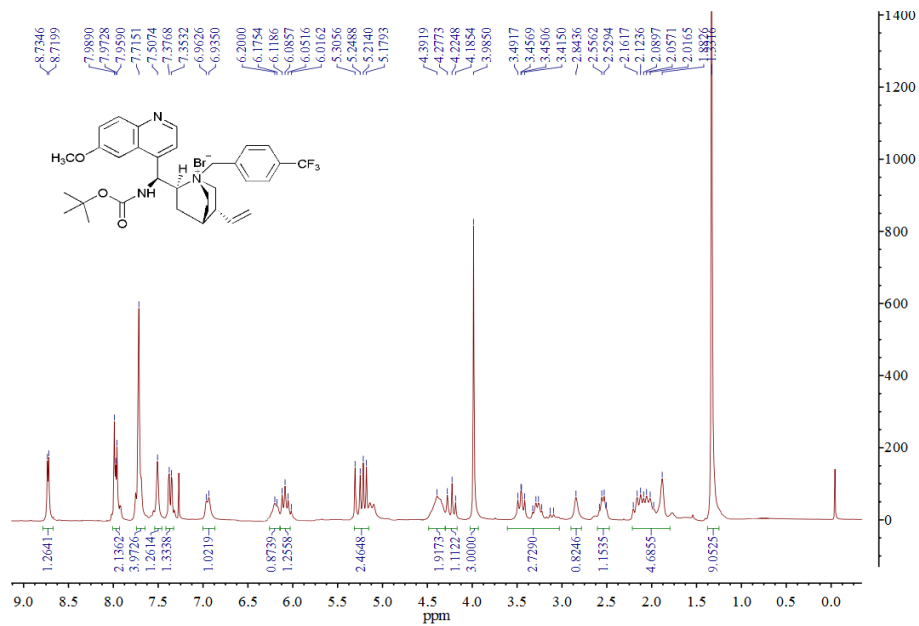


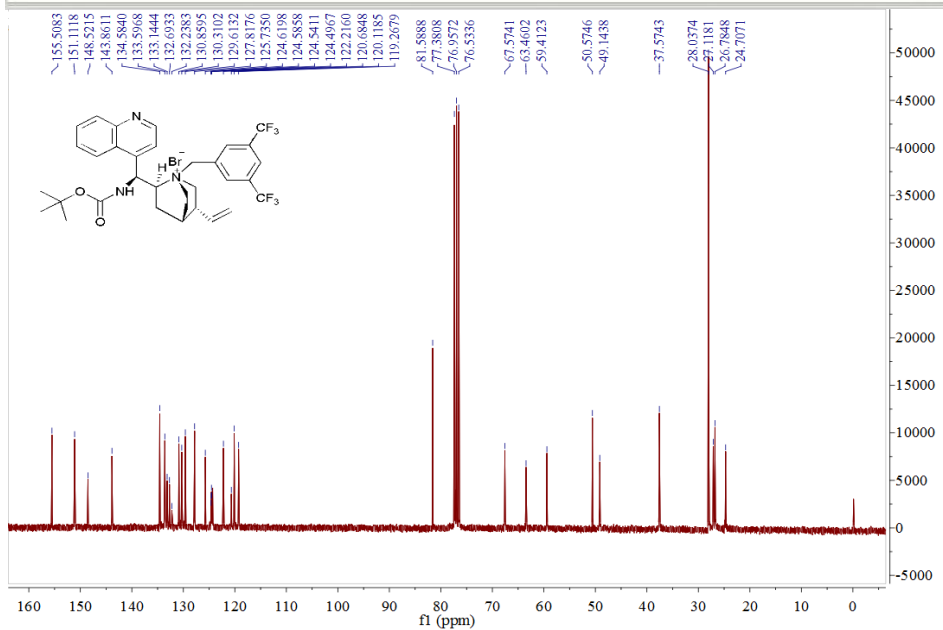
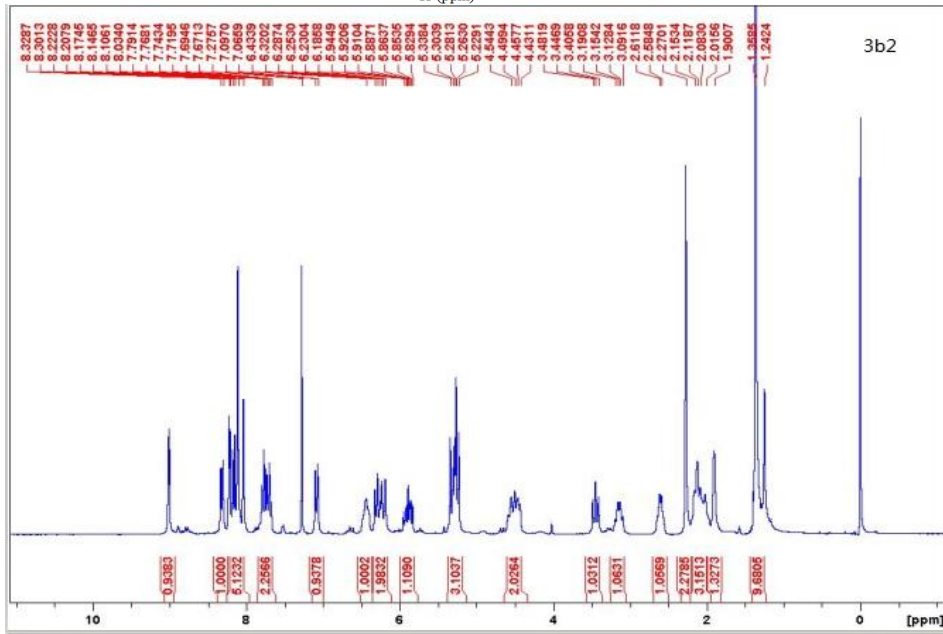
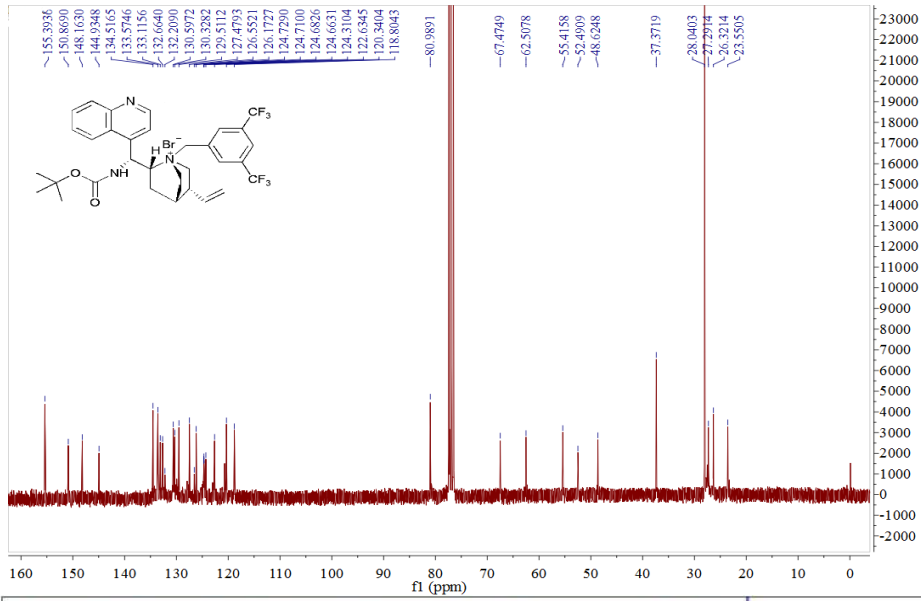


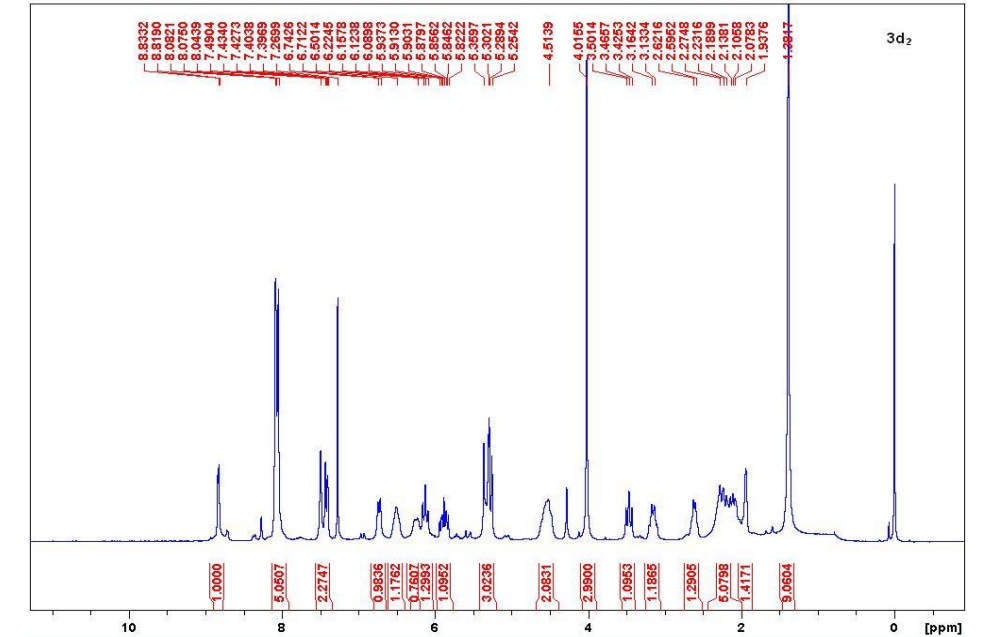
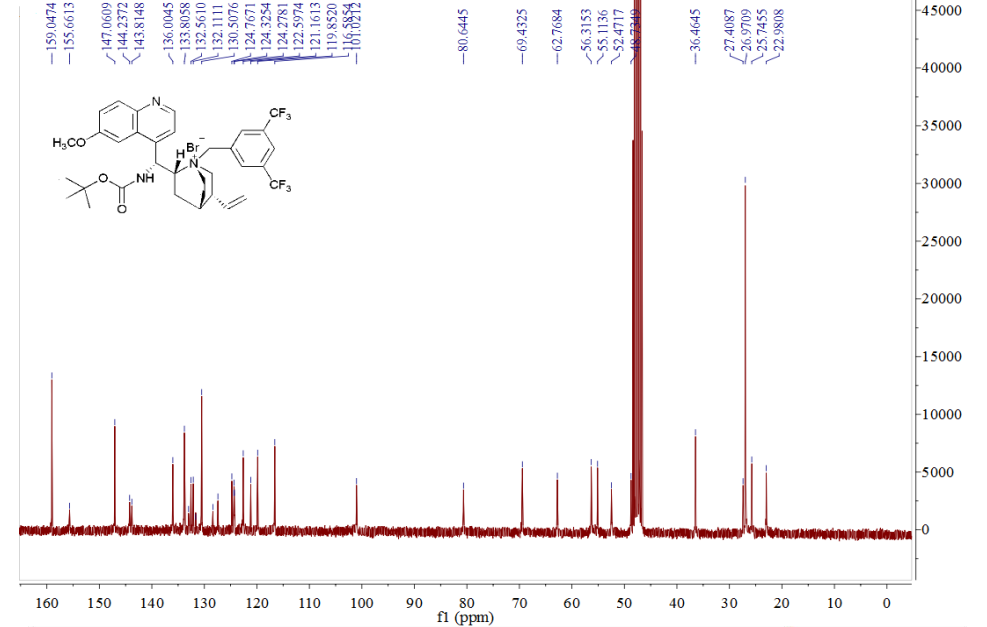
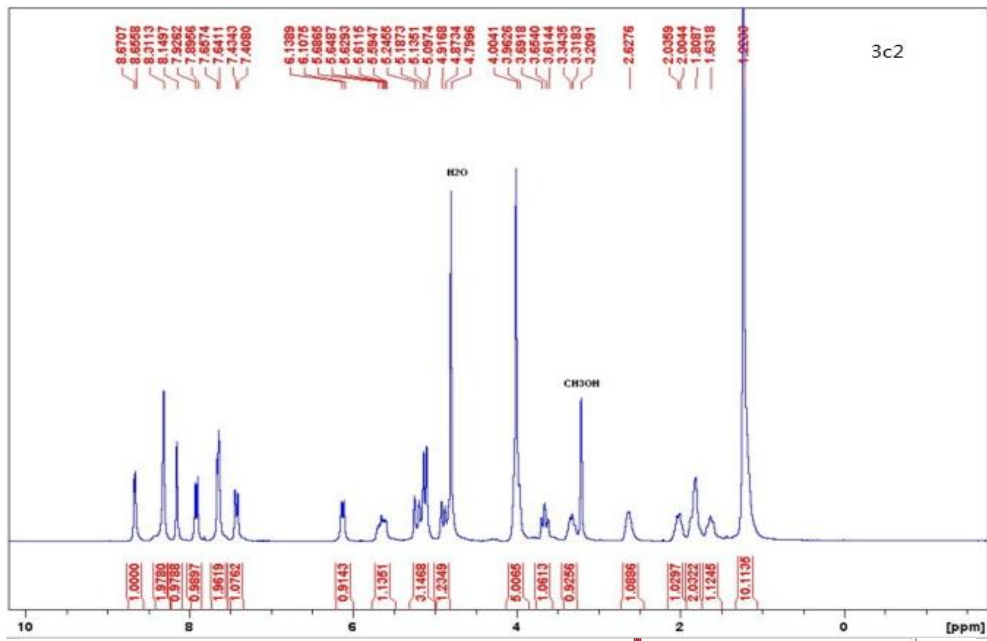
1.2 ^1H and ^{13}C NMR spectra of **3a₁-d₁** and **3a₂-d₂**

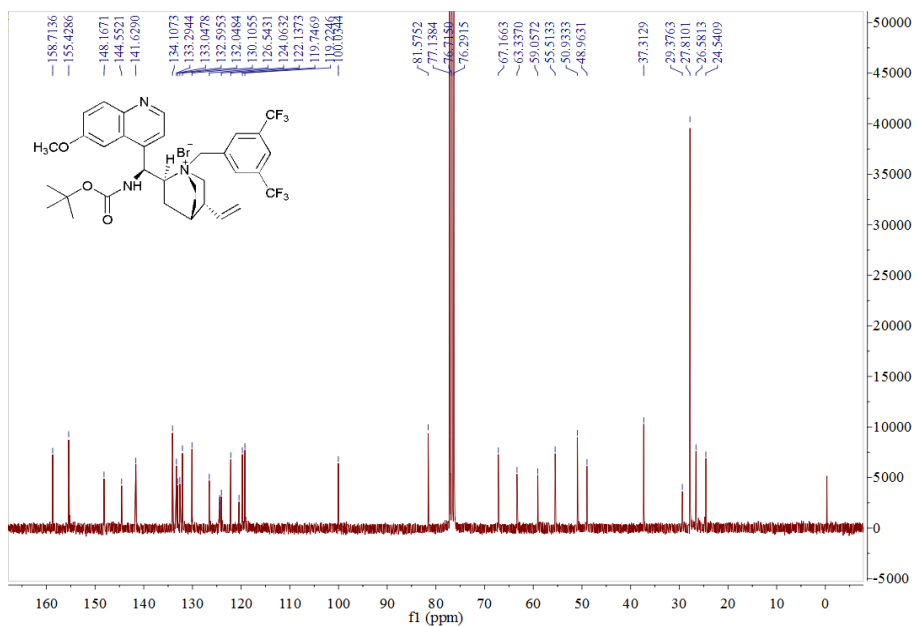




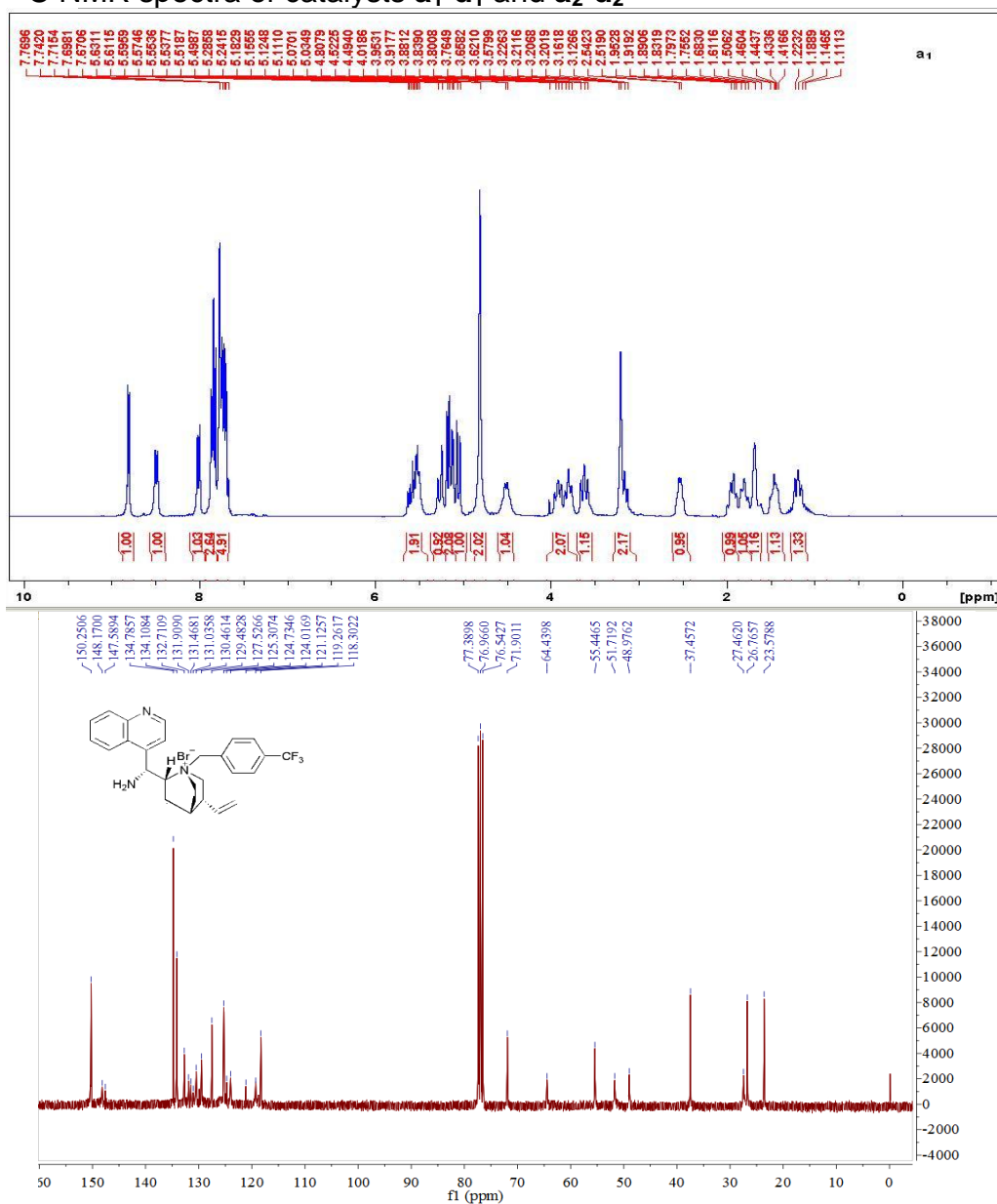


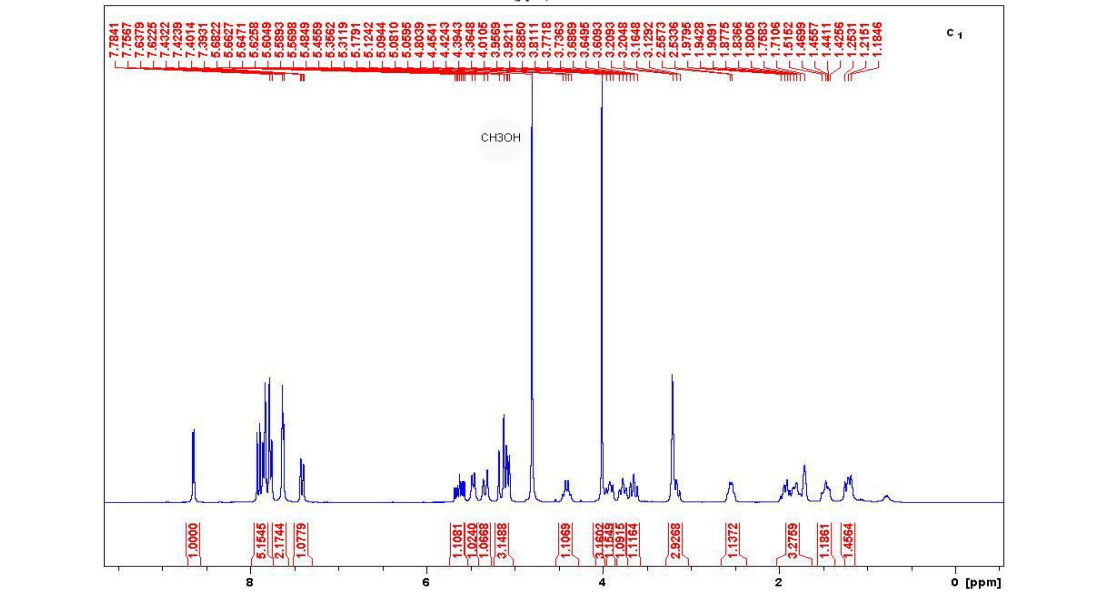
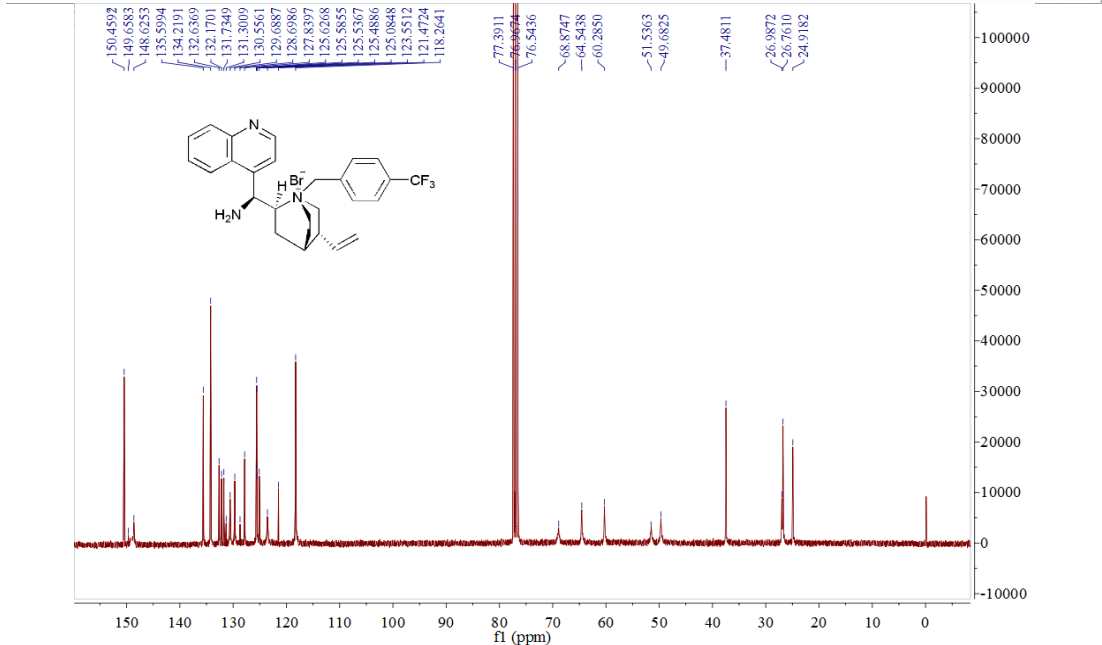
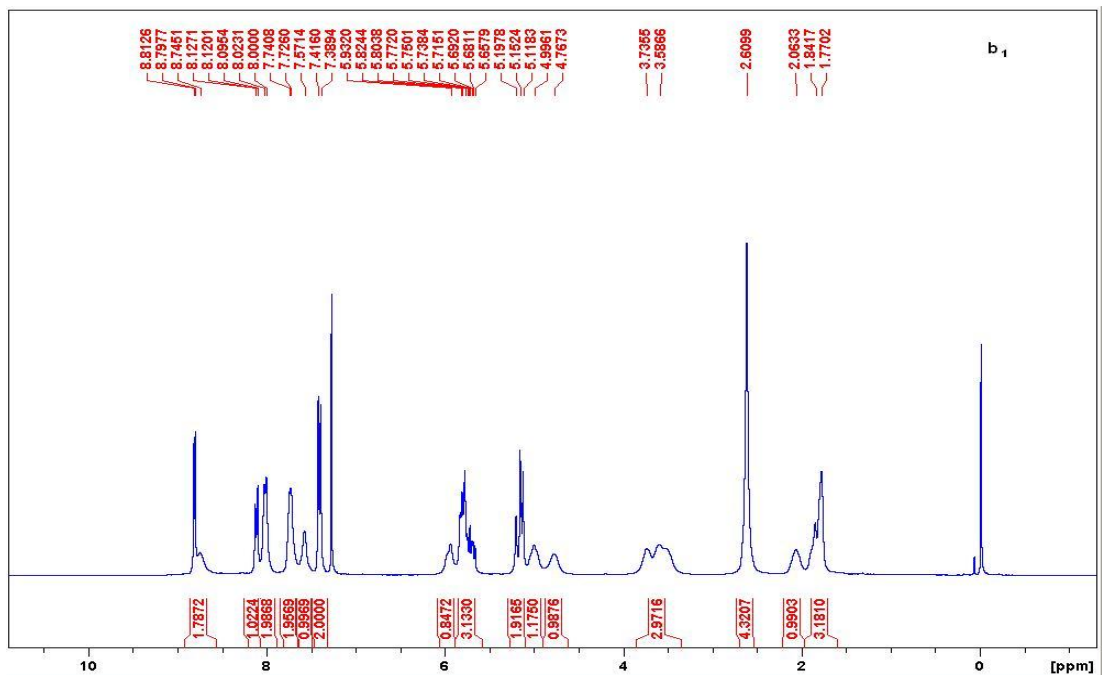


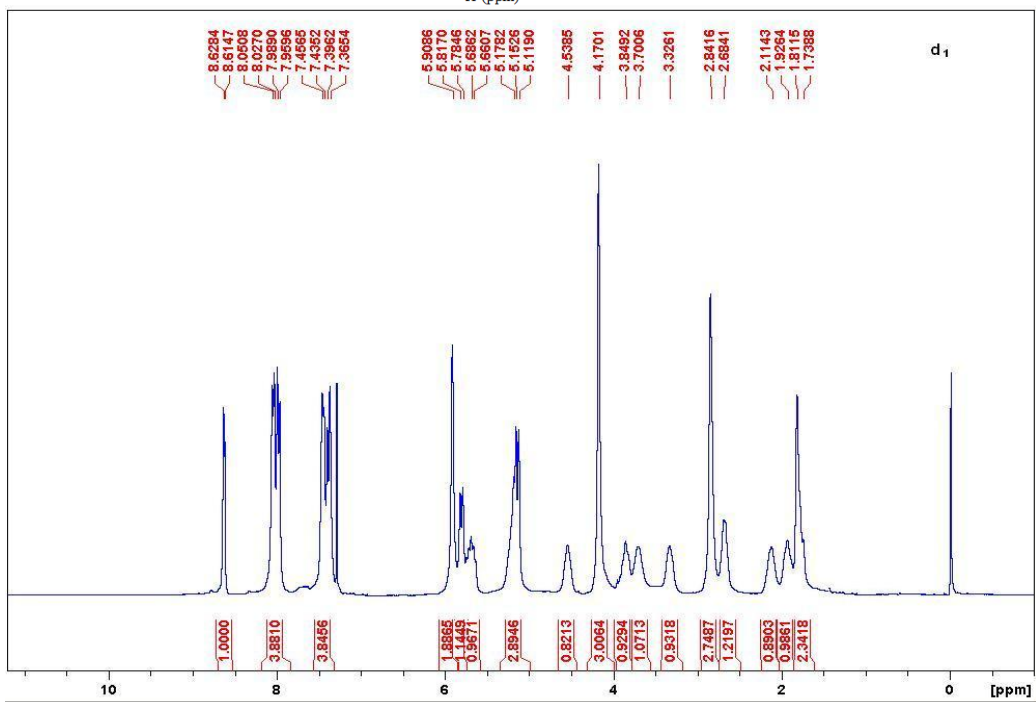
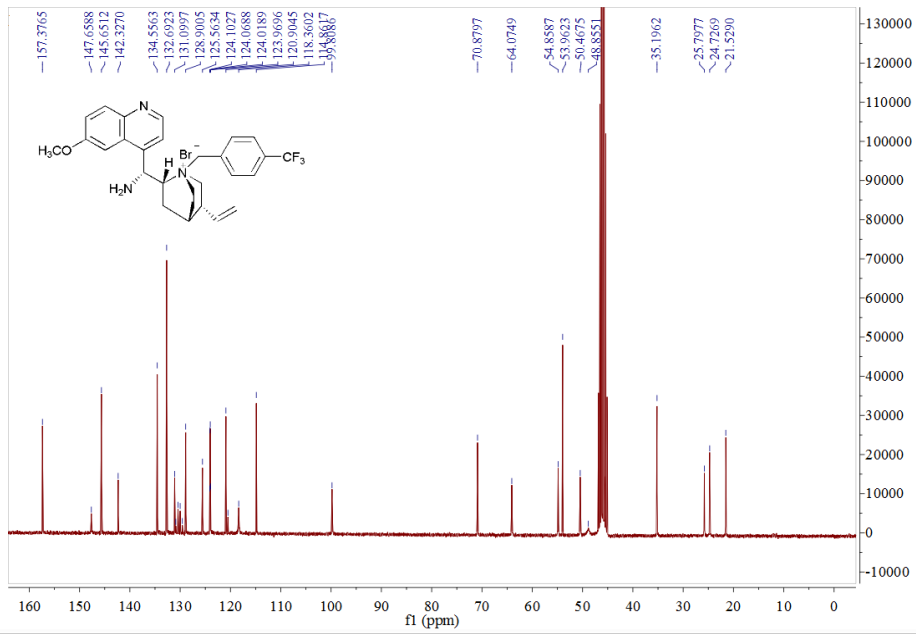


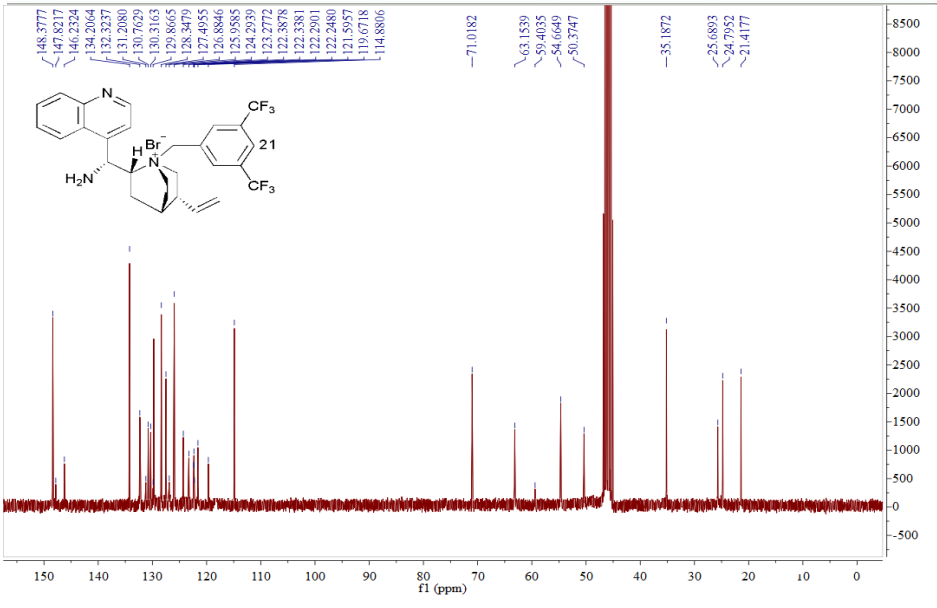
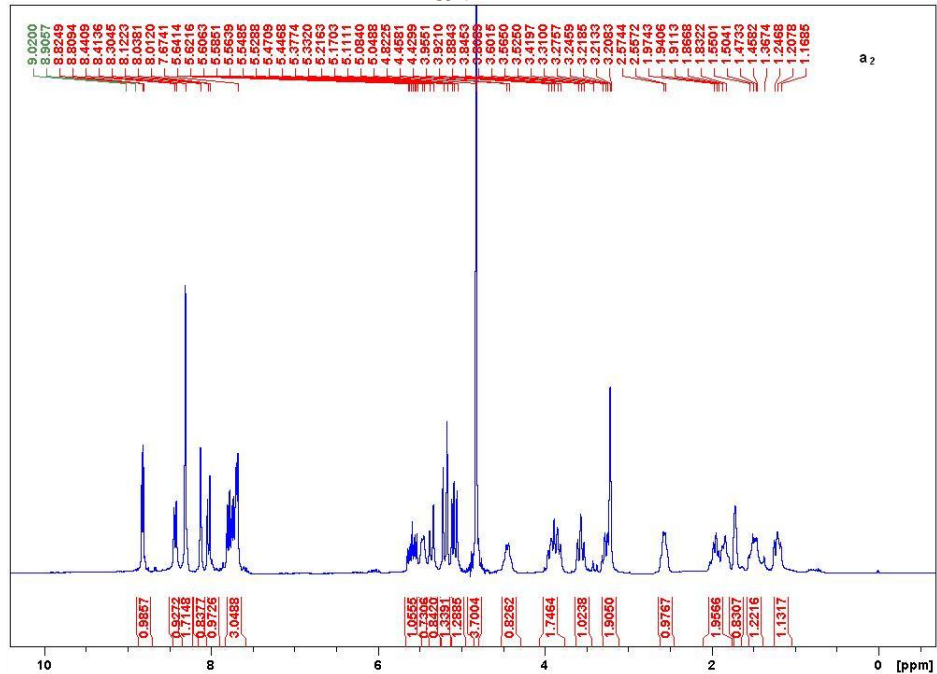
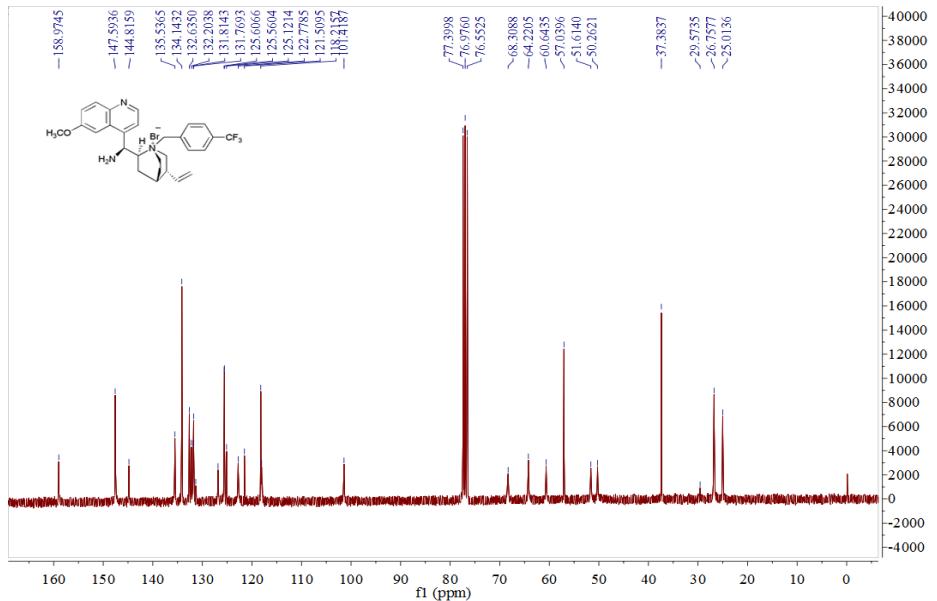


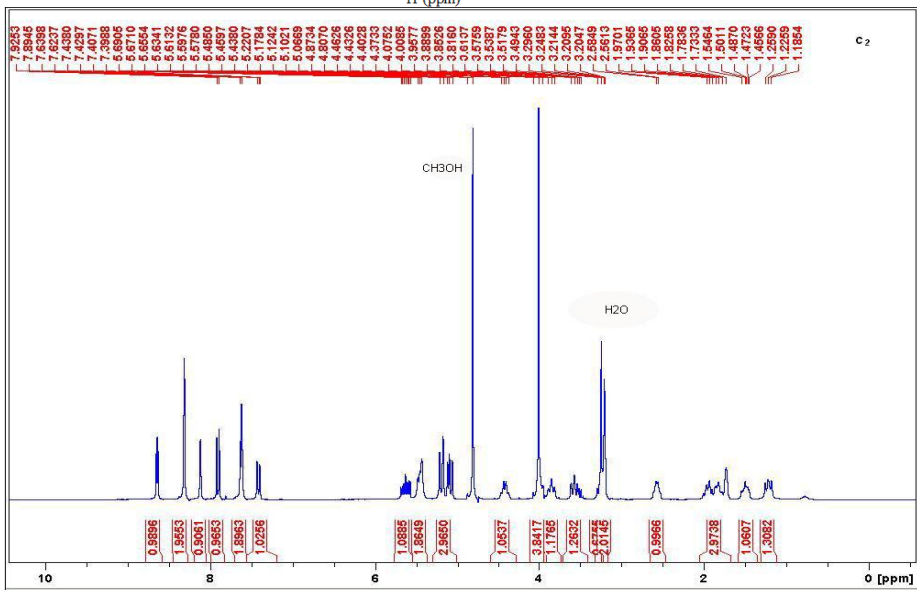
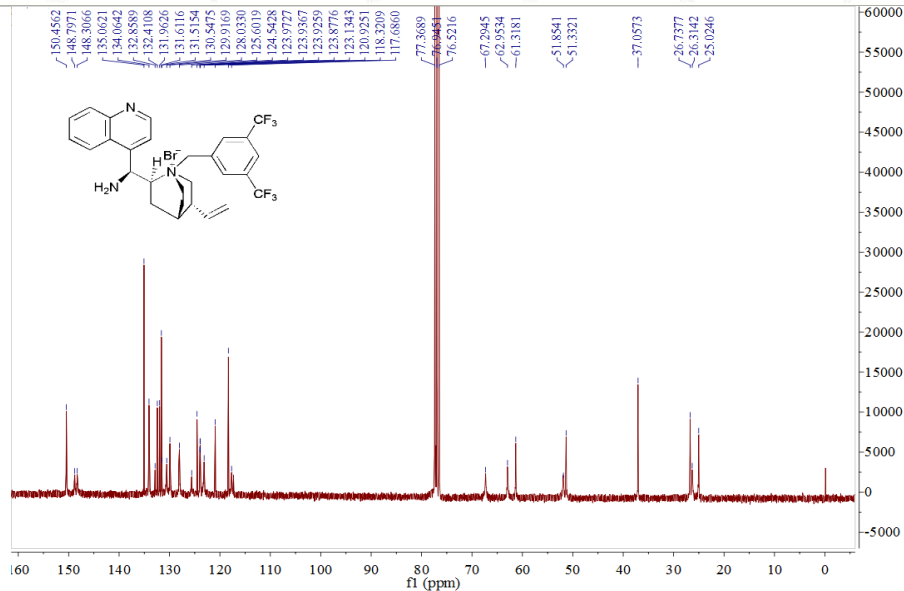
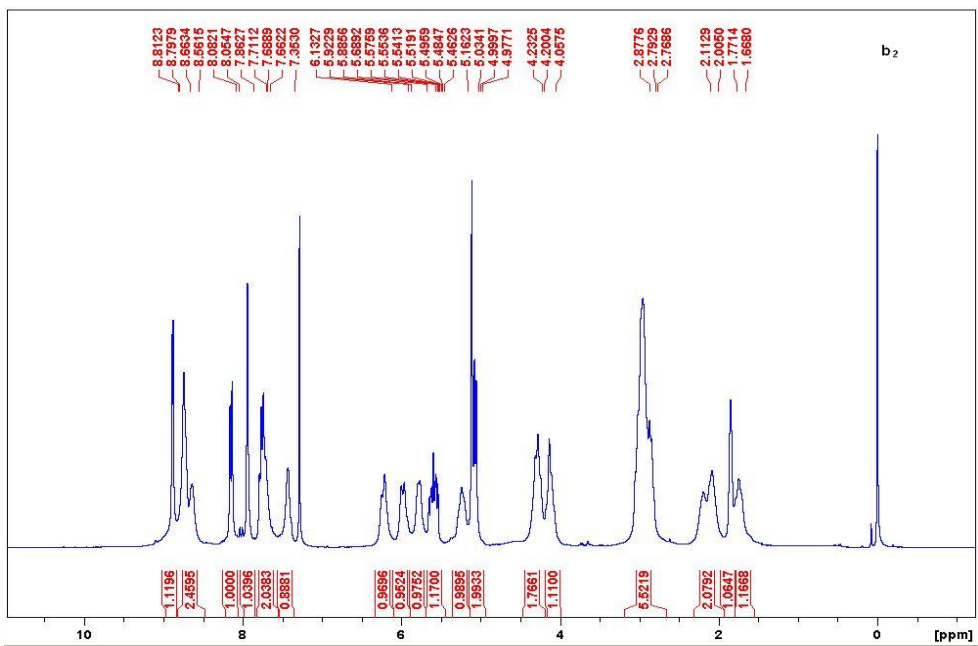
1.3 ¹H and ¹³C NMR spectra of catalysts **a_{1-d1}** and **a_{2-d2}**

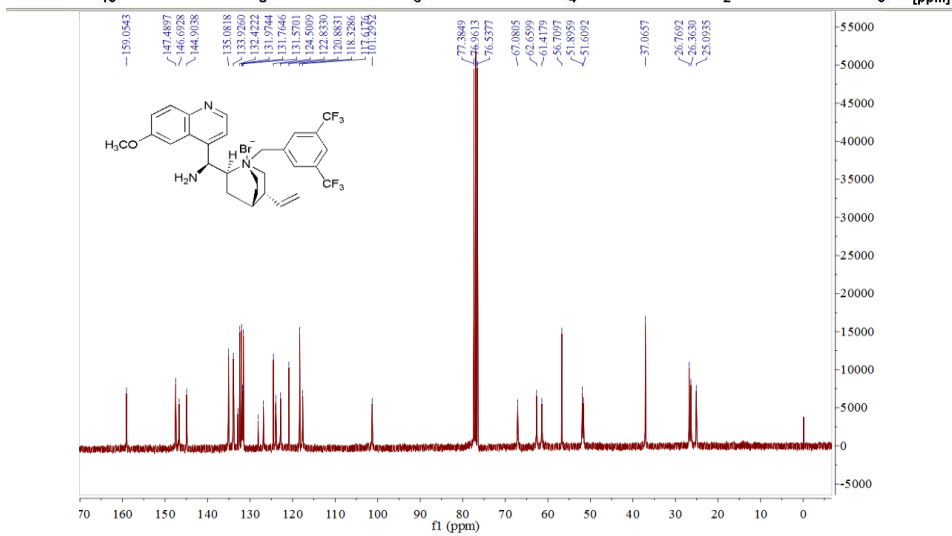
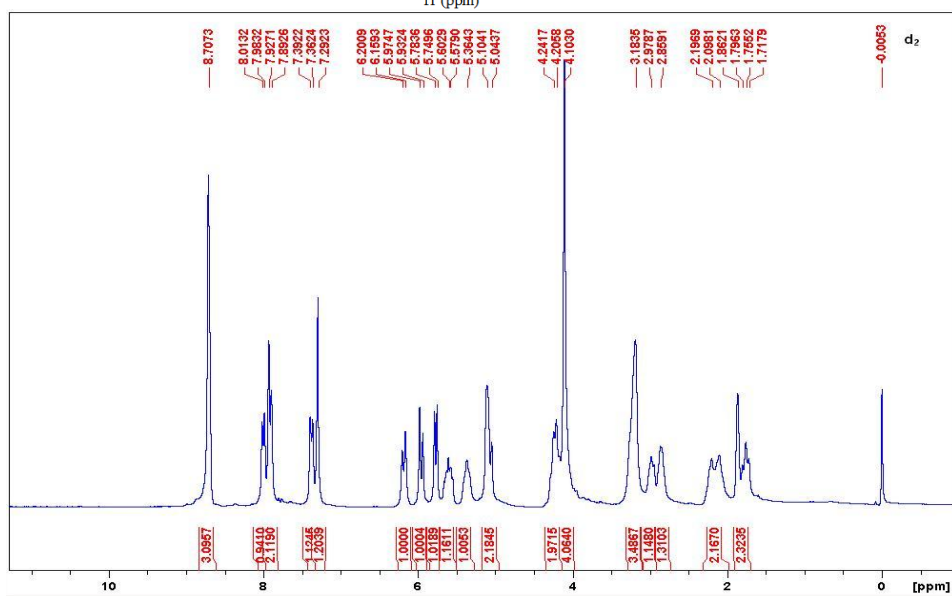
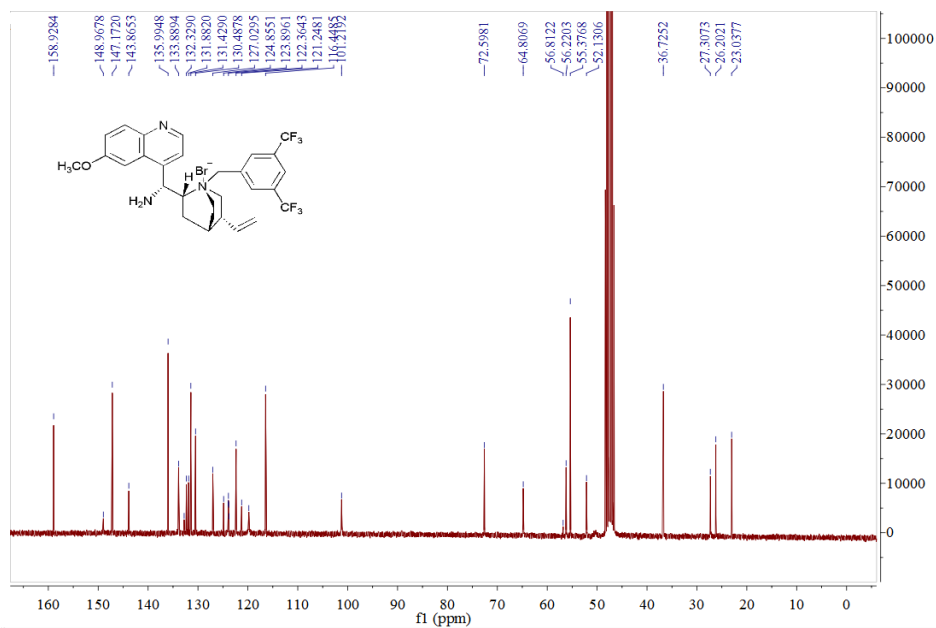










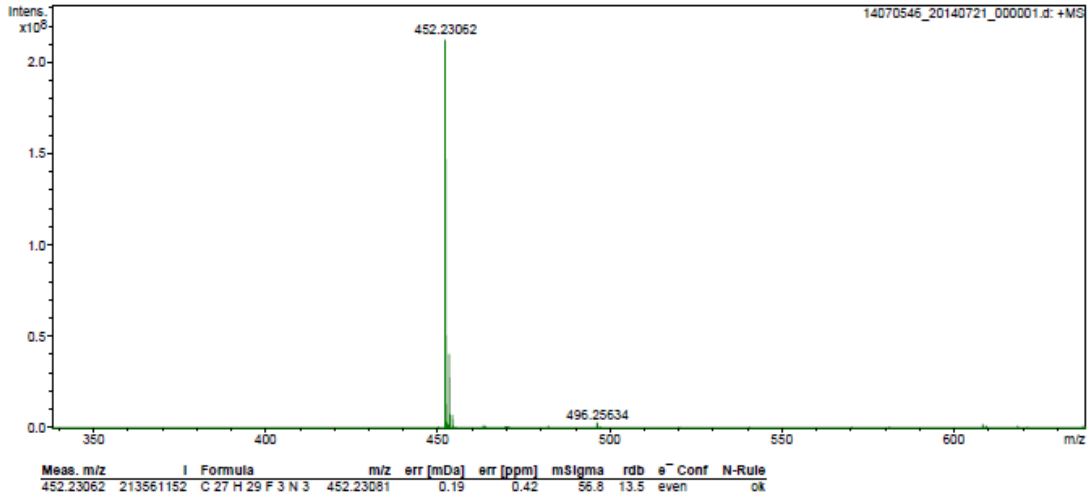


2. High resolution mass spectroscopy (HRMS)

Peking University Mass Spectrometry Sample Analysis Report

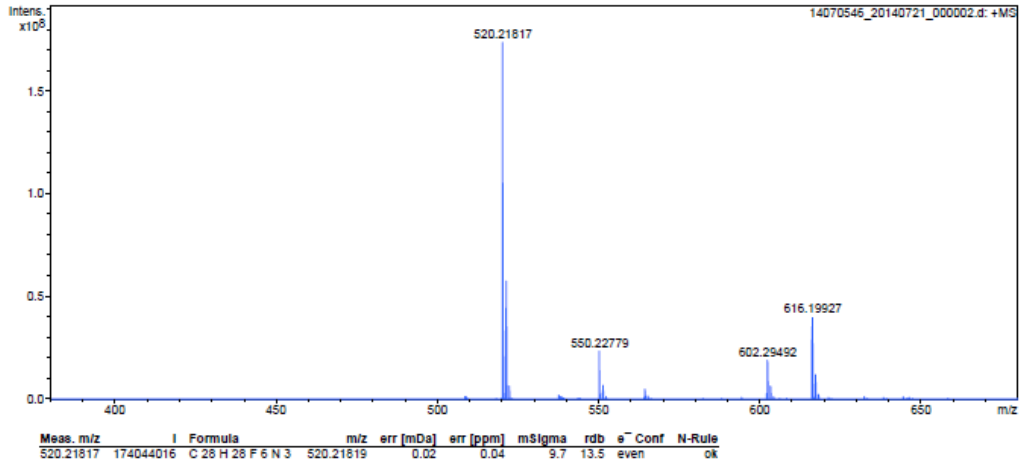
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Comment	ESI Positive	Operator	Peking University



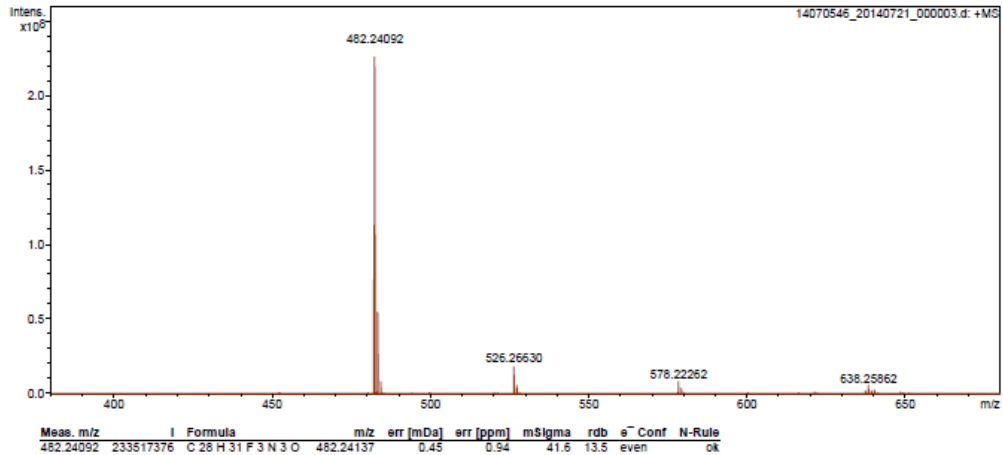
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Analysis Info

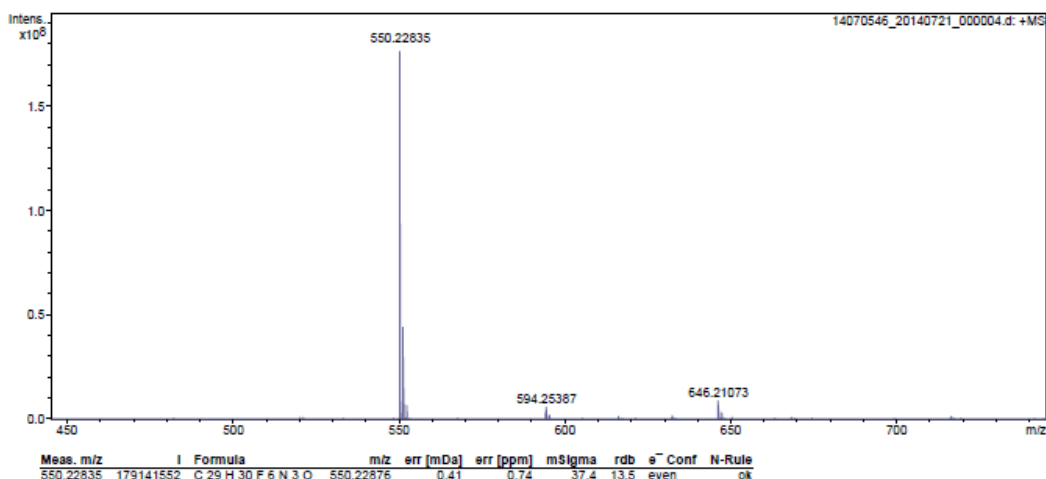
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Comment	ESI Positive	Operator	Peking University



Analysis Info

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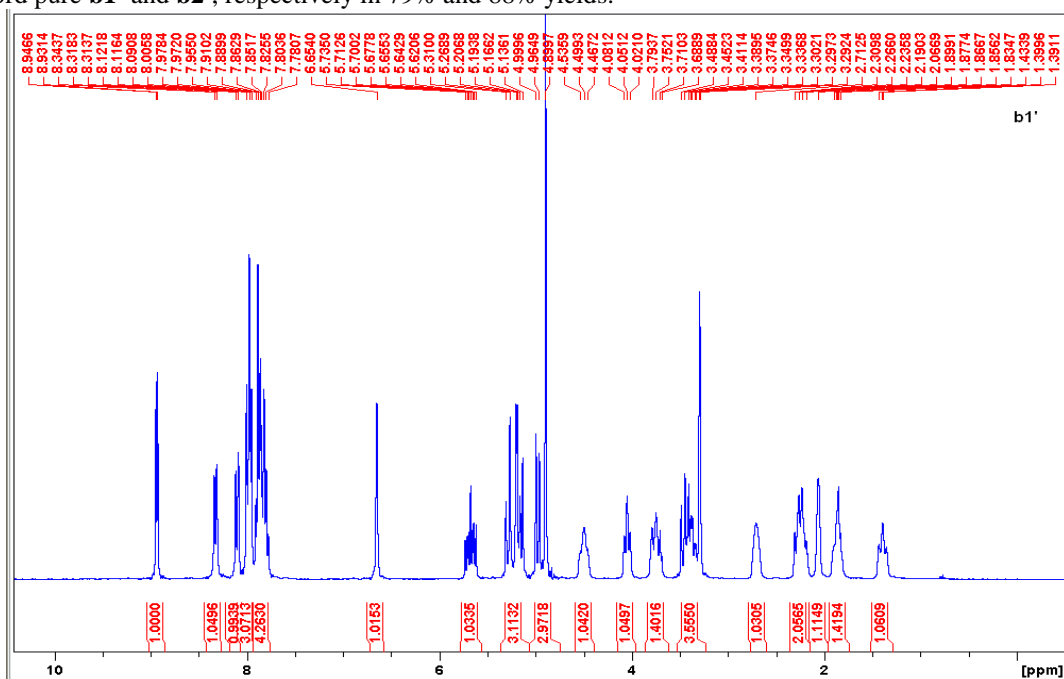
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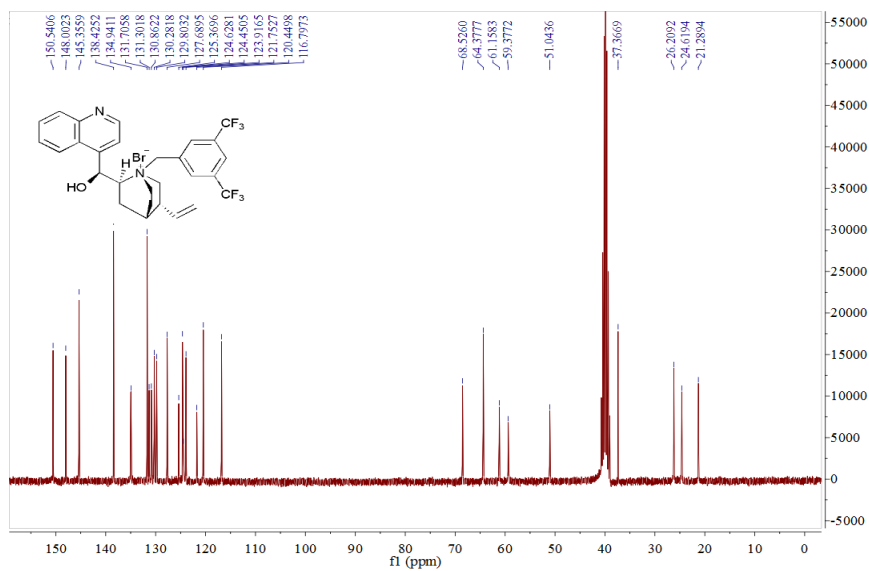
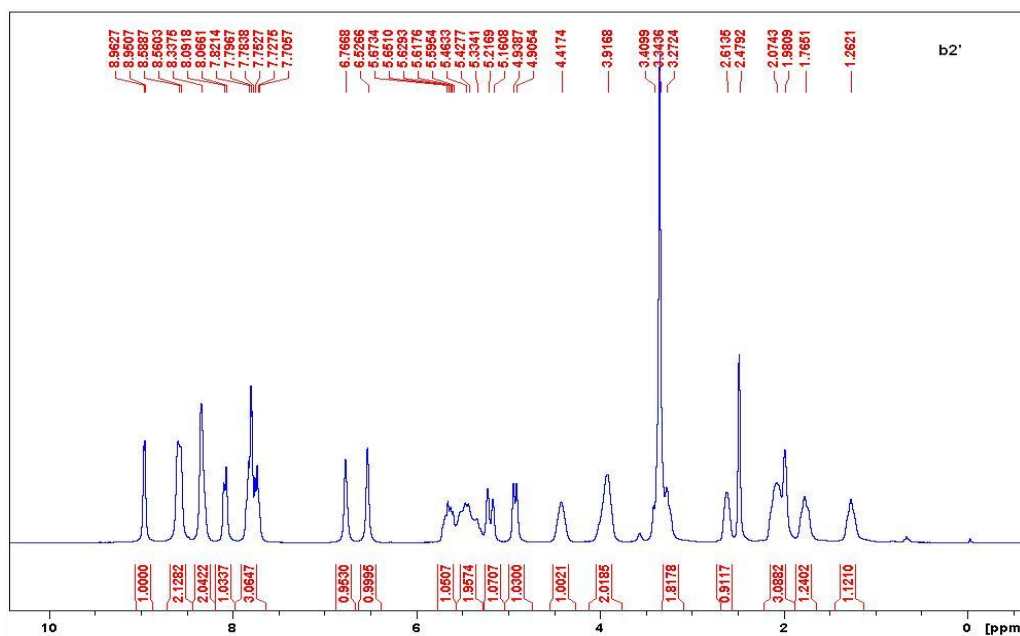
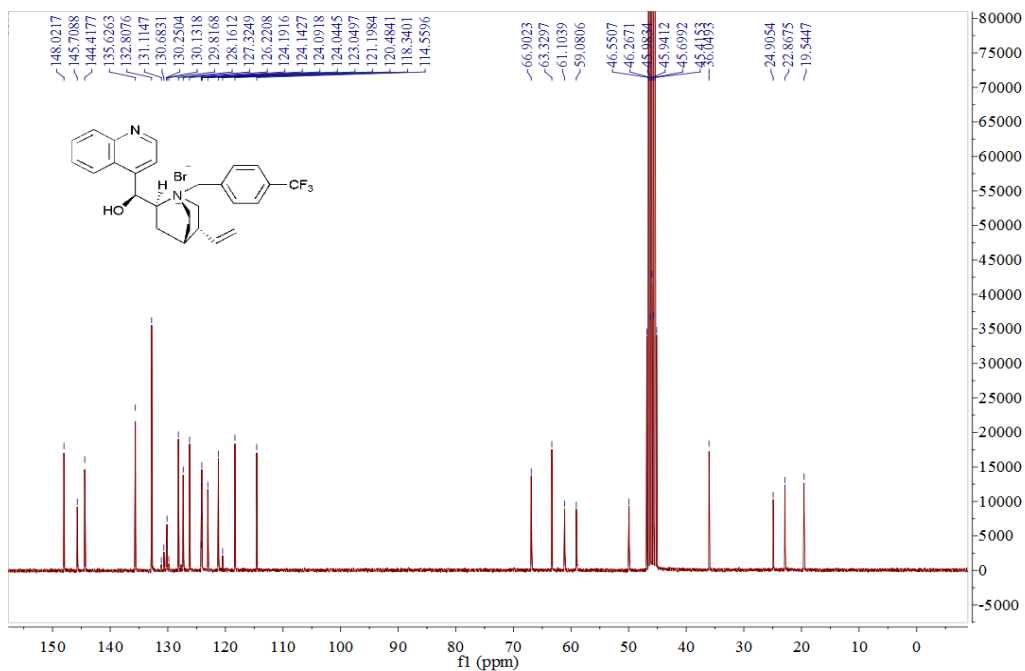


3. Synthesis of N-benzyl ammoniumsalts **b1'** and **b2'**

The first conversion of cinchonidine (CD) to 9-epimers of *cinchona* alkaloids (*epi*-CD) was achieved by one-pot inversion of Mitsunobu esterification–saponification (Ł. Sidorowicz, J. Skarzewski, *Synthesis*, 2011, **5**, 708-710).

The anhydrous THF solution (4 mL) containing CD (0.20 g, 0.68 mmol) and 3, 5-bis(trifluoromethyl)benzyl bromide (0.21 g, 0.68 mmol) or 4- trifluoromethyl benzyl bromide (0.16 g, 0.68 mmol) was stirred at 65 °C for 5 h. During this process, white solid gradually separate out. The white precipitate was filtered, washed with THF (3 mL ×2) and dried under reduced pressure to afford pure **b1'** and **b2'**, respectively in 79% and 88% yields.

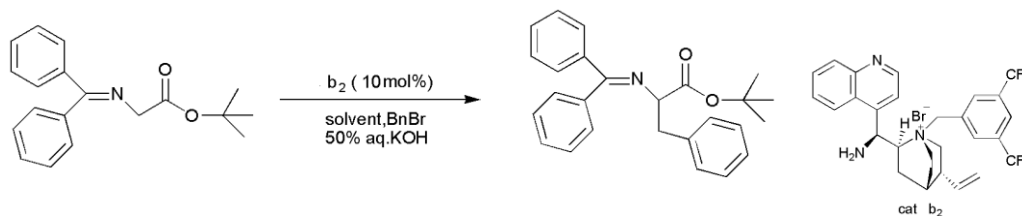




4. The optimization of catalytic conditions

The catalytic reaction conditions such as solvents, temperature, species and concentrations of base and used amounts of catalyst were carefully optimized and shown in Table 1-

Table 1 The influence of various solvents on the catalytic performances ^a



Entry	Solvent	Yield [%] ^b	% ee ^c
1	CH ₂ Cl ₂	85	-
2	<i>t</i> -BuOCH ₃	99	60.7(<i>R</i>)
3	CH ₃ OH	-	-
4	acetone	90	5(<i>R</i>)
5	toluene	98	73.9(<i>R</i>)
6	EAc	98	30.6(<i>R</i>)
7	CH ₃ CN	98	5.0(<i>R</i>)
8	DMF	99	-
9	CHCl ₃	-	-
10	DMSO	-	-
11	ether	99	82.5(<i>R</i>)

^a Reaction conditions: **b**₂, S=0.1 mmol, S/C=10, -20 °C, 12 h, Solvent: 2 mL, KOH: 50% aq 0.4mL.

^b Isolated yield.

^c Determined by charil HPLC.

Table 2 The influence of reaction temperature on catalytic performances^a

Entry	Temp.[°C]	Time	Yield [%] ^b	% ee ^c
1	25	5	99	49.0(<i>R</i>)
2	0	5	99	67.0(<i>R</i>)
3	-20	8	99	82.5(<i>R</i>)
4	-40	12	97	90.5(<i>R</i>)
5	-60	24	73	71.3(<i>R</i>)

^a Reaction conditions: **b**₂, S=0.1 mmol, S/C=10, ether: 2 mL, KOH: 50% aq 0.4 mL

^b Isolated yield.

^c Determined by charil HPLC.

Table 3 The influence of various base on catalytic performances ^a

Entry	Base	Yield [%] ^b	%ee ^c
1	LiOH	52	52.1(<i>R</i>)
2	NaOH	63	59.8(<i>R</i>)
3	KOH	97	90.5(<i>R</i>)
4	CsOH	60	65.0(<i>R</i>)

^a Reaction conditions: **b**₂, S=0.1 mmol, S/C=10, -40 °C, 12 h, ether: 2 mL, MOH: 50% aq 0.4 mL (LiOH:11%).

^b Isolated yield.

^c Determined by charil HPLC.

Table 4 The influence of concentration of the base on catalytic performances ^a

Entry	Wt (%)	Base	Yield [%] ^b	%ee ^c
1	10	KOH	13	85.3(<i>R</i>)
2	20	KOH	21	83.8(<i>R</i>)
3	30	KOH	58	88.7(<i>R</i>)
4	40	KOH	62	88.2(<i>R</i>)
5	50	KOH	97	90.5(<i>R</i>)

^a Reaction conditions: **b**₂, S=0.1 mmol, S/C=10, -40 °C, 12 h, ether: 2 mL, KOH: 0.4 mL

^b Isolated yield.

^c Determined by charil HPLC.

Table 5 The influence of loading amount on catalytic performances ^a

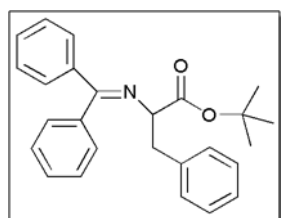
Entry	S/C	Cat. (mol%)	Yield [%] ^b	%ee ^c
1	20	5	98	87.9(<i>R</i>)
2	10	10	97	90.5(<i>R</i>)
3	7	15	84	87.9(<i>R</i>)
4	5	20	82	87.9(<i>R</i>)
5	4	25	78	85.7(<i>R</i>)

^a Reaction conditions: **b**₂, S=0.1 mmol, -40 °C, 12 h, ether: 2 mL, KOH: 50% aq. 0.4 mL.

^b Isolated yield.

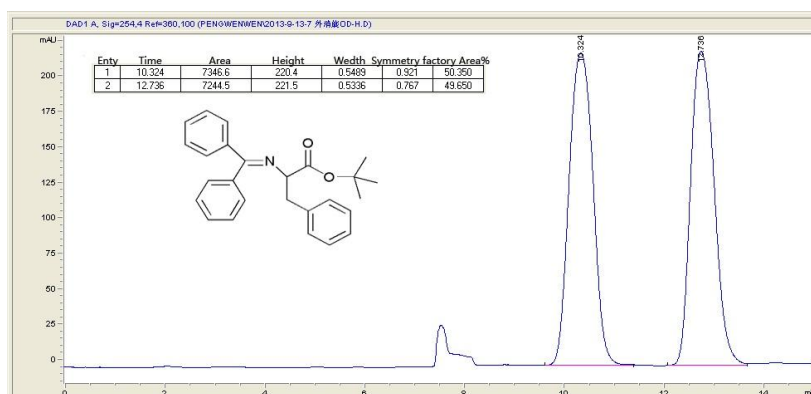
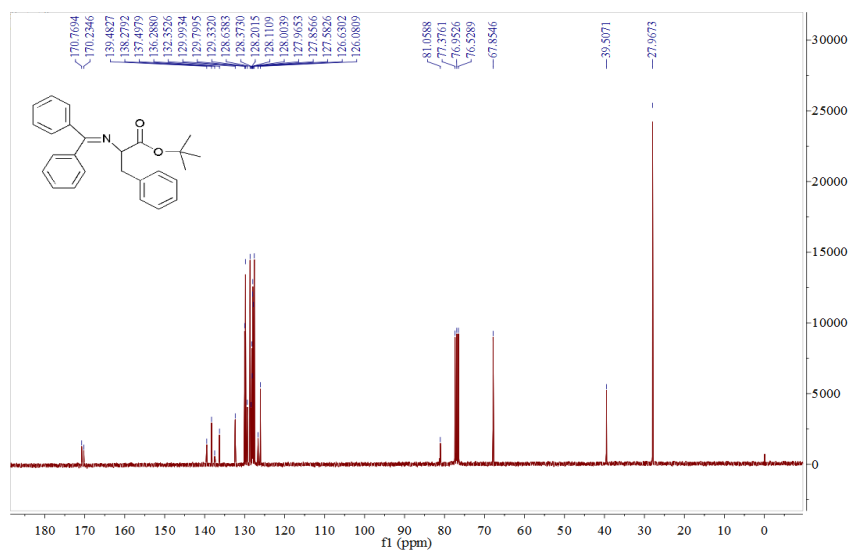
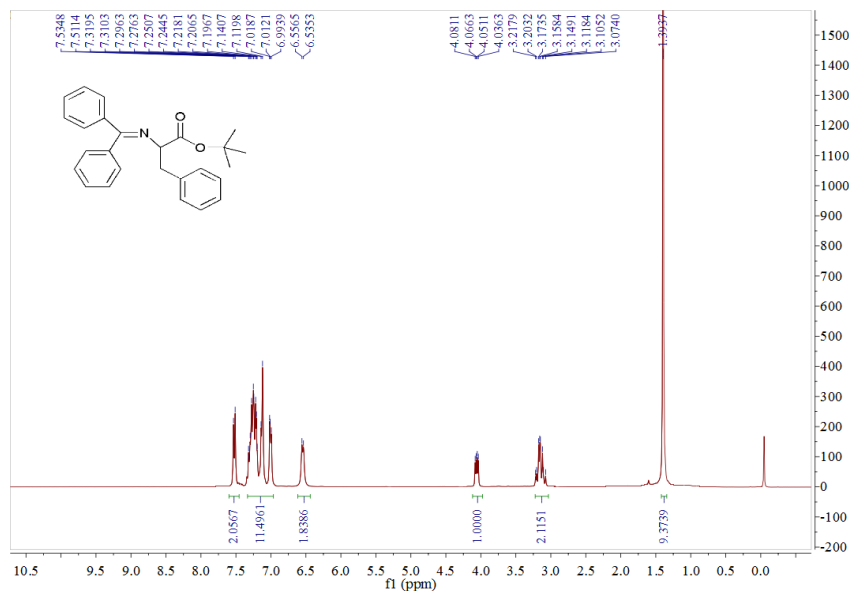
^c Determined by charil HPLC.

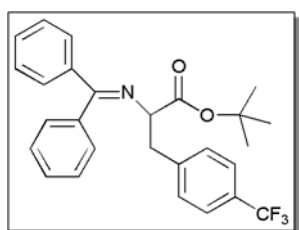
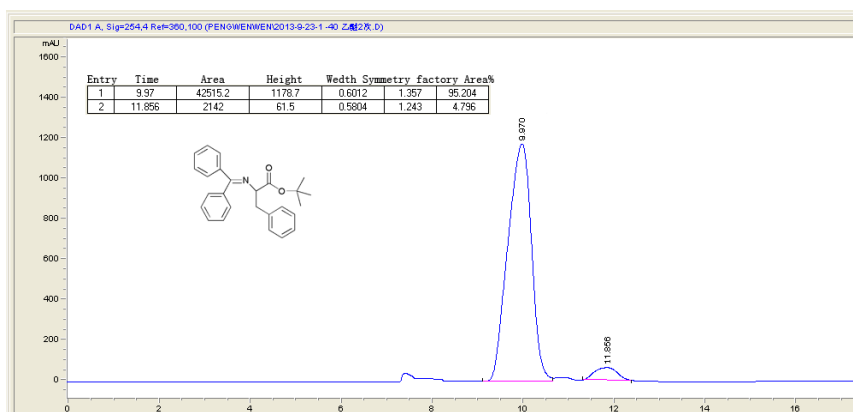
5. The data of catalytic products



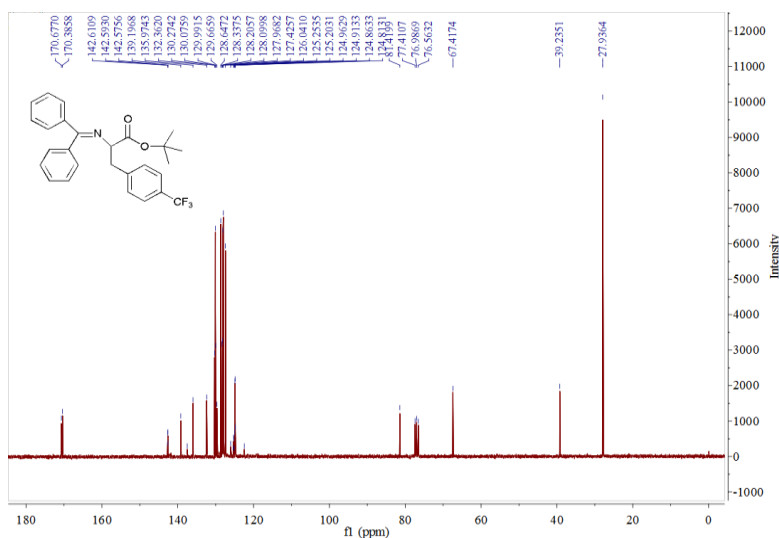
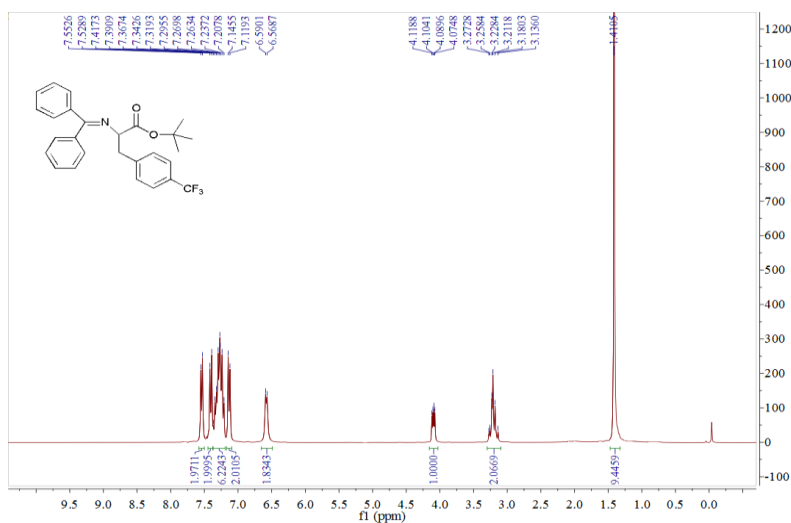
¹H NMR (300 MHz, CDCl₃, TMS): δ 7.52 (d, 2 H, ³J=7.0 Hz, Ph-H), 7.30–7.00 (m, 11 H, Ph-H), 6.54 (d, 2H, ³J=6.4 Hz, Ph-H), 4.06 (dd, 1 H, ³J = 4.4 Hz, 4.4 Hz, CH), 3.22–3.07 (m,

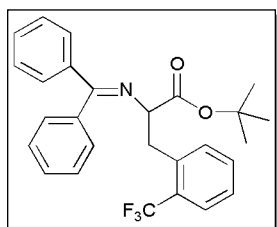
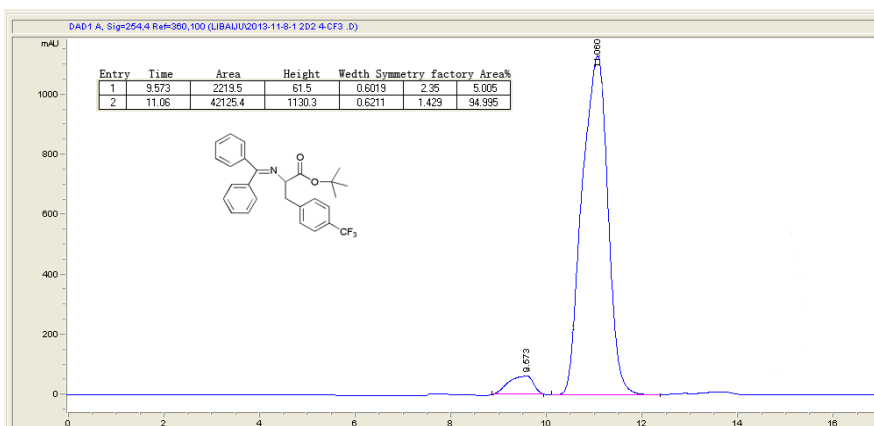
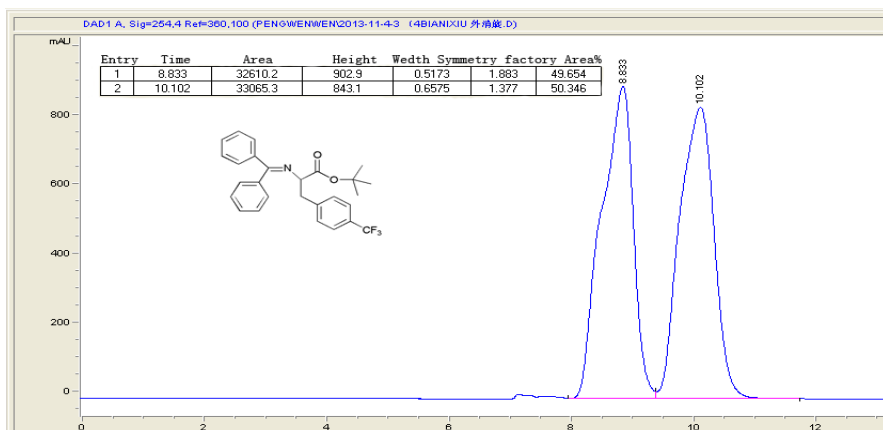
2 H, CH₂), 1.39 (s, 9 H, CH₃). ¹³C NMR (75 MHz, CDCl₃, TMS): δ 170.8, 170.2 (C=N, C=O), 139.5, 138.3, 137.5, 136.3, 132.4, 130.0, 129.8, 129.3, 128.6, 128.4, 128.2, 128.1, 128.0, 128.0, 127.9, 127.6, 126.6, 126.1, 81.1 (O-C), 67.9 (CH), 39.5 (CH₂), 28.0 (CH₃). Rt HPLC (Daicel Chiralpak OD-H, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.0 min (*R*-isomer), 11.9min (*S*-isomer).





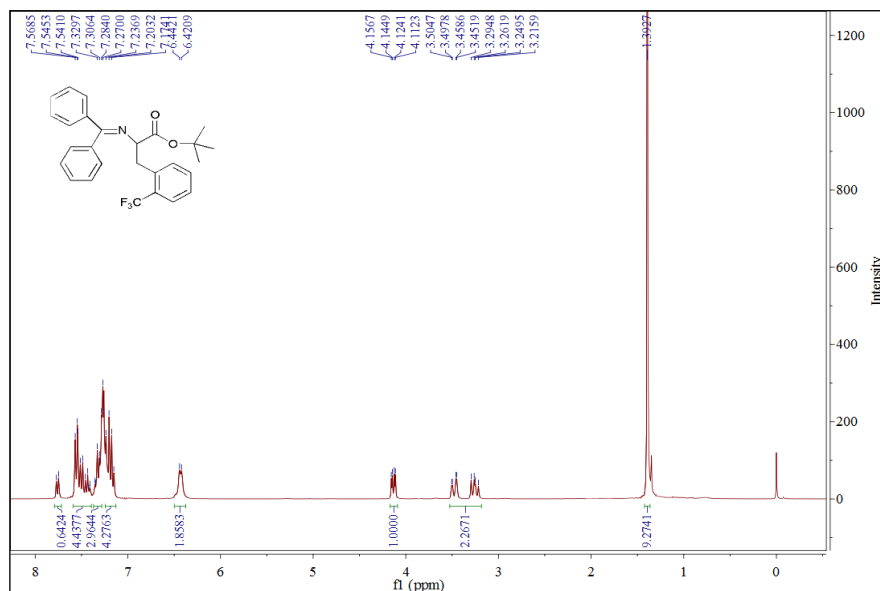
$^1\text{H NMR}$ (300 MHz, CDCl_3 , TMS): δ 7.54 (d, 2 H, $^3J = 7.1$ Hz, Ph-H), 7.40 (d, 2 H, $^3J = 7.9$ Hz, Ph-H), 7.37–7.21 (m, 6 H, Ph-H), 7.13 (d, 2 H, $^3J = 7.9$ Hz, Ph-H), 6.58 (d, 2 H, $^3J = 6.4$ Hz, Ph-H), 4.10 (dd, 1 H, $^3J = 4.4$ Hz, 4.4 Hz, CH), 3.27–3.14 (m, 2 H, CH_2), 1.41 (s, 9 H, CH_3). $^{13}\text{C NMR}$ (75 MHz, CDCl_3 , TMS): δ 170.7, 170.4 (C=N, C=O), 142.6 (q, $^3J_{\text{C-F}} = 1.3$ Hz), 139.2, 137.5, 136.0, 132.4, 130.3, 130.1, 130.0, 130.0, 129.7, 128.6, 128.3, 128.2, 128.1, 128.0, 127.4, 126.0, 125.2 (q, $^2J_{\text{C-F}} = 3.8$ Hz), 124.9 (q, $^1J_{\text{C-F}} = 3.8$ Hz, CF_3), 81.4 (O-C), 67.4 (CH), 39.2 (CH_2), 27.9 (CH_3). Rt HPLC (Daicel Chiralpak OD-H, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 9.6 min (*S*-isomer), 11.1min (*R*-isomer).

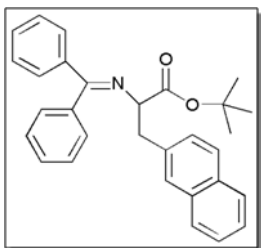
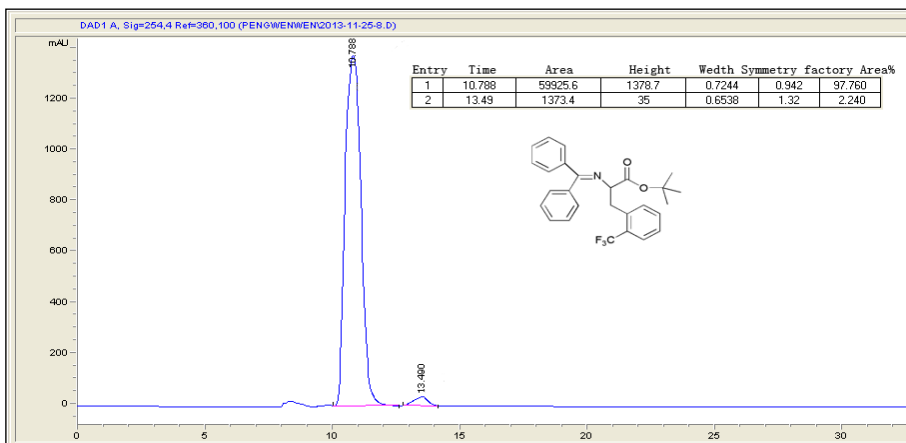
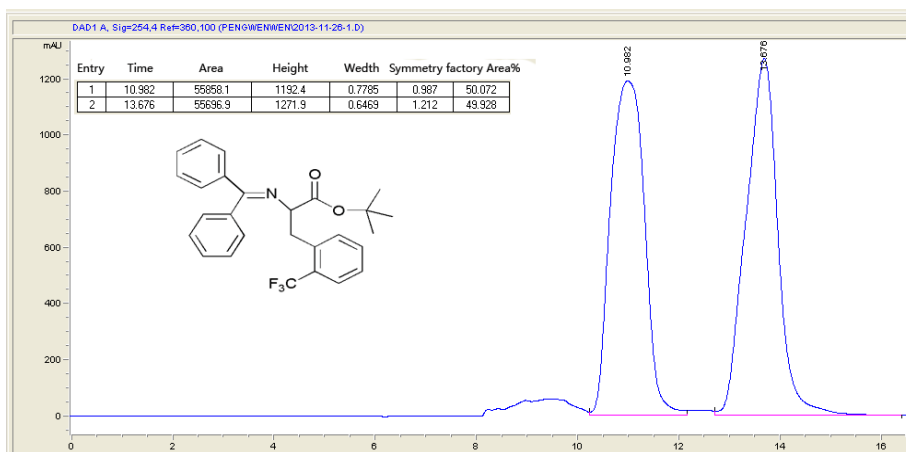
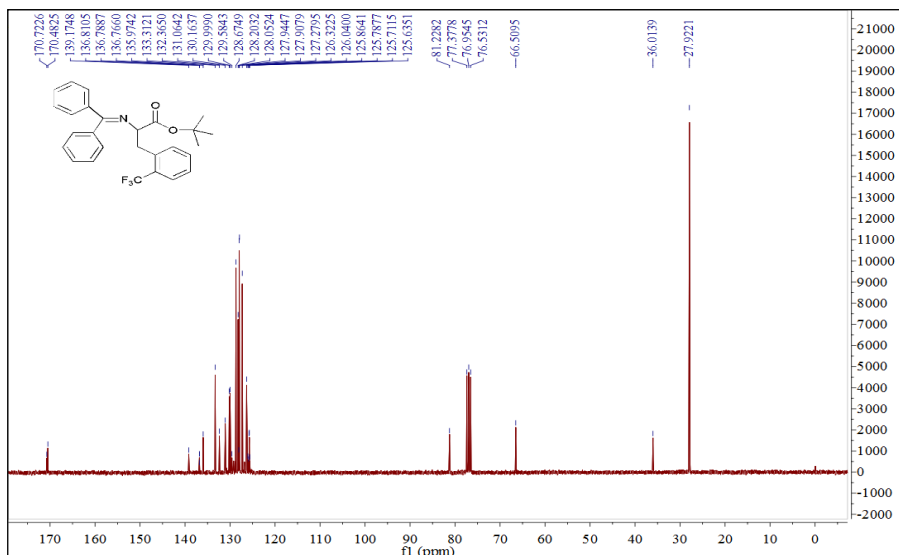




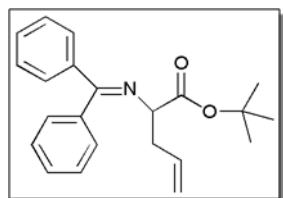
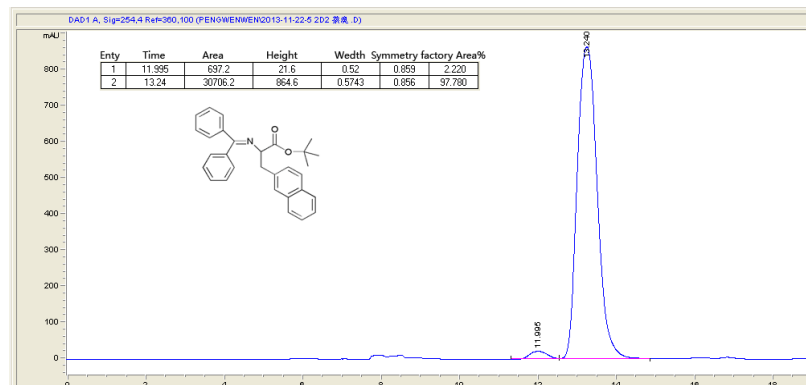
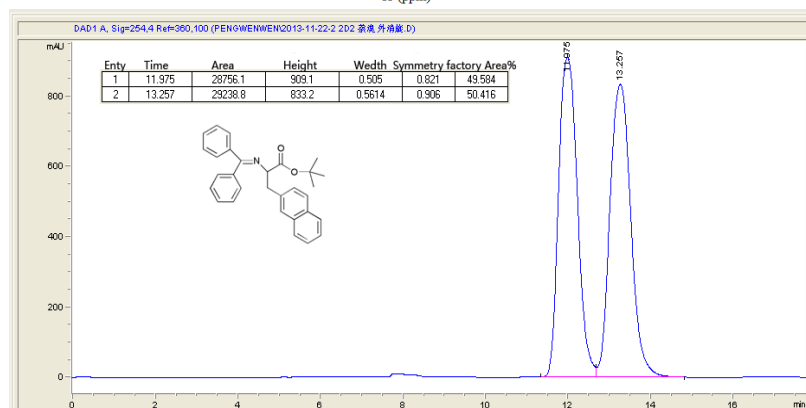
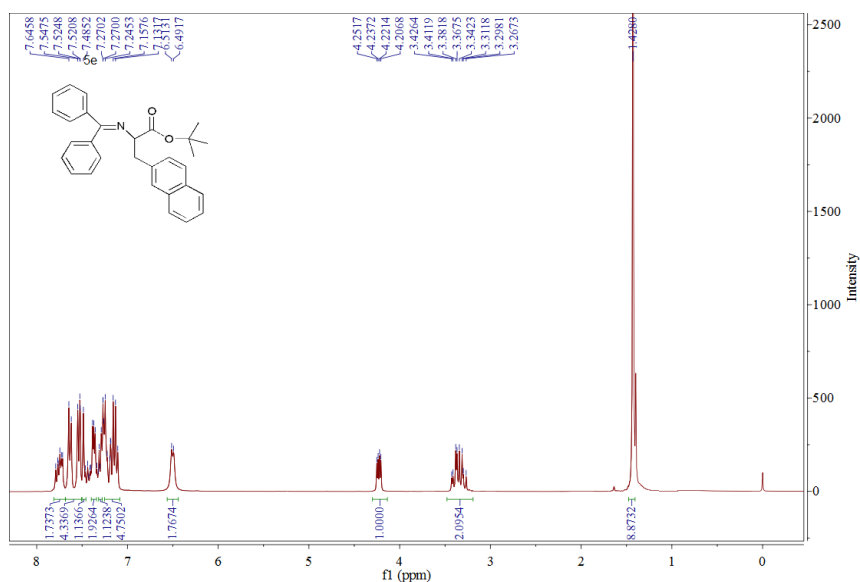
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.76 (d, 1 H, $^3J=7.1$ Hz, Ph-H), 7.57–7.41 (m, 4 H, Ph-H), 7.35–7.15 (m, 7 H, Ph-H), 6.43 (d, 2 H, $^3J=6.4$ Hz, Ph-H), 4.13 (dd, 1 H, $^3J=3.5$ Hz, 3.5 Hz, CH), 3.50–3.22 (m, 2 H, CH_2), 1.39 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.7, 170.5 (C=N, C=O), 139.2, 136.8 (q , $^2J_{\text{C-F}}=1.6$ Hz), 136.0, 133.3, 132.4, 131.1, 130.2, 131.0, 129.6, 128.7, 128.2, 128.1, 127.9, 127.9, 127.3, 126.3, 126.0 (Ph), 125.7

(q , $^1J_{\text{C-F}}=5.7$ Hz, CF_3), 81.2 (O-C), 66.5 (CH), 36.0 (CH_2), 27.9 (CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.8 min (*R*-isomer), 13.5min (*S*-isomer).

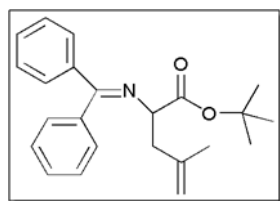
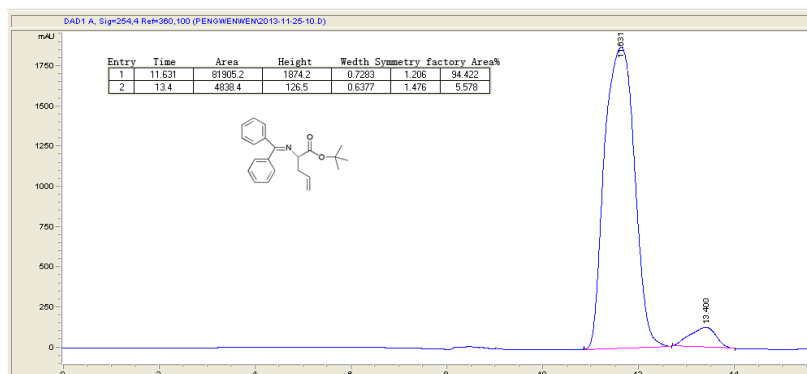
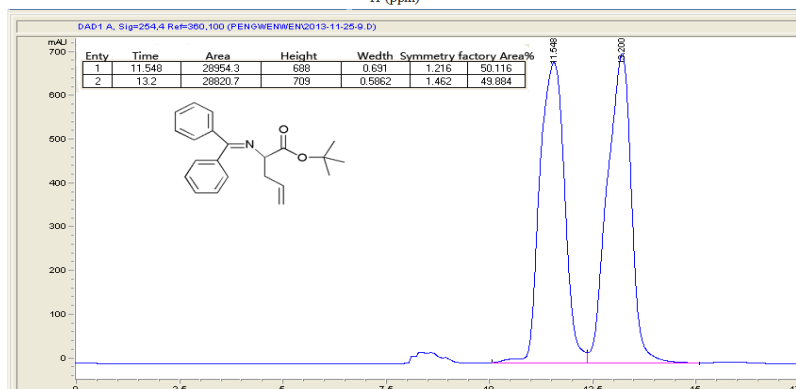
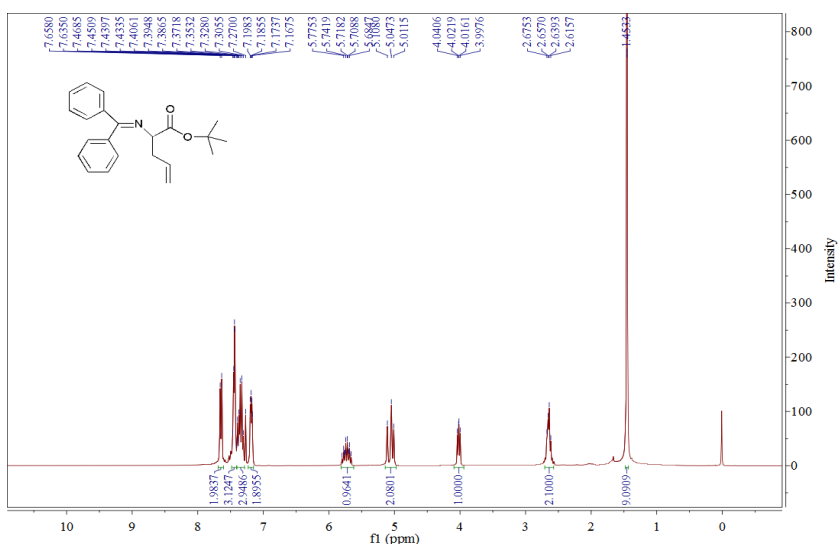




$^1\text{H NMR}$ (300 MHz, CDCl_3 , TMS): δ 7.79–7.11 (m, 15 H, Ar-H), 6.50 (d, 2 H, $^3J = 6.4$ Hz), 4.23 (dd, 1 H, $^3J = 4.4$ Hz, 4.4 Hz, CH), 3.43–3.27 (m, 2 H, CH_2), 1.43 (s, 9 H, CH_3). Rt HPLC (Daicel Chiralpak OD-H, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 12.0 min (*S*-isomer), 13.2 min (*R*-isomer).

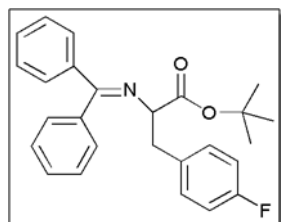
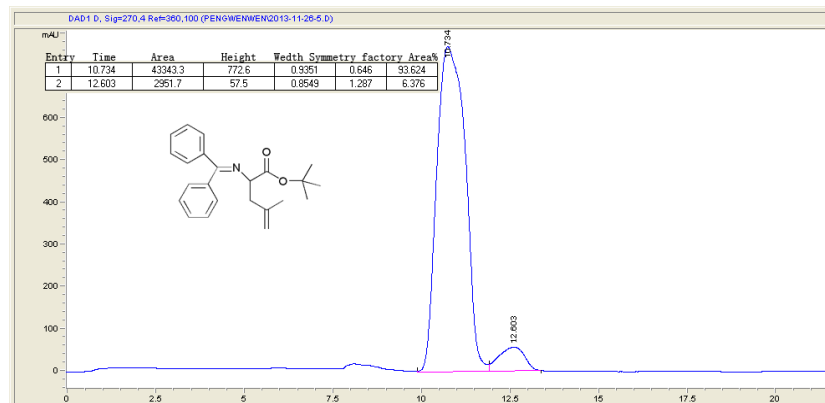
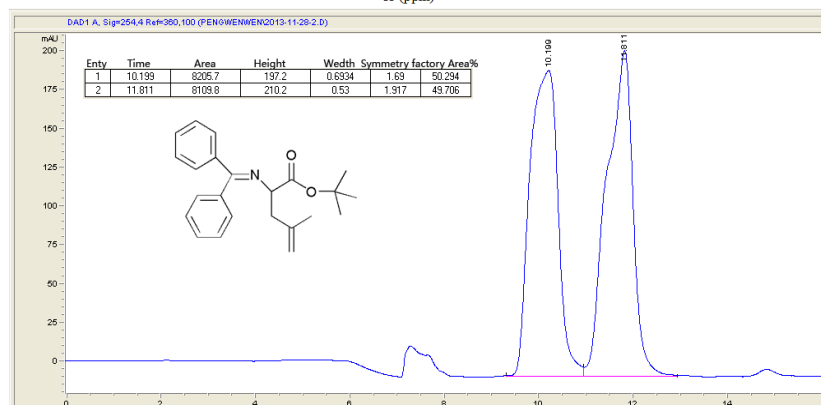
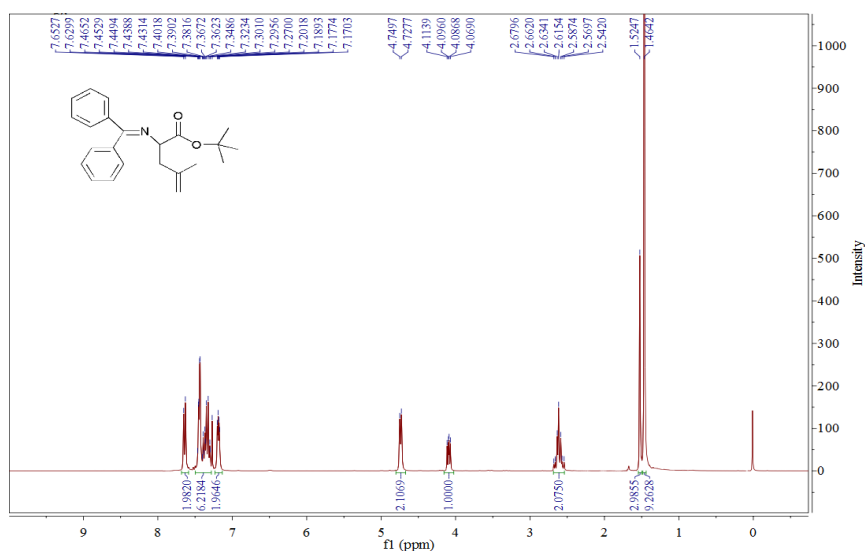


^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.64 (d, 2 H, $^3J = 6.9$ Hz, Ph-H), 7.47–7.43 (m, 3 H, Ph-H), 7.41–7.35 (m, 3 H, Ph-H), 7.20–7.17 (m, 2 H, Ph-H), 5.80–5.66 (m, 1 H, -CH=), 5.11–5.01 (m, 2 H, =CH₂), 4.02 (dd, 1 H, $^3J = 5.6$ Hz, 5.6 Hz, CH), 2.68–2.62 (m, 2 H, CH₂), 1.45 (s, 9 H, CH₃). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 11.6 min (*R*-isomer), 13.4 min (*S*-isomer).



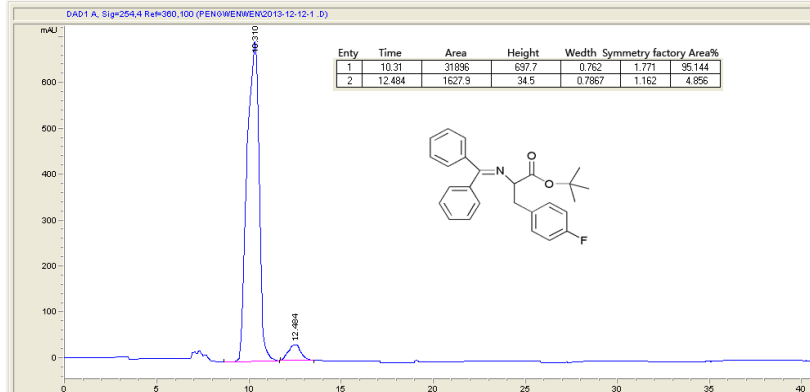
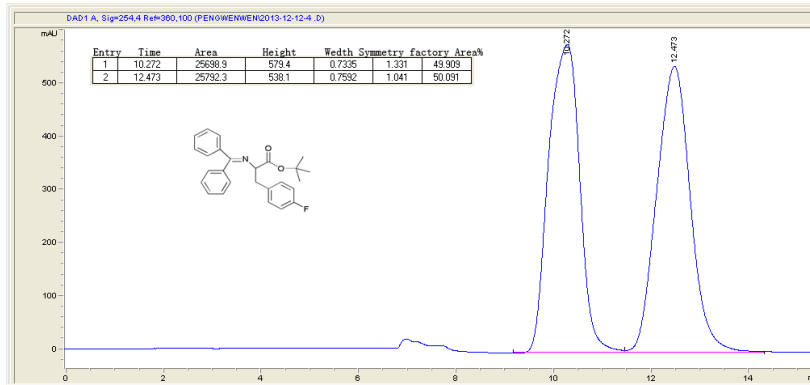
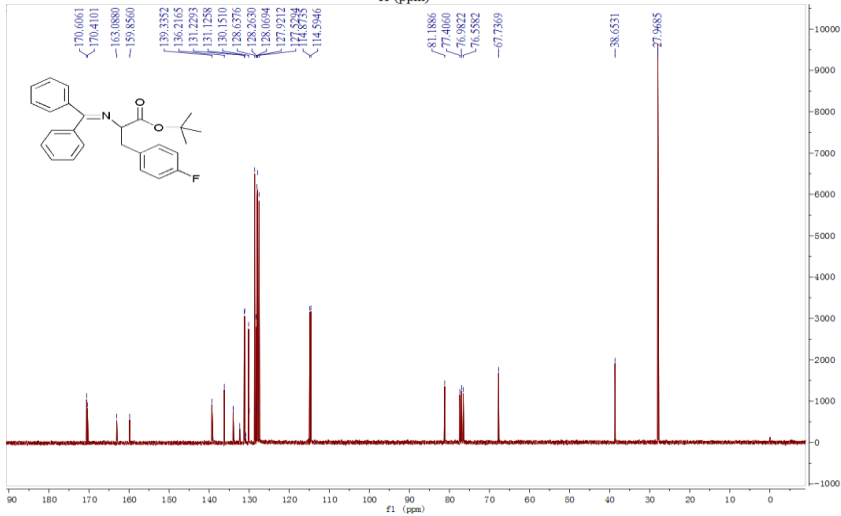
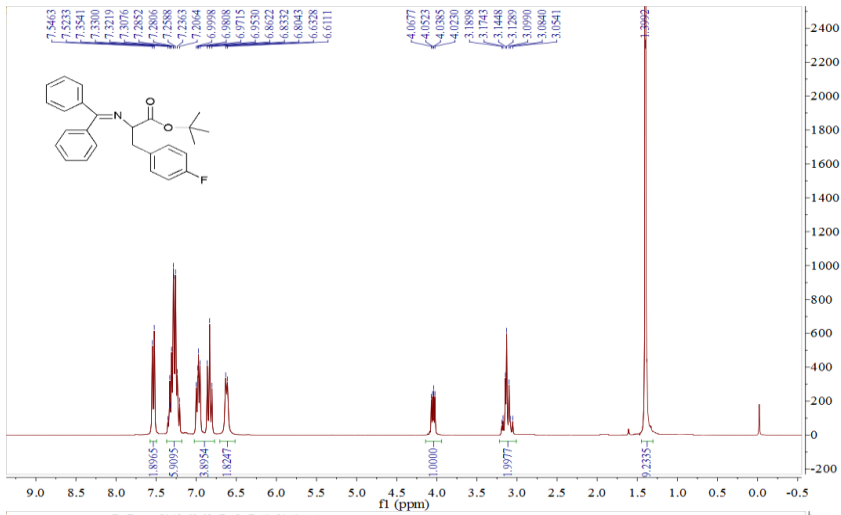
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.64 (d, 2 H, $^3J = 6.8$ Hz, Ph-H), 7.47–7.30 (m, 6 H, Ph-H), 7.20–7.17 (m, 2 H, Ph-H), 4.75 (d, 2 H, $^3J = 6.6$ Hz, $=\text{CH}_2$), 4.09 (dd, 1 H, $^3J = 5.4$ Hz, 5.3 Hz, CH), 2.68–2.54 (m, 2 H, CH_2), 1.52 (s, 3 H, CH_3), 1.46 (s, 9 H, CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.7 min

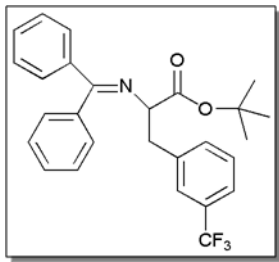
(*R*-isomer), 12.6 min (*S*-isomer).



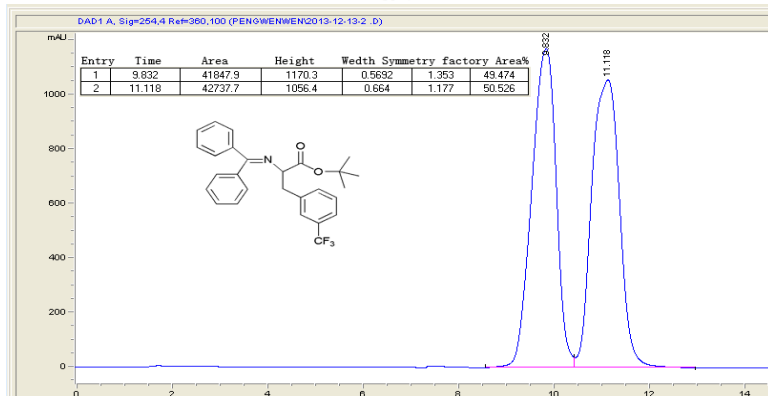
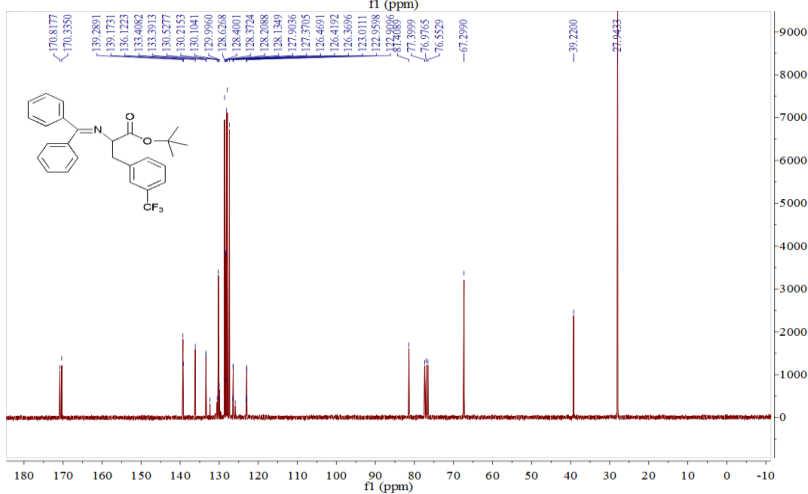
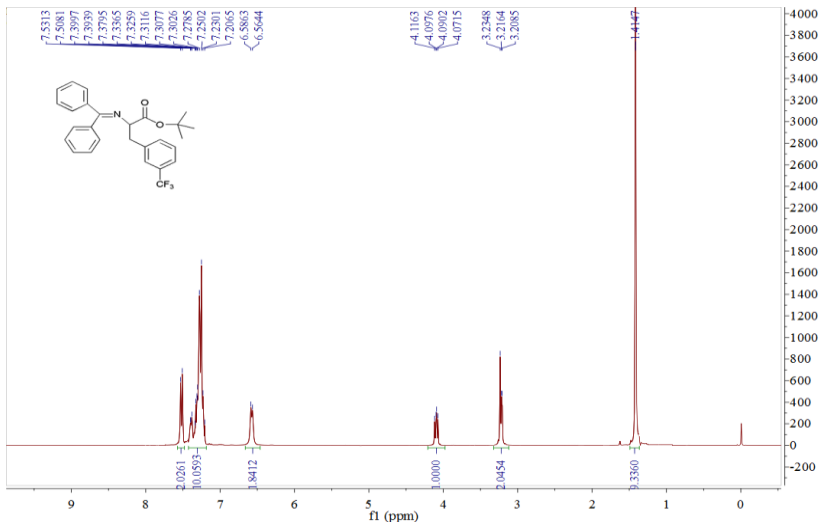
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.53 (d, 2 H, $^3J = 6.9$ Hz, Ph-H), 7.35–7.21 (m, 6 H, Ph-H), 7.00–6.80 (m, 4 H, Ph-H), 6.62 (d, 2 H, $^3J = 6.5$ Hz), 4.04 (dd, 1 H, $^3J = 4.6$ Hz, 4.7 Hz, CH), 3.19–3.05 (m, 2 H, CH_2), 1.40 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.6, 170.4 (C=N, C=O), 163.1, 160.0, 139.3, 134.0, 132.4, 131.2 (d, $^2J_{\text{C-F}} = 7.8$ Hz, F), 131.1, 130.9, 130.2, 130.0, 128.6, 128.3, 128.2, 128.1, 128.0, 127.5 (Ph), 114.7, (d, $^1J_{\text{C-F}} =$

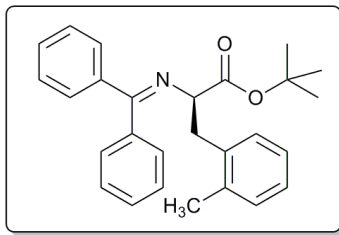
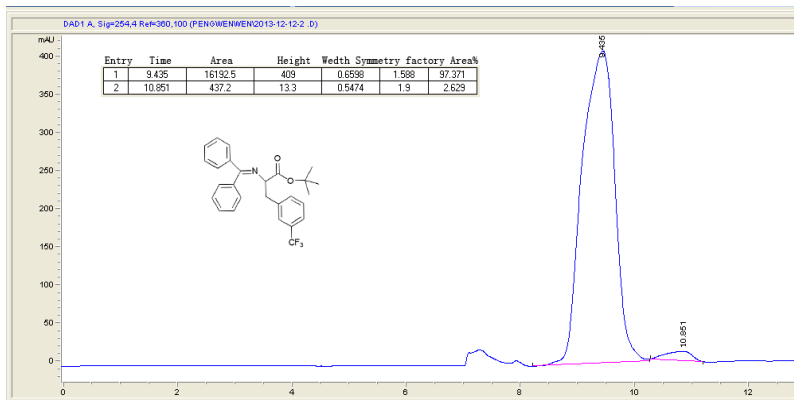
20.9 Hz, F), 81.2 (O-C), 67.7 (CH), 38.7 (CH_2), 28.0 (CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.3 min (*R*-isomer), 12.5 min (*S*-isomer).



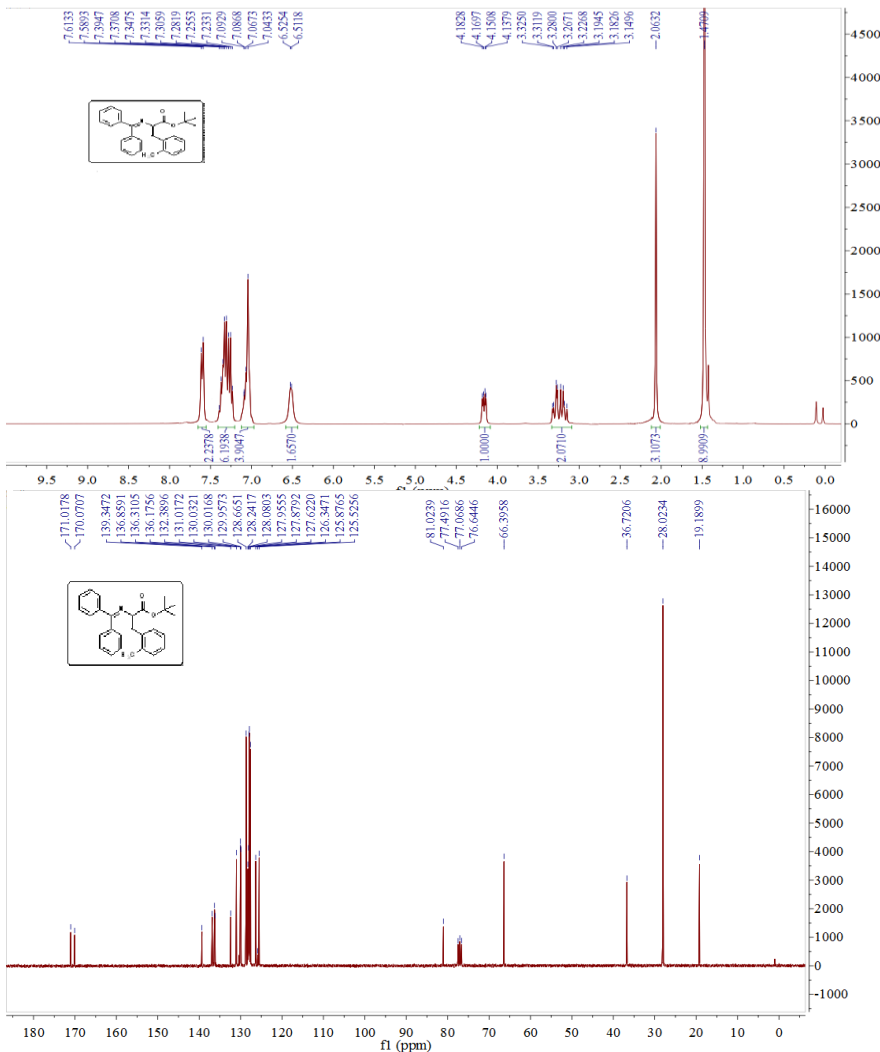


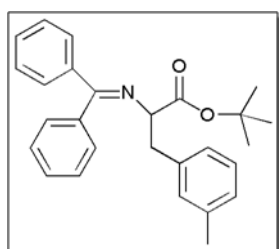
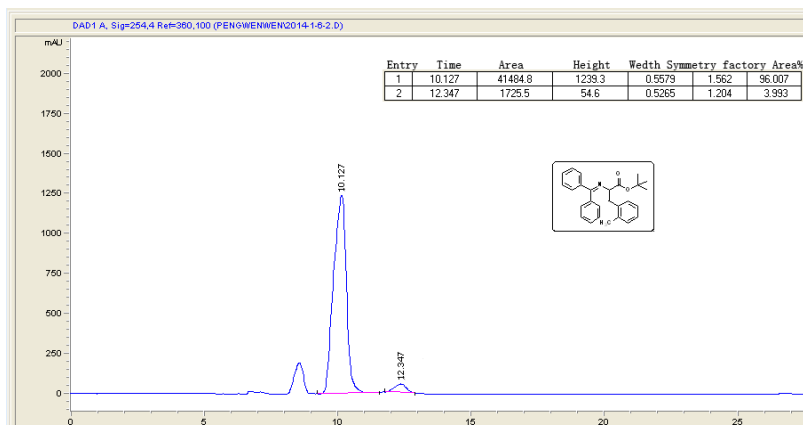
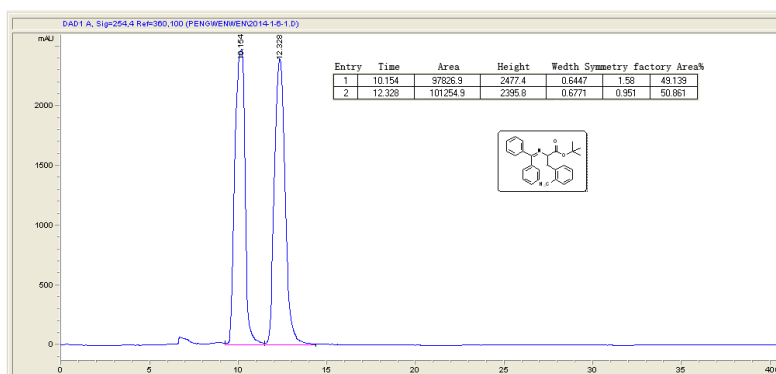
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.52 (d, 2 H, $^3J = 7.0$ Hz, Ph-H), 7.40–7.21 (m, 10 H, Ph-H), 6.57 (d, 2 H, $^3J = 6.6$ Hz, Ph-H), 4.09 (dd, 1 H, $^3J = 5.6$ Hz, 5.6 Hz, CH), 3.23–3.21 (m, 2 H, CH_2), 1.41 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.8, 170.3 (C=N, C=O), 139.3, 139.2, 136.1, 133.4, 133.4, 132.4, 130.5, 130.2, 130.1, 130.0, 128.6, 128.4, 128.3, 128.2, 128.1, 127.9, 127.4, 126.4 (q, $^2J_{\text{C-F}} = 3.7$ Hz, CF_3), 125.9 (Ph), 123.0 (q, $^1J_{\text{C-F}} = 3.8$ Hz, CF_3), 81.4 (O-C), 67.3 (CH), 39.2 (CH_2), 27.9 (CH_3). Rt HPLC (Daicel Chiralpak OD-H, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 9.4 min (*R*-isomer), 10.9 min (*S*-isomer).





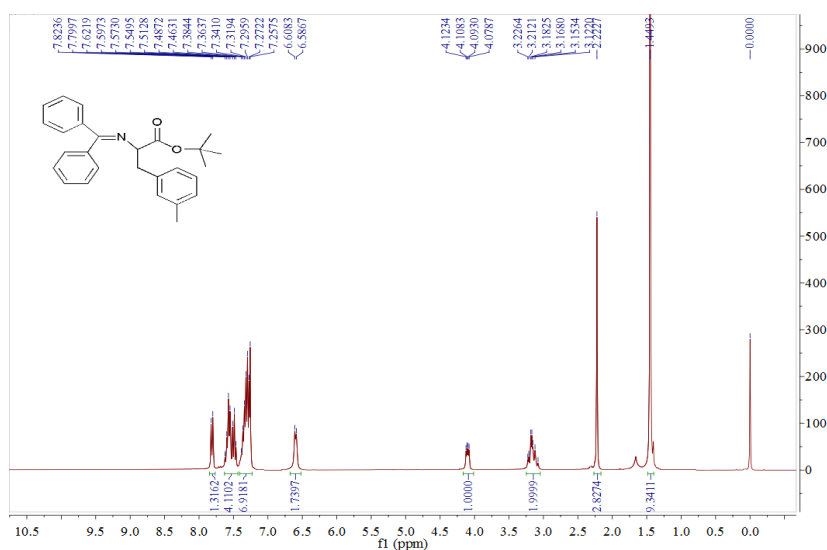
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.60 (d, 2 H, $^3J = 7.2$ Hz, Ph-H), 7.35–7.23 (m, 6 H, Ph-H), 7.09–7.04 (m, 4 H, Ph-H), 6.52 (d, 2 H, $^3J = 4.1$ Hz, Ph-H), 4.15 (dd, 1 H, $^3J = 3.9$ Hz, 3.9 Hz, CH), 3.33–3.15 (m, 2 H, CH_2), 2.06 (s, 3 H, Ph- CH_3), 1.39 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 171.0, 170.1 (C=N, C=O), 139.3, 136.9, 136.3, 136.2, 132.4, 131.0, 130.0, 130.0, 129.9, 128.7, 128.2, 128.1, 127.9, 127.8, 127.6, 126.3, 125.9, 125.5 (C-Ph), 81.0 (O-C), 66.4 (CH), 36.7 (CH_2), 28.0 (CH_3), 19.2 (Ph- CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.1 min (*R*-isomer), 12.3 min (*S*-isomer).

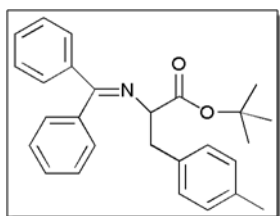
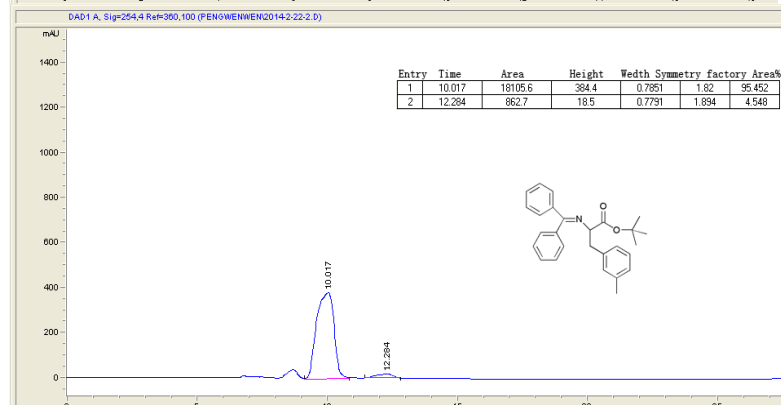
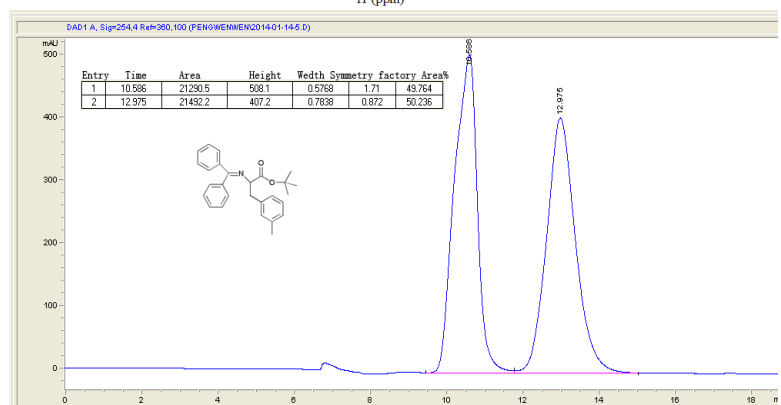
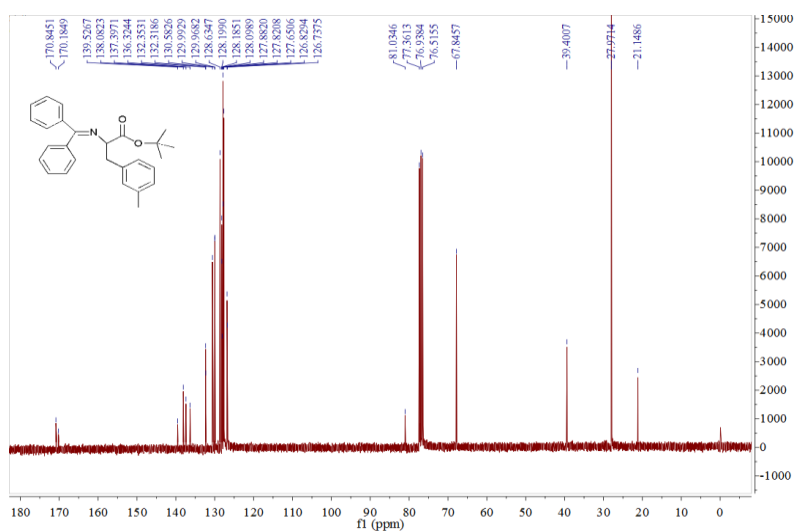




$^1\text{H NMR}$ (300 MHz, CDCl_3 , TMS): δ 7.81 (d, 1 H, $^3J = 7.2$ Hz, Ph-H), 7.62–7.46 (m, 4 H, Ph-H), 7.38–7.26 (m, 7 H, Ph-H), 6.59 (d, 2 H, $^3J = 6.5$ Hz, Ph-H), 4.09 (dd, 1 H, $^3J = 4.5$ Hz, 4.3 Hz, CH), 3.23–3.08 (m, 2 H, CH_2), 2.22 (s, 3 H, Ph- CH_3), 1.45 (s, 9 H, CH_3). $^{13}\text{C NMR}$ (75 MHz, CDCl_3 , TMS): δ 170.8, 170.2 (C=N, C=O), 139.5, 138.1, 137.4, 136.3, 132.4, 132.3, 130.6, 130.0, 130.0, 128.6, 128.2, 128.2, 128.1, 127.9, 127.8, 127.7, 126.8, 126.7 (C-Ph), 81.0 (O-C), 67.8 (CH), 39.4 (CH_2), 28.0 (CH_3), 21.1 (Ph- CH_3). Rt HPLC (Phenomenex Lux 5u

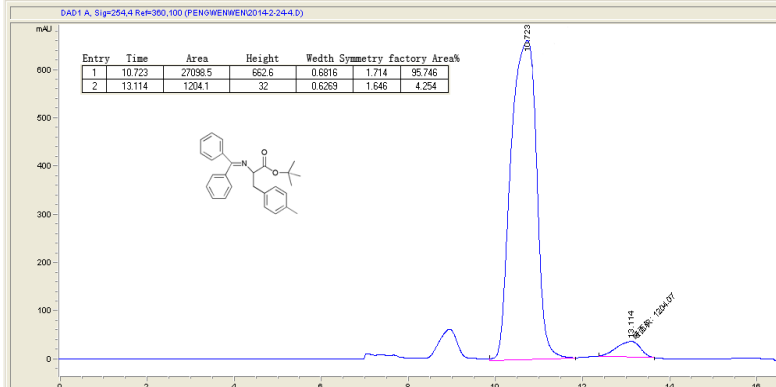
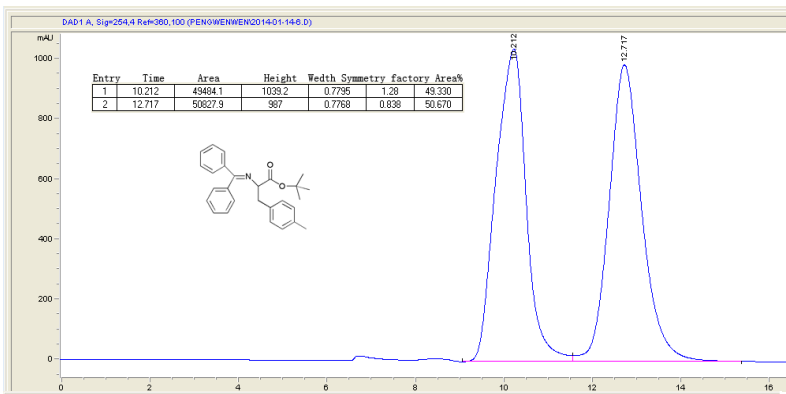
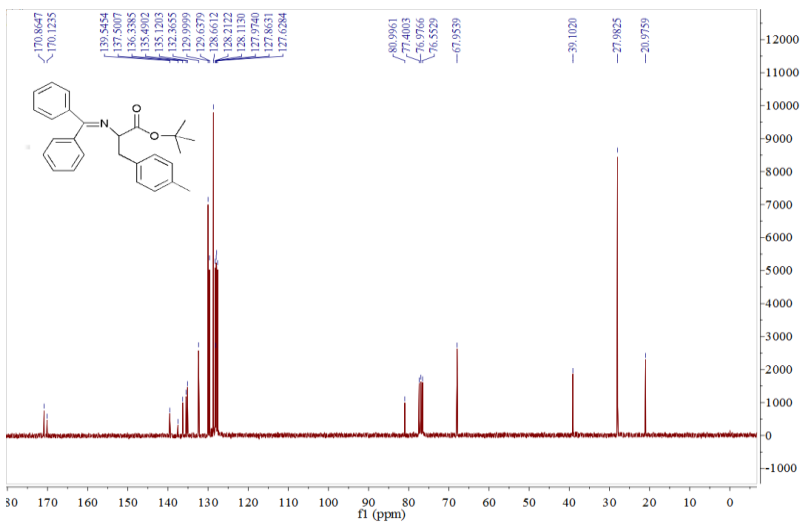
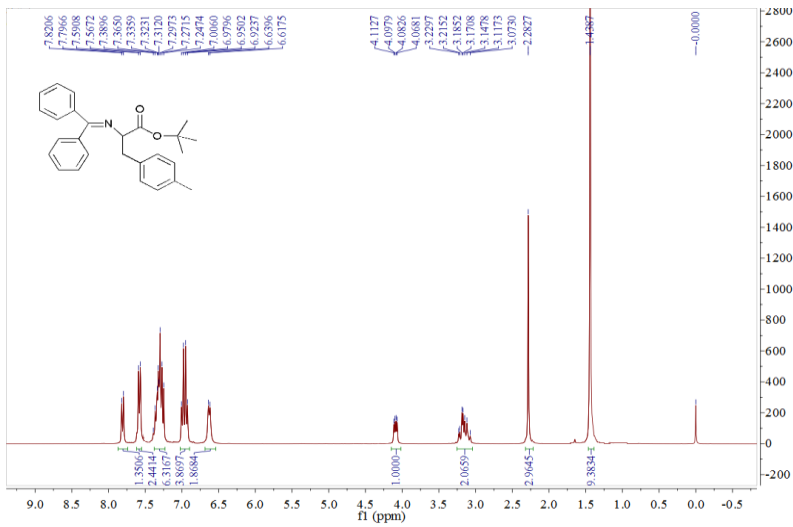
Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.0 min (*R*-isomer), 12.3 min (*S*-isomer).

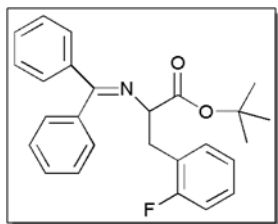




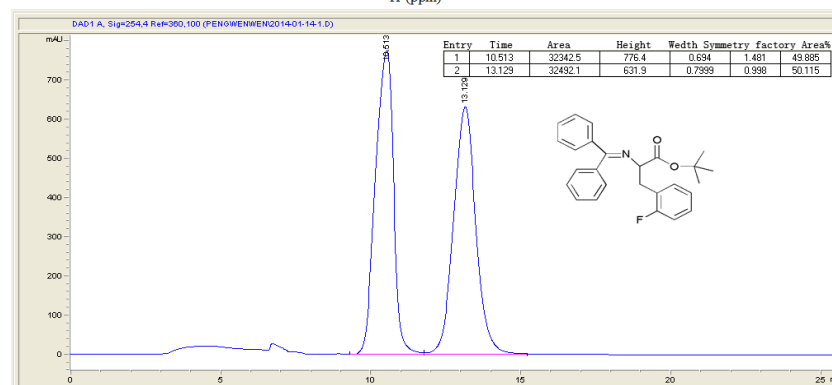
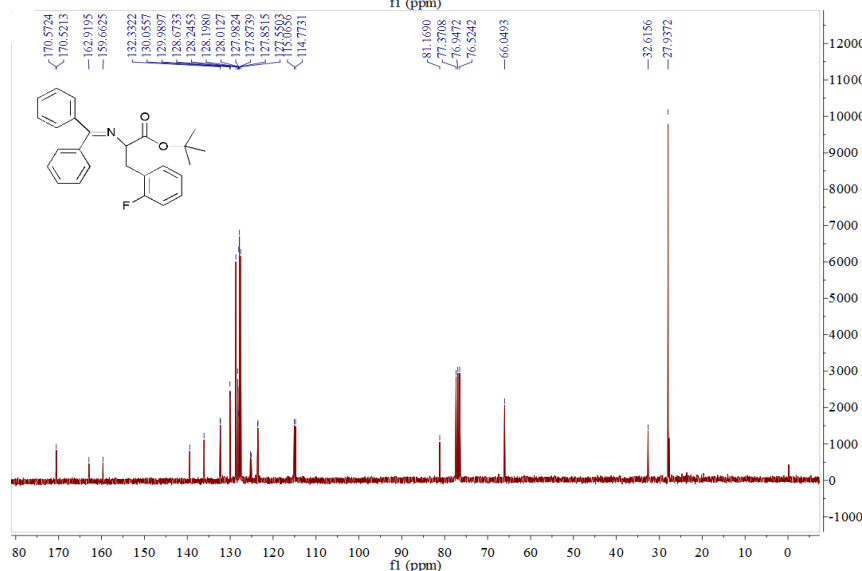
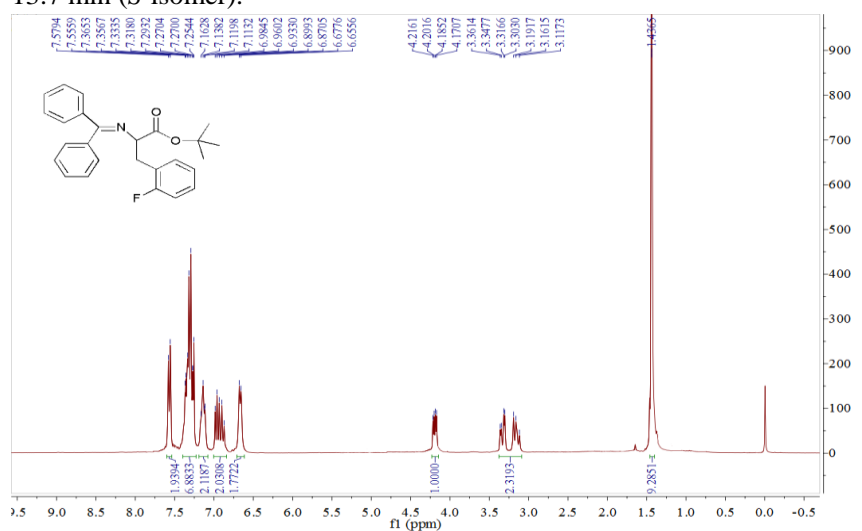
^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.81 (d, 1 H, $^3J = 7.2$ Hz, Ph-H), 7.58 (d, 2H, $^3J = 7.1$ Hz, Ph-H), 7.39–7.25 (m, 6 H, Ph-H), 6.96 (q, 4 H, $^3J = 7.9$ Hz, Ph-H), 6.62 (d, 2 H, $^3J = 6.6$ Hz, Ph-H), 4.09 (dd, 1 H, $^3J = 4.4$ Hz, 4.4 Hz, CH), 3.23–3.07 (m, 2 H, CH_2), 2.28 (s, 3 H, Ph- CH_3), 1.44 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.9, 170.1 (C=N, C=O), 139.5, 137.5, 136.3, 135.5, 135.1, 132.4, 130.0, 129.6, 128.7, 128.2, 128.1, 128.0, 127.9,

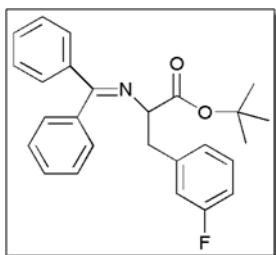
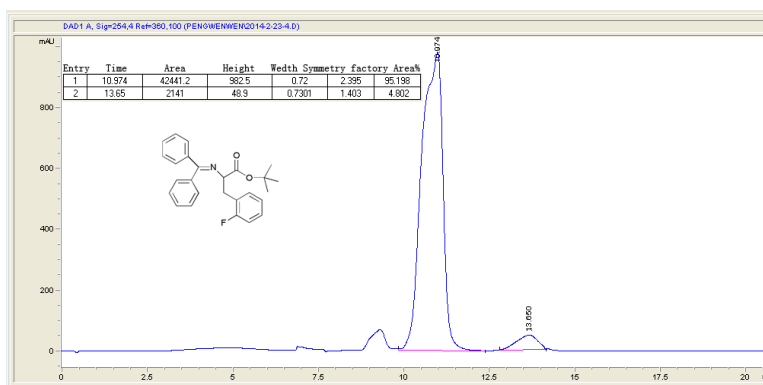
127.6 (C-Ph), 81.0 (O-C), 68.0 (CH), 39.1 (CH_2), 28.0 (CH_3), 21.0 (Ph- CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.7 min (*R*-isomer), 13.1 min (*S*-isomer).





^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.56 (d, 2 H, $^3J = 7.1$ Hz, Ph-H), 7.37–7.25 (m, 6 H, Ph-H), 7.16–7.11 (m, 2 H, Ph-H), 6.98–6.87 (m, 2 H, Ph-H), 6.66 (d, 2 H, $^3J = 6.6$ Hz, Ph-H), 4.19 (dd, 1 H, $^3J = 4.4$ Hz, 4.4 Hz, CH), 3.36–3.12 (m, 2 H, CH_2), 1.44 (s, 9 H, CH_3). ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.6, 170.5 (C=N, C=O), 162.9, 159.7, 139.4, 136.1, 132.3, 130.1, 130.0, 128.7, 128.2, 128.2, 128.0, 128.0, 127.9, 127.9, 127.6 (C-Ph), 125.2 (d, $^3J_{\text{C-F}} = 15.5$ Hz, F), 123.5 (d, $^2J_{\text{C-F}} = 3.5$ Hz, F), 114.9 (d, $^1J_{\text{C-F}} = 21.9$ Hz, F), 81.2 (O-C), 66.0 (CH), 32.6 (CH_2), 27.9 (CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 11.0 min (*R*-isomer), 13.7 min (*S*-isomer).

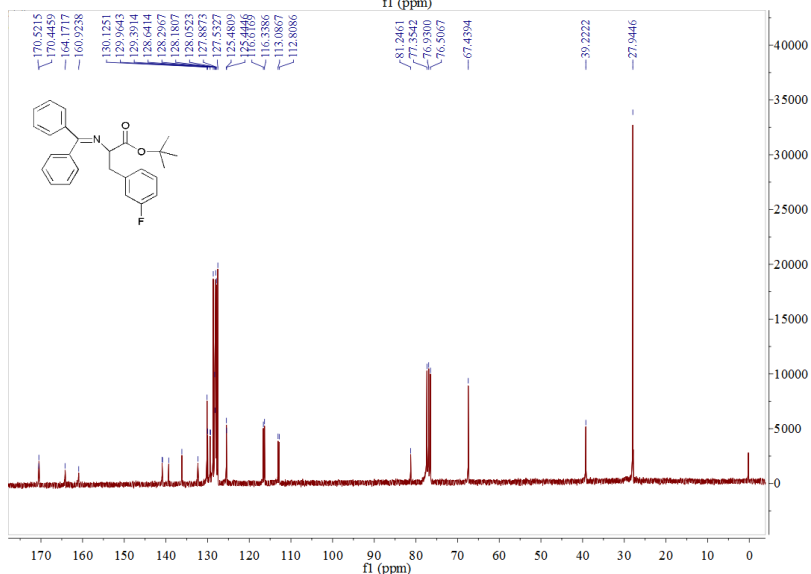
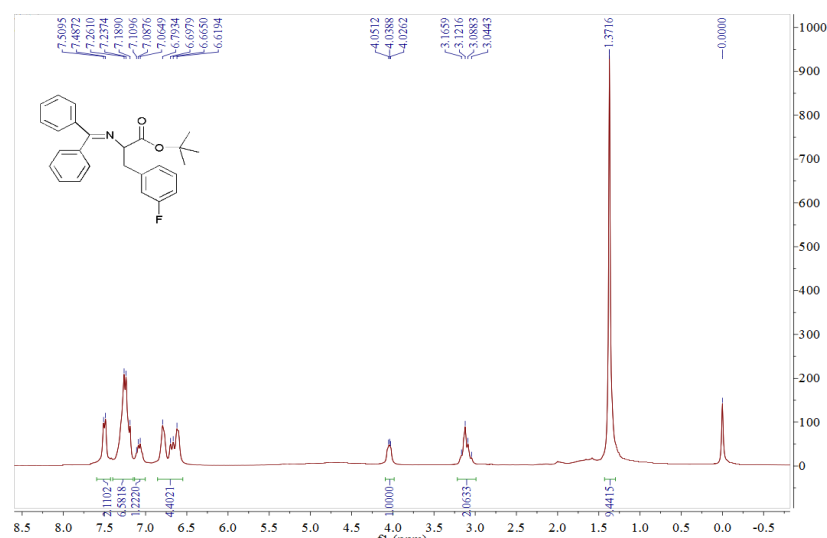


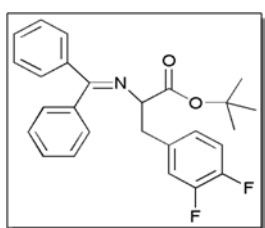
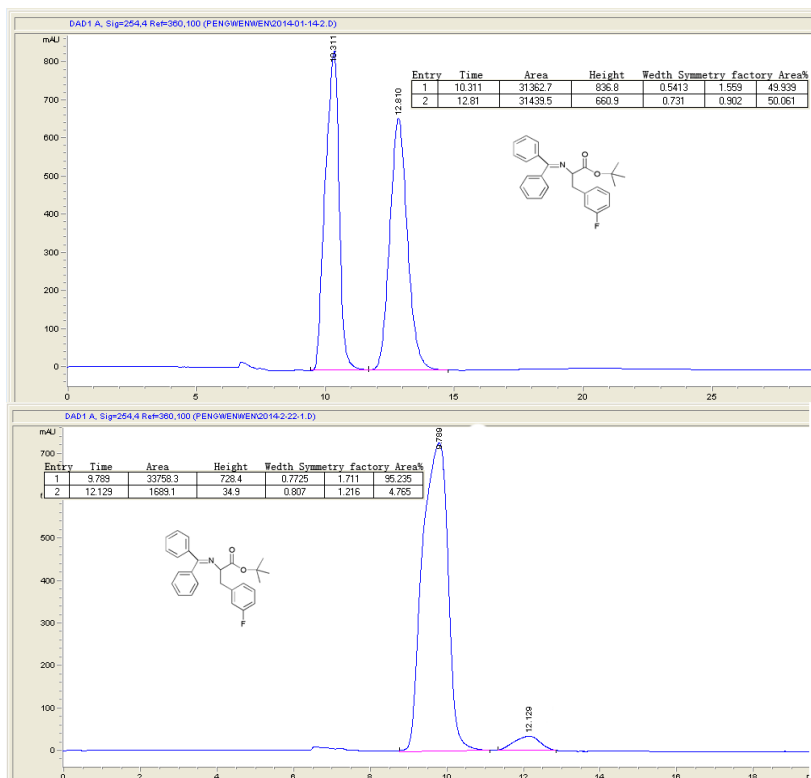


^1H NMR (300 MHz, CDCl_3 , TMS): δ 7.49 (d, 2 H, $^3J = 6.7$ Hz, Ph-H), 7.26–7.19 (m, 7 H, Ph-H), 7.08 (q, 1 H, $^3J = 6.6$ Hz, Ph-H), 6.79–6.62 (m, 4 H, Ph-H), 6.66 (d, 2 H, $^3J = 6.6$ Hz, Ph-H), 4.04 (dd, 1 H, $^3J = 3.8$ Hz, 3.8 Hz, CH), 3.17–3.04 (m, 2 H, CH_2), 1.37 (s, 9 H, CH_3).

^{13}C NMR (75 MHz, CDCl_3 , TMS): δ 170.5, 170.4 (C=N, C=O), 164.2, 160.9, 140.8 (d, $^3J_{\text{C-F}} = 7.4$ Hz, F), 139.3, 136.2, 132.3, 130.1, 130.0, 128.6, 128.3, 128.2, 128.1, 127.9, 127.5, 125.5, 125.4 (C-Ph), 116.4 (d, $^2J_{\text{C-F}} = 20.9$ Hz, F), 112.9 (d, $^1J_{\text{C-F}} = 20.9$ Hz, F), 81.2 (O-C),

67.4 (CH), 39.2 (CH_2), 27.9 (CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 9.8 min (*R*-isomer), 12.1 min (*S*-isomer).





$^1\text{H NMR}$ (300 MHz, CDCl_3 , TMS): δ 7.57 (d, 2 H, $^3J = 7.0$ Hz, Ph-H), 7.40–7.25 (m, 6 H, Ph-H), 7.02–6.73 (m, 5 H, Ph-H), 4.10 (dd, 1 H, $^3J = 4.7$ Hz, 4.7 Hz, CH), 3.21–3.07 (m, 2 H, CH_2), 1.44 (s, 9 H, CH_3). $^{13}\text{C NMR}$ (75 MHz, CDCl_3 , TMS): δ 170.7, 170.3 (C=N, C=O), 148.1 (d, $^3J_{\text{C-F}} = 12.5$ Hz, F), 147.3 (d, $^3J_{\text{C-F}} = 12.5$ Hz, F), 139.2, 136.1, 135.3 (dd, $^1J_{\text{C-F}} = 5.7$ Hz, F, $^2J_{\text{C-F}} = 3.9$ Hz, F), 132.3, 130.3, 130.0, 128.6, 128.4, 128.2, 128.1, 127.9, 127.5, 125.6 (dd, $^1J_{\text{C-F}} = 6.0$ Hz, F, $^2J_{\text{C-F}} = 3.6$ Hz, F), 118.4 (d, $^2J_{\text{C-F}} = 16.8$ Hz, F), 116.6 (d, $^2J_{\text{C-F}} = 16.8$ Hz, F), 114.4 (C-Ph), 81.4 (O-C), 67.3 (CH), 38.7 (CH_2), 27.9 (CH_3). Rt HPLC (Phenomenex Lux 5u Amylose-2, 95:5, hexane/dioxane, 254 nm, 0.5 ml/min), 10.3 min (*R*-isomer), 12.3 min (*S*-isomer).

