

# Pluronic L61 as a long-circulating modifier for enhanced liposomal delivery of cancer drugs

## Materials and Methods

### 1. Materials

Pluronic L61 was a product of BASF (USA). Soybean lipid (SPC) and DSPE-PEG2000 were purchased from Toshisun Enterprise Co., Ltd (Shanghai, China). Paclitaxel was obtained from Zhaolonghaiwang Biotechnology Co., Ltd (Xi'an, China). Cy5 fluorescence dye was acquired from Biotium, Inc. (USA). MCF-7 cell line was a generous gift of Pharmaceutical Institute, Zhejiang University, and was maintained with RPMI 1640 culture medium with 10% fetal bovine serum (FBS).

### 2. Pluronic L61 modification of paclitaxel loaded liposomes

Liposomes were prepared by using a film hydration method with sonication modified from our previous report.<sup>1</sup> In brief, SPC (2 mg) was dissolved in chloroform in a pear-shaped flask, and a certain quantity of paclitaxel and L61 ethanol solution was added. The organic solvent was removed by reduced pressure evaporation. The lipid/L61/paclitaxel film was then hydrated with 1ml of saline. The resultant suspension was sonicated using a sonifier (200 w, 2 min) equipped with ice-water bath. The final products contained paclitaxel (15 µg/ml) and SPC (2 mg/ml), with L61 concentration varying from 0% to 2% (w/w) in the different formulations.

DSPE-PEG modified liposome was also prepared as described above, with the DSPE-PEG concentration is 0.33 mg/ml (DSPE-PEG: SPC=1:5, w/w). The Cy5-liposomes were also prepared using the same “thin-film” procedures.

### **3. Particle size and morphology**

Particle size of the diluted samples was determined by using a Nano-S90 Zetasizer (Malvern, UK). A transmission electron microscope was utilized for the morphological observation of the liposomes (JEM-1230, JEOL, JP). Negative staining was processed using phosphotungstic acid.

### **4. Drug release study**

Release of paclitaxel from liposome was investigated using the modified method previously described by Yang et al.<sup>2</sup> In brief, 3 ml of liposomal solution with aliquots of paclitaxel about 300ug were sealed in dialysis bag (MW: 12,000 ) and be immersed into 300 ml PBS (pH 7.4) containing 0.1% (v/v) Tween 80. The application were kept on a shaker (180 rpm) at 37°C. With predetermined time interval, 4 ml solution was taken placed by fresh PBS for analysis. Cumulative release of free DOX was also conducted with the same procedure. Paclitaxel was determined by HPLC method (C18, 4.6×250 mm, 5µm, 40% acetonitrile, detection wavelength at 220 nm).

### **5. Liposome stability test**

Turbidity has been well employed as an index to assess serum protein binding efficiency of nanomaterials.<sup>1</sup> In brief, 20% (v/v) fetal blood serum was mixed with equivalent volume of liposome (1mg/ml). Turbidity was determined by UV absorbance under 400 nm at different time interval.

### **6. The protein-binding resistant effect of L61**

To access the effect of L61 on reduction of protein adsorption, the 96-well microplates were coated with different amounts of L 61. Then the FITC-labeled BSA was added and incubated at 37 °C for 2 hours for protein adsorption. The microplates were

washed with water for 5 times to remove unabsorbed protein. To quantify the surface adsorbed protein in the well, 200  $\mu$ l water was added and fluorescence was detected to quantify the amount of surface absorbed protein.

#### **4. *In vitro* cytotoxicity assay**

Human breast carcinoma cells (MCF-7) were cultured in RPMI 1640 medium containing 10% of fetal bovine serum and 2% of penicillin/streptomycin solution (10,000 IU/ml). The cytotoxicity was assessed by a routine MTT method. The cultured MCF-7 cells were exposed to the liposomal samples diluted with 1640 medium (equal to 75 ng paclitaxel per well). At 24 and 48hr after treatment, cell viability was measured by a standard MTT assay. Each sample was run quadruply.

#### **5. *In vivo* imaging study**

All the animal experiments were approved by the Animal Care and Use Committee of Zhejiang University. The anesthetized Balb/c nude mice were given 100  $\mu$ l of Cy5 liposome solution through tail vein injection. *In vivo* fluorescence optical evaluation of intratumor distribution was conducted by using the Maestro I system (CRi, USA) at 2, 5, 15, 30, 60, 180, 360, 720 and 1440 min postdose, respectively. The Cy5 excitation filter: 615–665 nm, emission filter: 700 nm. The data was analyzed with Maestro software (Version 2.5.42), and the autofluorescence of animals was subtracted. The *in vivo* fluorescence in the brain and liver was analyzed with Image J software, and the semi-quantitative comparison of various liposomes accumulated in the organs was conducted.

#### **6. *In vivo* tumor accumulation study**

Tumor-bearing animal model were established by injecting 200  $\mu$ l of MCF-7 cells ( $1 \times 10^6$ /ml) suspension to the armpits of the male Balb/c nude mice. The tumor-bearing

nude mice were injected the Cy5-labelled Pluronic L61-liposomes (100  $\mu$ l) for *in vivo* fluorescence imaging by Maestro I system with a procedure as described above. Liposomes without L61 modification were used as control.

### **7. Antitumor treatment in Mice**

The male ICR mice weighed from 18 to 20 g were randomly divided into 4 groups: i) saline, ii) SPC liposomes, iii) PEG-liposomes, and iv) 0.1% L61-liposomes. Each mouse injected subcutaneously with 100  $\mu$ l of cell suspension containing  $3.26 \times 10^7$  H22 cells at the axillary space. The pharmacodynamic test was started when the tumor volumes developed to approximately  $1.5 \times 1.5$  cm (L  $\times$  W) (L is the long diameter of the tumor and W is the short diameter). Liposomes were prepared and injected at a paclitaxel dose of 6 mg/kg into the mice through the tail vein. The body weight and tumor sizes of the mice were measured daily and the variation ratios were calculated.

### **8. Data analysis**

The results are presented as means  $\pm$  standard deviations (SD). Where applicable, multiple comparisons were evaluated by one-way ANOVA.

### Supplementary Table and Figures:

Table S1. Particle size and PDI of liposomes

Hydrodynamic particle size and PDI of liposome							
L61 Con. (%)	0	0.02	0.05	0.1	0.2	0.5	1
Size/nm	155±14.6	138±10.2	129±14.3	104±11.8	145±13.6	168±17.1	246±18.3
PDI	0.323	0.360	0.371	0.306	0.354	0.328	0.248

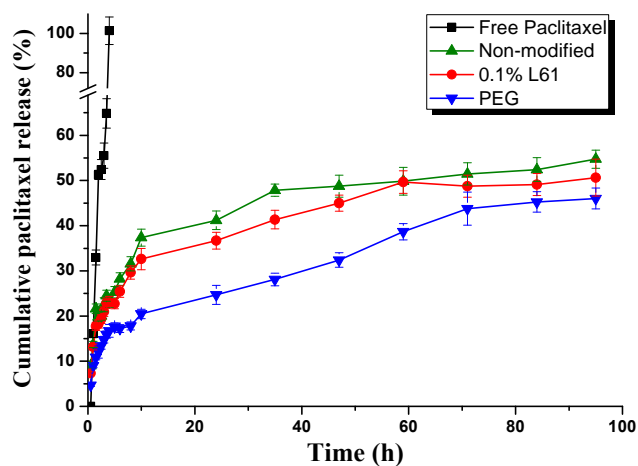


Fig. S1 Drug release profiles from various liposomes

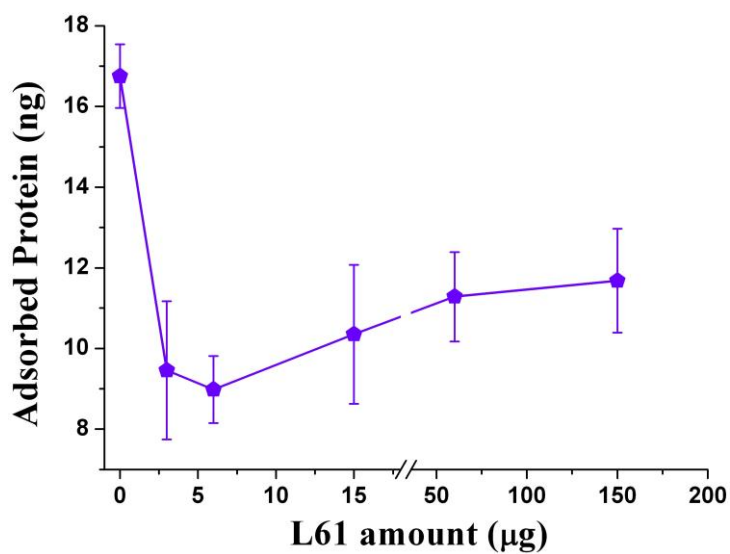
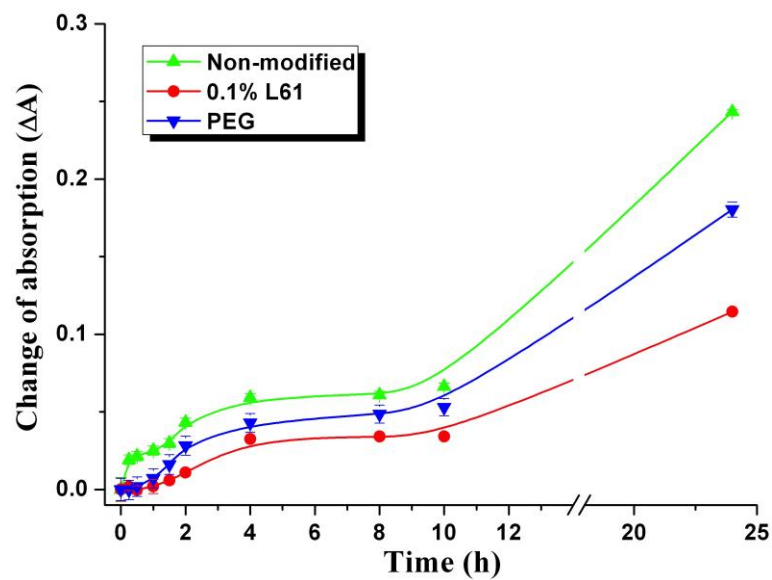
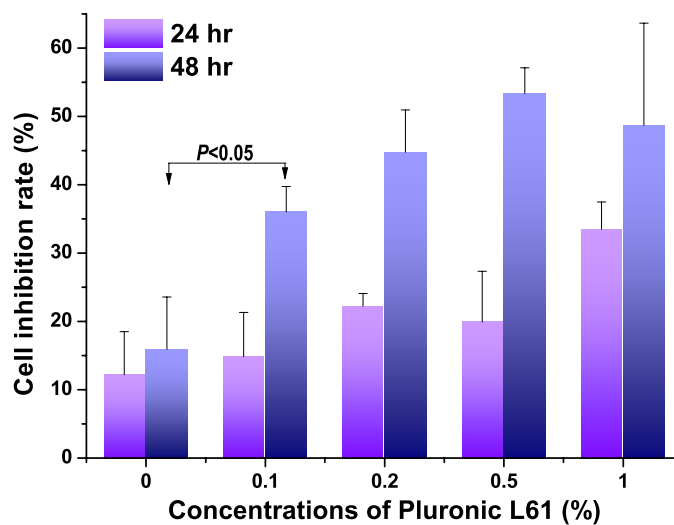


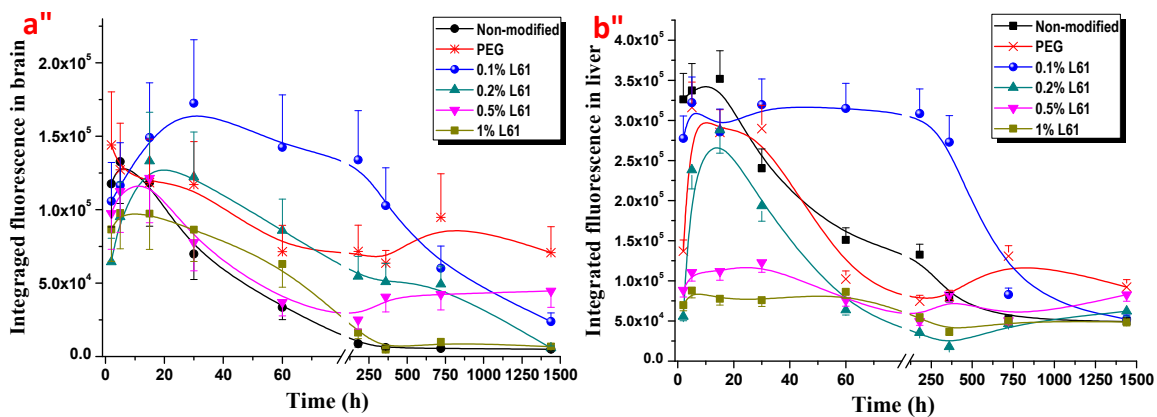
Fig. S2 Protein-binding resistant effect of Pluronic L61 coating



**Fig. S3** Turbidity change of the liposomes



**Fig. S4** Cell proliferation studies of paclitaxel L61-modified liposomes. It showed a slight trend that high concentration of L61 brought higher cytotoxicity at 48 h, presumably due to the inhibition effect of L61.



**Fig. S5** The *in vivo* fluorescence in the brain (a) and liver (b) was analyzed with Image J software and the semi-quantitative comparison of various liposomes accumulated in the organs.

## Reference:

1. Y. Huang, Y. Rao, J. Chen, V. C. Yang and W. Liang, *J Biomed Mater Res A*, 2011, **96**, 513-519.
2. S. Chung, C. Shim, D. Kim, *Int J Pharm*, 2007, **338**, 317-326.