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Electronic Supplementary Information

Attapulgite-doped electrospun poly(lactic-co-glycolic acid) nanofibers enable enhanced osteogenic differentiation of human mesenchymal stem cells

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Samples	Bulk density (g	Apparent density	Porosity (%)	Water contact angle (°)
	cm ⁻³)	$(g \text{ cm}^{-3})$		
PLGA	1.250	0.350±0.013	72.00±1.04	123.7±2.6
PLGA-1%ATT	1.255	0.403 ± 0.028	67.75±2.24	120.2±3.7
PLGA-2%ATT	1.261	0.412 ± 0.047	67.04±3.76	116.8±1.5
PLGA-3%ATT	1.266	0.422 ± 0.029	66.24±2.32	113.5±2.1

Table S1. Bulk density, apparent density, porosity, and water contact angle of PLGA and ATT-doped PLGA nanofibers.

Table S2. Hemolysis percentage (HP) of PLGA and ATT-doped PLGA nanofibers (all data are given as mean \pm SD, n=3).

Samples	PLGA	PLGA-1%ATT	PLGA-2%ATT	PLGA-3%ATT
HP (%)	1.2 ± 0.1	0.64 ± 0.13	0.65 ± 0.09	0.56 ± 0.21



Figure S1. TGA curves of ATT nanorods, and PLGA and ATT-doped PLGA nanofibers with different ATT doping levels.



Figure S2. Water contact angles of PLGA (a), PLGA-1%ATT (b), PLGA-2%ATT (c), and PLGA-3%ATT (d) fibrous mats.



Figure S3. Strain-stress curves of PLGA (a), PLGA-1%ATT (b), PLGA-2%ATT (c), and PLGA-3%ATT (d) nanofibrous mats.



Figure S4. Metabolic activity assay of hMSCs cultured onto TCP, cover slip, PLGA, PLGA-1%ATT, PLGA-2%ATT, and PLGA-3%ATT nanofibers at different culture times.



Figure S5. Anticoagulant assay of PLGA and ATT-doped PLGA nanofibers at different time intervals.



Figure S6. The picture of von Kossa staining of hMSCs cultured onto PLGA and PLGA-3%ATT

nanofibers in growth medium (without DEX) and osteogenic medium (with DEX) on day 21.