## **Electronic Supplementary Information for**

# Improved Analysis of Excited State Proton Transfer Kinetics by the Combination of Standard and Convolution Methods

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#### ESPT of 7-hydroxy-4-methylflavylium in water: 2 species

Given the reversible reactions represented by Scheme 1S:

#### Scheme 1S.



the time evolution of the species can be represented by the following differential equation:

$$\frac{d}{dt} \begin{bmatrix} AH^{+} * \\ A^{*} \end{bmatrix} = \begin{bmatrix} -X_{W} & k_{diff}[H^{+}] \\ k_{d} & -Y_{W} \end{bmatrix} \times \begin{bmatrix} AH^{+} * \\ A^{*} \end{bmatrix}$$
(S1)

where  $X_W = k_{AH^+} + k_d$  and  $Y_W = k_A + k_{diff}[H^+]$ . The initial concentrations of AH<sup>+\*</sup> and A<sup>\*</sup> are [AH<sup>+\*</sup>]<sub>0</sub> and 0, respectively, and the time-dependent concentrations,  $C_i(t)$ , of the species in the excited state can be written in integrated form as  $C_i(t) = P_i \otimes C_{i\delta}$  based on the concept of convolution (Reference 4 of the paper), where  $P_i$  is the rate of production of the species  $C_i$  and  $C_{i\delta}$  is the respective delta response function. For the flavylium cation and the quinonoidal base the respective rates of production and delta responses are:

$$P_{AH^{+*}} = [AH^{+*}]_0 \delta(t) + k_{diff} [H^{+}] [A^{*}]$$
(S2)

$$P_{A^*} = k_d [AH^{+*}] \tag{S3}$$

$$C_{AH^{*}*\delta} = [AH^{*}*]\delta(t) = e^{-X_{W}t}$$
(S4)

$$C_{A^*\delta} = [A^*]\delta(t) = \mathrm{e}^{-Y_W t}$$
(S5)

Thus, the temporal evolutions of the concentrations of AH<sup>+</sup>\* and A\* are:

$$[AH^{**}] = [AH^{**}]_0 e^{-X_W t} + k_{diff} [H^{*}] [A^{*}] \otimes e^{-X_W t}$$
(S6)

$$[A^*] = k_d [AH^{+*}] \otimes e^{-Y_W t}$$
(S7)

Equations S6 and S7 can be solved by Laplace transformation theory,  $\mathcal{L}{f(t)} = \int_{0}^{\infty} e^{-st} f(t) dt$ , and the

solution for reversible first order reactions is described in reference 4. Here we are interested only in the temporal evolution of [A\*] and do not need to solve equation S6 since  $[AH^{+*}]$  will serve as the effective Instrument Response Function (in practical terms,  $k_d[AH^{+*}]$  will operate as if it were a delta function). From the algebraic properties of convolution, the convolution of a delta function with some other function results in the function itself at the position of the delta function. Thus, the time evolution of the quinonoidal base using the signal of  $AH^{+*}$  as the IRF is:

$$[A^*] = \mathrm{e}^{-Y_W t} \tag{S8}$$

and the monoexponential lifetime obtained gives the two rate constants for the deactivation of the excited state of A\*, as experimentally observed for the ESPT of HMF in water.

#### ESPT of 7-hydroxy-4-methylflavylium in water-dioxane mixtures and functionalized flavylium

#### salts anchored in SDS micelles: 3 observable species involved in ESPT

Given Scheme 2S below:

#### Scheme 2S.

$$\begin{array}{c|c} AH^{+*} & \stackrel{k_{d}}{\longleftarrow} & (A^{*} \dots H^{+}) & \stackrel{k_{diss}}{\longleftarrow} & A^{*} + H^{+} \\ I_{0} & \downarrow \\ K_{AH^{+}} & \downarrow \\ AH^{+} & A & A \end{array}$$

the corresponding kinetics obey the differential equation,

$$\frac{d}{dt} \begin{bmatrix} AH^{+} * \\ (A^{*} \cdots H^{+}) \\ A^{*} \end{bmatrix} = \begin{bmatrix} -X & k_{rec} & 0 \\ k_{d} & -Y & k_{diff} [H^{+}] \\ 0 & k_{diss} & -Z \end{bmatrix} \times \begin{bmatrix} AH^{+} * \\ (A^{*} \cdots H^{+}) \\ A^{*} \end{bmatrix}$$
(S9)

where  $X = k_{AH^+} + k_d$ ,  $Y = k_A + k_{rec} + k_{diss}$  and  $Z = k_A + k_{diff}[H^+]$  represent the sums of the rate constants for all the decay processes of AH<sup>+</sup>\* and (A\*...H<sup>+</sup>), respectively. Since all of the processes indicated in Scheme 2S are first (or pseudo-first) order, it is also possible in this case to write the time-dependent concentration  $C_i(t)$  of the species directly in the integrated form, as done in the previous case. At pHs below 2, only the flavylium cation is present in solution and the initial concentrations of [AH<sup>+</sup>\*], (A\*...H<sup>+</sup>) and [A\*] are [AH<sup>+</sup>\*]\_0, 0 and 0, respectively. Thus, the Delta responses  $C_{i\delta}$  can be written as:

$$[AH^{*}]\delta(t) = e^{-Xt}$$
(S10)

$$[A^* \cdots H^+]\delta(t) = e^{-Yt}$$
(S11)

$$[A^*]\delta(t) = e^{-Zt}$$
(S12)

Because the initial concentrations  $[A^* \cdots H^+]_0$  and  $[A^*]_0$  are zero, the production rates  $P_i$  are:

$$P_{AH^{+*}} = [AH^{+*}]_{0} \delta(t) + k_{rec} [A^{*} \cdots H^{+}]$$
(S13)  

$$P_{A^{*} \cdots H^{+}} = [A^{*} \cdots H^{+}]_{0} \delta(t) + k_{d} [AH^{+*}] + k_{diff} [H^{+}] [A^{*}]$$
(S14)  

$$P_{A^{*} \cdots H^{+}} = k_{d} [AH^{+*}] + k_{diff} [H^{+}] [A^{*}]$$
(S14)  

$$P_{A^{*}} = [A^{*}]_{0} \delta(t) + k_{diss} [A^{*} \cdots H^{+}]$$
(S15)

Substituting the Delta responses and production rates of the species in the equation  $C_i(t) = P_i \otimes C_{i\delta}$ , the concentrations  $C_i(t)$  are given by:

$$[AH^{+*}] = [AH^{+*}]_{0} \delta(t) \otimes e^{-Xt} + k_{rec} [A^{*} \cdots H^{+}] \otimes e^{-Xt}$$

$$[AH^{+*}] = [AH^{+*}]_{0} e^{-Xt} + k_{rec} [A^{*} \cdots H^{+}] \otimes e^{-Xt}$$

$$[A^{*} \cdots H^{+}] = k_{d} [AH^{+*}] \otimes e^{-Yt} + k_{diff} [H^{+}] [A^{*}] \otimes e^{-Yt}$$
(S16)
(S17)

$$[A^*] = k_{diss}[A^* \cdots H^+] \otimes e^{-Zt}$$
(S18)

Thus, taking the decay of  $AH^{+*}$  measured at 480 nm as the effective IRF that generates the decay at 620 nm, the overall product [B\*] will be the sum of the concentrations of the geminate pair and the quinonoidal base, i.e.,  $[B^*] = [A^* \cdots H^+] + [A^*]$ . From the properties of Laplace transforms,  $\mathcal{L}{f \otimes g} = \mathcal{L}{f} \cdot \mathcal{L}{g}$  and from Laplace transform tables  $\mathcal{L}{e^{-at}} = 1/(s+a)$ . The Laplace transforms of S17 and S18 are, respectively:

$$\overline{[A^*\cdots H^+]} = \frac{k_d \overline{[AH^{+*}]}}{(s+Y)} + \frac{k_p \overline{[H^+]} \overline{[A^*]}}{(s+Y)}$$
(S19)

$$\overline{[A^*]} = \frac{k_{diss}[A^* \cdots H^+]}{(s+Z)}$$
(S20)

In this case, the temporal evolutions of  $[A^* \cdots H^+]$  and  $[A^*]$  are not independent and equations S19 and

S20 are coupled. The solution of S19 and S20 gives:

$$\overline{[A^* \cdots H^+]} = \frac{k_d [AH^{+*}](s+Z)}{(s+Y)(s+Z) - k_{diss} k_{diff} [H^+]}$$
(S21)

$$\overline{[A^*]} = \frac{k_{diss}k_d[\overline{AH^{+*}}]}{(s+Y)(s+Z) - k_{diss}k_{diff}[H^+]}$$
(S22)

Reorganizing the denominator in both equations leads to

$$\overline{[A^*\cdots H^+]} = \frac{k_d \overline{[AH^{+*}]}(s+Z)}{(s+\beta)(s+\gamma)}$$
(S23)

$$\overline{[A^*]} = \frac{k_{diss}k_d[\overline{AH^{**}}]}{(s+\beta)(s+\gamma)}$$
(S24)

where 
$$\beta = \frac{Y + Z - \sqrt{(Y - Z)^2 + 4k_{diss}k_{diff}[H^+]}}{2}$$
 and  $\gamma = \frac{Y + Z + \sqrt{(Y - Z)^2 + 4k_{diss}k_{diff}[H^+]}}{2}$ 

On the other hand, the inverse Laplace transform  $\mathcal{L}^{-1}\left\{\frac{s+\alpha}{(s+a)(s+b)}\right\}$  is  $\frac{(\alpha-a)}{b-a}e^{-at}-\frac{(\alpha-b)}{b-a}e^{-bt}$  and

the inverse of  $\mathcal{L}^{-1}\left\{\frac{1}{(s+a)(s+b)}\right\}$  is  $\frac{1}{b-a}e^{-at}-\frac{1}{b-a}e^{-bt}$ . With these pieces of information, the

solutions of S23 and S24 are:

$$[A^* \cdots H^+] = k_d [AH^{+*}] \otimes \left\{ \frac{1}{\gamma - \beta} \Big[ (Z - \beta)e^{-\beta t} - (Z + \gamma)e^{-\gamma t} \Big] \right\}$$
(S25)

$$[A^*] = k_d [AH^{+*}] \otimes \left[ \frac{k_{diss}}{\gamma - \beta} \left( e^{-\beta t} - e^{-\gamma t} \right) \right]$$
(S26)

$$[B^*] = [A^* \cdots H^+] + [A^*]$$
(S27)

$$[B^*] = k_d [AH^{+*}] \otimes \left\{ \frac{1}{\gamma - \beta} \Big[ (Z - \beta + k_{diss}) e^{-\beta t} + (\gamma - Z - k_{diss}) e^{-\gamma t} \Big] \right\}$$
(S28)

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where the term  $k_d[AH^{+*}]$  is the effective IRF. Equation S28 predicts a double-exponential decay with  $\tau_1 = 1/\beta$  and  $\tau_2 = 1/\gamma$  for a system of two reactions coupled in series, in agreement with the experimental results for HMF in aqueous-organic solvent mixtures and for functionalized flavylium salts anchored in SDS micelles.

#### Determining the individual rate constants

The differential Equation S9 (equivalent to equation 5 of the paper) predicts triple-exponential decays for all species,  $X_i^* = AH^+*$ ,  $(A^* \cdots H^+)$  and  $A^*$ :

$$\left[X_{i}^{*}\right] = a_{i,1}e^{-\lambda_{1}t} + a_{i,2}e^{-\lambda_{2}t} + a_{i,3}e^{-\lambda_{3}t}$$
(S29)

The reciprocals of the three decay times,  $\lambda_j = 1/\tau_j$ , are the eigenvalues of the constant matrix k, which can be expressed as a function of the diagonal reciprocal decay time matrix  $\lambda$ , and the pre-exponential coefficient ( $a_{i,j}$ ) matrix a, as:<sup>17,81</sup>

$$k = a\lambda a^{-1} \tag{S30}$$

However, Equation S30 cannot be solved analytically in the present case because the pre-exponential coefficients ( $a_{i,j}$ ) of the "matrix of concentrations" *a* cannot be equated to the experimental coefficients of the "matrix of fluorescence intensities" *A*,<sup>17</sup> because there are 3 kinetically discernible species but only 2 are spectroscopically observable. Thus, the matrix *A* is a 2 × 3 matrix with  $A_{2,j} = a_{2,j} + a_{3,j}$  (assuming that the radiative rate constants of the free base A\* and the geminate pair A\*...H<sup>+</sup> are equal). However, the analysis can be substantially improved by independently measuring the lifetime of the base ( $1/k_A = 1.46 \times 10^{-10}$  s) in solutions with pH > 9 and the lifetime of the acid in the absence of the excited state proton transfer process ( $1/k_{AH+} = 3.13 \times 10^{-9}$  s), taken to be equal to the lifetime of the parent compound 7-methoxy-4-methyl-flavylium ion (MMF). Since the 9 pieces of experimental information (3 decay times, 3 pre-exponential coefficients at 480 nm and 3 pre-exponential coefficients at 620 nm, listed in Table 1S) are sufficient to evaluate the remaining 4 unknowns, *k* can be estimated by minimization of the deviations between the calculated and experimental decay times and pre-exponential coefficients.<sup>17,9</sup>

The analysis of the results for the water:dioxane mixture with  $x_{Dx} = 0.18$ , based on Scheme 2S (equivalent to Scheme 3 of the paper), provides the individual values of  $k_d$ ,  $k_{rec}$ ,  $k_{diss}$  and  $k_{diff}$  (shown in Figure 1S) and separates the rate constants involved in the proton transfer processes (deprotonation and recombination) from the diffusional ones (pair dissociation and diffusion-controlled encounter).



**Figure 1S.** (a) Deprotonation, recombination and dissociation rate constants ( $k_d$ ,  $k_{rec}$ ,  $k_{diss}$ ) as a function of [H<sup>+</sup>]; (b) rate constants for diffusion-controlled formation of the geminate pair ( $k_{diff}a_{H^+}$ ) in water-1,4-dioxane ( $x_{Dx} = 0.18$ ), as a function of the activity of H<sup>+</sup>.

The deprotonation constant  $k_d$  (average value of  $2.1 \pm 0.1 \times 10^{11} \text{ s}^{-1}$ ) is smaller than that observed for HMF in water<sup>9</sup> ( $2.5 \pm 0.4 \times 10^{11} \text{ s}^{-1}$ ) and reflects the decrease in dielectric constant of the solution rather than a disruption of the hydrogen-bonding network, because water clusters are still present in the mixtures at  $x_{\text{Dx}} = 0.18$ .<sup>S2,S3</sup> The behavior of the recombination rate constant  $k_{\text{rec}}$  explains why the geminate pair is detected in H<sub>2</sub>O:Dx mixtures and not in water. In water,  $k_{\text{rec}}$  is negligible (expected<sup>9</sup> to be smaller than  $2 \times 10^9 \text{ s}^{-1}$ ) and the geminate pair is not detected because the efficiency of pair

recombination  $[k_{rec}/(k_{rec} + k_{diss})]$  is much too small. On the other hand, the addition of cosolvent increases  $k_{rec}$  to *ca*.  $1.7 \pm 0.3 \times 10^{10}$  s<sup>-1</sup> at  $x_{Dx} = 0.18$ , making it of the same order of magnitude as  $k_{diss}$   $(1.0 \pm 0.1 \times 10^{10} \text{ s}^{-1})$ , resulting in a pair recombination efficiency of 0.6.

The dissociation constant of the geminate pair,  $k_{diss}$ , depends on the viscosity  $\eta$  of the solution and can be estimated<sup>S4</sup> from the diffusion coefficient of the species  $D_i$ , given by  $D_i = k_B T / (6\pi \eta R_i f_i)$ , where

$$f_t = \left[1.5 \frac{R_{H_2O}}{R_i} + \left(1 + \frac{R_{H_2O}}{R_i}\right)^{-1}\right]^{-1}.$$

In this equation,  $k_{\rm B}$  is the Boltzmann constant and  $R_i$  is the van der Walls radius of the species, calculated with the program GEPOL.<sup>S5</sup> Assuming that the binding energy of the encounter complex is zero and taking  $R_{H_2O} = 1.4$  Å,  $R_{H_5O_2^+} + R_{HMF} = 5.9$  Å and  $\eta = 2.19$  cP (interpolated from data at T = 25 °C in reference S6), one obtains  $k_{\rm diss} = 3(D_{H_5O_2^+} + D_{HMF})/(R_{H_5O_2^+} + R_{HMF})^2 = 1.1 \times 10^{10}$  s<sup>-1</sup>, which is in good agreement with the experimental value.

**Table 1S.** Decay times ( $\tau_j \pm 10\%$ ) and pre-exponential coefficients ( $A_{i,j}$ ) of HMF in water:1,4-dioxane solutions ( $x_{Dx} = 0.18$ ) at T = 25 °C obtained from global analysis of the decays collected at 480 nm (emission of AH<sup>+</sup>\*, i = 1) and at 620 nm (emission of A\*, i = 2), with excitation at 434 nm. The reduced chi-squared ( $\chi_i^2$ ) of the global analyses is also presented.

$[\mathrm{H}^+]$ /	$ au_1$ /	$ au_2$ /	$ au_3$ /	$A_{1,1}$	$A_{1,2}$	$A_{1,3}$	$A_{2,1}$	$A_{2,2}$	$A_{2,3}$	$\chi_1^2$	$\chi_2^2$
$mol \cdot L^{-1}$	ps	ps	ps								
0.50	168	48.8	4.40	0.03	0.07	0.90	1.00	-0.08	-0.92	1.03	1.00
0.40	170	49.0	4.50	0.03	0.06	0.91	1.00	-0.07	-0.93	1.06	1.10
0.30	169	47.5	4.35	0.03	0.06	0.91	1.00	-0.08	-0.92	0.93	1.01
0.20	158	53.3	4.35	0.02	0.06	0.92	1.00	-0.09	-0.91	1.02	1.03
0.10	157	57.8	4.35	0.01	0.05	0.94	1.00	-0.07	-0.93	0.95	1.08
$1.0 \times 10^{-9}$	146	-	-	-	-	-	1.00	-	-	-	1.06

### References

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