## **Supporting Information.**

Includes synthesis, <sup>1</sup>H, <sup>13</sup>C, DEPT NMR of **1**, MGC data, FTIR, CD, DSC and <sup>1</sup>H-<sup>1</sup>H COSY of the gel before and after sonication, proposed model of assembly, Jobs plot and individual binding constant vs p plots for the <sup>1</sup>H NMR titration of **1** with TBAF and TBACl and other studies.



Scheme S1: Synthesis of gelator

- 20 Synthesis of the bolaaphiphilic peptide (1). (5)-2-benzamidopropanoic acid<sup>1</sup> (2.2 eq, 2.12 g, 11 mmol) and CDI (2.2 eq, 1.78 g, 11 mmol) were taken in dry THF and stirred for 5 min. The 1,12-diamino-dodecane (1 eq, 1 g, 5 mmol) in dry THF was added dropwise at 0 °C to the activated acid solution, and the reaction mixture was stirred for 1.5 h under N<sub>2</sub> atmosphere. The reaction was monitored by TLC (CHCl<sub>3</sub>/MeOH 9.5:0.5 v/v). THF was removed from the reaction mixture by vacuum and 50 mL of CHCl<sub>3</sub>/isopropanol (9:1) was added to it. The organic layer was washed with dil. HCl and sat. NaHCO<sub>3</sub>, and dried over anhyd. sodium sulphate. The solvent was then removed under rotary evaporation to yield the crude 1, which was purified by 2s column chromatography (stationary phase was silica 100-200 mesh and eluent was CHCl<sub>3</sub>/MeOH 9:1 v/v) to obtain 1 as a white solid (2.5 g, 90%). m.p. 160 °C. FTIR (KBr pellet, cm<sup>-1</sup>): 1637 (amide, C=O str.), 1535 (N-H bend), 3305 (N-H str.). <sup>1</sup>H NMR (400 MHz, DMSO-d6, ppm): δ= 1.12-1.24 (16H, (CH<sub>2</sub>)<sub>4</sub>), 1.25-1.31 (6H, CH<sub>3</sub>),1.32-1.40 (4H, NH-CH<sub>2</sub>-CH<sub>2</sub>), 3.29 (4H, NH-CH<sub>2</sub>), 4.38-4.41 (2H, C-H), 7.40-7.87(10H, Ph-H), 7.80-7.82 (NH<sub>b</sub>), 8.37-8.39
- $(NH_a)$ .<sup>13</sup>C NMR (400 MHz, DMSO)  $\delta$ = 172.56, 166.41, 134.60, 131.69, 128.59, 127.96, 49.43, 38.95, 29.47, 29.44, 29.20, 26.74, 18.58. C<sub>32</sub>H<sub>46</sub>N<sub>4</sub>O<sub>4</sub>+Na<sup>+</sup> calc: 573.3411, found: 573.3405.





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Fig. S2. <sup>13</sup>C NMR (100 MHz) spectra of 1 in DMSO-d6.



Fig. S3. DEPT (100 MHz) spectra of 1 in DMSO-d6.

Solvent	MGC (%w/v)	$T_m$ (°C)	_
THF	1.0	45±3	
1,4-Dioxane	1.5	50±2	5

Table S1. MGC values and melting temperatures (T\_m) at MGC  $^{\rm 10}$ 



Fig. S4. Optical microscopy images (100x magnification) of: a) the microcrystals of 1; b) short fibrils formed after 3 min sonication of the microcrystals



**Fig. S5**. Variable temperature <sup>1</sup>H NMR of the sonication-induced assembly in THF-d8. The physical gel-to-sol transition temperature (T<sub>m</sub>) around

45 °C is highlighted.

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Fig. S6. FTIR spectra of 1 recorded as KBr pellets. Dried microcrystals before sonication (red line), and dried fibrils formed after sonication (blue line).



Fig. S7. Proposed model for the assembly of 1 by intermolecular hydrogen bonding interactions between the amide groups.



Fig. S8. CD spectra of 1 recorded as a solution in THF (0.75 mg/mL) at 25 °C. Before sonication (red circles), and after sonication (blue triangles).



Fig. S9. DSC profiles of 1, as dried microcrystals before sonication (red line), and as dried fibrils formed upon sonication (blue line). 4 mg of pre-dried sample was heated at the scan rate of  $1^{\circ}$ C/min.

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Fig. S10. <sup>1</sup>H-<sup>1</sup>H COSY spectra of 1 (5.5 mM) in THF-d8. a) before sonication; b) after sonication.



**Fig. S11**. Relative rates of anion-induced disassembly of nanofibers (prepared through sonication of 1 at MGC in THF) by  $F^-$  or  $CI^-$  ions (4 eqv), as evaluated from the increase in %T at 600 nm.



**Fig. S12**. (a) Jobs plot for the chemical shift of NH<sub>a</sub> proton during the titration of TBAF with **1**. X = mole-fraction of **1**,  $\Delta\delta$  = change in the chemical shift of NH<sub>a</sub> proton. (b) *K*<sub>a</sub> vs. *p* for the NH<sub>a</sub> proton of **1** during NMR titration with TBAF; *p* is the probability of complexation (data fitted in polyfit using <sup>30</sup> Origin<sup>TM</sup> 8.0 (R<sup>2</sup>=0.999)).



**Fig. S13**. (a) Jobs plot for the chemical shift of NH<sub>a</sub> proton during titration of TBACl with **1**. X = mole-fraction of **1**,  $\Delta\delta$  = change in the chemical shift of NH<sub>a</sub> proton of **1**. (b) *K*<sub>a</sub> vs. *p* for the NH<sub>a</sub> proton of **1** during NMR titration with TBACl; *p* is the probability of complexation (data fitted in polyfit using Origin<sup>TM</sup> 8.0 (R<sup>2</sup>=0.999)).



35 Fig S14: Optical microscopy images (40x magnification) of the regenerated gel exhibiting fibrous architecture similar to the pristine gel (scale 50 µm).



Fig S15: CD profiles of TFA regenerated gel showing the regeneration of the CD signal similar to the pristine gelator.

 A.I. Vogel, A.R. Tatchell, B.S. Furnis, A.J. Hannaford, P.W.G. Smith, in Vogel's Textbook of Practical Organic Chemistry, (5th ed.) Longman Group Ltd., London 1989 pp. 815.

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