

Supporting Information

Stereoselective Synthesis of Conjugated α -Z/ γ -E – and α -Z/ γ -Z Dienoic acids. Kinetic torqueselectivity *versus* thermodynamic control

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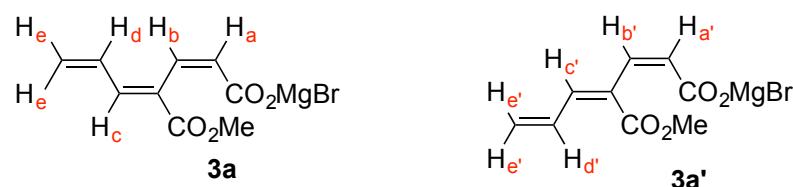
1- General Experimental Conditions

Unless otherwise noted, reactions were carried out under argon atmosphere with magnetic stirring in redistilled solvents when necessary. Reagents and chemicals were purchased from commercial sources and used as received. Merck 60F₂₅₄ silica gel was used for thin-layer chromatography (TLC) and Merck Geduran SI 60 Å silica gel 60 (40-63 µM) was used for flash column chromatography. IR spectra were recorded from a Bruker Tensor 27 ATR diamond PIKE spectrophotometer. NMR ¹H, ¹³C spectra were recorded at 400 and 100 MHz, respectively, using a Bruker AVANCE 400 spectrometer equipped with a BBFO probe. Some NMR ¹H, ¹³C spectra were recorded at 75 or 62.5 MHz using a Bruker AVANCE 300 or 250. Chemical shifts are reported in ppm, using, for ¹H and ¹³C, solvent residual peak as internal standard references. Coupling constants (*J*) are given in Hertz (Hz), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet). High resolution mass spectra (HRMS) were recorded on a LTQ-Orbitrap Mass Spectrometer [Thermo Scientific].

2- General Procedure for the Preparation of Carboxylates 3:

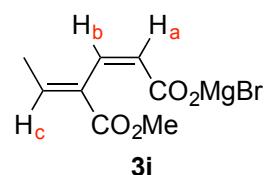
TMSCl (120 µL, 1 mmol) was added under argon to a solution of methylcoumalate (154 mg, 1 mmol) in dry THF (10 mL) at 0°C and the resulting mixture was stirred for 15 minutes. Then Grignard reagent (1.2 equiv.) was added dropwise, and the mixture was stirred for 15 minutes. The mixture was evaporated to get the anion.

4-Methoxycarbonyl-hepta-2,4,6-trienoic acid anion 3a/3a' (ratio 3a/3a': 60/40)



¹H NMR (400 MHz, MeOD) δ (ppm): 3.72 (3H, s, OCH₃) 5.46-5.65 (2H, m, He, He') 5.87 (0.4H, d, *J* = 12 Hz, Ha), 6.05 (0.6H, d, *J* = 12 Hz, Ha'), 6.47 (0.6H, d, *J* = 12 Hz, Hb), 6.58 (0.4H, d, *J* = 12 Hz, Hb'), 6.61-6.70 (1H, Hc, Hd'), 6.99-7.09 (1H, m, Hc', Hd).

4-Methoxycarbonyl-hexa-2,4-dienoic acid anion 3i



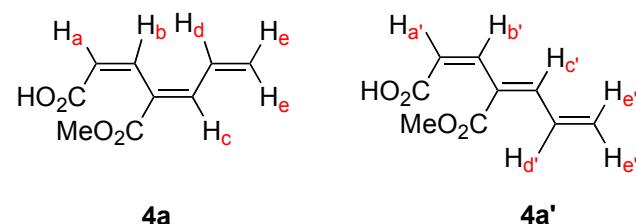
¹H NMR (250 MHz, MeOD) δ (ppm): 1.80 (3H, d, *J* = 7.2 Hz, Me), 3.35 (3H, s, CO₂CH₃), 6.02 (1H, d, *J* = 12 Hz, Ha) 6.43 (1H, dd, *J* = 12.0, 1.2 Hz, Hb), 6.75 (1H, qd, *J* = 7.2, 1.2 Hz, Hc).

3- General Procedure for the Preparation of Carboxylic Acids 4:

To a solution of methyl coumalate (2 mmol, 308 mg) in dry THF (20 mL) at 0 °C, under argon atmosphere, trimethylsilyl chloride (2 mmol, 0.25 mL) was added slowly. After 15 minutes stirring, Grignard reagent (1-2 equiv.) was added dropwise and the resulting solution was further stirred for 1-2 hr. Then, the reaction was quenched with saturated aq. sodium bicarbonate solution and washed with dichloromethane. The aqueous layer, containing the product as its sodium salt, was then acidified by adding dilute HCl and product was extracted with dichloromethane. The organic layers were pooled, dried over anhydrous MgSO₄ and evaporated to afford the pure product.

4-Allylidene-pent-2-enedioic acid 5-methyl ester 4a'/4a

Ratio 4a'/4a: 90:10

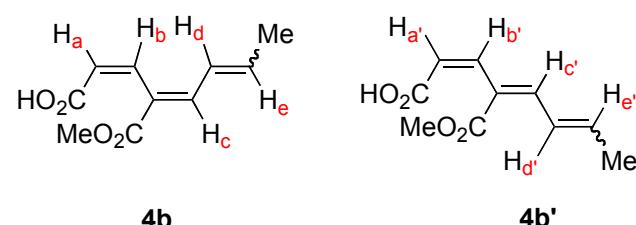


IR (KBr) ν_{max} : cm⁻¹: 2953, 1701, 1436, 1193. ¹H NMR (250 MHz, CDCl₃) δ (ppm): 3.77 (3H, s, OCH₃) 5.51-5.67 (2H, m, H_e & H_{e'}) 5.86 (0.9H, d, *J* = 12.2 Hz, H_{a'}) 6.06 (0.1H, d, *J* = 11.7 Hz, H_a) 6.50 (0.1H, d, *J* = 11.7 Hz, H_b) 6.60 (0.9 H, d, *J* = 12.2 Hz, H_{b'}) 6.64 (0.9H, d, *J* = 6.2 Hz, H_{d'}) 6.85 (0.1H, dd, *J* = 12.0, 7, 1 Hz, H_c) 7.02-7.23 (1H, m, H_{c'} & H_d) 10.43 (1H, br. s, CO₂H). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 51.8 (OCH₃) 52.2 (OCH₃) 119.7 (CH) 123.6 (CH) 127.1 (CH₂) 127.2 (CH₂) 128.0 (C(CO₂Me)) 129.1 (C(CO₂Me)) 131.8 (CH) 132.9 (CH) 138.7 (CH)

141.0 (CH) 142.7 (CH) 144.6 (CH) 166.0 (CO) 166.70 (CO) 170.3 (CO) 170.7 (CO). HRMS (ES+): for ($M^+ + Na$) $C_9H_{10}O_4Na$, calcd. 205.04713, found 205.04712.

4-Propenylidene-pent-2-enedioic acid 5-methyl ester 4b/4b'

Ratio **4a'(2 diastereoisomers)/4a (2 diastereoisomers)**: 50:50



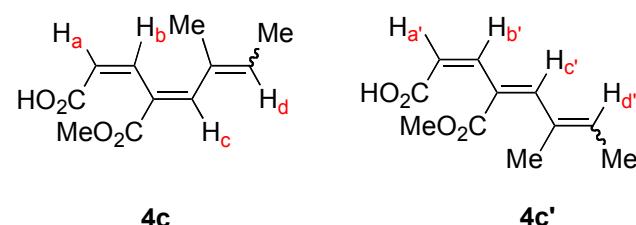
4b

4b'

IR (KBr) ν_{max} : cm⁻¹: 2952, 1692, 1630, 1435, 1235. ¹H NMR (250 MHz, CDCl₃) δ (ppm): 1.75 (3H, (4X) m, CH₃), 3.72 (0.75H, s, OCH₃), 3.74 (2.25H, s, OCH₃), 5.83 (0.5H, dd, J = 10.0, 7.5 Hz CH) 6.26-6.01 (1.75H, m, CH), 6.73-6.63 (2H, m, CH) 6.87 (0.25H, d, J = 11.7 Hz, H_b), 7.03 (0.25H, d, J = 10.0, Hz, CH), 7.57 (0.25H, d, J = 10.0, Hz, CH). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 13.72 (CH₃), 14.06 (CH₃), 18.97 (CH₃), 19.01 (CH₃), 51.48 (OCH₃), 51.55 (OCH₃), 51.87 (OCH₃), 51.94 (OCH₃), 118.41 (CH), 118.69 (CH), 122.92 (CH), 123.25 (CH), 124.29 (CH), 125.29 (CH), 126.70 (C(CO₂Me)), 126.96 (CH), 128.00 (C(CO₂Me)), 128.00 (C(CO₂Me)), 129.1 (C(CO₂Me)) 128.16 (CH), 135.28 (CH), 137.41 (CH), 138.56 (CH), 138.58 (CH), 138.71 (CH), 141.38 (CH), 141.43 (CH), 142.00 (CH), 142.99 (CH), 143.03 (CH), 145.30 (CH), 170.67 (CO), 170.74 (CO), 171.01 (CO), 171.10 (CO). HRMS (ES+): for ($M^+ + Na$) $C_{10}H_{12}O_4Na$, calcd. 219.06278, found 219.06283.

4-Prop-2'-methylidene-pent-2-enedioic acid 5-methyl ester 4c/4c'

Ratio **4c (2 diastereoisomers)/4c'(2 diastereoisomers)**: 70:30



4c

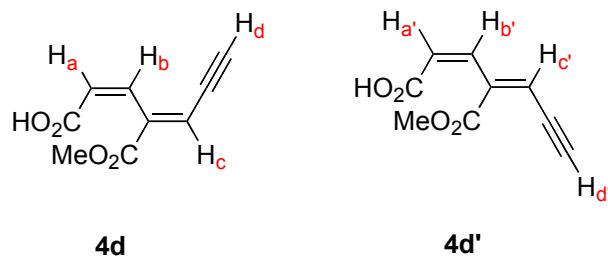
4c'

IR (KBr) ν_{max} : cm⁻¹: 2954, 1707, 1625, 1436, 1259, 1242, 1199. ¹H NMR (400 MHz, CDCl₃) δ (ppm): 1.55 (t, J = 1.6 Hz, 0.9 H, CH₃), 1.59 (d, J = 7.2 Hz, 2.1 H, CH₃), 1.81-1.77 (m, 0.9 H,

CH₃), 1.83 (t, *J* = 1.2 Hz, 2.1 H, CH₃), 3.68 (s, 0.9 H, OCH₃), 3.71 (s, 2.1 H, OCH₃), 5.66 (dd, *J* = 6.0, 13.2 Hz, 0.7 H, H_d), 5.86-5.78 (m, 0.3 H, H_{d'}), 5.96 (dd, *J* = 1.2, 12.0 Hz, 1 H, 0.7 H_a & 0.3 H_{a'}), 6.75 (dd, *J* = 2.0, 12.0 Hz, 0.7 H, H_b), 6.89 (dd, *J* = 1.6, 11.6 Hz, 0.3 H, H_{b'}), 7.29 (s, 0.7 H, H_c), 7.12 (s, 0.3 H, H_{c'}). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 14.6 (CH₃), 15.1 (CH₃), 16.0 (CH₃), 23.0 (CH₃), 51.9 (OCH₃), 52.0 (OCH₃), 121.8 (CH), 131.7 (CH), 122.0 (CH), 125.4 (C), 128.0 (C), 131.5 (C), 133.0 (C), 136.1 (CH), 140.6 (CH), 140.8 (CH), 140.9 (CH), 145.8 (CH), 167.5 (CO), 167.6 (CO), 171.2 (CO), 171.3 (CO). HRMS (ES+): for (M⁺+Na) C₁₁H₁₄O₄Na, calcd. 233.0784, found 233.0785.

4-Prop-2-ynylidene-pent-2-enedioic acid 5-methyl ester 4d/4d'.

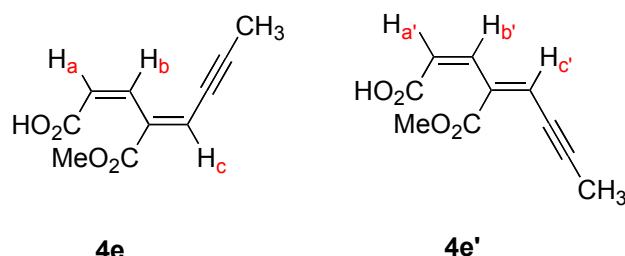
Ratio 4d'/4d' (Z,Z)/(Z,E): 50:50



IR (KBr) ν_{max}: cm⁻¹: 2959, 2162, 1685, 1433. ¹H NMR (300 MHz, CDCl₃) δ (ppm): 3.71 (0.5H, d, *J* = 2.7 Hz, H_d) 3.70 (0.5H, dd, *J* = 2.4, 0.6 Hz, H_d) 3.75 (1.5H, s, OCH₃) 3.79 (1.5H, s, OCH₃) 5.96 (0.5H, dd, *J* = 12.3, 0.6 Hz, H_a) 6.09 (0.5H, dd, *J* = 11.7, 0.9 Hz, H_a) 6.15-6.20 (0.5H, m, H_c) 6.61-6.73 (1H, m, 0.5 H_b & 0.5 H_c) 6.96 (0.5H, ddd, *J* = 12.0, 1.8, 0.6 Hz, H_b) 8.0-9.5 (1H, br. S, COOH). ¹³C NMR (75 MHz, CDCl₃) δ (ppm): 52.1 (OCH₃), 52.5 (OCH₃), 79.2 (C), 79.5 (C), 91.2 (C), 92.7 (C), 115.1 (CH), 121.1 (CH), 121.8 (CH), 121.9 (CH), 124.0 (CH), 138.0 (CH), 140.3 (C), 140.5 (C), 140.6 (CH), 165.0 (CO), 165.5 (CO), 170.3 (CO), 170.7 (CO). HRMS (ES+): for (M⁺+Na) C₉H₈O₄Na, calcd. 203.03147, found 203.03148.

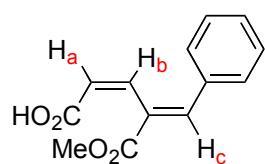
4-But-2-ynylidene-pent-2-enedioic acid 5-methyl ester 4e/4e'.

Ratio **4e'/4e** (Z,Z)/(Z,E): 70:30



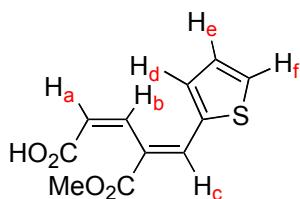
IR (KBr) ν_{\max} : cm⁻¹: 2950, 2213, 1723, 1685, 1609, 1436. ¹H NMR (400 MHz, CDCl₃) δ (ppm): 2.06 (0.9H, d, J = 2.8 Hz, CH₃) 2.08 (2.1 H, d, J = 2.8 Hz, CH₃) 3.72 (0.9H, s, OCH₃) 3.76 (2.1H, s, OCH₃) 5.86 (0.7H, d, J = 12.0 Hz, H_a) 6.03 (0.3H, dd, J = 12.0, 0.8 Hz, H_a) 6.18 (0.7H, d, J = 2.4 Hz, H_c) 6.68-6.54 (1H, m, 0.3 H_c & 0.7 H_b) 6.97 (0.3H, dd, J = 12.0, 0.8 Hz, H_b) 11.0 (1H, br. s, COOH). ¹³C NMR (75 MHz, CDCl₃) δ (ppm): 5.2 (CH₃), 5.4 (CH₃), 51.9 (OCH₃), 52.2 (OCH₃), 76.4 (C), 76.9 (C), 101.9 (C), 102.5 (C), 120.3 (CHA), 123.1 (CHA), 123.7 (CHc), 124.5 (CHc), 137.4 (CHb), 137.7 (C), 138.4 (C), 141.1 (CHb), 165.5 (CO), 166.1 (CO), 170.9 (CO), 171.4 (CO). HRMS (ES+): for (M⁺+Na) C₁₀H₁₀O₄Na, calcd. 217.0471, found 217.0469.

4-Benzylidene-pent-2-enedioic acid 5-methyl ester 4f



IR (KBr) ν_{\max} : cm⁻¹: 2951, 1708, 1629, 1434, 1256, 1175. ¹H NMR (400 MHz, CDCl₃) δ (ppm): 3.78 (s, 3 H, OCH₃), 6.17 (dd, J = 0.8, 11.6 Hz, 1 H, H_a), 6.89 (dd, J = 2.0, 11.6 Hz, 1 H, H_b), 7.45-7.35 (m, 5 H, Ph), 7.69 (d, J = 0.8 Hz, H_c). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 50.2 (OCH₃), 123.6 (CH), 128.4 (C), 128.5 (CH), 129.7 (CH), 130.6 (CH), 134.4 (C), 140.1 (CH), 141.4 (CH), 167.1 (CO), 171.2 (CO). HRMS (ES+): for (M⁺+Na) C₁₃H₁₂O₄Na, calcd. 255.0628, found 255.0627.

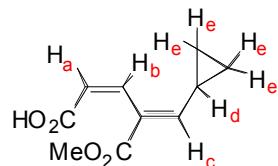
4-Thiophenyl-pent-2-enedioic acid 5-methyl ester 4g



4g

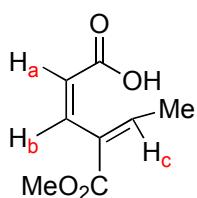
IR (KBr) ν_{max} : cm⁻¹: 2956, 1709, 1682, 1621, 1434, 1260, 1205. ¹H NMR (400 MHz, CDCl₃) δ (ppm): 3.75 (s, 3 H, OCH₃), 6.19 (dd, *J* = 0.8, 11.6 Hz, 1 H, H_a), 7.04 (dd, *J* = 2.0, 11.6 Hz, 1 H, H_b), 7.11 (dd, *J* = 3.6, 5.2 Hz, 1 H, H_e), 7.30 (d, *J* = 4.0 Hz, 1 H, H_d), 7.53 (d, *J* = 5.2 Hz, 1 H, H_f), 7.83 (dd, *J* = 0.8, 2.0 Hz, 1 H, H_c). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 52.2 (OCH₃), 124.7 (CH_a), 125.0 (CH_e), 127.9 (C), 131.4 (CH_f), 133.1 (CH_d), 133.2 (CH_c), 138.1 (C), 139.5 (CH_b), 166.7 (CO), 170.9 (CO). HRMS (ES+): for (M⁺+Na) C₁₁H₁₀O₄NaS, calcd. 261.0192, found 261.0189.

4-Cyclopropyl-pent-2-enedioic acid 5-methyl ester 4h



¹H NMR (300 MHz, CDCl₃) δ (ppm): 0.71 (m, 2 H, H_e), 1.00 (m, 2 H, H_e), 1.65-1.48 (m, 1 H, H_d), 3.70 (s, 3 H, OCH₃), 6.05 (d, *J* = 14.4 Hz, 1 H, H_a), 6.23 (d, *J* = 13.2 Hz, 1 H, H_c), 6.92 (dd, *J* = 1.5, 12.6 Hz, 1 H, H_b). ¹³C NMR (75 MHz, CDCl₃) δ (ppm): 9.25 (CH₂), 12.8 (CH), 51.8, (OCH₃), 122.5 (CH), 126.1 (C), 139.0 (CH), 151.7 (CH), 166.5 (CO), 171.0 (CO). HRMS (ES+): for (M⁺+Na) C₁₀H₁₂O₄Na, calcd. 219.06278, found 219.06282.

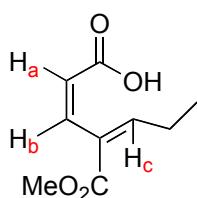
4-Ethylidene-pent-2-enedioic acid 5-methyl ester 4i



¹H NMR (250 MHz, CDCl₃) δ (ppm) : 1.82 (3H, d, *J* = 7.2 Hz, CH₃), 3.71 (3H, s, OCH₃), 6.02 (1H, d, *J* = 11.7 Hz, H_a), 6.71 (1H, d, *J* = 11.7 Hz, H_b), 6.92 (1H, q, *J* = 7.2 Hz, H_c), 9.08 (1H,

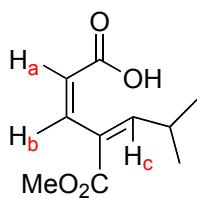
br. s, CO₂H). ¹³C NMR (75 MHz, CDCl₃) δ (ppm): 30.0 (CH₃), 51.8 (OCH₃), 122.8 (CH), 129.4 (C), 139.1 (CH), 142.3 (CH), 166.5 (CO), 172.1 (CO).

4-Ethyl-pent-2-enedioic acid 5-methyl ester 4j



IR (KBr) ν_{max}: cm⁻¹: 2954, 1708, 1432, 1349, 1258. ¹H NMR (250 MHz, CDCl₃) δ (ppm): 1.04 (t, *J* = 7.5 Hz, 3 H, CH₃), 2.17 (quintet, *J* = 7.5 Hz, 2 H, CH₂), 3.70 (s, 3 H, OCH₃), 6.01 (d, *J* = 12.5 Hz, 1 H, H_a), 6.85-6.75 (m, 2 H, H_b & H_c).

4-Isobutylidene-pent-2-enedioic acid 5-methyl ester 4k



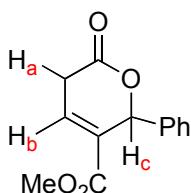
¹H NMR (250 MHz, CDCl₃) δ (ppm): 1.01 (3H, d, *J* = 6.8 Hz, CH(CH₃)₂) 1.20 (3H, d, *J* = 6.4 Hz, CH(CH₃)₂) 2.52-2.62 (1H, m, CH(CH₃)₂) 3.82 (3H, s, CO₂CH₃) 6.02 (1H, d, *J* = 12.4 Hz, Ha) 6.63 (1H, d, *J* = 10.4 Hz, Hc) 6.79 (1H, dd, *J* = 11.6, 2.0 Hz, Cb). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 17.50 (CH₃), 20.13(CH₃), 45.13 (CH), 52.24 (CO₂CH₃), 122.92 (CH), 127.42 (C), 138.87 (CH), 151.91 (CH), 165.77 (CO₂H), 169.15 (CO₂CH₃).

4- General Procedure for the Preparation of Unsaturated Lactones 5:

To a solution of methyl coumalate (2 mmol, 308 mg) in dry THF (20 mL) at 0 °C, under argon atmosphere, trimethylsilyl chloride (2 mmol, 0.25 mL) was added slowly. After 15 minutes stirring, Grignard reagent (1-2 equiv.) was added dropwise and the resulting solution was further stirred for 1-2 hr. Then the reaction was quenched with saturated aq. sodium bicarbonate solution and extracted twice with dichloromethane. The organic layers, were dried

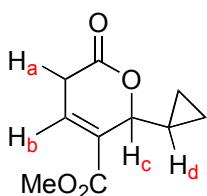
over anhydrous MgSO₄ and avaporated under reduced pressure. The residue was chromatographed on silica gel (Ethyl acetate / cyclohexane).

2-Phenyl-6-oxo-5,6-dihydro-2H-pyran-3-carboxylic acid methyl ester 5f



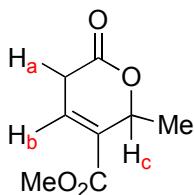
¹H NMR (250 MHz, CDCl₃) δ (ppm): 3.19 (td, *J* = 2.5, 22.2 Hz, H_a), 3.37 (ddd, *J* = 1.5, 5.5, 22.0 Hz, H_a), 3.77 (s, 3 H, OCH₃), 6.38 (t, *J* = 1.5 Hz, 1 H, H_c), 7.21 (dd, *J* = 2.7, 5.7 Hz, 1 H, H_b), 7.43-7.35 (m, 5 H, Ph). ¹³C NMR (62.5 MHz, CDCl₃) δ (ppm): 30.7 (CH₂), 52.2 (OCH₃), 79.8 (CH), 126.8 (CH), 128.8 (CH), 129.0 (CH), 129.7 (C), 134.1 (CH), 136.9 (C), 166.3 (CO), 167.4 (CO).

2-Cyclopropyl-6-oxo-5,6-dihydro-2H-pyran-3-carboxylic acid methyl ester 5h



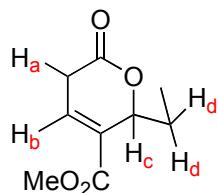
IR (KBr) ν_{max}: cm⁻¹: 1707, 1434, 1340, 1258. ¹H NMR (300 MHz, CDCl₃) δ (ppm): 0.68-0.48 (m, 4 H, CH₂), 1.14-1.01 (m, 1 H, H_d), 1.78-1.76 (m, 4 H, CH₂), 3.28 (m, H_a & H_{a'}), 3.81 (s, 3 H, OCH₃), 4.84 (td, *J* = 1.8, 9.9 Hz, 1 H, H_c), 7.01 (dd, *J* = 4.2, 5.7 Hz, 1 H, H_b). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 3.0 (CH₂), 3.2 (CH₂), 16.0 (CH), 30.7 (CH_a), 52.2 (OCH₃), 82.0 (CH_c), 131.0 (C), 133.1 (CH_b), 164.0 (CO), 168.3 (CO).

2-Methyl-6-oxo-5,6-dihydro-2H-pyran-3-carboxylic acid methyl ester 5i



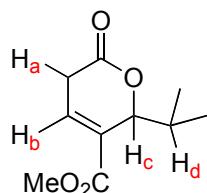
¹H NMR (400 MHz, CDCl₃) δ (ppm): 1.43 (3H, d, *J* = 6.4 Hz, CH₃), 3.10-3.29 (2H, m, H_a), 3.71 (3H, s, OCH₃) 5.31 (1H, qt, *J* = 6.8, 2.0 Hz, H_c) 6.88 (1H, dd, *J* = 5.0, 2.8 Hz, H_b). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 21.68 (CH₃), 30.02 (CH_a), 52.10 (OCH₃), 75.62 (CH_c), 131.60 (C(CO₂CH₃)), 132.56 (CH_b), 163.64 (CO), 167.53 (CO).

2-Ethyl-6-oxo-5,6-dihydro-2H-pyran-3-carboxylic acid methyl ester 5j

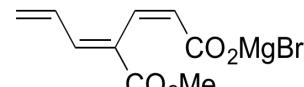


IR (KBr) ν_{max}: cm⁻¹: 2954, 1708, 1671, 1432, 1349, 1258, 1216, 1183. ¹H NMR (400 MHz, CDCl₃) δ (ppm): 0.10 (t, *J* = 7.6 Hz, 3 H, CH₃), 1.74 (m, 1 H, H_d), 1.96 (m, 1 H, H_d), 3.19 (td, *J* = 2.8, 22.8 Hz, H_a), 3.28 (ddd, *J* = 1.6, 5.2, 22.4 Hz, H_a), 3.32 (s, 3 H, OCH₃), 5.32-5.26 (m, 1 H, H_c) 7.00 (dd, *J* = 2.8, 5.2 Hz, 1 H, H_b). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 8.8 (CH₃), 28.7 (CH₂), 30.3 (CH_a), 52.2 (OCH₃), 80.3 (CH_c), 130.2 (C), 133.1 (CH_b), 163.8 (CO), 167.7 (CO). HRMS (ES+): for (M⁺+Na) C₉H₁₂O₄Na, calcd. 207.0628, found 207.0626.

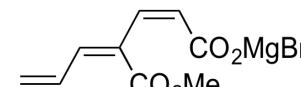
2-Isopropyl-6-oxo-5,6-dihydro-2H-pyran-3-carboxylic acid methyl ester 5k



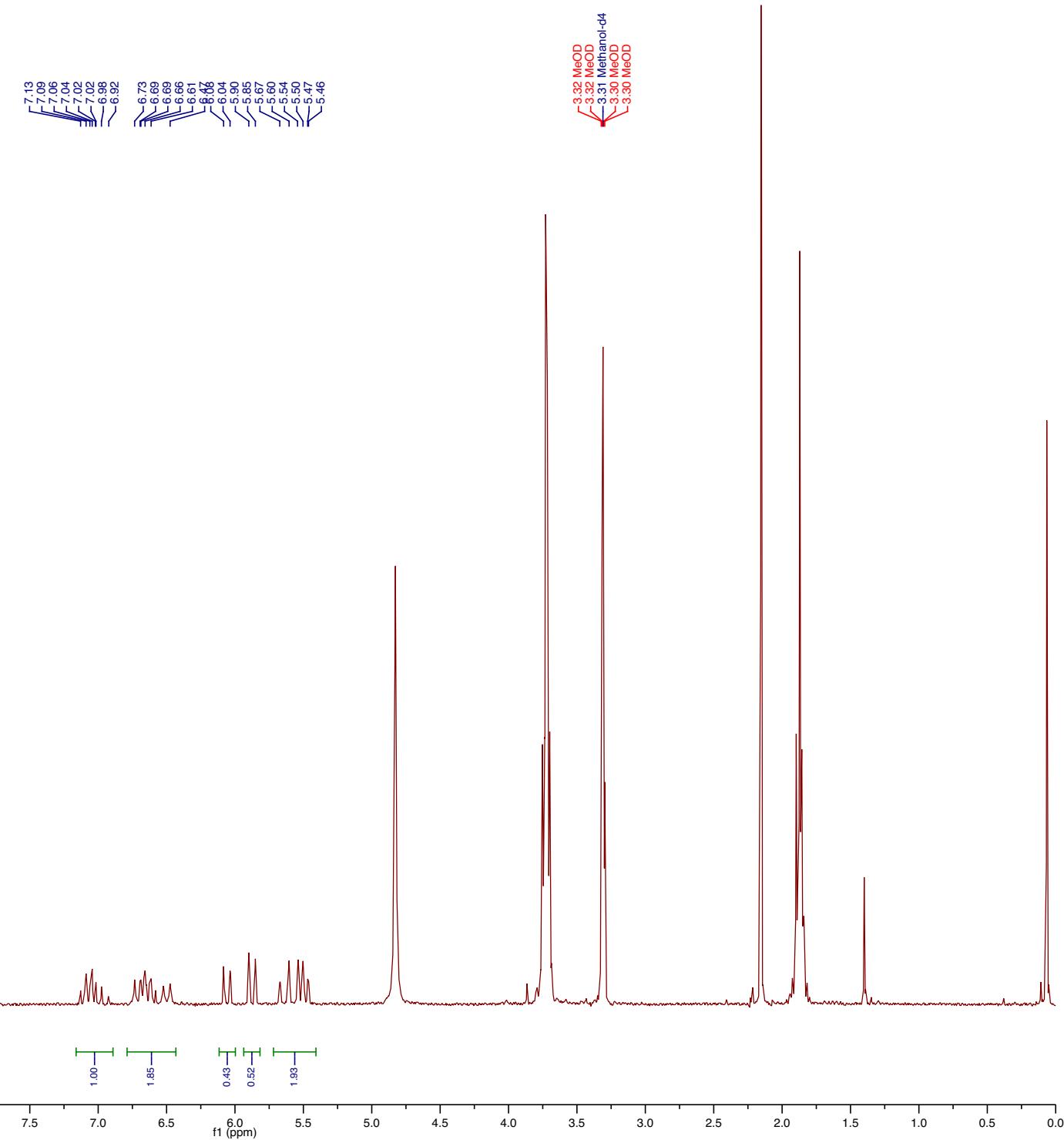
¹H NMR (400 MHz, CDCl₃) δ (ppm): 0.87 (3H, d, *J* = 6.8 Hz, CH(CH₃)₂), 1.09 (3H, d, *J* = 6.8 Hz, CH(CH₃)₂) 2.09-2.17 (1H, m, CH(CH₃)₂) 3.14-3.29 (1H, m, H_a) 3.79 (3H, s, OCH₃) 5.21 (1H, dt, *J* = 3.6, 2.4 Hz, H_c) 7.02 (1H, dd, *J* = 4.8, 2.8 Hz, H_b). ¹³C NMR (100 MHz, CDCl₃) δ (ppm): 15.7 (CH(CH₃)₂), 19.2 (CH(CH₃)₂), 30.7 (CH_a), 34.0 (CH(CH₃)₂), 52.3 (OCH₃), 84.0 (CH_c), 129.8 (C(CO₂CH₃)), 133.5 (CH_b), 164.3 (CO₂), 167.9 (CO₂). HRMS (ES+): for (M⁺+Na) C₁₀H₁₄O₄Na, calcd. 221.0777, found 221.0784.



3a



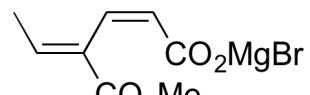
3a'



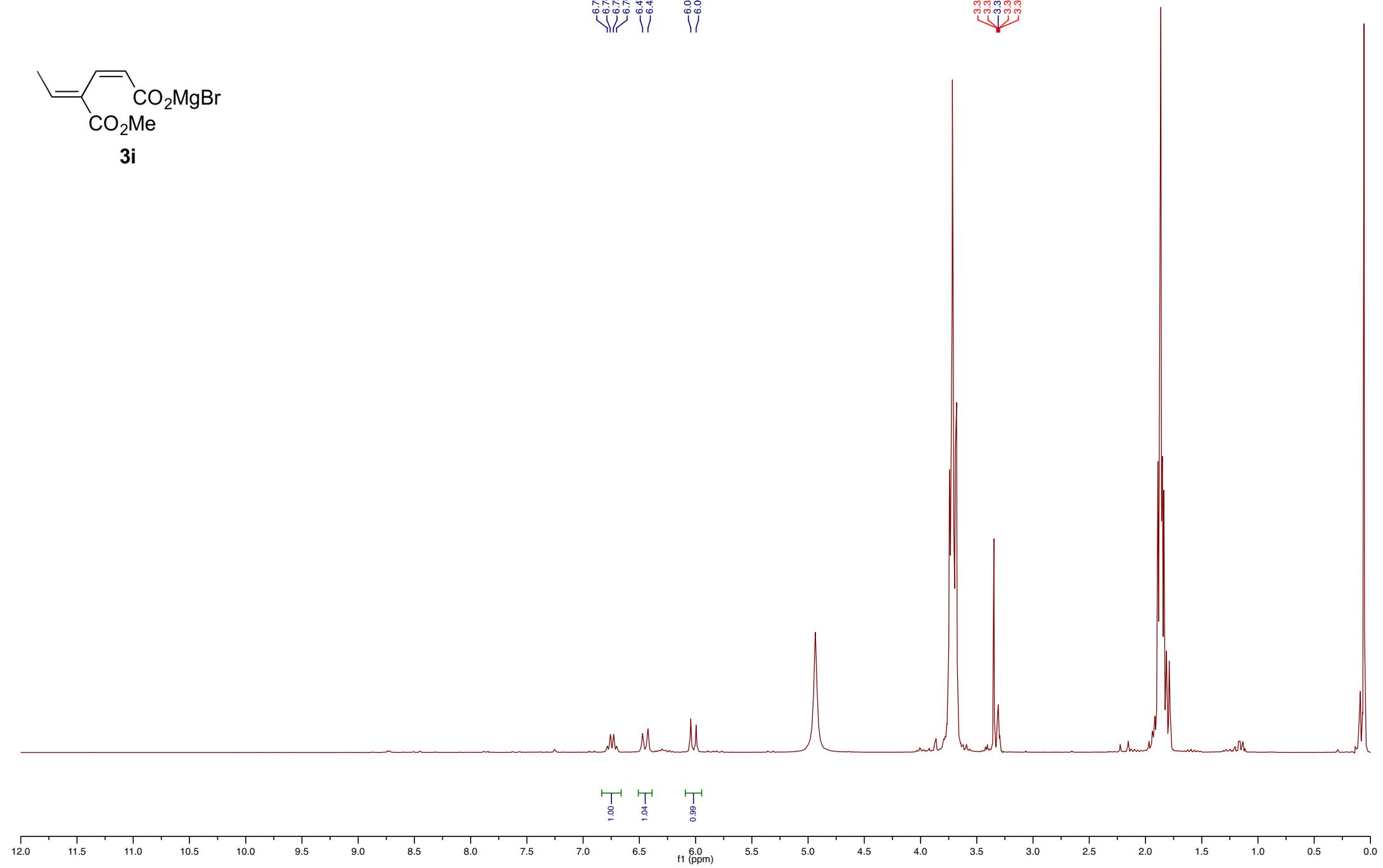
LDOB-16-25j/1

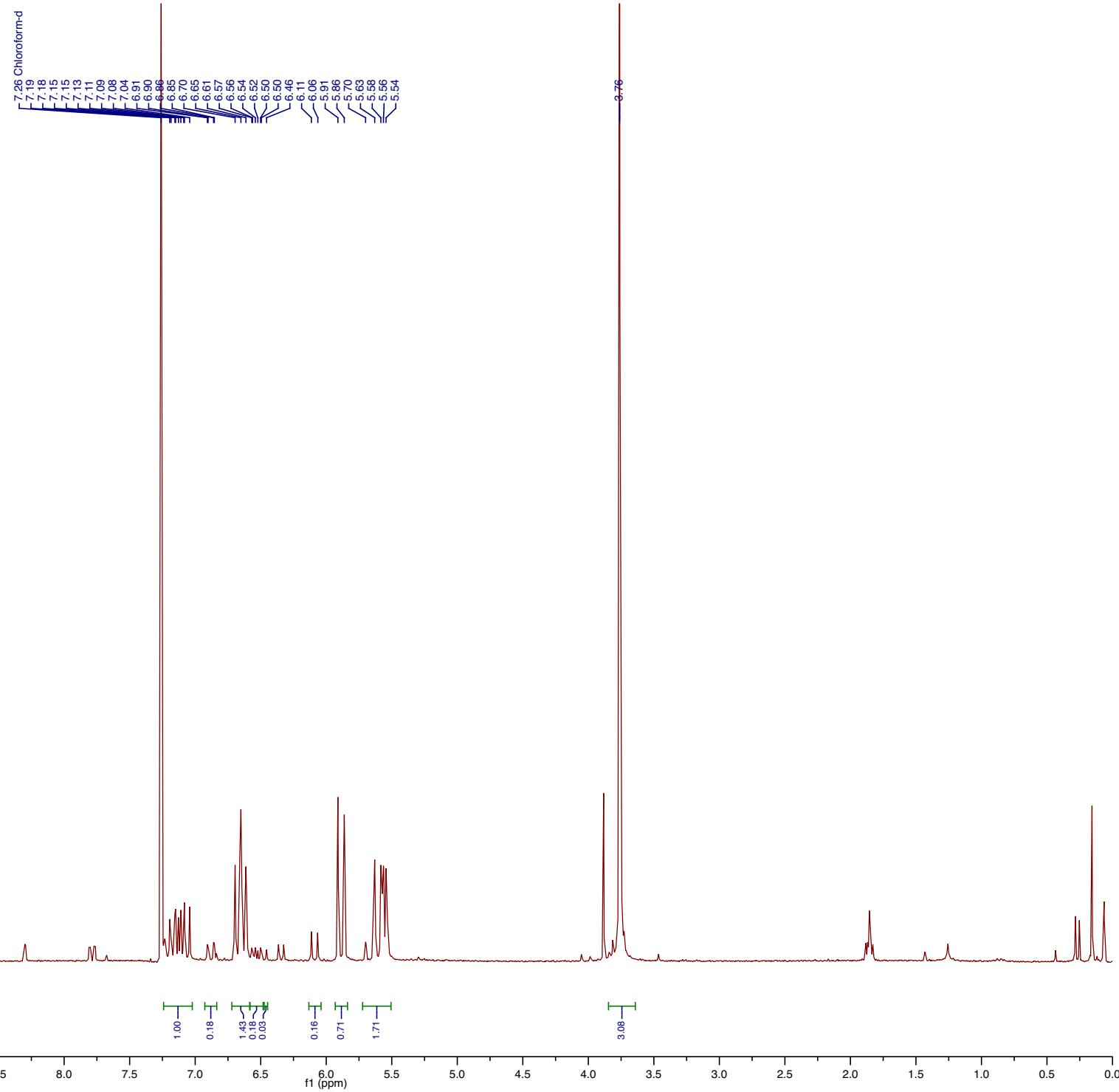
Spectre 1H routine en 8 Scan

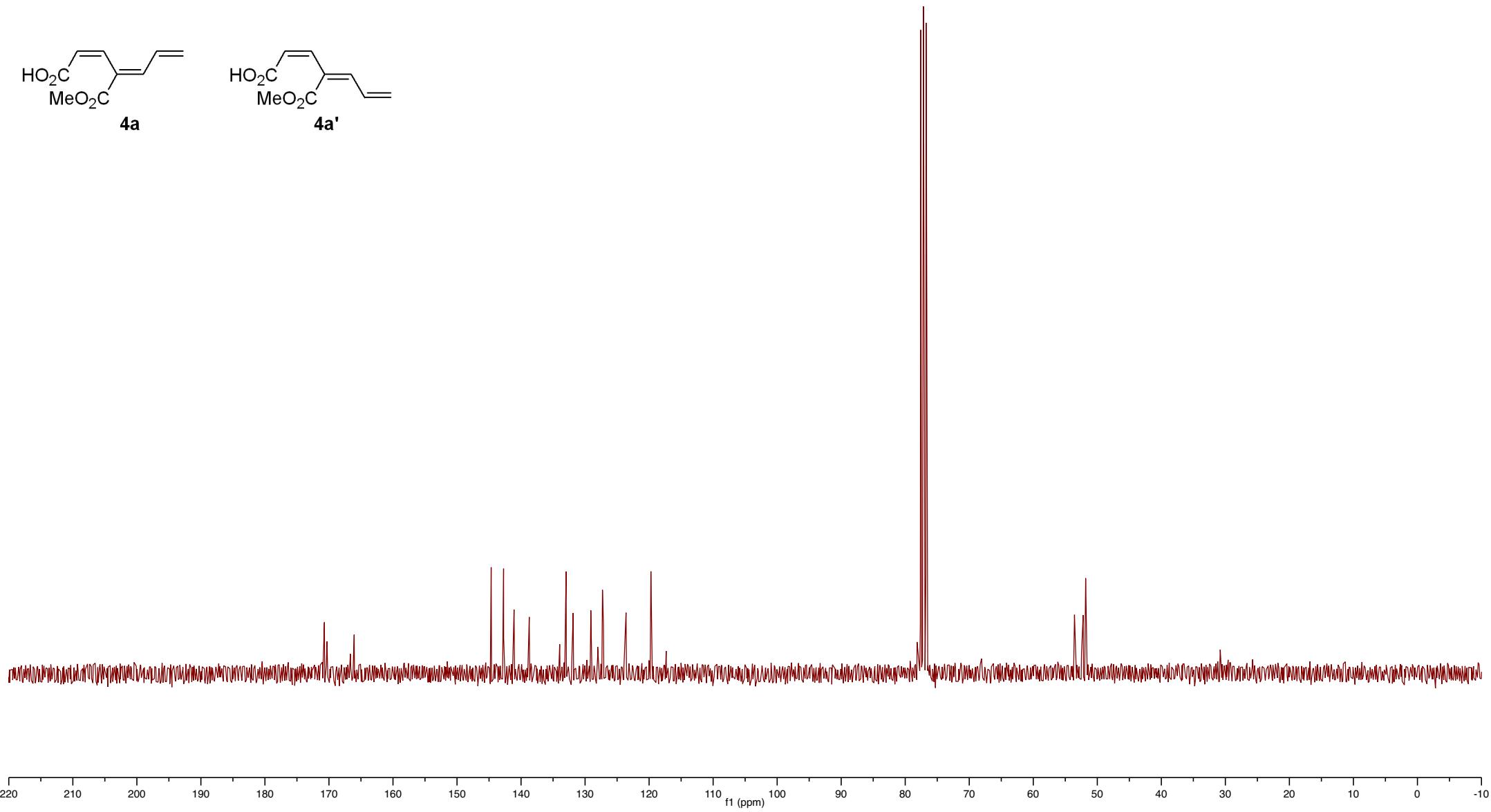
Sel dans MeOD

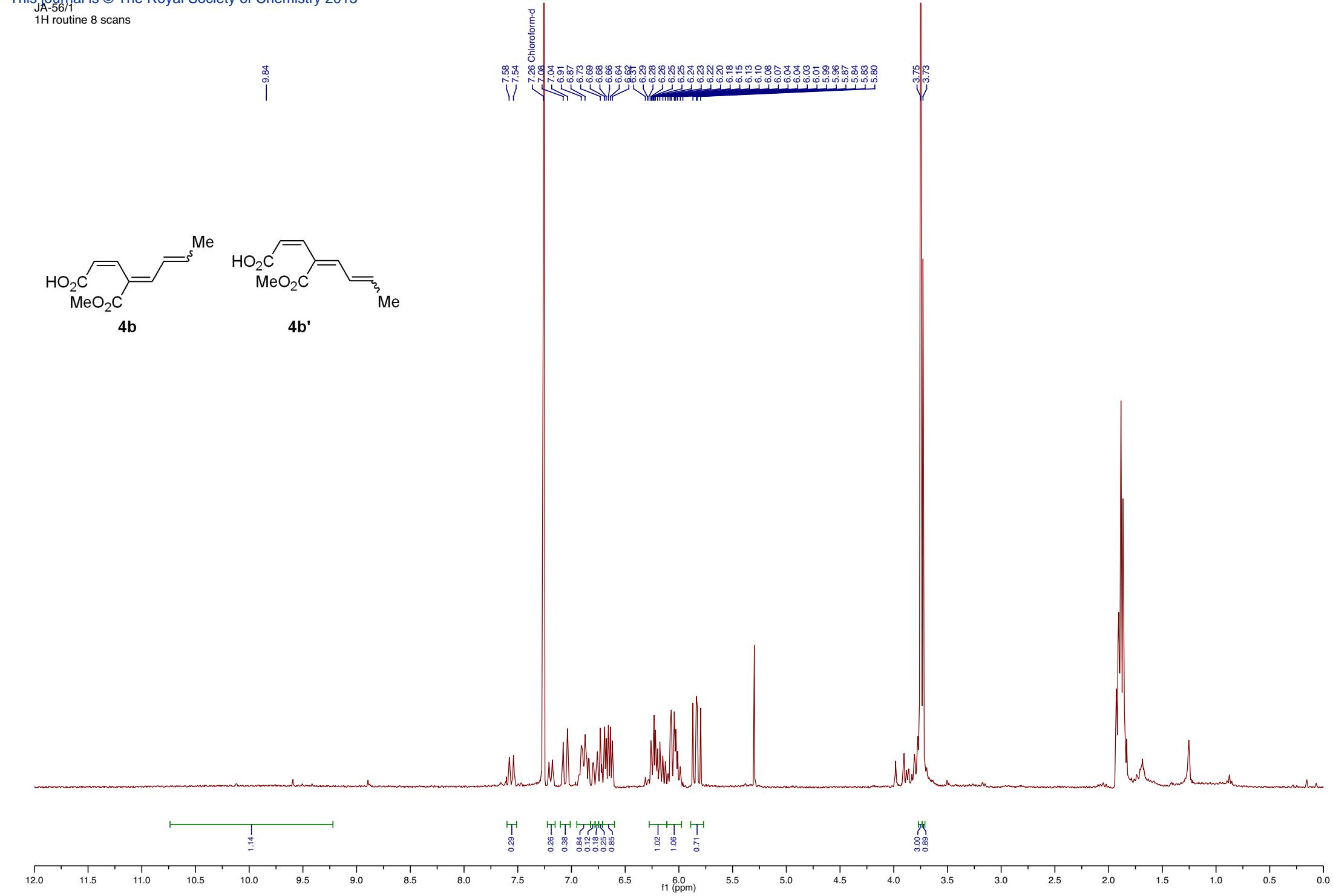
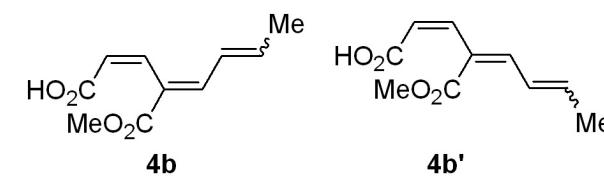


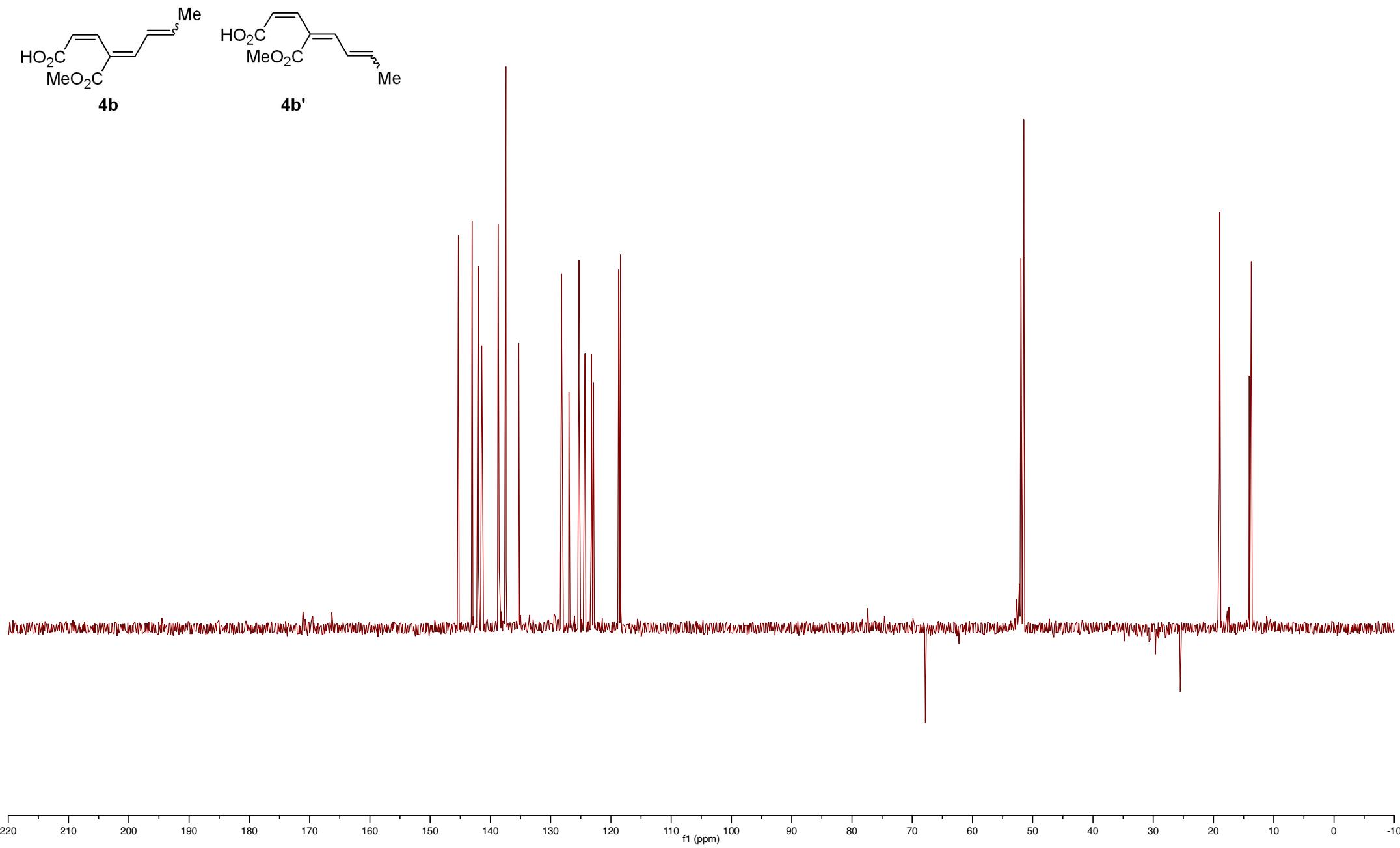
3i

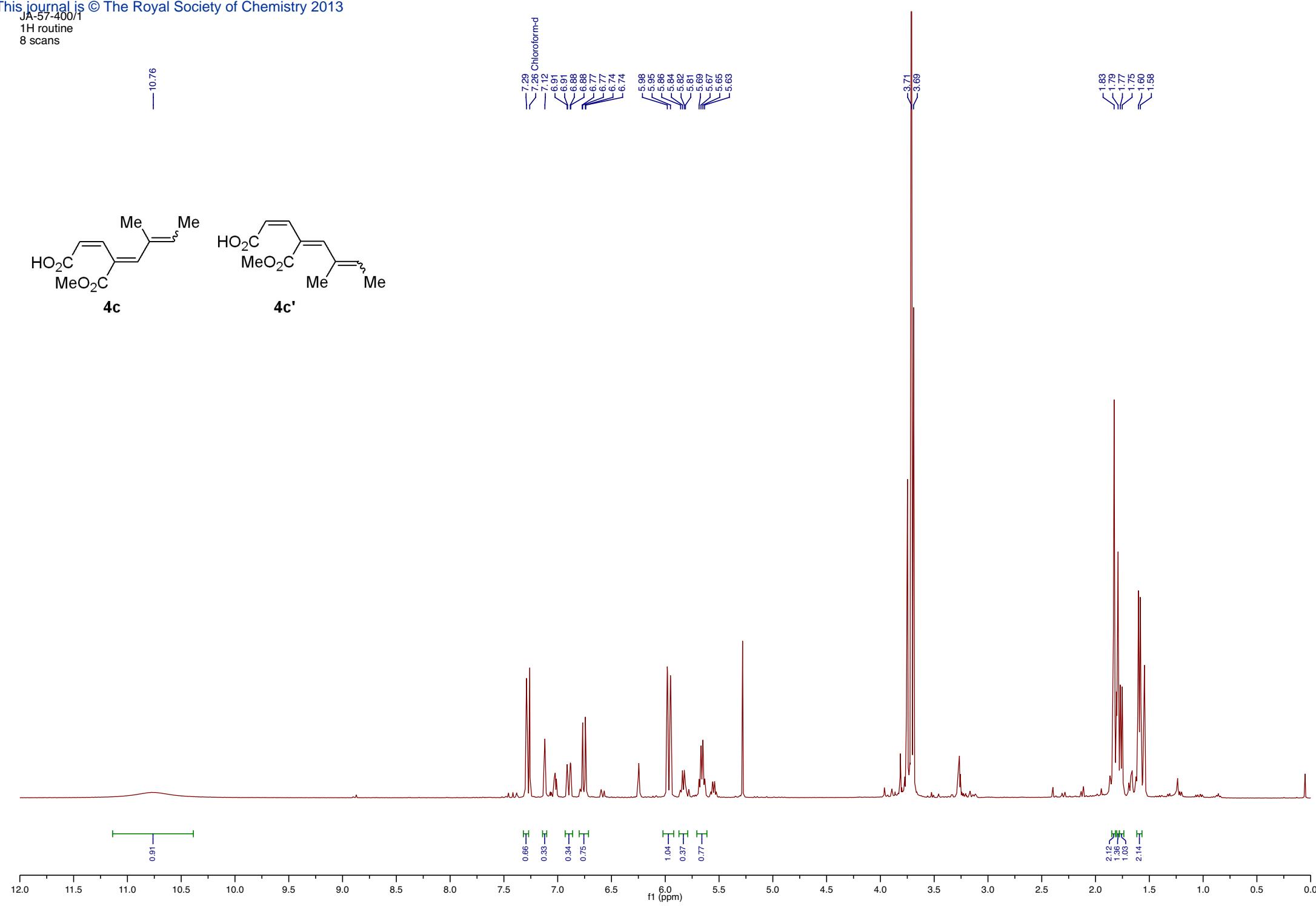




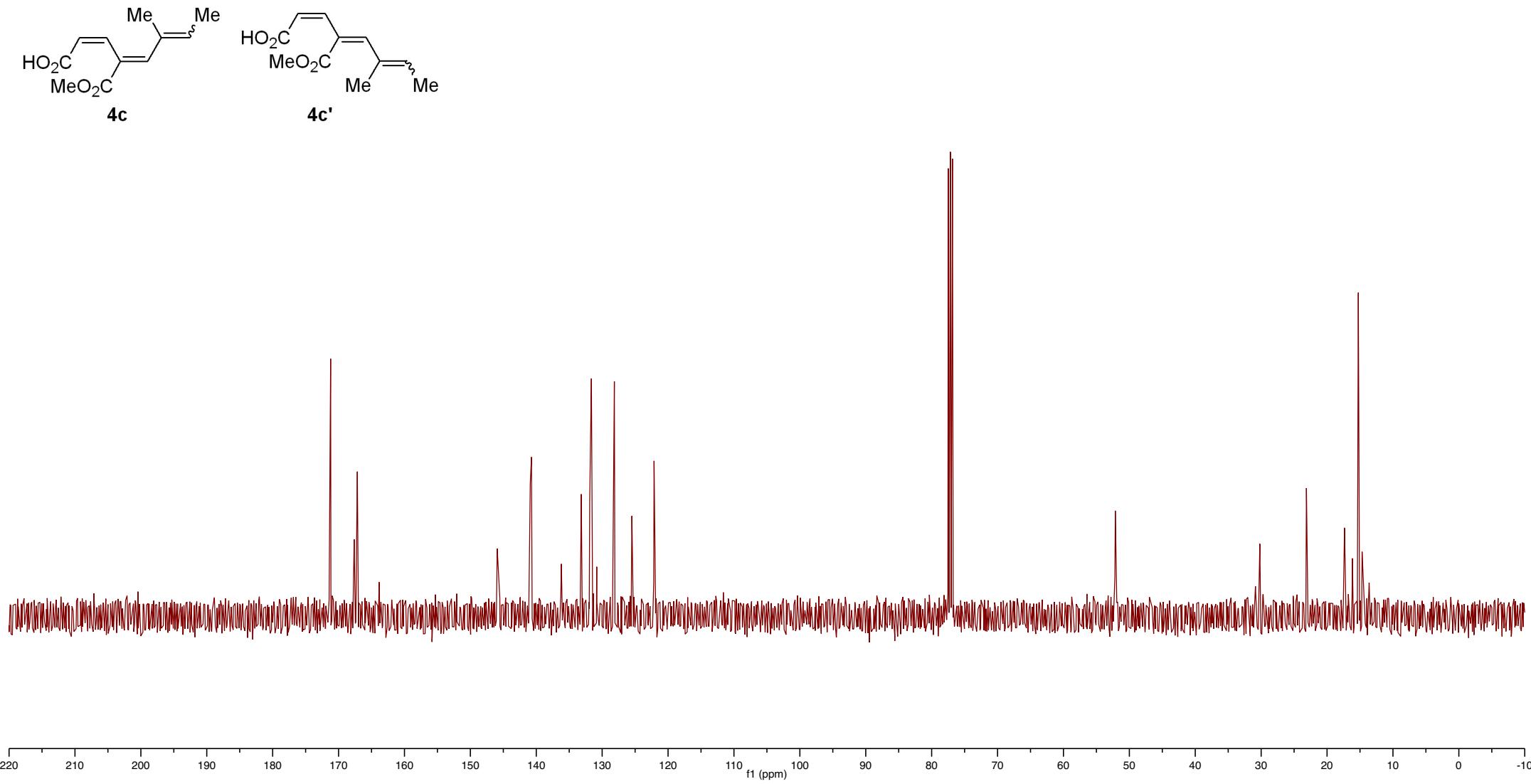


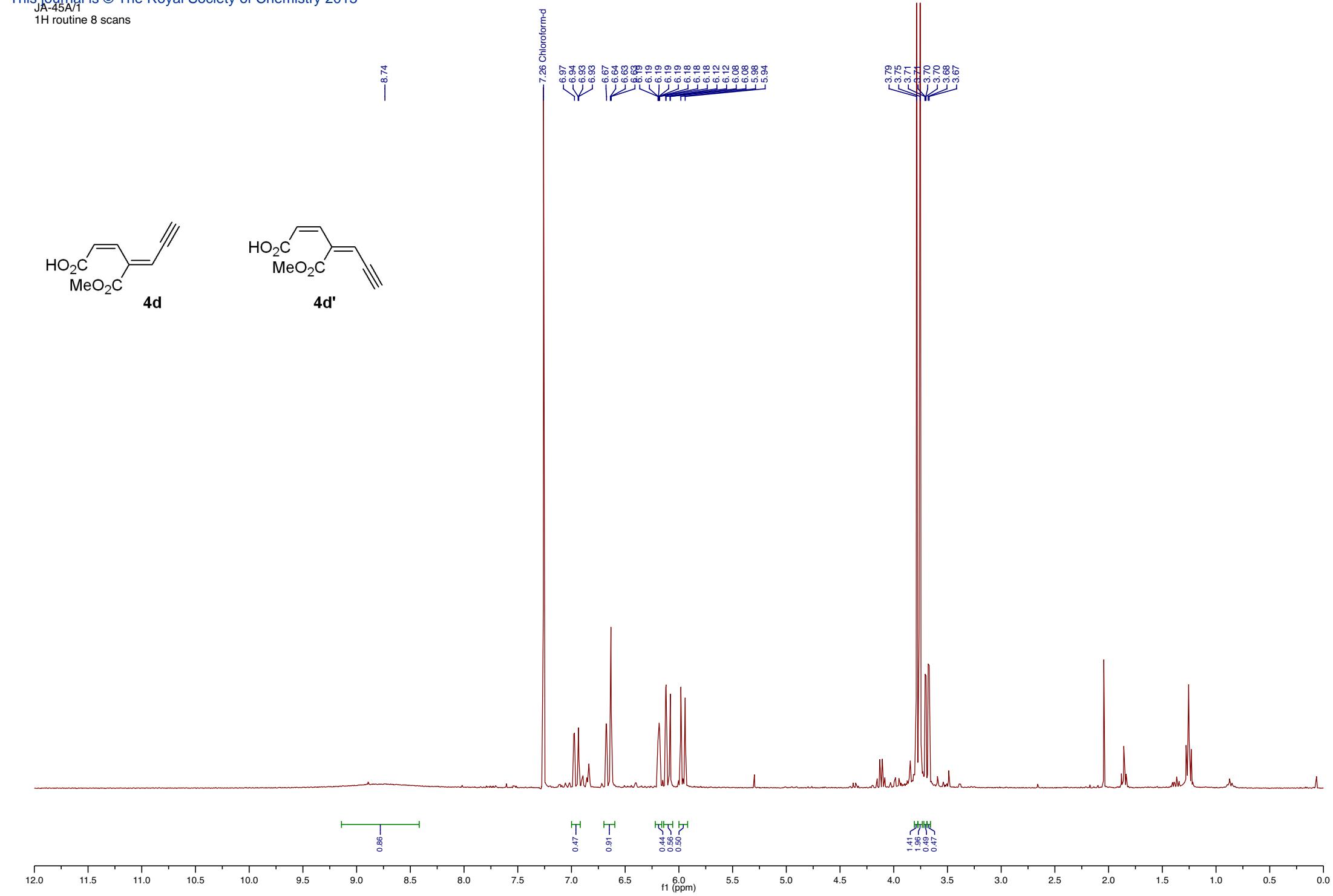
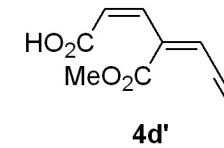
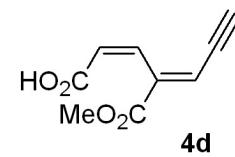


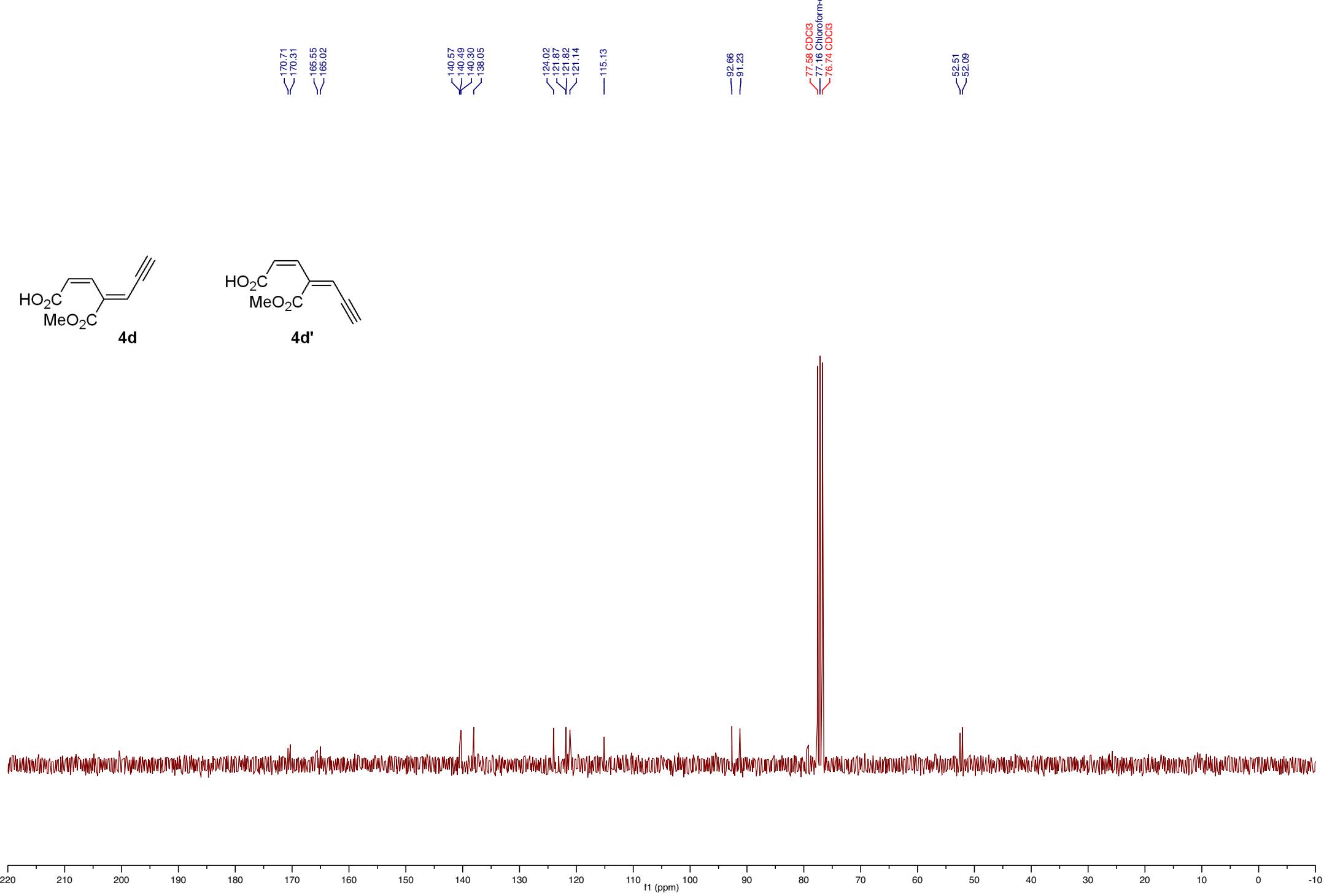


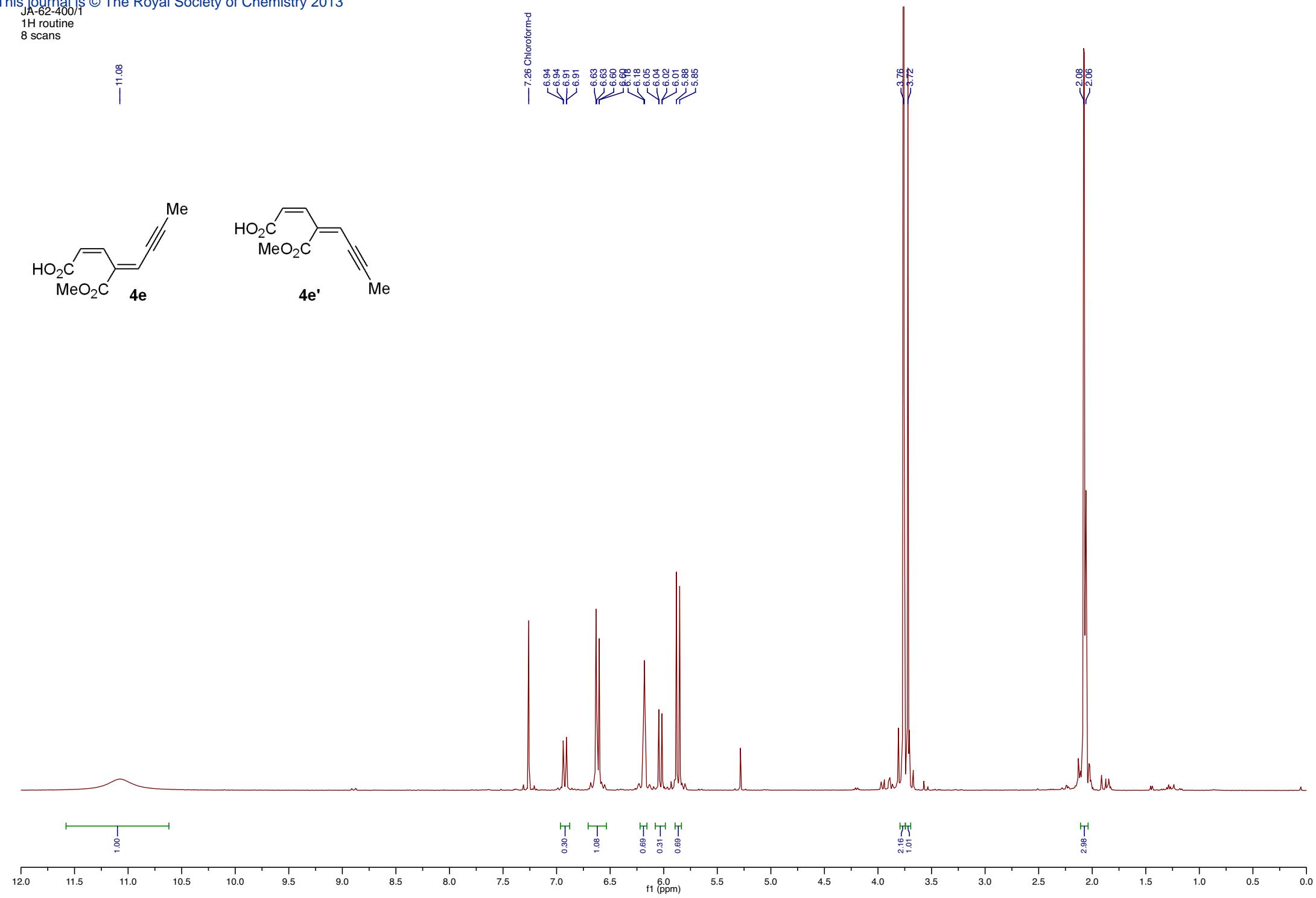
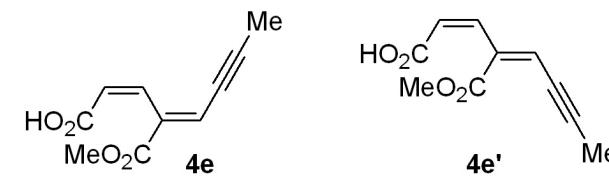


JA-57-400/3
13C routine
decouple 1H
32 scans

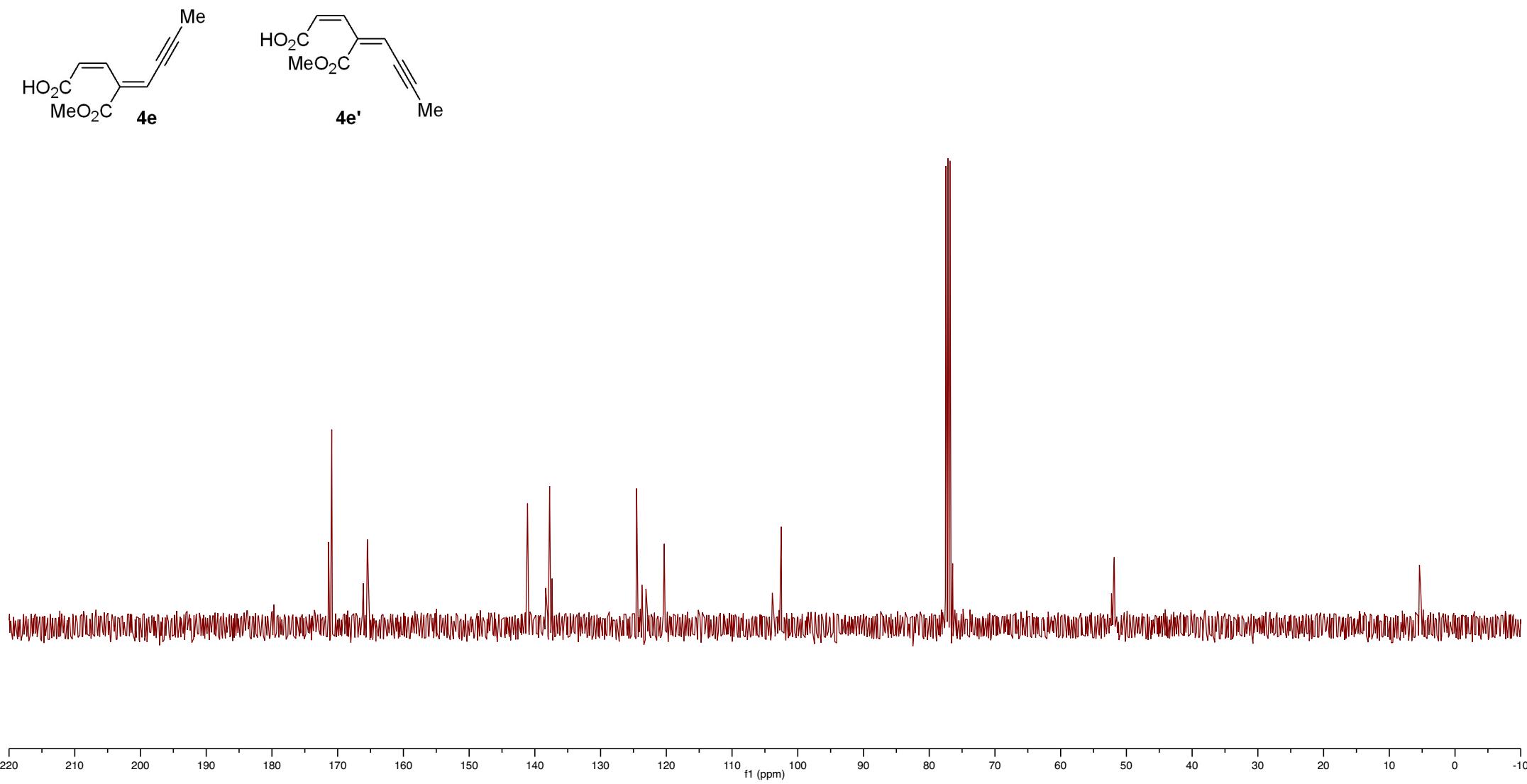


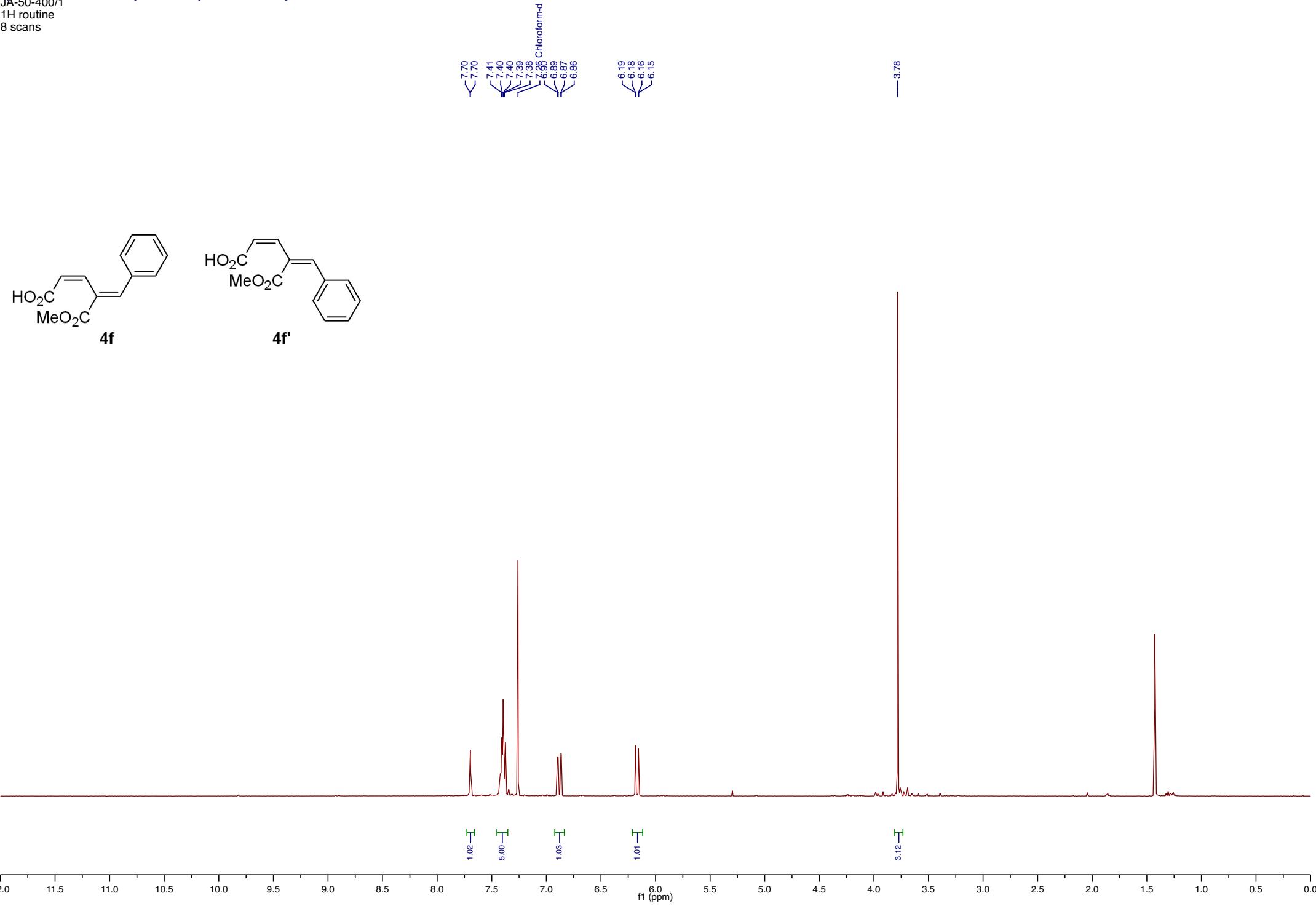


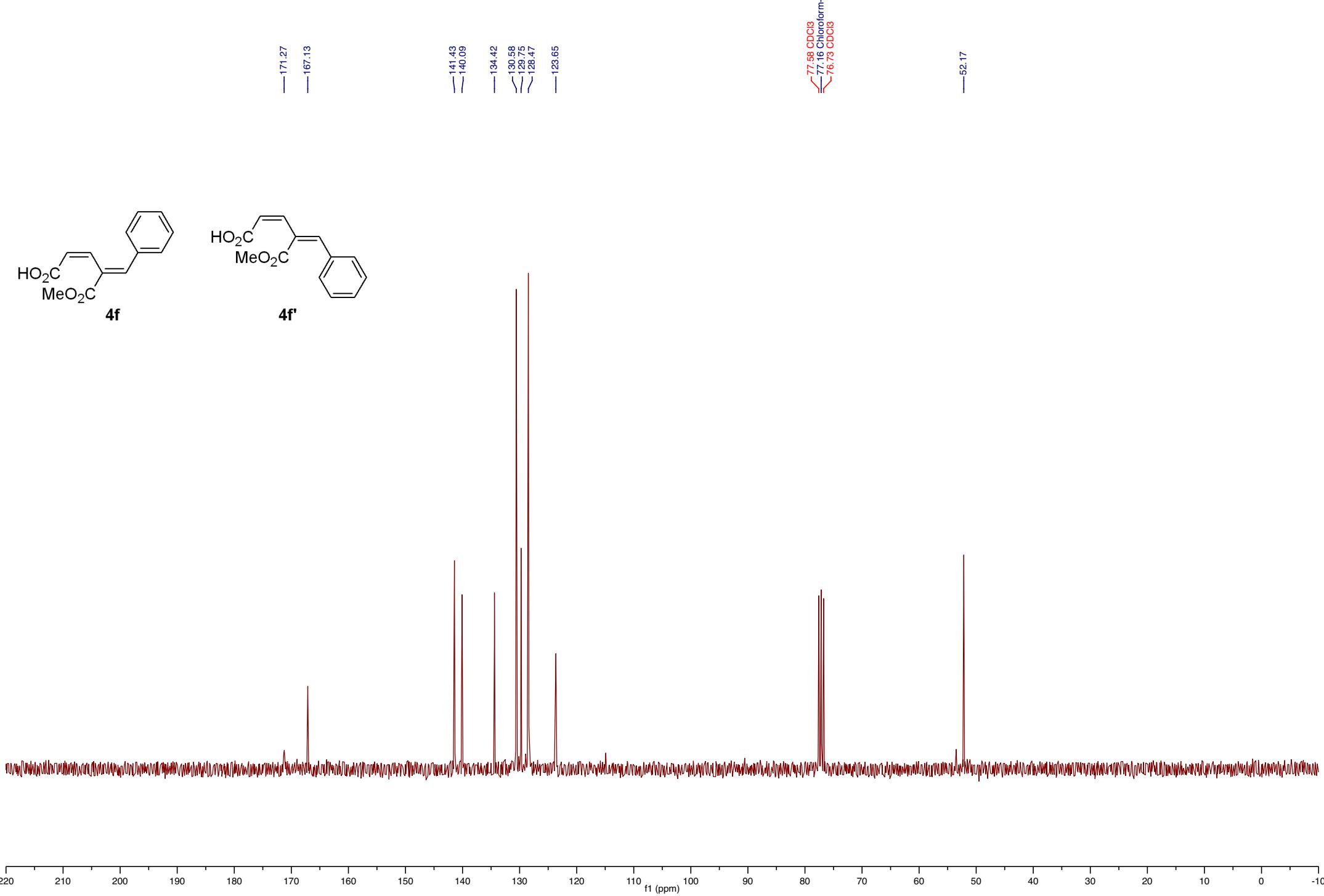


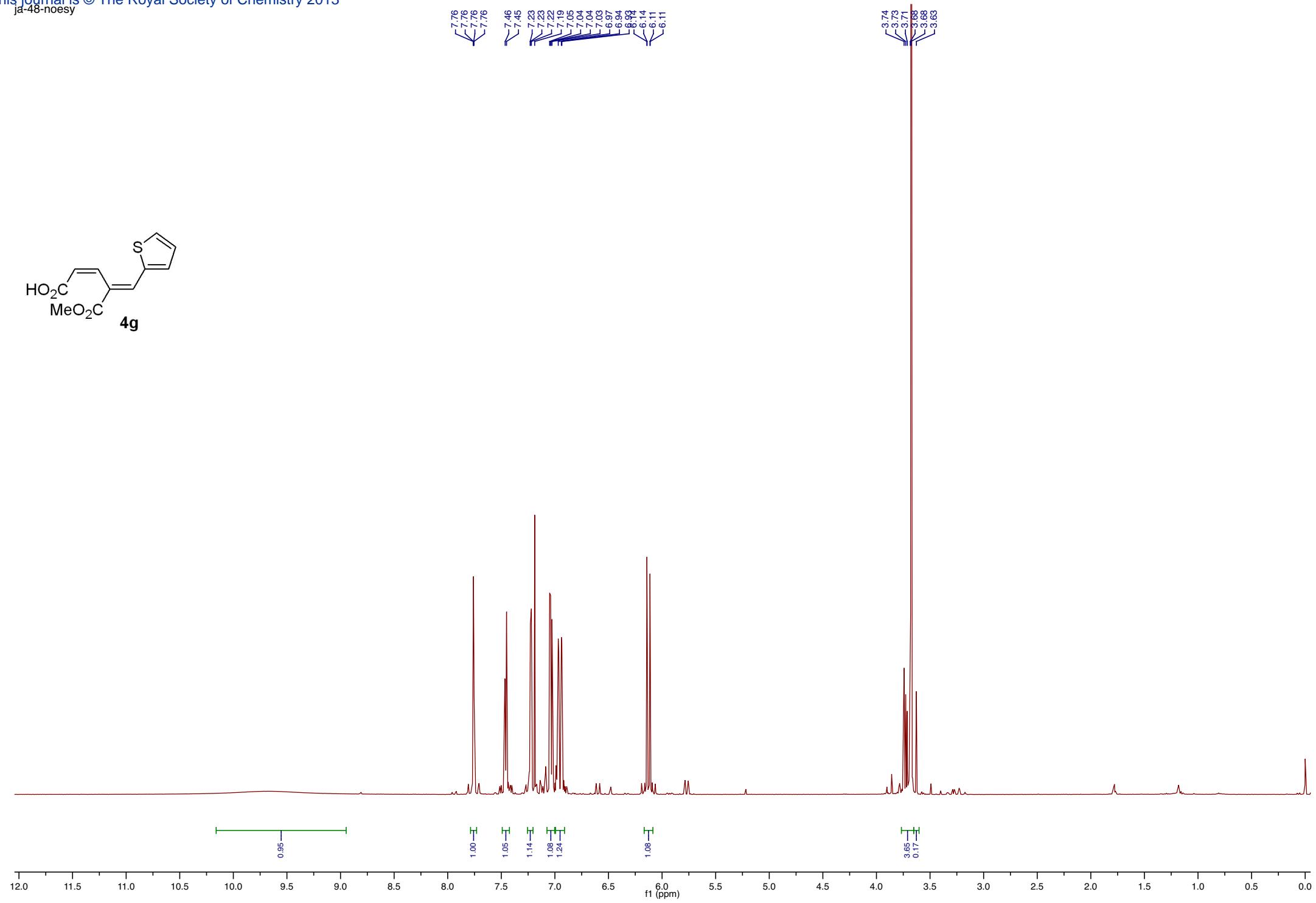
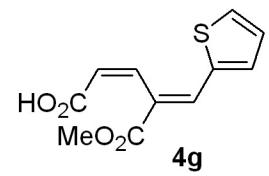


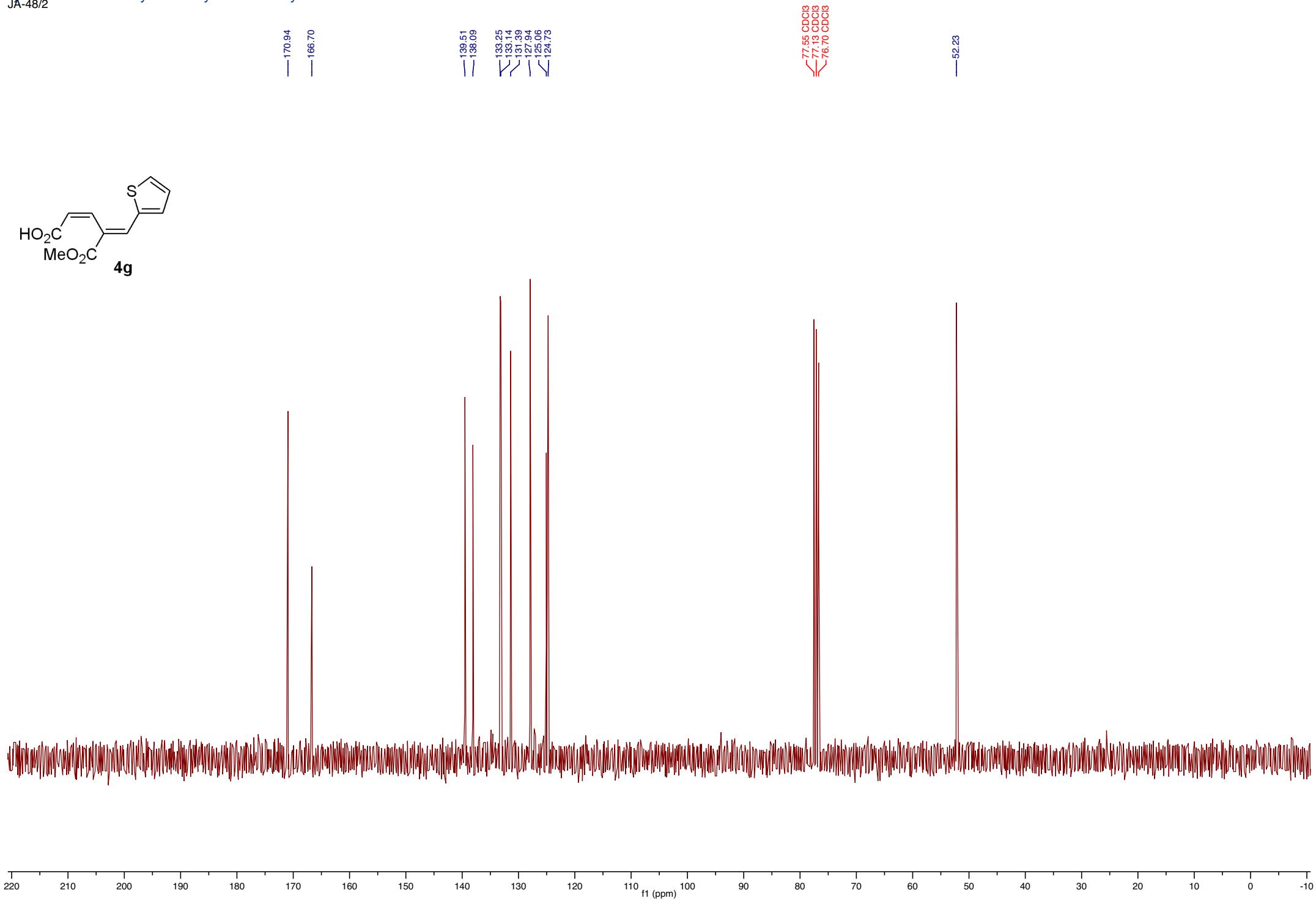
JA-62-400/2
13C routine
decouple 1H
32 scans

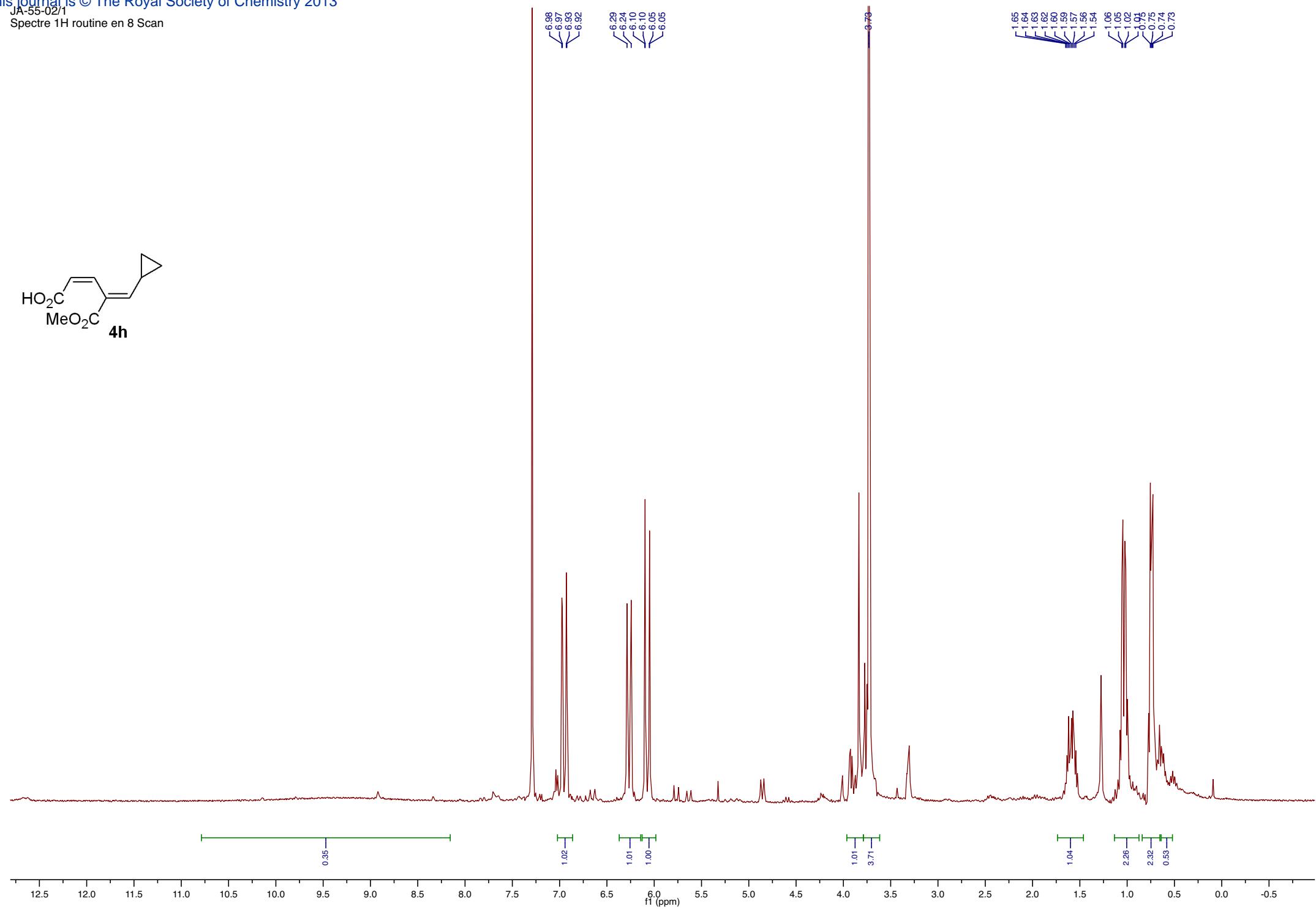
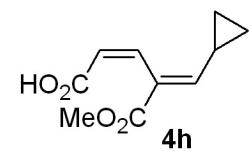


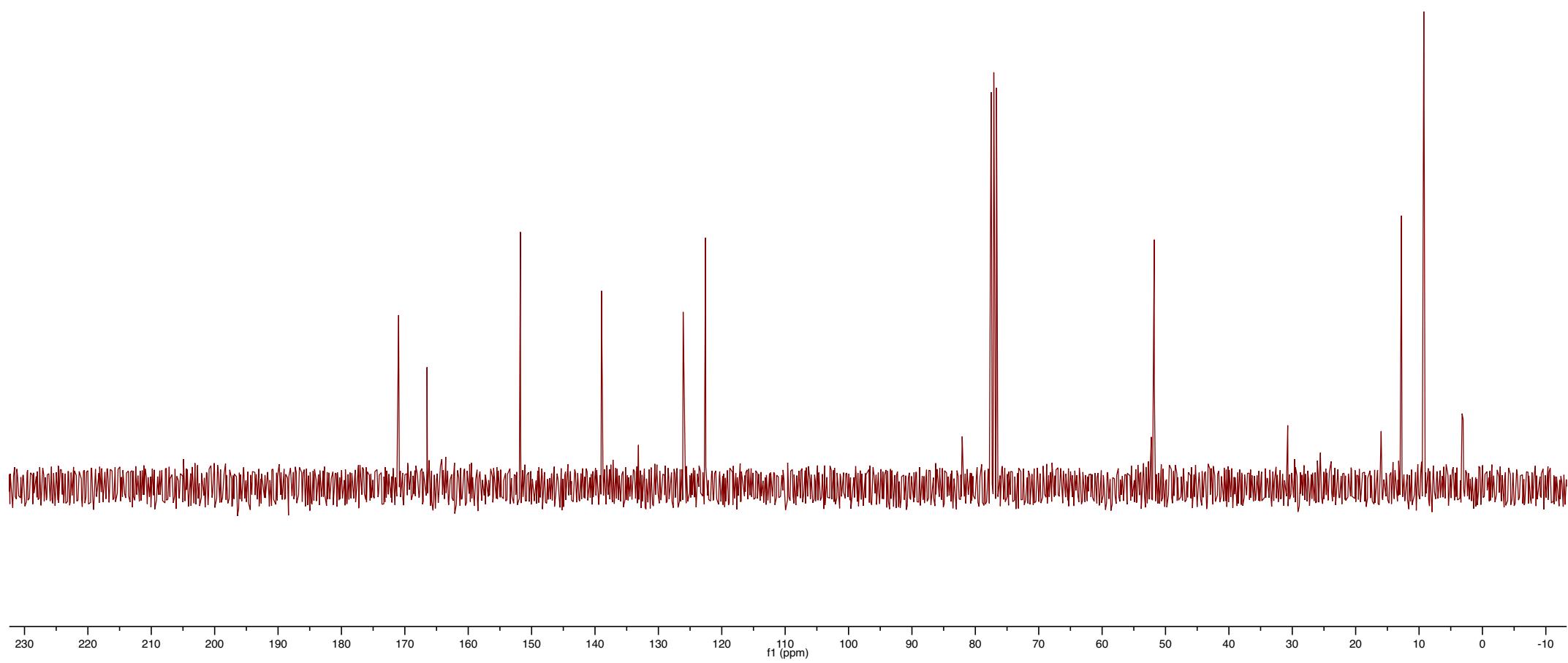
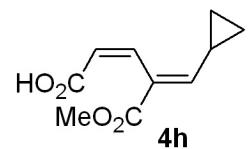


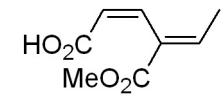




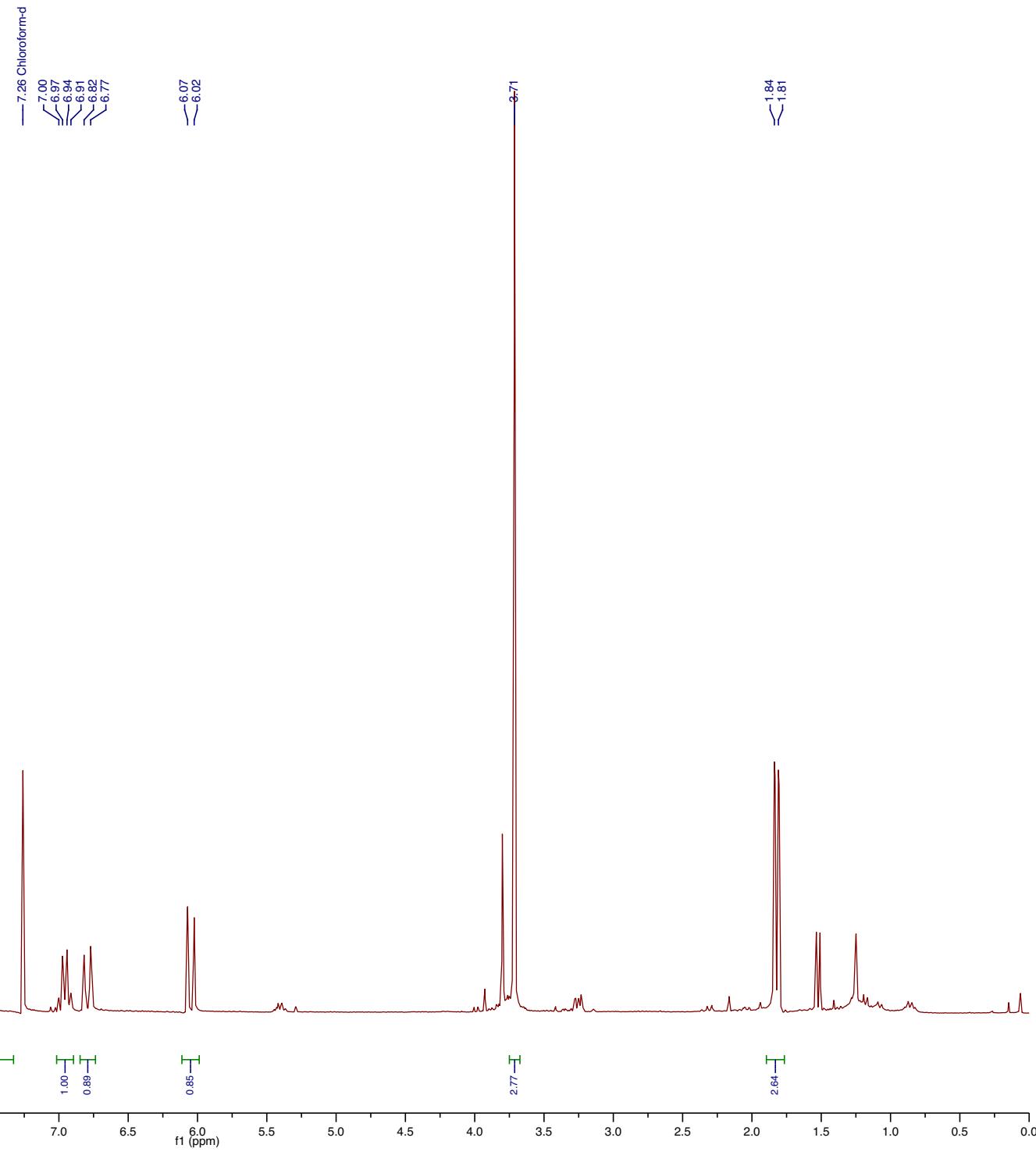


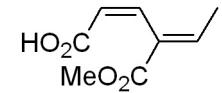




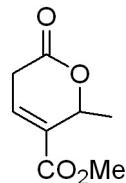


4i

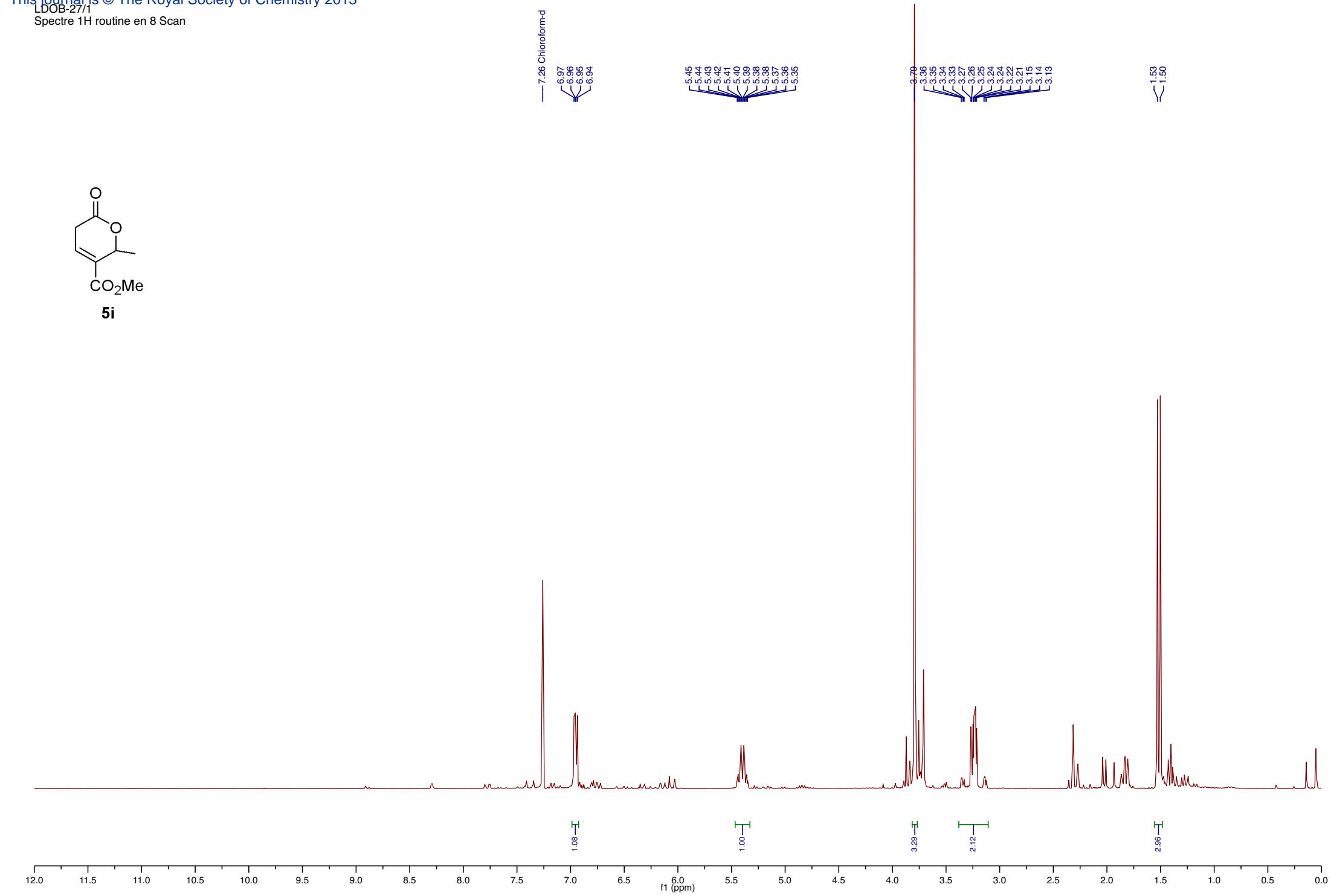


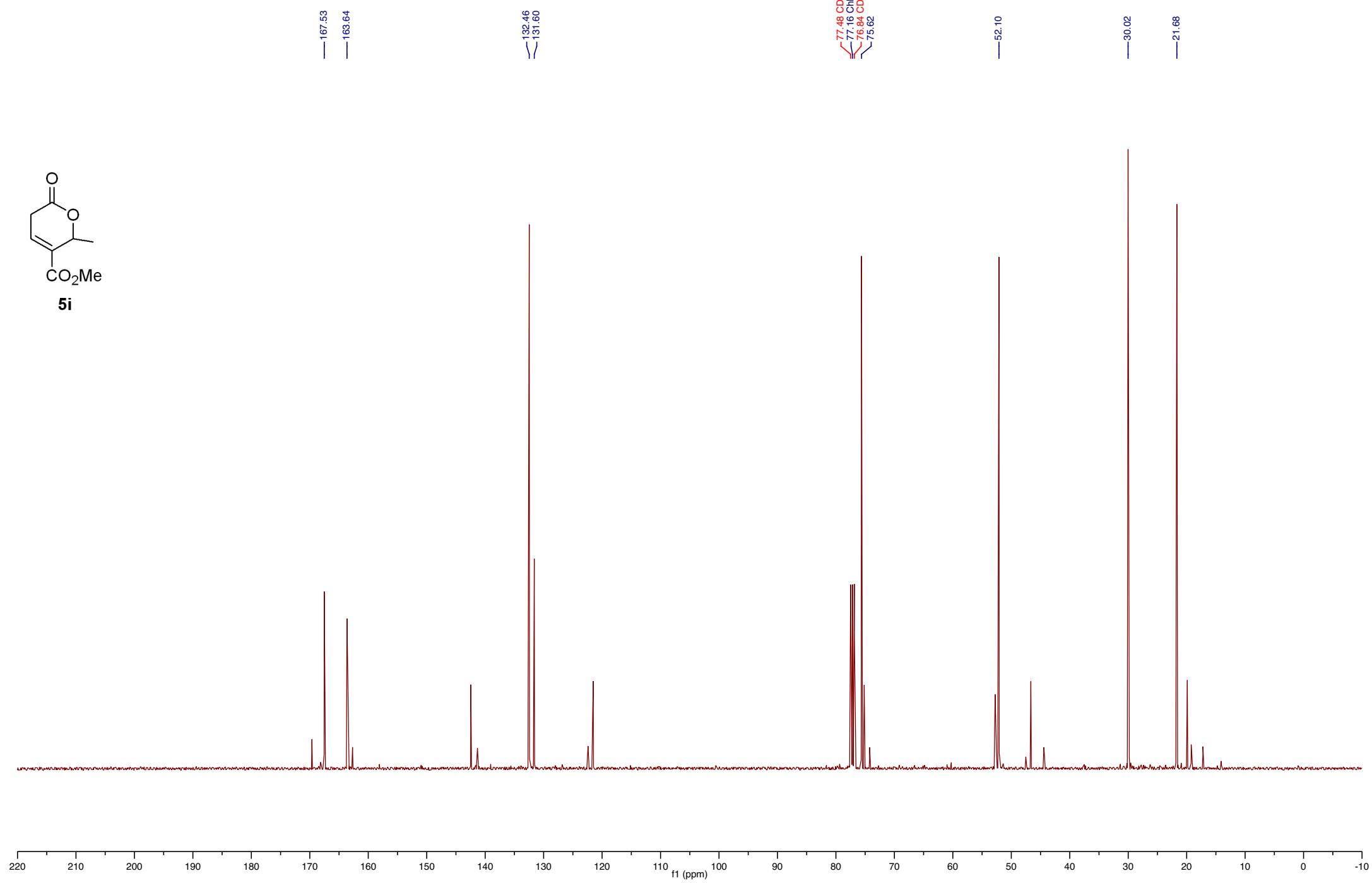
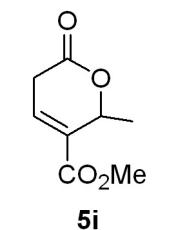


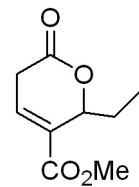
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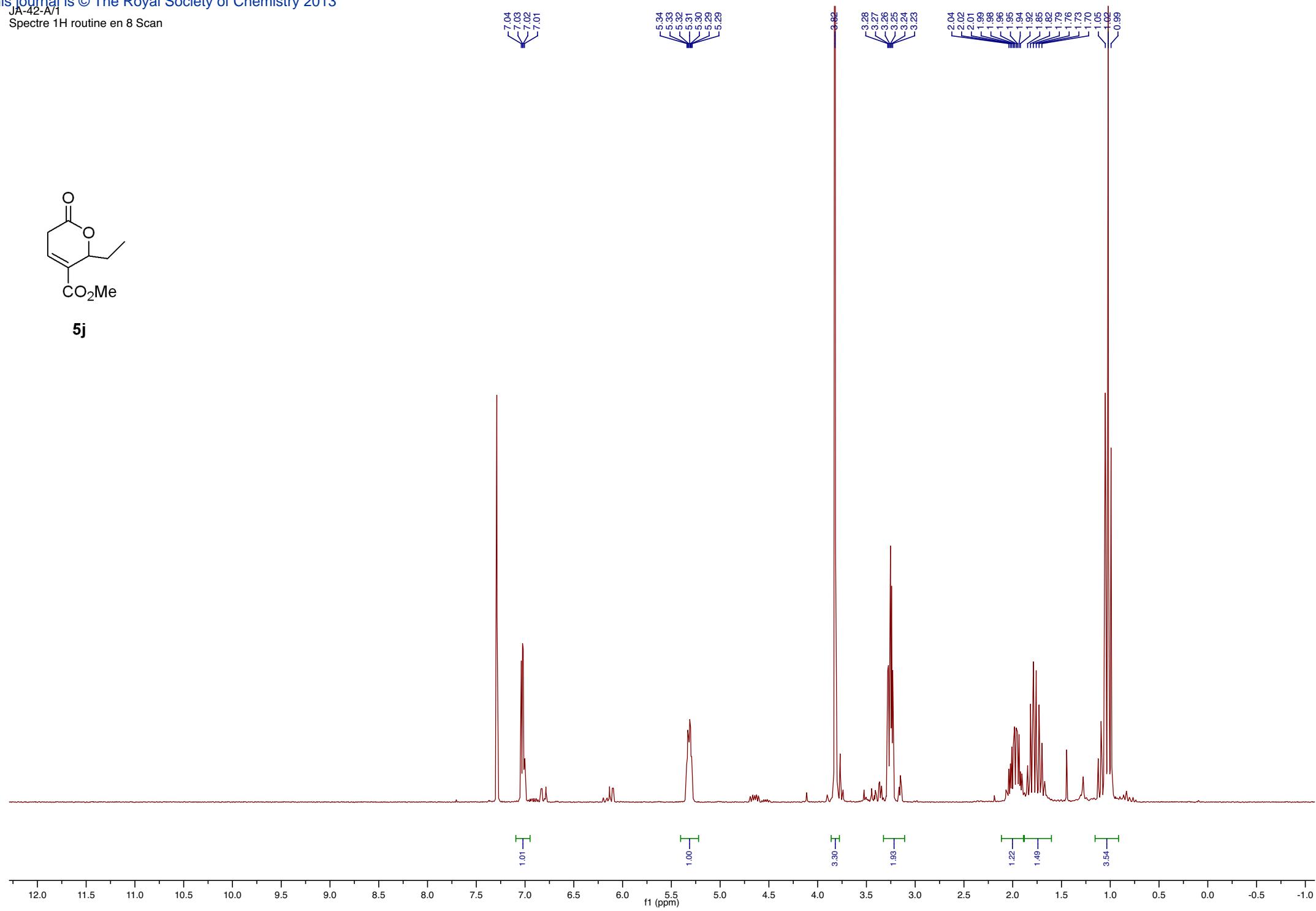
5i

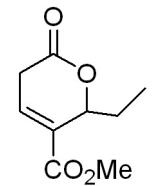




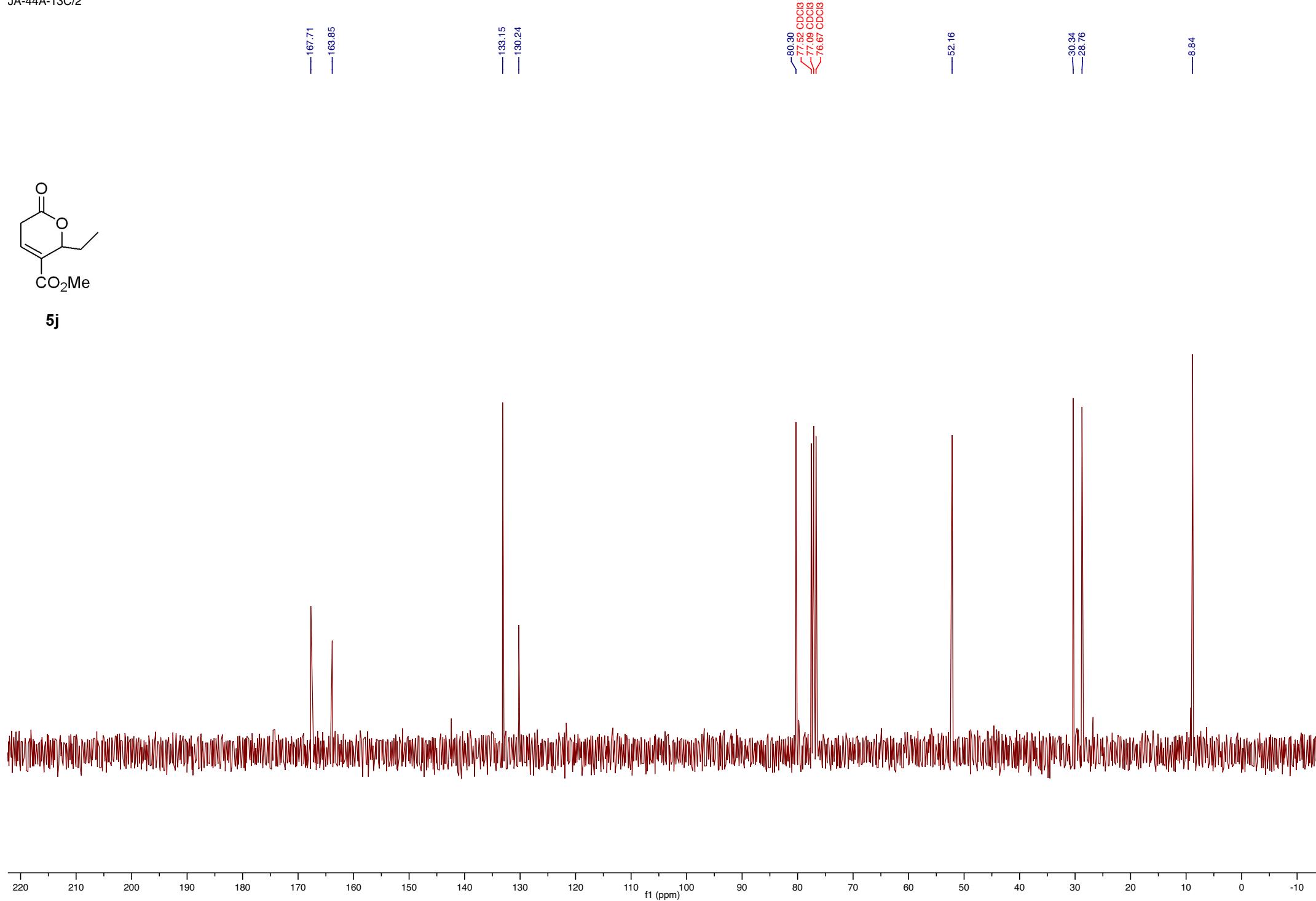


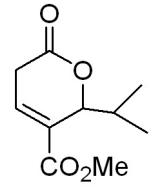
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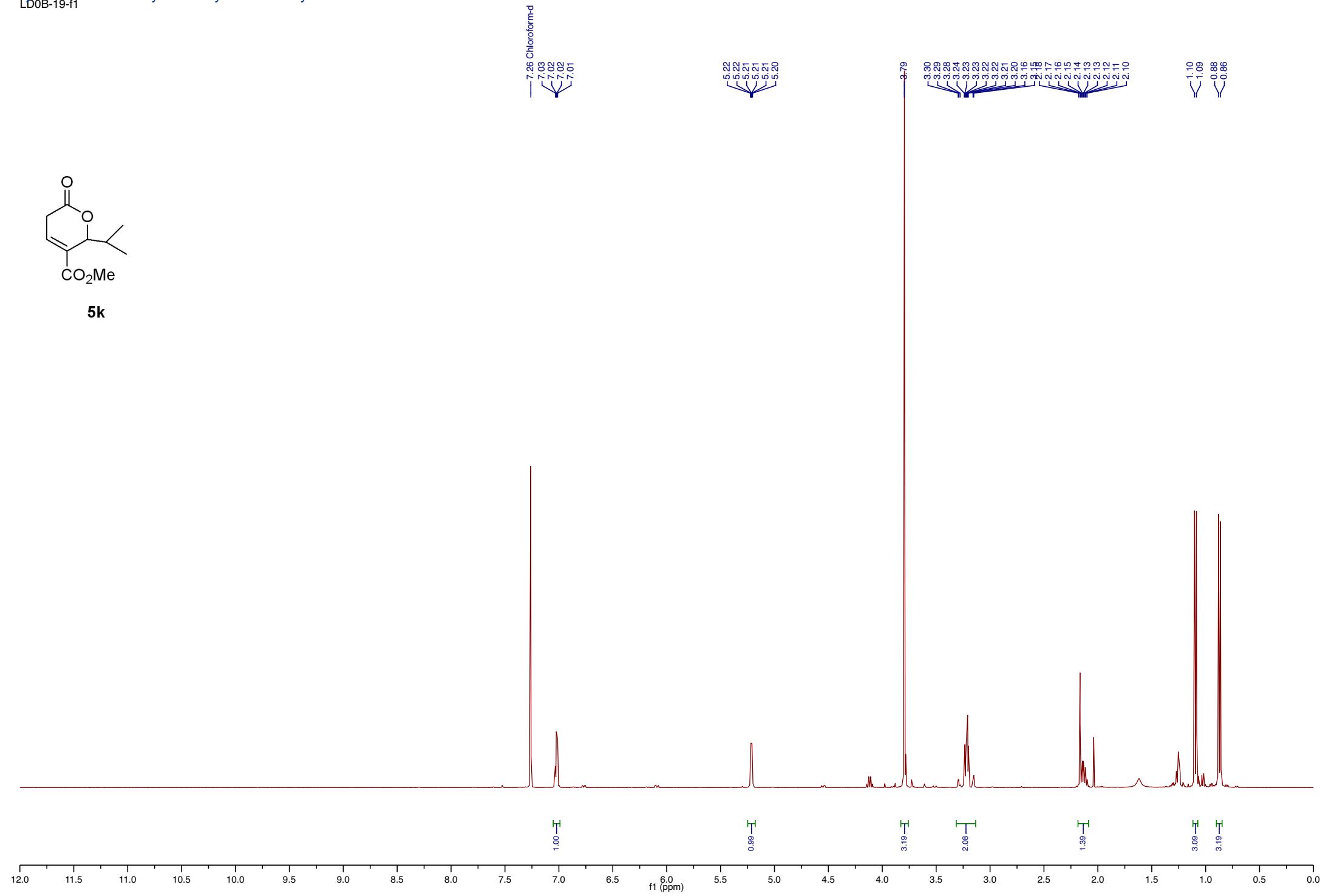


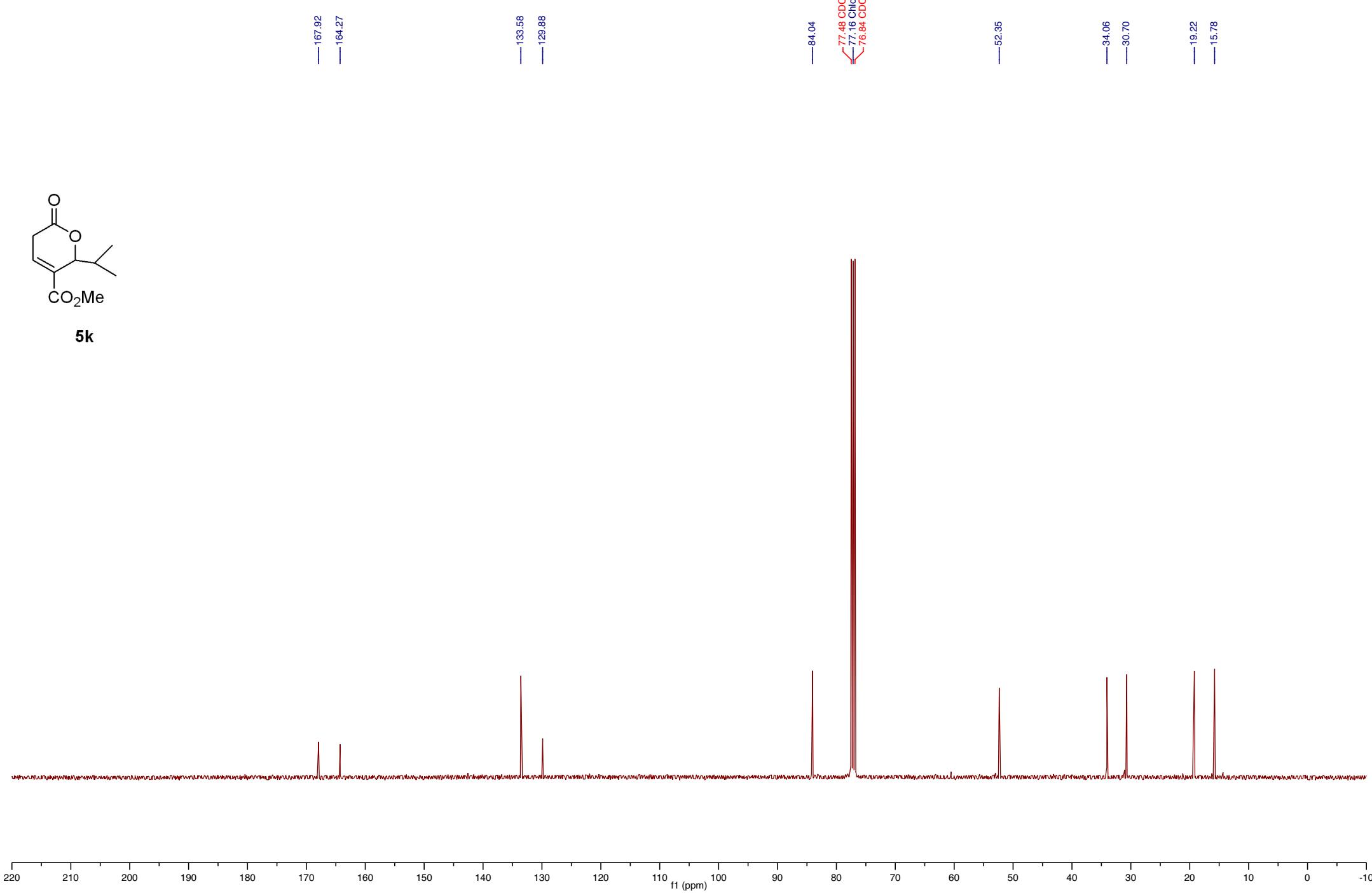
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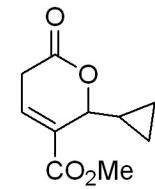




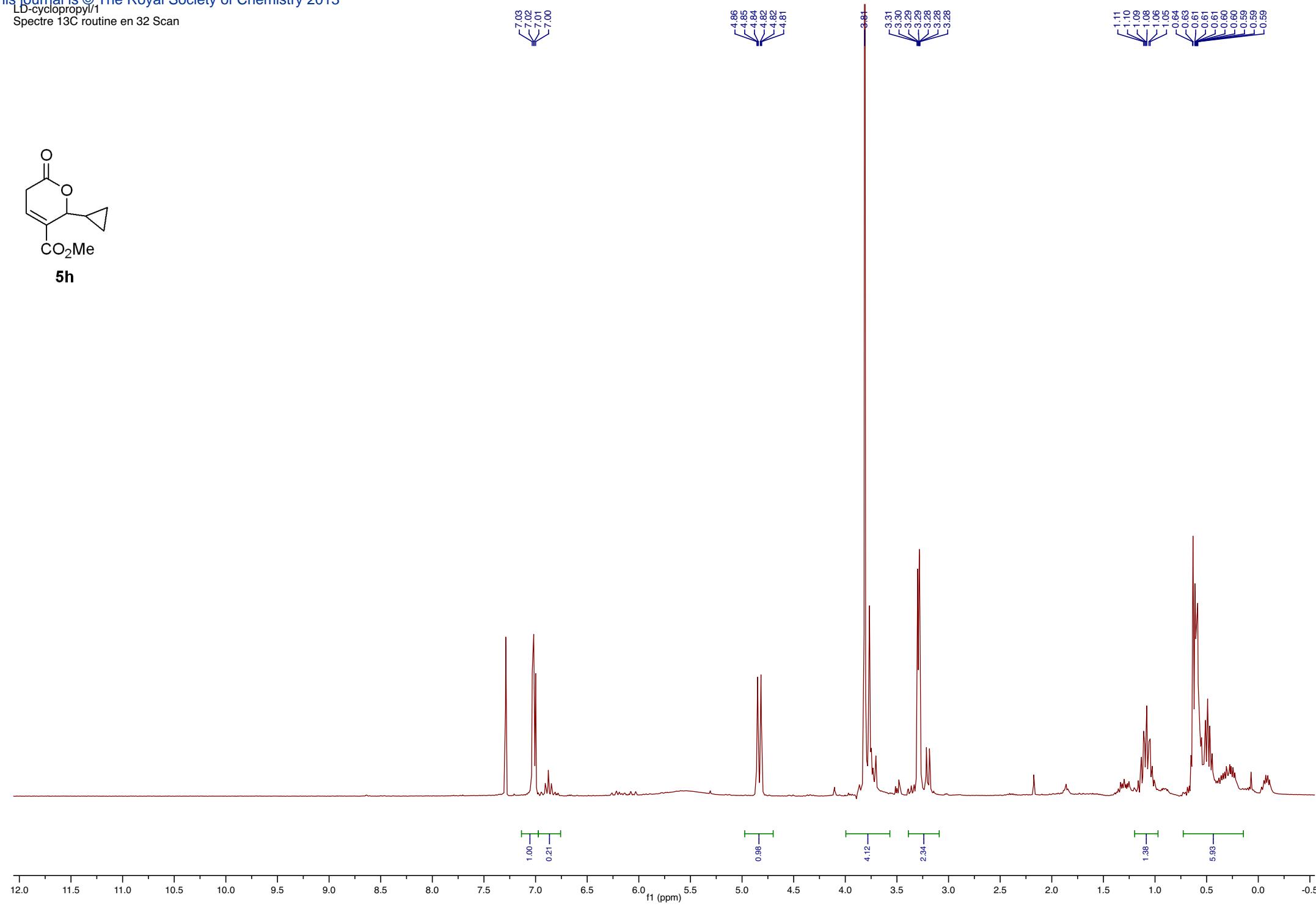
5k

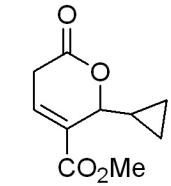






5h





5h

