

Electronic Supplementary Information for

**Hydrolysis of Cellobiose to Monosaccharide Catalyzed by Functional  
Lanthanum (III) Metallomicelle**

Xiao Peng, Xiang-Guang Meng\*, Chun Mi, Xiao-Hong Liao

*Key Laboratory of Green Chemistry and Technology, Ministry of Education, College of Chemistry,  
Sichuan University, Chengdu 610064, P. R. China. Tel: +86-28-85462979; Fax: +86-28-  
85412291. Email: [mengxgchem@163.com](mailto:mengxgchem@163.com).*

**Contents**

1. cmc of surfactant
2. Job plots for the ligand (DMBO) and  $\text{La}^{3+}$  ion
3. Effect of pH on conversion of cellobiose in the absence of catalyst
4. Comparison with other metal complexes
5. PAD-MS(E+) spectrum of reaction solution
6. Reaction kinetics

**1. cmc of surfactant**

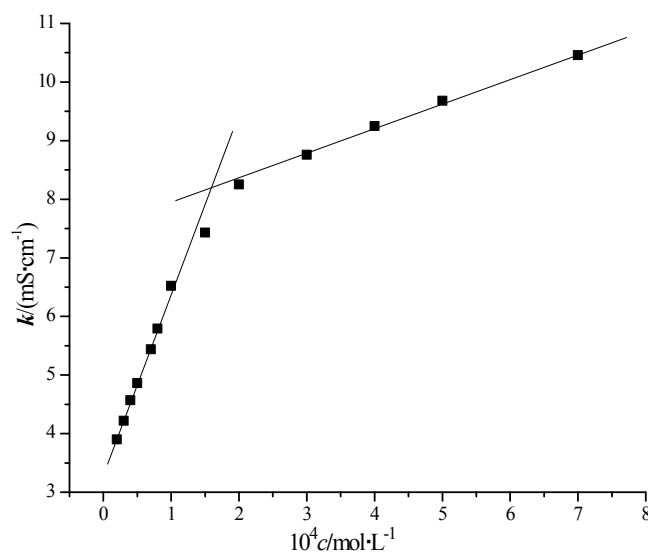


Fig.S1 Variation of electric conductivity with concentration of surfactant DMBO, 25°C

## 2 Determination of chelating ratio of $\text{La}^{3+}$ to ligand

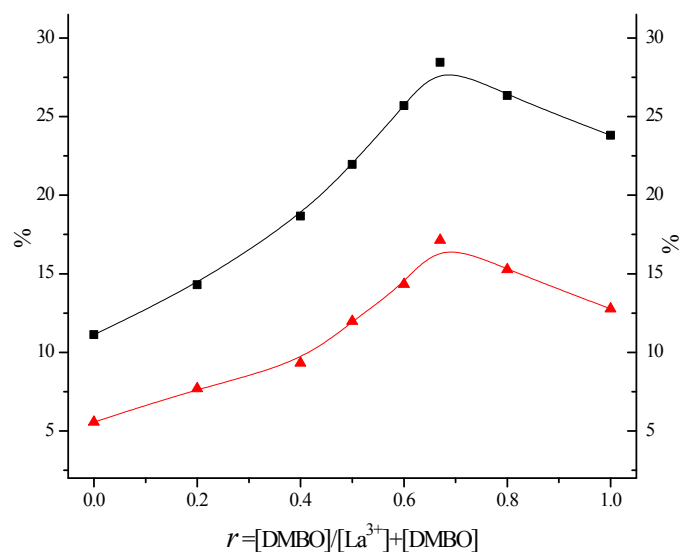


Fig.S2 Job plots for the ligand (DMBO) and  $\text{La}^{3+}$  ion complexation as measured by conversion of cellobiose (■) and yield of monosaccharide (▲)  
[Cellobiose]<sub>0</sub> = 0.02 mol·L<sup>-1</sup>, [DMBO] + [La<sup>3+</sup>] = 0.002 mol·L<sup>-1</sup>, pH 9.0, 90°C, 10h

## 3. Effect of pH

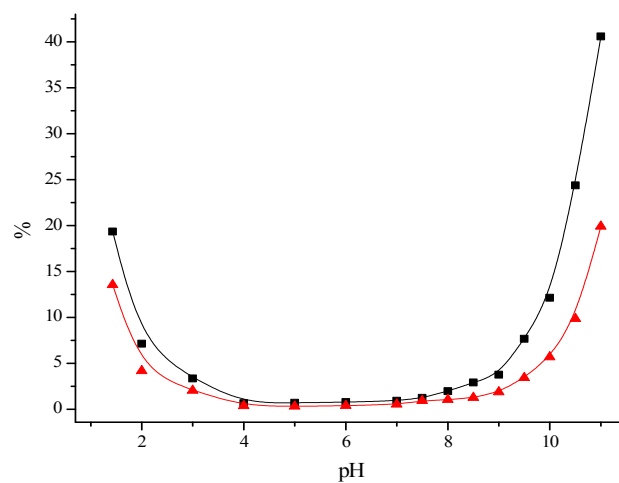


Fig.S3 Plots of conversion of cellobiose (■) and yield of monosaccharide (▲) with pH  
[Cellobiose]<sub>0</sub> = 0.02 mol·L<sup>-1</sup>, 90°C, 10h

Fig.S3 showed the effect of pH on conversion of cellobiose and yield of monosaccharide in the absence of catalyst in aqueous solution.

#### 4. Comparison with other metal complexes

To understand the effect of structure of metal complex on reaction activity, several metal complexes were employed as catalysts to catalyze cellobiose hydrolysis. The structures of these metal complexes were shown in Fig.S4 and the results were listed in table S1.

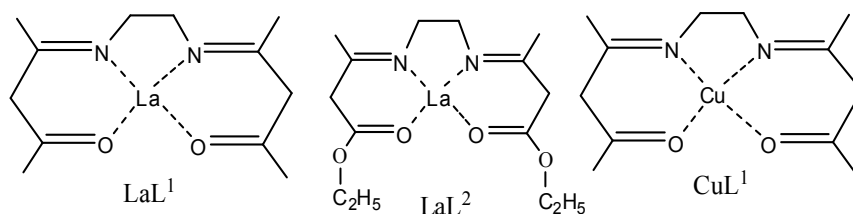


Fig.S4 Structures of metal complexes

Table S1 Conversion of cellobiose, yield of monosaccharide and selectivity of monosaccharide catalyzed by different metal complexes\*

Systems	Conversion of cellobiose /%	Yield of monosaccharide/%	Selectivity of monosaccharide/%
LaL <sup>1</sup>	9.8	4.8	49.0
LaL <sup>2</sup>	10.4	5.5	52.9
CuL <sup>1</sup>	8.5	4.4	51.8

\*[Cellobiose]=0.02mol·L<sup>-1</sup>, [Catalyst]=0.002mol·L<sup>-1</sup>, pH9.0, 10h

#### 5. PAD-MS (E+) spectrum of reaction solution

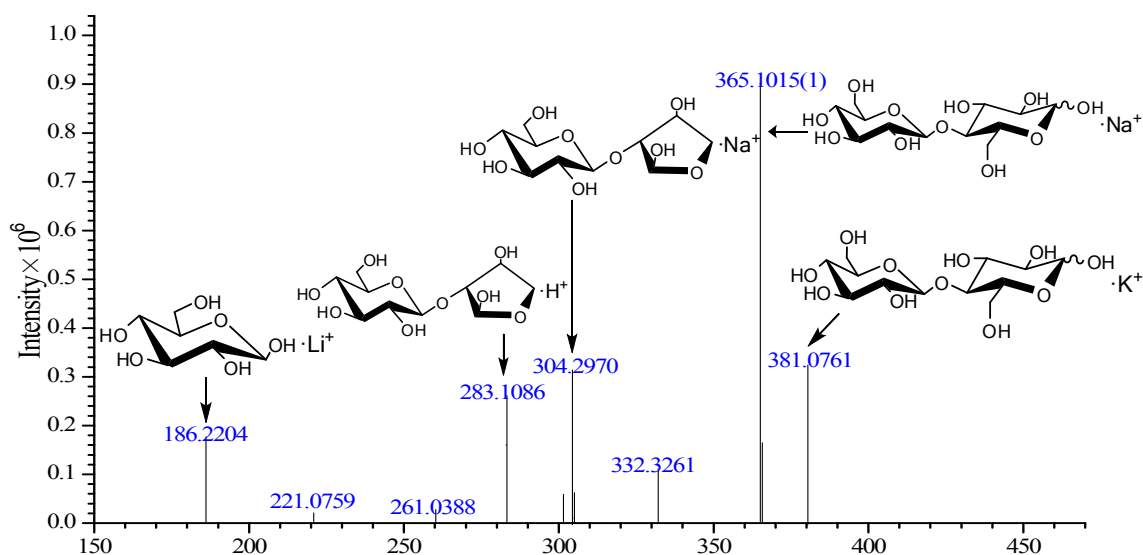


Fig.S5 PAD-MS (E+) spectrum of reaction solution of cellobiose hydrolysis catalyzed by metallomicelle La(DMBO)<sub>2</sub>

[Cellobiose]<sub>0</sub>=0.02 mol·L<sup>-1</sup>, [DMBO]=0.002 mol·L<sup>-1</sup> pH9.0, 10h

## 6 Reaction kinetics

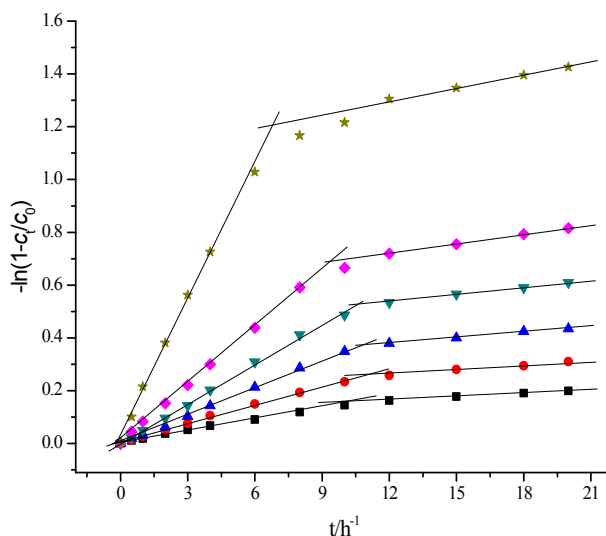
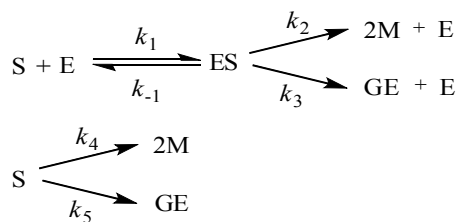


Fig.S6 Plots of  $-\ln(1-c_t/c_0)$  vs reaction time  $t$  at different temperature  
 $[\text{Cellobiose}]_0=0.02\text{mol}\cdot\text{L}^{-1}$ ,  $[\text{La}(\text{DMBO})_2]=0.002\text{mol}\cdot\text{L}^{-1}$ , pH9.0  
 (■)80 °C, (●)85°C, (▲)90°C, (▼)95°C, (◆)100°C, (★)110°C

According to our experimental results a more detailed reaction pathway for cellobiose hydrolysis reaction was proposed as shown in scheme S1: firstly cellobiose S combined with surfactant E to form an intermediate compound ES, and then ES further reacted to monosaccharide M and glucosyl-erythrose GE. In the first step, the surfactant E combined with substrate S reversibly with rate constants  $k_1$  and  $k_{-1}$ , then followed the rate-determining steps with rate constants  $k_2$  and  $k_3$  ( $k_{\text{cat}}=k_2+k_3$ ).  $k_4$  and  $k_5$  are the rate constant of cellobiose hydrolysis to M and GE in bulk solution, respectively ( $k_0=k_4+k_5$ ).



Scheme S1 □ The reaction pathway proposed for catalysis reaction

The cellobiose hydrolysis reaction rate equation is described as

$$r = \frac{d[\text{S}]}{dt} = k_{-1}[\text{ES}] - k_1[\text{S}][\text{E}] - k_0[\text{S}] \quad \text{S(1)}$$

And the production of monosaccharide rate equation could be described as

$$\frac{d[\text{M}]}{dt} = k_2[\text{ES}] + k_4[\text{S}] \quad \text{S(2)}$$

According to Steady State Approximation,

$$\frac{d[\text{ES}]}{dt} = k_1[\text{S}][\text{E}] - (k_2+k_3+k_{-1})[\text{ES}] = 0 \quad \text{S(3)}$$

Thus 
$$[ES] = \frac{k_1[E][S]}{k_2+k_3+k_{-1}} = \frac{[S][E]}{K_m} \quad S(4)$$

Where  $K_m = \frac{k_2+k_3+k_{-1}}{k_1}$ , is the Michaelis constant. Due to total concentration  $E_T$  of surfactant keeps constant,  $E_T=[E]+[ES]$ ,

Hence 
$$[E] = \frac{K_m * E_T}{K_m + [S]} \quad S(5)$$

Thus the equation S(1) and S(2) are transformed as equation S(6) and S(7).

$$r = \frac{d[S]}{dt} = -\frac{k_{cat}E_T[S]}{K_m + [S]} - k_0[S] \quad S(6)$$

$$\frac{d[M]}{dt} = k_2[ES] + k_4[S] = \left(\frac{k_2}{K_m}[E] + k_4\right)[S] = \left(\frac{k_2E_T}{K_m + [S]} + k_4\right)[S] \quad S(7)$$

And after integrating equation S(6) and rearrangement, the following equation S(8) is obtained.

$$K_mk_0 \ln \frac{c_t}{c_0} + k_{cat}E_T \ln \frac{k_{cat}E_T + K_mk_0 + k_0c_t}{k_{cat}E_T + K_mk_0 + k_0c_0} = -(k_{cat}E_T + K_mk_0) * k_0t \quad S(8)$$

Thus equation S(9) is obtained through transferring equation S(8).

$$t = \frac{K_mk_0}{(k_{cat}E_T + K_mk_0)k_0} \ln \frac{c_0}{c_t} + \frac{k_{cat}E_T}{(k_{cat}E_T + K_mk_0)k_0} \ln \frac{k_{cat}E_T + K_mk_0 + k_0c_0}{k_{cat}E_T + K_mk_0 + k_0c_t} \quad S(9)$$

Based on equation S(9),  $K_m$  and  $k_{cat}$  can be obtained by nonlinear fitting, as illustrated in Fig. S(7). All of the nonlinear correlation coefficients are above 0.97, this indicated that the kinetic model (Scheme S1 and equation S(9)) was reasonable.

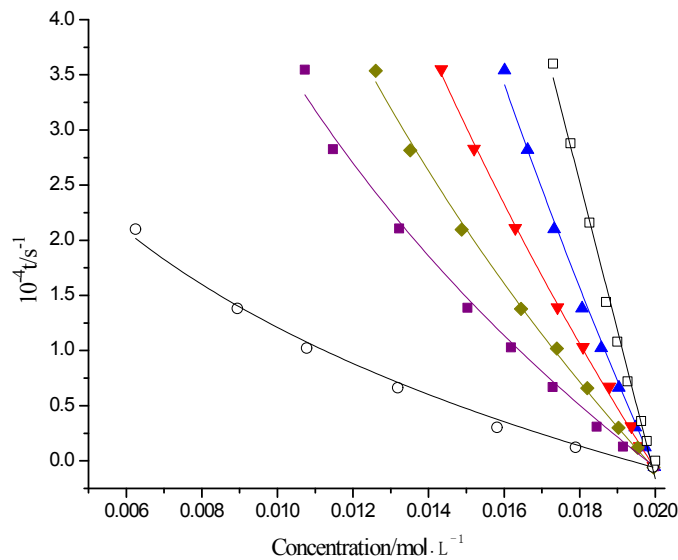


Fig.S7 Plots of reaction time vs. concentration of cellobiose  
 $[Cellobiose]_0=0.02\text{mol}\cdot\text{L}^{-1}$ ,  $[La(\text{DMBO})_2]=0.002\text{mol}\cdot\text{L}^{-1}$ , pH9.0  
 (□)80 °C, (▲)85 °C, (▼)90 °C, (◆) 95 °C, (■)100 °C, (○)110 °C

Base on the equation S(10):

$$\frac{d[M]}{d[S]} = \frac{d[M]/dt}{d[S]/dt} = \frac{\frac{k_2 E_T}{K_m + [S]} + k_4}{-\frac{k_{cat} E_T}{K_m + [S]} - k_0} = -\frac{k_2 E_T + k_4(K_m + [S])}{k_{cat} E_T + k_0(K_m + [S])} \quad S(10)$$

Hence the equation S(11) is obtained,

$$\int_0^{[M]} [M] = [M] = -\int_{[S]^0}^{[S]} \frac{k_2 E_T + k_4(K_m + [S])}{k_{cat} E_T + k_0(K_m + [S])} d[S] \quad S(11)$$

And from equation S(11) the following equation is obtained.

$$[M] = \frac{k_2 E_T + k_4 K_m}{k_0} \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} + \frac{k_4}{k_0} (c_0 - [S]) - \frac{k_4 (k_{cat} E_T + K_m k_0)}{k_0 * k_0} \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} \quad \dots S(12)$$

Therefore equation S(13) is obtained through transferring equation S(12).

$$k_0 [M] - k_4 (c_0 - [S]) + \frac{k_4 (k_{cat} E_T + K_m k_0)}{k_0} \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} = (k_2 E_T + k_4 K_m) \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} \quad \dots S(13)$$

Let 
$$Y = k_0 [M] - k_4 (c_0 - [S]) + \frac{k_4 (k_{cat} E_T + K_m k_0)}{k_0} \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} \quad S(14)$$

$$X = \ln \frac{k_{cat} E_T + K_m k_0 + k_0 c_0}{k_{cat} E_T + K_m k_0 + k_0 [S]} \quad S(15)$$

Thus

$$Y = (k_2 E_T + k_4 K_m) X \quad S(16)$$

Therefore according to the equation S(16),  $k_2$  can be calculated from the slope of straight line Y vs. X (see Fig.S8). All of the linear correlation coefficients are above 0.95.

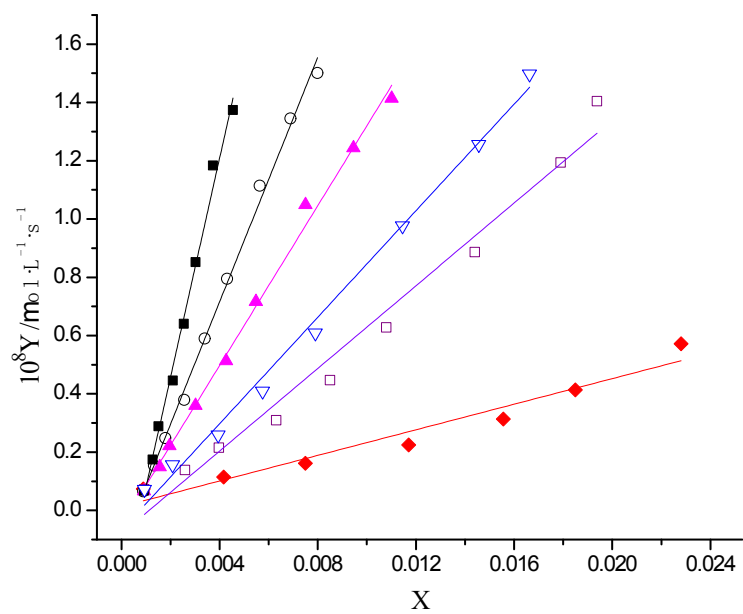


Fig.S8 Plots of Y vs. X

[Cellobiose]<sub>0</sub>=0.02mol·L<sup>-1</sup>, [La(DMBO)<sub>2</sub>]=0.002mol·L<sup>-1</sup>, pH9.0  
 (■)80 °C, (○)85°C, (▲)90°C, (▽) 95°C, (□)100°C, (◆)110°C