

Supporting Information

Organocatalytic stereoselective approach to the total synthesis of (-)-halosaline

Vishwajeet Jha^a and Pradeep Kumar*^a

^aDivision of Organic Chemistry, CSIR-NCL (National Chemical Laboratory), Pune 411008, India

*Corresponding Author: Telephone number: +91-20-25902050, Fax number: +91-20-25893614;

e-mail address: pk.tripathi@ncl.res.in

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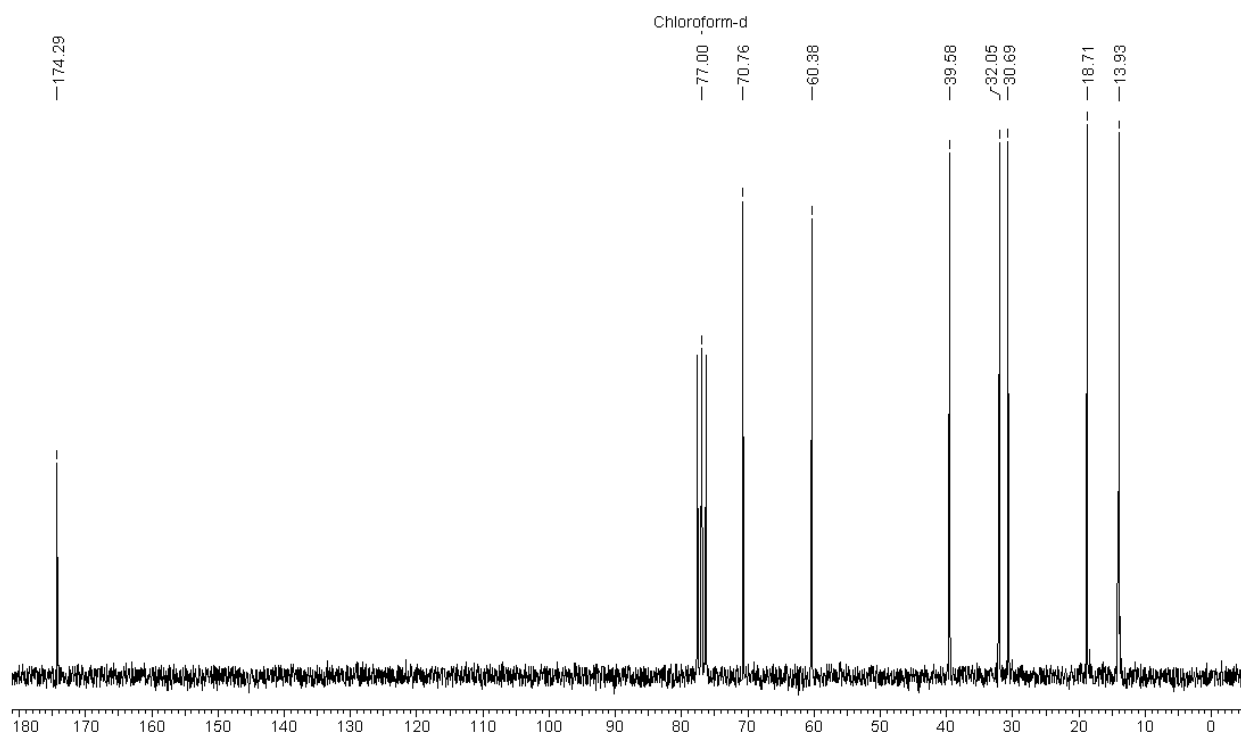
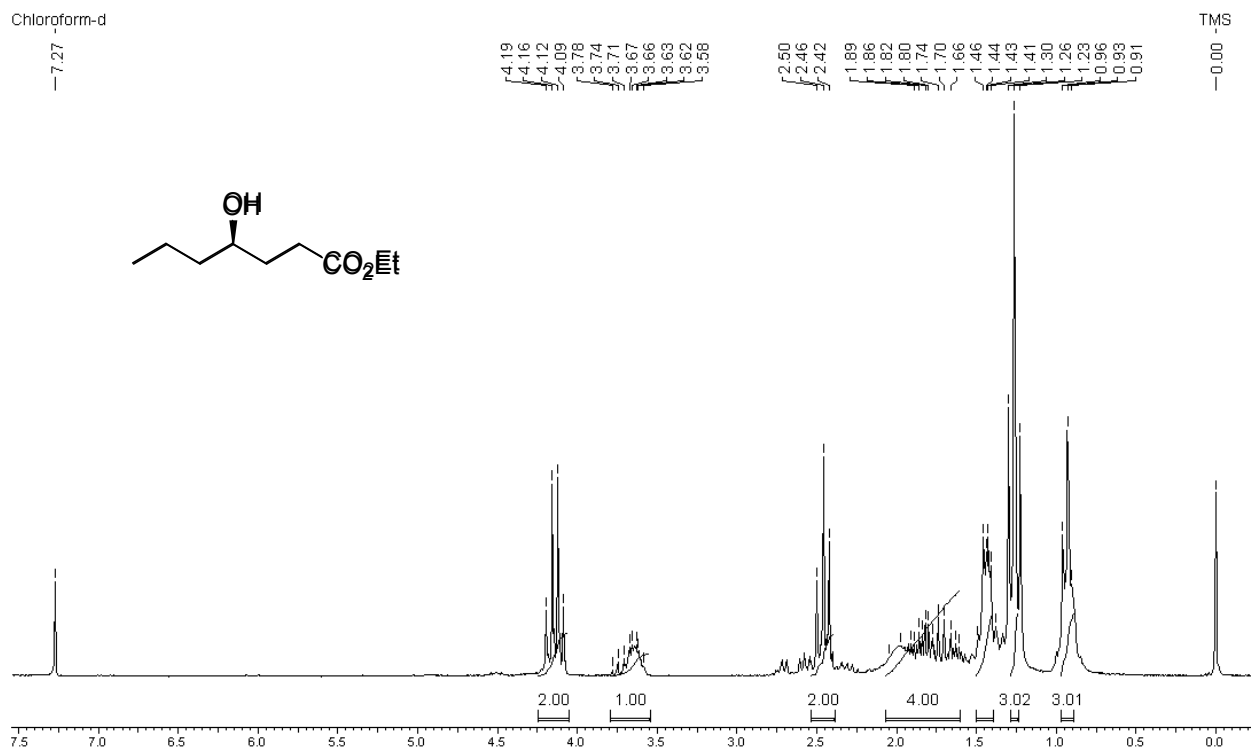
General Methods :

All reactions requiring anhydrous conditions were performed under a positive pressure of argon using oven-dried glassware (110 °C), which was cooled under argon. Solvents used for chromatography were distilled at respective boiling points using known procedures.

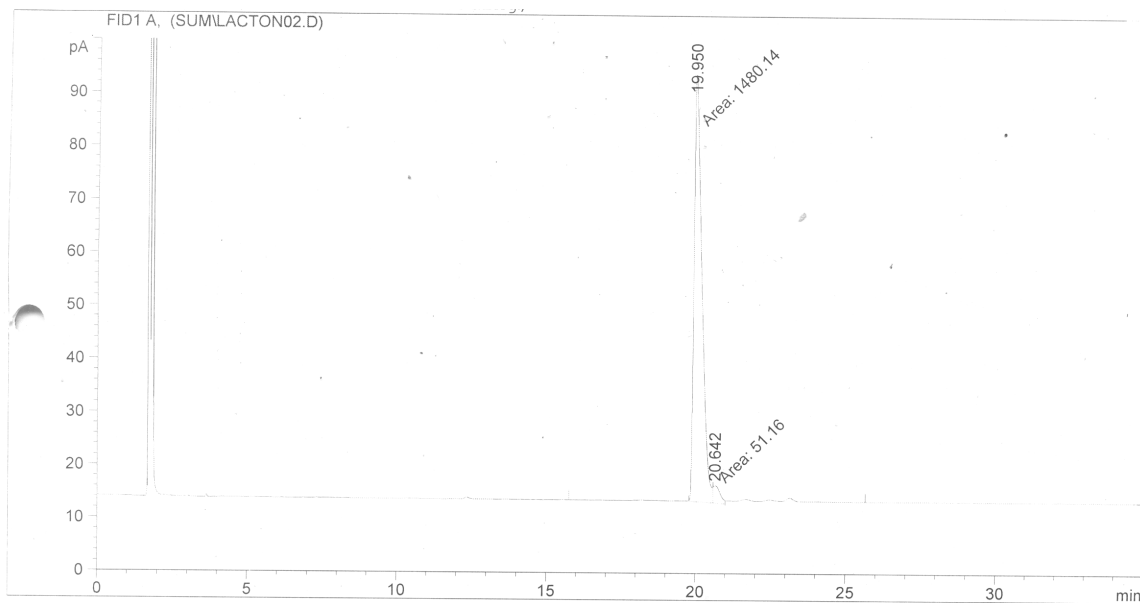
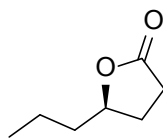
All commercial reagents were obtained from Sigma-Aldrich Chemical Co. and Alfa Aesar. Progress of the reactions was monitored by TLC using precoated aluminium plates (Merck kieselgel 60 F₂₅₄). Column chromatographies were performed on silica gel 60-120/ 100-200/ 230-400 mesh obtained from S. D. Fine Chemical Co. India or Spectrochem India. Typical syringe and cannula techniques were used to transfer air- and moisture-sensitive reagents.

IR spectra were recorded on a Perkin–Elmer infrared spectrometer model 599-B and model 1620 FT-IR. ¹H NMR spectra were recorded on Bruker AC-200, Bruker AV-400, Jeol-400 and Bruker DRX – 500 instruments using deuterated solvent. Chemical shifts are reported in ppm. Proton coupling constants (*J*) are reported as absolute values in Hz and multiplicity (br, broadened; s, singlet; d, doublet; t, triplet; m, multiplet). ¹³C NMR spectra were recorded on Bruker AC-200, Bruker AV- 400, Jeol-400 and Bruker DRX-500 instruments operating at 50 MHz, 100 MHz, and 125 MHz, respectively. ¹³C NMR chemical shifts are reported in ppm relative to the central line of CDCl₃ (δ 77.0). Elemental analyses were determined at Central Elemental Analysis Facility division at National Chemical Laboratory. All HPLC analyses used to determine enantiomeric purity were calibrated with sample of the racemate.

Ethyl (*R*)-4-hydroxyheptanoate (9):



Enantiomeric excess:

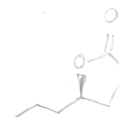


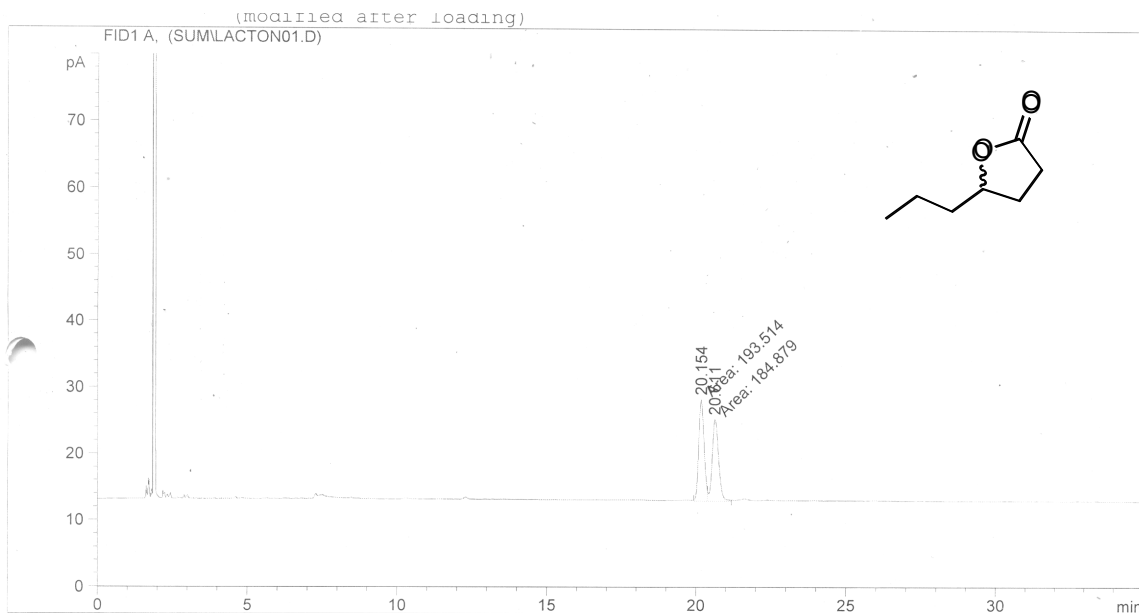
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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Concentration : 1.0000
Sample Amount : 2.00000 [ng/ul] (not used in calc.)

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	19.950	MF	0.3058	1480.13977	80.67734	96.65905
2	20.642	FM	0.2582	51.16001	3.30218	3.34095



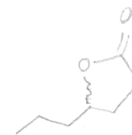


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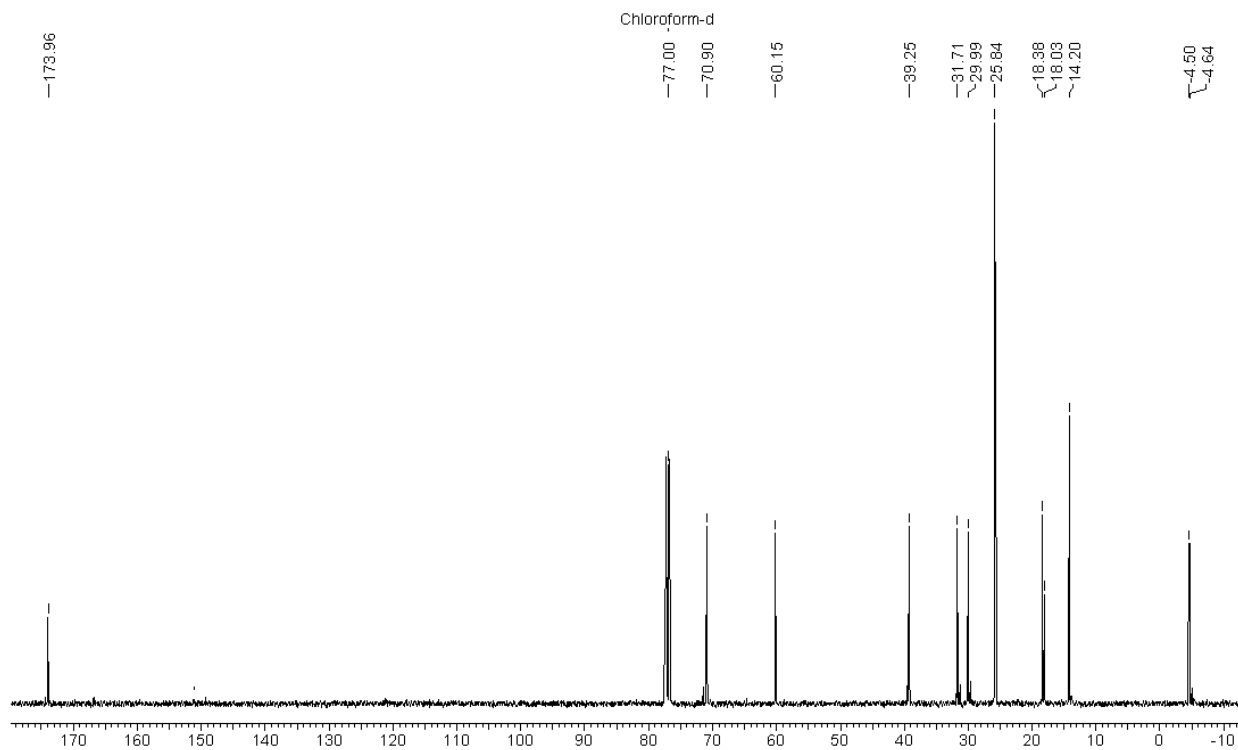
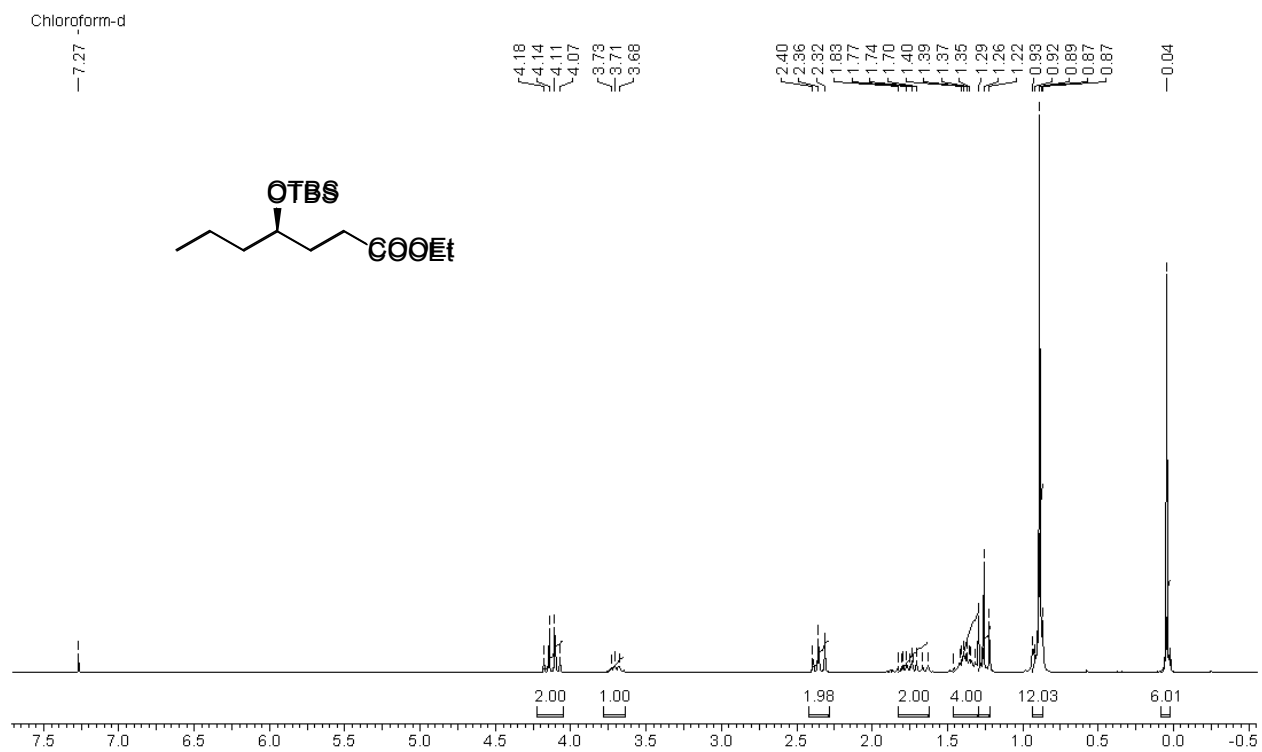
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Multiplier : 1.0000
Dilution : 1.0000
Sample Amount : 2.00000 [ng/ul] (not used in calc.)

Signal 1: FID1 A,

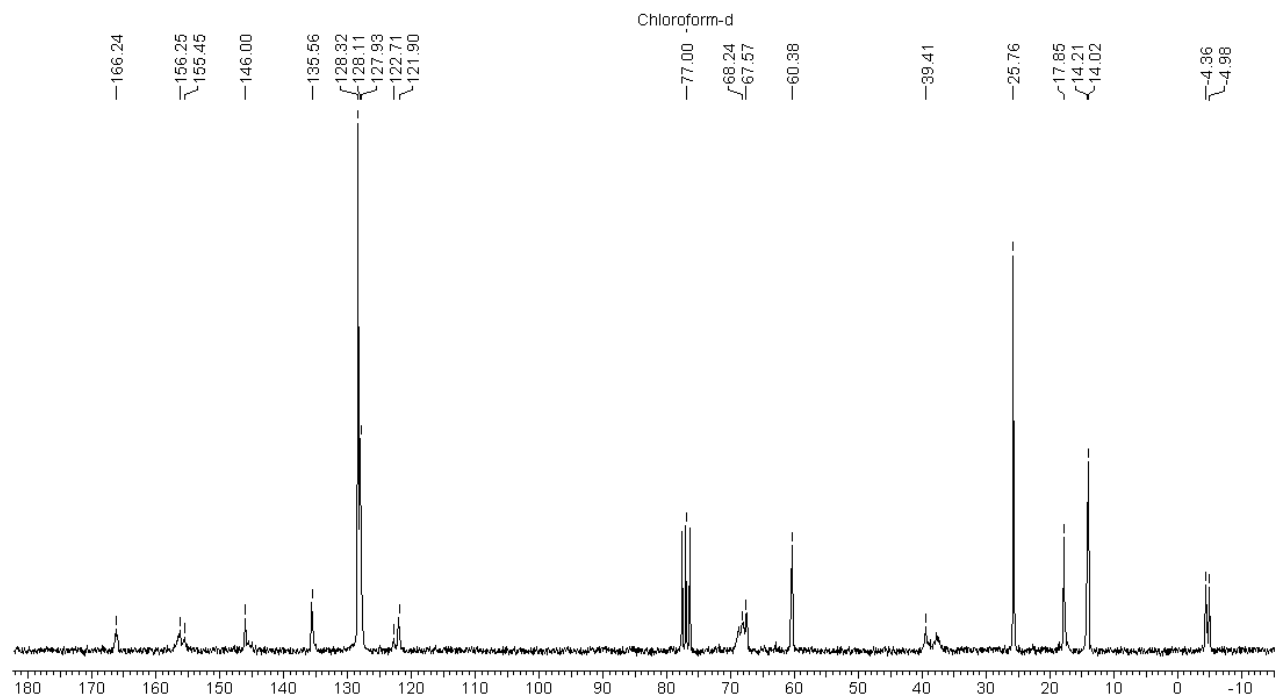
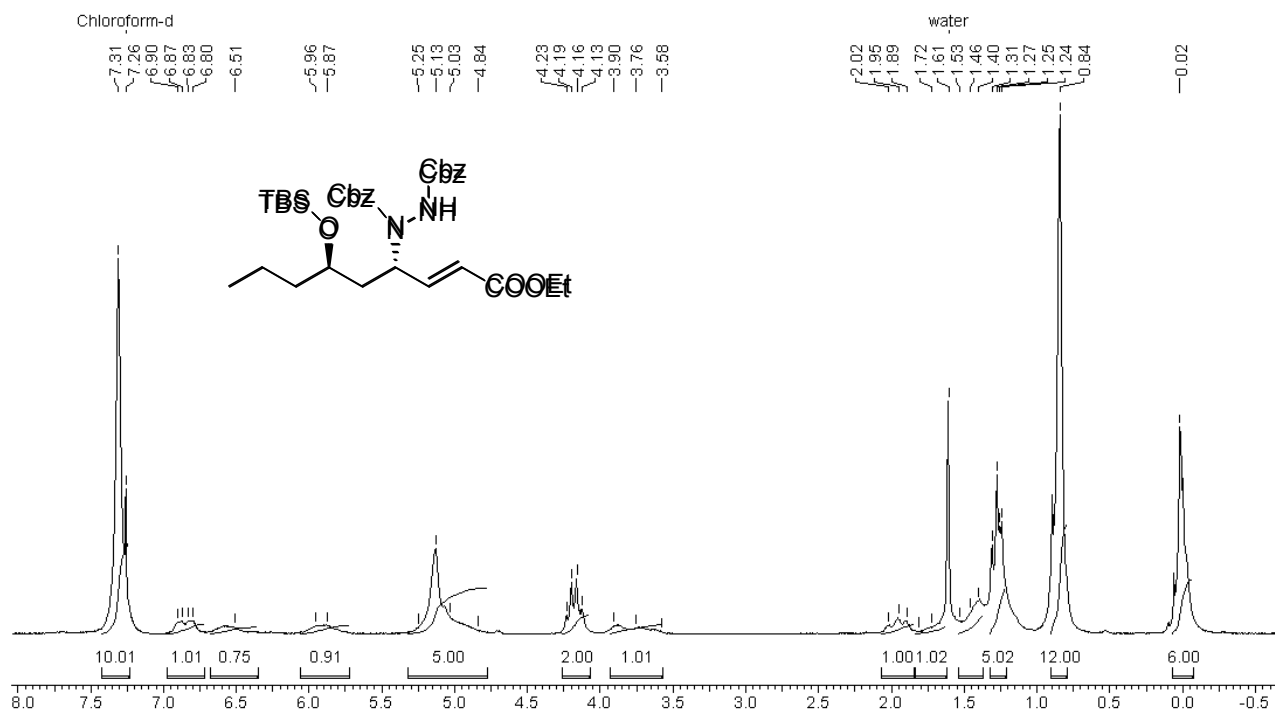
Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	20.154	MF	0.2097	193.51364	15.37672	51.14102
2	20.611	FM	0.2495	184.87860	12.35137	48.85898



Ethyl (*R*)-4-((*tert*-butyldimethylsilyloxy)heptanoate (11):

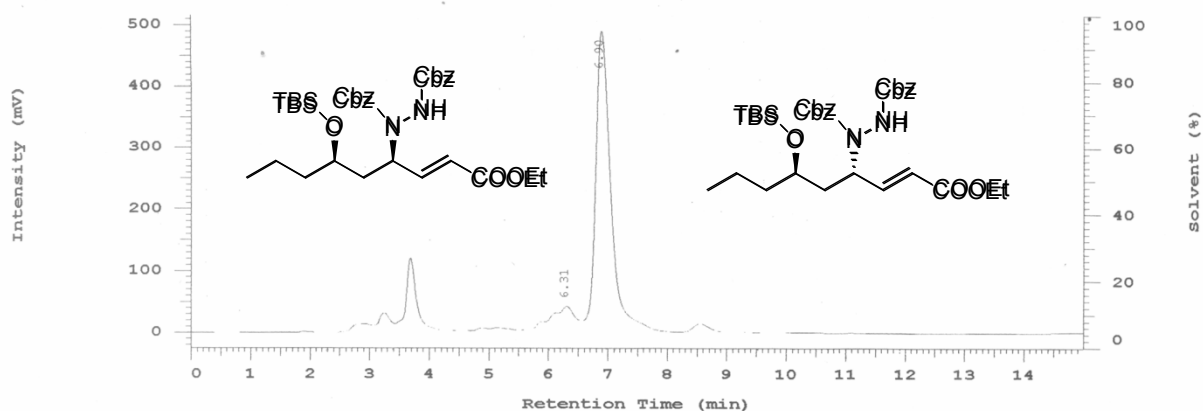


Dibenzyl 1-((4S,6R,E)-6-((*tert*-butyldimethylsilyl)oxy)-1-ethoxy-1-oxonon-2-en-4-yl)hydrazine-1,2-dicarboxylate (8):



Volume: 10.0 ul

Chrom Type: HPLC Channel : 1

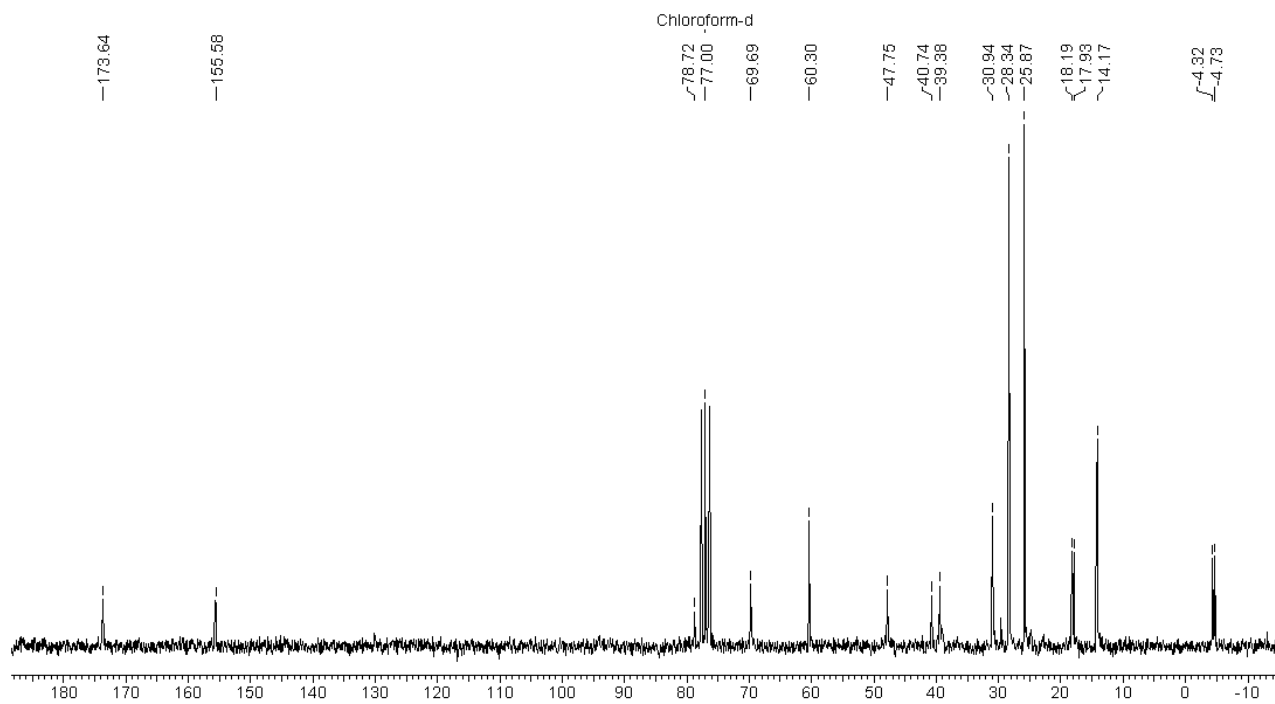
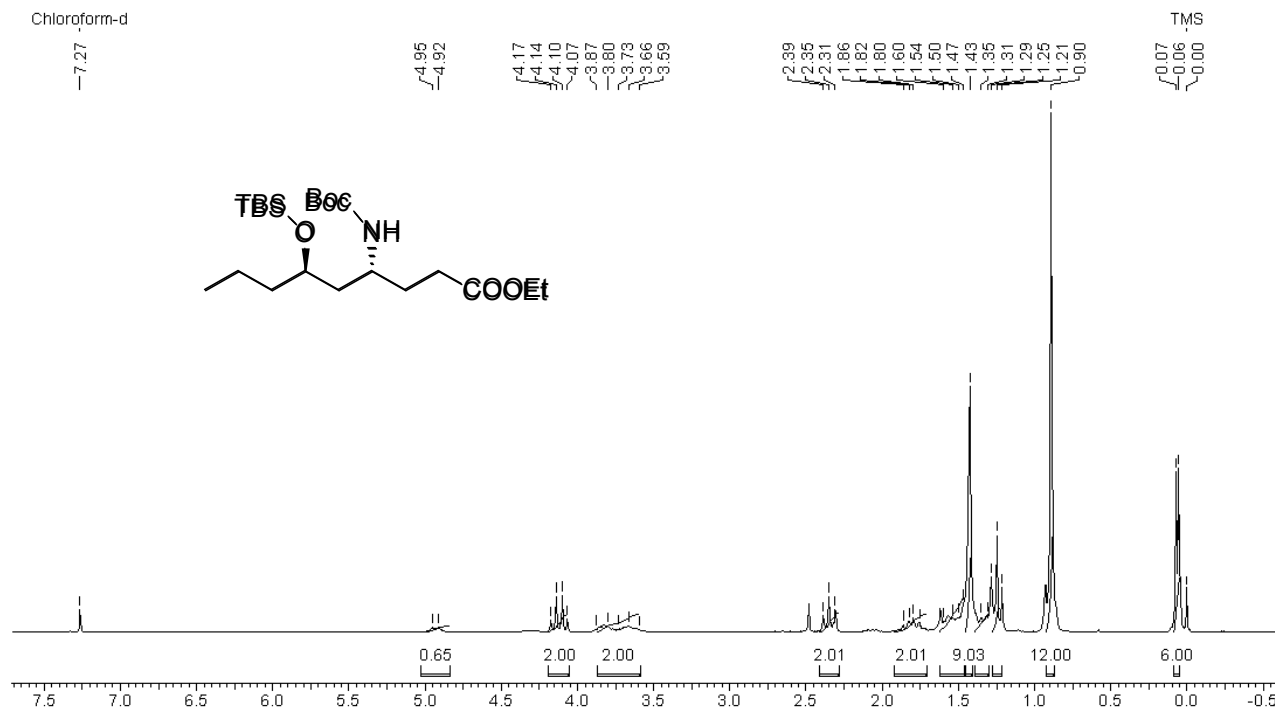


Peak Quantitation: AREA

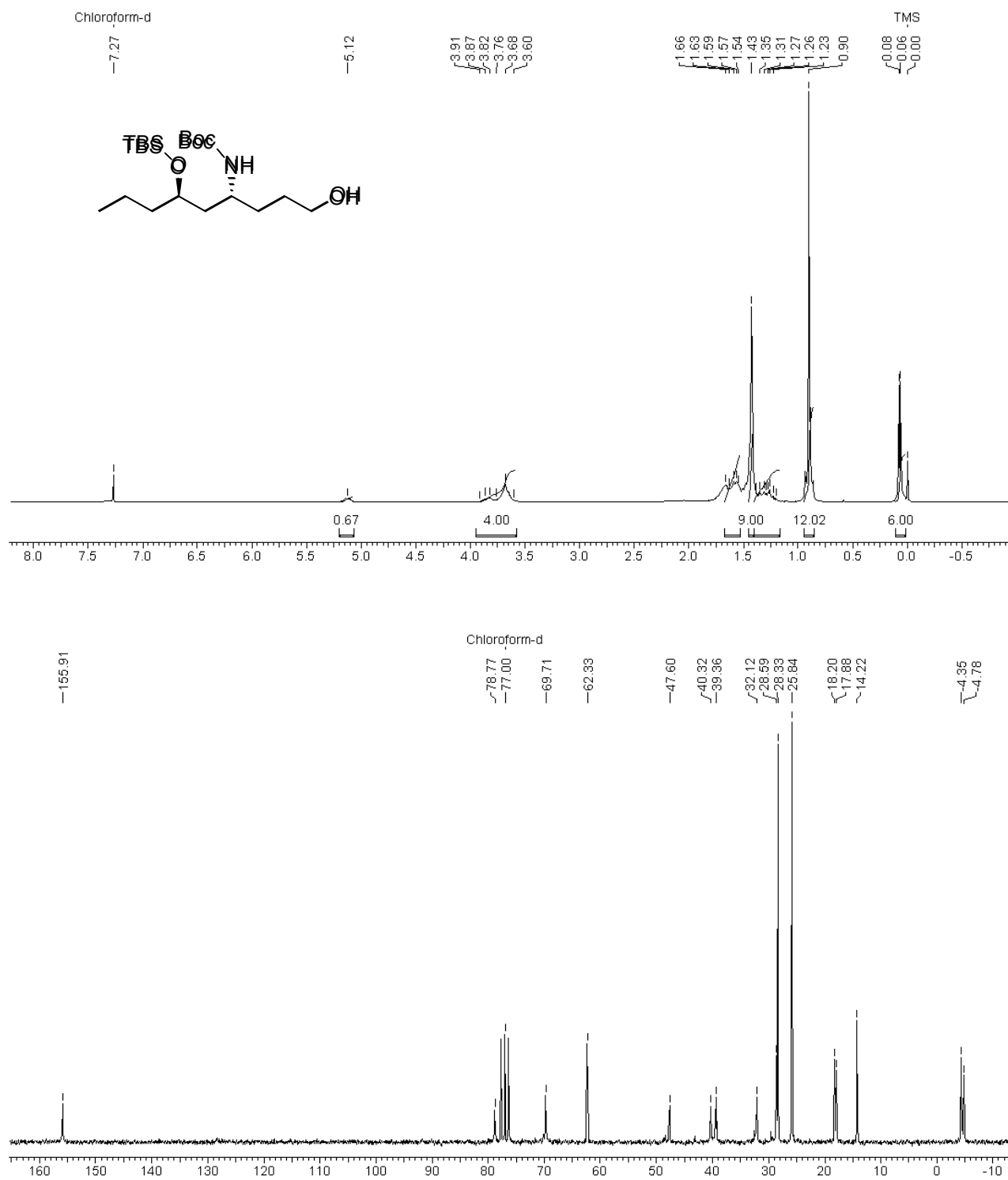
Calculation Method: AREA%

No.	RT	Height	Area	Area %
1	6.31	15473	153762	1.940
2	6.90	474211	7773316	98.060
		489684	7927078	100.000

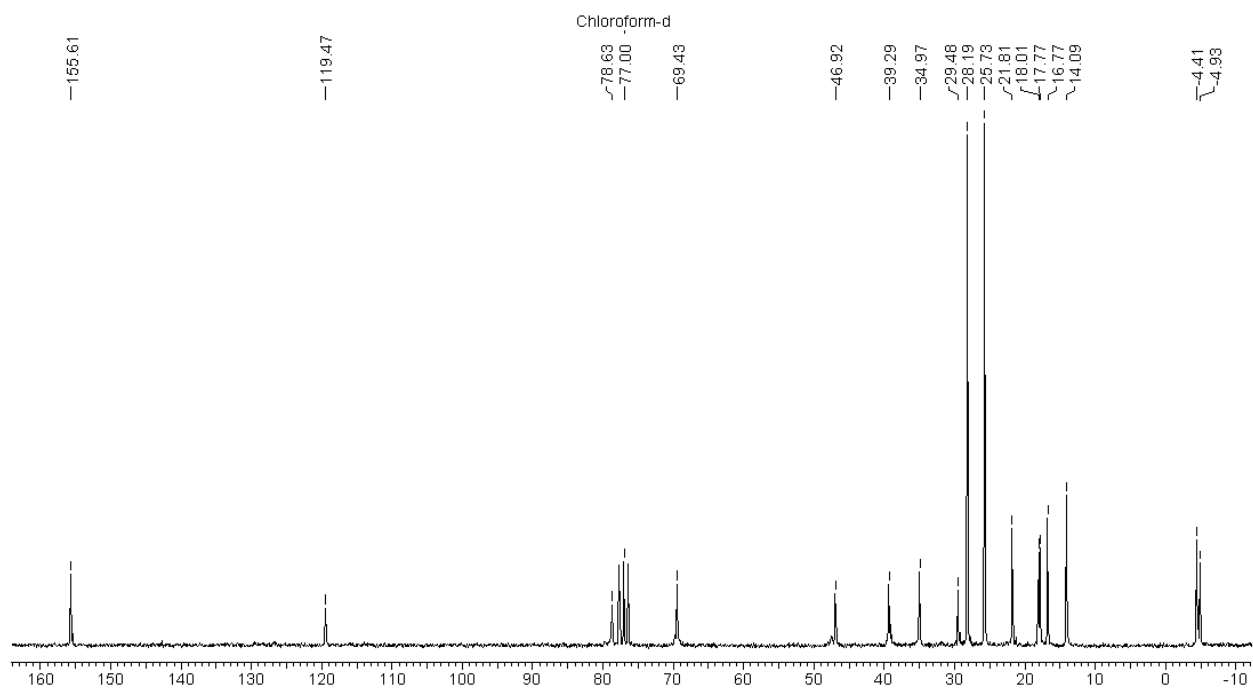
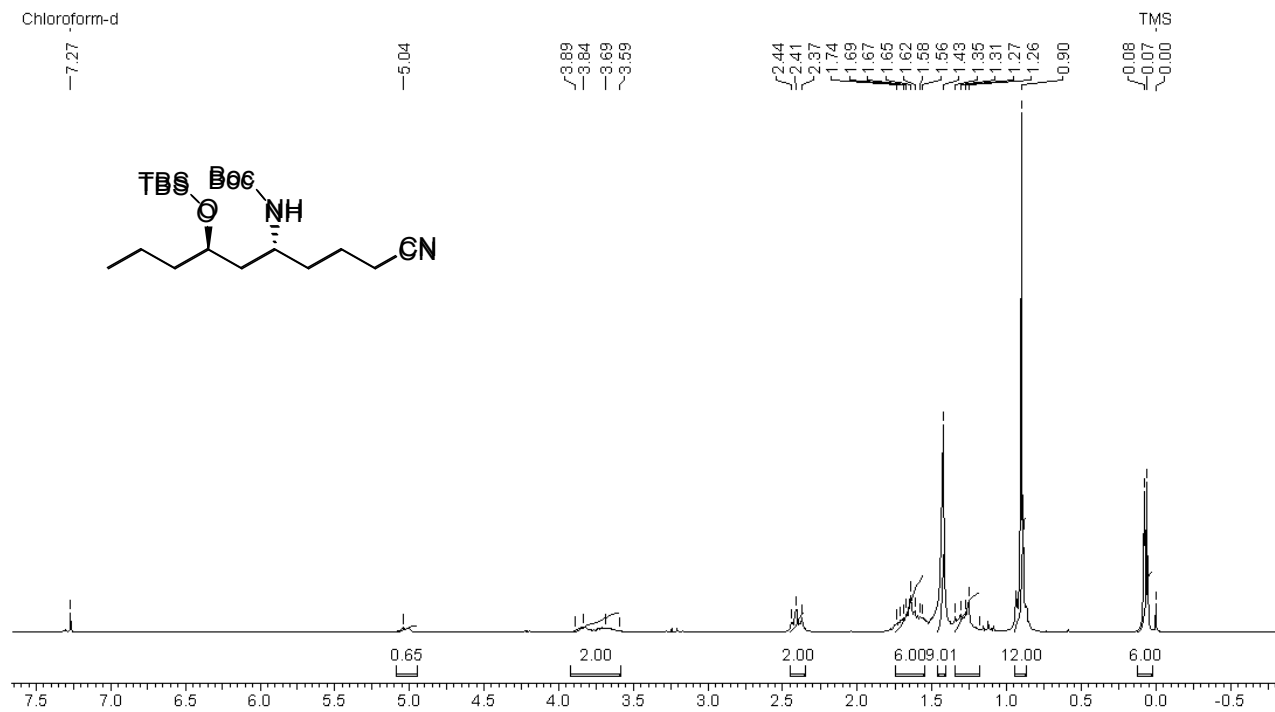
Ethyl (4*R*,6*R*)-4-((*tert*-butoxycarbonyl)amino)-6-((*tert*-butyldimethylsilyl)oxy)nonanoate (12):



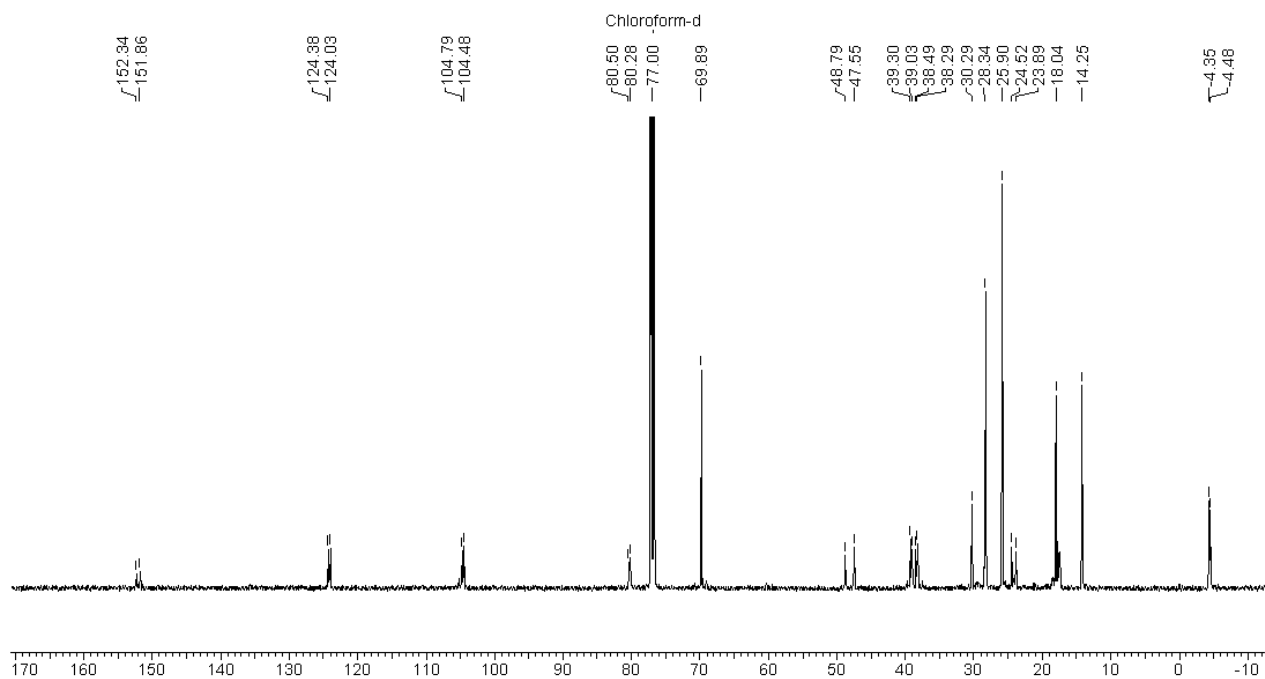
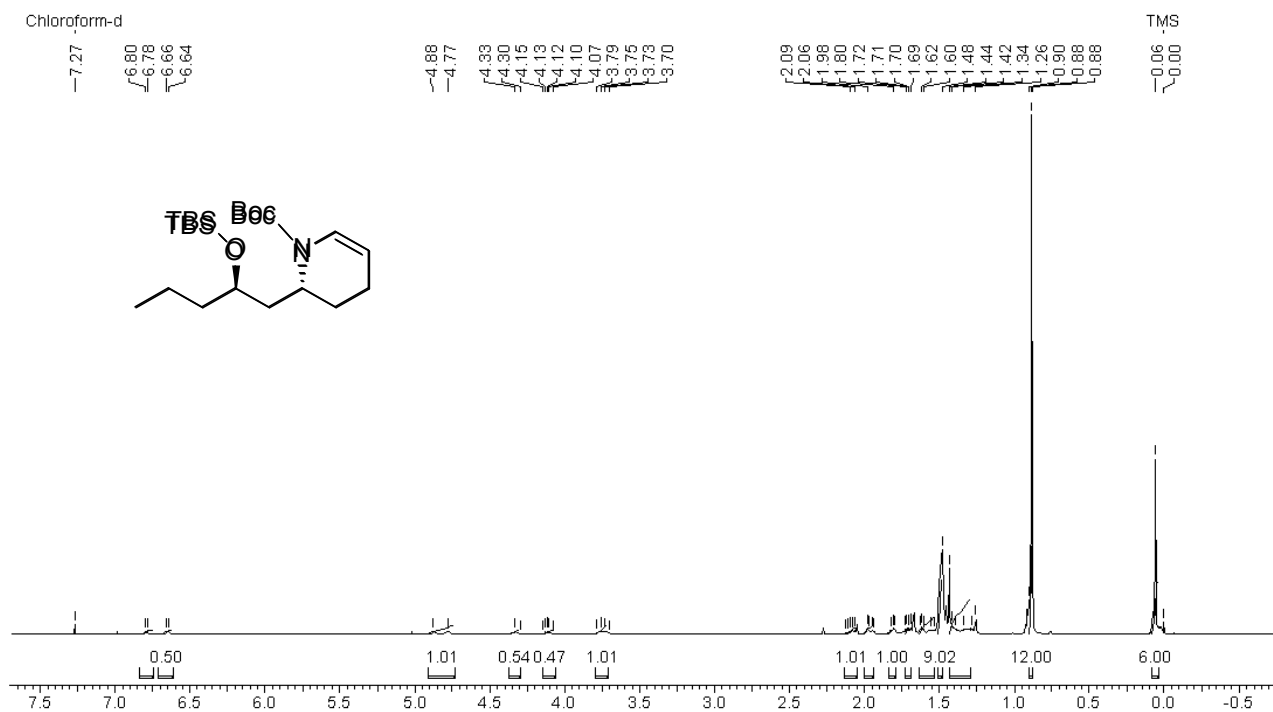
***tert*-Butyl ((4*R*,6*R*)-6-((*tert*-butyldimethylsilyl)oxy)-1-hydroxynonan-4-yl)carbamate (13):**



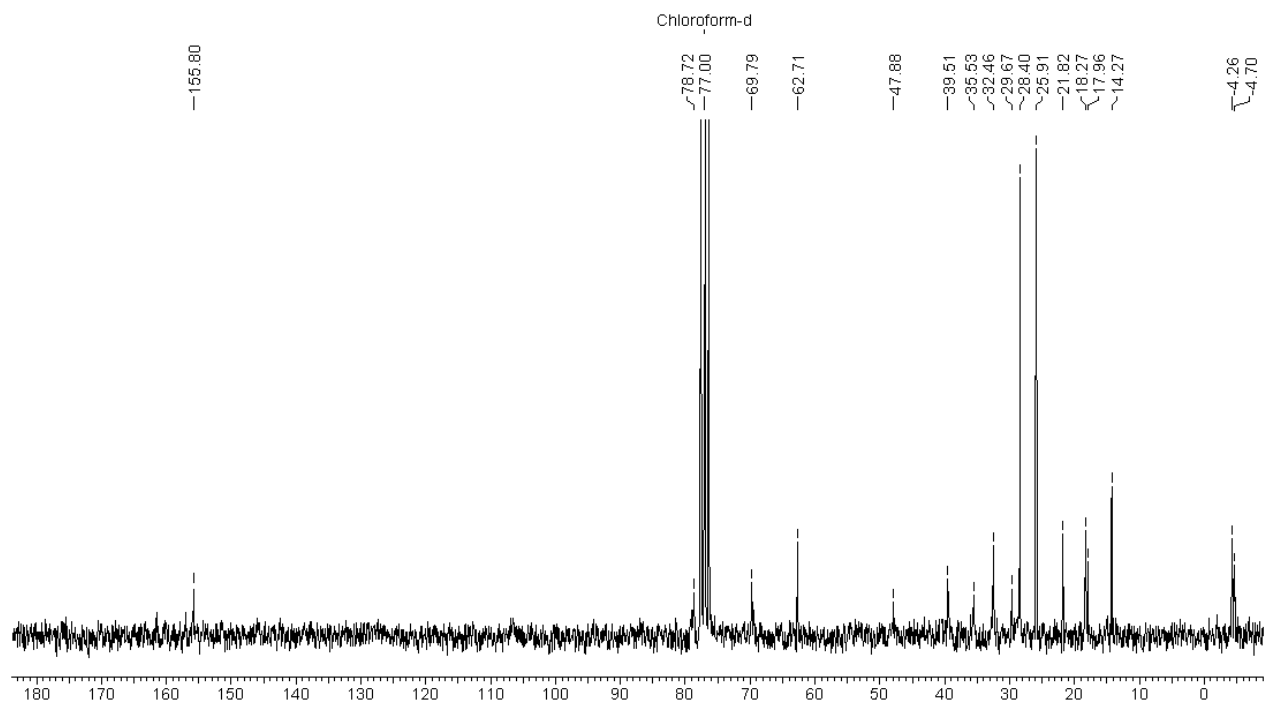
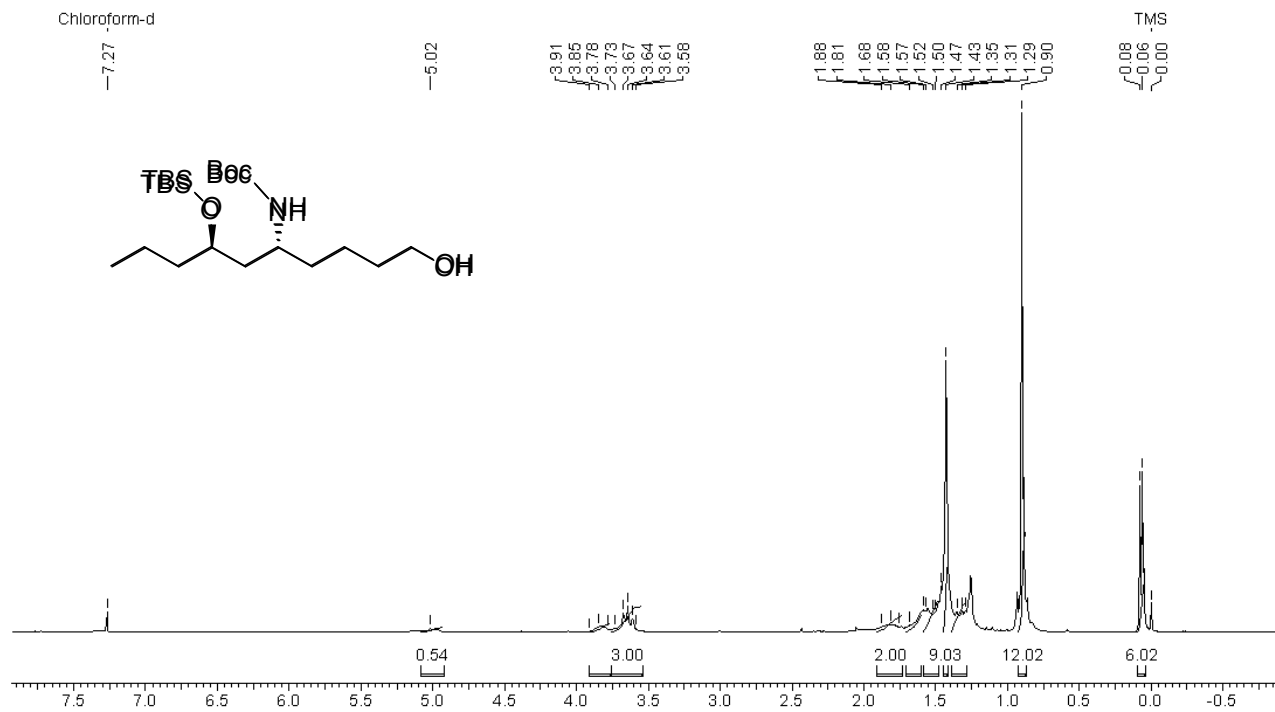
***tert*-Butyl ((4*R*,6*R*)-6-((*tert*-butyldimethylsilyl)oxy)-1-cyanononan-4-yl)carbamate (18):**



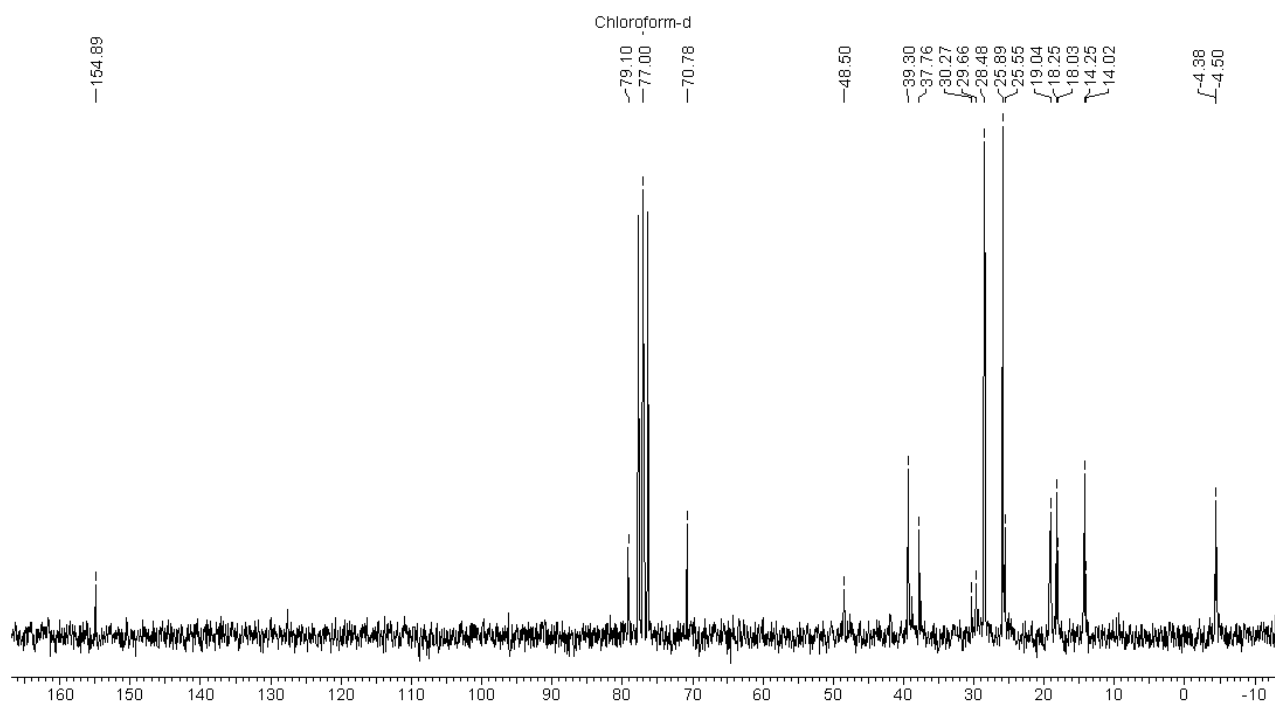
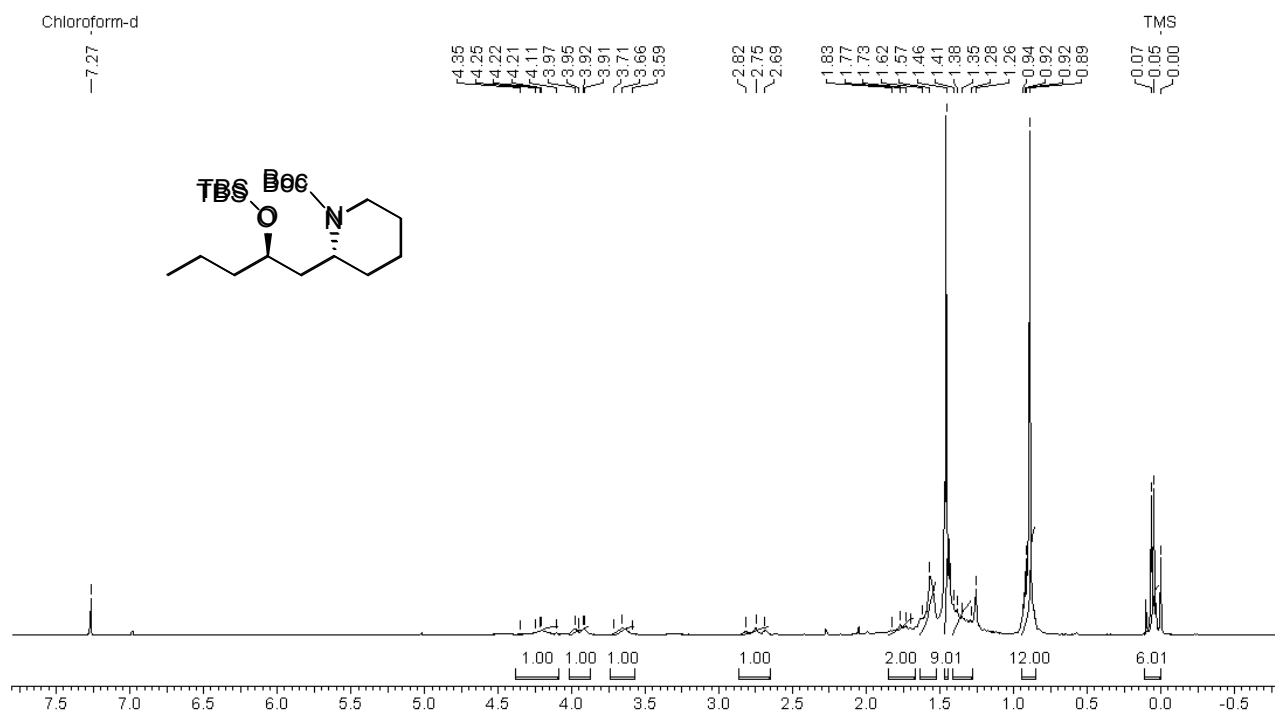
***tert*-Butyl (R)-2-((R)-2-((*tert*-butyldimethylsilyloxy)pentyl)-3,4-dihydropyridine-1(2H)-carboxylate (19):**



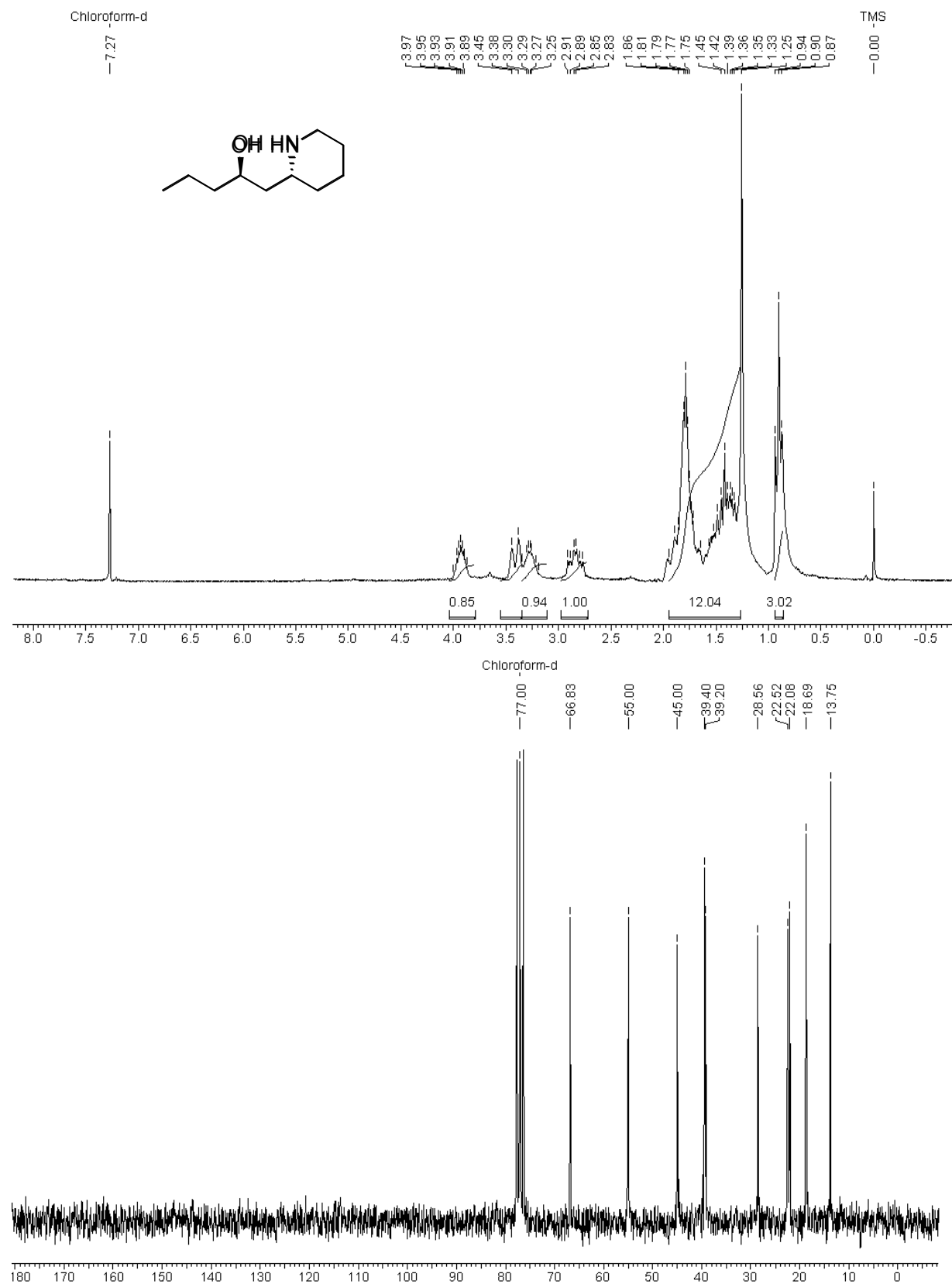
***tert*-Butyl ((5*R*,7*R*)-7-((*tert*-butyldimethylsilyl)oxy)-1-hydroxydecan-5-yl)carbamate (20):**



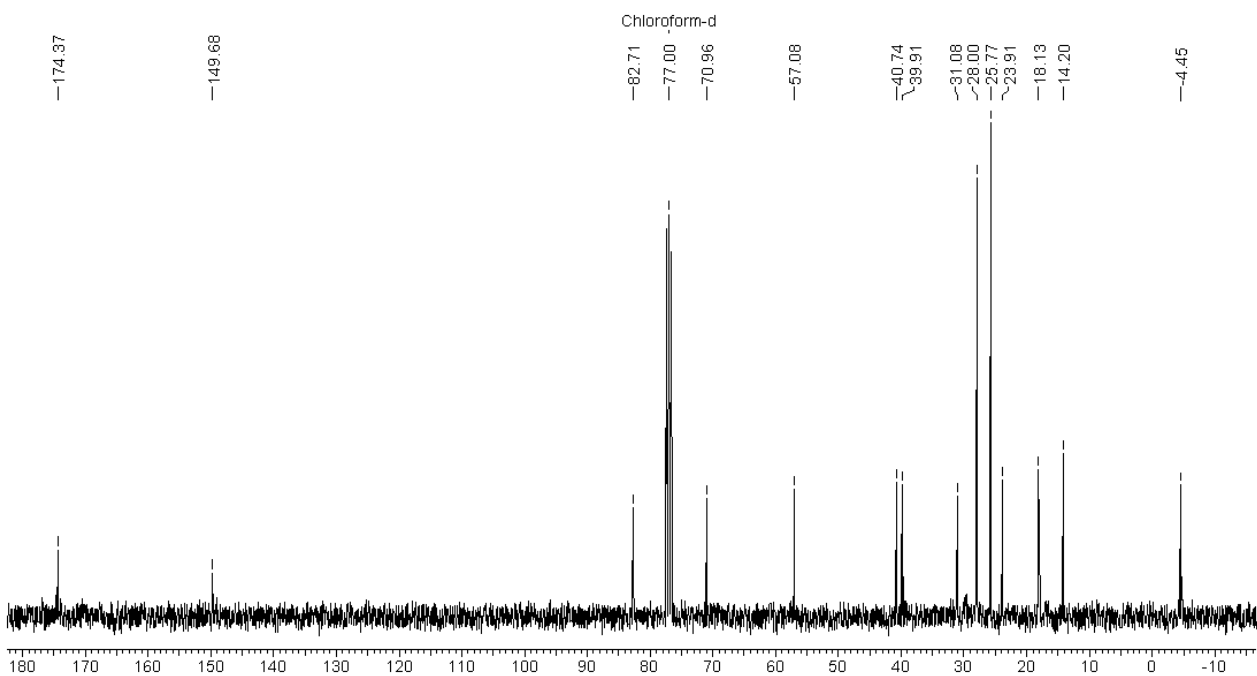
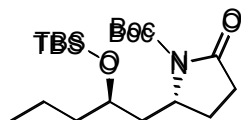
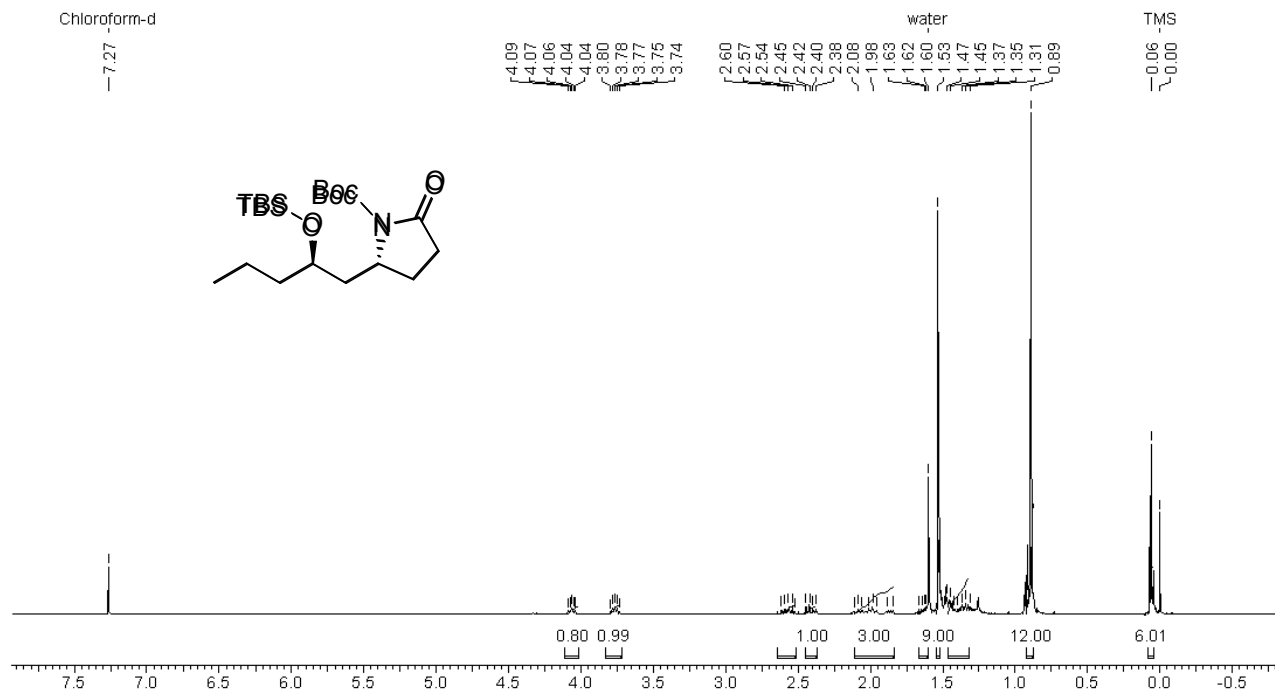
***tert*-Butyl (*R*)-2-((*R*)-2-((*tert*-butyldimethylsilyloxy)pentyl)piperidine-1-carboxylate (21):**



(-)-Halosaline (1):



***tert*-Butyl (R)-2-((R)-2-((*tert*-butyldimethylsilyl)oxy)pentyl)-5-oxopyrrolidine-1-carboxylate (17):**



***tert*-Butyl (*R*)-2-((*R*)-2-hydroxypentyl)pyrrolidine-1-carboxylate (15):**

