

1 **A Novel C18 Reversed Phase Organic-Silica Hybrid Cationic Monolithic**  
2 **Capillary Column with Ionic Liquid as Organic Monomer via “One-Pot”**  
3 **Approach for Capillary Electrochromatography**

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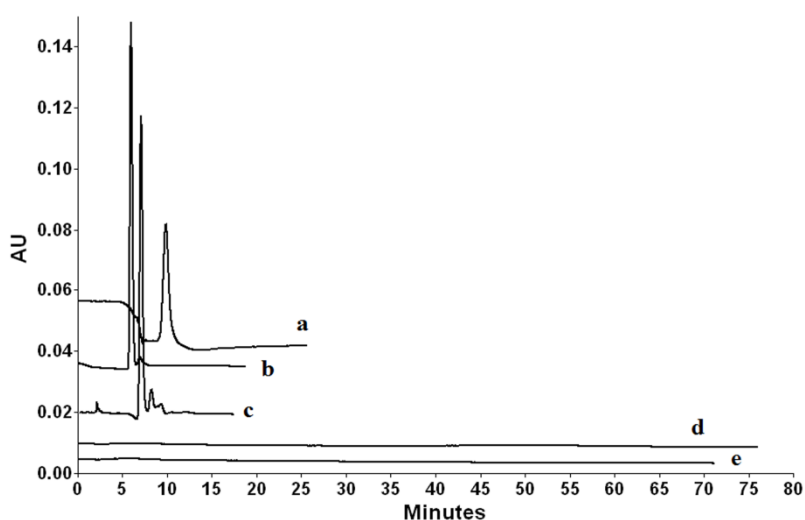
#### 14 Preparation of control columns

15 Preparation of control column 1 was similar to that of the VC<sub>18</sub>HIm<sup>+</sup>Br<sup>-</sup> hybrid  
16 monolithic capillary column except for removing the IL from the prepolymerization  
17 mixture.

18 The control column 2 was prepared with prepolymerization mixture containing i-  
19 PrOH (500 μL), n-BuOH (100 μL), AIBN (5 mg), VC<sub>18</sub>HIm<sup>+</sup>Br<sup>-</sup> (150 mg), and  
20 EGDMA (100 μL) in a 60 °C water bath for 24 h.

21 Preparation of control column 3 contained two steps. Firstly, a vinyl-functionalized  
22 monolith was obtained with prepolymerization mixture containing MeOH (600 μL),  
23 TEOS (300 μL), VTES (200 μL), CTAB (5 mg), water (110 μL), and 1.0 M ammonia  
24 water (50 μL) in a 40 °C water bath for 12 h. Then, a solution containing 250 mg/mL  
25 and 8 mg/mL AIBN was pumped through the column for 10 min. After that, the  
26 column was incubated in 60 °C water bath for another 12 h.

#### 27 The EOF of different columns

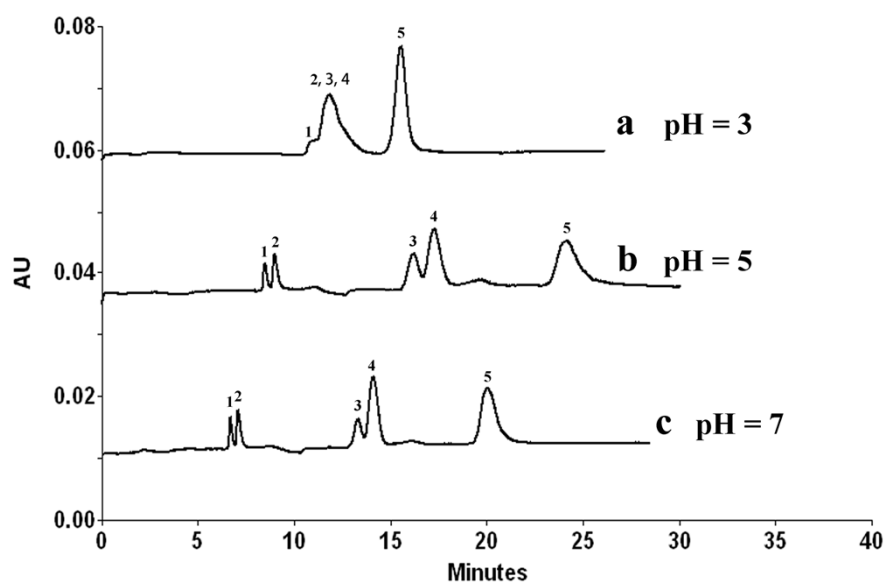


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29 **Supplementary Fig. 1.** The EOF of different columns; (a) control column 3 with -10  
30 kV voltage; (b) control column 2 with -10 kV voltage; (c) VC<sub>18</sub>HIm<sup>+</sup>Br<sup>-</sup> hybrid  
31 monolithic capillary column with -10 kV voltage; (d) control column 1 with -10 kV

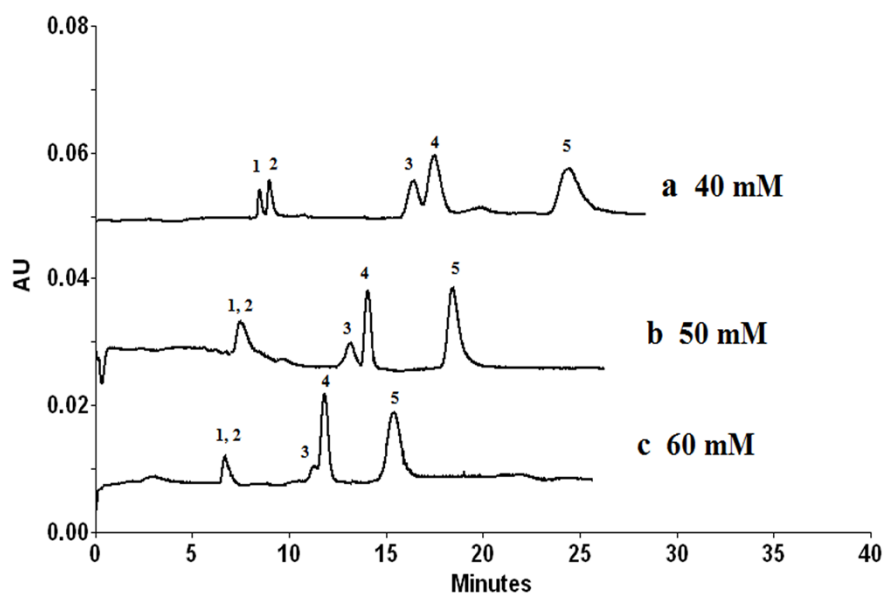
32 voltage; (e) control column 1 with 10 kV voltage. Experimental conditions: column  
33 dimension, 20 cm × 100 μm i.d.; injection, −0.5 psi for 5 s; detection wavelength, 214  
34 nm; mobile phase, ACN/30 mM acetic acid buffer at pH 3.0 = 40/60 (v/v).

### 35 Separation of amino acids in different condition



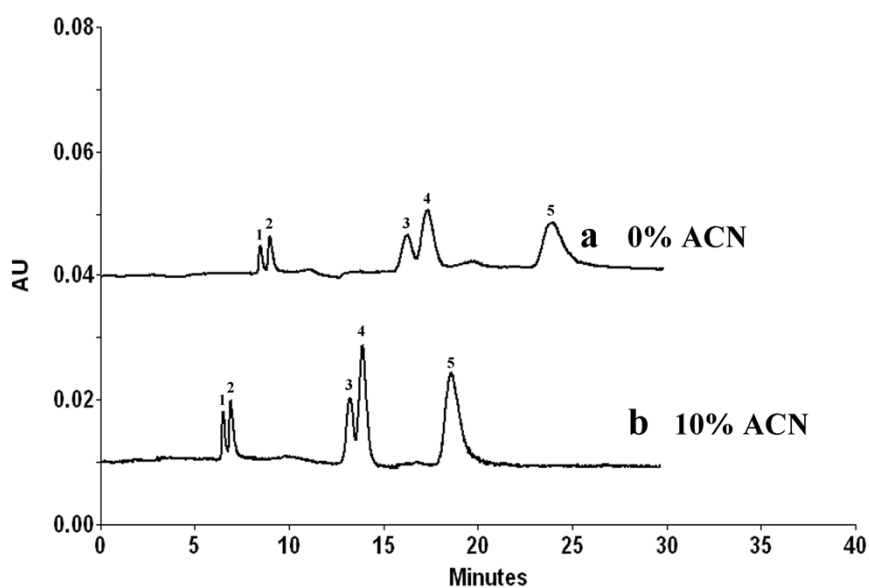
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37 **Supplementary Fig. 2.** Effect of pH on the separation of amino acids on the  
38 VC<sub>18</sub>HIm<sup>+</sup>Br<sup>-</sup> hybrid monolithic capillary column by CEC. Solutes, (1) aspartic acid,  
39 (2) glutamic acid, (3) glutamine, (4) L-proline, (5) L-phenylalanine. Experimental  
40 conditions: column dimension, 20 cm × 100 μm i.d.; injection, −1.0 psi for 15 s;  
41 separation voltage, −5 kV; detection wavelength, 190 nm; mobile phase, 40 mM  
42 H<sub>3</sub>PO<sub>4</sub>-Na<sub>2</sub>HPO<sub>4</sub> buffer at different pH.



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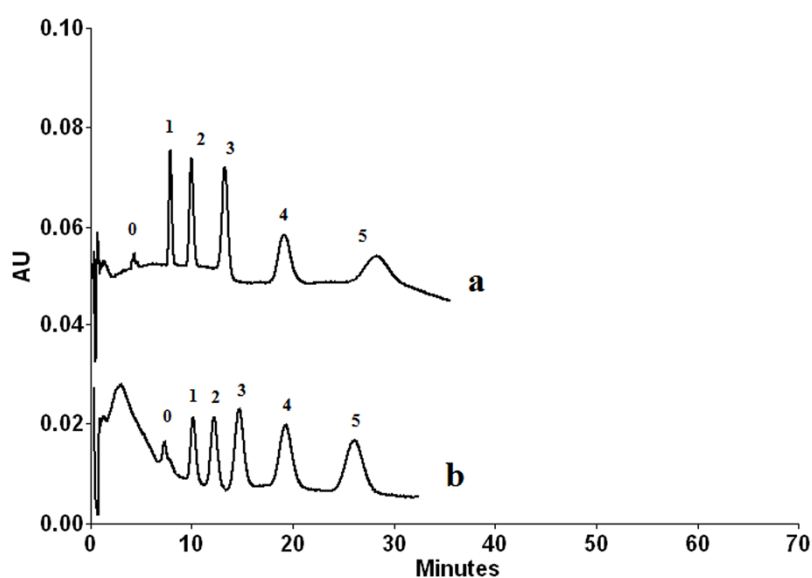
44 **Supplementary Fig. 3.** Effect of salt concentration on the separation of amino acids  
 45 on the VC<sub>18</sub>HIm<sup>+</sup>Br<sup>-</sup> hybrid monolithic capillary column by CEC. Solutes, (1) aspartic  
 46 acid, (2) glutamic acid, (3) glutamine, (4) L-proline, (5) L-phenylalanine.  
 47 Experimental conditions: column dimension, 20 cm × 100 μm i.d.; injection, -1.0 psi  
 48 for 15 s; separation voltage, -5 kV; detection wavelength, 190 nm; mobile phase,  
 49 different salt concentration buffer at pH 5.



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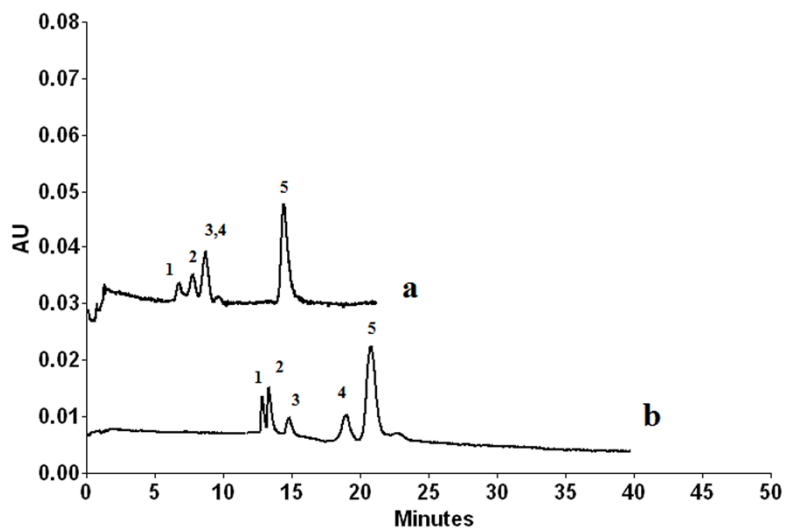
51 **Supplementary Fig. 4.** Effect of ACN content on the separation of amino acids on  
52 the VC18HIm+Br<sup>-</sup> hybrid monolithic capillary column by CEC. Solutes, (1) aspartic  
53 acid, (2) glutamic acid, (3) glutamine, (4) L-proline, (5) L-phenylalanine.  
54 Experimental conditions: column dimension, 20 cm × 100 μm i.d.; injection, -1.0 psi  
55 for 15 s; separation voltage, -5 kV; detection wavelength, 190 nm; mobile phase, 40  
56 mM H<sub>3</sub>PO<sub>4</sub>-Na<sub>2</sub>HPO<sub>4</sub> buffer at pH 5.

57 **Separation of control column**



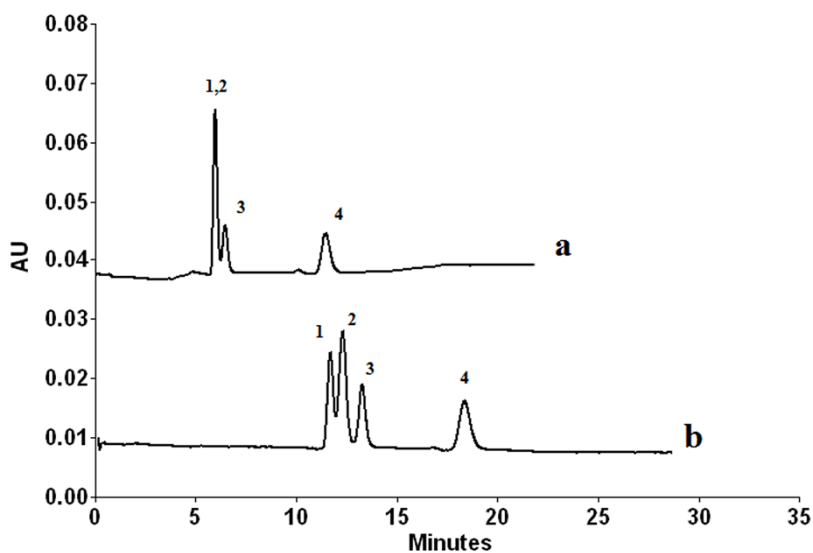
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59 **Supplementary Fig. 5.** Separation of alkylbenzenes on the control columns by CEC;  
60 a, control column 2; b, control column 3. Solutes: (0) thiourea, (1) benzene, (2)  
61 toluene, (3) ethylbenzene, (4) propylbenzene, (5) butylbenzene. Experimental  
62 conditions: mobile phase, ACN/30 mM acetic acid buffer at pH 3.0 = 45/55 (v/v);  
63 column dimension, 20 cm × 100 μm i.d.; injection, -0.5 psi for 5 s; separation voltage,  
64 -10 kV; detection wavelength, 214 nm.



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66 **Supplementary Fig. 6.** Separation of amino acids on the control columns by CEC; a,  
 67 control column 2; b, control column 3. Solutes: (1) aspartic acid, (2) glutamic acid, (3)  
 68 glutamine, (4) L-proline, (5) L-phenylalanine. Experimental conditions: column  
 69 dimension, 20 cm × 100 μm i.d.; injection, -1.0 psi for 15 s; separation voltage, -5  
 70 kV; detection wavelength, 190 nm; mobile phase, 40 mM H<sub>3</sub>PO<sub>4</sub>-Na<sub>2</sub>HPO<sub>4</sub> buffer at  
 71 pH 4.4.



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73 **Supplementary Fig. 7.** Separation of basic compounds on the control columns by  
 74 CEC; a, control column 2; b, control column 3. Solutes: (1) methimazole, (2) aniline,  
 75 (3) gramine, (4) 1,2-diphenyl hydrazine. Experimental conditions: column dimension,

76 20 cm × 100 μm i.d.; injection, -0.5 psi for 5 s; separation voltage, -10 kV; detection  
77 wavelength, 214 nm; mobile phase, ACN/30 mM H<sub>3</sub>PO<sub>4</sub>-Na<sub>2</sub>HPO<sub>4</sub> buffer at pH 5.0 =  
78 40/60 (v/v).